



**HAL**  
open science

## Presence of vitamin B<sub>12</sub> metabolism in the last common ancestor of land plants

Richard G Dorrell, Charlotte Nef, Setsen Altan-Ochir, Chris Bowler, Alison G Smith

### ► To cite this version:

Richard G Dorrell, Charlotte Nef, Setsen Altan-Ochir, Chris Bowler, Alison G Smith. Presence of vitamin B<sub>12</sub> metabolism in the last common ancestor of land plants. 2023. hal-04237983v1

**HAL Id: hal-04237983**

**<https://hal.science/hal-04237983v1>**

Preprint submitted on 11 Oct 2023 (v1), last revised 12 Apr 2024 (v2)

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

## Presence of vitamin B<sub>12</sub> metabolism in the last common ancestor of land plants

Richard G. Dorrell<sup>1,2</sup>, Charlotte Nef<sup>2</sup>, Setsen Altan-Ochir<sup>2</sup>, Chris Bowler<sup>2</sup>, and Alison G. Smith<sup>1</sup>

<sup>1</sup> Department of Plant Sciences, University of Cambridge, CB2 1ST, United Kingdom

<sup>2</sup> Institut de Biologie de l'ENS (IBENS), Département de Biologie, École Normale Supérieure, CNRS, INSERM, Université PSL, 75005 Paris, France

\*Corresponding authors: [richard.dorrell.algae@gmail.com](mailto:richard.dorrell.algae@gmail.com) , [as25@cam.ac.uk](mailto:as25@cam.ac.uk)

**Keywords:** bryophytes, Anthocerotophyta, Marchantiophyta, OneKp, *Chlamydomonas*, phylogenomics

**Vitamin B<sub>12</sub>, or cobalamin, (hereinafter B<sub>12</sub>) is an essential organic micronutrient, required by humans as a cofactor for methionine synthase (METH) and for methylmalonyl CoA mutase (MCM), involved in the propionate shunt. B<sub>12</sub> is a complex corrinoid molecule made only by a subset of bacteria. Plants and fungi have an alternative methionine synthase (METE) that does not need a B<sub>12</sub> cofactor, so these organisms are typically considered to neither synthesise nor utilise B<sub>12</sub>. In contrast many algal species utilise B<sub>12</sub> if it is available, because they encode both METE and METH. Moreover, a large proportion of algal species encode METH only, and so are like animals in being dependent on an external source of the vitamin. Here, we performed a detailed phylogenetic analysis of the distribution of METE, METH and eleven further proteins implicated in B<sub>12</sub> metabolism in eukaryotic cells across an exhaustive library of over 1,500 plant and algal genomes and transcriptomes. The results reveal the hitherto undetected existence of B<sub>12</sub>-associated metabolism deep into the streptophytes. The B<sub>12</sub>-dependent synthase METH, and the accessory proteins MTRR, CblB, CblC, CblD and CblJ were detected in the basally divergent plant lineage of hornworts, and CblB and CblJ were further identified in liverworts. Using phylogenetic and PFAM analysis we demonstrate this is due to retention of ancestral B<sub>12</sub>-metabolism pathways in the last common ancestor of land plants, followed by at least two independent complete losses in mosses and vascular plants. We further show more limited distributions of genes encoding B<sub>12</sub>-related proteins across the algal tree of life, including MCM and type II ribonucleotide reductase, alongside an obligate B<sub>12</sub>-dependency across several major marine algal orders. Finally, by considering the functional biology of early-diverging land plants, together with the collection sites of ten further algal species inferred to have lost B<sub>12</sub>-dependent metabolism, we propose freshwater-to-land transitions and symbiotic associations to have been major constraining factors in B<sub>12</sub> availability in early plant evolution.**

### Introduction

Vitamins are organic micronutrients that are taken up by organisms from the environment and are essential for central metabolic processes [1, 2]. In particular, B-vitamins are the precursors to enzyme cofactors, providing additional chemical reactivity to that of amino acids. Plants, fungi and microorganisms also require these

compounds for their metabolism, but are able to synthesise them *de novo*. The exception is for vitamin B<sub>12</sub>, a cobalt-containing corrinoid molecule that is not synthesised by any eukaryote [3, 4]. In humans, B<sub>12</sub> is essential for two enzymatic activities: B<sub>12</sub>-dependent methionine synthase (METH) in the C1 cycle [5, 6] and methylmalonyl-coA mutase (MCM), responsible for the metabolism of odd-chain fatty acids and branched-chain amino acids via the propionate shunt [7, 8]. B<sub>12</sub> deficiency and its associated pathologies (pernicious anaemia, methylmalonic acidemia) may be chronic in subsistence economies, with particularly adverse impacts on child development and during pregnancy [9-11]. Addressing how to mitigate B<sub>12</sub> deficiency in plant-based diets, which may have otherwise substantially lower environmental impacts than omnivorous nutrition, is therefore a key challenge to sustainably feeding a growing planetary population [12-14].

Historically, B<sub>12</sub> in the human diet has been obtained from different sources. These include from ruminant animals and their derivatives (i.e., dairy products) [15], the direct consumption of B<sub>12</sub> from soil particles (e.g., via geophagia [16], from fortified (nutritional yeast) and from edible seaweeds from across the algal tree of life [15, 17]. Indeed, many microalgae, microscopic photosynthetic eukaryotes, also encode METH and/or MCM and many are obligately dependent on B<sub>12</sub> [18, 19]. In one study for example, 171/326 sampled algal species were found to be unable to grow in the absence of supplemented B<sub>12</sub> [18]. B<sub>12</sub> auxotrophy has been subsequently documented to be widespread in specific algal phyla (e.g., haptophytes, dinoflagellates) across the Tree of Life [20, 21]. Some microalgal lineages may encode further B<sub>12</sub>-dependent enzymes (e.g. form II ribonucleotide reductase (RNR<sub>II</sub>), first documented in the green microalga *Euglena*) [7], which is not found in humans.

B<sub>12</sub> biosynthesis involves over twenty discrete enzymatic steps from the common tetrapyrrole intermediate uroporphyrinogen III [22, 23]. To date, complete B<sub>12</sub> biosynthesis has only been described in a subset of bacteria and archaea [22, 24]. Algae may acquire synthesised B<sub>12</sub> by scavenging from the environment [25], phagotrophic consumption of B<sub>12</sub>-containing organisms [26] or via symbiotic exchanges with B<sub>12</sub>-producing commensals [18]. Additionally, there are a number of B<sub>12</sub> variants with different axial ligands, which affects the bioavailability of the vitamin. Intrinsic factor, the B<sub>12</sub>-binding protein found in the human ileum, has a much higher affinity for cobalamin, where the lower axial ligand is 5,6-dimethylbenzimidazole (DMB), than for pseudocobalamin, which has adenine as the lower axial ligand [27]. Eukaryotic microalgae have a similar preference for cobalamin over pseudocobalamin [27], although a B<sub>12</sub> remodelling pathway has been documented in some species [27, 28], which enables them to convert pseudocobalamin to cobalamin if supplied with DMB.

Little is known of the uptake, cellular transport or metabolism of B<sub>12</sub> in algae, but the pathways are well documented in humans. Following its endocytic internalisation into the cell, cobalamin is released into the cytosol via the co-operative activity of a cytosolic chaperone CblC and the lysosomal transporters CblF/ CblJ [29] (**Fig. 1**). This involves the reductive removal of any preceding upper-axial ligands associated with the cob(III)-alamin and conversion into cob(I)- and cob(II)alamins complexed with CblC [29]. Cob(I,II)-alamins complexed with CblC can then be transferred to the conjugate protein CblD, which subsequently directs the synthesis of different cobalamin-dependent

enzymes [29, 30]. METH requires methyl-cobalamin, produced from CblD-associated B<sub>12</sub> by the activity of methionine synthase reductase (MTRR) [5] in the cytosol, whereas synthesis of MCM involves adenosyl-cobalamin, produced in the mitochondria via the conjugate activity of the probable B<sub>12</sub> transporter protein CblA [31] and the adenosyl-transferase CblB [7]. A small number of additional proteins (e.g., CblX, epi-CblC) have been shown to have epistatic effects on cobalamin uptake in humans, but are of unknown function [29]. Finally, in the algal species in which it occurs, RNR-II is cytosolic and utilises adenosyl-cobalamin.

B<sub>12</sub> produced by bacteria may be scarce in the environment, including in soil [32] and large portions of the contemporary ocean [33, 34], and uptake and processing of the vitamin is energetically costly [29, 35]. In contrast to the widespread utilisation of B<sub>12</sub> by aquatic algae, several ecologically successful groups of eukaryotes, including all previously documented land plants, have lost their metabolic dependence on B<sub>12</sub> [36]. This is possible because of a B<sub>12</sub>-independent methionine synthase (METE) that arose separately, which can replace METH, albeit with a lower catalytic activity [37]. The METE gene shows a discontinuous presence across the tree of life, and is also present in many METH-containing species, including a substantial proportion of eukaryotic algae, which renders them facultative B<sub>12</sub> users [36]. The most parsimonious explanation is an ancestral presence of both METE and METH genes in eukaryotes, with the repeated loss of METE from obligately B<sub>12</sub>-dependent species, and the occasional loss of METH from plants, fungi, and other species that do not have B<sub>12</sub>-associated metabolism.

In the last five years, the dramatic expansion in plant and algal genomic and transcriptomic resources, such as through the OneKp project [38, 39], have provided unprecedented insights into the foundational events underpinning plant evolution. This includes evidence for the stepwise accumulation of evolutionary innovations (e.g., homeobox domain transcription factors, auxin transport pathways and water-stress tolerance factors) associated with the colonisation of land in the closest streptophyte relatives of land plants [40-42], and the importance of gene losses as well as gene family expansions for the post-terrestrial diversification of both bryophyte (mosses, liverworts, and hornworts) and vascular plants [43, 44]. These expanded genomic resources prompted us to revise our current understanding of the distribution of B<sub>12</sub>-dependent metabolism across the photosynthetic eukaryotic tree of life. Here we investigate the present of B<sub>12</sub> uptake and associated metabolism in over 1,600 published plant and algal genome and transcriptome libraries, and consider in which ecological contexts loss of B<sub>12</sub>-dependent metabolism has occurred.

## Results

### *Distribution of METH/ METE genes indicate B<sub>12</sub> presence in hornworts*

We identified homologues of the thirteen B<sub>12</sub>-associated proteins, illustrated in **Fig. 1**, from the algae *Chlamydomonas reinhardtii*, *Euglena gracilis* and *Phaeodactylum tricorutum* and from humans (Table 1), and used these to search a composite library of 1,666 plant and algal genomes and transcriptomes. The library contained: 44 algal and plant genomes, including the recently published draft genomes of the hornworts *Anthoceros agrestis* and *A. punctigera* [44, 45]; decontaminated versions of the 1,000 plant transcriptomes (OneKp ; 1,292 libraries) [38, 46] and the Marine Microbial

Eukaryote Transcriptome Sequencing Project (MMETSP; 300 libraries) [47, 48]; and 26 further eukaryotic transcriptomes, in particular sampling ecologically abundant marine diatom genera [49] and freshwater chrysophyte lineages [50] (**Table 2**). Homologues were retrieved by reciprocal BLAST best-hit (RbH), single-gene RAxML trees, and PFAM domain analysis (Materials and Methods) [51-53], and were categorised taxonomically following recently published multigene phylogenies of plant and algal diversity [38, 49]. Full outputs are provided in **Dataset S1**.

The phylogenetic distributions of putative homologues are shown schematically in **Fig. 2** for each protein. The patterns underline the widespread occurrence of the B<sub>12</sub>-dependent form of methionine synthase, METH, across eukaryotic algae, found in all major marine groups, and in both early-branching (Klebsormidiaceae, Mesostigmatophyceae, Chlorokybophyceae) and close relatives of land plants (Zygnematophyceae, Charophyceae, Coleochaetophyceae) within the streptophyte lineage [38, 54]. METH was further detected in multiple distantly related hornwort genera, suggesting widespread conservation of B<sub>12</sub>-dependent methionine synthases across the class Anthocerotophyta (**Dataset S1**, sheet 5) [38, 55]. These included OneKp homologues from the hornwort genera *Megaceros* (*M. tosanus*, OneKp transcript- UCRN-2004435; *M. vincentianus*- TCBC-2004163), *Nothoceros* (*N. aenigmaticus* DXOU-2038410), *Paraphymatoceros* (*Paraphymato. hallii*: FAJB-2057847) and *Phaeoceros* (*Phaeo. carolinianus*, RXRQ-2022853, RXRQ-2022854), as well as probable METH genes from the *Anthoceros agrestis* Bonn (geneID: Sc2ySwM\_228\_5027); *A. agrestis* Oxford (geneID: utg000003l\_252) and *A. punctigera* (geneID: utg000098l\_165) genomes. METH was not found elsewhere within the land plant lineage, except for one potential METH homologue identified by RbH in the liverwort *Blasia sp.* (OneKp transcript: AEXY-2015053; **Dataset S1**, sheet 5), but that resolved during preliminary phylogenetic analyses with bacterial sequences and showed high sequence similarity with a *Mucilaginibacter* METH (NCBIId: WP\_129570389.1; >96% by BLASTp) [56]. Previous studies have documented high levels of bacterial contamination in the *Blasia* transcriptome, [46], and we therefore consider that this is unlikely to correspond to a liverwort METH.

The phylogeny of METH sequences robustly resolved a position for the hornwort sequences within the streptophytes (RAxML bootstrap support : 100%), in a weakly supported sister-group position to homologues from the charophytes / coleochaetales (*Chara vulgaris*, *Chaetosphaeridium globosum*, *Coleochaete scutata*, *Col. irregularis*) (**Fig. 3 ; Dataset S1**). This group was positioned more deeply within Viridiplantae sequences, as a sister to equivalent sequences from Chlorophytes (RAxML bootstrap support: 100% ; **Fig. 3**). Each of the phylogenetically validated *Megaceros*, *Nothoceros*, *Phaeoceros* and *Paraphymatoceros* hornwort OneKp sequences possessed all five PFAMs typically associated with METH (homocysteine S-methyltransferase, PF02574 ; pterin-binding, PF00809 ; B<sub>12</sub>-binding, PF02607 and PF02310 ; and the activation domain, PF02965) with similar e-values to functionally characterised equivalents from algae (**Fig. S1**) [21]. The three identified *Anthoceros* homologues possessed all PFAMs apart from the first B<sub>12</sub>-binding (PF02607) domain. However, this PFAM is detected in alternative gene models in each genome (*A. agrestis* Bonn: 362\_443; *A. agrestis* Oxford: utg000003l\_664; *A. punctigera*, utg000145l\_284), suggesting that *Anthoceros* is likely to also possess a functionally active METH. The hornwort sequences all had conserved

active sites (e.g., the substrate-binding pocket of the pterin-binding site) associated with METH activity (**Fig. S1**) [21, 57].

We also considered the distribution of the methionine synthase reductase (MTRR) necessary for METH activity in our dataset. Well-conserved homologues of MTRR were detected across hornwort genomes and transcriptomes, but were absent from other plant groups (**Fig. 2**). As per the situation for METH, the hornwort MTRR genes grouped with other streptophyte sequences within the Viridiplantae (**Fig. 3**), and had well-conserved PFAM domains (flavodoxin 1- PF00258, FAD-binding- PF00667, oxidoreductase- PF00175; **Fig. S1**), suggesting vertical origin and functionality. We therefore conclude that the hornwort METH sequences are likely to be catalytically functional, and vertically inherited from the streptophyte ancestor of terrestrial plants.

#### *Widespread occurrence of two distinct METE isoforms in plants*

All plant libraries studied possessed *METE* genes, including hornworts, indicating that both genes were present in the last common plant ancestor (**Fig. 2**). The hornwort *METE* genes resolved with other plant homologues (**Fig. 3**) and possessed well-conserved N-terminal (PF08267) and C-terminal catalytic (PF01717) PFAM domains (**Fig. S1**), indicating their likewise probable vertical origin and functionality.

The plant *METE* enzymes resolved phylogenetically in two discrete families: a conventional isoform with 1,768 recovered examples (labelled “Clade 1”); and a less frequently observed isoform (“Clade II”) with 125 observed examples (**Fig. 3**; **Dataset S1**). Both isoforms contained credible occurrences of both Meth\_synt\_1 (PF08267) and Meth\_synt\_2 (PF01717) domains, and thus are likely to function as methionine synthases (**Fig. S1**; **Dataset S1**). Both Clade I and Clade II isoforms were furthermore detected in higher plant, moss, liverwort, hornwort and streptophyte sequences (**Fig. 3**), suggesting their presence in the last plant common ancestor. All three characterised *Arabidopsis* methionine synthases (ATMS1- At5g17920; ATMS2 At3g03780; ATMS3- At5g20980) resolved within Clade 1, but the Clade II enzymes all showed greater proximity by BLASTp analysis to these than to the four closely related *Arabidopsis* adenosyl-methionine synthases (SAM1- At1g02500; SAM2- At4g01850; SAM3- At3g17390; and SAM4- At2g36880), implying that they likewise correspond to probable methionine synthase isoforms (**Dataset S1**, sheet 6) [58].

Both Clade I and Clade II isoforms distantly positioned in the phylogeny to chlorophyte *METE* sequences, which formed a sister-group to red algae, cryptomonads and chromerids (**Fig. 3**). The most parsimonious explanation for this distribution would be the ancestral replacement of the streptophyte *METE* sequence, still retained in chlorophytes, with the Clade I and Clade II isoforms, preceding the loss of METH within specific plant groups.

#### *Distribution of METE genes confirms widespread B<sub>12</sub> auxotrophy in algae*

In contrast to the widespread distribution of both *METE* isoforms in land plants, *METE* was absent from many of the algal libraries searched (**Fig. 2**). Transcriptome libraries may under-report the presence of *METE*, as it may be transcriptionally repressed in B<sub>12</sub>-supplemented cultures (most common algal growth media include

exogenous B<sub>12</sub>, either in the form of soluble vitamins or soil extract) [37, 59], and therefore we limit our consideration to lineages with at least one sequenced genome included [39]. Consistent with previous studies [20], we project B<sub>12</sub> auxotrophy in all haptophytes (including three genomes included in the study : *Emiliania huxleyi*, *Chrysochromulina tobinii*, and *Pavlova* sp. CCMP2436) [60]. Only one potential haptophyte METE homologue (from *Pavlova lutheri* UTEX LB1293) was found by RbH, but was excluded as a probable green algal contaminant (**Dataset S1**). No occurrences of METE were found in the prasinophyte class Mamiellophyceae (including the *Micromonas* and *Ostreococcus* sp. genomes) or the ochrophyte class Pelagophyceae (including two genomes : *Aureococcus anophagefferens*, and Pelagophyceae sp. CCMP2097) [60] (**Fig. 2**). The shared absence of METE from haptophytes, pelagophytes and Mamiellophyceae is particularly interesting given their environmental abundance as assessed e.g., via the *Tara* Oceans expedition [61, 62], underlining the importance of B<sub>12</sub> acquisition for marine photosynthesis.

Further instances of obligate B<sub>12</sub> requirements, due to the complete absence of METE, were found in less ecologically abundant algal groups and at more phylogenetically localised scales, including most immediate relatives of the model green algal species *C. reinhardtii* [63, 64] and the glaucophytes and Palmophyllphyceae, respectively considered as basally-divergent members of primary chloroplast-containing algae and the Viridiplantae [65, 66]. Finally, dinoflagellates (including three genomes from *Symbiodinium* sp.) were found to possess only partial METE sequences, which lacked the conventional N-terminal PFAM domain (MetH\_synt\_1 ; PF08267). The function of this partial METE sequence isoform awaits experimental validation, although B<sub>12</sub> dependency has been proposed to be widespread across the dinoflagellates [21, 67] (**Dataset S1**). We did, however, identify complete METE genes containing both PFAM domains in the chromerid algae *Chromera velia* and *Vitrella brassicaformis*, which are the closest obligately photosynthetic relatives of dinoflagellates within the alveolates [68]. The dinoflagellate METE-like genes were distantly related to all other equivalents and instead resolved with Archaea (**Fig. 3**), which likewise lacked the N-terminal PFAM domain. This may suggest a horizontal acquisition of the current dinoflagellate METE-like gene that may have been accompanied by the loss of the complete METE isoform.

Finally, the METE genes of the red algal genera *Porphyridium* and *Cyanidioschyzon*, as well as in the chrysophyte *Poteriospumella*, resolved with homologues from bacteria (**Dataset S1 ; Fig. 3**), independent to all other plant and algal proteins. In each case, these acquisitions were shared between multiple related species or strains, and are therefore likely to be genuine bacterial horizontal gene transfers as opposed to contaminants. Overall, these data indicate complex patterns of METE gene transfer and loss across the photosynthetic eukaryote tree of life.

#### *MCM and RNR II enzymes are restricted to individual algal lineages*

Alongside methionine synthase, we considered the distributions of the B<sub>12</sub>-dependent MCM and RNR II enzymes across our dataset (**Fig. 2**). Neither enzyme was detected in hornworts or more broadly across streptophytes, and were absent from the chlorophytes, except for some sporadic occurrences that are likely consistent with horizontal acquisition as opposed to an ancestral presence in Viridiplantae (discussed

below). Homologues of MCM were primarily detected in eukaryotic algae with secondary red chloroplasts (e.g., diatoms, cryptomonads, haptophytes, ochrophytes), where they typically possessed well-conserved PFAM domains, and in many cases predictable mitochondrial targeting sequences consistent with mitochondrial function (**Dataset S1 ; Fig. S2**). MCM is implicated in the catabolism of branched-chain amino acids in diatoms under nitrate exhaustion [69, 70], and has further been proposed to participate in the dissipation of excess diatom plastidial reducing potential [71]. We note that a previous RNAi study of *Phaeodactylum* MCM failed to identify a clear mutant phenotype [70], so it remains to be determined to what extent MCM activity is essential to algae with secondary red chloroplasts, and therefore whether it provides a reason alongside METH for widespread B<sub>12</sub> dependence across these groups in the environment.

We additionally detected MCM in a small number of glaucophytes, red algae (*Galdieria sulphuraria*) and green algae within the genera *Tetraselmis* and *Chlorella* (**Fig. 2**). In contrast to green algal methionine synthase-associated enzymes, whose phylogenies show typically vertical inheritances, the MCM phylogeny retrieve a sister-group relationship between green algae and haptophytes (**Fig. 3**), implying a probable ancestral loss of the propionate shunt within the Viridiplantae, and subsequent re-acquisition by horizontal gene transfer in specific lineages.

Similarly, the B<sub>12</sub>-dependent type II ribonucleotide reductase (RNR II), first detected in eukaryotes in the secondary green chloroplast-containing alga *Euglena gracilis* [36, 72] was also found in the distantly related haptophytes and chlorarachniophytes, which have secondary green chloroplasts, reflecting possible ancestral horizontal gene transfers inferred between all three lineages [73]. Intriguingly it is also present in the Chlamydomonadales (although not *C. reinhardtii*), where it has most likely been acquired by an independent horizontal gene transfer from bacteria (**Figs. 2, 3**).

#### *Distribution of uptake proteins suggests deeper retention of B<sub>12</sub>-dependent metabolism within the bryophytes*

We further considered the distribution of six proteins associated with the uptake and intracellular transport of B<sub>12</sub> (CblA, B, C, D, F, and J) and two known epistatic regulators in humans (CblX, epi-CblC) [29] across our dataset. Consistent with the distribution of METH and MTRR, we could identify widespread presence of CblC, D and J proteins in hornwort libraries (**Fig. 2**). The hornwort sequences resolved within clades of other Viridiplantae sequences, and typically contained well-conserved PFAM domains (**Fig. 4 ; Fig. S1**), suggesting vertical origin and conserved function. The exact location of these enzymes, and precise cellular trafficking pathway in which they may be involved remain to be determined experimentally, as very few carried credible signal peptides that might suggest targeting to the endomembrane system, and participation in a B<sub>12</sub> endosomal uptake pathway (**Fig. S2**).

The only protein for which we could identify consistent endomembrane and/ or vacuolar targeting predictions amongst our recovered homologues was for CblF (**Fig. S2**), but this was almost exclusively detected amongst dinoflagellates in our dataset (**Figs. 2, 4**). We therefore tentatively propose that the B<sub>12</sub> uptake and intracellular

trafficking pathways may not be identical across different branches of the eukaryotic tree of life. We could detect potential homologues of both CblX and epi-CblC in hornworts, but found these also in vascular plants and with no clear underlying phylogenetic signal, so we are not confident of the function of these enzymes in B<sub>12</sub>-associated hornwort metabolism (**Figs. 2, 4**).

Surprisingly, homologues of CblB, an enzyme responsible for the synthesis of adenosyl-cobalamin [7], were found in hornworts (**Figs. 2, S1**). This was despite the absence of MCM, which typically uses adenosyl-cobalamin, alongside the CblB partner enzyme CblA from the streptophytes (**Fig. 2**) [31]. We could further detect homologues of CblB and indeed CblJ in liverworts, and in both cases the homologues from these lineages grouped monophyletically with hornworts (**Fig. 4**) and possess well-conserved PFAM domains (**Fig. S1**). It is therefore possible that some form of adenosyl-cobalamin dependent metabolism persisted in the last common plant ancestor, and that this metabolism is retained not only by hornworts but by liverworts within the bryophyte lineage. The validation of this pathway will depend on the functional characterisation of putative adenosyl-cobalamin dependent enzymes, e.g., from published liverwort and hornwort genomes [44, 74].

#### *Rare occurrences of B<sub>12</sub>-independent algae are biased towards terrestrial and symbiotic species*

Finally, we considered across our global datasets which photosynthetic eukaryotes, other than plants, may have secondarily lost B<sub>12</sub>-dependent metabolism completely. Considering B<sub>12</sub> independence as the presence of METE but absence of METH/ MTRR, RNR II or MCM activities, we identify seven phylogenetically validated occurrences of the loss of B<sub>12</sub>-associated metabolism (**Table 2**). For four species, the same distributions could be identified from PFAM domains only. Finally, we identified four species that possess METE and either MCM or RNR II, but lack METH/MTRR activities, suggesting a loss of B<sub>12</sub>-related methionine synthesis (**Table 2**).

The widespread but seemingly random loss of METE led us to consider what environmental factors might be involved in driving the distribution of this trait, which might be considered *a priori* deleterious. An interactive map of all collection sites reported for our dataset is directly available at <https://tinyurl.com/3esfpkmv>. The B<sub>12</sub>-independent species in our dataset (blue points on interactive map, and listed in **Table 2**) show characteristic distributions considering their collection sites, mainly originating from freshwater (e.g., *Leptosira obovata*, *Porphyridium cruentum*, *Poteriospumella* JBC07) or terrestrial habitats (*Coccomyxa pringsheimii*, *Cyanidioschyzon merolae*, *Galdieria sulphuraria*). B<sub>12</sub>-producing bacteria are abundant in some freshwater habitats (e.g., in sediments, as fish gut commensals) but appear to be present in much lower quantities in the water column (e.g., 10<sup>3</sup>-fold less than in sediments), calling into question the bioavailability of B<sub>12</sub> to freshwater algae [75, 76]. It is possible in some habitats that this is due to cobalt scarcity [77], but is not universally the case. We similarly note that *Geminigera cryophila*, which not only lacks METH but is also the only cryptomonad within our dataset to encode METE, was collected from Antarctic water masses, which have been previously shown to be characterised by vitamin B<sub>12</sub>-limitation and co-limitation (**Fig. 1**) [78, 79]. Finally, many of the B<sub>12</sub>-independent algal species are endobiotic to other organisms (*Leptosira obovata* and two members of the

order Trentepohliales, isolated from lichens [80]; *Vitrella brassicaformis*, a coral symbiont [68]; and *Brandtodinium nutriculum*, a foraminiferan symbiont [81]), and may be subject to chronic B<sub>12</sub> limitation in their native habitats, or even potentially receive methionine and other key metabolites such as folate directly from their hosts.

## Discussion

In this study, we query the distribution of thirteen B<sub>12</sub>-associated proteins across more than 1,600 genomes and transcriptomes from plastid-containing eukaryotes (plants and eukaryotic algae), in particular profiting from the substantial advance in plant genomics made through the OneKp transcriptome project [38] (**Fig. 1, Table 2**). Our data suggest the hitherto undocumented retention of vertically inherited B<sub>12</sub>-associated methionine synthesis pathways (METH, MTRR **Figs. 2, 3**) and B<sub>12</sub> uptake-associated proteins (CblB, C D, J ; **Figs. 2, 4**) in hornworts, and the further retention of potential CblB and CblJ homologues in liverworts (**Fig. 4**). Formally, the active use of B<sub>12</sub> by hornworts awaits functional characterisation. This may be best performed by bioassay of B<sub>12</sub> content in hornwort tissue [23], but might also come from the experimental inference of B<sub>12</sub> use, via B<sub>12</sub>-dependent downregulation of hornwort METE expression [37], or the uptake of fluorescently-labelled B<sub>12</sub> analogues [82] by hornwort cells.

Considering the predicted functionality and vertical origin of the hornwort homologues, our data indicate the probable presence of B<sub>12</sub>-dependent metabolism in the last common plant ancestor following the transition to land, with subsequent losses in mosses, vascular plants, and (dependent on the functions of the detected CblB and CblJ homologues), liverworts (**Fig. 5**). It remains to be determined at what point B<sub>12</sub>-dependent metabolism was lost during land plant evolution. Some phylogenetic studies have proposed hornworts as the earliest-diverging land plant group [83, 84], which would imply a single loss of METH in a common ancestor of liverworts, mosses and vascular plants. Other studies, including the multigene phylogenomic analyses performed as part of the *Anthoceros* genome and OneKp transcriptome projects [38, 43, 44], recover well-supported monophyly of bryophytes, to the exclusion of vascular plants (**Fig. 5**). This topology would indicate independent losses of B<sub>12</sub>-dependent metabolism in vascular plants and mosses, with a potential third loss of use of methylcobalamin, but retention of MCM and its requirement for adenosyl-cobalamin, in liverworts (**Fig. 5**). In either case, the presence of a metabolic activity in hornworts previously considered unique to algae further blurs the biological distinctions between aquatic streptophytes and plants. This is alongside the retention of biophysical carbon concentrating mechanisms in hornworts [55, 85], and the preceding evolution of plant-like auxin transporters and homeobox transcription factors in charophytes [41, 42].

Finally, it remains to be determined why B<sub>12</sub>-dependent metabolism was lost in early plant lineages. Many hornwort species are characterised by the presence of cyanobacterial symbionts, which might produce pseudocobalamin; this is not bioavailable to microalgal species [27, 86]. It remains to be determined whether hornworts can assimilate pseudocobalamin and/ or remodel it into cobalamin, and if this is more bioavailable to them than other plant lineages from which METH has been lost. Or indeed if the plant microbiome has played a broader role in the loss of cobalamin-dependent metabolism. Considering the widespread distribution of B<sub>12</sub>-

independent metabolism across the algal tree of life (**Table 2**), we note repeated losses of METH and its associated enzymes in other freshwater, terrestrial and symbiotic lineages. Thus comparative genomic and physiological studies of these species may elucidate the ecological reasons for the loss of B<sub>12</sub>-associated metabolism both in algae and land plants. Understanding why plants do not utilise B<sub>12</sub>-dependent enzymes is particularly important due to the prevalence of vitamin B<sub>12</sub> insufficiency/deficiency in populations consuming plant-based diets [9]. Ultimately, understanding the significance of B<sub>12</sub> utilisation and acquisition across algal biology may facilitate the synthetic reintroduction of B<sub>12</sub> uptake into transformable crop species, or the mass cultivation of algae for dietary consumption [82, 87], sustainably feeding the human planetary population [13].

## Materials and Methods

**Homologue detection** : Potential homologues of thirteen Vitamin B<sub>12</sub>-associated enzymes (*Chlamydomonas* METE with GBID : AAC49178.1, METH : XP\_042923308.1; human CblA : NP\_001362573.1, CblB : NP\_443077.1, CblC : NP\_056321.2, CblD : NP\_056517.1, CblF : NP\_060838.3, CblJ : NP\_001340531.1, CblX : NP\_005325.2, epi-CblC : NP\_001189360.1, MTRR : AAF16876.1 ; *Phaeodactylum* MCM : XP\_002179504.1 ; *Euglena* RNR II : Q2PDF6.1) [36] were searched across a composite library of 1,663 non-redundant plant and algal genomes and transcriptomes (**Table 2**) [38, 47, 49, 50] by BLASTp with threshold e-value 10<sup>-05</sup> [88]. Potential matching sequences were extracted and searched by BLASTp against the complete *Arabidopsis thaliana* genome [89], which uniquely encodes METE, supplemented with the query sequences defined above. Sequences that retrieved a best-scoring match to a query sequence were retained for downstream phylogenetic analysis (**Dataset S1**). For *METE* which is retained in b-plants, genes that retrieved one of the three *Arabidopsis* methionine synthases (ATMS1- At5g17920; ATMS2 At3g03780; ATMS3- At5g20980) were likewise retained for downstream analysis [58, 90].

**Phylogeny** : Inferred homologues were aligned against the query sequence, and best-scoring homologues obtained from parallel BLASTp searches of 51 combined genome and transcriptome libraries corresponding to different prokaryotic and non-photosynthetic eukaryotic taxonomic groups from across the tree of life [49] by MAFFT v 7.487 using the -gt auto setting [91]. The resulting alignments were imported into GeneIOUS v 10.0.9 and initially screened using the in-built NJ tree function with 100 replicates and random starting seeds; and highly divergent branches (defined visually as branches with > 1.0 calculated substitutions/ site) were iteratively removed [92]. The curated alignment was then trimmed with trimal v 1.4 using the -gt 0.5 setting, prior to a second round of tree building and manual curation [93]. Curated trimmed alignments were then inspected with RAxML v 8.0 using the PROTGAMMAJTT substitution matrix and 350 bootstrap replicates, following the methodology of previous studies [49]; and the best-scoring tree was then inspected for branches of contaminant origin (defined as sequences from one single library that resolved within or as a sister-group to a distantly related clade, e.g. algal sequences that resolved with bacterial homologues) [51]. The full length sequences of homologues that passed the initial RAxML phylogenetic curation were finally passed through a second iteration of mafft alignment, manual curation, and RAxML phylogeny, and confirmed to resolve in

topologies coherent with protein function and taxonomy prior to enumeration of homologue presence/ absence (**Dataset S1**).

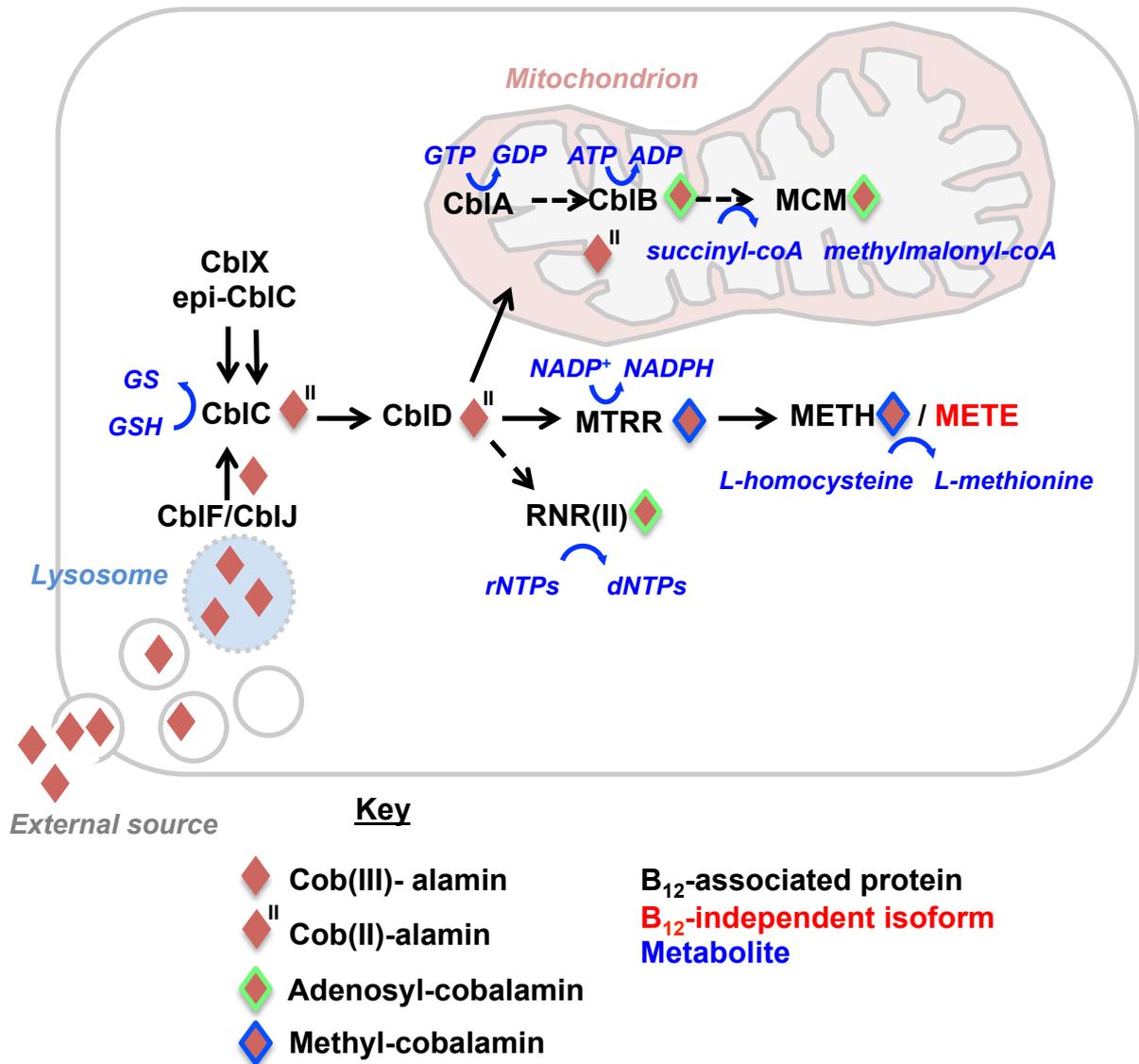
**Functional, targeting and biogeographical annotation** : PFAM domains were searched in each homologue retrieved by the initial RbH search by HMMER v 3.3.2 against the Pfam v 35.0 library [52, 94]. Sequences that retrieved PFAM domains associated with each query protein were recorded (METE : Meth\_synt\_1- PF08267, Meth\_synt\_2- PF01717; METH : B<sub>12</sub>-binding- PF02310, B<sub>12</sub>-binding\_2- PF02607, Met\_synt\_B<sub>12</sub>- PF02965, Pterin\_bind- PF00809, S-methyl\_trans- PF02574 ; CblA : MeaB- PF03308 ; CblB : Cob\_adeno\_trans- PF01923 ; CblC : MMACHC- PF16690 ; CblD : MMADHC- PF10229 ; CblF : LMBR1- PF04791 ; CblJ : ABC\_tran- PF00005 ; CblX : Kelch\_1- PF01344, Kelch\_2- PF07646, Kelch\_3- PF13415, Kelch\_4- PF13418, Kelch\_5- PF13854, Kelch\_6- PF13964 ; epi-CblC : AhpC\_TSA- PF00578, Redoxin- PF08534, 1-cysPrx\_C- PF10417MCM : MM\_CoA\_mutase- PF01642 ; MTRR : Flavodoxin\_1- PF00258, FAD\_binding\_1- PF00667, NAD\_binding\_1- PF00175 ; RNR II- Ribonuc\_red\_lgC- PF02867, RNR II\_Alpha- PF17975) with threshold e-value 10<sup>-05</sup>. Localisations for each protein were inferred, considering both the full length of the protein sequence and the protein sequence trimmed to the first annotated methionine, using WolfPSort v 1.0 with plant substitution matrix [95]; SignalP v 5.0 under eukaryotic settings [96]; TargetP v 2.0 with plant substitution matrix [97]; and HECTAR v 1.0 under default settings [98]. Collection sites were recorded for each algal species using the corresponding culture collection records considering strain synonyms, and where appropriate direct information from the literature or collector [99-107]. PFAM boxplots in **Fig. S1** are displayed using BoxPlotR [108].

**Data deposition** : All strain information, recovered homologues by RbH, alignments, phylogenetic topologies, PFAM and targeting annotations are provided in **Dataset S1**. An interactive map of all algal collection sites identified within the dataset, shadeable either via phylogenetic affiliation or inferred Vitamin B<sub>12</sub> metabolic status, is available via <https://tinyurl.com/3esfpkmv> .

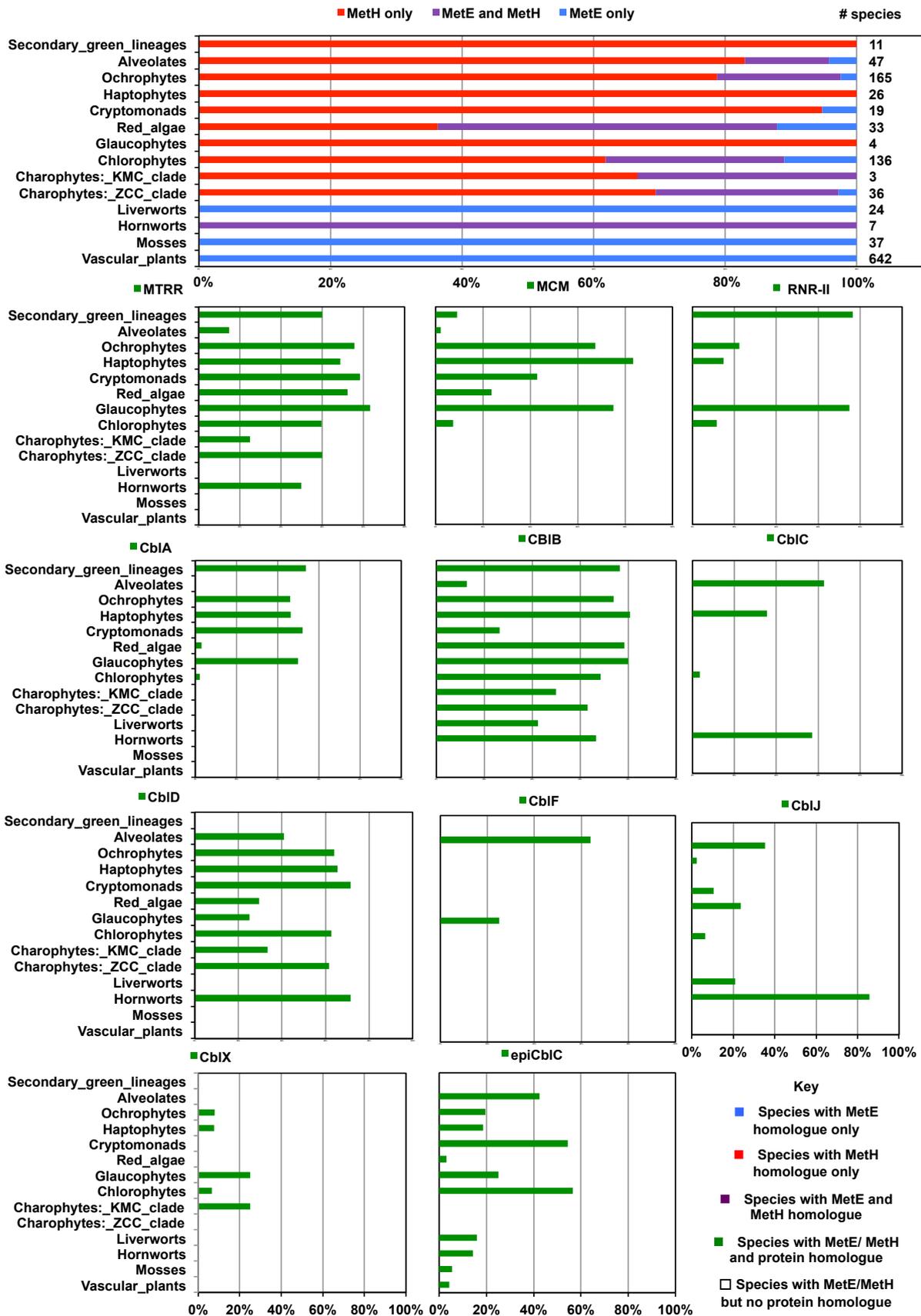
**Funding information** : RGD acknowledges an ERC Starting Grant (« ChloroMosaic » ; grant number 101039760), awarded 2023-2027. Work at the IBENS is supported by the Investissements d'Avenir programmes PSL and MEMOLIFE: MEMO LIFE (ANR-10-LABX-54), and PSL\*Research University (ANR-11-IDEX-0001-02).

**Acknowledgments** : The authors thank Dr. Fabio Rocha Jimenez Vieira (IBENS) for assistance with the decontamination of the OneKp transcriptome dataset; and Ms. Cathy Johnston (CSIRO), Dr. Maïke Lorenz (Georg-August-Universität Göttingen), Prof. Michael Melkonian (Universität Köln) and Mr. Stephen Peña (UTEX Culture Collection) for the assistance of strain isolation sites tabulated in **Dataset S1**.

## Figure Legends

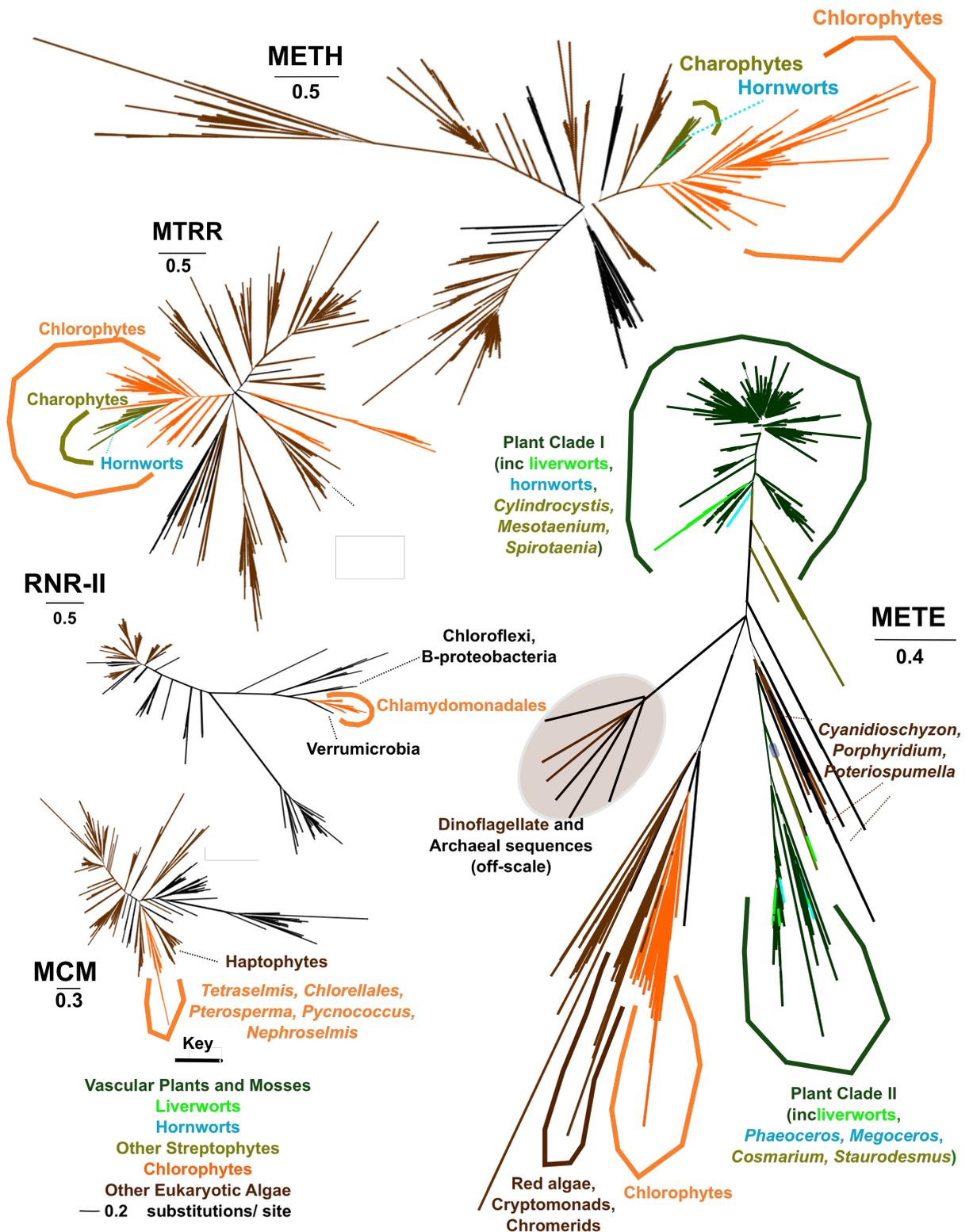


**Fig. 1. Known B<sub>12</sub> uptake and utilisation pathways in eukaryotes.** This figure shows a schematic eukaryotic cell, with potential B<sub>12</sub>-associated proteins detected in previous studies across the eukaryotes, labelled following [29] and [36]. The geneIDs, origins and PFAM domains associated with each query protein for the phylogenetic analysis are tabulated below.



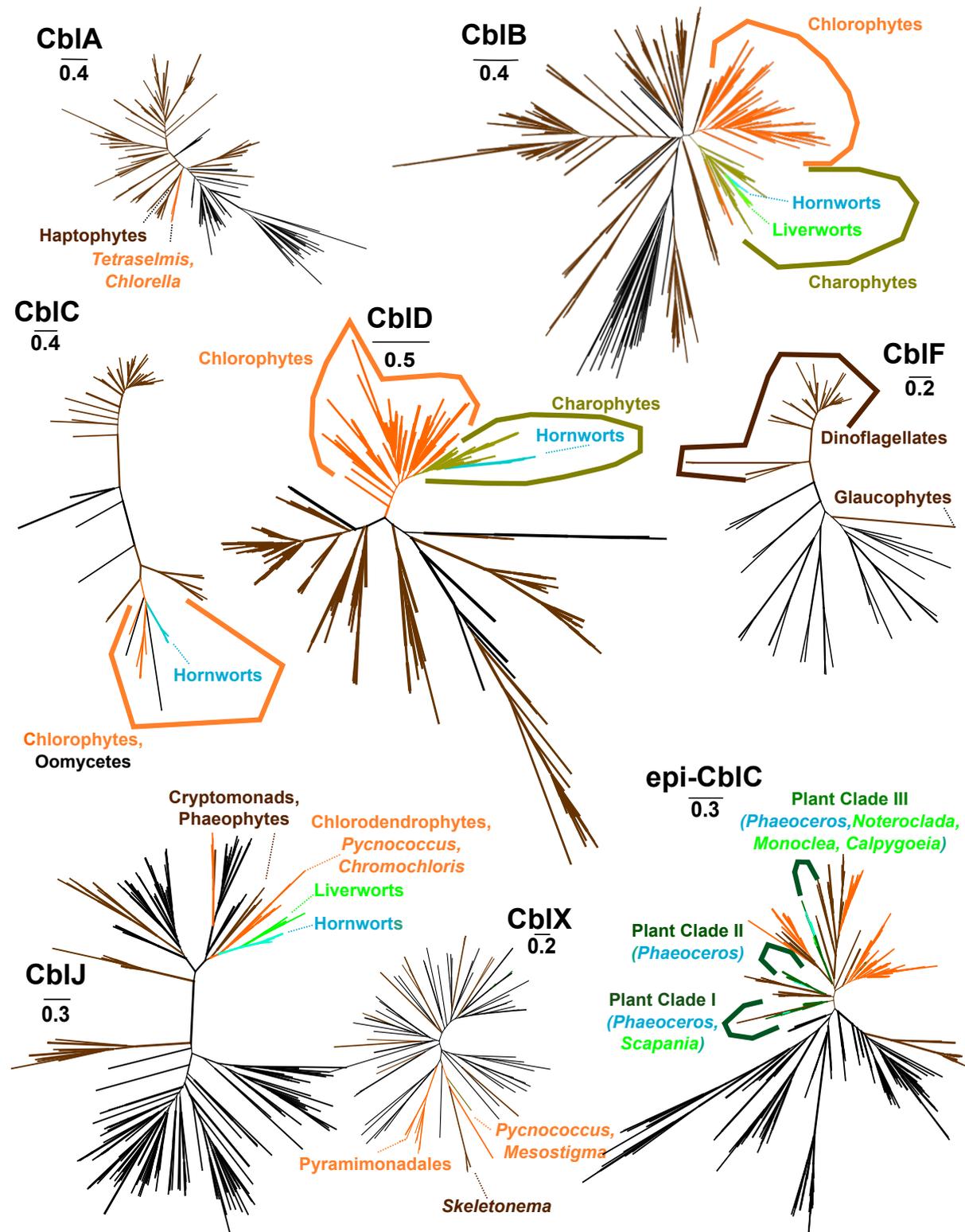
**Fig. 2. Bar plots of the occurrence of B<sub>12</sub>-associated metabolism across photosynthetic eukaryotes.** These graphs show the number of species from thirteen different algal phylogenetic or functional groups inferred to possess METH, METE, or

both METH and METE, by combined RbH, PFAM domain and single-gene phylogenetic analysis ; and the number of species from these groups for which at least one B<sub>12</sub>-associated enzyme was detected (the following, plus METE or METH) inferred to possess homologues of MTRR, MCM, RNR II, CBLA, CBLB, CBLC, CBLD, CBLF, CBLJ, CBLX or epi-CBLC via the same methodology. The total number of species assessable in each case is given to the right-hand side of each plot. Abbreviations : KMC- Klebsormidiaceae, Mesostigmatophyceae, Chlorokybophyceae ; ZCC- Zygnematophyceae, Charophyceae, Coleochaetophyceae. Complete tabulated occurrences per lineage and species are provided in **Dataset S1**, sheets 1-2 ; and individual homologue lists per gene in **Dataset S1**, sheets 5-17.



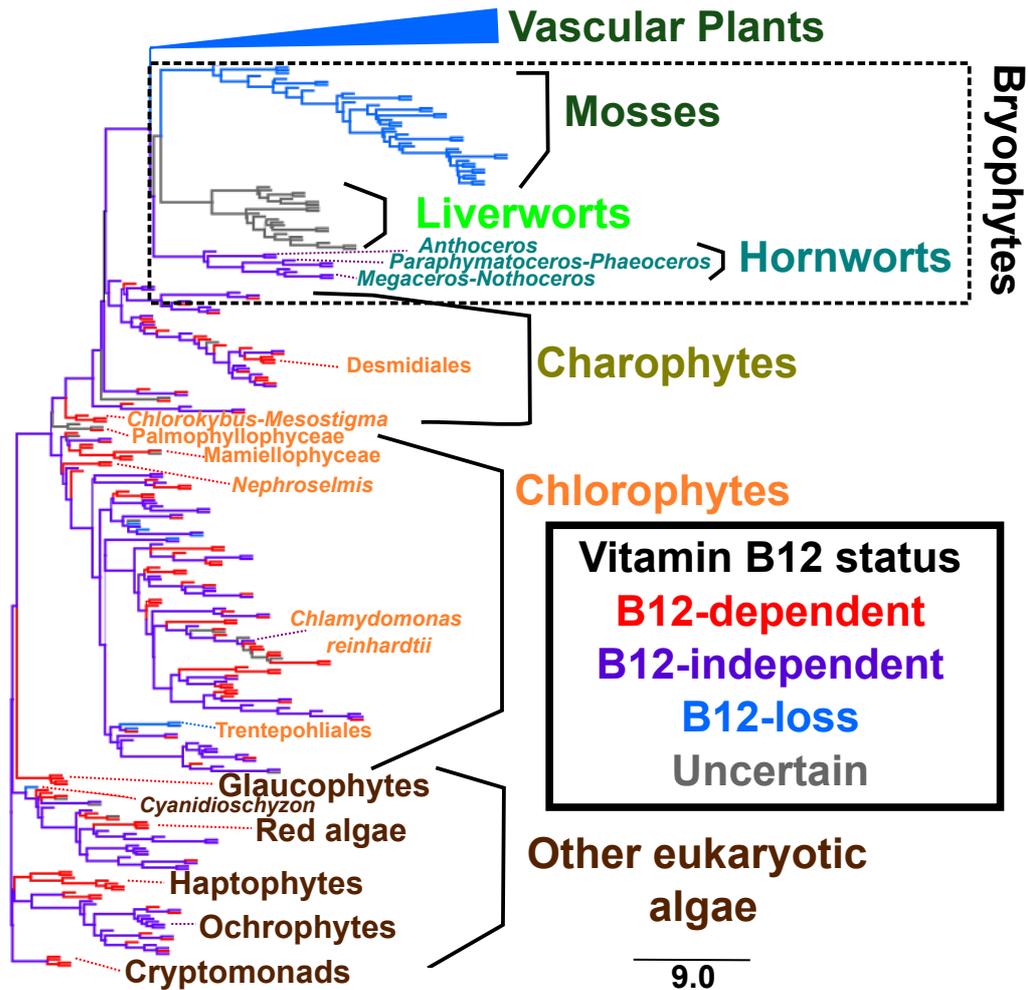
**Fig. 3. Phylogenetic trees of B<sub>12</sub>-dependent metabolism across photosynthetic eukaryotes.** This figure shows best-scoring RAxML trees realised across 350 bootstrap replicates with the PROTGAMMAJTT substitution matrix for METE, METH, MTRR, MCM and RNR II enzymes. Suspected dinoflagellate and archaeal METE sequences are shown in a separate panel to the remaining tree topology, due to the high divergence (> 2.0 substitutions per site) observed from all other aligned sequences. Taxa are coloured by phylogenetic origin: dark green- vascular plants and mosses; light green- liverworts;

cyan- hornworts; dark yellow- other streptophytes; orange- chlorophytes; dark brown- other algae; black- other lineages. Branch thickness corresponds to bootstrap support. Initial and curated alignment and tree topologies are provided in **Dataset S1**, sheets 18-19 ; and taxa inferred to be contaminants and thus excluded in **Dataset S1**, sheet 20.



**Fig. 4. Phylogenetic trees of intracellular transport pathways B<sub>12</sub> transport across photosynthetic eukaryotes.** This figure shows best-scoring RAxML trees realised across 350 bootstrap replicates with the PROT GAMMAJTT substitution matrix for CBLA,

CBLB, CBLC, CBLD, CBLF, CBLJ, CBLX and epi-CBLC proteins. Taxa are phylogenetic origin: dark green- vascular plants and mosses; light green- liverworts; cyan- hornworts; dark yellow- other streptophytes; orange- chlorophytes; dark brown- other algae; black- other lineages. Branch thickness corresponds to bootstrap support.



**Fig. 5. Vertical inheritance of hornwort B<sub>12</sub> metabolism.** This figure shows a concatenated multigene tree topology for all OneKp transcriptomes, taken from [38]. Taxa names are shown by taxonomic affiliation: dark green- vascular plants and mosses; light green- liverworts; cyan- hornworts; dark yellow- other streptophytes; orange- chlorophytes; dark brown- other algae. Branches are shaded by inferred B<sub>12</sub> status: red- presence of B<sub>12</sub>-dependent metabolism only; purple- presence of B<sub>12</sub>-dependent and -independent metabolism; light blue: B<sub>12</sub>-independent metabolism only; grey- unknown B<sub>12</sub> status. For concision, vascular plants (universally lacking B<sub>12</sub>-dependent metabolism) are collapsed to a single branch. Liverworts are marked as « unknown » due to the uncertain function of their encoded CblB and CblJ proteins. Major algal lineages are labelled, alongside individual species of interest and algal clades with inferred losses of B<sub>12</sub>-associated metabolism. ; and a nexus format of the schematic OneKp concatenated topology in **Dataset S1**, sheet 21.

**Fig. S1. Metabolic functions of Vitamin B<sub>12</sub>-associated metabolism in plants.** This figure shows boxplots, displayed using BoxPlotR [108], of the  $-\log_{10}$  of the calculated hmmer e-value scores of key PFAM domains observed for each protein [94] of all phylogenetically reconciled homologues of eight potential B<sub>12</sub>-associated proteins found in hornworts : METE, METH, MTRR, CBLB, CBLC, CBDL, CBLJ and epi-CBLC. Values are shown for four taxonomic groups: eukaryotic algae ; hornworts ; liverworts ; and mosses and vascular plants (grouped together by the absence of B<sub>12</sub> metabolism as

opposed to phylogeny). Greater values imply greater fidelity to the PFAM hmm ; groups for which no homologues were identified are plotted as a -1 value. For the METH sequences, an alignment is additionally provided of a 20 aa region of the first substrate binding pocket of the pterin binding domain of hornwort METH against the equivalent *Chlamydomonas* sequence following [21], and showing broad conservation of key residues and inferred functions. An overview of the PFAM values is given in **Dataset S1**, sheet 3; with individual scores for each homologue provided in **Dataset S1**, sheets 5-17.

**Fig. S2. Targeting predictions of individual B<sub>12</sub>-associated proteins.** **A** : bar plots of the proportion of phylogenetically identified homologues of each B<sub>12</sub>-associated protein to possess at least one mitochondrial, chloroplast or endomembrane targeting sequence, from combined WolfPSort, HECTAR, SignalP and TargetP predictions. Except for MCM which shows broadly mitochondrial targeting predictions and CblF with predominantly endomembrane localisations, no patterns are consistently observed. **B** : wheel plots of the taxonomic composition of mitochondria-targeted MCM sequences, showing broad conservation of this localisation across algae with secondary red chloroplasts ; and of endomembrane-targeted CblF sequences, which are restricted to dinoflagellates.

**Table 1.** Overview of the query sequences used for phylogenetic searches, as obtained from human [29] and algal gene sequences [36].

<b>Protein</b>	<b>GeneID</b>	<b>Query Source</b>	<b>PFAMs</b>
METH	XP_042923308.1	<i>Chlamydomonas</i>	PF00809, PF02310, PF02574, PF02607, PF02965
METE	AAC49178.1	<i>Chlamydomonas</i>	PF01717, PF08267
MCM	XP_002179504.1	<i>Phaeodactylum</i>	PF01642
RNR	Q2PDF6.1	<i>Euglena</i>	PF02867, PF17975
MTRR	AAF16876.1	Human	PF00175, PF00258, PF00667
CblA	NP_001362573.1	Human	PF03308
CblB	NP_443077.1	Human	PF01923
CblC	NP_056321.2	Human	PF16690
CblD	NP_056517.1	Human	PF10229
CblF	NP_060838.3	Human	PF04791
CblJ	NP_001340531.1	Human	PF00005
CblX	NP_005325.2	Human	PF01344, PF07646, PF13415, PF13418, PF13854, PF13964
epi-CblC	NP_001189360.1	Human	PF00578, PF08534, PF10417

**Table 2.** Overview of the libraries searched for presence of B<sub>12</sub>-containing homologues. Totals are provided for the unique numbers of species searched in different taxonomic groups, divided by library type (genome, 1kp transcriptome, MMETSP transcriptome, and other including single-cell genome and independent transcriptome libraries). Totals are provided by species as opposed to strain, and therefore e.g. three searched hornwort genomes (*Anthoceros agrestis* Oxford, *A. agrestis* Bonn, *A. punctigera*) [44] are tabulated as two distinct species. Species for which both genome and 1kp/ MMETSP transcriptomes have been sequenced (e.g., *A. agrestis*) are tabulated in both columns. Acronyms : ZCC- Zygnetophytes, Coleochaetales, Charophytes ; KMC- Klebsormidiophytes, Mesostigma, Chlorokybus.

<b><u>Taxonomic Group</u></b>	<b><u>1kp</u></b>	<b><u>Genome</u></b>	<b><u>MMETSP</u></b>	<b><u>Other</u></b>
Vascular plants	968	3	0	0
Mosses	48	1	0	0
Hornworts	11	2	0	0
Liverworts	34	1	0	0
Charophytes: ZCC clade	43	0	0	0
Charophytes: KMC clade	3	1	0	0
Chlorophytes	114	14	52	2
Glaucoephytes	5	3	1	0
Red algae	32	6	10	0
Cryptomonads	5	1	19	0
Haptophytes	4	2	29	0
Ochrophytes	24	12	133	30
Alveolates	1	5	46	0
Secondary green lineages	1	2	15	0

**Table 3.** Identified species inferred to have lost Vitamin B<sub>12</sub>-associated metabolism, based on the retention of METE and minimally the absence of METH and MTRR sequences as verified by phylogenetic and PFAM analysis. A detailed map of the recorded collection sites of all algae included in the dataset is provided at <https://tinyurl.com/3esfpkmv>.

Species	Lineage	Strain	Isolation site	Isolation latitude	Isolation longitude	Notes
Land Plants	Mosses, Vascular plants, liverworts?					
<i>Blastophysa cf. rhizopus</i>	Chlorophytes	M3368 / BEA 0074B	Epiphyte; Spain, Canary Islands, Fuerteventura, near the lighthouse. Marine	28,4052029	-14,4413182	Based on PFAM presence only
<i>Botryococcus braunii</i>	Chlorophytes	UTEX 2441	Lake Huaypo, Peru; Freshwater	-13.5	-72.1	Based on PFAM presence only
<i>Brandtodinium nutriculum</i>	Dinoflagellates	RCC3387	Marine; foraminiferan endosymbiont	43.7	7.3	
<i>Cephaleuros virescens</i>	Chlorophytes	SAG 28.93	Natal, parasite from leaves of Camellia	-29.9	31.0	Based on PFAM presence only; Trentepohliales
<i>Coccomyxa pringsheimii</i>	Chlorophytes	SAG 216.7	Finland: freshwater; phycobiont of lichen Botrydina vulgaris	N/A	N/A	
<i>Cyanidioschyzon merolae</i>	Red algae	10D	Hot spring; Italy; Sardinia	40.9	14.3	
<i>Galdieria sulphuraria</i>	Red algae	074W	Fumaroles, Mount Lawu, Indonesia	-7.6	11.2	Encodes MCM, CblA
<i>Geminigera cryophila</i>	Cryptomonads	CCMP2564	Marine	-77,8	-163	
<i>Ignatius tetrasporus</i>	Chlorophytes	UTEX20 <sub>12</sub>	Freshwater	N/A	N/A	May encode RNR II
<i>Leptosira obovata</i>	Chlorophytes	SAG 445.1	Basel, boggy water at Rosenau; freshwater	47.5	7.9	
<i>Porphyridium cruentum</i>	Red algae	UTEX 161	Basel, Switzerland; wet shaded tuff	47.6	7.6	Encodes CblB
<i>Poteriospumella JBC07</i>	Chrysophytes	JBC07	Peoples Republic of China, Lake Tai Hu	31.5	120.2	
<i>Pseudoscourfieldia marina</i>	Chlorophytes	SCCAP K/0017	Inner Oslofjord, Norway	59.4	10.6	Encodes CblB
<i>Trentepohlia annulata</i>	Chlorophytes	SAG 20.94	Damasská cesta pobl. Svratky, Ceskomoravská vrchovina	49.7	16.0	Based on PFAM presence only; Trentepohliales
<i>Vitrella brassicaformis</i>	Alveolates	CCMP3155	Marine, coral	-23.5	152	

## References

1. Schneider, L.K., et al., *A dataset on trophic modes of aquatic protists*. Biodivers Data J, 2020. **8**: p. e56648.
2. Croft, M.T., M.J. Warren, and A.G. Smith, *Algae need their vitamins*. Eukaryot Cell, 2006. **5**(8): p. 1175-83.
3. Warren, M.J., et al., *The biosynthesis of adenosylcobalamin (vitamin B12)*. Nat Prod Reports, 2002. **19**(4): p. 390-412.
4. Shelton, A.N., et al., *Uneven distribution of cobamide biosynthesis and dependence in bacteria predicted by comparative genomics*. ISME J, 2019. **13**(3): p. 789-804.
5. Yamada, K., et al., *Human methionine synthase reductase is a molecular chaperone for human methionine synthase*. Proc Natl Acad Sci USA, 2006. **103**(25): p. 9476-9481.
6. Banerjee, R.V. and R.G. Matthews, *Cobalamin-dependent methionine synthase*. The FASEB Journal, 1990. **4**(5): p. 1450-1459.
7. Dobson, C.M., et al., *Identification of the gene responsible for the cblB complementation group of vitamin B12-dependent methylmalonic aciduria*. Hum Mol Genet, 2002. **11**(26): p. 3361-9.
8. Banerjee, R. and S. Chowdhury, *Methylmalonyl-CoA mutase*. Chemi Biochem B12., 1999: p. 707-729.
9. Stabler, S.P. and R.H. Allen, *Vitamin B12 deficiency as a worldwide problem*. Ann Rev Nutr, 2004. **24**: p. 299.
10. Taneja, S., et al., *Cobalamin and folate status in infants and young children in a low-to-middle income community in India*. Am J Clin Nutr, 2007. **86**(5): p. 1302-9.
11. Obeid, R., et al., *Cobalamin Status from pregnancy to early childhood: lessons from global experience*. Adv Nutr, 2017. **8**(6): p. 971-979.
12. Niklewicz, A., et al., *The importance of vitamin B(12) for individuals choosing plant-based diets*, in *Eur J Nutr*. 2023. **62**(3): p. 1551-1559.
13. Pimentel, D. and M. Pimentel, *Sustainability of meat-based and plant-based diets and the environment*. Am J Clinical Nutrition, 2003. **78**(3): p. 660S-663S.
14. Marrero, A. and J. Mattei, *Reclaiming traditional, plant-based, climate-resilient food systems in small islands*. Lancet Planetary Health, 2022. **6**(2): p. e171-e179.
15. Watanabe, F. and T. Bito, *Vitamin B*. Exp Biol Med (Maywood), 2018. **243**(2): p. 148-158.
16. Reilly, C. and J. Henry, *Geophagia: why do humans consume soil?* Nutrition Bulletin, 2000. **25**(2): p. 141-144.
17. Marques de Brito, B., et al., *Vitamin B12 sources in non-animal foods: a systematic review*. Crit Rev Food Sci and Nutrition, 2022: p. 1-15.
18. Croft, M.T., et al., *Algae acquire vitamin B12 through a symbiotic relationship with bacteria*. Nature, 2005. **438**(7064): p. 90-93.
19. Tang, Y.Z., F. Koch, and C.J. Gobler, *Most harmful algal bloom species are vitamin B1 and B12 auxotrophs*. Proc Natl Acad Sci USA, 2010. **107**(48): p. 20756-61.
20. Nef, C., et al., *How haptophytes microalgae mitigate vitamin B12 limitation*. Sci Repo, 2019. **9**(1): p. 8417.

21. Lin, S., et al., *Vitamin B12-auxotrophy in dinoflagellates caused by incomplete or absent cobalamin-independent methionine synthase genes (metE)*. *Fundamental Res*, 2022 **2**(5): 727-737.
22. Raux, E., H.L. Schubert, and M.J. Warren, *Biosynthesis of cobalamin (vitamin B12): a bacterial conundrum*. *Cell Mol Life Sci*, 2000. **57**(13-14): p. 1880-93.
23. Raux, E., et al., *Salmonella typhimurium cobalamin (vitamin B12) biosynthetic genes: functional studies in S. typhimurium and Escherichia coli*. *J Bacteriol*, 1996. **178**(3): p. 753-767.
24. Roessner, C.A., P.J. Santander, and A.I. Scott, *Multiple biosynthetic pathways for vitamin B12: variations on a central theme*. *Vitam Horm*, 2001. **61**: p. 267-97.
25. Droop, M.R., *Vitamins, phytoplankton and bacteria: symbiosis or scavenging?* *J Plankton Res*, 2007. **29**(2): p. 107-113.
26. Sanders, R.W., K.G. Porter, and D.A. Caron, *Relationship between phototrophy and phagotrophy in the mixotrophic chrysophyte Poterioochromonas malhamensis*. *Microb Ecol*, 1990. **19**(1): p. 97-109.
27. Helliwell, K.E., et al., *Cyanobacteria and eukaryotic algae use different chemical variants of vitamin B12*. *Curr Biol*, 2016. **26**(8): p. 999-1008.
28. Vancaester, E., et al., *Comprehensive and functional analysis of horizontal gene transfer events in diatoms*. *Mol Biol Evol*, 2020. **37**(11): p. 3243-3257.
29. Hannibal, L. and D.W. Jacobsen, *Intracellular processing of vitamin B12 by MMACHC (CblC)*. *Vitamins And Hormones*, 2022. **119**: p. 275-298.
30. Coelho, D., et al., *Gene identification for the cblD defect of vitamin B12 metabolism*. *N Engl J Med*, 2008. **358**(14): p. 1454-64.
31. Dobson, C.M., et al., *Identification of the gene responsible for the cblA complementation group of vitamin B12-responsive methylmalonic acidemia based on analysis of prokaryotic gene arrangements*. *Proc Natl Acad Sci USA*, 2002. **99**(24): p. 15554-15559.
32. Hutner, S.H., et al., *Biochemical geography: some aspects of recent vitamin research*. *Geographical Review*, 1956. **46**(3): p. 404-407.
33. Bertrand, E.M., et al., *Influence of cobalamin scarcity on diatom molecular physiology and identification of a cobalamin acquisition protein*. *Proc Natl Acad Sci USA*, 2012. **109**(26): p. E1762-71.
34. Barber-Lluch, E., et al., *Variability of vitamin B12 concentrations in waters along the Northwest Iberian shelf*. *Regional Studies Mar Sci*, 2021. **42**: p. 101608.
35. Froese, D.S., B. Fowler, and M.R. Baumgartner, *Vitamin B12, folate, and the methionine remethylation cycle—biochemistry, pathways, and regulation*. *J Inherited Metabol Disease*, 2019. **42**(4): p. 673-685.
36. Helliwell, K.E., et al., *Insights into the evolution of vitamin B12 auxotrophy from sequenced algal genomes*. *Mol Biol Evol*, 2011. **28**(10): p. 2921-33.
37. Gonzalez, J. C., Banerjee, R. V., Huang, S., Sumner, J. S., and R.G. Matthews, *Comparison of cobalamin-independent and cobalamin-dependent methionine synthases from Escherichia coli: two solutions to the same chemical problem.* *Biochemistry*, 1992. **31**(26): p. 6045-6056.
38. One Thousand Plant Transcriptomes Initiative, *One thousand plant transcriptomes and the phylogenomics of green plants*. *Nature*, 2019. **574**(7780): p. 679-685.

39. Pierella Karlusich, J.J., et al., *Modèles biogéographiques et génomes des photoautotrophes aquatiques*. In *Planète bleue, photosynthèse rouge et verte*. S.C. Maberly, and B. Gontero, Editors. 2023, ISTE Editions: London 2023. p. 45-82.
40. Wang, S., et al., *Genomes of early-diverging streptophyte algae shed light on plant terrestrialization*. *Nat Plants*, 2020. **6**(2): p. 95-106.
41. Buschmann, H. and A. Holzinger, *Understanding the algae to land plant transition*. *J Exp Bot*, 2020. **71**(11): p. 3241-3246.
42. Skokan, R., et al., *PIN-driven auxin transport emerged early in streptophyte evolution*. *Nat Plants*, 2019. **5**(11): p. 1114-1119.
43. Harris, B.J., et al., *Divergent evolutionary trajectories of bryophytes and tracheophytes from a complex common ancestor of land plants*. *Nat Ecol Evol*, 2022. **6**(11): p. 1634-1643.
44. Li, F.-W., et al., *Anthoceros genomes illuminate the origin of land plants and the unique biology of hornworts*. *Nat Plants*, 2020. **6**(3): p. 259-272.
45. Grigoriev, I.V., et al., *PhycoCosm, a comparative algal genomics resource*. *Nucleic Acids Res*, 2021. **49**(D1): p. D1004-D1011.
46. Carpenter, E.J., et al., *Access to RNA-sequencing data from 1,173 plant species: The 1000 Plant Transcriptomes initiative (1KP)*. *Gigascience*, 2019. **8**(10).
47. Keeling, P.J., et al., *The Marine Microbial Eukaryote Transcriptome Sequencing Project (MMETSP): illuminating the functional diversity of eukaryotic life in the oceans through transcriptome sequencing*. *PLoS Biol*, 2014. **12**(6): p. e1001889.
48. Marron, A.O., et al., *The evolution of silicon transport in eukaryotes*. *Mol Biol Evol*, 2016. **33**(12): p. 3226-3248.
49. Dorrell, R.G., et al., *Phylogenomic fingerprinting of tempo and functions of horizontal gene transfer within ochrophytes*. *Proc Natl Acad Sci USA*, 2021. **118**(4): 2009974118.
50. Beisser, D., et al., *Comprehensive transcriptome analysis provides new insights into nutritional strategies and phylogenetic relationships of chrysophytes*. *PeerJ*, 2017. **5**: p. e2832.
51. Stamatakis, A., *RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies*. *Bioinformatics*, 2014. **30**(9): p. 1312-3.
52. Mistry, J., et al., *Pfam: the protein families database in 2021*. *Nucleic Acids Res*, 2021 **49** (D1): p. D412–D419.
53. Moreno-Hagelsieb, G. and K. Latimer, *Choosing BLAST options for better detection of orthologs as reciprocal best hits*. *Bioinformatics*, 2008. **24**(3): p. 319-324.
54. Hori, K., et al., *Klebsormidium flaccidum genome reveals primary factors for plant terrestrial adaptation*. *Nat Commun*, 2014. **5**: 3978.
55. Villarreal, J.C. and S.S. Renner, *Hornwort pyrenoids, carbon-concentrating structures, evolved and were lost at least five times during the last 100 million years*. *Proc Natl Acad Sci USA*, 2012. **109**(46): p. 18873-18878.
56. Dixon, M.M., et al., *The structure of the C-terminal domain of methionine synthase: presenting S-adenosylmethionine for reductive methylation of B12*. *Structure*, 1996. **4**(11): p. 1263-75.
57. Drennan, C.L., et al., *How a protein binds B12: a 3.0 Å X-ray structure of B12-binding domains of methionine synthase*. *Science*, 1994. **266**(5191): p. 1669-1674.
58. Joshi, V. and G. Jander, *Arabidopsis methionine gamma-lyase is regulated according to isoleucine biosynthesis needs but plays a subordinate role to threonine deaminase*. *Plant Physiol*, 2009. **151**(1): p. 367-78.

59. Andersen, R.A., *Algal Culturing Techniques*. Vol. 0-12-088426-7. 2005: Elsevier.
60. Dorrell, R.G., et al., *Convergent evolution and horizontal gene transfer in Arctic Ocean microalgae*. *Life Sci Alliance*, 2021. **6** (3): e202201833.
61. Penot, M., et al., *Genomic and meta-genomic insights into the functions, diversity and global distribution of haptophyte algae*. *Appl Phycol*, 2022. **3** (1): 340-359.
62. Pierella Karlusich, J.J., et al., *A robust approach to estimate relative phytoplankton cell abundances from metagenomes*. *Molecular Ecology Resources*, 2023. **23** (1): p. 16-40.
63. Grant, M.A., et al., *Direct exchange of vitamin B12 is demonstrated by modelling the growth dynamics of algal-bacterial cocultures*. *ISME J*, 2014. **8**(7): p. 1418-27.
64. Bunbury, F., et al., *Responses of a newly evolved auxotroph of Chlamydomonas to B12 deprivation*. *Plant Physiol*, 2020. **183**(1): p. 167-178.
65. Price, D.C., et al., *Cyanophora paradoxa genome elucidates origin of photosynthesis in algae and plants*. *Science*, 2012. **335**(6070): p. 843-847.
66. Li, L., et al., *The genome of Prasinoderma coloniale unveils the existence of a third phylum within green plants*. *Nat Ecol Evol*, 2020. **4**(9): p. 1220-1231.
67. Nef, C., et al., *Cobalamin scarcity modifies carbon allocation and impairs DMSP production through methionine metabolism in the haptophyte microalgae Tisochrysis lutea*. *Frontiers in Marine Science*, 2020. **7**: e569560.
68. Oborník, M., et al., *Morphology, ultrastructure and life cycle of Vitrella brassicaformis n. sp., n. gen., a novel chromerid from the Great Barrier Reef*. *Protist*, 2012. **163**(2): p. 306-23.
69. Kroth, P.G., et al., *A model for carbohydrate metabolism in the diatom Phaeodactylum tricornutum deduced from comparative whole genome analysis*. *PLoS One*, 2008. **3**(1): p. e1426.
70. Ge, F., et al., *Methylcrotonyl-CoA Carboxylase regulates triacylglycerol accumulation in the model diatom Phaeodactylum tricornutum*. *Plant Cell*, 2014. **26**(4): p. 1681-1697.
71. Ait-Mohamed, O., et al., *PhaeoNet: a holistic RNAseq-based portrait of transcriptional coordination in the model diatom Phaeodactylum tricornutum*. *Frontiers Plant Sci*, 2020. **11**: e590949.
72. Carell, E.F. and J. Seeger Jr, *Ribonucleotide reductase activity in vitamin B12-deficient Euglena gracilis*. *Biochem J*, 1980. **188**(2): p. 573-576.
73. Novák Vanclová, A.M., et al., *Metabolic quirks and the colourful history of the Euglena gracilis secondary plastid*. *New Phytol*, 2020. **225**(4): p. 1578-1592.
74. Bowman, J.L., et al., *Insights into land plant evolution garnered from the Marchantia polymorpha genome*. *Cell*, 2017. **171**(2): p. 287-304. e15.
75. Sugita, H., et al., *The vitamin B12-producing bacteria in the water and sediment of a carp culture pond*. *Aquaculture*, 1994. **119**(4): p. 425-431.
76. Tsuchiya, C., T. Sakata, and H. Sugita, *Novel ecological niche of Cetobacterium somerae, an anaerobic bacterium in the intestinal tracts of freshwater fish*. *Lett Appl Microbiol*, 2008. **46**(1): p. 43-48.
77. Facey, J.A., et al., *Assessing the importance of cobalt as a micronutrient for freshwater cyanobacteria*. *J Phycol*, 2022. **58**(1): p. 71-79.
78. Browning, T.J., et al., *Nutrient co-limitation at the boundary of an oceanic gyre*. *Nature*, 2017. **551**(7679): p. 242-246.

79. Taylor, D.L. and C.C. Lee, *A new cryptomonad from antarctica: Cryptomonas cryophila sp. nov.* Archiv für Mikrobiologie, 2004. **75**: p. 269-280.
80. Kosecka, M., et al., *Trentepohlialean Algae (Trentepohliales, Ulvophyceae) show preference to selected mycobiont lineages in lichen symbioses.* J Phycol, 2020. **56**(4): p. 979-993.
81. Probert, I., et al., *Brandtodinium gen. nov. and B. nutricula comb. Nov. (Dinophyceae), a dinoflagellate commonly found in symbiosis with polycystine radiolarians.* J Phycol, 2014. **50**(2): p. 388-399.
82. Lawrence, A.D., et al., *Construction of fluorescent analogs to follow the uptake and distribution of cobalamin (vitamin B12) in bacteria, worms, and plants.* Cell Chem Biol, 2018. **25**(8): p. 941-951. e6.
83. Wickett, N.J., et al., *Phylotranscriptomic analysis of the origin and early diversification of land plants.* Proc Natl Acad Sci USA, 2014. **111**(45): p. E4859-E4868.
84. Cox, C.J., *Land plant molecular phylogenetics: a review with comments on evaluating incongruence among phylogenies.* Crit Rev Plant Sci, 2018. **37**(2-3): p. 113-127.
85. Meyer, M., U. Seibt, and H. Griffiths, *To concentrate or ventilate? Carbon acquisition, isotope discrimination and physiological ecology of early land plant life forms.* Phil Trans R Soc B, 2008. **363**(1504): p. 2767-2778.
86. Adams, D.G., *Cyanobacteria in symbiosis with hornworts and liverworts*, in *Cyanobacteria in Symbiosis*, A.N. Rai, B. Bergman, and U. Rasmussen, Editors. 2002, Springer Netherlands: Dordrecht. p. 117-135.
87. Mozafar, A., *Enrichment of some B-vitamins in plants with application of organic fertilizers.* Plant and Soil, 1994. **167**(2): p. 305-311.
88. Altschul, S.F., et al., *Basic local alignment search tool.* J Mol Biol, 1990. **215**(3): p. 403-10.
89. Arabidopsis Genome Initiative, *Analysis of the genome sequence of the flowering plant Arabidopsis thaliana.* Nature, 2000. **408**(6814): p. 796-815.
90. Yan, X., et al., *METHIONINE SYNTHASE1 is involved in chromatin silencing by maintaining DNA and histone methylation.* Plant Physiol, 2019. **181**(1): p. 249-261.
91. Katoh, K. and D.M. Standley, *MAFFT multiple sequence alignment software version 7: improvements in performance and usability.* Mol Biol Evol, 2013. **30**(4): p. 772-80.
92. Kearse, M., et al., *Geneious Basic: An integrated and extendable desktop software platform for the organization and analysis of sequence data.* Bioinformatics, 2012. **28**(12): p. 1647-1649.
93. Capella-Gutiérrez, S., J.M. Silla-Martínez, and T. Gabaldón, *trimAl: a tool for automated alignment trimming in large-scale phylogenetic analyses.* Bioinformatics, 2009. **25**(15): p. 1972-3.
94. Potter, S.C., et al., *HMMER web server: 2018 update.* Nucleic Acids Res, 2018. **46**(W1): p. W200-W204.
95. Horton, P., et al., *WoLF PSORT: protein localization predictor.* Nucleic Acids Res, 2007. **35**: p. W585-W587.
96. Almagro Armenteros, J.J., et al., *SignalP 5.0 improves signal peptide predictions using deep neural networks.* Nature Biotechnol, 2019. **37**(4): p. 420-423.
97. Almagro Armenteros, J.J., et al., *Detecting sequence signals in targeting peptides using deep learning.* Life Sci Alliance, 2019. **2**(5).
98. Gschloessl, B., Y. Guermeur, and J.M. Cock, *HECTAR: a method to predict subcellular targeting in heterokonts.* BMC Bioinform, 2008. **9**: p. 393.

99. Gachon, C.M.M., et al., *The CCAP KnowledgeBase: linking protistan and cyanobacterial biological resources with taxonomic and molecular data*. *Systemat Biodiv*, 2013. **11**(4): p. 407-413.
100. Vaultot, D., et al., *The Roscoff Culture Collection (RCC): a collection dedicated to marine picoplankton*. *Nova Hedwigia*, 2004. **79**(1-2): p. 32.
101. Leya, T., *The CCCryo Culture Collection of Cryophilic Algae as a valuable bioresource for algal biodiversity and for novel, industrially marketable metabolites*. *App Phycol*, 2020. **3**(1): p. 167-188.
102. Kawai, H., et al., *The macroalgal culture collection in Kobe University (KU-MACC) and a comprehensive molecular phylogeny of macroalgae based on the culture strains*. *Appl Phycol*, 2020 **3**(1): p. 159-166.
103. Friedl, T. and M. Lorenz, *The Culture Collection of Algae at Göttingen University (SAG): a biological resource for biotechnological and biodiversity research*. *Procedia Environmental Sciences*, 2012. **15**: p. 110-117.
104. Surek, B. and M. Melkonian, *CCAC – Culture Collection of Algae at the University of Cologne: A new collection of axenic algae with emphasis on flagellates*. *Nova Hedwigia*, 2004. **79**: p. 77-92.
105. Edvardsen, B., R. Skulberg, and O. Skulberg, *NIVA Culture Collection of Algae - Microalgae for science and technology*. *Nova Hedwigia*, 2004. **79**: p. 99-114.
106. Kasai, F., et al., *NIES-Collection, List of Strains: Microalgae and Protozoa*. National Institute for Environmental Studies, 2004.
107. Blackburn, S.I., et al. *Australian National Algae Culture Collection-unique biodiversity informing ecosystem knowledge and bioresources*. in *IOth International Phycological Congress*. 2013.
108. Spitzer, M., et al., *BoxPlotR: a web tool for generation of box plots*. *Nature Methods*, 2014. **11**(2): p. 121-122.