



Defensive behavior is linked to altered surface chemistry following infection in a termite society

M. Alejandra Esparza-Mora, Tilottama Mazumdar, Shixiong Jiang, Renate Radek, Julian N. Thiem, Linshan Feng, Vesta Petrašiūnaitė, Ronald Banasiak, Marek Golian, Melanie Gleske, et al.

► To cite this version:

M. Alejandra Esparza-Mora, Tilottama Mazumdar, Shixiong Jiang, Renate Radek, Julian N. Thiem, et al.. Defensive behavior is linked to altered surface chemistry following infection in a termite society. Scientific Reports, In press, 13, pp.20606. 10.1038/s41598-023-42947-9 . hal-04288294

HAL Id: hal-04288294

<https://hal.science/hal-04288294>

Submitted on 15 Nov 2023

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Defensive behavior is linked to altered surface chemistry following infection in a termite society

M. Alejandra Esparza-Mora^{1,2}, Tilottama Mazumdar^{1,2}, Shixiong Jiang^{1,2}, Renate Radek¹, Julian N. Thiem¹, Linshan Feng¹, Vesta Petrašiūnaitė¹, Ronald Banasiak², Marek Golian³, Melanie Gleske³, Christophe Lucas⁴, Andreas Springer⁵, Jan Buellesbach³, Dino P. McMahon^{1,2,*}

¹ Institute of Biology, Freie Universität Berlin, Königin-Luise-Straße. 1-3, 14195 Berlin, Germany

² Department for Materials and Environment, BAM Federal Institute for Materials Research and Testing, Unter den Eichen 87, 12205 Berlin, Germany

³ Institute for Evolution and Biodiversity, University of Münster, Hüfferstraße. 1, 48149 Münster, Germany

⁴ Institute for Research on Insect Biology (UMR7261), CNRS – University of Tours, Tours, France

⁵ Core Facility BioSupraMol, Department of Biology, Chemistry and Pharmacy, Freie Universität Berlin, Takustraße. 3, 14195 Berlin, Germany

* Corresponding author

Abstract

The care-kill response determines whether a sick individual will be treated or eliminated from an insect society, but little is known about the underpinnings of this process. We exploited the stepwise infection dynamics of an entomopathogenic fungus in a termite to explore how care-kill transitions occur, and identify the chemical cues behind these shifts. We found collective responses towards pathogen-injected individuals to vary according to severity and timing of pathogen challenge, with elimination (via cannibalism) occurring sooner in response to a severe active infection. However, injection with inactivated fungal blastospores also resulted in increased albeit delayed cannibalism, even though injection did not universally cause host death. This indicates that the decision to eliminate an individual is triggered before pathogen viability or even terminal disease status is known. We then compared the surface chemistry of differently challenged individuals, finding increased amounts of long-chained methyl-branched alkanes with similar branching patterns in individuals injected with both dead or viable fungal blastospores, with the latter showing the largest increase. This coincided with the highest amounts of observed cannibalism as well as signs of severe moribundity. Our study provides new mechanistic insight into the emergent collective behaviours involved in the disease defence of a termite society.

Introduction

The evolutionary and ecological success of social insects is widely appreciated. Although sociality offers many benefits, there are also associated costs¹. On the one hand, cooperation facilitates division of labour, enabling the optimization of tasks such as brood care, foraging, nest building, information sharing and defense²⁻⁴. On the other hand, densely populated and closely related individuals inside a confined nest environment are at risk of disease transmission^{5,6}. The risk of repeated pathogen exposures have shaped the evolution of defence mechanisms across insect societies⁷. Social insect individuals are not only able to fight infections through their own individual immune systems^{8,9,10}, but also via social immune defenses^{2,3,11}. Social immunity can be viewed as a distributed organ, similar to the immune system of metazoan animals¹² that builds on two principal adaptive pillars – behaviour and physiology – that together act to prevent the entrance, establishment, and spread of pathogens in a colony^{2,4,6,9,13,14}.

Social insect groups, such as Hymenoptera (social bees, wasps, and ants) and Isoptera (termites) contain multiple lineages of species with advanced social living, often referred to as superorganisms¹⁵. Studies in these social groups have demonstrated the range of collective disease defenses that can be deployed against infected individuals^{3,16-20}. Infected individuals can be groomed, which in combination with the use of antimicrobial secretions, serve to cure or rescue infected individuals and reduce the likelihood of internal infection. In contrast, once an internal infection is successfully established, it can pose a threat to the colony and may need to be eliminated from the colony^{3,6,17,21}. Such radically different responses towards infected individuals have been described in terms of a care-kill dichotomy^{6,21,22}. Allogrooming is one of the first lines of defense against external pathogens, while elimination typically occurs at a later stage^{17,19,23-25}. Both strategies are thought to reduce the risk of disease outbreak and enhance colony survival^{3,6}, but the latter imposes an additional cost to the colony through the loss of workforce members, depending on worker age as well as colony size and maturity.

Collective disease defenses are known to vary within and between species and across different levels of social organization^{3,12,26,27}. Detection of pathogens and the regulation of the collective response depends on the ability of social organisms to accurately communicate disease-related information through behavioural or chemical cues^{6,28}. Chemical communication is thought to play a central role in social immunity⁶. For instance, in the ant *Lasius neglectus*, destructive disinfection behaviours are employed, whereby ants initially detect fungus-infected pupae via sickness cues emitted by the pupae, followed by elimination¹⁹. Signalling and chemical communication in social insects are often mediated by cuticular hydrocarbons (CHCs)²⁹. CHCs are the most abundant class of chemicals coating the cuticle of terrestrial insects and have a key role in inter- and intraspecific communication^{30,31} potentially also serving to discriminate between healthy and sick nestmates of a colony²⁹. CHC profiles can be altered

by activation of the immune system^{29,32-34} and altered CHC profiles have been implicated as signals to eliminate infected individuals in ants and honeybees^{19,29,32,35} in a manner analogous to body cells signalling their status to the immune system³⁶.

Termites also employ several collective mechanisms to prevent disease outbreaks, largely using two complementary behaviours: allogrooming and cannibalism, reflecting the care-kill strategy mentioned above. The switch in the response depends on the progression of the infection, with cannibalism occurring when infected individuals display signs of overt sickness, such as slow movement¹⁷. Application of antimicrobial secretions, in addition to behavioural responses, can effectively remove external pathogens from the cuticle of infected individuals and inhibit pathogen growth during the early-stage of infection^{23,37-44}. However, when a pathogen successfully initiates a potentially lethal internal infection⁴⁵, such weak⁴⁶ or moribund but not yet dead⁴⁷ individuals are typically cannibalized. Cannibalism is thought to be ancestral to all termites, which may have evolved as a prophylactic mechanism against disease from an initial role in nitrogen recycling⁴⁸. But the basis of the switch between social immune behaviours in termites remains largely unknown, and it is unclear what signals or chemical cues trigger elimination behaviours or to what extent they are linked with factors associated with moribundity¹⁷.

To address these gaps in our knowledge, we investigated how termites detect and respond to internal infection upon injection of either viable or dead blastospores of a generalist fungal pathogen. Fungal blastospores, which represent a within-host infectious form, play an important role in pathogenesis during disease development and their experimental injection can elicit a robust immune response⁴⁹⁻⁵². Blastospores are a vegetative cell state that can quickly multiply and colonize the insect body or a nutrient-rich liquid culture⁵³⁻⁵⁵. Furthermore, blastospores reflect an intermediate stage in the infection process, which begins with conidia adhesion and germination on the insect cuticle, followed by cuticular penetration via appressoria to colonize the host's body. Within the host, the fungus proliferates as single-celled blastospores or hyphal bodies in the hemolymph ultimately killing the infected insect^{45,53,56,57}. During infection, virulence factors are synthesized, such as destruxins (cyclodepsipeptidic mycotoxins) and even host miRNAs can be manipulated to inactivate the host and assist in avoidance of immune clearance^{49,52,54,58-60}.

Here, we combine blastospore infection and behavioural experiments with chemical analyses of termite CHC profiles and fungal destruxins to expand our understanding of termite social immunity. Our rationale is that to better study the care-kill switch, it is useful to separate direct external pathogen cues from indirect host-mediated cues, allowing one to better investigate how termites detect and respond to an internal pathogen challenge. We explore how defensive behaviours are triggered by experimentally varying internal pathogen challenges, and correlate these with alterations to termite

surface chemistry. Our study sheds new light on a key collective defensive mechanism of termite societies.

Results

Blastospores germinate quickly and synthesize destruxins

We applied *Metarhizium robertsii* blastospores via injection to exclude the presence of external pathogen cues (cuticular conidiospore exposure) from internal host activity (infection initiation). Prior to injection experiments, we examined rates of blastospore germination. We used a scanning electron microscope (SEM) to quantify germination (Figure 1A, B, C, D) on potato dextrose agar (PDA) plates. Germination was observed as early as 4 hours after inoculation, as evidenced by the formation of germ tubes (Figure 1B, Supplementary Figure S1). The earliest time reported in the literature is 2.7 hours in the unrelated fungus *Isaria fumosorosea*⁶¹. At 8 hours, germination and elongation of germ tubes continued and by 12 hours a thick mass of elongated hyphae, forming the mycelium (Figure 1C, D) could be observed. Our observed rate of *in vitro* blastospore germination is faster than either *in vitro* or *in vivo* (on cuticle) conidia germination^{17,45} (Supplementary Figure S1, S2). In addition to viable blastospores, we employed dead blastospores as an injection treatment. We confirmed the efficiency of inactivation following heat-treatment (autoclaving) by the absence of germination following plating. The presence of 5 major classes of destruxins: Destruxin A, B, C, D and Ed were also investigated in the filtered media (CD modified media) of cultured viable *M. robertsii* blastospores (Supplementary Figure S3). Comparisons of the UHPLC-MS/MS data with the elution order and the MS/MS schemes published in the literature^{62,63} verified the retention times for the respective destruxins found in the chromatograms, confirming their presence. By contrast, no destruxins were detected in the filtrate derived from heat-inactivated (dead) blastospores (Supplementary Figure S4), indicating that destruxins were destroyed by autoclaving.

Both viable and dead blastospore injection causes termite death

Before embarking on behavioural experiments in our study species, the eastern subterranean termite *Reticulitermes flavipes*, we first measured survival of the termite when kept in isolation, in order to measure treatment lethality in the absence of group dynamics. Ringer-injection had no negative effect on termite survival, whereas viable blastospores caused 100% mortality by 2.5 days post injection, and dead blastospores resulted in a plateau at 40% mortality by 5 days post injection (Supplementary Figure S5), remaining unchanged for 10 days, at which point observations were terminated.

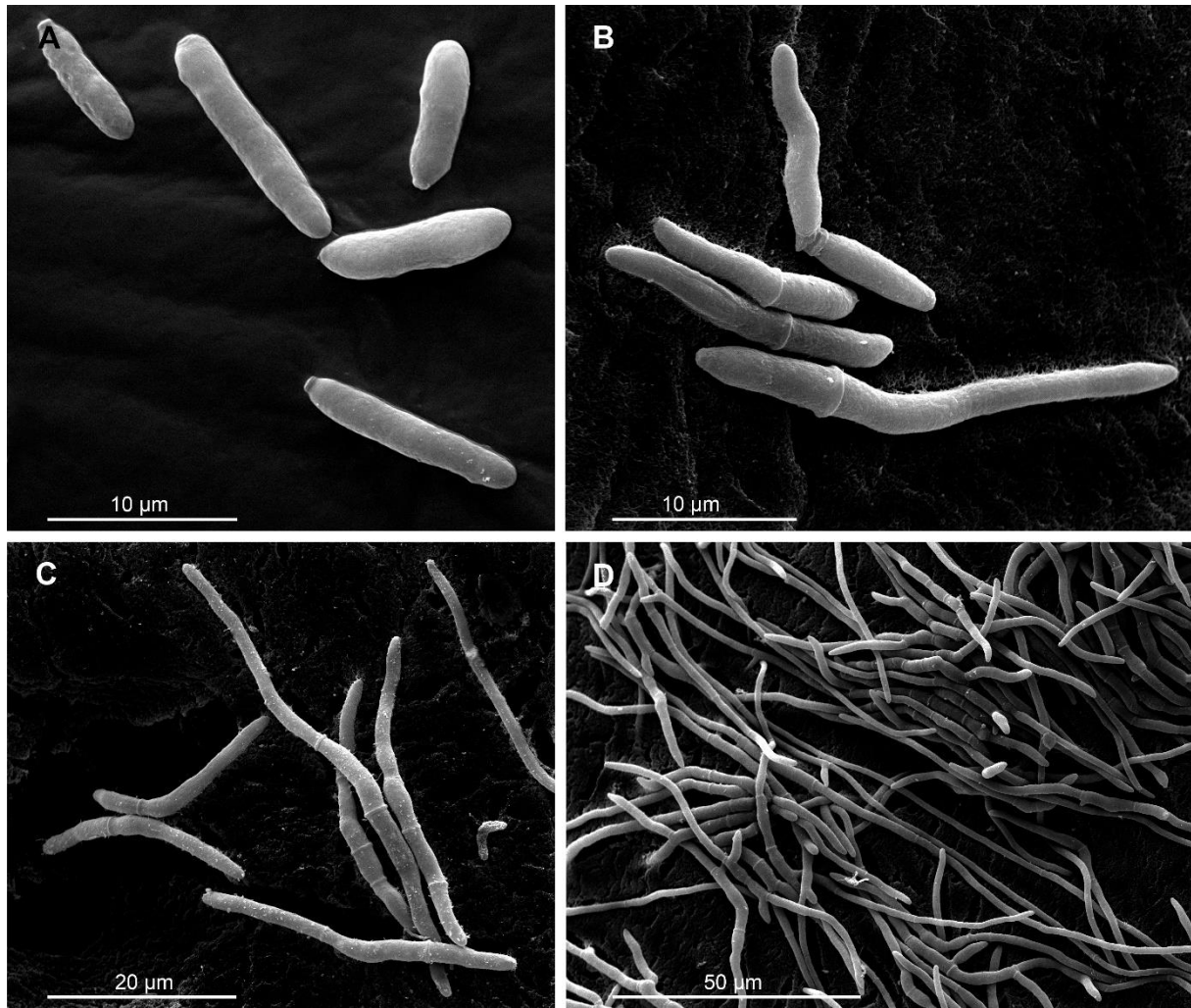


Figure 1. Germination process of *M. robertsii* 1490 blastospores. **A-D:** Scanning electron micrograph (SEM) images of blastospore germination at different time-points post-inoculation *in vitro* (PDA media). At 2 h blastospores are not yet germinated (A), while at 4 h, germination of blastospores can already be observed (B), which continues to an advanced stage via germ tube elongation at 8 h (C). By 12 h, elongation of the fungal hyphae has continued, and mycelial networks are now visible (D).

Termites slightly increase allogrooming following exposure to injected individuals

In the first behavioural observation experiment, we compared the effects of injection with Ringer's solution or blastospores (viable or dead). Injected focal termites were isolated for 2, 8, 12 and 15 hours prior to introduction into experimental colonies. Behavioural patterns were then recorded in response to Ringer's solution (Ringer), viable (Blastospores+) or dead blastospore (Blastospores-) injected focal termites at different stages post-challenge. In general, observed behaviours in response to blastospore injected focal termites consisted of low levels of grooming, high amounts of cannibalism, no observations of burial (Supplementary Figure S6), as well as other behavioural states unrelated to social immunity. Behavioural patterns in the control treatments were characterized mostly by other states, low levels of grooming and one incident of cannibalism in the 15 hours incubation time.

The overall proportion of allogrooming in both viable and dead blastospore treatments were significantly higher than control treatments, except at 8h and 15 hours (Supplementary Table S1) (2h, Blastospores- vs Ringer $z = 3.293$ $P = 0.0030$, Blastospores+ vs Ringer $z = 4.448$ $P = < 0.0001$; 12h, Blastospores- vs Ringer $z = 4.324$ $P = < 0.0001$, Blastospores+ vs Ringer $z = 4.515$ $P = < 0.0001$). The low proportion of allogrooming observed at the later 15 hour timepoint is linked with a high proportion of cannibalism (Figure 2, 3A, Supplementary Table S1, Supplementary Figure S7).

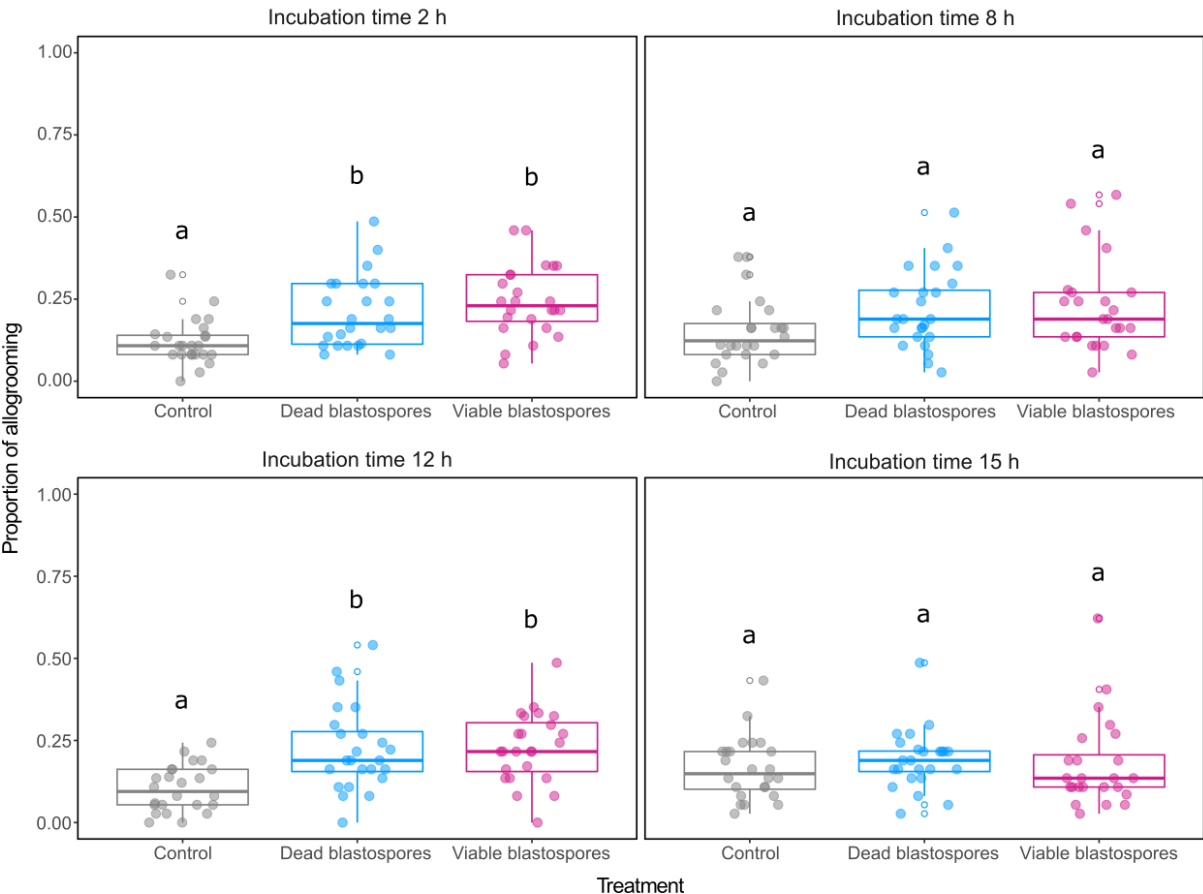


Figure 2. Allogrooming as a proportion of total states across treatments in the blastospore experiment. The proportion of allogrooming states depends significantly on treatment. Significant *post hoc* comparisons are indicated by different letters, derived from separate GLMMs from each time point. Note that *post hoc* comparisons do not apply between incubation time points. Lower and upper hinges correspond to first and third quartiles. Upper and lower whiskers extend to the largest/smallest value if no greater/smaller than 1.5 times the inter-quartile range from the hinge, respectively.

In a follow-up experiment, we compared injection of dead blastospores with injection of an equivalent solution filtered to remove blastospore debris (Filtrate). The overall proportion of allogrooming was at its highest at 12 hours post-challenge and decreased over time for the two fungal treatments (Blastospores- and Filtrate) while remaining unchanged for the control group, with no significant differences detected between any of the time points. The overall proportion of allogrooming in both dead blastospore and filtrate treatments was significantly higher over control treatments at two incubation time points (12h, Blastospores- vs Ringer $z = 7.636$ $P = 0.0001$, Filtrate vs Ringer $z = 5.349$ P

= 0.0001; 15h, Blastospores- vs Ringer $z = 6.626$ $P = 0.0001$, Filtrate vs Ringer $z = 4.634$ $P = 0.0001$) (Supplementary Figure S8). Allogrooming was significantly higher in dead blastospore versus filtrate treatment at 12 hours but at no other time point (12h, Blastospores- vs Filtrate $z = -2.510$ $P = 0.0363$). A progressively lower level of grooming corresponded with an increasingly higher proportion of cannibalism for the 15 and 18 hour time points, with a high number of termites already being eliminated by 21 hours post infection (Supplementary Table S3 and Supplementary Figure S9, S10).

Cannibalism does not completely depend on termite moribundity

In the first experiment, the highest amounts of cannibalism were observed at 15 hours post injection in the viable blastospores treatment, with cannibalism beginning shortly after introduction of the focal termite, and completely replacing grooming before the end of the observation period (Supplementary Figure S6). Interestingly, we observed that nestmates cannibalized dead blastospore-injected focal individuals as well (Figures 3A, Supplementary Table S2). However, cannibalism occurred sooner and more frequently when infected focal termites were visibly ill and close to death (i.e. viable blastospores at 15h). All focal termites were alive at the time of cannibalism (beginning < 24 h after challenge), tallying with the survival of injected individual termites when kept in isolation, where blastospore-induced death began between 24-48 h post injection (Supplementary Figure S5). Control focal termites were left unharmed.

In survival analyses, significant differences were detected between viable blastospore and Ringer-injected control treatments (2h, Blastospores+ vs Ringer $z = 3.312$ $P = 0.0028$; 8h, Blastospores+ vs Ringer $z = 3.226$ $P = 0.0038$; 12h, Blastospores+ vs Ringer $z = 2.808$ $P = 0.0150$; 15h, Blastospores+ vs Ringer $z = 4.822$ $P = < 0.0001$). Cannibalism was observed in all dead-blastospore treatments and comparisons with Ringer-injected controls were also significant for all incubation timepoints (2h, Blastospores- vs Ringer $z = 3.114$ $P = 0.0055$; 8h, Blastospores- vs Ringer $z = 2.397$ $P = 0.0497$; 12h, Blastospores- vs Ringer $z = 2.946$ $P = 0.0097$; 15h, Blastospores- vs Ringer $z = 2.684$ $P = 0.0218$) (Figure 3B, and Supplementary Table S2). Except for 1 individual in the 15h/Ringer treatment, cannibalism was not observed for any Ringer-injected focal termites (Figure 3A, and Supplementary Table S2).

In comparisons of viable and dead blastospore-injected treatments, the probability of remaining unharmed differed significantly only at the 15 hours incubation timepoint (15h, Blastospores+ vs Blastospores- $z = 4.054$ $P < 0.0002$). At this incubation time point, focal termites infected with viable blastospores treatment were quickly dismembered, whereas dead blastospore-injected individuals were cannibalized more slowly, but nonetheless these were completely cannibalized by 24 h after introduction (Figure 3B). Complete elimination of individuals quantified at 24 hours after introduction of the focal individual into experimental colonies was significantly higher for both viable and dead blastospore treatments compared with controls: (time in hours after injection) (26h, Blastospores- vs

Ringer $P = 0.0032$, Blastospores+ vs Ringer $P = 0.0018$; 32h, Blastospores- vs Ringer $P = 0.0044$, Blastospores+ vs Ringer $P = 0.0023$; 36h, Blastospores- vs Ringer $P = 0.0018$, Blastospores+ vs Ringer $P = 0.0012$; 39h, Blastospores- vs Ringer $P = 0.0012$, Blastospores+ vs Ringer $P = 0.0012$). Close to 100% of all focal individuals were eliminated in both viable and dead blastospore treatments. This is contrasted with the survival of the majority of termites following dead blastospore injection, when kept individually (Supplementary Figure S5). The percentage of eliminated focal individuals quantified at 24 hours did not significantly differ between viable and dead blastospore-injected individuals (26h, Blastospores+ vs Blastospores- $P = 0.5150$; 32h, Blastospores+ vs Blastospores- $P = 0.2244$; 36h, Blastospores+ vs Blastospores- $P = 1$; 39h, Blastospores+ vs Blastospores- $P = 1$) (Figure 3B).

In the blastospore filtrate experiment, significantly increased cannibalism was observed in both the dead blastospore and filtrate-injected treatments compared to the control (Blastospores- vs Ringer $z = 3.587$ $P = 0.000838$; Filtrate vs Ringer $z = 3.339$ $P = 0.0021$) (Supplementary Table S4 and Supplementary Figure S10). Termites challenged with dead blastospores were generally cannibalized at a higher rate, although the amount of cannibalism and overall survival probability was not significantly different between dead blastospore and filtrate injection (Supplementary Table S4).

Moribund termites display altered cuticular hydrocarbon profiles

A total of 90 individual equivalents of CHC extracts from worker pools (3 individuals each) taken from 3 different *R. flavipes* colonies were pre-defined into six groups for a discriminant analysis (DA) according to the different treatments: 12 and 15 hours after injection of viable and dead blastospores and Ringer's solution as a control, respectively (see above). Overall differentiation of the CHC profiles was significant according to the different treatments (Wilk's $\lambda < 0.05$, $P < 0.05$). Discriminant function 1 accounted for 47.36 % and discriminant function 2 for 26.11 % of the total variation, amounting to 73.47 % of total variance explained by the first two functions (Figure 4). Concerning the variations in specific CHC compounds according to treatments, 15 hours after injection, four methyl-branched alkanes consistently displayed significantly higher quantities in the viable blastospore treatment versus the control ($P < 0.05$, Benjamini-Hochberg corrected Mann-Whitney U tests, Figure 5). These compounds were 11-MeC35, 12-MeC36, 11-; 13-MeC37, and 11,15-DiMeC37. Injections with dead blastospores also led to a visible, albeit non-significant increase in overall quantities of these particular compounds compared to the controls. An overview of all identified CHCs including their respective absolute quantities per treatment is given in Table 1.

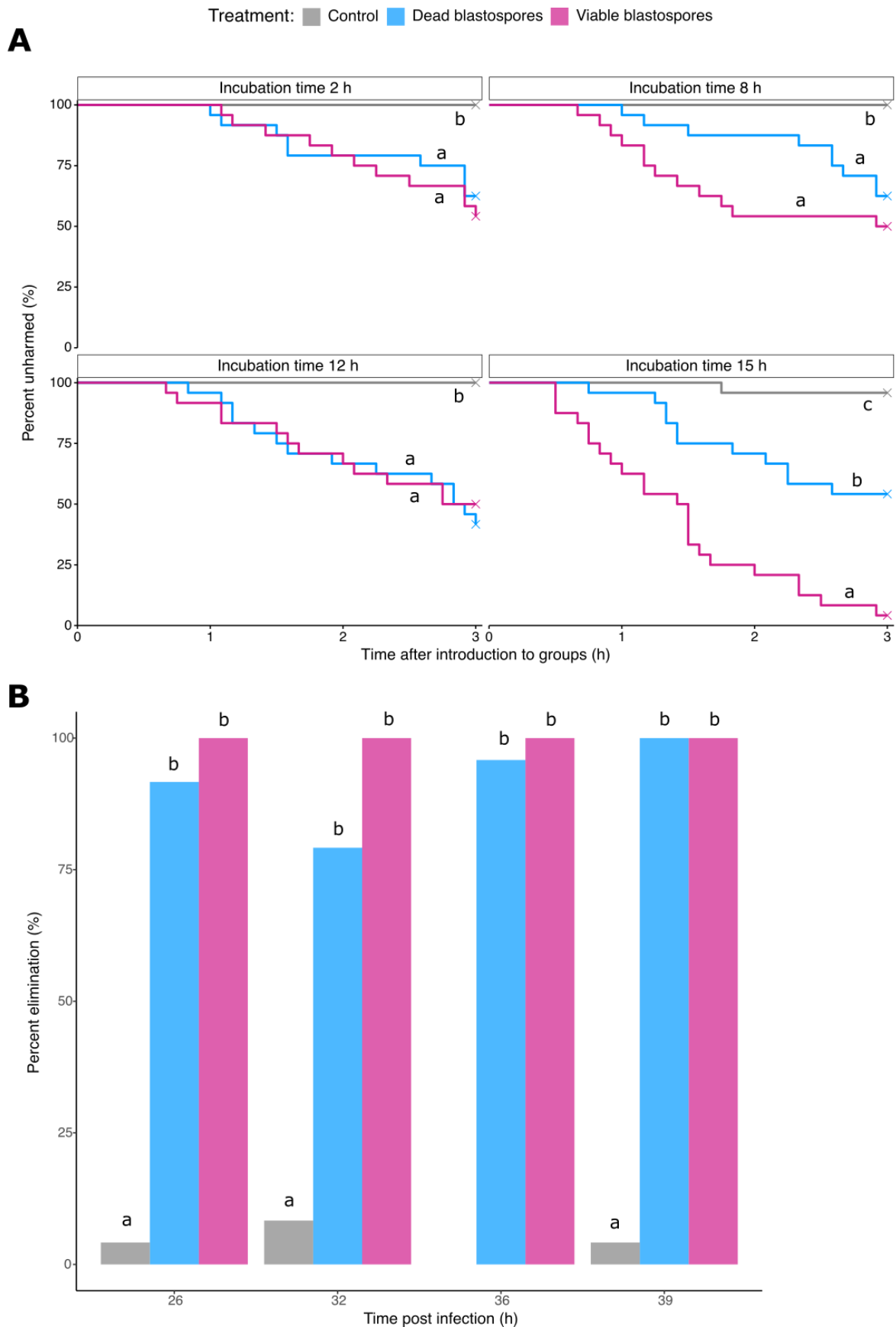


Figure 3. Survival of focal termites following introduction into experimental colonies. **A:** Percentage of focal termites that remain unharmed (due to cannibalism) after different incubation times (2, 8, 12, 15 h). Cannibalism states were recorded over a three-hour observation period following introduction into

experimental colonies. In both survival analyses, crosses indicate the presence of right-censored data (i.e. focal termites that were not dead (A) or harmed (B) during the observation period). Significant differences between treatments derived from survival models are indicated by different letters. Note that *post hoc* comparisons do not apply between incubation time points. **B:** Percentage of focal termites that had been eliminated at 24 h following their introduction into experimental colonies, corresponding to 26, 32, 36 and 39 h post-injection for incubation timepoints 2, 8, 12 and 15 h, respectively. Note that at equivalent timepoints for dead blastospore-injected termites kept in isolation (Supplementary Figure S5), mortality was lower than 20%. Treatments: Grey = Ringer (control), Blue = dead blastospores, Pink: viable blastospores.

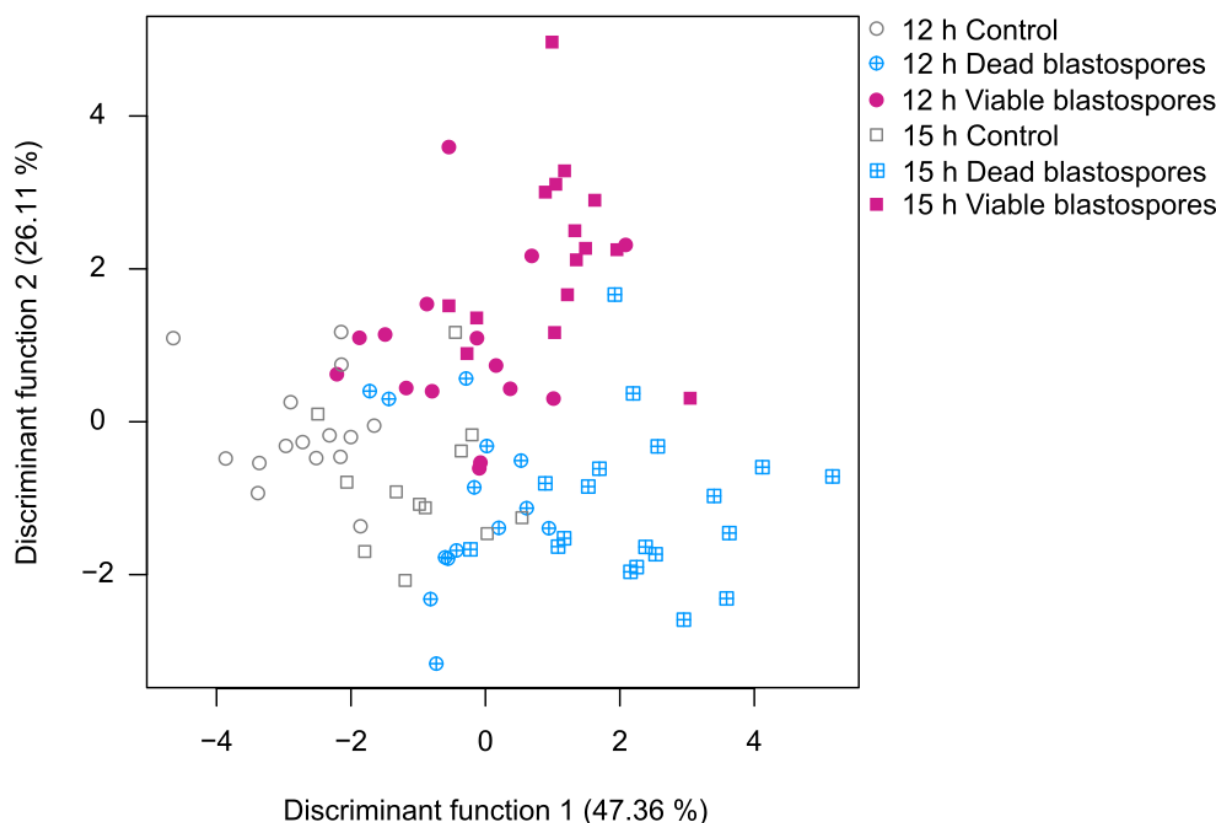


Figure 4. Linear discriminant analysis (LDA) based on 90 cuticular hydrocarbon (CHC) extracts from worker pools (3 individuals each) taken from 3 different *R. flavipes* colonies. The workers were treated with viable and dead blastospores, and Ringer solution as a control, and they were freeze-killed 12 or 15 h after injection. Overall differentiation was significant according to the different treatments (Wilk's $\lambda < 0.05$, $p < 0.05$), the respective contributions of the two discriminant functions to the total variation are indicated in percentages. Different colors and symbols indicate different treatments and time points, the different colony affiliations are not indicated due to them not resulting in consistently discernable differentiation.

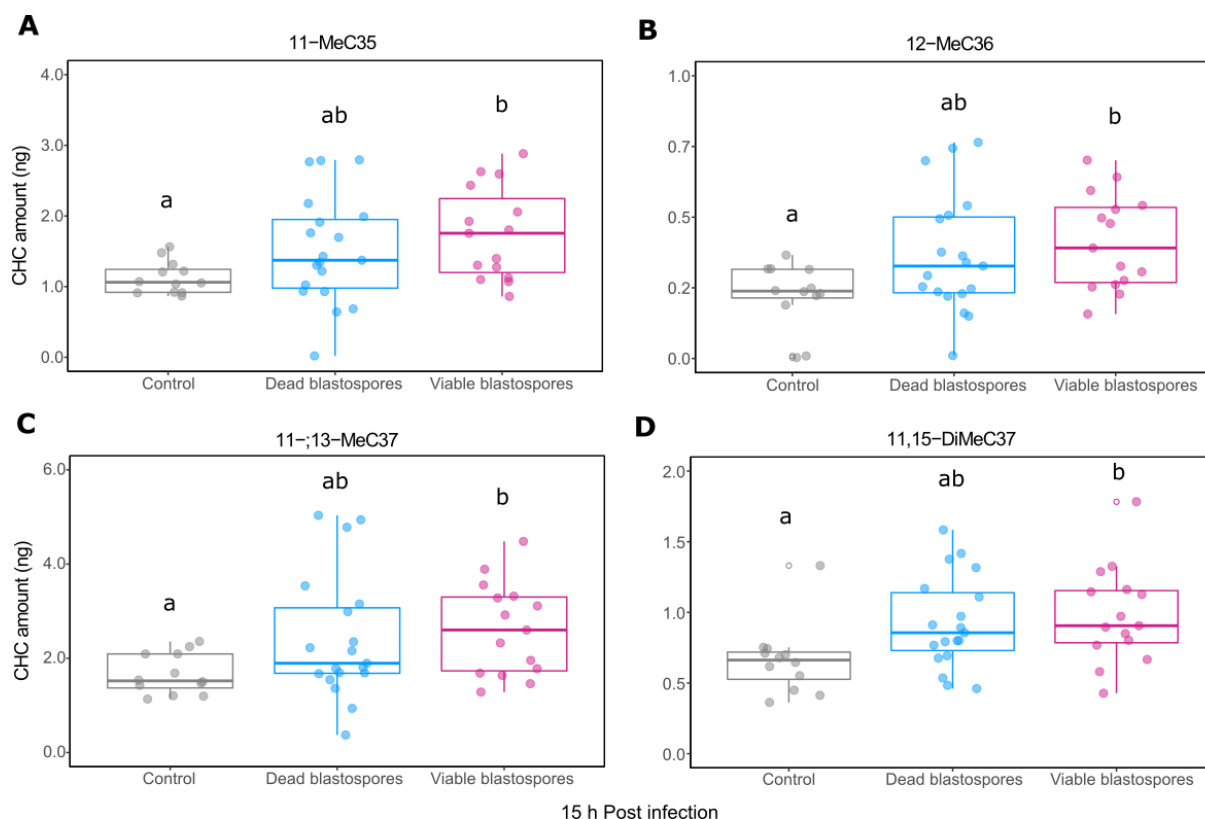


Figure 5. Quantitative comparison of a subset of CHC compounds (three mono- and one di-methyl-branched alkane) 15 h post injection of viable and dead blastospores, or Ringer solution as a control. CHC amounts were measured in ng as individual equivalents from extracts of worker pools (3 individuals each) taken from 3 different *R. flavipes* colonies. The absolute quantities of each of these compounds were significantly increased 15 h after injection of active blastospores as compared to the Ringer control (Benjamini-Hochberg corrected Mann-Whitney U tests). Although injection with inactive blastospores did not lead to significant increases after 15 h, an upward trend is observable.

Table 1. Retention indices (RI), compound identifications and the respective absolute quantities as well as standard deviations (in ng) for all CHCs compared between treatments of *R. flavipes* workers: 12 and 15 h post injection of viable and dead blastospores and Ringer solution as a control, respectively. Retention indices were calculated according to the position of detected *n*-alkanes in our samples and, where not available, with a C21-40 *n*-alkane standard run under the same conditions. Significantly different compound quantities are indicated in bold, accessed through Benjamini-Hochberg corrected Mann-Whitney U tests.

RI	Compound ID	12 h Dead blastospores	12 h Viable blastospores	12 h Control	15 h Dead blastospores	15 h Viable blastospores	15 h Control
2207	n-C22	0.12 ± 0.07	0.11 ± 0.08	0.09 ± 0.08	0.12 ± 0.06	0.15 ± 0.1	0.08 ± 0.05
2270	3-MeC22	0.09 ± 0.26	0.04 ± 0.05	0.02 ± 0.02	0.03 ± 0.02	0.04 ± 0.04	0.1 ± 0.27
2280	C23-ene_1	1.15 ± 0.69	0.96 ± 0.59	0.84 ± 0.69	1.4 ± 0.71	1.64 ± 1.35	0.86 ± 0.66
2287	C23-ene_2	0.07 ± 0.03	0.07 ± 0.04	0.05 ± 0.03	0.09 ± 0.03	0.11 ± 0.06	0.05 ± 0.02
2310	n-C23	9.2 ± 5.76	8.28 ± 5.93	8.04 ± 6.3	9.77 ± 4.21	11.69 ± 7.68	7.34 ± 4.79
2343	11-,13-MeC23	0.77 ± 0.47	0.66 ± 0.41	0.59 ± 0.44	0.92 ± 0.42	1.04 ± 0.72	0.63 ± 0.43
2371	4-MeC23	1.61 ± 1.08	1.54 ± 1.09	1.27 ± 0.87	1.87 ± 0.84	2.21 ± 1.46	1.33 ± 0.84
2380	C24-ene	0.57 ± 0.33	0.53 ± 0.37	0.43 ± 0.3	0.67 ± 0.23	0.82 ± 0.53	0.45 ± 0.29
2409	n-C24	5.66 ± 2.97	5.23 ± 3.11	5.03 ± 2.89	5.93 ± 1.61	6.89 ± 3.6	4.85 ± 2.28
2442	12-,14-MeC24	1.45 ± 0.77	1.36 ± 0.79	1.12 ± 0.68	1.82 ± 0.65	1.96 ± 1	1.27 ± 0.73
2463	C25-ene_1	0.1 ± 0.04	0.12 ± 0.04	0.1 ± 0.05	0.12 ± 0.04	0.16 ± 0.08	0.1 ± 0.03

2473	4-MeC24	16.46 ± 8.66	16.46 ± 9.71	13.92 ± 7.83	18.29 ± 5.39	22.33 ± 11.84	13.99 ± 6.49
2483	C25-ene_2	9.58 ± 5.23	9.19 ± 6.17	7.3 ± 4.86	11.7 ± 4.15	14.79 ± 8.41	7.7 ± 4.07
2483	C25-diene_1	6.02 ± 3.87	5.97 ± 4.53	4.49 ± 2.77	6.97 ± 3.69	8.9 ± 6.29	4.64 ± 3.12
2509	n-C25	25.88 ± 12.72	24.05 ± 13.28	23.74 ± 12.34	27 ± 6.45	31.66 ± 15.7	22.96 ± 9.55
2542	9-;11-;13-;15-MeC25	21.06 ± 11.28	19.47 ± 11.1	16.07 ± 9.25	24.94 ± 7.95	27.99 ± 14.37	18.62 ± 10.35
2555	C25-diene_2	3.29 ± 1.75	3.12 ± 1.81	2.72 ± 2.17	3.59 ± 1.61	4.74 ± 3.51	2.48 ± 1.39
2557	5-MeC25	2.58 ± 1.87	2.14 ± 1.68	1.68 ± 1.16	2.96 ± 1.99	4.09 ± 3.49	2.11 ± 1.52
2575	4-MeC25	3.37 ± 1.59	3.22 ± 1.59	2.7 ± 1.48	3.89 ± 1.05	4.67 ± 2.17	2.78 ± 1.19
2577	C25-diene_3	3.19 ± 1.95	3.15 ± 2.31	2.7 ± 1.5	3.66 ± 1.53	4.37 ± 2.72	2.86 ± 1.63
2581	3-MeC25	6.34 ± 3.15	6.24 ± 3.48	5.41 ± 3.11	7.06 ± 2.21	8.7 ± 4.72	5.59 ± 2.89
2641	12-;14-;16-MeC26	0.69 ± 0.45	0.63 ± 0.44	0.49 ± 0.31	0.73 ± 0.36	0.9 ± 0.51	0.58 ± 0.33
2651	6-MeC26	0.21 ± 0.09	0.2 ± 0.1	0.18 ± 0.12	0.25 ± 0.11	0.34 ± 0.28	0.2 ± 0.12
2669	4-MeC26	0.78 ± 0.42	0.71 ± 0.51	0.65 ± 0.39	0.89 ± 0.34	1.05 ± 0.56	0.69 ± 0.36
2681	3-MeC26 + C27ene	0.15 ± 0.1	0.14 ± 0.08	0.23 ± 0.2	0.18 ± 0.11	0.27 ± 0.17	0.14 ± 0.09
2711	n-C27	0.22 ± 0.11	0.18 ± 0.12	0.47 ± 0.36	0.28 ± 0.37	0.76 ± 2.11	0.28 ± 0.24
2742	11-;13-;15-MeC27	0.11 ± 0.05	0.08 ± 0.05	0.07 ± 0.04	0.12 ± 0.05	0.14 ± 0.07	0.1 ± 0.06
2744	C27diene1	0.29 ± 0.17	0.25 ± 0.17	0.21 ± 0.2	0.29 ± 0.18	0.45 ± 0.4	0.21 ± 0.15
2756	C27diene2	0.11 ± 0.05	0.09 ± 0.07	0.09 ± 0.05	0.13 ± 0.07	0.14 ± 0.06	0.11 ± 0.04
2792	5,17-DiMeC27	0.06 ± 0.03	0.05 ± 0.03	0.05 ± 0.04	0.07 ± 0.04	0.09 ± 0.11	0.05 ± 0.04
3531	11-MeC35	1.33 ± 0.62	1.44 ± 0.8	1.07 ± 0.41	1.52 ± 0.76	1.75 ± 0.65	1.13 ± 0.23
3629	12-MeC36	0.32 ± 0.15	0.34 ± 0.21	0.24 ± 0.12	0.37 ± 0.21	0.41 ± 0.17	0.22 ± 0.11
3729	11-;13-MeC37	2.12 ± 0.9	2.21 ± 1.16	1.7 ± 0.6	2.42 ± 1.33	2.62 ± 0.98	1.66 ± 0.43
3751	11,15-DiMeC37	0.77 ± 0.32	0.79 ± 0.38	0.6 ± 0.18	0.89 ± 0.38	0.96 ± 0.33	0.61 ± 0.25
3774	5,15-;5,17-DiMeC37	0.91 ± 0.47	0.93 ± 0.53	0.7 ± 0.22	1.07 ± 0.56	1.11 ± 0.48	0.67 ± 0.24
3927	13-MeC39	0.81 ± 0.39	0.83 ± 0.51	0.62 ± 0.22	0.94 ± 0.62	0.98 ± 0.46	0.61 ± 0.25
3949	11,15-DiMeC39	0.62 ± 0.31	0.63 ± 0.35	0.49 ± 0.12	0.73 ± 0.39	0.73 ± 0.3	0.52 ± 0.1
3973	5,15-;5,17-DiMeC39	0.54 ± 0.28	0.56 ± 0.34	0.4 ± 0.13	0.65 ± 0.39	0.63 ± 0.31	0.43 ± 0.1

Discussion

Our study shows that the collective response towards pathogen-exposed termites varies according to the severity and timing of internal challenge, and furthermore, that these are associated with alterations to the surface chemistry of the infected individual. Notably, we demonstrate that the decision to eliminate an individual via cannibalism is triggered at an early stage after pathogen exposure, before pathogen viability or even terminal disease status is known.

Individuals that were very moribund at 15 hours following viable blastospore injection were cannibalized at a higher rate than individuals injected with an equivalent dose of dead blastospores or a control solution. These observations complement previous findings from fungus-exposed individuals

that were cannibalized when *R. flavipes* termites were close to death¹⁷. Surprisingly, however, we observed increased cannibalism in response to both viable and dead blastospore challenge as early as 2 hours post injection (p.i.), when termites did not yet display overt signs of illness. This means that cannibalism is triggered in response to individuals that are not necessarily terminally ill. This can be seen in focal termites injected with dead blastospores, which induced 40% mortality at the individual level yet almost all were eventually cannibalized by 24 hours p.i. when placed inside experimental colonies. Dead blastospore-injected termites did not display severe signs of sickness even at later experimental time points, and generally died much later than viable blastospore-injected individuals when kept in isolation. A similar pattern was observed when termites were injected with a dead blastospore solution lacking both fungal cells and intact destruxins, showing that even subcellular fungal components lacking active toxins are sufficient to elicit a strong cannibalism response. The cause of dead blastospore induced death remains unclear but represents a worthy topic for future investigation.

These results reveal that cannibalism is triggered before the onset of moribundity. This could reflect the need to react rapidly to the natural course of disease progression of entomopathogenic fungi such as *Metarhizium*, indicating that *R. flavipes* might employ a better safe than sorry strategy towards individuals suspected of internal fungal infection. *Metarhizium* deploys a multi-component infection strategy to kill the host and complete the infection cycle, including toxin release after the fungus breaches the cuticular barrier⁴⁹, so once this stage has been reached, host death is likely unavoidable under normal circumstances. There are several points during internal infection that could activate endogenous signalling pathways, which in turn may communicate a change in disease status to interacting nestmates. The first may be as soon as when blastospores have gained access to the hemocoel, as this could feasibly be linked to the rapid activation of innate immune signalling pathways. The second stage is associated with fungal acquisition of nutrients for growth and reproduction, during which time, termite hosts gradually begin to show visible signs of sickness (from 12 h p.i.). Similar symptoms have been described from the higher termite *Nasutitermes exitiosus*⁵³. Our results indicate that elimination cues are transmitted to nestmates from an early stage following cuticular breach, but also that reinforcement cues may be generated that correlate with the progression and severity of infection. Correlated with the phase of fungal proliferation, is the ramping up of toxin biosynthesis^{49,54,57,59,64} which likely accelerates host death^{46,53,57}. Blastospores might evade innate insect immune responses through reduction of the number of pathogen-associated molecular patterns (PAMPs), by camouflaging or β -glucan modification^{65,66}, but in general their interaction with the termite immune system is poorly understood, and represents an interesting target for future research. With respect to inactivated blastospores, these harbour several components that could activate the immune system of the host, including denatured cellular contents and cell wall constituents. One

factor to consider is that heat-inactivated blastospores did not contain intact destruxins. Such missing factors could represent an important form of danger signal, either via direct interaction with signalling receptors, or indirectly via their negative influence on host cells, which may then produce signals of damage⁶⁷. It is possible that the faster cannibalism response to viable blastospore-injected termites is linked to the presence and active synthesis of virulence factors such as destruxins, although this also requires further testing.

We hypothesized that activation of endogenous host pathways would result in external changes that may be perceptible by interacting nestmates. We examined cuticular hydrocarbon (CHC) profiles of internal pathogen-challenged individuals to understand whether they could serve as chemical cues indicating infection. Injections with both viable and inactivated blastospores shifted overall CHC profiles, with a trend towards more clear differentiation after a longer incubation time. We detected significant increases in four exclusively methyl-branched CHCs 15 hours after injection with viable blastospores compared to control-injected individuals. Interestingly, these four compounds are structurally related, long-chained mono- and di-methyl-branched alkanes with a similar chain length range (C35-C37) and internal methyl-branches at positions 11, 12, 13 and 15. A trend towards an increase in these compounds was also clearly discernible albeit non-significant in individuals injected with dead blastospores. These results indicate a common pattern for encoding and conveying a chemical cue associated with infection status, which would be in line with the high potential of methyl-branched alkanes to encode chemical information⁶⁷⁻⁶⁹. However, whether these compounds are directly responsible for eliciting the observed cannibalism response requires further experimental validation. Similar findings have been reported in challenged *Lasius* ants where infection has been linked with changes in CHCs of a similar chain length range (C33-35), albeit of non-methyl branched compounds¹⁹ whereas in *Myrmica* ants, straight-chain alkane quantities increased upon infection while methyl-branched alkane quantities decreased³⁴. Studies reporting cuticular profile changes associated with disease have also been carried out in honeybees^{29,32,35}. In line with these findings, our study points to host condition-dependent cues as playing an important role, with elimination behaviours potentially being activated when changes in CHC compounds exceed a given quantitative threshold⁷⁰. Our observations of increased amounts of long-chained mono- and di-methyl-branched alkanes over time following viable blastospore injection, as well as a trend towards higher quantities of these compounds in dead blastospore injected termites, may be consistent with a threshold hypothesis.

With respect to other behaviours, in comparison to previous work employing conidia on the termite cuticle^{17,18,23,40,42,57,71,72}, our observed rates of allogrooming are lower across comparable time points, suggesting that direct contact with external pathogens are more important in generating an allogrooming response. The benefits of allogrooming for enhancing group survival in the presence of

external pathogens are well documented^{14,40-42}, which makes intuitive sense as allogrooming can remove potentially infectious microbes on the surface before they breach the cuticle. The general absence of burial in our experiments supports the idea that termites preferentially eliminate harmful individuals through cannibalism^{47,73}, although burial may be a more important defensive mechanism for older or decomposing cadavers⁷³⁻⁷⁵.

Overall, our results show that termites are capable of readily detecting and eliminating individuals displaying signs of internal fungal challenge. Our findings also indicate that elimination response behaviours are triggered before individuals enter a terminal disease state, and furthermore, that individuals that could theoretically recover will nonetheless be cannibalized by their nestmates. These findings may reflect the need for a vigilant social immune system that has evolved to recognize early signs of a typically highly virulent fungal infection. In terms of the care-kill paradigm, internal challenge with fungal blastospores appears to be uniformly associated with a kill response. Trade-offs in immune system vigilance have been explored in the context of beneficial versus pathogenic microbiota⁷⁶ and similar trade-offs may also operate with regards to social immunity. Here, the level of social immune vigilance might be determined by pathogen hazard on the one hand, and cost of social immunity (damage to the colony due to elimination of constituent members as well as risk to the survival of intervening nestmates), on the other. In comparisons across species, factors such colony size and maturity, as well as individual age could play a role in shaping this trade-off. As in individual immunity, a positive correlation between pathogen hazard and social immune vigilance may be expected. It is perhaps unsurprising that *R. flavipes* does not differentiate between viable and inactivated blastospores or associated moribundity effects arising from an internal fungal challenge, because such a situation would not arise under natural conditions. Although we have not experimentally verified the cuticular chemical cues or underlying mechanisms used to trigger elimination, we speculate that they may involve crosstalk between immune and other physiological processes such as metabolism and the regulation of biosynthetic pathways involved in cuticular chemistry variation. Expansion of chemical analyses to other volatile compound classes such as alcohols (e.g. Butanol, 2,3-Butanediol) and ketones (e.g. 2-Pentanone) released by entomopathogenic fungi^{77,78} may also be warranted. Another interesting question to consider is whether modification of the cuticular profile in pathogen-challenged individuals is sufficient to induce antifungal immunity in interacting nestmates^{19,29,32,35,79}. Other targets for future work could include functional experiments to test whether specific compounds, chemical combinations, or general alterations in overall CHC abundance are sufficient to trigger changes in social immune behaviour.

Methods

Termites

Five *R. flavipes* colonies were used in these experiments: colonies 1, 5, 11+13, E and X. Additional information regarding host organisms are given in the supplementary material.

Entomopathogenic fungus

We used the semelparous fungal entomopathogen *Metarhizium robertsii* (DSM 1490), previously classified as *Metarhizium anisopliae* by the German Collection of Microorganisms and Cell Cultures GmbH (DSMZ) (<https://www.dsmz.de/collection/catalogue/details/culture/DSM-1490>). *M. anisopliae* and *M. robertsii* are closely-related obligate-killing entomopathogens of *R. flavipes*, and are detected in soils in close proximity to colonies⁸⁰. Details on fungal conidia germination and harvesting methods are given in the supplementary material.

To prepare cultured blastospores, we added 1 mL of 1×10^8 conidia/mL suspension to 100 mL quantities of two different liquid media in 300 mL Erlenmeyer flasks: (i) 40g/L yeast extract, 80g/L glucose and 0.1% Tween 80; (ii) 40g/L yeast extract, 40g/L glucose, 30g/L corn steep liquor and 0.1% Tween 80⁸¹. Erlenmeyer flasks were incubated in a shaking incubator for 3 days (exponential phase) at 25 °C and 290 rpm. Blastospores were then harvested by combining the two cultured media and filtering them through two layers of a sterile miracloth (Merck KGaA, Darmstadt, D) to remove the mycelia. The filtrate was centrifuged for 5 min at 2000g at 4 °C and the pellet containing the blastospores was washed and resuspended three times in Ringer's $\frac{1}{4}$ solution. As with conidia, blastospore concentrations were determined in a Thoma counting chamber and adjusted to 5×10^8 blastospores/mL. Experimental injections consisted of two pathogen and one control treatment. The two pathogen treatments were: viable challenge, using live blastospores (5×10^8 blastospores/mL viable blastospores; 96% germination rate at 10 hours) or inactivated challenge, using dead blastospores (5×10^8 heat-inactivated blastospores/mL; 0% germination). The control treatment consisted of sterile Ringer's $\frac{1}{4}$ solution only. To prepare the dead blastospore treatment, an aliquot of the viable blastospore suspension was autoclaved. Germination was assessed on two PDA plates streaked with 50 μ L of the autoclaved blastospore solution, followed by incubation in the dark at 25 °C for 24 hours. As expected, no fungal growth was observed. Blastospores were considered to have germinated when an elongating germ tube of any size was visible at 200-400 \times magnification⁸².

Preparation of experimental colonies

Experimental *R. flavipes* colonies were set up inside Petri dishes as described elsewhere¹⁷ and as summarized in the supplementary material.

Injection of focal termites

We marked focal termites with Nile blue, a fat-soluble stain that has been used in several studies to mark termites and has not been reported to affect termite behaviour^{17,83}. It dyes the termites blue,

making them easily distinguishable from their otherwise colourless nestmates. Nile blue dyeing was carried out following a rapid method for marking termites as described previously^{17,84}. Each focal termite was anaesthetized with CO₂, then injected with 41.4 nL of either viable blastospores, dead blastospores, Ringer's solution or fungal filtrate directly into the hemocoel using a Nanoject II (Drummond Scientific Company, USA). The needle was inserted into the side of the thorax. With this method, the first stages of the *Metarhizium* infection process, that is, adhesion, germination of fungal conidia and cuticle penetration^{85,86} were bypassed. As described above, this allowed us to test the effect of indirect cues triggered by internal pathogen challenge. As expected from previous studies^{49,55,82}, germination and killing rates are faster when using blastospores as compared to conidia (Supplementary Figures S1, S11)¹⁷. By carrying out further experiments with dead blastospore and fungal filtrate treatments, we were able to test whether active infection and terminal sickness were necessary to induce cannibalism.

Injected focal termites were kept individually in small Petri dishes (35 Ø mm), each containing a Pall cellulose pad moistened with 1 mL of distilled water and then incubated for either 2, 8, 12 or 15 hours at 27 °C and 70% humidity post injection (p.i.) before use in behavioural observation experiments. Representative images and videos of focal termite and nestmate interactions inside experimental colonies were captured with a smartphone (Samsung galaxy S7) and adjusted for brightness and size (Adobe Photoshop [Adobe Systems, San Jose, California]) to highlight different social immune behaviours (Supplementary Videos). Pathogen dose and incubation times were determined based on blastospore germination and initial survival assays of termites injected with different concentrations of *M. robertsii* blastospores (Supplementary Figures S1, S11). The effect of the selected dose of viable and dead blastospores on termite survival when kept individually was examined by injecting one hundred and eighty *R. flavipes* termites from three different colonies, each assigned to one of the following treatments: (i) 60 individuals: Viable blastospores (injected 5 x 10⁸ blastospores/mL); (ii) 60 individuals: Dead blastospores (injected 5 x 10⁸ heat-killed blastospores/mL); (iii) 60 individuals: control (injected Ringer's solution). Survival of each individual was recorded every 12 hours for a total of 240 hours (Supplementary Figure S5).

Experimental design

In our observations, termites injected with the selected dose of viable blastospores were moribund by 16 hours p.i., with death beginning between 22 and 26 hours (Supplementary Figure S5, S10 and Table S5). Knowledge of the germination and infection timeline in *R. flavipes* (Supplementary Figure S1, S2 and Table S1) was used to select four incubation timepoints for behavioural observations experiments: (i) 2 h: blastospores have gained access to the hemocoel but have not yet germinated; (ii) 8 h: blastospores have germinated but the host appears healthy; (iii) 12 h: the host shows first signs of

sickness, as indicated by slower movements; and (iv) 15 h: the host is moribund (limited to no mobility). For individuals injected with dead blastospores, symptoms do not appear until 22 h p.i., and only among individuals that subsequently died (Supplementary Figure S1 and Table S1).

Blastospore experiment. Focal termites were injected with either a suspension containing 5×10^8 viable blastospores per mL, a suspension containing 5×10^8 dead blastospores per mL, or Ringer's solution as a control. Following experimental challenge with *M. robertsii* blastospores, focal termites were kept individually for the pre-assigned incubation times (2, 8, 12 and 15 h) prior to introduction into experimental colonies. For each of the four incubation time points, we conducted 24 replicates from 5 independent colonies for each treatment. Colonies X and E contributed 3 plate replicates per incubation time point and treatment, while colonies 1, 5 and 11+13 each contributed 6. Data for colonies X, E, 5 and 11+13 were collected over 2 experimental repeats (with colony E replacing X in the second repeat) while data for colony 1 were collected from a third experimental repeat. Plates from each incubation time were randomly assigned to one of the three treatment groups after the 15-day colony establishment period had elapsed (Total N=288 plates). Randomisation was achieved by using a random number generator to produce double-digit number combinations, where the first digit corresponded to a colony and the second to one of the treatments.

Blastospore filtrate experiment. To further narrow down the pathogen components that may be responsible for inducing termite social immunity, we tested the effect of heat-inactivated fungal filtrate (lacking blastospores) on nestmate behaviour. Filtrates consisted of the final Ringer's $\frac{1}{4}$ solution used to wash dead blastospores, so should contain subcellular components and metabolites but not larger blastospore-sized structural components, which were removed by filtration. Fifty-four experimental plates of *R. flavipes* from colonies 2, 5, and X (6 plates per colony) and the respective focal termites were randomly assigned to one of the three treatment groups (Ringer, dead blastospores, or fungal filtrate) after the 15-day colony establishment period had elapsed (N=18 per treatment). Focal termites were injected either with a suspension containing dead blastospores, fungal filtrate or Ringer's solution, which served as a control. Challenged termites were all isolated for 12 hours prior to introduction into the experimental colonies. Challenged and control termites were introduced individually into the experimental colonies, which were then resealed with parafilm. The observation period began immediately after the last experimental colony was sealed.

Behavioural recording

In the blastospore experiment, after 2, 8, 12, or 15 hours of incubation, injected focal termites were added individually to the Petri dish nests. To minimize vibrational stimuli that *R. flavipes* are known to be quite sensitive to⁸⁷, Petri dishes were not opened or moved after sealing. This process took approximately 10 minutes, and the observation period began immediately after the last dish was

sealed. For observations, we adopted a scan sampling method used previously¹⁷. In the blastospore experiment, we performed scan sampling of each Petri dish nest every 5 minutes for a total of 3 hours⁸⁸. For the blastospore filtrate experiment, behavioural responses were recorded later, due to the delayed onset of cannibalism in the dead blastospore treatment. Here scan sampling was carried out for 5 minutes every 2 hours every at 12, 15, 18 and 21 hours post-injection. Scanning typically took around 1 minute. When necessary, a magnifying glass was used (up to 3x magnification) to better distinguish between observed behaviours and a Samsung S7 smartphone was employed as a digital voice recorder. Observations were performed under constant overhead light at 27 °C and 70% humidity. To minimize observer bias, the experiment was blinded by randomly assigning a treatment to each Petri dish nest prior to the observation period as mentioned in the above section. In both experiments, focal termites were counted 24 hours after introduction into the groups to quantify survivorship. Behaviours of interest were defined *a priori* and classified into visually distinguishable and non-overlapping categories relevant to social immunity. We focused on five different behavioural categories:

Groomed by n (gb): Focal termite is being groomed by N nestmates with no evidence of biting.

Cannibalism (c): Focal termite is being bitten by one or more nestmates and/or focal termite body is no longer intact.

Buried (b): Focal termite has had pieces of paper or faeces placed on top of it. Although increasingly difficult to assess, the termite may still be alive. Note that this behaviour did not get exhibited once during the observation period, which led to it being excluded from the analysis.

Not visible (nv): Focal termite is in a part of the nest where it cannot be observed.

Other (o): Focal termite is alive, intact, and unburied, but nestmates are not interacting with it. This reflects behavioural states unrelated to social immunity.

Destruxin analysis

Blastospore cultures were prepared as described above, except that Czapek Dox liquid medium was used instead (modified, Oxoid, England)⁸⁹. Viable blastospore cultures were subjected to vacuum filtration (Thermo Scientific™ Nalgene™ Rapid-Flow™, USA) to remove all fungal cells and analysed directly. Dead blastospores were generated as described above. Autoclaved blastospores in Ringer's solution (as per the dead blastospore treatment) were then subjected to vacuum filtration to remove remaining fungal cells and used in the subsequent ultra-high pressure liquid chromatography coupled with tandem mass spectrometry (UHPLC-MS/MS) analysis for confirmation of the presence of respective destruxins. For this purpose, a Synapt G2-S HDMS, equipped with an Acquity UPLC system (Waters Co., Milford, MA, USA) was used. The conditions of the mass spectrometer were optimized for small peptides, using the following settings: (+)-ESI in sensitivity mode; capillary voltage: 3,3 kV,

sample cone voltage: 40; source offset: 60; source temperature: 90 °C; desolvation temperature: 250 °C; cone gas flow: 1 L/h; desolvation gas flow: 500 L/h; nebulizer gas flow: 6 bar. Throughout analysis, UPLC grade solvents by Biosolve BV, Valkenswaard, Netherlands. The UHPLC conditions were optimized to an injection volume of 7 µL; flow rate: 0,25 mL/min; eluents: A: water + 0,1 % formic acid; B: acetonitrile + 0,1 % formic acid; column thermostat: 40 °C; gradient: 0 min: 25 % B, 10 min: 40 % B, 13 min: 40 % B, 15 min: 95 % B, 20 min: 95 % B, 21 min: 25 % B, 25 min: 25 % B. This method was adopted following Taibon⁶³. Comparison with literature (retention times and fragmentation pattern) confirmed the retention times of the destruxins as noted in the chromatograms^{62,63}.

Statistical analysis

Survival of termites kept individually

Statistical analyses were carried out in R version 4.2.2 and RStudio version 2022.07.02.

Grooming

The amount of grooming (defined as the number of grooming states/total observed states) in each treatment was analysed by fitting a generalised linear mixed model (GLMM) to the data using the *glmer* function in the R package *lme4*⁹⁰. A binomial error distribution was used to model proportion data⁹¹. Models for each incubation time (2, 8, 12, 15 h) were composed of treatment and amount of cannibalism (defined as the number of cannibalism states/total observed states) as fixed effects. Petri dish nest ID and colony were used as nested random effects (Plate within colony). The blastospore filtrate models were constructed for each timepoint (12, 15, 18, 21 h) and contained treatment, amount of cannibalism and colony as fixed effects and Petri dish nest ID as random effect. All models were compared using Akaike information criterion (*AIC*-function in R), which revealed the full model to be the best fit. Model assumptions were checked using the functions *simulateResiduals*, *testDispersion* and *testZeroInflation* contained within the *DHARMa* package⁹². We performed post hoc pairwise comparisons using the *glht* function from the *multcomp* package v1.4-10 with Tukey tests using a Bonferroni correction^{93,94}.

Cannibalism

The *survfit*-function from the *survival* package⁹⁵ was used to model the onset of cannibalistic behaviour and the *ggsurvplot*-function from the *survminer* package⁹⁶ was used to plot the data. Survival curves were compared using a mixed effects Cox model (*coxme* function from the *coxme* package⁹⁷). For the blastospore experiment, models for each incubation time (2, 8, 12, 15 h) included treatment as a fixed effect, and Petri dish nest ID and colony as random effects. For the blastospore filtrate experiment, a single model included treatment and colony as fixed effects and Petri dish nest ID as a random effect. In the survival curve analysis, all control data was initially right-censored; in order to fit a mixed effects

Cox model to the data, it was necessary to uncensor one arbitrarily-selected control replicate from each incubation time^{17,24}. The *glht*-function was used to perform post-hoc pairwise comparisons using Tukey tests with Bonferroni corrections.

Mortality of focal termites at 24 hours after introduction into experimental colonies

Following the main blastospore experiment, survival of focal termites was assessed 24 hours after introduction into experimental colonies (corresponding to 26, 32, 36 and 39 hours after injection for each incubation time point, respectively). Mortality rates were calculated for each treatment and each timepoint. A Wilcoxon rank sum test was used to perform pairwise comparisons of mortality rate ranks between treatments, using a Bonferroni-correction to account for multiple comparisons.

Scanning electron micrographs (SEMs) of blastospores

Representative SEMs of germinating blastospores on PDA media at different time points (0, 2, 4, 8, 12 and 15 h (Supplementary Figure S2)) were taken to determine the *in vitro* germination rate. For SEM preparations, small and thin pieces of PDA with blastospores were mixed 1:1 with 25% glutaraldehyde in 0.1 M cacodylate buffer pH 7-7.2 for 60 minutes. After three 5-10 minutes washing steps with 0.1 M cacodylate buffer, the samples were fixed in 1% OsO₄ in distilled water for 60 minutes and washed three times in distilled water for 5-10 minutes each. Then the blastospores were transferred into vessels covered with planktonic gauze (mesh-size 10 µm) and were dehydrated in ascending ethanol concentrations (30%, 50%, 70%, 90%, 2 x 100%, 2 x 100% absolutely water free) about 10-15 minutes each. Critical point drying with CO₂ was performed with a Balzer CPD 030. The dried samples were transferred onto double-sided adhesive tape on specimen stubs before they were sputtered with gold in a Balzer SCD 040. Images were taken with a FEI Quanta 200 ESEM.

Cuticular Hydrocarbon analysis

Extractions of pools of three individual termites per treatment were performed in 2 ml glass vials (Agilent Technologies, Santa Clara, California, USA) on an orbital shaker (IKA KS 130 Basic, Staufen, Germany) for 10 minutes. Extracts were subsequently evaporated under a constant stream of gaseous carbon dioxide and then resuspended in 5 µL of a hexane solution containing 7.5 ng/µl dodecane (C12) as an internal standard. Following this, 3 µL of the resuspended extracts were injected in splitless mode with an automatic liquid sampler (ALS) (PAL RSI 120, CTC Analytics AG, Switzerland) into a gas-chromatograph (GC: 7890B) simultaneously coupled to a flame ionization detector (FID: G3440B) and a tandem mass spectrometer (MS/MS: 7010B, all provided by Agilent Technologies, Waldbronn, Germany). The system was equipped with a fused silica column (DB-5MS ultra inert; 30 m x 250 µm x 0.25 µm; Agilent J&W GC columns, Santa Clara, CA, USA) at a temperature of 300 °C with helium used as a carrier gas under a constant flow of 1.8 ml/min. The FID had a temperature of 300 °C and used

nitrogen with a 20 mL/min flow rate as make-up gas and hydrogen with a 30 mL/min flow rate as fuel gas. The column was split at an auxiliary electronic pressure control (Aux EPC) module into an additional deactivated fused silica column piece (0.9 m x 250 μ m x 0.25 μ m) with a flow rate of 0.8 mL/min leading into the FID detector, and another deactivated fused silica column piece (1.33 m x 250 μ m x 0.25 μ m) at a flow rate of 1.33 mL/min into the mass spectrometer. The column temperature program started at 60 °C and was held for 5 min, increasing 20 °C/min up to 200 °C and then increasing 3 °C/min to the final temperature of 325 °C, held for 5 min.

CHC peak detection, integration, quantification and identification were all carried out with Quantitative Analysis MassHunter Workstation Software (Version B.09.00 / Build 9.0.647.0, Agilent Technologies, Santa Clara, California, USA). CHCs were identified according to their retention indices, diagnostic ions, and mass spectra as provided by the total ion count (TIC) chromatograms, whereas their quantifications were achieved by the simultaneously obtained FID chromatograms, allowing for the combination of the best-suited method for hydrocarbon quantification (Agilent Technologies, Waldbronn, Germany, pers. comm.) with reliable compound identifications. Absolute CHC quantities (in ng) were obtained by standardizing all peak areas of the identified CHC compounds according to the concentration of the internal C12 standard, and additionally divided by three to obtain CHC amount equivalents per single individual. A discriminant analysis (DA) was performed with the R package MASS to test whether CHC profiles statistically differ between the investigated treatment groups and to visualize the degree of separation between them. Wilk's λ was calculated to measure the quality of the DA. Furthermore, quantitative differences between the treatments for each individual CHC compound were accessed with Benjamini-Hochberg corrected Man-Whitney U tests, and visualized with boxplots where significant differences were detected.

Data Availability Statement

All data generated or analysed during this study are included in this published article and its supplementary information files.

Acknowledgments

We would like to acknowledge the assistance of the Core Facility BioSupraMol supported by the DFG. M.A.E-M. and S.J. were supported by the DAAD (Deutscher Akademischer Austauschdienst) and the CSC (Chinese Scholarship Council), respectively.

References

- 1 Wilson, E. O. & Hölldobler, B. Eusociality: origin and consequences. *Proceedings of the National Academy of Sciences* 102, 13367-13371 (2005).

637 2 Aanen, D. K. Social immunity: the disposable individual. *Current Biology* 28, R322-R324 (2018).

638 3 Cremer, S., Armitage, S. A. & Schmid-Hempel, P. Social immunity. *Current Biology* 17, R693-
639 R702 (2007).

640 4 Schmid-Hempel, P. Parasites and their social hosts. *Trends in Parasitology* 33, 453-462 (2017).

641 5 Alexander, R. D. The evolution of social behavior. *Annual Review of Ecology and Systematics* 5,
642 325-383 (1974).

643 6 Cremer, S., Pull, C. D. & Fuerst, M. A. Social immunity: emergence and evolution of colony-level
644 disease protection. *Annual Review of Entomology* 63, 105-123 (2018).

645 7 Masri, L. & Cremer, S. Individual and social immunisation in insects. *Trends in Immunology* 35,
646 471-482 (2014).

647 8 Cotter, S. & Kilner, R. Personal immunity versus social immunity. *Behavioral Ecology* 21, 663-
648 668 (2010).

649 9 De Roode, J. C. & Lefèvre, T. Behavioral immunity in insects. *Insects* 3, 789-820 (2012).

650 10 Meunier, J. Social immunity and the evolution of group living in insects. *Philosophical
651 Transactions of the Royal Society B: Biological Sciences* 370, 20140102 (2015).

652 11 Wilson-Rich, N., Spivak, M., Fefferman, N. H. & Starks, P. T. Genetic, individual, and group
653 facilitation of disease resistance in insect societies. *Annual Review of Entomology* 54, 405-423 (2009).

654 12 Pull, C. D. & McMahon, D. P. Superorganism immunity: a major transition in immune system
655 evolution. *Frontiers in Ecology and Evolution* 8, 186 (2020).

656 13 Bulmer, M. S., Franco, B. A. & Fields, E. G. Subterranean termite social alarm and hygienic
657 responses to fungal pathogens. *Insects* 10, 240 (2019).

658 14 Liu, L., Zhao, X.-Y., Tang, Q.-B., Lei, C.-L. & Huang, Q.-Y. The mechanisms of social immunity
659 against fungal infections in eusocial insects. *Toxins* 11, 244 (2019).

660 15 Wheeler, W. M. The ant-colony as an organism. *Journal of Morphology* 22, 307-325 (1911).

661 16 Conroy, T. E. & Holman, L. Social immunity and chemical communication in the honeybee:
662 immune-challenged bees enter enforced or self-imposed exile. *bioRxiv* 2020.2008. 2021.262113
663 (2021).

664 17 Davis, H. E., Meconcelli, S., Radek, R. & McMahon, D. P. Termites shape their collective
665 behavioural response based on stage of infection. *Scientific Reports* 8, 14433 (2018).

666 18 Myles, T. G. Laboratory studies on the transmission of *Metarhizium anisopliae* in the eastern
667 subterranean termite, *Reticulitermes flavipes* (Isoptera: Rhinotermitidae), with a method for applying
668 appropriate doses of conidia to trapped termites for release. *Sociobiology* 40, 277-280 (2002).

669 19 Pull, C. D., Ugelvig, L. V., Wiesenhofer, F., Grasse, A. V., Tragust, S. et al. Destructive disinfection
670 of infected brood prevents systemic disease spread in ant colonies. *Elife* 7, e32073 (2018).

671 20 Wilson-Rich, N., Stuart, R. J. & Rosengaus, R. B. Susceptibility and behavioral responses of the
672 dampwood termite *Zootermopsis angusticollis* to the entomopathogenic nematode *Steinernema
673 carpocapsae*. *Journal of Invertebrate Pathology* 95, 17-25 (2007).

674 21 Cremer, S. & Sixt, M. Analogies in the evolution of individual and social immunity. *Philosophical*
675 *Transactions of the Royal Society B: Biological Sciences* 364, 129-142 (2009).

676 22 Schmid-Hempel, P. Parasites in social insects. Vol. 60 (Princeton University Press, 1998).

677 23 Esparza-Mora, M. A., Davis, H. E., Meconcelli, S., Plarre, R. & McMahon, D. P. Inhibition of a
678 secreted immune molecule interferes with termite social immunity. *Frontiers in Ecology and Evolution*
679 8, 75 (2020).

680 24 Tragust, S., Ugelvig, L. V., Chapuisat, M., Heinze, J. & Cremer, S. Pupal cocoons affect sanitary
681 brood care and limit fungal infections in ant colonies. *BMC Evolutionary Biology* 13, 1-10 (2013).

682 25 Ugelvig, L. V., Kronauer, D. J., Schrempf, A., Heinze, J. & Cremer, S. Rapid anti-pathogen
683 response in ant societies relies on high genetic diversity. *Proceedings of the Royal Society B: Biological*
684 *Sciences* 277, 2821-2828 (2010).

685 26 Rosengaus, R. B., Traniello, J. F. & Bulmer, M. S. in *Biology of termites: a modern synthesis*
686 165-191 (Springer, 2010).

687 27 Ugelvig, L. V. & Cremer, S. Social prophylaxis: group interaction promotes collective immunity
688 in ant colonies. *Current Biology* 17, 1967-1971 (2007).

689 28 Rosengaus, R., Jordan, C., Lefebvre, M. & Traniello, J. Pathogen alarm behavior in a termite: a
690 new form of communication in social insects. *Naturwissenschaften* 86, 544-548 (1999).

691 29 Hernández López, J., Riessberger-Gallé, U., Crailsheim, K. & Schuehly, W. Cuticular
692 hydrocarbon cues of immune-challenged workers elicit immune activation in honeybee queens.
693 *Molecular Ecology* 26, 3062-3073 (2017).

694 30 Blomquist, G. J. & Bagnères, A.-G. *Insect hydrocarbons: biology, biochemistry, and chemical*
695 *ecology*. (Cambridge University Press, 2010).

696 31 Blomquist, G. J. & Ginzl, M. D. Chemical ecology, biochemistry, and molecularbiology of insect
697 hydrocarbons. *Annual Review of Entomology* 66, 45-60 (2021).

698 32 Richard, F. J., Aubert, A. & Grozinger, C. M. Modulation of social interactions by immune
699 stimulation in honey bee, *Apis mellifera*, workers. *BMC Biology* 6, 50 (2008).

700 33 Salvy, M., Martin, C., Bagnères, A. G., Provost, E., Roux, M. et al. Modifications of the cuticular
701 hydrocarbon profile of *Apis mellifera* worker bees in the presence of the ectoparasitic mite *Varroa*
702 *jacobsoni* in brood cells. *Parasitology* 122, 145-159 (2001).

703 34 Csata, E., Timuş, N., Witek, M., Casacci, L. P., Lucas, C. et al. Lock-picks: fungal infection
704 facilitates the intrusion of strangers into ant colonies. *Scientific Reports* 7, 46323 (2017).

705 35 Baracchi, D., Fadda, A. & Turillazzi, S. Evidence for antiseptic behaviour towards sick adult bees
706 in honey bee colonies. *Journal of Insect Physiology* 58, 1589-1596 (2012).

707 36 Cremer, S. Social immunity in insects. *Current Biology* 29, R458-R463 (2019).

708 37 Hamilton, C. & Bulmer, M. S. Molecular antifungal defenses in subterranean termites: RNA
709 interference reveals in vivo roles of termicins and GNBPs against a naturally encountered pathogen.
710 *Developmental & Comparative Immunology* 36, 372-377 (2012).

711 38 Kramm, K. R., West, D. F. & Rockenbach, P. G. Termite pathogens: transfer of the
 712 entomopathogen *Metarhizium anisopliae* between *Reticulitermes* sp. termites. *Journal of Invertebrate*
 713 *Pathology* 40, 1-6 (1982).

714 39 Liu, L., Li, G., Sun, P., Lei, C. & Huang, Q. Experimental verification and molecular basis of active
 715 immunization against fungal pathogens in termites. *Scientific Reports* 5, 15106 (2015).

716 40 Rosengaus, R. B., Maxmen, A. B., Coates, L. E. & Traniello, J. F. A. Disease resistance: a benefit
 717 of sociality in the dampwood termite *Zootermopsis angusticollis* (Isoptera: Termopsidae). *Behavioral*
 718 *Ecology and Sociobiology* 44, 125-134 (1998).

719 41 Yanagawa, A., Fujiwara-Tsujii, N., Akino, T., Yoshimura, T., Yanagawa, T. et al. Behavioral
 720 changes in the termite, *Coptotermes formosanus* (Isoptera), inoculated with six fungal isolates. *Journal*
 721 *of Invertebrate Pathology* 107, 100-106 (2011).

722 42 Yanagawa, A. & Shimizu, S. Resistance of the termite, *Coptotermes formosanus* Shiraki to
 723 *Metarhizium anisopliae* due to grooming. *BioControl* 52, 75-85 (2007).

724 43 Yanagawa, A., Yokohari, F. & Shimizu, S. The role of antennae in removing entomopathogenic
 725 fungi from cuticle of the termite, *Coptotermes formosanus*. *Journal of Insect Science* 9, 6 (2009).

726 44 Yanagawa, A., Yokohari, F. & Shimizu, S. Influence of fungal odor on grooming behavior of the
 727 termite, *Coptotermes formosanus*. *Journal of Insect Science* 10, 141 (2010).

728 45 Syazwan, S. A. et al. Interaction between *Metarhizium anisopliae* and its host, the
 729 subterranean termite *Coptotermes curvignathus* during the infection process. *Biology* 10, 263 (2021).

730 46 Chouvenc, T., Su, N. Y. & Elliott, M. L. Interaction between the subterranean termite
 731 *Reticulitermes flavipes* (Isoptera: Rhinotermitidae) and the entomopathogenic fungus *Metarhizium*
 732 *anisopliae* in foraging arenas. *Journal of Economic Entomology* 101, 885-893 (2008).

733 47 Rosengaus, R. B. & Traniello, J. F. Disease susceptibility and the adaptive nature of colony
 734 demography in the dampwood termite *Zootermopsis angusticollis*. *Behavioral Ecology and*
 735 *Sociobiology* 50, 546-556 (2001).

736 48 Nalepa, C. A. Nourishment and the origin of termite eusociality. In: Hunt JH, Nalepa CA (eds)
 737 Nourishment and evolution in insect societies. Westview Press, Boulder, pp 57–104 (1994).

738 49 Gillespie, Bailey, A. M., Cobb, B. & Vilcinskas, A. Fungi as elicitors of insect immune responses.
 739 *Archives of Insect Biochemistry and Physiology* 44, 49-68 (2000).

740 50 Mullen, L. M. & Goldsworthy, G. J. Immune responses of locusts to challenge with the
 741 pathogenic fungus *Metarhizium* or high doses of laminarin. *Journal of Insect Physiology* 52, 389-398
 742 (2006).

743 51 Vertyporokh, L., Hułas-Stasiak, M. & Wojda, I. Host-pathogen interaction after infection of
 744 *Galleria mellonella* with the filamentous fungus *Beauveria bassiana*. *Insect Science* 27, 1079-1089
 745 (2020).

746 52 Vilcinskas, A., Matha, V. & Götz, P. Inhibition of phagocytic activity of plasmatocytes isolated
 747 from *Galleria mellonella* by entomogenous fungi and their secondary metabolites. *Journal of Insect*
 748 *Physiology* 43, 475-483 (1997).

749 53 Hänel, H. The life cycle of the insect pathogenic fungus *Metarhizium anisopliae* in the termite
 750 *Nasutitermes exitiosus*. *Mycopathologia* 80, 137-145 (1982).

54 Iwanicki, N. S. A., Júnior, I. D., Eilenberg, J. & De Fine Licht, H. H. Comparative RNAseq analysis
of the insect-pathogenic fungus *Metarhizium anisopliae* reveals specific transcriptome signatures of
filamentous and yeast-like development. *G3 Genes/Genomes/Genetics* 10, 2141-2157 (2020).

55 Alkhaibari, A. M., Carolino, A. T., Yavasoglu, S. I., Maffei, T., Mattoso, T. C. et al. *Metarhizium*
brunneum blastospore pathogenesis in *Aedes aegypti* larvae: attack on several fronts accelerates
mortality. *PLOS Pathogens* 12, e1005715 (2016).

56 Chouvenc, T., Su, N.-Y. & Robert, A. Cellular encapsulation in the eastern subterranean termite,
Reticulitermes flavipes (Isoptera), against infection by the entomopathogenic fungus *Metarhizium*
anisopliae. *Journal of Invertebrate Pathology* 101, 234-241 (2009).

57 Chouvenc, T., Su, N. Y. & Robert, A. Inhibition of *Metarhizium anisopliae* in the alimentary tract
of the eastern subterranean termite *Reticulitermes flavipes*. *Journal of Invertebrate Pathology* 101,
130-136 (2009).

58 Kershaw, M. J., Moorhouse, E. R., Bateman, R., Reynolds, S. E. & Charnley, A. K. The role of
destruxins in the pathogenicity of *Metarhizium anisopliae* for three species of insect. *Journal of*
Invertebrate Pathology 74, 213-223 (1999).

59 Singh, D., Son, S. Y. & Lee, C. H. Perplexing metabolomes in fungal-insect trophic interactions:
a terra incognita of mycobiocidal mechanisms. *Frontiers in Microbiology* 7, 1678 (2016).

60 Liu, L., Wang, D. H., Zhao, C. C., Yan, F. M., Lei, C. L., Su, L. J., Zhang, Y. C., Huang, Q. Y., Tang,
Q. B. Transcriptomics reveals the killing mechanism by which entomopathogenic fungi manipulate the
RNA expression profiles of termites and provides inspiration for green pest management. *Journal of*
Agricultural and Food Chemistry 71, 7152-7162 (2023).

61 Altre, J. & Vandenberg, J. Comparison of blastospores of two *Paecilomyces fumosoroseus*
isolates: In vitro traits and virulence when injected into fall armyworm, *Spodoptera frugiperda*. *Journal*
of Invertebrate Pathology 78, 170-175 (2001).

62 Jegorov, A., Havlíček, V. & Sedmera, P. Rapid screening of destruxins by liquid
chromatography/mass spectrometry. *Journal of Mass Spectrometry* 33, 274-280 (1998).

63 Taibon, J., Sturm, S., Seger, C., Parth, M., Strasser, H. et al. Development of a fast and selective
UHPLC-DAD-QTOF-MS/MS method for the qualitative and quantitative assessment of destruxin
profiles. *Analytical and Bioanalytical Chemistry* 406, 7623-7632 (2014).

64 Roberts, D. W. Toxins from the entomogenous fungus *Metarrhizium anisopliae*: II. Symptoms
and detection in moribund hosts. *Journal of Invertebrate Pathology* 8, 222-227 (1966).

65 Tartar, A., Shapiro, A. M., Scharf, D. W. & Boucias, D. G. Differential expression of chitin
synthase (CHS) and glucan synthase (FKS) genes correlates with the formation of a modified, thinner
cell wall in in vivo-produced *Beauveria bassiana* cells. *Mycopathologia* 160, 303-314 (2005).

66 Wang, C. & St. Leger, R. J. A collagenous protective coat enables *Metarhizium anisopliae* to
evade insect immune responses. *Proceedings of the National Academy of Sciences* 103, 6647-6652
(2006).

67 Chung, H. & Carroll, S. B. Wax, sex and the origin of species: dual roles of insect cuticular
hydrocarbons in adaptation and mating. *BioEssays* 37, 822-830 (2015).

68 Holman, L., Jørgensen, C. G., Nielsen, J. & d'Ettorre, P. Identification of an ant queen
pheromone regulating worker sterility. *Proceedings of the Royal Society B: Biological Sciences* 277,
3793-3800 (2010).

793 69 Holze, H., Schrader, L. & Buellesbach, J. Advances in deciphering the genetic basis of insect
794 cuticular hydrocarbon biosynthesis and variation. *Heredity* 126, 219-234 (2021).

795 70 Leclerc, J.-B. & Detrain, C. Ants detect but do not discriminate diseased workers within their
796 nest. *The Science of Nature* 103, 70 (2016).

797 71 Liu, L., Wang, W., Liu, Y., Sun, P., Lei, C. et al. The influence of allogrooming behavior on
798 individual innate immunity in the subterranean termite *Reticulitermes chinensis* (Isoptera:
799 Rhinotermitidae). *Journal of Insect Science* 19, 6 (2019).

800 72 Yanagawa, A. et al. Odor Aversion and Pathogen-Removal Efficiency in Grooming Behavior of
801 the Termite *Coptotermes formosanus*. *PLOS One* 7, e47412 (2012).

802 73 Chouvenc, T. & Su, N.-Y. When subterranean termites challenge the rules of fungal epizootics.
803 *PLOS one* 7, e34484 (2012).

804 74 López-Riquelme, G. & Fanjul-Moles, M. The funeral ways of social insects. Social strategies for
805 corpse disposal. *Trends in Entomology* 9, 71-129 (2013).

806 75 Sun, Q., Haynes, K. F. & Zhou, X. Dynamic changes in death cues modulate risks and rewards
807 of corpse management in a social insect. *Functional Ecology* 31, 697-706 (2017).

808 76 Metcalf, C. J. E. & Koskella, B. Protective microbiomes can limit the evolution of host pathogen
809 defense. *Evolution Letters* 3, 534-543 (2019).

810 77 Bojke, A., Tkaczuk, C., Stepnowski, P. & Gołębowski, M. Comparison of volatile compounds
811 released by entomopathogenic fungi. *Microbiological Research* 214, 129-136 (2018).

812 78 Hummadi, E. H. et al. Volatile organic compounds of *Metarhizium brunneum* influence the
813 efficacy of entomopathogenic nematodes in insect control. *Biological Control* 155, 104527-104527
814 (2021).

815 79 Nielsen, M. L. & Holman, L. Terminal investment in multiple sexual signals: immune-challenged
816 males produce more attractive pheromones. *Functional Ecology* 26, 20-28 (2012).

817 80 Denier, D. & Bulmer, M.S. Variation in subterranean termite susceptibility to fatal infections by
818 local *Metarhizium* soil isolates. *Insectes Sociaux* 62, 219-226 (2015).

819 81 Adamek, L. Submerge cultivation of the fungus *Metarhizium anisopliae* (Metsch.). *Folia*
820 *Microbiologica* 10, 255-257 (1965).

821 82 Bernardo, C. C., Barreto, L. P., e Silva, C. S. R., Luz, C., Arruda, W. et al. Conidia and blastospores
822 of *Metarhizium* spp. and *Beauveria bassiana* s.l.: their development during the infection process and
823 virulence against the tick *Rhipicephalus microplus*. *Ticks and Tick-borne Diseases* 9, 1334-1342 (2018).

824 83 Agüero, C. M., Eyer, P.-A. & Vargo, E. L. Increased genetic diversity from colony merging in
825 termites does not improve survival against a fungal pathogen. *Scientific Reports* 10, 4212 (2020).

826 84 Evans, T. A. Fast marking of termites (Isoptera : Rhinotermitidae). *Sociobiology* 36, 517-523
827 (2000).

828 85 Butt, T. M., Coates, C. J., Dubovskiy, I. M. & Ratcliffe, N. A. Entomopathogenic fungi: new
829 insights into host-pathogen interactions. *Advanced Genetics* 94, 307-364 (2016).

830 86 Gutierrez, A. C., Machado, J. A. R., Hubner-Campos. R., Pennisi, M. A., Rodrigues, J. et al. New
831 insights into the infection of the American cockroach *Periplaneta americana* nymphs with *Metarhizium*
832 *anisopliae* s.l. (Ascomycota: Hypocreales). *Journal of Applied Microbiology* 121, 1373-1383 (2016).

833 87 Hertel, H., Hanspach, A. & Plarre, R. Differences in alarm responses in drywood and
834 subterranean termites (Isoptera: Kalotermitidae and Rhinotermitidae) to physical stimuli. *Journal of*
835 *Insect Behavior* 24, 106-115 (2011).

836 88 Altmann, J. Observational study of behavior: sampling methods. *Behaviour* 49, 227-267 (1974).

837 89 Golo, P. S., Gardner, D. R., Grilley, M. M., Takemoto, J. Y., Krasnoff, S. B. et al. Production of
838 destruxins from *Metarhizium* spp. fungi in artificial medium and in endophytically colonized cowpea
839 plants. *PLOS One* 9, e104946 (2014).

840 90 Bates, D., Mächler, M., Bolker, B. & Walker, S. Fitting linear mixed-effects models using lme4.
841 *Journal of Statistical Software* 67, 1-48 (2015).

842 91 Crawley, M. J. Statistics: an introduction using R. (John Wiley & Sons, 2014).

843 92 Hartig, F. DHARMA: residual diagnostics for hierarchical (multi-level/mixed) regression models.
844 R package version 0.4.5. Available at: <https://CRAN.R-project.org/web/packages/DHARMA> (2022).

845 93 Korner-Nievergelt, F., Roth, T., von Felten, S., Guélat, J., Almasi, B. et al. Bayesian data analysis
846 in ecology using linear models with R, BUGS, and Stan. (Academic Press, 2015).

847 94 Hothorn, T., Bretz, F., Westfall, P. & Heiberger, R. M. Multcomp: simultaneous inference for
848 general linear hypotheses. R package version, 1.4-20. Available at: [http://CRAN.R-](http://CRAN.R-project.org/package=multcomp)
849 [project.org/package=multcomp](http://CRAN.R-project.org/package=multcomp) (2022).

850 95 Therneau T. A Package for Survival Analysis in R. R package version 3.4-0. Available at:
851 <https://CRAN.R-project.org/package=survival> (2022).

852 96 Kassambara, A., Kosinski, M., Biecek, P. & Fabian, S. Kassambara, A., Kosinski, M., Biecek, P., &
853 Fabian, S. survminer: Drawing Survival Curves using 'ggplot2'. R package version 3.4.0. Available at:
854 <http://CRAN.R-project.org/package=ggplot2> (2022).

855 97 Therneau TM. coxme: Mixed Effects Cox Models. R package version 2.2-18. Available at:
856 <https://CRAN.R-project.org/package=coxme> (2022).

857