

Identifying novel molecules targeting BKCa channels as treatment for auditory impairments

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BK channels as therapeutic targets for auditory impairments in neurodevelopmental disorders: preclinical findings from the CD mouse model of Williams-Beuren syndrome

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INTRODUCTION

Auditory impairments, such as hearing loss and hyperacusis (i.e., increased sound sensitivity), are commonly described in patients with neurodevelopmental disorders (NDDs) and may exacerbate the communication and social deficits typical of these pathological conditions. In particular, both hyperacusis and progressive hearing loss are frequently associated with Williams-Beuren syndrome (WBS), a rare genetic NDD, caused by an hemizygous deletion on chromosome 7, leading to the haploinsufficience of about 30 genes. However, the cause of auditory dysfunction in WBS is still largely unknown.

Big Potassium (BK) channels play a crucial role in multiple aspects of auditory signaling and are known to be quantitatively and functionally altered in several NDDs. Hence, we tested the hypothesis that a reduction in BK channel expression and functionality underlies acoustic dysfunction in WBS and that the pharmacological activation of these channels may therefore represent an effective novel pharmacological approach.



Fig.1 CD mice showed hypo-responsiveness in the ASR compared to WT (**a,b**). This phenotypic difference became significant with higher sound intensities. Chronic treatment with CHLOR rescued the hypo-acousis of CD mice (**b**).



Fig.3 (**a**) Immunostaining of the Organ of Corti in WT and CD mice. Red indicated oncomodulin staining of the three rows of outer hair cells (OHC); green indicated otoferlin staining of the one row of inner hair cells (IHC). Images were taken from the mid-basal turn of the cochlea and revealed impaired cytoarchitecture of OHC in CD mice; no morphological alterations were detected in IHC. (**b**) Immunostaining of cochlear BK channels in IHC in WT and CD mice. Red indicated BK channels at the apical end of IHC in dénse clusters above nuclei; green indicated myosin VII staining, a citoplasmic IHC marker. Images revealed a markedly reduced channel expression in IHC in CD mice.

RESULTS

AUDITORY BRAINSTEM RESPONSE (Click-ABR) AND DISTORSION PRODUCTS OTOACOUSTIC EMISSION (DPOAE)



Fig.2 CD mice showed increased hearing threshold (a,b) and decreased wave I amplitude (c,d), especially at lowest intensities, compared to WT littermates, which together indicate hearing loss and impaired peripheral synaptic transmission. CD mice also showed reduced DPOAEs (**e,f**), suggesting outer hair cells dysfunction. Both acute and chronic CHLOR rescued wave I amplitude deficits of CD mice, but not the increased hearing threshold. Chronic treatment with CHLOR, but not acute, rescued impaired DPOAEs in CD mice.



Fig.4 Whole cell patch clamp recording of BK current in IHC of adult WT and CD mice. Outward current were generated by depolarizing stimulation from -90mV to 10mV. No difference was observed between WT and CD mice (a). BK current patch clamp recordings in IHC before and after bath application of 24µM CHLOR in WT (b) and CD (c) mice. CHLOR potentiated the BK currents only in CD mice by shifting channel voltage dependence of 10mV.

METHODS



BK CURRENTS RECORDINGS IN MOUSE INNER HAIR CELLS

reduced acoustic startle CD displayed mice increased hearing thresholds, response and recapitulating the hearing loss of WBS patients. Electrophysiological recordings also revealed a reduced wave I amplitude, suggesting a defect in the synaptic transmission at the periphery of the auditory pathway (i.e. the synaptic connection between inner hair cells and auditory nerve fibers). Moreover, DPOAEs were significantly decreased in CD mutants, indicating a defect of outer hair cells. Such impairments were indeed associated with a desorganized structure of outer hair cells and reduced BK expression in inner hair cells, as revealed by immunohistological analysis.

Both the acute and chronic administration of CHLOR, a BK agonist, rescued the behavioral and electrophysiological alterations of CD mice. In particular, chronic treatment was able to rescue the auditory hypo-responsivness of CD mutants, as well as the reduced wave I amplitude and DPOAEs.

No differences were found in BK currents between CD and WT mice. However, CHLOR administration had a potentializing effect in CD mutants only, suggesting that a higher sensitivity of BK channels may be associated with CD mutation.

Taken togheter, our results provide preclinical evidence supporting the therapeutic potential of CHLOR to treat auditory impairments in WBS and highlight the value of BK channels as potential therapeutic targets for rare NDDs.





CONCLUSION







