

## Identification of critical effect factors for prediction of spatial and intra-annual variability of shallow groundwater nitrate in agricultural areas

Chunying Wang, Xinliang Wang, Gengchen Zhang, Feifei Zhang, Junfeng Li, Shuai Chen, Sabine Sauvage, José-Miguel Sánchez-Pérez, Yuping Han, Junguo

Liu

## ► To cite this version:

Chunying Wang, Xinliang Wang, Gengchen Zhang, Feifei Zhang, Junfeng Li, et al.. Identification of critical effect factors for prediction of spatial and intra-annual variability of shallow ground-water nitrate in agricultural areas. Science of the Total Environment, 2023, 891, pp.164342. 10.1016/j.scitotenv.2023.164342. hal-04278549

## HAL Id: hal-04278549 https://hal.science/hal-04278549

Submitted on 13 Nov 2023  $\,$ 

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

# Prediction of spatial and intra-annual variability of shallow groundwater nitrate in agricultural areas

3 Chunying Wang<sup>a\*</sup>, Xinliang Wang<sup>a</sup>, Gengchen Zhang<sup>a</sup>, Feifei Zhang<sup>a</sup>, Junfeng Li<sup>b</sup>, Shuai

- 4 Chen<sup>a</sup>, Sabine Sauvage<sup>c</sup>, José-Miguel Sánchez-Pérez<sup>c</sup>, Yuping Han<sup>a</sup>, Junguo Liu<sup>a,d</sup>
- 5 <sup>a</sup> School of Water Resources, North China University of Water Resources and Electric
- 6 Power, Zhengzhou 450045, PR China;
- <sup>b</sup> College of Water Conservancy and Architecture Engineering, Shihezi University, Shihezi
  832000, PR China;
- <sup>c</sup> Laboratory of Functional Ecology and Environment, University of Toulouse, CNRS, UPS,
   Toulouse INP, ENSAT campus, F-31326 Toulouse, France
- <sup>d</sup> School of Environmental Science and Engineering, Southern University of Science and
   Technology, Shenzhen 518055, PR China;

### 13 **ABSTRACT**

14 Shallow groundwater nitrate nitrogen (NO<sub>3</sub><sup>-</sup>-N) concentrations in agricultural areas usually show high spatial and intra-annual variability. It is hard to predict such concentrations due to 15 the complexity of influencing factors (e.g., different forms of N in soil, vadose zone 16 17 characteristics, and groundwater environmental conditions). Here, a large number of groundwater and soil samples were collected monthly over two years at 14 sites to analyze 18 the soil and groundwater physiochemical properties and the stable isotopes of  $\delta^{15}N$  and  $\delta^{18}O$ 19 of groundwater NO<sub>3</sub>-N in agricultural areas. Based on field observations, a random forest 20 (RF) model was used to predict the groundwater NO<sub>3</sub>-N concentrations and reveal the 21 importance of effect factors. The results show that there are large spatiotemporal variations 22 in NO<sub>3</sub><sup>-</sup>-N,  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup>, and  $\delta^{18}$ O-NO<sub>3</sub><sup>-</sup> in groundwater. NO<sub>3</sub><sup>-</sup>-N is the major dominant specie 23

24	of inorganic N in groundwater, and the groundwater NO <sub>3</sub> <sup>-</sup> -N concentration in 24% of the
25	samples failed to meet the drinking water standard of the WHO (10 mg $L^{-1}$ ). The RF model
26	satisfactorily predicted groundwater $NO_3^N$ concentrations with $R^2$ of 0.92–0.93, RMSE of
27	3.87-4.94, and MAE of 2.10-2.89. Groundwater nitrite and ammonium are the most
28	important factors related to NO <sub>3</sub> <sup>-</sup> -N removal and production in groundwater. Denitrification
29	and nitrification were further identified by the relationships among $\delta^{15}$ N-NO <sub>3</sub> <sup>-</sup> , $\delta^{18}$ O-NO <sub>3</sub> <sup>-</sup> ,
30	and NO <sub>3</sub> <sup>-</sup> -N, and by the ranges of $\delta^{15}$ N-NO <sub>3</sub> <sup>-</sup> , $\delta^{18}$ O-NO <sub>3</sub> <sup>-</sup> , temperature, pH, DO, and ORP in
31	groundwater. Soil-soluble organic nitrogen (S-SON) and the depth of groundwater table
32	were identified as vital factors related to N sourcing and leaching. Overall, as a first
33	approach to adopting a RF model for high spatiotemporal-resolution prediction of
34	groundwater NO <sub>3</sub> <sup>-</sup> -N variations, the findings of this study enable a better understanding of
35	groundwater N pollution in agricultural areas. Optimizing management of irrigation and N
36	inputs is anticipated to reduce S-SON accumulation and mitigate the threat to groundwater
37	quality in agricultural areas.

*Keywords:* Groundwater nitrate; Effect factor; Nitrogen transformation; Random forest;
42 Nitrogen and oxygen isotopes

#### 43 **1. Introduction**

44 Groundwater is currently facing the severe challenges of depletion and deteriorating quality due to natural and anthropogenic factors around the world (Liu et al., 2016; 45 46 Famiglietti and Ferguson, 2021). Among the many groundwater problems, nitrogen (N) pollution has become a global issue due to the use of N fertilizers and manures and elevated 47 48 atmospheric deposition (Vystavna et al., 2017). Groundwater N pollution reduces N use 49 efficiency and threatens the safety of the water supply in China (Gan et al., 2022; Gao et al., 50 2022). The nitrate nitrogen ( $NO_3$ -N) contamination of groundwater is a severe problem 51 threatening the environment and human health, especially in intensively irrigated 52 agricultural areas. Accurately predicting the variations of shallow groundwater NO<sub>3</sub><sup>-</sup>N 53 concentrations at high spatiotemporal resolution is very difficult due to the complex effect factors and processes (Hinkle and Tesoriero, 2014; Biddau et al., 2019; He et al., 2022). 54 55 Therefore, exploration of the spatiotemporal patterns and any associated effect factors and 56 processes is urgently needed to prevent widespread water-quality issues around the world.

57 Climate variables, soil texture, land use, and human activities are commonly 58 investigated and recognized as the main factors affecting the inter-annual variations of 59 groundwater  $NO_3^-$ -N pollution on the large or macroscopic scale (Pennino et al., 2020; El 60 Amri et al., 2022; He et al., 2022). Nonetheless, comprehensive identification of the effect 61 factors and evaluation of their impact on dynamics of groundwater  $NO_3^-$ -N has seldomly 62 been done at a high spatial and monthly temporal resolution, especially in irrigated 63 agricultural areas. Climate variables and agricultural management activities, such as

precipitation, irrigation, and fertilizer input, cause the high level of variability in the 64 65 different forms of N content in soil and subsequent leaching to groundwater. Vadose zone 66 thickness (i.e., the depth of the groundwater table) contributes significantly to the variations 67 of groundwater NO<sub>3</sub>-N concentrations by affecting the amounts of different forms of N 68 leaching and groundwater environmental conditions (Li et al., 2021; Weitzman et al., 2022). Meanwhile, groundwater environmental parameters influence the different forms of N 69 content by impacting transformation processes. Environmental factors related to N 70 71 groundwater include temperature, dissolved transformation in oxygen (DO), 72 oxidation-reduction potential (ORP), and dissolved organic carbon (DOC). The difficulty in 73 understanding groundwater NO<sub>3</sub><sup>-</sup>N variations stems from the ability to determine the most 74 critical explanatory variables among the many factors. This question is challenging when 75 using common traditional methods, such as correlation analysis, attribution analysis, cluster 76 analysis, principal component analysis, and numerical and distributed hydrological models. 77 Overall, groundwater  $NO_3^{-}N$  concentrations are controlled by diverse factors that are 78 linked through the complex interaction of multiple processes. These processes involve 79 inorganic and organic N transformation and transport in soil and groundwater (Zhang et al., 80 2019). Surface soil N leaching to groundwater and the groundwater environment are both 81 affected by characteristics of the vadose zone. The leaching of different forms of N content from the surface soil, as the source of N in groundwater, indirectly affects groundwater 82 NO<sub>3</sub>-N dynamics through N transformation in groundwater. The N transformation in 83 84 groundwater is affected by the groundwater environment, and it increases the importance

85 and difficulty of identifying the roles of N transformation in groundwater. In terms of the N 86 in groundwater, the widely reported mineralization, nitrification, and denitrification 87 processes in groundwater can modify different forms of N species balance (Liu et al., 2022; 88 Shen et al., 2023). For instance, the mineralization and nitrification processes in 89 groundwater can produce  $NO_3$ -N, while denitrification can reduce  $NO_3$ -N. These 90 processes in groundwater play a key influence in the spatiotemporal variations of NO<sub>3</sub><sup>-</sup>N 91 dynamics in addition to N leaching in soil. Therefore, identifying the main N 92 transformation processes in groundwater and evaluating their importance will further elucidate the mechanisms of groundwater  $NO_3$ -N pollution in agricultural areas. 93

94 The complex factors and processes influencing groundwater  $NO_3$ -N concentrations lead to great difficulty in making predictions at high spatial and temporal resolution. For 95 96 instance, physically based numerical and distributed hydrological models (e.g., HYDRUS, 97 AgriFlux, and SWAT [Soil & Water Assessment Tool]) need to be coupled with 98 groundwater flow models, such as MODFLOW-MT3D and the model of Lasserre et al. 99 (1999) linked to a GIS (geographic information system), to predict groundwater NO<sub>3</sub><sup>-</sup>N 100 dynamics (Wang et al., 2016; El Amri et al., 2022). The performance of these models basically depends on an adequate understanding of hydrological behaviors and 101 102 biogeochemical processes and the availability of detailed data on the properties of the 103 vadose zone and groundwater system. These data are usually difficult to measure and collect, resulting in unsatisfactory model performance (Coppola et al., 2005). Machine 104 105 learning methods, for example, artificial neural networks, multiple logistic regressions,

106 generalized additive models, generalized linear models, support vector regressions, and 107 random forest models, have been widely used to predict the N status in agricultural and 108 natural waste waters (Chlingaryan et al., 2018; Bagherzadeh et al., 2021; He et al., 2022). 109 They were gradually accepted by researchers due to their advantages, such as their high 110 generalization ability and low cost, but most of them have the problem of overfitting 111 (Castrillo and García, 2020). Among these machine learning methods, the random forest 112 model has the advantages of strong resistance to overfitting, no feature selection, and 113 automatic data filling. It has been proven to accurately predict inter-annual groundwater NO<sub>3</sub>-N concentrations on a large scale (Band et al., 2020; He et al., 2022). However, the 114 115 random forest model has not yet been applied to predict intra-annual variations of shallow 116 groundwater  $NO_3$ -N in irrigated agricultural areas.

117 The North China Plain (NCP) is the main crop production area in China, and groundwater resources in the NCP are generally dealing with the issue of N pollution. This 118 119 study was conducted in a semiarid heavily irrigated agricultural area located in the NCP, 120 that is characterized by a shallow groundwater table depth and, thus, vulnerable to 121 groundwater pollution. Monthly meteorological data were obtained from the Xinxiang 122 weather station, which is located near the study site. Two years' worth of monthly soil physiochemical data and groundwater quality parameters were measured in the field and 123 124 laboratory. The objectives of this study were to: (1) reveal the spatial and intra-annual variations of inorganic N species and isotopic signature of NO<sub>3</sub><sup>-</sup>N in groundwater, (2) 125 126 construct a random forest model to predict spatiotemporal variations of groundwater 127  $NO_3^--N$  using a simplified approach to the model, and (3) identify the main effect factors 128 and processes, and evaluate their relationships with groundwater  $NO_3^--N$  based on the 129 constructed random forest model and isotope approaches. The ability to prediction the 130 levels of groundwater  $NO_3^--N$  pollution and uncover the mechanism that underlies its high 131 spatial and temporal variability is important for future management of agricultural N input 132 and water-quality protection in all developing and developed countries.

133 2. Materials and Methods

#### 134 2.1. Site description

This study was conducted in an irrigated agricultural area (35°00′- 35°30′ N, 113°31′-135 136 114°25' E) located in the piedmont region of Taihang mountain, NCP, near the lower reaches of the Yellow River (Fig. 1). The study site covers about 1,500 km<sup>2</sup> and belongs to 137 138 the temperate continental monsoon climate. Weather data from Xinxiang station, located 139 near our study site, were obtained from the China Meteorological Data Service Center 140 (http://data.cma.cn/). From 2010 to 2019, the average annual temperature was 15.6°C, with a 141 maximum temperature of 40°C and a minimum temperature of -13.1°C. The average annual potential evaporation was 1,025 mm yr<sup>-1</sup>, and the average annual rainfall was about 500 mm 142 yr<sup>-1</sup>. The rainfall mainly occurred from June to September. According to the aquifer data 143 144 collected by field surveys, the average annual groundwater table depth ranged from 1.9 to 145 18 m, and the groundwater level ranged from 54.1 to 82.4 m (Fig. 1a–b). The particle size of 146 the soil was analyzed using a Malvern laser particle size analyzer as reported in our previous 147 study (Wang et al., 2021). According to the international soil texture classification standard, 148 the soil in the study area was classified into silt (8.3%), silty loam (61.1%), and sandy loam 149 (30.6%) (Fig. 1c). The main crops were winter wheat and summer maize (44%), winter 150 wheat and summer peanut (27%), and winter wheat and summer rice (11%) at the study site

151 (Fig. 1d). Other land uses, including rural residential areas, urban areas, and water body areas, 152 constitute the remaining 18% of the study site (Fig. 1d). Irrigation and fertilization play 153 important roles in ensuring stable crop production and the N applied on the agricultural 154 land surface was mainly derived from chemical fertilizers, manure, and crop residues. The 155 crop residues of winter wheat and summer corn/summer peanuts were all returned to the 156 fields. The large amount and temporal variability in N input causes the groundwater N 157 content to show high spatiotemporal variations and increase continuously, which poses a serious burden on groundwater N pollution and its management. 158



159 160

Fig. 1. Location of the study area in the North China Plain and the sampling sites, and maps of (a) the 161 depth of the groundwater table, (b) the groundwater level, (c) the soil type and (d) land use. The land use 162 map was obtained from the Geospatial Data Cloud (http://www.gscloud.cn/search)

#### 163 2.2. Field monitoring, sampling and chemical analysis

164 Fourteen representative sampling sites were selected according to the geographical location, land use, soil types, and the groundwater table depth to conduct field monitoring 165 and collect surface soil and groundwater samples (Fig. 1a and Table 1). The field 166

167 monitoring and sample collection were conducted monthly from May 2017 to April 2019 at 168 the 14 observation sites (Fig. 1a). The field monitoring involved the use of a multiparameter water quality probe (HORIBA, Ltd., Japan) to determine the basic physicochemical 169 170 parameters of groundwater, including water temperature (GW-Temp), pH (GW-pH), total 171 dissolved solids concentration (GW-TDS), oxidation-reduction potential (GW-ORP), and 172 dissolved oxygen (GW-DO). At the same time, the depth of the groundwater table 173 (GW-Dep) was measured and groundwater water samples were collected using a bailer tube. The data on irrigation were collected through a survey of the local farmers. 174

Sam	pling site	Soil texture	Crop rotation	Groundwater table depth (m)	Groundwater level (m)	
	SG1	Silt	Winter wheat-summer rice	2	82.4	
	SG2	Silty loam	Winter wheat-summer corn	3.9	65.5	
	SG3	Silty loam	Winter wheat-summer corn	4.9	65.9	
	SG4	Silty loam	Winter wheat-summer corn	6	69.3	
	SG5	Silty loam	Winter wheat-summer corn	9.4	74.5	
	SG6	Silty loam	Winter wheat-summer corn	9.9	64.5	
	SG7	Silty loam	Winter wheat-summer corn	10.5	63.6	
	SG8	Silty loam	Winter wheat-summer corn	10.7	71.5	
	SG9	Sandy loam	Winter wheat-summer peanuts	12.2	60.1	
1	SG10	Silty loam	Winter wheat-summer corn	12.3	60.8	
1	SG11	Sandy loam	Winter wheat-summer peanuts	12.9	58.8	
1	SG12	Sandy loam	Winter wheat-summer peanuts	13.3	55.2	
1	SG13	Silty loam	Winter wheat-summer corn	14.8	54.1	
1	SG14	Silty loam	Winter wheat-summer corn	18	60.1	

175 **Table 1.** Soil texture, crops, depth of groundwater table, and groundwater level at sampling sites

176	Groundwater and soil samples were collected at a frequency of once a month during the
177	two-year study period. The groundwater samples were collected from wells and filtered
178	using a 0.45 $\mu$ m filter. These groundwater samples were brought back to the laboratory, and
179	1 L of each sample was stored in a refrigerator at a temperature of 2°C until physiochemical
180	analysis. At the same time, 100 ml groundwater samples were immediately frozen until

isotope analysis. Moreover, soil samples were collected from farmland within 500 m of the 181 wells where the groundwater samples were collected. The soil samples were gathered at the 182 soil surface with a depth of 0–10 cm in the farmland. All the soil samples were randomly 183 184 collected from six sites and mixed thoroughly to obtain a representative soil sample (500 g) 185 at each sampling site. Meanwhile, additional soil samples were collected in aluminum boxes that were sealed and brought back to the laboratory. The mixed soil samples were dried 186 naturally, crushed through a 2 mm sieve, and stored in a cool and dry place until 187 188 physiochemical analysis.

The soil samples stored in the aluminum boxes were further used to analyze the soil 189 water content (SWC) through the oven-drying method. The parameters measured in the 190 191 laboratory include groundwater dissolved organic carbon (GW-DOC), groundwater dissolved organic N (GW-DON), groundwater ammonium (GW-NH<sub>4</sub><sup>+</sup>-N), groundwater 192 193 nitrate (GW-NO<sub>3</sub><sup>-</sup>-N), groundwater nitrite (GW-NO<sub>2</sub><sup>-</sup>-N), soil organic carbon (SOC), soil-soluble organic N (S-SON), soil nitrate (S-NO<sub>3</sub><sup>-</sup>-N), soil nitrite (S-NO<sub>2</sub><sup>-</sup>-N), and soil 194 ammonium (S-NH<sub>4</sub><sup>+</sup>-N). All these chemical parameters of soil and groundwater were 195 196 determined by the colorimetric method using a spectrophotometer (Thermo Fisher Scientific, Inc, USA) according to the procedures reported by Hood-Nowotny et al. (2010) and Wang 197 198 et al. (2021).

Since August 2018, the influence of N leaching on groundwater has been weaker in the following eight-month dry period than in the wet period. Therefore, in this study the isotope analysis was performed on groundwater samples to identify nitrification and denitrification in groundwater from August 2018 to April 2019. The stable isotope (<sup>15</sup>N and <sup>18</sup>O) abundance of groundwater  $NO_3$ <sup>-</sup>-N was determined using the denitrifying bacteria method. A seed solution (of glycerol 500 µL + bacteria 500 µL) was shaken for 12–15 hrs, purged with N<sub>2</sub> for 3 hrs, and then added to the sample and placed in a shaking table at 100 rpm overnight, shaking and as determined by an IRMS-100 mass spectrometer. USG32, USG34, and USG35 were used as standard samples, and the results were corrected based on a two-point calibration method. The measured isotope values correspond to the international standard substances, expressed as follows:

210 
$$\delta_{sample}(\%_0) = \frac{R_{sample} - R_{VSMOW}}{R_{VSMOW}} \times 1000$$

[1]

211

where  $\delta_{sample}$  is the isotope value of the corresponding sample,  $R_{sample}$  is the ratio of heavy and light isotopic abundance of elements in the sample, and  $R_{VSMOW}$  is the ratio of heavy and light isotopic abundance of Vienna Standard Mean Ocean Water (VSMOW).

#### 215 2.3. Random forest model

216 A random forest model was used to predict the intra-annual variations of groundwater 217 NO<sub>3</sub><sup>-</sup>N at the study site. The random forest model is a machine learning algorithm based on 218 the combination of the bagging integrated learning theory and the random subspace 219 algorithm, and it overcomes the drawbacks of overfitting and instability (Breiman, 2001). 220 The model constructs several regression trees (ntree) by setting nodes (mtry) on a random 221 subset of the original training dataset, according to Amit and Geman (1997). The random 222 forest model divides the data into training and test sets by setting a certain ratio (P), and the P 223 ratio of 2:1 was chosen in this study. The random forest model was then calibrated using the 224 training subset and validated using the test subset. A package of randomForest in Rstudio 225 (version 4.2.2) was adopted to construct the random forest model.

226 The model accuracy was made to meet the research needs by adjusting the three parameters ntree, mtry, and P. Three error metrics, the root mean square error (RMSE), the 227 coefficient of determination  $(R^2)$ , and the mean absolute error (MAE), were selected to 228 evaluate the random forest model's performance. The trained and validated random forest 229 230 model was used to analyze the importance of groundwater  $NO_3$ -N influencing factors. The 231 importance of an effect factor is defined as the increase in the predicted mean squared error 232 (MSE) after randomly permuting this factor (Breiman, 2001). It reflects the contribution of each effect factor to a groundwater nitrate concentration. The normalized increased MSE, 233 that is, the relative importance, for each effect factor was between 0% and 100%. The 234 235 partial dependence shows the marginal effect of each explanatory variable for the response 236 after considering the average effects of the other variables. The partial dependence plots of the important effect factors were used to analyze the relationship between a single 237 independent variable and the groundwater NO<sub>3</sub>-N subject to the influence of the other 238 239 independent variables.

240 *2.4.* Data analysis

The spatial distribution of the measured data was interpolated using inverse distance weighting in ArcGIS to analyze the spatial distribution characteristics of different N species in groundwater from the study site. The Spearman coefficient (r), paired t-tests, and a one-way analysis of variance (ANOVA) were used to calculate the correlation coefficient and test the significance using SPSS software (version 25.0, IBM Corporation, Chicago, Illinois). Denitrification was identified based on NO<sub>3</sub><sup>-</sup>-N isotopes and their relationships, that is, the enrichment of  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> and  $\delta^{18}$ O-NO<sub>3</sub><sup>-</sup> with a slope of 0.5–1.0 ( $\Delta\delta^{18}$ O/ $\delta^{15}$ N), and a decrease of  $NO_3^--N$  concentrations. An increase of  $NO_3^--N$  concentrations and decrease of  $\delta^{15}N-NO_3^-$  and  $\delta^{18}O-NO_3^-$  were analyzed to identify nitrification.

#### 250 **3. Results and Discussion**

#### 251 3.1. Statistical analysis of groundwater nitrate concentration and its effect factors

252 Table 2 lists the variables analyzed in this study, that is, the precipitation, irrigation, soil sand content (SSC), SWC, SOC, S-SON, S-NO<sub>3</sub><sup>-</sup>-N, S-NH<sub>4</sub><sup>+</sup>-N, S-NO<sub>2</sub><sup>-</sup>-N, 253 GW-NO<sub>3</sub><sup>-</sup>-N, GW-NH<sub>4</sub><sup>+</sup>-N, GW-NO<sub>2</sub><sup>-</sup>-N, GW-DON, GW-DOC, GW-Dep, GW-Temp, 254 GW-TDS, GW-pH, GW-DO, and GW-ORP. The 19 predictor variables in Table 2 were 255 considered as effect factors that could potentially influence groundwater GW-NO<sub>3</sub><sup>-</sup>N. The 256 257 maximum, minimum and mean values; standard deviation (SD); coefficient of variation 258 (CV); and quartile of these variabiles are shown in Table 2. Different forms of N content 259 from surface soil were considered instead of N input due to the complex sources and 260 transformation processes of N input. The statistical characteristics of N input were not 261 analyzed. As shown in Table 2, all the CV values, which can reflect the size of dispersion of 262 the measured data, ranged from 0.08 to 2.06, and most of the factors had small CVs and 263 low variability. However, the CV values of groundwater inorganic N were all greater than 1, 264 indicating that spatial and temporal patterns of groundwater inorganic N concentrations were highly variable in the study area. The SD and CV of groundwater NO<sub>3</sub><sup>-</sup>N 265 concentrations were the largest compared with NH<sub>4</sub><sup>+</sup>-N and NO<sub>2</sub><sup>-</sup>-N, indicating that the 266 267 spatiotemporal variability of groundwater NO<sub>3</sub>-N concentrations was the highest among 268 the three forms of inorganic N.

269

Table 2 Summary statistics of the variables analyzed in this study (May 2017-April 2019)

	X7 · 11	M	Ma	Maaa	GD	CN	Quartiles		8
	variables	Max	Min	Mean	SD	CV	25th	50th	75th
	Precipitation (mm)	98.20	0.20	35.47	33.07	0.93	4.78	26.65	67.95
	Irrigation (mm)	300.0	0.00	96.10	93.25	0.97	0.00	105.0 0	200.0 0
	GW-Dep	19.00	0.10	10.03	4.48	0.45	5.82	10.90	13.03
	SSC (%)	71.26	12.45	31.10	16.60	0.53	21.4 0	25.70	28.2
	$\frac{SWC}{(mg mg^{-1})}$	0.42	0.01	0.16	0.09	0.54	0.09	0.16	0.22
	SOC (g kg <sup>-1</sup> )	37.19	0.75	16.80	6.83	0.41	11.5 7	16.19	21.91
	$\frac{\text{S-SON}}{(\text{mg kg}^{-1})}$	805.5 0	34.55	333.6 2	146.3 2	0.44	209. 80	335.0 6	440.3 3
Predictor variables i e	$\frac{\text{S-NO}_3 - \text{N}}{(\text{mg kg}^{-1})}$	265.1 0	6.87	53.89	47.22	0.88	25.5 7	38.00	59.64
the effect	$S-NO_2^N$ (mg kg <sup>-1</sup> )	11.40	0.01	2.74	1.80	0.66	1.68	2.28	3.15
could coutally	S-NH <sub>4</sub> <sup>+</sup> -N (mg kg <sup>-1</sup> )	62.80	6.41	18.99	7.12	0.38	14.8 9	18.49	22.25
influence	GW-pH	8.70	3.42	7.14	0.58	0.08	6.84	7.13	7.52
GW-NO <sub>3</sub> -N	GW-Temp (°C)	31.20	6.95	18.40	4.07	0.22	15.5 6	17.85	20.80
	GW-DO (mg L <sup>-1</sup> )	50.00	2.65	11.78	7.28	0.62	7.31	9.91	13.37
	GW-ORP (mV)	308.0	-136. 0	137.2 4	120.6 7	0.88	46.2 5	172.5 0	235.0 0
	$\frac{\text{GW-DOC}}{(\text{mg } \text{L}^{-1})}$	280.3 0	<lo D</lo 	42.84	43.27	1.01	13.3 5	25.83	64.60
	GW-TDS	3.17	0.22	1.15	0.65	0.56	0.73	0.95	1.30
	$\frac{(g L^{-1})}{GW-DON}$ (mg L <sup>-1</sup> )	808.3 8	<lo D</lo 	69.40	78.20	1.13	25.7 0	69.40	73.30
	$\frac{\text{GW-NO}_2-\text{N}}{(\text{mg }\text{L}^{-1})}$	0.52	<lo D</lo 	0.08	0.11	1.49	0.01	0.03	0.07
	$\frac{\text{GW-NH}_4^+-\text{N}}{(\text{mg }\text{L}^{-1})}$	22.80	0.04	1.52	1.61	1.05	0.77	1.23	1.93
Response variable	$GW-NO_3^{-1}-N$ (mg L <sup>-1</sup> )	132.2 0	0.09	8.31	14.77	1.78	1.60	3.60	9.24

270 Note: standard deviation (SD), coefficient of variation (CV), depth of groundwater table (GW-Dep), 271 soil sand content (SSC), soil water content (SWC), soil organic carbon content (SOC), soil soluble 272 organic N content (S-SON), soil nitrate content (S-NO<sub>3</sub><sup>-</sup>-N), soil nitrite content (S-NO<sub>2</sub><sup>-</sup>-N), soil 273 ammonium content (S-NH<sub>4</sub><sup>+</sup>-N), groundwater pH (GW-pH), groundwater temperature (GW-Temp), 274 groundwater dissolved oxygen content (GW-DO), groundwater oxidation-reduction potential 275 (GW-ORP), groundwater dissolved organic carbon concentration (GW-DOC), groundwater total 276 dissolved solids (GW-TDS), groundwater dissolved organic N concentration (GW-DON), 277 groundwater ammonium concentration (GW-NH<sub>4</sub><sup>+</sup>-N), groundwater nitrite concentration

278

(GW-NO<sub>2</sub><sup>-</sup>-N), groundwater nitrate concentration (GW-NO<sub>3</sub><sup>-</sup>-N), Limit of detection (LOD).

279 *3.2. Spatiotemporal variations of inorganic N species in groundwater* 

280 The temporal and spatial variations of groundwater inorganic N concentrations in the study area are shown in Figs. 2–3. There is a large difference in the temporal variations of 281 groundwater inorganic N concentrations (Fig. 2a-c). The groundwater NO3-N 282 concentrations ranged from 0.09 to 132.16 mg  $L^{-1}$ . The groundwater NO<sub>3</sub><sup>-</sup>-N concentrations 283 were higher from May to August and usually decreased from September to April of the 284 following year. In this study, the peaks of the groundwater NO<sub>3</sub>-N concentrations were 285 286 caused by high level of NO<sub>3</sub>-N leaching after fertilization, irrigation, and heavy rain in the rainy season (May-August). This result is consistent with the previous findings of Biddau 287 et al. (2019), who reported the high levels of  $NO_3^{-}$ -N concentrations (up to 162 mg L<sup>-1</sup>) in 288 289 shallow groundwater also occur when NO<sub>3</sub>-N leaching is high, particularly during fertilization and irrigation periods. The groundwater NH<sub>4</sub><sup>+</sup>-N concentrations ranged from 290 0.04 to 22.8 mg L<sup>-1</sup>. They showed a fluctuating increasing trend from May 2017 to May 291 292 2018, and the values were relatively high and stable from September 2018 to April 2019. The concentrations of groundwater  $NO_2^-$ -N ranged from 0.01 to 0.52 mg L<sup>-1</sup>, and they were 293 higher from May to October than in other months for both 2017 and 2018. Comparing the 294 295 three inorganic forms of N, the concentrations of groundwater  $NO_3$ -N were the highest, while NO<sub>2</sub>-N was the lowest among them. Overall, all three forms of groundwater 296 297 inorganic N in the second year were generally higher than in the first year (Fig. 2d-f). This 298 increase in annual NO<sub>3</sub>-N indicates that the shallow groundwater system might not be able to rapidly recover from N pollution. 299

300 The spatial variations of inorganic N concentrations in groundwater at the 14 sampling sites are shown in Fig. 3 a-f. The GW-NO<sub>3</sub><sup>-</sup>-N concentrations were high and followed the 301 302 order of SG3, SG4, SG6, SG7, and SG2, while they were lower at the other sites. Groundwater NH<sub>4</sub><sup>+</sup>-N concentrations were the highest at SG3, followed by SG2, SG7. 303 304 Groundwater NO<sub>2</sub><sup>-</sup>N concentrations were the highest at SG6, followed by SG4, SG5, SG7. 305 Overall, all the forms of groundwater inorganic N concentrations varied spatially in the 306 study area. Among them, the spatial variations of GW-NO<sub>3</sub><sup>-</sup>N concentrations were highly variable in the study area. Site SG1 showed the lowest GW-Dep and low GW-NO<sub>3</sub><sup>-</sup>N 307 concentrations (Table 1). This is likely because the soil NO<sub>3</sub>-N source was lost by 308 309 denitrification under intermittent ponding irrigation during the summer rice season. Thus, 310 the NO<sub>3</sub>-N leaching was reduced at SG1. Compared with sites SG8-SG14, the sites 311 SG2-SG4 and SG6-SG7 had lower GW-Dep and showed higher GW-NO3-N concentrations (Table 1). This could be because NO<sub>3</sub>-N leaching decreased with the 312 313 increase of vadose zone thickness (Weitzman et al., 2022). Sites SG5 and SG8-SG14 showed similar GW-NO<sub>3</sub><sup>-</sup>-N concentrations but were characterized by different GW-Dep. It 314 315 indicates that in addition to N source and leaching, other groundwater environmental factors might affect GW-NO<sub>3</sub>-N. Overall, for the GW-NO<sub>3</sub>-N concentrations, about 24% 316 of samples exceeded the World Health Organization (WHO)'s water quality standard of 10 317  $mg L^{-1}$ . 318



Fig. 2 Temporal variations of concentrations of (a) monthly nitrate, (b) monthly ammonium, (c) monthly nitrite, (d) annual nitrate, (e) annual ammonium, and (f) annual nitrite in groundwater. The different letters above the error bars in (d), (e), and (f) indicate significant difference (p<0.05) between the different years.</li>



Fig. 3 Spatial distribution of concentrations of (a) nitrate, (b) ammonium, and (c) nitrite in groundwater

332 333

#### 3.3. Spatiotemporal variations of groundwater nitrate isotopes

The  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> values ranged from -20.16‰ to 107.88‰ with a mean value of 334 11.16±16.30‰. The  $\delta^{18}$ O-NO<sub>3</sub><sup>-</sup> values ranged from -15.69‰ to 56.56‰ with a mean 335 value of 8.39±12.40‰. The temporal variations of the  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> and  $\delta^{18}$ O-NO<sub>3</sub><sup>-</sup> 336 337 values in groundwater from August 2018 to April 2019 are shown in Fig. 4a. The  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> and  $\delta^{18}$ O-NO<sub>3</sub><sup>-</sup> values showed similar temporal variation trends. 338 Groundwater NO<sub>3</sub>-N in August was heavily affected by leaching because of the 339 340 application of fertilizer and manure and wheat residue return to the field in the wet 341 season when heavy and high-frequency precipitation happens. Meanwhile, in the 342 following months (September to April of the following year), the leaching amount was reduced due to less precipitation in the dry seasons. The measured mean  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> 343 value was 26.36‰ in August 2018, and the mean  $\delta^{18}$ O-NO<sub>3</sub><sup>-</sup> value was 17.83‰ in the 344 same month. While the mean  $\delta^{15}$ N-NO<sub>3</sub> values ranged from 3.54‰ to 19.73‰ and the 345 mean  $\delta^{18}$ O-NO<sub>3</sub><sup>-</sup> values ranged 2.91% to 10.79% from September 2018 to April 2019, 346 both are lower than the measured values obtained in August 2018. During the periods 347 August-October 2018, December 2018-January 2019, and March-April 2019, the 348  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> and  $\delta^{18}$ O-NO<sub>3</sub><sup>-</sup> showed decreasing trends. The slight increase in the 349  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> and  $\delta^{18}$ O-NO<sub>3</sub><sup>-</sup> values after October 2018 (during wheat sowing) and 350 351 February 2019 (during wheat regeneration) can be attributed to the application of fertilizer, crop residue returning to the field, and irrigation. These management practices 352 caused an increase in NO<sub>3</sub><sup>-</sup>N denitrification rates due to the increase of soil NO<sub>3</sub><sup>-</sup>N, 353

354 soil water, and organic carbon in the vadose zone.

355	The spatial distribution of the $\delta^{15}$ N-NO <sub>3</sub> <sup>-</sup> and $\delta^{18}$ O-NO <sub>3</sub> <sup>-</sup> values in groundwater
356	are shown in Fig. 4b–c. The $\delta^{15}$ N-NO <sub>3</sub> <sup>-</sup> values from the SG5 (mean value of 29.55‰)
357	were obviously higher than those of the other sites (mean values ranged from 0.92‰ to
358	14.35‰) (Fig. 4d). The maximum $\delta^{15}$ N-NO <sub>3</sub> value of 107.88 ‰ was observed in
359	August 2018 at SG5. This could be because the silt loam soil could have preserved soil
360	water, and the manure application could have provided a carbon source for
361	denitrification during wet season. Thus, the NO <sub>3</sub> -N denitrification rates could be higher
362	during $NO_3^{-}$ -N leaching in the vadose zone at this silt loam site during the late wet
363	season of August. Meanwhile, the $\delta^{18}$ O-NO <sub>3</sub> values from the SG5, SG6, SG9, and
364	SG14 (mean values ranged from 13.97‰ to 18.03‰) were higher than those of the
365	other sites (mean values ranged from 2.05‰ to 8.67‰). At these sites, the $\delta^{18}$ O-NO <sub>3</sub> <sup>-</sup>
366	values were as high as $33.93\%$ to $50.69\%$ in August 2018, indicating that NO <sub>3</sub> <sup>-</sup> N in
367	precipitation was a direct source of recharge to the groundwater. This agrees well with
368	the measured $\delta^{18}$ O-NO <sub>3</sub> <sup>-</sup> values in precipitation which ranged from 35‰–59‰
369	(VSMOW) as reported by Spoelstra et al. (2001).





<sup>20</sup> 



Fig. 4 (a) temporal variations of  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> and  $\delta^{18}$ O-NO<sub>3</sub><sup>-</sup>values, spatial variations of (b)  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> and (c)  $\delta^{18}$ O-NO<sub>3</sub><sup>-</sup> values, and (d) box plots of  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> and  $\delta^{18}$ O-NO<sub>3</sub><sup>-</sup> values at 14 sampling sites

<sup>375</sup> *3.4. Prediction of spatiotemporal variations of groundwater nitrate* 

<sup>376</sup> In this study, the 19 effect factors in Table 2 were set as predictor variables, and GW-NO<sub>3</sub><sup>-</sup>N was set as a response variable when applying random forest model. Fig. 5 377 378 shows the comparison between the measured and random forest model predicted GW-NO<sub>3</sub>-N for the training dataset and test dataset. For the training dataset, the 379 simulation results yielded an  $R^2$  of 0.93, an RMSE of 4.94, and an MAE of 2.10. After 380 training, the model was validated using the test dataset, which yielded an  $R^2$  of 0.92, an 381 RMSE of 3.87, and an MAE of 2.89, indicating that the model simulation results were 382 383 good. Compared with previous results from random forest models reported by Ouedraogo et al. (2019), Pennino et al. (2020) and He et al. (2022), the model 384

constructed in this study achieved better performance in predicting  $NO_3^--N$ concentrations in groundwater. In addition, the result of this study showed better model accuracy ( $R^2$  range of 0.92–0.93) compared to the studies of Knoll et al. (2019) and El Amri et al. (2022), which used other machine learning models,e.g., classification and regression trees and artificial neural network, to assess groundwater  $NO_3^--N$  and yielded a model accuracy ( $R^2$ ) range of 0.39–0.90.



Fig. 5 Comparison between the observed and random forest model predicted groundwater nitrate
 concentrations in groundwater for the (a) training and (b) test datasets

393 The spatial and temporal distribution of the observed and random forest model 394 predicted groundwater NO<sub>3</sub>-N concentrations are shown in Fig. 6. Comparing the measured and predicted spatial and temporal distributions of groundwater NO<sub>3</sub>-N 395 concentrations, the results showed that the predicted values were generally close to the 396 measured values. Despite its remarkable performance, the random forest model may 397 398 result in a loss when predicting the extreme ends or responses beyond the boundaries of 399 the training data (Smarra et al., 2018). Therefore, predicting values beyond the range in the training data is not recommended. A representative training dataset is important for 400

assuring model performance when constructing a random forest model. In addition, it
was found that the performance of the model may be influenced by other factors, such
as the choice of a dependent variable, and independent variables, and the size of the
dataset.







409

407 Fig. 6 Comparison between (a) observed and (b) the random forest model predicted spatiotemporal
408 variation of nitrate in groundwater

410 *3.5. Importance of effect factors on spatiotemporal variations of groundwater nitrate* 

411 The random forest importance and partial dependency analysis were used to 412 calculate the importance of each influential factor on the spatiotemporal variations of 413 GW-NO<sub>3</sub><sup>-</sup>N, and to comprehensively analyze the relationship between environmental 414 factors and GW-NO<sub>3</sub><sup>-</sup>N (Figs. 7–8). As shown in Fig. 7, the key factors affecting GW-NO<sub>3</sub><sup>-</sup>-N concentrations were GW-NO<sub>2</sub><sup>-</sup>-N, GW-NH<sub>4</sub><sup>+</sup>-N, S-SON, and GW-Dep, 415 416 with relative importance of 21.46%, 6.92%, 6.91%, and 6.01%, respectively, which are 417 all higher than 5.0%. Meanwhile the relative importance of the other factors to GW-NO<sub>3</sub>-N concentrations were all less than 5.0%. The GW-DON as the substrate of 418 419 mineralization contributed an importance of 3.83%, which was less important than 420 GW-NO<sub>2</sub><sup>-</sup>-N and GW-NH<sub>4</sub><sup>+</sup>-N. Among the different surface soil N species, SON was the main influential factor on GW-NO<sub>3</sub><sup>-</sup>-N, followed by the S-NH<sub>4</sub><sup>+</sup>-N (3.32%) and 421 S-NO<sub>3</sub><sup>-</sup>N (0.69%), whereas the importance of S-NO<sub>2</sub><sup>-</sup>N (0.13%) was the smallest. The 422







434 Fig. 7 Relative importance of potential effect factors to groundwater nitrate concentrations

#### 435 *3.6. Relationship between effect factors and groundwater nitrate*

The relationships between the observed mean values of each explanatory variable,
i.e., each effect factor (Table 2) and GW-NO<sub>3</sub><sup>-</sup>-N at the 14 sites are shown in Fig. S1 in the
supporting information. Significant correlations were found between GW-NO<sub>2</sub><sup>-</sup>-N and

GW-NO<sub>3</sub><sup>-</sup>-N (r=0.84, p<0.01), between GW-NH<sub>4</sub><sup>+</sup>-N and GW-NO<sub>3</sub><sup>-</sup>-N (r=0.59, p<0.05), 439 between GW-Dep and GW-NO<sub>3</sub>-N (r=-0.68, p<0.01), between GW-TDS and and 440 GW-NO<sub>3</sub><sup>-</sup>-N (r=0.56, p<0.05), and between GW-ORP and GW-NO<sub>3</sub><sup>-</sup>-N (r=0.57, 441 442 p<0.05). However, the interactions of all the effect factors resulted in nonsignificant 443 correlations between other factors and GW-NO<sub>3</sub><sup>-</sup>-N, and led to difficulty in reflecting 444 their impacts on groundwater NO<sub>3</sub><sup>-</sup>N concentrations. Therefore, the partial dependence plots based on the random forest model were used to show the complex nonlinear 445 relationships between GW-NO<sub>3</sub>-N and each effect factor, along with the frequency 446 447 distribution (Fig. 8a-s).

#### 448 3.6.1 Different forms of N in groundwater

449 The GW-NO<sub>2</sub><sup>-</sup>-N and GW-NH<sub>4</sub><sup>+</sup>-N were significantly positively correlated with the GW-NO<sub>3</sub><sup>-</sup>-N, with an r of 0.92 (p<0.01) and 0.56 (p<0.01), respectively (Fig. 8a-b). A 450 451 positive correlation (r = 0.42, p<0.01) was identified between GW-DON and 452 GW-NO<sub>3</sub><sup>-</sup>N (Fig. 5h). However, a negative correlation was found when DON was below 100 mg L<sup>-1</sup>. The GW-NO<sub>2</sub><sup>-</sup>-N, an intermediate product of nitrification and 453 454 denitrification, is of great importance in the N transformation process and is the most 455 crucial factor as an indicator of the NO<sub>3</sub><sup>-</sup>N transformation rate. In this study, the presence or accumulation of NO<sub>2</sub>-N showed the highest impacts on groundwater 456 457 NO<sub>3</sub>-N concentrations compared to the other factors (Fig. 7). This could be due to 458 denitrification as a heterotrophic process. The previous results of Du et al. (2016) 459 indicated that denitrifying bacteria in the system preferred using NO<sub>3</sub><sup>-</sup>-N as an electron acceptor rather than  $NO_2^{-}N$ . Thus, the accumulation and production of  $NO_2^{-}N$  can 460 occur during denitrification, and NO<sub>2</sub><sup>-</sup>-N could be regarded as an index of the activity of 461 462 denitrifying bacteria. The high-throughput sequencing analysis of Du et al. (2016)

463 revealed that the genus of Thauera bacteria was dominant in the denitrifying community with high  $NO_2^-$ -N accumulation. According to the previous study, the DON retained by 464 the soil accounts for 25%-35% of the total DON, and the remaining amount enters the 465 466 groundwater with leaching (Zhou et al., 2003). The leached DON undergoes mineralization (ammonification and nitrification) and subsequently causes an increase of 467 both NH<sub>4</sub><sup>+</sup>-N and NO<sub>3</sub><sup>-</sup>-N in groundwater (Liu et al., 2022). The GW-DON can produce 468 GW-NH<sub>4</sub><sup>+</sup>-N through ammonification and then indirectly influence GW-NO<sub>3</sub><sup>-</sup>-N 469 through nitrification of GW-NH<sub>4</sub><sup>+</sup>-N (Wang et al., 2020; Liu et al., 2022). In this study, 470 the GW-NH<sub>4</sub><sup>+</sup>-N was the second important factor influencing GW-NO<sub>3</sub><sup>-</sup>-N. This could 471 472 be due to the nitrification processes, where the  $GW-NH_4^+-N$  are the substrates for 473 NO<sub>3</sub><sup>-</sup>N production by the nitrification process. During the nitrification processes, 474 because the oxidation of NO<sub>2</sub><sup>-</sup>-N to NO<sub>3</sub><sup>-</sup>-N is rapid in natural systems, the slower oxidation of  $NH_4^+$ -N to  $NO_2^-$ -N is the main process that controls  $NO_2^-$ -N production 475 476 (Nikolenko et al., 2018). Both NO<sub>3</sub><sup>-</sup>-N and NO<sub>2</sub><sup>-</sup>-N appeared when organic N 477 mineralization occurred in groundwater. The GW-NH<sub>4</sub><sup>+</sup>-N showed higher importance 478 and a greater correlation coefficient than GW-DON, which might indicate that NH<sub>4</sub><sup>+</sup>-N can be rapidly converted into  $NO_3^{-}N$ . Thus, it is likely that the GW-NO<sub>2</sub><sup>-</sup>N is an 479 480 intermediate product of denitrification in this study. Overall, the availability of  $GW-NO_2^{-}-N$  and  $GW-NH_4^{+}-N$  are indicated them as the primary factors reflecting the 481 482 denitrification and nitrification rates. Their highly ranked importance showed clear evidence that N transformation played a critical role in controlling groundwater NO<sub>3</sub>-N 483 484 variations.



489 Fig. 8 Partial dependence plots of constructed random forest model. Blue lines show the partial dependence function, and red lines show the x-axis factor frequency

491 The spatiotemporal variations of GW-NO<sub>3</sub>-N depended on different forms of N content in the soil and leaching. In terms of N forms in the soil, there are positive 492 correlations bwtween SON and GW-NO<sub>3</sub><sup>-</sup>-N (r = 0.97, p<0.01, Fig. 8c), bwtween 493 S-NO<sub>2</sub><sup>-</sup>-N and GW-NO<sub>3</sub><sup>-</sup>-N (r = 0.59, p<0.01, Fig. 8s), and bwtween S-NH<sub>4</sub><sup>+</sup>-N and 494 GW-NO<sub>3</sub><sup>-</sup>N (r = 0.33, p<0.05, Fig. 8i). This is mainly because mineralization and 495 496 nitrification play critical roles in the production of NO<sub>3</sub><sup>-</sup>N and the subsequent NO<sub>3</sub><sup>-</sup>N 497 leaching. Moreover, a higher S-SON might indicate more accumulated organic and 498 inorganic N in the vadose zone. On the other hand, another possible explanation is that the soil-leached SON,  $NH_4^+$ -N, and  $NO_2^-$ -N has undergone a transformation and 499 produced  $NO_3$ -N in the vadose zone before entering into the groundwater. Despite 500 501 the fact that S-NO<sub>3</sub><sup>-</sup>N has a direct impact on GW-NO<sub>3</sub><sup>-</sup>N, no significant correlation was found between them (r = 0.26, p>0.01) (Fig. 8r). The S-NO<sub>3</sub><sup>-</sup>-N could be divided 502 into two groups by the threshold around 30 mg kg<sup>-1</sup>. A negative correlation was found 503 when S-NO<sub>3</sub><sup>-</sup>N was below 30 mg kg<sup>-1</sup>, indicating that the lower S-NO<sub>3</sub><sup>-</sup>N could be 504 attributed to more NO<sub>3</sub>-N leaching loss. In contrast, their relationship changed to be 505 positive when S-NO<sub>3</sub><sup>-</sup>N was above 30 mg kg<sup>-1</sup>, indicating that more NO<sub>3</sub><sup>-</sup>N in 506 507 surface soil resulted in more  $NO_3$ -N leaching loss into the groundwater.

The depth from the soil surface to the groundwater table plays an essential role in the accumulation and reduction of N leaching loss in the vadose zone. The GW-Dep was significantly negatively correlated with the GW-NO<sub>3</sub><sup>-</sup>-N (r = -0.99, p<0.01) (Fig. 8d). Our results showed that a shallower groundwater table leads to an increasing amount of NO<sub>3</sub><sup>-</sup>-N in groundwater, consistent with many previous studies (Awais et al., 2021; Li et al., 2021; El Amri et al., 2022; He et al., 2022). Other characteristics of the vadose zone also affected N transformation and subsequent 515 leaching of NO3-N, including SWC, SSC, and SOC. A significant negative 516 correlation (r = -0.80, p<0.01) was found between SWC and GW-NO<sub>3</sub><sup>-</sup>-N (Fig. 81). The increase of SWC caused the decrease of NO<sub>3</sub><sup>-</sup>-N production by nitrification and 517 518 enhanced denitrification removal of NO<sub>3</sub>-N in the soil (Sexstone et al., 1985). However, it is notable that a positive correlation was identified when the SWC was 519 520 above 0.30. This could be because higher SWC increases  $NO_3$ -N leaching into the groundwater. No significant negative correlation (r = -0.56, p>0.05) was found 521 522 between irrigation and  $GW-NO_3^{-}-N$  (Fig. 8n). Meanwhile, a significant positive 523 correlation (r = 0.92, p<0.01) was identified between precipitation and GW-NO<sub>3</sub><sup>-</sup>N (Fig. 8g). Irrigation and precipitation influenced both soil water content and soil water 524 525 percolation. Irrigation mainly results in the increase of SWC but might not increase N 526 leaching loss because the higher SWC could enhance the denitrification rate, which 527 reduces NO<sub>3</sub>-N content in root zone soil (Sexstone et al., 1985). However, precipitation is more likely to increase soil-soluble N leaching into the groundwater 528 529 due to the increase of antecedent SWC by irrigation (Razzaghi et al., 2012). Thus, 530 precipitation exhibited higher importance than irrigation and SWC, as shown in Fig. 7. 531 No significant correlation (p>0.05) was observed between SSC and GW-NO<sub>3</sub><sup>-</sup>-N (Fig. 80). However, it should be noted that when SSC is around 20%, the GW-NO<sub>3</sub><sup>-</sup>N was 532 533 obviously higher; that is, the GW-NO<sub>3</sub><sup>-</sup>-N decreased with SSC when the SSC was less 534 than 60%. In contrast, the GW-NO<sub>3</sub>-N showed a slight increase when the SSC was higher than 60%. The SSC can influence SWC and the subsequent N leaching in soil. 535 Less sand content might be associated with higher antecedent SWC and more 536 537 preferential flow (Van Es et al., 2004; Razzaghi et al., 2012). Thus, the N leaching might be increase with the decrease of SSC. On the other hand, the nitrification rate 538 539 increased when the SSC was higher than 60% in sandy loam soil. Thus, the soil

540  $NO_3^--N$  content and leaching correspondingly increased. A significant positive 541 correlation (r = 0.79, p <0.01) was identified between SOC and GW-NO<sub>3</sub><sup>-</sup>-N (Fig. 8f). 542 This could be due to the higher SOC, which usually associated with higher SON in 543 soil and corresponds to more N leaching. Nevertheless, when SOC was below 15 g 544 kg<sup>-1</sup>, it had a minor impact on GW-NO<sub>3</sub><sup>-</sup>-N.

### 545 3.6.3 Groundwater environmental conditions

The DO and ORP are indicators of redox status in the groundwater environment. 546 In this study, DO usually ranged from 4 to 20 mg  $L^{-1}$ . It is generally believed that the 547 appropriate DO concentration for denitrification is 2 mg  $L^{-1}$  (Peng et al., 2020). 548 Denitrification still exists when the DO concentration of groundwater is  $2 \sim 6 \text{ mg L}^{-1}$ , 549 550 but the rate is reduced with higher DO (Peng et al., 2020). The average monthly DO concentration in the study area from May 2017 to April 2019 was 11.78 mg L<sup>-1</sup>, and 551 there was no record of below 2 mg  $L^{-1}$  values in any of the samples. Instead, the DO 552 values in 14.6% of the 336 samples were between about 2 and 6 mg  $L^{-1}$ , mainly in 553 April and August–October. Overall, a significant positive correlation (r = 0.96, 554 p<0.01) was identified between GW-DO and GW-NO<sub>3</sub><sup>-</sup>-N (Fig. 8m), which could be 555 due to nitrification. Meanwhile a negative correlation was identified when DO was 556 less than 10 mg  $L^{-1}$  (Fig. 8m). This could be due to stronger denitrification than 557 nitrification. An increase of GW-NO<sub>3</sub><sup>-</sup>-N with ORP is observed when ORP is above 558 559 200 mV. This could be due to the positive correlation between ORP and actual nitrification (Bohrerova et al., 2004). No significant correlation (p>0.05) was found 560 between GW-ORP and GW-NO<sub>3</sub><sup>-</sup>N (Fig. 8k). When ORP was less than -100 mV, it 561 562 is notable that there was a negative correlation, while a positive correlation was identified when ORP was great than 100 mV. Nevertheless, when the ORP was 563 between -100 mV and 100 mV, it had a minor impact on GW-NO<sub>3</sub><sup>-</sup>N. According to 564

565 the relationships between GW-NO<sub>3</sub><sup>-</sup>N and DO and between GW-NO<sub>3</sub><sup>-</sup>N and ORP based on the partial dependence plots of the random forest model (Fig. 8), the 566 environment of groundwater can be classified as being under nitrate-reducing 567 conditions if the DO is less than  $10 \text{ mg L}^{-1}$  and the ORP is less than -100 mV. On the 568 other hand, the groundwater environment can be described as being and under 569 nonreducing conditions if the DO is greater than 10 mg  $L^{-1}$  and the ORP is greater 570 than 100 mV. This is in accordance with previous studies (Rivett et al., 2008; Jahangir 571 et al., 2017; Thayalakumaran et al., 2008). Groundwater DO and ORP are shown to 572 be pH dependent. Thus, the nitrification and denitrification rates could vary with 573 changes in pH (Bohrerova et al., 2004). The GW-pH showed a significantly negative 574 575 correlation with GW-NO<sub>3</sub><sup>-</sup>-N (r = -0.28, p<0.05) (Fig. 8q). The groundwater NO<sub>3</sub><sup>-</sup>-N 576 decreased with an increasing trend of pH from 6.0 to 7.0, which could be attributed to denitrification. Meanwhile groundwater NO<sub>3</sub>-N increased with pH from 7.0 to 8.0, 577 578 which could be attributed to nitrification. The results of this study are consistent with 579 previous studies, which reported that the highest denitrification removal of NO<sub>3</sub>-N was at a pH of 6.0–7.5 and the most adapted pH range for nitrifying bacteria is 7.0 to 580 9.0 (Ghafari et al., 2009; Hu et al., 2018; Bergamasco et al., 2019). A drastic increase 581 of NO<sub>3</sub>-N is observed when pH was above 8.0. This could be attributed to soil water 582 percolation to shallow groundwater following fertilization and irrigation during the 583 584 rainy season (Jendia et al., 2020).

The DOC is considered as the source of electron donors for denitrification (Thayalakumaran et al., 2008). Overall, there was no significant correlation (r = -0.05, p>0.05) observed between GW-DOC and GW-NO<sub>3</sub><sup>-</sup>-N (Fig. 8p). A negative correlation was found when DOC was below 75 mg L<sup>-1</sup>, while a positive correlation was found when DOC was above 75 mg L<sup>-1</sup>. A concentration value of DOC > 1.0 mg

 $L^{-1}$  can indicate the presence of electron donors significant enough to support 590 denitrification (Rivett et al., 2008; Thayalakumaran et al., 2008). The DOC 591 concentrations of the 336 samples observed in this study ranged from 0.42 to 280.26 592 mg  $L^{-1}$ , with more than 99% of samples exceeding 1.0 mg  $L^{-1}$ . The negative 593 relationship between GW-NO<sub>3</sub><sup>-</sup>N and DOC indicates the role of DOC as a carbon 594 source in the reduction processes (Rivas et al., 2017). The other electron donors, such 595 as  $Fe^{2+}$ ,  $Mn^{2+}$ , and  $S^{2-}$ , might also exist in groundwater and influence N 596 transformation, but their roles are not dependent on temperature, DO, and ORP (Pang 597 598 and Wang, 2021). A significant positive correlation (r = 0.94, p<0.01) was found between GW-TDS and GW-NO<sub>3</sub>-N (Fig. 8i). The GW-TDS was usually higher than 599  $0.5 \text{ g L}^{-1}$ ; however, when it was lower than this value, there was a negative correlation 600 between them. The other electron donors, rather than DOC, might have played critical 601 roles when TDS was below 0.50 g  $L^{-1}$ , causing the negative correlation between 602 GW-NO<sub>3</sub><sup>-</sup>N and TDS. However, when it was higher than 0.5 g  $L^{-1}$ , a positive 603 604 correlation was found between  $NO_3$ -N and TDS. This could be because the growth of denitrification bacteria is reduced with the increase of salinity. A previous study 605 reported that denitrification was more sensitive to salt compared to nitrification 606 607 (Dincer and Kargi, 1999). Besides, it could also be because the groundwater TDS was increased by a large amount of  $NO_3$ -N leaching. 608

The groundwater temperature controls microbial activity. Overall, a positive correlation (r = 0.62, p<0.01) was found between GW-Temp and GW-NO<sub>3</sub><sup>-</sup>-N (Fig. 8e), which could be due to higher N leaching to groundwater in the summer period. When the GW-Temp was below 20°C, a negative correlation was found between GW-Temp and GW-NO<sub>3</sub><sup>-</sup>-N. This could be because the increased temperature

enhances the growth and activity of denitrifiers, thus, reducing NO<sub>3</sub>-N in 614 groundwater (Singh et al., 2010). The groundwater temperature also impacts other 615 environmental factors, such as DO and DOC availability (Thayalakumaran et al., 616 2008; Nikolenko et al., 2018). The DOC can be stimulated, and the DO can be 617 618 depleted by increased groundwater temperature (Guo et al., 2017). The suitable temperature for the nitrification rate is 10°C-35°C, with the optimum temperature 619 ranging between 25°C-30°C (Hayatsu and Kosuge, 1993). In contrast, the suitable 620 temperature for denitrification is 20°C–43°C (Strous et al., 1999). In general, higher 621 temperatures result in stronger denitrification and nitrification activity in the 622 groundwater environment with temperatures less than 25°C. The optimum 623 624 temperature for denitrification is relatively higher than that for nitrification 625 (Nikolenko et al., 2018).

Although the relative importance of groundwater environmental factors (i.e., 626 temperature, DO, ORP, DOC, TDS, and pH) to groundwater NO<sub>3</sub><sup>-</sup>N were less than 627 5.0% (Fig. 7), the partial dependence plots generated by the random forest model 628 suggested that they play important roles in influencing nitrification and/or 629 denitrification in groundwater (Fig. 8). The groundwater NO<sub>3</sub><sup>-</sup>-N,  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> and 630  $\delta^{18}$ O-NO<sub>3</sub> exhibited tremendous changes in the summer periods in this study, 631 probably due to simultaneous leaching and denitrification (Figs. 2-4). The 632 nitrification rate also increases with an increase in temperature. In November, January, 633 and February, when the temperature is low, the DO and ORP increase, and less DOC 634

is consumed, indicating a more oxidizing condition. During these months, denitrifying
bacteria growth might be limited by the low temperature (Nikolenko et al., 2018).
Thus, the denitrification is low, and nitrification might superimpose on denitrification
during these low-temperature periods.

#### 639 3.7. Identification of key N transformation processes in groundwater using isotopes

The simultaneous increase of  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> and  $\delta^{18}$ O-NO<sub>3</sub><sup>-</sup> with a slope of 0.5-1.0 640  $(\delta^{18}O/\delta^{15}N)$  is attributed to denitrification (Osaka et al., 2010; Wells et al., 2016) (Fig. 641 9). If strong denitrification occurs in groundwater, the slope of the linear relationship 642 (k) between  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> and  $\delta^{18}$ O-NO<sub>3</sub><sup>-</sup> is close to 0.5, and  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> in groundwater is 643 significantly negatively correlated with ln[NO<sub>3</sub><sup>-</sup>] (Zhang et al., 2019). Among the 14 644 observation sites in the study area, the linear relationship slope (k) of  $\delta^{15}$ N-NO<sub>3</sub> and 645  $\delta^{18}$ O-NO<sub>3</sub> in 8 sites, that is, SG1 (k = 0.98, R<sup>2</sup> = 0.85), SG3 (k = 0.83, R<sup>2</sup> = 0.98), 646 SG4 (k = 0.50,  $R^2 = 0.047$ ), SG6 (k = 0.87,  $R^2 = 0.87$ ), SG8 (k = 0.81,  $R^2 = 0.12$ ), 647 SG13 (k = 0.59,  $R^2$  = 90), SG14 (k = 0.82,  $R^2$  = 0.70), and SG11 (k = 1.05,  $R^2$  = 0.50), 648 ranged from 0.5 to 1.0 and with high  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> composition, strongly supporting the 649 occurrence of denitrification in the NO<sub>3</sub>-N source and/or transformation in 650 groundwater (Fig. 9). The k value at SG7 (0.30,  $R^2 = 0.64$ ), SG5 (0.44,  $R^2 = 0.84$ ), 651 and SG10 (0.34,  $R^2 = 0.22$ ) did not fall into the denitrification line. However, 652 relatively higher  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> and  $\delta^{18}$ O-NO<sub>3</sub><sup>-</sup> than the potential NO<sub>3</sub><sup>-</sup>-N sources 653 indicated that weak denitrification still existed. The  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> and  $\delta^{18}$ O-NO<sub>3</sub><sup>-</sup> at SG1, 654 SG4, SG10, SG11, and SG12 were relatively smaller, indicating weaker 655 denitrification. In this study, the groundwater  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> was barely found to be 656 657 negatively correlated with  $\ln[NO_3]$ . It is likely that the isotopic fingerprint of 658 denitrification was simply masked by the pulse inputs of leached NO<sub>3</sub><sup>-</sup>-N from the

vadose zone. In addition, it might also be because shallower groundwater has a short residence time. Thus, transport of  $NO_3^--N$  affects the isotopic fingerprint of denitrification.





664

665

Fig. 9. Relationship between  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> and  $\delta^{18}$ O-NO<sub>3</sub><sup>-</sup> in groundwater (the \* indicates the range of nitrified  $\delta^{18}$ O-NO<sub>3</sub><sup>-</sup> within the dotted line, the 1:2 broken line indicates strong denitrification, and the 1:1 broken line indicates nitrification and denitrification)

666 Nitrification will simultaneously increase NO<sub>3</sub><sup>-</sup>N concentrations and reduce the abundance of  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> due to the isotope fractionation. The  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> and 667  $\delta^{18}$ O-NO<sub>3</sub> vary along the 1:1 line and, with an R<sup>2</sup> closer to 1, indicates that the 668 nitrification occurred concurrently with denitrification (Fig. 9). Moreover, 669 nitrification in the groundwater environment resulted in the final  $\delta^{18}$ O-NO<sub>3</sub><sup>-</sup> values 670 being mostly between -10‰ and 10‰ (Xue et al., 2009; Ji et al., 2021). In this study, 671 the  $\delta^{18}$ O-NO<sub>3</sub> values of 64.28% of the 182 measured data were between -10% and 672 10‰, indicating that nitrification also existed in the shallow groundwater of the study 673 area. The NO<sub>3</sub><sup>-</sup>-N is expected to be mainly from nitrifying processes since  $\delta^{18}$ O-NO<sub>3</sub><sup>-</sup> 674 values in 80% of the samples the ranged from -5% to 5%, which is fits within the 675 range of -6.1‰-5.2‰ (Osaka et al., 2010). The relationship between the monthly 676

 $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> and NO<sub>3</sub><sup>-</sup>-N (from September 2018 to April 2019) in the 14 sites is shown 677 in Fig. 10. Nitrification along the groundwater flow path was identified, including in 678 the flow path along L1 surrounded by sites SG5, SG7, SG3, and SG1 and in the flow 679 680 path L2 surrounded by sites SG8, SG10, SG11, SG12, and SG13 (Fig. 10). Data with negative  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> abundance in September 2018, January 2019, and April 2019 681 682 were excluded from SG11 because the site was irrigated and fertilized in those months. The more negative  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> and  $\delta^{18}$ O-NO<sub>3</sub><sup>-</sup> ranges measured fell within the 683 expected range for  $NO_3$ -N produced from urea/urine (Wells et al., 2016). Wang et al. 684 (2020) has reported that nitrification occurred in the groundwater, which is consistent 685 with results of this study. 686

687 Along the groundwater flow path L1, nitrification increased NO<sub>3</sub><sup>-</sup>-N concentrations by about 10 mg  $L^{-1}$ . Meanwhile along the groundwater flow path L2, 688 nitrification increased NO<sub>3</sub><sup>-</sup>-N concentrations by about 2 mg  $L^{-1}$ . Higher nitrification 689 690 rates along flow path L1 were observed. This is due to the shallower and decreased 691 depth of the groundwater table along the groundwater flow direction (from 9.37 to 3.86 m). On the other hand, along flow path L2, the depth of the groundwater table 692 increased from 2.0 to 14.8 m. The denitrification rates along flow path L1 were also 693 stronger, as evidenced by the higher  ${}^{15}$ N-NO<sub>3</sub> at sites around flow path L1. This is 694 because the temperature and contents of electron donors (DOC and TDS) at sites 695 696 around flow path L1 were higher than those around flow path L2. Therefore, both denitrification and nitrification were higher in shallower groundwater around flow 697 path L1 than flow path L2. Overall, based on the DO, ORP, different forms of N 698 699 concentrations, and nitrate isotopic composition, it can be concluded that nitrification 700 occurred concurrently with denitrification in this shallow groundwater system. The 701 co-occurrence of denitrification and nitrification found in this study is consistent with







Fig. 10. (a) Locations of sampling sites along the groundwater flow path lines (L1 and L2) or the
groundwater-level contour line (L3) and the relationship between monthly average (from
September 2018–April 2019) groundwater δ<sup>15</sup>N-NO<sub>3</sub><sup>-</sup> and NO<sub>3</sub><sup>-</sup>-N concentrations at the 14
sampling sites along (b) L1, (c) L2, and (d) L3 (SG11\*: data with negative δ<sup>15</sup>N-NO<sub>3</sub><sup>-</sup> abundance
in September 2019, January 2019, and April 2019 were excluded from SG11)

Site SG1 was located upstream of groundwater flow with the shallowest depth of the groundwater table. However,  $NO_3$ -N concentration at SG1 ranged between SG8 and SG12 as shown in Fig. 10.This is because the irrigation water source was from the Yellow River at SG1, while groundwater was abstracted for irrigation at other

sites. Moreover, the groundwater at SG1 might have received NO<sub>3</sub>-N from vertical 716 717 and lateral recharge of the Yellow River. It is notable, though, that the irrigation schedule during the summer rice growth period at SG1 was different from that of the 718 other sites. High  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> and low NO<sub>3</sub><sup>-</sup> concentration values were found at SG14. 719 720 The groundwater table depth of SG14 was the deepest and was caused by groundwater abstraction for paper production in factories, according to the Xinxiang 721 City Water Resources Bulletin in 2017 (XCWRB, 2017). The deep groundwater table 722 723 meant that NO<sub>3</sub>-N might have already been denitrified before arriving in the groundwater (Vidon and Hill, 2004). As for SG9, there was a river nearby where 724 725 surface water could recharge the groundwater. The mean DO concentration was about 14.6 mg  $L^{-1}$  at SG9, while it ranged from 10.2 to 12.8 mg  $L^{-1}$  at the other sites. The 726 relatively higher DO at SG9 than the other sites could support the reasoning that 727 surface water existed recharge to the groundwater. The denitrification occurred 728 underneath the riverbed, and the denitrified NO<sub>3</sub>-N continuously flowed to the 729 groundwater at SG9. In addition, the DOC was lower at SG9 (41.9 mg  $L^{-1}$ ) than at the 730 neighboring sites of SG11 (45.8 mg  $L^{-1}$ ) and SG 12 (45.2 mg  $L^{-1}$ ) because DOC was 731 consumed by denitrification underneath the riverbed. Therefore, the isotopic 732 733 composition and proportion of NO<sub>3</sub><sup>-</sup>N sources at SG9 differed from that of the other sites. The sites SG6 and SG4 near L3 in Fig. 10 lay along the groundwater water-level 734 contour line. The value of  $\delta^{15}$ N-NO<sub>3</sub><sup>-</sup> at SG6 was higher than that at SG4. The 735 736 groundwater table depth at SG6 was slightly higher than that of SG4. In addition, the concentration of DOC in the groundwater of SG6 was higher than that at SG4. 737 Therefore, the stronger denitrification at SG6 could be attributed to the sufficient 738 739 carbon sources in the groundwater to support denitrification. Furthermore, the deeper vadose zone thickness could provide enough time for denitrification before enteringthe groundwater.

#### 742 **4. Implications and Limitations**

#### 743 *4.1. Groundwater nitrate pollution risk and attenuation in agricultural areas*

The spatiotemporal variations of groundwater NO3-N concentrations can be 744 745 explained by inorganic and organic N leaching and subsequent N transformation in 746 groundwater. A shallower groundwater table is more vulnerable to N contamination. 747 The deeper the vadose zone, the less likely contaminants are coming from leaching. 748 However, continuous organic and inorganic fertilizer application under the combined effects of irrigation and precipitation can result in massive N accumulation in the deep 749 750 vadose zone, and the peak of N content in soil contentiously moves downward to 751 groundwater (Weitzman et al., 2022). The inorganic and organic N in groundwater 752 located in a region with both a shallower groundwater table and high groundwater 753 level can be transported to a lower groundwater water-level region. In addition, the 754 groundwater is undergoing nitrification along the groundwater-level gradient. Thus, 755 slow DON mineralization and nitrification might continuously increase NO<sub>3</sub><sup>-</sup>-N along 756 the groundwater flow path, threatening groundwater quality for a long time. Therefore, the soil DON and NH<sub>4</sub><sup>+</sup>-N leaching and the subsequent transformation in groundwater 757 758 should not be overlooked when evaluating groundwater N pollution and making 759 nonpoint source control policies. In our research, the groundwater flow could export 760 the N out of the study area, thus, contributing to the temporal decrease of  $NO_3$ -N in the local groundwater while also posing as a groundwater pollution threat to 761 762 neighboring areas.

763 The rapid decrease of  $NO_3^--N$  during May–August indicated that the leached 764  $NO_3^--N$  was consumed immediately by denitrification. The temporal increases of

40

765 NO<sub>3</sub><sup>-</sup>N were observed from September 2017 to January 2018 and from January 2019 766 to March 2019. In addition, the increase of NO<sub>3</sub><sup>-</sup>N occurred along the groundwater 767 flow during September 2018–April 2019. These results suggested that the NO<sub>3</sub>-N 768 accumulation might occur under low groundwater temperature conditions. This is mainly due to the denitrification rate decreasing at lower temperatures in groundwater 769 770 (Singh et al., 2010). Therefore, sufficient DOC must be provided for the efficient 771 removal of groundwater NO<sub>3</sub>-N when the groundwater temperature is suitable or 772 optimum for denitrification, particularly in late spring, summer, and early autumn. In 773 the future, quantifying nitrification and denitrification rates at different time scales 774 (e.g., daily, monthly, seasonally, or annually) is necessary to put forward N 775 attenuation measures. Agronomists recommend the application of organic fertilizer 776 and crop residue return with reduced chemical fertilizer in China and around the 777 world to increase soil quality (Zhao et al., 2016). Management of organic fertilizers 778 and crop residue is as crucial as chemical fertilizer application for reducing the risk of 779 SON accumulation and subsequent groundwater and surface water pollution while providing enough DOC for denitrification. Furthermore, scientists and policy makers 780 781 should pay attention to the leaching risk of stocked N and evaluate its transport delay time to protect groundwater quality in agricultural areas. 782

783 4.2. Implications of groundwater nitrate prediction and limitations of this study

The performance of the random forest model demonstrates that it can be well applied for predicting spatial and intra-annual variations of groundwater nitrate concentrations. The importance analysis for the influencing factors likewise provides strong evidence for determining the main influencing factors and processes of  $NO_3^-$ -N in groundwater. In the NCP irrigation area, the effect of N transformation in 789 groundwater is as important as soil N leaching on the variations of NO<sub>3</sub><sup>-</sup>N in shallow groundwater. The findings of this study can provide technical support for the rapid 790 prediction and evaluation of N pollution in shallow groundwater through readily 791 792 available effect factors at higher spatial and temporal resolution. The simplified 793 approach of the random forest model can be applied in irrigation regions around the 794 world to predict the spatial and temporal variations of NO<sub>3</sub><sup>-</sup>-N in groundwater. Unlike  $NO_3^-N$ , adsorption/desorption processes dominate DON and  $NH_4^+-N$  transport in the 795 796 soil-groundwater systems (Vandenbruwane et al., 2007; Wang et al., 2021). Therefore, other factors related to adsorption/desorption processes are required to predict 797 spatiotemporal variations of DON and NH<sub>4</sub><sup>+</sup>-N concentrations in groundwater using 798 799 machine learning models (Wang et al., 2016; Wang et al., 2021).

The N leaching and transformation in groundwater, especially denitrification and 800 nitrification, must be considered in other numerical and/or distributed groundwater 801 802 flow models to accurately predict NO<sub>3</sub><sup>-</sup>N dynamics. The NO<sub>3</sub><sup>-</sup>N in groundwater tends to be the most sensitive to NO<sub>2</sub>-N, indicating that biotic factors play critical 803 804 roles in controlling the N transformation (Yang et al., 2012). These biochemical processes cause the difficulty and challenge of building and verifying groundwater 805 flow and solute transport models. It is essential to study microbial communities and 806 807 growth response to changes in N and C substrates and environmental factors to gain 808 better insights into N transformation processes and rates (Nikolenko et al., 2018). It should be noted that except for the commonly considered nitrification and 809

810 denitrification processes identified in this study, other N transformation processes in shallow groundwater are also possible. For instance, NO2-N accumulation in 811 denitrification can provide the substrate for anammox (Smith et al., 2015), DNRA 812 813 (Jahangir et al., 2017), and shortcut nitrification-denitrification (Hu et al., 2018). Therefore, it is necessary to measure the N transformation rates using the <sup>15</sup>N tracing 814 815 method, and analyze microbial communities based on in situ experiments and 816 laboratory simulation of the groundwater environment to precisely understand the fate of N in groundwater in future studies. 817

#### 818 **5.** Conclusions

819 Nitrate as the primary source of inorganic N pollutants in shallow groundwater, 820 showed highly variable spatiotemporal patterns in an irrigated agricultural area, the NCP. Monthly soil physiochemical data, groundwater quality data, and groundwater 821 822 NO3<sup>-</sup>N isotopic composition data were measured and analyzed from May 2017 to 823 April 2019 in the study area. This research focused on predicting spatial and 824 intra-annual variations of shallow groundwater NO<sub>3</sub>-N and evaluating their main 825 effect factors and processes. The random forest model, as a simplified approach compared to numerical and distributed hydrological models, performed well in 826 predicting the spatial and intra-annual variations of  $NO_3^{-}N$  in groundwater with  $R^2$ 827 value of 0.93 and 0.92, RMSE value of 4.94 and 3.87, and MAE values of 2.10 and 828 829 2.89 for the training and test datasets, respectively. The application of the random forest model may have important implications for groundwater NO<sub>3</sub>-N prediction in 830 831 other regions worldwide. Evaluation of the importance of the effect factors emphasized the roles of N transformation, including denitrification and nitrification, 832

833 in controlling spatiotemporal variations of NO<sub>3</sub><sup>-</sup>-N in addition to N leaching. The 834 impacts of soil SON on groundwater  $NO_3$ -N was the greatest among different forms 835 of N in surface soil. The temporal increase of NO<sub>3</sub>-N was mainly attributed to 836 NO<sub>3</sub>-N leaching, while the temporal decrease of NO<sub>3</sub>-N can be attributed to denitrification. Although the N leaching usually decreases with the increase of 837 838 groundwater table depth, nitrification along the groundwater flow path will adversely 839 affect groundwater quality in deep vadose zone areas. Monitoring and management of 840 SON in soil and DON in groundwater will become increasingly important since 841 agronomists and policy makers widely recommend the use of manure instead of chemical fertilizers in developing and developed countries. The groundwater NO<sub>3</sub>-N 842 843 is significantly related to nitrite which is an indicator of biotic factors, but 844 groundwater NO<sub>3</sub><sup>-</sup>N is less sensitive to other environmental factors. The prediction of 845 groundwater NO<sub>3</sub>-N dynamics using hydrological or biochemical models should 846 address all forms of N leaching and transformation in groundwater, which is still a 847 considerable challenge due to the involvement of biological factors. The ability to identify the N transformation process and quantify the N transformation rates is 848 urgently needed and deserves further attention through the adoption of 849 microbiological techniques and <sup>15</sup>N tracing methods based on in situ and laboratory 850 851 groundwater experiments.

852 **Declaration of Competing Interest:** The authors declare no conflict of interest.

**Data availability:** The data analyzed and presented in this study are openly available.

Acknowledgment: This research was funded by the National Natural Science
Foundation of China (No. 51609084). The authors sincerely thank the anonymous
reviewers for their insightful comments.

44

#### 857 **References**

858 [1] Amit Y, Geman D. 1997. Shape quantization and recognition with randomized
859 trees. Neural Comput.; 9: 1545-1588. https://doi.org/10.1162/neco.1997.9.7.1545

- [2] Awais M, Aslam B, Maqsoom A, Khalil U, Ullah F, Azam S, et al. 2021.
  Assessing nitrate contamination risks in groundwater: A machine learning approach. Appl. Sci.; 11: 10034. https://doi.org/10.3390/app112110034
- 863 [3] Bagherzadeh F, Mehrani M-J, Basirifard M, Roostaei J. 2021. Comparative study
  864 on total nitrogen prediction in wastewater treatment plant and effect of various
  865 feature selection methods on machine learning algorithms performance. J. Water
  866 Process. Eng.; 41: 102033. https://doi.org/10.1016/j.jwpe.2021.102033
- 867 [4] Band SS, Janizadeh S, Pal SC, Chowdhuri I, Siabi Z, Norouzi A, et al. 2020.
  868 Comparative analysis of artificial intelligence models for accurate estimation of
  869 groundwater nitrate concentration. Sensors; 20.
  870 https://doi.org/10.3390/s20205763
- [5] Bergamasco MAM, Braos LB, Guidini Lopes I, Cruz MCP. 2019. Nitrogen mineralization and nitrification in two soils with different pH levels. Commun.
  Soil Sci. Plant Anal.; 50: 2873-2880. https://doi.org/10.1080/00103624.2019.1689250
- 875 [6] Biddau R, Cidu R, Da Pelo S, Carletti A, Ghiglieri G, Pittalis D. 2019. Source
  876 and fate of nitrate in contaminated groundwater systems: Assessing spatial and
  877 temporal variations by hydrogeochemistry and multiple stable isotope tools. Sci.
  878 Total Environ.; 647: 1121-1136. https://doi.org/10.1016/j.scitotenv.2018.08.007
- 879 [7] Bohrerova Z, Stralkova R, Podesvova J, Bohrer G, Pokorny E. 2004. The relationship between redox potential and nitrification under different sequences of
  881 crop rotations. Soil Tillage Res.; 77: 25-33.
  882 https://doi.org/10.1016/j.still.2003.10.006
- 883 [8] Breiman L. 2001. Random Forests. Mach. Learn.; 45: 5–32.
  884 https://doi.org/10.1023/A:1010933404324
- [9] Castrillo M, García ÁL. 2020. Estimation of high frequency nutrient
  concentrations from water quality surrogates using machine learning methods.
  Water Res.; 172: 115490. https://doi.org/10.1016/j.watres.2020.115490
- [10] Chlingaryan A, Sukkarieh S, Whelan B. 2018. Machine learning approaches for
  crop yield prediction and nitrogen status estimation in precision agriculture: A
  review. Comput. Electron. Agric.; 151: 61-69.
  https://doi.org/10.1016/j.compag.2018.05.012
- [11]Coppola Jr EA, Rana AJ, Poulton MM, Szidarovszky F, Uhl VW. 2005. A neural
  network model for predicting aquifer water level elevations. GROUNDWATER;
  43: 231-241. https://doi.org/10.1111/j.1745-6584.2005.0003.x
- [12] Dinçer AR, Kargi F. 1999. Salt inhibition of nitrification and denitrification in
  saline wastewater. Environ. Technol.; 20: 1147-1153.
  https://doi.org/10.1080/09593332008616912

- [13] Du R, Peng Y, Cao S, Li B, Wang S, Niu M. 2016. Mechanisms and microbial structure of partial denitrification with high nitrite accumulation. Appl. Microbiol.
  Biotechnol.; 100: 2011-2021. https://doi.org/10.1007/s00253-015-7052-9
- 901 [14]El Amri A, M'Nassri S, Nasri N, Nsir H, Majdoub R. 2022. Nitrate concentration
  902 analysis and prediction in a shallow aquifer in central-eastern Tunisia using
  903 artificial neural network and time series modelling. Environ. Sci. Pollut. Res. Int.;
  904 29: 43300-43318. https://doi.org/10.1007/s11356-021-18174-y
- 905 [15]Famiglietti JS, Ferguson G. 2021. The hidden crisis beneath our feet. Science;
  906 372: 344-345. https://doi.org/10.1126/science.abh2867
- 907 [16]Foulquier A, Malard F, Mermillod-Blondin F, Montuelle B, Dolédec S, Volat B,
  908 et al. 2011. Surface water linkages regulate trophic interactions in a groundwater
  909 food web. Ecosystems; 14: 1339-1353. http://doi.org/10.1007/s10021-011-9484-0
- [17]Gan L, Huang G, Pei L, Gan Y, Liu C, Yang M, et al. 2022. Distributions, origins,
  and health-risk assessment of nitrate in groundwater in typical alluvial-pluvial
  fans, North China Plain. Environ. Sci. Pollut. Res. Int.; 29: 17031-17048.
  https://doi.org/10.1007/s11356-021-17067-4
- [18]Gao Z, Han C, Yuan S, Liu J, Peng Y, Li C. 2022. Assessment of the
  hydrochemistry, water quality, and human health risk of groundwater in the
  northwest of Nansi Lake Catchment, north China. Environ. Geochem. Health; 44:
  961-977. https://doi.org/10.1007/s10653-021-01011-z
- [19]Guo H, Ye C, Zhang H, Pan S, Ji Y, Li Z, et al. 2017. Long-term nitrogen &
  phosphorus additions reduce soil microbial respiration but increase its
  temperature sensitivity in a Tibetan alpine meadow. Soil Biol. Biochem.; 113:
  26-34. https://doi.org/10.1016/j.soilbio.2017.05.024
- [20] Hayatsu M, Kosuge N. 1993. Autotrophic nitrification in acid tea soils. Soil Sci.
  Plant Nutr.; 39: 209-217. https://doi.org/10.1080/00380768.1993.10416992
- 924 [21]He S, Wu J, Wang D, He X. 2022. Predictive modeling of groundwater nitrate
  925 pollution and evaluating its main impact factors using random forest.
  926 Chemosphere; 290: 133388. https://doi.org/10.1016/j.chemosphere.2021.133388
- 927 [22]Hinkle SR, Tesoriero AJ. 2014. Nitrogen speciation and trends, and prediction of
  928 denitrification extent, in shallow US groundwater. J. Hydrol.; 509: 343-353.
  929 https://doi.org/10.1016/j.jhydrol.2013.11.048
- 930 [23] Hood-Nowotny R, Umana NH-N, Inselbacher E, Oswald- Lachouani P, Wanek W. 931 2010. Alternative methods for measuring inorganic, organic, and total dissolved 932 Soc. J.: 74: 1018-1027. nitrogen in soil. Soil Sci. Am. https://doi.org/10.2136/sssaj2009.0389 933
- [24]Hu W, Zhou Y, Min X, Liu J, Li X, Luo L, et al. 2018. The study of a pilot-scale aerobic/Fenton/anoxic/aerobic process system for the treatment of landfill
  leachate. Environ. Technol.; 39: 1926-1936.
  https://doi.org/10.1080/09593330.2017.1344325
- [25] Jahangir MMR, Fenton O, Müller C, Harrington R, Johnston P, Richards KG.
  2017. In situ denitrification and DNRA rates in groundwater beneath an
  integrated constructed wetland. Water Res.; 111: 254-264.

- 941 https://doi.org/10.1016/j.watres.2017.01.015
- 942 [26] Jendia AH, Hamzah S, Abuhabib AA, El-Ashgar NM. 2020. Removal of nitrate
  943 from groundwater by eggshell biowaste. Water Supply; 20: 2514-2529.
  944 http://doi.org/10.2166/ws.2020.151
- [27] Ji W, Xiao J, Toor GS, Li Z. 2021. Nitrate-nitrogen transport in streamwater and
  groundwater in a loess covered region: Sources, drivers, and spatiotemporal
  variation. Sci. Total Environ.; 761: 143278.
  https://doi.org/10.1016/j.scitotenv.2020.143278
- [28]Knoll L, Breuer L, Bach M. 2019. Large scale prediction of groundwater nitrate
  concentrations from spatial data using machine learning. Sci. Total Environ.; 668:
  1317-1327.https://doi.org/10.1016/j.scitotenv.2019.03.045
- [29] Lasserre F, Razack M, Banton O. 1999. A GIS-linked model for the assessment of
  nitrate contamination in groundwater. J. Hydrol.; 224: 81-90.
  https://doi.org/10.1016/S0022-1694(99)00130-4
- [30] Li Z, Zhang Q, Qiao Y, Leng P, Zhang Q, Du K, et al. 2021. Influence of the
  shallow groundwater table on the groundwater N<sub>2</sub>O and direct N<sub>2</sub>O emissions in
  summer maize field in the North China Plain. Sci. Total Environ.; 799: 149495.
  https://doi.org/10.1016/j.scitotenv.2021.149495
- [31]Liu J, Liu Q, Yang H. 2016. Assessing water scarcity by simultaneously
  considering environmental flow requirements, water quantity, and water quality.
  Ecol. Indic.; 60: 434-441. https://doi.org/10.1016/j.ecolind.2015.07.019
- [32] Liu Y, Xin J, Wang Y, Yang Z, Liu S, Zheng X. 2022. Dual roles of dissolved organic nitrogen in groundwater nitrogen cycling: Nitrate precursor and denitrification promoter. Sci. Total Environ.; 811: 151375.
  https://doi.org/10.1016/j.scitotenv.2021.151375
- [33]Nikolenko O, Jurado A, Borges AV, Knöller K, Brouyère S. 2018. Isotopic composition of nitrogen species in groundwater under agricultural areas: A review.
  Sci. Total Environ.; 621: 1415-1432. https://doi.org/10.1016/j.scitotenv.2017.10.086
- 970 [34]Osaka Ki, Ohte N, Koba K, Yoshimizu C, Katsuyama M, Tani M, et al. 2010.
  971 Hydrological influences on spatiotemporal variations of δ<sup>15</sup>N and δ<sup>18</sup>O of nitrate
  972 in a forested headwater catchment in central Japan: Denitrification plays a critical
  973 role in groundwater. Journal of Geophysical Research: Biogeosciences; 115.
  974 https://doi.org/10.1029/2009JG000977
- 975 [35]Ouedraogo I, Defourny P, Vanclooster M. 2019. Validating a continental-scale
  976 groundwater diffuse pollution model using regional datasets. Environ. Sci. Pollut.
  977 Res. Int.; 26: 2105-2119. https://doi.org/10.1007/s11356-017-0899-9
- 978 [36]Pang Y, Wang J. 2021. Various electron donors for biological nitrate removal: A
  979 review. Sci. Total Environ.; 794: 148699.
  980 https://doi.org/10.1016/j.scitotenv.2021.148699
- [37]Peng B, Liang H, Wang S, Gao D. 2020. Effects of DO on N<sub>2</sub>O emission during
  biological nitrogen removal using aerobic granular sludge via shortcut
  simultaneous nitrification and denitrification. Environ. Technol.; 41: 251-259.

984 https://doi.org/10.1080/09593330.2018.1494757

- [38] Pennino MJ, Leibowitz SG, Compton JE, Hill RA, Sabo RD. 2020. Patterns and predictions of drinking water nitrate violations across the conterminous United
  States. Sci. Total Environ.; 722: 137661.
  https://doi.org/10.1016/j.scitotenv.2020.137661
- [39] Razzaghi F, Plauborg F, Jacobsen S-E, Jensen CR, Andersen MN. 2012. Effect of nitrogen and water availability of three soil types on yield, radiation use
  efficiency and evapotranspiration in field-grown quinoa. Agric. Water Manag.;
  109: 20-29. https://doi.org/10.1016/j.agwat.2012.02.002
- [40]Rivas A, Singh R, Horne D, Roygard J, Matthews A, Hedley MJ. 2017.
  Denitrification potential in the subsurface environment in the Manawatu River
  catchment, New Zealand: Indications from oxidation-reduction conditions,
  hydrogeological factors, and implications for nutrient management. J. Environ.
  Manag.; 197: 476-489. https://doi.org/10.1016/j.jenvman.2017.04.015
- 998 [41]Rivett MO, Buss SR, Morgan P, Smith JWN, Bemment CD. 2008. Nitrate
  999 attenuation in groundwater: A review of biogeochemical controlling processes.
  1000 Water Res.; 42: 4215-4232. https://doi.org/10.1016/j.watres.2008.07.020
- 1001 [42]Sexstone AJ, Parkin TB, Tiedje JM. 1985. Temporal response of soil
  1002 denitrification rates to rainfall and irrigation. Soil Sci. Soc. Am. J.; 49: 99-103.
  1003 https://doi.org/10.2136/sssaj1985.03615995004900010020x
- [43]Shen Z, Xin J, Wu H, Jiang Z, Peng H, Xu F, et al. 2023. Kinetic and molecular
  evidence for DON transformation in the deep vadose zone: Important
  implications for soil nitrogen budgeting and groundwater nitrate management. J.
  Hydrol.; 616: 128782. https://doi.org/10.1016/j.jhydrol.2022.128782
- 1008 [44]Singh BK, Bardgett RD, Smith P, Reay DS. 2010. Microorganisms and climate
  1009 change: terrestrial feedbacks and mitigation options. Nat. Rev. Microbiol.; 8:
  1010 779-790. http://doi.org/10.1038/nrmicro2439
- 1011 [45]Smarra F, Jain A, de Rubeis T, Ambrosini D, D'Innocenzo A, Mangharam R.
  1012 2018. Data-driven model predictive control using random forests for building
  1013 energy optimization and climate control. Appl. Energy; 226: 1252-1272.
  1014 https://doi.org/10.1016/j.apenergy.2018.02.126
- 1015 [46]Smith RL, Böhlke JK, Song B, Tobias CR. 2015. Role of anaerobic ammonium
  1016 oxidation (anammox) in nitrogen removal from a freshwater aquifer. Environ. Sci.
  1017 Technol.; 49: 12169-12177. https://doi.org/10.1021/acs.est.5b02488
- 1018 [47]Spoelstra J, Schiff S, Elgood R, Semkin R, Jeffries D. 2001. Tracing the sources
  1019 of exported nitrate in the Turkey Lakes Watershed using <sup>15</sup>N/<sup>14</sup>N and <sup>18</sup>O/<sup>16</sup>O
  1020 isotopic ratios. Ecosystems; 4: 536-544.
  1021 https://doi.org/10.1007/s10021-001-0027-y
- [48] Strous M, Kuenen JG, Jetten MS. 1999. Key physiology of anaerobic ammonium
  oxidation. Appl. Environ. Microbiol.; 65: 3248-50.
  https://doi.org/10.1128/aem.65.7.3248-3250.1999
- 1025 [49] Thayalakumaran T, Bristow KL, Charlesworth PB, Fass T. 2008. Geochemical
  1026 conditions in groundwater systems: Implications for the attenuation of

- 1027
   agricultural nitrate.
   Agric.
   Water
   Manag.;
   95:
   103-115.

   1028
   https://doi.org/10.1016/j.agwat.2007.09.003

   103-115.
- 1029 [50] van Es HM, Schindelbeck RR, Jokela WE. 2004. Effect of manure application
  1030 timing, crop, and soil type on phosphorus leaching. J. Environ. Qual.; 33:
  1031 1070-1080. https://doi.org/10.2134/jeq2004.1070a
- 1032 [51] Vandenbruwane J, De Neve S, Qualls R G, Sleutel S, Hofman G. 2007.
  1033 Comparison of different isotherm models for dissolved organic carbon (DOC)
  1034 and nitrogen (DON) sorption to mineral soil. Geoderma, 139(1-2), 144-153.
  1035 https://doi.org/10.1016/j.geoderma.2007.01.012
- 1036 [52] Vidon P, Hill AR. 2004. Denitrification and patterns of electron donors and
  1037 acceptors in eight riparian zones with contrasting hydrogeology. Biogeochemistry;
  1038 71: 259-283. http://doi.org/10.1007/s10533-004-9684-1
- [53] Vystavna Y, Diadin D, Valeriy Y, Hejzlar J, Vadillo I, Huneau F, et al. 2017.
  Nitrate contamination in a shallow urban aquifer in East Ukraine: Evidence from hydrochemical, stable nitrate isotope, and land use analysis. Environ. Earth Sci.;
  76. https://doi.org/10.1007/s12665-017-6796-1
- 1043 [54] Wang C, Jiang R, Boithias L, Sauvage S, Sánchez-Pérez JM, Mao X, Han Y,
- Hayakawa A, Kuramochi K, Hatano R. 2016. Assessing potassium environmental
  losses from a dairy farming watershed with the modified SWAT model. Agr.
  Water Manage.; 175, 91-104. https://doi.org/10.1016/j.agwat.2016.02.007
- 1047 [55] Wang C, Wu D, Mao X, Hou J, Wang L, Han Y. 2021. Estimating soil ammonium adsorption using pedotransfer functions in an irrigation district of the North 1049 China Plain. Pedosphere; 31: 157-171. 1050 https://doi.org/10.1016/S1002-0160(20)60054-6
- [56] Wang Y, Peng J, Cao X, Xu Y, Yu H, Duan G, et al. 2020. Isotopic and chemical
  evidence for nitrate sources and transformation processes in a plateau lake basin
  in Southwest China. Sci. Total Environ.; 711: 134856.
  https://doi.org/10.1016/j.scitotenv.2019.134856
- 1055 [57] Weitzman JN, Brooks JR, Compton JE, Faulkner BR, Mayer PM, Peachey RE, et
  1056 al. 2022. Deep soil nitrogen storage slows nitrate leaching through the vadose
  1057 zone. Agr Ecosyst Environ; 332: 107949.
  1058 https://doi.org/10.1016/j.agee.2022.107949
- [58] Wells NS, Baisden WT, Horton T, Clough TJ. 2016. Spatial and temporal variations in nitrogen export from a New Zealand pastoral catchment revealed by
  stream water nitrate isotopic composition. Water Resour. Res.; 52: 2840-2854.
  https://doi.org/10.1002/2015WR017642
- 1063 [59]Xinxiang City Water Resources Bureau (XCWRB), 2017. Xinxiang City Water
   1064 Resources Bulletin. http://slj.xinxiang.gov.cn/ggl/1983.html
- 1065 [60] Xue D, Botte J, De Baets B, Accoe F, Nestler A, Taylor P, et al. 2009. Present
  1066 limitations and future prospects of stable isotope methods for nitrate source
  1067 identification in surface- and groundwater. Water Res.; 43: 1159-1170.
  1068 https://doi.org/10.1016/j.watres.2008.12.048

- 1069 [61] Yang X, Wang S, Zhou L. 2012. Effect of carbon source, C/N ratio, nitrate and dissolved oxygen concentration on nitrite and ammonium production from denitrification process by Pseudomonas stutzeri D6. Bioresour. Technol.; 104: 65-72. https://doi.org/10.1016/j.biortech.2011.10.026
- 1073 [62]Zhang Y, Xu B, Guo Z, Han J, Li H, Jin L, et al. 2019. Human health risk
  1074 assessment of groundwater arsenic contamination in Jinghui irrigation district,
  1075 China. J. Environ. Manag.; 237: 163-169.
  1076 https://doi.org/10.1016/j.jenvman.2019.02.067
- 1077 [63]Zhao B, Zhang J, Yu Y, Karlen DL, Hao X. 2016. Crop residue management and 1078 fertilization effects on soil organic matter and associated biological properties.
  1079 Environ. Sci. Pollut. Res.; 23: 17581-17591.
  1080 http://doi.org/10.1007/s11356-016-6927-3
- 1081 [64]Zhou J, Green M, Shaviv A. 2003. Mineralization of organic N originating in
  1082 treated effluent used for irrigation. Nutr. Cycl. Agroecosyst.; 67: 205-213.
  1083 https://doi.org/10.1023/B:FRES.0000003599.60911.a2

1084