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# Numerical simulation of cardiac pacing devices: modeling and validation issues

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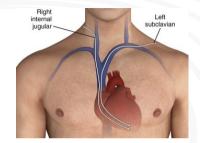


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#### Pacemakers and electrical questions



(a) Diverse pacemaker leads



(b) Focus: bradychardia leads

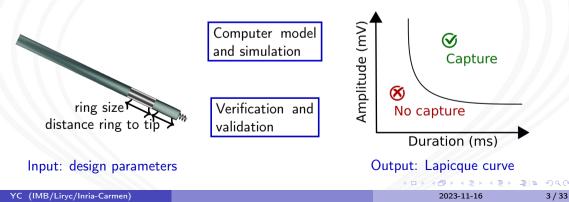
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Intervention	Lifetime of the device	10-20 years later			
Insertion, navigation to target, deformation, constraints ?	Energy to pace the heart ? Sensing evoked potentials ?	Rupture due to fatigue ?			
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#### Question of interest

What are the stimulation pulse characteristics (voltage amplitude in V and pulse duration in ms) required for a bradychardia lead in bipolar (tip/ring) mode to capture (stimulate) healthy cardiac tissue ?

Context: threshold test in animal experiments. Influence of the design of the lead, like geometry and contact properties ?



## Web-based computational platform

Ideal objective: run trials on a web based platform, and obtain credible results



## InSilicoTrials

Credible according to VV40<sup>1</sup> : Qol, Cou, Verification, Validation...

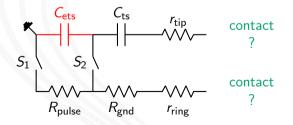
<sup>1</sup>40-2018, Assessing Credibility of Computational Modeling through Verification and Validation: Application to Medical Devices.

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2023-11-16 4 / 33

- **O Pacing**: for duration T (ms), with charge Q (eq. V)
- Interruption (switches): 122µs
- **③** Output Capacitor Discharge (OCD):  $\simeq 13$ ms

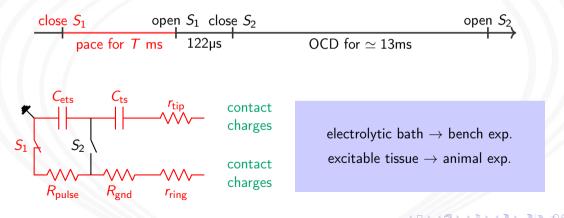




electrolytic bath  $\rightarrow$  bench exp. excitable tissue  $\rightarrow$  animal exp.

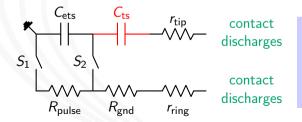
5/33

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- **O Pacing**: for duration T (ms), with charge Q (eq. V)
- Interruption (switches): 122µs
- **3** Output Capacitor Discharge (OCD):  $\simeq 13$ ms



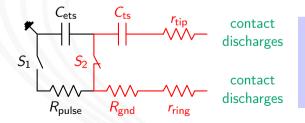


electrolytic bath  $\rightarrow$  bench exp. excitable tissue  $\rightarrow$  animal exp.

5/33

- **O Pacing**: for duration T (ms), with charge Q (eq. V)
- Interruption (switches): 122µs
- **Output Capacitor Discharge (OCD)**:  $\simeq$  13ms

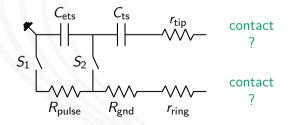




electrolytic bath  $\rightarrow$  bench exp. excitable tissue  $\rightarrow$  animal exp.

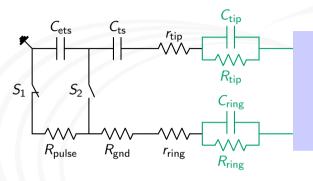
- **O Pacing**: for duration T (ms), with charge Q (eq. V)
- Interruption (switches): 122µs
- **3** Output Capacitor Discharge (OCD):  $\simeq 13$ ms





electrolytic bath  $\rightarrow$  bench exp. excitable tissue  $\rightarrow$  animal exp.

## Contact impedance model



electrolytic bath  $\rightarrow$  bench exp. excitable tissue  $\rightarrow$  animal exp.

6/33

- At pacing, creation of a "double layer" impedance
- Boundary layer related to the Debye length
- Literature is huge<sup>2</sup>

#### Parallel R-C equivalent circuit

<sup>2</sup>Grimnes and Martinsen, *Bioimpedance and Bioelectricity Basics – Third Edition.* 

## Bath experiment / model

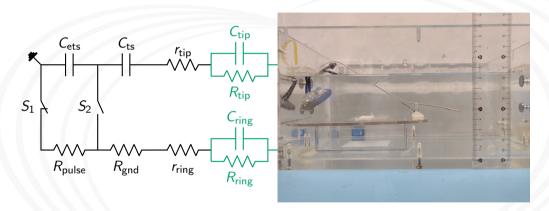


Figure: Passive bath  $\rightarrow$  def of contact model and parameter identification

## Bath experiment / model

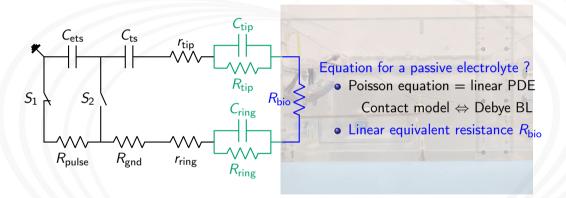
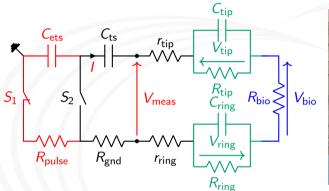
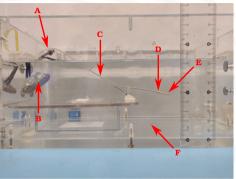


Figure: Passive bath  $\rightarrow$  def of contact model and parameter identification

#### The passive bath model



$$\begin{split} R_{\rm tot} &= R_{\rm pulse} + R_{\rm gnd} + r_{\rm ring} + r_{\rm tip} + R_{\rm bio} \\ \frac{1}{C_{\rm tot}} &= \frac{1}{C_{\rm ets}} + \frac{1}{C_{\rm ts}} + \frac{1}{C_{\rm tip}} + \frac{1}{C_{\rm ring}} \end{split}$$



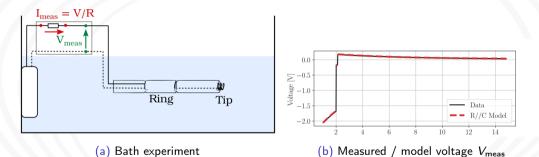
$$\frac{dI}{dt} + \frac{1}{\tau}I = \frac{1}{R_{\text{tot}}} \left(\frac{V_{\text{tip}}}{\tau_{\text{tip}}} + \frac{V_{\text{ring}}}{\tau_{\text{ring}}}\right)$$
$$\frac{dV_i}{dt} + \frac{V_i}{\tau_i} = \frac{I}{C_i}, \quad i \in \{\text{tip}, \text{ring}\}$$

8/33

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#### The passive bath experiments

- MICROPORT CRM BOREA pacemaker + VEGA leads
- Saline solution measured at  $\sigma = 2.85 \mathrm{mS/cm}$



• 9 datasets:  $(0.25, 0.5, 1) \operatorname{ms} \times (1, 2, 4) \operatorname{V}$ 

#### Method to calibrate the contact parameters

• Bath equivalent resistance computed from the initial data ( $V_{tip} = V_{ring} = 0$ ):

$$R_{ ext{bio}} = rac{V^{\star}_{ ext{meas}}(t=0)}{I^{\star}(t=0)} - r_{ ext{tip}} - r_{ ext{ring}}$$

• 4 parameters (device components are known):

$$\theta = (R_{tip}, R_{ring}, \tau_{tip} := R_{tip}C_{tip}, \tau_{ring} := R_{tip}C_{tip})$$

• Cost function weighted by step  $s \in \{$ pulse, interruption, OCD $\}$ 

$$J(\theta) = \sum_{s} \omega_{s} \mathbb{1}_{s}(t) \left( \frac{|V_{\mathsf{meas}}(\theta) - V_{\mathsf{meas}}^{\star}|_{\ell^{2}}}{|V_{\mathsf{meas}}^{\star}|_{\ell^{2}}} + \frac{|I(\theta) - I^{\star}|_{\ell^{2}}}{|I^{\star}|_{\ell^{2}}} \right), \quad \sum_{s} \omega_{s} = 1$$

- L-BFGS-B from Python Scipy, restricted to a subset of  $\mathbb{R}^4$ 
  - ODE solved semi-analytically (i.e. no approx. error)
  - ▶ First step in Log scale,  $R \in (1, 1 \times 10^5)$ ,  $\tau \in (1 \times 10^{-6}, 1)$
- 1 optimisation for each dataset  $\rightarrow$  9 parameter sets

#### Calibration results - contact parameters

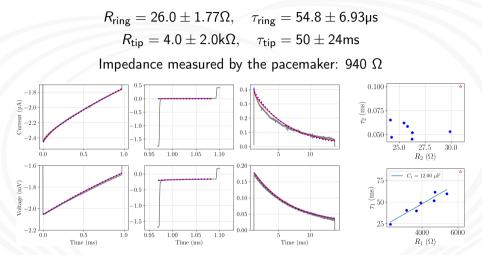


Figure: Current and voltage in each step, measures and model (left), and parameters (right)

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#### Tissue experiment / model

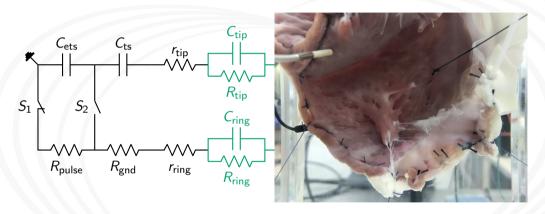


Figure: Animal experiment  $\rightarrow$  validation data

2023-11-16 12/33

#### Tissue experiment / model

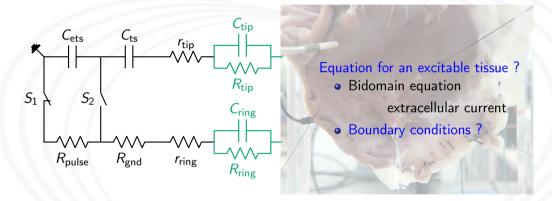
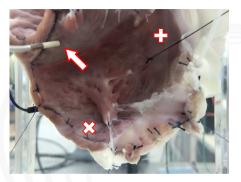


Figure: Animal experiment  $\rightarrow$  validation data

2023-11-16 12/33

#### Thresehold detection tests in animal experiments



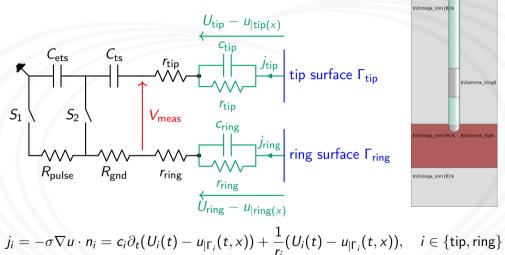
• Ventricular wedge preparation, 2 sheeps (now 3)

- 3 sites: apex (X), septum ( $\rightarrow$ ), base (+)
- screw fully deployed, perpendicular
- Recording of  $V^{\star}_{\mathrm{meas}}$ , like in the bath
- Recording of distant EGMs with needle electrodes
- Optical signals + 9.4T MRI

Automatic threshold search by the pacemaker:

- For a fixed duration T (ms) 5 pulses at 1.5 Hz
- $\bullet$  Decrease voltage to locate the interval : last capturing  $\rightarrow$  first non capturing
- Capture is decided by the pacemaker, and controlled through EGMs

#### Bidomain coupled to the pacemaker – CEM<sup>3</sup>



<sup>&</sup>lt;sup>3</sup>Somersalo, Cheney, and Isaacson, "Existence and Uniqueness for Electrode Models for Electric Current Computed Tomography".

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2023-11-16	14 /	33
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## Complete system of equations

$$\tau_{\text{tip}} \frac{dU_{\text{ring}}}{dt} + U_{\text{ring}} = \tau \frac{dU_{\text{tip}}}{dt}, \qquad -\operatorname{div} (\sigma_i \nabla u_i) = -c_m \partial_t v_m - l_{\text{ion}}(v_m, h) \quad \text{in } \Omega_M$$
  

$$\tau_{\text{tip}} = r_{\text{tip}} C \ll \tau = RC, \qquad -\operatorname{div} (\sigma \nabla u) = \begin{cases} c_m \partial_t v_m + l_{\text{ion}}(v_m, h) & \text{in } \Omega_M \\ 0 & \text{in } \Omega_B \end{cases}$$
  

$$\frac{1}{C} = \frac{1}{C_{\text{ets}}} + \frac{1}{cts} \qquad \partial_t h + g_{\text{ion}}(v_m, h) = 0 \quad \text{in } \Omega_M$$

With the CEM transmission conditions

$$j_i = -\sigma 
abla u \cdot n_i = c_i \partial_t (U_i - u_{|\Gamma_i}) + rac{1}{r_i} (U_i - u_{|\Gamma_i}), \quad i \in \{ ext{tip}, ext{ring}\}$$

#### Energy estimate

• There is an inequality if energy, which states the energy associated to each (equivalent) dipole in the overall electrical circuit

For test functions  $w_i(x), w(x) \in H^1$ , and  $W_{tip}, W_{ring} \in \mathbb{R}$ ,

$$\begin{split} \int_{\Omega_M} \sigma_i \nabla u_i \cdot \nabla w_i &+ \int_{\Omega} \sigma \nabla u \cdot \nabla w + \int_{\Omega_M} \left( c_m \partial_t v_m + I_{\text{ion}} \right) w_m \\ &+ \sum_{k \in \{ \text{tip,ring} \}} \int_{\Gamma_k} \left( c_k \partial_t (U_k - u_{|\Gamma_k}) + \frac{1}{r_k} (U_k - u_{|\Gamma_k}) \right) (W_i - w_{|\Gamma_i}) \\ &+ C \frac{dU_{\text{tip}}}{dt} W_{\text{tip}} + \frac{1}{R} U_{\text{ring}} W_{\text{ring}} = 0 \end{split}$$

•  $\rightarrow$  adapt the standard<sup>4</sup> existence proof (article in preparation), and error estimate for FEM

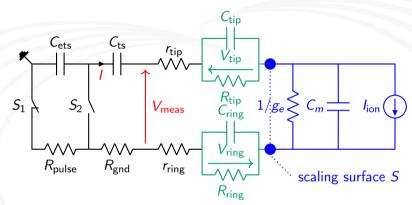
<sup>4</sup>Bourgault, Coudière, and Pierre, "Existence and uniqueness of the solution for the bidomain model used in cardiac electrophysiology".

16 / 33

2023-11-16

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## A surrogate 0D model



We need to rescale the current per unit of surface (ionic) to a current:

$$I = S\left(C_m \frac{dV_m}{dt} + I_{\rm ion} + g_e V_m\right)$$

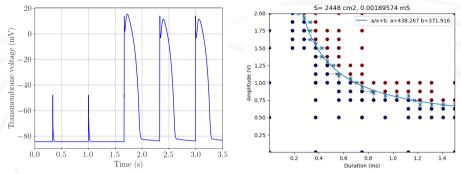
#### Surrogate 0D model ODEs

$$\frac{dI}{dt} + \frac{1}{\tau}I = \frac{1}{R_{\text{tot}}} \left(\frac{V_{\text{tip}}}{\tau_{\text{tip}}} + \frac{V_{\text{ring}}}{\tau_{\text{ring}}} + \frac{I_{\text{ion}}(V_m, h)}{C_m} + \frac{V_m}{\tau_m}\right)$$
(1)  
$$\frac{dV_i}{dt} + \frac{V_i}{\tau_i} = \frac{I}{C_i}, \quad i \in \{\text{tip}, \text{ring}\}$$
(2)  
$$\frac{dV_m}{dt} + \frac{I_{\text{ion}}(V_m, h)}{C_m} + \frac{V_m}{\tau_m} = \frac{I}{SC_m},$$
(3)  
$$\frac{dh}{dt} + g(V_m, h) = 0,$$
(4)

• How to choose S and  $g_e$ ?

• Our starting choice:  $S = 100 {
m cm}^2$  and  $g_e = 10^{-2} {
m mS}$ , obtained from a volume of 1mm<sup>3</sup>

#### Threshold detection in the model



- Capture := more than 50% successful captures, seen on  $V_m$
- a 2D dichotomy algorithm  $\rightarrow$  intervals  $(V_{\min}(T), V_{\max}(T))$
- Rheobase  $V_{\star}$  (0.372 V) and chronaxie  $T_{\star}$  (1.18 ms) from the Lapicque law (LS fitting):

$$V_{\text{Lapicque}}(T) := V_{\star}\left(1 + \frac{T_{\star}}{T}\right), \text{ an minimize } \left|V_{\text{Lapicque}}(t) - \frac{1}{2}\left(V_{\min}(t) + V_{\max}(t)\right)\right|_{\ell^{2}}^{2}$$

19/33

## First comparisons – experiments and 0D model, see<sup>5</sup>

- Beeler-Reuter ionic model, with parameters from CellML
- Contact parameters fixed from the bath experiments

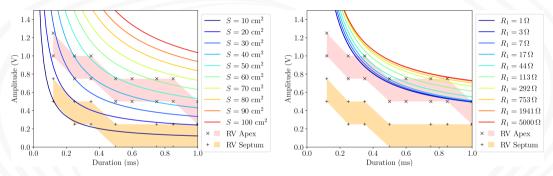


Figure: Comparison of Lapicque curves (solid lines) computed with different values of S (left) and  $R_{tip}$  (right). The shaded regions indicate where the experimental capture threshold is located.

315

<sup>&</sup>lt;sup>5</sup>Pannetier, Leguèbe, Coudière, Walton, Feuerstein, and Amaro, "Modeling cardiac stimulation by a pacemaker, with accurate tissue-electrode interface".

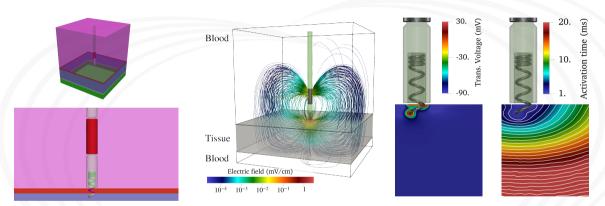
#### Remarks

• Pacemakers have very sparse covering of the Lapicque plane, but we model the pacemaker

 $T \in \{0.12, 0.25, 0.35, 0.5, 0.75, 0.85, 1.00\}$ ms,  $V \in \{2.0, 1.75, 1.5, 1.0, 0.75, 0.5, 0.25\}$ V

- Lapicque law fits well the model
- Lapicque law do not fit within the experimental regions
- (Too ?) many modeling simplification ?
  - ▶ 0D 1 cell
  - $g_e$  and S: influence of  $g_e \ll$  influence of S
  - Contact properties in bath versus tissue ? Influence of  $R_{tip}$
  - BR model with CellML std parameters
- What about the V&V40 guidelines
- Fast computations, OK for prototyping ideas / computational procedures

## Some preliminary 3D results I



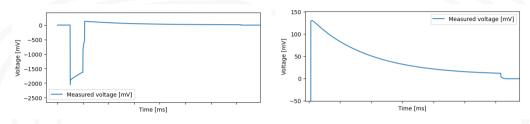
Mesh resolution 6µm - 1.5mm, time-step 10µs, P1-Lagrange FEM, BDF2+RL2, BiCGStab

Supra-threshold pacing, T = 1ms and V = 2.5V

Standard CellML BR model + electrical conductivity coefficients

22/33

#### Some preliminary 3D results II



(a) Simulated voltage  $V_{meas}$ , capture case

(b) voltage during OCD, capture case

Figure: We obtain the expected behavior for  $V_{meas}$ 

#### The V&V40 framework

Next: how to make it useful and credible ?

Qol: What is the energy level needed to capture a heart beat ?

CoU: As it would be reported by threshold detection procedures during animal experiments

		Decision consequence		
		low	medium	high
	low	1	2	3
influence	medium	2 COU	3	4
Model	high	3	4	5

Figure 2: Model Risk Matrix (cf. ASME VV40) evaluating the COU included in UC1.

(a) Risk matrix foll. ASME VV40

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Tochnical Report D 6.2, VALIDATION OF THE INSERTIONS FOR THE LISE CASES OF 1875.6 Ward Pockaps (1876) VERIFICATION, VALIDATION (CENTRICATION & CERTIFICATION

Task Lead: MPC, France WP Lead: MPC, France

(b) V: 231 pages

(c) V: 151 pages

#### Work on verification

Table 6: Verification Credibility Factors Coverage Level for Use Case 1 (cf. ASME VV40); \* indicates verification activities not yet completed.

Model Ris	k		х			
Credibility Factor Coverage Leve	el	1	2	3	4	Ę
Code Verification: Software Quality Assurance	V		Х			
Code Verification: Numerical Code Verification - NCV	V		X			
Calculation Verification - Discretization Error *	V		X			
Calculation Verification - Numerical Solver Error *	V		X			
Calculation Verification - Use Error	IV		х			

SQA: software code CEPS, thorough CI, SonarQube analysis, ...

NCV, discretization error, solver error: that is numerical analysis.

We provide many automatic tests (interpolation, quadrature, ODE solvers) and convergence tests on PDEs (Laplace, heat, reaction-diffusion) w/r to manufactured and reference solutions

Use error: on the web-based platform + within SQA

We cover credibility up to 4 or 5, while 2 is needed (risk analysis).

25 / 33

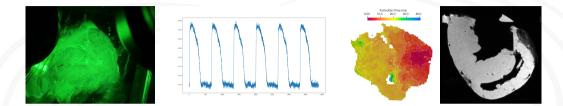
#### On-going work on validation

Table 6 : Validation and Applicability Credibility Factors Coverage Level for Use Case 1 (cf. ASME VV40); \* indicates validation activities not yet completed; N/A indicates activities which requires completion to be evaluated.

Model Risk				<			
Credibility Factor Coverage Level			1 2	2	3	4	5
Validation - Model [Form]	- 111		)	<			
Validation - Model [Inputs] *	- 111			<			
Validation - Comparator [Test Samples] *	IV			<			
Validation - Comparator [Test Conditions] *	11		)	<			
Validation - Assessment [Input Parameters] *	- 111		)	<			
Validation - Assessment [Output Comparison]	N.A.	Г	)	<			
Applicability: Relevance of the Quantities of Interest *	V		)	<			
Applicability: Relevance of the Validation Activities to the COU *	V		)	<			

- To be done in view of the QoI and CoU
- For UC1 (Simcardiotest), we could design and run our own experiments

## Path to calibration, before validation



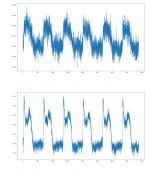
Calibrate model's parameters

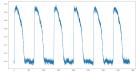
- Contact parameters → done (bath experiments)
- Ionic parameters  $\rightarrow$  optical signal, pixel-wise, done
- Conductivity coefficients  $\rightarrow$  9.4T MRI (300 µm) + activation maps, partially done

Compare to experiments for validation

• Lapicque curves are within the experiment Lapicque regions ? Computational challenge

#### Ionic parameters for the BR model





- BR has 2 parameters for the Na<sup>+</sup> current:  $g_{Na}$  and  $g_{Na,c}$
- Optimisation technique developped in<sup>a</sup>: compass search algorithm
- Automatic search in each pixel of each camera (2 camera)
- Using 6 consecutive pacings, beginning of the experiment
- Data has to be normalized (either baseline only or baseline + peak)

<sup>a</sup>Pongui Ngoma, Bourgault, Pop, and Nkounkou, "Adjustment of Parameters in Ionic Models Using Optimal Control Problems".

#### Identification of parameters for 1 pixel

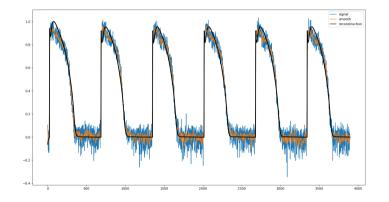


Figure: Exemple identification match. Raw signal (blue), filtered one (orange), and model (black).

## Spatial distribution of $g_{Na}$ and $g_{Na,c}$

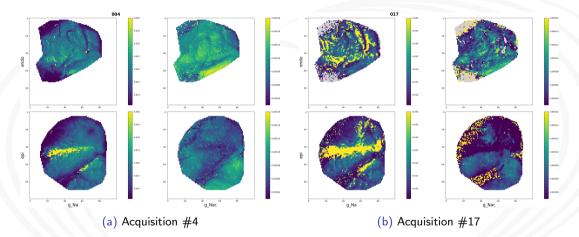


Figure:  $g_{Na}$  (left) and  $g_{Na,c}$  (right) on endo (top) and epi (bottom) – 2 acquisitions.

2023-11-16 30 / 33

#### Endo / epi distribution of parameters

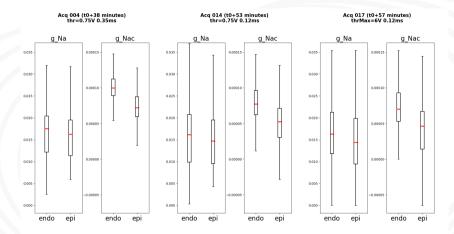
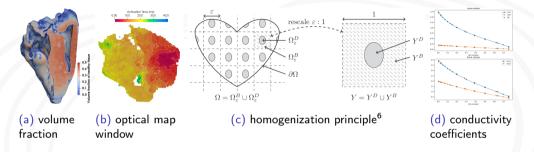


Figure: Statistical distribution of identified  $g_{Na}$  and  $g_{Na,c}$ 

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## Microstructure around the tip : conductivity coefficients



- Segmentation and fiber direction  $\rightarrow$  done (thanks to V. Ozenne, J. Magat)
- Volume fraction  $\rightarrow done^7$
- How to register the optical window to the the MR image (folded sample) ?

<sup>6</sup>Davidović, Coudière, and Bourgault, "Modelling the action potential propagation in a heart with structural heterogeneities: From high-resolution MRI to numerical simulations". <sup>7</sup>Ibid.

2023-11-16

#### Conclusion

Bridging between bioimpedance (signal processing) and bidomain (PDE modeling)

2023-11-16

33 / 33

- The computational pipeline is complete
- The software did undergo a thorough verification process
- Validation experiment were specifically designed, that is on-going work
- It remains possible improvements (numerics) to study
- Overall, the V&V process was certainly helpful

# Thanks for your attention!

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