# Machine learning approach to assess the association between anthropometric, metabolic and nutritional status and semen parameters

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- 21 **Running title:** Modeling, metabolic status and semen parameters.
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### 25 Abstract:

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27 Many lifestyle factors, such as nutritional imbalance leading to obesity, metabolic disorders and nutritional deficiency, have been identified as potential risk factors for male infertility. The aim 28 29 of this study was to evaluate the relationship between semen parameters and both anthropometric, 30 metabolic and nutritional parameters individually and then after the application of a previously constructed and validated machine learning score that allows their combination. Anthropometric, 31 32 metabolic, antioxidant, micronutrient, and sperm parameters from 75 men suffering from 33 idiopathic infertility recruited from four infertility centers in France (Jean-Verdier ART center Hospital, Bondy; North Hospital ART center, Saint-Étienne; Navarre Polyclinic ART center, 34 35 Pau: Cochin Hospital ART center, Paris) between September 2009 and December 2013 were 36 collected. After assessing standard correlation analysis, a previously built machine learning model, providing a score ranging from 0 (poorest) to 1 (most favorable) was calculated for each 37 38 man in the study cohort. This machine learning model, that separates infertile/fertile men with 39 unexplained infertility on the basis of their bioclinical signature, providing a more holistic evaluation of the influence of the considered markers (anthropometric, metabolic and oxidative 40 status). We observed a significant correlation of some anthropometric, metabolic and nutritional 41 42 disorders with some sperm characteristics. Moreover, unfavorable machine learning score was associated with a high level of sperm DNA fragmentation. Favorable anthropometric, metabolic 43 and oxidative patterns, which may reflect an appropriate lifestyle, appear to positively impact 44 overall health, in particular reproductive function. This study, consistent with previous 45 46 publications, suggests that beyond semen quality parameters, in an essential assessment of male 47 fertility, other key factors should be taken into account. In this regard, the application of 48 emerging artificial intelligence techniques may provide a unique opportunity to integrate all these 49 parameters and deliver personalized care.

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52 Keywords: lifestyle ; machine learning ; metabolism ; nutrition ; sperm DNA fragmentation

#### 53 Introduction

As recently published by Levine *et al.*, fertility has been declining worldwide for the past half 54 century, especially sperm quality<sup>1</sup>. Many lifestyle factors have been identified as potential risk 55 56 factors for infertility in men. Among them, overweight and obesity have been particularly studied <sup>2</sup>. A global increase in the prevalence of obesity has been observed over the last three decades 57 58 and now more than half of men of reproductive age are currently considered to be overweight or 59 obese. It has been well established that overweight and obesity are associated with male infertility <sup>3</sup> and have a negative impact on semen parameters including sperm count <sup>2,4</sup> and sperm DNA 60 fragmentation <sup>5,6</sup>. In addition, metabolic disorders that are often the consequence of an 61 62 unbalanced diet and/or insufficient physical activity, also contribute to impaired reproductive function<sup>7</sup>. 63

Recently our team has highlighted, that metabolic syndrome, but also metabolic disorders (increased fasting blood glucose) and anthropometric disorders (increased body mass index (BMI) and abdominal obesity) may be risk factors for idiopathic infertility <sup>8</sup>. We had also observed that male partners of infertile couples were less physically active than fertile men <sup>9</sup>. They also had lower plasma levels of antioxidant vitamins <sup>10</sup>.

69 There are many complex mechanisms involved in these phenomena. Overweight, obesity, and 70 metabolic disorders can lead to impaired functioning of the hypothalamic-pituitary-gonadal 71 (HHG) axis. The process of converting steroids into estrogens in peripheral adipose tissue leads 72 to an elevation in estradiol levels, which exerts a negative feedback loop at the hypothalamic-73 pituitary level. In these men, secondary hypogonadism is therefore observed, characterized by a 74 decrease in serum concentrations of follicle-stimulating hormone (FSH), luteinizing hormone (LH) and testosterone <sup>11,12</sup>. Hyperinsulinemia leads to a decrease in hepatic production of Sex 75 Hormone Binding Globulin (SHBG)<sup>13</sup>. Consequences are increasing level of free testosterone 76

77 available for conversion to estradiol. Chronic systemic inflammation, which plays a critical role in the development of metabolic syndrome, is associated with obesity <sup>14</sup>. Levels of inflammatory 78 cytokines (Interleukin 1-alpha, Interleukin-6, Tumor necrosis factor- $\alpha$ , Activin a, etc.), which are 79 essential for normal spermatogenesis, are disturbed <sup>15</sup>. Systemic oxidative stress is also associated 80 with increased body weight and obesity <sup>16</sup>. Oxidative stress and lipid peroxidation are increased 81 in the testicular microenvironment following the accumulation of adipose tissue <sup>17</sup>. Furthermore, 82 83 high blood glucose levels have been shown to increase oxidative stress through mitochondrial oxidation of glucose, which releases a substantial amount of free radicals into the cytosol<sup>18</sup>. In 84 85 addition, oxidative stress leads to the production of nitric oxide (NO) which oxidizes sperm membrane lipids. Oxidative sperm impair spermatogenesis and is detrimental to sperm motility 86 87 <sup>19</sup>. Germ cells are thought to be more sensitive to free radical oxidation than somatic cells, 88 because their plasma membranes contain a greater quantity of polyunsaturated fatty acids leading to lipid peroxidation<sup>20</sup>. Oxidative stress increases sperm damage and results in increased sperm 89 90 DNA fragmentation, which affects early embryo development, future child health, and increases the risk of miscarriage  $^{21,22}$ . 91

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93 Thus, finally, the mechanisms involved in the impact of obesity and metabolic disorders on male 94 reproductive functions are numerous, but strongly interlinked. Until recently, most of the studies 95 have independently investigated the relationship between infertility and anthropometric, 96 metabolic and dietary elements. Therefore, identifying the parameter(s) involved in male or 97 female infertility is challenging. Artificial intelligence, especially machine learning, has been 98 adopted in scientific and medical research in the last few years, providing access to innovative 99 and more powerful tools. We previously built and evaluated a Machine Learning (ML) model to identify and stratify fertile couples as well as those with idiopathic infertility. This score was 100

based on a panel of lifestyle-influenced parameters, including the anthropometric, metabolic and antioxidant status of both partners. This tool combines several easily measurable parameters and generates a score to assess the impact of anthropometric, metabolic and antioxidant factors on the couple's risk of infertility. Although less efficient, we have also shown that this score, in its version built on male parameters alone, could identify almost 70% of men who were at risk of infertility <sup>10</sup>.

However, a score combining different elements influenced by lifestyle has never been used toidentify the risk of impaired sperm parameters.

109 The objective of this study was to evaluate the relationship between semen parameters and 110 anthropometric, metabolic, and nutritional parameters. First, we analyzed each parameter 111 individually, and then we applied a previously constructed and validated machine learning score 112 to combine them.

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#### 114 Material and methods

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116 Subjects:

117 "ALImentation et FERTilité" (ALIFERT) is a cross-sectional case-control study enrolling 118 infertile men between September 2009 and December 2013 (National biomedical research ID no. 119 P071224; ethics committee approval ('Comité de Protection des Personnes') ID no. AOM 2009-120 A00256-51; NEudra CT ID no. 08180; clinicaltrials.gov ID no. NCT01093378). The design of ALIFERT was multicenter. The study cohort included 75 infertile men recruited from four 121 122 infertility centers in France (Jean-Verdier ART center Hospital, Bondy (JV); North Hospital ART 123 center, Saint-Étienne (SE); Navarre Polyclinic ART center, Pau (PAU); Cochin Hospital ART 124 center, Paris (CCH)).

Men were between 18 to 45 and exhibiting idiopathic infertility for over 12 months. Eligibility criteria have been described previously and include the provision of written informed consent <sup>10</sup>. Eligibility criteria for subjects were as follows: (i) primary idiopathic infertility >12 months, (ii) age between 18 and 45 years, (iii) absence of severe oligozoospermia (<5 million/ml), (iv) absence of male reproductive tract abnormalities such as undescended testis, varicocele or infection, (v) written informed consent. Subjects with any current known or previous metabolic or digestive disease and smokers were excluded.

132 <u>Assessments:</u>

Anthropometric, metabolic, antioxidant, and micronutrient parameters were assessed as describedbelow:

Height, weight, visceral fat (Tanita BC-420MA analyzer, TANITA USA, Issaquah, WA, USA), and waist circumference measured at the narrowest point between the lower edge of the ribs and the iliac crest were measured by the investigator. The measurements were carried out by a trained investigator in the morning under fasting conditions.

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Blood samples were obtained after 12 hours of fasting for measurement of total
cholesterol, high-density lipoprotein (HDL) cholesterol, low-density lipoprotein (LDL)
cholesterol, triglycerides, and glucose in fresh plasma. Both serum and plasma were frozen at 80°C until further analysis.

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After 5 minutes of rest, systolic and diastolic blood pressures were assessed with a sphygmomanometer cuff around the patient's forearm while lying down. The values for systolic and diastolic pressures were calculated as the mean of the right and left measurements.

Finally, concentrations of serum vitamin D (ng/mL), vitamin B9 (folic acid-erytho) 148 149 (nmol), vitamin B9 (nmol/L), vitamin B12 (pmol/L), alpha-tocopherol (vitamin E) (µmol/L), zinc 150 (mmol/L), selenium (mmol/L), vitamin C (mg/mL), alpha-carotene (µmol/L), beta-carotene (µmol/L), Lycopene (µmol/L), lutein (µmol/L), beta-kryptoxanthin (µmol/L), and retinol 151 152 (µmol/L) (vitamin A) levels were determined at the Department of Integrated Biology -Nutritional Biology and Oxidative Stress (Grenoble Hospital, Grenoble, France). Ascorbic acid 153 154 (vitamin C) in serum was assessed by an automated continuous-flow method. The concentrations 155 of retinol, tocopherol, and carotenoids (lutein,  $\beta$ -cryptoxanthin, lycopene,  $\alpha$ -carotene, and  $\beta$ -156 carotene) in serum were obtained by HPLC (Biotek-Kontron, Montigny-le-Bretonneux, France). Concentrations of serum zinc were measured by flame atomic absorption spectrometry (model 157 158 3110; Perkin Elmer, Norwalk, CT, USA) and selenium by atomic absorption spectrometry (4100 159 ZL; Perkin Elmer).

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#### 161 <u>Semen quality parameters:</u>

The semen specimens were collected in the laboratory by masturbation into a sterile plastic cup after an abstinence period of 2 to 5 days. The samples were liquefied at room temperature for 30 minutes and conventional semen quality (semen volume, sperm concentration, vitality and motility) was assessed according to the 2010 WHO guidelines <sup>23</sup>. David's criteria were used to evaluate sperm morphology <sup>24,25</sup>.

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<u>Sperm DNA fragmentation</u>: The TUNEL (terminal deoxynucleotidyl transferase dUTP nick end
 labeling) technique with an In Situ Cell Death Detection Kit (Fluorescein, Roche Applied
 Science, Meylan, France) was used to determine sperm DNA fragmentation.

172 <u>Machine learning model:</u>

173 In previous work, our team developed and evaluated three machine learning models, capable of 174 separating infertile/fertile (1) couples, (2) men and (3) women through their bioclinical signature, in order to facilitate the management of couples with unexplained infertility <sup>10</sup>. This signature 175 176 was the result of a precise selection of biomarkers among hundreds of parameters investigated 177 and is based on the findings of a large-scale clinical study evaluating the impact of lifestyle and 178 nutrition on couples' infertility. A major contribution of lifestyle parameters to the discriminatory 179 power of the model had been demonstrated. The result generated and calculated by the model 180 produces a score ranging from 0 (the highest risk of infertility with involvement of nutritional and 181 metabolic factors) to 1 (the lowest risk, therefore rather healthy nutritional and metabolic profile). 182 We therefore calculated the model score for each man in the study cohort, using the formula obtained previously<sup>10</sup>. A score between 0 and 1 was therefore calculated for each man in the 183 184 cohort, with the aim of assessing more globally the nutritional and metabolic status on fertility 185 (Figure 1) and comparing its results with sperm parameters.

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187 <u>Statistical analysis</u>

Python 3.9.12 was used to carry out the statistical analysis, and the SciPy package version 1.9.3
was used. Data are shown as means and standard deviation. Correlation analysis of metabolic
and sperm parameters was investigated by parametric Pearson test. P-value < 0.05 was</li>
considered statistically significant.

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193 **Results:** 

194 Data are summarized as mean and standard deviation and shown in Table 1.

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Association between anthropometric, metabolic, and dietary parameters and semen parameters ispresented in Table 2.

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In this population of idiopathic infertile men, we observed a correlation between some anthropometric characteristics and semen parameters. A decrease in sperm motility was observed with high BMI (r = -0.233, p < 0.05). In addition, high visceral fat was associated with a decrease in sperm vitality and motility (r = -0.285, p < 0.01 and r = -0.27, p < 0.05).

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Regarding the correlation between metabolic characteristic and sperm parameters, high levels of total cholesterol and serum LDL were associated with lower sperm count (r = -0.272 and r = -0.249, p < 0.05). In addition, lower sperm vitality was observed in patients with high total cholesterol and serum LDL (r = -0.311 and r = -0.326 p < 0.01). Finally, sperm DNA fragmentation was negatively correlated with serum HDL concentration (r = -0.364, p < 0.01).

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A high concentration of certain antioxidants in the serum is associated with better sperm parameters. Correlation between sperm count and plasma Glutathione peroxidase activity (r = 0.23, p < 0.05), as well as between sperm motility and vitamin B12 (cobalamin) concentration was observed (r = 0.275, p < 0.05).

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216 Unfavorable machine learning score, based on a combination of anthropometric, metabolic and 217 oxidative status, was associated with a high level of sperm DNA fragmentation (r = -0.263, p-218 value < 0.05). 220 Discussion

The aim of this study was to explore the impact of factors such as anthropometric and metabolic status along with the nutritional environment on semen parameters in a population of patients with unexplained infertility. This study explore the relationship between a circulating metabolic and nutritional signature and sperm DNA fragmentation, a qualitative marker of spermatogenesis.

226 We observed a significant correlation of some anthropometric, metabolic and nutritional 227 disorders with some sperm characteristics. These associations were not as pronounced as those described in other publications<sup>2,4-6</sup>. This difference may be related to the fact that men in our 228 229 study did not have major alterations in conventional semen parameters. Moreover, traditional 230 sperm parameters are known to be highly variable between ejaculates, meaning intra-individual 231 variability. DNA fragmentation, on the other hand, appears to be more stable across ejaculates. 232 Traditional univariate analyses, particularly simple pairwise correlations, may have limitations in 233 this work, given this particular population, and may lack the power to capture more complex 234 associations. The contribution and integration of new methods from artificial intelligence may 235 provide an interesting way to better process available clinical and biological data.

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The fertility score previously developed and published by our team was built on anthropometric, metabolic, and antioxidative factors, parameters close to those studied in this work. As shown in figure 1, the result of the score was put into perspective with sperm parameters. Interestingly, this score is the only feature, along with HDL level, that correlates significantly with DNA fragmentation level. Notwithstanding controversial findings in the

literature, DNA fragmentation level is widely recognized as a marker of intrinsic semen quality.
Its levels seem to affect embryo quality, implantation and miscarriage risk <sup>26</sup>. Many factors can
increase the level of sperm DNA fragmentation, including obesity, high local temperature, drug
treatment, exposure to environmental pollutants, and smoking.

Using the strength of multivariate statistical modelling, the score may reveal underlying phenomena that are difficult to identify using conventional univariate methods alone. By correlating significantly with DNA fragmentation, the score may provide a more comprehensive way of assessing male semen quality, particularly in patients with normal or subnormal semen. This assumption must be demonstrated by further studies. Furthermore, from an analytical and technical point of view, DNA fragmentation is a more challenging and less standardized biomarker than other semen analysis parameters.

Although only men with no severe sperm alteration were recruited, a negative correlation between BMI, visceral fat and sperm motility was observed. Obesity in males has been found to be a risk factor for infertility and increased time to conception <sup>3</sup>. In men, many studies, supported by a meta-analysis <sup>27</sup>, have reported an alteration of sperm parameters in case of overweight and obesity, more specifically a decrease in sperm concentration and sperm count <sup>28–30</sup>. Sperm motility also appears to be affected by overweight or obesity <sup>31</sup>.

Dyslipidemia has been reported to negatively affect testicular and epididymal function, sperm maturation and quality, and ejaculatory function <sup>32</sup>. Consistent with these findings, in this population of patients with unexplained infertility, we observed an association between lipid levels and some semen parameters. Indeed, despite the absence of patients with dyslipidemia, unfavorable lipid composition (high total and LDL cholesterol, and/or low HDL) may be negatively associated with semen parameters. However, while blood glucose was a strongly

discriminating factor between fertile vs infertile couples (as studied previously <sup>10</sup>), it does not 265 266 appear to be correlated with sperm parameters in this population of infertile men. The molecular 267 mechanisms involved in this phenomenon are unclear, but hypotheses have been proposed. Impairment of steroidogenesis has been reported. Cholesterol may modulate the renin-268 269 angiotensin system in the testes, leading to inhibition of steroidogenesis. This in turn leads to a decrease in testosterone production <sup>33</sup>. Post-testicular sperm maturation may also be affected: 270 271 hypercholesterolemia would induce changes in the epididymal epithelium such as an 272 accumulation of cholesterol ester lipid droplets in the smooth muscle of the epididymal tract. 273 Epididymal peristalsis contractions would be impaired, compromising sperm progression inside the epididymal lumen, and thus the maturation process  $^{34}$ . Eventually, an increase of oxidative 274 275 stress and an excess of free radical production could also be the consequence of dyslipidemia<sup>35</sup>. 276 This last hypothesis is consistent with our findings. Indeed, in our cohort, the HDL level and thus the "good" cholesterol level was correlated with the DNA fragmentation level, directly related to 277 278 oxidative stress.

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281 Besides overweight and lipid and carbohydrate profile, some microelements and vitamins 282 were positively correlated with sperm count and motility. Vitamin B12 levels and glutathione 283 peroxidase activity (GPX) were correlated with sperm motility and total sperm count, respectively, in this cohort. A meta-analysis by Banihani <sup>36</sup> highlighted the importance of 284 vitamin B12 in sperm physiology and quality, particularly on total sperm count <sup>37</sup>. Indeed, lower 285 vitamin B12 levels have been reported in infertile men<sup>38</sup>. Several beneficial mechanisms of 286 287 vitamin B12 have been described, including its link to homocysteine metabolism and its antioxidant properties <sup>39</sup>. 288

Numerous studies have demonstrated the importance of the GPX during spermatogenesis and its relationship with male fertility <sup>40,41</sup>. Similarly, previous studies have reported higher GPX activity in seminal plasma in patients with better sperm quality <sup>42,43</sup>. GPX plays a key function as reactive oxygen species (ROS) scavengers for spermatozoa, to maintain the balance between ROS production and recycling <sup>44</sup>.

A limitation of this study was the lack of measurement of antioxidant status in seminal plasma for the entire cohort of patients. In addition, and in accordance with international recommendations <sup>45–47</sup>, exploratory andrological assessments (hormone tests, ultrasound, etc.) were not carried out in our patients.

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299 This study focused on a particular population of men from couples with unexplained 300 infertility, and a limited population size of 75 patients. However, measurement of sperm 301 parameters alone, which were normal or subnormal in this population, does not appear to be 302 sufficient to evaluate and stratify these patients. These findings suggest that despite a normal or 303 subnormal semen parameters, the metabolic and oxidative status factors considered in this study, 304 which are probably in part linked to the lifestyle and behavior of these patients, should be 305 integrated into a holistic and comprehensive diagnostic and therapeutic strategy. Unlike other 306 factors or causes of infertility (genetic or hormonal abnormalities, chemotherapy, etc.), in this 307 case patients can be involved in the management of their health, especially given the reversible 308 character of these indicators. Indeed, an improvement in metabolic and oxidative status could be associated with a reduced risk of infertility, and even a better chance of success when in vitro 309 310 fertilization (IVF) treatment is still required.

Improving lifestyle for these patients therefore appears to be essential, and even beyond reproductive health alone, for overall physical and mental well-being. Such improvement should be advised by healthcare professionals, using dietary and physical activity coaching or guidance properly designed and validated by international guidelines. Interventional studies are needed to confirm the positive impact of lifestyle improvement <sup>48</sup>.

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#### 318 Conclusion

Favorable anthropometric, metabolic and oxidative patterns, which may reflect an appropriate lifestyle, appear to positively impact overall health, in particular reproductive function. Despite normal or subnormal semen parameters, findings reported in this work using patients from a cohort of idiopathic infertile couples show that altered body composition, as well as unfavorable lipid balance and vitamin and microelement deficiency were associated with lower semen parameters.

This study, consistent with previous publications in this area, suggests that beyond semen quality parameters, in an essential assessment of male fertility, other key factors should be taken into account. A more comprehensive evaluation of the patients' health, affecting their nutritional, metabolic and oxidative status, seems to be an indispensable element to consider for a more holistic care. In this regard, the application of emerging artificial intelligence techniques may provide a unique opportunity to integrate all these parameters and deliver personalized care.

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#### 333 **Author contributions:**

- 334 GB participated in the study's conception and design, performed statistical analysis, and drafted the manuscript. RL
- 335 supervised the study, participated in patient recruitment, interpreted the data, and critically revised the manuscript for
- 336 intellectual content. CF was involved in study conception and design, patient recruitment, interpretation of data,
- 337 critical revision of the manuscript for intellectual content. SC and AL both worked on the study conception and
- 338 design and critically revised the manuscript for intellectual content. CD contributed to study conception and design,
- 339 patient recruitment, interpretation of data, and drafting of the manuscript.
- 340 The ALIFERT collaborative group members contributed to the study design and were involved in patient recruitment
- 341 or analysis. The final manuscript was approved by all authors.
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505 <u>Table 1</u>: Baseline characteristics: clinical, biological and sperm analysis. Data reported as mean 506  $\pm$  standard deviation. BMI : Body mass index ; LDL : Low-density lipoprotein ; HDL : High-

507 density lipoprotein

508 § : 4 patients with unknown systolic blood pressure (mmHg) and diastolic blood pressure (mmHg), 6 patients with unknown ascorbic acid

510

Parameters	n = 75			
Baseline, metabolic and biological parameters				
Age (years)	$33.6 \pm 5.3$			
BMI (kg/m <sup>2</sup> )	$26.1 \pm 4.3$			
Waist measurement (cm)	$92.6 \pm 11.3$			
Visceral fat (%)	$7.0 \pm 4.2$			
Systolic Blood pressure (mmHg) §	$126.8 \pm 12.2$			
Diastolic Blood pressure (mmHg) <sup>§</sup>	$80.8 \pm 10.0$			
Glycemia (mmol/L)	$5.0 \pm 0.7$			
Cholesterol (mmol/L)	$5.2 \pm 1.0$			
LDL (mmol/L)	$3.3 \pm 1.0$			
HDL (mmol/L)	1.3 ±0.4			
Triglyceride (mmol/L)	$1.4 \pm 0.9$			
Micronutrients and vitamins				
Vitamin D (ng/mL)	$23.2 \pm 11.3$			
Folate (nmol/L)	$12.7 \pm 7.2$			
Cobalamin (pmol/L)	$300.0 \pm 111.0$			
Retinol (µmol/L)	$2.1 \pm 0.5$			
Alpha-Tocopherol (µmol/L)	$25.0 \pm 6.1$			
Zinc (µmol/L)	$12.8 \pm 1.9$			
Selenium (mmol/L)	$1.2 \pm 0.2$			
Glutathione peroxidase (mUI/mL)	$391.6 \pm 61.0$			
Ascorbic acid (mg/mL) §	$42.3 \pm 18.8$			
Alpha-Carotene (µmol/L)	$0.1 \pm 0.1$			
Lycopene (µmol/L)	$0.6 \pm 0.3$			
Lutein (µmol/L)	$0.3 \pm 0.2$			
β-Cryptoxanthin (µmol/L)	$0.2 \pm 0.2$			
Glutathione (µmol/L)	$829.6 \pm 200.0$			
Beta-Carotene (µmol/L)	$0.4 \pm 0.3$			
Machine learning model score	0.5 ±0.1			

Semen quality parameters

Semen volume (ml)	$3.4 \pm 1.6$
Sperm concentration (10 <sup>6</sup> /ml)	$44.2 \pm 29.8$
Total sperm count (TSC) (10 <sup>6</sup> )	$143.7 \pm 109.4$
Progressive Motility a+b (%)	37.5 ±11.6
Vitality (%)	$63.4 \pm 15.4$
Normal morphology (%)	$20.7\pm9.9$
DNA fragmentation rate (%)	$28.6\pm18.4$

<u>Table 2:</u> Correlations (Pearson's parametric test) between the studied parameters and sperm quality. BMI : Body mass index ; LDL : Low-density lipoprotein ; HDL : High-density 

lipoprotein

- \*\* : P< 0.01 ; \*: P < 0.05 ; n.s., not significant.

Parameters	Semen volume (ml)	Sperm concentration (10 <sup>6</sup> /ml)	Total sperm count (TSC) (10 <sup>6</sup> )	Progressive Motility a+b (%)	Vitality (%)	Normal morphology (%)	DNA fragmentation rate (%)
	r (P value)	r (P value)	r (P value)	r (P value)	r (P value)	r (P value)	r (P value)
baseline, metabolic and biological parameters							
Age (years)	-0.142 (n.s.)	0.243 (n.s.)	0.199 (n.s.)	-0.066 (n.s.)	-0.119 (n.s.)	0.045 (n.s.)	0.107 (n.s.)
BMI (kg/m²)	-0.307 (*)	-0.08 (n.s.)	-0.136 (n.s.)	-0.233 (*)	-0.217 (n.s.)	-0.068 (n.s.)	0.078 (n.s.)
Waist measurement (cm)	-0.286 (*)	-0.102 (n.s.)	-0.157 (n.s.)	-0.191 (n.s.)	-0.169 (n.s.)	0.015 (n.s.)	0.152 (n.s.)
Visceral fat (%)	-0.221 (n.s.)	-0.06 (n.s.)	-0.062 (n.s.)	-0.27 (*)	-0.285 (**)	-0.067 (n.s.)	0.152 (n.s.)
Blood pressure Systolic(mmHg)	-0.194 (n.s.)	0.129 (n.s.)	0.053 (n.s.)	0.087 (n.s.)	-0.045 (n.s.)	0.073 (n.s.)	-0.12 (n.s.)
Blood pressure Diastolic(mmHg)	-0.23 (n.s.)	0.187 (n.s.)	0.078 (n.s.)	-0.011 (n.s.)	-0.107 (n.s.)	0 (n.s.)	-0.088 (n.s.)
Glycemia (mmol/L)	0.059 (n.s.)	0.127 (n.s.)	0.132 (n.s.)	-0.147 (n.s.)	-0.166 (n.s.)	0.018 (n.s.)	-0.064 (n.s.)
Cholesterol (mmol/L)	-0.189 (n.s.)	-0.155 (n.s.)	-0.272 (*)	-0.211 (n.s.)	-0.311 (**)	-0.117 (n.s.)	-0.034 (n.s.)
LDL (mmol/L)	-0.13 (n.s.)	-0.201 (n.s.)	-0.249 (*)	-0.208 (n.s.)	-0.326 (**)	-0.022 (n.s.)	0.053 (n.s.)
HDL (mmol/L)	0.031 (n.s.)	-0.045 (n.s.)	-0.054 (n.s.)	-0.003 (n.s.)	0.011 (n.s.)	-0.186 (n.s.)	-0.364 (**)
Triglycerides (mmol/L)	-0.152 (n.s.)	0.088 (n.s.)	-0.04 (n.s.)	-0.016 (n.s.)	-0.041 (n.s.)	-0.066 (n.s.)	0.1 (n.s.)
vitamins							
Vitamin D (ng/mL)	0.208 (n.s.)	-0.028 (n.s.)	0.095 (n.s.)	-0.077 (n.s.)	0.002 (n.s.)	0.12 (n.s.)	0.149 (n.s.)
Folate (nmol/L)	0.09 (n.s.)	-0.121 (n.s.)	0 (n.s.)	0.197 (n.s.)	0.027 (n.s.)	0.155 (n.s.)	0.161 (n.s.)
Cobalamin (pmol/L)	-0.173 (n.s.)	0.057 (n.s.)	-0.08 (n.s.)	0.275 (*)	-0.063 (n.s.)	0.106 (n.s.)	0.074 (n.s.)
Retinol (µmol/L)	-0.049 (n.s.)	-0.046 (n.s.)	-0.076 (n.s.)	0.025 (n.s.)	0.159 (n.s.)	-0.037 (n.s.)	0.126 (n.s.)
Alpha-Tocopherol (µmol/L)	-0.042 (n.s.)	-0.194 (n.s.)	-0.226 (n.s.)	0.05 (n.s.)	-0.035 (n.s.)	-0.105 (n.s.)	-0.043 (n.s.)
Zinc (µmol/L)	-0.039 (n.s.)	-0.001 (n.s.)	0.008 (n.s.)	-0.122 (n.s.)	0.124 (n.s.)	0.043 (n.s.)	0.027 (n.s.)
Selenium (mmol/L)	-0.031 (n.s.)	0.022 (n.s.)	0.015 (n.s.)	0.051 (n.s.)	-0.105 (n.s.)	0.03 (n.s.)	0.07 (n.s.)
Glutathione peroxidase (mUI/mL)	0.055 (n.s.)	0.204 (n.s.)	0.23 (*)	0.017 (n.s.)	-0.182 (n.s.)	-0.053 (n.s.)	0.002 (n.s.)
Ascorbic acid (mg/mL)	-0.114 (n.s.)	-0.077 (n.s.)	-0.145 (n.s.)	0.141 (n.s.)	0.067 (n.s.)	0.091 (n.s.)	0.129 (n.s.)
Alpha-Carotene (µmol/L)	-0.082 (n.s.)	-0.003 (n.s.)	-0.05 (n.s.)	-0.042 (n.s.)	0.041 (n.s.)	-0.05 (n.s.)	-0.041 (n.s.)
Lycopene (µmol/L)	0.13 (n.s.)	-0.111 (n.s.)	-0.074 (n.s.)	-0.032 (n.s.)	-0.221 (n.s.)	-0.117 (n.s.)	-0.077 (n.s.)
Lutein (µmol/L)	-0.086 (n.s.)	-0.148 (n.s.)	-0,.872 (n.s.)	0.149 (n.s.)	-0.05 (n.s.)	0.009 (n.s.)	0.073 (n.s.)
$\beta$ -Cryptoxanthin	-0.146 (n.s.)	-0.028 (n.s.)	-0.112 (n.s.)	0.069 (n.s.)	-0.018 (n.s.)	0.045 (n.s.)	-0.086 (n.s.)

Glutathione (µmol/L)	0.137 (n.s.)	-0.041 (n.s.)	0.067 (n.s.)	-0.003 (n.s.)	-0.078 (n.s.)	-0.035 (n.s.)	0.135 (n.s.)
Beta-Carotene (µmol/L)	0.022 (n.s.)	-0.078 (n.s.)	-0.111 (n.s.)	-0.104 (n.s.)	-0.14 (n.s.)	0.044 (n.s.)	-0.059 (n.s.)
Machine learning model score	0.091 (n.s.)	0.008 (n.s.)	-0.008 (n.s.)	0.132 (n.s.)	0.167 (n.s.)	-0.06 (n.s.)	-0.263 (*)
520							



525 Figure 1: A complex relationship between lifestyle, diet, antioxidants and sperm DNA

# 526 <u>fragmentation.</u>

527 An unbalanced lifestyle and diet lead to anthropometric, metabolic and nutritional changes, 528 which can lead to an imbalance in oxidative stress. These consequences are multiple and 529 interrelated, and the use of artificial intelligence represents a unique opportunity to provide a 530 more holistic evaluation of the influence of the considered marker.

531 This oxidative stress could be one of the main drivers leading to a decline in male reproductive

532 functions and consequently in sperm parameters, in particular sperm DNA fragmentation.

533

