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Travelling with a parasite: the evolution of resistance and dispersal syndromes during experimental range expansion

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1 **Travelling with a parasite: the evolution of resistance and**
2 **dispersal syndromes during experimental range**
3 **expansion**

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21

22 **Abstract**

23 Rapid evolutionary changes during range expansions can lead to the divergence between range core and
24 front populations, with the emergence of dispersal syndromes. Besides intraspecific effects, range
25 expansions may be impacted by interspecific interactions such as parasitism. Yet, despite the potentially large
26 impact of parasites imposing additional selective pressures on the host, their role on range expansions
27 remains largely unexplored. Here, we investigated whether parasites affect the evolution of host dispersal
28 syndromes during spatial spread. Using populations of the ciliate *Paramecium caudatum* and its bacterial
29 parasite *Holospira undulata* spreading in microcosm landscapes, we studied experimental range expansions
30 under the presence or absence of the parasite. We found that range expansion and parasite treatments acted
31 independently on the evolution of host dispersal syndromes. Front host populations showed different trait
32 associations between swimming behaviour and population growth parameters than core populations,
33 indicating divergent evolution. The presence of parasites reshaped these trait associations and divergence
34 between front and core, with hosts exposed to the parasite exhibiting overall reduced dispersal, but
35 increased resistance. Our work shows that host-parasite interactions during range expansions can shift the
36 contenders to novel evolutionary trajectories and may influence the speed of the range expansion itself and
37 of spreading epidemics.

38

39 **Keywords**

40 Dispersal syndromes, eco-evolutionary dynamics, host-parasite interactions, invasion front, range expansion,
41 resistance

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49 Introduction

50 Species range expansions are increasing in frequency in response to rapidly changing environments [1] and
51 becoming crucial to reducing the risk of extinction [2]. While ecological and evolutionary processes have been
52 well explored for single species, we are still lacking a good community perspective of range shifts [3,4].
53 Species might rarely "travel alone", and it is conceivable that interactions with other organisms facilitate or
54 slow down spatial spread, and add new selection pressures while a species is spreading.

55 Antagonistic species interactions have been suggested a major factor defining species ranges [5,6].
56 Parasitism, for example, are ubiquitous and exert strong demographic and evolutionary pressures on the
57 host, which can limit geographic ranges as has been shown in some theoretical models [7–9]. In fact, parasites
58 may re-enforce range limits by reducing dispersal and/or by imposing additional mortality in already small
59 populations at the front. This might be the case in the cane toad [10,11] and in other natural systems [12–
60 14]. Adaptation to parasite attack may remove such range limits, but potentially also affect the evolution of
61 other traits known to be selected at range fronts. This could produce eco-evolutionary feedbacks that will
62 not only influence rates of spatial spread of the host species, but also impact parasite propagation in the
63 target species, or spill-over of disease to other species along the way [15,16]. Further, parasites can cause
64 knock-on effects on interactions with other species in the community [17,18] and affect ecosystem
65 functioning [19].

66 Concomitant with antagonistic species interactions, range expansions often involve stochastic genetic
67 changes in small vanguard front populations (via drift or founder bias), but also targeted evolution of
68 dispersal due to spatial selection, spatial assortative mating and kin competition [20,21]. Dispersal evolution
69 likely entails concerted changes in life-history, behavioural or physiological traits, leading to so-called
70 dispersal syndromes [22,23]. Thus, the underlying genetic architecture and covariation among these traits
71 will be critical to responses to selection at range fronts. Indeed, it has been shown for various biological
72 systems that dispersal is a heritable trait and that covariation with other traits has a genetic basis [24], making
73 it possible for selection to drive the divergence between core and front populations [25,26]. This is illustrated
74 by the introduced cane toad (*Rhinella marina*) in Australia, where the evolution of dispersal and other life-
75 history traits promoted the speed of spread at the invasion front [27–29]. Similar results were found for the
76 invasive ladybird *Harmonia axyridis* [30], and in laboratory range expansions using experimental evolution
77 approaches [31–33].

78 We currently know relatively little about parasite-mediated selection at range fronts and its consequences
79 for dispersal syndrome evolution [3,4]. While a considerable bulk of literature exists on resistance evolution
80 in host-parasite metapopulations [34,35], there is no specific theory in the context of range expansion.

81 Intuitively, we may expect that virulent parasites select for increased resistance and that this removes a
82 parasite-imposed range speed limit. Such a case of resistance evolution has been found for populations of
83 the cane toad infested by lung-worms at the range margin [36]. Theory further suggests that natural enemies
84 select for changes in host dispersal. For example, parasite-induced fluctuations in host population dynamics
85 can modify environmental predictability and fitness expectations, and consequently favour increased host
86 dispersal [37,38]. Parasites may also select for plastic responses in dispersal, with sometimes counter-
87 intuitive consequences for the spread of infection [39].

88 However, few if any studies have considered the joint action of parasite-mediated selection and spatial
89 selection, and hence the interplay between the evolution of resistance and dispersal-related traits. Selection
90 for resistance can be rapid and strong, but also modify genetic correlations with other life-history traits
91 [40,41], often involving trade-offs [42,43] that constrain evolutionary trajectories. Such constraints may also
92 concern dispersal syndromes. Our hypothesis is that dispersal syndrome might be different in the presence
93 of the parasite, especially if there are trade-offs between dispersal and resistance.

94 Using experimental evolution, we investigated the effect of a parasite on the emergence of dispersal
95 syndromes. In a long-term experiment, we mimicked range expansions in interconnected microcosms of the
96 freshwater protist *Paramecium caudatum* and its bacterial parasite *Holospora undulata*. In a range front
97 treatment, the paramecia were constantly selected to disperse into a new microcosm, whereas in the range
98 core treatment, only the non-dispersing fraction of the population was maintained. Range and core
99 treatments were established for infected and uninfected populations. Evolved hosts were assayed for six
100 traits, including dispersal, resistance, population growth characteristics and swimming behaviour. We found
101 substantial divergence in multiple phenotypic trait space, attributable to the combined additive effect of our
102 experimental treatments. A dispersal syndrome emerged, with higher dispersal, equilibrium density and
103 movement tortuosity in the front treatment, and higher growth rate and swimming speed in the core
104 treatment. Parasite-mediated selection affected these trait relationships. Namely, front populations evolving
105 with the parasite were more resistant, but also dispersed less than their parasite-free counterparts. The
106 parasite-driven evolution of such novel characteristics may influence the speed of range expansions, but also
107 the risk of spreading epidemics, which can have important implications for biological control, conservation
108 and management decisions.

109 **Material and methods**

110 **Study system**

111 *Paramecium caudatum* is a freshwater filter-feeding ciliate with a world-wide distribution [44]. Ciliates
112 typically exhibit a nuclear dimorphism: The “germ-line” micronucleus is active during the sexual stage, while

113 the highly polyploid “somatic” macronucleus regulates gene expression during the asexual stage, when
114 replication occurs through mitotic division. In this experiment, clonal populations were maintained asexually
115 (max. 1-2 population doublings per day at constant 23°C) in 50 mL tubes, using a sterilised lettuce medium
116 (1g dry weight of organic lettuce per 1.5l of Volvic™ mineral water), supplemented *ad libitum* with the
117 bacterium *Serratia marcescens* as a food resource (referred to as “bacterised medium” or “medium”,
118 hereafter). The gram-negative bacterium *Holospira undulata* is an obligate parasite, infecting the
119 micronucleus of *P. caudatum* [45]. The infection life cycle comprises both horizontal and vertical
120 transmission. *Paramecium* ingest infectious forms from the aquatic environment, which subsequently
121 colonise the micronucleus and differentiate into multiplying reproductive forms; these reproductive forms
122 are vertically transmitted to the daughter cells of mitotically dividing hosts. The infection life cycle is
123 completed when reproductive forms differentiate into infectious forms, which are then released during host
124 cell division or upon host death. Infection reduces cell division and survival of the *Paramecium*, as well as
125 dispersal [46–48]. Experimental evolution of resistance to this parasite was demonstrated in previous long-
126 term experiments and can come at reproductive costs [49,50].

127 **Experimental protocols**

128 **(i) Long-term range expansion experiment**

129 Dispersal in two-patch systems

130 We used two-patch systems for this selection experiment (Supplementary Information, Fig. S1). The systems
131 were built from two 14 mL plastic tubes (“core patch” and “front patch”) interconnected by 5-cm silicon
132 tubing (0.6 mm inner diameter) serving as a corridor through which the *Paramecium* can actively swim. We
133 define dispersal as the active displacement of *P. caudatum* from the core patch to the front patch. In the
134 long-term experiment, short episodes of dispersal (3h) alternated with periods of population growth and
135 maintenance (1 week). For a dispersal episode, we filled the two-patch system with 9.5 mL of fresh bacterised
136 medium and then blocked the corridor with a clamp. The core patch was filled with c. 8ml of culture
137 containing *Paramecium* (\approx 2000 individuals), whereas the front patch was only topped up with non-
138 bacterised medium and was thus “empty”. After removal of the clamp, *Paramecium* could freely disperse to
139 the front patch or to stay in the core. After three hours, we blocked the corridor and estimated the cell
140 density in the core and front patch, by sampling up to 1mL from each tube and counting the number of
141 individuals under a dissecting microscope. The dispersal rate is thus the number of dispersers divided by the
142 total number of individuals in the two-patch systems, divided by 3 hours.

143 Range expansion treatments

144 Two selection treatments were imposed. In the range front treatment, only *Paramecium* that had dispersed
145 into the front patch were maintained and allowed to grow for 1 week until the next episode of dispersal.

146 Conversely, in the core selection treatment, only the non-dispersing *Paramecium* were maintained and
147 allowed to regrow. These contrasting selection protocols were continued for a total of 26 cycles. The front
148 selection treatment mimics the leading front of range expansion or a biological invasion, with populations
149 continuously dispersing into a new microcosm. Populations from the core selection treatment stay in place
150 and continuously lose emigrants. Each new growth cycle was started by placing on average 200 paramecia
151 from front and core selection treatments in 20 mL of fresh bacterised medium, and equilibrium density was
152 then reached within the following 3-4 days. The experiment was conducted with a single host line [47]. This
153 line (63D) had undergone three years of parasite-free core selection prior to the present experiment; initially
154 started from a mix of strains, it has become fixed for a single haplotype [51].

155 Parasite treatment

156 Core and front treatments were established for both infected and uninfected populations. Starting from a
157 63D laboratory culture, infected and uninfected selection lines were established. The parasites were taken
158 from an ongoing experiment [47] that had already been imposing core and front selection on infected 63D
159 populations for about 8 months (30 cycles). Using standard protocols (see S2), we extracted infectious forms
160 of the parasite from 5 core selection lines and from 5 front selection lines, which were then used to inoculate
161 our new, naive 63D hosts. In other words, we continued core and front selection treatments for the parasite,
162 but replaced the previous hosts by new unselected hosts. In addition to these 10 infected selection lines, we
163 established 3 uninfected front-selection lines and 3 uninfected core-selection lines as controls for a total of
164 16 selection lines.

165 **(ii) Phenotypic trait assays**

166 At the end of the long-term experiment, phenotypic trait assays for *Paramecium* from all 16 selection lines
167 were performed under common-garden conditions. Using a micropipette, we arbitrarily picked 4 uninfected
168 paramecia from each selection line and placed them individually in single 1.5 mL Eppendorf tubes filled with
169 bacterised medium, where they were allowed to grow for 2 weeks until small monoclonal lines had
170 established (c. 7-8 asexual generations). Each monoclonal line was then split into three technical replicates
171 and grown for a second common-garden period of 10 days in 50-mL tubes to obtain mass cultures for the
172 phenotype assays (16 selection lines x 4 monoclonal lines x 3 technical replicates = 64 monoclonal lines and
173 192 replicates, Fig. S3). Of the 64 monoclonal lines, 60 were available to measure 6 phenotypic traits, as
174 follows.

175 Resistance

176 To measure resistance, the *Paramecium* were confronted with parasites from core-selection and front-
177 selection lines. We prepared the inocula by mixing the 5 infected core and 5 infected front selection lines,
178 and extracting infectious forms from the two mixes (for details of the extraction protocol, see S2). For

179 inoculation, c. 5000 paramecia were placed in a volume of 25 mL in a 50-mL tube, to which we added $4.5 \times$
180 10^5 infectious forms (core-parasite or front-parasite inoculum). In this way, we set up 4-8 inoculated tubes
181 per host selection line, balanced between the two parasite inocula (16 selection lines \times 2-4 monoclonal lines
182 \times 2 technical replicates = 120 inoculated tubes). Four days post-inoculation, we fixed c. 20 individuals from
183 each inoculated replicate with lacto-aceto-orcein [45] and inspected them for absence or presence of
184 infection using a phase-contrast microscope (1000x magnification). We define resistance as the proportion
185 of uninfected individuals in the sample. Preliminary analysis showed that *Paramecium* from the four different
186 selection treatments did not differ in their resistance to the mixes of front or core parasites ($F_{3,12} = 0.44$, n.s.);
187 we therefore combined the two inoculum sources into a single “infected” category for the main analysis.

188 Dispersal rate

189 Dispersal was measured in linear 3-patch systems (50 mL tubes; Fig. S4), where the *Paramecium* dispersed
190 from the middle tube into the two outer tubes (see SI for detailed protocol). This system configuration
191 allowed us to use bigger volumes of culture and thus obtain higher numbers of dispersers than in 2-patch
192 systems. Connections were opened for 3 h, dispersal rates were then estimated by counting the *Paramecium*
193 in samples from the central tube (500 μ l) and from the combined two outer tubes (3 mL). We employed
194 technical replicates that had not been used for the resistance assay, and were kept in 30 mL of fresh medium
195 for several days prior to the dispersal test. One replicate per monoclonal line was tested (60 dispersal tests).

196 Population growth rate and equilibrium density

197 For the population growth assay, we placed groups of 5 arbitrarily picked *Paramecium* in 15-mL tubes filled
198 with 10 mL of bacterised medium. Over 9 days, we tracked densities in 24-h intervals, estimated from the
199 number of individuals present in 200- μ L samples. We set up 6-12 tubes per host selection line (3 tubes per
200 monoclonal line), with a total of 180 tubes tested. For each tube, estimates of intrinsic population growth
201 rate (r_0) were obtained by fitting a Beverton-Holt population growth model to each density time series, using
202 a Bayesian approach [52], see section S5 for details). For certain tubes we obtained unsatisfactory fits of
203 equilibrium density; we therefore decided to use the mean density over the second half of the assay (day 5-
204 9) as a proxy for equilibrium density (\bar{N}). A small fraction of 19 tubes failed to produce a coherent growth
205 pattern and remained at very low density for unknown reasons; it was not possible to fit our population
206 growth model to these data, and the tubes were therefore excluded from analysis.

207 Swimming speed and tortuosity

208 At the end of the above population growth assay, we analysed swimming behaviour, using an established
209 pipeline of computer vision and automated video analysis to collect this data [53,54]. From a given tube, one
210 sample of 119 μ L was imaged under a Perflex Pro 10 stereomicroscope, using a Perflex SC38800 camera (15
211 frames per second; duration: 10 s; total magnification: 10x). Videos were analysed using the bemovi R-

212 package [54], which provided individual-based data on swimming speed and the tortuosity of swimming
213 trajectories (standard deviation of the turning angle distribution). Details of the script video analysis are given
214 in S6. Swimming speed and tortuosity were averaged over all individuals in a sample prior to analysis. A total
215 of 60 samples (1 per monoclonal line) was used for analysis, giving 2-4 observations per host selection line.

216 **Statistical analysis**

217 All statistical analyses were performed with R v 4.1.2 [55]. Focusing on the analysis of trait associations, we
218 constructed a data matrix with the measurements of the 6 traits for 60 monoclonal lines (for resistance, the
219 mean over the two technical replicates was calculated). To impute 10 missing observations in population
220 growth rate and equilibrium density we used the “missMDA” package version 1.16 [56]. Trait distributions
221 were normalised by picking the best transformation (“bestNormalize” package version 1.4.2, [57]). We then
222 performed a multivariate analysis of variance (MANOVA), with range expansion treatment (core vs front
223 selection lines), parasite treatment (infected vs uninfected control selection lines) and their interaction
224 considered as fixed factors. Selection line identity was included as error term. From this same data set, we
225 also performed univariate analyses for each of the 6 traits. In a second step, to better understand the
226 multivariate results, we performed a principal component analysis (PCA), based on the means per selection
227 line and thus a data matrix of $16 \times 6 = 96$ observations. To complement graphical inspection of results, we
228 used the first and second components (PC1, PC2) for ad-hoc comparisons between treatments. Finally, we
229 conducted correlations among monoclonal lines analysis using a Bayesian approach with the “rstan” package
230 version 2.19.3 [58], details in S7). We considered the three biologically relevant pairs of traits: dispersal-
231 resistance, $r_0\bar{N}$ and speed-tortuosity.

232 **Results**

233 We found signatures of selection history in the observed phenotypic variation and covariation (Table 1).
234 Univariate analyses showed significant effects of range expansion treatment and/or parasite treatment on
235 all 6 traits (Table 1). Multivariate analysis of the combination of these traits (MANOVA, Table 1) revealed
236 significant effects of the range expansion ($F_{1,12} = 8.69$, $p = 0.005$) and parasite treatment ($F_{1,12} = 8.82$, $p =$
237 0.005), but not their interaction ($F_{1,12} = 2.68$, $p = 0.111$). Thus, both selection treatments acted jointly, but
238 largely independently, to produce phenotypic differentiation in the *Paramecium*. All examined traits
239 contribute to this divergence, with three pairs of traits forming opposite vectors, pulling treatments in
240 different directions in multi-trait space: dispersal-resistance, $r_0\bar{N}$ and speed-tortuosity. The presence of
241 parasites changed the intensity and/or sign of the among-line correlation coefficient (Table S1) between
242 these pairs of traits as shown in Figure 2 (full dots vs empty dots).

243 **Table 1** MANOVA and ANOVAs for the effect of range expansion treatment (core vs. front) and parasitism
 244 treatment (infected vs. parasite-free populations) on six traits, measured for 16 *P. caudatum* selection
 245 lines. P-values < 0.05 highlighted in bold.

	Range expansion		Parasitism		Range x Parasitism	
MANOVA	$F_{1,12}$	p-value	$F_{1,12}$	p-value	$F_{1,12}$	p-value
All traits	8.69	0.005	8.82	0.005	2.68	0.111
ANOVA	$F_{1,12}$	p-value	$F_{1,12}$	p-value	$F_{1,12}$	p-value
Dispersal	1.08	0.318	11.43	0.005	1.86	0.197
Resistance	9.91	0.008	3.85	0.073	4.44	0.056
Speed	10.03	0.008	0.05	0.827	0.14	0.713
Tortuosity	14.82	0.002	3.29	0.094	2.29	0.156
r_0	3.72	0.077	0.68	0.425	5.25	0.040
\bar{N}	2.79	0.120	12.17	0.004	4.76	0.049

246 Considering the PCA components, we observed along the horizontal PC1 axis a clear effect of the range
 247 expansion treatment (front vs core selection), separating the respective clouds of points (red vs blue Fig. 1).
 248 This front / core separation was significant both for the parasite-free selection lines (lower two clouds in Fig.
 249 1, $F_{1,4} = 16.47$, $p = 0.015$), and for selection lines evolving in the presence of the parasite (upper two clouds,
 250 $F_{1,4} = 5.83$, $p = 0.042$). The direction and length of the different arrows in Fig. 1 show that these patterns were
 251 mainly driven by opposing trends in movement (speed / tortuosity; highest PC 1 loadings, Table S3). Overall,
 252 *Paramecium* from the front treatment generally had a lower swimming speed (-12%) and more non-linear
 253 movement trajectories (+23% tortuosity) than *Paramecium* from the core treatment (main effects of range
 254 expansion treatment: $p < 0.009$; Table 1). The PCA also indicates opposing trends in demography-related
 255 traits (Table S3), with lower population growth rate and higher equilibrium density for front selection lines
 256 (Fig. 1). However, univariate analyses show that these trends mainly hold for parasite-free control lines
 257 (range x parasite interaction: $p < 0.05$; Table 1).

258 Along the vertical PC2 axis, phenotypic differentiation was mainly determined by the parasite selection
 259 treatment (Fig. 1), thus separating selection lines that had been exposed to the parasite and parasite-free
 260 lines. The separation was more pronounced for the range front treatment (upper vs lower red clouds, $F_{1,4} =$
 261 9.74 , $p = 0.020$) than for the core treatment (upper vs lower blue points, $F_{1,4} = 3.22$, $p = 0.122$). Resistance
 262 and dispersal are the key traits associated with the PC2 axis (Table S2), with arrow trajectories indicating
 263 higher dispersal for parasite-free lines and higher resistance for lines that had evolved with the parasite (Fig.
 264 1). Univariate analyses confirmed these trends (effects of parasite treatment: $p < 0.07$; Table 1), but also
 265 indicate interactions with the range expansion treatment. Thus, the PCA signal for resistance was mainly due

266 to increased resistance in the front treatment (+ 15%), but not in the core (Fig. 2B; parasite x range treatment:
267 $p = 0.056$; Table 1). The details of the PCA are provided in Tables S2 and S3. In summary, the experimental
268 treatment combining new front selection conditions with new exposure to parasites produced the strongest
269 phenotypic divergence relative to the negative control treatment (no parasite, no selection for dispersal).

270 **Discussion**

271 Natural populations are currently forced to shift their ranges due to human-induced climatic and
272 environmental changes, which often involves exposure to novel abiotic conditions or confrontation with
273 biotic selection pressures such as parasitism [59,60]. Understanding how ecological dynamics affect the
274 evolution and the trajectory of phenotypic traits in a host-parasite context is of major interest and relevance
275 for conservation, controlling infectious disease and epidemic outbreaks [61,62], as well for the design of
276 proper invasive species management strategies [63]. Importantly, parasites can change the availability of
277 genetic variance to selection as well as the strength and direction of selection. Together with the selective
278 pressures due to range expansion, parasitism will likely produce diverse patterns of divergence and
279 syndromes, leading to changes in the whole host phenotype with trait modifications and the emergence of
280 trade-offs. A multivariate perspective is therefore essential to shed light on how complex traits, like dispersal
281 and resistance, respond to eco-evolutionary pressure in natural populations.

282 In our study, we find substantial multi-trait divergence between core and front populations of an expanding
283 range. Maximum divergence was observed when both new selection pressures were additively combined
284 (front and parasite treatment). Parasites accentuated the difference between core and front populations for
285 some traits, while reducing it for others. Three pairs of traits, typically involved in differentiation at range
286 fronts, acted as opposing vectors in multi-trait space defining the phenotypic divergence between
287 treatments: dispersal - resistance, $r_0 - \bar{N}$ and swimming speed - tortuosity. In particular, front populations
288 evolved with the parasite were more resistant, but dispersed less than their uninfected counterparts. Even
289 though we may currently not know the precise physiology or morphology underlying these trait relationships,
290 our results illustrate how spatial selection and parasite-mediated selection act jointly to produce major shifts
291 in phenotypic trait space, resulting in multi-faceted dispersal syndromes. Below we focus on the
292 evolutionary forces driving the three trait associations and discuss possible feedbacks of such evolutionary
293 changes on range expansion speed.

294 **Parasitism and dispersal syndromes: effects of the parasite at the front**

295 *Dispersal and resistance*

296 In line with theoretical and empirical results on range expansion dynamics, we found evidence for the
297 emergence of dispersal syndromes, with higher levels of dispersal at the range front compared to core
298 populations [23,29,51]. However, the parasite imposed a strong additional selection pressure involving

299 resistance as key component. Resistance is indeed a major trait selected under parasitism in the *Paramecium*
300 - *Holospira* system [49,64] as well as in many others [65,66], but has so far received relatively little attention
301 in a range expansion context. Here, front populations exposed to the parasite exhibited overall increased
302 resistance, but reduced dispersal, relative to parasite-free populations. Additionally, parasites exacerbated
303 the negative correlation between dispersal and resistance at the front (Figure 2, Table 2), indicating the
304 presence of a trade-off.

305 Although exceptions are known [67,68], resistance [69] and dispersal [70] are usually considered costly traits,
306 suggesting the potential for a trade-off between the two traits. Thus, the energetic costs of mounting an
307 effective (constitutive) defence against the parasite may reduce the resources available for host dispersal.
308 Similarly, the trade-off may have a behavioural component, linking resistance and feeding activity. Paramecia
309 are filter-feeders and become infected when they ingest infectious spores of the parasite together with other
310 food particles. Hence, resistance may be increased by reducing contact rates with the parasite through
311 reduced feeding activity [71]. Such behavioural aspects of resistance may also include altered swimming
312 behaviour, which could, in turn, affect feeding rates as well as dispersal (see below).

313 The nature of resistance - dispersal trade-offs may vary from system to system. For example, in bacteria, cell
314 surface modification conferring resistance to bacteriophages might come at the cost of reduced motility
315 [72,73]. Results from selection experiments indicate that the presence of phage can indeed favour bacterial
316 dispersal evolution [72,74]. However, unlike in our study, there was no clear link with resistance, implying
317 that the two traits evolved independently. This suggests that, apart from mechanical or physiological
318 constraints, the emergence of evolutionary trade-offs also depends on the underlying genetic architecture
319 and relationships with fitness, or on the environmental conditions [39,48,75,76].

320 r_0 and \bar{N}

321 On the demographic axis of the multi-trait space, populations at the range front were characterized by high
322 \bar{N} (equilibrium density) and low r_0 (population growth rate), indicating a trade-off typical of classic r-K
323 selection theory (Fig. 1). This negative correlation was particularly pronounced in the parasite treatment (Fig.
324 2, Table 2). At first sight, this finding appears in contradiction with common ideas about dispersal syndrome
325 evolution (Burton et al. 2010), predicting competitive strategies in the core (high K or \bar{N}) and opportunistic
326 strategies at range fronts (high r_0). However, in our experiment competitive ability is characterised by growth
327 rate, rather than equilibrium density [51,53]. Indeed, the authors showed that rapacious high-growth
328 variants are favoured in the core treatment and more slowly growing variants at the front. This was further
329 corroborated in another study on *P. caudatum*, using both experimental evolution and predictive modelling,
330 again showing the emergence of high \bar{N} and low r_0 at the range front [51]. Comparative evidence suggests

331 that this may actually be a widespread pattern [77]. Parasite-mediated selection reinforced this pattern and
332 further accentuated the divergence between front and core populations (Fig. 1).

333 Swimming speed and tortuosity

334 We found decreased swimming speed for *Paramecium* from the range front treatment, consistent with
335 previous observations [51]. This is opposite to results in a very similar experiment with another ciliate [53]
336 and also seems to contradict observations in other biological systems, where populations at the range front
337 show enhanced movement ability [29,32]. However, in our case, speed reduction was associated with
338 increased swimming tortuosity, i.e., the ability of the individuals to change directions while swimming. This
339 could be considered as an upregulated exploratory behaviour that potentially increases the probability to
340 find the dispersal corridor in our dispersal systems. The negative relationship between these swimming
341 behaviours was accentuated by the presence of the parasite at the range front (Figure 2, Table 2). Thus,
342 similarly to the reinforcement of the above dispersal - resistance and $r_0 - \bar{N}$ relationships, parasite-mediated
343 selection also seems to affect the trait architecture of movement behaviour, i.e., the mechanistic basis of
344 dispersal.

345 **Evo-to-eco feedbacks: Is the evolution with a parasite slowing down range expansions?**

346 The evolution of dispersal syndromes typically produces ecological feedbacks that speed up range expansions
347 [4]. Parasites, in contrast, may keep in check host populations and thereby limit or slow down range fronts,
348 as generically shown for interspecific (antagonistic) interactions [5,9,78]. As discussed above, one of our main
349 results is the emergence of a trade-off between resistance and dispersal. Thus, an intuitive prediction is that,
350 due to lower dispersal, evolution with the parasite slows down the range expansion of infected populations
351 compared to parasite-free populations. This is all the more conceivable given the negative ecological effect
352 of the parasite, via reductions in host density or reduced dispersal [46]. In natural range expansions, instances
353 of resistance evolution against parasites have been described, e.g., at the range front of the cane toad [36].
354 It is unknown how the resistance evolution in the toad affects dispersal traits, but it seems obvious that, in
355 the face of a detrimental parasite (a lung worm), having the resistance will benefit the range expansion more
356 than not having it.

357 Beyond these simple considerations, however, answers may be more complex [79], especially when we
358 compare different "strategy sets" (or, dispersal syndromes) in the presence of parasites. Our results suggest
359 that consequences for range expansion speed depend on how the trade-off between dispersal and resistance
360 plays out precisely. If contact rates are high and the parasite virulent, higher levels of resistance may indeed
361 compensate lower dispersal, such that front populations with an evolutionary parasite past might advance
362 faster than their naive, parasite-free counterparts. On the other hand, if enemy-free space can be reached,
363 less resistant variants may actually outrun the parasite and fully profit from their higher dispersal capacity.

364 These scenarios may, for example, relate to biological invasions, where invaders bring along a parasite to
365 which they have already adapted or, like the cane toad, encounter new parasites at the range front.

366 For our experimental system, predictions from the present data remain speculative, because relevant
367 measurements (growth, dispersal, movement) were taken in the absence of infection. A full picture would
368 require estimates of virulence/tolerance or dispersal assays that include infected hosts [47,48]. Moreover,
369 future experiments can use multi-microcosm landscapes to actually measure range expansion speeds for the
370 different types of hosts and/or epidemiological scenarios. Ideally, these would be accompanied by theory
371 exploring the simultaneous evolution of dispersal and interaction traits [80] or accounting for additional
372 factors, such as parasite evolution [47], dispersal plasticity [39,48] or the general upregulation of immune
373 responses, known to occur during range expansion [81,82].

374 **Conclusion**

375 Our selection experiment illustrates the possible impact of biotic interactions on evolutionary trajectories
376 during range expansions. Parasite-mediated selection changed the structure of dispersal syndromes, shaping
377 the phenotypic divergence between front and core populations for multiple traits, including dispersal, life-
378 history and resistance. Specifically, increased resistance traded off with a decrease in dispersal, which could
379 potentially slow down range expansion. This study focused on the host, but dispersal syndromes may also
380 evolve in the parasite, or even coevolve. This may further complexify the interplay between spatial selection
381 and biotic (co-)evolution, and make it even more difficult to predict the accompanying ecological feedbacks
382 on range expansion dynamics or disease spread. Replicated range expansions under laboratory conditions
383 represent one tool to disentangle these processes. Although being simplified abstractions of the real world,
384 such experiments may also provide baseline information for applied issues of biocontrol or the monitoring
385 of emerging diseases.

386 **Ethics**

387 Not applicable

388 **Data accessibility**

389 The data from this study are available at <https://doi.org/10.5281/zenodo.5907048>

390 **Authors' contributions**

391 OK, LN and GZ conceived the study. GZ, LN, MH, EAF, and OK designed the experiments. GZ, LN, CGB and
392 OK performed the experimental work. GZ, LN, OK and EAF performed the statistical analysis. All authors
393 interpreted the results. GZ and OK wrote the first draft of the manuscript and all authors commented on
394 the final version.

395 **Competing interests**

396 The authors declare no competing financial interests.

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591

592 **Figure captions**

593 **Figure 1** PCA for six traits measured for 16 long-term selection lines of *P. caudatum*, showing the first two
594 principal component axes, PC1 and PC2. The length of the thin arrows inside the graph represents the loading
595 values; the longer the arrow the higher the correlation with the PC axis and the variance explained by that
596 trait. Each point represents the average phenotypic value of a given selection line in multivariate space, with
597 long-term treatment origins specified: range core (blue) vs. range front (red) treatment; evolved in the
598 presence (full circles) vs. absence (empty circles) of the parasite *H. undulata*. The blue empty circles with
599 dotted ellipsis represent the ancestral host populations. The ellipses represent the containment probability
600 region of the multivariate space for the four treatments.

601 **Figure 2** Trait correlations for *P. caudatum* evolved in range core and range front treatments: (A-B) dispersal
602 - resistance, (C-D) growth rate (r_0) - equilibrium density (\bar{N}) and (E-F) swimming speed - swimming tortuosity.
603 For each panel, the parasitism treatment is specified: evolved in the presence of the parasite (full dots and
604 lines) vs. parasite-free (dashed dots and lines). The points are the means for 60 monoclonal cultures, isolated
605 from a total of 16 long-term selection lines (for details, see text). Correlations were obtained with Bayesian
606 inference, separately for each combination of range expansion and parasitism treatments. Ellipses are bound
607 to non-linear space and represent the 10, 25, 50, 75 and 90 % CI of the correlation of pairs of traits.
608 Distributions in the small panels represent the posterior distributions of the correlation coefficients, the
609 dashed black line highlight the 0 value, and thus the absence of correlation.

610



