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Molecular insights into the activation of Mre11-Rad50 endonuclease activity by Sae2/CtIP

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Summary

1 In *S. cerevisiae*, the Mre11-Rad50-Xrs2 (MRX)-Sae2 nuclease activity is required for the resection of
2 DNA breaks with secondary structures or protein blocks, while in humans, the MRE11-RAD50-NBS1
3 (MRN) homologue with CtIP is needed to initiate DNA end resection of all breaks. Phosphorylated
4 Sae2/CtIP stimulates the endonuclease activity of MRX/N. Structural insights into the activation of the
5 Mre11 nuclease are available only for organisms lacking Sae2/CtIP, so little is known how Sae2/CtIP
6 activates the nuclease ensemble. Here, we uncover the mechanism of Mre11 activation by Sae2 using
7 a combination of AlphaFold2 structural modeling, biochemical and genetic assays. We show that Sae2
8 stabilizes the Mre11 nuclease in a conformation poised to cleave substrate DNA. Several designs of
9 compensatory mutations establish how Sae2 activates MRX *in vitro* and *in vivo*, validating the structural
10 model. Last, our study uncovers how human CtIP, despite considerable sequence divergence, employs
11 a similar mechanism to activate MRN.

Keywords

DNA repair, Double-strand breaks, Nuclease, Meiosis, Homologous Recombination, AlphaFold2

Introduction

1 DNA double-strand breaks (DSBs) represent a threat to cellular survival and genome stability. Cells
2 possess two main DSB repair pathways: template-independent and potentially mutagenetic non-
3 homologous end-joining (NHEJ) and template-directed, hence more accurate, homologous
4 recombination (HR). HR is initiated by a process termed DNA end resection. In resection, the 5'-
5 terminated DNA strand at a DSB site is nucleolytically degraded, resulting in the formation of a 3'-
6 terminated single-stranded DNA (ssDNA) overhang. The overhang is then bound by the RAD51 strand
7 exchange protein to form a nucleoprotein filament with the capacity to identify and pair with
8 homologous DNA, which serves as a repair template. Once the DNA strand exchange takes place, the
9 invaded 3'-terminated end can prime DNA synthesis. There are multiple HR sub-pathways leading to
10 diverse genetic outcomes, however they all share the initial DNA end resection process ^{1,2}.

11 There are at least three conserved nucleases or nuclease complexes that catalyze DNA end resection
12 in eukaryotic cells. Their usage and relative importance are dependent on the nature of the DNA break,
13 cell cycle stage and physiological context. Resection pathways are generally divided into initial short-
14 range and downstream long-range resection steps ¹. Short-range resection is catalyzed by the
15 Mre11/MRE11 nuclease, which functions within the Mre11-Rad50-Xrs2 (MRX) complex in the budding
16 yeast model *Saccharomyces cerevisiae*, or MRE11-RAD50-NBS1 (MRN) in human cells ^{3,4}. Further
17 downstream, either EXO1 or DNA2 nuclease extend the resection tracks to perform long-range
18 resection ^{3,5,6}. Beyond the initiation of DNA end resection, the MRX/N proteins also have nuclease-
19 independent functions to promote NHEJ, or to activate DNA damage checkpoint through the control
20 of the apical Tel1/ATM kinase ⁷.

21 MRN is generally required to initiate DNA end resection in human cells under all conditions, while MRX
22 may be dispensable in budding yeast for the processing of chemically "clean" DSBs without protein
23 adducts or secondary structures, such as hairpins ⁸⁻¹⁰. The unique feature of MRX/N is the ability to
24 resect protein-blocked DSBs ¹¹. Both yeast and human MRX/N help remove the NHEJ factor Ku from
25 DNA ends, and in this way to channel non-productive NHEJ complexes into HR ¹²⁻¹⁵. MRX/N is also
26 necessary to process meiotic DNA breaks, catalyzed in a programmed manner by Spo11 during the
27 prophase of the first meiotic division ¹⁶⁻¹⁸. Spo11 remains covalently attached to the DNA end post-
28 cleavage and needs to be removed by MRX before further processing and repair. The MRX/N complex
29 employs a unique strategy, as it cuts the 5'-terminated DNA strand endonucleolytically some distance
30 away from the DSB ^{11,18}. The incision site is thus located on the internal side of the protein block, and
31 can serve as an entry site for the long-range resection nucleases acting downstream. The endonuclease
32 activity of the MRX/N complex is strongly stimulated by a phosphorylated co-factor, Sae2 in yeast and

1 CtIP in human cells ^{11,19}. Sae2/CtIP are phosphorylated in a cell cycle- and DNA damage-dependent
2 manner ²⁰⁻²³. Particularly, cyclin-dependent kinase (CDK) phosphorylation of Sae2/CtIP at S267/T847
3 occurs in the S and G2 phases, which allows DNA end resection only in the cell cycle phases when sister
4 chromatids are available as templates for repair. Mutations in components of the MRN complex or
5 CtIP lead to genome instability and several human diseases ^{24,25}.

6 Due to the complex phenotypes associated with MRX/N defects, resulting from impaired
7 recombination, end-joining and checkpoint signaling, biochemical and structural insights are necessary
8 to uncover the underlying mechanisms of Mre11 nuclease activation. The Mre11-Rad50 proteins are
9 heterodimers with a M₂R₂ stoichiometry ²⁶⁻²⁹. The nuclease, ATPase and DNA binding sites are a part
10 of globular domains that form a base, from which protrude two Rad50 coiled-coils, joined by a Zn-hook
11 at the apex of the structure ²⁹⁻³¹. Biochemical assays revealed that in yeast phosphorylated Sae2 is
12 bound by Rad50, albeit with a low affinity, and Xrs2 is largely dispensable for resection ^{7,23}. However,
13 these studies did not explain how Sae2/CtIP activates the Mre11/MRE11 endonuclease. Attempts to
14 obtain high resolution structures of the MRX/N complexes and Sae2/CtIP were unsuccessful to date,
15 particularly as both MRX/N and Sae2/CtIP oligomerize, making the sample very heterogeneous for
16 traditional structural biology approaches ³². Insights into a potential mode of Mre11 nuclease
17 activation came from studies employing bacteria or archaea, which contain homologues of Mre11 and
18 Rad50, but neither Sae2/CtIP nor Xrs2/NBS1 ³³⁻³⁹. From these studies, we learned that the MR
19 homologues exist in at least two basic conformations. In the nuclease inactive, or resting state, ATP-
20 bound Rad50 (SbcC) blocks the Mre11 (SbcD) active site, preventing the nuclease activity. Upon ATP
21 hydrolysis, the complex undergoes a major structural change, which places Mre11 to the side of the
22 dimer, enabling its endonuclease activity ³⁸. Nevertheless, due to the lack of Sae2/CtIP in these
23 organisms, it is not known how Sae2 participates in these structural transitions to activate the
24 endonuclease of the MRX/N complex.

25 Here, we modeled the structures of Mre11-Rad50 in its various states, and provide insights into the
26 mechanism of Mre11 endonuclease activation by phosphorylated Sae2. The data yielded structural
27 details of key protein-protein interfaces that assemble the enzyme complex in the nuclease active
28 state. The model was substantiated by the design of individual mutations that disrupt these interfaces,
29 and of combinations of the mutations on each side of the three interfaces that compensate each other
30 and allow the assembly of an active Mre11-Rad50-Sae2 ternary complex. These mutants were
31 extensively characterized biochemically *in vitro* and in genetic assays that necessitate the action of the
32 MRX nuclease, which validates the structural models. The data explain how Sae2 phosphorylation at
33 the S267 CDK site activates the nuclease ensemble, and we also designed a Rad50 mutant that partially
34 bypasses this activation. Sae2 and CtIP homologs are evolutionarily related although their sequence

1 diverged drastically during evolution. The size of the homologs can vary from less than 200 amino acids
2 in *P. tetraurelia* to more than 900 residues in *H. sapiens*⁴⁰, but also the sequence identity in the most
3 conserved C-terminal region can be as low as 10% between *S. cerevisiae* and *H. sapiens*⁴¹. Our analysis
4 revealed how these protein interfaces evolved in higher eukaryotic systems and how, in human cells,
5 CtIP is able to perform the same regulatory function despite its high sequence divergence with Sae2.

Results

1 **Alphafold2 predicts two conformations for ScMre11 and ScRad50 complex consistent with an auto-** 2 **inhibited and an activated state.**

3 Using AlphaFold2-multimer (AF2) algorithm, we predicted the structure of the (ScMre11)₂-(ScRad50)₂
4 assembly (M₂R₂) adapting the length of ScRad50 coiled-coil to match the size limits imposed by AF2.
5 The models converged into two distinct classes of structures differing by a major conformational
6 change at the interface between the Mre11 catalytic domains and the Rad50 dimer (Figure 1A and 1B).
7 The class 1 model is similar to the auto-inhibited form of the nuclease as observed in the experimental
8 structures of archaeal, bacterial and eukaryotic homologs (Figure 1A, Figure S1D) ^{29,34,35}. In the auto-
9 inhibited state, the Rad50 ATPase domain obstructs access to the Mre11 active site. Class 2 model
10 places the catalytic domain of Mre11 in an entirely different orientation (Figure 1B, Figure S1E), which
11 is highly asymmetric, and more similar to the arrangement experimentally observed with the active
12 state of the *E. coli* EcMre11 homologs in the context of a DNA bound to the EcRad50 groove (Figure
13 1C) ³⁸.

14 In the *E. coli* active state MR complex bound to DNA, the coiled-coils of EcRad50 are wrapped around
15 the DNA in a closed state, which brings the EcMre11 active site into the proper geometry to cleave
16 dsDNA substrates. The closed conformation of the ScRad50 coiled-coils was only generated by AF2 for
17 models integrating longer lengths of the Rad50 coiled-coil. Using a longer construct of (ScRad50)₂
18 alone, embedding up to 711 residues out of 1312, and without adding ScMre11, we could generate a
19 symmetrical structural model of ScRad50 coiled-coil forming a long rod using AF2 (Figure 1D, Figure
20 S1A and S1C). This closed conformation with contact points regularly distributed along the coiled-coil
21 sequence is similar to the rod structure observed in the low resolution cryoEM map of the *Escherichia*
22 *coli* MR homolog ³⁸ and in the recently reported structure of the *Chaetomium thermophilum* MRN
23 complex ²⁹ (Figure S1B). Thus, despite the different means of achieving MR nuclease activation
24 between organisms, in particular with or without a Sae2/CtIP homolog, our multiple sequence
25 alignment led AF2 models to converge on distinct classes of states likely to represent key evolutionarily
26 encoded functional steps.

27

28 **Addition of ScSae2 to ScMre11 and ScRad50 favors the activated state conformation.**

29 In most eukaryotes, stimulation of the endonuclease activity of the MR complex requires the binding
30 of a phosphorylated co-factor, Sae2 in *S. cerevisiae* or CtIP in *H. sapiens* ^{11,19}. To probe the mechanisms

1 underlying this process, we predicted the structure of the complex formed between ScMre11, ScRad50
2 and ScSae2 using AF2. For simplicity, we omitted ScXrs2, because it is largely dispensable for DNA
3 resection *in vitro* and *in vivo*⁷. We also focused on the C-terminal domain of Sae2, which was previously
4 shown to bind Rad50²³. We did not include the N-terminal domain of Sae2 that is involved in
5 tetramerization^{23,42,43}. Different constructs and lengths were explored ranging from a large
6 (ScMre11)₂-(ScRad50)₂-ScSae2(180-345) (M₂R₂S) assembly to a smaller one as shown in Figure 2A (see
7 Methods). In contrast to the (M₂R₂) assembly in which two classes of conformations were generated,
8 the presence of ScSae2 generated only the class 2 (Figure 2A), resembling the activated form of the *E.*
9 *coli* MR nuclease (Figure 1B and 1C), while the class 1, corresponding to the auto-inhibited form, was
10 no longer generated. The stimulatory function of scSae2 may thus be explained by shifting the
11 equilibrium of the MR structure towards the active conformation.

12 **A ternary complex forms between ScSae2, ScMre11 and ScRad50.**

13 The structure of ScSae2 in its state bound to M₂R₂ (Figure 2A and Figure S2A) is distinct from any known
14 fold in the PDB as established by the DALI server⁴⁴ (Table S1). In the model, the C-terminal domain of
15 ScSae2 is predicted to bind one Rad50 monomer on its outer β sheet (Figure 2A). It also contacts the
16 subunit of ScMre11 positioned to cleave dsDNA in the activated form, tightening the asymmetric
17 arrangement of the Mre11 dimer on that of Rad50 (Figure 1C). Therefore, ScSae2 is predicted to create
18 a trimeric interface connecting the outer β -sheet of ScRad50 and ScMre11 (Figure 2A and Figure S2D).

19 The region of ScMre11 in contact with ScSae2 and ScRad50 involves a short loop protruding from the
20 catalytic domain and folded as a helix (Figure 2B). This ScMre11 helix makes only a few direct contacts
21 with ScRad50 such as between residues E113^{ScRad50} and R181^{ScMre11}/R184^{ScMre11} (Figure 2C, Figure S2B).
22 In prokaryotes, a similar but larger helix, assigned as the fastener loop³⁸ (Figure 1C), was found in the
23 EcMre11 homologue, and was shown to play an important role to connect EcMre11 and EcRad50 by
24 binding the outer β sheet of Rad50 through a large network of polar and ionic interactions. Compared
25 to the structure of the *E. coli* homologs, the interface of the eukaryotic ScMre11-ScRad50 subunits
26 involving the helix exhibits a much smaller interaction surface (Figure 1B), suggesting that a part of the
27 fastener function in eukaryotes was taken up by Sae2 (Figure S2B)³⁹. Indeed, the AF2 structural model
28 of ScSae2 sheds light on this set of interactions, and Sae2 may strengthen the interaction of the small
29 interface between ScMre11 and ScRad50 as will be seen below.

30

31 **Anchoring of ScSae2 to the ScRad50 outer β -sheet**

1 Focus on the predicted interface between ScSae2 and ScRad50 identified an extensive network of
2 interactions involving side-chain and backbone atoms of ScSae2 (Figure 2, Figure S2B). The most
3 central residue involved in these interactions is S267^{ScSae2}, which specifically needs to be
4 phosphorylated for optimal stimulation of the MRX endonuclease ^{11,21}. The non phosphorylatable
5 S267A^{ScSae2} mutant was shown to be inefficient in MR stimulation whereas phospho-mimicking
6 S267E^{ScSae2} mutant partially retained the activity of phosphorylated wild-type Sae2. In our model,
7 S267^{ScSae2} is positioned near R77^{ScRad50} enabling the formation of a salt bridge between both residues
8 when S267^{ScSae2} is phosphorylated (Figure 2B). Downstream of S267^{ScSae2}, another salt bridge between
9 D275^{ScSae2} and R125^{ScRad50} was also predicted (Figure 2B). The K81^{ScRad50} ammonium moiety can form up
10 to three hydrogen bonds with the carbonyl backbone atoms of the triad R273^{ScSae2}-D275^{ScSae2}. The
11 involvement of K81^{ScRad50} is consistent with the original report of absence of meiotic DNA break
12 processing, sporulation defects and spore lethality in cells bearing the K81I (termed *rad50S*) mutation
13 ⁴⁵ (Figure 2B). This was later explained by the inability to remove Spo11 from break ends ¹⁸. In
14 biochemical assays, the K81I mutation in ScRad50 was found to impair ScMre11 endonuclease activity
15 by disrupting the physical interaction between phosphorylated ScSae2 C-terminus and ScRad50 ^{11,23}.
16 Last, the ScSae2-ScRad50 interface is potentially further stabilized by the interaction of the highly
17 conserved F271^{ScSae2} and F276^{ScSae2} aromatic rings packing against the carbon moieties of
18 aforementioned R77^{ScRad50} and K81^{ScRad50} sidechains, respectively (Figure 2B).

19

20 **Inspection of the interface between ScSae2 and ScMre11 in the M₂R₂S ternary complex.**

21 Several contacts can be distinguished between ScSae2 and ScMre11, which likely favor the cooperative
22 binding of ScSae2 and ScRad50 and the assembly with ScMre11 in a suitable geometry for nuclease
23 activation. First, the phosphorylated form of S267^{ScSae2} can form a salt-bridge with R187^{ScMre11} (Figure
24 2B). Three residues upstream in ScSae2, a second salt-bridge can be formed between R264^{ScSae2} and
25 E183^{ScMre11} located in the short helix protruding from ScMre11 (Figure 2C). The model also predicts the
26 formation of a small hydrophobic cluster involving L261^{ScSae2} and F186^{ScMre11} with a potential
27 contribution of surrounding side-chains from F229^{ScMre11} and Y254^{ScSae2} (Figure 2D), which likely further
28 stabilizes the complex. Altogether, a larger network of interactions was found in the two interfaces
29 involving ScSae2 with ScRad50 and ScMre11, while direct contacts between the Rad50 and Mre11
30 subunits appear rather limited compared to the complex formed in the bacterial homologs ^{38,39}. This
31 property may explain the need for a central role of phosphorylated ScSae2 in stabilizing a functional
32 and active nuclease. The global integration of the models shown in Figures 1B, 1D and 2A with the
33 positioning of the DNA shown in Figure 1C could be achieved without any major structural

1 rearrangement and converged on a coherent composite model (Figure 2E) representing the global
2 architecture of the ScRad50-ScMre11-ScSae2 assembly and the DNA in the activated form of the
3 complex.

4

5 **Assessment of the *S. cerevisiae* M₂R₂S structural model by targeted mutagenesis in biochemical and** 6 ***in vivo* assays**

7 Our structural model of the activated form of the ScM₂R₂-Sae2 predicts that phosphorylation of
8 S267^{ScSae2} stabilizes a conformation of ScSae2 that promotes the positioning of ScMre11 on ScRad50 in
9 its active state. To verify the predictions of this active structural arrangement, we designed 5 sets of
10 mutants to probe the various interfaces predicted by AF2 (Figure 2F). We tested each mutation alone
11 or in combination, to test for potential compensatory effects. To provide a simple quantitative readout
12 for the compensatory effect of the mutations, we calculated the theoretical impact of the mutations
13 assuming they were independent (i.e. on separate interfaces). Dividing the observed efficacy by the
14 theoretical value gave us a rescue coefficient (R). A R value >1 suggests that the mutations compensate
15 each other, and hence likely involve spatially close positions, while values ≈ 1 suggest no interaction.

16 We expressed and purified the wild-type and mutant recombinant proteins and protein complexes and
17 tested for their capacity to endonucleolytically cleave protein-blocked DNA in reconstituted
18 biochemical assays (Figure 3A and 3B and Figure S3). Additionally, we mutagenized *S. cerevisiae* cells
19 and determined the effects of the individual and combined mutations *in vivo* in processes previously
20 established to rely on the endonuclease activity of the MRX complex in conjunction with Sae2. To this
21 point, we first employed the assay developed by Lobachev et al., which measures the recombination
22 rate of DNA breaks terminated by hairpins formed by inverted repeated sequences, which require
23 MRX-Sae2 activity for initial hairpin processing (Figure 3C) ¹⁰. Second, we probed for meiotic
24 phenotypes. In *S. cerevisiae*, the Mre11 and Rad50 proteins are essential to promote DSB formation
25 by the Spo11 transesterase, independently of the MRX nuclease activity and thus likely by a structural
26 role (reviewed in ¹⁷). A complete loss of Mre11 or Rad50 therefore results in meiotic cells capable of
27 sporulation, but forming only unviable spores, due to the absence of meiotic DSBs (see graph Figure
28 3D). However, the Mre11 nuclease activity is absolutely required for the removal of Spo11
29 oligonucleotides from meiotic DSB ends to allow for DSB resection and subsequent repair by HR,
30 producing crossovers (Figure 3D). Mutants where the MRX is present but deficient in the Mre11
31 nuclease activity, such as *rad50*^{K81I} (called *rad50S*⁴⁵⁻⁴⁷) or *sae2Δ*, therefore allow meiotic DSB formation
32 but exhibit greatly impaired sporulation efficiency and spore viability, reduced or absent Spo11
33 oligonucleotides and a strong reduction in meiotic crossovers.

1 Set 1 of mutants was designed to probe the predicted small interface between ScRad50 and ScMre11
2 as suggested by the model in Figure 2C. In this interface, R181^{ScMre11} and R184^{ScMre11} make a contact
3 with E113^{ScRad50} (Figure 2C). We replaced the positively charged arginines R181^{ScMre11} and R184^{ScMre11}
4 with negatively charged glutamic acid residues, creating R181E^{ScMre11} and R184E^{ScMre11} mutants.
5 Reciprocally, on the Rad50 side, we generated the E113R^{ScRad50} charge reversal variant. *In vitro*, the
6 E113R^{ScRad50} mutation alone strongly reduced the endonuclease activity (Figure 3A and 3B). The
7 negative effects of the E113R^{ScRad50} mutation could be partially rescued by either R181E^{ScMre11} or
8 R184E^{ScMre11}, which had a moderate inhibitory effect in isolation (with ScRad50), with a rescue
9 coefficient of 3.1 and 4.2, respectively. The observed rescue effect confirmed that the mutants impact
10 the same interface, validating thus the structural model. In contrast, no rescue was observed in
11 combination with R187E^{ScMre11}, which is positioned at the other end of the scMre11 helix containing
12 R181^{ScMre11} and R184^{ScMre11}, and is thus less likely to interact with E113^{ScRad50} (rescue coefficient of 1.2),
13 indicating no interaction.

14 The hairpin cleavage genetic assays exhibited a much higher dynamic range (Figure 3C). The single
15 mutations E113R^{ScRad50}, R181E^{ScMre11} and R184E^{ScMre11} reduced the recombination rates 560-, 120- and
16 660-fold with respect to the wild-type strain, respectively. The recombination rates of the double
17 mutants between E113R^{ScRad50} and R181E^{ScMre11} or R184E^{ScMre11} were no worse than those of the single
18 mutant strains, suggesting an interaction (rescue coefficients of 561 and 293, respectively). Finally, the
19 meiotic phenotype, as assessed by the sporulation efficiency, confirmed these results, with the three
20 single mutants strongly affected, to levels similar to the *sae2Δ* mutation, whereas the combination of
21 E113R^{ScRad50} with R181E^{ScMre11} or R184E^{ScMre11} revealed rescue coefficients of 44 and 9, respectively,
22 validating the relevance of the interaction also in the context of meiotic recombination (Figure 3D).
23 Our biochemical and genetic experiments thus support the AF2 model predicting contacts between
24 E113R^{ScRad50} and R181E^{ScMre11}/R184E^{ScMre11}.

25 Set 2 of mutants was designed to probe the ScRad50 and ScSae2 interface by evaluating the
26 contribution of a potential salt bridge connecting R125^{ScRad50} and D275^{ScSae2} (Figure 2B). As above, we
27 generated charge reversal mutations and evaluated the D275R^{ScSae2} and R125D^{ScRad50} mutants
28 individually and combined. The rescue coefficient in biochemical experiments was rather modest
29 (R=1.7) (Figure 3E), while in the genetic hairpin cleavage assays much more apparent (R=1751) (Figure
30 3F). Indeed, while the R125D^{ScRad50} mutation decreased recombination rate 760-fold in otherwise wild-
31 type background, it increased recombination in the D275R^{ScSae2} cells, indicating a strong genetic
32 interaction. Similarly, in meiosis, while both single mutants showed strongly decreased sporulation,
33 similar to *sae2Δ*, the R125D^{ScRad50} mutation rescued the defect of D275R^{ScSae2} (R=58), confirming the

1 strong genetic interaction (Figure 3G). These *in vivo* results support the structural AF2 model for the
2 interface between ScRad50 and ScSae2.

3

4 **R77E substitution in Rad50 partially bypasses the requirement for co-activation of MRX by Sae2**

5 Phosphorylation of the conserved S267 residue in Sae2 by cyclin-dependent kinase (CDK) strongly
6 stimulates the endonuclease activity of MRX^{21,23}. While a non-phosphorylatable S267A^{ScSae2}
7 substitution mutant is strongly impaired, the phosphomimetic S267E^{ScSae2} retains some activity and
8 partially bypasses the requirement for CDK in resection²¹. The AF2 model predicted that
9 phosphorylated S267^{ScSae2} forms salt bridges with neighboring R187^{ScMre11} and R77^{ScRad50} (Figure 2B). In
10 this way, phosphorylated S267^{ScSae2} promotes the positioning of the Mre11 and Rad50 subunits to
11 specifically stabilize the nuclease in its active conformation. The third set of our mutants was designed
12 to probe specifically this crucial trimeric interface. We designed S267R^{ScSae2}; the positive charge of this
13 normally negatively charged residue (in its phosphorylated form) was anticipated to confer a strong
14 resection defect. We then set out to test this mutant in combination with charge reversal substitutions
15 R187E^{ScMre11} and R77E^{ScRad50}. As anticipated, the S267R^{ScSae2} mutant conferred a strongly impaired
16 endonuclease activity to MRX *in vitro* (Figure 4A). A combination of S267R^{ScSae2} and R77E^{ScRad50} rescued
17 resection *in vitro* (R=1.7) (Figure 4A). Likewise, the R187E^{ScMre11} and S267R^{ScSae2} mutations compensated
18 each other *in vitro* (R=1.9). Hairpin cleavage assays verified these interactions in a cellular setting: the
19 S267R^{ScSae2} substitution was as defective as the *sae2Δ* strain, and the R77E^{ScRad50} mutation increased
20 recombination in S267R^{ScSae2} cells (R=1330) (Figure 4B). Likewise, a combination with R187E^{ScMre11}
21 partially rescued recombination in S267R^{ScSae2} (R=552). Corroborating this effect on hairpin cleavage,
22 in meiosis, the S267R^{ScSae2} mutant was as defective as the *sae2Δ*, and this defect was compensated by
23 the R77E^{ScRad50} mutation, increasing sporulation efficiency (R=16) (Figure 4C), although not to a level
24 sufficient to ensure spore viability (Figure S4). It is important to note that Spo11 catalyzes 150-200
25 DSBs in each meiotic *S. cerevisiae* cell. In order to achieve a full spore viability, all of these breaks have
26 to be resected and repaired. Therefore, even a substantial rescue of DNA end resection, with only a
27 few unrepaired breaks remaining, is not expected to result in spore viability. R187E^{ScMre11} also
28 compensated the single S267R^{ScSae2}, to a more modest extent, as measured by sporulation efficacy
29 (R=9) (Figure 4C). These results converge towards S267^{ScSae2} closely and functionally interacting with
30 R77^{ScRad50} and R187^{ScMre11}.

31 Biochemical assays revealed that the R77E^{ScRad50} mutant was able to partially support resection without
32 Sae2, more than any other variant constructed in this study (Figure 4D). Additionally, R77E^{ScRad50}
33 promoted resection with pSae2 variants non-phosphorylatable at S267, but phosphorylated at other

1 Sae2 sites, which is necessary to prevent protein aggregation (Figure 4E) ²³. In *sae2Δ* cells, the
2 recombination rate in the presence of R77E^{ScRad50} increased notably (R=2489), showing that R77E^{ScRad50}
3 can partially bypass the requirement for Sae2 even in a cellular setting (Figure 4F). This was confirmed
4 in meiosis, where R77E^{ScRad50} strongly compensated the *sae2Δ* mutant for sporulation (R=13) (Figure
5 4G). However, the functional rescue was likely not sufficient to ensure the repair of all meiotic DSBs,
6 because spores remained unviable in the double mutant (Figure S4).

7 According to the AF2 model, the negative charge of R77E^{ScRad50} may place it in a favorable conformation
8 with respect to positively charged R187^{ScMre11}, and thus partially bypass the need for phosphorylated
9 S267^{ScSae2} normally sandwiched in between. To our knowledge, R77E^{ScRad50} is the first reported mutation
10 in the MRX complex that partially bypasses the requirement for Sae2 or its phosphorylation at the S267
11 site to co-activate its endonuclease activity *in vitro* and *in vivo*.

12

13 **Assessing the existence of a physical contact between ScSae2 and ScMre11.**

14 The AlphaFold2 model suggests that the ScSae2, ScMre11 and ScRad50 ternary assembly could be
15 additionally cooperatively stabilized by direct contact between ScSae2 and ScMre11. Three residues
16 upstream of S267^{ScSae2}, we identified a salt bridge between the positively charged R264^{ScSae2} and
17 negatively charged E183^{ScMre11} (Figure 2C) that could contribute to this tripartite stabilization. We set
18 out to test a charge reversal mutation R264E^{ScSae2} in combination with E183R^{ScMre11}. Both point mutants
19 exhibited defects in the reconstituted endonuclease assay *in vitro*. A notable rescue effect was
20 observed with combination of R264E^{ScSae2} and E183R^{ScMre11} *in vitro* (R= 1.8) (Figure 5A). In the hairpin
21 cleavage recombination assay, the compensatory effect was quite dramatic (R= 2670, the strongest of
22 any mutant combination in this study) (Figure 5B). Confirming these compelling *in vitro* and *in vivo*
23 results, the meiotic sporulation efficiency defect of the E183R^{ScMre11} was almost entirely rescued by the
24 R264E^{ScSae2} mutation, reaching nearly wild-type levels (R=10) (Figure 5C). This high rescue of
25 sporulation prompted us to investigate if other readouts of meiotic recombination were also rescued.
26 Indeed, this was the case, since spore viability of the double mutant was also nearly at a wild-type
27 level, suggesting that most meiotic DSBs are repaired to ensure meiotic progression and viable spores
28 (Figure 5D). Consistent with these dramatic compensatory effects, the amount of Spo11
29 oligonucleotides was nearly wild-type in the double mutant (Figure 5E). The combination of the
30 biochemical and cellular assays validates the structural model that identified a close and specific
31 interaction between the R264E^{ScSae2} and E183R^{ScMre11} residues, promoting the Mre11 nuclease.

32

1 **The small hydrophobic cluster between Sae2 and Mre11 favors the Mre11 nuclease**

2 Finally, we tested the functional relevance of a hydrophobic cluster our model identified at the
3 interface between Mre11 and Sae2, by mutating the L261^{ScSae2} residue, conserved in
4 *Saccharomycetaceae* species (mutant 5, Figure 2F). In both the biochemical and *in vivo* meiotic assays,
5 this mutant was defective, showing decreased *in vitro* resection (Figure S5A), as well a reduced
6 sporulation (Figure S5B) and spore viability (Figure S5C). These results indicate that this hydrophobic
7 patch supports optimal Mre11 conformation and nuclease activity.

Discussion

1 In all eukaryotes, the nuclease activity of the MR machinery is strongly dependent on a conserved
2 partner from the Sae2/CtIP family, whose sequences have diverged dramatically during evolution. The
3 physical interaction between MR and Sae2 was reported to be weak and transient ^{11,23}, likely due to
4 the necessity to keep the MR endonuclease under a tight control. Nevertheless, the AlphaFold2
5 generated models reported here predict a ternary complex formation that was validated by
6 mutagenesis in a combination of biochemical experiments *in vitro* and genetic assays in both
7 vegetative and meiotic yeast cells.

8 Our structural, biochemical and genetics study in *S. cerevisiae* revealed that the transition from the
9 inactive state of Mre11 to its active state, is triggered by the establishment of a cooperative set of
10 interactions centered on the phosphorylated status of S267^{ScSae2} (Figure 6). The addition of a negative
11 charge on S267^{ScSae2} by CDK-mediated phosphorylation switches the balance between unfavorable and
12 favorable interactions that can form at the outer β -sheet of Rad50 with Mre11. In ScSae2, the
13 conserved residues L261^{ScSae2}, R264^{ScSae2} and D275^{ScSae2} provide key complementary contributions to
14 the stabilization of the tripartite complex with ScMre11 and ScRad50. The validated structural model
15 provides further insights on how Mre11 could be released from its auto-inhibitory conformation
16 (Figure 1A) upon S267^{ScSae2} phosphorylation. In the Sae2 bound state, the backbones of residues R320-
17 Q322 (in helix α 9) (Figure S6A) are predicted to induce a steric clash with the capping domain of
18 ScMre11 in its inactive state, thus favoring its relocation toward the active state. Interestingly, the
19 telomeric protein Rif2 was proposed to antagonize this active conformation to specifically inhibit the
20 endonuclease activity of MRX at telomeres ⁴⁸⁻⁵⁰.

21 Sae2 and CtIP homologs are evolutionarily related although their sequence diverged drastically during
22 evolution. Intriguingly, the residues L261^{ScSae2}, R264^{ScSae2} and D275^{ScSae2} whose importance for the
23 formation of the tripartite complex with Mre11 and Rad50 have been shown, are essentially conserved
24 in the small subset of *Saccharomycetaceae* species (Figure 7A and Figures S2C and S7D) questioning
25 the generality of the unraveled mechanisms. In ScSae2, the C-terminal domain adopts a combination
26 of two small β -strands packing against α 2 and α 7- α 9 helices (Figure S2A) that seems completely absent
27 in human CtIP (Figure 7A). On another hand, in most eukaryotes CtIP contains a highly conserved
28 “CxxC...RHR” motif (termed C-rich-H domain) that is completely absent in ScSae2 and in related species
29 (Figure 7A). Interestingly, in *Yarrowia lipolytica*, a yeast belonging to the same subphylum as *S.*
30 *cerevisiae* (*Ascomycotina*), the features specific to either ScSae2 (2 β -strands domain) or HsCtIP (C-
31 rich-H domain) appear to coexist in the Sae2/CtIP hybrid ortholog, noted YICtIP (Figure 7A).

1 A model of the structure of the *Y. lipolytica* Rad50-Mre11-CtIP complex generated with AF2 provides
2 key insights to understand how the C-rich-H domain could contribute to the activation of the MR
3 nuclease. The C-rich-H domain is predicted to fold into a small two-helix domain, whose geometry is
4 constrained by the highly conserved cysteine and histidine residues forming a cluster (Figure 7B and
5 Figures S7A and S7B). The structural arrangement of the YIMR-CtIP ternary complex is very similar to
6 that obtained for the ScMR-Sae2 complex. Of utmost interest, as in the structure of ScMR-Sae2, an
7 apolar cluster is also formed between YICtIP and YIMre11 involving the short fastener helix of YIMre11
8 (Figure 7C). In the case of *Y. lipolytica*, one of the helices of the C-rich-H domain mediates this
9 interaction by exposing conserved apolar residues that pack against aromatic residues of the YIMre11
10 fastener helix, as in the MR-Sae2 complex (purple triangles in Figure 7A). In *S. cerevisiae*, deprived of
11 C-rich-H domain, this interaction was in part mediated by L261^{ScSae2} as measured by ‘mutant set 5’
12 assays (Figure 2D and Figure S5). Thus, the model of the *Y. lipolytica* assembly suggests that a role of
13 the highly conserved C-rich-H domain is to create an anchor between Mre11 and CtIP to stimulate the
14 active conformation of the nuclease. This orientation would additionally place conserved basic
15 residues in favorable position to interact with DNA.

16 We further compared the evolutionary properties of Sae2 and CtIP orthologs by performing AF2
17 structural predictions between human HsRad50, HsMre11 and HsCtIP (Figure 7D and Figure S7C). AF2
18 models of the human assembly lead to a conformation between the C-rich-H^{HsCtIP} domain and HsMre11
19 that was not as compact as for the *Y. lipolytica* proteins, but the two domains come close to each other
20 in a similar orientation. Conserved hydrophobic residues also protrude from the C-rich-H^{HsCtIP} and could
21 also favorably make contacts with the aromatic residues of the fastener HsMre11 with little
22 conformational change required. The contribution of DNA or other unmodeled partners in the
23 assembly may be necessary to reach the fully active geometry with the human proteins. Just
24 downstream of the RHR motif, T847^{HsCtIP}, equivalent to S267^{ScSae2}, is also phosphorylated by CDK²². In
25 this phosphorylated loop, the architecture of the state bound to the outer β sheet of HsRad50 is very
26 similar to that modeled for the *S. cerevisiae* complex, with T847^{HsCtIP} contacting R79^{HsRad50}, an arginine
27 in the position equivalent to R77^{ScRad50} (Figure 7E; compare with Figure 2B). The possibility of
28 establishing a charged salt-bridge interaction upon phosphorylation may therefore explain the
29 stabilization of the MR-CtIP complex as in ScSae2. In humans, phosphorylation on another residue,
30 T859^{HsCtIP}, by ATM/ATR kinase was shown to also promote MR activity⁵¹. In the structural model,
31 T859^{HsCtIP} contacts a basic residue, K6^{HsRad50}, which may additionally support the interaction with Rad50
32 upon CtIP phosphorylation (Figure 7E). The variety of ways in which the physico-chemical
33 complementarity is encoded at the interface between the homologs of Sae2/CtIP, Rad50 and Mre11
34 is also highlighted when looking at the case of SpCtp1, the ortholog of HsCtIP in *Schizosaccharomyces*

1 *pombe*. A specific polypeptide containing the conserved 15 amino acids at the C terminus of SpCtp1 is
2 sufficient to stimulate Mre11 endonuclease activity in the absence of phosphorylation by CDK⁵². In
3 SpCtp1, the position corresponding to the phosphorylated residues S267^{ScSae2} and T847^{HsCtIP} is occupied
4 by a hydrophobic amino-acid I283^{SpCtp1}. On the other hand, in SpRad50, the position equivalent to the
5 basic residues R77^{ScRad50} and R79^{HsRad50} is a L77^{SpRad50}. This physico-chemical switch from an electrostatic
6 to an apolar interaction could account for Ctp1 stabilizing the interaction with SpRad50 in a CDK-
7 independent manner and promoting the formation of a tri-partite interface with SpMre11 to activate
8 its endonuclease activity.

9 In conclusion, the comparative analysis of several interologous complexes has provided key insights
10 into functional configurations and mechanisms for interpreting the role of the conserved motifs such
11 as the C-rich-H domain in CtIP homologs. The CtIP and Sae2 families appear to have undergone
12 considerable structural reorganization during evolution, while maintaining an ability to stabilize the
13 active conformation of the Mre11 nuclease domain so that it can act on DNA clamped by Rad50.

14

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22

23 **Author Contributions**

24 R.G., P.C. and V.B. designed the study. H.B. and R.G. performed the structural modeling and designed
25 all the mutations. E.C. and P.C. performed all biochemical experiments. J.N. and V.B. designed and
26 carried out the genetic assays in yeast. R.G., P.C. and V.B. wrote the manuscript. All authors
27 contributed to prepare the final version of the manuscript.

28

29 **Declaration of interests**

30 The authors declare no competing interests.

Figures Legends

1 **Figure 1: Structural modeling of *S. cerevisiae* Mre11-Rad50 assembly.**

2 (A) Structure of the *S. cerevisiae* Mre11 (1-533) (green) - Rad50 (1-215+1106-1312) (blue) complex
3 generated in the first class of AF2 models in a 2:2 stoichiometry representative of an inactive form. (B)
4 Structure of the same *S. cerevisiae* complex as in panel (A) generated in the second class of AF2 models
5 more similar to an active form. The close-up view highlights that the fastener loop region is much
6 shorter in eukaryotic Mre11 than in bacterial Mre11. (C) Experimental structure (PDB: 6S85) of the
7 activated form of the *E. coli* Mre11-Rad50 complex in a 2:2 stoichiometry bound to a double-stranded
8 DNA substrate. Close-up view of the fastener loop folding as a helix that stabilizes the active state. (D)
9 Structural model of the *S. cerevisiae* Rad50 homodimer generated by AF2 (delimitations shown 1-400
10 and 950-1312), in which the coiled-coils assembled into a rod-like structure. The regions in contact
11 between two coiled-coils involve highly conserved glycines (Figure S1A) that are labeled in brown at
12 different positions along the coiled-coils. The G1083^{ScRad50} and G349^{ScRad50} are reminiscent of the
13 G849^{EcRad50} in *E. coli*, whose mutation to alanine was shown to affect significantly the endonuclease
14 activity of the bacterial enzyme ³⁸.

15

16 **Figure 2: Structural modeling of *S. cerevisiae* Sae2-Mre11-Rad50 assembly.**

17 (A) Structural model of the *S. cerevisiae* Sae2 (180-345) (orange) - Mre11 (1-303) (green) - Rad50 (1-
18 166+1206-1312) (blue) complex generated by AF2 in a 1:1:1 stoichiometry representative of the active
19 conformation form. (B) Close-up view of (A) highlighting the sidechains of interfacial residues forming
20 an interaction between the loop of ScSae2 phosphorylated at S267. (C) Second close-up view
21 corresponding to a different orientation with respect to panel (B) obtained after rotating the structure
22 by 180° around a vertical axis and highlighting the network of salt-bridges formed at the interface of
23 the trimer. (D) Third close-up view corresponding to a different orientation with respect to panel (B)
24 obtained after rotating the structure by 90° around a horizontal axis and highlighting the cluster of
25 apolar side-chains packing at the interface between ScSae2 and ScMre11. (E) General composite model
26 generated by integrating models from Figure 1B, 1C, 1D and 2A of the *S. cerevisiae* Sae2 (180-345)
27 (orange) - Mre11 (1-533) (green) - Rad50 (1-364+966-1312) (blue) in a 1:2:2 stoichiometry bound to
28 DNA and shown as surface representation. (F) Summary of the five sets of disruptive and compensatory
29 mutants designed to validate the importance of each of the contact points at the interface of the
30 complex assembly.

31

1 **Figure 3: Validation of the *S. cerevisiae* M₂R₂S structural model by targeted mutagenesis**

2 (A) Representative endonuclease assays with indicated proteins. Mre11 variants were co-expressed
3 with Xrs2 (used at 25 nM), Rad50 variants were used at 50 nM, and pSae2 at 100 nM. (B) Quantitation
4 of the assays such as shown in panel A. N=3-7; Error bars, SEM. The rescue coefficient (R), a value
5 obtained by dividing the observed by the theoretical efficacy, assuming the mutations were
6 independent, is shown in red. (C) Recombination frequencies of strains with the *lys2-AluIR* and *lys2-*
7 $\Delta 5'$ ectopic reporter system (illustrated by a cartoon). The rate of Lys⁺ recombinants is derived from
8 the mean recombination frequency determined from six different isolates of each strain. Error bars
9 represent confidence interval at 95% (see Methods). The red bar represents the expected
10 recombination rate if we consider the two mutations as independent (i.e, if their effects were
11 cumulative). The number in red indicates the rescue frequency (R), calculated as in (B). (D) Sporulation
12 efficiency of diploid strains bearing the indicated Mre11 complex and *SAE2* homozygous alleles. As
13 illustrated, a strain deficient for Mre11 nuclease does not process Spo11-dependent DSBs and shows
14 high defect in sporulation. As in (A) and (C), the rescue coefficient R in the double mutants is indicated
15 in red. Red bar: as in (C). Error bars represent the standard deviation (n=2). The total number of cells
16 counted for each genotype is indicated. Statistical test: Fisher test. P-value $\geq 0,05$ = NS ; P-value $< 0,05$
17 = * ; P-value $< 0,01$ = ** ; P-value $< 0,001$ = *** ; P-value $< 0,0001$ = ****. (E) Nuclease assays with the
18 indicated variants (Mre11-Xrs2, 25 nM; Rad50, 50 nM; pSae2, 100 nM). Bottom, a representative
19 experiment; top, a quantitation. N=6; Error bars, SEM.

20

21 **Figure 4: R77E substitution in Rad50 partially bypasses the requirement for co-activation of MRX by**
22 **Sae2**

23 (A) Nuclease assays with the indicated variants (Mre11-Xrs2, 25 nM; Rad50, 50 nM; pSae2, 100 nM).
24 Bottom, a representative experiment; top, a quantitation. N=4; Error bars, SEM. (B) Recombination
25 frequencies of strains with the *lys2-AluIR* and *lys2- $\Delta 5'$* ectopic reporter system, as in Figure 3C. (C)
26 Sporulation efficiency, as in Figure 3D. (D) Quantitation of the nuclease assays with the indicated
27 variants (Mre11-Xrs2, 25 nM; Rad50, 50 nM; pSae2, 100 nM). N=5; Error bars, SEM. (E) Quantitation
28 of assays with either wild-type MRX (assembled from Mre11-Xrs2 heterodimer, 25 nM, and Rad50, 50
29 nM), or MR^{R77E}X (assembled from Mre11-Xrs2 heterodimer, 25 nM, and Rad50^{R77E}, 50 nM), and either
30 mock-treated or λ phosphatase (λ) treated pSae2 variants. All pSae2 variants were prepared with
31 phosphatase inhibitors, and hence were initially phosphorylated at multiple residues, before
32 dephosphorylation by λ phosphatase, where indicated. N=3; Error bars, SEM. (F) Recombination

1 frequencies of strains with the *lys2-AluIR* and *lys2-Δ5'* ectopic reporter system, as in Figure 3C. (G)
2 Sporulation efficiency, as in Figure 3D.

3

4 **Figure 5: Validation of the interface between ScSae2 and ScMre11**

5 (A) Nuclease assays with the indicated variants (Mre11-Xrs2, 25 nM; Rad50, 50 nM; pSae2, 100 nM).
6 Bottom, a representative experiment; top, a quantitation. N=4; Error bars, SEM. (B) Recombination
7 frequencies of strains with the *lys2-AluIR* and *lys2-Δ5'* ectopic reporter system, as in Figure 3C. (C)
8 Sporulation efficiency, as in Figure 3D. (D) Spore viability of strains with the indicated mutations. The
9 number n of tetrad dissections is indicated for each. The statistical test used was a Fisher test between
10 the observed and expected spore viability (P-value < 0,0001 = ****). Red bar and red number: as in
11 Figure 3. (D) Spo11 oligonucleotide assay on strains with the indicated genotype. Two major bands
12 correspond to Spo11 oligonucleotides. The asterisk marks a non-specific terminal deoxynucleotidyl
13 transferase band.

14

15 **Figure 6: Schematic model for the molecular mechanism of activation of the nuclease complex.**

16 General model of the activation of the *S. cerevisiae* Mre11-Rad50-Sae2 complex upon phosphorylation
17 of S267 by CDK. In step 1, the states of Sae2 and Mre11 are off. In step 2, binding of the phosphorylated
18 loop to the outer β-sheet is incompatible with Mre11 in off-state. The added negative charge is
19 represented by a circle with a minus sign and the red star reports steric hindrance. In step 3, one of
20 the Mre11 dimer forms a trimeric assembly positioning Mre11 catalytic nuclease site in a position
21 poised to cut DNA (represented by Mn atoms as yellow circles).

22

23 **Figure 7: Evolution at the sequence and structural level of the interactions between Sae2/CtIP** 24 **orthologs and the Mre11-Rad50 complex.**

25 (A) Multiple sequence alignment of Sae2 and CtIP orthologs in the conserved C-terminal domain
26 involved in the Mre11 nuclease activation. Orange boxes indicate species with a 2 β-stranded globular
27 domain and the conserved positions associated with the presence of this domain. Purple boxes
28 indicate species with a cysteine-rich and histidine domain in their Sae2/CtIP orthologs and the
29 positions defining this domain. Two boxes indicate the positions of the helical segment that the models
30 identify as contributing to the formation of the apolar cluster between Sae2 or CtIP and Mre11 in their
31 active conformations. Species in bold correspond to those for which a structural model of the complex

1 with Mre11 and Rad50 was generated and analyzed. Purple triangles indicate the positions involved in
2 the formation of apolar clusters with Mre11 in the active conformation in the orthologs having a C-
3 rich-H domain. (B) Structural model of the *Y. lipolytica* CtIP/Sae2 (255-407) (orange+purple) - Mre11
4 (1-296) (green) - Rad50 (1-167+1179-1292) (blue) complex generated by AF2 in a 1:1:1 stoichiometry
5 representative of the active conformation form. (C) The close-up view of the model from panel B is
6 rotated by 90° with respect to panel B and highlights the relative orientation of the RHR, CxxC and
7 apolar cluster motifs when the C-rich-H domain interacts with the Mre11 fastener helix in the active
8 form. (D) Structural model of the *H. sapiens* CtIP (792-869) (purple) – 2 Mre11 (1-304 and 1-515)
9 (green) - Rad50 (1-220+1098-1312) (blue) complex generated by AF2 in a 1:1:1:1 stoichiometry
10 representative of the active conformation form. (E) Close-up view of the model from panel D showing
11 the sidechains of the threonine residues of CtIP phosphorylated by CDK and ATM and which interact
12 with basic sidechains of Rad50 outer β -sheet surface.

13

Methods

1 Structural modeling

2 Sequences of *S. cerevisiae*, *Y. lipolytica* and *H. sapiens* for the Mre11, Rad50 and Sae2/CtIP were
3 retrieved from UniProt database ⁵³ (Table S2). Mre11, Rad50 and CtIP from *H. sapiens* or *Y. lipolytica*
4 were used as input of mmseqs2 homology search program ⁵⁴ to generate a multiple sequence
5 alignment (MSA) against the UniRef30 clustered database ⁵⁵. Homologs sharing less than 25%
6 sequence identity with their respective query and less than 50% of coverage of the aligned region were
7 not kept. For Sae2, we used HHblits ⁵⁶ for its enhanced sensitivity with 8 iterations to retrieve remote
8 homologs against the UniRef30_2020_06 database. In every MSA, in case several homologs belonged
9 to the same species, only the one sharing highest sequence identity to the query was kept. Full-length
10 sequences of the orthologs were retrieved and re-aligned with mafft ⁵⁷. To model the structures of the
11 Mre11-Rad50-Sae2/CtIP complexes, the corresponding MSAs were concatenated. In the concatenated
12 MSAs, when homologs of different subunits belonged to the same species, they were aligned in a
13 paired manner, otherwise they were left unpaired ⁵⁸. The number of paired and unpaired sequences
14 in the final concatenated MSAs are reported in Table S2. Concatenated MSAs were used as inputs to
15 generate 25 structural models (5 different seeds) for each of the conditions using a local version of the
16 ColabFold v1.3 interface ⁵⁹ running 6 iterations of the AlphaFold2 v2.2.0 algorithm ⁶⁰ trained on the
17 multimer dataset ⁶¹ on a local HPC equipped with NVIDIA Ampere A100 80Go GPU cards. Four scores
18 were provided by AlphaFold2 to rate the quality of the models, the pLDDT, the pTMScore, the
19 ipTMScore and the model confidence score (weighted combination of pTM and ipTM scores with a
20 20:80 ratio) ⁶¹. The scores obtained for all the generated models are reported in Table S3 for the model
21 ranked first according to the confidence score. The models ranked first for each complex were relaxed
22 using rosetta relax protocols to remove steric clashes ⁶² under constraints (std dev. of 2 Å for the
23 interatomic distances) and were deposited in the ModelArchive database (<https://modelarchive.org/>)
24 (Table S4). The global composite model of the (Mre11)₂-(Rad50)₂-Sae2 in complex with DNA was built
25 by combining the *S. cerevisiae* models of (i) the long Rad50 dimer (Figure 1D, ModelArchive ID: ma-
26 i9sv6), (ii) the Mre11 dimers in the active conformation with Rad50 (Figure 1B, ModelArchive ID: ma-
27 jfoh4) and (iii) the trimeric assembly between Sae2, Mre11 and Rad50 (Figure 2A, ModelArchive ID:
28 ma-qhmpy). Model (iii) was first superimposed on model (i) to recover the relative position of Sae2
29 and of one Mre11 nuclease domain in the context of the closed Rad50 coiled-coil, taking the outer β-
30 sheet of Rad50 binding to Sae2 as a reference. The Mre11 nuclease domain from (iii) was then taken
31 as reference to model the relative position of the second Mre11 nuclease domain using model (ii). The
32 orientation of Mre11 (1-412) regions were then kept fixed while Mre11 domains (429-533) binding to
33 Rad50 coiled-coils as in model (ii) were modeled on the closed conformation of the Rad50 coiled-coil

1 using model (i) as reference for superposition. The flexible linkers Mre11 (412-429) connecting the two
2 Mre11 domains of each subunit were finally remodeled using RCD+⁶³. The positions of DNA,
3 nucleotides and Mn²⁺ ions were modeled using the superposition of the composite global model with
4 the bacterial assembly in PDB 6S85 taking the position of the Rad50 ATPase domains as a reference.
5 The resulting global model was relaxed using rosetta under the same conditions as described above.
6 The Molecular graphics and analyses were performed with UCSF ChimeraX⁶⁴. Multiple sequence
7 alignments were represented using JalView⁶⁵.

8 **Yeast strains**

9 *S. cerevisiae* strains used for all the meiotic experiments are diploids of the efficiently sporulating SK1
10 background⁶⁶. Strains used for the hairpin-opening assays are derived from the haploid HS21 strain,
11 of the CGL background^{10,67}. Their genotypes are listed in Table S5. Cells were grown and all assays were
12 performed at 30°C. Strains were transformed by the lithium acetate method. Spo11-His6-Flag3::NatMX
13 allele was amplified by PCR. Haploid *mre11Δ*, *rad50Δ* and *sae2Δ* strains for hairpin cleavage assays
14 were provided by K. Lobachev. *mre11Δ* and *sae2Δ* in the SK1 diploid strains were obtained by replacing
15 the entire coding sequence by the KanMX resistance marker⁶⁸. Introduction of all the *mre11*, *rad50* or
16 *sae2* point mutations were done in two steps: first, the regions surrounding the aminoacids to mutate
17 were replaced by the KanMX cassette by PCR with long flanking homology. The regions deleted were
18 *mre11Δ113-291*; *rad50Δ1-192* (which we also used as the “*rad50Δ*” for the meiotic experiments, since
19 it deletes the start codon and the first 192 residues of the protein); *sae2Δ193-345*. Then, these strains
20 were co-transformed with a NatMX selectable plasmid encoding both Cas9 and a guide RNA against
21 the KanMX sequence and a mutagenized PCR fragment spanning the deletion as a healing fragment.
22 Transformants were selected on clonNAT, and all transformants were sequenced for correct
23 introduction of the mutation and preservation of the remaining of the targeted gene.

24 **Yeast hairpin cleavage assays**

25 The rate of Lys+ recombinants was computed as the number of colonies growing on -Lys plates
26 compared to the total number of colonies growing on YPD rich medium¹⁰. The mean recombination
27 rate was calculated from six different colonies of each strain using the FalcOR website tool using the
28 MSS-MLE⁶⁹ and (<https://lianglab.brocku.ca/FALCOR/index.html>).

29 **Yeast meiotic assays**

30 Meiosis was induced synchronously by pre-growing cells in SPS pre-sporulation medium as previously
31 described⁷⁰. Sporulation efficiency was measured after 24 h in sporulation medium as the percentage
32 of cells that showed cells with two or more spores. At least two independent experiments were

1 performed and at least 400 cells were counted for each genotype. For spore viability, cells were put
2 on solid SPO plates for 48 h at 30°C. Spo11 oligonucleotide analysis was performed by
3 immunoprecipitating Spo11-His6-Flag3 cells at t = 4hr in sporulation medium essentially as described
4 in ⁷¹, with the following modifications: Dynabeads Protein G beads (Thermo Fisher Scientific : 10003D)
5 and mouse anti-FLAG M2 monoclonal antibody (Sigma-Aldrich : F3165) were used. Samples were run
6 on an Invitrogen NuPAGE Novex Bis Tris gel 4-12% gradient SDS-PAGE gel and dried under vacuum
7 before exposing to a Phosphorimager screen.

8 **MRX and Sae2 purification**

9
10 All proteins were expressed in *Spodoptera frugiperda* 9 (*Sf9*) cells. The Mre11 construct (vector pTP391
11 provided by T. Paull) contained a His-tag at the C- terminus, and was co-expressed and co-purified with
12 Xrs2, tagged at the C-terminus with FLAG (pTP694 vector from T. Paull). The heterodimer was purified
13 using affinity chromatography using NiNTA resin (Qiagen) followed by anti-FLAG M2 affinity resin
14 (Sigma) ¹¹. Rad50, FLAG-tagged at the C-terminus (vector pFB-Rad50-FLAG), was purified by using anti-
15 FLAG M2 affinity resin (Sigma) ⁷². Phosphorylated Sae2 (pSae2), MBP-tagged at the N-terminus and
16 his-tagged at the C-terminus, was prepared as described previously in the presence of phosphatase
17 inhibitors ²³ (vector pFB-MBP-Sae2-his). The MBP tag was cleaved and removed during protein
18 purification. The λ phosphatase treatment was carried out as described previously ²³. Point
19 mutagenesis of the expression vectors was carried out using the QuikChange site directed mutagenesis
20 kit (Agilent) according to standard procedures. The mutant proteins were then expressed and purified
21 in the same way as the wild type proteins.

22

23 **Endonuclease assays**

24 Nuclease assays (15 µl volume) were carried out with oligonucleotide-based DNA substrate blocked at
25 all ends with streptavidin (1 nM molecules, oligonucleotides, 210:

26 (GTAAGTGCCGCGGTGCGGGTGCCAGGGCGTGCCCTTGGGCTCCCCGGGCGCGTACTCCACCTCATGCATC

27 and 211:

28 GATGCATGAGGTGGAGTACGCGCCCGGGGAGCCCAAGGGCACGCCCTGGCACCCGCACCGCGGCACTTAC)

29 ¹¹, in a buffer containing 25 mM Tris-acetate, pH 7.5, 1 mM dithiothreitol, 5 mM magnesium acetate,
30 1 mM manganese acetate, 1 mM ATP, 80 U/ml pyruvate kinase (Sigma), 1 mM phosphoenolpyruvate,
31 0.25 mg/ml bovine serum albumin (New England Biolabs). 30 nM streptavidin (Sigma), was added to
32 the substrate containing the reaction mixture and incubated for 5 min at room temperature.
33 Recombinant proteins were then added on ice and incubated for 30 min at 30°C. Reactions were
34 terminated and the DNA products were separated by denaturing gel electrophoresis and detected by

- 1 autoradiography⁷³. The data were quantitated using Fiji (Image J). The signal below the substrate band
- 2 in no protein lane was removed from all lanes as a background. DNA end resection was then calculated
- 3 as the fraction of the endonucleolytic DNA cleavage products in each lane. The data were plotted using
- 4 GraphPad Prism (V8).

Supplementary material

Supplementary Figures legends

1 **Figure S1: Details on the structural modeling of *S. cerevisiae* Mre11-Rad50 assembly. Related to**

2 **Figure 1**

3 (A) Structural model of a homodimer of ScRad50 obtained using AlphaFold2 with delimitations 1-434
4 and 920-1312 in blue and purple cartoons with the positions of the conserved glycine residues in
5 contact with each other in the closed coiled-coils conformation highlighted in orange. (B)
6 Superimposition between the *S. cerevisiae* Rad50 model shown in panel A and the cryoEM structure
7 of the Rad50 homodimer from *Chaetomium thermophilum* (PDB: 7ZR1) highlighting the similarity and
8 reliability of the AF2 model. (C) Predicted Alignment Error (PAE) map calculated by AlphaFold2 showing
9 the confidence of the distances between residues of the ScRad50 model shown in panel A. (D) PAE
10 map calculated for the model with highest AF2 confidence score for the *S. cerevisiae* Mre11 (1-533)
11 (green) - Rad50 (1-215+1106-1312) (blue) complex generated in the first class of AF2 models in a 2:2
12 stoichiometry representative of an inactive form. (E) PAE map calculated for the model with highest
13 AF2 confidence score for the *S. cerevisiae* Mre11 (1-533) (green) - Rad50 (1-215+1106-1312) (blue)
14 complex generated in the first class of AF2 models in a 2:2 stoichiometry representative similar to an
15 active form.

16 **Figure S2: Details on the structural modeling of *S. cerevisiae* Mre11-Rad50-Sae2 assembly. Related**
17 **to Figure 2**

18 (A) Cartoon representation of the ScSae2 domain (180-345) as predicted in the bound conformation
19 to the ScMre11 (1-303) and ScRad50 (1-166 + 1206-1312) domains. The secondary structure elements
20 are labelled together with the position of the phosphorylated residue S267. (B) Close-up view of the
21 structural model of the *S. cerevisiae* Sae2 (180-345) (orange) - Mre11 (1-303) (green) - Rad50 (1-
22 166+1206-1312) (blue) complex generated by AF2 in a 1:1:1 stoichiometry representative of the active
23 conformation form as shown in Figure 2 with most of the amino-acids involved in the tripartite
24 interface labelled. (C) Multiple sequence alignment of the ScSae2 domain (180-345) with closely
25 related homologs from *Saccharomycetaceae* yeast species. Secondary structure elements as in panel
26 A are labelled together with the residues mutated in this study. The grey dots indicate the conserved
27 hydrophobic residues forming the hydrophobic core buried in this domain. Dotted lines represent
28 insertion regions in the homologs of ScSae2 which are not shown for clarity (D) Predicted Alignment
29 Error (PAE) map calculated by AlphaFold2 showing the confidence of the distances between the
30 residues in contact in the ScMre11 (1-303) - ScRad50 (1-166 + 1206-1312) - ScSae2 (180-345) modeled
31 complex.

1 **Figure S3: Purified recombinant proteins used in this study. Related to Figure 3**

2 Mre11 and Xrs2 proteins were co-expressed together and purified as a complex. Rad50 variants were
3 purified separately. Phosphorylated Sae2 and variants (indicated as pSae2) were expressed and
4 purified in the presence of phosphatase inhibitors, resulting in an electrophoretic mobility shift. Sae2
5 prepared without phosphatase inhibitors (right lane, Sae2) was included as a reference. The
6 polyacrylamide gels were stained with Coomassie blue.

7 **Figure S4: R77E substitution in Rad50 does not restore spore viability of *sae2S267R* or *sae2Δ*.**

8 **Related to Figure 4**

9 Spore viability of strains for the indicated mutations is shown. The number of tetrad dissections (n) is
10 indicated for each.

11

12 **Figure S5: The small hydrophobic cluster between Sae2 and Mre11 favors Mre11 nuclease. Related**
13 **to Figure 5**

14 (A) Nuclease assays with the indicated variants (Mre11-Xrs2, 25 nM; Rad50, 50 nM; pSae2, 100 nM).
15 Bottom, a representative experiment; top, a quantitation. N=3; Error bars, SEM. (B) Sporulation
16 efficiency, as in Figure 3D. (C) Spore viability of strains with the indicated mutations. The number of
17 tetrad dissections (n) is indicated for each.

18

19 **Figure S6: Models illustrating the molecular events occurring the transition between the inactive**
20 **and active conformations. Related to Figure 6**

21 (A) Superimposition between the structural model of the *S. cerevisiae* Mre11 (1-533) (green) - Rad50
22 (1-215+1106-1312) (blue) complex generated in the first class of AF2 models in a 2:2 stoichiometry
23 representative of an inactive form and the ScSae2 domain in complex with ScMre11 and ScRad50 in
24 the active conformation (as shown in Figure 2A) taking the outer β -sheet of Rad50 as a common
25 reference for the superimposition. (B) Cartoon illustrating in human the transition from an inactive to
26 an active state triggered by CtIP phosphorylation by CDK kinases on threonine 847 suggested from the
27 structural arrangement of the CxxC/RHR motif in the predicted model from *Y. lipolytica*.

28

29 **Figure S7: Details on the structural modeling of *Y. lipolytica* and *H. sapiens* Mre11-Rad50-Sae2**
30 **assembly. Related to Figure 7**

1 (A) Predicted Alignment Error (PAE) map calculated by AlphaFold2 showing the confidence of the
2 distances between the residues in contact in the *Y. lipolytica* CtIP/Sae2 (255-407) (orange+purple) -
3 Mre11 (1-296) (green) - Mre11 (1-486) (lime) - Rad50 (1-202+1097-1292) (blue) complex generated by
4 AF2 in a 1:1:1:1 stoichiometry representative of the active conformation form. (B) PAE map calculated
5 by AlphaFold2 showing the confidence of the distances between the residues in contact in the *Y.*
6 *lipolytica* CtIP/Sae2 (255-407) (orange+purple) - Mre11 (1-296) (green) - Rad50 (1-167+1182-1292)
7 (blue) complex generated by AF2 in a 1:1:1 stoichiometry representative of the active conformation
8 form. (C) PAE map calculated by AlphaFold2 showing the confidence of the distances between the
9 residues in contact in the *H. sapiens* CtIP (792-869) (purple) – 2 Mre11 (1-304 and 1-515) (green) -
10 Rad50 (1-220+1098-1312) (blue) complex generated by AF2 in a 1:1:1:1 stoichiometry representative
11 of the active conformation form. (D) Multiple sequence alignment of the ScSae2 domain (190-284) a
12 larger number of representative species than in Figure 6. Secondary structure elements as in panel A
13 are labelled together with the residues mutated in this study. The grey dots indicate the conserved
14 hydrophobic residues forming the hydrophobic core buried in this domain. Dotted lines represent
15 insertion regions in the homologs of ScSae2 which are not shown for clarity

Supplementary Tables legends

- 1 **Table S1: Ten top hits from the DALI server analysis.**
- 2 **Table S2: Description of conditions used to generate models.**
- 3 **Table S3: Summary of evaluation scores for models generated by AlphaFold2.**
- 4 **Table S4: Links to the ModelArchive database containing model coordinates, multiple sequence**
- 5 **alignments and evaluation scores.**
- 6 **Table S5: Yeast strains used in this study.**

References

1. Cejka, P., and Symington, L.S. (2021). DNA End Resection: Mechanism and Control. *Annu Rev Genet* 55, 285-307. 10.1146/annurev-genet-071719-020312.
2. Kawale, A.S., and Sung, P. (2020). Mechanism and significance of chromosome damage repair by homologous recombination. *Essays Biochem* 64, 779-790. 10.1042/EBC20190093.
3. Mimitou, E.P., and Symington, L.S. (2008). Sae2, Exo1 and Sgs1 collaborate in DNA double-strand break processing. *Nature* 455, 770-774. 10.1038/nature07312.
4. Paull, T.T., and Gellert, M. (1999). Nbs1 potentiates ATP-driven DNA unwinding and endonuclease cleavage by the Mre11/Rad50 complex. *Genes Dev* 13, 1276-1288. 10.1101/gad.13.10.1276.
5. Zhu, Z., Chung, W.H., Shim, E.Y., Lee, S.E., and Ira, G. (2008). Sgs1 helicase and two nucleases Dna2 and Exo1 resect DNA double-strand break ends. *Cell* 134, 981-994. 10.1016/j.cell.2008.08.037.
6. Gravel, S., Chapman, J.R., Magill, C., and Jackson, S.P. (2008). DNA helicases Sgs1 and BLM promote DNA double-strand break resection. *Genes Dev* 22, 2767-2772. 10.1101/gad.503108.
7. Oh, J., Al-Zain, A., Cannavo, E., Cejka, P., and Symington, L.S. (2016). Xrs2 Dependent and Independent Functions of the Mre11-Rad50 Complex. *Mol Cell* 64, 405-415. 10.1016/j.molcel.2016.09.011.
8. Llorente, B., and Symington, L.S. (2004). The Mre11 nuclease is not required for 5' to 3' resection at multiple HO-induced double-strand breaks. *Mol Cell Biol* 24, 9682-9694. 10.1128/MCB.24.21.9682-9694.2004.
9. Shibata, A., Moiani, D., Arvai, A.S., Perry, J., Harding, S.M., Genoio, M.M., Maity, R., van Rossum-Fikkert, S., Kertokallio, A., Romoli, F., et al. (2014). DNA double-strand break repair pathway choice is directed by distinct MRE11 nuclease activities. *Mol Cell* 53, 7-18. 10.1016/j.molcel.2013.11.003.
10. Lobachev, K.S., Gordenin, D.A., and Resnick, M.A. (2002). The Mre11 complex is required for repair of hairpin-capped double-strand breaks and prevention of chromosome rearrangements. *Cell* 108, 183-193. 10.1016/s0092-8674(02)00614-1.
11. Cannavo, E., and Cejka, P. (2014). Sae2 promotes dsDNA endonuclease activity within Mre11-Rad50-Xrs2 to resect DNA breaks. *Nature* 514, 122-125. 10.1038/nature13771.
12. Chanut, P., Britton, S., Coates, J., Jackson, S.P., and Calsou, P. (2016). Coordinated nuclease activities counteract Ku at single-ended DNA double-strand breaks. *Nat Commun* 7, 12889. 10.1038/ncomms12889.
13. Reginato, G., Cannavo, E., and Cejka, P. (2017). Physiological protein blocks direct the Mre11-Rad50-Xrs2 and Sae2 nuclease complex to initiate DNA end resection. *Genes Dev* 31, 2325-2330. 10.1101/gad.308254.117.
14. Wang, W., Daley, J.M., Kwon, Y., Krasner, D.S., and Sung, P. (2017). Plasticity of the Mre11-Rad50-Xrs2-Sae2 nuclease ensemble in the processing of DNA-bound obstacles. *Genes Dev* 31, 2331-2336. 10.1101/gad.307900.117.
15. Deshpande, R.A., Myler, L.R., Soniat, M.M., Makharashvili, N., Lee, L., Lees-Miller, S.P., Finkelstein, I.J., and Paull, T.T. (2020). DNA-dependent protein kinase promotes DNA end processing by MRN and CtIP. *Sci Adv* 6, eaay0922. 10.1126/sciadv.aay0922.
16. Keeney, S., Giroux, C.N., and Kleckner, N. (1997). Meiosis-specific DNA double-strand breaks are catalyzed by Spo11, a member of a widely conserved protein family. *Cell* 88, 375-384. 10.1016/s0092-8674(00)81876-0.
17. Borde, V. (2007). The multiple roles of the Mre11 complex for meiotic recombination. *Chromosome Res* 15, 551-563. 10.1007/s10577-007-1147-9.
18. Neale, M.J., Pan, J., and Keeney, S. (2005). Endonucleolytic processing of covalent protein-linked DNA double-strand breaks. *Nature* 436, 1053-1057. 10.1038/nature03872.
19. Anand, R., Ranjha, L., Cannavo, E., and Cejka, P. (2016). Phosphorylated CtIP Functions as a Co-factor of the MRE11-RAD50-NBS1 Endonuclease in DNA End Resection. *Mol Cell* 64, 940-950. 10.1016/j.molcel.2016.10.017.

20. Ira, G., Pelliccioli, A., Balijja, A., Wang, X., Fiorani, S., Carotenuto, W., Liberi, G., Bressan, D., Wan, L., Hollingsworth, N.M., et al. (2004). DNA end resection, homologous recombination and DNA damage checkpoint activation require CDK1. *Nature* *431*, 1011-1017. [10.1038/nature02964](https://doi.org/10.1038/nature02964).
21. Huertas, P., Cortes-Ledesma, F., Sartori, A.A., Aguilera, A., and Jackson, S.P. (2008). CDK targets Sae2 to control DNA-end resection and homologous recombination. *Nature* *455*, 689-692. [10.1038/nature07215](https://doi.org/10.1038/nature07215).
22. Huertas, P., and Jackson, S.P. (2009). Human CtIP mediates cell cycle control of DNA end resection and double strand break repair. *J Biol Chem* *284*, 9558-9565. [10.1074/jbc.M808906200](https://doi.org/10.1074/jbc.M808906200).
23. Cannavo, E., Johnson, D., Andres, S.N., Kissling, V.M., Reinert, J.K., Garcia, V., Erie, D.A., Hess, D., Thoma, N.H., Enchev, R.I., et al. (2018). Regulatory control of DNA end resection by Sae2 phosphorylation. *Nat Commun* *9*, 4016. [10.1038/s41467-018-06417-5](https://doi.org/10.1038/s41467-018-06417-5).
24. Qvist, P., Huertas, P., Jimeno, S., Nyegaard, M., Hassan, M.J., Jackson, S.P., and Borglum, A.D. (2011). CtIP Mutations Cause Seckel and Jawad Syndromes. *PLoS Genet* *7*, e1002310. [10.1371/journal.pgen.1002310](https://doi.org/10.1371/journal.pgen.1002310).
25. Bian, L., Meng, Y., Zhang, M., and Li, D. (2019). MRE11-RAD50-NBS1 complex alterations and DNA damage response: implications for cancer treatment. *Mol Cancer* *18*, 169. [10.1186/s12943-019-1100-5](https://doi.org/10.1186/s12943-019-1100-5).
26. Hopfner, K.P., Karcher, A., Craig, L., Woo, T.T., Carney, J.P., and Tainer, J.A. (2001). Structural biochemistry and interaction architecture of the DNA double-strand break repair Mre11 nuclease and Rad50-ATPase. *Cell* *105*, 473-485. [10.1016/s0092-8674\(01\)00335-x](https://doi.org/10.1016/s0092-8674(01)00335-x).
27. de Jager, M., Trujillo, K.M., Sung, P., Hopfner, K.P., Carney, J.P., Tainer, J.A., Connelly, J.C., Leach, D.R., Kanaar, R., and Wyman, C. (2004). Differential arrangements of conserved building blocks among homologs of the Rad50/Mre11 DNA repair protein complex. *J Mol Biol* *339*, 937-949. [10.1016/j.jmb.2004.04.014](https://doi.org/10.1016/j.jmb.2004.04.014).
28. Seifert, F.U., Lammens, K., Stoehr, G., Kessler, B., and Hopfner, K.P. (2016). Structural mechanism of ATP-dependent DNA binding and DNA end bridging by eukaryotic Rad50. *EMBO J* *35*, 759-772. [10.15252/embj.201592934](https://doi.org/10.15252/embj.201592934).
29. Rotheneder, M., Stakyte, K., van de Logt, E., Bartho, J.D., Lammens, K., Fan, Y., Alt, A., Kessler, B., Jung, C., Roos, W.P., et al. (2023). Cryo-EM structure of the Mre11-Rad50-Nbs1 complex reveals the molecular mechanism of scaffolding functions. *Mol Cell* *83*, 167-185. [e169. 10.1016/j.molcel.2022.12.003](https://doi.org/10.1016/j.molcel.2022.12.003).
30. Hopfner, K.P., Craig, L., Moncalian, G., Zinkel, R.A., Usui, T., Owen, B.A., Karcher, A., Henderson, B., Bodmer, J.L., McMurray, C.T., et al. (2002). The Rad50 zinc-hook is a structure joining Mre11 complexes in DNA recombination and repair. *Nature* *418*, 562-566. [10.1038/nature00922](https://doi.org/10.1038/nature00922).
31. Park, Y.B., Hohl, M., Padjasek, M., Jeong, E., Jin, K.S., Krezel, A., Petrini, J.H., and Cho, Y. (2017). Eukaryotic Rad50 functions as a rod-shaped dimer. *Nat Struct Mol Biol* *24*, 248-257. [10.1038/nsmb.3369](https://doi.org/10.1038/nsmb.3369).
32. Kissling, V.M., Reginato, G., Bianco, E., Kasaciunaite, K., Tilma, J., Cereghetti, G., Schindler, N., Lee, S.S., Guerois, R., Luke, B., et al. (2022). Mre11-Rad50 oligomerization promotes DNA double-strand break repair. *Nat Commun* *13*, 2374. [10.1038/s41467-022-29841-0](https://doi.org/10.1038/s41467-022-29841-0).
33. Lammens, K., Bemeleit, D.J., Mockel, C., Clausing, E., Schele, A., Hartung, S., Schiller, C.B., Lucas, M., Angermuller, C., Soding, J., et al. (2011). The Mre11:Rad50 structure shows an ATP-dependent molecular clamp in DNA double-strand break repair. *Cell* *145*, 54-66. [10.1016/j.cell.2011.02.038](https://doi.org/10.1016/j.cell.2011.02.038).
34. Lim, H.S., Kim, J.S., Park, Y.B., Gwon, G.H., and Cho, Y. (2011). Crystal structure of the Mre11-Rad50-ATPgammaS complex: understanding the interplay between Mre11 and Rad50. *Genes Dev* *25*, 1091-1104. [10.1101/gad.2037811](https://doi.org/10.1101/gad.2037811).
35. Mockel, C., Lammens, K., Schele, A., and Hopfner, K.P. (2012). ATP driven structural changes of the bacterial Mre11:Rad50 catalytic head complex. *Nucleic Acids Res* *40*, 914-927. [10.1093/nar/gkr749](https://doi.org/10.1093/nar/gkr749).
36. Sung, S., Li, F., Park, Y.B., Kim, J.S., Kim, A.K., Song, O.K., Kim, J., Che, J., Lee, S.E., and Cho, Y. (2014). DNA end recognition by the Mre11 nuclease dimer: insights into resection and repair of damaged DNA. *EMBO J* *33*, 2422-2435. [10.15252/embj.201488299](https://doi.org/10.15252/embj.201488299).

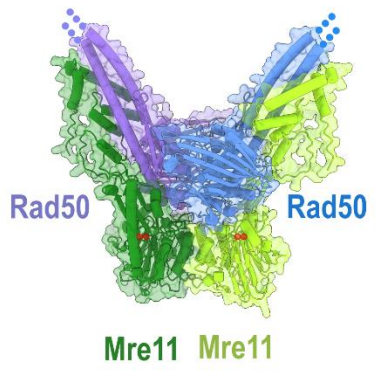
37. Liu, Y., Sung, S., Kim, Y., Li, F., Gwon, G., Jo, A., Kim, A.K., Kim, T., Song, O.K., Lee, S.E., and Cho, Y. (2016). ATP-dependent DNA binding, unwinding, and resection by the Mre11/Rad50 complex. *EMBO J* 35, 743-758. [10.15252/embj.201592462](https://doi.org/10.15252/embj.201592462).
38. Kashammer, L., Saathoff, J.H., Lammens, K., Gut, F., Bartho, J., Alt, A., Kessler, B., and Hopfner, K.P. (2019). Mechanism of DNA End Sensing and Processing by the Mre11-Rad50 Complex. *Mol Cell* 76, 382-394 e386. [10.1016/j.molcel.2019.07.035](https://doi.org/10.1016/j.molcel.2019.07.035).
39. Gut, F., Kashammer, L., Lammens, K., Bartho, J.D., Boggusch, A.M., van de Logt, E., Kessler, B., and Hopfner, K.P. (2022). Structural mechanism of endonucleolytic processing of blocked DNA ends and hairpins by Mre11-Rad50. *Mol Cell* 82, 3513-3522 e3516. [10.1016/j.molcel.2022.07.019](https://doi.org/10.1016/j.molcel.2022.07.019).
40. Godau, J., Ferretti, L.P., Trenner, A., Dubois, E., von Aesch, C., Marmignon, A., Simon, L., Kapusta, A., Guerois, R., Betermier, M., and Sartori, A.A. (2019). Identification of a miniature Sae2/Ctp1/CtIP ortholog from *Paramecium tetraurelia* required for sexual reproduction and DNA double-strand break repair. *DNA Repair (Amst)* 77, 96-108. [10.1016/j.dnarep.2019.03.011](https://doi.org/10.1016/j.dnarep.2019.03.011).
41. Sartori, A.A., Lukas, C., Coates, J., Mistrik, M., Fu, S., Bartek, J., Baer, R., Lukas, J., and Jackson, S.P. (2007). Human CtIP promotes DNA end resection. *Nature* 450, 509-514. [10.1038/nature06337](https://doi.org/10.1038/nature06337).
42. Andres, S.N., Appel, C.D., Westmoreland, J.W., Williams, J.S., Nguyen, Y., Robertson, P.D., Resnick, M.A., and Williams, R.S. (2015). Tetrameric Ctp1 coordinates DNA binding and DNA bridging in DNA double-strand-break repair. *Nat Struct Mol Biol* 22, 158-166. [10.1038/nsmb.2945](https://doi.org/10.1038/nsmb.2945).
43. Forment, J.V., Jackson, S.P., and Pellegrini, L. (2015). When two is not enough: a CtIP tetramer is required for DNA repair by Homologous Recombination. *Nucleus* 6, 344-348. [10.1080/19491034.2015.1086050](https://doi.org/10.1080/19491034.2015.1086050).
44. Holm, L. (2022). Dali server: structural unification of protein families. *Nucleic Acids Res* 50, W210-W215. [10.1093/nar/gkac387](https://doi.org/10.1093/nar/gkac387).
45. Alani, E., Padmore, R., and Kleckner, N. (1990). Analysis of wild-type and rad50 mutants of yeast suggests an intimate relationship between meiotic chromosome synapsis and recombination. *Cell* 61, 419-436. [10.1016/0092-8674\(90\)90524-i](https://doi.org/10.1016/0092-8674(90)90524-i).
46. de Jager, M., and Kanaar, R. (2002). Genome instability and Rad50(S): subtle yet severe. *Genes Dev* 16, 2173-2178. [10.1101/gad.1025402](https://doi.org/10.1101/gad.1025402).
47. Nairz, K., and Klein, F. (1997). mre11S--a yeast mutation that blocks double-strand-break processing and permits nonhomologous synapsis in meiosis. *Genes Dev* 11, 2272-2290. [10.1101/gad.11.17.2272](https://doi.org/10.1101/gad.11.17.2272).
48. Marsella, A., Gobbin, E., Cassani, C., Tisi, R., Cannavo, E., Reginato, G., Cejka, P., and Longhese, M.P. (2021). Sae2 and Rif2 regulate MRX endonuclease activity at DNA double-strand breaks in opposite manners. *Cell Rep* 34, 108906. [10.1016/j.celrep.2021.108906](https://doi.org/10.1016/j.celrep.2021.108906).
49. Khayat, F., Cannavo, E., Alshmary, M., Foster, W.R., Chahwan, C., Maddalena, M., Smith, C., Oliver, A.W., Watson, A.T., Carr, A.M., et al. (2021). Inhibition of MRN activity by a telomere protein motif. *Nat Commun* 12, 3856. [10.1038/s41467-021-24047-2](https://doi.org/10.1038/s41467-021-24047-2).
50. Roisne-Hamelin, F., Pobiega, S., Jezequel, K., Miron, S., Depagne, J., Veaute, X., Busso, D., Du, M.L., Callebaut, I., Charbonnier, J.B., et al. (2021). Mechanism of MRX inhibition by Rif2 at telomeres. *Nat Commun* 12, 2763. [10.1038/s41467-021-23035-w](https://doi.org/10.1038/s41467-021-23035-w).
51. Wang, H., Shi, L.Z., Wong, C.C., Han, X., Hwang, P.Y., Truong, L.N., Zhu, Q., Shao, Z., Chen, D.J., Berns, M.W., et al. (2013). The interaction of CtIP and Nbs1 connects CDK and ATM to regulate HR-mediated double-strand break repair. *PLoS Genet* 9, e1003277. [10.1371/journal.pgen.1003277](https://doi.org/10.1371/journal.pgen.1003277).
52. Zdravkovic, A., Daley, J.M., Dutta, A., Niwa, T., Murayama, Y., Kanamaru, S., Ito, K., Maki, T., Argunhan, B., Takahashi, M., et al. (2021). A conserved Ctp1/CtIP C-terminal peptide stimulates Mre11 endonuclease activity. *Proc Natl Acad Sci U S A* 118. [10.1073/pnas.2016287118](https://doi.org/10.1073/pnas.2016287118).
53. UniProt, C. (2021). UniProt: the universal protein knowledgebase in 2021. *Nucleic Acids Res* 49, D480-D489. [10.1093/nar/gkaa1100](https://doi.org/10.1093/nar/gkaa1100).
54. Steinegger, M., and Soding, J. (2017). MMseqs2 enables sensitive protein sequence searching for the analysis of massive data sets. *Nat Biotechnol* 35, 1026-1028. [10.1038/nbt.3988](https://doi.org/10.1038/nbt.3988).

55. Mirdita, M., von den Driesch, L., Galiez, C., Martin, M.J., Soding, J., and Steinegger, M. (2017). Uniclust databases of clustered and deeply annotated protein sequences and alignments. *Nucleic Acids Res* *45*, D170-D176. 10.1093/nar/gkw1081.
56. Remmert, M., Biegert, A., Hauser, A., and Soding, J. (2011). HHblits: lightning-fast iterative protein sequence searching by HMM-HMM alignment. *Nat Methods* *9*, 173-175. 10.1038/nmeth.1818.
57. Katoh, K., and Standley, D.M. (2013). MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Mol Biol Evol* *30*, 772-780. 10.1093/molbev/mst010.
58. Bryant, P., Pozzati, G., and Elofsson, A. (2022). Improved prediction of protein-protein interactions using AlphaFold2. *Nat Commun* *13*, 1265. 10.1038/s41467-022-28865-w.
59. Mirdita, M., Schutze, K., Moriwaki, Y., Heo, L., Ovchinnikov, S., and Steinegger, M. (2022). ColabFold: making protein folding accessible to all. *Nat Methods* *19*, 679-682. 10.1038/s41592-022-01488-1.
60. Jumper, J., Evans, R., Pritzel, A., Green, T., Figurnov, M., Ronneberger, O., Tunyasuvunakool, K., Bates, R., Zidek, A., Potapenko, A., et al. (2021). Highly accurate protein structure prediction with AlphaFold. *Nature* *596*, 583-589. 10.1038/s41586-021-03819-2.
61. Evans, R., O'Neill, M., Pritzel, A., Antropova, N., Senior, A.W., Green, T., Židek, A., Bates, R., Blackwell, S., Yim, J., et al. (2022). Protein complex prediction with AlphaFold-Multimer. *bioRxiv*.
62. Lemay, J.K., Weitzner, B.D., Lewis, S.M., Adolf-Bryfogle, J., Alam, N., Alford, R.F., Aprahamian, M., Baker, D., Barlow, K.A., Barth, P., et al. (2020). Macromolecular modeling and design in Rosetta: recent methods and frameworks. *Nat Methods* *17*, 665-680. 10.1038/s41592-020-0848-2.
63. Lopez-Blanco, J.R., Canosa-Valls, A.J., Li, Y., and Chacon, P. (2016). RCD+: Fast loop modeling server. *Nucleic Acids Res* *44*, W395-400. 10.1093/nar/gkw395.
64. Pettersen, E.F., Goddard, T.D., Huang, C.C., Meng, E.C., Couch, G.S., Croll, T.I., Morris, J.H., and Ferrin, T.E. (2021). UCSF ChimeraX: Structure visualization for researchers, educators, and developers. *Protein Sci* *30*, 70-82. 10.1002/pro.3943.
65. Waterhouse, A.M., Procter, J.B., Martin, D.M., Clamp, M., and Barton, G.J. (2009). Jalview Version 2--a multiple sequence alignment editor and analysis workbench. *Bioinformatics* *25*, 1189-1191. 10.1093/bioinformatics/btp033.
66. Kane, S.M., and Roth, R. (1974). Carbohydrate metabolism during ascospore development in yeast. *J Bacteriol* *118*, 8-14. 10.1128/jb.118.1.8-14.1974.
67. Lobachev, K.S., Stenger, J.E., Kozyreva, O.G., Jurka, J., Gordenin, D.A., and Resnick, M.A. (2000). Inverted Alu repeats unstable in yeast are excluded from the human genome. *EMBO J* *19*, 3822-3830. 10.1093/emboj/19.14.3822.
68. Longtine, M.S., McKenzie, A., 3rd, Demarini, D.J., Shah, N.G., Wach, A., Brachat, A., Philippsen, P., and Pringle, J.R. (1998). Additional modules for versatile and economical PCR-based gene deletion and modification in *Saccharomyces cerevisiae*. *Yeast* *14*, 953-961. 10.1002/(SICI)1097-0061(199807)14:10<953::AID-YEA293>3.0.CO;2-U.
69. Hall, B.M., Ma, C.X., Liang, P., and Singh, K.K. (2009). Fluctuation analysis CalculatOR: a web tool for the determination of mutation rate using Luria-Delbruck fluctuation analysis. *Bioinformatics* *25*, 1564-1565. 10.1093/bioinformatics/btp253.
70. Murakami, H., Borde, V., Nicolas, A., and Keeney, S. (2009). Gel electrophoresis assays for analyzing DNA double-strand breaks in *Saccharomyces cerevisiae* at various spatial resolutions. *Methods Mol Biol* *557*, 117-142. 10.1007/978-1-59745-527-5_9.
71. Neale, M.J., and Keeney, S. (2009). End-labeling and analysis of Spo11-oligonucleotide complexes in *Saccharomyces cerevisiae*. *Methods Mol Biol* *557*, 183-195. 10.1007/978-1-59745-527-5_12.
72. Cannavo, E., Reginato, G., and Cejka, P. (2019). Stepwise 5' DNA end-specific resection of DNA breaks by the Mre11-Rad50-Xrs2 and Sae2 nuclease ensemble. *Proc Natl Acad Sci U S A* *116*, 5505-5513. 10.1073/pnas.1820157116.
73. Pinto, C., Anand, R., and Cejka, P. (2018). Methods to Study DNA End Resection II: Biochemical Reconstitution Assays. *Methods Enzymol* *600*, 67-106. 10.1016/bs.mie.2017.11.009.

Figure 1

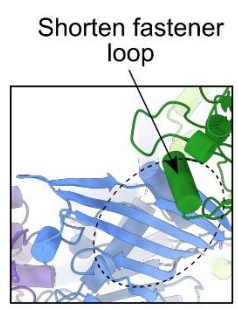
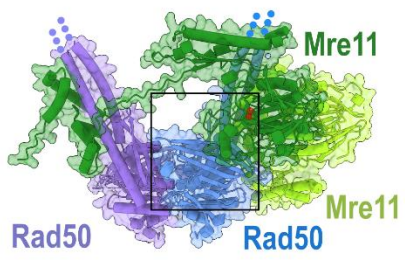
A

Mre11-Rad50 (S. cerevisiae)
class 1 = 'inactive'

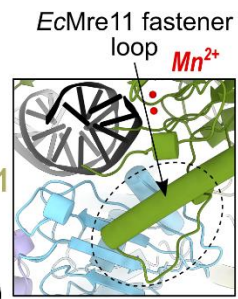
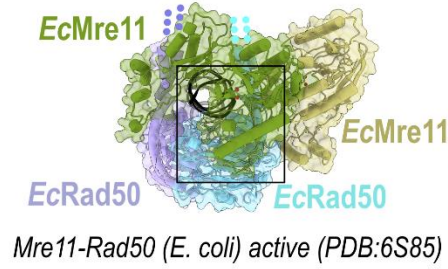


B

Mre11-Rad50 (S. cerevisiae)
class 2 = 'active'



C



D

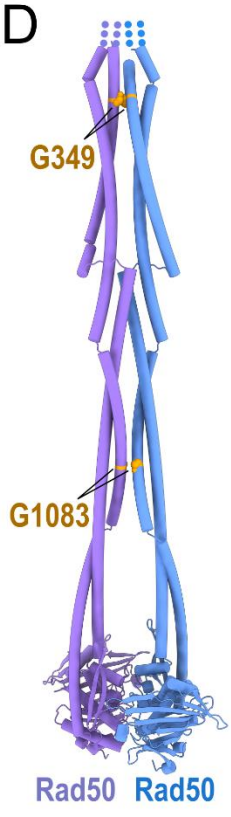


Figure 2

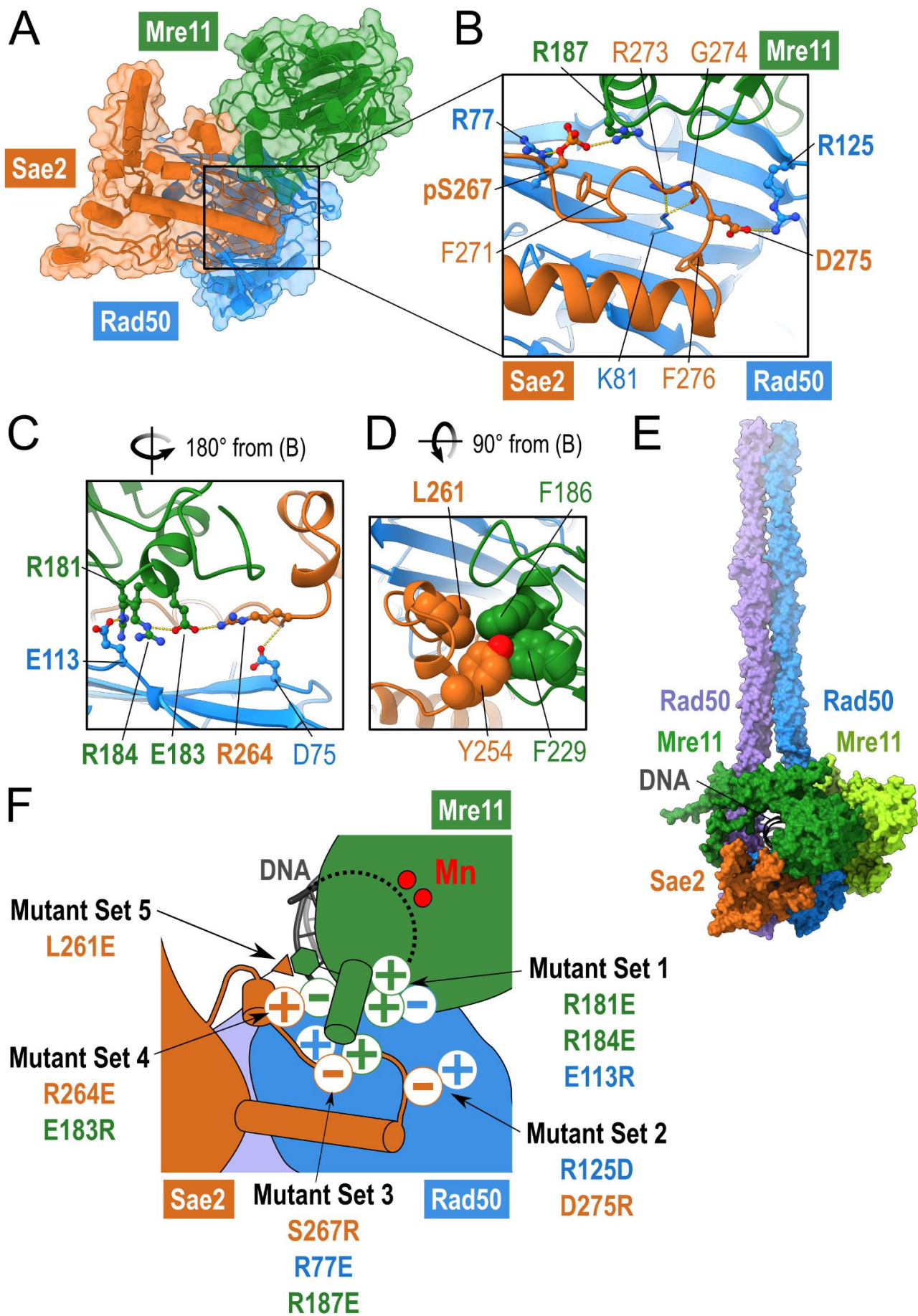
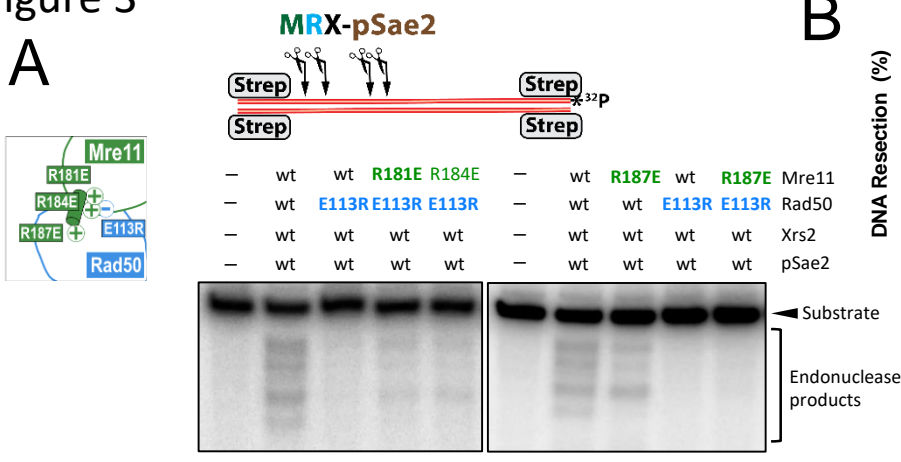
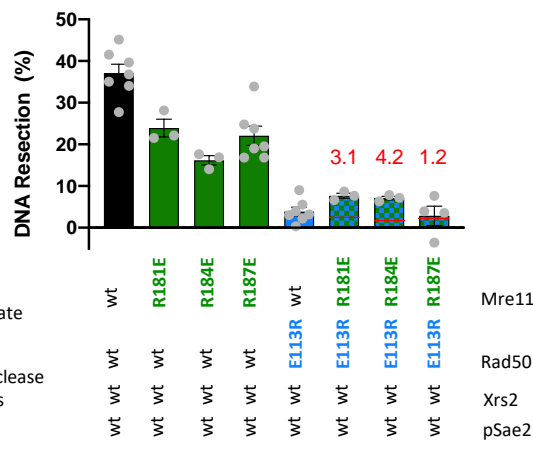


Figure 3

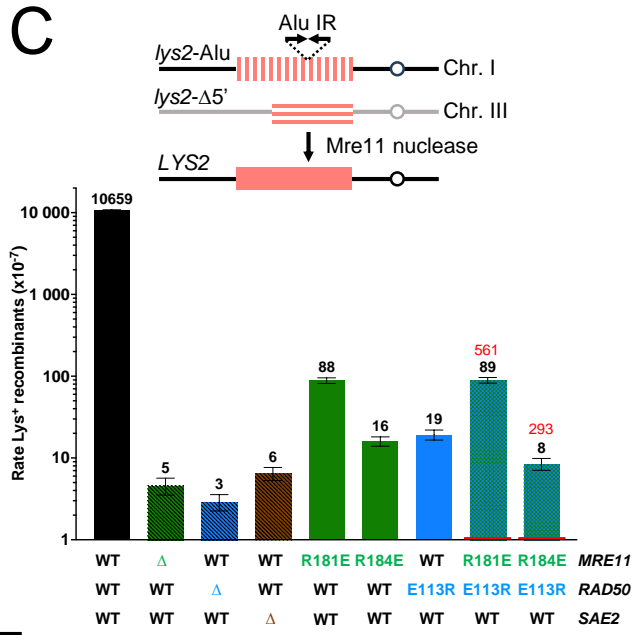
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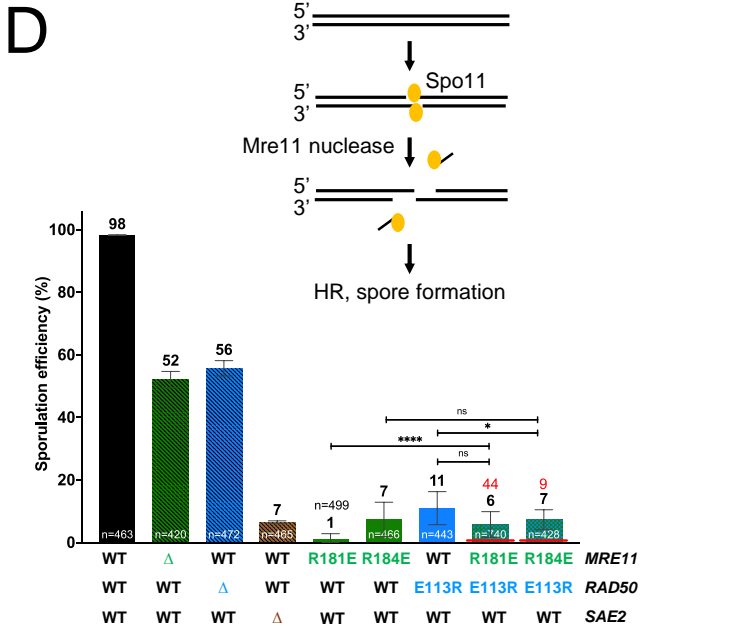
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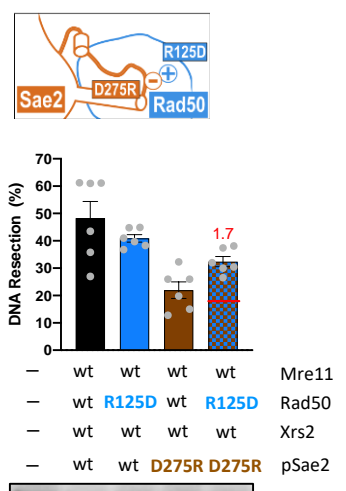
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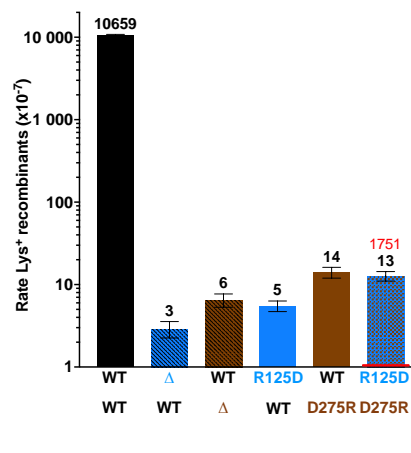
D



E



F



G

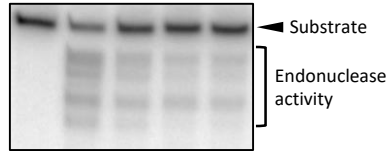
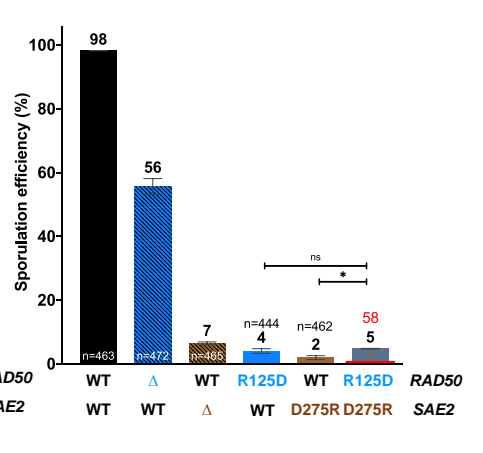
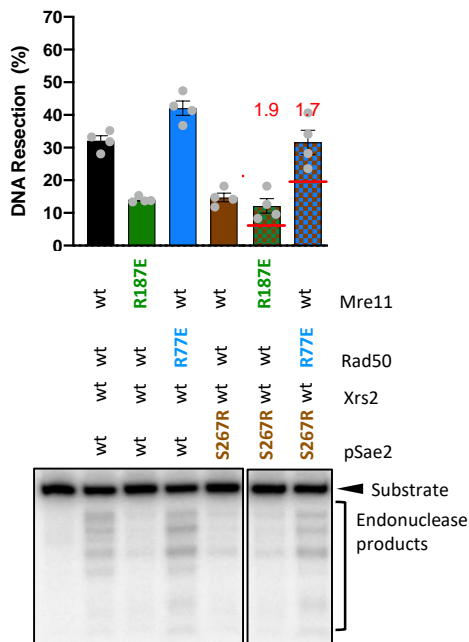
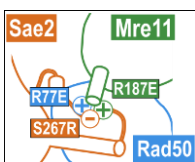
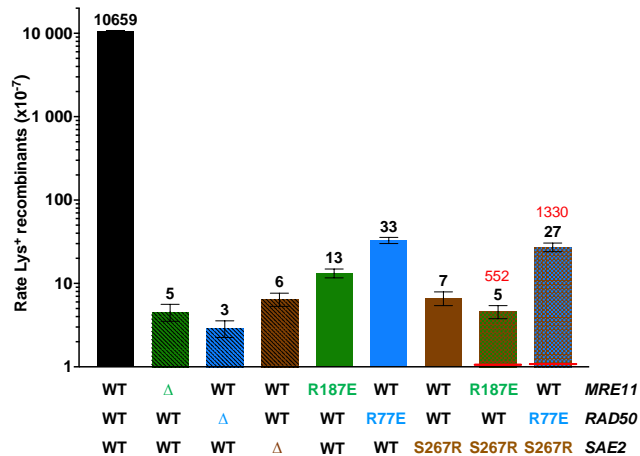


Figure 4

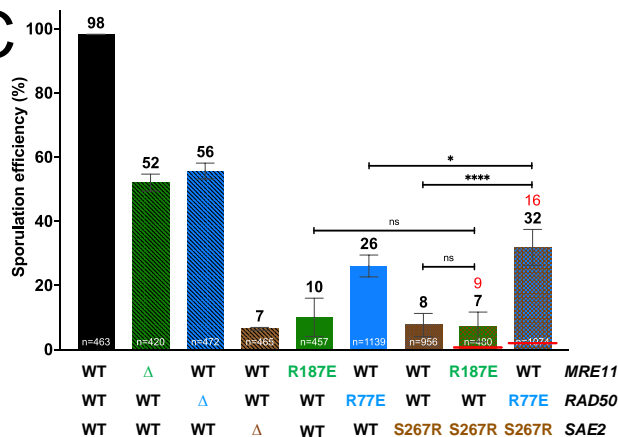
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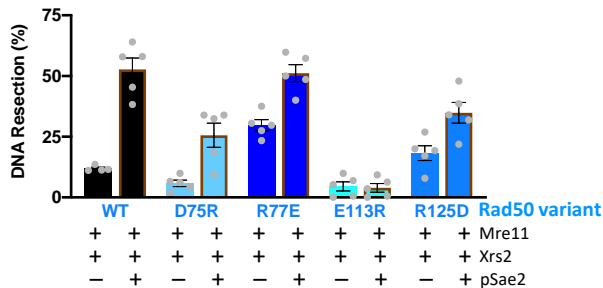
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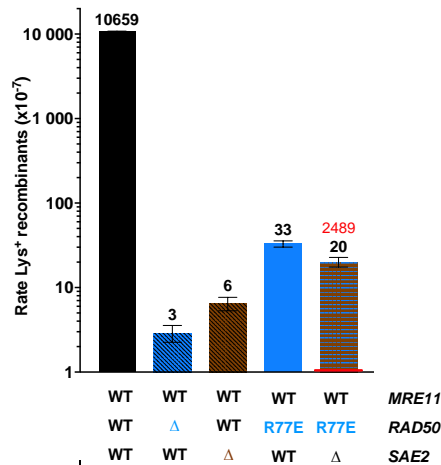
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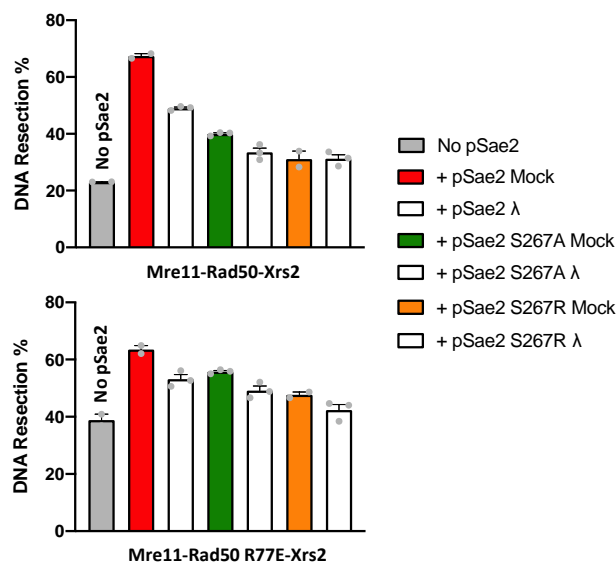
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F



E



G

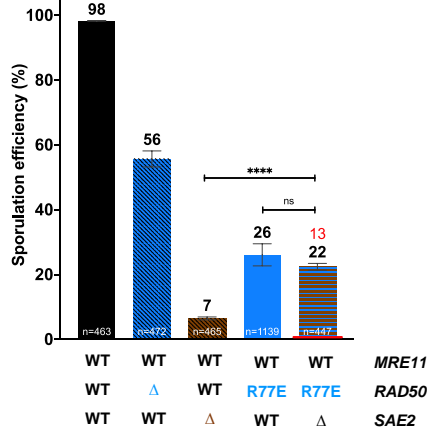
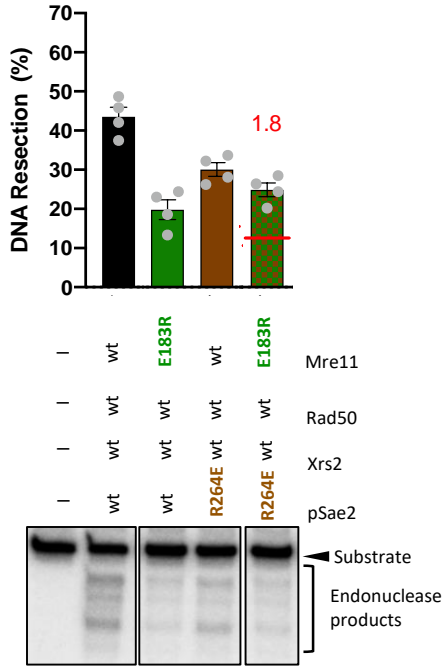
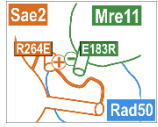
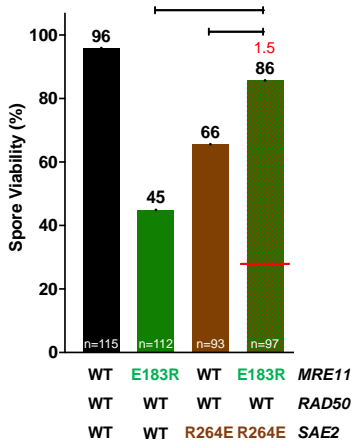


Figure 5

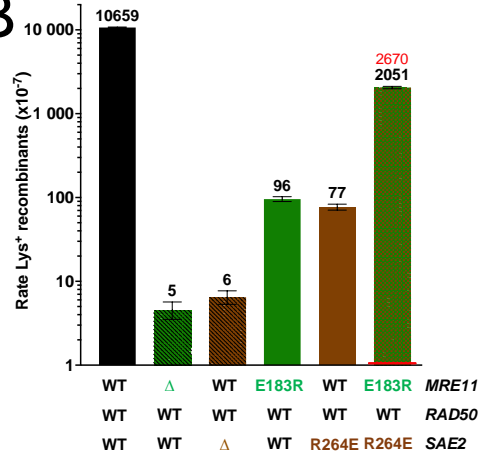
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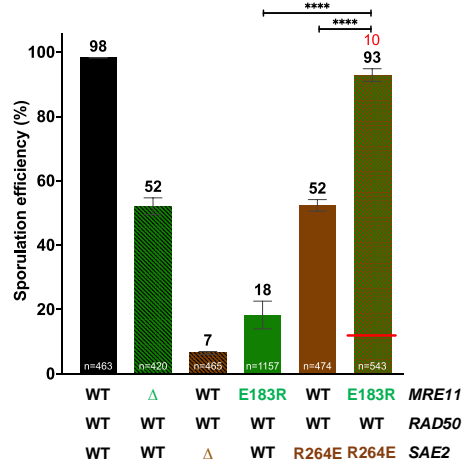
D



B



C



E

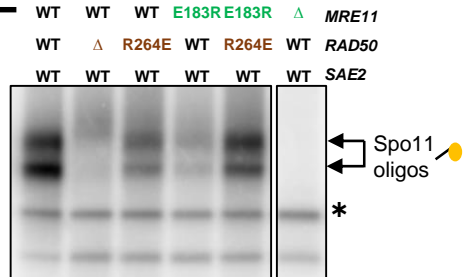


Figure 6

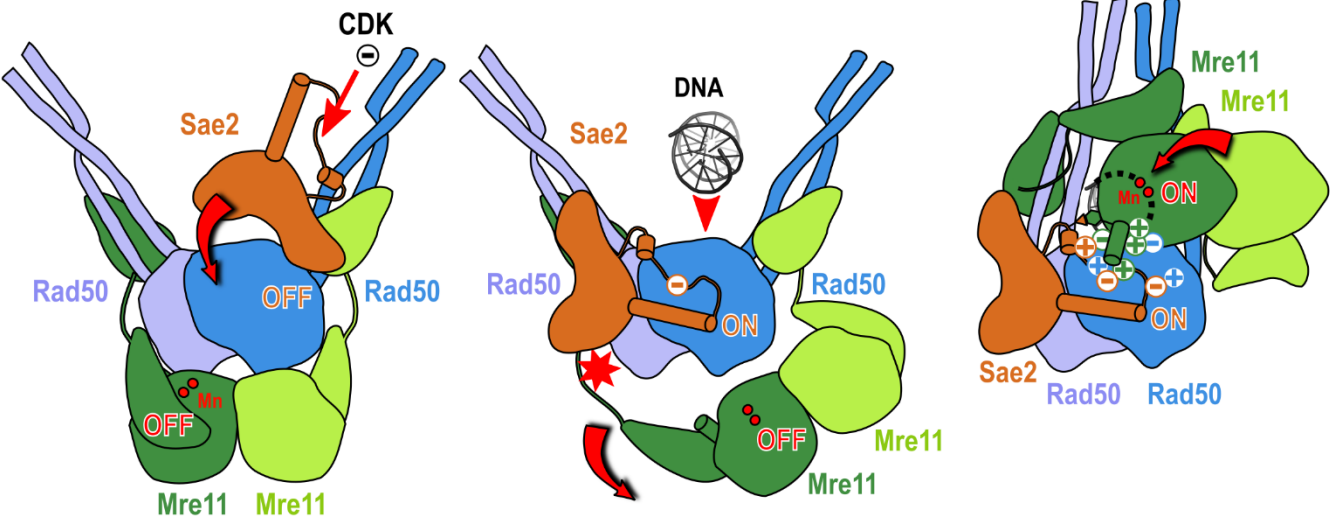
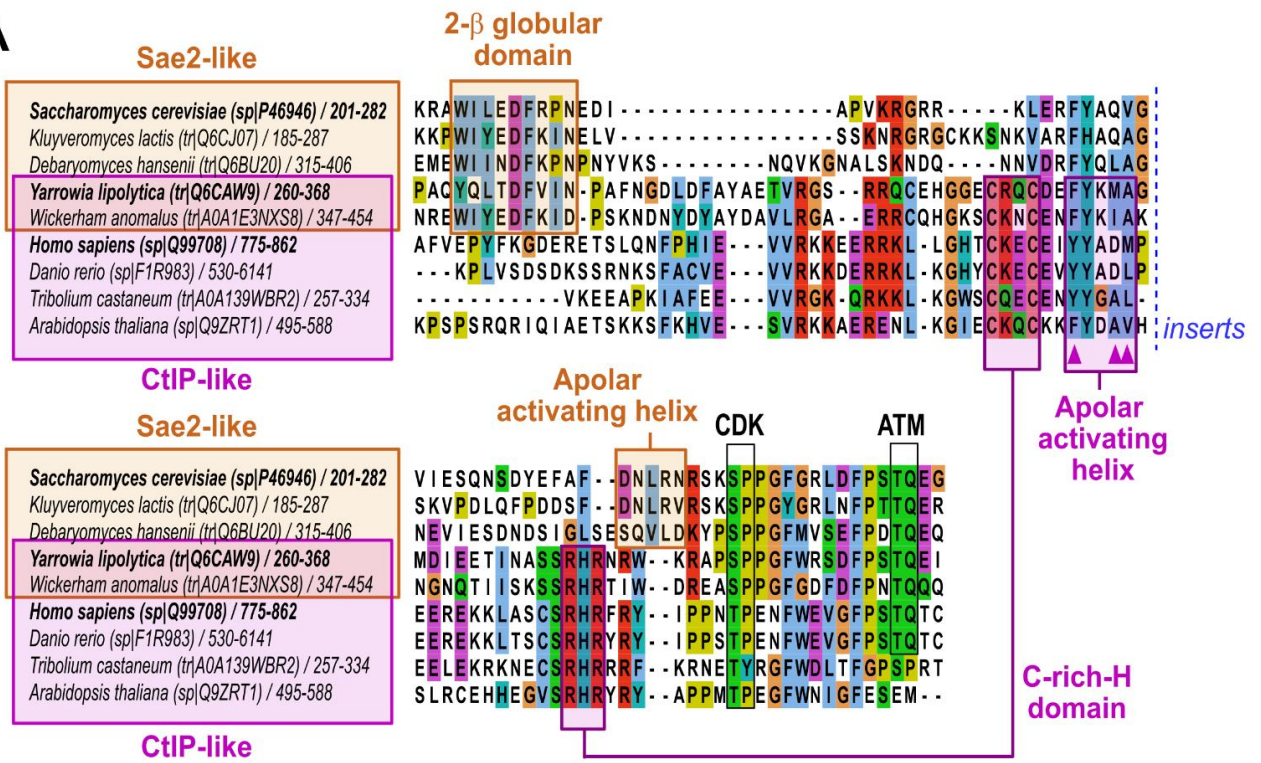


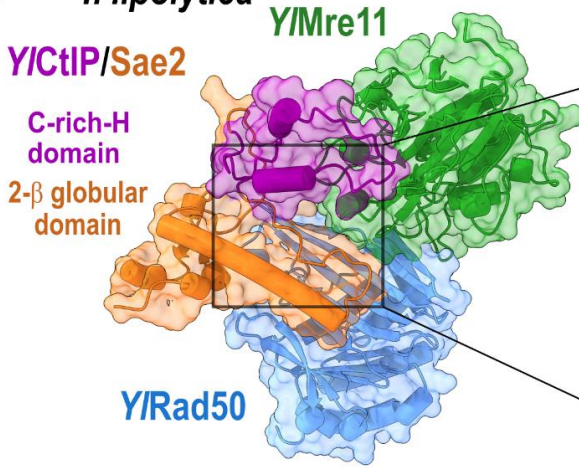
Figure 7

A

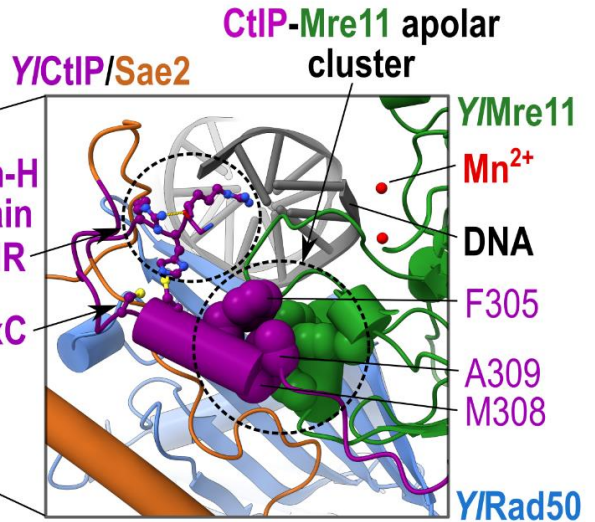


B

Y. lipolytica

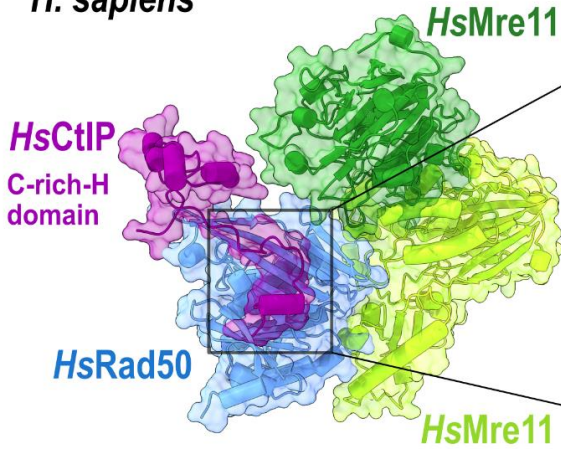


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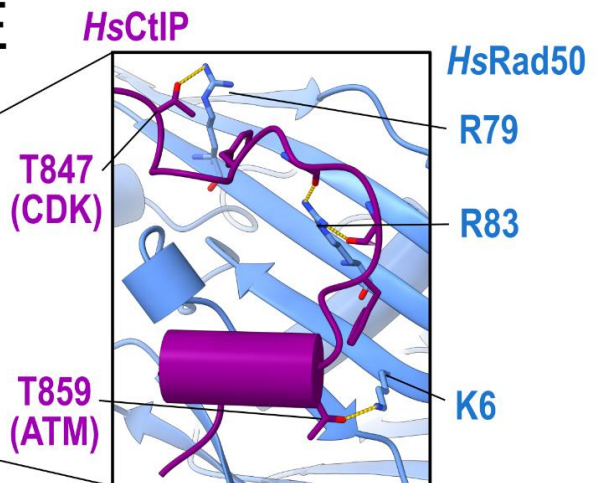


D

H. sapiens



E



7 Supplementary Figures and 5 Supplementary Tables

Legends of the Supplementary Figures are in the main manuscript.

Figure S1: Details on the structural modeling of *S. cerevisiae* Mre11-Rad50 assembly. Related to Figure 1

Figure S2: Details on the structural modeling of *S. cerevisiae* Mre11-Rad50-Sae2 assembly. Related to Figure 2

Figure S3: Purified recombinant proteins used in this study. Related to Figure 3

Figure S4: R77E substitution in Rad50 does not restore spore viability of *sae2S267R* or *sae2Δ*. Related to Figure 4

Figure S5: The small hydrophobic cluster between Sae2 and Mre11 favors Mre11 nuclease. Related to Figure 5

Figure S6: Models illustrating the molecular events occurring the transition between the inactive and active conformations. Related to Figure 6

Figure S7: Details on the structural modeling of *Y. lipolytica* and *H. sapiens* Mre11-Rad50-Sae2 assembly. Related to Figure 7

Table S1: Ten top hits from the DALI server analysis.

Table S2: Description of conditions used to generate models.

Table S3: Summary of evaluation scores for models generated by AlphaFold2.

Table S4: Links to the ModelArchive database containing model coordinates, multiple sequence alignments and evaluation scores.

Table S5: Yeast strains used in this study.

Figure S1

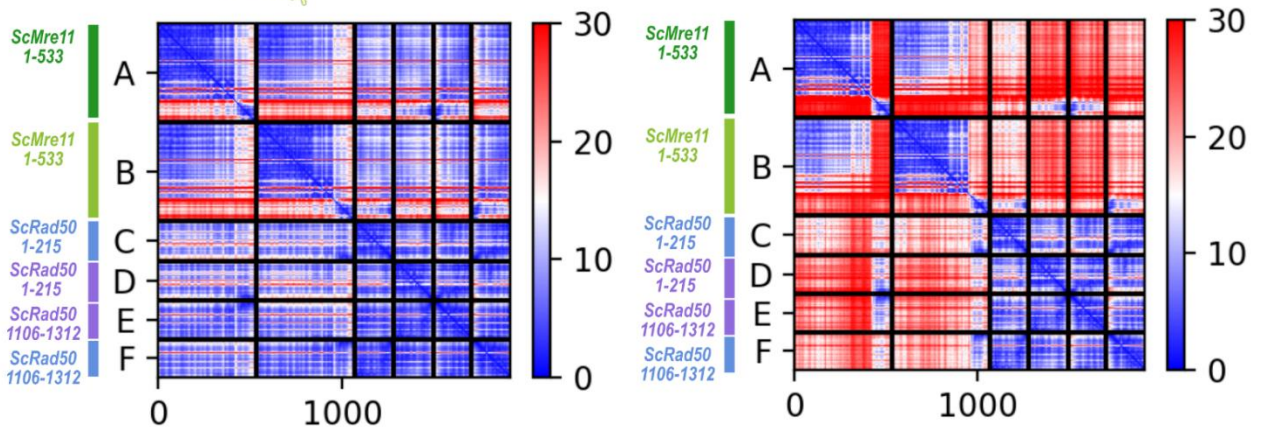
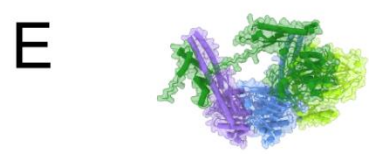
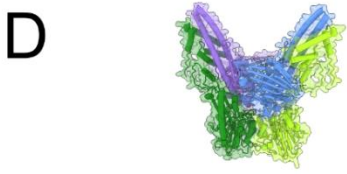
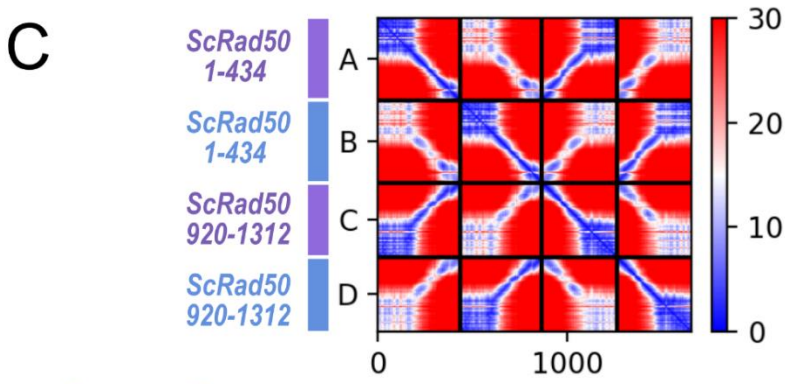
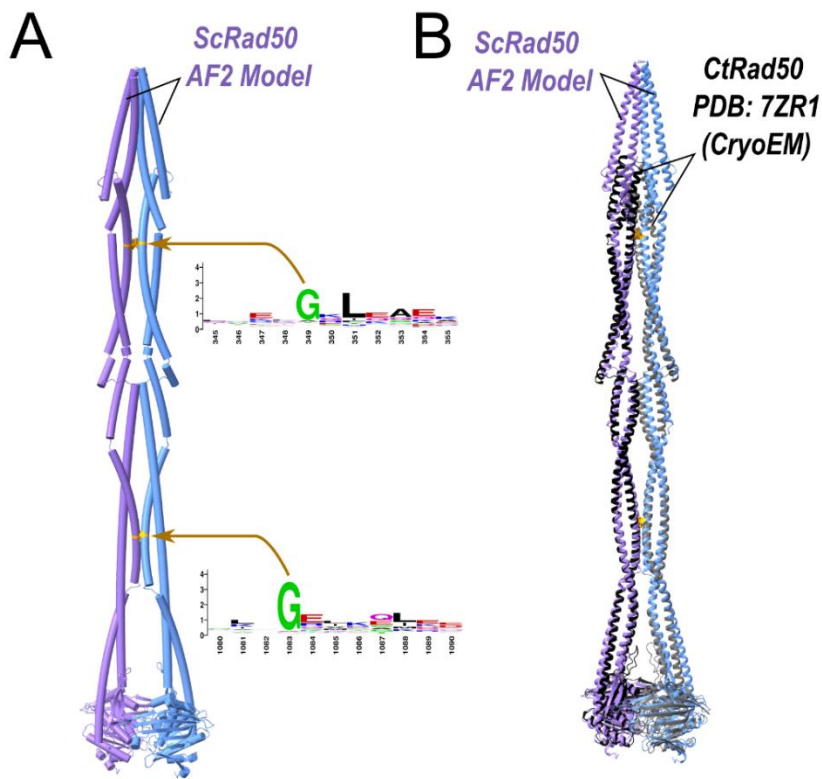


Figure S2

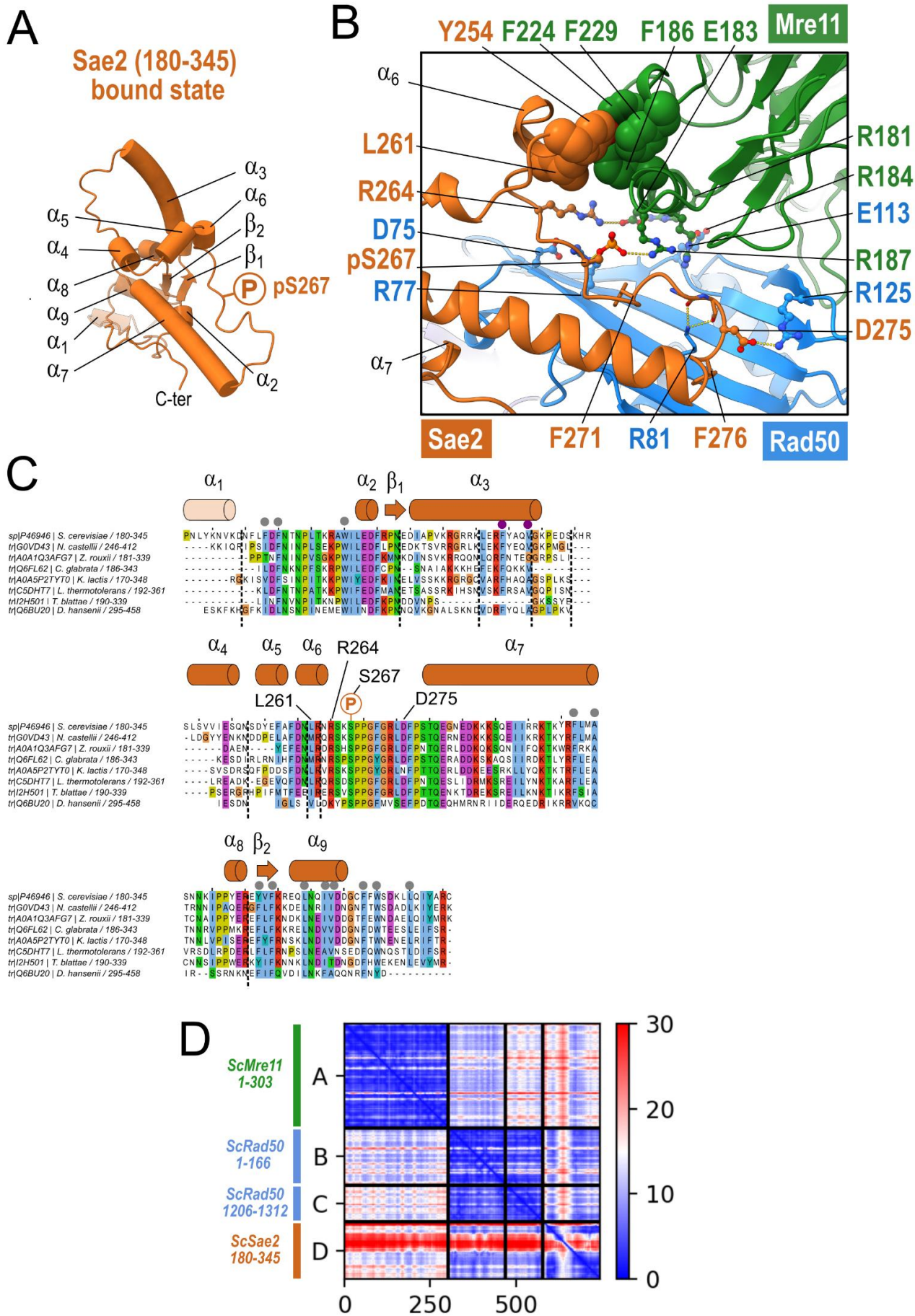


Figure S3

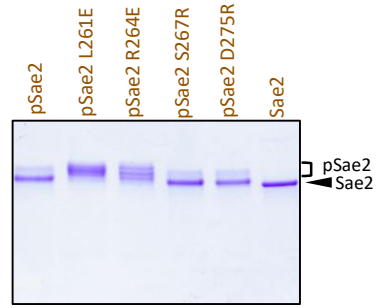
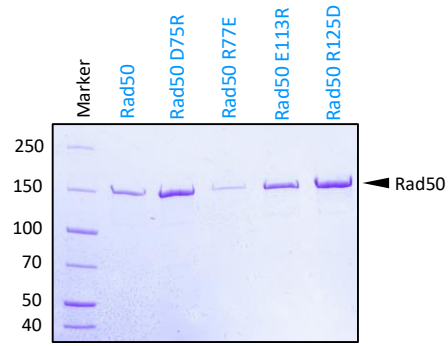
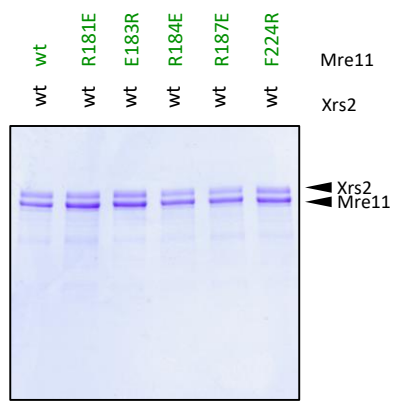


Figure S4

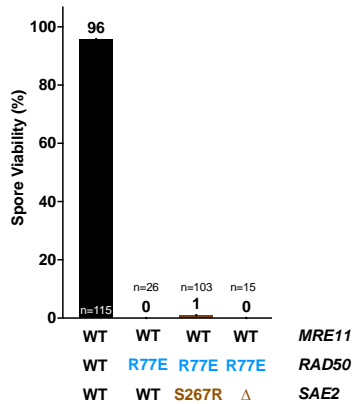


Figure S5

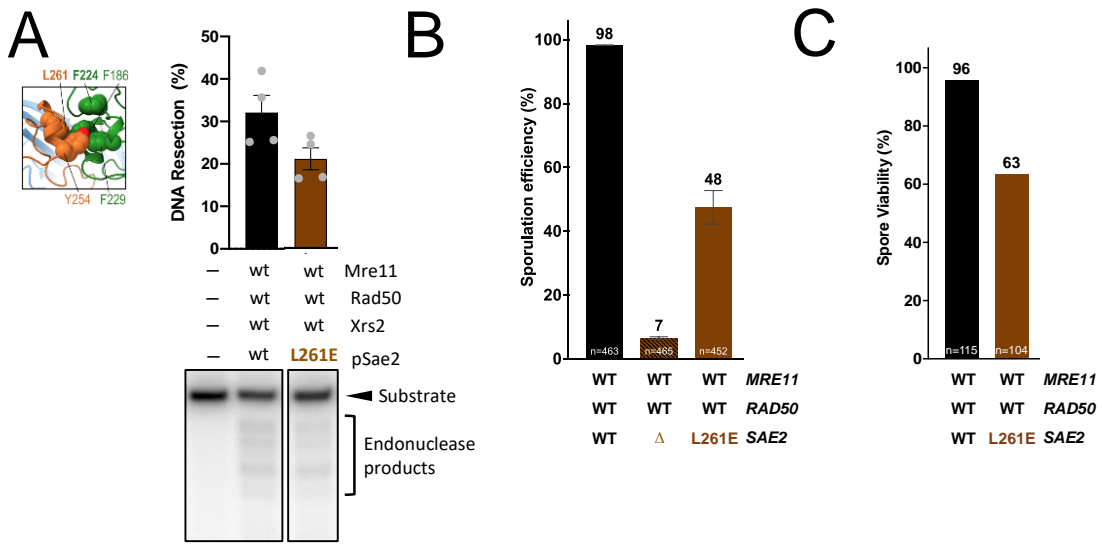


Figure S6

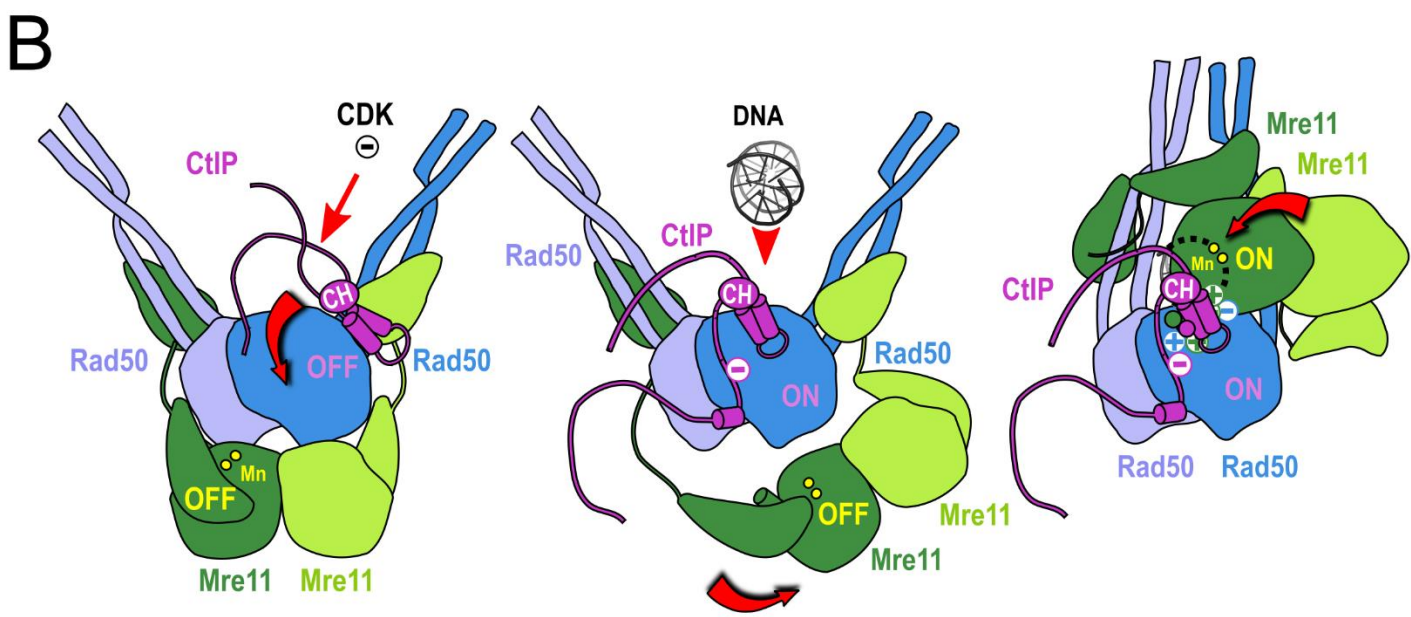
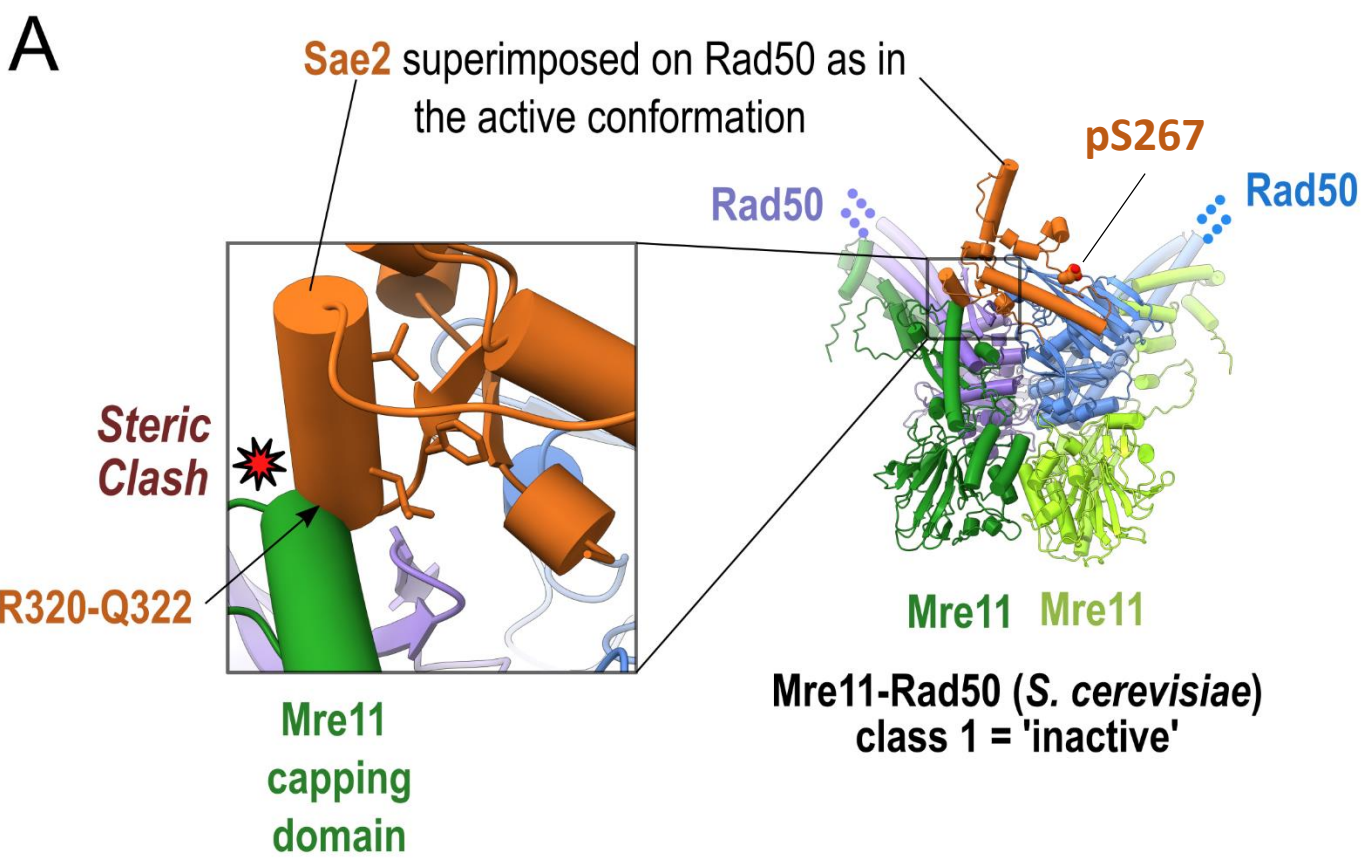


Figure S7

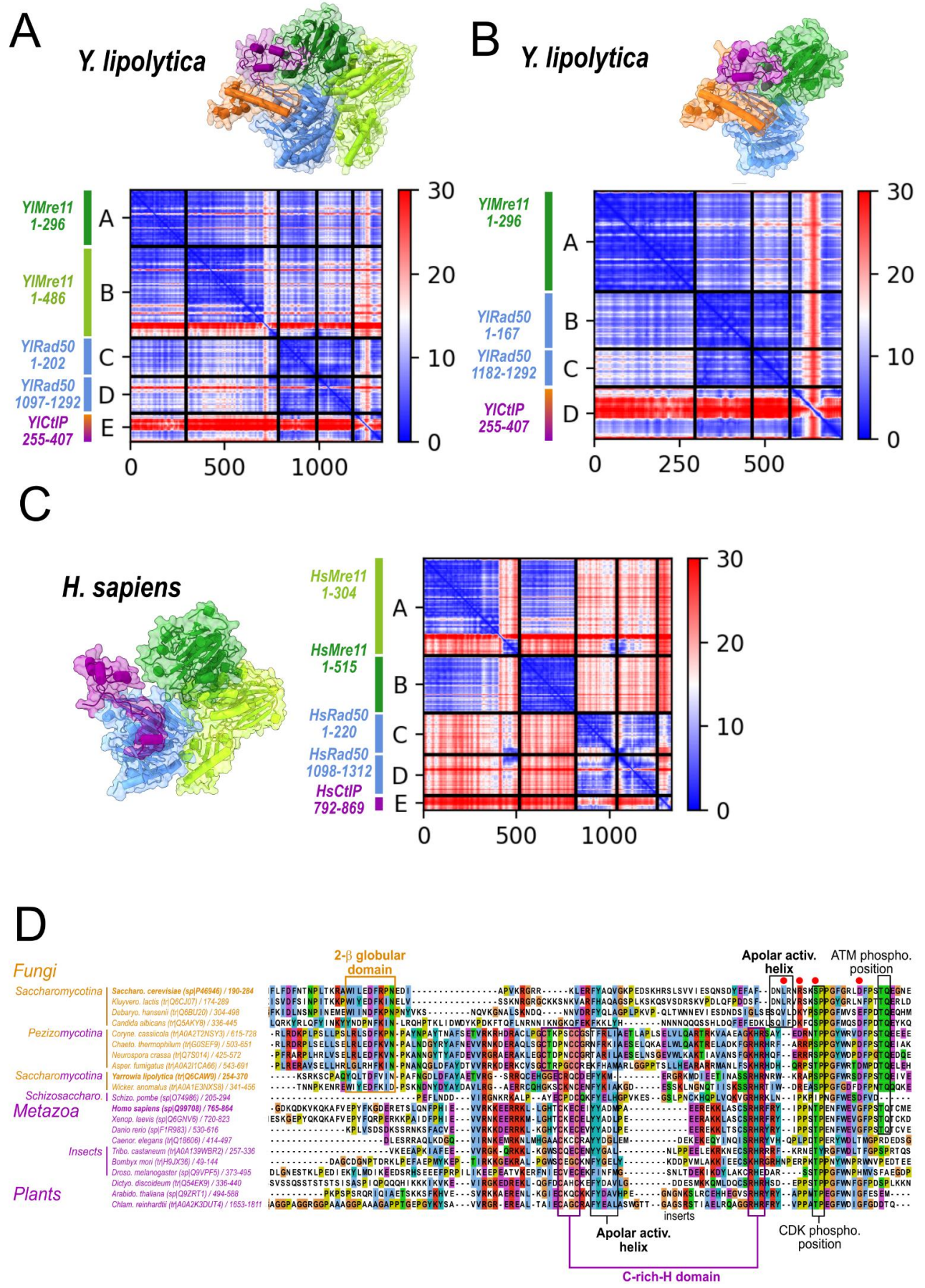


Table S1: Top ten hits retrieved from the DALI server analysis of PDB structures potentially similar to the C-terminal domain of ScSae2. The first hit with a Z-score of 3.1 aligned with a fraction of the fold (mainly the long α 7 helix) without significant resemblance to the structure of the ScSae2 bound domain.

No:	Chain	Z	rmsd	lali	nres	%id	PDB	Description
1:	5l1m-C	3.1	3.9	72	90	1	PDB	MOLECULE: TRANSCRIPTIONAL REPRESSOR FRMR;
2:	4uig-A	2.7	4.6	70	91	6	PDB	MOLECULE: COPPER SENSITIVE OPERON REPRESSOR;
3:	2pms-C	2.7	4.8	77	109	9	PDB	MOLECULE: LACTOTRANSFERRIN;
4:	5we2-A	2.5	6.1	90	229	11	PDB	MOLECULE: PROTECTION OF TELOMERES PROTEIN POZ1;
5:	4h8s-C	2.5	4.9	100	382	6	PDB	MOLECULE: DCC-INTERACTING PROTEIN 13-BETA;
6:	6ezn-B	2.4	3.9	80	110	5	PDB	MOLECULE: DOLICHYL-DIPHOSPHOLIGOSACCHARIDE--PROTEIN
7:	6d03-E	2.4	4.4	85	466	12	PDB	MOLECULE: TRANSFERRIN RECEPTOR PROTEIN 1;
8:	3k6q-A	2.3	3.7	60	139	15	PDB	MOLECULE: PUTATIVE LIGAND BINDING PROTEIN;
9:	1x04-A	2.3	4.5	90	200	10	PDB	MOLECULE: SH3-CONTAINING GRB2-LIKE PROTEIN 2;
10:	5ihf-B	2.3	4.2	72	109	14	PDB	MOLECULE: VIRG-LIKE PROTEIN;

Table S2: Description of conditions used to generate models.

Index	Model name	Species	Subunits	Uniprot	Start	Stop	Size	Nb sequences
1	ScR2M2S composite	<i>S. cerevisiae</i>	ScRAD50	P12753	1	363	2654	-
			ScRAD50	P12753	965	1312		
			ScRAD50	P12753	1	363		
			ScRAD50	P12753	965	1312		
			ScMRE11	P32829	1	533		
			ScMRE11	P32829	1	533		
			ScSAE2	P46946	180	345		
2	ScR2medium	<i>S. cerevisiae</i>	ScRAD50	P12753	1	363	1422	p=1727
			ScRAD50	P12753	965	1312		
			ScRAD50	P12753	1	363		
			ScRAD50	P12753	965	1312		
3	ScR2long	<i>S. cerevisiae</i>	ScRAD50	P12753	1	434	1654	p=1727
			ScRAD50	P12753	920	1312		
			ScRAD50	P12753	1	434		
			ScRAD50	P12753	920	1312		
4	ScM2R2on	<i>S. cerevisiae</i>	ScRAD50	P12753	1	215	1910	p=1625,u=413
			ScRAD50	P12753	1106	1312		
			ScRAD50	P12753	1	215		
			ScRAD50	P12753	1106	1312		
			ScMRE11	P32829	1	533		
			ScMRE11	P32829	1	533		
5	ScM2R2off	<i>S. cerevisiae</i>	ScRAD50	P12753	1	215	1910	p=1625,u=413
			ScRAD50	P12753	1106	1312		
			ScRAD50	P12753	1	215		
			ScRAD50	P12753	1106	1312		
			ScMRE11	P32829	1	533		
			ScMRE11	P32829	1	533		
6	ScMRS	<i>S. cerevisiae</i>	ScRAD50	P12753	1	166	742	p=1055,u=1038
			ScRAD50	P12753	1206	1312		
			ScMRE11	P32829	1	303		
			ScSAE2	P46946	180	345		
7	YIMRC	<i>Y. lipolytica</i>	YIRAD50	Q6C910	1	167	727	p=494,u=2023
			YIRAD50	Q6C910	1182	1292		
			YIMRE11	Q6CEM3	1	296		
			YICtIP	Q6CAW9	255	407		
8	HsMRC	<i>H. sapiens</i>	HsRAD50	Q92878	1	220	1332	p=491,u=1879
			HsRAD50	Q92878	1098	1312		
			HsMRE11	P49959	1	304		
			HsMRE11	P49959	1	515		
			HsCtIP	Q99708	792	869		

Table S3: Summary of evaluation scores for the model generated with the highest confidence score for all the models generated by AlphaFold2.

Index	Model name	Species	pLDDT	pTMscore	ipTMscore	confidence_score	PAE
2	ScR2medium	<i>S. cerevisiae</i>	79.8	0.54	0.51	0.52	
3	ScR2long	<i>S. cerevisiae</i>	81.2	0.48	0.46	0.46	Fig. S1C
4	ScM2R2on	<i>S. cerevisiae</i>	81.2	0.68	0.65	0.66	Fig. S1E
5	ScM2R2off	<i>S. cerevisiae</i>	85.5	0.87	0.86	0.86	Fig. S1D
6	ScMRS	<i>S. cerevisiae</i>	84.7	0.75	0.69	0.70	Fig. S2D
7	YIMRC	<i>Y. lipolytica</i>	83.5	0.8	0.76	0.77	Fig. S7B
8	HsMRC	<i>H. sapiens</i>	83.7	0.71	0.63	0.65	Fig. S7C

Table S4: Links to the ModelArchive database containing model coordinates, multiple sequence alignments and evaluation scores.

Index	Model name	ModelArchive
1	ScR2M2S composite	https://modelarchive.org/doi/10.5452/ma-jf8hv
2	ScR2medium	https://www.modelarchive.org/doi/10.5452/ma-i9sv6
3	ScR2long	https://modelarchive.org/doi/10.5452/ma-vje4m
4	ScM2R2on	https://modelarchive.org/doi/10.5452/ma-jfoh4
5	ScM2R2off	https://modelarchive.org/doi/10.5452/ma-lbblz
6	ScMRS	https://www.modelarchive.org/doi/10.5452/ma-qhmpy
7	YIMRC	https://www.modelarchive.org/doi/10.5452/ma-ks31s
8	HsMRC	https://www.modelarchive.org/doi/10.5452/ma-qtbol

Table S5: Yeast strains used in this study

Diploid strains used for meiotic studies (SK1 background)

VBD2335

Mata/α ura3/" ho::hisG/" leu2::hisG HIS4::LEU2-(BamHI; +ori)/his4-X::LEU2-(BamHI; +ori)—URA3 SPO11-His6-Flag3::NatMX/"

Used in Figures 3D, 3G, 4C, 4G, 5C, 5D, 5E

VBD2338

Same as VBD2335 but *rad50Δ(1-192)::KanMX/"*

Used in Figures 3D, 3G, 4C, 4G

VBD2371

Same as VBD2335 but *mre11Δ::KanMX/"*

Used in Figure 3D

VBD2372

Same as VBD2335 but *sae2Δ::KanMX/"*

Used in Figures 3D, 3G

VBD2339

Same as VBD2335 but *mre11R181E/"*

Used in Figure 3D

VBD2341

Same as VBD2335 but *mre11R184E/"*

Used in Figure 3D

VBD2350

Same as VBD2335 but *rad50E113R/"*

Used in Figure 3D

VBD2362

Same as VBD2335 but *mre11R181E/" rad50E113R/"*

Used in Figure 3D

VBD2364

Same as VBD2335 but *mre11R184E/*" *rad50E113R/*"
Used in Figure 3D

VBD2347
Same as VBD2335 but *sae2D75R/*"
Used in Figure 3G

VBD2351
Same as VBD2335 but *rad50R125D/*"
Used in Figure 3G

VBD2369
Same as VBD2335 but *rad50R125D/*" *sae2D75R/*"
Used in Figure 3G

VBD2342
Same as VBD2335 but *mre11R187E/*"
Used in Figure 4C

VBD2346
Same as VBD2335 but *sae2S267R/*"
Used in Figure 4C

VBD2349
Same as VBD2335 but *rad50R77E/*"
Used in Figure 4C, 4G

VBD2365
Same as VBD2335 but *mre11R187E/*" *sae2S267R/*"
Used in Figure 4C

VBD2368
Same as VBD2335 but *rad50R77E/*" *sae2S267R/*"
Used in Figure 4C

VBD2370
Same as VBD2335 but *rad50R77E/*" *sae2Δ::KanMX/*"

Used in Figure 4G

VBD2340

Same as VBD2335 but *mre11E183R*/'

Used in Figures 5C, 5D, 5E

VBD2345

Same as VBD2335 but *sae2R264E*/'

Used in Figures 5C, 5D, 5E

VBD2363

Same as VBD2335 but *mre11E183R*/' *sae2R264E*/'

Used in Figures 5C, 5D, 5E

VBD2344

Same as VBD2335 but *sae2L261E*/'

Used in Figure Supp Figure 5

Haploid strains used for hairpin cleavage assays (CGL background)

VBH3709 (HS21 strain, from K. Lobachev, (Lobachev et al., 2000))

MATa ade5-1 his7-2 leu2-3,112:: p305L3 (LEU2) trp1-289 ura3-D lys2::AluIR

Used in Figures 3C, 3F, 4B, 4F, 5B

VBH3713 (ALE104 strain, from K. Lobachev)

Same as VBH3709 but *rad50Δ::HphMX*

Used in Figures 3C, 3F, 4B, 4F

VBH3711 (ALE102 strain, from K. Lobachev)

Same as VBH3709 but *mre11Δ::HphMX*

Used in Figure 3C

VBH3716 (ALE109 strain, from K. Lobachev)

Same as VBH3709 but *sae2Δ::HphMX*

Used in Figures 3C, 3F

VBH3723

Same as VBH3709 but *mre11R181E*
Used in Figure 3C

VBH3725
Same as VBH3709 but *mre11R184E*
Used in Figure 3C

VBH3823
Same as VBH3709 but *rad50E113R*
Used in Figure 3C

VBH3826
Same as VBH3709 but *mre11R181E rad50E113R*
Used in Figure 3C

VBH3827
Same as VBH3709 but *mre11R184E rad50E113R*
Used in Figure 3C

VBH3801
Same as VBH3709 but *sae2D275R*
Used in Figure 3F

VBH3824
Same as VBH3709 but *rad50R125D*
Used in Figure 3F

VBH3830
Same as VBH3709 but *sae2D275R rad50R125D*
Used in Figure 3F

VBH3726
Same as VBH3709 but *mre11R187E*
Used in Figure 4B

VBH3799
Same as VBH3709 but *sae2S267R*

Used in Figure 4B

VBH3822

Same as VBH3709 but *rad50R77E*

Used in Figures 4B, 4F

VBH3933

Same as VBH3709 but *mre11R187E sae2S267R*

Used in Figure 4B

VBH3829

Same as VBH3709 but *sae2S267R rad50R77E*

Used in Figure 4B

VBH3825

Same as VBH3709 but *sae2Δ rad50R77E*

Used in Figure 4F

VBH3724

Same as VBH3709 but *mre11E183R*

Used in Figure 5B

VBH3798

Same as VBH3709 but *sae2R264E*

Used in Figure 5B

VBH3803

Same as VBH3709 but *mre11E183R sae2R264E*

Used in Figure 5B

VBH3798

Same as VBH3709 but *sae2L261E*

Used in Supp Figure 5

Bibliography

Lobachev, K.S., Stenger, J.E., Kozyreva, O.G., Jurka, J., Gordenin, D.A., and Resnick, M.A. (2000). Inverted Alu repeats unstable in yeast are excluded from the human genome. *EMBO J* 19, 3822-3830.