



HAL
open science

Respiratory rhythm modulates membrane potential and spiking of non-olfactory neurons

Maxime Juventin, Mickael Zbili, Nicolas Fourcaud-Trocmé, Samuel Garcia, Nathalie Buonviso, Corine Amat

► **To cite this version:**

Maxime Juventin, Mickael Zbili, Nicolas Fourcaud-Trocmé, Samuel Garcia, Nathalie Buonviso, et al.. Respiratory rhythm modulates membrane potential and spiking of non-olfactory neurons. *Journal of Neurophysiology*, 2023, 130 (6), pp.1552-1566. 10.1152/jn.00487.2022 . hal-04335818

HAL Id: hal-04335818

<https://hal.science/hal-04335818>

Submitted on 11 Dec 2023

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

1 RESEARCH ARTICLE

2

3 RUNNING HEAD: Respiratory modulation in the brain: deep in the cells

4

5 ARTICLE TITLE: Respiratory rhythm modulates membrane potential and spiking of non-
6 olfactory neurons

7

8 AUTHORS: Maxime Juventin¹, Mickael Zbili^{1,2}, Nicolas Fourcaud-Trocmé¹, Samuel
9 Garcia¹, Nathalie Buonviso¹ and Corine Amat¹

10

11 AFFILIATION:

12 ¹ Université Claude Bernard Lyon 1, CNRS, INSERM, Centre de Recherche en
13 Neurosciences de Lyon CRNL U1028 UMR5292, F-69500, Bron, France

14 ² Université Clermont Auvergne, CHU Clermont-Ferrand, INSERM, Neuro-Dol, F-63100,
15 Clermont-Ferrand, France

16

17

18

19 CORRESPONDING AUTHOR: Corine Amat: corine.amat@univ-lyon1.fr

20 UCBL, CNRS, INSERM, CRNL U1028 UMR5292
21 Centre Hospitalier Le Vinatier - Bâtiment 462 - Neurocampus Michel Jouvét
22 95 boulevard Pinel
23 69675 Bron Cedex
24 France

25

27 ABSTRACT

28 In recent years, several studies have shown a respiratory drive of the local field potential
29 (LFP) in numerous brain areas so that the respiratory rhythm could be considered as a master
30 clock promoting communication between distant brain locations. However, outside of the
31 olfactory system it remains unknown whether the respiratory rhythm could shape membrane
32 potential (MP) oscillations. To fill this gap, we co-recorded MP and LFP activities in different
33 non-olfactory brain areas: medial prefrontal cortex (mPFC), primary somatosensory cortex
34 (S1), primary visual cortex (V1), and hippocampus (HPC), in urethane-anesthetized rats.
35 Using respiratory cycle-by-cycle analysis, we observed that respiration could modulate both
36 MP and spiking discharges in all recorded areas during episodes that we called Respiration-
37 Related oscillations (RRO). Further quantifications revealed that RRO episodes were transient
38 in most neurons (5 consecutive respiratory cycles in average). RRO development in MP was
39 largely correlated with the presence of respiratory modulation in the LFP. By showing that the
40 respiratory rhythm influenced brain activities deep to the MP of non-olfactory neurons, our
41 data support the idea that respiratory rhythm could mediate long-range communication
42 between brain areas.

43

44 NEW & NOTWORTHY

45 In this study, we evidenced strong respiratory driven oscillations of neuronal membrane
46 potential and spiking discharge in various non-olfactory areas of the mammal brain. These
47 oscillations were found in medial prefrontal cortex, primary somatosensory cortex, primary
48 visual cortex and hippocampus. These findings support the idea that respiratory rhythm could
49 be used as a common clock to set the dynamics of large-scale neuronal networks on a same
50 slow rhythm.

51

52 KEY WORDS: freely breathing anesthetized rat, intracellular activities, local field potential,
53 membrane potential, respiration-related oscillations.

54

55

56

57

58 INTRODUCTION

59 In recent years, a resurgence of publications has proposed respiration as a master clock for
60 brain rhythms, following many observations that reported the breathing rhythm as being able
61 to modulate complex behaviors or cognitive process (1–4). The exploratory behavior of
62 rodents, consisting in rhythmical sampling of the environment, is a perfect example since the
63 onset of each respiratory cycle initiates a “snapshot” of the orofacial sensory environment,
64 comprising sniffing, whisking and head movements (5–7). Similarly, licking synchronizes
65 with breathing (8, 9). In humans, respiration modulates memory consolidation (10),
66 associative learning (11), cognitive performances (12–14), sensory perception (15, 16), and
67 motor behaviors (17, 18). Respiratory phase influences the initiation of a voluntary action (19)
68 or a cognitive task and its performance (13). All these data therefore suggest that the brain
69 could have at least one processing mode depending on the respiratory phase.

70 Parallel to this literature on the effects of breathing on behavior has appeared an equally
71 important literature on a respiratory drive of brain activity, both in human and rodent. In
72 humans, intracerebral EEG-breathing coherence has been observed particularly in the
73 hippocampus, amygdala, insula and the parietal lobe (14, 20) and the inhalation phase is
74 correlated with an increase in the power of delta oscillations (14). In rodents, a respiration-
75 locked local field potential (LFP) oscillation has been described in the hippocampus (21), the
76 somatosensory cortex (22), the prefrontal cortex (23, 24), neocortical regions as posterior as
77 visual cortex and even subcortical areas (25, 26). Breathing also modulates other rhythms in
78 several brain regions. In the hippocampus, the timing of hippocampal sharp-waves and ripples
79 is respiration-coupled (27, 28). In the somatosensory cortex, delta oscillations are respiration-
80 locked whereas gamma power is respiration-modulated (22). In the prefrontal cortex, fast
81 gamma LFP oscillations (29) and unitary activities (23) are phase-coupled with respiration.

82 This respiratory drive, described in such a large cerebral network, is at the moment assumed
83 to arise via the activation of the olfactory bulb (OB) *via* mechanical stimulation of olfactory
84 receptor cells by nasal respiratory airflow (30). Indeed, respiration-brain activity coupling
85 dissipates in human when subjects breathe through the mouth (14) or in rodents when OB is
86 silenced (21, 22, 28, 31). This respiration-related activity was first found in the OB where it
87 has been largely described (For review, see (1)): respiration modulates both LFP and
88 individual cells activities, including spiking discharge and membrane potential dynamics. In
89 LFP signal, fast rhythms occurrence is shaped by the slow respiration-related oscillation
90 (RRo), beta and gamma waves alternating on expiratory and inspiratory phases, respectively
91 (33, 34). Mitral/tufted cells spiking discharge is strongly patterned by respiratory rhythm both
92 in anesthetized (34–37) and awake rodent (38, 39). Importantly, even the membrane potential
93 of mitral/tufted cells is influenced by respiration (35, 40, 41). Particularly, RRo of membrane
94 potential constrain fast oscillations to occur concomitantly with LFP signal, hence favoring
95 interaction between fast LFP and spiking (41). We and others have demonstrated that most of
96 OB respiratory modulation is related to the amplitude of the inspiratory airflow (36, 42–44).
97 Respiratory influence is also present, to a lesser extent, in the olfactory cortex both at the LFP
98 and cellular levels (Piriform cortex: (1–4); Olfactory tubercle: (49)). Directly in contact with
99 nasal respiratory airflows, the olfactory system is thus strongly impacted by respiration
100 dynamics.

101 Oppositely, while LFP activity in non-olfactory regions has been described as respiration-
102 modulated, very little is known about respiratory modulation of individual cell activities. Only
103 a few reports described spiking discharge (extracellularly recorded) modulated by respiration
104 (23, 24). Only one article reported intracellular recordings in parietal cortex (50). The authors
105 showed membrane potential oscillations that were coherent with respiration in 6 out of 29
106 parietal neurons, no further studies were done. Here we asked to what extent respiratory

107 modulation could shape cellular activity, including MP and spiking activity, in non-olfactory
108 areas. To this purpose, we co-recorded membrane potential and LFP activities in widespread
109 brain areas, namely the medial prefrontal cortex (mPFC), the primary somatosensory cortex
110 (S1), primary visual cortex (V1), and hippocampus (HPC), in urethane-anesthetized rats.
111 Using respiratory cycle-by-cycle analysis, we tracked respiratory modulation of cellular
112 activities. We observed that both MP and spiking discharges could be respiration-modulated
113 in all recorded areas, but more transiently than what we previously described in the OB (35).
114 Particularly, respiratory modulation of MP was highly variable both between structures and
115 between cells, very transient for most of cells and largely influenced by LFP state. Finally,
116 moderate hyperpolarization of MP reduced the probability of RRo.

117

118 MATERIALS AND METHODS

119 Experimental procedures

120 *Animal care*

121 The 25 male Wistar rats (325.2±32.1g, Charles River) used were housed in groups of four in a
122 temperature (22±1 °C) and humidity (55±10%) controlled room and exposed to a 12/12 h
123 light/dark cycle (light onset, 6:00 am). Experiments were conducted during the light period
124 (between 9:00 am and 3:00 pm). Food and water were available *ad libitum*. All experiments
125 were carried out in accordance with Directive 2010/63/EU of the European Parliament and of
126 the Council of the European Union regarding the protection of animals used for scientific
127 purposes and in compliance with the ARRIVE guidelines. The experimental protocols were
128 approved by the National Ethics Committee “Animal Experimentation Committee of Univ.
129 Claude Bernard Lyon 1—CEEA-55” (Agreement APAFIS #17088).

130 *Animal preparation*

131 Rats were anesthetized with an intraperitoneal injection of urethane (1.5 g/kg), then, placed in
132 a stereotaxic apparatus. Respiration was recorded with a homemade bidirectional airflow
133 sensor placed at the entrance of the right nostril. Positive and negative flows corresponded
134 respectively to inspirations and expirations. Body temperature was kept at 37 °C using a
135 heating pad (Harvard Apparatus, Holliston, MA USA). A craniotomy of about 3x3 mm was
136 performed above the site of recording: medial prefrontal cortex, primary visual cortex,
137 somatosensory barrel cortex or hippocampus. Dura was removed, and Ringer’s lactate
138 solution was regularly applied onto the brain to prevent from drying. At the end of the
139 experiments, rats were euthanized by intracardiac injection of Dolethal©.

140 *In vivo Electrophysiological Recordings*

141 We made simultaneous intra- and extracellular recordings in freely breathing anesthetized
142 rats. Intracellular recordings (n = 69 cells) were performed in four different areas of the left
143 hemisphere: the medial prefrontal cortex (mPFC, AP: 3.1 ± 0.4 mm, MD: 0.7 ± 0.2 mm), the
144 primary somatosensory cortex (S1, AP: -2.4 ± 0.4 mm, MD: 5.0 ± 0.1 mm), the primary visual
145 cortex (V1, AP: -5.3 ± 0.2 mm, MD: 4.5 ± 0.8 mm) and the hippocampus (HPC, -3.5 ± 0.7 mm,
146 MD: 2.6 ± 0.7 mm). Input resistance of the neurons was 52.7 ± 4.7 M Ω (Supplemental Figure
147 S1). Borosilicate glass capillaries (o.d. = 1.5 mm; i.d. = 0.86 mm, Harvard Apparatus,
148 Holliston, MA, USA) were pulled with a horizontal puller (model P-97, Sutter Instruments,
149 Novato, CA, USA). The micropipettes were filled with a 2 M potassium acetate solution, and
150 their resistances ranged from 50 to 120 M Ω . Microelectrodes were lowered using a piezo
151 manipulator (PM-10, World Precision Instruments, Sarasota, FL, USA). The
152 electrophysiological signal was amplified and low-pass filtered at 10 kHz by an intracellular
153 amplifier (Axoclamp 2B, Axon Instruments, Foster City, CA, USA). Local field potential
154 (LFP) was recorded simultaneously with intracellular signal using silicon probes (Neuronexus
155 Technology, Ann Arbor, MI, USA) placed as close as possible to the glass microelectrode.
156 The signal was amplified, and low-filtered at 5 kHz with a homemade amplifier. Respiration
157 was recorded with an airflow sensor (described in (51)). Positive and negative values
158 corresponded to expiration and inspiration respectively. All signals were then digitalized at 25
159 kHz (NI USB-6211, National Instruments, Austin, TX, USA) and stored on a computer using
160 Neurolabscope, a homemade software.

161 *Internal polarity state*

162 Internal polarity states were encoded based on the sign (positive or negative) and intensity of
163 direct current (DC) injection, amplitude of MP variation and firing activity. We identified 4
164 states: strong hyperpolarization (Hyp2), light hyperpolarization (Hyp1), basal level (Base),
165 depolarization (Dep). A given cell was not systematically recorded in each of the 4 states.

166 *Post hoc* verification confirmed our classification (Supplemental Figure S2). Whatever the
167 structure, basal level (Base) represented the state where no (N = 42 cells) or a very low
168 stabilizing current (-0.07 ± 0.06 nA, N = 14 cells) was injected. Hyp1 level corresponded to a
169 MP hyperpolarization of around 7 mV relative to the basal level, at which the spike discharge
170 decreased by around 60%. This level was reached with a -0.29 ± 0.04 nA (N = 25 cells) DC
171 injection. Hyp2 level corresponded to a MP hyperpolarization of around 12 mV relative to the
172 basal level, at which the spike discharge decreased by 90% to 100%. This level was reached
173 with a -0.42 ± 0.04 nA (N = 31 cells) DC injection. Dep level corresponded to a MP
174 depolarization of around 7 mV relative to the basal level, at which the spike discharge
175 increased by at least 10%. This level was reached with a 0.19 ± 0.03 nA (N = 21 cells) DC
176 injection. For the cell-by-cell analysis, in order to limit the influence of external factors, we
177 selected pair of recordings close in time. In the mPFC, cells (N = 9 cells) received -0.17 ± 0.02
178 nA for Hyp1, resulting in a hyperpolarization of 7.6 ± 2.5 mV and a decrease in spike
179 discharge of 74.4 ± 7.5 %. Latency between Base and Hyp1 recordings was 3.2 ± 1 min. In S1,
180 cells (N = 4 cells) received -0.19 ± 0.06 nA for Hyp1 resulting in a hyperpolarization of
181 8.4 ± 5.2 mV and a decrease spike discharge decrease of 50.0 ± 26.6 %. Latency between Base
182 and Hyp1 recordings was 3.3 ± 1.4 min. In HPC, cells (N = 3 cells) received -0.26 ± 0.05 nA for
183 Hyp1 resulting in a hyperpolarization of 7.6 ± 3.0 mV and a decrease in spike discharge of
184 50.5 ± 10.0 %. Latency between Base and Hyp1 recordings was 1.3 ± 0.2 min.

185 Data analysis

186 All following analyses have been made using homemade Python scripts.

187 *Respiratory signal processing*

188 Respiratory cycles were detected as previously described in (51). Respiration was monitored
189 with an airflow sensor. Positive, respectively negative, values corresponded to expiration,

190 respectively inspiration. Null values corresponded to transition between inspiration and
191 expiration. Briefly, respiratory signal was low-pass filtered (8Hz) to reduce noise, then zero-
192 crossing points were detected to delineate the start points of inspiration and expiration.
193 Finally, the time of each respiratory cycle was linearly converted to phases ranging from 0 to
194 1. In summary, all respiratory cycles were detected, inspiration and expiration phases were
195 well determined, and respiratory phases were computed.

196 *Rescaling time basis of signals with respiratory phase*

197 We wanted to track variations in the signals of interest (LFP, MP, AP discharge) in relation to
198 respiratory rhythm. However, respiratory cycles exhibited variability in duration, and the
199 inspiration-expiration ratio in two cycles with similar duration may be different. The
200 respiratory phase is therefore the appropriate time scale for reducing jitter due to respiratory
201 cycle variability. Following (51), we rescaled the signals of interest as a function of the
202 respiratory phase. Briefly, after detecting the respiratory cycles, the signal for each cycle was
203 interpolated into a 2000-points vector. To account for the asymmetry of respiratory cycles, the
204 average inspiration ratio (inspiration duration/cycle duration) was computed for each
205 recording. Cycle by cycle interpolation preserved the ratio, *i.e.*, for a 0.33 ratio, interpolation
206 attributed 33% of the 2000 points to the inspiration period and the remainder to the expiration
207 period. This rescaling allowed the construction of respiration-triggered MP average (Figure
208 2C bottom) or respiration-triggered AP histogram (Figure 2C top). We made one exception to
209 construct Figure 4B as we wanted to compare the preferred phase of depolarization between
210 cells, so we rescaled all recordings with the same ratio 0.4 in order to align all cell respiratory
211 phases. The average inspiration-expiration ratio of our dataset was 0.41 ± 0.01 .

212 *Time dynamics of respiration-related oscillations*

213

214 In this section, we will describe how we detected **respiration-related oscillations (RRO)** in
215 oscillatory signals (membrane potential and LFP). We proceeded in 5 steps (Figure 1A).

216 (i) Preprocessing and deformation to respiratory phase. For the membrane potential,
217 preprocessing consisted in removing the APs, in order to focus on membrane potential
218 oscillations. As the duration of APs in our population of neurons was homogeneous
219 (Supplemental Figure S3), we used the same cutting values for all our neurons. To remove
220 APs, single APs were cut from 4 ms before to 5 ms after the peak while bursts (defined by
221 interspike interval < 20ms) were cut from 30 ms before the first AP to 30 ms after the last AP.
222 The cut signal was replaced by a linear interpolation between the two extremities. For the
223 LFP, preprocessing consisted in high-pass filtering (0.1 Hz). Once preprocessed the signal of
224 interest was deformed to respiratory phase (see *Rescaling time basis of signals with*
225 *respiratory phase*).

226 (ii) Detection of synchronization with respiration in local windows. As we wanted to track the
227 temporal dynamics of respiration-related oscillations, we scanned the recordings with
228 windows of 4 respiratory cycles, with an overlap of 3 cycles between consecutive windows
229 (Figure 1B.2). For each window, we computed the median signal from the 4 respiratory cycles
230 (example in Figure 1C.2 left, green trace), which represented a proxy for estimating a local
231 oscillation at respiratory frequency. To assess the importance of the respiratory modulation,
232 we measured the amplitude of this median signal (example in Figure 1C.2 left, red bar), which
233 is expected to be high when the oscillations are synchronized with the respiration. To test the
234 significance of such a local respiration-related oscillation, we compared the actual amplitude
235 with amplitudes of 500 surrogates (example in Figure 1C.2 and C.3). We considered the
236 oscillation of the windows to be respiration-related if the actual amplitude was larger than 95th
237 percentile of surrogate amplitudes (example in Figure 1C.3). The construction of the
238 surrogates (see Figure 1C.1) consisted in creating pairs of cycles (cycles 118&119, and

239 121&120 in our example Figure 1C.1), and then phase-shifting the signal. For one pair, we
240 shifted the first cycle with a random phase (φ_1). The other cycle of the pair was shifted in
241 phase opposition ($\varphi_1+\pi$). We repeated this transformation for the second pair of cycles with
242 another random phase shift (φ_2). This manipulation neutralized the respiratory component of
243 the signal.

244 (iii) Verification of the similarity between individual cycles and local oscillation. From the
245 windows previously identified as respiration-related (windows 118 to 120 in our example
246 Figure 1B.2), we wanted to identify the cycles that actually contributed to this respiration-
247 related oscillation. To verify that individual cycles within the window were similar to the
248 local waveform, we computed the scalar product of each cycle with the window median signal
249 (example for one cycle in Figure 1D.1). Since we focused on the waveform, not on the
250 amplitude, we z-scored the signals before performing the scalar product. We assessed the
251 significance by comparing the scalar product of actual data with the scalar product from the
252 500 surrogate window signals (Figure 1D.1 and D.2). We considered the individual cycle
253 significantly similar to the local waveform if the scalar product value was greater than the 95th
254 percentile of surrogated scalar product values (detailed histogram for cycle 118 with window
255 118 in Figure 1D.2).

256 (iv) Cycle by cycle encoding. To switch toward a cycle-by-cycle encoding, we wanted to
257 make sure that the individual cycles, constituting the windows previously identified as
258 respiration-related, actually displayed respiration-related oscillations. To do so we scored
259 each cycle as follows. Each time the Vm of a cycle was similar (black rectangle in Figure
260 1B.3) to the waveform of a respiration-related window (red windows in Figure 1B.2), the
261 score was incremented by one. For example, the cycle 121 (Figure 1B.3) participated in three
262 windows identified as respiration-related (windows 118-120). The cycle 121 was similar to
263 the waveforms of these 3 respiration-related windows and was not tested for the non-

264 modulated (window 121). The cycle 121 reached the score of 3. Since a cycle can be scanned
265 up to four times (in four consecutive windows), score of a cycle ranged from 0 to 4. We
266 classified cycles in three respiration-related categories as follows: 0 and 1: non-modulated
267 cycles, 2: undetermined, 3 and 4: modulated cycles. In our example (see Figure 1B.4) cycles
268 120 and 121, with a score of 3, were then considered as RRo.

269 (v) Curation of isolated events. Finally, we eliminated all single or doublet cycles identified as
270 RRo, and they were reassigned to the undetermined category. Indeed, we considered that
271 respiratory related episodes lasting less than 3 consecutive cycles were not statistically
272 relevant and too much susceptible to false detection.

273 For each recording, we were then able to compute the probability of RRo (expressed in
274 percent), defined as the ratio between the number of respiratory cycles with respiration-related
275 oscillations and the total number of cycles. This probability was not biased by the recording
276 duration (Supplemental Figure S4A, which varies from cell to cell), the absolute respiratory
277 frequency (Supplemental Figure S4B, proxy for depth of anesthesia), the input resistance of
278 the cells (Supplemental Figure S4C). On the contrary, the RRo probability was correlated
279 with coherence to respiration (Supplemental Figure S4D, $p = 5.6 \cdot 10^{-16}$).

280 We performed a last refinement to identify **respiration-modulated cells or LFP**: For further
281 quantification of membrane potential or LFP respiration-related dynamics, we wanted to keep
282 only **cells or LFP** recordings that were significantly synchronized with respiration. If we
283 considered every recording for which RRo were detected as an overall respiration-modulated
284 recording, longer recordings would present greater chance to be defined as “respiration-
285 modulated”. Therefore, to counterbalance this bias, we computed a cell-by-cell threshold on
286 the proportion of required RRo cycles in order to classify the cell or LFP recording as
287 respiration-modulated. To do so, for each recording, we first estimated the respiratory

288 frequency by taking the peak frequency from the respiration power spectrum. The breathing
289 rhythm being extremely stable during anesthesia, a narrow notch filter (mne.notch filter,
290 'spectrum_fit' method, notch width = 1 Hz, mne version 0.20.1) applied on the signal of
291 interest (membrane potential or LFP) removed the respiratory component of the signal, while
292 preserving the other oscillations. The signal was then randomized using Fourier phase
293 randomization technique. Spurious respiration-related oscillations were detected with the
294 previously described algorithm. These steps were repeated 200 times. The number of cycles
295 with respiration-related oscillations detected out of the 200 respiration-deprived signals were
296 gathered and, the 95th percentile was computed. This percentile represented our threshold of
297 significance. If the number of cycles with respiration-related oscillation detected on the true
298 signal was above the threshold, we considered the recording as respiration-modulated. As a
299 result, this selection excluded one V1 neuron from our Base polarity population, (indicated
300 with the black cross on cell 3 from V1 in Figure 3A). One cell in HPC was excluded from
301 Hyp1 population. Two cells in HPC were excluded from Hyp2 population.

302 *Respiration-triggered membrane potential*

303 Using the classification method described previously, we could isolate membrane potential
304 cycles related to respiration. These cycles were then stacked, and we computed the median
305 signal. For each respiration-modulated cell, the amplitude of respiration-related oscillations
306 was defined as the maximum/minimum difference of this median signal. To analyze the
307 depolarization preferred phase, we computed the median respiration-triggered signal, then we
308 extracted the mean vector from which we finally measured the angle. For this latter analysis
309 we only used RRo cycles of respiration-modulated cells.

310 *Respiration-triggered spiking pattern*

311 AP times were rescaled to a respiratory cycle phase basis. Therefore, AP were identified by
312 the number of the respiratory cycle they belong to, and their respiratory phase within that
313 cycle. Once respiratory cycles were classified in respiratory-related categories, we could sort
314 AP according to these categories. Then, for each cell and each respiratory-related category we
315 built a respiration-triggered discharge histogram. For each cell, three experts, independently,
316 visually encoded spiking patterns associated to each respiratory-related category. Spiking
317 patterns were classified as: respiration-related, not respiration-related, or undetermined (not
318 enough spikes to conclude). This classification was inspired from (32).

319 *Power spectrum*

320 For the 4 examples in Figure 2, Morlet wavelet time-frequency maps (0.4-6Hz) for respiratory
321 signal and AP-removed membrane potential were computed for whole recordings. We then
322 extracted the 4-second time windows showed in the figure and averaged across time to
323 estimate the power spectrum. Spectra were normalized by their sum to obtain a density.

324 *Extracellular signal (LFP) processing*

325 The LFP signal was 0.1 Hz high pass filtered. Then, similarly to membrane potential
326 oscillations, LFP-respiration relationship was classified cycle by cycle. Using this
327 classification, the relationship to respiratory rhythm was then compared between membrane
328 potential and LFP.

329 *Conditional probabilities*

330 For this analysis, we kept recordings for which both membrane potential and LFP exhibited
331 significant respiration-modulation (as described in *Time dynamics of respiration-related MP*
332 *oscillations*). In Figure 5D, we examined the conditional probability to observe membrane
333 potential RRo in relation to LFP dynamics. In other words, we split recording according to the

334 LFP RRo encoding. On the one hand we computed membrane potential RRo probability
335 among cycles with LFP RRo. On the other hand, we again computed the membrane potential
336 RRo probability among cycles during which LFP activity was not related to respiration (other
337 LFP activity). Figure 5E is the complementary view, since we computed the conditional
338 probability to observe LFP RRo according to membrane potential dynamics (RRo versus
339 other activity).

340 *Statistics*

341 Statistics were made with Scipy (version 1.4.1) python package (scipy.stats.wilcoxon,
342 scipy.stats.mannwithneyu, scipy.stats.kruskal). All averages are presented under the format
343 mean \pm SEM. Where this is not the case, it is explicitly stated.

344

345

346 RESULTS

347 **Evidence of respiratory modulation of cell membrane potential in non-olfactory**
348 **structures**

349 *In vivo* intracellular activities recordings were obtained from 4 non-olfactory structures in 25
350 freely breathing anesthetized rats; all polarity states combined, 34 cells were recorded in
351 medial prefrontal cortex (mPFC, N = 9 rats), 13 cells in somatosensory cortex (S1, N = 4
352 rats), 5 cells in visual cortex (V1, N = 4 rats), and 17 cells in hippocampus (HPC, N = 8 rats).
353 The respiratory frequencies ranged from 1.44 Hz to 2.08 Hz (1.78 ± 0.03 Hz, N = 25 rats).

354 In all these four areas, we found cells exhibiting respiration-related oscillations (RRO) in their
355 membrane potential (MP) and respiration-related (RR) spiking discharge, as shown in the raw
356 traces examples (Figure 2A). To highlight the RRO, the filtered respiratory signal (light gray)
357 was superimposed on the raw traces of the intracellular MP oscillations. In Figure 2B, the
358 power spectral density analysis of the MP signal (black PSD) showed a clear power peak at
359 the respiratory frequency (gray PSD) confirming that MP follows the respiratory rhythm in
360 these examples.

361 In order to properly characterize the relation between MP and respiratory rhythm, we
362 developed a cycle-by-cycle method for detecting the respiratory cycles during which MP
363 oscillated at the respiratory frequency (see Figure 1 and materials and methods for details).
364 We plotted the median of respiration-triggered membrane potential (Figure 2C bottom panels)
365 and the histograms of spiking occurrence during these cycles (Figure 2C top panels). In the
366 examples in Figure 2, MP RRO were large (median amplitude \pm 95% confidence interval:
367 mPFC = 5.5 ± 0.9 mV, S1 = 14.5 ± 2.1 mV, V1 = 8.8 ± 2.2 mV, HPC = 3.1 ± 0.8 mV; Figure 2C
368 bottom panels), and APs occurrences were clearly related to respiratory cycle (top panels in
369 Figure 2C). Moreover, on these examples, MP RRO and firing pattern were similarly phase-

370 locked to respiration, suggesting a strong correlation between MP RRo and the associated
371 firing. Indeed, spikes appeared on the top of the oscillation, where the MP exhibited the
372 greatest depolarization.

373

374 In this way, we evidenced that a respiratory modulation could be expressed in the intracellular
375 activity (MP and spiking) of cells in the four non-olfactory areas we recorded. The next step
376 aimed to better characterize this MP respiratory modulation in each structure.

377

378 We first quantified the percentage of cells exhibiting MP RRo at resting membrane potential
379 (Base polarity). As explained in Methods, we considered RRo only if the episode lasted at
380 least 3 consecutive respiratory cycles. We measured in each cell, and in the 4 structures, the
381 percentage of respiratory cycles during which an MP RRo was detected (Figure 3A), then we
382 plotted the cumulative percentage of MP RRo cycles across cells for mPFC, S1 and HPC
383 (Figure 3B). Since too few V1 recordings were available, V1 will not be included in the
384 following comparative analyses. Additional shuffling statistics (see methods) revealed
385 respiration-modulated cells represented 65.6% of cells (21/32) in mPFC, 70% (7/10) in S1,
386 and 80% (8/10) in HPC. Therefore, at least 65% of the recorded cells in each structure
387 presented a respiratory modulation. Additionally, the two cortical structures (mPFC and S1)
388 presented slightly lower proportions of respiration-modulated cells than HPC. Specifically,
389 the cumulative percentage curves (Figure 3B) showed that 20% of the cells in S1 and mPFC
390 had more than 20% of their MP cycles modulated by respiration, about 20% of HPC cells had
391 more than 75% of their cycles modulated.

392 As shown in Figure 3A1-4, the percentages of MP RRo cycles varied greatly from cell to cell
393 within the same structure, from one structure to another. Indeed, if only the respiration-

394 modulated cells were considered (black bars without black cross), the percentage of cycles
395 presenting an MP RRo ranged from 1.3% to 90.5% in mPFC ($15.3\pm 4.7\%$, $N = 21$ cells) and
396 from 3.2% to 88.9% in HPC ($29.5\pm 11.4\%$, $N = 8$ cells), while it only extended from 3.9 to
397 33.3% in S1 ($13.4\pm 3.9\%$, $N = 7$ cells) (Figure 3C). Three neurons (one in mPFC and two in
398 HPC) exhibited an extreme proportion of MP RRo cycles (more than 90%). The low
399 percentage of MP RRo cycles for some cells was certainly due to our highly selective method
400 for detecting RRo cycles (see material and methods). Due to the high variability between
401 cells, no significant difference in proportion of MP RRo cycles between structures was
402 observed (Figure 3C, Kruskal-Wallis test, $p = 0.53$, $H(2) = 1.24$).

403 To go one step further, we quantified for each respiration-modulated cell the average duration
404 of RRo episodes (*i.e.*, the number of consecutive cycles in an MP RRo). As shown in Figure
405 3D, there was no significant difference in the duration of RRo episodes between structures
406 (Figure 3D), (Kruskal-Wallis test, $p = 0.58$, $H(2) = 1.07$). In fact, the average number of
407 consecutive MP RRo cycles was around five regardless of the structure (mPFC: 5.0 ± 0.7 , $N =$
408 21 cells; S1: 5.3 ± 1.4 , $N = 7$ cells; HPC: 5.8 ± 1.2 , $N = 8$ cells).

409

410 Taken together, these results showed that at least 65% of cells of each recorded structures
411 could synchronize their MP oscillations with respiration. However, this respiratory
412 modulation of MP was 1) highly variable both between structures and between cells, 2) very
413 transient for most cells (with a mean episode duration of 5 cycles).

414 Next, to further explore the intrinsic dynamics of these RRo episodes, we studied both their
415 amplitudes and phases. We first measured the amplitude of the median respiration-triggered
416 MP (see material and methods for details) for each modulated cell (Figure 4A). Figure 4A
417 revealed that amplitude of MP RRo was higher in S1 (10.7 ± 2.4 mV, $N = 7$ cells) compared to

418 mPFC (4.0 ± 0.5 mV, N = 21 cells) and HPC (4.4 ± 0.9 mV, N = 8 cells) (Kruskal-Wallis test, p
419 = 0.01, $H(2) = 9.1$, *post hoc* Mann-Whitney U test S1–mPFC: p = 0.002 and S1-HPC: p =
420 0.02).

421

422 To pursue, we characterized the respiratory phase of the MP RRo. As already suggested by
423 the illustrative examples in Figure 2, the MP RRo phases were not unique. Indeed, while MP
424 exhibited maximal depolarized values around the transition phase between inspiration and
425 expiration in S1 and HPC examples (Figure 2C2 and 2C4), MP of mPFC and V1 examples
426 exhibited hyperpolarized values at this same respiratory phase (Figure 2C1 and 2C3). On the
427 whole dataset, we extracted, for each cell the phase of the most depolarized value of the
428 median MP RRo (see Figure 2, panels C-bottom for examples of median MP RRo). Figure 4B
429 shows for each structure the preferred phase for each cell. For mPFC, S1 and HPC these
430 phases seemed to be distributed according to two preferential values, one between 130-170
431 degree and the other between 315-340 degree. These ranges of values approximately
432 corresponded respectively to a period between the end of inspiration and the beginning of
433 expiration, *i.e.* near inspiration to expiration (I/E) transition (at 140 degree) and another
434 around the end of expiration, before expiration to inspiration (E/I) transition (at 0 degree).

435 Finally, we examined in detail the relationship between MP RRo and spiking discharge
436 (Respiration-Related (RR) or not). Table 1 presented the different association's possibilities
437 between MP (RRo or no-RRo) and discharge (RR discharge and no-RR discharge). To
438 construct this table, we considered the distribution of spikes as a function of respiratory phase
439 (see Figure 2C, top panels) using previously described method used in the olfactory bulb (34,
440 35) (see Material and Methods for details). For a given cell, we could compute two spike
441 distributions on the respiratory phase, one with the spikes occurring during period of MP RRo

442 and the other with the spikes occurring during MP no-RRo. The two spike distributions were
443 then classified as either respiration-related (RR) discharge or non-respiration-related (no-RR)
444 discharge. However, if the cell had no MP RRo period or if there were too few AP, spike
445 distribution could not be classified, which explains the N in table 1 are lower than those
446 observed in Figure 3. Table 1 summarized this classification across cells for each structure. It
447 showed the percentages of each association between spiking discharge patterns (columns) and
448 respiration-triggered MP types (lines). In mPFC and S1, the MP RRo was mostly associated
449 with the RR discharge pattern (80% for both structures) and conversely the MP no-RRo was
450 mostly associated with the No RR discharge pattern (71.4% and 85.7% respectively). The
451 results for HPC did not show the same association profile. Indeed, only 25% of the MP RRo
452 was associated with RR discharge pattern.

453

454 In summary, we showed that MP RRo were larger for cells in S1. The respiratory phases were
455 mainly around I/E and E/I transitions for mPFC, S1 and HPC cells. Moreover, respiratory
456 coupling between MP and spiking discharge was much more frequent in mPFC and S1 (80%
457 each) than in HPC (25%).

458 Given the transient nature of MP RRo and their variability in amplitude and phase, we
459 wondered whether parameters such as cell intrinsic state or local network state could
460 influence respiratory modulation of MP.

461

462 **Influence of intrinsic excitability and network activity on the respiratory modulation of**
463 **membrane potential**

464 Our second aim was then to examine to what extent MP RRo could be influenced by (i)
465 intracellular excitability, and (ii) by local field potential (LFP) state.

466 (i) Effect of intracellular excitability.

467 First, we investigated the effect of excitability level on the probability for MP to be modulated
468 by the respiration. Briefly, we tested two hyperpolarization levels (Hyp1 and Hyp2) and one
469 depolarized level (Dep) by progressive direct intracellular DC current injection. As described
470 in materials and methods, we used 3 criteria (firing, MP value, and injected current) to
471 classify these excitability states.

472 The proportions of respiration-modulated cells (mPFC, S1 and HPC were pooled) were
473 relatively stable between the four different excitability levels (Supplemental Figure S5A,
474 Hyp2: 58% - 18 out 31 cells, Hyp1: 52% - 13 out of 25 cells, Base: 69% - 36 out of 52 cells,
475 Dep: 62% - 13 out of 21 cells). Thus, considering the whole population, excitability level did
476 not clearly influence the proportion of respiration-modulated cells.

477 Next, we compared the percentages of MP RRo cycles for a same cell when it was
478 hyperpolarized from Base to Hyp1 level (Supplemental Figure S5B). This cell-by-cell
479 analysis was necessary because of the great variability of MP RRo cycles proportion at base
480 level within a same structure (*cf.* Figure 3). This analysis indicated that, the percentage of MP
481 RRo cycles was reduced upon hyperpolarizing the cell (Base: $19.6 \pm 6.8\%$, Hyper1:
482 $13.3 \pm 7.0\%$, Wilcoxon test $p = 0.01$, $Z = 7.0$, $N = 16$ cells)

483 These data showed that, although the internal excitability state had a weak influence on the
484 proportion of respiration-modulated cells at the whole population level, moderate
485 hyperpolarization of a cell decreased its percentage of MP RRo cycle.

486 (ii) Effect of LFP state.

487 As it is now well established that LFP in these brain structures can be modulated by
488 respiration (25, 26), we then wondered how MP and LFP respiratory rhythms could interact.
489 The raw traces in Figure 5A1 showed an example where neither MP nor LFP displayed RRo.
490 Conversely, for the same recording, a few minutes later (Figure 5A2, after 2 min), respiratory
491 frequency appeared simultaneously in MP and LFP signals. The similarity between both
492 signals is highlighted when superimposing filtered respiratory signal (light gray traces on MP
493 and LFP raw signals).

494 LFP RRo cycles were detected in the same way than MP RRo cycles, in order to be able to
495 compare their temporal dynamics (see Methods). LFP RRo were found in the three structures.
496 Figure 5B showed the average percentage of LFP RRo per recording (gray dots) and the
497 percentages per structure (dark-gray squares). mPFC showed a significantly higher percentage
498 of LFP RRo cycles than S1 (Kruskal-Wallis test, $p = 0.03$, $H(2) = 6.96$, *post hoc* Mann-
499 Whitney U test mPFC-S1, $p = 0.01$). The same tendency was observed with HPC, but
500 significance was not reached (Mann-Whitney U test mPFC-H, $p = 0.07$). In the same way as
501 for the MP recordings, we quantified for each LFP recording the mean duration of LFP RRo
502 episodes. The mean number of consecutive LFP RRo cycles (Figure 5C) was significantly
503 higher for mPFC (10.6 ± 2.2 cycles, $N = 21$ recordings) than for S1 (3.8 ± 0.2 cycles, $N = 5$
504 recordings), and HPC (4.2 ± 0.4 cycles, $N = 4$ recordings). (Kruskal Wallis $p = 7.10^{-3}$, $H(2) =$
505 10.0 , *post hoc* Mann-Whitney U test mPFC-S1: $p = 0.005$, mPFC-HPC: $p = 0.02$)

506 We then investigated the extent to which LFP and MP respiration-modulation could be
507 correlated. To do this, we calculated the percentage MP RRo cycles when the LFP exhibited
508 RRo or not (Figure 5D). It became clear that, irrespective of the structure considered, the
509 probability of the MP displaying RRo was higher when the LFP displayed RRo. For mPFC,
510 the increased MP RRo probability when LFP switched from no RRo to RRo was significant
511 (Wilcoxon test $p = 0.02$, $Z = 44$, $N = 20$ cells). HPC exhibited the same tendency (Wilcoxon

512 test, $p = 0.07$, $Z = 0$, $n = 4$), and, to a lesser extent S1 also (Wilcoxon test, $p = 0.14$, $Z = 1$, $n = 4$).
513 Reciprocally, we wondered whether the probability of the LFP displaying RRo could be
514 different when the MP displayed RRo or not (Figure 5E). We found that in mPFC, S1 and
515 HPC, the probability of the LFP displaying RRo was higher when the MP displayed RRo, but
516 this result was statistically significant only for mPFC (Wilcoxon test mPFC: $p = 0.001$, $Z =$
517 21 , $N = 20$ cells). This confirmed that, most of the time, the respiratory modulation of a cell
518 MP is coherent with the respiratory modulation of its network environment. Note that the lack
519 of statistical significance in regions others than mPFC is probably due to the small sample
520 size.

521 To summarize, we observed that MP and LFP are often respiratory co-modulated.

522

523

524 DISCUSSION

525 While many studies have recently evidenced a respiration-related activity in LFPs of many
526 brain regions (21–26, 28, 29), our study is the first to describe respiratory-related oscillations
527 in the MP activity in widespread brain areas, namely mPFC, S1, and HPC. We showed
528 respiration-related oscillations of MP in a large proportion of cells (65.6% in mPFC, 70% in
529 S1, and 80% in HPC). Before us, very sparse and scarce data existed reporting MP
530 oscillations coherent with LFP respiration-related oscillation ((26, 29): 1 cell recorded in
531 parietal cortex; (21): 3 cells out of 7 recorded in dentate gyrus after brainstem stimulation,
532 (50) : 6 cells in parietal cortex). Our study goes further by comparing the probability of MP
533 respiration-related oscillations in three different brain regions and by detailing how such
534 oscillation could be influenced by excitability state or network activity. These results suggest
535 that respiratory rhythm can entrain brain activity down to the membrane potential. In addition,
536 we recorded in a preliminary study two prefrontal neurons whose membrane potential RRo
537 was suppressed by ipsilateral nostril obstruction (Supplemental Figure S6). If extended, such
538 results would definitively show that MP RRo in mPFC are entrained by respiratory nasal
539 airflow. Contrarily to anesthetized animals, awake animals display various pattern of
540 respiration. It has been shown that respiratory modulation is stronger for low frequency
541 respiration (52–54), more precisely for short and deep inspirations (25, 55). During
542 anesthesia, respiration tends to have a slow frequency and the inspiration tends to be shorter
543 than the expiration. In our dataset, the mean respiratory frequency was 1.78Hz and the mean
544 inspiration duration was 230ms. This inspiration duration is close to the optimal inspiration
545 duration (100-200ms) previously observed during quiet wake (25, 55). Since the anesthesia
546 brought the animal's respiration in a pattern close to the optimum, it could explain the extent
547 of the respiratory modulation that we observed in this study. Finally, it would also be
548 interesting to test whether the MP of cells in some brain structures could still be respiration

549 modulated in the absence of any OB afferents since it has been shown that cerebral activity
550 can also be modulated due to the presence of an intracerebral respiratory corollary discharge
551 originating from the brainstem (27).

552 Accumulating data showed that respiration can influence LFP from almost all brain areas (20,
553 25, 26), the unitary activity of several areas (22, 24, 27) and cognition (10, 13, 14). We
554 complementarily showed that membrane potential and action potential can be modulated by
555 respiration, and thus we reinforce the idea that respiratory rhythm could act as a common
556 clock for the brain. In this view, we expected the respiratory rhythm to synchronize
557 hyperpolarization and depolarization from distant neurons, facilitating their communication.
558 However, we rather observed that RRo displayed variable phase-locking to the respiration
559 (Figure 4B). Indeed, independently from the brain areas, neurons' depolarization tended to
560 phase-lock either to the inspiration-expiration transition or to the expiration-inspiration
561 transition. However, is this finding completely against the hypothesis of a communication
562 through respiratory rhythm? The same variability in phase-locking was observed in the
563 olfactory bulb (35, 40). In addition, Ackels and colleagues (40) showed mitral cells tended to
564 be depolarized during inspiration, while tufted cells depolarization occurred during expiration,
565 suggesting that the phase of respiratory rhythm could mediate information multiplexing. The
566 variability in phase-locking observed here (Figure 4B) suggests this multiplexing continues
567 beyond the olfactory bulb.

568 We first evidenced that the proportions of cells expressing MP respiration-related oscillations
569 were disparate between brain regions. In fact, cells were more likely to display RRo in HPC
570 than in S1 or mPFC (Figure 3B). Moreover, the amplitude of MP RRo was larger in S1 than
571 in other brain regions, at least in a sub-population of cells (see Figure 4A). Interestingly,
572 extracellularly recorded spiking activity in S1 has been previously described as rhythmically
573 correlated with respiration (22). This stronger relation between respiration and S1 barrel

574 cortex cells could have something to do with the fact that respiration binds different orofacial
575 rhythms as whisking and sniffing particularly during olfactory search (7, 56, 57). Even if there
576 is no whisker movement in the anesthetized preparation, olfactory and whisker cortices could
577 be functionally linked since a crosswire between piriform and barrel cortices has been
578 revealed in mouse brain slices after whiskers are trimmed, leading to a cross-modal sensory
579 plasticity (58). Such a link could explain our observation.

580 We expected cells in mPFC to display more clearly respiration-related oscillations in their
581 MP, given the strong modulation of LFP described in this region in the freely moving animal
582 (25, 27, 59, 60). However, neither the duration of MP respiration-modulated episodes nor the
583 amplitude of MP RRo is more pronounced in mPFC than in the three other areas. The fact that
584 anesthetics have voltage-dependent effects on resting MP (61) could lower MP sensitivity to
585 respiration. However, even in the awake head-fixed mouse, (24) reported that spiking
586 discharges of neurons in mPFC are much less coupled to breathing than neurons in orbito-
587 frontal cortex. This different probability of respiration-related oscillation between LFP and
588 intracellular signals raises the question of a possible contamination of LFP signals by volume
589 conduction coming from distant generators (62, 63).

590 Oppositely, we showed a very clear respiration-related activity in MP of HPC cells (80% of
591 cells expressed RRo and 20% of them displayed at least 75% of modulated cycles). This
592 probably reflects the strong relationship between olfactory and limbic structures. Very
593 recently, Zhou et al. (64) reported that human hippocampal connectivity is stronger with
594 olfactory cortex than other sensory cortices and that olfactory-hippocampal connectivity
595 oscillates with nasal breathing. Interestingly, several recordings of HPC neurons exhibiting
596 MP RRo revealed that, when emitted in bursts, action potentials were locked to the I/E
597 transition point of respiratory cycle while single action potentials were more loosely
598 distributed over the respiratory cycle (data not shown). It is commonly admitted that bursts

599 could facilitate synaptic transmission (65) and induce synaptic plasticity, LTP or LTD,
600 depending on the theta oscillatory cycle phase of the burst (66). Furthermore, bursts increase
601 detection of presynaptic events (67), thanks to their higher signal to noise ratio compared to
602 single action potential. The fact that the MP of HPC cells could be paced by breathing could
603 thus influence place cells coding notably during sniffing where respiratory and theta rhythms
604 are in the same frequency range (25, 26).

605 We only recorded 4 cells in V1, 2 of them showed episodes of MP RRo. In these cells, the
606 percentage of MP RRo did not exceed more than about 25% of total cycles. This confirms the
607 observation that LFP in posterior areas are less susceptible to respiratory drive (25, 26). It is
608 nevertheless interesting to observe that, even in a region as posterior as V1 and so improbably
609 connected to the olfactory input, respiration can affect subthreshold activity.

610 We previously showed that the MP of mitral/tufted cells, the principal neurons of the
611 olfactory bulb, is strongly respiration-modulated (35). We cannot compare the percentages of
612 modulated cells between the non-olfactory structures recorded here and olfactory bulb
613 neurons because we did not use the same experimental paradigm, neither the same detection
614 method nor data processing. However, it can be noted that hyperpolarization of mPFC and S1
615 cells appears to reduce the number of cells exhibiting MP RRo. On the contrary, in the
616 olfactory bulb, hyperpolarization induced an increase in the proportion of respiratory
617 modulated cells, which could suggest a reversal potential of MP RRo more hyperpolarized in
618 non-olfactory structures than in olfactory bulb. It would be necessary to undertake a fine
619 characterization of the different types of RRo that may exist (as in mitral/tufted cells) in order
620 to be able to formulate hypotheses. However, we could postulate that a greater inhibition
621 would be implicated in the oscillatory dynamics of these non-olfactory cells, which is in line
622 with recent findings (50). Indeed, it has been shown that inhibitory interneurons appear to be
623 more respiratory modulated than pyramidal neurons (23, 27). It would be of great interest to

624 test the possibility that the consequence of a hyperpolarization (increase or a decrease of the
625 respiratory modulation) would come from the fact that the respiratory influence could rather
626 impinge on inhibitory interneurons (as it could be in mPFC) or on a balanced excitatory-
627 inhibitory network (as in OB).

628 LFP RRo episodes were variable between regions, with duration up to 10 cycles in mPFC and
629 between 3 and 6 cycles for other structures. This longer LFP RRo episode duration in mPFC
630 is not surprising since several studies observed a particularly strong respiratory modulation in
631 the prelimbic cortex. Then, the respiratory rhythm may take over more often than other
632 rhythms. Alternating populations of cells could transiently express respiratory modulation in
633 their MP resulting in a longer duration of LFP respiratory modulation. In other structures, the
634 competition between several other LFP rhythms (such as slow rhythm or theta) probably does
635 not favor the emergence of this rhythm. Additionally, for all the structures, the sensitivity of
636 each individual cell MP to LFP respiratory modulation likely differs according to its
637 excitability state, limiting episodes of MP RRo to short durations. This could be supported by
638 our observation that cells could show less MP RRo when hyperpolarized (see Figure S5B).

639 Although our study focused on the respiratory rhythm, the influence of other LFP rhythms on
640 cellular activity (MP and discharge) should not be ignored. For examples, in few cases, we
641 observed a spiking discharge that was RR while MP displayed no RRo. We could suppose
642 that if the spiking discharge is influenced by a rhythm twice slower or twice faster than the
643 respiratory rhythm, they will appear locked on respiratory cycle on spike histograms. We also
644 observed more complicated patterns with several rhythms nested. For example, on the MP
645 spectrum analysis of Figure 2B3 we can distinguish a slow oscillation (around 1 Hz) which
646 could compete with the respiratory rhythm.

647 Overall, our results evidenced a consequent proportion of RRo within electrophysiological
648 signals, which could only have been revealed thanks to respiration co-recording. We
649 evidenced a strong respiratory coupling between MP and spike discharge in PFC, S1 and to a
650 lesser extent in HPC. Furthermore, MP and LFP RRo dynamics showed strong covariations.
651 Even if our experiments in anesthetized animal are purely descriptive, our results show that, at
652 very different places in the brain, the MP of neurons can be depolarized and hyperpolarized
653 according to the respiratory rhythm. These findings contribute to the evidence that the
654 respiratory rhythm could be used as a common clock to set the dynamics of large-scale
655 networks on the same slow rhythm.

656

657

658 SUPPLEMENTAL MATERIAL

659 Figure S1: **Input resistance**

660 Figure S2: **Polarity states**

661 Figure S3: **AP waveform**

662 Figure S4: **Verification of possible correlations**

663 Figure S5: **Effect of excitability changes**

664 Figure S6: **Obstruction of the nostril**

665

666 Supplemental figures availability: <https://doi.org/10.5281/zenodo.8165176>

667

668 ACKNOWLEDGMENTS

669 We would like to thank Marc Thévenet and Belkacem Messaoudi for providing technical

670 assistance.

671

672 AUTHOR CONTRIBUTIONS

673 C.A. and N.B. conceived and designed research; M.J. and M.Z. performed experiments; M.J.,
674 N.F-T., S.G., C.A developed the RRo detection method; M.J analyzed data; M.J, N.F-T., N.B.
675 and C.A. interpreted results of experiment; M.J. and C.A. prepared figures. M.J., N.B. and
676 C.A. drafted manuscript; M.J., M.Z., N.F-T., N.B. and C.A. edited and revised manuscript;
677 M.J., M.Z., N.F-T., S.G., N.B. and C.A approved final version of manuscript.

678

679

680

681

682 REFERENCES

- 683 1. **Heck DH, McAfee SS, Liu Y, Babajani-Feremi A, Rezaie R, Freeman WJ,**
684 **Wheless JW, Papanicolaou AC, Ruzsinkó M, Sokolov Y.** Breathing as a fundamental
685 rhythm of brain function. *Front Neural Circuits* 10: 115, 2017.
- 686 2. **Heck DH, Kozma R, Kay LM.** The rhythm of memory: how breathing shapes
687 memory function. *J Neurophysiol* 122: 563–571, 2019.
- 688 3. **Maric V, Ramanathan D, Mishra J.** Respiratory regulation & interactions with
689 neuro-cognitive circuitry. *Neurosci Biobehav Rev* 112: 95–106, 2020.
- 690 4. **Zaccaro A, Piarulli A, Laurino M, Garbella E, Menicucci D, Neri B, Gemignani**
691 **A.** How breath-control can change your life: a systematic review on psycho-physiological
692 correlates of slow breathing. *Front Hum Neurosci* 12: 353, 2018.
- 693 5. **Cao Y, Roy S, Sachdev RN, Heck DH.** Dynamic correlation between whisking and
694 breathing rhythms in mice. *J Neurosci* 32: 1653–1659, 2012.
- 695 6. **Kurnikova A, Moore JD, Liao S-M, Deschênes M, Kleinfeld D.** Coordination of
696 orofacial motor actions into exploratory behavior by rat. *Curr Biol* 27: 688–696, 2017.
- 697 7. **Moore JD, Deschênes M, Furuta T, Huber D, Smear MC, Demers M, Kleinfeld**
698 **D.** Hierarchy of orofacial rhythms revealed through whisking and breathing. *Nature* 497:
699 205–210, 2013.
- 700 8. **Lu L, Cao Y, Tokita K, Heck DH, Boughter Jr JD.** Medial cerebellar nuclear
701 projections and activity patterns link cerebellar output to orofacial and respiratory behavior.
702 *Front Neural Circuits* 7: 56, 2013.
- 703 9. **Welzl H, Bureš J.** Lick-synchronized breathing in rats. *Physiol Behav* 18: 751–753,
704 1977.
- 705 10. **Arshamian A, Iravani B, Majid A, Lundström JN.** Respiration modulates olfactory
706 memory consolidation in humans. *J Neurosci* 38: 10286–10294, 2018.
- 707 11. **Waselius T, Wikgren J, Penttonen M, Nokia MS.** Breathe out and learn: expiration-
708 contingent stimulus presentation facilitates associative learning in trace eyeblink conditioning.
709 *Psychophysiology* 56: e13387, 2019.
- 710 12. **Nakamura NH, Fukunaga M, Oku Y.** Respiratory modulation of cognitive
711 performance during the retrieval process. *PloS One* 13: e0204021, 2018.
- 712 13. **Perl O, Ravia A, Rubinson M, Eisen A, Soroka T, Mor N, Secundo L, Sobel N.**
713 Human non-olfactory cognition phase-locked with inhalation. *Nat Hum Behav* 3: 501–512,
714 2019.
- 715 14. **Zelano C, Jiang H, Zhou G, Arora N, Schuele S, Rosenow J, Gottfried JA.** Nasal

- 716 respiration entrains human limbic oscillations and modulates cognitive function. *J Neurosci*
717 36: 12448–12467, 2016.
- 718 15. **Grund M, Al E, Pabst M, Dabbagh A, Stephani T, Nierhaus T, Gaebler M,**
719 **Villringer A.** Respiration, heartbeat, and conscious tactile perception. *J Neurosci* 42: 643–
720 656, 2022.
- 721 16. **Iwabe T, Ozaki I, Hashizume A.** The respiratory cycle modulates brain potentials,
722 sympathetic activity, and subjective pain sensation induced by noxious stimulation. *Neurosci*
723 *Res* 84: 47–59, 2014. doi: 10.1016/j.neures.2014.03.003.
- 724 17. **Li S, Park W-H, Borg A.** Phase-dependent respiratory-motor interactions in reaction
725 time tasks during rhythmic voluntary breathing. *Motor Control* 16: 493–505, 2012.
- 726 18. **Li S, Laskin JJ.** Influences of ventilation on maximal isometric force of the finger
727 flexors. *Muscle Nerve* 34: 651–655, 2006. doi: <https://doi.org/10.1002/mus.20592>.
- 728 19. **Park H-D, Barnoud C, Trang H, Kannape OA, Schaller K, Blanke O.** Breathing is
729 coupled with voluntary action and the cortical readiness potential. *Nat Commun* 11: 1–8,
730 2020.
- 731 20. **Herrero JL, Khuvis S, Yeagle E, Cerf M, Mehta AD.** Breathing above the brain
732 stem: volitional control and attentional modulation in humans. .
- 733 21. **Yanovsky Y, Ciatipis M, Draguhn A, Tort AB, Brankač J.** Slow oscillations in
734 the mouse hippocampus entrained by nasal respiration. *J Neurosci* 34: 5949–5964, 2014.
- 735 22. **Ito J, Roy S, Liu Y, Cao Y, Fletcher M, Lu L, Boughter JD, Grün S, Heck DH.**
736 Whisker barrel cortex delta oscillations and gamma power in the awake mouse are linked to
737 respiration. *Nat Commun* 5: 3572, 2014. doi: 10.1038/ncomms4572.
- 738 23. **Biskamp J, Bartos M, Sauer J-F.** Organization of prefrontal network activity by
739 respiration-related oscillations. *Sci Rep* 7: 1–11, 2017.
- 740 24. **Köszeghy Á, Lasztóczy B, Forro T, Klausberger T.** Spike-timing of orbitofrontal
741 neurons is synchronized with breathing. *Front Cell Neurosci* 12: 105, 2018.
- 742 25. **Girin B, Juventin M, Garcia S, Lefèvre L, Amat C, Fourcaud-Trocmé N,**
743 **Buonviso N.** The deep and slow breathing characterizing rest favors brain respiratory-drive.
744 *Sci Rep* 11: 1–15, 2021.
- 745 26. **Tort ABL, Ponsel S, Jessberger J, Yanovsky Y, Brankač J, Draguhn A.** Parallel
746 detection of theta and respiration-coupled oscillations throughout the mouse brain. *Sci Rep* 8:
747 6432, 2018. doi: 10.1038/s41598-018-24629-z.
- 748 27. **Karalis N, Sirota A.** Breathing coordinates cortico-hippocampal dynamics in mice
749 during offline states. *Nat Commun* 13: 1–20, 2022.
- 750 28. **Liu Y, McAfee SS, Heck DH.** Hippocampal sharp-wave ripples in awake mice are

- 751 entrained by respiration. *Sci Rep* 7: 1–9, 2017.
- 752 29. **Zhong W, Ciatipis M, Wolfenstetter T, Jessberger J, Müller C, Ponsel S,**
753 **Yanovsky Y, Brankač J, Tort AB, Draguhn A.** Selective entrainment of gamma subbands
754 by different slow network oscillations. *Proc Natl Acad Sci* 114: 4519–4524, 2017.
- 755 30. **Grosmaître X, Santarelli LC, Tan J, Luo M, Ma M.** Dual functions of mammalian
756 olfactory sensory neurons as odor detectors and mechanical sensors. *Nat Neurosci* 10: 348–
757 354, 2007.
- 758 31. **Lockmann AL, Laplagne DA, Leão RN, Tort AB.** A respiration-coupled rhythm in
759 the rat hippocampus independent of theta and slow oscillations. *J Neurosci* 36: 5338–5352,
760 2016.
- 761 32. **Buonviso N, Amat C, Litaudon P.** Respiratory modulation of olfactory neurons in
762 the rodent brain. *Chem Senses* 31: 145–154, 2006.
- 763 33. **Buonviso N, Amat C, Litaudon P, Roux S, Royet J-P, Farget V, Sicard G.** Rhythm
764 sequence through the olfactory bulb layers during the time window of a respiratory cycle. *Eur*
765 *J Neurosci* 17: 1811–1819, 2003.
- 766 34. **Cenier T, David F, Litaudon P, Garcia S, Amat C, Buonviso N.** Respiration-gated
767 formation of gamma and beta neural assemblies in the mammalian olfactory bulb. *Eur J*
768 *Neurosci* 29: 921–930, 2009.
- 769 35. **Briffaud V, Fourcaud-Trocmé N, Messaoudi B, Buonviso N, Amat C.** The
770 relationship between respiration-related membrane potential slow oscillations and discharge
771 patterns in mitral/tufted cells: what are the rules? *PLoS One* 7: e43964, 2012.
- 772 36. **Courtiol E, Amat C, Thevenet M, Messaoudi B, Garcia S, Buonviso N.** Reshaping
773 of bulbar odor response by nasal flow rate in the rat. *PLoS One* 6: e16445, 2011.
- 774 37. **David FO, Hugues E, Cenier T, Fourcaud-Trocmé N, Buonviso N.** Specific
775 entrainment of mitral cells during gamma oscillation in the rat olfactory bulb. *PLoS Comput*
776 *Biol* 5: e1000551, 2009.
- 777 38. **Cury KM, Uchida N.** Robust odor coding via inhalation-coupled transient activity in
778 the mammalian olfactory bulb. *Neuron* 68: 570–585, 2010.
- 779 39. **Gschwend O, Beroud J, Carleton A.** Encoding odorant identity by spiking packets
780 of rate-invariant neurons in awake mice. *PLoS One* 7: e30155, 2012.
- 781 40. **Ackels T, Jordan R, Schaefer AT, Fukunaga I.** Respiration-locking of olfactory
782 receptor and projection neurons in the mouse olfactory bulb and its modulation by brain state.
783 *Front Cell Neurosci* 14: 220, 2020.
- 784 41. **Fourcaud-Trocmé N, Briffaud V, Thévenet M, Buonviso N, Amat C.** In vivo beta
785 and gamma subthreshold oscillations in rat mitral cells: origin and gating by respiratory
786 dynamics. *J Neurophysiol* 119: 274–289, 2018.

- 877 42. **Courtiole E, Hegoburu C, Litaudon P, Garcia S, Fourcaud-Trocme N, Buonviso**
878 **N.** Individual and synergistic effects of sniffing frequency and flow rate on olfactory bulb
879 activity. *J Neurophysiol* 106: 2813–2824, 2011.
- 790 43. **Esclassan F, Courtiole E, Thevenet M, Garcia S, Buonviso N, Litaudon P.** Faster,
791 deeper, better: the impact of sniffing modulation on bulbar olfactory processing. *PloS One* 7:
792 e40927, 2012.
- 793 44. **Oka Y, Takai Y, Touhara K.** Nasal airflow rate affects the sensitivity and pattern of
794 glomerular odorant responses in the mouse olfactory bulb. *J Neurosci* 29: 12070–12078,
795 2009.
- 796 45. **Courtiole E, Buonviso N, Litaudon P.** Odorant features differentially modulate
797 beta/gamma oscillatory patterns in anterior versus posterior piriform cortex. *Neuroscience*
798 409: 26–34, 2019.
- 799 46. **Litaudon P, Amat C, Bertrand B, Vigouroux M, Buonviso N.** Piriform cortex
800 functional heterogeneity revealed by cellular responses to odours. *Eur J Neurosci* 17: 2457–
801 2461, 2003.
- 802 47. **Litaudon P, Garcia S, Buonviso N.** Strong coupling between pyramidal cell activity
803 and network oscillations in the olfactory cortex. *Neuroscience* 156: 781–787, 2008.
- 804 48. **Miura K, Mainen ZF, Uchida N.** Odor representations in olfactory cortex:
805 distributed rate coding and decorrelated population activity. *Neuron* 74: 1087–1098, 2012.
- 806 49. **Carlson KS, Dillione MR, Wesson DW.** Odor-and state-dependent olfactory tubercle
807 local field potential dynamics in awake rats. *J Neurophysiol* 111: 2109–2123, 2014.
- 808 50. **Jung F, Witte V, Yanovsky Y, Klumpp M, Brankack J, Tort AB, Dr. Draguhn A.**
809 Differential modulation of parietal cortex activity by respiration and β -oscillations. .
- 810 51. **Roux SG, Garcia S, Bertrand B, Cenier T, Vigouroux M, Buonviso N, Litaudon**
811 **P.** Respiratory cycle as time basis: An improved method for averaging olfactory neural
812 events. *J Neurosci Methods* 152: 173–178, 2006. doi: 10.1016/j.jneumeth.2005.09.004.
- 813 52. **Pager J.** Respiration and olfactory bulb unit activity in the unrestrained rat: statements
814 and reappraisals. *Behav Brain Res* 16: 81–94, 1985.
- 815 53. **Kay LM, Laurent G.** Odor-and context-dependent modulation of mitral cell activity
816 in behaving rats. *Nat Neurosci* 2: 1003–1009, 1999.
- 817 54. **Sheriff A, Pandolfi G, Nguyen VS, Kay LM.** Long-range respiratory and theta
818 oscillation networks depend on spatial sensory context. *J Neurosci* 41: 9957–9970, 2021.
- 819 55. **Juventin M, Ghibaud V, Granget J, Amat C, Courtiole E, Buonviso N.**
820 Respiratory influence on brain dynamics: the preponderant role of the nasal pathway and deep
821 slow regime. *Pflug Arch - Eur J Physiol* 475: 23–35, 2023. doi: 10.1007/s00424-022-02722-
822 7.

- 823 56. **Kleinfeld D, Moore JD, Wang F, Deschênes M.** The brainstem oscillator for
824 whisking and the case for breathing as the master clock for orofacial motor actions. In: *Cold*
825 *Spring Harbor symposia on quantitative biology*. Cold Spring Harbor Laboratory Press, 2014,
826 p. 29–39.
- 827 57. **Ranade S, Hangya B, Kepecs A.** Multiple Modes of Phase Locking between Sniffing
828 and Whisking during Active Exploration. *J Neurosci* 33: 8250–8256, 2013. doi:
829 10.1523/JNEUROSCI.3874-12.2013.
- 830 58. **Ye B, Huang L, Gao Z, Chen P, Ni H, Guan S, Zhu Y, Wang J-H.** The functional
831 upregulation of piriform cortex is associated with cross-modal plasticity in loss of whisker
832 tactile inputs. .
- 833 59. **Bagur S, Lefort JM, Lacroix MM, de Lavilléon G, Herry C, Chouvaeff M,**
834 **Billand C, Geoffroy H, Benchenane K.** Breathing-driven prefrontal oscillations regulate
835 maintenance of conditioned-fear evoked freezing independently of initiation. *Nat Commun*
836 12: 1–15, 2021.
- 837 60. **Dupin M, Garcia S, Boulanger-Bertolus J, Buonviso N, Mouly A-M.** New insights
838 from 22-kHz ultrasonic vocalizations to characterize fear responses: relationship with
839 respiration and brain oscillatory dynamics. *Eneuro* 6, 2019.
- 840 61. **MacIver MB, Kendig JJ.** Anesthetic effects on resting membrane potential are
841 voltage-dependent and agent-specific. *Anesthesiology* 74: 83–88, 1991.
- 842 62. **Kajikawa Y, Schroeder CE.** How local is the local field potential? *Neuron* 72: 847–
843 858, 2011.
- 844 63. **Parabucki A, Lampl I.** Volume conduction coupling of whisker-evoked cortical LFP
845 in the mouse olfactory bulb. *Cell Rep* 21: 919–925, 2017.
- 846 64. **Zhou G, Olofsson JK, Koubeissi MZ, Menelaou G, Rosenow J, Schuele SU, Xu P,**
847 **Voss JL, Lane G, Zelano C.** Human hippocampal connectivity is stronger in olfaction than
848 other sensory systems. *Prog Neurobiol* 201: 102027, 2021.
- 849 65. **Miles R.** Synaptic excitation of inhibitory cells by single CA3 hippocampal pyramidal
850 cells of the guinea-pig in vitro. *J Physiol* 428: 61–77, 1990.
- 851 66. **Huerta PT, Lisman JE.** Bidirectional synaptic plasticity induced by a single burst
852 during cholinergic theta oscillation in CA1 in vitro. *Neuron* 15: 1053–1063, 1995.
- 853 67. **Sherman SM.** Tonic and burst firing: dual modes of thalamocortical relay. *Trends*
854 *Neurosci* 24: 122–126, 2001.

855

856

857 FIGURE LEGENDS:

858 **Figure 1 Description of the RRo detection method.** Common color code: real data
859 measurements are in red, median signals are in green, surrogate measurements are in light
860 blue. **A)** Schematic description of the processing chain. **B)** Detailed description of the
861 processing chain. **B.1)** Preprocessed membrane potential (APs removal and deformation to
862 respiratory phase) along 7 respiratory cycles. **B.2)** Encoding of the 4 windows associated to
863 the 7 previous cycles (B.1). The encoding of the window 118 is detailed in the panel C.
864 Windows 118 to 120 are found to be related to respiration (in red), while the window 121 is
865 not (blue). The median signals associated to each window are shown in green. **B.3)** Encoding
866 of the similarity associated with the 4 windows presented above (B.2). The encoding of the
867 cycle 118 as part of the windows 118 is detailed in panel D. Individual cycles similar to the
868 window they belong to are shown in black. Dissimilar cycles are in gray. Cycles that are not
869 tested (because the associated window is not respiration-related) are in light gray. **B.4)** Cycle-
870 by-cycle encoding. The score of a cycle corresponds to the number of times this cycle is
871 found similar to the waveform of a respiration-related window. For example, the cycle 121 is
872 found similar to 3 respiration-related window waveforms and thus reached the score of 3
873 (B.3). Cycles with score of 3 or 4 are classified as RRo. Cycles with score of 2 are classified
874 as undetermined. Cycles with score of 0 or 1 are classified as no RRo. **C)** Detailed detection
875 on local windows. **C.1)** Construction of surrogates. The left-hand column corresponds to
876 actual cycles. The right-hand column corresponds to surrogates. Surrogates are constructed in
877 two steps. First, the cycles are randomly paired (here cycle 118 with 119, and 121 with 120).
878 Then, the cycles are phase-shifted, so that they are two by two in phase opposition. **C.2)** The
879 median signal of the 4 actual cycle, on the left, or of the 4 surrogates, on the right. The red,
880 respectively the light-blue, line indicates the amplitude of the actual signal, respectively the
881 surrogate **C3.)** Distribution of the surrogate amplitudes. Blue bars represent the distribution of
882 surrogate amplitudes. Black arrow highlights the p-values (95th percentiles). Red arrow

883 highlights the amplitude of actual data (C.2 left). **D)** Detailed verification of the similarity.
884 **D.1)** To assess the similarity between individual cycles (gray) and the local window
885 oscillation (green), we compute the scalar product of these z-scored signals. **D.2.)** Distribution
886 of the surrogate scalar products. Blue bars represent the distribution of surrogate scalar
887 products. The black arrow highlights the p-values (95th percentiles). The red arrow highlights
888 the scalar product of actual data (D.1 left).

889

890 **Figure 2 : Intracellular activity of widespread brain areas correlates with respiratory**
891 **rhythm. A)** Raw traces of intracellular activity from mPFC, S1, V1 and HPC (respectively
892 subpanels 1, 2, 3, 4). Top trace: intracellular raw trace in black, overlaid with a filtered (3 Hz
893 low pass) respiratory signal, in gray, which has been time-shifted to highlight MP RRo. Mid
894 trace: Respiratory signal. Bottom: Respiratory phase, with gray bars representing inspiration
895 phases, white bars representing expiration phases. Time scale bar = 1s, amplitude scale bar =
896 10 mV. E = expiration, I = inspiration. AP have been truncated. **B)** MP (black) and respiratory
897 signal (gray) power density spectrum over the 4s of signal presented in A. Arrow heads
898 highlight 1st harmonic of the spectrum. **C)** Respiration-triggered AP firing histogram (top) and
899 respiration triggered MP (bottom). Data of respiration-triggered figures are extracted from
900 several cycles during which MP is identified as correlated with respiration. Cycles and AP
901 numbers are indicated on the right top. To visually reinforce oscillatory behavior, plots have
902 been duplicated. Respiratory phase is reminded in between the two respiration-triggered
903 figures. Bottom: respiration triggered MP where black line corresponds to the median, shaded
904 areas delineate the 25th and 75th percentiles.

905

906 **Figure 3 : A significant number of cells in each structure exhibit MP RRo, yet individual**
907 **episodes are short.** Only recordings at basal MP are considered. **A)** Distribution of MP en-
908 coding by cell in mPFC, S1, V1 and HPC (respectively subpanels 1, 2, 3, 4). Black, gray and
909 white, bars correspond to the probability of RRo cycles, undetermined cycles and no RRo
910 cycles respectively. The total number of respiratory cycles recorded in each cell is indicated at
911 the top of the corresponding bar. Black crosses indicated cells which are not considered as
912 modulated by respiration despite showing some MP RRo (see methods) **B)** Cumulative distri-
913 bution of RRo percentages of each structure. Blue, orange and pink curves correspond to
914 mPFC, S1 and HPC respectively. **C)** MP RRo probabilities across structures. Gray dots repre-
915 sent MP RRo probability for individual cells, blue squares represent the mean by structure and
916 black lines represent SEM. Only respiration-modulated cells are considered. No significant
917 differences are observed (Kruskal-Wallis test, $p = 0.53$, $H(2) = 1.24$, $N_{\text{mPFC}} = 21$, $N_{\text{S1}} = 7$,
918 $N_{\text{HPC}} = 8$). **D)** MP RRo episodes duration across structures. Gray dots represent mean MP
919 RRo episode duration of individual cells, blue squares represent the mean by structure and
920 black lines represent SEM. The same cell pool as in B is used. No significant differences are
921 observed (Kruskal-Wallis test, $p = 0.58$, $H(2) = 1.07$, $N_{\text{mPFC}} = 21$, $N_{\text{S1}} = 7$, $N_{\text{HPC}} = 8$).
922

923 **Figure 4 : Phase and amplitude characterization of MP RRo.** Only recordings at basal MP
924 and respiration-modulated cells are considered. **A)** Amplitude of MP RRo by structures. Gray
925 dots represent MP RRo amplitude of individual cells, dark gray squares represent the mean by
926 structure and black lines represents SEM. Significant differences are observed (Kruskal-
927 Wallis test, $p = 0.01$, $H(2) = 9.1$, $N_{\text{mPFC}} = 21$, $N_{\text{S1}} = 7$, $N_{\text{HPC}} = 8$). *Post hoc* two-sided Mann-
928 Whitney U test reveals amplitude of MP RRo is higher in S1 cells than in mPFC ($p = 0.002$)
929 and HPC ($p = 0.02$). **B)** Preferred respiratory phase MP RRo by structures. Preferred phases

930 are presented on a trigonometric circle, phase progression is anticlockwise. Shaded and white
931 areas, correspond respectively to inspirations and expirations.

932

933 **Figure 5: MP RRo probability increases when RRo are concomitantly present in LFP.**

934 Only recordings at basal MP are considered. In addition, for B and C, only recording with
935 respiration-modulated LFP are considered. While for D and E, only recordings with both
936 respiration-modulated LFP and cells are considered. **A)** MP (Top), respiratory signal (mid)
937 and LFP (bottom) raw traces of the same cell when LFP and MP do not display RRo (A1) and
938 two minutes later when LFP and MP display RRo (A2). On A2 panel, light gray traces show
939 filtered and time-shifted respiratory signal superimposed on MP and LFP traces. Time scale
940 bar: 1s. **B)** LFP RRo probability by structures. Gray points represent LFP RRo probability,
941 dark gray squares represent the mean by structure and black lines represent SEM. Significant
942 differences are observed (Kruskal-Wallis test, $p = 0.03$, $H(2) = 6.96$, $N_{\text{mPFC}} = 21$, $N_{\text{S1}} = 5$,
943 $N_{\text{HPC}} = 4$). The LFP in mPFC presents more RRo than S1 (Mann-Whitney U test, $p = 0.01$).
944 **C)** LFP RRo episode duration by structures. Gray dots represent mean LFP RRo duration of
945 individual traces, dark gray squares represent the mean by structure and black lines represent
946 SEM. The same pool of traces as in B is used. Significant differences are observed (Kruskal-
947 Wallis test, $p = 7.10^{-3}$, $H(2) = 10.0$, $N_{\text{mPFC}} = 21$, $N_{\text{S1}} = 5$, $N_{\text{HPC}} = 4$). The mPFC presents
948 longer LFP RRo episodes than S1 (Mann-Whitney U test, $p = 0.005$) and HPC (Mann-
949 Whitney U test, $p = 0.02$). **D)** Conditional probability of MP RRo given LFP dynamics. The
950 columns “LFP RRo” represent the probability to observe MP RRo when LFP is also RRo. On
951 the other hand, the columns “other LFP” represent MP RRo probability when LFP is no RRo.
952 In the mPFC, the probability of MP RRo increases when LFP is RRo (Wilcoxon test, $p =$
953 0.02 , $Z = 44$, $n = 20$). S1 (Wilcoxon test, $p = 0.14$, $Z = 1$, $n = 4$) and HPC (Wilcoxon test, $p =$
954 0.07 , $Z = 0$, $n = 4$) presented the same tendency. **E)** Conditional probability of LFP RRo given

955 MP dynamics. The columns “MP RRo” represent the probability to observe LFP RRo when
956 MP is also RRo. On the other hand, the columns “other MP” represent the probability of LFP
957 RRo when MP is no RRo. In the mPFC, LFP RRo probability increases when MP is RRo
958 (Wilcoxon test, $p = 0.002$, $Z=21$, $n=20$). S1 (Wilcoxon test, $p = 0.14$, $Z= 1$, $n=4$) and HPC
959 (Wilcoxon test, $p = 0.07$, $Z= 0$, $n=4$) presented the same tendency.

960

961

962 TABLE:

963 **Table 1: mPFC and S1 show stronger coupling between MP oscillations and spiking**
964 **than hippocampus.**

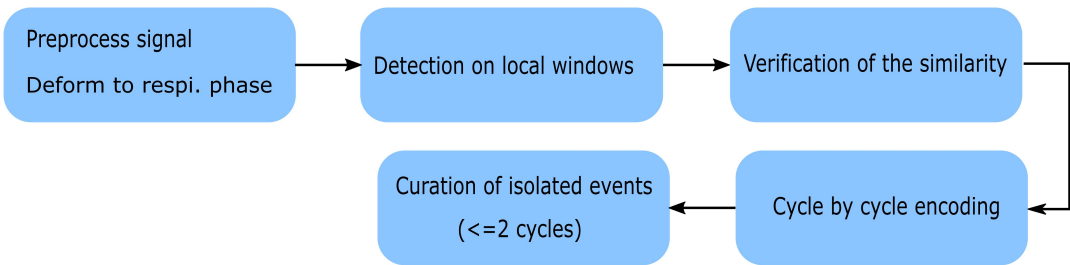
Area	MP activity		RR discharge	No RR discharge	Total cells
mPFC	RRo	N _{cell} %	4 80%	1 20%	5
	no RRo	N _{cell} %	6 28.6%	15 71.4%	21
S1	RRo	N _{cell} %	4 80%	1 20%	5
	no RRo	N _{cell} %	1 14.3%	6 85.7%	7
HPC	RRo	N _{cell} %	1 25%	3 75%	4
	no RRo	N _{cell} %	3 37.5%	5 62.5%	8

965

966 For each cell, spikes are sorted according to whether they occurred during MP RRo or no MP
967 RRo. Respiration-triggered discharge histograms are computed for both categories. The
968 histograms are then classified as either RR discharge or no RR discharge. Results are
969 presented for each area and each MP activity. The line “N_{cell}” represents the number of cells in
970 each discharge class. It should be noted that spike distributions with too few spikes were
971 not considered here or some cells never displayed MP RRo with spikes. Consequently, the
972 total number of cells in the lines ‘RRo’ are lower than the number of respiration-modulated
973 cells (showed in Figure 3) and the total number of cells in the lines ‘no RRo’ are lower than
974 the total number of cells. The line “%” represents the percentage of cells in each discharge
975 category relative to the total number of cells associated to an MP RRo category.

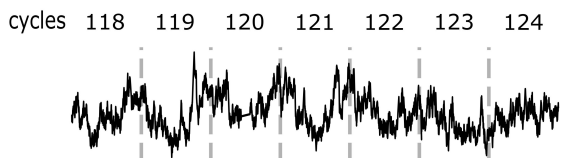
976

A Processing chain

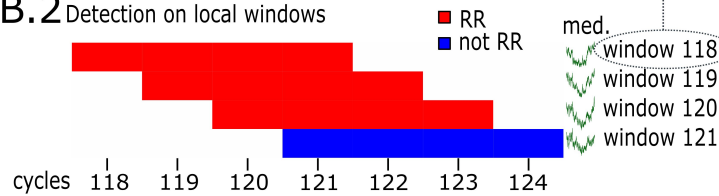


B Detailed processing chain

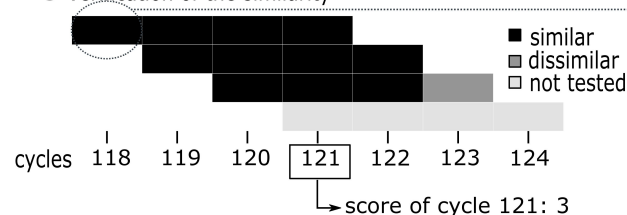
B.1 Preprocessing and deformation



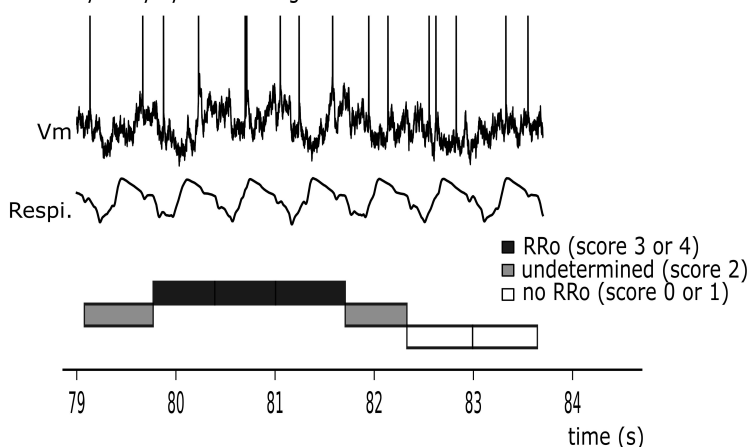
B.2 Detection on local windows



B.3 Verification of the similarity

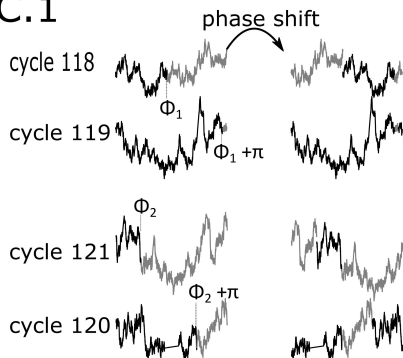


B.4 Cycle by cycle encoding



C Detection on local windows

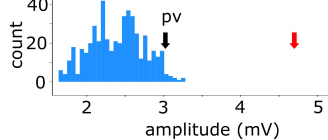
C.1



C.2

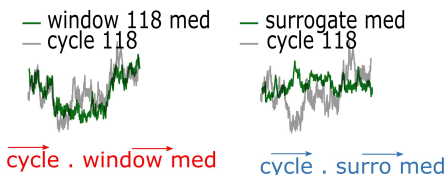


C.3

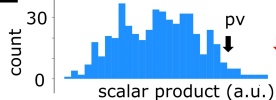


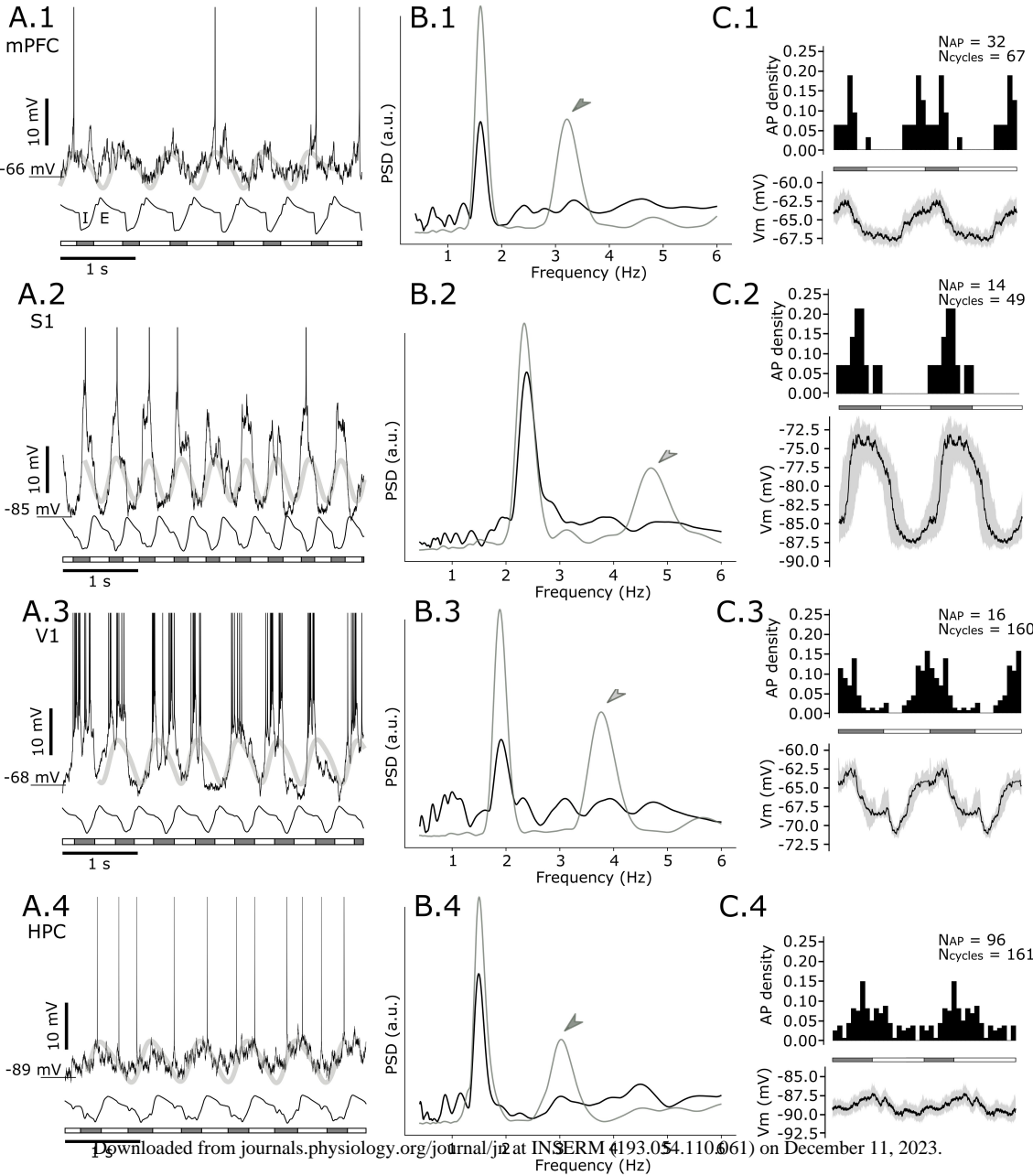
D Verification of the similarity

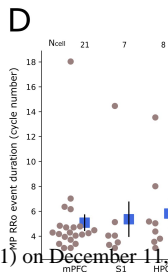
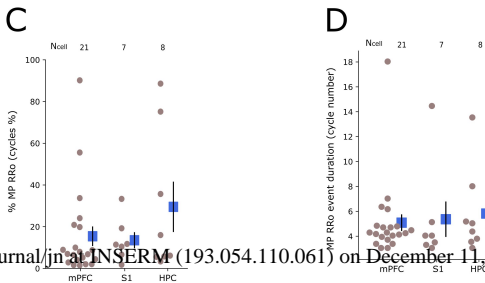
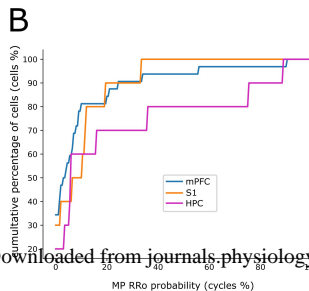
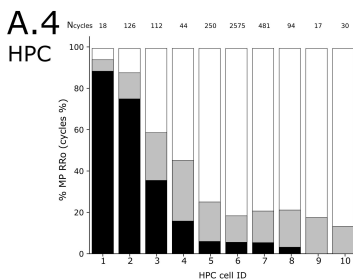
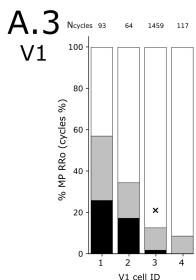
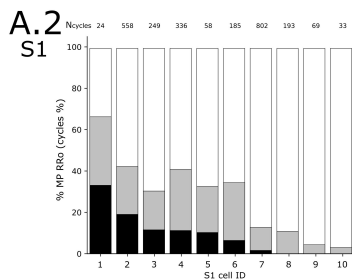
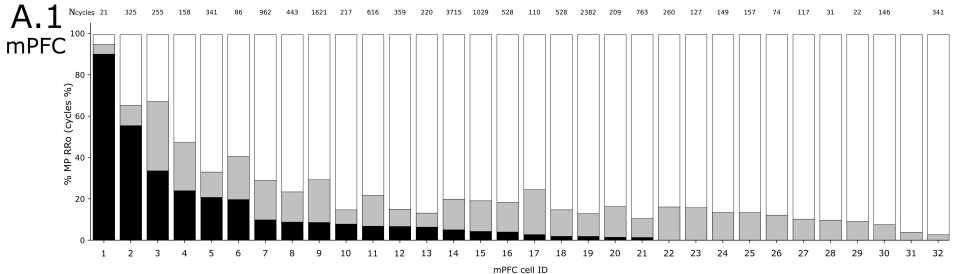
D.1

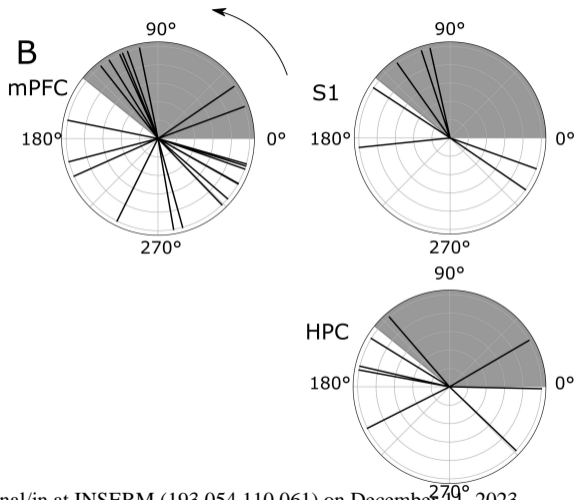
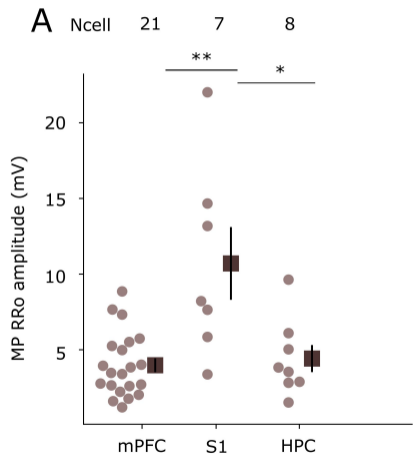


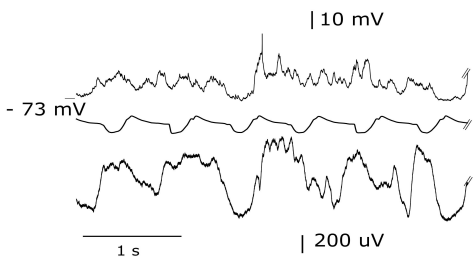
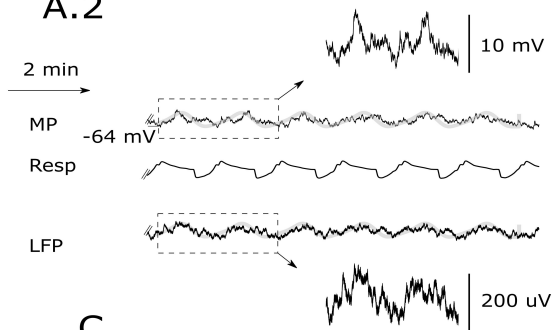
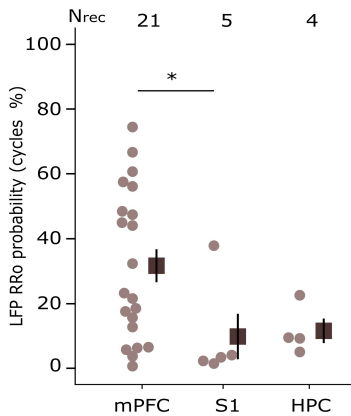
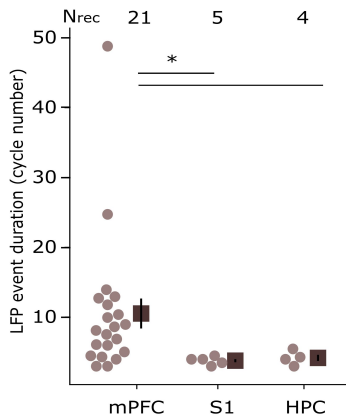
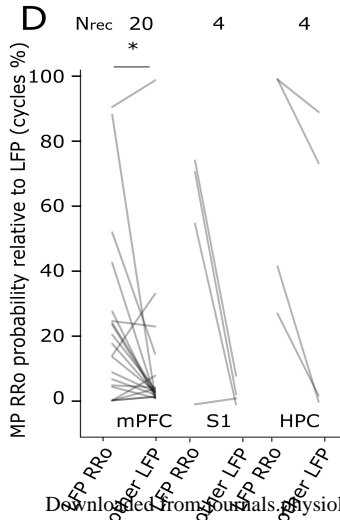
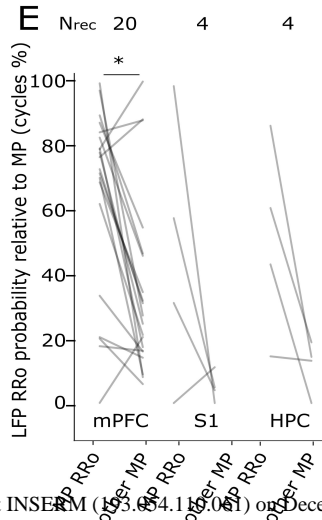
D.2









A.1**A.2****B****C****D****E**

1 **Table 1: mPFC and S1 show stronger coupling between MP oscillations and spiking**
 2 **than hippocampus.**

Area	MP activity		RR discharge	No RR discharge	Total cells
mPFC	RRo	N _{cell} %	4 80%	1 20%	5
	no RRo	N _{cell} %	6 28.6%	15 71.4%	21
S1	RRo	N _{cell} %	4 80%	1 20%	5
	no RRo	N _{cell} %	1 14.3%	6 85.7%	7
HPC	RRo	N _{cell} %	1 25%	3 75%	4
	no RRo	N _{cell} %	3 37.5%	5 62.5%	8

3

4 For each cell, spikes are sorted according to whether they occurred during MP RRo or no MP
 5 RRo. Respiration-triggered discharge histograms are computed for both categories. The
 6 histograms are then classified as either RR discharge or no RR discharge. Results are
 7 presented for each area and each MP activity. The line “N_{cell}” represents the number of cells in
 8 each discharge class. It should be noted that spike distributions with too few spikes were
 9 not considered here or some cells never displayed MP RRo with spikes. Consequently, the
 10 total number of cells in the lines ‘RRo’ are lower than the number of respiration-modulated
 11 cells (showed in **Error! Reference source not found.**) and the total number of cells in the
 12 lines ‘no RRo’ are lower than the total number of cells. The line “%” represents the
 13 percentage of cells in each discharge category relative to the total number of cells associated
 14 to an MP RRo category.

15

