

A novel synonymous variant in exon 1 of GNAS gene results in a cryptic splice site and causes pseudohypoparathyroidism type 1A and pseudo-pseudohypoparathyroidism in a French family

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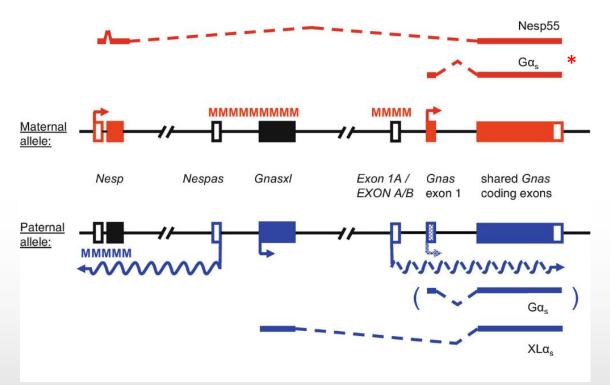
A novel synonymous variant in exon 1 of *GNAS* gene results in a cryptic splice site and causes pseudohypoparathyroidism type 1A and pseudo-pseudohypoparathyroidism in a French family

Andreea Apetrei¹, Arnaud Molin¹, Nicolas Gruchy¹, Manon Godin¹, Claire Bracquemart¹, Antoine Resbeut¹, Gaëlle Rey², Gwenaël Nadeau², Nicolas Richard¹

1 - Normandy University, UNICAEN, Caen University Hospital, Department of Genetics, Reference Center of Rare Diseases of Calcium and Phosphorus Metabolism, EA 7450 BioTARGen, Caen, France

2 - Metropole Savoie Hospital Center, Genetics Department, Chambéry, France





The main imprinted sense and antisense transcripts from the GNAS complex locus.

*Gαs is predominantly expressed from the maternal allele in the renal proximal tubules, parathyroid, thyroid, pituitary gland and ovaries and has a biallelic expression in the other tissues.

M – methylated region

Adapted from Pulix M., Plagge A. (2020) Imprinted Genes and Hypothalamic Function

GNAS – associated phenotype

	Genetic defect	Parental origin	Hormonal resistance (PTH, TSH) [§]	AHO Phénotype [#]
PHP1A	Coding sequence variant	Maternal	Yes	Yes
РРНР	Coding sequence variant	Paternal		Yes 🗶

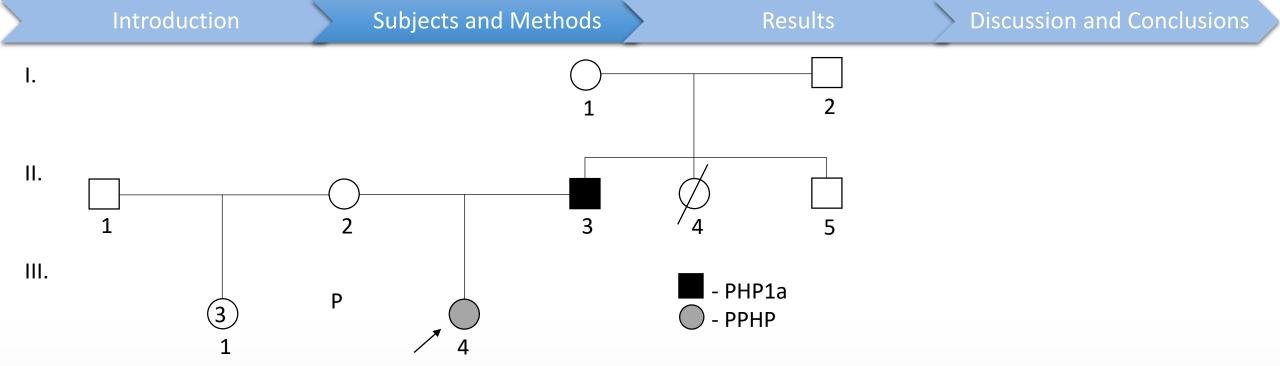
The main disease subtypes caused by inactivating GNAS gene mutations.

[§]The resistance to parathormone (PTH) is characterized by high serum PTH, hypocalcemia and hyperphosphatemia and it can be associated with resistance to other hormones, such as thyroid stimulating hormone (TSH).

[#]Described Albright's Hereditary Osteodystrophy (AHO) features are obesity, round face, short stature, brachydactyly type E, subcutaneous ossification and mild mental retardation.

The crossed alleles correspond to the inactivated alleles of the GNAS gene. Adapted from Turan S., Bastepe M. (2015) Curr Osteoporos Rep

Maternaly inherited loss of function mutations lead to **PHP1A**, whereas paternaly inherited loss of function mutations lead to **PPHP**. Therefore, the two phenotypes usually coexist in the same family, but never in the same sibship.



Clinical manifestation	111.4	II.3
AHO features	Yes	Yes
Intrauterine growth retardation	Yes	No
 Serum calcium Serum phosphate Serum PTH Thyroid dysfunction 	N N N No	N* N 个 Yes
Clinical diagnosis	РРНР	PHP1a

N – normal level

*At the low limit of the normal range

GNAS sequencing

GNAS exons 1- 13 were sequenced and interpreted following the routine protocol.

In silico splicing prediction algorithms (SPiP and Alamut) were used.

GNAS expression analysis

Quantification

qRT-PCR was used to assess the expression of *GNAS* in cDNA in the proband and in a healthy, sex and aged-paired control. Primers spanned the coding region of exons 1 and 2.

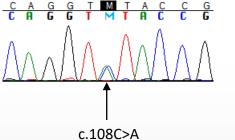
Transcript study and sequencing

RT-PCR using a gene-specific primer localized in the exon 5 was followed by a PCR using primers localized in exons 1 and 2.

A capillary electrophoresis analysis allowed to isolate and characterize the PCR products before sequencing.

GNAS sequencing

A heterozygous variation Chr20(GRCh37):g.57466889C>A, NM 001077488.2:c.108C>A p.(Val36=) localized in the exon 1 of GNAS gene, absent from the local and international databases was identified in III.4., inherited from her father (II.3):



Chromatogram from DNA sequencing of a nucleotide substitution in the proband (111.4)

1.50 -

1.00

0.50 م م

0.00

control

GNAS CDNA levels y qRT-PCR

Relative

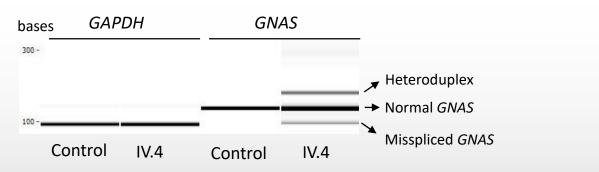
GNAS expression analysis

111.4

The expression quantification showed a reduction of GNAS expression by 35% in comparison with a normal control:

> Relative GNAS expression level measured by qRT-PCR using oligo(dT)18 primer, normalized to GAPDH expression level in the proband (III.4) and a healthy control.

A second RT-PCR using a gene-specific primer allowed to amplify the expected PCR product and 2 additional products in smaller quantities, a shorter one and a longer one, respectively corresponding to the wild-type, the mutant allele and a heteroduplex (a mixture of wild-type and mutant allele) product:

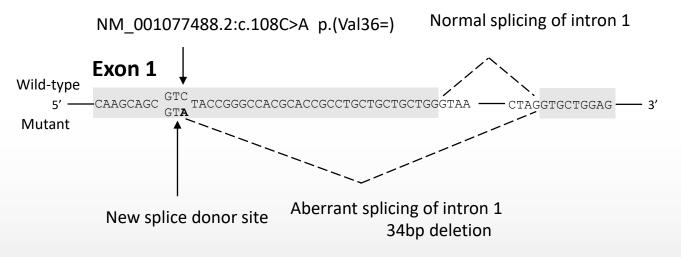


Capillary electrophoresis performed on the cDNA obtained by RT-PCR using GNAS gene specific primer.

The sequencing of these PCR products showed a 34bp deletion in the mutant allele.

We describe the first family with PHP1a/PPHP presenting a pathogenic synonymous variant in the GNAS gene, responsible for a splicing defect leading to the premature splicing of the first exon and to a probably unstable RNA or truncated protein:

GNAS



This observation supports the increasing interest in the identification of splicing variants, particularly those predicted as synonymous, which are often systematically categorized as benign on this sole element.

Splice-prediction algorithms could be added to the bioinformatic pipelines for massive parallele sequencing data in routine analysis to improve the detection of such variations.