

# Polyploidization as an opportunistic mutation: The role of unreduced gametes formation and genetic drift in polyploid establishment

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2	and genetic drift in polyploid establishment		
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16	and wrote the initial draft of the manuscript. J.C. N. P-G. and F.K. edited the manuscript.		
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27			
28 29	<b>Data availability:</b> The simulation code has been deposited on Github (https://github.com/JosselinCLO/Drift_polyploidy).		
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Abstract: It is broadly assumed that polyploidy success reflects an increase in fitness 30 31 associated with whole-genome duplication (WGD), due to higher tolerance to stressful 32 conditions. Nevertheless, WGD also arises with several costs in neo-polyploid lineages, like genomic instability, or cellular mis-management. In addition to these costs, neo-polyploid 33 individuals also face frequency dependent selection, because of frequent low-fitness triploids 34 formed by cross-ploidy pollinations when tetraploids are primarily rare in the population. 35 Interestingly, the idea that polyploidy can be fixed by genetic drift, as a neutral or deleterious 36 mutation is currently underexplored in the literature. To test how and when polyploidy can fix 37 in a population by chance, we built a theoretical model in which autopolyploidization occurs 38 39 through the production of unreduced gametes, a trait modeled as a quantitative trait that is 40 allowed to vary through time. We found that when tetraploid individuals are less or as fit as their diploid progenitors, fixation of polyploidy is only possible when genetic drift is stronger 41 42 than natural selection. The necessity of drift for tetraploid fixation holds even when polyploidy confers a selective advantage, except for scenarios where tetraploids are much 43 fitter than diploids. Finally, we found that self-fertilization is less beneficial for tetraploid 44 establishment than previously thought, notably when polyploids harbour an initial decrease in 45 fitness. Our results bring a novel, non-exclusive explanation for the unequal temporal and 46 47 spatial distribution of polyploid species.

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50 <u>Keywords:</u> Polyploid establishment, self-fertilization, unreduced gametes, genetic drift
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## 54 INTRODUCTION

55

56	Whole-genome duplication is present throughout the evolution of eukaryotes (Mable,
57	2004; Gregory & Mable, 2005), with a strong representation in angiosperms for which 15% of
58	speciation events are associated with a ploidy increase (Ramsey & Schemske, 2002; Soltis et
59	al., 2007; Wood et al., 2009; Parisod et al., 2010; Barker et al., 2016). Polyploidy has been
60	demonstrated to have a broad range of effects on plant phenotypes and genomes.
61	Polyploidization is hypothesized to be both an evolutionary dead-end (Stebbins, 1971;
62	Mayrose et al., 2011), and an important driver of plant adaptation and speciation (Van de Peer
63	et al., 2017). Notably, the apparent non-random establishment probability of genome doubling
64	events has suggested a link with environmental conditions (Van de Peer et al., 2021).
65	Polyploidization is known to confer a selective advantage under several stressful conditions
66	(Bomblies, 2020; Van de Peer et al., 2021). For example, polyploidy has been associated with
67	higher tolerance to salt (Chao et al., 2013) and drought stress (Ruiz et al., 2016), and higher
68	resistance to pathogens and herbivory (Hannweg et al., 2016; Hias et al., 2018; Wang et al.,
69	2018). All these advantages tend to explain why successful polyploidization events are
70	associated with periods of dramatic global change (Vanneste et al., 2014a), or harsher
71	environments (Rice et al., 2019).

As shown before, polyploidization can be favored during unstable and global change periods. Nevertheless, it has been suggested that genome doubling should be counter-selected in stable periods and environments (Comai, 2005; Oberlander *et al.*, 2016; Kreiner *et al.*, 2017a; Van de Peer *et al.*, 2017). First, because polyploidization arises with several costs in neo-polyploid lineages, like genomic instability, mitotic and meiotic abnormalities, and a reduction in fitness (Comai, 2005; Otto, 2007; Doyle & Coate, 2019; Porturas *et al.*, 2019;

Clo & Kolář, 2021). In addition to these intrinsic costs of polyploidization, neo-polyploid 78 79 individuals are also facing frequency dependent selection. Newly formed polyploid individuals are primarily in minority within populations, and most of their reproductive events 80 with diploid individuals lead to low-fitness triploids, enhancing the extinction probability of 81 82 polyploids (a phenomenon called Minority Cytotype Exclusion, Levin, 1975). Second, the production of polyploid individuals is often a costly mechanism for the diploid ancestral 83 populations (Kreiner et al., 2017a). It is recognized that the production of unreduced gametes 84 is the primary mechanism of polyploid formation (Harlan & deWet, 1975; Ramsey & 85 Schemske, 1998; Tayalé & Parisod, 2013; Mason & Pires, 2015). The production of 86 87 unreduced gametes is expected to be counter-selected in stable environments, because for 88 each unreduced gamete formed, two normal gametes are lost (Kreiner et al., 2017a), which can potentially reduce the contribution of an individual to fertilization events. In addition, if 89 90 the production of unreduced gametes is low in a population, most fertilization events involving unreduced gametes will lead to the production of triploid individuals, which 91 92 generally harbor low fitness (Ramsey & Schemske, 1998; Burton & Husband, 2000; Husband & Sabara, 2003). The expectation of low unreduced gamete production in natural populations 93 94 under stable environmental conditions is generally supported by empirical studies (Kreiner et 95 al., 2017a). Other factors, like (partial) asexuality, can however reduce the strength of 96 selection against unreduced gametes (see for example Bohutínská et al., 2021). The maintenance of unreduced gamete production in stable environments is likely to be a balance 97 98 between the rate of mutations favoring their synthesis, and selective forces acting against them. 99

100 It appears that the production of unreduced gametes and polyploid individuals and 101 their maintenance in populations is more likely, or even only possible, if polyploidy comes 102 with a fitness advantage. The different theoretical models exploring under which conditions

polyploidy can spread and fix within a diploid population generally support this expectation. 103 104 The establishment of polyploid individuals within the diploid progenitor population is more likely when polyploid individuals benefit from a higher fitness compared to their diploid 105 progenitors, both in stable conditions (Felber, 1991), or during environmental changes 106 107 (Oswald & Nuismer, 2011; Yao et al., 2019; Van Drunen & Friedman, 2021), and when other factors limit gene flow among cytotypes, like assortative mating, self-fertilization or clonality 108 109 (Oswald & Nuismer, 2011; Griswold, 2021; Van Drunen & Friedman, 2021) or restricted pollen and seed dispersals (Li et al., 2004; Baack, 2005). 110

111 Theoretical models in which polyploidy can fix neutrally or with an initial decrease in 112 fitness received less attention (Felber, 1991; Husband, 2004; Li *et al.*, 2004; Baack, 2005; 113 Meyers & Levin, 2006; Oswald & Nuismer, 2011). In these conditions, polyploidy can spread 114 or fix only when the population size is small and stochasticity is high (Li *et al.*, 2004), when 115 there is pre-zygotic isolation between cytotypes (Rausch & Morgan, 2005; Oswald & 116 Nuismer, 2011) or when there is a niche differentiation among cytotypes (Rodriguez, 1996).

Surprisingly, the above-mentioned models make the strong assumption that the 117 production of unreduced gametes in the population is a high (much higher than what is found 118 in natural populations, Kreiner et al., 2017a; b) and fixed quantity, that does not vary through 119 120 time (Felber, 1991; Husband, 2004; Oswald & Nuismer, 2011; Van Drunen & Friedman, 2021). It is known that the production of unreduced gametes is determined genetically (Parrott 121 & Smith, 1986; Tavoletti et al., 1991), varies among individuals within and between 122 population scales (Kreiner et al., 2017a), and is likely to be polygenic (Brownfield & Köhler, 123 2011; De Storme & Geelen, 2013). It has been pointed out that we are currently lacking clear 124 125 theoretical and empirical frameworks for the effect of the genetic architecture of unreduced gamete production on the spread and fixation probability of polyploidy (Kreiner et al., 126 2017a). 127

In this paper, we investigated theoretically the effect of the genetic architecture of 128 129 unreduced gametes production in the spread and fixation of polyploidy, when tetraploid individuals are more or less fit than their diploid progenitors. We first built a simple analytical 130 model which allowed us to predict in which conditions the production of unreduced gametes 131 132 and the formation of tetraploid individuals can evolve by genetic drift. We then built an individual-based simulation model in which the production of unreduced gametes is a 133 quantitative trait that evolves through time by mutation, selection and genetic drift, to test our 134 predictions, and to study the effect of the genetic architecture of unreduced gametes 135 production in the fixation of polyploidy when tetraploids are more or less fit than their diploid 136 137 progenitors. Overall, we found that when polyploid individuals are less or as fit as their 138 diploid progenitors, the fixation of polyploidy is only possible when genetic drift is stronger than natural selection. Surprisingly, this result remains true even when polyploidy confers a 139 140 selective advantage, with genetic drift being necessary for tetraploidy to be established, except when polyploids are considerably more fit than their diploid progenitors. Finally, we 141 found that self-fertilization is less beneficial than previously thought, as it increases the 142 selection against unreduced gamete production compared to outcrossing populations when 143 polyploids harbour an initial decrease in fitness. 144

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## 146 MATERIAL & METHODS

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148 Verbal prediction
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We consider a diploid population in which the proportion of unreduced gametes canevolve through time. The genetic architecture of the trait "proportion of unreduced gametes"

is considered to be the same in male and in female functions. It is hard to judge if the abovementioned assumption is realistic or not. Male and female functions can slightly vary in their
average production of unreduced gametes, but the among-individual variation is high and
overlapping in both sexes (Kreiner *et al.*, 2017a).

Consider that the number of unreduced gametes is coded by L bi-allelic loci. The allele 156 A being the ancestral allele and coding for no unreduced gametes, and the allele M at the *i*th 157 locus increasing the production of unreduced gametes by a value  $h.x_i$  and  $x_i$  respectively in its 158 heterozygous and homozygous states. In this model, h is the coefficient of dominance of the 159 M allele, and is fixed for all loci. The multi-locus probability of producing an unreduced 160 gamete,  $p_{UG}$  is inferred as the sum of genotypic values over the L loci, and depends on the 161 162 number of mutations carried by an individual (see eq. 1 for details). We assume that the 163 environment effects on unreduced gamete production are null, on average (Falconer & Mackay, 1996). First, we will infer  $s_{0,i}$ , the coefficient of selection against the allele M at the 164 165 *i*th locus at t = 0, when the population is uniform and produces no unreduced gametes. We assume that an individual produces  $\gamma_{tot}$  gametes in total. The wildtype homozygote genotype 166  $G_{AA,i}$  (homozygote for the allele A at the *i*th locus) only produces haploid gametes, such that 167 168 its absolute fitness is equal to  $\gamma_{tot}$ . To infer  $s_{0,i}$ , we will consider  $G_{AA,i}$  as our reference, and fix 169 its relative fitness to  $\omega_{AA} = 1$ , while the relative fitness of the homozygote genotype  $G_{MM,i}$ (homozygote for the allele *M* at the *i*th locus) is equal to  $\omega_{MM} = 1 - s_{0,i}$ . The number of 170 171 gametes of  $G_{MM,i}$  can be divided as the proportion of haploid gametes  $\gamma_n$  it produces at frequency  $p_n = 1 - x_i$ , with  $\gamma_n = p_n \cdot \gamma_{tot}$ , and the proportion of unreduced gametes  $\gamma_{2n}$  produced at 172 173 frequency  $p_{2n} = x_i$ , such that  $\gamma_{2n} = p_{2n}.\gamma_{tot}$ .

Assuming that, initially, the probability of forming a tetraploid offspring is so low due to minority cytotype exclusion, unreduced gametes will not contribute to fitness, so that the relative fitness of  $G_{MM,i}$  is easily seen to be  $1 - x_i$ . It appears that  $s_{0,i} = x_i$ , and that the smaller the effect of the modifier of unreduced gametes production, the less it will be counterselected. The average coefficient of selection against unreduced gamete production  $(\bar{s_0})$  is the mean of the selection coefficients among loci  $(\bar{s_0} = \frac{1}{L}\sum_{i}^{L} x_i = \bar{x_i})$ . Now we have a way to quantify the strength of selection, we can deduce in which conditions the evolution of unreduced gametes production will be influenced by genetic drift. If we consider that the dynamic of the modifier is mainly driven by genetic drift when  $N_{\rm E}$ .  $\bar{s_0} < 1$ , then  $N_{\rm E}$  has to be strictly smaller than  $1/\bar{s_0}$ , where  $N_{\rm E}$  is the effective population size.

When selection is strong compared to drift  $(N_E > 1/\overline{s_0})$ , the frequency of the allele M is 184 expected to remain low at all loci, such that the probability of generating tetraploid 185 individuals remains low, and the population is expected to stay diploid. When genetic drift is 186 187 stronger than selection ( $N_{\rm E} < 1/\overline{s_0}$ ), the allele M can reach higher frequencies, or can even be fixed by chance, such that the probability of generating tetraploid individuals becomes high, 188 and tetraploid individuals have a chance to replace their diploid progenitors. In addition, it has 189 to be noted that the coefficient of selection against unreduced gametes is not fixed, and will 190 evolve through time. Notably, the selection pressure will decrease when the frequency of 191 unreduced gametes (from diploids and tetraploids) will increase, and it can eventually become 192 negative (and then the production of unreduced gametes will be positively selected) when 193 unreduced gametes become dominant in a population. Then,  $\overline{s_0}$ , and the associated  $N_{\rm E}$  for 194 which the trait is expected to mainly evolve by drift, are the critical values that initiate the 195 system. Any factor decreasing the effective population size, like self-fertilization, is expected 196 197 to facilitate the increase in the frequency of the deleterious alleles M by genetic drift. Selffertilization (or any other form of assortative mating) also promotes polyploidization by 198 helping polyploids escape the minority cytotype exclusion. 199

200

In order to test our analytical prediction about the effect of drift in the formation of 203 tetraploid populations, we wrote an individual-based simulation model. We simulated 204 populations with a constant population size of N individuals (N contains both D diploid and T 205 206 tetraploid individuals), and a constant selfing rate  $\sigma$ . The trait "unreduced gametes production" is coded by L freely recombining bi-allelic loci, with the allele A coding for no 207 unreduced gametes, and the allele *M* increasing the production of unreduced gametes at the *i*th 208 209 locus by a fraction  $x_i = 1/L$  in its homozygous state, and by  $hx_i = h/L$  in its heterozygous state, h being the coefficient of dominance of the allele M, and being fixed among loci. With this 210 simplification, if an individual carries respectively n and m mutations respectively at 211 212 heterozygous and homozygous states, the probability of producing an unreduced gamete,  $p_{UG}$ , become 213

214 
$$p_{UG} = \frac{nh+m}{L}$$
 , (eq. 1)

and varied between 0 (the individual is homozygote for the allele A at all loci) and 1 (the 215 individual is homozygote for the allele *M* at all loci). As  $x_i$  is set to 1/L at all loci,  $\overline{s_0} = 1/L$ , 216 and genetic drift is stronger than selection when  $N_{\rm E} < L$ . For each individual and at each 217 218 generation, an environmental effect is drawn in a Gaussian distribution of mean 0 and 219 variance  $V_{\rm E}$  of 0.005, in order to introduce variation in unreduced gametes production. The fitness of diploid individuals is fixed to  $W_{2x} = 1$ , while the fitness of neo-tetraploid individuals 220 is fixed within a simulation, but can vary from  $W_{4x} = 0.75$  to 1.25 among simulations, to fit 221 222 with empirical data demonstrating initial fitness decrease in neo-polyploids due to their problems in cellular management and meiosis (Porturas et al., 2019; Clo & Kolář, 2021), or 223 higher fitness in stressful conditions (Chao et al., 2013; Ruiz et al., 2016). We assume random 224

mating opportunities among individuals of any ploidy. Only diploid and tetraploid cytotypes 225 226 are viable, triploids and higher polyploids are assumed to be not viable (Ramsey & Schemske, 1998; Burton & Husband, 2000; Husband & Sabara, 2003). Note that this hypothesis is 227 conservative, as viable triploid individuals are expected to favour the establishment of 228 229 tetraploid populations (Husband, 2004). However, a (virtual) lack of triploids in many mixedploidy systems (Kolář et al., 2017) demonstrates that such a conservative model is realistic for 230 many species. The population evolves during 20K generations, and the frequencies of the 231 allele *M*, of unreduced gametes and of tetraploid individuals are followed. 232

We initiated each simulation from a population of genetically identical diploid 233 234 individuals that are homozygous for the allele A at all loci ( $p_{UG} = 0$  at t = 0). The life cycle can be summarized in five successive events. First, there is a fitness-dependent choice of the 235 first parent (selection), followed by mating-type choice (selfing versus outcrossing at rates  $\sigma$ 236 237 and  $[1-\sigma]$ , respectively). In the case of outcrossing, the choice of the second parent is also fitness-dependent. Selection takes place as follows: an individual is sampled randomly among 238 239 the N individuals, but its sampling probability is weighted by its fitness. The probabilities of 240 sampling a diploid  $(p_{2X})$  or a tetraploid  $(p_{4X})$  individual are respectively

241 
$$p_{2X} = \frac{D.W_{2x}}{D.W_{2x} + T.W_{4x}}, \text{ (eq 2.1)}$$

242 and

243 
$$p_{4X} = \frac{T.W_{4x}}{D.W_{2x} + T.W_{4x}} . \text{ (eq 2.2)}$$

With these equations, it appears that tetraploids are as likely to be sampled as diploids when they represent respectively 57%, 50% or 44% of the population when  $W_{4x}$  is respectively equal to 0.75, 1 or 1.25. For mixed mating regime, a number is sampled in a uniform law comprised between 0 and 1, if the number is smaller (respectively higher) than 0.5, the

selected individual reproduced by outcrossing (respectively by selfing). Once the two parents 248 249 are chosen, the type of gametes (reduced or unreduced) they will produce is chosen. For each reproducer, one number is sampled in a uniform distribution between 0 and 1. If  $p_{UG}$  is higher 250 than the sampled value, the reproducer generates an unreduced gamete (note that a tetraploid 251 252 individual can also produce unreduced gametes, see below for details). For reduced (n)gametes (n = x and n = 2x gametes for diploid and tetraploid individuals, respectively), one 253 254 allele per locus of each sister chromatid is sampled randomly. This phase is then followed by the introduction of mutations from allele A to allele M and vice versa, the number of which is 255 sampled from a Poisson distribution with parameter U (with  $U = \mu L$ , where  $\mu$  is the per-locus 256 257 mutation rate and L is the number of loci underlying the quantitative trait). If the resulting 258 offspring is not diploid or tetraploid, it will not contribute to the next generation. The reproduction phase stops when N viable offspring are formed. 259

260

## 261 Parameter sets tested

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The number of loci underlying the production of unreduced gametes can vary from L =263 264 2 to 200. The population size can be equal to N = 50, 100 or 200. The associated effective population size is equal to  $N_{\rm E} = N(2-\sigma)/2$  (Glémin & Ronfort, 2013). Bigger population sizes 265 (e.g. N > 2000) do not change the results qualitatively, but the simulation time becomes 266 limiting, and the number of loci needed for the trait to evolve by genetic drift becomes 267 unrealistically high. We tested three mutation rates, with the diploid genomic mutation rate 268 269 (U) being equal to 0.5, 0.05 or 0.005, which is the range of what is found in natural populations (Halligan & Keightley, 2009). For simplicity, the coefficient of dominance of the 270 alleles *M* can be equal to 0 (fully recessive: none of the heterozygote genotypes in diploids or 271

272	tetraploids produce unreduced gametes), 0.5 (co-dominant, the diploid and tetraploid
273	heterozygotes genotypes at the <i>i</i> th locus are: $G_{AM,i}$ , $G_{AAAM,i}$ , $G_{AAMM,i}$ and $G_{AMMM,i}$ produce
274	unreduced gametes at frequencies $0.5/L$ , $0.25/L$ , $0.5/L$ and $0.75/L$ , respectively) or 1 (fully
275	dominant: any heterozygote genotypes in diploids or tetraploids produce unreduced gametes
276	at a frequency $1/L$ at a given locus). Finally, we tested three selfing rates with $\sigma$ being equal to
277	0, 0.5 or 1. The selfing rate is the same in diploids and tetraploids. As mentioned before,
278	tetraploid individuals can be more or less fit compared to their diploid progenitors, reflecting
279	the fitness (dis)advantage associated with tetraploidy in standard vs disturbed environments.
280	For each parameter, we performed $n = 100$ replicates. A summary of the parameters tested
281	and their values can be found in table 1.
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_	Parameter	Abbreviation	Value(s)	
296	simulations.			
295	Table 1. Description of the model	odel parameters, their abbreviation	ons, and tested value	s in

	ADDIEviation	v aluc(s)
Population size	Ν	50, 100, 200
Number of loci	L	From 2 to 200
Diploid genomic mutation rate	U	0.005, 0.05 or 0.5
Fitness of diploid individuals	$W_{2x}$	1
Fitness of tetraploid individuals	$W_{4\mathrm{x}}$	0.75, 1, 1.25 or 2
Selfing rate	σ	0, 0.5 or 1
Probability of sampling an	<i>p</i> UG, <i>p</i> <sub>2x</sub> , <i>p</i> <sub>4x</sub>	From 0 to 1
tetraploid individual		
Coefficient of selection at $t = 0$	$\overline{S_0}$	From 0.5 to 0.005
Dominance's coefficient of allele M	h	0, 0.5, 1
$V_{\rm E}$ of environmental effects	$V_{ m E}$	0.005

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## 306 The effects of fitness and genetics on the probability of fixation of polyploidy

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As predicted analytically, when considering tetraploids being less fit than diploids the 308 309 fixation of polyploidy is only possible when genetic drift is stronger than natural selection 310 against the unreduced gamete formation (Figure 1, left panel). This finding remains true, even when tetraploid individuals are as fit as their diploid progenitors (Figure 1, middle panel). If 311 312 tetraploids are slightly fitter than their diploid progenitors, fixation of the tetraploidy occurs either when drift is stronger or as strong as selection, but never when selection is stronger than 313 314 drift (Figure 1, right panel). Furthermore, when tetraploid individuals are as fit as or slightly fitter than diploids, the fixation of polyploids is faster compared to the case in which 315 316 tetraploids are less fit than diploids (Figure 1 middle and right panels compared to the left 317 panel). Polyploidy can fix without drift only when it comes with an important fitness 318 advantage ( $W_{\text{tetra}} = 2W_{\text{diplo}}$ , Figure 2). More dominant mutations and higher mutation rates increased the production of unreduced gametes and then the probability of fixing 319 320 polyploidization (Figure S1 & S2).

321

### 322 The production of unreduced gametes within populations

323

The production of unreduced gametes in diploid individuals strongly depends on two parameters, the mutation rate for unreduced gamete production, and the strength of drift compare to selection (Figure 3). As expected, higher mutation rates lead to higher production

327	of unreduced gametes within populations (Figure 3, $U = 0.005 vs$ . $U = 0.05$ ). For example,
328	when selection is stronger than drift, the production of unreduced gametes in diploid
329	individuals is on average 2.8% when $U = 0.005$ , and is on average 17.1% when $U = 0.05$ . As
330	predicted, the higher the level of genetic drift, the higher the number of unreduced gametes
331	produced by individuals (Figure 3). For example, for $U = 0.05$ , the production of unreduced
332	gametes in diploids is on average 17.1% when selection is stronger than drift, and around
333	30.4% when drift is stronger than selection. The same pattern is observable for smaller
334	mutation rates (Figure 3). The fitness of tetraploid individuals slightly affects the production
335	of unreduced gametes in diploids, the fitter the tetraploids, the higher the production of
336	unreduced gametes in diploids (Figure S3). The production of unreduced gametes remains
337	comparable in diploids and tetraploid individuals, on average (Figure S4).
338	
220	Effect of the mating system
339	Effect of the mating system
340	
341	Self-fertilization appears to increase the strength of selection against unreduced
342	gamete formation when polyploidy is associated with a fitness cost, despite its positive effect
343	on the strength of genetic drift. When genetic drift is stronger than selection and polyploidy
344	has a fitness cost, only obligate outcrossing populations are able to fix polyploidy (Figure 4
345	left panel). When polyploidy comes with a fitness advantage over diploidy, we confirmed the
346	
	classical expectation that self-fertilization increases the probability of fixing polyploidy, due

348 middle panel) or weaker (Figure 4 right panel) than natural selection.

# **DISCUSSION**

### 352 Genetic drift is a major component of the probability of fixation of polyploidy

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One major result of our theoretical model is that genetic drift is important for the evolution of polyploidy, even when autotetraploids are fitter than their diploid progenitors. This result indicates that the initial frequency-dependent selection is the strongest barrier to polyploids establishment.

Our predictions offer a new and alternative view on the establishment of polyploidy in 358 populations. Even if polyploid lineages are more likely to survive given their potential fitness 359 advantage over their diploid progenitors (Van de Peer et al., 2021), this does not necessarily 360 361 mean such a slight adaptive advantage has been the cause for their establishment. Harsher environment conditions and/or periods of drastic environmental changes, extreme latitudes, 362 and range margins are also expected to decrease the demographic and effective population 363 364 sizes of species (Brown, 1984), offering the possibility for polyploidization to fix by genetic 365 drift. It can explain why polyploid species are contemporarily more frequently found at range margins (Levin, 1975; Fowler & Levin, 1984; Felber, 1991; Rojas-Andrés et al., 2020), 366 367 extreme environments (Brochmann et al., 2004), extreme latitudes (Rice et al., 2019) and why they are frequently invasive (Te Beest et al., 2012). It can also help to understand why 368 polyploidy can sometimes be associated with higher extinction rates compared to diploidy 369 (Mayrose et al., 2011, 2015; Arrigo & Barker, 2012), if polyploidy has been initially fixed 370 despite an initial decrease in fitness. It is also important to note that drift and natural selection 371 372 are not exclusive explanations, and both forces may increase the probability of establishment of polyploid lineages under certain scenarios. Genetic drift can help in increasing the 373 frequency of genes favouring unreduced gametes production and escaping minority cytotype 374

exclusion, while selection will then maintain them at high frequency if polyploid individualsare more fit than their diploid progenitors.

377 Direct evidence of the role of drift in the evolution of polyploidy is however hard to 378 find and could be hard to produce in the future. Nevertheless, and as a support of our model, the few experiments testing for local adaptation of polyploid lineages led to mitigated results, 379 with either no (Baack & Stanton, 2005; Buggs & Pannell, 2007; Martin & Husband, 2013; 380 381 McIntyre & Strauss, 2017) or a weak (Raabová et al., 2008; Duchoslav et al., 2017) fitness advantages of polyploids in their local sites. Note that testing for local adaptation patterns can 382 be a weak way of testing for the effect of drift or selection in polyploidy establishment, 383 because a fitness advantage is not always sufficient for polyploidy to fix, and because 384 polyploidization can fix with an initial decrease in fitness with the help of genetic drift, and 385 adaptation can occur later after establishment. 386

387

### 388 Comparing the number of unreduced gametes with empirical results

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In our simulations, the average number of unreduced gametes within diploid 390 populations that tetraploids are unable to establish (*i.e.* population in which selection is 391 stronger than drift) strongly depends on the mutation rate for unreduced gametes production. 392 393 Empirical estimates made in diploid populations range from 0.2% to 11.6% (Bretagnolle, 394 2001; Ramsey, 2007; Kron & Husband, 2009; Sheidai et al., 2009; Kovalsky & Solís Neffa, 2012, 2016; Herben et al., 2016; Kreiner et al., 2017b), with an average number of 2.6% 395 396 (Kreiner et al., 2017a). In our simulations, we are obtaining similar values only when the mutation rate for unreduced gametes is low (Figure 3 & S3) and when selection is stronger 397

than drift, suggesting that it is also probably the case in the few natural populations that havebeen studied.

400 However, the number of unreduced gametes needed for polyploidy to fix in an initially 401 diploid population is much higher in our simulations (at least 15%) than what is found in natural diploid populations (Kreiner et al., 2017a). Our results suggest that, even if polyploid 402 individuals are fitter than their diploid progenitors, the number of unreduced gametes found in 403 404 natural diploid populations would theoretically not allow polyploid individuals to invade. This result challenges the idea that the production of unreduced gamete in diploid populations may 405 be selected to enhance polyploid production in stressful conditions or in habitats where they 406 407 are most expected to thrive (Stebbins, 1950; Vanneste et al., 2014b; Mason & Pires, 2015). The production of unreduced gametes remains similar in diploid and tetraploid individuals 408 after the fixation of polyploidy. This is mainly because of the assumptions we made in our 409 model, notably that after fixation of tetraploidy, there are no other cytotypes possible, making 410 intermediate rates of unreduced gamete production not costly in tetraploid populations, in our 411 412 simulation model.

413

## 414 Pre-mating isolation among cytotypes and polyploidy establishment

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We found that pre-mating isolation among cytotypes promotes polyploidization only when tetraploid individuals are as fit or fitter than their diploid progenitors. This stems from the fact that selfing lineages that tend to produce more unreduced gametes will also produce more tetraploid offspring with a lower fitness, compared to other selfing lineages that produce more frequently diploid offspring with a higher fitness. As neo-polyploid lineages are generally less fit than their diploid progenitors (Porturas *et al.*, 2019; Clo & Kolář, 2021), it

could be expected that the evolution of self-fertilization particularly, and other forms of 422 423 assortative mating in general, should be limited in empirical data. As a confirmation, Husband et al. (2008) found that autopolyploid lineages generally have smaller selfing rates than 424 diploid species. Other forms of assortative mating, such as delayed phenology among 425 426 cytotypes, are also rarely observed at a large phylogenetic scale (Porturas et al., 2019; Clo & Kolář, 2021). Our results also fit well with empirical data that did not find an effect of the 427 mating system on the frequency of unreduced gametes in natural populations (Kreiner et al., 428 2017b). 429

430

### 431 Considerations related to the genetic architecture of unreduced gametes production

432

It is first important to notice that little is known about the genetic architecture of the 433 production of unreduced gametes in natural populations (Brownfield & Köhler, 2011; De 434 435 Storme & Geelen, 2013). As a consequence, we cannot precisely quantify for which effective 436 population sizes the production of unreduced gametes can occur by genetic drift when polyploidization is detrimental to individuals' fitness. It is known, however, that there are 437 438 several pre-meiotic (Fernández & Neffa, 2004; Kim et al., 2009; Mason et al., 2011), meiotic (Bretagnolle & Thompson, 1995; Pécrix et al., 2011; De Storme et al., 2012) and post-meiotic 439 (Bastiaanssen et al., 1998; Ramanna & Jacobsen, 2003) pathways that lead to unreduced 440 gametes production. For genetic drift to have a chance to play a role in polyploidization 441 events, it is necessary that these physiological pathways are not all governed by a few major 442 443 effect mutations (Fig. 2). If there are enough loci with small effects, then it becomes more likely that genetic drift can play a role in the production of unreduced gametes and 444 polyploidization events, independently of the (mal)adaptive value of polyploid individuals. 445

### 447 *Limitations of the model*

448

Although we tried to select realistic parameter settings (e.g. for the fitness of 449 450 polyploids compared to diploids, the mutation rate of the trait), due to limited empirical 451 proofs, some assumptions could be unrealistic. This could make our predictions and further 452 empirical research unreliable. The first limitation is that we considered that the genetic architecture of unreduced gametes production is the same for the male and female functions. 453 454 While the few empirical data available do not reject this assumption (Kreiner et al., 2017a), it 455 may not be true in some species (De Storme & Geelen, 2013). Our predictions would become unreliable if the production of unreduced gametes is coded by a few numbers of major effect 456 loci in one sex, and by a large number of small effect loci in the other. In such a case, 457 polyploidization is unlikely to evolve by genetic drift, as it would be constrained by one of the 458 459 two sexes. Another simplification is that we hypothesized that triploid and higher polyploid levels are not viable. While the hypothesis is justified empirically for multiple plant species 460 (Ramsey & Schemske, 1998; Burton & Husband, 2000; Husband & Sabara, 2003; Kolář et 461 462 al., 2017), there are also species with frequent natural triploidy (Trávníček et al., 2011; Čertner et al., 2019), and it has been shown theoretically that viable triploids favour the 463 establishment of polyploid populations (Husband, 2004). Relaxing our hypothesis would not 464 change our conclusions qualitatively, since it would decrease the selection pressure against 465 the production of unreduced gametes, and favour the evolution of polyploidy by drift for a 466 467 wider set of parameters. Finally, we considered a fixed population size in our simulations, but the evolution toward polyploidy can affect the populations' survival in different ways. First, if 468 the fitness of polyploids is smaller than the one of diploids, the mean population fitness will 469 decrease, and the growth rate can become smaller than one, leading to the population's 470

decline and extinction. Second, genetic drift can also lead to the fixation of deleterious
mutations, if the average coefficient of selection of deleterious mutations is similar or smaller
to the one of mutations involved in the production of unreduced gametes. In such a case,
diploids and/or tetraploids can accumulate deleterious mutations, potentially leading to the
extinction of populations by mutational meltdown (Lynch *et al.*, 1995).

476

## 477 CONCLUSIONS

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479 In this paper, we investigated theoretically the effect of genetic drift on the probability of fixation of polyploidy in a population under a scenario assuming selection against 480 481 unreduced gamete production, a generally maladaptive trait. The main conclusions of our model are that, for polyploidization to fix, genetic drift is necessary, independently of the 482 selective (dis)advantage of tetraploid individuals over their diploid progenitors. Thus, our 483 484 simulation models suggest that the cost of being the minority cytotype in the population of 485 origin is stronger than a slight increase in fitness associated with polyploidy. The effective population size necessary for the production of unreduced gametes to evolve by genetic drift 486 487 depends on the average effects of the loci coding for the trait. This implies that if unreduced gamete production is coded by a few major effect loci, or if a species has a large effective 488 population size, polyploidization is unlikely to occur without a strong selective advantage. 489 Selfing rate, and probably other forms of assortative mating, seems to be deterministic in the 490 fixation of polyploidy only when polyploids do not suffer from an initial decrease in fitness. 491 492 However, we cannot discard that other reproductive interactions, such as fertile intermediate hybrids, can affect the outcome. On the opposite, any factors reducing the demographic and 493 effective population size are expected to favour polyploidization, independently of the 494

selective (dis)advantages of tetraploids compared to diploids. Our model thus offers new and
non-exclusive explanations of why polyploidization is associated with periods of extinction
and environmental turmoil, and harsh and/or disturbed environments (Rice *et al.*, 2019; Van
de Peer *et al.*, 2021).

Testing our predictions empirically will remain however quite challenging. Although 499 we know that the formation of unreduced gametes is determined genetically and that it 500 exhibits heritable variance (Parrott & Smith, 1986; Tavoletti et al., 1991), little is known 501 about the number of genes underlying the trait (Brownfield & Köhler, 2011; De Storme & 502 Geelen, 2013; Kreiner et al., 2017a). In order to test our hypothesis, one of the next steps 503 would be to investigate at a large taxonomic scale the genetic architecture of unreduced 504 gamete production. The other challenge is that, when polyploidization occurs, the fixation 505 time of polyploidy is really small, increasing the probability of missing the evolution of 506 polyploidy. Invasive species could be interesting model species, as founding events are 507 expected to decrease the effective population size of populations, and it is more likely for 508 509 polyploidy to evolve during the invasion process. Simón-Porcar et al. (2017) recently found a 510 mixed-ploidy population of the invasive diploid Mimulus guttatus in the UK that could be a good system for following our investigation of the role of drift in polyploidy evolution. 511

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## 513 **BIBLIOGRAPHY**

514

Arrigo, N. & Barker, M.S. 2012. Rarely successful polyploids and their legacy in plant genomes.
 *Current opinion in plant biology* 15: 140–146. Elsevier.

Baack, E.J. 2005. To succeed globally, disperse locally: effects of local pollen and seed dispersal on
 tetraploid establishment. *Heredity* 94: 538–546. Nature Publishing Group.

- Baack, E.J. & Stanton, M.L. 2005. Ecological factors influencing tetraploid speciation in snow
   buttercups (Ranunculus adoneus): niche differentiation and tetraploid establishment.
   *Evolution* 59: 1936–1944. Wiley Online Library.
- 522 Barker, M.S., Arrigo, N., Baniaga, A.E., Li, Z. & Levin, D.A. 2016. On the relative abundance of 523 autopolyploids and allopolyploids. *New Phytologist* **210**: 391–398. Wiley Online Library.
- BASTIAANSSEN<sup>+</sup>, VAN DEN BERG, LINDHOUT, JACOBSEN, & RAMANNA<sup>\*</sup>. 1998. Postmeiotic
   restitution in 2n-egg formation of diploid potato. *Heredity* 81: 20–27. John Wiley & Sons, Ltd.
- Bohutínská, M., Alston, M., Monnahan, P., Mandáková, T., Bray, S., Paajanen, P., et al. 2021. Novelty
   and convergence in adaptation to whole genome duplication. *Molecular biology and* evolution 38: 3910–3924. Oxford University Press.
- Bomblies, K. 2020. When everything changes at once: finding a new normal after genome
   duplication. *Proceedings of the Royal Society B* 287: 20202154. The Royal Society.
- Bretagnolle, F. 2001. Pollen production and spontaneous polyploidization in diploid populations of
   Anthoxanthum alpinum. *Biological Journal of the Linnean Society* 72: 241–247. Oxford
   University Press.
- Bretagnolle, F. & and THOMPSON, J.D. 1995. Gametes with the somatic chromosome number:
   mechanisms of their formation and role in the evolution of autopolyploid plants. *New Phytologist* 129: 1–22. Wiley Online Library.
- Brochmann, C., Brysting, A.K., Alsos, I.G., Borgen, L., Grundt, H.H., Scheen, A.-C., *et al.* 2004.
  Polyploidy in arctic plants. *Biological journal of the Linnean society* 82: 521–536. Oxford
  University Press.
- Brown, J.H. 1984. On the relationship between abundance and distribution of species. *The american naturalist* 124: 255–279. University of Chicago Press.
- Brownfield, L. & Köhler, C. 2011. Unreduced gamete formation in plants: mechanisms and prospects.
   Journal of experimental botany 62: 1659–1668. Oxford University Press.
- Buggs, R.J. & Pannell, J.R. 2007. Ecological differentiation and diploid superiority across a moving
   ploidy contact zone. *Evolution* 61: 125–140. Wiley Online Library.
- Burton, T.L. & Husband, B. 2000. Fitness differences among diploids, tetraploids, and their triploid
   progeny in Chamerion angustifolium: mechanisms of inviability and implications for polyploid
   evolution. *Evolution* 54: 1182–1191. Wiley Online Library.
- Čertner, M., Kúr, P., Kolář, F. & Suda, J. 2019. Climatic conditions and human activities shape diploid–
   tetraploid coexistence at different spatial scales in the common weed Tripleurospermum
   inodorum (Asteraceae). *Journal of Biogeography* 46: 1355–1366. Wiley Online Library.
- Chao, D.-Y., Dilkes, B., Luo, H., Douglas, A., Yakubova, E., Lahner, B., *et al.* 2013. Polyploids exhibit
  higher potassium uptake and salinity tolerance in Arabidopsis. *Science* 341: 658–659.
  American Association for the Advancement of Science.
- Clo, J. & Kolář, F. 2021. Short- and long-term consequences of genome doubling: a meta-analysis.
   *American Journal of Botany* 108: 2315–2322. John Wiley & Sons, Ltd.

- 557 Comai, L. 2005. The advantages and disadvantages of being polyploid. *Nature Reviews Genetics* 6:
   558 836–846.
- 559 De Storme, N., Copenhaver, G.P. & Geelen, D. 2012. Production of diploid male gametes in
   560 Arabidopsis by cold-induced destabilization of postmeiotic radial microtubule arrays. *Plant* 561 *physiology* 160: 1808–1826. American Society of Plant Biologists.
- De Storme, N. & Geelen, D. 2013. Sexual polyploidization in plants cytological mechanisms and
   molecular regulation. *New Phytologist* **198**: 670–684. John Wiley & Sons, Ltd.
- 564 Doyle, J.J. & Coate, J.E. 2019. Polyploidy, the nucleotype, and novelty: the impact of genome
  565 doubling on the biology of the cell. *International Journal of Plant Sciences* 180: 1–52.
  566 University of Chicago Press Chicago, IL.
- 567 Duchoslav, M., Fialová, M. & Jandová, M. 2017. The ecological performance of tetra-, penta-and
   568 hexaploid geophyte Allium oleraceum in reciprocal transplant experiment may explain the
   569 occurrence of multiple-cytotype populations. *Journal of Plant Ecology* 10: 569–580. Oxford
   570 University Press.
- Falconer, D.S. & Mackay, T.F.C. 1996. Introduction to quantitative genetics. Essex. UK: Longman
   *Group*.
- Felber, F. 1991. Establishment of a tetraploid cytotype in a diploid population: effect of relative
   fitness of the cytotypes. *Journal of evolutionary biology* 4: 195–207. Wiley Online Library.
- Fernández, A. & Neffa, V.G.S. 2004. Genomic relationships between Turnera krapovickasii (2x, 4x)
  and T. ulmifolia (6x)(Turneraceae, Turnera). *Caryologia* 57: 45–51. Taylor & Francis.
- Fowler, N.L. & Levin, D.A. 1984. Ecological constraints on the establishment of a novel polyploid in
   competition with its diploid progenitor. *The American Naturalist* 124: 703–711. University of
   Chicago Press.
- Glémin, S. & Ronfort, J. 2013. Adaptation and maladaptation in selfing and outcrossing species: new
   mutations versus standing variation. *Evolution: International Journal of Organic Evolution* 67:
   225–240. Wiley Online Library.
- 583 Gregory, T.R. & Mable, B.K. 2005. Polyploidy in animals. In: *The evolution of the genome*, pp. 427–
  584 517. Elsevier.
- Griswold, C.K. 2021. The effects of migration load, selfing, inbreeding depression, and the genetics of
   adaptation on autotetraploid versus diploid establishment in peripheral habitats. *Evolution* 75: 39–55. Wiley Online Library.
- Halligan, D.L. & Keightley, P.D. 2009. Spontaneous mutation accumulation studies in evolutionary
   genetics. *Annual Review of Ecology, Evolution, and Systematics* 40: 151–172. Annual Reviews.
- Hannweg, K., Steyn, W. & Bertling, I. 2016. In vitro-induced tetraploids of Plectranthus esculentus are
   nematode-tolerant and have enhanced nutritional value. *Euphytica* 207: 343–351. Springer.
- Harlan, J.R. & deWet, J.M. 1975. On Ö. Winge and a prayer: the origins of polyploidy. *The botanical review* 41: 361–390. Springer.

594	Herben, T., Trávníček, P. & Chrtek, J. 2016. Reduced and unreduced gametes combine almost freely
595	in a multiploidy system. <i>Perspectives in Plant Ecology, Evolution and Systematics</i> 18: 15–22.
596	Elsevier.
597	<ul> <li>Hias, N., Svara, A. &amp; Keulemans, J.W. 2018. Effect of polyploidisation on the response of apple</li></ul>
598	(Malus\$\times\$ domestica Borkh.) to Venturia inaequalis infection. <i>European Journal of Plant</i>
599	<i>Pathology</i> 151: 515–526. Springer.
600 601	Husband, B.C. 2004. The role of triploid hybrids in the evolutionary dynamics of mixed-ploidy populations. <i>Biological Journal of the Linnean Society</i> <b>82</b> : 537–546.
602	Husband, B.C., Ozimec, B., Martin, S.L. & Pollock, L. 2008. Mating consequences of polyploid
603	evolution in flowering plants: current trends and insights from synthetic polyploids.
604	International Journal of Plant Sciences <b>169</b> : 195–206. The University of Chicago Press.
605 606 607	<ul> <li>Husband, B.C. &amp; Sabara, H.A. 2003. Reproductive isolation and their diploid autotetraploids</li> <li>Chamerion progenitors in fireweed, Chamerion angustifolium (Onagraceae). New Phytologist</li> <li>161: 703–713.</li> </ul>
608	Kim, J.S., Oginuma, K. & Tobe, H. 2009. Syncyte formation in the microsporangium of
609	Chrysanthemum (Asteraceae): a pathway to infraspecific polyploidy. <i>Journal of Plant</i>
610	<i>Research</i> 122: 439–444. Springer.
611	Kolář, F., Čertner, M., Suda, J., Schönswetter, P. & Husband, B.C. 2017. Mixed-ploidy species:
612	progress and opportunities in polyploid research. <i>Trends in Plant Science</i> <b>22</b> : 1041–1055.
613	Elsevier.
614	Kovalsky, I.E. & Solís Neffa, V.G. 2012. Evidence of 2n microspore production in a natural diploid
615	population of Turnera sidoides subsp. carnea and its relevance in the evolution of the T.
616	sidoides (Turneraceae) autopolyploid complex. <i>Journal of plant research</i> <b>125</b> : 725–734.
617	Springer.
618 619 620	Kovalsky, I.E. & Solís Neffa, V.G. 2016. Evidence of the production of 2n eggs in diploid plants of the autopolyploid complex Turnera sidoides L.(Passifloraceae). <i>Plant systematics and evolution</i> <b>302</b> : 357–366. Springer.
621 622	Kreiner, J.M., Kron, P. & Husband, B.C. 2017a. Evolutionary dynamics of unreduced gametes. <i>Trends in Genetics</i> <b>33</b> : 583–593. Elsevier.
623	Kreiner, J.M., Kron, P. & Husband, B.C. 2017b. Frequency and maintenance of unreduced gametes in
624	natural plant populations: associations with reproductive mode, life history and genome size.
625	<i>New Phytologist</i> 214: 879–889. Wiley Online Library.
626 627	Kron, P. & Husband, B.C. 2009. Hybridization and the reproductive pathways mediating gene flow between native Malus coronaria and domestic apple, M. domestica. <i>Botany</i> <b>87</b> : 864–874.
628 629	Levin, D.A. 1975. Minority cytotype exclusion in local plant populations. <i>Taxon</i> <b>24</b> : 35–43. Wiley Online Library.
630 631 632	Li, BH., Xu, XM. & Ridout, M.S. 2004. Modelling the establishment and spread of autotetraploid plants in a spatially heterogeneous environment. <i>Journal of Evolutionary Biology</i> <b>17</b> : 562–573. Wiley Online Library.

- Lynch, M., Conery, J. & Burger, R. 1995. Mutation accumulation and the extinction of small
   populations. *The American Naturalist* 146: 489–518. University of Chicago Press.
- Mable, B.K. 2004. 'Why polyploidy is rarer in animals than in plants': myths and mechanisms.
   *Biological Journal of the Linnean Society* 82: 453–466. Oxford University Press.
- Martin, S.L. & Husband, B.C. 2013. Adaptation of diploid and tetraploid Chamerion angustifolium to
   elevation but not local environment. *Evolution* 67: 1780–1791. Wiley Online Library.
- Mason, A.S., Nelson, M.N., Yan, G. & Cowling, W.A. 2011. Production of viable male unreduced
   gametes in Brassica interspecific hybrids is genotype specific and stimulated by cold
   temperatures. *BMC plant biology* **11**: 1–13. BioMed Central.
- Mason, A.S. & Pires, J.C. 2015. Unreduced gametes: meiotic mishap or evolutionary mechanism?
   *Trends in Genetics* 31: 5–10. Elsevier.
- Mayrose, I., Zhan, S.H., Rothfels, C.J., Arrigo, N., Barker, M.S., Rieseberg, L.H., *et al.* 2015. Methods
  for studying polyploid diversification and the dead end hypothesis: a reply to Soltis et
  al.(2014). *New Phytologist* 206: 27–35. JSTOR.
- Mayrose, I., Zhan, S.H., Rothfels, C.J., Magnuson-Ford, K., Barker, M.S., Rieseberg, L.H., *et al.* 2011.
  Recently formed polyploid plants diversify at lower rates. *Science* 333: 1257–1257. American
  Association for the Advancement of Science.
- McIntyre, P.J. & Strauss, S. 2017. An experimental test of local adaptation among cytotypes within a
   polyploid complex. *Evolution* **71**: 1960–1969. Wiley Online Library.
- Meyers, L.A. & Levin, D.A. 2006. On the abundance of polyploids in flowering plants. *Evolution* 60:
   1198–1206. Wiley Online Library.
- Oberlander, K.C., Dreyer, L.L., Goldblatt, P., Suda, J. & Linder, H.P. 2016. Species-rich and polyploid poor: Insights into the evolutionary role of whole-genome duplication from the Cape flora
   biodiversity hotspot. *American Journal of Botany* **103**: 1336–1347. Wiley Online Library.
- Oswald, B.P. & Nuismer, S.L. 2011. A unified model of autopolyploid establishment and evolution.
   *The American Naturalist* 178: 687–700. University of Chicago Press Chicago, IL.
- 659 Otto, S.P. 2007. The evolutionary consequences of polyploidy. *Cell* **131**: 452–462. Elsevier.
- Parisod, C., Holderegger, R. & Brochmann, C. 2010. Evolutionary consequences of autopolyploidy.
   *New phytologist* 186: 5–17. Wiley Online Library.
- Parrott, W.A. & Smith, R.R. 1986. Evidence for the existence of endosperm balance number in the
   true clovers (Trifolium spp.). *Canadian journal of genetics and cytology* 28: 581–586. NRC
   Research Press Ottawa, Canada.
- Pécrix, Y., Rallo, G., Folzer, H., Cigna, M., Gudin, S. & Le Bris, M. 2011. Polyploidization mechanisms:
   temperature environment can induce diploid gamete formation in Rosa sp. *Journal of experimental botany* 62: 3587–3597. Oxford University Press.
- Porturas, L.D., Anneberg, T.J., Curé, A.E., Wang, S., Althoff, D.M. & Segraves, K.A. 2019. A metaanalysis of whole genome duplication and the effects on flowering traits in plants. *American journal of botany* 106: 469–476. Wiley Online Library.

- Raabová, J., Fischer, M. & Münzbergová, Z. 2008. Niche differentiation between diploid and
   hexaploid Aster amellus. *Oecologia* 158: 463–472. Springer.
- Ramanna, M.S. & Jacobsen, E. 2003. Relevance of sexual polyploidization for crop improvement–A
   review. *Euphytica* 133: 3–8. Springer.
- Ramsey, J. 2007. Unreduced gametes and neopolyploids in natural populations of Achillea borealis
   (Asteraceae). *Heredity* 98: 143–150. Nature Publishing Group.
- Ramsey, J. & Schemske, D.W. 2002. Neopolyploidy in flowering plants. *Annual review of ecology and systematics* 33: 589–639. Annual Reviews 4139 El Camino Way, PO Box 10139, Palo Alto, CA
   94303-0139, USA.
- Ramsey, J. & Schemske, D.W. 1998. Pathways, mechanisms, and rates of polyploid formation in
  flowering plants. *Annual review of ecology and systematics* 29: 467–501. Annual Reviews
  4139 El Camino Way, PO Box 10139, Palo Alto, CA 94303-0139, USA.
- Rausch, J.H. & Morgan, M.T. 2005. THE EFFECT OF SELF-FERTILIZATION, INBREEDING DEPRESSION,
   AND POPULATION SIZE ON AUTOPOLYPLOID ESTABLISHMEN. *Evolution* 59: 1867–1875. Wiley
   Online Library.
- Rice, A., Šmarda, P., Novosolov, M., Drori, M., Glick, L., Sabath, N., *et al.* 2019. The global
   biogeography of polyploid plants. *Nature Ecology & Evolution* 3: 265–273. Nature Publishing
   Group.
- Rodriguez, D.J. 1996. A model for the establishment of polyploidy in plants. *The American Naturalist* 147: 33–46. University of Chicago Press.
- Rojas-Andrés, B.M., Padilla-García, N., de Pedro, M., López-González, N., Delgado, L., Albach, D.C., *et al.* 2020. Environmental differences are correlated with the distribution pattern of cytotypes
   in Veronica subsection Pentasepalae at a broad scale. *Annals of botany* 125: 471–484. Oxford
   University Press US.
- Ruiz, M., Quiñones, A., Martínez-Cuenca, M.-R., Aleza, P., Morillon, R., Navarro, L., *et al.* 2016.
   Tetraploidy enhances the ability to exclude chloride from leaves in carrizo citrange seedlings.
   *Journal of plant physiology* 205: 1–10. Elsevier.
- Sheidai, M., Jafari, S., Taleban, P. & Keshavarzi, M. 2009. Cytomixis and unreduced pollen grain
  formation in Alopecurus L. and Catbrosa Beauv.(Poaceae). *Cytologia* 74: 31–41. Japan
  Mendel Society, International Society of Cytology.
- Simón-Porcar, V.I., Silva, J.L., Meeus, S., Higgins, J.D. & Vallejo-Marín, M. 2017. Recent
   autopolyploidization in a naturalized population of Mimulus guttatus (Phrymaceae).
   *Botanical Journal of the Linnean Society* 185: 189–207. Oxford University Press.
- Soltis, D.E., Soltis, P.S., Schemske, D.W., Hancock, J.F., Thompson, J.N., Husband, B.C., *et al.* 2007.
   Autopolyploidy in angiosperms: have we grossly underestimated the number of species?
   *Taxon* 56: 13–30. Wiley Online Library.
- 707 Stebbins, G.L. 1971. Processes of organic evolution.
- 708 Stebbins, G.L. 1950. *Variation and evolution in plants*. Columbia University Press.

- Tavoletti, S., Mariani, A. & Veronesi, F. 1991. Phenotypic recurrent selection for 2n pollen and 2n egg
   production in diploid alfalfa. *Euphytica* 57: 97–102. Springer.
- Tayalé, A. & Parisod, C. 2013. Natural pathways to polyploidy in plants and consequences for genome
   reorganization. *Cytogenetic and Genome Research* 140: 79–96. Karger Publishers.
- Te Beest, M., Le Roux, J.J., Richardson, D.M., Brysting, A.K., Suda, J., Kubešová, M., *et al.* 2012. The
  more the better? The role of polyploidy in facilitating plant invasions. *Annals of botany* 109:
  19–45. Oxford University Press.
- Trávníček, P., Dočkalová, Z., Rosenbaumová, R., Kubátová, B., Szeląg, Z. & Chrtek, J. 2011. Bridging
   global and microregional scales: ploidy distribution in Pilosella echioides (Asteraceae) in
   central Europe. Annals of Botany 107: 443–454. Oxford University Press.
- Van de Peer, Y., Ashman, T.-L., Soltis, P.S. & Soltis, D.E. 2021. Polyploidy: an evolutionary and
   ecological force in stressful times. *The Plant Cell* 33: 11–26. Oxford University Press.
- Van de Peer, Y., Mizrachi, E. & Marchal, K. 2017. The evolutionary significance of polyploidy. *Nature Reviews Genetics* 18: 411. Nature Publishing Group.
- Van Drunen, W.E. & Friedman, J. 2021. Autopolyploid establishment depends on life history strategy
   and the mating outcomes of clonal architecture. *bioRxiv* 2021.10.21.465190.
- Vanneste, K., Baele, G., Maere, S. & Van de Peer, Y. 2014a. Analysis of 41 plant genomes supports a
   wave of successful genome duplications in association with the Cretaceous–Paleogene
   boundary. *Genome research* 24: 1334–1347. Cold Spring Harbor Lab.
- Vanneste, K., Maere, S. & Van de Peer, Y. 2014b. Tangled up in two: a burst of genome duplications
   at the end of the Cretaceous and the consequences for plant evolution. *Philosophical Transactions of the Royal Society B: Biological Sciences* 369: 20130353. The Royal Society.
- Wang, W., He, Y., Cao, Z. & Deng, Z. 2018. Induction of tetraploids in impatiens (Impatiens walleriana) and characterization of their changes in morphology and resistance to downy mildew. *HortScience* 53: 925–931. American Society for Horticultural Science.
- Wood, T.E., Takebayashi, N., Barker, M.S., Mayrose, I., Greenspoon, P.B. & Rieseberg, L.H. 2009. The
   frequency of polyploid speciation in vascular plants. *Proceedings of the national Academy of sciences* 106: 13875–13879. National Acad Sciences.
- Yao, Y., Carretero-Paulet, L. & Van de Peer, Y. 2019. Using digital organisms to study the evolutionary
   consequences of whole genome duplication and polyploidy. *PloS one* 14: e0220257. Public
   Library of Science San Francisco, CA USA.
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**Figure 1.** Dynamics of the proportion of tetraploid individuals through time for different strengths of genetic drift when tetraploid individuals are less fit (left panel), as fit (middle panel) or fitter (right panel) than their diploid progenitors. The thin lines represent individual simulations (n = 100), and the thick line is the mean proportion of polyploids among simulations. Other parameters are L = 100 (and then  $\overline{s_0} = 0.01$ ), h = 0.5,  $\sigma = 0$ , U = 0.1 and Ncan be equal to 50 (Drift > Selection), 100 (Drift = Selection) or 200 (Drift < Selection).



**Figure 2.** Probability of fixation of polyploidy as a function of the number of loci underlying the trait and the relative fitness of diploid and tetraploid individuals ( $W_{\text{tetra}} = 0.75$  in blue, 1.25 in yellow, or 2 in red). *N* is equal to 100, and *L* varies from 2 to 200, genetic drift is stronger than selection when L > 100. Other parameters are h = 0.5,  $\sigma = 0$ , U = 0.1,  $\bar{s_0}$  varies from 0.005 (L = 200) to 0.5 (L = 2). The probability is computed on n = 100 replicates.



775 Figure 3. Production of unreduced gametes in diploid individuals, as a function of strength of 776 selection compared to genetic drift, for different mutation rates (left panel U = 0.005, right panel U = 0.05), when tetraploid individuals are less fit than diploid ones. The white points 777 778 represent the average production of unreduced gametes and is estimated with respectively n =5000, 10.000 and 20.000 estimates respectively when N = 50, 100 or 200 (100 replicates 779 times the population size). Other parameters are L = 100 (and then  $\overline{s_0} = 0.01$ ), h = 0.5,  $\sigma = 0$ 780 781 and N can be equal to 50 (Drift > Selection), 100 (Drift = Selection) or 200 (Drift < Selection). 782



Figure 4. Dynamics of the proportion of tetraploid individuals in time, as a function of the 786 strength of the selfing rate in the population ( $\sigma = 0$ : obligate outcrossers in orange,  $\sigma = 0.5$ : 787 non-obligate outcrossers in light green,  $\sigma = 1$ : obligate selfers in blue). (Left panel) Polyploids 788 are less fit than diploids, and genetic drift is stronger than natural selection. (Middle panel) 789 Polyploids are fitter than diploids, and genetic drift is stronger than natural selection. (Right 790 panel) Polyploids are fitter than diploids, and genetic drift is weaker than natural selection. 791 792 The thin lines represent individual simulations (n = 100), and the thick line is the mean proportion of polyploids among simulations. Other parameters are L = 100 (and then  $\overline{s_0} =$ 793 0.01), h = 0.5, U = 0.1 and N can be equal to 50 (Drift > Selection) or 200 (Drift < Selection). 794