

Cognitive and hippocampal effects of adult male mice exposure to environmentally relevant doses of phthalates

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- 3

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21 Abstract

22 We recently showed that chronic exposure of adult male mice to environmental doses of DEHP alone 23 or in a phthalate mixture altered blood brain barrier integrity and induced an inflammatory profile in 24 the hippocampus. Here, we investigate whether such exposure alters hippocampus-dependent 25 behavior and underlying cellular mechanisms. Adult C57BL/6J male mice were continuously 26 exposed orally to the vehicle or DEHP alone (5 or 50 µg/kg/d) or to DEHP (5 µg/kg/d) in a phthalate 27 mixture. In the Morris water maze, males showed reduced latencies across days to find the platform 28 in the cue and spatial reference memory tasks, regardless of their treatment group. In the probe test, 29 DEHP-50 exposed males displayed a higher latency to find the platform quadrant. In the temporal 30 order memory test, males exposed to DEHP alone or in a phthalate mixture were unable to 31 discriminate between the most recently and previously seen objects. They also displayed reduced 32 ability to show a preference for the new object in the novel object recognition test. These behavioral alterations were associated with a lowered dendritic spine density and protein levels of glutamate 33 34 receptors and postsynaptic markers, and inceased protein levels of the presynaptic synaptophysin in 35 the hippocampus. Metabolomic analysis of the hippocampus indicated changes in amino acid levels 36 including reduced tryptophan and L-kynurenine and elevated NAD+ levels, respectively, a precursor, 37 intermediate and endproduct of the kynurenine pathway of tryptophan metabolism. Interestingly, the 38 protein amounts of the xenobiotic aryl hydrocarbon receptor, a target of this metabolic pathway, were 39 elevated in the CA1 area. These data indicate that chronic exposure of adult male mice to 40 environmental doses of DEHP alone or in a phthalate mixture impacted hippocampal function and 41 structure, associated with modifications in amino acid metabolites with a potential involvement of 42 the kynurenine pathway of tryptophan metabolism.

43

44 Keywords

45 Phthalates; Plasticizers; Nervous system; Behavior; Hippocampus; Learning and memory; Endocrine
46 Disruptors.

48 Introduction

49 Plasticizers such as phthalates are widely spread in the environment. Di(2-ethylhexyl)phthalate (DEHP), the most abundant substance of this family, was identified as an endocrine disruptor for both 50 51 human health and environment due to its ability to reduce fetal testosterone production (European 52 Chemical Agency (ECHA), 2014; European Food Safety Agency (EFSA), 2005; EFSA et al., 2019). 53 Other phthalates such as benzyl butyl phthalate (BBP), dibutyl phthalate (DBP), diisobutyl phthalate 54 (DIBP) also detected in the environment were also recognized as substances of very high concern 55 due to their reprotoxicity and endocrine disrupting activity (ECHA site; EFSA et al., 2019). We have previously shown that exposure of adult male mice to DEHP at environmental doses reduced the 56 57 expression of male sexual behavior possibly through an endocrine mode of action involving downregulation of the androgen receptor (AR), without any effects on the estrogen receptor α (ER α), in 58 59 the neural circuitry underlying this behavior (Dombret et al., 2017). In the male nervous system, 60 testosterone can act directly through AR activation or be metabolized into neural estradiol, which then 61 stimulates ERs. These receptors are expressed in several brain areas including the hippocampus and cortex (Raskin et al., 2009; Simerly et al., 1990). Several studies using pharmacological or genetic 62 63 tools that impacted either testosterone levels or AR and ER activation or expression showed that these 64 signaling pathways modulate spatial memory and temporal processing of objects through molecular 65 and cellular mechanisms involved in synaptic plasticity in male rodents (Liu et al., 2008; Packard et 66 al., 1996; Picot et al., 2016). These behaviors could thus be influenced by exposure to endocrine 67 disrupting compounds such as phthalates, which can impact either gonadal production of testosterone 68 and/or their neural signaling pathways.

69

The effects of phthalates on learning and memory and related hippocampal processes have been widely studied in particular for the developmental period of exposure (Barakat et al., 2018; Holahan et al., 2018; Kougias et al., 2018; Lv et al., 2022; Safarpour et al., 2021; Sun et al., 2014). Whether adult exposure to these substances also triggers adverse effects was less addressed. Previous studies

74 analyzed the effects of acute or sub-chronic exposure to dibutyl phthalate (DBP) or diisononyl phthalate (DINP) at doses ranging from 0.2 to 250 mg/kg/d on learning and memory in young adult 75 76 mice (Ding et al., 2017; Farzanehfar et al., 2016; Peng, 2015; Song et al., 2021). Exposure for 9 or 77 14 days to DINP at 150 or 200 mg/kg/d resulted in increased latency to find the platform during the 78 training and test phases and reduced the time spent in the target quadrant in the retention test 79 performed two days later in the Morris water maze (Peng, 2015). No effects were observed at lower 80 DINP doses (0.2, 1.5 or 2 mg/kg/d). Comparable data were reported for an exposure period of 28 81 days to DINP at 200 mg/kg/d (Song et al., 2021). Using the same task, DBP exposure for 5 weeks 82 was shown to induce a higher escape latency at the last day of the training phase and reduce time 83 spent in the target quadrant and swimming speed in the retention task (Ding et al., 2017). These 84 effects were observed only for the group exposed to DBP at 250 mg/kg/d but not at lower doses (10 85 and 50 mg/kg/d). Male mice exposed for 14 days to DBP at 6.25 to 200 mg/kg/d exhibited a lower 86 latency in the passive avoidance test and reduced time spent in the open arms of the elevated plus 87 maze, but only at the doses of 25 and 200 mg/kg/d (Farzanehfar et al., 2016). It thus appears from 88 these studies that acute/sub-chronic exposure to phthalates induced effects on learning and memory 89 at high doses (25 mg/kg/d to 250 mg/kg/d), which are not relevant for the environmental exposure. We have recently shown that chronic exposure of adult male mice to environmentally relevant doses 90 91 of DEHP alone or in a phthalate mixture increased blood brain barrier permeability and affected 92 endothelial accessory tight junction proteins in the hippocampal CA1 and CA3 areas (Ahmadpour et 93 al., 2021). This was associated with an inflammatory profile including microglial activation and 94 induced cyclooxygenase-2 expression. Whether these cellular modifications triggered by low doses 95 of DEHP aone or in phthalate mixture are accompanied with impaired hippocampus-dependent 96 behaviors remains to be investigated.

97

98 The present study was therefore undertaken to investigate the effects of chronic exposure to DEHP 99 alone or in a phthalate mixture on learning and memory in male mice. Adult C57BL/6J male mice 100 were exposed orally to the vehicle (controls), DEHP alone at 5 or 50 µg/kg/d, or to a phthalate mixture 101 as previously described (Adam et al., 2021; Ahmadpour et al., 2021). Exposure started eight weeks 102 before the begining of behavioral analyses and treatment was maintained throughout the period of 103 analyses. Males were assessed for spatial memory, temporal order memory and novel object 104 recognition. As the execution of these behaviors can be altered by the anxiety-state level of animals, 105 anxiety-related behavior was also assessed. Analyses of dendritic spine density, pre- and post-106 synaptic components, synapse ultrastructure and metabolome were conducted in the hippocampus. 107 General physiological parameters such as body and androgen-sensitive organ weights, and 108 hippocampal protein amount levels of AR, ERα and the xenobiotic aryl hydrocarbon-receptor (AhR) 109 were also determined.

111 Materials and Methods

112 Animals and treatments

113 The study was conducted according to the European legal requirements (Decree 2010/63/UE) and 114 received the approval of the «Charles Darwin» Ethical committee (project number 01490-01). Male 115 C57BL/6J mice (Janvier Labs) were housed at 22°C and 12:12 h light-dark cycle (the dark phase was 116 scheduled from 1:30 P.M. to 1:30 A.M.) and treated as detailed in Supplemental Material and Methods. Briefly, male mice of eight-week-old were fed with chow containing the vehicle (control 117 118 group), DEHP (CAS 117-81-7) at the tolerable daily intake dose of 50 µg/kg/d (EFSA, 2005, 2019), 119 DEHP at the dose of 5 µg/kg/d in the range of the estimated environmental exposure, or to a phthalate 120 mixture containing DEHP at 5 µg/kg/d, DBP (CAS 8474-2) at 0.5 µg/kg/d, BBP (CAS 85-68-7) at 0.5 µg/kg/d, DIBP (CAS 84-69-5) at 0.5 µg/kg/d and di-ethylphthalate (DEP; CAS 84-66-2) at 0.25 121 µg/kg/d. The phthalate doses were weekely adjusted to male body weights. The exposure started eight 122 123 weeks before the beginning of analyses and was maintained until sacrifice as described in Figure S1. 124 Two cohorts of male mice, each including four treated groups, were used in the behavioral tests 125 (Figure S1). At the end of the tests, animals were euthanized using pentobarbital injection (120 126 mg/kg). The brains were collected for analyses by Golgi-cox staining, Western blot and transmission 127 electron microscopy. A third cohort was used for metabolomic analysis. The collected seminal vesicles and testes from these cohorts were weighed, and the results were expressed as percentage of 128 129 body weight.

130

131 Behavioral tests

Male mice were subjected to the behavioral tests described below and in Figure S1. Tests, which were
conducted during the light phase from 6:00 to 11:00 A.M, were recorded and the videos were analysed
as previously described (Dombret et al., 2020; Picot et al., 2016).

135

136 Morris Water Maze

The tasks were performed in a large circular pool (2.0 m diameter, 0.5 m height) filled with water, 137 138 which was maintained at a temperature of $25^{\circ}C \pm 1^{\circ}C$ and made opaque with 400 ml of latex liquid. 139 After each trial, the animals were dried off and returned to cages placed on heating pads. The test 140 consisted of two training phases (cue task and spatial reference memory) before the probe test. In the 141 cue task, mice were trained to locate a cue placed on a visible platform (13 cm diameter) on four trials per day for three days. Each trial lasted 90 sec, with an inter-trial interval of around 20 to 25 min. The 142 143 platform was located in different quadrants of the pool and the mice were released at various points 144 around the swimming pool. The location of the quadrant and point of release were counterbalanced 145 every day. The animals were placed on the platform for 30 sec if they did not find it within the 90 sec 146 trial.

In the spatial reference memory task, mice were trained to locate a hidden platform, with visible spatial landmarks, on four trials per day for five days. Each trial lasted 90 sec, with an inter-trial interval of around 10 min. The platform was located in one quadrant of the pool and the mice were released at various points around the swimming pool. The animal was placed on the platform for 30 sec if it did not find it within the 90 sec trial. Ten minutes after the last trial, the platform was removed; the probe test lasted for 60 seconds. Measures included the latency to find the platform and swimming velocity.

154

155 *Object recognition tasks*

The temporal order task was conducted in a round arena (45 cm diameter) filled with sawdust as previously described (Picot et al., 2016). During the habituation phase, the animals were put for two days in the arena in the absence of objects (one 30 min session followed by two daily 10 min sessions) and then exposed for one day to objects in place in the arena to decrease neophobia. During the familiarization phase of three sessions, the mice were allowed to explore two objects (a–a, b–b and c–c) for at least 15 seconds per object, with an interval of 3 min between the sessions. Ten minutes later, the males were subjected to a 5-min testing phase in the arena, where they were placed in the presence of the first (a) and last (c) object seen. The discrimination index (DI) between objects was defined as the difference in the time (t) spent exploring the first (tFi) and last objects (tL) divided by the total time spent exploring the two objects [DI = (tFi - tL)/(tFi + tL)].

In the novel object recognition task, the habituation and familiarization phases were performed as described for temporal order processing. In the testing phase, the animals were exposed to a familiar object (a) they had seen before and a new object (d). The discrimination index between objects was defined as the difference in the time (t) spent exploring the novel (tN) and the familiar objects (tFa) divided by the total time spent exploring the objects [DI = (tN - tFa)/(tN + tFa)].

171

172 Analysis of anxiety-related behavior

The anxiety state level was measured in the elevated zero maze as previously described (Dombret et al., 2020; Trouillet et al., 2022). Male mice were first placed in one of the closed arms of the maze. The test ended after 9 min of free exploration of the maze. The entry in the open arm was taken into account when all four paws of the mouse were inside this area. The analyses included the number of entries and the time spent in the open arms.

178

179 *Golgi-cox staining*

180 The brains were fixed for eight days in Golgi-cox solution (NovaUltra Golgi-cox Stain kit, Interchim, 181 Montluçon, France) as previously described (Jean et al., 2017), and then cut into 30 µm-thick coronal 182 sections with a vibratome (Leica Microsystems VTC1000S, Wetzlar, Germany). Sections including 183 the hippocampus, identified using neuroanatomical landmarks of the Mouse Brain Atlas (Paxinos and 184 Franklin, 2001), were immersed in post-impregnation solution for 10 min, washed in phosphate buffer 185 saline and mounted with Mowiol. Observations were made using the Zeiss Axiovert 200M 186 microscope equipped with an Axiocam MRm camera. For each animal, 10 neurons homogeneously 187 distributed throughout the CA1 (Bregma 2.18, plate 49) or CA2/3 (Bregma 2.30, plate 50) areas were 188 analyzed. For each neuron, the analyses were performed on a dendritic length of 50 µm localized at 189 30 µm from the start of the dendrite. Thin, stubby and mushroom subtypes of dendritic spines were

190 determined by applying criteria described by Harris et al. (1992).

191

192 Western blotting

193 Protein extracts from the CA1 or CA2/3 areas were prepared as previously described for AR and ERa 194 immunodetection (Capela et al., 2018; Picot et al., 2016) and pre- and post-synaptic proteins (Jean et 195 al., 2017). Protein samples (20 to 30 µg) were separated on 4-12% polyacrylamide gels (Invitrogen) 196 and transferred onto nitrocellulose membranes. Blots were incubated overnight at 4°C with 197 polyclonal anti-AR (1:400, Santa-Cruz Biotechnology), anti-ER α (1:400, Santa-Cruz 198 Biotechnology), anti-AhR (1:500, Santa-Cruz Biotechnology), anti-spinophilin (1:500, Millipore), 199 anti-GluN2A (1:500, Millipore) or monoclonal anti-glyceraldehyde 3-phosphate dehydrogenase 200 (GAPDH; 1:10000, Santa-Cruz Biotechnology), anti-postsynaptic density protein 95 (PSD95; 1:500, 201 Santa-Cruz Biotechnology), anti-GluN1 (1:500, Millipore), anti-GluN2B (1:500, Millipore) and anti-202 synaptophysin (1:500, Abcam). Incubation with secondary peroxidase-conjugated monoclonal or 203 polyclonal antibodies diluted at 1:5000 (Amersham Bioscience) was performed for 1 h at room 204 temperature. Signals were revealed with the Pico or Super Signal detection kits (Thermo Scientific) 205 and quantified using ImageJ software (NIH). The data were normalized with respect to GAPDH 206 protein amounts.

207

208 Transmission electron microscopy

Three male mice per treatment group were subjected to anesthesia using a lethal dose of pentobarbital (120 mg/kg, i.p.) before transcardiac perfusion with a 0.9% saline solution followed by a 2.5% glutaraldehyde solution in 0.1 M cacodylate buffer pH 7.4. Brains were removed and post-fixed in 2.5% glutaraldehyde/0.1 M cacodylate buffer overnight at 4°C. The brains were washed in cacodylate buffer and cut with a vibratome (Leica VT1000S, Leica Biosystems, Wetzlar, Germany) into four coronal sections including the hippocampus (100-µm thickness), which were post-fixed in 2%

215 osmium tetroxide diluted in cacodylate buffer (v/v) for 1 h at RT. After dehydration in an alcohol 216 ascending series, sections were embedded in epoxy resin (Epoxy-Embedding Kit, cat. Nº 45359, Sigma Aldrich, Switzerland). The polymerization step was performed at 60°C for 48 h between two 217 218 silicone-coated glass slides. Sections containing the hippocampal CA1 areas, selected using 219 neuroanatomical landmarks of the Mouse Brain Atlas (Paxinos and Franklin, 2001) indicated above, 220 ultrathin sections (70 nm) with an ultramicrotome (Reichert/Leica Ultracut S were cut in 221 ultramicrotome, Leica Microsystems, Bannockburn, IL, USA): four ultrathin sections were collected 222 per copper mesh grid with four grids per animal. Then, these ultrathin sections were contrasted with 223 2.5% uranyl salts and Reynold's lead citrate and observed under a transmission electron microscope 224 (80–120 kV EM 912 Omega ZEISS) equipped with a digital camera (Veleta Olympus).

225

226 Targeted LC/MS metabolites analyses

The metabolomic analysis was conducted as described previously (de la Calle Arregui et al., 2021). Hippocampal samples were extracted in a solution containing 50% methanol, 30% acetonitrile (ACN) and 20% water, and the collected supernatants were stored at -80 °C. The LC/MS analysis was performed as detailed in Supplemental Materials and Methods. Data were acquired with Thermo Xcalibur software (Thermo), and raw data were analyzed using MetaboAnalyst 5.0 software. The peak area of each metabolite was normalized to the total peak areas of all metabolites. The top 40 heatmap corresponds to the 40 most significant metabolites modified between the treatment groups.

234

235 *Statistics*

Data were expressed as means ± S.E.M. and analyzed by GraphPad Prism 6 (GraphPad Software).
Normality tests were performed (Kolmogorov-Smirnov and Shapiro-Wilks tests). Two-way analysis
of variance (ANOVA) was used for the main effect of exposure on the latency to reach the platform
and swimming distance in the Cue and Spatial reference memory tasks of the Morris water maze.
Bonferroni's post hoc tests were used to determine group differences. Kruskal-Wallis test was used

241	to analyze the effect of exposure on the latency to reach the platform and swimming distance in the
242	Probe test of the Morris water maze. Dunn's post hoc tests were used to determine group differences.
243	One way ANOVA and Fisher's LSD post hoc test were used to analyze metabolomic results.
244	Metabolic fold changes between two groups were quantified by using Student t-test. One way
245	ANOVA was used to analyze the effect of exposure on the remaining data. Bonferroni's post hoc
246	tests were used to determine group differences. P values < 0.05 were considered to be significant.

248 **Results**

249 Behavioral effects of adult exposure to DEHP alone or in a phthalate mixture

250 Spatial learning and memory assessed in the Morris water maze

251 In the cue task with a visible platform (Figure 1A-B), two-way ANOVA shows an effect of time ($F_{(2)}$) 252 $_{88} = 112.8$, p < 0.0001) but not of treatment (F_(3,44) = 0.33, p = 0.803). Male mice exhibited improved 253 performance in finding the platform across days, regardless of their treatment group. Comparable 254 data were found in the spatial reference memory task with hidden platform and visible spatial 255 landmarks (Figure 1C-D). There was an effect of time ($F_{(4,176)} = 14.95$, p < 0.0001) but not of treatment 256 $(F_{(3,44)} = 1.60, p = 0.203)$, with males again being able to exhibit increased performance over the 257 training period without any significant difference between the treated groups. In these two tasks, the 258 distance travelled did not differ between the treatment groups (Figure S2A-B). Ten minutes after the 259 end of training, males were assessed for their ability to find the platform location (Figure 1E-F). There 260 was an effect of treatment (p = 0.002) on the latency to find the platform quadrant. Post hoc analyses 261 show a significantly higher latency for the DEHP-50 group (+371% versus the control group). 262 Although not significant, a tendency of increase was also observed for the latency of DEHP-5 and 263 Mix groups. The distance travelled was also comparable between the treatment groups (Figure S2C). 264

265 *Temporal order memory and novelty detection tasks*

266 In the temporal order task for visual objects (Figure 2A), the total time spent exploring the objects 267 did not differ between the treated groups (55.84 \pm 4.72 sec for the Veh group, 54.32 \pm 4.64 sec for the DEHP-5 group, 53.74 ± 4.83 sec for the DEHP-50 group, 59.68 ± 4.69 for the Mix group). Mice 268 269 from the vehicle group were more interested (p < 0.0001) in exploring the first object seen than the last one (Figure 2B). By contrast, males from DEHP-5, DEHP-50 and Mix-groups did not 270 271 discriminate between the two presented objects (p > 0.05). One way ANOVA revealed a significant 272 main effect of treatment (p = 0.0012), with a lower mean discrimination index for DEHP-5, DEHP-273 50 and Mix-groups (-94%, -87% and -75% versus the vehicle group, respectively).

274 In the novelty detection task (Figure 2C), the total time spent exploring the objects was comparable 275 between the treatment groups $(55.43 \pm 4.64 \text{ sec for the Veh group}, 58.85 \pm 3.69 \text{ sec for the DEHP-5}$ 276 group, 63.88 ± 5.39 sec for the DEHP-50 group, 63.57 ± 4.12 for the Mix group). Analysis of mean 277 discrimination index showed that males from the vehicle and Mix groups spent more time exploring the new object than the familiar one (p < 0.001 for the vehicle group and p < 0.01 for the Mix-group) 278 279 as shown in Figure 2D. One-way ANOVA indicated an effect of treatment (p = 0.0057), with a lower 280 discrimination index for DEHP-5, DEHP-50 and Mix-groups (-68%, -67% and -59% versus the 281 vehicle group, respectively).

282

283 Anxiety-related behavior

In the elevated O-maze, an effect of treatment was observed on the number of entries (p = 0.0358) and time spent (p < 0.05) into the open arms (Figure 2E-F). Reduced number of entries for the DEHP-5 and Mix groups (-24% and -25% versus the Veh group, respectively) and time spent in the open arms for the Mix group (-25% versus the Veh group) were observed, but a tendency to decrease was also seen for the other groups.

289

290 Dendritic spine density and protein amounts for pre- and post-synaptic markers in the 291 hippocampus of adult male mice exposed to DEHP alone or in a phthalate mixture

292 Behavioral processing and reorganization of circuitries during learning and memory relies on several 293 cellular processes involved in neuroplasticity. The formation and maturation of spines emanating 294 from neuronal dendrites and known to constitute excitatory postsynapses are considered to be 295 essential for neuroplasticity. We therefore assessed whether the behavioral alterations induced by 296 exposure of adult male mice to DEHP alone or in phthalate mixture were associated with 297 modifications in the density and morphology of dendritic spines in the hippocampus. For this purpose, 298 brains from males exposed to the vehicle or DEHP alone or in a phthalate mixture were processed in 299 Golgi-Cox staining and dendritic spines were analyzed and classified in stubby, thin and mushroom

300 types according to their level of maturation as previously described (Jean et al., 2017). Figure 3A-B 301 shows that there was an effect of treatment on the number of stubby (p = 0.0008), thin (p = 0.0014)302 and mushroom spines (p = 0.0003) in the CA1 area. The density was reduced in almost all treatment 303 groups compared to vehicle-treated males (-35% to -43% for stubby spines, -64% to -78% for thin 304 spines and -64% to -69% for mushroom spines). This resulted in an average reduction in the total 305 number of dendritic spines in males exposed to DEHP alone or in phthalate mixture (-46% to -55%) 306 compared to the vehicle group. These effects were not restricted to the CA1 area since reduced spine 307 dendritic density was also apparent in the CA2/3 region as shown in Figure S3A-B.

308 Quantification by Western blotting of proteins extracted from the CA1 area showed modifications in 309 the protein levels of postsynaptic markers. Figure 3C-F shows a significant effect of treatment on the 310 protein amounts of PSD-95 (p = 0.0007), spinophilin (p = 0.035), GluN1(p = 0.0006) and GluN2A 311 receptors (p = 0.0014). No significant treatment effect was found on the protein amount of GluN2B 312 (p = 0.25), although a tendency towards a decrease was also observed (Figure 3G). Posthoc analyses 313 show that the protein levels were reduced for PSD-95 in the DEHP-50 and Mix groups (-53% and -314 55%, respectively), for spinophilin in the DEHP-50 group (-50%), for GluN1 in the DEHP-50 and 315 Mix groups (-85% and -95%, respectively), and for GluN2A in the DEHP-5, DEHP-50 and Mix 316 groups (-61%, -62% and -45%, respectively) in comparison to the Veh group.

317 The protein levels for the presynaptic vesicle membrane protein synaptophysin were also quantified 318 in the CA1 area (Figure S4A). There was an effect of treatment (p = 0.04), with a higher amount in 319 the Mix group (+63% versus the vehicle group). We also conducted a qualitative analysis of ultrastructural morphology of synapses corresponding to dendritic spines in the hippocampal CA1 320 321 area. Qualitative random electron microscopy observations were thus performed with 33 to 42 images 322 for each experimental group including at least one synapse associated with dendrite. Representative 323 images in Figure S4B show that the synapses observed in the CA1 region of the hippocampus were clearly identifiable for the vehicle group with, on the one hand, a clean, homogeneous, compact and 324 electron-dense postsynaptic membrane and, on the other hand, the presence of vesicles with low dense 325

electron density content in the presynaptic element. This ultrastructural profile corresponds to that of
functional synapses. The observation of synapses in the CA1 area of DEHP-5 treated mice showed a
similar ultrastructure (Figure S4B). For mice treated with DEHP-50 alone or in a phthalate mixture,
vesicles were not identifiable, so making the distinction between the pre- and postsynaptic element
hazardous (Figure S4B). The ultrastructure of the mitochondria and myelinated axons attests to the
absence of artefacts linked to the chemical fixation of the brains or to the procedure for preparing
samples for electron microscopy (Figure S4B).

333

334 Metabolomic analysis of the hippocampus of male mice exposed to DEHP alone or in a 335 phthalate mixture

336 We have previously shown that adult exposure to DEHP triggered modifications in the levels of proteins involved in cellular metabolism in the medial preoptic area in male mice (Dombret et al., 337 338 2017). As metabolic activity is mandatory for the cellular processes involved in neural function and 339 behavior, we conducted a metabolomic analysis by LC/MS on the hippocampus of adult mice exposed to the vehicle, DEHP alone or in a phthalate mixture. The heatmap of the top 40 modified 340 341 metabolites is shown in Figure 4. Among them, ANOVA analysis shows that several metabolites 342 were significantly modified by exposure to DEHP alone or in a phthalate mixture (FDR p-adjusted 343 value < 0.05). Most of them are amino acids, which were down-regulated, and may have an important 344 role in neurotransmission. This included tyrosine (FDR p-adjusted value = 0.022), valine (FDR p-345 adjusted value = 0.022), proline (FDR p-adjusted value = 0.044) and tryptophan (FDR p-adjusted 346 value = 0.022) which are involved either in catecholamine, serotonin or glutamate metabolism 347 (Dalangin et al., 2020; Fernstrom & Fernstrom, 2007; Höglund et al., 2019; Jones et al., 2021; 348 Schwarcz & Stone, 2017). Tryptophan is particularly interesting as it is also involved in the synthesis 349 of neuroactive compounds via the kynurenine pathway. Tryptophan was down-regulated with a fold 350 change of 0.563 in the DEHP-50 group (p = 0.007) and 0.61 for the Mix group (p = 0.009), while 351 NAD, the endproduct of this metabolic pathway (O'Mahony et al., 2015; Ramos-Chávez et al., 2018),

was overproduced with a fold change of 1.325 (p = 0.003) for the DEHP-50 group (Table S1). Lkynurenine, an intermediate of this pathway, was also downregulated in the Mix group (FC = 0.275, p = 0.043; Table S1), suggesting that tryptophan was actively metabolized via the kynurenine pathway following phthalate exposure.

356

357 Protein amounts of AR, ERα and AhR in the hippocampus of adult male mice exposed to DEHP 358 alone or in a phthalate mixture

359 Our previous data showed that adult DEHP exposure did not impact the gonadotropic axis or circulating levels of testosterone, but rather impacted AR expression levels (Dombret et al., 2017). 360 361 Accordingly, analyses of body weight and relative weights of androgen-sensitive tissues such as the 362 testis and seminal vesicles did not show any effect of the treatment with DEHP alone or in a phthalate 363 mixture (Figure 5A-C). We thus quantified the protein amounts of the AR and ERa by Western 364 blotting in the CA1 area. Data illustrated in Figure 5D-E show an effect of treatment on AR protein 365 amounts (p = 0.041), but not on ER α (p = 0.425), with significantly reduced protein levels in the 366 DEHP-50 group. An effect of treatment (p = 0.014) was also found on the protein amount of the 367 xenobiotic receptor AhR (Figure 5F), a target of the kynurenine pathway of tryptophan metabolism 368 (Ramos-Chávez et al., 2018) and a negative regulator of AR expression (Arabnezhad et al., 2020; 369 Ghotbaddini et al., 2018). Significantly higher protein levels were observed for the DEHP-5 group (+78%, p < 0.01) compared to the vehicle group. Although not significant, a tendency to increase was 370 371 also seen for the other treated groups.

373 Discussion

The present study shows that chronic exposure of adult male mice to environmental doses of DEHP alone or in a phthalate mixture impaired short-term memory in spatial, temporal order and novelty tasks. These behavioral alterations were associated with reduced dendritic spine density and protein levels of postsynaptic markers in the hippocampus. Metabolomic modifications including changes in the levels of metabolites involved in tryptophan metabolism via the kynurenine pathway were also observed together with higher protein levels of the AhR, a xenobiotic receptor target of this metabolic pathway.

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382 Adult male mice chronically exposed to DEHP at 50 µg/kg/d exhibited altered spatial memory in the Morris Water Maze. A tendency to increase for the latency to find the platform quadrant was 383 384 also observed for DEHP-5 and Mix-treated groups, despite a comparable training phase between the 385 control and the three treated groups. Males exposed to DEHP alone at 5 or 50 µg/kg/d or in a phthalate 386 mixture displayed also altered temporal processing of visual objects as they were unable to discriminate between the first and last objects seen. This could be related to either reduced temporal 387 388 order processing and/or diminished capacity to discriminate between new and familiar objects since 389 these male groups exhibited a reduced performance in the novelty task. The higher anxiety state level 390 of male mice exposed to DEHP alone or in a phthalate mixture compared t the vehicle group could 391 only partially account for the observed alterations since a session to decrease neophobia towards 392 objects was included in the two object recognition tasks.

Further analyses of the hippocampus, which plays a key role in the processing of spatial and temporal order memory, showed a lower density of dendritic spines in the CA1 region of males exposed to DEHP alone or in a phthalate mixture compared to the vehicle group. The effect of exposure on these excitatory postsynapses was observed for all spine types ranging from the transient (thin) to the most mature and stable spine (mushroom). In agreement with this reduced dendritic spine density, analysis of postsynaptic proteins in the CA1 area showed lower levels for PSD-95 and spinophilin as well as

for glutamate receptors (GluN1, GluN2A) in males exposed to DEHP alone or in a phthalate mixture. Interestingly, a low dendritic spine density was also observed in the CA2/3 area, suggesting that these structural alterations were not restricted to the CA1 area. As male mice were exposed to DEHP alone or in a phthalate mixture during eight weeks before the beginning of behavioral tests, with exposure maintained during the whole period of analyses until sacrifice, we suggest that such exposure prevented structural modifications necessary for neuroplasticity processes related to dendritic spines to occur in the CA areas, thus resulting in a low neuronal connectivity and behavioral processing.

406 Qualitative ultrastructural analysis of synapses corresponding to dendritic spines in the hippocampal 407 CA1 area showed normal morphology, but the presynaptic vesicles appeared hardly identifiable in 408 DEHP-50 and Mix-exposed male mice. In addition, higher protein levels for the presynaptic synaptophysin were observed in the CA1 area of males exposed to DEHP alone or in a phthalate 409 410 mixture. While the exact role of synaptophysin in neurotransmission is still not clear, it was recently 411 suggested that the synaptophysin/gyrin family might negatively regulate excitocytosis (Raja et al., 412 2019). It thus remains to determine whether adult exposure to DEHP alone or in a phthalate mixture 413 at low doses affects also neurotransmission processes.

414

415 The metabolomic approach highlighted changes in several metabolites involved in 416 neurotransmitter synthesis or aminoacid metabolism following exposure to DEHP alone or in a 417 phthalate mixture. The reduced tryptophan and L-kynurenine levels associated with the elevated 418 levels of NAD are of particular interest, Tryptophan and L-kynurenine levels depend not only on 419 tryptophan abundance but also on the activity of the enzymes involved in this pathway. This could 420 explain the discrepancy observed between the concentrations of the metabolites and the endproduct, 421 NAD (Covarrubias et al., 2021). Hence, the reduced tryptophan and L-kynurenine suggests that 422 tryptophan concentration is limited and that both tryptophan and L-kynurenine are fed into NAD+ 423 synthesis.

424 Altogether, these observations suggest that exposure to DEHP alone or in a phthalate mixture 425 triggered modifications in the kynurenine pathway of tryptophan metabolism. First, exposure to high 426 doses of phthalates was previously reported to disturb the kynurenine pathway of tryptophan 427 metabolism in both male rats (Fukuwatari et al., 2002a; 2002b; 2004) and men (Nassan et al., 2019). Phthalates share, indeed, structural similarities with tryptophan metabolites, which allow binding and 428 429 inhibition of α -amino- β -carboxymuconate- ϵ -semialdehyde decarboxylase, an enzyme that prevents 430 the accumulation of kynurenine pathway metabolites (Singh et al., 2018). Second, disruption of the 431 kynurenine pathway is suggested to be involved, through its neuroactive metabolites, in glutamatergic toxicity and inflammatory processes via a mechanism involving activation of the xenobiotic receptor 432 433 AhR (Ramos-Chávez et al., 2018). The relevance of environmental doses of phthalates to induce such 434 metabolic modifications is supported by a recent study showing that urinary concentrations of several 435 phthalate metabolites were positively associated with kynurenine-derived neuroactive metabolites in 436 women (Nassan et al., 2020). Furthermore, we have recently shown that exposure of adult male mice 437 to environmental doses of DEHP alone or in a phthalate mixture, under similar experimental 438 conditions, induced microglial activation and increased BBB permeability in the hippocampus 439 (Ahmadpour et al., 2021). Based on all these observations, we suggest that chronic exposure to low 440 doses of phthalates may primarily induce several metabolic changes including but not limited to the 441 dysregulation of tryptophan metabolism, which may then trigger microglial activation and BBB 442 impairment, leading to reduced neuronal connectivity and neurotransmission and thus to behavioral 443 deficits.

444

The present data indicate that exposure to low environmental doses of phthalates impacted the hippocampus and related cognitive function. This contrasts with previous studies showing effects of phthalates at high but not at low doses (Ding et al., 2017; Peng, 2015; Song et al., 2021). The discrepancy with our data is probably due to the short duration of exposure used in these studies compared to the chronic continuous exposure used here. The curently described cognitive effects 450 following exposure of male mice to environmental doses of DEHP alone or in a phthalate mixture 451 can be relevant for humans. A recent human study indicates that exposure to phthalates is associated 452 with poor performance in immediate and delayed recall tests in the U.S. elderly (Weng et al., 2022). 453 The effects induced by exposure to DEHP alone or in a phthalate mixture are not restricted to the 454 hippocampus and its related cognitive function. Indeed, reduced sexual behavior in male mice 455 exposed to DEHP alone or in a phthalate mixture, under similar experimental conditions, was also 456 associated with alterations in metabolic activity and structural plasticity in the preoptic area (Ducroq et al., under revision). The underlying mode of action in this hypothalamic region seemed to involve 457 458 reduced AR protein amounts, associated with AhR up-regulation, a known negative regulator of AR 459 expression (Arabnezhad et al., 2020; Ghotbaddini et al., 2018) and receptor target of the kynurenine 460 pathway of tryptophan metabolism (Ramos-Chávez et al., 2018). By comparison, the data obtained 461 in the hippocampus show reduced AR protein levels but only for the DEHP-50 group, while AhR protein levels were significantly higher in the DEHP-5 group with a tendency to increase in DEHP-462 463 50 and Mix-groups. This suggests that AR down-regulation in the hippocampus possibly participated 464 to the behavioral effects observed in male mice exposed to DEHP alone or in a phthalate mixture, but 465 other mechanisms were probably also involved. In support of this hypothesis the fact that male mice 466 deleted for the neural AR exhibited impaired temporal order memory but displayed normal novel 467 object recognition (Picot et al., 2016), while male mice exposed to DEHP alone or in a phthalate 468 mixture exhibited deficits for both behaviors.

469

470 Conclusion

471 Chronic exposure of adult male mice to DEHP alone at doses equivalent to or below the TDI 472 dose or in a phthalate mixture disrupted spatial and temporal order memory. The comparable effects 473 of DEHP alone and in a phthalate mixture found for the majority of data suggest that DEHP drives 474 the effect in the mixture. Possible underlying mechanisms of these behavioral deficits could be the 475 metabolomic alteration observed in the hippocampus and including changes in the kynurenine 476 pathway of tryptophan metabolism. This could participate to induce microglial activation and BBB 477 impairment (Ahmadpour et al., 2021), and alter structural neuroplasticity (present study) in the 478 hippocampus. It is possible that exposure to phthalates altered other brain regions involved in 479 cognitive processes such as the cortex or related to other neural functions. We have recently described 480 reduced expression of sexual behavior and altered cellular and molecular processes in the 481 hypothalamus of male mice exposed in similar experimental conditions to equivalent doses of DEHP 482 and phthalate mixture (Dombret et al., 2017; Ahmadpour et al., 2021; Ahmadpour et al., 2022). This 483 highlights the vulnerability of the adult nervous system to continuous exposure to phthalates at doses 484 equivalent and below the established reference doses for other endpoints (EFSA, 2005; EFSA et al., 485 2019; US EPA, 1987).

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673 Figures

674 Figure 1. Effects of adult exposure to DEHP alone or in a phthalate mixture on spatial learning 675 and memory assessed in the Morris water maze. Adult male mice were exposed to the vehicle (Veh), DEHP at 5 (DEHP-5) or 50 µg/kg/d (DEHP-50 or to a phthalate mixture (Mix). A-B. Latency 676 677 of to reach the visible platform in the Cue task. Data are as means \pm S.E.M. for 11-12 males per treatment group; there was an effect of training ##p < 0.001. C-D. Latency to reach the hidden 678 679 platform whith spatial cues around the maze during the Spatial reference memory. The effect of 680 training is indicated (##p < 0.001). E-F. Latency to reach the platform quadrant with spatial cues around the maze during the Probe test. Effect of treatment (#p < 0.01) and post hoc comparisons (*p 681 682 < 0.001) are indicated.

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684 Figure 2. Effects of adult exposure to DEHP alone or in a phthalate mixture on temporal order 685 memory, novel object recognition and anxiety-related behavior. A-B. The testing sequence of 686 object presentation and testing phase of the temporal order task are indicated (A). The discrimination 687 index is shown as means \pm S.E.M. for 11-12 males per treatment group (**B**). Only the Veh group 688 displayed more interest in exploring the first object (a) seen, $^{+++}p < 0.001$. The effect of treatment 689 (#p < 0.01) and post hoc analyses (*p < 0.05, **p < 0.01, ***p < 0.001) versus the Veh group are 690 indicated. C-D. The testing sequence of object presentation and testing phase of the novelty task are 691 indicated (C). Only the Veh and Mix groups displayed more interest in exploring the new object (d), $^{+++}p < 0.001$, $^{++}p < 0.01$ (**D**). The effect of treatment (##p < 0.01) and post hoc analyses (*p < 0.05) 692 693 versus the Veh group are indicated. E-F. Total number of entries (E) and time spent in the open arms 694 (F) of the elevated O-maze. The effect of treatment (# p < 0.05) and post hoc comparisons (*p < 0.05) 695 are shown.

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Figure 3. Effects of adult male mice exposure to DEHP alone or in a phthalate mixture on neural structure in the CA1 area of the hippocampus. A. Representative images of dendritic spines in the

699 CA1 area of the hippocampus of male mice treated with the vehicle (Veh), DEHP at 5 (DEHP-5) or 700 50 (DEHP-50) µg/kg/d, or to a phthalate mixture (Mix). Thin (t), stubby (s) and mushroom (m) 701 dendritic spines type are indicated. Scale bar: 10 µm **B**. Quantification of total and subtype dendritic 702 spine (10 neurons per mouse). Data are presented as means \pm S.E.M. from 4 males per treatment 703 group. Treatment effects (##p < 0.001, #p < 0.01) and post hoc comparisons (***p < 0.001, **p < 704 0.01) are indicated. C-H. Quantification of protein amounts for PSD95 (C), spinophilin (D), GluN1 (E), GluN2A (F) and GluN2B (G) normalized to GAPDH in the CA1 area of the hippocampus. Data 705 706 are expressed as means \pm S.E.M. from 4-6 males per treatment group. Treatment effects (###p < 707 0.001, #p < 0.01, #p < 0.05) and post hoc comparisons (**p < 0.01, *p < 0.05) are indicated.

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Figure 4. Pan-metabolomic analysis in the hippocampus of males mice exposed to DEHP alone or in a mixture of phthalates. Data are means of 7-8 males per treatment group. Relative levels vary from high (red) to low (blue). ADP, adenosine diphosphate; CDP, cytosine diphosphate; CMP, cytosine monophosphate; GDP, guanosine diphosphate; GSSG, oxidized glutathione; NAD⁺, nicotinamide adenine dinucleotide; NADP⁺, nicotinamide adenine dinucleotide phosphate; UDP, uridine diphosphate; UMP, uridine monophosphate.

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716 Figure 5. Effects of adult male mice exposure to DEHP alone or in a phthalate mixture on 717 physiological parameters and the protein amounts of androgen (AR) and estrogen receptor 718 (ERα), and aryl hydrocarbon receptor (AhR) in the CA1 area. A. Growth curve for the total 719 duration of exposure for male mice exposed to the vehicle (Veh), DEHP at 5 or 50 µg/kg/day, or to a 720 phthalate mixture (Mix). The blue line indicates the duration of the behavioral analyses. Data are 721 expressed as means \pm S.E.M. of 23-24 males per treatment group. **B-C**. The weight of testes (**B**) and 722 seminal vesicles (C) are expressed as percentage of body weight (bw). D-E-F. Quantification of the 723 protein amounts for the AR (D), ERa (E), and AhR (F) normalized to GAPDH. Data are expressed

- as the means \pm S.E.M. of 4-6 males per treatment group. Treatment effect (#p < 0.05) and post hoc
- 725 analyses (*p < 0.05, **p < 0.01) are indicated.



Figure 1



Figure 2



Figure 3



Figure 4



Figure 5

Supplemental Materials and Methods

734 *Animals and treatments.*

Male mice used in this study were produced from the breeding of male and female C57BL/6J mice 735 736 (Janvier Labs). Animals were housed in a nest-enriched environment using polysulfone cages and bottles, at 22°C and 12:12 h light-dark cycle (the dark time began at 1:30 P.M. and ended at 1:30 737 A.M.), and ad libitum access to food (A03-10; Safe-diets) and water. Male offspring from 11 litters 738 739 for cohort 1, 10 litters for cohort 2 and 9 litters for cohort 3 were mixed at weaning. The exposure 740 started at the adult age of 8 weeks. All the analyses were conducted according to the European legal 741 requirements (Decree 2010/63/UE) and received the approval of the «Charles Darwin» Ethical 742 committee (project number 01490-01).

Oral exposure was performed through incorporation of phthalates (Sigma-Aldrich) into food as previously described (Adam et al., 2021; Adam et al., 2022; Ahmadpour et al., 2021; Ahmadpour et al., 2022; Dombret et al., 2017). Phthalates (Sigma-Aldrich, Saint-Quentin Fallavier, France) were dissolved in ethanol and water (1% and 40% of prepared food, respectively) and then incorporated into powdered food (A03–10; Safe-diets), which was then reconstituted into pellets.

748 Male mice of eight-week-old had ad libitum access to chow containing the vehicle (control group), 749 DEHP (CAS 117-81-7) at the tolerable daily intake dose of 50 µg/kg/d (EFSA, 2005, 2019), DEHP 750 at the dose of 5 µg/kg/d in the range of the estimated environmental exposure, or to a phthalate 751 mixture containing DEHP at 5 µg/kg/d, DBP (CAS 8474-2) at 0.5 µg/kg/d, BBP (CAS 85-68-7) at 0.5 µg/kg/d, DiBP (CAS 84-69-5) at 0.5 µg/kg/d and DEP (CAS 84-66-2) at 0.25 µg/kg/d. The 752 753 composition of the mixture was based on French and European studies reporting external co-exposure 754 to these substances (Martine et al., 2013) and presence of their metabolites in urinary samples (Anses 2015; Dewalque et al., 2014). The phthalate doses were weekly adjusted to male body weights, and 755 calculated for a daily food intake of 5 g per animal (Dombret et al., 2017). 756

757 The exposure started eight weeks before the beginning of analyses and was maintained until sacrifice.

758 Two cohorts of male mice, each including four treated groups, were used in the behavioral tests

(Figure S1). At the end of the tests, male mice were sacrificed and brains and tissues were collected
for analyses by Golgi-cox staining, Western blot and transmission electron microscopy. An additional
cohort was used for metabolomic analysis.

- The analyses were performed by blind observers using only the numbers attributed at weaning.
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764 Targeted LC/MS metabolites analyses

765 Animals were sacrified by cervical dislocation and the isolated hippocampus from collected brains 766 were frozen in liquid nitrogen. Metabolomic analysis was conducted as described previously (de la 767 Calle Arregui et al., 2021). The extraction solution (50% methanol, 30% acetonitrile (ACN) and 20% water) adjusted to the weight of the tissue (1 ml per 50 mg), was added to hippocampal samples, 768 769 which were then vortexed for 5 min at 4 °C and centrifuged at 16,000 g for 15 min at 4 °C. The 770 collected supernatants were stored at -80 °C. The LC/MS step was performed using QExactive Plus Orbitrap mass spectrometer equipped with an Ion Max source and a HESI II probe coupled to a 771 772 Dionex UltiMate 3000 UPLC system (Thermo). A standard calibration mixture was used for external 773 mass calibration every seven days. The samples (5 µl) were injected onto a ZIC-pHILIC column (150 774 mm \times 2.1 mm; i.d. 5 µm) with a guard column (20 mm \times 2.1 mm; i.d. 5 µm) (Millipore). Buffer A 775 (20 mM ammonium carbonate and 0.1% ammonium hydroxide (pH 9.2)) and buffer B (CAN) were 776 used. The chromatographic gradient was run at a flow rate of 0.200 μ l min⁻¹ as follows: 0-20 min, linear gradient from 80% to 20% of buffer B; 20–20.5 min, linear gradient from 20% to 80% of buffer 777 778 B; 20.5–28 min, 80% buffer B. The mass spectrometer was operated in full scan, polarity switching 779 mode with the spray voltage set to 2.5 kV and the heated capillary held at 320 °C. The sheath gas 780 flow was set to 20 units, the auxiliary gas flow to 5 units and the sweep gas flow to 0 units. Metabolites 781 were detected across a mass range of 75–1,000 m/z at a resolution of 35,000 (at 200 m/z) with the automatic gain control target at 10⁶ and the maximum injection time at 250 ms. Lock masses were 782 783 used to ensure mass accuracy below 5 ppm. Data were acquired with Thermo Xcalibur software 784 (Thermo). The peak areas of metabolites were determined using Thermo TraceFinder software

(Thermo), identified by the exact mass of each singly charged ion and by the known retention timeon the HPLC column.

The raw data were analyzed with MetaboAnalyst 5.0 software. The peak area of each metabolite was normalized for each sample to the total peak areas of all metabolites. The top 40 heatmap served to evaluate the 40 most significant metabolites changing between the treatment groups. Metabolites presenting equivalent abundance patterns were positioned close together.

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Figure S1. A-B. Male mice from the first (A) and second cohorts (B) were exposed since 8 weeks of
age. The duration of exposure is indicated in green, with males exposed for 8 weeks before the first
analyses and the exposure maintained until sacrifice. AR: androgen receptor; ER: estrogen receptor;
ARB: Anxiety-related behavior; MWM: Morris Water Maze; TOM: Temporal Order Memory; NOR:
Novel Object Recognition; WB: Western blotting.



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Figure S2. Effects of adult exposure to DEHP alone or in a phthalate mixture on spatial learning and memory in the Morris water maze. Male mice were exposed to the vehicle (Veh), DEHP at 5 or 50 μ g/kg/d, or to a phthalate mixture (Mix). A-C. Distance to reach the platform in the Cue (A) and spatial reference memory tasks (B), and at the probe test (C). Distance (meters) is shown as means ± S.E.M. for 11-12 males per treatment group; ###p < 0.001: effect of time.





855 Figure S3. Effects of adult male mice exposure to DEHP alone or in a phthalate mixture on 856 dendritic spine density in the hippocampal CA2/3 area. A. Representative images of dendritic 857 spines in the CA2/3 area of the hippocampus of males mice treated with the vehicle (Veh), DEHP at 858 5 (DEHP-5) or 50 (DEHP-50) µg/kg/d, or to a phthalate mixture (Mix). Thin (t), stubby (s) and 859 mushroom (m) dendritic spines type are indicated. B. Quantification of total and subtype dendritic 860 spines (10 neurons per mouse). Data are presented as means \pm S.E.M. from 4 males per treatment group. Treatment effects (#p < 0.01, #p < 0.05) and post hoc comparisons (**p < 0.01, *p < 0.05) 861 862 are indicated.



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866 Figure S4. Effects of adult male mice exposure to DEHP alone or in a phthalate mixture on 867 synaptophysin levels and ultrastructural morphology in the CA1 area of the hippocampus. A. 868 Quantification of protein amount of synaptophysin normalized to GAPDH in the CA1 area of male 869 mice exposed to vehicle (Veh), DEHP at 5 (DEHP-5) or 50 (DEHP-50) µg/kg/d, or to a phthalate 870 mixture (Mix). Data are expressed as means ± S.E.M. from 4 males per treatment group. Treatment 871 effects (# p < 0.05) and post hoc comparisons (* p < 0.05) are indicated. **B**. Representative electron micrographs of three independent experiments per treatment showing synapses in the CA1 area. For 872 873 the Veh, synapses are clearly identifiable with the presynaptic element containing vesicles, and the 874 compact and electron-dense postsynaptic membrane (purple bracket), delimiting the synaptic cleft. 875 For DEHP-5, the ultrastructure of the synapses appears similar to that of the vehicle group. For

DEHP-50 and Mix, the presynaptic vesicles are not observable and it is difficult to distinguish the
presynaptic element from the post-synaptic element. Scale bar: 500 nm. Mit: mitochondria; Ax:
myelinated axon. Purple squares: inserts showing at higher magnification the corresponding synapses
indicated by the purple brackets.