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# Estimates of absolute crown strength and bite force in the lower postcanine dentition of *Gigantopithecus blacki*

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#### **Abstract**

Gigantopithecus blacki is hypothesized to have been capable of processing mechanically challenging foods, which likely required this species to have high dental resistance to fracture and/or large bite force. To test this hypothesis, we used two recently developed approaches to estimate absolute crown strength and bite force of the lower postcanine dentition. Sixteen Gigantopithecus mandibular permanent cheek teeth were scanned by micro-computed tomography. From virtual mesial cross-sections, we measured average enamel thickness and bicervical diameter to estimate absolute crown strength, and cuspal enamel thickness and dentine horn angle to estimate bite force. We compared G. blacki with a sample of extant great apes (Pan, Pongo, and Gorilla) and australopiths (Australopithecus anamensis, Australopithecus afarensis, Australopithecus africanus, Paranthropus robustus, and Paranthropus boisei). We also evaluated statistical differences in absolute crown strength and bite force between the premolars and molars for G. blacki. Results reveal that molar crown strength is absolutely greater, and molar bite force absolutely higher, in G. blacki than all other taxa except P. boisei, suggesting that G. blacki molars have exceptionally high resistance to fracture and the ability to generate exceptionally high bite force. In addition, G. blacki premolars have comparable absolute crown strength and larger bite force capabilities compared with its molars, implying possible functional specializations in premolars. The dental specialization of G. blacki could thus represent an adaptation to further facilitate the processing of mechanically challenging foods. While it is currently not possible to determine which types of foods were actually consumed by G. blacki through this study, direct evidence (e.g. dental chipping and microwear) left by the foods eaten by G. blacki could potentially lead to greater insights into its dietary ecology. Keywords: Chuifeng Cave; Mohui Cave; Dietary adaption; Mechanically challenging foods; Fallback foods

#### 1. Introduction

In 1935, von Koenigswald described a huge, high-crowned molar that he purchased from a Chinese drugstore in Hong Kong and found this molar to present distinctive features compared to other known primate teeth and erected the species *Gigantopithecus blacki* for it (Von Koenigswald, 1935). Evidence accumulated since then reveals that this extinct giant ape is the largest primate ever discovered (Zhang and Harrison, 2017). Most of the *G. blacki* specimens come from South China (Pei and Woo, 1956; Woo, 1962; Zhang, 1982; Wang, 2009; Jin et al., 2009, 2014; Zhao and Zhang, 2013; Wang et al., 2017; Zhang and Harrison, 2017), while a few isolated teeth from North Vietnam (Ciochon et al., 1996), North Thailand (Bocherens et al., 2017), and Java (Noerwidi et al., 2016; Zanolli et al., 2019) have also been reported. Over the

past six decades, nearly two thousand isolated teeth as well as four partial mandibles have been unearthed from ongoing cave excavations across South China (Zhang and Harrison, 2017). The estimated age of *G. blacki* ranges from ~2.0 Ma to ~300 ka (initial Early Pleistocene to Middle Pleistocene), as indicated by associated faunal assemblages and direct dating methods (Sun et al., 2014; Shao et al., 2014, 2015, 2017). Since the initial discovery of *G. blacki*, its phylogenetic relationship has been questioned. *Gigantopithecus blacki* was once considered to be a hominin (Weidenreich, 1945; Broom and Schepers, 1946; Gelvin, 1980; Zhang and Zhao, 2013). However, some also considered it to be a specialized great ape, with a possible close affinity to the *Sivapithecus*—*Indopithecus* clade (Pei and Woo, 1956; Pilbeam, 1970; Miller et al., 2008; Olejniczak et al., 2008a; Begun, 2010). Recently, Welker et al. (2019) successfully extracted dental proteins from *G. blacki* molar enamel and demonstrated that this species was an early diverging pongine.

Attempts have been made to determine the dietary behavior of G. blacki through a variety of approaches, such as dentognathic morphology (e.g. enamel thickness, tooth root length, occlusal area, and mandibular corpus depth; Woo, 1962; Olejniczak et al., 2008a; Kupczik and Dean, 2008; Zhang and Zhao, 2013; Kono et al., 2014; Zhang and Harrison, 2017), stable isotope analysis (Nelson, 2014; Qu et al., 2014; Bocherens et al., 2017; Jiang et al., 2021; Hu et al., 2022), dental microwear (Daegling and Grine, 1994; Zhao and Zhang, 2013), incidence of dental caries (Han and Zhao, 2002; Wang, 2009), analysis of phytoliths (Ciochon et al., 1990), and analysis of starch grains (Qu, 2014). These studies provided indications—e.g. enhanced enamel stability reflected by high calcium (Ca) isotope values, possibly being adapted to the consumption of hard foods (Hu et al., 2022)—that G. blacki may have been capable of processing mechanically challenging foods (MCFs), i.e. tough foods (Dickson, 2003; Olejniczak et al., 2008a; Kono et al., 2014) and/or hard foods (Kupczik and Dean, 2008; Qu, 2014; Hu et al., 2022). Feeding on tough or hard foods may require high dental resistance to fracture associated with repetitive and/or intense loading, respectively, while feeding on hard foods may further require relatively large bite force (BF). However, relatively few quantitative studies have investigated these dental biomechanical properties for G. blacki (e.g. Kupczik and Dean, 2008). Here, we address the following questions: 1) can the teeth of G. blacki resist breakage when processing MCFs and 2) does G. blacki have absolutely larger BF than extant apes and australopiths?

Schwartz et al. (2020) and Chai (2018) recently derived methods to estimate tooth resistance to fracture and BF, respectively. Schwartz et al. (2020) proposed a new metric, absolute crown strength (ACS), as a proxy for assessing tooth resistance to fracture. This parameter can be readily measured on the mesial section of a tooth. Estimating ACS involves two dental anatomical variables, i.e. average enamel thickness (AET) and bi-cervical diameter. In the Schwartz et al. (2020) study, tooth crown strength was reflected by two indices. One is the critical force (denoted by PRF) needed to fully propagate a radial-median fracture from cusp to cervix, and the other is the critical force (denoted by PMF) needed to fully propagate a margin fracture from cervix to cusp. Dental radial-median fractures can only be caused by biting on hard food items, while margin fractures can be induced by biting on hard or soft objects (Schwartz et al., 2020). The higher the tooth crown strength, the larger the critical force needed to cause radial-median fractures or margin fractures, and thus the larger the PRF and PMF. Both PRF (Supplementary Online Material [SOM] S1) and P<sub>MF</sub> (SOM S1) can be estimated according to the formulae derived by Lawn and Lee (2009). It is worth mentioning that in the biomechanical work of Lawn and Lee (2009), a tooth was modeled as a simplified dome-like structure with an equalthickness enamel layer sitting atop a semicircular dentine base. To test whether ACS is a

significant predictor of tooth crown strength (i.e.  $P_{RF}$  and  $P_{MF}$ ), Schwartz et al. (2020) estimated ACS,  $P_{RF}$ , and  $P_{MF}$  for a sample of 139 extant hominoid and extinct hominin mandibular molars and then performed least squares regressions between ACS and  $P_{RF}$ , and between ACS and  $P_{MF}$ . Results showed that ACS exhibited significant linear correlations with both  $P_{RF}$  and  $P_{MF}$  (with  $r^2 > 0.86$ ), indicating that ACS is strongly correlated with the Lawn and Lee (2009) force estimates of tooth fracture.

In the numerical model proposed by Chai (2018), a tooth cusp was modeled as a truncated cone characterized by equal-thickness enamel layer that rests on conical-like dentine base (Chai, 2018: Fig. 4), which is different from the scheme for deriving ACS (Lawn and Lee, 2009; Schwartz et al., 2020). In addition, occlusal loading was simulated as a tooth cusp biting on a hard particle entrapped at the central fossa of the opposing tooth. With these simplifications on cusp structure and occlusal loading form, Chai (2018) conducted a series of finite element analyses (FEAs) to evaluate the conditions for cusp failure under occlusal loading. Simulation results showed that 1) tensile stresses were greater at the enamel-dentine junction (EDJ) and the peak stress occurred slightly below the dentine horn tip and 2) BF was correlated with two dental anatomical parameters: cusp enamel thickness and dentine horn angle (DHA). In parallel with the numerical simulations, Chai (2018) carried out tooth fracture tests to examine the reliability of the numerical model. Specifically, the polished cusps of human maxillary molars were subjected to axial occlusal force loaded by a tungsten carbide ball. After unloading, teeth were cut open to observe internal crack patterns. Internal cracking patterns showed general consistency with the numerical simulation results. For instance, in both cases, the enamel cracks extended from the EDJ and the peak stress occurred near the EDJ. Thus, despite underlying assumptions and limitations, the simplifications in the numerical model were deemed acceptable (Chai, 2018, 2020). Although there are other methods to estimate BF, most require cranial material (e.g. Demes and Creel, 1988; O'Connor et al., 2005; Wroe et al., 2010; Constantino et al., 2010; Eng et al., 2013) and are not suitable for the current study because no cranial remains of G. blacki have been found.

To test the hypothesis that *G. blacki* was capable of processing MCFs, we adopted the methods of Schwartz et al. (2020) and Chai (2018) to estimate ACS and BF, respectively. We then compared ACS and BF between *G. blacki* and a sample of extant great apes and extinct australopiths. The dietary profiles of some of the extant great apes have been well documented (e.g. Elgart-Berry, 2004; Vogel et al., 2008, 2009, 2014; Yamagiwa and Basabose, 2009; Coiner-Collier et al., 2016). We thus use these profiles to determine whether *G. blacki* had processing capability comparable to those of extant great apes. For instance, if *G. blacki* has higher ACS and absolutely larger BF than *Pongo*, then *G. blacki* teeth were likely capable of withstanding the stresses associated with biting on hard foods, given that some *Pongo* species have been observed to consume hard seeds in the wild (i.e. *Pongo abelii* and *Pongo pygmaeus wurmbii* in the wild; Vogel et al., 2014) and in captivity (e.g. *P. pygmaeus*; Lucas et al., 1994). Australopiths and *G. blacki* have dentognathic traits that have been adaptively linked to processing MCFs, such as robust jaws and thickly-enameled cheek teeth (Teaford and Ungar, 2000; Olejniczak et al., 2008b), yet it is not clear how they differ in their resistance to tooth fracture and BF potential. Therefore, we also compared ACS and BF between *G. blacki* and australopiths.

# 2. Materials and methods

# 2.1. Samples

In this study, we focused on mandibular permanent postcanine teeth of *G. blacki*, including four P<sub>3</sub>s, four P<sub>4</sub>s, four M<sub>1</sub>s or M<sub>2</sub>s, and four M<sub>3</sub>s (Table 1). All samples are isolated teeth and none

show obvious dental pathology. According to the wear stage identification method by Molnar (1971), all teeth studied here are slightly worn, i.e. no wear facets (wear stage = 1) or small wear facets without dentine exposure (wear stage = 2). Only slightly worn teeth (maximum wear stage 2) were deemed suitable for this study as the worn enamel portion needed to be reconstructed in the mesial section prior to estimating ACS and BF, and it would become difficult and less reliable to reconstruct the missing enamel for moderately or heavily worn teeth (wear stage >2). The high degree of morphological similarity between the  $M_1s$  and  $M_2s$  made it difficult to confidently distinguish between them and thus both positions were provisionally placed into one analytical category ( $M_{1/2}$ ). All teeth were excavated in situ from Bubin basin, Guangxi province, China. Two of these teeth (Z010670 and Z010071) are from Mohui Cave (Wang et al., 2005) and the remaining teeth are from Chuifeng Cave (Wang, 2009). These teeth are now stored in the Anthropology Museum of Guangxi, China.

Comparative data for molar ACS and molar BF were compiled from the literature (Skinner et al., 2015; Chai, 2020; Schwartz et al., 2020) and are provided in SOM Tables S1 and S2. The samples of extant great apes include *Pan paniscus*, *Pan troglodytes*, *Pongo* sp., and *Gorilla* sp. Australopiths include specimens assigned to *Australopithecus anamensis*, *Australopithecus africanus*, *Paranthropus robustus*, and *Paranthropus boisei*. Species classification and sample sizes (number of permanent molar teeth) are presented in Table 2. Note that the premolar results (ACS and BF) of these comparative taxa were not provided in previous studies, so we only compared the differences for molars. We also did not carry out comparisons for each molar position due to the small sample sizes for many taxa, and instead, M<sub>1</sub>–M<sub>3</sub> were pooled for comparisons.

#### 2.2. Data collection

The methods used in this study rely on mesial sections to estimate ACS and BF. To obtain mesial sections, each tooth was scanned using  $\mu$ CT with a 225 kV- $\mu$ CT scanner at the Institute of Vertebrate Paleontology and Paleoanthropology (IVPP), Chinese Academy of Sciences, with the following parameters: 140 kV, 120  $\mu$ A, 0.5 angular increment one step, and 360 degrees of rotation. The resultant isometric voxel size (spatial resolution) ranged from 20.39 to 25.90  $\mu$ m (Table 1). Using IVPP225kVCT\_Recon v. 1.0 (IVPP, Beijing), raw projections were converted into image stacks, which were then segmented with the watershed algorithm in Avizo v. 8.0 (www.thermofisher.com). Surface models were generated using the constrained smoothing algorithm with a kernel size of three.

The surface model of a P<sub>3</sub> (Z008545) is shown in Figure 1a to illustrate how the virtual mesial sections were produced. This process was performed in Geomagic Design v. X 64 (www.3dsystems.com). Following the protocols of Tafforeau (2004) and extended by Benazzi et al. (2014), a spline curve was digitized along the cervical line path and then a best fitting plane was produced according to the digitized cervical line. Once this has been accomplished, a mesial section, which is perpendicular to the cervical plane and passing through the two mesial dentine horn tips, can be determined. The resultant mesial section is shown in Figure 1b. For specimens showing slightly worn enamel, the missing enamel was manually reconstructed (Fig. 1c) based on the profiles of unworn teeth (Smith et al., 2012). Skinner et al. (2015) and Lockey et al. (2020) demonstrated that manual enamel reconstruction for slightly worn teeth is generally associated with relatively small error (2.2% and 2.0%, respectively) in 2D enamel thickness measurements. We, therefore, deem the manual reconstruction method to be acceptable in the current study since our main purpose for restoring missing enamel is to measure 2D enamel thickness. Nevertheless, given the subjective nature of enamel reconstruction, different researchers may obtain different

results for a given mesial section. For this reason, the mesial sections prior to reconstruction are provided in SOM Figure S1 to enable other researchers to evaluate the results shown in this work. Following enamel reconstruction, AET (mm) and bi-cervical diameter (BCD, mm) were measured. According to Martin (1985), the expression of AET is shown as:

$$AET = A_{enamel}/L_{edj}$$
 (1)

where  $A_{enamel}$  and  $L_{edj}$  are the enamel area (mm<sup>2</sup>) and the length (mm) of the EDJ (green dashed line in Fig. 1b), respectively. The calculation of ACS (mm) is shown in Eq. (2) (Schwartz et al., 2020):

$$ACS = (AET \times BCD/2)^{0.5}$$
 (2)

We measured DHA and cuspal enamel thickness (CET) to estimate BF. Figure 1c illustrates how DHA  $(\phi)$  is measured. CET  $(d_c, mm)$  is the average of three enamel thickness parameters: cuspal inner enamel thickness  $(d_i, occlusal basin side)$ , cuspal outer enamel thickness  $(d_o, buccal side for protoconid)$ , and cuspal tip enamel thickness  $(d_t, the maximum distance between the dentine horn tip and enamel cusp apex; see the illustration in Fig. 1c).$ 

Following Chai (2018), BF (N) can be estimated using the following equation:

$$BF=sA0q^{-1}dc^2$$

where 's' is a safety factor, defined as the ratio of BF to fracture force (the critical pressure needed to fracture a tooth). Since the safety factor is assumed to be independent of taxon (Chai, 2018), this parameter remains a constant. Chai (2018) calculated the safety factor from a sample of modern human teeth. In the case of modern human molars, in vivo measured average BF was reported as 738 N (Braun et al., 1995; O'Connor et al., 2005) and the estimated average fracture force based on FEA is 1359 N (Chai, 2018). Therefore, the safety factor is approximately equal to 0.55 (s = 738/1359). The parameter  $A_0$  is related to tooth material properties and is expressed as:

$$A_0 = \sigma_F/B \tag{4}$$

$$B \equiv (1 - 2v_e) / 4\pi + 0.72 \log \left[ E_e \left( 1 - v_d^2 \right) / E_d \left( 1 - v_e^2 \right) \right]$$
 (5)

where  $\sigma_F$  is the peak fracture stress, which occurs slightly below the dentine horn tip,  $E_e$  and  $E_d$  are the elasticity modulus for enamel and dentine, respectively, and  $\upsilon_e$  and  $\upsilon_d$  are the Poisson's ratios for enamel and dentine, respectively. The experimental measurements of human tooth properties revealed that the mean values of  $\sigma_F$ ,  $E_e$ ,  $E_d$ ,  $\upsilon_e$ , and  $\upsilon_d$  were 118.7 MPa, 90 GPa, 18 GPa, 0.28, and 0.31, respectively (Lees and Rollins, 1972; Xu et al., 1998; Chai, 2014). Substituting these material parameters into Eq. (4) yields  $A_0 = 224$  MPa. To determine the expression of q, Chai (2018) applied FEA to perform a series of numerical simulations, where the force loaded on the tooth cusp remained constant (1 N) while  $\phi$  varied from 15° to 175° and  $d_c$  had four variable choices (0.55, 1.0, 1.5, and 1.9 mm, respectively). By controlling variables, the relationship between q and  $\phi$  for several choices of  $d_c$  can be obtained from the FEA results (Chai, 2018: Fig. 5a). Results showed that q has a polynomial relationship with DHA (Eq. (6)) regardless of the choices of  $d_c$ .

$$q=0.212(1+a\phi+b\phi^2+c\phi^3)/(1+d\phi+e\phi^2+f\phi^3+g\phi^4) \tag{6}$$

[a, b, c, d, e, f, g]=
$$10^{-6}$$
[-23974, 227.95, -0.6569, -26535, 227.95, -1.3058, 0.0023] (7)

When the DHA is  $<150^{\circ}$ , q follows a positive trend with increasing dentine angle (Chai, 2018: Fig. 5). Given that the DHA is  $<150^{\circ}$  for most hominid teeth (Chai, 2020), it is assumed that a

positive relationship between  $\phi$  and q holds in most cases. Since both safety factors and the material-related parameter  $A_0$  are assumed to be independent of taxon, Chai (2018) assumed that BF is mainly controlled by DHA and CET.

For the studied *G. blacki* mesial sections, the buccal dentine horns show conical-like shape while the lingual dentine horns generally do not (e.g. Z008583 in SOM Fig. S1). The method proposed by Chai (2018) is limited to the tooth cusps having conical-like dentine horns, so we measured DHA and CET only on the buccal cusps. Supplementary Online Material Figure S1 displays the buccal cusps in mesial sections and illustrates how DHA and CET were measured for each sample.

# 2.3. Data analysis

To test for pairwise statistical differences in molar ACS and molar BF between *G. blacki*, extant great apes, and australopiths, we performed nonparametric one-tailed Mann-Whitney (M-W) Utests, evaluating all possible pairwise differences. We used two-tailed M-W U tests to test for statistical differences in ACS, CET, DHA, q, and BF between premolars (results of P<sub>3</sub> and P<sub>4</sub> were pooled) and molars (results of M<sub>1/2</sub> and M<sub>3</sub> were pooled) in *G. blacki*. Here, we did not carry out pairwise comparisons for each tooth position because the sample size per tooth position was too small. The Benjamini–Hochberg (B-H) correction was adopted to minimize Type I error for all comparisons (Benjamini and Hochberg, 1995) according to the following two steps. First, all comparisons were ranked by their *p*-values from the smallest to the largest. Second, the original *p*-value for each comparison was corrected by the adjusted *p*-value using the following Eq. (8):

adjusted 
$$p$$
-value = original  $p$ -value  $\times n/k$  (8)  
adjusted  $p$ -value = original  $p$ -value  $\times n$  (9)  
adjusted  $p$ -value = original  $p$ -value  $\times (n-k+1)$  (10)

where n is the total number of comparisons and k is the rank of a given comparison. Compared with the B-H correction, the Bonferroni-type corrections (e.g. standard Bonferroni correction using Eq. (9) and sequential Bonferroni correction using Eq. (10); Holm, 1979; Rice, 1989) are more stringent and may result in rejection of true positive results when there is a large number of comparisons (Benjamini and Hochberg, 1995). As this is likely to be the case in this study, the B-H correction was adopted here. The significance level for all tests was set at p < 0.05. All statistical procedures were carried out using Origin v. 9.0 (OriginLab, Northampton).

#### 3. Results

# 3.1. Absolute crown strength estimates

The results of AET, BCD, and the estimates of ACS for *G. blacki* are provided in Table 3 and Figure 2. Qualitatively, from anterior to posterior along the postcanine tooth row, ACS and AET show subtle increases while BCD basically remains unchanged. Mean P<sub>3</sub> ACS is slightly smaller than that of other tooth positions, while mean P<sub>4</sub> ACS is comparable to that of M<sub>1/2</sub> and M<sub>3</sub>. The average ACS of the premolars (pooled estimate for P<sub>3</sub> and P<sub>4</sub> = 3.811 mm) is approximately equal to that of the molars (pooled estimate for M<sub>1/2</sub> and M<sub>3</sub> = 3.990 mm). No statistical differences were found between the premolars and molars (M-W U statistic = 25, Z = -0.683, p = 0.505). Thus, *G. blacki* premolars and molars have comparable resistance to fracture. There are significant differences in molar ACS among extant great apes, with molar ACS of *Gorilla* being significantly higher than that for *Pongo* and *Pan*, and molar ACS of *Pongo* being significantly higher than that for *Pan* (Fig. 3; Table 4). Within australopiths, all pairwise comparisons of molar ACS show significant differences (Table 4). In ascending order of molar

ACS, these australopiths species are ranked as follows: *A. anamensis* < *A. afarensis* <

#### 3.2. Bite force estimates

The results of CET, DHA, q, and the estimates of BF potential at each tooth position along the postcanine tooth row for *G. blacki* are provided in Table 5 and Figure 4. Results indicate that BF potential is larger at the premolars than at the molars (Table 5) and is significantly larger at the premolars (pooled estimates) compared with the molars (pooled estimates; M-W U = 55, Z = 2.363, p = 0.015). Moreover, statistical comparisons between the premolars (P<sub>3</sub> and P<sub>4</sub> pooled) and molars (M<sub>1/2</sub> and M<sub>3</sub> pooled) for *G. blacki* revealed: 1) no significant CET difference (M-W U = 19, Z = -1.315; p = 0.185), 2) significant DHA difference (M-W U = 1, Z = -3.203, p < 0.001), and 3) significant q difference (M-W U = 1, Z = -3.203, p < 0.001). Therefore, from anterior to posterior along the postcanine tooth row, CET shows subtle increases (Fig. 4b) and DHA shows significant increases (Fig. 4c). The latter indicates that the dentine horns of premolars are sharper than that of the molars. Corresponding to the variation of DHA, q increases distally (Fig. 4d).

Molar BF comparisons among the nine comparative taxa are provided in Figure 5 and Table 6. Among the extant great apes, *Gorilla gorilla* exhibits the significantly largest molar BF potential while BF potential is significantly larger in *P. pygmaeus* than *P. troglodytes*. All pairwise comparisons among australopiths in molar BF potential are significantly different except between the two *Paranthropus* species, and between *A. anamensis* and *A. afarensis*. The molar BF potential of *Paranthropus* is significantly larger than that of all *Australopithecus* species studied. *Gigantopithecus blacki* shows significantly larger molar BF potential than all extant great apes, *Australopithecus*, and *P. robustus* (Table 6). *Gigantopithecus blacki* and *P. boisei* do not significantly differ in molar BF (p > 0.05; Table 6).

#### 4. Discussion and conclusions

# 4.1. Adaptions of Gigantopithecus blacki lower postcanine dentition

Our results show that *G. blacki* has high molar ACS, indicating that its teeth can resist damage caused by intense and/or repeated loading. In addition, the estimated molar BF of *G. blacki* is significantly larger than all species compared except *P. boisei*, suggesting the potential capability of breaching hard objects. Moreover, compared with its molars, *G. blacki* premolars have comparable ACS and larger BF potential. The molarized premolar morphology, with low cusps and relatively flat occlusal basins (Zhang and Harrison, 2017), and the structural properties of crown tissues of *G. blacki* premolars (reflected by ACS and BF, as well as by enamel thickness; Zhang and Zhao, 2013; Pan et al., 2021) may suggest some functional specializations in the premolars. Similar phenomena (but possibly through different evolutionary paths) can be observed in extant primates that primarily feed on tough foods (such as folivores) or hard foods (such as hard-object feeders). For instance, compared to frugivores and insectivores, folivorous primates have statistically larger premolar rows relative to palate area and hard-object feeders generally have larger fourth premolars relative to the first molar (Daegling et al., 2011; Scott et al., 2018).

High ACS and the capacity to generate absolutely large BF along the postcanine dentition, as well as other distinctive dentognathic features, such as an absolutely deep mandibular corpus and relatively thickly-enameled cheek teeth with absolutely large occlusal areas and absolutely long

roots (Woo, 1962; Kupczik and Dean, 2008; Zhang et al., 2014, 2015, 2016) all indicate that *G. blacki* is characterized by a robust masticatory apparatus that has been interpreted as a potential adaptation to processing MCFs.

# 4.2. Differentiating between tough and hard foods?

Our results allow us to infer that G. blacki was capable of processing a wide range of food types including MCFs, but cannot provide direct clues to distinguishing between tough and hard foods or can we say whether G. blacki fed opportunistically on MCFs, i.e. as fallback foods when preferred foods were scarce (e.g. Constantino and Wright, 2009; Constantino et al., 2009; Marshall et al., 2009), or whether G. blacki fed regularly on MCFs. Differentiation between tough and hard foods can be better appreciated in our study through data on the feeding ecology of extant great apes and through paleodietary inferences of australopiths. First, although Gorilla molars show significantly higher ACS and larger estimated BF potential than Pongo (Table 4, Table 6; Figure 3, Figure 5), the overall diet of *Gorilla* is generally less resistant to deformation and fracture than that of *Pongo*, as demonstrated by previous studies on food material properties (Elgart-Berry, 2004; Taylor et al., 2008; Vogel et al., 2008, 2014; but see also van Casteren et al., 2019). Second, it is well appreciated that all extant great apes prefer ripe fruits and they switch to more mechanically resistant fallback foods (e.g. tough leaves, bark, and pith) when preferred foods are not available (Vogel et al., 2008, 2009; Yamagiwa and Basabose, 2009; Harrison and Marshall, 2011). This phenomenon suggests that primates may actually avoid those plant tissues to which their masticatory systems are adapted when more nutritious, less mechanically challenging food items are available (Wich et al., 2006; Ungar, 2011, 2019; Grine et al., 2012). Third, our results show that Australopithecus and Paranthropus have high molar ACS and large estimated molar BF (Figure 3, Figure 5), higher even than Pongo (Table 4, Table 6) who are known to feed on hard foods (e.g. P. pygmaeus wurmbii; Vogel et al., 2014). The high molar ACS and large estimated BF in Australopithecus and Paranthropus could thus possibly reflect adaptations for consuming hard foods. However, direct evidence in the form of microwear signatures and tooth chipping left by actual foods eaten generally do not support the routine mastication of hard objects by Australopithecus or P. boisei (e.g. Ungar et al., 2008, 2010; Ungar, 2011; Ungar and Sponheimer, 2011; Cerling et al., 2011, 2013; Wynn et al., 2013; Constantino and Konow, 2021; Towle et al., 2021). Thus, while our results indicate that P. boisei was likely well suited to comminuting hard foods, P. boisei teeth have low microwear texture complexity, low to moderate microwear anisotropy values, and low levels of dental chipping, all of which are at odds with the regular consumption of hard objects (Ungar et al., 2008; Cerling et al., 2011; Constantino and Konow, 2021; Towle et al., 2021).

Collectively, high ACS and large BF do not necessarily predict a tougher or harder diet, and they do not necessarily indicate frequent or opportunistic feeding on MCFs. Species with specialized morphologies can have generalized diets or eat more preferred food items when available. This phenomenon, originally documented in a study of African cichlid fishes by Liem (1980), is known as Liem's Paradox (Robinson and Wilson, 1998). It is possible that this phenomenon also characterizes *G. blacki* and australopiths.

4.3. Potential future directions to gain insights into the dietary ecology of Gigantopithecus blacki Carbon (C) and oxygen (O) isotopic analyses of tooth enamel have suggested that G. blacki foraged in densely forested habitat and consumed C<sub>3</sub> plants exclusively (Zhao et al., 2011; Zhao and Zhang, 2013; Nelson, 2014; Qu et al., 2014; Bocherens et al., 2017; Jiang et al., 2021). In this paleoecological context, potential food sources would have been plentiful, leading to speculation

about the foods possibly consumed by *G. blacki*, such as fruits (Woo, 1962; Ciochon et al., 1990; Han and Zhao, 2002; Wang, 2009), tough items (Daegling and Grine, 1994; Zhao and Zhang, 2013), and/or hard objects (Ciochon et al., 1990; Kupczik and Dean, 2008; Qu, 2014; Hu et al., 2022). Previous studies of enamel carbon isotopes and starch grains that bonded to the enamel surface suggest that *G. blacki* may have had a broad dietary range (Qu, 2014; Qu et al., 2014; Nelson, 2014; Bocherens et al., 2017). Our results, specifically the high ACS and large postcanine BF potential, further suggest that *G. blacki* was able to process a broad range of food items including MCFs.

Substantial efforts have been made in previous studies to improve our understanding of G. blacki dietary ecology. A recent study presented the Ca isotope data of G. blacki and compared it with the data of other 19 taxa, including fauna from the Liucheng Gigantopithecus Cave, extant primates, and fossil hominins (Hu et al., 2022). Results revealed that G. blacki, P. boisei, and the giant panda (*Ailuropoda* sp.) had the highest  $\delta^{44/42}$ Ca values among the studied taxa. Hu et al. (2022) hypothesized that foraging on mineral licks could be one of the main factors contributing to the high  $\delta^{44/42}$ Ca values in the enamel of G. blacki, P. boisei, and the giant panda, which might reflect physiological demands to adapt to feeding on hard foods. While the thick enamel of P. boisei was initially interpreted as evidence for durophagy (e.g. Rak, 1983), more recent analyses of dental chipping and microwear patterns of P. boisei teeth reveal no evidence of regular hard food consumption (Ungar et al., 2008; Constantino and Konow, 2021; Sponheimer et al., in press). Ailuropoda feeds on bamboo, which is a tough food to process (even if some parts of the stem can be hard, they are eaten less often than leaves; Yamashita et al., 2009; King, 2014). Given the paleodietary inferences of *P. boisei* and the tough diet of the giant panda, the high  $\delta^{44/42}$ Ca values could also reflect feeding on tough foods. Whether G. blacki fed on hard foods (regularly or occasionally), tough foods, or both, is still an open question. Further studies of the incidence of dental chipping and microwear analyses in G. blacki teeth could provide new insights into this issue.

# **Declaration of competing interest**

The authors declare no conflict of interest.

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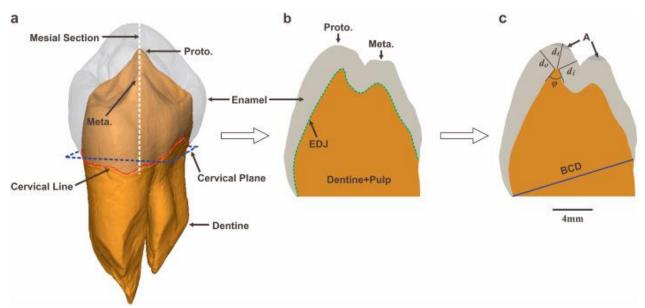
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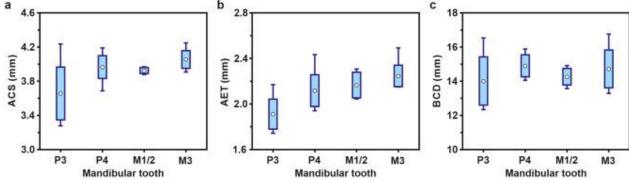
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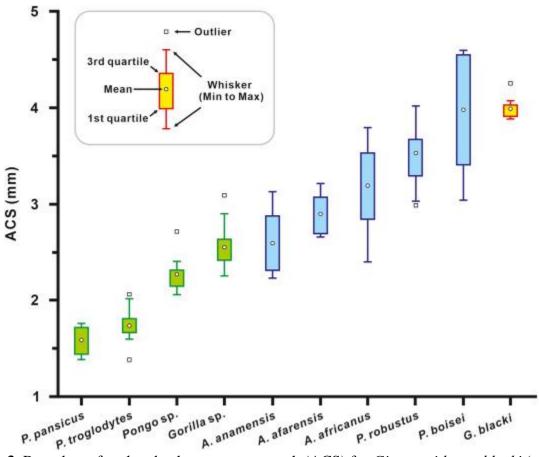
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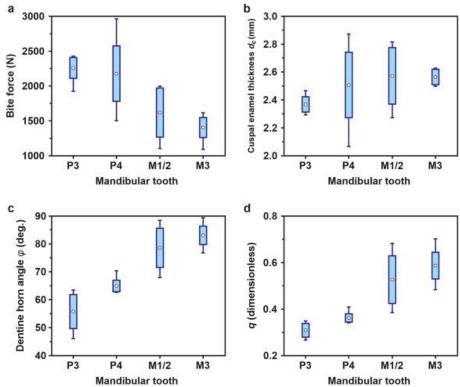
**Figure 1**. a) Virtual three-dimensional reconstruction of a *Gigantopithecus blacki* third premolar (Z008545). The enamel is rendered in semitransparency, showing the underlying enamel-dentine junction surface. The white dashed line indicates mesial section position. b) Virtual mesial section is produced following the general protocol (Benazzi et al., 2014; Tafforeau, 2004). c) Illustration of restoring missing enamel (dark zone pointed by arrows from A), measuring bicervical diameter (BCD), dentine horn angle ( $\varphi$ ), and cuspal enamel thickness parameters ( $d_i$ ,  $d_o$ , and  $d_i$ ). Abbreviations: Proto. = protoconid; Meta. = metaconid.



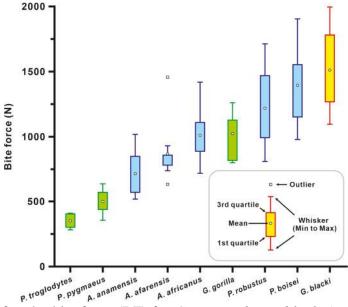
**Figure 2**. Box plots of a) absolute crown strength (ACS), b) average enamel thickness (AET), and c) bi-cervical diameter (BCD) for *Gigantopithecus* mandibular premolars and molars. The premolars (especially P<sub>4</sub>) have ACS that are comparable to the molars.



**Figure 3**. Box plots of molar absolute crown strength (ACS) for *Gigantopithecus blacki* (yellow box), australopiths (blue boxes), and extant great apes (green boxes). Except for *G. blacki*, the ACS results for the other taxa were extracted from Schwartz et al. (2020). *Gigantopithecus blacki* has significantly higher ACS than most taxa (see Table 4).



**Figure 4**. Box plots of a) bite force, b) cuspal enamel thickness, c) dentine horn angle, and d) configuration parameter q for *Gigantopithecus* mandibular premolars and molars. The premolars have larger BF than the molars do.



**Figure 5.** Box plots of molar bite force (BF) for *Gigantopithecus blacki* (yellow box), australopiths (blue boxes), and extant great apes (green boxes). Except for *G. blacki*, the BF data for the other taxa were extracted from Chai (2020). *Gigantopithecus blacki* has significantly larger BF than most taxa (see Table 6).

Table 1 Sample of Gigantopithecus blacki teeth studied in this work.

Specimen	Tooth	Wear <sup>a</sup>	Voxel size (µm)	Provenance <sup>b</sup>	Age (Ma)	References		
Z008545	LP <sub>3</sub>	2	25.90	CF		Wang (2009); Shao et al. (2014)		
Z008547	$LP_3$	2	23.50	CF		Wang (2009); Shao et al. (2014)		
Z008549	$LP_3$	2	25.90	CF		Wang (2009); Shao et al. (2014)		
Z008551	$LP_3$	2	25.90	CF		Wang (2009); Shao et al. (2014)		
Z008553	$LP_4$	2	25.90	CF	1.07 - 0.10 1.20 - 0.17	Wang (2009); Shao et al. (2014)		
Z008562	$RP_4$	1	$1.97 \pm 0.19 - 1.38 \pm 0.17$	Wang (2009); Shao et al. (2014)				
Z008558	$LP_4$	1	25.90	CF	(Early Pleistocene)	Wang (2009); Shao et al. (2014)		
Z008563	$RP_4$	2	25.90	CF		Wang (2009); Shao et al. (2014)		
Z008564	$LM_{1/2}$	2	25.09	CF		Wang (2009); Shao et al. (2014)		
Z008566	$LM_{1/2}$	2	25.09	CF		Wang (2009); Shao et al. (2014)		
Z008572	$LM_{1/2}$	2	25.90	CF		Wang (2009); Shao et al. (2014)		
Z010670	$LM_{1/2}$	2	20.39	МН	$1.69 \pm 0.22 - 1.29 \pm 0.11$	Wang et al. (2005); Shao et al. (2015)		
Z010071	$LM_3$	2	20.39	МН	(Early Pleistocene)	Wang et al. (2005); Shao et al. (2015)		
Z010654	$LM_3$	2	23.50	CF	$1.97 \pm 0.19 - 1.38 \pm 0.17$	Wang (2009); Shao et al. (2014)		
Z008581	$LM_3$	2	23.50	CF	(Early Pleistocene)	Wang (2009); Shao et al. (2014)		
Z008583	$RM_3$	2	23.50	CF	(Larry 1 leistocelle)	Wang (2009); Shao et al. (2014)		

Abbreviations: L = left; R = right; CF = Chuifeng Cave; MF = Mohui Cave.

<sup>a</sup> Occlusal wear stage was assessed in line with Molnar (1971).

<sup>b</sup> Both Chuifeng and Mohui are located in Tiandong, Guangxi province, China.

**Table 2**Extant ape and australopith comparative data for molar absolute crown strength (ACS) and molar bite force (BF) included in this study<sup>a</sup>.

Taxon	Sample size	Estimated ACS/BF (Mean ± standard deviation)	Reference		
	ACS (m	m) comparisons			
Pan paniscus	8	$1.59 \pm 0.15$			
Pan troglodytes	16	$1.74 \pm 0.16$			
Pongo sp.	9	$2.28 \pm 0.20$			
Gorilla sp.	13	$2.56 \pm 0.23$			
Australopithecus anamensis	13	$2.60\pm0.29$	Schwartz et al. (2020)		
Australopithecus afarensis	8	$2.90\pm0.21$	(===)		
Australopithecus africanus	31	$3.20\pm0.37$			
Paranthropus robustus	24	$3.53\pm0.27$			
Paranthropus boisei	7	$3.98 \pm 0.58$			
	BF (N	) comparisons			
Pan troglodytes <sup>b</sup>	4	$354 \pm 53$			
Pongo pygmaeus	9	$500 \pm 83$			
Gorilla gorilla	5	$1023 \pm 184$			
Australopithecus anamensis	8	$715\pm182$	Chai (2020)		
Australopithecus afarensis	9	$865 \pm 222$	Chai (2020)		
Australopithecus africanus	15	$1017 \pm 197$			
Paranthropus robustus	17	$1220\pm284$			
Paranthropus boisei	9	$1396 \pm 307$			

<sup>&</sup>lt;sup>a</sup> Measurement details of ACS and BF are provided in Schwartz et al. (2020) and Chai (2020), respectively.

Table 3

<sup>&</sup>lt;sup>b</sup> Updated results (Chai, pers. commun., 2022).

Results of enamel area, enamel-dentine junction (EDJ) length, average enamel thickness (AET), and bi-cervical diameter (BCD) used to compute the absolute crown strength (ACS) index for

Gigantopithecus blacki lower premolars and molars.

Specimen	Tooth	Enamel area (mm²)	EDJ length (mm)	AET (mm)	BCD (mm)	ACS (mm)	Mean ACS (mm)
Z008545		52.156	28.702	1.817	12.847	3.416	
Z008547	D.	51.846	29.739	1.743	12.338	3.279	2 657
Z008549	P <sub>3</sub>	76.770	35.385	2.170	16.524	4.234	3.657
Z008551		60.604	31.639	1.915	14.288	3.699	
Z008553		77.133	38.178	2.020	15.878	4.005	
Z008562	$\mathbf{P}_4$	60.164	31.062	1.937	14.050	3.689	2.064
Z008558	<b>P</b> 4	83.262	34.230	2.432	14.441	4.191	3.964
Z008563		63.444	30.542	2.077	15.200	3.973	
Z008564		67.566	29.284	2.307	13.556	3.954	
Z008566	$M_{1/2}$	62.280	27.699	2.248	13.987	3.965	3.925
Z008572	IVI 1/2	53.073	25.718	2.064	14.567	3.877	3.923
Z010670		60.983	29.843	2.043	14.908	3.903	
Z010071		56.448	26.259	2.150	14.878	3.999	
Z010654	M	61.537	24.707	2.491	13.285	4.067	4 055
Z008581	<b>M</b> 3	64.322	29.855	2.154	16.746	4.247	4.055
Z008583		58.985	26.960	2.188	13.946	3.906	

**Table 4**Results of pairwise comparisons for differences in molar absolute crown strength (ACS) among species. *P*-values are presented below the diagonal and directional differences are presented above the diagonal<sup>a, b</sup>.

Taxon	Pan pansicus	Pan troglodytes	Pongo	Gorilla	A. anamensis	A. afarensis	A. africanus	P. robustus	P. boisei	G. blacki
Pan pansicus		Ppa < Ptr	Ppa < Pon	Ppa < Gor	Ppa < Aan	Ppa < Aafa	Ppa < Aafr	Ppa < Pro	Ppa < Pbo	Ppa < Gbl
Pan troglodytes	0.037		Ptr < Pon	Ptr < Gor	Ptr < Aan	Ptr < Aafa	Ptr < Aafr	Ptr < Pro	Ptr < Pbo	Ptr < Gbl
Pongo	<0.001	<0.001		Pon < Gor	Pon < Aan	Pon < Aafa	Pon < Aafr	Pon < Pro	Pon < Pbo	Pon < Gbl
Gorilla	<0.001	<0.001	0.002		NSD	Gor < Aafa	Gor < Aafr	Gor < Pro	Gor < Pbo	Gor < Gbl
A. anamensis	<0.001	<0.001	0.004	0.381		Aan < Aafa	Aan < Aafr	Aan < Pro	Aan < Pbo	Aan < Gbl
A. afarensis	<0.001	<0.001	<0.001	0.001	0.014		Aafa < Aafr	Aafa < Pro	Aafa < Pbo	Aafa < Gbl
A. africanus	<0.001	<0.001	<0.001	<0.001	<0.001	0.013		Aafr < Pro	Aafr < Pbo	Aafr < Gbl
P. robustus	< 0.001	<0.001	<0.001	< 0.001	< 0.001	< 0.001	0.001		Pro < Pbo	Pro < Gbl
P. boisei	< 0.001	<0.001	<0.001	< 0.001	< 0.001	< 0.001	0.002	0.029		NSD
G. blacki	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	0.732	

<sup>&</sup>lt;sup>a</sup> Bold p-values indicate significant differences in molar ACS following the Benjamini-Hochberg adjustment. NSD = nonsignificant difference prior to the Benjamini-Hochberg adjustment.

<sup>&</sup>lt;sup>b</sup> Ppa = Pan pansicus; Ptr = Pan troglodytes; Pon = Pongo; Gor = Gorilla; Aan = Australopithecus anamensis; Aafa = Australopithecus afarensis; Aafr = Australopithecus africanus; Pro = Paranthropus robustus; Pbo = Paranthropus boisei; Gbl = Gigantopithecus blacki.

**Table 5** Results of cuspal enamel thickness (d<sub>c</sub>) and dentine horn angle ( $\varphi$ ) used to estimate bite force (BF) for *Gigantopithecus blacki* lower premolars and molars<sup>a</sup>.

Specimen	Tooth	d <sub>c</sub> (mm)	$\varphi$ (degree)	q	BF (N)	Mean BF (N)
Z008545		2.34	63.48	0.35	1925.52	
Z008547	<b>P</b> <sub>3</sub>	2.29	46.06	0.27	2427.25	2250 21
Z008549		2.47	60.19	0.33	2292.48	2258.31
Z008551		2.38	53.32	0.29	2388.01	
Z008553		2.07	63.69	0.35	1502.20	
Z008562	$P_4$	2.61	70.39	0.41	2059.13	2177.57
Z008558	Γ4	2.87	62.68	0.34	2960.74	21/7.37
Z008563		2.48	63.00	0.34	2188.20	
Z008564		2.73	75.05	0.46	1996.60	
Z008566	$M_{1/2}$	2.82	88.47	0.68	1430.04	1619.53
Z008572	1411/2	2.27	82.79	0.58	1104.49	1017.33
Z010670		2.47	67.98	0.38	1946.98	
Z010071		2.63	82.76	0.58	1478.15	
Z010654	$M_3$	2.61	83.38	0.59	1431.23	1405.20
Z008581	1 <b>V1</b> 3	2.52	76.76	0.48	1617.65	1403.20
Z008583		2.50	89.41	0.70	1093.76	

 $<sup>^{\</sup>rm a}$  q is a parameter that has a polynomial relationship with dentine horn angle.

**Table 6**Results of pairwise comparisons for differences in molar bite force (BF) among species. *P*-values are presented below the diagonal and directional differences are presented above the diagonal<sup>a, b</sup>.

Taxon	Pan troglodytes	Pongo pygmaeus	A. anamensis	A. afarensis	A. africanus	Gorilla gorilla	P. robustus	P. boisei	G. blacki
Pan troglodytes		Ptr < Ppy	Ptr < Aan	Ptr < Aafa	Ptr < Aafr	Ptr < Ggo	Ptr < Pro	Ptr < Pbo	Ptr < Gbl
Pongo pygmaeus	0.009		Ppy < Aan	Ppy < Aafa	Ppy < Aafr	Ppy < Ggo	Ppy < Pro	Ppy < Pbo	Ppy < Gbl
A. anamensis	0.003	0.003		NSD	Aan < Aafr	Aan < Ggo	Aan < Pro	Aan < Pbo	Aan < Gbl
A. afarensis	0.003	<0.001	0.057		Aafa < Aafr	NSD	Aafa < Pro	Aafa < Pbo	Aafa < Gbl
A. africanus	<0.001	<0.001	0.002	0.040		NSD	Aafr < Pro	Aafr < Pbo	Aafr < Gbl
Gorilla gorilla	0.012	0.001	0.013	0.073	0.359		NSD	Ggo < Pbo	Ggo < Gbl
P. robustus	<0.001	<0.001	<0.001	0.002	0.028	0.109		NSD	Ggo < Gbl
P. boisei	0.003	<0.001	< 0.001	0.002	0.002	0.038	0.132		NSD
G. blacki	0.003	<0.001	<0.001	0.001	<0.001	0.030	0.050	0.212	

<sup>&</sup>lt;sup>a</sup> Bold p-values indicate significant differences in molar BF following the Benjamini-Hochberg adjustment. NSD = nonsignificant difference prior to the Benjamini-Hochberg adjustment.

<sup>&</sup>lt;sup>b</sup>Ptr = Pan troglodytes; Pon = Pongo; Gor = Gorilla; Aan = Australopithecus anamensis; Aafa = Australopithecus afarensis; Aafr = Australopithecus africanus; Pro = Paranthropus robustus; Pbo = Paranthropus boisei; Gbl = Gigantopithecus blacki.