



HAL
open science

Filming DNA repair at the atomic level

Marten Vos

► **To cite this version:**

Marten Vos. Filming DNA repair at the atomic level. 2023, pp.996-997. 10.1126/science.adl3002 . hal-04383261

HAL Id: hal-04383261

<https://hal.science/hal-04383261v1>

Submitted on 16 Oct 2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Filming DNA repair at the atomic level

Dissection of multistep catalysis by a photoenzyme could inspire green chemistry applications

By Marten H. Vos

Enzymes catalyze biochemical reactions at high rates and specificity. Structural characterization of transition states and intermediates of enzymatic catalysis at the atomic level provides fundamental understanding and could reveal bioengineering possibilities. For most enzymes, real time studies are limited by substrate binding kinetics. By contrast, in natural photoenzymes, enzyme-substrate complexes can be pre-formed and catalysis initiated by ultrashort light pulses. The photoenzyme CPD DNA photolyase repairs the potentially mutagenic DNA cyclobutane pyrimidine dimers (CPDs) that are induced by ultraviolet light. On pages XXX and YYY of this issue, Maestre-Reyna *et al.* (1) and Christou *et al.* (2), respectively, report time-resolved crystallographic studies of photolyase, characterizing a wealth of functional steps spanning the picosecond to microsecond timescale, including electron transfer, substrate bond breaking and product migration. Beyond establishing the order of these events, these findings may inspire engineering of sunlight driven biobased catalysis.

Natural photoenzymes form stable complexes with substrate molecules in the dark (3). Catalytic transformation of substrates to products then occurs upon photon absorption by a molecule within this complex. Very few natural photoenzymes are known, but these are important in several aspects. Using sunlight directly to drive catalysis, they are intrinsically “sustainable” bioreactors. Moreover, the breadth of photoproducts can be expanded by engineering photoenzymes for green chemistry photocatalysis, or by constructing enzyme variants inspired by photoenzymes. On a more fundamental level, using ultrashort light pulses allows visualization of photoenzyme catalytic intermediates in real-time with femtosecond resolution, i.e., down to the timescale of intrinsic nuclear motions. Dissecting such natural catalytic pathways is also informative about non-photochemical enzymes.

Three different photoenzymes are known to date. Catalysis by light-dependent protochlorophyllide oxidoreductase (involved in chlorophyll biosynthesis) is initiated by excitation of

the substrate itself. The two other photoenzymes, photolyase and the algal enzyme fatty acid photodecarboxylase (FAP), both harbor a blue-light absorbing flavin-adenine dinucleotide (FAD) co-enzyme, but their photochemical mechanisms are markedly different. FAD can adopt three different redox states (4), and both enzymes produce semi-reduced FAD radical intermediates. FAP-mediated photoreduction of the oxidized flavin leads to oxidation of the nearby substrate followed by bond breaking, whereas photolyase-mediated photooxidation of the fully reduced FADH⁻ leads to CPD reduction followed by (double) bond breaking (5). Of the three enzymes, only photolyase is also found in non-photosynthetic organisms, although not in placental mammals.

The mechanisms of the three photoenzymes have been studied by time-resolved optical spectroscopy. However, structural snapshots of intermediates with atomic resolution have hitherto only been obtained in FAP (6). Upon oxidation of the fatty acid substrate by the excited flavin, the bond between the hydrocarbon chain and the CO₂ moieties breaks. These structural studies helped to reveal that this bond breaking occurs in hundreds of picoseconds, concomitant with the preceding electron transfer, in full agreement with spectroscopy.

The method of choice for time-resolved structural studies is serial femtosecond crystallography (TR-SFX) using X-ray Free Electron Lasers (XFEL) (7), which became available about a decade ago. Five facilities offering TR-SFX are now operational worldwide, three of which have contributed to the work of Maestre-Reyna *et al.* and Christou *et al.* XFEL sources provide extremely bright and short (sub-10 fs) X-ray pulses allow diffraction from protein microcrystals before any radiation damage (8) (diffract-before-destroy). Using many individual microcrystal diffraction patterns enables the determination of reliable steady-state structures of proteins that are prone to alteration by X-ray radiation (1, 6, 9). When combined with ultrafast visible pulses, intermediate structures in light-induced processes can be determined.

Finding appropriate conditions for the experiments described by Maestre-Reyna *et al.* and Christou *et al.* was challenging. Photolyase is often unstable, and the enzyme from the

anaerobic archaeon *Methanosarcina mazei* appears most suitable to form photolyase-DNA complex microcrystals, which must be produced at the XFEL facilities, under anaerobic and light-shielded conditions. Time-resolved spectroscopy on photocatalysis in solution has been previously performed on other photolyases (10, 11), but not on the *M. mazei* enzyme, presumably due to its low solubility. It is remarkable that these TR-SFX studies are not preceded by any time-resolved spectroscopic characterization of the same process, as with other colored proteins.

The experiments were performed in an intensity regime of visible light just above the level where differences with the structures obtained without illumination become assignable. Yet this regime is far above the linear, physiological, regime where less than one photon is absorbed per protein. The necessity of working in this regime, common to nearly all TR-SFX studies in proteins, is not fully understood and presently of intense interest (12-14). Maestre-Reyna *et al.* and Christou *et al.* argue that in their studies this issue has limited effect, because the catalytic processes are slower than the expected non-linear effects and in general kinetic agreement with linear-regime spectroscopic studies in other photolyases.

The CPD substrate, which consists of a pair of doubly cross-linked adjacent thymine or cytosine bases, initially bulges from the damaged DNA strand into the enzyme. Although the emphasis of the findings differs, both studies come up with a coherent picture of the order and timing of events leading to the restoration of the intact DNA double helix (see the figure). Upon population of the FADH^{*} excited state, four chemical reactions are distinguished within the ~10⁻¹⁰ – 10⁻⁹ s timespan. Flavin to CPD electron transfer occurs. Reducing the CPD breaks its two crosslinks (also without photolyase present). In the *M. mazei* photolyase, the CPD crosslinks were found to break one after the other with sizeable delay both upon reduction and between the bond ruptures. Subsequently the electron returns to the flavin. These events occurred non-concomitantly, which allowed the structural reorganizations associated with the subsequent processes to be distinguished. Whether the two bonds break simultaneously or sequentially has been intensely discussed based on spectroscopy and quantum

Laboratoire d'Optique et Biosciences, Centre National de la Recherche Scientifique, Institut National de la Santé et de la Recherche Médicale, École Polytechnique, Institut Polytechnique de Paris, 91120 Palaiseau, France. Email: marten.vos@polytechnique.edu

1 yields (QY, the number of products formed per
2 absorbed photon; sub-unity QY arises from
3 back reactions) for other photolyases (10, 11).
4 The results of Maestre-Reyna *et al.* and Chris-
5 tou *et al.* settle the issue for the *M. mazei* en-
6 zyme; yet, given the close timescales, and the
7 enzyme-dependent QY, the situation could
8 vary between different photolyases.

9 Photolyase catalysis in the strict chemical
10 sense is completed within a few nanoseconds.
11 Two distinct steps occur on a much slower
12 timescale ($>10^7$ s), corresponding to sequential
13 migration out of the active site and partial dis-
14 ordering of the two DNA bases. As pointed out
15 in both studies, this trajectory may be influ-
16 enced by the crystalline rather than solution
17 environment, hindering full restoration of the
18 repaired DNA double helix structure.

19 Intriguingly, Christou *et al.* reported a
20 striking 'flip' of the three-ring flavin chromo-
21 phore along the butterfly bend angle at 3 ps,
22 suggested to functionally stabilize the FADH*
23 excited state. At such short times, the heat
24 from multiple-photon absorption is not dissi-
25 pated and future studies should investigate this
26 feature under linear excitation conditions. A re-
27 lated open question is the origin of the sub-
28 unity QY, reported by Maestre-Reyna *et al.* at
29 ~ 0.25 , lower than for other photolyases. In nei-
30 ther of the two studies a corresponding strong
31 decay of the product states is reported. This
32 suggests that wasteful back reactions compete
33 with the primary catalytic electron transfer on
34 the picosecond timescale, an issue that can be
35 addressed by time-resolved spectroscopy.

36 Together, the studies of Maestre-Reyna *et al.*
37 *et al.* and Christou *et al.* give detailed structural
38 insight in a complex catalytic mechanism and
39 at the same time open up important new ave-
40 nues for structural, spectroscopic as well as ex-
41 tended quantum computational investigations
42 to map out the entire reaction pathway and en-
43 ergetics based on the observations. Moreover,
44 they are expected to promote further develop-
45 ment of engineered flavin-dependent light-
46 driven biocatalysts inspired by photolyase pho-
47 tochemistry initiated by excitation of reduced
48 flavin. These green chemistry systems catalyze
49 a rapidly expanding range of reactions, includ-
50 ing highly specific radical cyclizations and
51 polymerizations (15), but are yet far less effi-
52 cient than native systems (4).

53 REFERENCES AND NOTES

- 54 1.M. Maestre-Reyna *et al.*, *Science* **382**, XXX
55 (2023).
56 2.N. Christou *et al.*, *Science* **382**, YYY (2023).
57 3.A. Taylor, D. J. Heyes, N. S. Scrutton, *Curr. Opin.*
58 *Struct. Biol.* **77**, 102491 (2022).
59 4.B. Zhuang, U. Liebl, M. H. Vos, *J. Phys. Chem. B*
126, 3199 (2022).
5. A. Sancar, *J. Biol. Chem.* **283**, 32153 (2008).
6.D. Sorigué *et al.*, *Science* **372**, eabd5687 (2021).

7. A. M. Orville, *Curr. Opin. Struct. Biol.* **65**, 193
(2020).
8. R. Neutze, R. Wouts, D. van der Spoel, E. Weck-
ert, J. Hajdu, *Nature* **406**, 752 (2000).
9. H. Lebrette *et al.*, *Science* **382**, 109 (2023).
10. Z. Liu *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* **108**,
14831 (2011).
11. V. Thiagarajan, M. Byrdin, A. P. M. Eker, P. Mül-
ler, K. Brettel, *Proc. Natl. Acad. Sci. U.S.A.* **108**,
9402 (2011).
12. T. R. M. Barends *et al.*, *bioRxiv*,
2022.2011.2022.517513 (2022).
13. J. E. Besaw, R. J. D. Miller, *Curr. Opin. Struct. Biol.*
81, 102624 (2023).
14. M. L. Grünbein *et al.*, *Nature Methods* **17**, 681
(2020).
15. M. A. Emmanuel *et al.*, *Chem. Rev.* **123**, 5459
(2023).

ACKNOWLEDGMENTS

Thanks to U. Liebl and M. Sliwa for critical reading of
this Perspective. M.H.V. receives support from
ANR grants ANR-18-CE11-0021 and ANR-23-
CE29-0010.

10.1126/science.adl3002

PHOTO CREDIT GOES HERE

Light-induced DNA repair monitored by TR-SFX
Figure Legend: Structures of catalytic intermedi-
ates of were determined by photolyase micro-
crystals diffracting intense X-ray pulses arriving at
known delays after visible light excitation pulses.
At least six distinct intermediates are distin-
guished between the photolyase-CPD enzyme-
substrate complex (left) and the recovered intact
double helix (right) spanning more than six orders
of magnitude in time. Cartoons representing the
four temporally distinct chemical steps occurring
on the hundreds of picoseconds to a few nano-
seconds time range are shown. These are based
on PDB 5ZCW and figures in (1), but do not cor-
respond directly to the structural models. Also re-
ported Intermediates on the microsecond time-
scale along the trajectory of the repaired bases
out of the active site towards the intact double
helix are not represented. Details of the interme-
diates in (1, 2).