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Optical control of adenosine A_{2A} receptor using istradefylline photosensitivity

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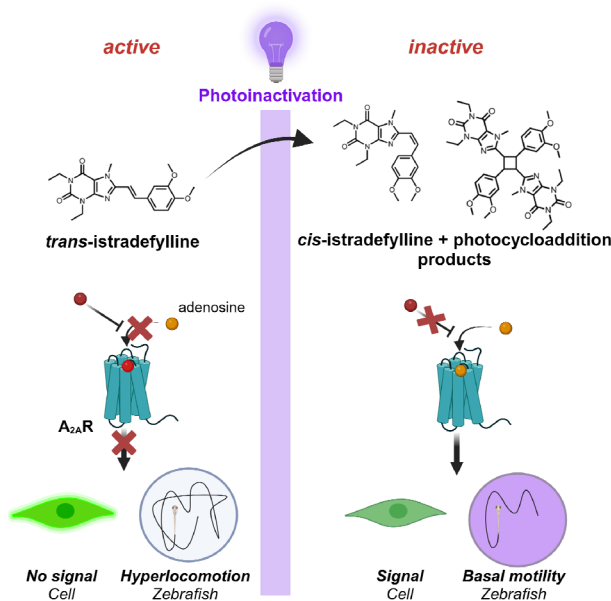
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Summary

In recent years, there has been growing interest in the potential therapeutic use of inhibitors of adenosine A_{2A} receptors (A_{2A}R) for the treatment of neurodegenerative diseases and cancer. Nevertheless, the widespread expression of A_{2A}R throughout the body emphasizes the importance of temporally and spatially selective ligands. Photopharmacology is an emerging strategy that utilises photosensitive ligands to attain high spatiotemporal precision and regulate the function of biomolecules using light. In this study, we combined photochemistry, cellular and *in vivo* photopharmacology to investigate the light sensitivity of the FDA-approved antagonist istradefylline and its potential use as an A_{2A}R photopharmacological tool. Our findings reveal that istradefylline exhibits rapid *trans* to *cis* isomerization under near UV light, and prolonged exposure results in the formation of photocycloaddition products. We demonstrate that exposure to UV light triggers a time-dependent decrease in the antagonistic activity of istradefylline in A_{2A}R-expressing cells and enables real-time optical control of A_{2A}R signaling in living cells and zebrafish. Together, these data demonstrate that istradefylline is a photoinactivatable A_{2A}R antagonist and that this property can be utilized to perform photopharmacological experiments in living cells and animals.



Keywords: photopharmacology, GPCR, photoswitch, purinergic, adenosine

Introduction

G Protein-Coupled Receptors (GPCRs) are a superfamily of more than 800 membrane proteins, which are key players in cell communication¹⁻². With more than a third of the drugs on the market directed against them, GPCRs are attractive therapeutic targets³⁻⁴.

Among GPCRs, the adenosine receptor family has attracted a lot of interest for their therapeutic potential in neurodegenerative, inflammatory and autoimmune diseases, cardiac ischemic diseases as well as chronic pain, sleep disorders and cancer⁵⁻⁶. The family is composed of four members (A₁R, A_{2A}R, A_{2B}R, A₃R), with different pharmacological, signaling and expression profiles⁵. More particularly, the A_{2A} subtype is a recognized target for the treatment of Parkinson disease (PD)⁷⁻⁸ and cardiovascular diseases⁹ and is a promising target for cancer immunotherapy¹⁰⁻¹¹, pain¹² and Alzheimer's disease¹³. So far, three drugs targeting this receptor received FDA approval. The endogenous ligand adenosine¹⁴ and the synthetic selective A_{2A}R agonist regadenoson¹⁵ are used in clinics as vasodilator agents, while the A_{2A}R antagonist istradefylline (also named KW-6002) has been approved as an adjunctive treatment for PD in combination with levodopa in Japan and USA¹⁶⁻¹⁷. Istradefylline is a styryl-xanthine analog related to caffeine. It is a selective and potent competitive A_{2A}R antagonist¹⁸. In addition, several studies reported that istradefylline could be also be therapeutically used as a potential drug for Alzheimer disease¹³, since it reduces memory deficits in aging mice with amyloid pathology¹⁹ and enhances amyloid- β generation and γ -secretase activity²⁰.

A common difficulty in pharmacology comes from the broad expression of a given target, which emphasizes the need for temporally and spatially selective drugs to reduce the risk of side effects. This is well illustrated by A_{2A}R that is not only present in the central nervous system, where it is highly expressed in the striatum and at lower expression levels in the cerebral cortex and in the hippocampus, but also in the immune system, heart and lungs⁵. One way to circumvent this difficulty is photopharmacology, an emerging light-based strategy to manipulate biological processes with high spatiotemporal precision²¹⁻²². This relies on the use of photosensitive ligands, which allows the photocontrol of drug/target interaction and its consequent biological activity. Two major categories of photosensitive ligands are available, photoactivable ligands (also named caged ligands)²³, which are irreversibly activated by light, and photochromic or photoswitchable ligands²⁴, which can be reversibly switched from an active to an inactive conformation. Combined with optical technologies, photopharmacology is a powerful fundamental research tool to explore the physiopathological function of endogenous regulatory systems, with micrometric spatial resolution and millisecond temporal resolution, opening new pharmacotherapeutic opportunities towards precision medicine²⁵⁻²⁶. Increasing interest is being shown in GPCR photopharmacology²⁷⁻²⁸, which offers the possibility to modulate and understand GPCR physiology and pathology with a higher spatiotemporal control than classical pharmacological approaches. Photopharmacology has been applied to many GPCRs including dopamine²⁹, cannabinoid³⁰⁻³¹, opioid³²⁻³³, glutamate³⁴ and adrenergic receptors³⁵⁻³⁶. Concerning adenosine receptors, a photoactivatable A_{2A}R antagonist, MRS7145³⁷, a non-selective photoswitchable partial agonist, MRS5543³⁸, and an adenosine-based photoswitchable agonist, AA-3³⁹ have been reported. Exploration of the pathophysiological function of A_{2A}R and its therapeutic potential may benefit from additional selective A_{2A}R photochromic ligands.

In this study, seeking novel photopharmacological tools for A_{2A}R, we combined photochemistry, cellular and *in vivo* photopharmacology to carry out an in-depth investigation of the light sensitivity of the well-known A_{2A}R antagonist istradefylline and its potential use in photopharmacology. Previous studies have shown that a prolonged exposure of istradefylline to daylight induces a isomerization of its heterostilbene olefinic bond from *trans* to *cis*, resulting in a strong reduction of its affinity for the receptor⁴⁰. Here, we confirm the sensitivity of istradefylline to illumination and, in particular, to near UV light, which rapidly turns it from its active *trans* isomer to the inactive *cis* isomer, and irreversibly leads towards four fully A_{2A}R inactive photoproducts. We then demonstrate that illumination by UV light can rapidly suppress the istradefylline-induced blockade of A_{2A}R cellular signaling and abolish the istradefylline-mediated hyperlocomotor activity induced by A_{2A}R blockade in zebrafish larvae. Taken together, our experiments indicate that istradefylline is a photoinactivatable A_{2A}R antagonist and that this property can be used for the optical control of A_{2A}R in cells and living organisms.

Results

Istradefylline exhibits rapid *trans* to *cis* isomerization under near UV light, while prolonged exposure leads to the generation of photocycloaddition products

Istradefylline, along with other styryl-xanthenes, are known to photoisomerize from *trans* to the corresponding *cis* isomer and vice versa following prolonged illumination with visible light, with a photostationary state obtained between 1.5 and 24 hr depending on the solvent and the concentration used⁴¹⁻⁴².

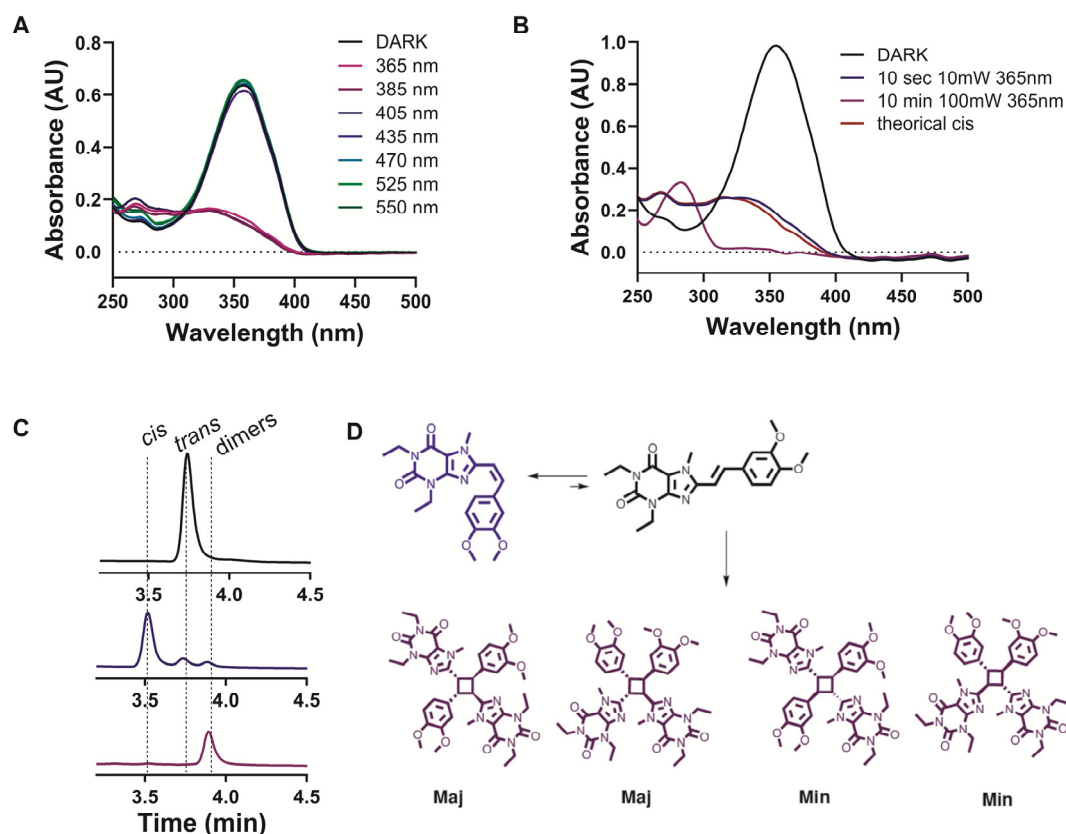


Figure 1: Photochemical characterization of Istradefylline light sensitivity. (A) Absorption spectrum of istradefylline (100 μ M in PBS, 1% DMSO) after 10 s illumination with different wavelength at 10 mW (B) Absorption spectrum of istradefylline, 25 μ M in PBS (20% DMSO) in the dark and after different time and power of 365 nm UV-light illumination (C) HPLC of the corresponding samples of panel B; top: DARK, middle: 10 sec 10mW 365nm, bottom: 10 min 100 mW 365nm. (D) Chemical structures of the photoreaction products of istradefylline after illumination with near-UV light: *trans*-istradefylline in black (corresponding to the peak at $t=3.73$ min in the HPLC chromatograms), *cis*-istradefylline in blue (HPLC $t=3.52$ min) and the group of dimers (HPLC $t=3.89$ min).

First, we determined whether the illumination time to reach a photostationary state could be reduced by the use of wavelength-specific light. To this end, we illuminated 100 μ M istradefylline samples for 10 s with different wavelengths, ranging from 365 to 550 nm at 10 mW, and measured its UV-visible absorption spectrum in PBS (20% DMSO). This revealed that near-UV light drastically affects the absorption spectrum, in the direction of the disappearance of the *trans* isomer (**Figure 1A**). In order to better define the kinetics and nature of the photoreaction, we then monitored the absorption spectrum of istradefylline after exposure to UV light at different times and power levels. Interestingly, we observed a substantial change in the spectrum coherent with a mixture of *trans* and *cis* isomers, with an isosbestic point found at 304nm (**Figure S1**), after only five seconds of illumination with a 365 nm LED light at 10 mW. Illuminating for 5 more seconds slightly changed the absorption spectrum, mainly by reducing the absorbance between 350-370 nm due to the decrease of the *trans* isomer (**Figure 1B**). Based on HPLC-MS analysis (**Figure S2**), it was confirmed that these conditions of illumination mainly led to the *cis*-istradefylline (~93%), with only few remaining *trans* isomers (~5%) (**Figure 1C**). Of note, a

new peak of low intensity at higher retention time was obtained. When illuminating for a longer period and with a higher power (100 mW), the UV-Vis absorption spectrum changed again with a blue-shift of the absorption maximum of 283 nm (**Figure 1B**). The HPLC-MS showed the total disappearance of the peaks corresponding to the *cis* and the *trans* isomers but not the peak at higher retention time that corresponds to a mixture of products (**Figure 1C**), all of them having a molecular mass (*m/z*) of 769.5. This mass is consistent with the dimerization of istradefylline as a result of a [2+2] photocycloaddition of the *trans*-isomer to form several cyclobutane adducts (**Figure 1D**), as previously reported in solid state⁴⁰. The formation of stilbene photocycloaddition products from the *cis*-isomers is generally discarded because the *cis*-stilbene has an S₁ state that is much too short-lived to complete the cycloaddition reaction⁴³. HPLC-MS analysis of the separated cyclobutane analogues obtained in our conditions indicated the existence of at least three different cyclobutane isomers. To identify the structure of the cyclobutane dimers obtained in solution, we illuminated a concentrated solution of istradefylline in DMSO. Preparative HPLC allowed us to separate the three main HPLC peaks corresponding to the dimers. The first fraction (**dimer 1**) appears to be a mixture of two dimers that account for 7% of the total mass of dimers obtained after the purification. The second (**dimer 2**) and third fractions (**dimer 3**) correspond to two pure photocycloaddition products, accounting respectively for 45% and 48% respectively of the total mass of dimers after the purification. The structure of the compounds obtained from the two major fractions were determined by ¹H-NMR and ¹³C-NMR (**Figure S3**). **Dimer 2** corresponds to the *syn* head to tail cyclobutane isomers, previously identified in solid state by the group of Müller⁴⁰ whereas the others have a distinct structure. According to literature, both head to head and head to tail isomers are favored, therefore we assumed that that **dimer 3** is the *syn* head to head cyclobutane derivative. Besides, NMR spectra of similar [2+2] photocycloaddition products⁴⁴⁻⁴⁵, confirms our hypothesis (**Figure S4**). The small quantity obtained for **dimer 1** only allowed us to record an ¹H-NMR spectrum. Based on the comparison between the ¹H-NMR signals obtained for the cyclobutane protons and cyclobutanes obtained in the literature, we tentatively identify them as originating from the less-favored [2+2] photocyclization *anti* head to head and head to tail dimeric pairing of two istradefylline molecules.

UV-light exposure time-dependently decreases the antagonistic activity of istradefylline on A_{2A}R

We next assessed the activity of istradefylline at A_{2A}R at different stages of the photo-transformation. The geometric isomers (i.e. *cis/trans*) and the different irreversible cycloalkane photoproducts were obtained from istradefylline samples exposed to different UV-illumination times. The conditions of illumination were set up in order to be compatible with an analysis of the samples by HPLC-MS. Stock samples in DMSO (2 μL at 10 mM in DMSO) were illuminated with a 365 nm LED source set at 10 mW for 1, 10, 20 and 40 min. Then, these samples were tested in CHO cells stably expressing the human A_{2A}R. Since A_{2A}R couple G_{as} family members, which activate adenylyl cyclase, the cytosolic cAMP concentration was recorded using an endpoint TR-FRET-based cAMP accumulation assay to account for A_{2A}R activation. To assess the antagonistic activity of istradefylline at different times of illumination, A_{2A}R-expressing cells were co-incubated with a fixed concentration of the agonist NECA (1 μM) and increasing concentrations of the different istradefylline samples. A large difference in the IC₅₀ values were observed between the istradefylline sample kept in the dark and the illuminated samples, as we observed a large rightward shift of the inhibition curve (**Figure 2A**). The 1 min -pre-illuminated sample displays a modest difference of potency (3-fold) whereas the 20 minute-illuminated sample shows potency (41-fold), as compared to the istradefylline sample in the dark (**Figure 2C; Table 1**).

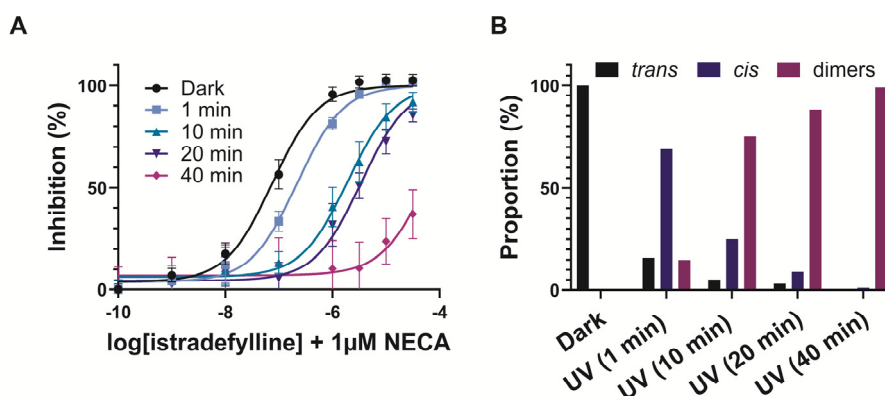


Figure 2: Influence of UV light on istradefylline photoisomerization and photoreaction and its capacity to inhibit $A_{2A}R$ function in cells. (A) The influence of UV light on the ability of istradefylline to inhibit agonist-induced activity of adenosine A_{2A} receptor was determined by measuring the production of cAMP in CHO cells stably expressing the human $A_{2A}R$. Experiments were performed in the presence of a fixed dose of the agonist NECA (1 μ M) and different doses of istradefylline, in the dark or after 365 nm illumination at 10 mW for different exposure times (1 to 40 minutes, as indicated). Data are mean \pm SEM of $n=5$ and $n=3$ independent experiments in dark and UV light conditions respectively. (B) Estimation of the proportion of *trans*, *cis* and dimers after illumination in the same conditions than in A, determined by HPLC from chromatograms displayed at the wavelength of the two isosbestic points *trans-cis* and *trans-dimers*.

To estimate the proportion of the mixture of dimers, *trans* and *cis* isomers in each sample, an analysis was performed from the HPLC-MS istradefylline samples illuminated in the same illumination conditions as for the cell assay (Figure S5, Table S1). As previously shown in a diluted aqueous solution, we first observed by HPLC-MS analysis a fast decrease of the *trans* isomer fraction and concomitant increase in the *cis* isomer. Extending the photoreaction time results in the accumulation of irreversible [2+2]-cycloaddition photoproducts (Figure 2B). After 10 min of illumination, only 5% of the initial *trans* istradefylline remains in the solution (Table 1). We estimate the theoretical IC_{50} of the 5% of remaining $A_{2A}R$ active istradefylline *trans* isomer, to an apparent IC_{50} of 1.4 μ M, close to the potency determined experimentally ($IC_{50} = 1.8 \pm 0.5 \mu$ M). It is also the case for the 20 min illumination sample, where the 3% of remaining *trans*-istradefylline leads to a theoretical IC_{50} of 2.3 μ M, located in the range of the experimental data ($3.1 \pm 0.8 \mu$ M). These results suggest that almost all the remaining antagonistic activity of the pre-illuminated samples is due to the residual *trans* isomer, meaning that the *cis* isomer has a lower antagonistic potency for $A_{2A}R$. It is consistent with the results obtained by the group of Suzuki⁴¹ in which they reported a 800-fold higher activity of the *trans* isomer compared to the *cis* isomer, in a styryl-xanthine molecule similar to istradefylline. Of note, we attempted to obtain purified *cis* isomer, which turns out to have a low bistability and relaxes thermally to the *trans* isomer. Therefore, the isolated sample contained a small amount (14%) of the *trans* isomer, precluding its use in a pharmacological assay to precisely determine the *cis* isomer activity (Figure S6).

	IC_{50} (μ M)	$IC_{50}^{illum}/IC_{50}^{dark}$	% <i>trans</i>	% <i>cis</i>	% dimers
dark	0.07 \pm 0.02		100	0	0
1min	0.24 \pm 0.01	3	16	69	15
10min	1.9 \pm 0.6	27	5	25	70
20min	2.7 \pm 1.1	39	3	9	88
40 min	25.7 \pm 5.1	367	<0.5	1	99

Table 1. Potencies and proportion of isomers and photoproducts in the dark and following different exposure times at 365 nm. IC_{50} and ratio of IC_{50} of istradefylline were determined using a cAMP accumulation assay in CHO cells stably expressing the human $A_{2A}R$. Experiments were performed in the presence of a fixed dose of the agonist NECA (1 μ M) and different doses of istradefylline that have been protected from light (dark) or illuminated with a 365 nm LED (10 mW) for different exposure times (1 to 40 minutes, as indicated). Data are mean \pm SEM of at least 3 independent experiments. Percentage of *trans*, *cis* and dimers entity were calculated from the HPLC analysis of the illuminated samples (365 nm; 10 mW), from chromatograms displayed at the wavelength of the two isosbestic points *trans-cis* and *trans-dimers*.

All these experiments show that istradefylline is antagonizing A_{2A}R only in its *trans* configuration and that light can be used to convert it to the inactive *cis* isomer or dimeric photoproducts.

Istradefylline enables real-time optical control of A_{2A}R cellular signaling

Next, we wanted to take advantage of this rapid light-induced loss of istradefylline activity to dynamically regulate A_{2A}R activity in cells. To assess real time A_{2A}R cell signaling, a double stable cell line containing both the A_{2A}R and the cAMP fluorescent indicator *Flamindo2* was developed. *Flamindo2* is a yellow fluorescent protein-based cAMP indicator⁴⁶ that has an absorption peak at 480 nm and emission maximum at 510 nm, which are fully compatible with istradefylline photochemistry. *Flamindo2* fluorescence is inversely correlated with cAMP concentration, decreasing as cAMP concentration increases (**Figure 3A**). As shown in **Figure 3B**, a dynamic and dose-dependent decrease of fluorescence can be measured following activation of A_{2A}R by the agonist NECA. The potency of NECA extrapolated from experimental analysis (pEC₅₀ = 21 ± 6 nM) is consistent with the results obtained with other cell-based assays (**Figure 3C**).

We then tested whether **dimer 1**, **dimer 2** and **dimer 3** have an antagonistic activity at A_{2A}R. The three dimeric products isolated from a photoreaction (10 μM), *trans*-istradefylline (10 μM, kept in the dark), the antagonist XAC (10 μM) or buffer were applied 3 min prior to the addition of 1 μM of NECA to initiate the production of cAMP. The signals were analyzed by integrating the area under each curve and were normalized to the NECA induced response alone (0% inhibition) and the signal with the vehicle alone (100% inhibition) (**Figure 3D**). We did not observe an inhibition of the NECA induced response by any of the photoreaction dimers at 10 μM while *trans*-istradefylline and XAC at the same concentration reached 86% and 98% of inhibition, respectively.

Next, we assessed the light-dependent activity of istradefylline. Cells were pre-incubated with the photochemically stable antagonist XAC, with istradefylline or vehicle (baseline) for 4.5 min. During this period, the cells were kept in the dark or illuminated for 30 sec at 395 nm. Then the agonist NECA was added to induce cAMP production following A_{2A}R activation. The fluorescence intensity (FI) was recorded during all the experiment, except during the 30 sec of illumination (**Figure 3E**). The signal was normalized and corrected to the baseline to compensate for the loss of fluorescence over time caused by photobleaching. In dark conditions, we can clearly observe a reversion of NECA induced-signal in similar proportions for istradefylline and XAC (**Figure 3F**) while the antagonistic activity of istradefylline is abolished following UV illumination. On the other hand, the antagonistic effect of XAC remains unaltered in both dark or illuminated conditions. This demonstrates that it is possible to dynamically abolish the antagonistic effect of istradefylline on A_{2A}R cell signalling by controlling the UV light-induced phototransformation of istradefylline.

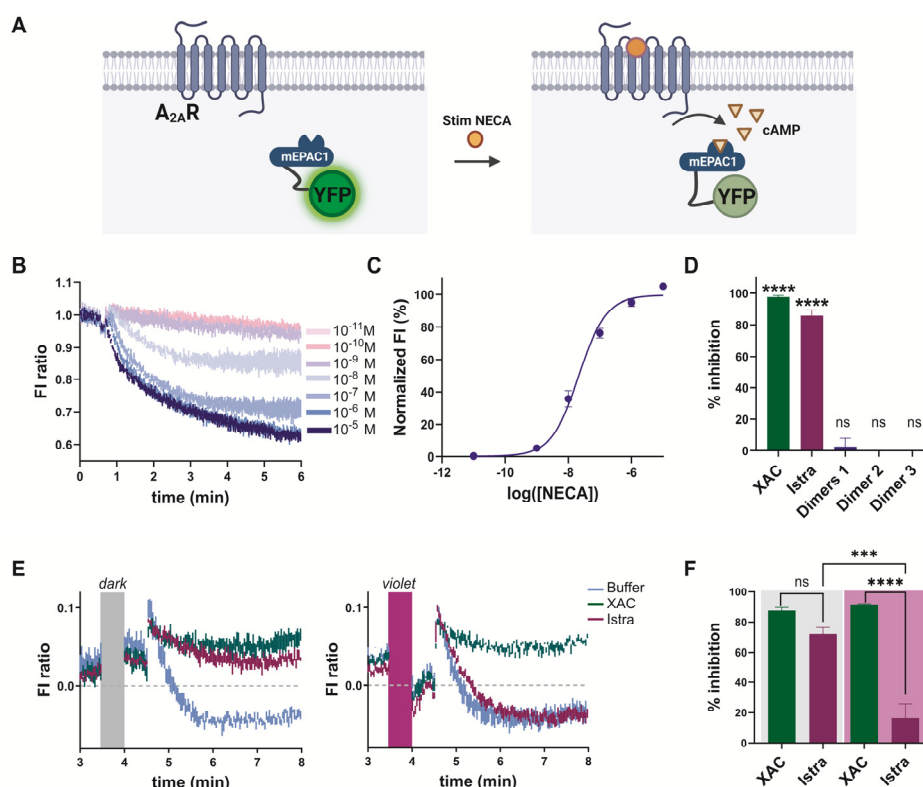


Figure 3: Influence of UV light on istradefylline inhibition on real-time $A_{2A}R$ cellular signaling. (A) Principle of the fluorescence-based assay in CHO- $A_{2A}R$ -Flamindo2 cell line. Fluorescence intensity (FI) of Flamindo2 decreases as the intracellular cAMP concentration increases. (B) Time-course of FI following $A_{2A}R$ stimulation with different concentrations of the agonist NECA in CHO- $A_{2A}R$ -Flamindo2 cell line. (C) Dose-response curve of NECA, as determined from panel B. The peak of FI in the absence of NECA was normalized to 0%, and the peak in the presence of 10 μ M NECA was normalized to 100%. All data points represent the mean \pm SEM of four independent experiments. (D) Quantification of the percentage of receptor inhibition with 10 μ M of XAC, istradefylline and the three purified dimers. Data are mean \pm SEM of at least four independent experiments. (E) Time-course of FI induced by 1 μ M NECA in CHO- $A_{2A}R$ -Flamindo2 cells, alone (blue trace) or in presence of 1 μ M of istradefylline (violet trace) or 1 μ M of the non-photosensitive A_{2A} antagonist XAC (green trace), with 30 seconds dark or violet light (395 nm, 14 mW) treatment (left and right panel respectively). This experiment is representative of three individual experiments. (F) Quantification of the percentage of receptor inhibition with antagonists following dark or UV treatment. Data are presented as mean \pm SEM of $n=5$ and $n=3$ experiments for istradefylline and XAC respectively. Data were analyzed by a One-way ANOVA followed by Dunnett's post hoc test. *** $p < 0.005$, **** $p < 0.001$.

UV light abolishes istradefylline-induced hyperlocomotion in zebrafish larvae

We next assessed whether we can use istradefylline photosensitivity to control $A_{2A}R$ *in vivo* with light using zebrafish larvae. Their transparency makes them ideal models for manipulating photosensitive compounds *in vivo*⁴⁷. Zebrafish possess two $A_{2A}R$ genes named *adora2a.1* and *adora2a.2*, which share 62 and 74% amino acid identity to mammalian $A_{2A}R$ ⁴⁸. The key residues involved in the binding of istradefylline on human $A_{2A}R$ ⁴⁹ are conserved in zebrafish orthologs. Since blockade of $A_{2A}R$ receptors has been previously reported to increase zebrafish locomotion⁵⁰, we evaluated the influence of light on the effect of istradefylline on the free-swimming behavior of naïve 5-day-old zebrafish larvae.

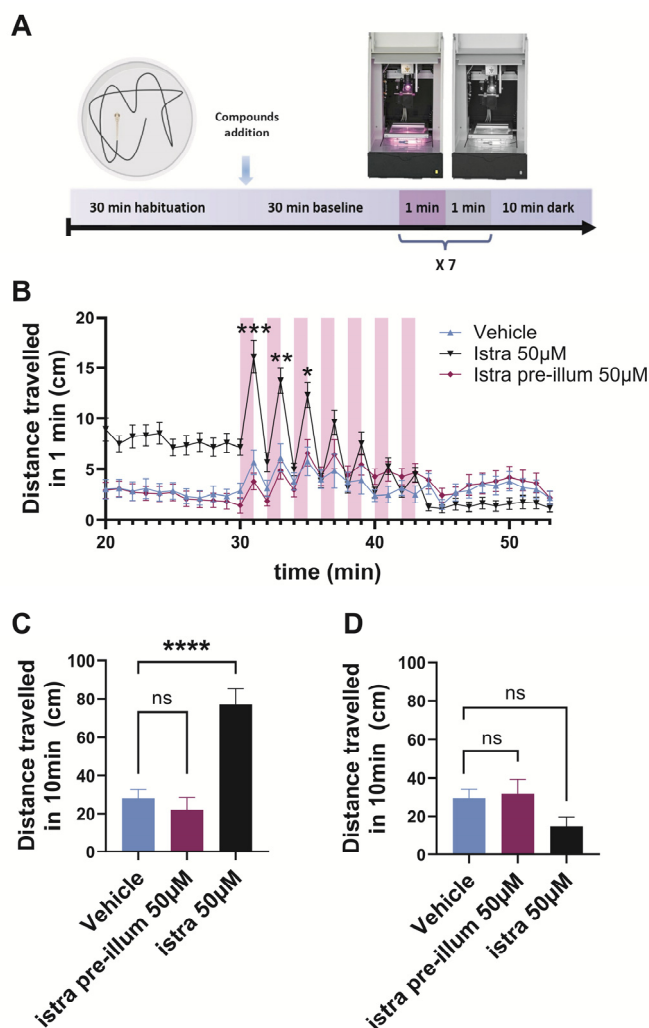


Figure 4: Influence of UV light on istradefylline-induced hyperlocomotion in zebrafish (A) Experimental timeline: One 5 day-old zebrafish larva was placed in a well of a 96-well plate. Following 30 minutes of habituation, larvae were exposed for 30 minutes to vehicle (DMSO 0.5%), 50 μ M istradefylline (kept in the dark) or 50 μ M UV-pretreated istradefylline (Istra pre-illum), directly added to the well. After 30 minutes, zebrafish received seven UV/dark illumination cycles (1 minute UV (365 nm, 8 mW) and 1 minute dark). (B) Integration of the total free-swimming distance for every minute. Each data point corresponds to the mean \pm SEM of the total distance swum during 1 minute by 15-16 zebrafish larvae. (C, D) Cumulative distance travelled in 10 minutes by the zebrafish larva treated with vehicle, istradefylline or pre-illuminated istradefylline, before (C) and after (D) the illumination cycles. Each data point corresponds to the mean \pm SEM of the total distance swum during 10 minutes by 15-16 zebrafish larvae. Data shown correspond to a representative experiment from 3 independent experiments. Statistics were performed with a one-way (panel C) or a two-way (panel B) ANOVA followed by Dunnett's multiple comparison test. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.005$.

We used a conventional locomotion assay on zebrafish larvae coupled to a LED illumination system to measure locomotion after direct addition of the compounds in the medium, either in the dark or under illumination with violet light (385 nm) (**Figure 4A**). Istradefylline in its *trans* configuration increased larvae motility 2 fold, while the istradefylline sample that has been pre-illuminated (365 nm, 10 mW, 2 min) had no effect on motility (**Figure 4C, left**) in agreement with the $A_{2A}R$ activity that we recorded in cell-based assays. Under these illumination conditions, HPLC-MS analysis indicated that the solution contains a majority of dimers (~75%), some *cis* isomers (~24%) and 1% of *trans* isomer (**Figure S7, Table S2, S3**). UV light by itself enhanced the swimming behavior of the larvae (**Figure 4B**), as shown in experiments with zebrafish and light in previous reports^{47, 51}. This effect is exacerbated in the presence of the active *trans* isomer, where UV-induced hyperlocomotion and istradefylline-induced hyperlocomotion are additive, while maintaining a ratio of around 2.2 between the distance travelled

under illumination vs the distance without illumination (**Figure 4B**). However, the istradefylline-induced hypermobility is lost after four UV illumination cycles (385 nm, 1 min at 8mW each cycle) (**Figure 4C, right**). The conditions of illumination after each of the four first illumination cycles were reproduced and analyzed by HPLC-MS to obtain the ratio of istradefylline isomers and other photo-products (**Figure S8, Table S2**). After the first cycle, only 10% of *trans* isomer remains which decreased to 2% after cycle 2 and 3 and further decreased to 1% after cycle 4, the rest being a mixture of *cis* and dimer photo-products (**Table S3**). The loss of istradefylline activity is consistent with a conversion from the *trans* A_{2A}R-active form to the less active or inactive *cis*/photoreaction.

In this series of experiments, we provide evidence that istradefylline activity can be rapidly switched-off *in vivo* in living zebrafish larvae by the application of UV-light.

Discussion

In this study, we explored the influence of UV light on istradefylline at the molecular and functional level. We showed that, in solution, near UV light rapidly triggers the reversible isomerization of the ligand from its active *trans* isomer to the less active *cis* isomer and that UV light also mediates its irreversible transformation into fully inactive dimeric photo-products. Finally, we demonstrated that UV-triggered photoinactivation of istradefylline is compatible with real-time photopharmacological experiments both *in vitro* and *in vivo*.

We elaborated the present study based on previous work showing that istradefylline, along with others styryl-xanthenes, is photosensitive. Istradefylline was known to isomerize following prolonged illumination with light^{40, 42}. Moreover, in solid state, Hockemeyer and colleagues reported the occurrence of a [2+2] photocycloaddition product after illumination of the sample for 100 minutes that they identified to be a cyclobutane derivative originating from the *syn* head to tail dimeric pair of the *trans* isomers⁴⁰. One of the findings of our study is that, after sustained near-UV exposure in solution, we were able to identify four different dimeric cyclobutane photoproducts of istradefylline (**Figure 1D, S4**). **Dimer 2** corresponds to the previously identified product⁴⁰ whereas **dimers 1** and **dimer 3** have distinct structures, that we propose to be a mixture of *anti* head to head and head to tail cyclobutane isomers and the pure *syn* head to head cyclobutane isomer respectively. In a study from Matsuura group on the [2+2] photocyclization of 4-cyanostilbene, the different cyclobutane isomers were obtained in a similar ratio⁵². All together, these results demonstrate that light promotes a dynamic equilibrium between *trans* and *cis* istradefylline isomers, which under sustained exposure to light mediates the production of four photo-products which correspond to the different dimeric cyclobutane analogues of istradefylline.

One of the interesting outcomes from this study is that we can harness the photosensitivity of istradefylline to our advantage and use it as photoinactivatable antagonist tool in cells or living animals. We demonstrate that the dimeric photo-products are fully inactive towards A_{2A}R and that exposure to near-UV light rapidly and dramatically decreases the antagonistic activity of istradefylline to A_{2A}R making it compatible with *in vivo* photopharmacology experiments. This photochemical property can be exploited to obtain localized optically control of A_{2A}R activity with a high temporal precision in biological media. In combination with the photoactivatable A_{2A}R antagonist MRS7145³⁷, the photoinactivatable A_{2A}R antagonist istradefylline provides a complementary light-controlled molecular probe to dynamically study A_{2A}R function. Of interest, istradefylline is a readily available well-characterized clinically proven drug. This powerful and accessible tool could find useful applications to decipher the physiopathological mechanisms related to A_{2A}R activation and inactivation during defined times and at precise locations in cells, ex-vivo and in animal models. For example, it could be possible to block all the A_{2A} receptors in an organism and, with very precise control in space and time, to remove the block at a specific site and observe the associated biological, physiological or behavioral responses. This study also demonstrates that istradefylline has to be strictly protected from light by users to avoid the photodegradation of the active drug into (*trans/cis*/photodimer)-istradefylline mixtures. Moreover, photosensitivity has to be considered for the handling, packaging and storage of this drug. Future studies should be aimed at developing a selective photoswitchable A_{2A}R antagonist that could be reversibly activated and inactivated by light, which would provide an additional temporal dimension to the control of this receptor and expand the range of A_{2A} photocontrollable molecules. Based on our data, we speculate that istradefylline appears to be a good scaffold candidate for its transformation into a selective photoswitchable A_{2A}R antagonist.

The concept of photo-repurposing was recently defined by Kobauri et al²⁴ as the study of light-induced changes in activity of compounds that were not initially designed for photopharmacology. There are several examples of

photo-repurposing, such as Plinabulin⁵³, an tubulin polymerization inhibitor, and GL479⁵⁴, a dual PPAR α /y agonist, to cite two recent ones, now joined by istradefylline. Considering styryl-based compounds, two studies examined the difference of activity between the *trans* and the *cis* isomers. The Thorn-Seshold group successfully developed a light-specific styryl-benzothiazole (SBT)⁵⁵, a tubulin polymerization inhibitor, by photoconverting the *trans* inactive isomer into the 200-fold more active *cis* isomer with fast application of violet light in cells, whole organ and *in vivo*. More similarly to our study, axitinib, a -styryl heteroarene tyrosine kinase inhibitor marketed for second-line therapy of renal cell carcinoma, displays a phototransformation with UV-light⁵⁶. In a vascular endothelial growth factor receptor 2 assay, isolated *trans*-axitinib was more active than the *cis* isomer, but the generation of an inactive [2+2] photocycloaddition product with UV light prevented reversible switching, leading to a reduction in the activity of the molecule after illumination. Taken together, these data suggest a proof of concept to use certain styryl-based molecules that can undergo a fast isomerization and/or [2+2] photocycloaddition with UV light as a way to irreversibly induce a reduction in activity of the molecule following illumination. It also opens a question on the photostability of stilbene or other functional group containing drugs and the possible deleterious effects of light exposure on drug therapeutic treatments⁵⁷⁻⁵⁹. Beyond these examples, there are very likely other photoswitch-containing ligands that have the potential to become photopharmacological tools. Notably, considering that the styryl arene or heteroarene group is widely represented in medicinal chemistry⁶⁰, this category could be expanded to include other styryl-arene-based compounds. With its light-induced irreversible loss of activity due to photocycloaddition products, the styryl-containing istradefylline has exactly the opposite property to photoactivatable ligands. If more examples of this type were to be discovered, they could therefore constitute a new class of photopharmacological ligands, i.e. photoinactivatable ligands that are irreversibly inactivated by light, allowing rapid, local neutralization of their pharmacological activity, in complement of photoactivatable (caged) and photoswitchable ligands.

In conclusion, following an in-depth understanding of the photosensitivity of istradefylline at the molecular level, we propose a new application for istradefylline as a photopharmacological tool. We have established the proof of concept that the blockade of A_{2A}R function by istradefylline can be rapidly abolished by near UV light in living cells and organisms, in a spatiotemporal-controlled manner. Istradefylline behaves as a *bone fide* photoinactivatable A_{2A}R antagonist, which could find useful applications to explore the role of localized A_{2A}R in biological, physiological or behavioral responses in various pathophysiological contexts.

Methods

Photochemistry - UV-Visible absorption spectroscopy.

For the determination of the optimal illumination wavelengths, UV-Visible absorption spectra were recorded using a Spark microplate reader. Istradefylline samples were prepared 100 μ M in PBS with 0.1% DMSO at room temperature. Samples were measured between 500 nm and 250 nm with 2 nm fixed intervals in 96-well transparent plates (200 μ L of compound solution/well). Illumination at the different wavelengths was achieved using a 96-LED plate (LEDA-x, BRC), set at 10 mW.

For the determination of the kinetics and nature of the photoreaction, UV-Visible spectra were recorded using a Thermofisher UV-vis spectrophotometer. Samples were prepared with 25 μ M of istradefylline in PBS with 20% DMSO. Samples were measured between 250 and 600 nm with 2 nm fixed intervals in a quartz cuvette (200 μ L of compound solution/cuvette). Illumination at 365 nm was achieved using the CoolLED pE4000 light source, set at 10% intensity (10 mW) or 100% (100 mW).

Estimation of the Cis/Trans/photoproduct content by HPLC

For the quantitative analysis of the HPLC chromatogram obtained upon different illumination conditions of the istradefylline samples, the first step was to identify the three isosbestic points (*trans-cis*, *trans*-dimers and *cis*-dimers). The *trans-cis* isosbestic point was obtained from the UV-Vis spectra of a 25 μ M sample of a mixture of only *trans* and *cis* isomer, obtained after 5 seconds of illumination of the sample with 365 nm light (CoolLED pE4000) set at 10mW. The *trans*-dimers isosbestic point was obtained from the UV-Visible absorption spectrum of *trans* isomer and pure dimers, obtained respectively in the dark and after 10 minutes of illumination of the same sample with high power light (100 mW). To obtain the theoretical pure *cis* absorption spectrum, a 25 μ M sample was illuminated for 10 seconds with 365 nm light set at 10 mW, conditions that mainly lead to the *cis* isomer. To estimate the proportion of each species in this sample, it was first analyzed by HPLC-MS (see conditions and gradient at *Analytical HPLC-MS & NMR* section) and two chromatograms were generated at the

wavelengths of the two isosbestic points *trans-cis* and *trans-dimers*. For each one, the two peaks corresponding to the two products of the corresponding isosbestic point were integrated to obtain the relative proportion of each. Respecting the *trans/cis* and the *trans/dimers* proportion, an estimation of the ratio of dimers and *trans* and *cis* isomer was calculated. The pure *cis* spectrum was then extrapolated, implementing the percentage found for the *cis* and subtracting the effect given by the remaining *trans* and the percentage of dimers produced in these conditions. From this theoretical *cis* isomer absorption spectrum, we obtained the *cis*-dimers isosbestic point used for the calculation of the estimation of *cis/trans/dimers* content from the different HPLC chromatograms obtained in different illumination conditions.

Analytical HPLC-MS & NMR

Analytical HPLC was performed on a Thermo Ultimate 3000SD (Thermo Scientific Dionex) coupled to a PDA detector and Mass Spectrometer LTQ XL ESI-ion trap (Thermo Scientific) (HPLC-PDA-MS) or on a Waters 2795 Alliance coupled to a DAD detector (Agilent 1100) and an ESI Quattro Micro MS detector (Waters); HPLC columns used were ZORBAX Eclipse Plus C18 (4.6x150mm; 3.5µm) and ZORBAX Extend-C18 (2.1 x 50 mm, 3.5 µm) respectively.

HPLC purity was determined using the following binary solvent system: A water +0.05% formic acid, B acetonitrile (ACN) with 0.05% formic acid: 5%B for 0.5 min, from 5- 100%B in 5 min, 100%B for 1.5 min, 100-5%B in 1 min and 5%B for 2 min (total runtime 10 min). The flow rate was 0.5 mL/min, column temperature was fixed to 35°C and wavelengths from 210-600 nm were registered.

Nuclear Magnetic Resonance (NMR) spectroscopy was performed using a Brüker Avance NEO instrument 400 MHz spectrometer. Chemical shifts are reported in δ (ppm) relative to an internal standard (non-deuterated solvent signal) [Chloroform (CDCl₃) δ = 7.26, 1.56 ppm (¹H), δ = 77.16 ppm (¹³C)].

Purification of photoreaction istradefylline products

A vial with 10 mg of istradefylline in DMSO (20 mM) was illuminated with 365 nm light (100 mW) for 9 hr. The sample was purified by preparative HPLC using a phenomenex column (00G-4435-N0 Gemini 5u C18 110A 250 x 10.00 mm 5 micron 534461-1), using a flow of 10 mL/min and water/acetonitrile as solvents (+0.05% formic acid) and using a low gradient from 40 to 50% acetonitrile. Three main fractions were obtained named **Dimers1**, **Dimer2** and **Dimer3**.

Dimers1, mixture of the *anti* head to tail and head to head cyclobutane isomers: 8,8'-((1*r*,2*r*,3*r*,4*r*)-2,4-bis(3,4-dimethoxyphenyl)cyclobutane-1,3-diyl)bis(1,3-diethyl-7-methyl-1*H*-purine-2,6(3*H*,7*H*)-dione) and 8-((1*S*,2*R*,3*R*,4*S*)-2,3-bis(3,4-dimethoxyphenyl)-4-(3-ethyl-1,7-dimethyl-2,6-dioxo-2,3,6,7-tetrahydro-1*H*-purin-8-yl)cyclobutyl)-1,3-diethyl-7-methyl-1*H*-purine-2,6(3*H*,7*H*)-dione (0,2 mg, 0,130 µmol, 2 % yield); ¹H NMR (400 MHz, CDCl₃) δ 7.09 (d, J = 2.0 Hz, 2H), 6.93 – 6.88 (m, 2H), 6.84 (d, J = 8.3 Hz, 2H), 6.72 (d, J = 2.0 Hz, 2H), 6.68 (dd, J = 8.3, 2.1 Hz, 2H), 6.55 (d, J = 8.2 Hz, 2H), 5.76 (t, J = 10.6 Hz, 2H), 4.43 (t, J = 9.7 Hz, 2H), 4.24 (app. t, J = 9.9 Hz, 4H), 4.21 – 4.13 (m, 8H), 4.00 (q, J = 6.9 Hz, 8H), 3.87 (s, 6H), 3.84 (s, 6H), 3.75 (s, 6H), 3.64 (s, 12H), 3.58 (s, 6H), 1.33 (t, J = 7.0 Hz, 12H), 1.21 (t, J = 7.0 Hz, 12H).

Dimer2, *syn* head to tail cyclobutane isomer : 8-((1*S*,2*S*,3*S*,4*S*)-2,4-bis(3,4-dimethoxyphenyl)-3-(1-ethyl-3,7-dimethyl-2,6-dioxo-2,3,6,7-tetrahydro-1*H*-purin-8-yl)cyclobutyl)-3-ethyl-1,7-dimethyl-1*H*-purine-2,6(3*H*,7*H*)-dione (2,7 mg, 3,51 µmol, 27% yield); ¹H NMR (400 MHz, CDCl₃) δ 6.86 (dd, J = 8.3, 2.1 Hz, 2H), 6.71 (d, J = 8.3 Hz, 2H), 6.66 (d, J = 2.1 Hz, 2H), 5.06 (dd, J = 9.6, 7.6 Hz, 2H), 4.30 (dd, J = 10.0, 7.3 Hz, 2H), 4.21 (q, J = 7.0 Hz, 4H), 4.02 (q, J = 6.9 Hz, 4H), 3.82 (s, 6H), 3.72 (s, 6H), 3.60 (s, 6H), 1.36 (t, J = 7.0 Hz, 6H), 1.21 (t, J = 7.0 Hz, 6H).

Dimer3, *syn* head to head cyclobutane isomer: 8,8'-((1*R*,2*S*,3*R*,4*S*)-3,4-bis(3,4-dimethoxyphenyl)cyclobutane-1,2-diyl)bis(1,3-diethyl-7-methyl-1*H*-purine-2,6(3*H*,7*H*)-dione) (2,9 mg, 3,77 µmol, 29 % yield); ¹H NMR (400 MHz, CDCl₃) δ 6.75 (d, J = 8.3 Hz, 2H), 6.71 (dd, J = 8.3, 1.9 Hz, 2H), 6.41 (d, J = 1.9 Hz, 2H), 4.76 (app. d, J = 6.4 Hz, 2H), 4.35 (app. d, J = 6.4 Hz, 2H), 4.08 – 3.97 (m, 8H), 3.83 (s, 6H), 3.71 (s, 6H), 3.64 (s, 6H), 1.22 (t, J = 7.0 Hz, 6H), 1.17 (t, J = 7.0 Hz, 6H).

Cell culture and stable transfection

Chinese hamster ovary FlpIn (FlpIn-CHO) cells obtained from ThermoFisher Scientific were grown in Ham's F12 medium supplemented with 10% fetal bovine serum (previously inactivated at 55 °C for 30 min). Manipulation and cell culture were done in a biological safety cabinet class 1 and in an incubator at 37°C, 5% CO₂. Absence of mycoplasma was checked regularly to guarantee the use of mycoplasma-free cells. CHO-A_{2A}R

stable cells have been generated using the FlpIn system following manufacturer's instructions, which allows the generation of isogenic stable cell line through the guided insertion of the gene at a unique FRT-site. Briefly, DNA coding for human A_{2A} was inserted in the pcDNA5/FRT-plasmid using cloning sites HindIII and Xho. The ligated DNA has been transformed in *Escherichia coli* DH5 α competent cells and extracted from different colonies. The integrity of the full plasmid has been checked by sequencing. CHO Flp-In cells have been co-transfected with the pcDNA5/FRT-A_{2A} plasmid and the Flp recombinase vector pOG44 using lipofectamine, according to manufacturer instructions. The cells having integrated the plasmid into their genome has been selected with hygromycin treatment at 400 μ g/mL for two weeks. CHO-FlpIn cells stably expressing the human A_{2A}R were used in pharmacological experiments.

Cell-based cAMP accumulation assay

CHO cells stably expressing A_{2A}R were seeded in black clear-bottom 96-well plates at a concentration of 10 000 cells/well and incubated at 37°C and 5% CO₂ for 24 h. cAMP accumulation was estimated using the cAMP Gs HTRF kit (Revvity) according to the manufacturer's instructions. Briefly, 2 μ L of istradefylline aliquots were illuminated for different time at 10 mW at 365 nm using a pE4000 CoolLED illumination system. Cells were stimulated with the agonist NECA to induce cAMP accumulation while being treated with istradefylline pre-illuminated under the different light conditions for 20 min, at 37°C. The stimulation buffer also contained adenosine deaminase (0.5 U/mL) to avoid the response induced by ambient adenosine and the PDE IV inhibitor Ro-20-1724, to avoid cAMP degradation. Cells were lysed and then transferred to a 384-well plate. The d₂-labeled cAMP analog and the europium cryptate-labeled anti-cAMP antibody were added and the plate was incubated 1 hr at room temperature. The plate was read using a PheraStar fluorimeter (BMG Labtech) microplate reader. Data were fitted by non-linear regression and normalized to istradefylline maximal inhibition response (100%) using GraphPad Prism 10 software.

Visualization of intracellular cAMP dynamics

To perform these experiments, CHO-A_{2A}R cells were infected by lentivirus encoding the yellow fluorescent cAMP indicator Flamindo2 and a puromycin resistance gene to obtain a polyclonal stable cell line expressing both A_{2A}R and Flamindo2, after selection with 6 μ g/mL puromycin. Flamindo2 was a gift from Tetsuya Kitaguchi (Addgene plasmid #73938; <http://n2t.net/addgene:73938>; RRID:Addgene_73938)⁴⁶. Twenty-four hours before the experiment, CHO-A_{2A}-Flamindo2 were plated in black clear-bottom 96-well plates at a concentration of 100 000 cells/well and incubated at 37°C and 5% CO₂. The day after, medium was changed with the appropriate volume of FDSS (Functional Drug Screening System) buffer containing 1 mM MgSO₄, 20 mM HEPES, 3.3 mM Na₂CO₃, 1.3 mM CaCl₂ and 0.1% of BSA and cells were incubated for 10 min. The decrease of fluorescence due to cAMP production was assessed in a 96-wells plate containing 100 000 cells/well using a FDSS/ μ CELL kinetic plate imager (Hamamatsu) implemented with a custom LED illumination module that allows illuminating 6 \times 3 wells at 395 nm directly inside the reading chamber. For the characterization of the CHO-A_{2A}-Flamindo2 stable cell line, 20 μ L of 10X NECA solution at increasing concentrations were added after 1min of baseline and the subsequent decrease of fluorescence was followed for 6 min. Fluorescence intensity (FI) signal was monitored at 540 nm after an excitation at 480 nm. A similar protocol was made to obtain the activity of the three purified fractions of the [2+2]-photocycloaddition products obtained. After 1 min of baseline, a 100 μ M solution (10X) of the three purified fractions, istradefylline, XAC or vehicle was injected. FI signal was recorded for 3 min then a 10 μ M (10X) of the agonist NECA was injected and FI was recorded for an additional 8 min. For photopharmacological experiments, the cells were accommodated 10 min at 37°C with 160 μ L/well of FDSS buffer. The experiment started with one minute of baseline followed by the addition of 20 μ L of a 10 μ M solution containing either the A_{2A}R antagonists istradefylline or XAC. FI was recorded for 3 min then, recording was stopped during 30 s, during which illumination with 395 nm LED was applied in half of the wells. FI was recorded for 20 sec then 20 μ L of a 10 μ M solution of the agonist NECA was added and the fluorescence was recorded for 8 min. For quantitative analysis, the area under the curve after the injection of NECA was determined using GraphPad Prism software.

Behavioral tests in zebrafish larvae (*Danio rerio*)

Fish locomotion was measured using a video tracking system (Daniovision, Noldus), equipped with a LED illumination module (385 nm) placed above the observation chamber. We used 5-days post-fertilization fish larvae. One zebrafish larva was placed in each well of a 96-well plate (corning Costar) with 75 μ L of E3 medium. Thirty min prior to the measurement, the plate was placed inside the observation chamber, previously set at 28°C, for acclimatization of the fish to the environment. After this period, 75 μ L of 2X solution of the different

compounds with 1% DMSO (0.5% DMSO final) was then applied directly into the wells. The pre-illuminated istradefylline sample was prepared from the 2X solution with an illumination for 2 min at 365 nm at 10 mW (CoolLED, pE4000). The locomotion was recorded 2 min after the addition of the molecules for the duration of the experiment. Zebrafish larvae remained in the dark during the first 30 min then were submitted to seven cycles of 1 min UV illumination (385 nm, 8 mW) and 1 min in the dark, followed by 10 min in the dark.

Ligands and chemicals

All chemicals were reagent grade (Merck, or Sigma, Germany or Fluorochem, Ireland). Istradefylline was purchased from MedChemExpress while NECA and XAC were purchased from Bio-technie Tocris. Except when stated otherwise, istradefylline was carefully protected from light during the experiments.

Statistics

All data are reported as mean \pm standard error of the mean (\pm SEM). Number of experiments/animals and statistical tests that were performed on datasets are indicated in Figure Legends. Data were analyzed with Microsoft excel 2016 and GraphPad Prism softwares v10, using one-way or two-way analysis of variance (ANOVA) and the appropriate post-hoc tests for multiple comparisons. Data were considered significant when $p < 0.05$.

Supporting Information

UV-visible absorption spectra; HPLC chromatograms; ¹H-NMR spectra; Figure showing the preorganization of istradefylline for the auto-photocycloaddition reaction in solution and the stereochemistry of the products obtained; Summary tables of relative percentage of *cis* and *trans* istradefylline isomers and photocycloaddition products in the illumination conditions corresponding to Fig. 2 and Fig. 4.

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Author contributions

Conceptualization, AL, GL and CG; Methodology, XGS, FM, CJ, DM, GL, AL and CG; Investigation, AD, XGS, FM and CG; Ressources: AL, CJ, DM, GL, and CG; Writing – Original Draft, AD and CG; Writing – Review & Editing, all authors; Visualization, AD, XGS and CG; Funding Acquisition, CG, GL and AL; Supervision, XGS, CJ, GL, AL and CG

Declaration of interests

The authors declare no competing interests.

Bibliography

1. Alexander, S. P. H.; Christopoulos, A.; Davenport, A. P.; Kelly, E.; Mathie, A.; Peters, J. A.; Veale, E. L.; Armstrong, J. F.; Faccenda, E.; Harding, S. D.; Pawson, A. J.; Sharman, J. L.; Southan, C.; Davies, J. A.; Collaborators, C.; Abbracchio, M. P.; Alexander, W.; Al-hosaini, K.; Bäck, M.; Beaulieu, J. M.; Bernstein, K. E.; Bettler, B.; Birdsall, N. J.; Blaho, V.; Bousquet, C.; Bräuner-Osborne, H.; Burnstock, G.; Caló, G.; Castaño, J. P.; Catt, K. J.; Ceruti, S.; Chazot, P.; Chiang, N.; Chun, J.; Cianciulli, A.; Clapp, L. H.; Couture, R.; Csaba, Z.; Dent, G.; Singh, K. D.; Douglas, S. D.; Dournaud, P.; Eguchi, S.; Escher, E.; Filardo, E.; Fong, T. M.; Fumagalli, M.; Gainetdinov, R. R.; Gasparo, M. d.; Gershengorn, M.; Gobeil, F.; Goodfriend, T. L.; Goudet, C.; Gregory, K. J.; Gundlach, A. L.; Hamann, J.; Hanson, J.; Hauger, R. L.; Hay, D.; Heinemann, A.; Hollenberg, M. D.; Holliday, N. D.; Horiuchi, M.; Hoyer, D.; Hunyady, L.; Husain, A.; Ijzerman, A. P.; Inagami, T.; Jacobson, K. A.; Jensen, R. T.; Jockers, R.; Jonnalagadda, D.; Karnik, S.; Kaupmann, K.; Kemp, J.; Kennedy, C.; Kihara, Y.; Kozielowicz, P.; Kreienkamp, H. J.; Kukkonen, J. P.; Langenhan, T.; Leach, K.; Lecca, D.; Lee, J. D.; Leeman, S. E.; Leprince, J.; Lolait, S. J.; Lupp, A.; Macrae, R.; Maguire, J.; Mazella, J.; Mc Ardle, C. A.; Melmed, S.; Michel, M. C.; Miller, L.; Mitolo, V.; Mouillac, B.; Murphy, P. M.; Nahon, J. L.; Norel, X.; Nyimanu, D.; O'Carroll, A. M.; Offermanns, S.; Panaro, M. A.; Pertwee, R. G.; Pin, J. P.; Prossnitz, E.; Ramachandran, R.; Reinscheid, R. K.; Rondard, P.; Rovati, G. E.; Ruzza, C.; Sanger, G.; Schöneberg, T.; Schulte, G.; Schulz, S.; Segaloff, D. L.; Serhan, C. N.; Stoddart, L. A.; Sugimoto, Y.; Summers, R.; Tan, V.; Thomas, W.; Timmermans, P. B.; Tirupula, K.; Tulipano, G.; Unal, H.; Unger, T.; Vanderheyden, P.; Vaudry, D.; Vaudry, H.; Vilardaga, J. P.; Walker, C. S.; Ward, D. T.; Wester, H. J.; Willars, G. B.; Williams, T. L.; Woodruff, T. M.; Yao, C., THE CONCISE GUIDE TO PHARMACOLOGY 2019/20: G protein-coupled receptors. *British Journal of Pharmacology* **2019**, *176* (S1).
2. Hilger, D.; Masureel, M.; Kobilka, B. K., Structure and dynamics of GPCR signaling complexes. *Nature Structural & Molecular Biology* **2018**, *25* (1), 4-12.
3. Hauser, A. S.; Attwood, M. M.; Rask-Andersen, M.; Schiöth, H. B.; Gloriam, D. E., Trends in GPCR drug discovery: new agents, targets and indications. *Nature Reviews Drug Discovery* **2017**, *16* (12), 829-842.
4. Santos, R.; Ursu, O.; Gaulton, A.; Bento, A. P.; Donadi, R. S.; Bologa, C. G.; Karlsson, A.; Al-Lazikani, B.; Hersey, A.; Oprea, T. I.; Overington, J. P., A comprehensive map of molecular drug targets. *Nature Reviews Drug Discovery* **2017**, *16* (1), 19-34.
5. Ijzerman, A. P.; Jacobson, K. A.; Müller, C. E.; Cronstein, B. N.; Cunha, R. A., International Union of Basic and Clinical Pharmacology. CXII: Adenosine Receptors: A Further Update. *Pharmacological Reviews* **2022**, *74* (2), 340-372.
6. Saini, A.; Patel, R.; Gaba, S.; Singh, G.; Gupta, G. D.; Monga, V., Adenosine receptor antagonists: Recent advances and therapeutic perspective. *European Journal of Medicinal Chemistry* **2022**, *227*, 113907.
7. Berger, A. A.; Winnick, A.; Welschmeyer, A.; Kaneb, A.; Berardino, K.; Cornett, E. M.; Kaye, A. D.; Viswanath, O.; Urits, I., Istradefylline to Treat Patients with Parkinson's Disease Experiencing "Off" Episodes: A Comprehensive Review. *Neurology International* **2020**, *12* (3), 109-129.
8. Schwarzschild, M. A.; Agnati, L.; Fuxe, K.; Chen, J.-F.; Morelli, M., Targeting adenosine A2A receptors in Parkinson's disease. *Trends in Neurosciences* **2006**, *29* (11), 647-654.
9. Boknik, P.; Eskandar, J.; Hofmann, B.; Zimmermann, N.; Neumann, J.; Gergs, U., Role of Cardiac A2A Receptors Under Normal and Pathophysiological Conditions. *Frontiers in Pharmacology* **2021**, *11*, 627838.
10. Sun, C.; Wang, B.; Hao, S., Adenosine-A2A Receptor Pathway in Cancer Immunotherapy. *Frontiers in Immunology* **2022**, *13*, 837230.
11. Yu, F.; Zhu, C.; Xie, Q.; Wang, Y., Adenosine A_{2A} Receptor Antagonists for Cancer Immunotherapy: Miniperspective. *Journal of Medicinal Chemistry* **2020**, *63* (21), 12196-12212.
12. Sawynok, J., Adenosine receptor targets for pain. *Neuroscience* **2016**, *338*, 1-18.

13. Merighi, S.; Borea, P. A.; Varani, K.; Vincenzi, F.; Jacobson, K. A.; Gessi, S., A2A Adenosine Receptor Antagonists in Neurodegenerative Diseases. *Current Medicinal Chemistry* **2022**, *29* (24), 4138-4151.
14. Iskandrian, A. S., Adenosine Myocardial Perfusion Imaging.
15. Elkholy, K. O.; Hegazy, O.; Okunade, A.; Aktas, S.; Ajibawo, T., Regadenoson Stress Testing: A Comprehensive Review With a Focused Update. *Cureus* **2021**.
16. Chen, J.-F.; Cunha, R. A., The belated US FDA approval of the adenosine A2A receptor antagonist istradefylline for treatment of Parkinson's disease. *Purinergic Signalling* **2020**, *16* (2), 167-174.
17. Dungo, R.; Deeks, E. D., Istradefylline: First Global Approval. *Drugs* **2013**, *73* (8), 875-882.
18. Shimada, J.; Koike, N.; Nonaka, H.; Shiozaki, S.; Yanagawa, K.; Kanda, T.; Kobayashi, H.; Ichimura, M.; Nakamura, J.; Kase, H.; Suzuki, F., Adenosine A2A antagonists with potent anti-cataleptic activity. *Bioorganic & Medicinal Chemistry Letters* **1997**, *7* (18), 2349-2352.
19. Orr, A. G.; Lo, I.; Schumacher, H.; Ho, K.; Gill, M.; Guo, W.; Kim, D. H.; Knox, A.; Saito, T.; Saïdo, T. C.; Simms, J.; Toddes, C.; Wang, X.; Yu, G.-Q.; Mucke, L., Istradefylline reduces memory deficits in aging mice with amyloid pathology. *Neurobiology of Disease* **2018**, *110*, 29-36.
20. Lu, J.; Cui, J.; Li, X.; Wang, X.; Zhou, Y.; Yang, W.; Chen, M.; Zhao, J.; Pei, G., An Anti-Parkinson's Disease Drug via Targeting Adenosine A2A Receptor Enhances Amyloid- β Generation and γ -Secretase Activity. *PLOS ONE* **2016**, *11* (11), e0166415.
21. Hoorens, M. W. H.; Szymanski, W., Reversible, Spatial and Temporal Control over Protein Activity Using Light. *Trends in Biochemical Sciences* **2018**, *43* (8), 567-575.
22. Hüll, K.; Morstein, J.; Trauner, D., *In Vivo* Photopharmacology. *Chemical Reviews* **2018**, *118* (21), 10710-10747.
23. Ellis-Davies, G. C., Caged compounds: photorelease technology for control of cellular chemistry and physiology. *Nat Methods* **2007**, *4* (8), 619-28.
24. Kobauri, P.; Dekker, F. J.; Szymanski, W.; Feringa, B. L., Rational Design in Photopharmacology with Molecular Photoswitches. *Angew Chem Int Ed Engl* **2023**, *62* (30), e202300681.
25. Fuchter, M. J., On the Promise of Photopharmacology Using Photoswitches: A Medicinal Chemist's Perspective. *Journal of Medicinal Chemistry* **2020**, *63* (20), 11436-11447.
26. Welleman, I. M.; Hoorens, M. W. H.; Feringa, B. L.; Boersma, H. H.; Szymański, W., Photoresponsive molecular tools for emerging applications of light in medicine. *Chemical Science* **2020**, *11* (43), 11672-11691.
27. Ricart-Ortega, M.; Font, J.; Llebaria, A., GPCR photopharmacology. *Molecular and Cellular Endocrinology* **2019**, *488*, 36-51.
28. Wijnmans, M.; Josimovic, I.; Vischer, H. F.; Leurs, R., Optical control of Class A G protein-coupled receptors with photoswitchable ligands. *Curr Opin Pharmacol* **2022**, *63*, 102192.
29. Donthamsetti, P. C.; Winter, N.; Schönberger, M.; Levitz, J.; Stanley, C.; Javitch, J. A.; Isacoff, E. Y.; Trauner, D., Optical Control of Dopamine Receptors Using a Photoswitchable Tethered Inverse Agonist. *Journal of the American Chemical Society* **2017**, *139* (51), 18522-18535.
30. Westphal, M. V.; Schafroth, M. A.; Sarott, R. C.; Imhof, M. A.; Bold, C. P.; Leippe, P.; Dhopeswarkar, A.; Grandner, J. M.; Katritch, V.; Mackie, K.; Trauner, D.; Carreira, E. M.; Frank, J. A., Synthesis of Photoswitchable Δ^9 -Tetrahydrocannabinol Derivatives Enables Optical Control of Cannabinoid Receptor 1 Signaling. *Journal of the American Chemical Society* **2017**, *139* (50), 18206-18212.
31. Rodriguez-Soacha, D. A.; Steinmuller, S. A. M.; Isbilir, A.; Fender, J.; Deventer, M. H.; Ramirez, Y. A.; Tutov, A.; Sotriffer, C.; Stove, C. P.; Lorenz, K.; Lohse, M. J.; Hislop, J. N.; Decker, M., Development of an Indole-Amide-Based Photoswitchable Cannabinoid Receptor Subtype 1 (CB(1)R) "Cis-On" Agonist. *ACS Chem Neurosci* **2022**, *13* (16), 2410-2435.

32. Banghart, Matthew R.; Sabatini, Bernardo L., Photoactivatable Neuropeptides for Spatiotemporally Precise Delivery of Opioids in Neural Tissue. *Neuron* **2012**, 73 (2), 249-259.
33. Schönberger, M.; Trauner, D., A Photochromic Agonist for μ -Opioid Receptors. *Angewandte Chemie International Edition* **2014**, 53 (12), 3264-3267.
34. Gomez-Santacana, X.; Panarello, S.; Rovira, X.; Llebaria, A., Photoswitchable allosteric modulators for metabotropic glutamate receptors. *Curr Opin Pharmacol* **2022**, 66, 102266.
35. Duran-Corbera, A.; Faria, M.; Ma, Y.; Prats, E.; Dias, A.; Catena, J.; Martinez, K. L.; Raldua, D.; Llebaria, A.; Rovira, X., A Photoswitchable Ligand Targeting the β_1 Adrenoceptor Enables Light Control of the Cardiac Rhythm**. *Angewandte Chemie International Edition* **2022**, 61 (30).
36. Bosma, R.; Dijon, N. C.; Zheng, Y.; Schihada, H.; Hauwert, N. J.; Shi, S.; Arimont, M.; Riemens, R.; Custers, H.; van de Stolpe, A.; Vischer, H. F.; Wijtman, M.; Holliday, N. D.; Kuster, D. W. D.; Leurs, R., Optical control of the beta(2)-adrenergic receptor with opto-prop-2: A cis-active azobenzene analog of propranolol. *iScience* **2022**, 25 (9), 104882.
37. Taura, J.; Nolen, E. G.; Cabré, G.; Hernando, J.; Squarcialupi, L.; López-Cano, M.; Jacobson, K. A.; Fernández-Dueñas, V.; Ciruela, F., Remote control of movement disorders using a photoactive adenosine A_{2A} receptor antagonist. *Journal of Controlled Release* **2018**, 283, 135-142.
38. Bahamonde, M. I.; Taura, J.; Paoletta, S.; Gakh, A. A.; Chakraborty, S.; Hernando, J.; Fernández-Dueñas, V.; Jacobson, K. A.; Gorostiza, P.; Ciruela, F., Photomodulation of G Protein-Coupled Adenosine Receptors by a Novel Light-Switchable Ligand. *Bioconjugate Chemistry* **2014**, 25 (10), 1847-1854.
39. Hüll, K.; Fernández-Dueñas, V.; Schönberger, M.; López-Cano, M.; Trauner, D.; Ciruela, F., Optical Control of Adenosine-Mediated Pain Modulation. *Bioconjugate Chemistry* **2021**, 32 (9), 1979-1983.
40. Hockemeyer, J.; Burbiel, J. C.; Müller, C. E., Multigram-Scale Syntheses, Stability, and Photoreactions of A_{2A} Adenosine Receptor Antagonists with 8-Styrylxanthine Structure: Potential Drugs for Parkinson's Disease. *The Journal of Organic Chemistry* **2004**, 69 (10), 3308-3318.
41. Nonaka, Y.; Shimada, J.; Nonaka, H.; Koike, N.; Aoki, N.; Kobayashi, H.; Kase, H.; Yamaguchi, K.; Suzuki, F., Photoisomerization of a potent and selective adenosine A₂ antagonist, (E)-1,3-dipropyl-8-(3,4-dimethoxystyryl)-7-methylxanthine. *Journal of Medicinal Chemistry* **1993**, 36 (23), 3731-3733.
42. Petzer, J. P.; Steyn, S.; Castagnoli, K. P.; Chen, J.-F.; Schwarzschild, M. A.; Van der Schyf, C. J.; Castagnoli, N., Inhibition of monoamine oxidase B by selective adenosine A_{2A} receptor antagonists. *Bioorganic & Medicinal Chemistry* **2003**, 11 (7), 1299-1310.
43. Meier, H., The Photochemistry of Stilbenoid Compounds and Their Role in Materials Technology. *Angewandte Chemie International Edition* **1992**, 31 (11), 1399-1420.
44. Vedernikov, A. I.; Kuz'mina, L. G.; Sazonov, S. K.; Lobova, N. A.; Loginov, P. S.; Churakov, A. V.; Strelenko, Y. A.; Howard, J. A. K.; Alfimov, M. V.; Gromov, S. P., Styryl dyes. Synthesis and study of the solid-state [2+2] autophotocycloaddition by NMR spectroscopy and X-ray diffraction. *Russian Chemical Bulletin* **2007**, 56 (9), 1860-1883.
45. Green, D. A. B.-A. a. B. S., The use of mid-points or average NMR chemical shifts in stereochemical assignments. *Tetrahedron* 21 January 1974, 1974, pp 2357-2364.
46. Odaka, H.; Arai, S.; Inoue, T.; Kitaguchi, T., Genetically-Encoded Yellow Fluorescent cAMP Indicator with an Expanded Dynamic Range for Dual-Color Imaging. *PLoS ONE* **2014**, 9 (6), e100252.
47. Gómez-Santacana, X.; Pittolo, S.; Rovira, X.; Lopez, M.; Zussy, C.; Dalton, J. A. R.; Faucherre, A.; Jopling, C.; Pin, J.-P.; Ciruela, F.; Goudet, C.; Giraldo, J.; Gorostiza, P.; Llebaria, A., Illuminating Phenylazopyridines To Photoswitch Metabotropic Glutamate Receptors: From the Flask to the Animals. *ACS Central Science* **2017**, 3 (1), 81-91.
48. Boehmler, W.; Petko, J.; Woll, M.; Frey, C.; Thisse, B.; Thisse, C.; Canfield, V. A.; Levenson, R., Identification of zebrafish A₂ adenosine receptors and expression in developing embryos. *Gene Expression Patterns* **2009**, 9 (3), 144-151.

49. Ohno, Y.; Suzuki, M.; Asada, H.; Kanda, T.; Saki, M.; Miyagi, H.; Yasunaga, M.; Suno, C.; Iwata, S.; Saito, J.-i.; Uchida, S., In Vitro Pharmacological Profile of KW-6356, a Novel Adenosine A_{2A} Receptor Antagonist/Inverse Agonist. *Molecular Pharmacology* **2023**, *103* (6), 311-324.
50. Santos, L. C.; Ruiz-Oliveira, J.; Silva, P. F.; Luchiari, A. C., Caffeine Dose-Response Relationship and Behavioral Screening in Zebrafish. In *The Question of Caffeine*, Latosinska, J. N.; Latosinska, M., Eds. InTech: 2017.
51. Rovira, X.; Trapero, A.; Pittolo, S.; Zussy, C.; Faucherre, A.; Jopling, C.; Giraldo, J.; Pin, J.-P.; Gorostiza, P.; Goudet, C.; Llebaria, A., OptoGluNAM4.1, a Photoswitchable Allosteric Antagonist for Real-Time Control of mGlu 4 Receptor Activity. *Cell Chemical Biology* **2016**, *23* (8), 929-934.
52. Ito, Y.; Kajita, T.; Kunimoto, K.; Matsuura, T., Accelerated photodimerization of stilbenes in methanol and water. *The Journal of Organic Chemistry* **1989**, *54* (3), 587-591.
53. Kirchner, S.; Leistner, A. L.; Godtel, P.; Seliwjorstow, A.; Weber, S.; Karcher, J.; Nieger, M.; Pianowski, Z., Hemipiperazines as peptide-derived molecular photoswitches with low-nanomolar cytotoxicity. *Nat Commun* **2022**, *13* (1), 6066.
54. Willems, S.; Morstein, J.; Hinnah, K.; Trauner, D.; Merk, D., A Photohormone for Light-Dependent Control of PPARalpha in Live Cells. *J Med Chem* **2021**, *64* (14), 10393-10402.
55. Gao, L.; Meiring, J. C. M.; Varady, A.; Ruider, I. E.; Heise, C.; Wranik, M.; Velasco, C. D.; Taylor, J. A.; Terni, B.; Weinert, T.; Standfuss, J.; Cabernard, C. C.; Llobet, A.; Steinmetz, M. O.; Bausch, A. R.; Distel, M.; Thorn-Seshold, J.; Akhmanova, A.; Thorn-Seshold, O., In Vivo Photocontrol of Microtubule Dynamics and Integrity, Migration and Mitosis, by the Potent GFP-Imaging-Compatible Photoswitchable Reagents SBTubA4P and SBTub2M. *J Am Chem Soc* **2022**, *144* (12), 5614-5628.
56. Schmidt, D.; Rodat, T.; Heintze, L.; Weber, J.; Horbert, R.; Girreser, U.; Raeker, T.; Bußmann, L.; Kriegs, M.; Hartke, B.; Peifer, C., Axitinib: A Photoswitchable Approved Tyrosine Kinase Inhibitor. *ChemMedChem* **2018**, *13* (22), 2415-2426.
57. Onoue, S.; Tsuda, Y., Analytical studies on the prediction of photosensitive/phototoxic potential of pharmaceutical substances. *Pharm Res* **2006**, *23* (1), 156-64.
58. Kim, W. B.; Shelley, A. J.; Novice, K.; Joo, J.; Lim, H. W.; Glassman, S. J., Drug-induced phototoxicity: A systematic review. *J Am Acad Dermatol* **2018**, *79* (6), 1069-1075.
59. El Ouardi, M.; Tamarit, L.; Vaya, I.; Miranda, M. A.; Andreu, I., Cellular photo(geno)toxicity of gefitinib after biotransformation. *Front Pharmacol* **2023**, *14*, 1208075.
60. Bhurta, D.; Bharate, S. B., Styryl Group, a Friend or Foe in Medicinal Chemistry. *ChemMedChem* **2022**, *17* (7).

Supplementary figures and tables

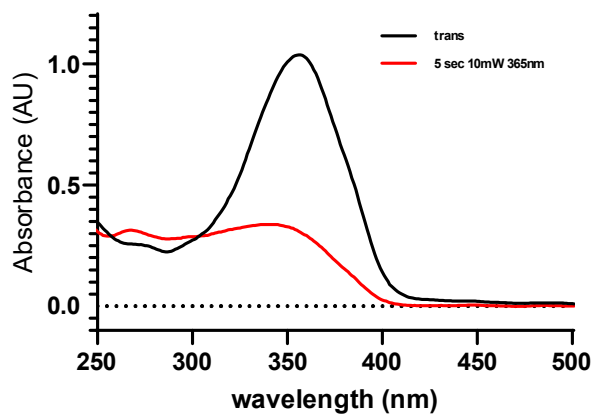


Figure S1: UV-visible absorption spectrum of istradefylline, 25 μ M in 80% PBS (20% DMSO) in the dark and after 5 seconds illumination with 365nm UV-light (10mW)

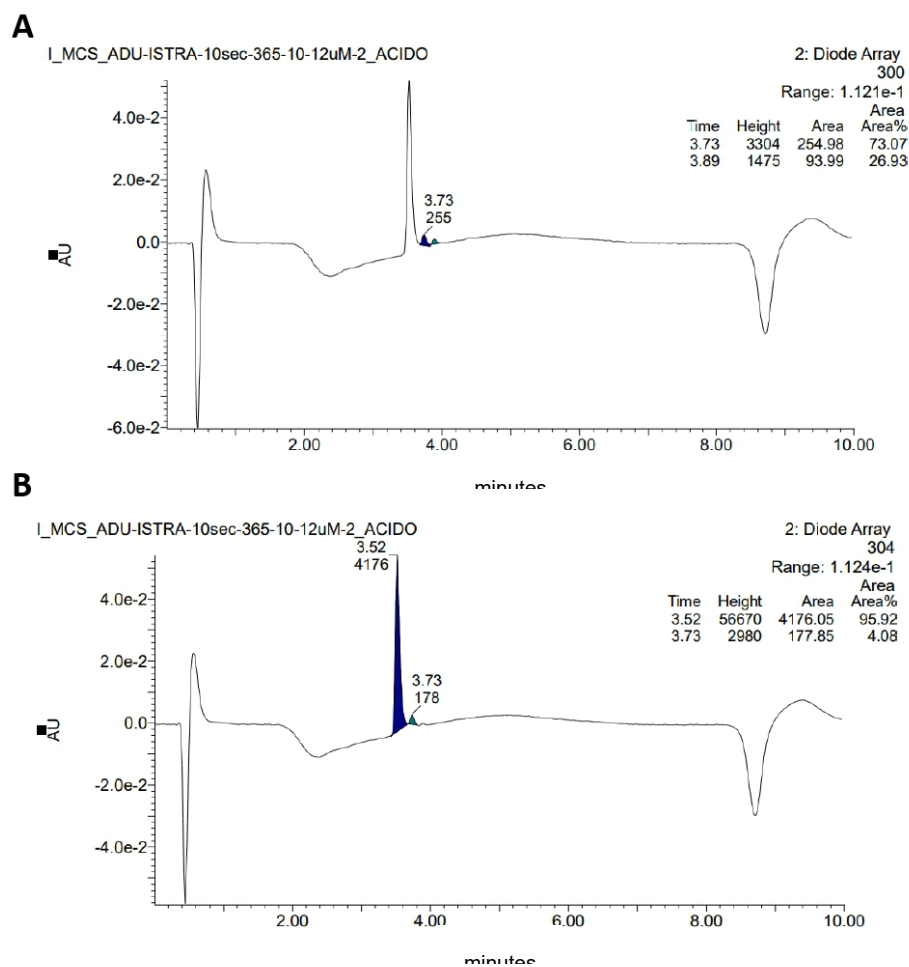
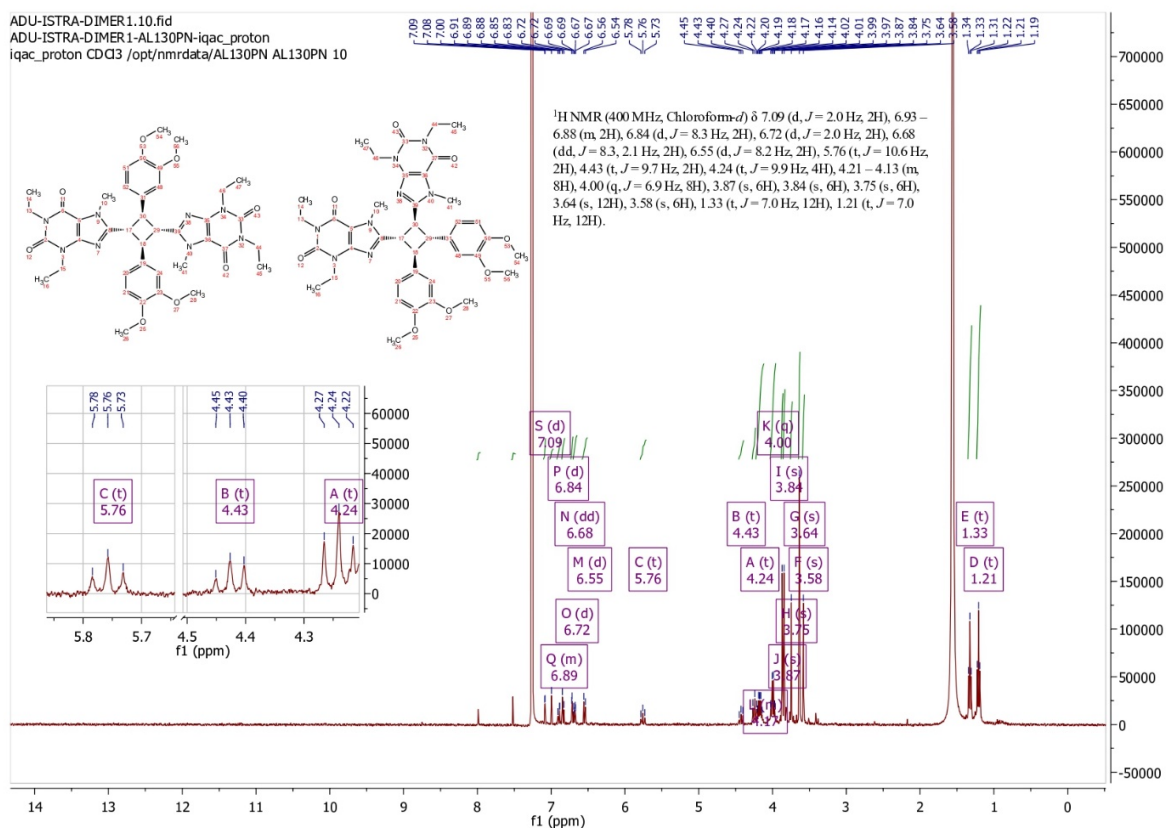
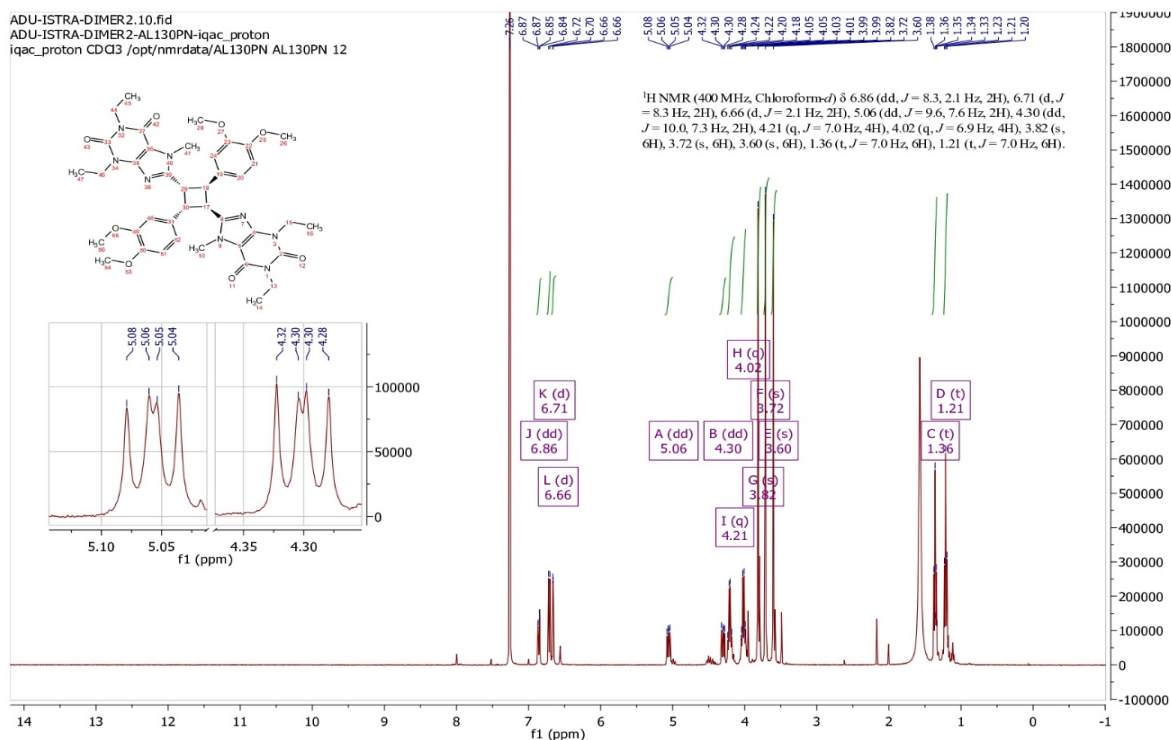


Figure S2: HPLC chromatograms of istradefylline, 25 μ M in PBS (20% DMSO) illuminated for 10 seconds with 365nm LED light (10mW), displayed at the isosbestic point of *trans*-istradefylline and the dimers (300nm) (A) and at the isosbestic point of *cis*- and *trans*-istradefylline isomers (304nm) (B) with the integration of the two species involved. The peak at t=3.52 min corresponds to *cis*-istradefylline, the peak at t=3.73 corresponds to *trans*-istradefylline and the peak at t=3.89 corresponds to the dimers.



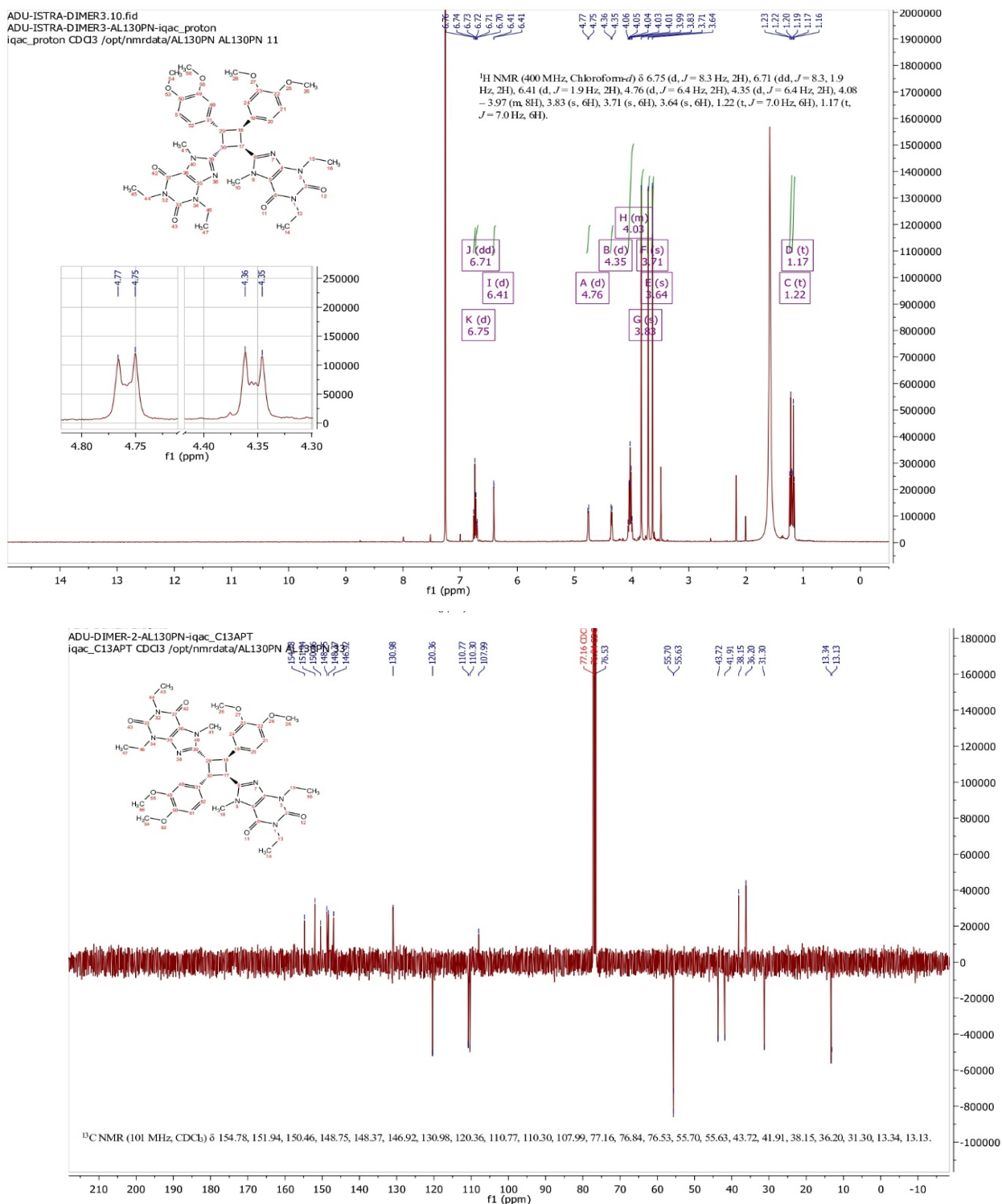


Figure S3: ¹H-NMR of dimers 1, dimer 2 and dimer 3 and ¹³C-NMR of dimer 2 and dimer 3

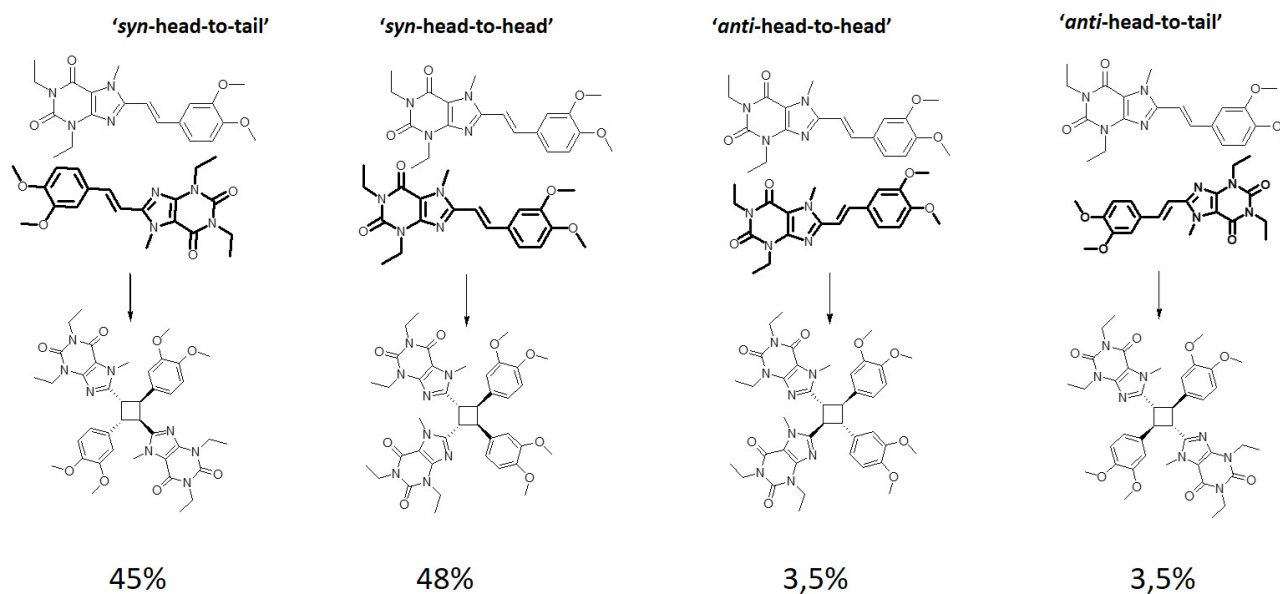
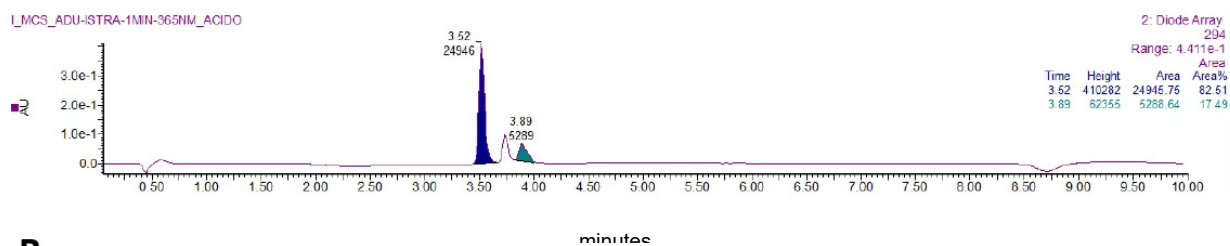
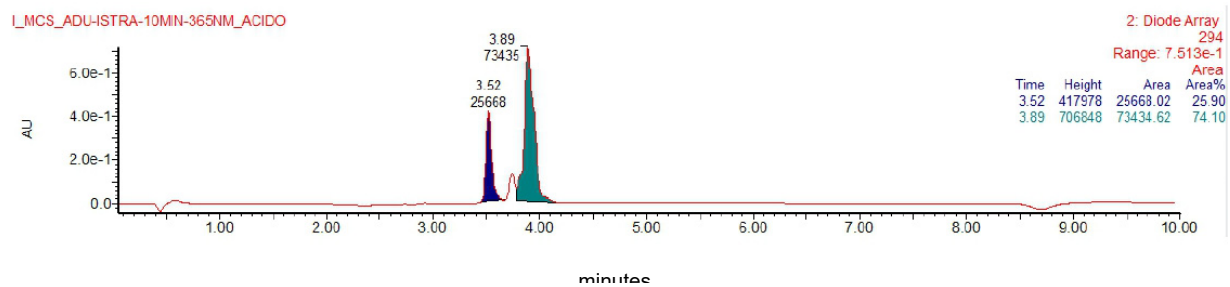


Figure S4: Preorganization of istradefylline for the auto-photocycloaddition reaction in solution and the stereochemistry of the products obtained.

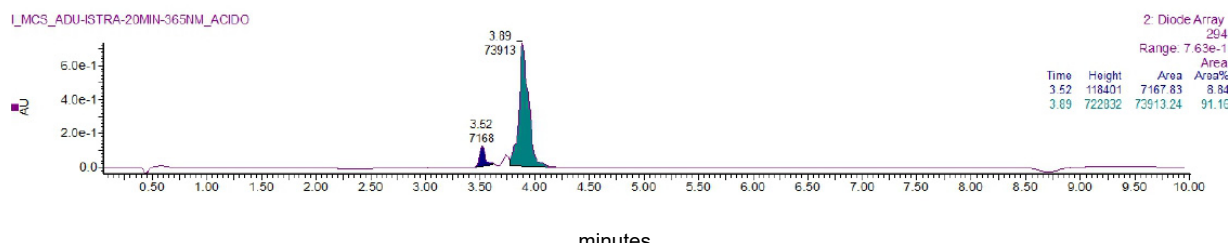
A



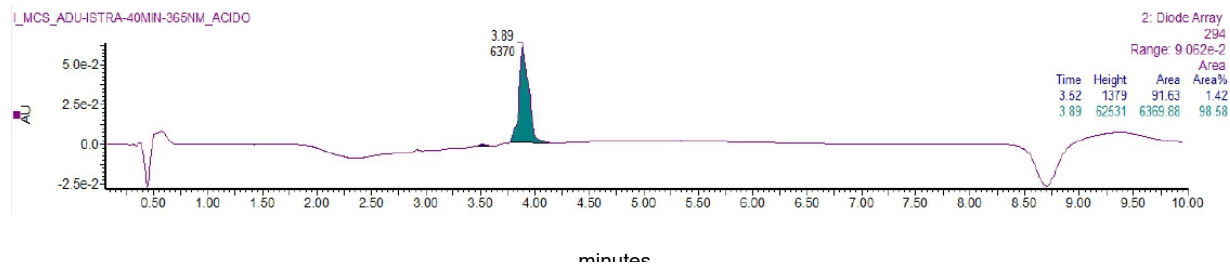
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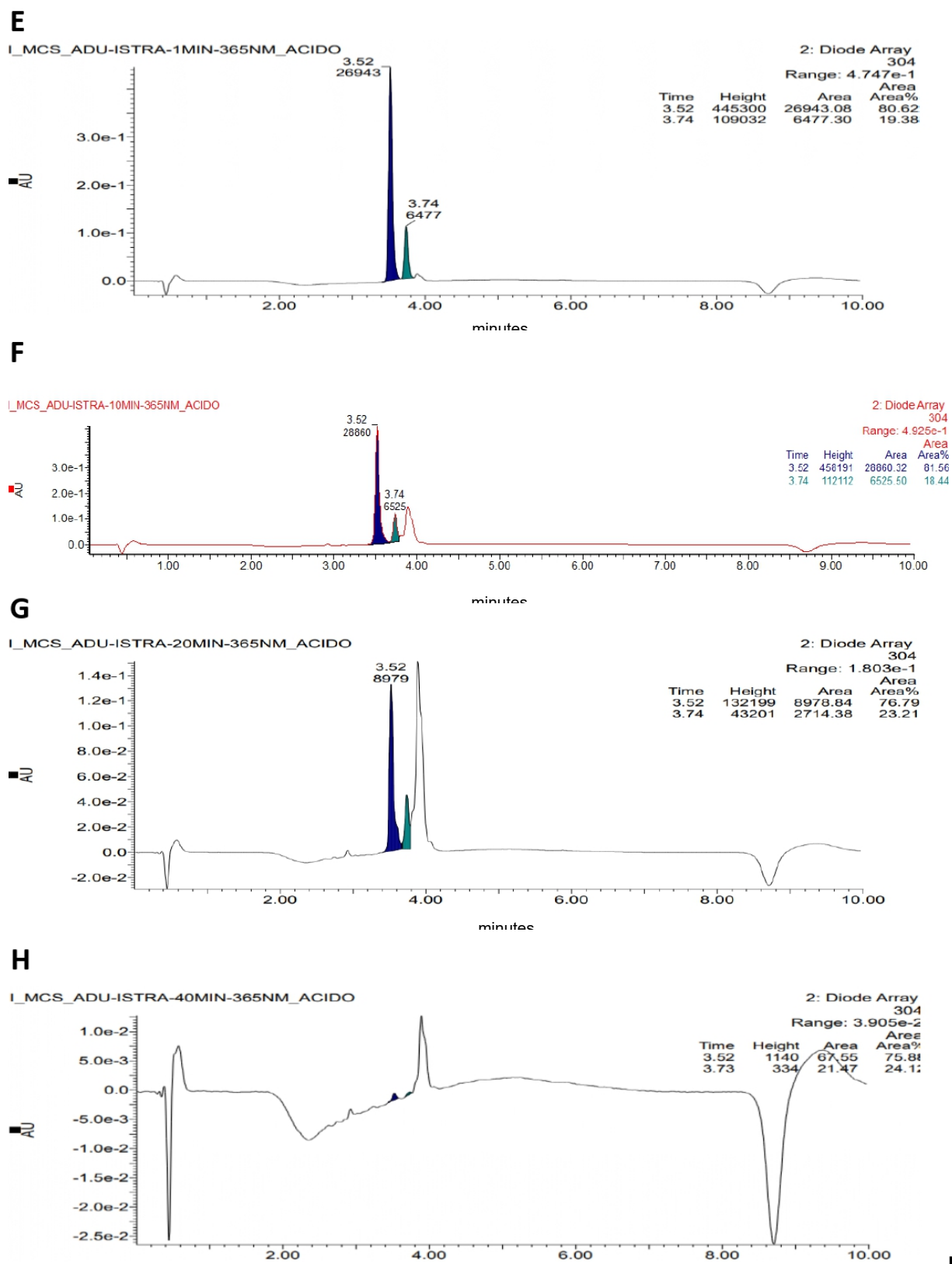


C



D





Figure

S5: HPLC chromatograms of the samples of istradefylline (2µL of 10mM in DMSO) illuminated with a 365nm

LED light (100mW) and displayed at the isosbestic point of the *cis* isomer and the dimers (294nm) (A-D) with an illumination time of 1min (A), 10min (B), 20min (C) and 40 min (D) and displayed at the isosbestic point of the *trans* and the *cis* istradefylline isomers (304nm) (E-H) with the same illumination times of 1min (E), 10min (F), 20min (I) and 40 min (J). The integration of the two species involved is displayed for each chromatogram. The peak at t=3.52 min corresponds to *cis*-istradefylline, the peak at t=3.73 corresponds to *trans*-istradefylline and the peak at t=3.89 corresponds to the dimers.

Illumination time	Cis (304nm)	Trans (304nm)	Cis (294nm)	Dimers (294nm)
1 min	82.5 %	17.5 %	80.6 %	19.4 %
10 min	25.9 %	74.1 %	81.6 %	18.4 %
20 min	8.8 %	91.2 %	76.8 %	23.2 %
40 min	1.4 %	98.6 %	75.8 %	24.1 %

Table S1: Summary table with the relative percentage of *cis* and *trans* istradefylline isomers (in blue) and the relative percentage *cis* isomer and dimers (in violet) of 2 μ L samples of 10mM istradefylline illuminated for 1, 10, 20 and 40min with 365nm light set at 100mW, obtained from the HPLC chromatograms displayed in Figure S5.

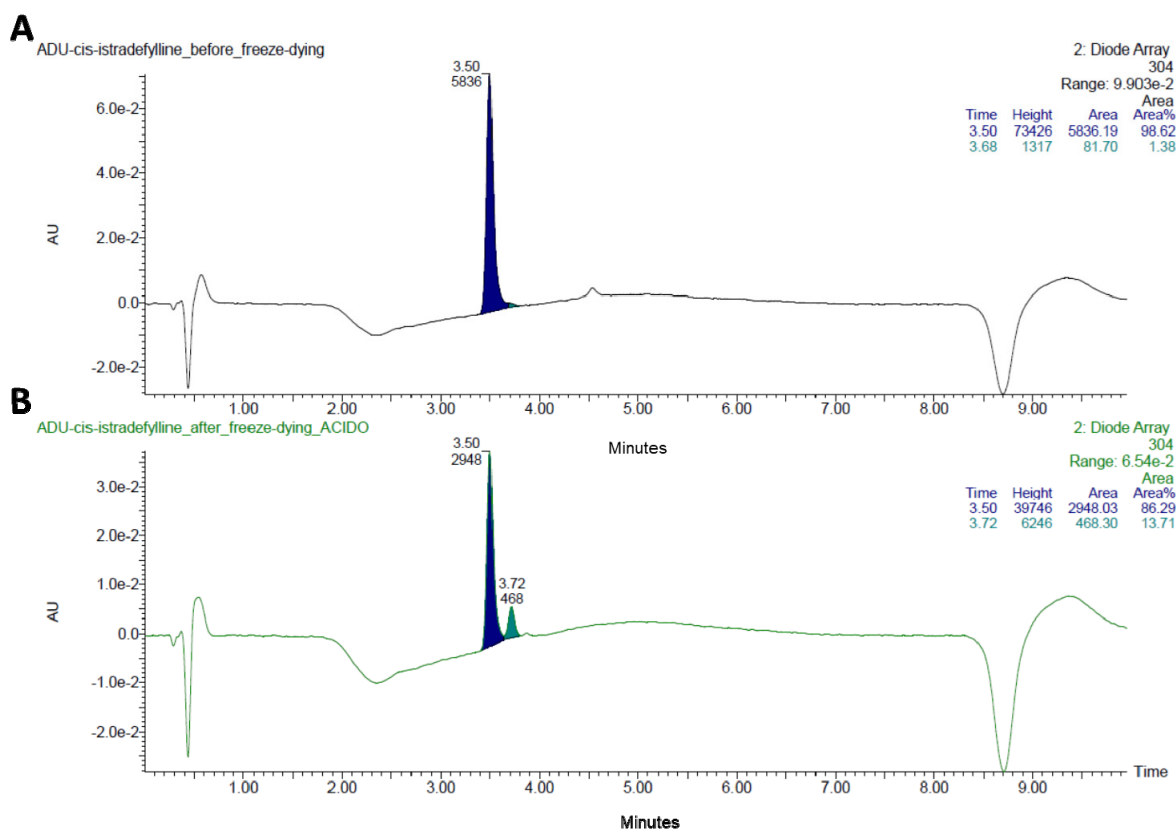


Figure S6: HPLC chromatograms of the fraction corresponding to the *cis*-istradefylline obtained by preparative HPLC before solvent removal by freeze-drying (A) and after solvent removal by freeze-drying (B). The chromatograms are displayed at the isobestic point of the *trans* and the *cis* istradefylline isomers (304nm) and the peak at t=3.50 min corresponds to *cis*-istradefylline and the peak at t=3.72 corresponds to *trans*-istradefylline. These results show an increase of the *trans*-isomer proportion from a 1.4% to 13.7% during the mild isolation process through evaporation of acetonitrile under reduced pressure in the dark and with no heating followed by freeze drying. This demonstrates that the *cis*-istradefylline displays a low bistability and relaxes thermally to the *trans*-isomer.

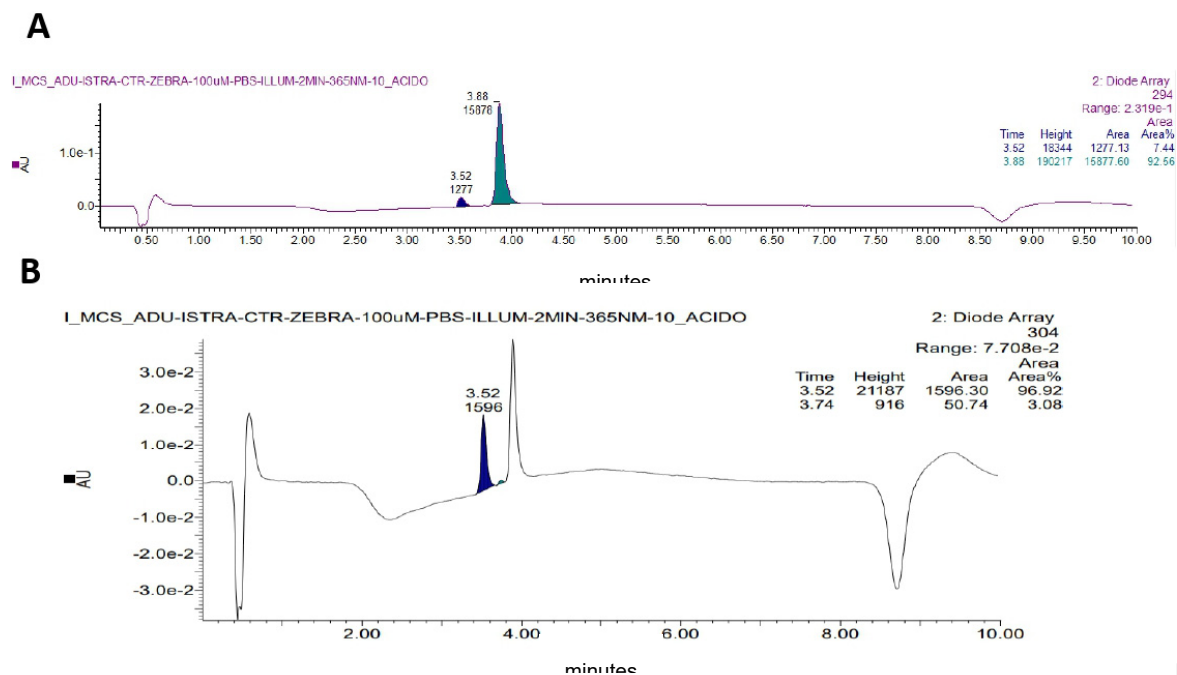
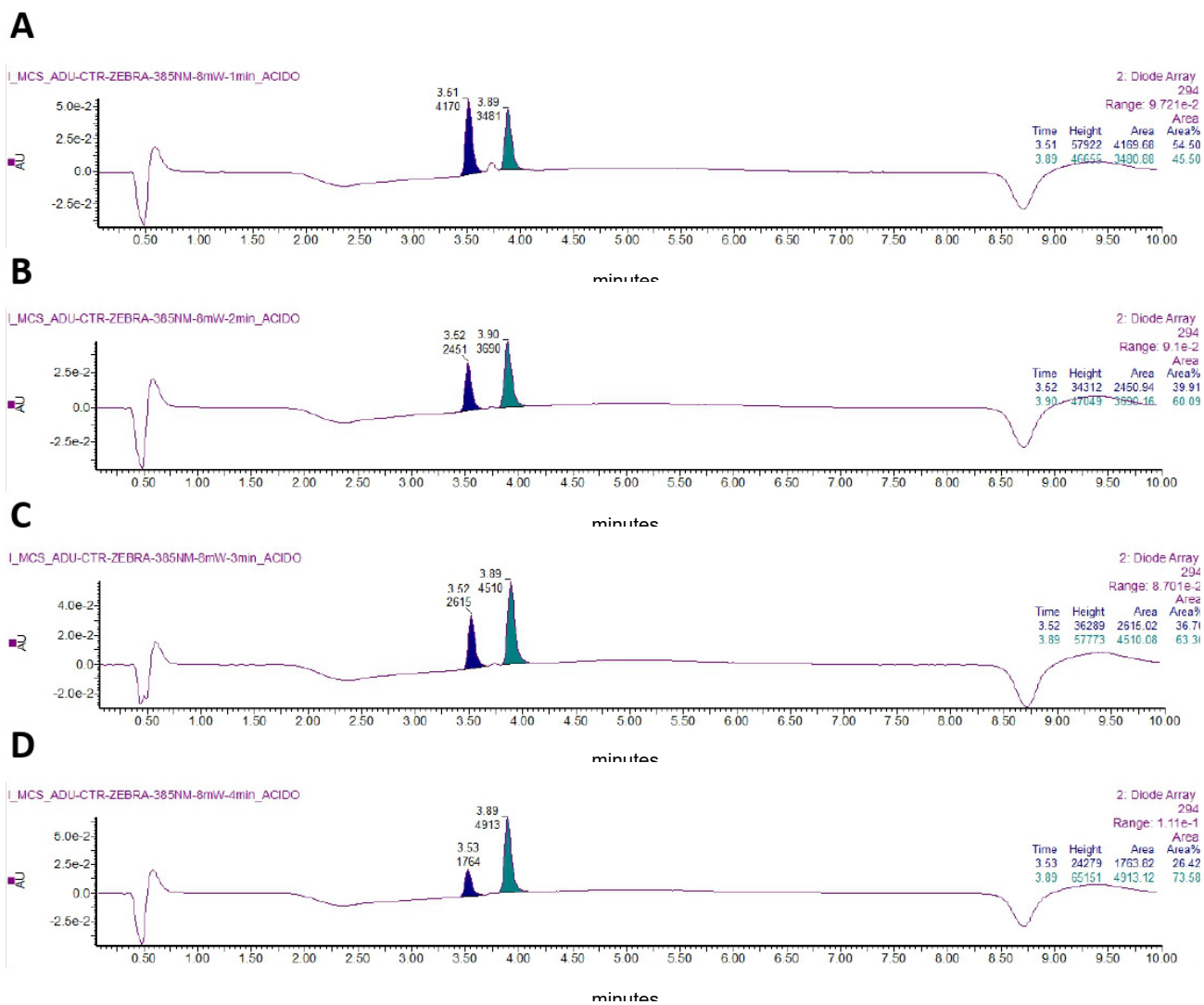


Figure S7:

HPLC chromatograms of a sample of istradefylline (100 μ M in PBS) illuminated for 2 min with 365nm LED light (10mW), displayed at the isosbestic point of the *cis* isomer and the dimers (294nm) (A) or at the isosbestic point of the *trans* and the *cis* istradefylline isomer (304nm) (B) with the integration of the two species involved. This condition corresponds to the one used for the sample of istradefylline pre-illuminated in Figure 4. The peak at t=3.52 min corresponds to *cis*-istradefylline, the peak at t=3.73 corresponds to *trans*-istradefylline and the peak at t=3.89 corresponds to the dimers.



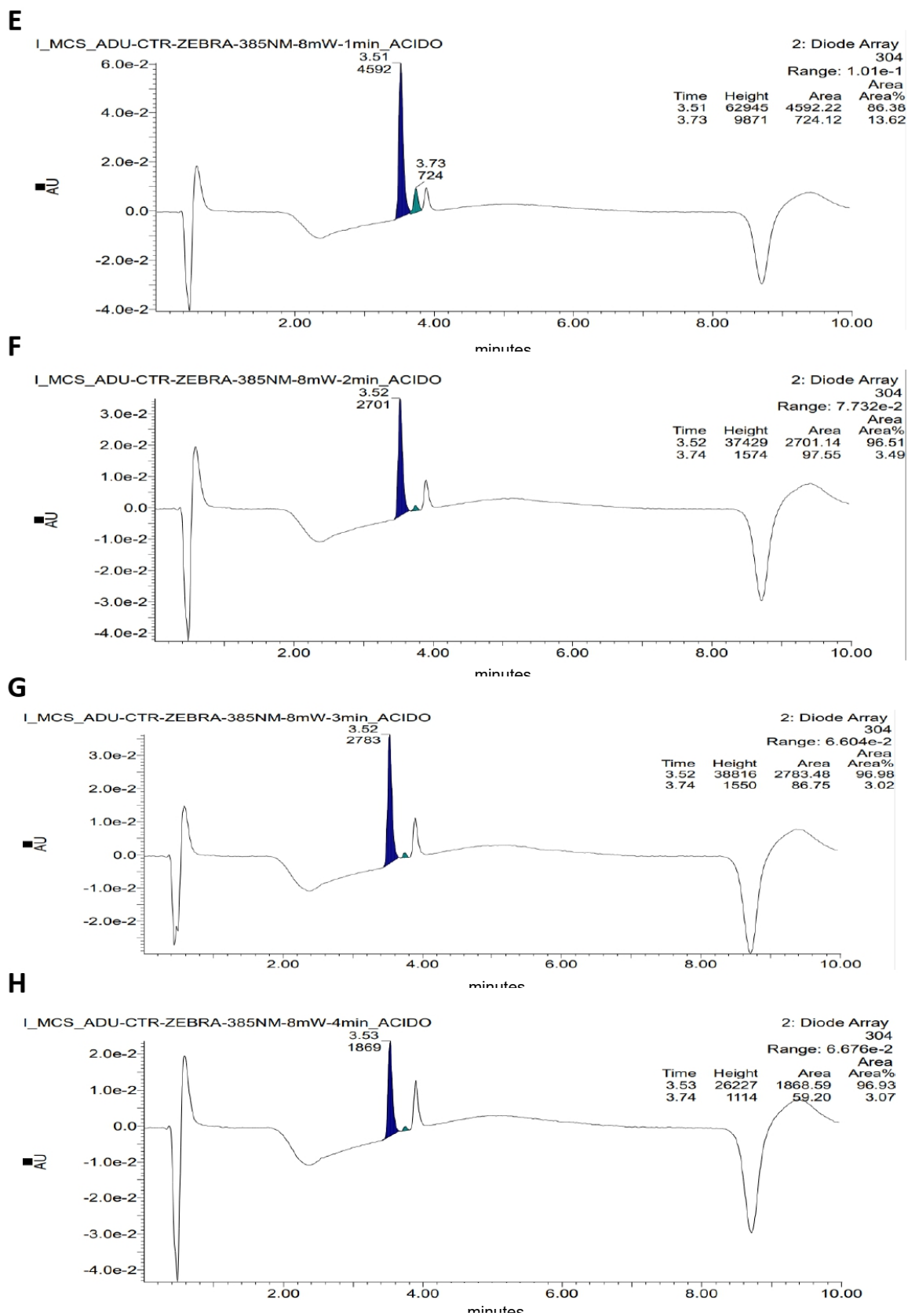


Figure S8:

HPLC chromatograms of a sample of istradefylline (50 μ M in PBS) illuminated with a 385 nm LED light (8 mW), displayed at the isosbestic point of the *cis* isomer and the dimers (294nm) (A-D) in PBS with an illumination of 1min (A), 2min (B), 3min (C) or 4min (D) and at the isosbestic point of the *trans* and the *cis* istradefylline isomer (304nm) (E-H) with an illumination of 1min (E), 2min (F), 3min (G) or 4min (H) with the integration of the two species involved. These conditions correspond to the same conditions used in the four first cycles of zebrafish experiment Figure 4B. The peak at t=3.52 min corresponds to *cis*-istradefylline, the peak at t=3.73 corresponds to *trans*-istradefylline and the peak at t=3.89 corresponds to the dimers.

Sample	<i>Cis</i> (304nm)	<i>Trans</i> (304nm)	<i>Cis</i> (294nm)	Dimers (294nm)
2 min pre-illum	96.9 %	3.1 %	7.4 %	92.6 %
1 st cycle	86.4 %	13.6 %	54.5 %	45.5 %
2 nd cycle	96.5 %	3.5 %	39.9 %	60.1 %
3 rd cycle	97.0 %	3.0 %	36.7 %	63.3 %
4 th cycle	96.9 %	3.1 %	26.4 %	73.6 %

Table S2: Summary table with the relative percentage of *cis* and *trans* istradefylline isomers (in blue) and the relative percentage of *cis* isomer and dimers (in violet) of samples illuminated in the same conditions than for during the zebrafish experiments from Figure 4, obtained from the chromatograms shown in Figure S7 and Figure S8.

Sample	Trans	Cis	Dimers
Pre-illum. 2min 365nm 10 mW	1 %	24 %	75 %
1 st cycle	10 %	60 %	30 %
2 nd cycle	2 %	56 %	42 %
3 rd cycle	2 %	49 %	49 %
4 th cycle	1 %	39 %	60 %

Table S3: Percentage of *cis* and *trans* istradefylline and dimers obtained from the analysis of the HPLC chromatograms (Figure S6 and Figure S7) of samples illuminated in the same conditions than the ones used for the pre-illumination of the istradefylline sample from Figure 4B-D and the first 4 cycles of illumination inside the zebrafish chamber from Figure 4B.