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Structure and seasonal variability in fish food webs in a small macrotidal estuary (Canche estuary, Eastern English Channel) based on stable carbon and nitrogen isotope analysis

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Abstract:

Estuaries are highly important nursery habitats for a range of fish species because they provide refuge and food, especially for juveniles. However, the importance of specific salinity zones and adjacent shallow marine habitats (subtidal and surf zones) for fish feeding is not well understood, particularly in small macrotidal estuaries. Using the example of the Canche estuary, which is considered a reference for small macrotidal estuaries in France, we investigated the structure and seasonal variability in fish food webs based on stable carbon and nitrogen isotope analysis. Our results provide a new vision of the Canche estuary, which has been considered a major feeding ground for all marine fish that spend time there. Based on C results, our results revealed that organic matter of terrestrial origin has little influence on functioning of the Canche food web, except for flounders in the upstream area of the estuary. Conversely, microphytobenthos and marine particulate organic matter contribute most to the food web of fish in the estuary. Our study also revealed that some fish species visit the estuary for reasons other than feeding, such as to avoid predation or because they are carried by the tide. This work confirmed the suitability of using stable isotopes to trace fish fidelity to feeding grounds less than 10 km apart.

Keywords: Food web, macrotidal estuary, stable isotopes, seasonal variations, nursery

1. Introduction

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Using shallow marine coastal zones and estuaries as nursery areas is an important phase in the 36 life history of many marine organisms, including commercially valuable species (Amara, 37 2003; Vasconcelos et al., 2007). In temperate ecosystems, juvenile fish concentrate in nursery 38 grounds from late spring to early fall, which is the peak of macrobenthic prey biomass 39 (Amara and Paul, 2003; Pasquaud et al., 2010; Selleslagh et al., 2015). Refuge and feeding 40 areas for young fish in estuaries are considered important for fish survival and replenishing 41 42 coastal fish stocks (McLusky and Elliott, 2004). However, many authors have shown that estuaries are only one component of broader nursery-use patterns and that shallow marine 43 44 coastal areas and estuaries may provide a mosaic of vital nursery habitat types for juvenile marine fish (Beck et al., 2001). Although opportunism is widely reported for estuary-45 associated fish, as well as for fish in general (Amara et al., 2001; Elliott et al., 2007; Elliott 46 47 and Hemingway, 2002), the importance of these different habitats (specific salinity zones inside estuaries and adjacent marine subtidal and surf zones) for fish feeding is not well 48 understood (Selleslagh et al., 2015; Vinagre et al., 2008). In addition, there is still a lack of 49 information and confusing conclusions about the main origin of the organic matter that 50 sustains juvenile fish food webs in nursery grounds (Le Pape et al., 2013). For estuaries, some 51 studies (Darnaude et al., 2004; Leakey et al., 2008; Vinagre et al., 2008) indicated that 52 53 allochthonous organic matter of terrestrial origin predominated, while other studies suggested 54 that marine organic matter predominated (Pasquaud et al., 2008; Selleslagh et al., 2015). Most 55 of these studies were conducted in large estuaries (e.g. Tagus, Gironde, Thames, Rhone, Vilaine). However, in small estuaries with little freshwater influence, in situ primary 56 production can override other food sources and contribute significantly to juvenile food webs 57 58 (Kostecki et al., 2012). Understanding the main ecological processes in ecosystems (e.g. food 59 sources, trophic transfer through the food web) and identifying juvenile fish feeding areas and their spatial use of shallow nursery habitats are fundamental issues for effective conservation 60 and management of these essential fish habitats (Hobson et al., 1999). 61 62 Reconstructing marine food webs is largely limited by methodological difficulties. A 63 traditional approach to describe fish feeding ecology and determine their feeding niche has been stomach content analysis (Amara et al., 2001; Besyst et al., 1999). Although it may 64 provide high taxonomic resolution, this method can be biased due to the difficulty in 65 determining the origin of partially digested food items. Previous studies of estuarine fish food 66 webs highlighted that stable isotope analysis (SIA), based on nitrogen (δ^{15} N) and carbon 67 $(\delta^{13}C)$ signatures of various food-web compartments, can be a powerful tool. This technique, 68

69 which identifies trophic pathways and energy sources in a variety of ecosystems (Fry, 2006; 70 Peterson and Fry, 1987), is particularly successful in coastal and estuarine systems, in which the fate of different sources of both freshwater and marine particulate organic matter (POM) 71 can be distinguished (Fry, 1999; Riera et al., 1999; Yokoyama et al., 2008). 72 δ 13C and δ 15N are typically increased from prey to consumers by 3‰–4‰ for δ 15N and 1‰ 73 for δ13C (De Niro and Epstein, 1981; Minagawa and Wada, 1984; Cabana and Rasmussen, 74 1994). Thus, δ15N can indicate the trophic position of an organism within the food web and 75 76 δ13C values of the food sources (primary producers as phytoplankton or microphytobenthos) 77 are also responsible for differences in the isotopic compositions of their consumers (Riera et 78 al. 1999), allowing distinctions to be made between pelagic and benthic food webs (Hobson et al. 2002) or fresh vs marine waters (Fry, 2006). Consequently, δ13C can be used as a tracer of 79 organic trophic sources for benthic consumers (Peterson, 1999). 80 81 Given the diversity of juvenile nursery habitats, an isotopic approach to tracing fish movement is particularly appealing as it increases the likelihood of finding habitat-specific 82 isotopic signatures (Herzka, 2005). Several authors have successfully used stable isotopes to 83 study, for example, the connectivity of habitats (Fry et al., 2003; Selleslagh et al., 2015; 84 85 Vinagre et al., 2008). In the present study, we analysed for the first time the fish food web in a small macrotidal 86 estuary on the French coast of the Eastern English Channel (EEC) - the Canche estuary -87 based on stable carbon and nitrogen isotopes analysis. The Canche estuary is considered a 88 reference for small macrotidal estuaries because it is subject to low human disturbance and is 89 an important fish nursery ground (Selleslagh et al., 2009; Selleslagh and Amara, 2015) that 90 supports species with high commercial and recreational value during their juvenile stage. The 91 92 role and importance of small macrotidal estuaries as feeding grounds for marine juvenile fish is not well understood. Along the French coast of the EEC, juvenile marine fish are known to 93 use both estuaries and shallow marine coastal waters, but the exact areas where they feed 94 95 remain unknown (Amara and Paul, 2003; Selleslagh and Amara, 2015). 96 The main objectives of the present study were to i) distinguish the origin of sources in the 97 marine juvenile fish food web, ii) investigate seasonal variations in fish food-web structure and iii) assess the feeding-ground fidelity of marine juvenile fish that inhabit the Canche 98 estuary and adjacent coastal nursery grounds. 99

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2. Materials and Methods

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The Canche estuary (50°50'-50°56' N, 1°57'-1°67' E) is located in northern France along the 104 105 coast of the Eastern English Channel (EEC). The Canche estuary is 12 km long and has a maximum width of 1 km at its mouth. The estuary has a semi-diurnal tide, with an average 106 tidal range of ca. 1 m at neap tides and 6 m at spring tides, and is considered a macro/hyper-107 108 tidal estuary according to the McLusky and Elliott (2004) classification. Water circulation depends mainly on the tide and small freshwater inputs, with a mean annual rate of ca. 13 109 110 m³.s⁻¹. The Canche estuary is little impacted by human disturbances (Amara et al., 2007), it belongs to the special conservation zone "Bay of Canche and corridor of the 3 estuaries" and 111 112 is classified as a "Natura 2000" site because it accounts as a major area of hosting juvenile 113 fish in the Eastern English Channel.

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2. 2 Sampling strategy

- 116 Samples of particular organic matter (POM), sediment organic matter (SOM),
- 117 microphytobenthos (MPB) as well as fish and their main potential prey (i.e. benthic
- organisms, shrimp and crabs) were collected in the fall (October-November 2017) and spring
- 119 (May-June 2018) at three sites inside the Canche estuary along a salinity gradient (upstream,
- 120 middle and downstream).
- To better understand the estuary's feeding role and connectivity with adjacent marine habitats,
- three additional sites were sampled outside the Canche estuary: two in the surf zone on both
- 123 sides of the mouth of the estuary (Sainte Cécile beach and Le Touquet beach, respectively)
- 124 and one in the subtidal zone in the plume of the estuary (Figure 1). For technical reasons,
- 125 these sites were sampled only in spring (May-June 2018). We also described benthic
- communities and their biomass at all six sites only in spring (May-June 2018).
- 127 The sampling sites consist of a variety of habitats. The middle part of the Canche estuary is a
- muddy-sand shore that contains polychaetes and bivalves (EUNIS classification A2.24; Rolet
- 129 et al., 2015). The downstream part is characterized by a medium fine-sand benthic community
- 130 (EUNIS classification A2.223) and is dominated by amphipods and *Scolelepis spp*. (Rolet et
- al, 2015) such as the surf zone (Sainte Cécile and Le Touquet beaches). The subtidal site is
- muddy fine sand and contains polychaetes (Magelona jonhsoni, Nephtys spp.) and bivalves
- 133 (Donax vittatus) (Desroy et al., 2003).

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135	2.3 Particulate organic matter and sediment organic matter sampling
136	The POM was sampled at the surface at high tide using sterile pots and the

a cool box. In the laboratory, the water was filtered through pre-combusted Whatman GF/F

then was conserved in

- 138 filters (0.45 µm pore size, 47 mm diameter), with 3 replicates per site. The SOM was sampled
- at low tide by scraping the top first cm of sediment. Three replicates were performed at each
- 140 site. The POM and SOM samples were conserved at -20 °C in the laboratory until transfer for
- 141 SIA.

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2.4 Microphytobenthos sampling

144 Benthic diatoms were collected at the sites inside the estuary and in the surf zone at low tide by scraping the surface of the sediment and were then extracted according to the protocol of 145 Riera et al. (1999). In the laboratory, we allowed MPB to migrate by leaving the sediment 146 containing benthic diatoms in flat trays to form a layer 1 cm thick. A nylon screen (60 µm 147 mesh) was placed on top of the sediment and covered with a layer of combusted sand powder 148 149 (60-200 µm) 5 mm thick. The trays were illuminated until the first dense brown mats appeared on the surface. Meanwhile, the sand was kept moist by spraying filtered (GF/F) 150 151 seawater from the sampling site. The top 2 mm of sand were removed and sieved over a 60 µm mesh nylon screen to separate the diatoms from the remaining sand and nematodes or 152 copepods. The benthic diatoms were then collected on precombusted GF/F filters. Samples 153

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2.5 Benthic organism sampling

were conserved at -20 °C until transfer for SIA.

In the surf zone (on both sides) and inside the estuary, macrobenthic fauna was sampled during low tide with a hand corer (0.025 m^2 area, 20 cm depth, 10 replicates for density and biomass, and as many as needed for SIA). At the subtidal site, macrobenthic organisms were sampled using a Van Veen grab (sampling an area of 0.1 m^2 , 10 replicates for density and biomass and as many as needed for SIA). Samples were washed, sieved through a 1 mm mesh size and then washed again with milli-Q water to avoid contamination. In the laboratory, benthic fauna was sorted and identified to the species level when is possible. Ash-free dry weight (AFDW) of benthic invertebrates was determined using the method recommended by the Benthos Ecology Working Group of the ICES (Hamilton and Kingston, 1985). The fauna was dried in an oven at 60 ± 1 °C for at least 48 hours until a constant weight was obtained. It

was then weighed (with 10^{-1} mg precision) and placed in an oven at $520 \pm 20^{\circ}$ C for 6 h to 167 calcinate the organic matter completely without altering the mineral matter, and the calcinated 168 fauna was weighed again. The AFDW equalled the difference between the two weights. 169 170 2.6 Fish, crab and shrimp sampling 171 Fish, crabs and shrimps were sampled during daylight hours using a 1.5 m beam trawl, with 172 one tickler chain and 5 mm mesh size in the cod end, towed by a semi-rigid boat against the 173 174 current at 2 knots for 15 min. Fish, crabs and shrimp were identified to the species level, counted, and then measured (total length, with 1 mm precision). 175 176 177 2.7 Stable isotope analyses Species selected for SIA were dominant in both abundance and biomass to obtain a synthetic 178 image of the trophic structure within each community. As lipids are depleted in δ^{13} C 179 compared to carbohydrates and proteins (DeNiro and Epstein, 1977; Griffiths, 1991), which 180 indicates that fatty tissues tend to be isotopically lighter than leaner ones, trophic 181

compared to carbohydrates and proteins (DeNiro and Epstein, 1977; Griffiths, 1991), which indicates that fatty tissues tend to be isotopically lighter than leaner ones, trophic interpretations based on δ^{13} C composition may be confounded by lipid effects (Bodin et al., 2007; Wada et al., 1993). To minimize these effects, mega- and macrofaunal (except for polychaetes) low-lipid muscle tissue was used for SIA. Polychaetes were analysed after removing their viscera by dissection. The valve muscle of bivalves, the abdomen muscle of shrimp, the muscle in crab pincers and the white dorsal muscle of fish (even small ones) were dissected and analysed for SIA. For other benthic organisms, the entire organism was analysed after removing the digestive tract, jaws and cerci. The tissues were then washed with

As fish size can influence isotope values, especially $\delta^{15}N$, due to ontogeny (Galván et al., 2010; Wilson et al., 2009), we carefully selected individuals of similar size across species in order to be sure that we select G0 juveniles. Before $\delta^{13}C$ analyses, POM and MPB filters were divided into two subsamples: one was exposed to HCl vapour for 4 h to remove residual carbonates (Cresson et al., 2012) before being placed in tin cups (Lorrain et al., 2003), while the other was not treated and was used to measure $\delta^{15}N$. Sediment samples were dried at 60°C for 24 h. They were divided into two subsamples: one was treated with HCl, to remove

milli-Q water to prevent contamination and freeze-dried before being encapsulated. For small

benthic organisms (Bathyporeia pilosa, Eurydice pulchra, Gastrosaccus spinifer, Haustorius

arenarius), each sample represented a combination of 2-4 individuals.

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199	carbonate, then rinsed three times with distilled water, and freeze-dried for 48 hours before
200	encapsulation, while the other was encapsulated immediately after drying.
201	$\delta^{13}C$ and $\delta^{15}N$ were measured using an elemental analyser Flash EA 2000 (Thermo
202	Scientific), connected to an Isotope Ratio Mass Spectrometer (Delta V+) with a ConFlo IV
203	interface (Thermo Scientific) at the Pôle Spectrométrie Océan in Plouzané, France. Replicate
204	analyses of international IAEA and laboratory USGS standards provided analytical errors
205	<0.20% for both $\delta^{13}C$ and $\delta^{15}N$. Stable isotope ratios were expressed as parts per mil (‰) in
206	the δ notation relative to the Pee Dee Belemnite standard for carbon and atmospheric N_2 for
207	nitrogen using the formula:
208	$\delta X(\%) = [(R sample/R standard) - 1] \times 1000$
209	where X is ¹³ C or ¹⁵ N, and R is the ratio of ¹³ C: ¹² C or ¹⁵ N: ¹⁴ N.
210	
211	2.8 Data analysis
212	2.8.1 Statistical analysis
213	We first tested the hypothesis that potential sources and prey for fish had significantly
214	different isotopic compositions along the salinity gradient, and then that compositions at sites
215	inside vs. outside of the estuary differed significantly. For all samples, two-way ANOVAs
216	were performed separately for each ratio, factors are sites and seasons (after verifying that the
217	assumptions of parametric tests were met). ANOVA was used to test differences in $\delta^{13}\text{C}$ and
218	$\delta^{15}N$ in the POM from the water sources, considering site and season effects. Biplots of $\delta^{13}C$
219	vs. $\delta^{15}N$ were used to represent graphically means and standard deviations of isotopic
220	compositions of all compartments of the entire food web in each habitat. Besides, we
221	performed a Kruskal-Wallis test to test for fish species length homogeneity between sites and
222	seasons.
223	
224	2.8.2 Layman metrics
225	We assessed isotopic niches of fish by calculating the three Layman metrics (Layman et al.,
226	2007): nitrogen range (NR), carbon range (CR) and total area (TA). We used a Bayesian
227	approach based on multivariate ellipse-based metrics (Jackson et al., 2011), in which the
228	location of the centroid represents the centre of the trophic niche in isotopic space. To
229	describe the spread of data points, we calculated the parameters developed by Layman et al.

(2007). We constructed convex hulls to estimate the smallest TA that contained all individuals in the isotopic space. The TA can be interpreted as a measure of the total isotopic niche of a population (Layman et al., 2007). We estimated the niche width in each season using multivariate ellipse-based metrics (Jackson et al., 2011). The analysis generates standard ellipse areas (SEA), which are bivariate equivalents of standard deviations in univariate analyses.

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3. Results

3.1 Benthic communities and biomass

The upstream site was too muddy to sample benthic organisms. Among the five sites sampled in spring, we identified 1288 individuals that belonged to 51 taxa (Table 1). Mean macrobenthic species richness, abundance and biomass were higher outside than inside the estuary (Wilcoxon test, p < 0.0001). Species richness was lowest in the middle of the estuary (6 species) but higher at the downstream site (16), Sainte Cécile beach (13), Le Touquet beach (11) and the subtidal site (32). The main species observed in the middle of the estuary were the polychaetes Hedistes diversicolor (326 ind.m-2; 2011 mg.m-2) and the bivalve Scorbicularia plana (565 mg.m⁻²). The polychaete Scolelepis squamata (823 ind.m⁻²; 3 445 mg.m⁻²) and the bivalves Limecola balthica (1 282 mg.m⁻²) and Cerastoderma edule (640 mg.m⁻²) dominated the downstream site. The two surf zone sites (Le Touquet and Sainte Cécile) were characterized by the polychaetes S. squamata (48-300 ind.m⁻²; 145-835 mg.m⁻²), Nephtys cirrosa (124-160 ind.m⁻²; 559-831 mg.m⁻²) and Lanice conchylega (1 022 mg.m⁻² at Sainte Cécile) and the bivalves M. balthica (153-1336 mg.m⁻²), C. edule (693 mg.m⁻² at Sainte Cécile) and Fabulina fabula (706 mg.m⁻² at Le Touquet). More diverse taxa (N = 32) and huge benthic biomass were observed at the subtidal site, which was dominated mainly by the bivalves D. vittatus (516 ind.m⁻²; 108 035 mg.m⁻²), F. fabula (412 ind.m⁻²; 12 906 mg.m⁻²) and Ensis leei (36 ind.m⁻²; 9 064 mg.m⁻²).

Table 1. Mean Biomass (B) (mg.m⁻²) and Density (D) (ind.m⁻²) of each benthic species inside and outside the Canche estuary in spring. (Abbreviations correspond to species analysed for stable isotope composition)

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\	Inside the Canche estuary	Canc]	he estu	ıary		Outsi	Outside the Canche estuary	he estua	ry	
Species	Middle	Ω	Downstream	eam	Sainte Céci	le beach	Sainte Cécile beach Le Touquet beach	t beach	Subtidal	al
	B D		В	D	В	D	В	D	В	D
Hediste diversicolor (H div)	2011.7 32	326								
Scrobicularia plana (Spla)	565.8	14								
Carcinus maenas (C mae)	108.6		85.5	_						
Oligochaeta		2	1.4	5						
Gammarus zaddachi		4								
Bathyporeia pilosa	0.4		15.1	61						
Scolelepis squamata (S squ)		347	3445.6	823	835.8	300	145.9	48		
Limecola balthica (L bal)		128	1282.7	21	153.6	4	1336.6	12	8990.1	72
Cerastoderma edule (C edu)		-64	640.5	1	693.6	4			2709.8	4
Eurydice pulchra (E pul)		24	246.5	κ			3.3	16		
Nephtys cirrosa (N cir)		14	146.2	119	559.4	124	831.72	160		
Heteromastus filiformis		5	29.1	32						
Bathyporeia sarsi		4	5.	36	5					
Eteone longa (E lon)			11.6	∞						
Glycera tridactyla		∞	5.5	3	62.1	4			7.8	4
Haustorius arenarius (H are)		4	4.1	4	42.4	28	9.5	4		
Urothoe poseidonis		_	5.	4	17.0	40	3.4	%	2.7	4
Pygospio elegans		_	0.4	4						
Mytilus edulis		_	.3							
Lanice conchilega (L con)					1022.8	16			416.2	4196
Portumnus latipes					447.9	8	257.4	∞	7	7
Nepthys hombergi					295.6	24			981.6	124
Gastrosaccus spinifer (G spi)					57.7	28	59.6	16		
Crangon crangon (C cra)					28.0	52	1.8	4	2.2	4
Spio martinensis					7.1	16			7	
Fabulina fabula (F fab)							706.4	12	12906.6	412
Cumopsis goodsiri							0.4	4		
Donax vittatus (D vit)									108035.2	216
Ensis leei (E lee)									9604.8	36
Tritia reticulata									2630.4	40
Ophiura albida									9.868	∞
Abra alba (A alb)									603.6	89
Arenicola marina (A mar)									586.0	24

Liocarcinus vernalis								465.9	36	
Sigalion squamosus								443.1	4	
Anapagurus hyndmanni								377.4	12	
Nephtys assimilis								244.3	4	
Lagis koreni								218.7	36	
Phyllodoce mucosa (P muc)								144.4	244	
Planarian								118.8	4	
Pinnotheres pisum	A							74.6	12	
Nemertians								42.2	32	
Eteone picta								9.7	4	
Schistomysis kervillei								8.4	4	
Diastylis rugosa								5.8	4	
Capitella capitate (C cap)								2.8	20	
Leucothoe incisa		,						2.4	∞	
Eumida sanguinea						4		1.1	∞	
Nototropis swammerdamei								9.0	4	
Pariambus typicus				7				0.5	4	
TOTAL	2 698.1	348	5 922.4	1 404	2 698.1 348 5 922.4 1 404 4 223.0	648	3 356.0	292 150 536.3 5 992	5 992	

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3.2 FISH	sambled for	stable is	otobe ana	uvses

260	Nine fish species (eight in spring and six in fall) were collected for SIA: Buglossidium luteum,
261	Dicentrarchus labrax, Limanda limanda, Platichthys flesus, Pleuronectes platessa, Solea
262	solea, Sprattus sprattus, Pomatoschistus microps and Pomatoschistus minutus (Table 2).
263	Most individuals were G0 juveniles, except P. microps, P. minutus, S. solea and P. platessa
264	(at the subtidal site in spring) and P. flesus (at Sainte Cécile beach in spring). S. solea, L.
265	limanda and B. luteum were collected only outside the Canche estuary.
266	In the fall, fish of the same species did not differ significantly in size among sites, except P.
267	flesus ($p = 0.020$,) and S. sprattus ($p = 0.006$), which were smaller in the middle of the estuary
268	than upstream or downstream. In spring, P. platessa were significantly longer at the subtidal
269	site (p < 0.0001). P. flesus were also significantly longer at Sainte Cécile than at the other
270	sites (p = 0.003). Inside the estuary, fish size varied between the two seasons for the four
271	species caught in both spring and fall (D. labrax, P. flesus, S. sprattus and P. platessa; p <
272	0.0001)

Table 2. Total length (mean ± standard error, cm) and samples size (n) of the nine fish species collected in the Canche estuary in two seasons (fall and spring) analysed for carbon and nitrogen stable isotope composition. 273 274

Sussing	Coogn	Upstream	u	Middle		Downstr	eam	Sainte Céc	ile beach	See Upstream Middle Downstream Sainte Cécile beach Le Touquet Beach Subtidal	t Beach	Subtidal	
Species	Season	Length 1	u	Length	u	Length	u	Length	u	Length n Length n Length n Length	u	n Length	u
Dicentrarchus. labrax (D lab)		5.9±0.9 1	10	5.2±0.9	10	5.9±0.9 10 5.2±0.9 10 6.3±0.5	11						
Platichtthys flesus (P fle)		6.0 ± 3.0 11 3.8 ± 0.2 10 6.9 ± 1.3	11	3.8 ± 0.2	10	6.9 ± 1.3	6						
Pleuronectes platessa (P pla)	110					6.6 ± 1.3	S						
Sprattus sprattus (S spr)	Lall	6.9 ± 2.8 11 3.9 ± 0.2 6 6.4 ± 1.2	11	3.9 ± 0.2	9	6.4 ± 1.2	6						
Pomatochistus microps (P mic)		4.4±0.2 5 3.8±0.2 5	5	3.8 ± 0.2		4.6 ± 0.7	S						
Pomatochistus minutus (P min)		5.3±0.5 ₺	5	4.1 ± 0.1	S	5.3±0.5 5 4.1±0.1 5 5.9±0.3	S						
Dicentrarchus labrax (D lab)	7			8.2±1.1	4	8.2±1.1 4 8.4±0.5	3						
Platichtthys flesus (P fle)		7.8±2.8 4 6.4±2.8 5	4	6.4 ± 2.8		8.7 ± 5.2	4	13.0 ± 1.2	9	10.6 ± 3.1	5	7.0 ± 5.4	\subseteq
Pleuronectes platessa (P pla)				3.9±4.8	5	$.9\pm4.8$ 5 4.6 ± 1.0	4	5.5±7.4	2	7.1 ± 0.9	6	14.5 ± 5.2	4
Sprattus sprattus (S spr)	C	4.3±3.0 10 4.8±3.7 5	10	4.8 ± 3.7	2	4.5±0.4	8	4		6.4 ± 1.5	5		
Pomatoschistus sp. (P sp)	Smile		1	4.3±2.8 4		4.8±2.5	S						
Solea solea (S sol)								3.5±2.4	7	3.7±2.2	7	13.1 ± 2.1	∞
Buglossidium luteum (B lut)) 			7.6±3.8	5	4.1 ± 2.3	S	5.9 ± 4.2	5
Limanda Limanda (L lim)						1						13.4 ± 1.0	9

3.3 Results of stable isotope analyses

276 3.3.1 Food sources

275

The stable isotope composition of POM ranged from -29.70% to -20.15% for δ¹³C and 5.20-277 7.30% for δ^{15} N (Table 3). POM δ^{13} C differed significantly among sites (F = 9.536; p < 278 0.0001, ANOVA)) and between seasons (F = 9.448; p < 0.0001, ANOVA). In spring, mean 279 POM δ^{13} C was significantly lower inside the estuary (F = 9.536; P < 0.0001) and higher at the 280 subtidal site (-20.89 \pm 0.10%) (Table 3). Mean $\delta^{15}N$ also differed significantly among sites (F 281 = 3.745; P < 0.0001) and between seasons (F = 3.762; P < 0.0001). POM δ^{15} N was 282 significantly higher at Le Touquet beach $(7.30 \pm 0.01\% \text{ P} = 0.144, \text{Kruskal-Wallis})$ (Table 3). 283 The stable isotope composition of SOM ranged from -24.18‰ to -19.62‰ for δ¹³C and 4.67-284 7.19‰ for $\delta^{15}N$ (Table 3) and differed significantly among sites and between seasons for both 285 δ^{13} C and δ^{15} N. The ANOVA revealed a site effect on δ^{15} N, as well as a season effect, with the 286 lowest values in spring, and a significant effect of the site × season interaction (F = 10.548, p 287 < 0.0001). SOM δ^{13} C showed significant enrichment from upstream to downstream along the 288 estuary in both spring (F = 9.352, p < 0.0001) and fall (F=4.531, p < 0.0001) (Table 3). The 289 MBP had the most depleted $\delta^{15}N$ ratios of the food sources sampled. The stable isotope 290 composition of MPB ranged from -17.1% to -13.1% for δ^{13} C and 3.66-6.44% for δ^{15} N 291 (Table 3). MBP δ^{13} C and δ^{15} N did not differ significantly among sites or between seasons. 292

Table 3. Carbon and nitrogen stable isotope compositions (mean ± standard error, %) of particulate organic matter (POM), sediment organic matter (SOM) and microphytobenthos (MPB) inside (Upstream, Middle, Downstream) and outside (Sainte Cécile and Le Touquet beaches, Subtidal) the Canche estuary in fall and spring. *only one replicate.

		•		•	4							
000	Ups	tream	Mi	Middle	Down	Downstream	Sainte Co	Sainte Cécile beach Le Touquet beach	Le Touc	quet beach	Subtidal	idal
scaso	N ₅₁ 8	813C	N ₅₁ 8	813C	N ₂₁ 8	§13C	$N^{15}N$	813C	N ₂₁ 8		\$15N 813C	813C
POM	6.61*	-29.70*	6.00 ± 0.05	6.00 ± 0.05 -24.20 ± 0.14 6.96 ± 0.35 -27.43 ± 1.37	6.96 ± 0.35	-27.43 ± 1.37						
SOM Fall	96.00 ± 00.9	-24.18 ± 0.46	7.19 ± 0.62	$0.46 7.19 \pm 0.62 -23.04 \pm 0.36 6.07 \pm 0.57 -22.09 \pm 0.80$	6.07 ± 0.57	-22.09 ± 0.80						
MPB	6.44 ± 1.90 -17.11 ± 0.8	-17.11 ± 0.85	3.66 ± 1.75	-0.85 3.66 ± 1.75 -16.18 ± 0.44 5.94 ± 0.75 -16.12 ± 0.48	5.94 ± 0.75	-16.12 ± 0.48						
POM	5.20 ± 0.66	-29.16 ± 0.21	6.57 ± 0.07	-20.89 ± 0.10	5.96 ± 1.24	-22.43 ± 1.24	6.08 ± 0.72	-20.15 ± 0.23	7.30 ± 0.01	$0.21 6.57 \pm 0.07 -20.89 \pm 0.10 5.96 \pm 1.24 -22.43 \pm 1.24 6.08 \pm 0.72 -20.15 \pm 0.23 7.30 \pm 0.01 -20.89 \pm 0.10 6.50 \pm 0.40 -20.50 \pm 0.17 -20.10 \pm 0.00 -20.10 \pm 0.00 $	6.50 ± 0.40	20.50 ± 0.17
SOM Spring	Spring 5.46 ±0.01	-24.18 ± 0.16	5.39 ± 0.06	-19.62 ± 0.05	4.67 ± 0.94	$0.16 \ \ 5.39 \pm 0.06 \ \ -19.62 \pm 0.05 \ \ \ 4.67 \pm 0.94 \ \ \ -19.98 \pm 0.20 \ \ \ \ 4.80 \pm 0.72 \ \ \ -19.95 \pm 0.92$	4.80 ± 0.72	-19.95 ± 0.92				
MPB			6.40 ± 0.59	-16.13 ± 0.34	5.12 ± 0.37	$6.40 \pm 0.59 -16.13 \pm 0.34 5.12 \pm 0.37 -15.71 \pm 1.41 6.42 \pm 0.16 -15.52 \pm 0.65 4.88 \pm 0.49 -13.14 \pm 0.56 4.88 \pm 0.49 -13.14 \pm$	6.42 ± 0.16	-15.52 ± 0.65	4.88 ± 0.49	-13.14 ± 0.56		

297	3.3.2 Isotopic compositions of benthic communities
298	Inside the estuary, the benthic invertebrate community followed the same isotopic trend in
299	both seasons. The bivalve Scrobicularia plana and polychaetes H. diversicolor and N. cirrosa
300	had the lowest δ^{13} C values (-20% to -18%), while the crustaceans <i>Carcinus maenas</i> and
301	Crangon crangon had the highest δ^{13} C values (-16‰ to -15‰) (Figures 2 and 3). The
302	suspension feeder L. balthica (7.9-9.9%) and the deposit feeders S. squamata (8.2-10.2%)
303	and S. plana (8.4-9.2‰) had the lowest $\delta^{15}N$ values, while the crustaceans C. crangon and C.
304	maenas (11.3-12.6‰) had the highest δ^{15} N values (Appendix 1, Appendix 2).
305	Outside the estuary, S. squamata (-18.8%), L. conchylega (-18.6±0.4%) and E. pulchra
306	(-20.4±0.9‰) had the lowest δ^{13} C values, while the predatory <i>C. maenas</i> , <i>C. crangon</i> and <i>N.</i>
307	cirrosa (-17.1‰ to -15.2‰) had the highest δ^{13} C values (Appendix 1). The suspension
308	feeders D. vittatus, L. balthica and L. conchylega had the lowest $\delta^{15}N$ values (7.1-9.6‰),
309	while the predatory shrimp <i>C. crangon</i> had the highest δ^{15} N values (13.6‰) (Appendix 1).
310	In spring, the mean δ^{13} C values for <i>C. crangon</i> and <i>C. maenas</i> were significantly lower inside
311	the estuary (C. crangon: -18.3±0.9‰ at the upstream site to -15.5±0.7‰ at Sainte Cécile
312	beach; C. maenas: -17.4±0.5‰ at the upstream site to -15.2±0.2‰ at the intertidal site).
313	
314	3.3.3 Isotopic composition of fish
315	Inside the estuary in both seasons, <i>P. flesus</i> had the lowest δ^{13} C values (-24.2% to -16.8%),
316	while in the fall, P . microps had the highest $\delta^{13}C$ values (-15.9% to -14.8%). Inside the
317	
	estuary in the fall, all δ^{15} N values exceeded 13‰, except for <i>P. flesus</i> (12.5±0.6‰) and <i>P.</i>
318	estuary in the fall, all δ^{15} N values exceeded 13‰, except for <i>P. flesus</i> (12.5±0.6‰) and <i>P. plastessa</i> (12.4±0.2‰), both downstream. Fish δ^{15} N was lower in spring (11.5-13.1‰),
318 319	
	plastessa (12.4±0.2‰), both downstream. Fish δ^{15} N was lower in spring (11.5-13.1‰),
319	plastessa (12.4±0.2‰), both downstream. Fish δ^{15} N was lower in spring (11.5-13.1‰), except for <i>D. labrax</i> (ca. 14‰). Thus, inside the estuary we observed enrichment of
319 320	plastessa (12.4±0.2‰), both downstream. Fish δ^{15} N was lower in spring (11.5-13.1‰), except for <i>D. labrax</i> (ca. 14‰). Thus, inside the estuary we observed enrichment of organisms from upstream to downstream (Figure 2). Outside the estuary in spring, carbon
319 320 321	plastessa (12.4±0.2‰), both downstream. Fish δ^{15} N was lower in spring (11.5-13.1‰), except for <i>D. labrax</i> (ca. 14‰). Thus, inside the estuary we observed enrichment of organisms from upstream to downstream (Figure 2). Outside the estuary in spring, carbon ratios displayed the same patterns, with δ^{13} C values lowest for <i>P. flesus</i> (-20.7‰ to -17.2‰)
319 320 321 322	plastessa (12.4±0.2‰), both downstream. Fish $\delta^{15}N$ was lower in spring (11.5-13.1‰), except for <i>D. labrax</i> (ca. 14‰). Thus, inside the estuary we observed enrichment of organisms from upstream to downstream (Figure 2). Outside the estuary in spring, carbon ratios displayed the same patterns, with $\delta^{13}C$ values lowest for <i>P. flesus</i> (-20.7‰ to -17.2‰) and highest for <i>P. microps</i> (-15.6±0.3‰). Fish had a wider range of $\delta^{15}N$ (10.2-13.8‰)
319320321322323	plastessa (12.4±0.2‰), both downstream. Fish $\delta^{15}N$ was lower in spring (11.5-13.1‰), except for <i>D. labrax</i> (ca. 14‰). Thus, inside the estuary we observed enrichment of organisms from upstream to downstream (Figure 2). Outside the estuary in spring, carbon ratios displayed the same patterns, with $\delta^{13}C$ values lowest for <i>P. flesus</i> (-20.7‰ to -17.2‰) and highest for <i>P. microps</i> (-15.6±0.3‰). Fish had a wider range of $\delta^{15}N$ (10.2-13.8‰)
319 320 321 322 323 324	plastessa (12.4±0.2‰), both downstream. Fish δ^{15} N was lower in spring (11.5-13.1‰), except for <i>D. labrax</i> (ca. 14‰). Thus, inside the estuary we observed enrichment of organisms from upstream to downstream (Figure 2). Outside the estuary in spring, carbon ratios displayed the same patterns, with δ^{13} C values lowest for <i>P. flesus</i> (-20.7‰ to -17.2‰) and highest for <i>P. microps</i> (-15.6±0.3‰). Fish had a wider range of δ^{15} N (10.2-13.8‰) outside the estuary than inside.
319 320 321 322 323 324 325	plastessa (12.4±0.2‰), both downstream. Fish $\delta^{15}N$ was lower in spring (11.5-13.1‰), except for <i>D. labrax</i> (ca. 14‰). Thus, inside the estuary we observed enrichment of organisms from upstream to downstream (Figure 2). Outside the estuary in spring, carbon ratios displayed the same patterns, with $\delta^{13}C$ values lowest for <i>P. flesus</i> (-20.7‰ to -17.2‰) and highest for <i>P. microps</i> (-15.6±0.3‰). Fish had a wider range of $\delta^{15}N$ (10.2-13.8‰) outside the estuary than inside.

the estuary, total fish SEAc (corrected SEA) were larger in spring (21.79%) than in fall (13.96%). In spring, fish isotopic niches were smaller outside the estuary than inside.

Table 5. The core isotopic niche area (SEAc, %), carbon range (CR) and nitrogen range (NR)

for fish species sampled inside and outside the Canche estuary in fall and spring.

		Fall				Sp	ring		
		Inside			Inside			Outside	
Species	SEAc	CR	NR	SEAc	CR	NR	SEAc	CR	NR
Pomatoschistus minutus	17.58	9.01	2.63				7		
Platichthys flesus	16.41	7.67	3.59	23.49	8.50	4.74	12.79	4.77	3.95
Pomatoschistus microps	14.74	7.62	3.10						
Dicentrarchus labrax	13.28	7.19	2.68	17.60	7.30	4.17			
Pleuronectes platessa	5.45	6.99	2.86	25.04	8.86	4.67	14.65	6.50	3.59
Pomatoschistus sp				20.05	7.95	4.56			
Sprattus sprattus				18.92	8.43	3.89			
Buglossidium luteum							12.95	5.26	3.84
Limanda limanda							12.44	5.06	3.99
Solea solea				7			14.14	5.09	4.29
Total	13.96	8.00	2.39	21.79	8.34	4.42	12.75	5.41	3.55

4. Discussion

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4.1 Origin of and variability in organic matter in the Canche estuary

As estuaries are complex and changing ecosystems, it is challenging to distinguish the sources 340 of organic matter at the base of food webs (Pasquaud et al., 2008; Selleslagh and Amara, 341 2015). Organic matter is a major component of suspended particles and fine sediment 342 particles that determine many biogeochemical processes in marine environments (Bernasconi 343 et al., 1997; Chen et al., 2012). Two natural sources of organic matter are generally 344 345 considered in coastal ecosystems: allochthonous inputs and autochthonous production (Antonio and Richoux, 2014; Luo et al., 2016). Major sources of autochthonous organic 346 347 matter include phytoplankton and aquatic macrophytes (Dalu and Froneman, 2016; Pearson et al., 2015). In estuarine ecosystems, organic matter of terrestrial origin is a major contributor 348 to allochthonous organic matter (Duan et al., 2014; Lu et al., 2013). Analysis of the carbon 349 and nitrogen stable isotopic compositions of estuarine organic matter can identify their 350 contributions to the food web (Darnaude et al., 2004; Evans et al., 2019). Generally, POM in 351 estuaries is composed of river POM (mixture of terrestrial POM and freshwater 352 phytoplankton), estuarine-produced and marine phytoplankton, resuspended 353 microphytobenthos and diverse detritus (e.g. faeces, macrophytes), which can make 354 interpretation difficult (Kang et al., 2006). 355 In the Canche estuary, the POM stable isotopic signatures indicated a mixed organic matter 356 composition that included freshwater/estuarine phytoplankton (δ¹³C ranging from -29% 357 to -20%), with a clear increase in POM δ^{13} C values from fresh to marine waters, similar to 358 previous estuarine studies in the Bay of Marennes-Oléron (Riera and Richard, 1996), the 359 Vilaine estuary (Kostecki et al., 2010) and the Gironde estuary (Selleslagh et al., 2015). In the 360 Canche estuary, salinity exhibits short-term changes, with a large amplitude from 0-35 due to 361 the small size of the estuary, tide conditions, season and weather conditions (Amara et al., 362 2009; Selleslagh and Amara, 2008). MPB that live in intertidal flats in estuaries can 363 contribute much of the total primary production in estuaries (Underwood and Kromkamp, 364 365 1999). MPB on intertidal flats is composed mainly of benthic diatoms (Méléder et al., 2007) 366 and several studies have emphasised its key role in sustaining intertidal food webs 367 (Christianen et al., 2017; Herman et al., 2000; Thrush et al., 2012). In the Canche estuary, the MPB is one of the main primary producers and could therefore be an important source of 368 organic matter for benthic invertebrates. As in other European estuaries, the Canche MPB had 369 the highest enriched carbon ratios (-17% to -15%) among food sources, which allowed it to 370 371 be traced in its consumers (Moens et al., 2002; Moncreiff and Sullivan, 2001; Riera and

372 Richard, 1996). SOM is a mixture of benthic and deposited pelagic microalgae, bacteria, aquatic and terrestrial plant debris and meiofauna. In the Canche estuary, SOM and POM 373 δ¹³C followed the same trend, with the lowest values upstream and highest values 374 downstream due to the presence of freshwater phytoplankton, as measured in other nearby 375 estuaries (Lambert et al., 2017; Middelburg and Nieuwenhuize, 1998). 376 377 In estuaries, variations in amounts and origins of nutrients such as nitrogen are common along salinity gradients, with a decrease in concentrations from fresh to marine waters due to 378 379 mixing, which can be traced in food webs (Baeta et al., 2009; Connolly et al., 2013). The extent of nutrient mixing in estuaries varies spatially according to estuary size, and temporally 380 381 at the seasonal and daily scales due to changing tides, wind, precipitation and temperature (Baeta et al., 2009; Hoeinghaus et al., 2011; Lautenschlager et al., 2014). δ¹⁵N can be an 382 383 accurate tracer for nitrogen inputs that originate from untreated domestic, industrial and/or 384 agricultural activities that are incorporated in the food web through assimilation by primary producers (Fry, 2002). In the Canche estuary, POM, SOM and MPB δ^{15} N were similar along 385 the salinity gradient during the same season, highlighting the relatively low nitrogen input 386 from the watershed (Guelinckx et al., 2006) due to the short length of the Canche river and 387 the low human modification of its catchment (Amara et al., 2007; Durou et al., 2007). 388

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4.2 Structure of invertebrate communities

Two benthic communities were found in the Canche estuary: i) S. squamata/E. 391 pulchra/Bathyporeia spp. (EUNIS A2.223), which corresponds to estuarine mid-shore 392 393 medium-fine sand, and ii) H. diversicolor/S. plana on the upper shore mud banks (EUNIS 394 A2.24; Rolet et al., 2015). Outside the estuary, Sainte Cécile and Le Touquet beaches were characterized by a low-shore fine sand N. cirrosa/S. squamata community (EUNIS A2.23), 395 while the subtidal site had a muddy fine sand Abra alba/D. vittatus/F. fabula community 396 (Desroy et al., 2003; Rolet et al., 2015). This distribution of benthic invertebrate communities 397 is also present in the nearby estuaries of the Authie and Somme Rivers (Rolet et al., 2015). 398 Biomasses inside the estuary and on adjacent beaches were lower (2.6-6.0 g AFDW.m⁻²) than 399 in the subtidal A. alba community (150 g AFDW.m⁻²) dominated by the bivalve D. vittatus. In 400 the EEC, benthic biomass within the A. alba community is heterogeneous, with a mean of 8.1 401 g AFDW.m⁻² (Desroy et al., 2003), and a higher biomass ranging from 23.5-27.5 g AFDW.m⁻ 402 ² in the Seine Bay (Thiébaut et al., 1997) and from 45-3 000 g AFDW.m⁻² in Gravelines 403 (Desroy et al., 2003; Dewarumez et al., 1992; Ghertsos et al., 2000). This high subtidal 404

405 biomass of potential flatfish prey (mainly bivalves) outside the estuary created a major feeding ground for flatfish species. 406 One objective of the study was to assess the origin of the food source consumed by potential 407 prey throughout the estuary, from the intertidal zone upstream of the Canche estuary, with 408 low benthic species diversity and biomass, to the subtidal zone outside the estuary, with 409 higher species diversity and biomass. In the Canche estuary, suspension and deposit feeders 410 dominated the benthic invertebrate biomass. Their feeding activity is an important connection 411 412 between suspended and sedimented organic matter originating from POM, SOM or MPB (Little, 2000; Mann and Wetzel, 2000). However, it is often difficult to distinguish food 413 414 sources of macrozoobenthos in estuaries due to spatio-temporal variability in the isotopic compositions of food sources along the salinity gradient, and because macrofauna feed on 415 different food sources and have plastic feeding behaviour depending on the environmental 416 conditions (Daggers et al., 2020; Herman et al., 2000). Nevertheless, benthic primary 417 consumers had higher d¹³C (around – 20 to -16‰) than fresh water POM (around -30‰), 418 revealing the latter's low contribution to the trophic functioning of the estuary. This finding, 419 even in the upstream of the estuary, may be due to the relatively weak flow of the Canche 420 River and consequently the small amount of organic matter that it carries (Selleslagh and 421 Amara, 2008). Thus, we can hypothesize that the marine POM, SOM and MPB which d¹³C 422 423 composition are around -22 to -15% are the main food sources for the benthic community 424 inside the estuary. However, it may be difficult to distinguish suspension and deposit feeders 425 isotopically as their feeding behaviour does not provide information about the origin of their food; for example, suspension feeders can consume resuspended MPB, while deposit feeders 426 can consume sediment POM (Kang et al., 2015). 427

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4.3 Fish structure and seasonal variations

Two fish assemblages were observed, one inside the Canche estuary (P. minutus, P. microps and P. Labrax juveniles) and the other outside the estuary (P. luteum, P. solea and P. limanda juveniles). P. flesus and P. platessa juveniles occurred in both assemblages (Selleslagh and Amara, 2008). In both seasons in upstream Canche, P. flesus juveniles had the lowest δ^{13} C values, δ^{13} C values, δ^{13} C values, δ^{13} C values are from freshwater. This could be due to the diet of δ^{13} C values, which is composed of meiofauna (nematodes, harpacticoides and ostracods (Aarnio et al., 1996; Selleslagh and Amara, 2015)) that may consume SOM. Conversely, δ^{13} C values, which were similar to those of MPB, revealing the contribution of MPB

439	to the feeding of <i>P. microps</i> prey. Previous studies indicated that <i>P. microps</i> feeds mainly on
440	amphipods, polychaetes and meiofauna (Leclerc et al., 2014; Selleslagh and Amara, 2015).
441	MPB production is high on the intertidal mud banks of the Canche estuary and provides food
442	for the potential prey of P. microps. Inside the estuary, the other fish species (e.g. D. labrax,
443	P. minutus or S. sprattus) had $\delta^{13}C$ values from -19 to -17‰ upstream to -16% downstream,
444	close to those of marine POM (around -20%) and MPB (-16%) revealing a food source
445	originating from both marine POM and MPB.
446	The seasonal comparison of fish SEA in the Canche estuary is informative, as the isotopic
447	space occupied is smaller in spring than in fall. Thus, a wider range of prey appears to be
448	consumed in the fall. This could be due to the higher biomass and diversity of coastal benthic
449	invertebrates at the end of summer compared to the lower benthic biomass in spring (Rauch
450	and Denis, 2008). In the fall, SEA of the two Pomatoschistus sp. did not overlap, unlike
451	Platichthys flesus and Pleuronectes platessa that inhabit the Canche estuary, which confirms
452	that their diet may differ (Salgado et al., 2004; Selleslagh and Amara, 2015). Conversely, the
453	SEA of P. flesus and P. platessa overlapped in spring, perhaps due to the smaller amount of
454	available prey (Pape and Bonhommeau, 2015).
455	Outside the estuary, the $\delta^{13}C$ values of flatfish juveniles (B. luteum, S. solea, L. limanda) were
456	ca16‰, revealing MPB and POM to be a major basic food source, except for P. platessa,
457	which had much lower $\delta^{13}C$ values, similar to those found in the estuary. This suggests that P .
458	platessa individuals caught outside the Canche estuary did not feed exclusively in the habitat
459	in which they were collected, which indicates that this species has high mobility and habitat
460	connectivity.
461	Flatfish SEA indicates a slight isotopic niche overlap of S. solea, B. luteum and L. limanda,
462	which suggests trophic segregation of the three species. Conversely, the SEA of P. flesus
463	completely overlapped those of juveniles of these flatfish species, which indicates that P .
464	flesus consumes a wider range of prey and may have trophic competition with the three other
465	flatfish species. Juvenile fish in estuaries usually follow an opportunistic feeding strategy,
466	which is driven by intra- and inter-specific competition (Brown et al., 2019; Post et al., 1999)
467	and prey availability.
468	
469	5. Conclusion
470	We showed a significant difference in invertebrate biomass between subtidal and intertidal

sites, which influences the quality of the feeding ground for juvenile fish. This is a classic

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472	situation in European estuaries (Dubois et al., 2014). This result provides a new vision of the
473	Canche estuary, which has been considered an important feeding ground for marine fish.
474	Our study revealed that these continental inputs have a minor role in the functioning of the
475	Canche estuary and that fish species might visit the estuary for reasons other than feeding,
476	such as to avoid predation or because they are carried by the tide. We highlighted the need to
477	take into account the whole small macrotidal estuary and adjacent ecosystems to better
478	describe the flatfish nursery. This work demonstrated that potential prey and feeding sources
479	for fish had habitat-specific compositions, which confirms the suitability of SIA for tracing
480	fish movements, fidelity and connectivity inside and outside the Canche estuary for sites less
481	than 10 km apart. Estuarine nursery feeding grounds, even in small estuaries, appear to be
482	complex due to the mosaic of benthic communities (potential prey), which are related to the
483	habitat (e.g. sediment type, foreshore position, salinity fluctuations) and to trophic
484	competition and predation.

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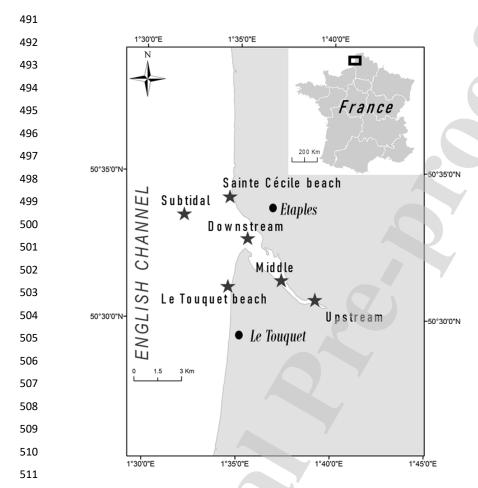


Figure 1. Map of the Canche estuary showing the sampling sites (stars) inside the estuary (upstream, middle, and downstream) and outside the estuary (surf zone (Sainte Cécile and Le Touquet beaches) and subtidal site).

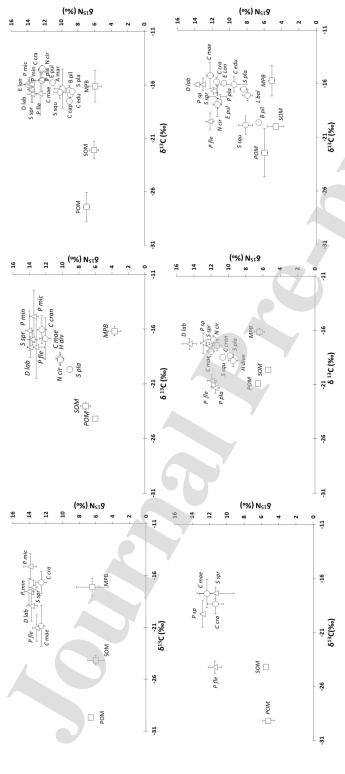


Figure 2. Mean (\pm standard deviation) $\delta^{13}C$ vs. $\delta^{15}N$ biplots of the food sources (\Box), invertebrates (\circ), and fish (Δ) inside the Canche estuary in the fall (A: upstream, B: middle, C: downstream) and spring (D: upstream, E: middle, F: downstream). See Tables 2 and 4 for abbreviations.

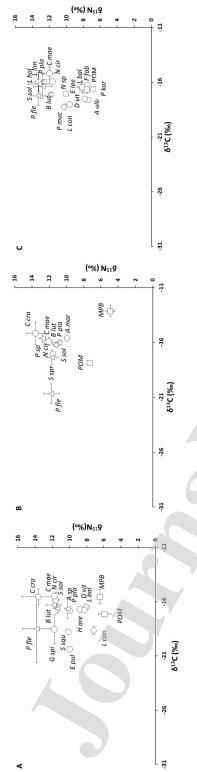


Figure 3. Mean (\pm standard deviation) $\delta^{13}C$ vs. $\delta^{15}N$ biplots of the food sources (\Box), invertebrates (\circ), and fish (Δ) outside the Canche estuary in the fall (A: Sainte Cecile beach, B: Le Touquet beach, C: subtidal site). See Tables 2 and 4 for abbreviations.

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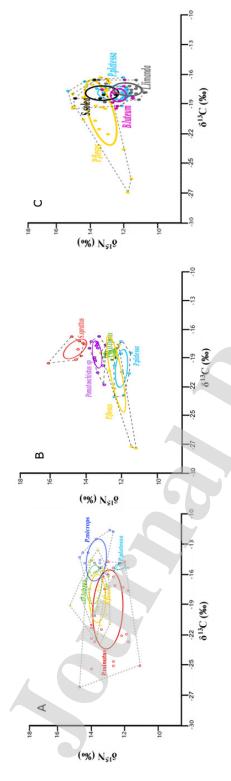


Figure 4. Convex hull polygons and corrected Standard Ellipse Areas (SEAc) of the fish community. Each convex hull is a measure of the total area, while each SEAc is a proxy of the mean isotopic niche of the core population, which is insensitive to sample size (A: inside the Canche estuary in the fall, B: inside the Canche estuary in spring, C: outside the Canche estuary in spring).

Appendix 1. Macrobenthic carbon and nitrogen stable isotope compositions (mean ± standard error, %), and number of individuals analysed (N) 531

532 in spring

N 8 ¹⁵ N 6 11.3 ±0.9 -1 6 12.3 ±1.3 -1.	- Pour curr	Milaale	e	1100	Downsti cam	S	Saint Cecile Beach	Beach:	Fe	Le Touquet Beach	Beach		Subtidal	lal
N 6 5 N 6 6 N 6 6 N 6 6 12.3 ± 1.3 - 1 1 1 1 1 1 1 1 1 1														
div) div) div) (squ) (h) (n) (n) (r) (e) (e) (f) (f) (f) (f) (f) (f) (f) (f) (f) (f	δ^{13} C N	N ₂ 18N	813C N 815N	N ₂ 15N	δ^{13} C N	Z	N ₂₁ 8	δ^{13} C	Z	N ₂₁ 8	δ^{13} C	Z	8 ¹⁵ N	δ^{13} C
divy (qqu) (h) (h) (h) (h) (axi) (x) (x) (y) (y) (y) (y) (y) (y) (y) (y) (y) (y	3	9.2 ± 0.1	-18.9 ± 0.1 3	8.4 ± 0.4	-16.5 ± 0.1									
(1) (1) (2) (3) (4) (4) (5) (6) (6) (7) (9) (9) (9) (9) (9) (9) (9) (9) (9) (9	3	9.4 ± 0.5	-18.8 ± 0.5											
(i) (i) (ii) (iii)	2	10.7	-18.4	8.2 ± 0.8		7	10.2	-18.8						
are) (a) (b) (c) (c) (d) (e) (e) (f)			4	6.0 ± 0.7	-17.1 ± 0.5	2	8.2	-16.7				3	8.1 ± 0.6	-16.4 ± 0.3
are) (7) (9) (9) (9) (9) (9) (9) (9) (9) (9) (9			v.	9.5 ± 1.5										
are) (r) (r) (r) (r) (r) (r) (r) (r) (r) (r						4	7.1 ± 0.1	-18.6 ± 0.4				7	9.6	-18.0
(r) (6 (c)						S	8.1 ± 1.3	-16.5 ± 0.5				ж	7.8 ± 0.2	-17.5 ± 0.3
(c) (d) (d) (d) (d) (d) (d) (d) (d) (d) (d						7	8.8	-16.7						
9 (0)						2	6.6	-16.8	-	6.6	-15.6			
6 (6 (6 (7 (7 (7 (7 (7 (7 (7 (7 (7 (7 (7 (7 (7												7	7.4	-17.6
6) 6	,											3	7.5 ± 0.1	-16.2 ± 0.2
6) 6												5	8.2 ± 0.2	-16.6 ± 0.2
9 (9	-18.3 ± 0.9 5	11.3 ± 0.5	-17.4 ± 0.5 5	11.5 ± 0.2	2 -15.8 ± 0.5	9 !	13.6 ± 1.9	-15.5 ± 0.7	7 1	13.6 ± 1.0	-15.6 ± 0.3			
Nephtys cirrosa (N cir) secondary Eteone longa (E lon) consumers Eurydice pulchra (E pul)	-17.4 ± 0.5 5	12.0 ± 0.4	-17.9 ± 1.1 5	12.3 ± 0.4	4 -15.3 ± 1.2	4	11.6 ± 0.3	-15.8 ± 0.7	4	12.0 ± 0.3	-15.9 ± 1.0	5	11.9 ± 0.5	-15.2 ± 0.2
secondary Eteone longa (E lon) consumers Eurydice putchra (E pul)	3	11.7 ± 0.6	-16.8 ± 0.4 2	11.4	-17.9	7	11.8	-15.5	7	11.6	-17.1	_	11.6	-15.9
consumers Eurydice pulchra (E pul)			_	10.7	-15.9									
			9	11.3 ± 0.1	$1 - 21.9 \pm 0.6$	3	9.1 ± 0.9	-20.4 ± 0.9						
Gastrosaccus spinifer (G spi)			7				11.7 ± 0.1	-18.5 ± 1.3						
Nemertes sp				/								т	10.0 ± 0.2	-17.0 ± 0.5
Phyllodoce mucosa (P muc)												5	10.2 ± 0.3	-18.2 ± 1.5

Appendix 2. Macrobenthic carbon and nitrogen stable isotope compositions (mean ± standard error, ‰), and the number of individuals analysed (N) in the fall

			Upstrea	ım		Midd	lle		Downst	ream
Trophic position	Species									
F		N	$\delta^{15}N$	$\delta^{13}C$	N	$\delta^{15}N$	$\delta^{13}C$	N	$\delta^{15}N$	$\delta^{13}C$
	Scrobicularia plana (S pla)				5	9.1 ± 0.2	-19.7 ± 0.2	3	8.7 ± 0.2	-16.6 ± 0.1
	Hediste diversicolor (H div)				10	10.3 ± 0.5	-18.7 ± 0.6			
primary consumers	Scolelepis squamata (S squ)							4	10.2 ± 0.7	-16.7 ± 0.6
	Limecola balthica (L bal)							5	9.9 ± 0.3	-16.1 ± 0.5
	Cerastoderma edule (C edu)							7	8.9 ± 0.1	-16.9 ± 0.3
	Crangon crangon (C cra)	10	12.6 ± 0.7	-16.7 ± 1.0	9	12.5 ± 0.5	-16.0 ± 1.2	8	12.4 ± 0.9	-14.6 ± 0.4
	Carcinus maenas (C mae)	5	12.7 ± 0.4	-20.9 ± 1.9	6	12.1 ± 0.7	-16.9 ± 2	7	12.3 ± 0.8	-15.6 ± 1.0
secondary consumers	Nephtys cirrosa (N cir)				5	10.3 ± 0.4	-18.8 ± 0.5	3	12.3 ± 1.1	-14.5 ± 1.2
consumers	Eteone longa (E lon)							2	13.9	-15.9
	Eurydice pulchra (E pul)							4	11.0 ± 0.3	-15.8 ± 0.2

537 Appendix 3. Fish carbon and nitrogen stable isotope compositions (mean ± standard error, ‰) in fall and spring

	Inside												Outside					
	Upstream			-	Middle				Downstream				Saint Cécile Beach		Le Touquet Beach	Beach	Subtidal	
Species (abbreviation)	fall		spring		fall		spring		fall		spring		spring		spring		spring	
7	§¹3℃	N 518	8 ¹³ C	N 518	8 ¹³ C	815 N	8 ¹³ C	N 518	8 ¹³ C	N 515 N	8 ¹³ C	815 N	8 ¹³ C	N 515 N	8 ¹³ C	N 218	8¹³ C	N 515
Pomatoschistus microps (P mic) -15.1 ± 1.2 13.9 ± 0.8	-15.1 ± 1.2	13.9 ± 0.8	A		-14.8 ± 2.4	13.4 ± 0.5			13.8 ± 0.5	13.8 ± 0.6								
Spratus sprattus (S spr)	-17.1 ± 0.7	13.5 ± 0.5	-17.1 ± 0.7 13.5 ± 0.5 -17.3 ± 0.5	11.3 \pm 2.1 -16.6 \pm 0.8		13.7 ± 0.6 -17.4 ± 0.3		12.2 ± 0.3	-16.8 ± 0.8	13.5 ± 0.8	$13.5 \pm 0.8 - 16.7 \pm 0.3$	11.9 ± 0.1		•	-17.4 ± 0.6 11.7 ± 0.7	11.7 ± 0.7		
Pomatoschistus minutus (P min) -17.6 ± 1.3 14.0 ± 0.6	$\textbf{-}17.6\pm1.3$	14.0 ± 0.6		Ŝ	-16.1 ± 1.8	13.7 ± 0.6			$\textbf{-15.9} \pm 0.2$	13.6 ± 0.1								
Dicentrarchus labrax (D lab)	$\textbf{-}18.9 \pm 1.5$	-18.9 ± 1.5 13.7 ± 0.7			-17.7 ± 1.0	13.9 ± 0.3 -17.2 ± 1.2		14.5 ± 0.8	-16.4 ± 1.2	13.6 ± 0.3	13.6 ± 0.3 -16.1 ± 0.2	13.8 ± 0.1						
Platichthys flesus (P fle)	-21.1 ± 1.3	13.1 ± 0.5	-21.1 ± 1.3 13.1 ± 0.5 -24.2 ± 4.5 11.4 ± 0.8 -17.6 ± 2.8	11.4 ± 0.8	-17.6 ± 2.8	13.2 ± 1.0 -20.6 ± 0.5		11.9 ± 0.5	-16.8 ± 2	12.5 ± 0.6	12.5 ± 0.6 -19.5 ± 0.9	12.3 ± 0.5 -18.5 ± 3.1	-18.5 ± 3.1	13.8 \pm 1.6 -20.7 ± 0.5		11.8 ± 0.9	11.8 \pm 0.9 -17.2 \pm 0.5	13.3 ± 0.8
Buglossidium luteum (B lut)				7									-16.2 ± 0.3	11.8 ± 0.9 -16.2 ± 0.2	-16.2 ± 0.2	11.3 ± 0.1	11.3 \pm 0.1 -17.1 \pm 0.4	11.9 ± 0.2
Pomatoschistus sp (P sp)			-19.3 ± 1.2	12.8 ± 0.3			-17.27 ± 0.9 12.9 ± 0.3	12.9 ± 0.3			$\textbf{-}16.1 \pm 0.6$	13.1 ± 0.3		•	-15.6 ± 0.3	12.7 ± 0.3		
Limanda Limanda (L lim)																	$\textbf{-}16.4 \pm 0.6$	12.8 ± 1
Pleuronectes platessa (P pla)							-21.3 ± 0.5	11.5 \pm 0.5 \pm 0.2	$\textbf{-15.5} \pm \textbf{0.2}$	12.4 ± 0.2	12.4 ± 0.2 -17.2 ± 1.0 11.4 ± 1.0 -16.6 ± 0.9	11.4 ± 1.0	-16.6 ± 0.9	10.2 ± 0.3	10.2 ± 0.3 -16.0 ± 0.6	10.8 ± 0.3	10.8 ± 0.3 -15.9 ± 0.7	12.7 ± 0.2
Solea solea (S sol)							,						-16.6 ± 0.4	$-16.6 \pm 0.4 \qquad 11.4 \pm 0.2 \qquad -16.3 \pm 0.5 \qquad 11.3 \pm 0.2 \qquad -16.2 \pm 0.6 \qquad 13.2 \pm 1.2$	-16.3 ± 0.5	11.3 ± 0.2	$\textbf{-}16.2 \pm 0.6$	13.2 ± 1.2
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AUTHORSHIP STATEMENT

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All persons who meet authorship criteria are listed as authors, and all authors certify that they have participated sufficiently in the work to take public responsibility for the content, including participation in the concept, design, analysis, writing, or revision of the manuscript. Furthermore, each author certifies that this material or similar material has not been and will not be submitted to or published in any other publication before its appearance in **Regional Studies in Marine Science**

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Conflict of interest

Structure and seasonal variability in fish food webs in a small macrotidal estuary
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The authors declare they don't have any conflict of interest.