

Diagnostic and the rapeutic approach to adult central nervous system vasculitis

Ahmad Nehme, Marion Boulanger, Achille Aouba, Christian Pagnoux,

Mathieu Zuber, Emmanuel Touzé, Hubert de Boysson

► To cite this version:

Ahmad Nehme, Marion Boulanger, Achille Aouba, Christian Pagnoux, Mathieu Zuber, et al.. Diagnostic and the rapeutic approach to adult central nervous system vasculitis. Revue Neurologique, 2022, 178 (10), pp.1041-1054. 10.1016/j.neurol.2022.05.003 . hal-04240101

HAL Id: hal-04240101 https://hal.science/hal-04240101v1

Submitted on 13 Oct 2023

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial - NoDerivatives 4.0 International License

Diagnostic and therapeutic approach to adult central nervous system vasculitis

Ahmad Nehme MD^{1,2,3}, Marion Boulanger MD PhD^{1,2,3}, Achille Aouba MD PhD^{1,4}, Christian Pagnoux MD MPH⁵, Mathieu Zuber MD PhD⁶, Emmanuel Touzé, MD PhD^{1,2,3}, Hubert de Boysson MD PhD^{1,4}

¹Normandie University,
²Department of Neurology, Caen University Hospital, Caen, France
³INSERM UMR-S U1237 PhIND/BB@C
⁴Department of Internal Medicine, Caen University Hospital, Caen, France
⁵Division of Rheumatology, Mount Sinai Hospital, University of Toronto, Toronto, Ontario, Canada.
⁶Department of Neurology, Saint-Joseph Hospital, Paris, France

Characters in title: 78 Abstract count: 252 Total word count: 4359 References: 105 Tables: 4 Figures: 3

Keywords: Central nervous system vasculitis, Primary angiitis of the central nervous system, Vasculitis

Address correspondence to :

Ahmad Nehme, Department of Neurology, Centre Hospitalier Universitaire de Caen-Normandie, Avenue de la Côte de Nacre, 14000 Caen, France E-mail: <u>nehme-a@chu-caen.fr</u> Phone: 00 33 2 31 06 31 86 Fax: 00 33 2 31 06 51 16

ABSTRACT

The clinical manifestations of central nervous system (CNS) vasculitis are highly variable. In the absence of a positive CNS biopsy, CNS vasculitis is particularly suspected when markers of both vascular disease and inflammation are present. To facilitate the clinical and therapeutic approach to this rare condition, CNS vasculitis can be classified according to the size of the involved vessels. Vascular imaging is used to identify medium vessel disease. Small vessel disease can only be diagnosed with a CNS biopsy. Medium vessel vasculitis usually presents with focal neurological signs, while small vessel vasculitis more often leads to cognitive deficits, altered level of consciousness and seizures. Markers of CNS inflammation include cerebrospinal fluid pleocytosis or elevated protein levels, and vessel wall, parenchymal or leptomeningeal enhancement. The broad range of differential diagnoses of CNS vasculitis can be narrowed based on the disease subtype. Common mimickers of medium vessel vasculitis include intracranial atherosclerosis and reversible cerebral vasoconstriction syndrome. The diagnostic workup aims to answer two questions: is the neurological presentation secondary to a vasculitic process, and if so, is the vasculitis primary (i.e., primary angiitis of the CNS) or secondary (e.g., to a systemic vasculitis, connective tissue disorder, infection, malignancy or drug use)? In primary angiitis of the CNS, glucocorticoids and cyclophosphamide are most often used for induction therapy, but rituximab may be an alternative. Based on the available evidence, all patients should receive maintenance immunosuppression. A multidisciplinary approach is necessary to ensure an accurate and timely diagnosis and to improve outcomes for patients with this potentially devastating condition.

Abbreviations : ABRA – amyloid-beta related angiitis, CAA – cerebral amyloid angiopathy, CSF – cerebrospinal fluid, CNS – central nervous system, ICAD – intracranial atherosclerotic disease, MVV – medium vessel vasculitis, PACNS – primary angiitis of the central nervous system, RCVS –

reversible cerebral vasoconstriction syndrome, SVV – small vessel vasculitis, VZV – Varicella Zoster virus. **Introduction**

The diagnosis and treatment of primary angiitis of the central nervous system (PACNS) are challenging. Most clinicians only see a few patients with PACNS during their career, and no randomized controlled studies or consensual recommendations exist to guide clinical management. Our knowledge of the disease is based on single¹⁻⁵ and multi-center cohorts⁶ of limited size. Multiple conditions can mimic central nervous system (CNS) vasculitis. ⁷⁻⁹ Even after an appropriate workup, in the absence of a positive CNS biopsy, the diagnosis of PACNS can remain uncertain, with various treatment strategies reported. ¹⁰⁻¹²

The existence of PACNS subtypes contributes to the polymorphous clinical and radiological presentations of the disease. ¹³ Multiple cohorts refer to large, medium, and small vessel PACNS, but definitions vary across publications. ^{5, 14, 15} For intracranial arteries, we consider that medium vessel disease is diagnosed on vascular imaging, while small vessel disease can only be identified with a CNS biopsy. Based on this classification, the reported proportions of PACNS subtypes is isolated medium vessel vasculitis (MVV) in 45-69% of patients, isolated small vessel vasculitis (SVV) in 23-30%, and both MVV and SVV in 6-32%. ^{4, 16, 17} We use this classification to provide clinicians with an updated approach to the diagnosis and treatment of PACNS.

In this review, we aimed to: (1) describe the clinicoradiological manifestations and differential diagnoses of CNS vasculitis; (2) discuss the etiologies of CNS vasculitis, and (3) provide an up-to-date review on PACNS treatment.

Section 1: When should I suspect CNS vasculitis?

No single clinical manifestation is sensitive or specific for CNS vasculitis. MVV tends to present with focal neurological signs, while SVV is more likely to lead to cognitive deficits, altered level of consciousness, and seizures. ^{4, 16-18} Headaches are reported in 51% of patients with PACNS. ¹⁸ A history of unusual subacute or chronic headaches prior to stroke onset increases the likelihood of CNS vasculitis. The clinical course of CNS vasculitis is most often subacute, but catastrophic presentations with rapidly successive infarcts or chronic presentations with aseptic meningitis and recurrent infarcts are possible. ^{19, 20} In a patient with otherwise unexplained neurological findings, CNS vasculitis is particularly suspected when both vascular disease and markers of CNS inflammation are present.

CNS vascular disease

Although not specific for a vasculitic process, multifocal arterial stenoses – sometimes accompanied by post-stenotic dilation or beading – are the most common direct signs of arterial disease in MVV. ²¹ Rarely, MVV leads to arterial occlusion or development of aneurysms. ²² Steno-occlusive arterial lesions involve at least two cerebral arteries in 72% of patients with medium vessel PACNS. ²³ Computed tomography angiography (CTA) and magnetic resonance angiography (MRA) can directly visualize medium vessel disease, but digital subtraction angiography (DSA) has superior sensitivity for detecting distal or posterior arterial lesions. ²³⁻²⁵ The sensitivity of time-of-flight MRA for vasculitic arterial stenoses is higher at 3T than 1.5T. ²⁶ Importantly, normal vascular imaging does not rule out CNS vasculitis, as CTA, MRA and DSA do not image small vessels, which have a diameter

of less than 500 μ m. ²⁷ Direct visualization of small vessels is only possible with a CNS biopsy. (Fig. 1)

Cerebral vasculitis typically presents with both acute and chronic multi-territorial infarcts. In the French PACNS cohort, 75% of patients had acute infarcts and 92% had subacute or chronic infarcts on brain magnetic resonance imaging (MRI) at the time of diagnosis. ²³ Hemorrhagic manifestations are present on MRI in up to 64% of cases and may result from hemorrhagic transformation of ischemic lesions or rupture of inflamed vessels. ²³ When the disease is restricted to small vessels, brain MRI can reveal small punctate infarcts with normal vascular imaging. SVV can also lead to white matter hyperintensities and cerebral microbleeds. PACNS presents as a tumor-like lesion in 7-12% of patients. Pseudotumoral PACNS most often results from isolated SVV. ^{28, 29} Spinal cord involvement is documented in 5% of patients in the Mayo Clinic PACNS cohort and tends to occur simultaneously or several months after cerebral vasculitis, ³⁰

CNS inflammatory findings

Definite diagnosis of cerebral vasculitis requires the visualization of CNS vessel wall inflammation, which is only possible with a CNS biopsy and is mainly limited to small vessels. In patients with isolated medium vessel involvement and/or a negative biopsy, the combination of vascular disease and markers of CNS inflammation allows the clinician to infer the presence of CNS vasculitis.

In medium vessel disease, smooth and concentric enhancement of the vessel wall on MRI can result from vascular inflammation and can support the diagnosis of MVV. ^{31, 32} In the context of vasculitis, enhancement is thought to result from increased permeability of endothelial cells, with

secondary leakage of gadolinium into the vessel wall. ³² However, overreliance on vessel wall MRI findings can lead to overdiagnosis of CNS vasculitis. Non-vasculitic arteriopathies (e.g. atherosclerosis, dissection, vasospasm), partially recanalized thrombi and mechanical thrombectomy can lead to vessel wall enhancement. ³²⁻³⁴ Conversely, misidentification of vessel wall enhancement can result from incomplete blood flow suppression, crossing arteries, microhemorrhages, or adjacent venous, meningeal or parenchymal enhancement. ³³ The significance of vessel wall enhancement in the absence of arterial stenoses is unknown. The specificity of vessel wall enhancement has not been established in comparison with a pathological gold standard. ³¹ Leptomeningeal or non-ischemic parenchymal gadolinium enhancement suggest the presence of CNS inflammation and are more frequent in SVV (73%) than in MVV (36%). ¹⁶

Cerebrospinal fluid (CSF) pleocytosis and elevated protein levels can result from vasculitisrelated CNS inflammation. In a systematic review and meta-analysis of patients with PACNS, 48% had a CSF white blood cell count > 4/mm³ and 65% had a CSF protein level > 450 mg/l. ¹⁸ SVV is more likely to present with abnormal CSF findings than MVV. ^{14, 16} Mild CSF anomalies should be interpreted with caution, as ischemic stroke unrelated to vasculitis is associated with CSF pleocytosis in up to 13-18% of patients, with white blood cell counts usually ranging from 5 to 15/mm³. ^{35, 36} Based on our experience and reported cohorts, the CSF white blood cell count seldom exceeds 200/mm³ in PACNS. ^{1, 12, 17} In addition, minor elevations in CSF protein levels are not specific to CNS vasculitis, as they can occur with all strokes. ³⁷ Oligoclonal bands are detected in the CSF of 21-26% of patients with PACNS but can also occur in all etiologies of ischemic stroke. ^{17, 36, 38}

Section 2: What disorders can mimic CNS vasculitis?

The differential diagnosis of CNS vasculitis is broad but can be narrowed based on the clinical and radiological presentation.

Mimics of CNS MVV

Table 1 details a selection of the main non-vasculitic mimics of CNS MVV.

While reversible cerebral vasoconstriction syndrome (RCVS) is often compared to CNS vasculitis, the presence of thunderclap headaches in > 90% of patients with RCVS usually helps to differentiate the two disorders. ^{39, 40} Specific triggers (e.g. nasal decongestants, postpartum, sexual activity) are associated with RCVS. In CNS vasculitis, initial brain MRI is always abnormal, and patients more often present focal neurological signs and ischemic stroke. In RCVS, initial brain MRI is normal in > 50% of patients and can identify convexity subarachnoid hemorrhage or vasogenic edema, which are infrequent in CNS vasculitis. Arterial stenoses peak two weeks after the onset of RCVS symptoms and resolve within three months, while they usually persist in CNS vasculitis. ^{41, 42}

In our experience, intracranial atherosclerotic disease (ICAD) is commonly misdiagnosed as CNS vasculitis, particularly when it presents in young adults with recurrent strokes. The likelihood of ICAD increases with the number and severity of vascular risk factors, as well as with the presence of related complications (e.g. left ventricular hypertrophy, hypertensive/diabetic retinopathy or nephropathy). ICAD is more common in Asian, Black and Hispanic populations. ⁴³ Atherosclerotic stenoses tend to develop at arterial bifurcations and can be associated with calcifications. ⁴⁴ However, the absence of extracranial atherosclerosis or calcifications does not rule out ICAD. On vessel wall MRI, the presence of a lipid core, an intraplaque hemorrhage or negative remodeling can support the

diagnosis of ICAD. ³² Eccentric vessel wall enhancement is more common in ICAD but can also occur in MVV. ^{32, 45}

Mimics of CNS SVV

Table 2 details a selection of the main non-vasculitic mimics of CNS SVV.

The suspicion for a non-inflammatory mimic of CNS SVV increases with a normal lumbar puncture: small vessel PACNS is associated with elevated CSF white blood cells (> 4/mm³) or protein levels (> 450 mg/L) in at least 75% of cases. ¹⁸ Both vascular and non-vascular diseases can mimic CNS SVV. Sudden-onset neurological deficits, small infarcts and microbleeds suggest a vascular disease. Etiologies include severe forms of sporadic small vessel disease (e.g. deep perforator arteriopathy), hereditary pathologies (e.g. cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy [CADASIL]), drugs (e.g. cocaine, amphetamines), prothrombotic states (e.g. antiphospholipid syndrome, malignancies, disseminated intravascular coagulation), and neoplastic vasculopathies (e.g. intravascular lymphoma).

Non-vascular inflammatory (e.g. autoimmune encephalitis) or metabolic (e.g. mitochondrial encephalopathy with lactic acidosis and stroke) diseases can present with headaches, rapidly evolving neurological symptoms and white matter lesions, mimicking CNS SVV. Susac syndrome leads to ischemic strokes and is associated with markers of CNS inflammation. ⁴⁶ However, Susac syndrome is not classified as a CNS vasculitis, as its clinical manifestations are thought to result from immune-mediated endothelial cell injury rather than vessel wall inflammation. ⁴⁷

Section 3: What are the etiologies of secondary CNS vasculitis?

Table 3 lists the etiologies of secondary CNS vasculitis, which can result from infectious, autoimmune, auto-inflammatory, paraneoplastic, genetic and drug-induced disorders.

Infections can lead to CNS vasculitis as a result of septic emboli (e.g. infectious endocarditis), contiguous spread from the subarachnoid space (e.g. bacterial meningitis), direct invasion of endothelial cells (e.g. varicella Zoster virus [VZV]), or an immune response to the infectious agent (e.g. hepatitis B-induced polyarteritis nodosa). ⁴⁸ Infectious endocarditis can present with intracranial arterial stenoses and markers of inflammation in the CSF. ⁴⁹ Two-thirds of VZV vasculitis cases occur after an episode of shingles, and one-third are isolated to small vessels. ^{50, 51} An elevated CSF to serum ratio of anti-VZV IgG antibodies can suggest the diagnosis in patients with a recent history of shingles and a negative CSF VZV PCR. ⁵² (Fig. 2) Vasculitis frequently occurs in patients with tuberculous meningitis and is often associated with hydrocephalus and low CSF glucose. ⁵³ Aspergillosis and mucormycosis mainly develop in immunocompromised patients and can lead to hemorrhagic CNS vasculitis. ⁴⁸ One biopsy-proven case of pediatric pseudotumoral CNS vasculitis has been reported after COVID-19. ⁵⁴

CNS involvement can occur in primary vasculitides that affect large, medium and small vessels. Giant cell arteritis is associated with an increased risk of stroke, particularly in the posterior circulation. ^{55, 56} Giant cell arteritis preferentially affects the V3 and proximal V4 segments of the vertebral artery, possibly due to the higher amount of elastic tissue in their media and aventitia in comparison with other intracranial arteries. ^{57, 58} In granulomatosis with polyangiitis, two patterns of CNS involvement are recognized: a granulomatous pattern, with cerebral and/or spinal cord pachymeningitis, and a vasculitic pattern, where CNS vasculitis frequently co-exists with necrotizing glomerulonephritis. ⁵⁹ Neuro-Behçet causes meningoencephalitis and cerebral venous thrombosis more often than arteritis and can present with neutrophilic pleocytosis in the CSF. Retinal vasculitis can be associated with neuro-Behçet and tends to involve both arteries and veins. ⁶⁰

Many systemic auto-immune or auto-inflammatory conditions are associated with CNS disease through parenchymal or vascular inflammation, hypercoagulability, involvement of other organs, or complications of immunosuppressive treatments. Systemic lupus erythematosus can lead to ischemic stroke through accelerated atherosclerosis, antiphosholipid syndrome, Libman-Sacks endocarditis, and, more rarely, vasculitis. ⁶¹ Similarly, neurological symptoms can occur in Sjogren's syndrome due to CNS vasculitis but also to non-vasculitic mechanisms or co-existant auto-immune conditions (e.g. neuromyelitis optica). ^{62, 63} Neurosarcoidosis can present with granulomatous CNS vasculitis and is more likely than PACNS to affect the brainstem, basal citern meninges, hypothalamus, and spinal cord. ⁶⁴

Paraneoplasic CNS vasculitis is suspected when CNS vasculitis occurs simultaneously with lymphoma, leukemia, or solid cancers. ^{65, 66} Treatment with immune checkpoint inhibitors can be complicated by CNS vasculitis. ⁶⁷ There are rare reports of biopsy-proven CNS vasculitis in relation with the use of cocaine, amphetamines, and cannabis, often with hemorrhagic manifestations. ^{68, 69} However, in patients using those drugs, other mechanisms such as accelerated atherosclerosis or vasospasm are more frequent.

In cerebral amyloid angiopathy (CAA)-related inflammation, an auto-immune reaction targets cerebrovascular amyloid-beta deposits. Amyloid-beta-related angiitis (ABRA) refers to a subset of patients with CAA-related inflammation, where this auto-immune reaction involves the vessel wall. ABRA is diagnosed when vasculitic changes colocalize with amyloid-beta peptides on CNS biopsy. ⁷⁰ This pattern was found in 32% of patients with a previous diagnosis of biopsy-confirmed PACNS in the Mayo Clinic cohort. ⁷¹ When compared to biopsy-confirmed PACNS, patients with ABRA are older and more likely to present with leptomeningeal enhancement, intracerebral hemorrhage, and a granulomatous vasculitic pattern. ⁷² On MRI, ABRA is suspected when markers of CNS SVV co-occur with signs of CAA, such as cortical microbleeds or intracerebral hemorrhage, superficial siderosis, dilated perivascular spaces, and posterior predominant white-matter hyperintensities. ^{70, 73} (Fig. 3) CAA-related inflammation typically leads to asymetrical vasogenic edema and is more common in patients with a homozygous APOE4 genotype. ^{74, 75}

Section 4 : What is the workup for suspected CNS vasculitis?

The diagnostic workup in suspected CNS vasculitis aims to answer two questions: is the neurological presentation due to a vasculitic process, and if so, is the vasculitis primary (i.e., PACNS) or secondary? ⁷⁶ As defined by Birnbaum and Hellman, a definite diagnosis of PACNS can only be made with a positive CNS biopsy. ⁷⁷ In the absence of a tissue confirmation, the diagnosis of PACNS is at best probable.

A thorough multidisciplinary clinical evaluation should be performed, looking for signs of extra-CNS involvement. An ophthalmological exam should be considered in all patients, as many mimickers and secondary etiologies of CNS vasculitis can involve the eye. Fluorescein angiography can reveal peripheral retinal vascular disease in patients with an otherwise normal fundus examination. ⁷⁸ An audiogram can demonstrate patterns suggestive of specific immune-mediated disorders affecting the brain, eye and ear, such as low to mid frequencies sensorineural hearing loss in Susac syndrome. ⁷⁹ Table 4 lists a suggested workup and tests that can be considered based on the clinical presentation. Intravascular lymphoma can be isolated to the CNS and should be considered when serum lactate dehydrogenase levels are elevated. ⁸⁰ In the absence of a contraindication, a lumbar puncture should always be performed to evaluate for the presence of CNS inflammation and alternative infectious or neoplastic conditions. A CSF VZV PCR should systematically be obtained. CSF cytology, flow cytometry and T-cell receptor clonality assays can be considered when the clinical presentation is also compatible with CNS lymphoma. ⁸¹

Head CTA or MRA should always be obtained to document the presence and burden of medium vessel disease at initial presentation. In patients with medium vessel disease on non-invasive vascular imaging, DSA provides limited additional information and carries a small risk of neurological complications. ⁸² In patients with a normal CTA or MRA, DSA can identify distal or posterior medium vessel disease. ²³⁻²⁵ Repeat vascular imaging is sometimes necessary to clarify the diagnosis. Arterial stenoses are completely reversible in RCVS and post-SAH vasospasm, and often progress in untreated CNS vasculitis. Whole body imaging can reveal signs of extra-neurological findings (e.g. infection, cancer, systemic vasculitis), which may narrow the differential diagnosis and provide an extra-CNS target to biopsy.

Section 5 : When should CNS biopsy be considered?

The histopathological diagnosis of CNS vasculitis requires the presence of inflammatory cells within the vessel wall. Perivascular inflammation is not specific for vasculitis. ⁸³ Histological PACNS subtypes include granulomatous, lymphocytic and necrotizing CNS vasculitis. Although these subtypes may overlap, necrotizing CNS vasculitis tends to be associated with more severe presentations and hemorrhagic manifestations. ¹³ However, intracerebral hemorrhage can lead to secondary necrosis within the vessel wall. Necrotizing vasculitis can only be diagnosed in biopsy areas without hemorrhage or parenchymal necrosis. ¹³ Biopsy specimens should be analyzed for infectious etiologies of CNS vasculitis. In CNS lymphoma, immunostains can reveal atypical lymphocytes with high mitotic activity both within and around blood vessels. ⁸¹ Congo red staining differentiates PACNS from ABRA. ⁷⁰

CNS biopsies in patients with suspected PACNS are associated with a low risk of severe complications (ranging from 0-4%). ⁸⁴⁻⁸⁷ This risk should be weighed with the potential complications of immunosuppression in patients with alternative non-inflammatory disorders. In patients with SVV, the diagnosis of PACNS is only possible with a positive CNS biopsy. Moreover, patients with positive biopsies tend to have normal angiograms, likely because their disease is isolated to small vessels. ⁸⁸ In patients with unremitting or relapsing disease, a positive biopsy decreases diagnostic uncertainty and facilitates the decision to intensify treatment.

A CNS biopsy can reveal an alternative diagnosis in 30-39% of patients with suspected PACNS. ^{84, 85} In a meta-analysis of three observational studies, CNS biopsies for suspected PACNS led to the diagnosis of vasculitis or an alternative disorder in 75% of patients. ⁸⁶ A biopsy may be negative due to the segmental nature of vasculitic lesions, or to isolated involvement of proximal medium vessels, which are not sampled per procedure. Strategies to increase the yield of CNS biopsies include: (1) targeting radiologically abnormal sites; (2) selecting patients with radiological evidence of small vessel involvement (small infarcts, leptomeningeal or non-ischemic parenchymal enhancement, microbleeds and/or pseudotumoral lesions); (3) targeting a peripherally located and enhancing artery on vessel wall imaging; (4) sampling at least 1 cm³ of tissue; and (5) assessing tissue representativeness during the procedure. ^{13, 86, 89, 90}

Section 6 : What are the treatments and outcomes of cerebral vasculitis?

In secondary etiologies of cerebral vasculitis, the treatment is tailored to the underlying disease. For most systemic auto-immune or inflammatory conditions, CNS involvement predicts a worse prognosis and justifies more aggressive immunosuppression. ^{91, 92} In the absence of prospective and randomized trials, the treatment of PACNS is based on retrospective cohort studies and the well-established drug regimens that are used in systemic vasculitides. Infectious etiologies of CNS vasculitis should be ruled out prior to introducing immunosuppressants. In patients with mild or chronic presentations, we suggest completing the workup prior to initiating treatment. The impact of corticosteroids on CNS biopsy results in PACNS is unknown. However, corticosteroids increase the likelihood of false negative biopsies in certain PACNS mimics, such as giant cell arteritis and CNS lymphoma. ^{93, 94}

The treatment of PACNS should probably include an induction (to achieve remission) followed by a maintenance phase (to maintain remission and prevent relapses). Remission may be defined as the absence of new or worsening disease activity attributable to PACNS, after initiation of treatment. The minimum period needed to define remission after treatment introduction varies across cohorts. ⁴⁻⁶ Similarly, there is no consensual definition for a PACNS relapse. A relapse may be defined as the association of new clinical PACNS manifestations that occur after remission, are associated with new abnormalities on imaging, and lead to a change in treatment. ⁶ Whether isolated clinical (e.g. recrudescence of headaches with a stable brain MRI) or radiological events (e.g. silent brain infarct or asymptomatic new leptomeningeal enhancement) should define a relapse is unknown.

Induction therapy consists of high-dose glucocorticoids, usually accompanied by an immunosuppressive agent. Intravenous methlyprednisolone pulses (500-1000 mg/day) are sometimes used for 3-5 days, with no evidence to suggest they are superior to oral corticosteroids alone. Oral glucocorticoids (mainly prednisone or an equivalent) are initiated at 1 mg/kg per day (not exceeding 80 mg/day). This dose is maintained for 15-21 days, followed by a slow tapering over at least a oneyear period. In the two largest PACNS cohorts, induction with cyclophosphamide was not associated with higher odds of remission or lower relapse rates.^{4, 6} However, patients with or without cyclophosphamide were probably not comparable, i.e. patients who received cyclophosphamide probably had a more severe presentation. In light of its benefit in systemic vasculitides, it is reasonable to use cyclophosphamide in the induction phase of PACNS. ⁹⁵⁻⁹⁷ We usually prefer intravenous to oral cyclophosphamide, due to its association with reduced cumulative drug doses and lower odds of myelosuppression. ⁹⁸ Intravenous cyclophosphamide can be prescribed monthly at $0.7 \text{ g/m}^2/\text{month}$ (not exceeding 1200 mg) for at least six months and is adjusted for the leukocyte nadir and adverse events. In patients with a severe initial presentation, the first three infusions can be delivered every 15 days at 0.6 g/m^2 followed by three to six infusions every three weeks at 0.7 g/m².

Preliminary evidence suggests that rituximab may be effective in inducing remission when PACNS is refractory to glucocorticoids and cyclophosphamide, or when cyclophosphamide is contraindicated. ^{10, 99} In systemic vasculitides, rituximab is nowaday more frequently used than cyclophosphamide to achieve remission. Two rituximab regimens can be used: 375 mg/m² once weekly for four doses, or 1000 mg every two weeks for two doses. In systemic vasculitides, there is no evidence to suggest that either regimen is superior in terms of efficacy or safety. ¹⁰⁰

Maintenance therapy should be initiated after completing the induction treatment and achieving remission, usually one month after the last infusion of cyclophophamide or rituximab. Azathioprine (2 mg/kg/day in two oral intakes, not exceeding a total of 150 mg/day) is the most commonly used agent, followed by methotrexate (0.3 mg/kg/week, usually 20-25 mg/week, associated with folic acid) and mycophenolate mofetil (2000 to 3000 mg/day in two oral intakes). Similarly to systemic vasculitides, one infusion of 500 to 1000 mg of rituximab can be used for maintenance every six months, although data are scarce in PACNS.

In the French PACNS cohort, maintenance therapy was associated with higher odds of prolonged remission and good functional status. ⁶ The optimal duration of maintenance therapy (excluding glucocorticoids) is unknown. Maintenance therapy should probably be prescribed for at least 24 months to reduce the risk of a relapse. The rapidity of the glucocorticoid taper and the total duration of treatment should be adjusted based on the severity of the initial presentation, disease evolution, and drug tolerability.

Relapses are reported in 30-59% of patients with PACNS. ⁴⁻⁶ New-onset neurological manifestations may result from an undiagnosed PACNS mimic, PACNS sequelae (e.g. remote

seizure), a PACNS relapse, or complications of immunosuppression (e.g. CNS infection). ¹⁰¹ If the disease relapses after discontinuation of treatment, new induction and maintenance regimens should be initiated, and a longer course of maintenance immunosuppression is reasonable. If the disease relapses while the patient is receiving maintenance therapy, glucocorticoids should be re-initiated or increased, and a change of maintenance agent should be discussed. In cases where the relapse is clinically severe, a new induction regimen can be considered. In the rare cases refractory to cyclophosphamide or rituximab, the diagnosis should be reconsidered. In isolated case reports, PACNS responded to TNF-alpha inhibitors or tocilizumab (anti-IL6 receptor monoclonal antibody). ¹⁰²⁻¹⁰⁴

The benefit of aspirin in PACNS has not been demonstrated. ⁶ Aspirin should probably be avoided in patients with hemorrhagic manifestations and may be considered in the presence of medium vessel disease. All patients treated with glucocorticoids should receive adequate prophylaxes for osteoporosis (calcium, vitamin D \pm biphosphonates) and gastric ulcers (proton pump inhibitors). Co-trimoxazole is frequently prescribed for *Pneumocystic jiroveci* prophylaxis in patients initially treated with cyclophosphamide or rituximab. This treatment should be avoided in patients who receive maintenance therapy with methotrexate, due to the risk of drug interaction and methotrexate toxicity.

In the largest PACNS cohorts, induction therapy is associated with remission in 68-95% of patients. ⁴⁻⁶ Absence of remission should encourage clinicians to re-examine the diagnosis of PACNS and increases the justification for a CNS biopsy. In unremitting patients, a second biopsy may also be considered, as CNS lymphoma can initially present with a vasculitic pattern on histopathology. ⁸¹

Section 7 : How should I follow PACNS?

During the induction and maintenance phases, patients are closely monitored for signs of persistent disease activity or treatment-related complications. In absence of clear recommendations, we suggest controlling the brain MRI in the 3-4 months after introducing induction therapy, to document the presence of new lesions and establish a new baseline prior to initiating maintenance therapy. Afterwards, a brain MRI can be repeated every 6-12 months, or earlier if a relapse is suspected. Vascular imaging should be included in the follow-up of all patients. If medium vessel disease was demonstrated on non-invasive vascular imaging, a CTA or an MRA are sufficient for routine follow-up. Lumbar puncture can be discussed for patients with a potential relapse. Vessel wall enhancement can persist on MRI after remission, and is hypothesized to result from subclinical vasculitis, vascular remodeling, or post-inflammatory mural neovascularization. ¹⁰⁵ Persistant vessel wall enhancement without other signs of active CNS vasculitis probably does not justify intensifying immunosuppressive therapy. In these patients, closer clinical and radiological follow-up is reasonable.

During follow-up visits, patients should be screened for complications of CNS vasculitis, such as cognitive decline, epilepsy, or neuropsychiatric symptoms. Clinical evaluation and imaging can reveal signs of a PACNS relapse, which is more likely to occur in patients with isolated SVV, gadolinium enhancement at presentation, and absence of maintenance therapy. ^{6, 16} Increasing age and cerebral infarcts at diagnosis are associated with higher odds of disability or death during follow-up. ⁴ Overall mortality was 8% in the French PACNS cohort. ⁶ However, mortality after CNS vasculitis is likely underestimated, as patients with catastrophic presentation are more likely to die prior to diagnosis and not be included in retrospective cohort studies.

Conclusion

The diagnostic approach to PACNS is complex, due to the rarity of the disease, its multiple potential presentations and its numerous differential diagnoses. The distinction between medium and small vessel vasculitis is useful to guide the clinical approach. Cerebral vasculitis is more often suspected than confirmed, and alternative diagnoses should always be considered prior to exposing patients to the potential toxicity of immunosuppressants. Multidisciplinary collaboration (neurology, internal medicine, rheumatology, neuroradiology) is essential during the workup and the follow-up of the disease. Further studies are needed to identify and validate new diagnostic tests (biomarkers, advanced imaging), better define patient subgroups (monophasic versus recurrent PACNS), and allow the design of prospective therapeutic trials.

Reversible cerebral	Thunderclap headaches
vascoconstriction syndrome	Triggers (e.g. vasoconstrictive drugs, postpartum, sexual
	activity)
	Convexity subarachnoid hemorrhage or vasogenic edema
	Resolution of arterial stenoses at three months
Intracranial atherosclerosis	Vascular risk factors
	Arterial calcifications on CT
	Extracranial atherosclerosis
	Lipid core or intraplaque hemorrhage on vessel wall imaging
Intracranial dissection	Intramural hematoma (hyperintensity on T1 fat-saturated
	MRI)
	Double lumen or intimal flap
	Subarachnoid hemorrhage
	Pseudoaneurysm
Moya-moya disease or	Stenosis of the intracranial internal carotid artery and its
syndrome	proximal branches
	Dilated collaterals (puff of smoke pattern)
	Ivy sign on MRI
Intracranial fibromuscular	String of beads pattern
dysplasia	Extracranial fibromuscular dysplasia
	Cerebral aneurysms
Cerebral embolism	Partially recanalized cerebral emboli (e.g. atrial myxoma or
	other cardiac tumors, non-bacterial thrombotic endocarditis,
	aortic atherosclerosis)
Post-subarachnoid	History of recent thunderclap headache
hemorrhage vasospasm	Cerebrospinal fluid xanthochromia
Lymphoma	Intravascular lymphoma (elevated lactate dehydrogenase,
	skin lesions, small > medium vessel disease)
Prothrombotic state	Antiphospholipid syndrome
	Sneddon syndrome (livedo racemosa)
	Malignancy-associated hypercoagulability
Drug-induced vasospasm	Amphetamines, cannabis, cocaine
Radiation induced	History and other complications of cranial radiotherapy (e.g.
vasculopathy	small vessel disease, cavernoma, telangiectasia, meningioma)

Table 1. Selected non-vasculitic mimics of central nervous system medium vessel vasculitis

Hereditary disorders	with	ACTA2 mutation (abnormally straight course of intracranial
intracranial		arteries, periventricular white matter hyperintensities, patent
medium vessel disease		ductus arteriosus)
		Divry-van-Bogaert syndrome (juvenile ischemic stroke and dementia, arterial occlusions, cerebromeningeal angiomatosis, livedo racemosa) Hereditary moya-moya disease

Table 2. Selected non-vasculitic mimics of central nervous system small vessel vasculitis

Sporadic small vessel disease	Deep perforator arteriopathy
	Cerebral amyloid angiopathy
Hereditary disorders with	Cerebral autosomal dominant arteriopathy with subcortical
small vessel disease	infarcts and leukoencephalopathy (CADASIL)
	Cerebral autosomal recessive arteriopathy with subcortical
	infarcts and leukoencephalopathy (CARASIL)
	COL4A1 small-vessel disease
	Retinal vasculopathy with cerebral leukodystrophy (TREX1
	mutation)
	Fabry disease
	Homocystinuria
	Hereditary cerebral amyloid angiopathy
Other causes of small vessel	Drugs (amphetamines, cocaine)
disease	Radiation induced small vessel disease
Neoplastic vasculopathy	Intravascular lymphoma
	Primary central nervous system lymphoma
	Vascular infiltration by leptomeningeal neoplastic cells
Immune-mediated	Susac syndrome
endotheliopathy	
Prothrombotic state	Antiphospholipid syndrome
	Sneddon syndrome
	Myeloproliferative disorders (polycythemia vera, essential
	thrombocythemia)
	Thrombotic thrombocytopenic purpura
	Disseminated intravascular coagulation
	Malignancy-associated hypercoagulability
Cerebral embolism	Cholesterol embolization syndrome
Non-vascular disorders	Autoimmune encephalitis (primary or secondary to systemic
	disorder)
	Acute demyelinating encephalomyelitis
	Metabolic disease (ex: mitochondrial encephalopathy with
	lactic acidosis and stroke)

Infectious	Infectious endocarditis
	Bacterial (Streptococcus Pneumoniae, Haemophilus influenzae,
	Neisseria Meningitidis, syphilis, brucellosis, leptospirosis,
	Lyme disease, rickettsia)
	Mycobacterial (tuberculosis)
	Viral (varicella-zoster, cytomegalovirus, dengue, Epstein-Barr,
	hepatitis B and C, herpes simplex, West Nile)
	Fungal (aspergillosis, candidiasis, coccidiomycosis,
	cryptococcus, histoplasmosis, mucormycosis,
	paracoccidiomycosis)
	Parasitic (neurocysticercosis, malaria, schistosomiasis,
	toxoplasmosis)
Systemic vasculitides	Large vessel vasculitis (giant cell arteritis, Takayasu)
	Medium vessel vasculitis (Kawasaki, polyarteritis nodosa)
	Small vessel vasculitis (ANCA-associated vasculitis,
	cryoglobulinemic vasculitis, Goodpasture's syndrome, IgA
	vasculitis)
	Behcet's disease
	Buerger's disease
	Degos disease
Vasculitis associated with	Connective tissue disorder (systemic lupus erythematosus,
systemic disease	Sjogren, dermatomyositis, mixed connective tissue disease,
	systemic sclerosis)
	Graft versus host disease
	Inflammatory bowel disorder (Crohn, ulcerative colitis)
	Relapsing polychondritis
	Rheumatoid vasculitis
	Sarcoidosis
	Sweet syndrome
Vasculitis with ear, nose and	Acute posterior multifocal placoid pigment epitheliopathy
throat involvement	Cogan syndrome
Cancer	Paraneoplasic cerebral vasculitis (Hodgkin's lymphoma, non-
	Hodgkin's lymphoma, leukemia, solid cancers)
Drugs	Amphetamine, cannabis, cocaine

Table 3. Etiologies of central nervous system vasculitis

	Immune checkpoint inhibitors
	Propylthiouracil
Hereditary disorders	Adenosine deaminase 2 deficiency
	Common variable immunodeficiency
CNS-isolated vasculitis	Amyloid-beta related angiitis
	Primary angiitis of the CNS

Abbreviations: CNS - Central Nervous System

Table 4. Workup of central nervous system vasculitis

Pland and uning	Suggested
blood and urme	Suggesteu
	Complete blood count, routine coaguration testing (protironibin
	time, partial thromboplastin time)
	Creatinine, urine protein test, liver function tests
	C-reactive protein
	Anti-nuclear antibodies, anti-dsDNA antibodies, antineutrophil
	cytoplasmic antibodies, complement levels, rheumatoid factor
	Anti-cardiolipin antibodies, anti-B2GP1 antibodies, lupus
	anticoagulant
	Serum protein electrophoresis
	Lactate dehydrogenase
	Syphilis, HIV, hepatitis B, hepatitis C, Lyme, hemocultures,
	Quantiferon or tuberculin skin test
	Optional
	Cryoglobulins
	Street or urine drug test
	Genetic workup
CSF	Suggested
	Oligoclonal bands and IgG index
	Cytopathology
	Varicella Zoster PCR
	Bacterial and mycobacterial cultures
	Optional
	Flow cytometry, T-cell receptor clonality assay,
	immunoglobulin heavy chain gene rearrangement studies
	CSF/serum quantitative anti-VZV IgG/IgM ratio
	PCRs for Herpes Simplex. Cytomegalovirus. Epstein-Barr.
	Mycobacterium Tuberculosis
	Fungal cultures
	Cytokine panel
	Metagenomics next generation sequencing
Imaging	Suggested

MRI (including T1, T2, FLAIR, diffusion, MR-
aphy, gadolinium injected sequences)
nd neck CT-angiography
oracic or transesophageal echocardiogram
examination
body imaging (CT or PET)
al
solution MR vessel-wall imaging
subtraction angiography
cord MRI
cein angiography
ram
Nando Contra Con

Figure legends

Figure 1 – Imaging findings in cerebral vasculitis

A-D: Medium vessel vasculitis findings in different patients. A: Large and multiterritorial acute infarcts. B: Arterial stenoses in the proximal left anterior and middle cerebral arteries. C: Arterial stenoses (black arrows) with post-stenotic dilatation (white arrow) in the left anterior cerebral artery. D: Concentric enhancement of the second segment of the left middle cerebral artery on MRI vessel wall imaging.

E-H: Small vessel vasculitis findings in different patients. E: Multiple areas of punctiform nonischemic parenchymal enhancement. F: Numerous cortical microbleeds. The arrow shows the biopsy site. G: Cerebral and spinal parenchymal enhancement. H: Pseudotumoral cerebral vasculitis.

<u>Figure 2 – Imaging and histopathological findings in amyloid-beta related angiitiis (small vessel vasculitis)</u>

A: MRI diffusion weighted-imaging shows multiple punctiform acute and predominantly cortical infarcts. B: FLAIR sequence demonstrates predominantly posterior white-matter hyperintensities (circled), which can occur in cerebral amyloid angiopathy. C: Susceptibility-weighted imaging reveals multiple and exclusively cortical cerebral microbleeds (arrows). D: Hematoxyilin and eosin stain shows eosinophilic vessel walls, with mononucleate (arrow) and giant cell (triangle) infiltration of the vessel wall. E: Congo red staining is positive for amyloid (arrow). Giant cells are also seen in the perivascular space (triangle). Inflammatory changes colocalize with vascular amyloid deposits. F: Anti-amyloid β immunostaining reveals amyloid fibrils in the cortical and leptomeningeal vessel walls (arrows). Case courtesy of Dr Émile Lemoine (Centre Hospitalier de l'Université de Montréal).

<u>Figure 3 – Varicella Zoster medium vessel vasculitis in a patient with multiple sclerosis on</u> <u>fingolimod</u>

A: MRI diffusion-weighed imaging reveals acute infarction of the right parieto-occipital region. B: FLAIR sequence shows periventricular multiple sclerosis-related white matter lesions and infarction of the right parieto-occipital region. C: MRI vessel wall imaging demonstrates concentric enhancement of the right supraclinoid carotid artery (white arrow). D: MR-angiography with gadolinium reveals multifocal intracranial arterial stenoses (black arrows). Reproduced with permission from Elsevier (Muccilli et al. Varicella-zoster virus vasculopathy in a multiple sclerosis patient on fingolimod, J Neurol Sci 2019;403:119-21).

References

1. Sundaram S, Menon D, Khatri P, Sreedharan S, Jayadevan E, Prabhakaran S, et al. Primary angiitis of the central nervous system: Clinical profiles and outcomes of 45 patients. Neurology India 2019;67:105-12. https://http://dx.doi.org/10.4103/0028-3886.253578

2. Singhal AB, Topcuoglu MA, Fok JW, Kursun O, Nogueira RG, Frosch MP, et al. Reversible cerebral vasoconstriction syndromes and primary angiitis of the central nervous system: clinical, imaging, and angiographic comparison. Ann Neurol 2016;79:882-94. <u>https://10.1002/ana.24652</u>

3. Chang HB, Gao M, Zhang JN, Cao WD, Guo SL, Wang P, et al. Retrospective Analysis of 28 Cases Confirmed for Primary Angiitis of the Central Nervous System by Biopsy. Journal of Stroke and Cerebrovascular Diseases 2020;29(12) (no

pagination):https://http://dx.doi.org/10.1016/j.jstrokecerebrovasdis.2020.105400

4. Salvarani C, Brown RD, Jr., Christianson TJH, Huston J, 3rd, Giannini C, Hunder GG. Long-term remission, relapses and maintenance therapy in adult primary central nervous system vasculitis: A single-center 35-year experience. Autoimmun Rev 2020;19:102497. <u>https://10.1016/j.autrev.2020.102497</u>

5. Schuster S, Ozga AK, Stellmann JP, Deb-Chatterji M, Häußler V, Matschke J, et al. Relapse rates and long-term outcome in primary angiitis of the central nervous system. J Neurol 2019;266:1481-9. https://10.1007/s00415-019-09285-1

6. de Boysson H, Arquizan C, Touzé E, Zuber M, Boulouis G, Naggara O, et al. Treatment and Long-Term Outcomes of Primary Central Nervous System Vasculitis. Stroke 2018;49:1946-52. https://10.1161/strokeaha.118.021878

7. Zarka F, Veillette C, Makhzoum JP. A Review of Primary Vasculitis Mimickers Based on the Chapel Hill Consensus Classification. Int J Rheumatol 2020;2020:8392542. <u>https://10.1155/2020/8392542</u>

8. Cho TA, Jones A. CNS vasculopathies: Challenging mimickers of primary angiitis of the central nervous system. Best Pract Res Clin Rheumatol 2020;34:101569. <u>https://10.1016/j.berh.2020.101569</u>

9. Néel A, Auffray-Calvier E, Guillon B, Fontenoy AM, Loussouarn D, Pagnoux C, et al. Challenging the diagnosis of primary angiitis of the central nervous system: a single-center retrospective study. J Rheumatol 2012;39:1026-34. <u>https://10.3899/jrheum.110707</u>

10. Salvarani C, Brown RD, Jr., Muratore F, Christianson TJH, Galli E, Pipitone N, et al. Rituximab therapy for primary central nervous system vasculitis: A 6 patient experience and review of the literature. Autoimmun Rev 2019;18:399-405. <u>https://10.1016/j.autrev.2018.12.002</u>

11. Salvarani C, Brown RD, Christianson TJH, Huston J, Giannini C, Miller DV, et al. Mycophenolate mofetil in primary central nervous system vasculitis. Seminars in Arthritis and Rheumatism 2015;45(1):55-9. https://http://dx.doi.org/10.1016/j.semarthrit.2015.02.008 12. de Boysson H, Parienti JJ, Arquizan C, Boulouis G, Gaillard N, Régent A, et al. Maintenance therapy is associated with better long-term outcomes in adult patients with primary angiitis of the central nervous system. Rheumatology (Oxford) 2017;56:1684-93. https://10.1093/rheumatology/kex047

13. Giannini C, Salvarani C, Hunder G, Brown RD. Primary central nervous system vasculitis: pathology and mechanisms. Acta Neuropathol 2012;123:759-72. <u>https://10.1007/s00401-012-0973-9</u>

14. Salvarani C, Brown RD, Calamia KT, Christianson TJH, Huston J, Meschia JF, et al. Angiographynegative primary central nervous system vasculitis: A syndrome involving small cerebral vessels. Medicine 2008;87(5):264-71. <u>https://http://dx.doi.org/10.1097/MD.0b013e31818896e1</u>

15. de Boysson H, Zuber M, Naggara O, Neau JP, Gray F, Bousser MG, et al. Primary angiitis of the central nervous system: description of the first fifty-two adults enrolled in the French cohort of patients with primary vasculitis of the central nervous system. Arthritis Rheumatol 2014;66:1315-26. https://10.1002/art.38340

16. de Boysson H, Boulouis G, Aouba A, Bienvenu B, Guillevin L, Zuber M, et al. Adult primary angiitis of the central nervous system: Isolated small-vessel vasculitis represents distinct disease pattern.

Rheumatology (United Kingdom) 2017;56(3):439-44. <u>https://http://dx.doi.org/10.1093/rheumatology/kew434</u>
17. Schuster S, Bachmann H, Thom V, Kaufmann-Buehler AK, Matschke J, Siemonsen S, et al. Subtypes of primary angiitis of the CNS identified by MRI patterns reflect the size of affected vessels. J Neurol Neurosurg Psychiatry 2017;88:749-55. <u>https://10.1136/jnnp-2017-315691</u>

18. Beuker C, Strunk D, Rawal R, Schmidt-Pogoda A, Werring N, Milles L, et al. Primary Angiitis of the CNS: A Systematic Review and Meta-analysis. Neurol Neuroimmunol Neuroinflamm 2021:8:https://10.1212/nxi.00000000001093

19. de Boysson H, Pagnoux C. Catastrophic primary angiitis of the central nervous system. Eur J Neurol 2018;25:e3. https://10.1111/ene.13469

20. Salvarani C, Brown RD, Jr., Morris JM, Huston J, 3rd, Hunder GG. Catastrophic primary central nervous system vasculitis. Clin Exp Rheumatol 2014;32:S3-4.

21. Edgell RC, Sarhan AE, Soomro J, Einertson C, Kemp J, Shirani P, et al. The Role of Catheter Angiography in the Diagnosis of Central Nervous System Vasculitis. Interv Neurol 2016;5:194-208. https://10.1159/000445255

22. Küker W. Cerebral vasculitis: imaging signs revisited. Neuroradiology 2007;49:471-9. https://10.1007/s00234-007-0223-3

23. Boulouis G, de Boysson H, Zuber M, Guillevin L, Meary E, Costalat V, et al. Primary Angiitis of the Central Nervous System: Magnetic Resonance Imaging Spectrum of Parenchymal, Meningeal, and Vascular Lesions at Baseline. Stroke 2017;48:1248-55. <u>https://10.1161/strokeaha.116.016194</u>

24. de Boysson H, Boulouis G, Parienti JJ, Touzé E, Zuber M, Arquizan C, et al. Concordance of Timeof-Flight MRA and Digital Subtraction Angiography in Adult Primary Central Nervous System Vasculitis. AJNR Am J Neuroradiol 2017;38:1917-22. <u>https://10.3174/ajnr.A5300</u>

25. Eleftheriou D, Cox T, Saunders D, Klein NJ, Brogan PA, Ganesan V. Investigation of childhood central nervous system vasculitis: magnetic resonance angiography versus catheter cerebral angiography. Dev Med Child Neurol 2010;52:863-7. <u>https://10.1111/j.1469-8749.2009.03591.x</u>

26. Cosottini M, Canovetti S, Pesaresi I, Desideri I, Pizzanelli C, Catarsi E, et al. 3-T magnetic resonance angiography in primary angiitis of the central nervous system. Journal of computer assisted tomography 2013;37(4):493-8.

27. Wang J, Fleischmann D. Improving Spatial Resolution at CT: Development, Benefits, and Pitfalls. Radiology 2018;289:261-2. <u>https://10.1148/radiol.2018181156</u>

28. de Boysson H, Boulouis G, Dequatre N, Godard S, Néel A, Arquizan C, et al. Tumor-Like Presentation of Primary Angiitis of the Central Nervous System. Stroke 2016;47:2401-4. https://10.1161/strokeaha.116.013917

29. Salvarani C, Brown RD, Jr., Christianson TJH, Huston J, 3rd, Morris JM, Giannini C, et al. Primary central nervous system vasculitis mimicking brain tumor: Comprehensive analysis of 13 cases from a single institutional cohort of 191 cases. J Autoimmun 2019;97:22-8. <u>https://10.1016/j.jaut.2018.10.001</u>

30. Salvarani C, Brown RD, Jr., Calamia KT, Christianson TJ, Huston J, 3rd, Meschia JF, et al. Primary CNS vasculitis with spinal cord involvement. Neurology 2008;70:2394-400. https://10.1212/01.wnl.0000314687.69681.24

31. Obusez EC, Hui F, Hajj-Ali RA, Cerejo R, Calabrese LH, Hammad T, et al. High-resolution MRI vessel wall imaging: spatial and temporal patterns of reversible cerebral vasoconstriction syndrome and central nervous system vasculitis. AJNR Am J Neuroradiol 2014;35:1527-32. https://10.3174/ajnr.A3909

32. Mandell DM, Mossa-Basha M, Qiao Y, Hess CP, Hui F, Matouk C, et al. Intracranial Vessel Wall MRI: Principles and Expert Consensus Recommendations of the American Society of Neuroradiology. AJNR Am J Neuroradiol 2017;38:218-29. <u>https://10.3174/ajnr.A4893</u>

33. Kang N, Qiao Y, Wasserman BA. Essentials for Interpreting Intracranial Vessel Wall MRI Results: State of the Art. Radiology 2021;300:492-505. <u>https://10.1148/radiol.2021204096</u>

34. Renú A, Laredo C, Lopez-Rueda A, Llull L, Tudela R, San-Roman L, et al. Vessel Wall Enhancement and Blood-Cerebrospinal Fluid Barrier Disruption After Mechanical Thrombectomy in Acute Ischemic Stroke. Stroke 2017;48:651-7. <u>https://10.1161/strokeaha.116.015648</u>

35. Ramirez-Lassepas M, Patrick BK. Cerebrospinal fluid cellular response in uncomplicated acute ischemic stroke. J Stroke Cerebrovasc Dis 1992;2:168-72. <u>https://10.1016/s1052-3057(10)80229-0</u>

36. Prüss H, Iggena D, Baldinger T, Prinz V, Meisel A, Endres M, et al. Evidence of intrathecal immunoglobulin synthesis in stroke: a cohort study. Arch Neurol 2012;69:714-7. https://10.1001/archneurol.2011.3252

37. Britton M, Hultman E, Murray V, Sjöholm H. The diagnostic accuracy of CSF analyses in stroke. Acta Med Scand 1983;214:3-13. <u>https://10.1111/j.0954-6820.1983.tb08563.x</u>

38. Kraemer M, Berlit P. Primary central nervous system vasculitis: clinical experiences with 21 new European cases. Rheumatol Int 2011;31:463-72. <u>https://10.1007/s00296-009-1312-x</u>

39. Rocha EA, Topcuoglu MA, Silva GS, Singhal AB. RCVS(2) score and diagnostic approach for reversible cerebral vasoconstriction syndrome. Neurology 2019;92:e639-e47. https://10.1212/wnl.00000000006917

40. de Boysson H, Parienti JJ, Mawet J, Arquizan C, Boulouis G, Burcin C, et al. Primary angiitis of the CNS and reversible cerebral vasoconstriction syndrome: A comparative study. Neurology 2018;91:e1468-e78. https://10.1212/wnl.00000000006367

41. Ducros A. Reversible cerebral vasoconstriction syndrome. Lancet Neurol 2012;11:906-17. https://10.1016/s1474-4422(12)70135-7

42. Mandell DM, Matouk CC, Farb RI, Krings T, Agid R, terBrugge K, et al. Vessel wall MRI to differentiate between reversible cerebral vasoconstriction syndrome and central nervous system vasculitis: preliminary results. Stroke 2012;43:860-2. <u>https://10.1161/strokeaha.111.626184</u>

43. Sacco RL, Kargman DE, Gu Q, Zamanillo MC. Race-ethnicity and determinants of intracranial atherosclerotic cerebral infarction. The Northern Manhattan Stroke Study. Stroke 1995;26:14-20. https://10.1161/01.str.26.1.14

44. Hurford R, Wolters FJ, Li L, Lau KK, Küker W, Rothwell PM. Prevalence, predictors, and prognosis of symptomatic intracranial stenosis in patients with transient ischaemic attack or minor stroke: a population-based cohort study. Lancet Neurol 2020;19:413-21. <u>https://10.1016/s1474-4422(20)30079-x</u>

45. Swartz RH, Bhuta SS, Farb RI, Agid R, Willinsky RA, Terbrugge KG, et al. Intracranial arterial wall imaging using high-resolution 3-tesla contrast-enhanced MRI. Neurology 2009;72:627-34. https://10.1212/01.wnl.0000342470.69739.b3

46. Marrodan M, Acosta JN, Alessandro L, Fernandez VC, Carnero Contentti E, Arakaki N, et al. Clinical and imaging features distinguishing Susac syndrome from primary angiitis of the central nervous system. J Neurol Sci 2018;395:29-34. <u>https://10.1016/j.jns.2018.09.029</u>

47. Gross CC, Meyer C, Bhatia U, Yshii L, Kleffner I, Bauer J, et al. CD8(+) T cell-mediated endotheliopathy is a targetable mechanism of neuro-inflammation in Susac syndrome. Nat Commun 2019;10:5779. <u>https://10.1038/s41467-019-13593-5</u>

48. Younger DS, Coyle PK. Central Nervous System Vasculitis due to Infection. Neurol Clin 2019;37:441-63. <u>https://10.1016/j.ncl.2019.01.002</u>

49. Berlit P. Isolated angiitis of the CNS and bacterial endocarditis: similarities and differences. J Neurol 2009;256:792-5. <u>https://10.1007/s00415-009-5018-5</u>

50. Nagel MA, Cohrs RJ, Mahalingam R, Wellish MC, Forghani B, Schiller A, et al. The varicella zoster virus vasculopathies: clinical, CSF, imaging, and virologic features. Neurology 2008;70:853-60. https://10.1212/01.wnl.0000304747.38502.e8

51. Muccilli A, Nehme A, Labrie M, Girard M, Odier C, Poppe AY. Varicella-zoster virus vasculopathy in a multiple sclerosis patient on fingolimod. J Neurol Sci 2019;403:119-21. <u>https://10.1016/j.jns.2019.06.025</u>

52. Nagel MA, Forghani B, Mahalingam R, Wellish MC, Cohrs RJ, Russman AN, et al. The value of detecting anti-VZV IgG antibody in CSF to diagnose VZV vasculopathy. Neurology 2007;68:1069-73. https://10.1212/01.wnl.0000258549.13334.16

53. Javaud N, Certal Rda S, Stirnemann J, Morin AS, Chamouard JM, Augier A, et al. Tuberculous cerebral vasculitis: retrospective study of 10 cases. Eur J Intern Med 2011;22:e99-104. https://10.1016/j.ejim.2011.04.004

54. Poisson KE, Zygmunt A, Leino D, Fuller CE, Jones BV, Haslam D, et al. Lethal Pediatric Cerebral Vasculitis Triggered by Severe Acute Respiratory Syndrome Coronavirus 2. Pediatr Neurol 2021;127:1-5. https://10.1016/j.pediatrneurol.2021.11.003

55. Ungprasert P, Wijarnpreecha K, Koster MJ, Thongprayoon C, Warrington KJ. Cerebrovascular accident in patients with giant cell arteritis: A systematic review and meta-analysis of cohort studies. Semin Arthritis Rheum 2016;46:361-6. https://10.1016/j.semarthrit.2016.07.005

56. de Boysson H, Liozon E, Larivière D, Samson M, Parienti JJ, Boutemy J, et al. Giant Cell Arteritisrelated Stroke: A Retrospective Multicenter Case-control Study. J Rheumatol 2017;44:297-303. https://10.3899/jrheum.161033

57. Beuker C, Wankner MC, Thomas C, Strecker JK, Schmidt-Pogoda A, Schwindt W, et al. Characterization of Extracranial Giant Cell Arteritis with Intracranial Involvement and its Rapidly Progressive Subtype. Ann Neurol 2021;90:118-29. <u>https://10.1002/ana.26101</u>

58. Wilkinson IM, Russell RW. Arteries of the head and neck in giant cell arteritis. A pathological study to show the pattern of arterial involvement. Arch Neurol 1972;27:378-91.

https://10.1001/archneur.1972.00490170010003

59. De Luna G, Terrier B, Kaminsky P, Le Quellec A, Maurier F, Solans R, et al. Central nervous system involvement of granulomatosis with polyangiitis: clinical-radiological presentation distinguishes different outcomes. Rheumatology (Oxford) 2015;54:424-32. <u>https://10.1093/rheumatology/keu336</u>

60. Akman-Demir G, Serdaroglu P, Tasçi B. Clinical patterns of neurological involvement in Behçet's disease: evaluation of 200 patients. The Neuro-Behçet Study Group. Brain 1999;122 (Pt 11):2171-82. https://10.1093/brain/122.11.2171

61. Weiner DK, Allen NB. Large vessel vasculitis of the central nervous system in systemic lupus erythematosus: report and review of the literature. J Rheumatol 1991;18:748-51.

62. Carvalho DC, Tironi TS, Freitas DS, Kleinpaul R, Talim NC, Lana-Peixoto MA. Sjögren syndrome and neuromyelitis optica spectrum disorder co-exist in a common autoimmune milieu. Arq Neuropsiquiatr 2014;72:619-24. <u>https://10.1590/0004-282x20140097</u>

63. Jobling K, Ledingham D, Ng WF, Guadagno J. Positive anti-MOG antibodies in a patient with Sjögren's syndrome and transverse myelitis. Eur J Rheumatol 2019;6:102-4. https://10.5152/eurjrheum.2018.18041

64. Saygin D, Jones S, Sundaram P, Calabrese LH, Messner W, Tavee JO, et al. Differentiation between neurosarcoidosis and primary central nervous system vasculitis based on demographic, cerebrospinal and imaging features. Clin Exp Rheumatol 2020;38 Suppl 124:135-8.

65. Salvarani C, Brown RD, Jr., Christianson TJH, Huston J, 3rd, Ansell SM, Giannini C, et al. Primary central nervous system vasculitis associated with lymphoma. Neurology 2018;90:e847-e55. https://10.1212/wnl.000000000005062

66. Taccone FS, Salmon I, Marechal R, Blecic SA. Paraneoplastic vasculitis of central nervous system presenting as recurrent cryptogenic stroke. Int J Clin Oncol 2007;12:155-9. <u>https://10.1007/s10147-006-0631-0</u>

67. Daxini A, Cronin K, Sreih AG. Vasculitis associated with immune checkpoint inhibitors-a systematic review. Clin Rheumatol 2018;37:2579-84. <u>https://10.1007/s10067-018-4177-0</u>

68. Merkel PA, Koroshetz WJ, Irizarry MC, Cudkowicz ME. Cocaine-associated cerebral vasculitis. Semin Arthritis Rheum 1995;25:172-83. <u>https://10.1016/s0049-0172(95)80029-8</u>

69. Citron BP, Halpern M, McCarron M, Lundberg GD, McCormick R, Pincus IJ, et al. Necrotizing angiitis associated with drug abuse. N Engl J Med 1970;283:1003-11. <u>https://10.1056/nejm197011052831901</u>

70. Moussaddy A, Levy A, Strbian D, Sundararajan S, Berthelet F, Lanthier S. Inflammatory Cerebral Amyloid Angiopathy, Amyloid-β-Related Angiitis, and Primary Angiitis of the Central Nervous System: Similarities and Differences. Stroke 2015;46:e210-3. <u>https://10.1161/strokeaha.115.010024</u>

71. Salvarani C, Hunder GG, Morris JM, Brown RD, Jr., Christianson T, Giannini C. Aβ-related angiitis: comparison with CAA without inflammation and primary CNS vasculitis. Neurology 2013;81:1596-603. https://10.1212/WNL.0b013e3182a9f545

72. Salvarani C, Brown RD, Jr., Calamia KT, Christianson TJ, Huston J, 3rd, Meschia JF, et al. Primary central nervous system vasculitis: comparison of patients with and without cerebral amyloid angiopathy. Rheumatology (Oxford) 2008;47:1671-7. <u>https://10.1093/rheumatology/ken328</u>

73. Scolding NJ, Joseph F, Kirby PA, Mazanti I, Gray F, Mikol J, et al. Abeta-related angiitis: primary angiitis of the central nervous system associated with cerebral amyloid angiopathy. Brain 2005;128:500-15. https://10.1093/brain/awh379

74. Antolini L, DiFrancesco JC, Zedde M, Basso G, Arighi A, Shima A, et al. Spontaneous ARIA-like Events in Cerebral Amyloid Angiopathy-Related Inflammation: A Multicenter Prospective Longitudinal Cohort Study. Neurology 2021;97:e1809-e22. https://10.1212/wnl.000000000012778

75. Eng JA, Frosch MP, Choi K, Rebeck GW, Greenberg SM. Clinical manifestations of cerebral amyloid angiopathy-related inflammation. Ann Neurol 2004;55:250-6. <u>https://10.1002/ana.10810</u>

76. Calabrese LH, Mallek JA. Primary angiitis of the central nervous system. Report of 8 new cases, review of the literature, and proposal for diagnostic criteria. Medicine (Baltimore) 1988;67:20-39. https://10.1097/00005792-198801000-00002

77. Birnbaum J, Hellmann DB. Primary angiitis of the central nervous system. Arch Neurol 2009;66:704-9. <u>https://10.1001/archneurol.2009.76</u>

78. Martinet N, Fardeau C, Adam R, Bodaghi B, Papo T, Piette JC, et al. Fluorescein and indocyanine green angiographies in Susac syndrome. Retina 2007;27:1238-42. <u>https://10.1097/IAE.0b013e31809ff824</u>

79. Triplett JD, Buzzard KA, Lubomski M, Riminton DS, Barnett MH, Welgampola MS, et al. Immunemediated conditions affecting the brain, eye and ear (BEE syndromes). J Neurol Neurosurg Psychiatry 2019;90:882-94. <u>https://10.1136/jnnp-2018-319002</u>

80. Ponzoni M, Ferreri AJ, Campo E, Facchetti F, Mazzucchelli L, Yoshino T, et al. Definition, diagnosis, and management of intravascular large B-cell lymphoma: proposals and perspectives from an international consensus meeting. J Clin Oncol 2007;25:3168-73. <u>https://10.1200/jco.2006.08.2313</u>

81. Feldman L, Li Y, Ontaneda D. Primary CNS lymphoma initially diagnosed as vasculitis. Neurol Clin Pract 2020;10:84-8. <u>https://10.1212/CPJ.0000000000693</u>

Leffers AM, Wagner A. Neurologic complications of cerebral angiography. A retrospective study of complication rate and patient risk factors. Acta Radiol 2000;41:204-10. <u>https://10.1080/028418500127345299</u>
 Parisi JE, Moore PM. The role of biopsy in vasculitis of the central nervous system. Semin Neurol

Parisi JE, Moore PM. The role of biopsy in vasculitis of the central nervous system. Se 1994;14:341-8. <u>https://10.1055/s-2008-1041093</u>

84. Alrawi A, Trobe JD, Blaivas M, Musch DC. Brain biopsy in primary angiitis of the central nervous system. Neurology 1999;53:858-60. <u>https://10.1212/wnl.53.4.858</u>

85. Torres J, Loomis C, Cucchiara B, Smith M, Messé S. Diagnostic Yield and Safety of Brain Biopsy for Suspected Primary Central Nervous System Angiitis. Stroke 2016;47:2127-9. https://10.1161/strokeaha.116.013874

86. Bai HX, Zou Y, Lee AM, Lancaster E, Yang L. Diagnostic Value and Safety of Brain Biopsy in Patients With Cryptogenic Neurological Disease: A Systematic Review and Meta-analysis of 831 Cases. Neurosurgery 2015;77:283-95; discussion 95. <u>https://10.1227/neu.000000000000756</u>

87. Rice CM, Gilkes CE, Teare E, Hardie RJ, Scolding NJ, Edwards RJ. Brain biopsy in cryptogenic neurological disease. Br J Neurosurg 2011;25:614-20. <u>https://10.3109/02688697.2010.551677</u>

88. Raghavan A, Wright JM, Huang Wright C, Shammassian BH, Onyewadume L, Momotaz H, et al. Concordance of angiography and cerebral biopsy results for suspected primary central nervous system vasculitis: A multi-center retrospective review. Clin Neurol Neurosurg 2019;185:105482. https://10.1016/j.clineuro.2019.105482

89. Zeiler SR, Qiao Y, Pardo CA, Lim M, Wasserman BA. Vessel Wall MRI for Targeting Biopsies of Intracranial Vasculitis. AJNR American journal of neuroradiology 2018;39:2034-6. https://10.3174/ajnr.A5801

90. Krawczyk M, Barra LJ, Sposato LA, Mandzia JL. Primary CNS vasculitis: A systematic review on clinical characteristics associated with abnormal biopsy and angiography. Autoimmun Rev 2021;20:102714. https://10.1016/j.autrev.2020.102714

91. Guillevin L, Lhote F, Gayraud M, Cohen P, Jarrousse B, Lortholary O, et al. Prognostic factors in polyarteritis nodosa and Churg-Strauss syndrome. A prospective study in 342 patients. Medicine (Baltimore) 1996;75:17-28. https://10.1097/00005792-199601000-00003

92. Rennebohm RM, Asdaghi N, Srivastava S, Gertner E. Guidelines for treatment of Susac syndrome - An update. Int J Stroke 2020;15:484-94. <u>https://10.1177/1747493017751737</u>

93. Jakobsson K, Jacobsson L, Mohammad AJ, Nilsson J, Warrington K, Matteson EL, et al. The effect of clinical features and glucocorticoids on biopsy findings in giant cell arteritis. BMC Musculoskelet Disord 2016;17:363. <u>https://10.1186/s12891-016-1225-2</u>

94. Giannini C, Dogan A, Salomão DR. CNS lymphoma: a practical diagnostic approach. J Neuropathol Exp Neurol 2014;73:478-94. <u>https://10.1097/nen.00000000000076</u>

95. Hoffman GS, Kerr GS, Leavitt RY, Hallahan CW, Lebovics RS, Travis WD, et al. Wegener granulomatosis: an analysis of 158 patients. Ann Intern Med 1992;116:488-98. <u>https://10.7326/0003-4819-116-6-488</u>

96. Salvarani C, Brown RD, Jr., Christianson TJ, Huston J, 3rd, Giannini C, Miller DV, et al. Adult primary central nervous system vasculitis treatment and course: analysis of one hundred sixty-three patients. Arthritis Rheumatol 2015;67:1637-45. <u>https://10.1002/art.39068</u>

97. Guillevin L, Jarrousse B, Lok C, Lhote F, Jais JP, Le Thi Huong Du D, et al. Longterm followup after treatment of polyarteritis nodosa and Churg-Strauss angiitis with comparison of steroids, plasma exchange and cyclophosphamide to steroids and plasma exchange. A prospective randomized trial of 71 patients. The Cooperative Study Group for Polyarteritis Nodosa. J Rheumatol 1991;18:567-74.

98. de Groot K, Harper L, Jayne DR, Flores Suarez LF, Gregorini G, Gross WL, et al. Pulse versus daily oral cyclophosphamide for induction of remission in antineutrophil cytoplasmic antibody-associated vasculitis: a randomized trial. Ann Intern Med 2009;150:670-80. <u>https://10.7326/0003-4819-150-10-200905190-00004</u>

99. De Boysson H, Arquizan C, Guillevin L, Pagnoux C. Rituximab for primary angiitis of the central nervous system: Report of 2 patients from the French COVAC cohort and review of the literature. Journal of Rheumatology 2013;40(12):2102-3. <u>https://http://dx.doi.org/10.3899/jrheum.130529</u>

100. Bénard V, Farhat C, Zarandi-Nowroozi M, Durand M, Charles P, Puéchal X, et al. Comparison of Two Rituximab Induction Regimens for Antineutrophil Cytoplasm Antibody-Associated Vasculitis:

Systematic Review and Meta-Analysis. ACR Open Rheumatol 2021;3:484-94. <u>https://10.1002/acr2.11274</u> 101. Bose G, Fitzpatrick T, Yogendrakumar V, Zwicker JC, Jansen G. Clinical Reasoning: A 73-year-old man with recurrent aphasia, headaches, and confusion. Neurology 2020;95:e2595-e9. https://10.1212/wnl.000000000010475

102. Salvarani C, Brown RD, Jr., Calamia KT, Huston J, 3rd, Meschia JF, Giannini C, et al. Efficacy of tumor necrosis factor alpha blockade in primary central nervous system vasculitis resistant to immunosuppressive treatment. Arthritis Rheum 2008;59:291-6. https://10.1002/art.23337

103. Fernandes BM, Abreu P, Bernardes M, Reis C, Azevedo E. Tocilizumab-A New Therapeutic Option for Primary Angiitis of the Central Nervous System? J Clin Rheumatol 2021;27:S735-s6. https://10.1097/rhu.00000000001535

104. Cabreira V, Dias L, Fernandes B, Aires A, Guimarães J, Abreu P, et al. Tocilizumab for severe refractory primary central nervous system vasculitis: A center experience. Acta Neurol Scand 2022;145:479-83. <u>https://10.1111/ane.13563</u>

105. Patzig M, Forbrig R, Küpper C, Eren O, Saam T, Kellert L, et al. Diagnosis and follow-up evaluation of central nervous system vasculitis: an evaluation of vessel-wall MRI findings. J Neurol 2022;269:982-96. https://10.1007/s00415-021-10683-7







