

The unexpected role of the STING protein in lipid metabolism

Isabelle K Vila, Nadine Laguette

▶ To cite this version:

Isabelle K Vila, Nadine Laguette. The unexpected role of the STING protein in lipid metabolism. Comptes Rendus. Biologies, 2023, 346, pp.29 - 33. 10.5802/crbiol.110 . hal-04241169

HAL Id: hal-04241169 https://hal.science/hal-04241169v1

Submitted on 17 Oct 2023

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 The unexpected role of the STING protein in lipid metabolism.

2 Isabelle K Vila^{1*} & Nadine Laguette^{1*}

3 ¹ IGH, Université de Montpellier, CNRS, Montpellier, France

4 * Correspondence to: isabelle.vila@igh.cnrs.fr and nadine.laguette@igh.cnrs.fr

5

6 1. STING, a central protein of the immune response associated with cytosolic nucleic

7 acids.

8 The innate immune system is the first line of defence against pathogens. One of the central 9 pathways for pathogen detection is the recognition of their nucleic acid content by intracellular 10 receptors. A large panel of receptors involved in the initiation of the innate immune response has been 11 described [1]. Their interaction with pathological nucleic acids leads to the activation of signalling 12 pathways that induce the production of inflammatory cytokines. This inflammatory response mobilises 13 effector cells of the adaptive immune system and is therefore a crucial step in the initiation, 14 maintenance and clatexoordination of immune responses.

15 Recently, a central pathway for the detection of cytosolic nucleic acids has been described 16 (Figure 1). This pathway was originally identified in immune cells as activated by cytosolic double-17 stranded DNAs during viral or bacterial infections [2]. This pathway relies on the cyclic GMP-AMP 18 synthase (cGAS) receptor and the Stimulator of Interferon Genes (STING) adaptor protein. The cGAS 19 receptor recognises cytosolic nucleic acids, including double-stranded DNA (dsDNA), single-stranded 20 DNA (ssDNA) or DNA:RNA hybrids [3–5].Binding of cGAS to cytosolic nucleic acids induces its activation 21 and the synthesis of 2'3'-cGAMP, a cyclic dinucleotide [6]. This secondary messenger interacts with the 22 adaptor protein STING, promoting the assembly of a signalosome in which STING serves as an anchoring site for the various partners required for the induction of the inflammatory response. Those partners 23 24 notably include the protein TANK binding kinase 1 (TBK1) and the interferon regulatory factor 3 (IRF3) 25 and/or nuclear factor KB (NF-KB) transcript factors [7,8]. Activation of the cGAS-STING signalling axis is characterised by the production of type I interferons, besides other pro-inflammatory cytokines and
 chemokines. Thus, this signalling pathway is particularly critical in the response to pathogens, as the
 production of type I interferons leads to the establishment of an antiviral state through induction of the
 expression of interferon response genes.

Recent work suggests that cGAS and STING proteins, beyond their canonical function in innate
 immunity, are also involved in various cellular processes such as the induction and maintenance of
 senescence or the regulation of cell proliferation [9–11].

33 2. STING in fatty acid metabolism: an unexpected role.

34 In 2022, work performed by the team of Dr Nadine Laguette in Montpellier, revealed a central role 35 of STING in metabolic homeostasis [12](figure 2). Using a mouse model deleted for the STING protein 36 [13], this study showed that STING inhibits the fatty acid desaturase 2 (FADS2) enzyme essential for the 37 desaturation of dietary polyunsaturated fatty acids (PUFAs). Therefore, ablation or agonist-induced 38 degradation of STING led to increased FADS2 activity leading to the accumulation of PUFA derivatives 39 and driving increased thermogenic gene activation, which results in browning of white fat deposits in 40 vivo. PUFAs were also shown to bind STING and inhibit its activity. Modulation of PUFA production thus 41 regulates STING-dependent antiviral responses while contributing to the resolution of STING-associated 42 inflammation. Finally, this study also revealed that STING agonists can bind and modulate the enzymatic activity of FADS2. Thus, this work demonstrates the existence of a negative regulatory loop between 43 44 STING and FADS2, opening many questions pertaining to their respective roles in human pathologies 45 presenting with a chronic STING-associated inflammation (Vila, Guha, et al., 2022).

Interestingly, alleles of STING not competent for the induction of interferon responses have been
documented in human populations. These include the STING HAQ allele that bears 3 amino acid
substitutions: R71H-I229A-R292Q [15]. Analysis of STING haplotypes revealed a high prevalence of the
HAQ allele in East Asian populations, compared to sub-Saharan African populations while sub-Saharan
African populations show a high prevalence of an AQ allele (G230A, R293Q) which is competent for the

51 recognition of cyclic dinucleotides and the induction of interferon responses [16]. The authors propose 52 that these STING alleles (HAQ and AQ) underwent natural selection during migration out of Africa 53 50,000~70,000 years ago. Using mouse models expressing the HAQ or AQ human STING alleles, they 54 showed that AQ mice store less lipid than HAQ mice, while displaying improved fatty acid oxidation. 55 Furthermore, the visceral white adipose tissue of HAQ mice was infiltrated by regulatory T cells and anti-56 inflammatory macrophages, unlike that of AQ mice. Thus, the HAQ haplotype promotes fat storage, 57 while the AQ haplotype promotes immune tolerance. The study by Vila et al showed that the HAQ allele 58 of STING strongly interacts with FASD2 [12], which is consistent with the description of a prominent role 59 of this allele in fatty acid metabolism [16]. Thus, the study of the evolution of STING haplotypes suggests 60 that the function of STING in modulating fatty acid metabolism may have been critical during human migration out of Africa. 61

62 In support of this hypothesis, work in Drosophila [17] confirmed the conservation of the functional 63 link between STING and fatty acid metabolism. The authors showed that the Drosophila STING 64 orthologue interacts with enzymes involved in the biosynthesis of long-chain fatty acids such as acetyl-65 CoA carboxylase (ACC) and fatty acid synthetase (FASN) within a multiprotein complex. Furthermore, 66 absence of STING in Drosophila led to decreased lipid storage and decreased expression of genes related 67 to lipid metabolism. Thus, absence of STING increased sensitivity to food deprivation and oxidative 68 stress in Drosophila. These results showing profound lipid metabolism alteration in Drosophila support 69 a conserved role for STING in the regulation of lipid metabolism.

70

3. Regulation of lipid metabolism by STING: a link to innate immune regulation?

Although the role of STING in host defence mechanisms against pathogens is well established, these recent findings on the link between STING and fatty acid metabolism demonstrate that this protein has important functions beyond the detection of cytosolic nucleic acids. Evolutionary analysis of STING suggests that the modulation of lipid metabolism is ancient, conserved and may be the primordial function of STING. Metabolic pathways that interact with STING, such as those leading to the production of PUFAs, are also involved in the control of STING-independent inflammatory responses. Indeed, while PUFAs can directly inhibit STING [12], they can also be processed into lipid mediators (oxylipins) that possess, among others, immunomodulatory properties [18]. Thus, it can be hypothesised that STING alleles expressed in organisms lacking an interferon signalling pathway could participate to the control of inflammatory responses by regulating the generation of these lipid metabolites.

Altogether, these recent studies [12,16,17] highlight a tight link between innate immunity and lipid metabolism, paving the way for research into the role of metabolic alterations in human pathologies associated with aberrant STING activation [19]. Those notably include cancer, where a crucial role of a functional cGAS-STING axis has been recently shown to be essential for driving tumour immunogenicity [20,21].The role of STING-dependent metabolic regulation remains to be investigated in this context.

87 Conflicts of interest: Authors have no conflict of interest to declare.

88





Figure 1: Overview of the cytosolic DNA sensing via the cGAS-STING pathway. dsDNA presence in the 91 92 cytosol can result from pathogen infection or self-DNA after cellular stress. GAS binds dsDNA leading to cGAS activation and synthesis of 2,3'-cGAMP from ATP and GTP. cGAMP binds to ER-resident STING, this 93 94 binding will trigger STING translocation to the Golgi apparatus. In the Golgi, STING binds to and activates 95 TBK1 (by autophosphorylation) as well as the transcription factor NF-KB via IKK. TBK1 then phosphorylates IRF3 which undergoes dimerization and translocates to the nucleus to induce 96 transcription of type I IFNs and several inflammatory chemokines. NF-kB induces transcription of 97 98 inflammatory cytokines.



99

100 Figure 2: STING plays a role in lipid metabolism. Convergent data in 2022 showed that STING regulates 101 lipid metabolism. In Drosophila, STING form a multi-protein complex with two enzymes involves in long-102 chain fatty acid synthesis FASN and ACC. In mouse STING regulates the activity of the lipid desaturase 103 FADS2 modulating the synthesis of long-chain polyunsaturated fatty acid. The production of those lipid 104 species will in turn modulate STING activity and narrow its activation in presence of its agonist cGAMP. 105 In human populations several alleles exist and were already known to be more or less potent in the 106 induction of IFN response. The study of AQ and HAQ alleles showed that they also differ in lipid 107 metabolism modulation as HAQ promotes more fat storage than AQ.

108

109

- 110 [1] O. Takeuchi, S. Akira, Pattern recognition receptors and inflammation., Cell. 140 (2010) 805–
 111 20. https://doi.org/10.1016/j.cell.2010.01.022.
- 112 [2] J. Tao, X. Zhou, Z. Jiang, cGAS-cGAMP-STING: The three musketeers of cytosolic DNA sensing
- and signaling., IUBMB Life. 68 (2016) 858–870. https://doi.org/10.1002/iub.1566.
- 114 [3] A.K. Mankan, T. Schmidt, D. Chauhan, M. Goldeck, K. Höning, M. Gaidt, A. v Kubarenko, L.
- 115 Andreeva, K.-P. Hopfner, V. Hornung, Cytosolic RNA:DNA hybrids activate the cGAS-STING axis.,
- 116 EMBO J. 33 (2014) 2937–46. https://doi.org/10.15252/embj.201488726.
- 117 [4] P.J. Kranzusch, A.S.-Y. Lee, J.M. Berger, J.A. Doudna, Structure of human cGAS reveals a
- 118 conserved family of second-messenger enzymes in innate immunity., Cell Rep. 3 (2013) 1362–
- **119** 8. https://doi.org/10.1016/j.celrep.2013.05.008.
- 120 [5] L. Sun, J. Wu, F. Du, X. Chen, Z.J. Chen, Cyclic GMP-AMP synthase is a cytosolic DNA sensor that
- activates the type I interferon pathway., Science. 339 (2013) 786–91.
- 122 https://doi.org/10.1126/science.1232458.
- 123 [6] A. Ablasser, M. Goldeck, T. Cavlar, T. Deimling, G. Witte, I. Röhl, K.-P. Hopfner, J. Ludwig, V.
- Hornung, cGAS produces a 2'-5'-linked cyclic dinucleotide second messenger that activates

125 STING., Nature. 498 (2013) 380–4. https://doi.org/10.1038/nature12306.

- 126 [7] S. Liu, X. Cai, J. Wu, Q. Cong, X. Chen, T. Li, F. Du, J. Ren, Y.-T. Wu, N. v Grishin, Z.J. Chen,
- Phosphorylation of innate immune adaptor proteins MAVS, STING, and TRIF induces IRF3
 activation., Science. 347 (2015) aaa2630. https://doi.org/10.1126/science.aaa2630.
- 129 [8] T. Abe, G.N. Barber, Cytosolic-DNA-mediated, STING-dependent proinflammatory gene
- induction necessitates canonical NF-κB activation through TBK1., J Virol. 88 (2014) 5328–41.
- 131 https://doi.org/10.1128/JVI.00037-14.

- 132 [9] H. Jiang, X. Xue, S. Panda, A. Kawale, R.M. Hooy, F. Liang, J. Sohn, P. Sung, N.O. Gekara,
- 133 Chromatin-bound cGAS is an inhibitor of DNA repair and hence accelerates genome
- destabilization and cell death., EMBO J. 38 (2019) e102718.
- 135 https://doi.org/10.15252/embj.2019102718.
- 136 [10] S. Glück, B. Guey, M.F. Gulen, K. Wolter, T.-W. Kang, N.A. Schmacke, A. Bridgeman, J.
- **137** Rehwinkel, L. Zender, A. Ablasser, Innate immune sensing of cytosolic chromatin fragments
- through cGAS promotes senescence., Nat Cell Biol. 19 (2017) 1061–1070.
- 139 https://doi.org/10.1038/ncb3586.
- 140 [11] S. Cerboni, N. Jeremiah, M. Gentili, U. Gehrmann, C. Conrad, M.-C. Stolzenberg, C. Picard, B.
- 141 Neven, A. Fischer, S. Amigorena, F. Rieux-Laucat, N. Manel, Intrinsic antiproliferative activity of
- the innate sensor STING in T lymphocytes., J Exp Med. 214 (2017) 1769–1785.
- 143 https://doi.org/10.1084/jem.20161674.
- 144 [12] I.K. Vila, H. Chamma, A. Steer, M. Saccas, C. Taffoni, E. Turtoi, L.S. Reinert, S. Hussain, J.
- 145 Marines, L. Jin, X. Bonnefont, M. Hubert, O. Schwartz, S.R. Paludan, G. van Simaeys, G.
- 146 Doumont, B. Sobhian, D. Vlachakis, A. Turtoi, N. Laguette, STING orchestrates the crosstalk
- 147 between polyunsaturated fatty acid metabolism and inflammatory responses., Cell Metab. 34
- 148 (2022) 125-139.e8. https://doi.org/10.1016/j.cmet.2021.12.007.
- 149 [13] L. Jin, A. Getahun, H.M. Knowles, J. Mogan, L.J. Akerlund, T.A. Packard, A.-L. Perraud, J.C.
- 150 Cambier, STING/MPYS mediates host defense against Listeria monocytogenes infection by
- regulating Ly6C(hi) monocyte migration., J Immunol. 190 (2013) 2835–43.
- 152 https://doi.org/10.4049/jimmunol.1201788.
- **153** [14] I.K. Vila, S. Guha, J. Kalucka, D. Olagnier, N. Laguette, Alternative pathways driven by STING:
- 154 From innate immunity to lipid metabolism., Cytokine Growth Factor Rev. (2022).
- 155 https://doi.org/10.1016/j.cytogfr.2022.08.006.

- 156 [15] L. Jin, L.-G. Xu, I. v Yang, E.J. Davidson, D.A. Schwartz, M.M. Wurfel, J.C. Cambier, Identification
 157 and characterization of a loss-of-function human MPYS variant., Genes Immun. 12 (2011) 263–
 158 9. https://doi.org/10.1038/gene.2010.75.
- 159 [16] S. Mansouri, H. Gogoi, S. Patel, D.S. Katikaneni, A. Singh, A. Aybar-Torres, G. de Lartigue, L. Jin,
- 160 MPYS Modulates Fatty Acid Metabolism and Immune Tolerance at Homeostasis Independent
- 161 of Type I IFNs., J Immunol. 209 (2022) 2114–2132. https://doi.org/10.4049/jimmunol.2200158.
- 162 [17] K. Akhmetova, M. Balasov, I. Chesnokov, Drosophila STING protein has a role in lipid
 163 metabolism., Elife. 10 (2021). https://doi.org/10.7554/eLife.67358.
- 164 [18] M. Gabbs, S. Leng, J.G. Devassy, M. Monirujjaman, H.M. Aukema, Advances in Our
- 165 Understanding of Oxylipins Derived from Dietary PUFAs., Adv Nutr. 6 (2015) 513–40.

166 https://doi.org/10.3945/an.114.007732.

- 167 [19] I.K. Vila, M. Fretaud, D. Vlachakis, N. Laguette, C. Langevin, Animal Models for the Study of
 168 Nucleic Acid Immunity: Novel Tools and New Perspectives., J Mol Biol. 432 (2020) 5529–5543.
 169 https://doi.org/10.1016/j.jmb.2020.08.016.
- 170 [20] C. Taffoni, J. Marines, H. Chamma, M. Saccas, A. Bouzid, S. Guha, A.-L.C. Valadao, K. Polak, M.
- del Rio, C. Gongora, D. Pineau, J.-P. Hugnot, K. Kissa, L. Fontenille, F.P. Blanchet, I.K. Vila, N.
- 172 Laguette, The crosstalk between DNA-PK and cGAS drives tumor immunogenicity, BioRxiv.

173 (2022) 2022.06.08.495278. https://doi.org/10.1101/2022.06.08.495278.

- 174 [21] C. Hong, M. Schubert, A.E. Tijhuis, M. Requesens, M. Roorda, A. van den Brink, L.A. Ruiz, P.L.
- 175 Bakker, T. van der Sluis, W. Pieters, M. Chen, R. Wardenaar, B. van der Vegt, D.C.J. Spierings, M.
- de Bruyn, M.A.T.M. van Vugt, F. Foijer, cGAS-STING drives the IL-6-dependent survival of
- 177 chromosomally instable cancers., Nature. 607 (2022) 366–373.
- 178 https://doi.org/10.1038/s41586-022-04847-2.

179