

# **Applications of ultrasound elastography to hand and upper limb disorders**

Jessica Billy, Sabine F. Bensamoun, Julie Mercier, Sébastien Durand

## **To cite this version:**

Jessica Billy, Sabine F. Bensamoun, Julie Mercier, Sébastien Durand. Applications of ultrasound elastography to hand and upper limb disorders. Hand Surgery and Rehabilitation, 2024, 43 (2), pp.101636. 10.1016/j.hansur.2024.101636. hal-04397515

## **HAL Id: hal-04397515 <https://hal.science/hal-04397515>**

Submitted on 16 Jan 2024

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



[Distributed under a Creative Commons Attribution - NoDerivatives 4.0 International License](http://creativecommons.org/licenses/by-nd/4.0/)

**Applications of ultrasound elastography to hand and upper limb disorders.**

## **Applications de l'élastographie ultrasonore dans les pathologies de la main et du**

## **membre supérieur.**

## **Authors:**

Jessica Billy<sup>1</sup>, Sabine F Bensamoun<sup>2</sup>, Julie Mercier<sup>1</sup>, Sébastien Durand<sup>1</sup>.

<sup>1</sup> Department of Hand Surgery, Lausanne University Hospital, University of Lausanne, 1011 Lausanne, Switzerland

<sup>2</sup> Sorbonne University, Université de Technologie de Compiègne, CNRS UMR 7338, Biomechanics and Bioengineering, Compiègne, France

## **Corresponding author:**

Sébastien Durand, e-mail : [sebastien.durand@chuv.ch](mailto:sebastien.durand@chuv.ch)

The content has not been published or submitted for publication elsewhere, except as a brief abstract in the proceedings of a scientific meeting or symposium.

All authors have contributed significantly and all authors are in agreement with the content of the manuscript.

There is no potential conflicts of interest.

The authors received no financial support.

This study has been performed in accordance with the ethical standards as laid down in the

declaration of Helsinki.

## **Abstract**

Ultrasound elastography is a recently developed method for accurate measurements of soft tissue stiffness in addition to the subjective clinician's evaluation. The present review briefly describes the ultrasound elastography techniques and outlines clinical applications for tendon, muscle, nerve, skin and others soft tissues of the hand and upper limb.

## **Résumé**

L'élastographie ultrasonore est une méthode récemment développée pour déterminer la rigidité des tissus mous en complément de l'appréciation subjective du clinicien. Cet article de synthèse décrit brièvement les techniques d'élastographie par ultrasons et leurs applications cliniques pour les tendons, muscles, nerfs, peau et autres tissus mous de la main et du membre supérieur.

*Mots-clés* **:** Elastographie; Ultrason; Main; Chirurgie; tendon; muscle; nerf; peau.

*Keywords***:** Elastography; Ultrasound; Hand; Surgery; tendon; muscle; nerve; skin.

## **1. Ultrasound Elastography**

Ultrasound (US) elastography method noninvasively measures the stiffness of the tissue by producing and measuring tissue deformation [1]. Indeed, the stiffness represents the tendency of the tissue to resist at a deformation induced by an applied force and is characterized by the modulus of elasticity (Young's modulus; Pa) defined as follows:

E = σ/ε

where  $\sigma$  is the stress (Pa), corresponding to the normalized force, and  $\epsilon$  is the strain (unitless) which represents the deformation of the tissue due to an applied force. This US technique was first introduced in the early 1990s with *in vitro* experiments [2], then gradually expanded into clinical practice for diagnostic and sometimes prognostic purposes in the fields of senology, hepatology, thyroid diseases, prostatic diseases, and musculoskeletal conditions [3, 4]. Since 1990, different ultrasound elastography (UE) techniques have been used:

*The strain imaging* of the tissue, which revealed the physical tissue displacement, is estimated by its axial deformation parallel to the externally applied force exerted on body surface using the ultrasound probe (Fig. 1a). This method provides a qualitative evaluation of the stiffness [5]. Two approaches for strain imaging using ultrasound techniques were developed: the strain elastography (SE) and the acoustic radiation force impulse (ARFI) strain imaging. For the strain elastography, tissue displacement is generated by manual external compression with ultrasound transducer [2] or by an internal physiologic motion (cardiovascular) to assess more deeper located organ [6]. For ARFI technique, the force is produced by acoustic "pushing pulse". Different methods exist to measure displacement, such as radiofrequency echo correlation-based tracking or Doppler processing [7]. As the manual or physiological stresses are not quantifiable, the measured strain provides a qualitative assessment of the stiffness, or a pseudo-quantitative measure (strain ratio) represented with a color map called an elastogram.

*The Shear wave imaging* uses a dynamic stress to generate shear wave inside the tissue and the measurement of shear wave speed allow the determination of the stiffness. Three approaches exist to obtain shear wave imaging. Firstly, 1D transient elastography performed with the FibroScan® machine (Echosens, Paris, France) composed of a vibration-controlled probe [8]. Secondly, ARFI (Acuson S2000; Siemens Medical Solutions, Berlin, Germany) can be used and a portion of its longitudinal waves is converted to shear waves perpendicular to the longitudinal waves [9]. With the hypothesis of an isotropic, homogeneous, incompressible medium, shear wave velocity (SWV) is measured and converted to Young's modulus (E) using the equation:  $E = 3 \rho c^2$  where  $\rho$  is the density and c the shear wave velocity. Thirdly, in 2D shear wave elastography, acoustic radiation force is also used (Fig. 1b), with the Aixplorer ultrasound system (Supersonic Imagine, Aix-en-Provence, France) but multiple focal zones are stimulated and this creates a near cylindrical shear wave cone [10], permitting real-time monitoring of shear waves in 2D and generating quantitative elastograms superimposed on a B-mode image (Fig. 1c).

#### **2. Elastography and tendon**

Tendon rupture and histological modifications (e.g. disintegration of collagen fibers or mucoid degeneration) in degenerative tendinopathy are expected to lead to a localized decrease in tendon stiffness [11] and can be detected by UE earlier than conventional ultrasonography (US) [12]. Decreased mechanical properties (stiffness values) in degenerative tendinopathy and/or tendon injury compared to healthy tendon have been demonstrated in several studies [12-16] but in some cases of tendinopathy (long-head biceps tendon), UE analysis provides conflicting results [17]. Despite the different data, shear wave elastography (SWE) seems to be a reproducible technique to evaluate tendon elasticity with low inter-observer differences [21,22].

SWV of human hand tendons in physiologic in vivo conditions (Figs. 2a, 2b, 2c) has been analyzed in various studies [23]. Experimental studies using flexor tendons of porcine demonstrated that the stiffness of a partially teared tendon is lower in the vicinity of the injury and is dependent on the size of the tear [21]. Moreover, the stiffness of a damaged tendon induced by collagenase solution is significantly lower than a normal tendon [24]. Fluids accumulation and hematoma associated with acute ruptures appear as signal void areas on SWE [13,15]. After flexor tendon repair, re-rupture can be much more difficult to assess in B- mode. SWE of the flexor tendon proximally to the lesion demonstrated systematically a significant decrease of SWV compared to healthy side (figs. 2g, 2h, 2i). Injured flexor hand tendons after surgical repair showed altered regional mechanical elastic properties and appeared locally stiffer compared to the healthy side using SWE. The clinical significance of these findings was that the tendon would glide less on rigid local zone and authors propose to focus rehabilitation on these stiffer zones, aiming at improving gliding [24].

In their study, Turkay et al. [13] compared SWE acquisitions performed on 40 patients suffering from De Quervain tenosynovitis, with 40 healthy volunteers. They have demonstrated that the Young's modulus values of healthy tendons of the first extensor compartment were  $69.17 \pm 22.45$  kPa whereas pathologic tendons showed lower values (29.75±8.02 kPa).

Medial and lateral epicondylitis have also been analyzed in several studies, all suggesting that pathological tendons are softer than healthy tendons [14,16,23-25]. Park et al. [25] concluded that UE accuracy was better than US by 7.1% for epicondylitis diagnosis, sensibility and specificity being of 96% and 89%, respectively. Moreover, it has been demonstrated that SWV increases after conservative treatment in patients with lateral epicondylitis [14]. The combined modalities of UE and B-mode ultrasonography provided a statistically significant improvement in the agreement between imaging and histologic results compared with each modality alone [16] in common flexor tendinopathy at the medial side of the elbow.

In a recent systematic review concerning rotator cuff tears [26], SWE has been successfully used to identify the localization and degree of supraspinatus tendon tear, improving the value of ultrasound. Indeed, patients with rotator cuff tears had a lower mean SWV of muscle and tendon under active conditions. Intraclass correlation coefficient was excellent (0.96). Supraspinatus tendon Stiffness increased between 8 days and 24 weeks after repair [26].

## **3. Elastography and muscle**

With a Young's modulus ranging from 5-40 kPa at rest and up to 300 kPa during passive stretching or active contraction, SWE is a validated method to assess muscle stiffness [27,28]. The reliability of the SWE for measuring muscle stiffness has been proven for several muscle groups including lower limb, trunk, upper limbs muscles including the intrinsic muscles of the hand [29,30]. Furthermore, studies suggested using SWE to monitor the muscle activity or muscle force and to demonstrate contraction of accessory muscles [29, 31]. SWE was also found to be a reliable method to evaluate the functional muscle activity compared to the physiological activity recorded with conventional surface electromyography [32]. SWV of muscle seems to be influenced by sex, age, and dominance of the patient [32, 33]. Moreover, studies have highlighted the importance of the probe orientation, as the stiffness may decrease in a perpendicular position compared to the parallel orientation to the muscle fibers [34,35]. This result showed the anisotropic behavior of the skeletal muscle. The parallel orientation of the probe with the main direction of the muscle fiber is more valid and reliable for SWV measurements.

Increased stiffness values, related to muscle spasticity, were shown in patients with neuromuscular diseases, such as cerebral palsy [36], Duchenne muscular dystrophy [37] in passive state, Parkinson's disease or neonatal brachial plexus palsy [38]. SWE provides an additional objective tool for spasticity evaluation in targeting the right skeletal muscle in a spastic upper limb [39] and could be helpful to assess results of Botulinum Toxin injection [40] or surgery.

In the study of Li et al., SWE of thenar muscle in hemiplegic patients was performed. The plegic side showed decreased stiffness compared to the healthy side. Changes in muscle stiffness in the follow-up could be used as an objective assessment of rehabilitation degree [41].

In 2017, Lamouille et al. reported the assessment of in vivo muscle tension during transfer of the extensor indicis proprius to the extensor pollicis longus (Fig. 3). SWE measurements were obtained at different stages of surgery including at rest before transferring, during active extension, and after tendon transfer at rest. The investigators showed differences in stiffness values at the various stages of the tendon transfer procedure, providing new insights to improve therapeutic treatments of tendon transfers [42].

Due to the frequency of rotator cuff degenerative lesion, shoulder muscles are of interest, and the most analyzed structures in the upper limb. In non-pathologic shoulders, SWE supraspinatus muscle values gradually decreased with increased passive abduction of the shoulder. In case of large to massive tears, the stiffness of supraspinatus muscle did not vary from adduction to abduction conditions [43]. After rotator cuff surgical repair, the contractile behavior of the supraspinatus muscle increased from 6 weeks to 3 months after surgery and the stiffness reached a steady-state after 3 months. Opposite variation was found for deltoid muscle activity which reached the same level as healthy muscle after 6 months [44]. This phenomenon is explained by a compensatory role of the deltoid during rotator cuff injury. In addition to human muscle, SWE technique is developed to characterize the muscle stiffness of small rodents to follow the effect of treatment [45].

#### **4. Elastography and nerve**

For many years, electroneuromyography (ENMG) has been considered as the main diagnostic tool for peripheral neuropathies. Then, B-mode ultrasound image and doppler examination were found to be of interest in Carpal Tunnel Syndrome (CTS) diagnosis showing 1) an increase in nerve cross sectional area (CSA) proximal to the region where the nerve is compressed, 2) a reduced mobility of the nerve, 3) a modification of the structural properties trough a variation in echogenicity, and 4) an increased vascularity of the nerve. SWE in adjunction to B-mode ultrasound seems to be reliable for entrapment neuropathy diagnosis, with the advantages of being a noninvasive test, accessible and fast for the surgeon. Most SWE studies related to the upper limb nerves were performed on the median nerve (MN) for CTS. Authors have demonstrated that long term edema or high carpal tunnel pressure could lead to increase median nerve stiffness [46]. Reliability and feasibility of the stiffness

measurements obtained with SWE realized in healthy MN ( $N = 40$ ) subjects showed excellent interobserver and intra-observer agreement (0.852–0.930), with no difference between bilateral forearm measurements [47]. Attention must be paid to limb positioning during SWE measurements, as it directly affects nerve tension and the stiffness values [48]. A metaanalysis, composed of 17 studies, using sonoelastography to image the MN at wrist level (N = 1401 wrists) confirmed that the nerve is consistently stiffer in patients with CTS than healthy subjects regardless of the ultrasound elastography technique [49]. Moreover, studies have successively identified the severity of the nerve disease trough different stages [50,51]. Thus, SWE allowed for a classification of CTS similar as the gold-standard electrodiagnostic, with cases stratified as mild, moderate or severe based on the stiffness of the MN. This was not achievable using B-mode US alone. Both median nerve CSA at the wrist and shear wave velocity were reduced one week after surgical carpal tunnel release, reflecting nerve recovery [52]. However, cut-off stiffness values proposed for CTS diagnosis are different according to authors, varying from 40.4 kPa to 79 kPa [46,53]. Also, Sugiyama et al. [54] proposed SWE technique as an objective, quantitative and noninvasive evaluation test for MN follow-up after volar locking plate osteosynthesis of distal radius fracture, helping to decide on material removal and MN neurolysis timing.

For ulnar nerve neuropathy at the elbow and at the Guyon's canal, SWE appears as a new reliable method to support this diagnosis [55,56]. Ulnar nerve stiffness with a Young's modulus over 61 kPa and the stiffnesses ratios of the cubital tunnel to distal arm and to mid-arm were 1.68 and 1.75, respectively, provided 100% specificity and sensitivity for the detection of ulnar neuropathy at the elbow [55]. Also, SWE can be used to differentiate ulnar neuropathy at the cubital tunnel from asymptomatic ulnar nerve with medial epicondylitis and healthy uncompressed ulnar nerve [57]. Patients with unilateral ulnar tunnel syndrome showed higher CSA and stiffness in the affected side for all positions, i.e., extension, 45° flexion, 90° flexion and at the maximum flexion of the elbow where the stiffness increased [58]. In patients with ulnar nerve decompression associated with anterior transposition, post-operative ulnar nerve stiffness increased with elbow joint extension (Fig. 4). However, on the non-operative side, ulnar nerve stiffness increased with elbow flexion [59].

Recently, SWE was used before ultrasound-guided perineural hydrodissection to identify the level of stiffness of the scar surrounding the radial nerve in two cases of radial nerve palsy after humeral shaft fractures [60].

#### **5. Elastography and skin**

The use of high frequency ultrasonographic transducers has made elastographic assessment of the skin possible [61]. Most of the previous studies using elastography for skin evaluation included patients with cancers, connective tissue diseases, chronic systemic inflammation, lipodermatosclerosis or patients at risk of ulceration [62] but it may also find applications in aesthetic medicine.

SWE distinguishes normal skin from scars and could be used to evaluate scar severity, which could be important for patient care and treatment. Additionally, excellent intra-observer and inter-observer reliability was noted, even when the test is performed by a novice clinician versus an experienced sonographer. A direct linear relationship was established between scar thickness, scar pliability and SWV measurements [63]. In a similar way, SWE was also used to quantify keloid response to treatment after intralesional corticosteroid injection [64]. The results showed no statistical differences in thickness between normal skin and treated keloids. SWE values of treated keloids significantly decreased but the data were still higher than normal skin.

Conservative treatment of fingertip amputation using occlusive dressings can lead to soft tissue regeneration. Ultrasonography and SWE were performed on the regenerated fingertips [65]. Compared to collateral uninjured finger, there were no difference in pulp thickness, but vascularization and stiffness were both significantly higher after fingertip regeneration.

SWE appears also as an objective tool to assess elasticity of the skin after flap reconstruction. In our clinic, we used the UE to measure the stiffness the skin of the hand after fascia superficialis flap surgery following a severe hand trauma (Fig. 5). To our knowledge, this is the first case described. To date, a unique study conducted by a team of plastic surgeons focused on elastography applied to this field [66]. They used SWE on subcutaneous fat after deep inferior epigastric perforator (DIEP) flap and found a positive correlation between flap weight and fat tissue stiffness. This result is particularly relevant because fat induration and necrosis are common complications after breast reconstruction with DIEP flaps but are currently only evaluated clinically.

### **6. Elastography and other indications in hand and upper extremity.**

Extrapolating results from the pre-mentioned tissues, it was felt that UE may be a useful way to assess ligament health and integrity. Decreased SWV in the ulnar collateral ligament of the elbow in the dominant side was observed in baseball pitchers at midseason [67]. This functional decline of the ulnar collateral ligament was probably associated with structural modification. Another study described the use of SWE to quantify the central band stiffness of interosseous membrane located in the forearm placed in different positions of pronosupination [68]. The results demonstrated that the stiffness measurements were reproducible between examinators. Authors raised that elastography could help guiding diagnostic and therapy in the management of interosseous membrane lesions.

B-mode US image for trigger finger diagnosis shows A1 pulley thickening. Additional UE acquisitions showed A1 pulley stiffness. These two parameters tend to alleviate after corticosteroid injection, being a valid asset for treatment follow-up [69].

 To date, measurement methods of acute compartment syndrome are invasive and clinical diagnosis may be challenging. According to Zhang et al. [70], muscle stiffness indirectly reflects intra-compartmental pressure revealing that SWE may be a noninvasive quantitative diagnostic tool for compartment syndrome diagnosis. In a short study [70], muscle stiffness was found to be significantly higher than the unaffected side in four patients with actual acute compartment syndrome, but more data are needed before a quantitative cut-off can be established. Chronic compartment syndrome can also be explored using SWE (Fig. 6).

 In the oncological field, higher stiffness values are noted for malignant lesions (breast, thyroid, prostate, skin, lymph node) compared to benign masses [71-74]. SWE shows higher sensitivity and specificity than traditional B-mode US image to identify malignant lesion [72] and could avoid invasive risky procedures such as biopsies. Musculoskeletal tumors encompass a vast array of distinct tumor types, the majority of which are usually benign. Malignant musculoskeletal tumors may express a wider range of stiffness values due to its more heterogeneous structure compared to benign lesions [75]. With only a few studies on this topic [75-79], no evident correlation was found between stiffness and malignancy detection of musculoskeletal tumors [75-77].

In a recent retrospective study [80], the combination of elastography with 2D imaging and color flow imaging achieved an excellent diagnostic accuracy in schwannomas for patients with softtissues masses located in the limb.

Finally, elastography could be also used for vessels stiffness evaluation. For example, SWE can distinguish between acute and chronic clots by characterizing tissue stiffness in deep vein thrombosis. It was also demonstrated that variation of stiffness in brachial arteries was statistically inferior in patients with known cardiovascular disease when compared to the healthy control group. This could be an early sign of atherosclerosis [81].

#### **7. Conclusion**

Ultrasound Elastography is an accessible non-invasive imaging modality that can be used to measure stiffness in a variety of soft biological tissue. With the growing interest in developing new elastography applications in hand and upper extremity, one should keep in mind technical limitations concerning reproducibility and repeatability. This technique has a promising future but a significant amount of work remains to be done. Indeed, protocol must be standardized as positioning of the joint affects SWV measurement of the surrounding soft tissues. In addition, the selection of region of interest (ROI) is operator dependent and may introduce variability. Also, several commercial systems exist leading to the use of different probes with different ranges of frequencies which can impact SWV values.

SE provides only qualitative values of stiffness and the external force is difficult to reproduce, or is variable through time, and can generate artifacts. SWE takes a simplistic view on the soft tissues mechanical properties (isotropic, homogeneous, linear elastic, incompressible) to facilitate the process of imaging, but tissues are anisotropic, heterogeneous, viscoelastic and skeletal muscle, for example, accounting for the associated vascular and lymphatic components, is compressible.

Despite the above cited limitations, UE has demonstrated important correlation to diffuse and focal disease states in multiple soft tissues of the upper limb. UE holds huge clinical applications in MSK tissues. Continued development of systems and increased formation of clinicians to UE will lead to expand our knowledge of Elastography and its clinical applications in the future.

#### *Authors declare no conflicts of interest.*

#### **References**

[1] Giambini H, An KN. Ultrasound Elastography for Hand Soft Tissue Assessment. Hand Clin 2022;38(1):119-28.

[2] Ophir J, Cespedes I, Ponnekanti H, Yazdi Y, Li X. Elastography: a quantitative method for imaging the elasticity of biological tissues. Ultrasonic imaging 1991;13:111-34.

[3] Drakonaki EE, Allen GM, Wilson DJ. Ultrasound elastography for musculoskeletal applications. Br J Radiol 2012;85:1435–45.

[4] Klauser AS, Miyamoto H, Bellmann-Weiler R, Feuchtner GM, Wick MC, Jaschke WR. Sonoelastography: musculoskeletal applications. Radiology 2014;272:622–33.

[5] Debernard L, Robert L, Charleux F, Bensamoun SF. A possible clinical tool to depict muscle elasticity mapping using magnetic resonance elastography (MRE). Muscle & Nerve 2013; 47(6):903- 8.

[6] Gennisson JL, Deffieux T, Fink M, Tanter M. Ultrasound elastography: principles and techniques. Diagnostic and interventional imaging. 2013;94:487-95.

[7] Sigrist RMS, Liau J, Kaffas AE, Chammas MC, Willmann JK. Ultrasound Elastography: Review of Techniques and Clinical Applications. Theranostics 2017;7(5):1303-29.

[8] Pouletaut P, Boussida S, Ternifi R, Miette V, Audière S, Fournier C, Sandrin L, Charleux F, Bensamoun SF. Impact of Hepatic Iron Overload in the Evaluation of Steatosis and Fibrosis in Patients With Nonalcoholic Fatty Liver Disease Using Vibration Controlled Transient Elastography (VCTE) and MR Imaging Techniques: a Clinical Study. Innovation and Research in BioMedical engineering (IRBM), 2023;44(3):100750. [hal-03937041](https://hal.science/hal-03937041) doi[:10.1016/j.irbm.2022.100750](https://doi.org/10.1016/j.irbm.2022.100750)

[9] Nightingale K. Acoustic Radiation Force Impulse (ARFI) Imaging: a Review. Current medical imaging reviews 2011;7:328-39.

[10] Bamber J, Cosgrove D, Dietrich CF, Fromageau J, Bojunga J, Calliada F, et al. EFSUMB guidelines and recommendations on the clinical use of ultrasound elastography. Part 1: Basic principles and technology. Ultraschall in der Medizin 2013;34:169-84.

[11] Balaban M, Cilengir AH, Idilman IS. Evaluation of Tendon Disorders with Ultrasonography and Elastography. J Ultrasound Med 2021;40(7):1267–86.

[12] Chen XM, Cui LG, He P, Shen WW, Qian YJ, Wang JR. Shear wave elastographic characterization of normal and torn achilles tendons: a pilot study. J Ultrasound Med 2013;32(3):449– 55.

[13] Turkay R, Inci E, Aydeniz B, Vural M. Shear wave elastography findings of de Quervain tenosynovitis. European Journal of Radiology 2017;95:192–6.

[14] Zhu B, You Y, Xiang X, Wang L, Qiu L. Assessment of common extensor tendon elasticity in patients with lateral epicondylitis using shear wave elastography. Quant Imaging Med Surg 2020;10(1):211–9.

[15] Aubry S, Nueffer JP, Tanter M, Becce F, Vidal C, Michel F. Viscoelasticity in Achilles tendinopathy: quantitative assessment by using real-time shear-wave elastography. Radiology. 2015;274(3):821–9.

[16] Klauser AS, Pamminger MJ, Halpern EJ, Abd Ellah MMH, Moriggl B, Taljanovic MS, et al. Sonoelastography of the Common Flexor Tendon of the Elbow with Histologic Agreement: A Cadaveric Study. Radiology 2017;283(2):486–91.

[17] Sahan MH, Inal M, Burulday V, Kultur T. Evaluation of Tendinosis of the Long Head of the Biceps Tendon by Strain and Shear Wave Elastography. Med Ultrason 2018;20:192–8.

[18] Prado-Costa R, Rebelo J, Monteiro-Barroso J, Preto AS. Ultrasound elastography: compression elastography and shear-wave elastography in the assessment of tendon injury. Insights Imaging 2018;9(5):791–814.

[19] Şendur HN, Cindil E, Cerit M, Demir NB, Şendur AB, Oktar SÖ. Interobserver variability and stiffness measurements of normal common extensor tendon in healthy volunteers using shear wave elastography. Skeletal Radiol 2019 Jan;48(1):137–41.

[20] Chen PY, Yang TH, Kuo LC, Shih CC, Huang CC. Characterization of Hand Tendons Through High-Frequency Ultrasound Elastography. IEEE Trans Ultrason Ferroelectr Freq Control. 2020 Jan;67(1):37-48.

[21] Dewall RJ, Jiang J, Wilson JJ, Lee KS. Visualizing tendon elasticity in an ex vivo partial tear model. Ultrasound Med Biol. 2014;40(1):158-67.

[22] Yeh CL, Kuo PL, Gennisson JL, Brum J, Tanter M, Li PC. Shear Wave Measurements for Evaluation of Tendon Diseases. IEEE Trans Ultrason Ferroelectr Freq Control 2016;63(11):1906-21.

[23] De Zordo T, Lill SR, Fink C, Feuchtner GM, Jaschke W, Bellmann-Weiler R, et al. Real-time sonoelastography of lateral epicondylitis: comparison of findings between patients and healthy volunteers. AJR Am J Roentgenol 2009;193(1):180–5.

[24] Chen PY, Yang TH, Kuo LC, Hsu HY, Su FC, Huang CC. Evaluation of Hand Tendon Elastic Properties During Rehabilitation Through High-Frequency Ultrasound Shear Elastography. IEEE Trans Ultrason Ferroelectr Freq Control 2021;68(8):2716–26.

[25] Park G, Kwon D, Park J. Diagnostic confidence of sonoelastography as adjunct to greyscale ultrasonography in lateral elbow tendinopathy. Chin Med J (Engl) 2014;127(17):3110–5.

[26] Seth I, Hackett LM, Bulloch G, Sathe A, Alphonse S, Murrell GAC. The application of shear wave elastography with ultrasound for rotator cuff tears: a systematic review. JSES Rev Rep Tech 2023;3(3):336-42.

[27] Shinohara M, Sabra K, Gennisson JL, Fink M, Tanter M. Real-time visualization of muscle stiffness distribution with ultrasound shear wave imaging during muscle contraction. Muscle Nerve 2010;42(3):438–41.

[28] Koo TK, Hug F. Factors that influence muscle shear modulus during passive stretch. J Biomech 2015;48(12):3539–42.

[29] Kim K, Hwang HJ, Kim SG, Lee JH, Jeong WK. Can Shoulder Muscle Activity Be Evaluated With Ultrasound Shear Wave Elastography? Clin Orthop Relat Res 2018;476(6):1276–83.

[30] Watanabe Y, Iba K, Taniguchi K, et al. Assessment of the passive tension of the first dorsal interosseous and first lumbrical muscles using shear wave elastography. J Hand Surg Am 2019;44(12):1092.e1–18.

[31] Durand S, Collinot JA, Christen T, Becce F, Voser T. Morphological and functional assessment of the flexor carpi radialis brevis using conventional ultrasound and elastography. Surg Radiol Anat 2021;43(5):721–6.

[32] Xie Y, Thomas L, Hug F, Johnston V, Coombes BK. Quantifying cervical and axioscapular muscle stiffness using shear wave elastography. J Electromyogr Kinesiol Elsevier Ltd; 2019;48:94– 102.

[33] Chen J, O'Dell M, He W, Du LJ, Li PC, Gao J. Ultrasound shear wave elastography in the assessment of passive biceps brachii muscle stiffness: influences of sex and elbow position. Clin Imaging Elsevier Inc. 2017;45:26–9.

[34] Eby SF, Song P, Chen S, Chen Q, Greenleaf JF, An KN. Validation of shear wave elastography in skeletal muscle. J Biomech 2013;46: 2381–2387.

[35] Gennisson JL, Deffieux T, Mace E, et al. Viscoelastic and anisotropic mechanical properties of in vivo muscle tissue assessed by supersonic shear imaging. Ultrasound Med Biol 2010;36(5):789–801.

[36] Brandenburg JE, Eby SF, Song P, Kingsley-Berg S, Bamlet W, Sieck GC, et al. Quantifying passive muscle stiffness in children with and without cerebral palsy using ultrasound shear wave elastography. Dev Med Child Neurol 2016;58:1288–94.

[37] Bensamoun SF, Charleux F, Debernard L, Themar-Noel C, Voit T. Elastic properties of skeletal muscle and subcutaneous tissues in Duchenne muscular dystrophy by magnetic resonance elastography (MRE): a feasibility study. Innovation and Research in BioMedical engineering (IRBM) 2015;36(1):4-9.

[38] Riquier-Le Chatelier M, Giai J, Lallement-Dudek P, Herisson O, Fitoussi F. Muscle elasticity in patients with neonatal brachial plexus palsy using shear-wave ultrasound elastography. Preliminary results. J Pediatr Orthop B 2021;30(4):385–92.

[39] Lehoux MC, Sobczak S, Cloutier F, Charest S, Bertrand-Grenier A. Shear wave elastography potential to characterize spastic muscles in stroke survivors: Literature review. Clin Biomech (Bristol, Avon) 2020;72:84–93.

[40] Gao J, Rubin JM, Chen J, O'Dell M. Ultrasound Elastography to Assess Botulinum Toxin A Treatment for Post-stroke Spasticity: A Feasibility Study. Ultrasound Med Biol 2019;45(5):1094–102. [41] Li R, Zheng S, Zhang Y, Zhang H, Du L, Cheng L, Li H, Zhang W, Du K, He W, Zhang W. Quantitative assessment of thenar to evaluate hand function after stroke by Bayes discriminant. BMC Musculoskelet Disord 2023;24(1):682.

[42] Lamouille J, Müller C, Aubry S, Bensamoun S, Raffoul W, Durand S. Extensor indicis proprius tendon transfer using shear wave elastography. Hand Surgery and Rehabilitation. 2017;36(3):173–80.

[43] Hatta T, Giambini H, Uehara K, Okamoto S, Chen S, Sperling JW, et al. Quantitative assessment of rotator cuff muscle elasticity: Reliability and feasibility of shear wave elastography. J Biomech 2015;48(14):3853–8.

[44] Ishikawa H, Muraki T, Morise S, Kurokawa D, Yamamoto N, Itoi E, et al. Changes in shoulder muscle activities and glenohumeral motion after rotator cuff repair: an assessment using ultrasound real-time tissue elastography. J Shoulder Elbow Surg 2021;30(11): 2577–86.

[45] Ternifi R, Kammoun M, Pouletaut P, Subramaniam M, Hawse JR, Bensamoun SF. Ultrasound image processing to estimate the structural and functional properties of mouse skeletal muscle. Biomedical Signal Processing and Control 2020 Feb;56:101735. doi: 10.1016/j.bspc.2019.101735.

[46] Kantarci F, Ustabasioglu FE, Delil S, Olgun DC, Korkmazer B, Dikici AS et al. Median nerve stiffness measurement by shear wave elastography: a potential sonographic method in the diagnosis of carpal tunnel syndrome. Eur Radiol 2014;24(2):434-40. doi: 10.1007/s00330-013-3023-7

[47] Zhu B, Yan F, He Y, et al. Evaluation of the healthy median nerve elasticity: Feasibility and reliability of shear wave elastography. Medicine (Baltimore) 2018;97(43):e12956.

[48] Rugel CL, Franz CK, Lee SSM. Influence of limb position on assessment of nerve mechanical properties by using shear wave ultrasound elastography. Muscle Nerve 2020;61(5):616–22.

[49] Lin CP, Chen IJ, Chang KV, Wu WT, Özçakar L. Utility of Ultrasound Elastography in Evaluation of Carpal Tunnel Syndrome: A Systematic Review and Meta-analysis. Ultrasound Med Biol 2019;45(11):2855–65.

[50] Wee TC, Simon NG. Shear wave Elastography in the Differentiation of Carpal Tunnel Syndrome Severity. PM R 2020;12(11):1134–9.

[51] Sernik RA, Pereira RFB, Cerri GG, Damasceno RS, Bastos BB, Leão RV. Shear wave elastography is a valuable tool for diagnosing and grading carpal tunnel syndrome. Skeletal Radiol 2023;52(1):67–72.

[52] Wu H, Zhao HJ, Xue WL, Wang YC, Zhang WY, Wang XL. Ultrasound and elastography role in pre- and post-operative evaluation of median neuropathy in patients with carpal tunnel syndrome. Front Neurol 2022;13:1079737.

[53] Paluch Ł, Pietruski P, Walecki J, Noszczyk BH. Wrist to forearm ratio as a median nerve shear wave elastography test in carpal tunnel syndrome diagnosis. J Plast Reconstr Aesthet Surg 2018;71(8):1146–52.

[54] Sugiyama Y, Naito K, Miyamoto H, Goto K, Kinoshita M, Nagura N, et al. A Survey of the Median Nerve Elasticity after Volar Locking Plate Fixation Using Ultrasound Elastography. J Hand Microsurg 2020;12(2):95–9.

[55] Paluch Ł, Noszczyk B, Nitek Ż, Walecki J, Osiak K, Pietruski P. Shear-wave elastography: a new potential method to diagnose ulnar neuropathy at the elbow. Eur Radiol 2018;28(12):4932–9.

[56] Paluch Ł, Noszczyk BH, Walecki J, Osiak K, Kiciński M, Pietruski P. Shear-wave elastography in the diagnosis of ulnar tunnel syndrome. J Plast Reconstr Aesthet Surg 2018;71(11):1593–9.

[57] Kim S, Lee GY. Evaluation of the ulnar nerve with shear-wave elastography: a potential sonographic method for the diagnosis of ulnar neuropathy. Ultrasonograph. 2021;40(3):349–56.

[58] Wolny T, Fernández-de-Las-Peñas C, Granek A, Linek P. Changes in Ultrasound Measurements of the Ulnar Nerve at Different Elbow Joint Positions in Patients with Cubital Tunnel Syndrome. Sensors (Basel) 2022;22(21):8354.

[59] Durand S, Raffoul W, Christen T, Pedrazzi N. Post-Operative Assessment of Ulnar Nerve Tension Using Shear-Wave Elastography. Neurol Int 2021;13(3):469–76.

[60] Su DC, Chang KV, Lam SKH. Shear Wave Elastography to Guide Perineural Hydrodissection: Two Case Reports. Diagnostics (Basel) 2020;10(6):348. doi: 10.3390/diagnostics10060348.

[61] Xiang X, Yan F, Yang Y, Tang Y, Wang L, Zeng J, Qiu L. Quantitative Assessment of Healthy Skin Elasticity: Reliability and Feasibility of Shear Wave Elastography. Ultrasound Med Biol 2017;43(2):445-452.

[62] Deprez JF, Brusseau E, Fromageau J, Cloutier G, Basset O. On the potential of ultrasound elastography for pressure ulcer early detection. Med Phys 2011;38(4):1943-50.

[63] DeJong H, Abbott S, Zelesco M, Spilsbury K, Martin L, Sanderson R, et al. A Novel, Reliable Protocol to Objectively Assess Scar Stiffness Using Shear Wave Elastography. Ultrasound Med Biol 2020;46(7):1614–29.

[64] Huang SY, Xiang X, Guo RQ, Cheng S, Wang LY, Qiu L. Quantitative assessment of treatment efficacy in keloids using high-frequency ultrasound and shear wave elastography: a preliminary study. Sci Rep 2020;10(1):1375.

[65] Jafari P, Muller C, Grognuz A, Applegate LA, Raffoul W, di Summa PG, Durand S. First Insights into Human Fingertip Regeneration by Echo-Doppler Imaging and Wound Microenvironment Assessment. Int J Mol Sci 2017;18(5):1054.

[66] Sowa Y, Yokota I, Fujikawa K, Morita D, Taguchi T, Numajiri T. Objective evaluation of fat tissue induration after breast reconstruction using a deep inferior epigastric perforator (DIEP) flap. Journal of Plastic Surgery and Hand Surgery 2019;53(3):125–9.

[67] Gupta N, Taylor RE, Lambert B, Dong D, Phillips P, Jack RA 2nd, Goble HM, Labis JS, Trakhtenbroit MA, McCulloch PC. Shear wave elastography of the ulnar collateral ligament in division IA pitchers across a competitive collegiate season. JSES Int 2023;7(4):703-708.

[68] Rougereau G, Marty-Diloy T, Vigan M, Vialle R, Soubeyrand M, Langlais T. Biomechanical assessment of the central band of the interosseous membrane using shear wave elastography: reliability and reproducibility. J Hand Surg Eur Vol 2022;47(11):1134–41.

[69] Miyamoto H, Miura T, Isayama H, Masuzaki R, Koike K, Ohe T. Stiffness of the first annular pulley in normal and trigger fingers. J Hand Surg Am 2011;36(9):1486–91. others.

[70] Zhang J, Zhang W, Zhou H, Sang L, Liu L, Sun Y, et al. An exploratory study of two-dimensional shear-wave elastography in the diagnosis of acute compartment syndrome. BMC Surg 2021;21(1):418.

[71] Berg WA, Cosgrove DO, Doré CJ, Schäfer FKW, Svensson WE, Hooley RJ, et al. Shear-wave elastography improves the specificity of breast US: the BE1 multinational study of 939 masses. Radiology 2012;262(2):435–49.

[72] Tang GX, Xiao XY, Xu XL, Yang HY, Cai YC, Liu XD, et al. Diagnostic value of ultrasound elastography for differentiation of benign and malignant axillary lymph nodes: a meta-analysis. Clin Radiol 2020;75(6):481.e9-481.e16.

[73] Wang F, Chang C, Gao Y, Chen YL, Chen M, Feng LQ. Does Shear Wave Elastography Provide Additional Value in the Evaluation of Thyroid Nodules That Are Suspicious for Malignancy? Journal of Ultrasound in Medicine 2016;35(11):2397–404.

[74] Han X, Li J, Zeng F, Liu H, He Y. Differential diagnosis of basal cell carcinoma by high-resolution ultrasound elastography. Skin Res Technol 2022;28(2):350–4.

[75] Li A, Peng XJ, Ma Q, Dong Y, Mao CL, Hu Y. Diagnostic performance of conventional ultrasound and quantitative and qualitative real-time shear wave elastography in musculoskeletal soft tissue tumors. J Orthop Surg Res 2020;15(1):103.

[76] Pass B, Jafari M, Rowbotham E, Hensor EMA, Gupta H, Robinson P. Do quantitative and qualitative shear wave elastography have a role in evaluating musculoskeletal soft tissue masses? Eur Radiol 2017;27(2):723–31.

[77] Winn N, Baldwin J, Cassar-Pullicino V, Cool P, Ockendon M, Tins B, et al. Characterization of soft tissue tumours with ultrasound, shear wave elastography and MRI. Skeletal Radiol 2020;49(6):869–81.

[78] Tavare AN, Alfuraih AM, Hensor EMA, Astrinakis E, Gupta H, Robinson P. Shear-Wave Elastography of Benign versus Malignant Musculoskeletal Soft-Tissue Masses: Comparison with Conventional US and MRI. Radiology 2019;290(2):410–7.

[79] Schivo D, Gjika E, Traverso A, Durand S. Shear Wave Elastography in the Diagnosis of Hand Tumours. Case Rep Orthop. 2019 Feb 24;2019:2736529.

[80] Yuan Y, Gao J, Xiong G, Guo L. Diagnostic accuracy of multiparametric ultrasound for peripheral nerve schwannoma. Acta Radiol 2023;64(4):1608-1614. doi: 10.1177/02841851221125109.

[81] Gülşen F, Samanci C, Memis Durmaz ES, Durmaz E, Tel C, Gencturk M, et al. Brachial Artery Wall Stiffness Assessment by Shear Wave Elastography: A Promising New Diagnostic Tool for Endothelial Dysfunction Detection. J Ultrasound Med 2018;37(8):1977–83.

## **Figures legends**

Fig. 1: Ultrasound elastography represented by strain elastography (a) and shear wave elastography (b). The colored region represents the 2D quantitative elastogram superimposed on a B-mode image with a color scale (see top right). The software allowed us to measure the mean stiffness (Young's modulus in kPa) value and the shear wave velocity,  $(m.s<sup>-1</sup>)$  of the flexor pollicis longus (asterisk) inside a circular region of interest (ROI), (c).

Fig. 2: Shear wave elastography of human flexor pollicis longus tendon in physiological conditions at rest (210.1 kPa, 8.4 m.s<sup>-1</sup>) (a), active flexion (490 kPa, 12.7 m.s<sup>-1</sup>) (b) and passive extension (486.6 kPa, 12.7 m.s<sup>-1</sup>) (c). Patient with anterior interosseous nerve palsy (second row). Shear wave velocity (SWV) and stiffness does not increase between the rest position (175.7 kPa, 7.7 m.s<sup>-1</sup>) (d) and the active flexion (172.5 kPa, 7.6 m.s<sup>-1</sup>) (e). Hyperintensity in flexor pollicis longus and pronator quadratus muscles (arrow) are revealed using 3T MRI acquisitions (f). Patient with tendon rupture after flexor digitorum profondus of the index repair (third row). Re-rupture is difficult to identify in B-mode (g) proximally to the rupture. The cartographies of stiffness show lower values for the flexor digitorum profondus of the index (66.1 kPa, 4.7 m.s<sup>-1</sup>) (h) compared to the flexor digitorum superficialis of the index (378.5 kPa, 11.2 m.s −1 ) (i).

Fig. 3: Intra operative photographs of the transfer of the extensor indicis proprius to the extensor pollicis longus using ultrasound shear wave elastography (a). Differences in stiffness values at the various stages of surgery were obtained from the rest (b) to the transferring active extension (c) and after tendon transfer at rest (d).

Fig. 4: Decompression of the ulnar nerve at the elbow with subcutaneous transposition and anterior stabilization with a fascial sling (a). Box plot of the stiffness of the ulnar nerve (kPa) at 0 and 120 degrees of elbow flexion on the operative side (red) and non-operative side (blue) (b). Elastogram of the ulnar nerve after anterior transposition at rest (c) and at 120 degrees elbow flexion (d).

Fig. 5: Temporoparietal fascia free flap (a) for coverage of a large defect of dorsal aspect of the hand (b). Combination of Matriderm® and skin graft. Clinical outcomes at 6 months post operative (c, d). Similar skin shear wave velocity and elasticity observed in the pathologic (e) and contralateral side (f), respectively (17.3 kPa, 2.4 m.s<sup>-1</sup>) and (16.8 kPa, 2.4 m.s<sup>-1</sup>).

Fig. 6: Patient with chronic compartmental syndrome of the right forearm confirmed using intra compartmental pressure monitor (Compass®). B-mode of the transversal / axial image of the right forearm (a). Shear wave elastography measurement of the antebrachial fascia showed a stiffness at 36.7 kPa at rest (b) which increase after 10 min and 20 min muscle contraction exercise at 85.6 kPa (b) and 189.4 kPa (c), respectively. Then, patient stopped because of pain and tingling in the fingers (d).

Fig.1



Fig.2



Fig.3



 $\mathbf{b}^{\mathrm{in}}$ a lulus 15 (kPa) elbow flexion  $0^\circ$ elbow flexion 120° left elbow 120 left elbow 0  $\overline{\mathbf{e}}$ 





Fig.4

