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Exploring carbonate rock wettability across scales: role of (bio)minerals

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Abstract

Hypothesis: The wettability of carbonate rocks is expected to be affected by the organic components of biominerals which are complex, nanostructured organo-mineral assemblages, whose surfaces have a potential influence on wetting properties. Elucidating the nanoscale mechanisms driving the wettability of solid surfaces will enable a better understanding and/or control of various geological, environmental and industrial processes.

Experiments: Using Atomic Force Microscopy and Spectroscopy (AFM/AFS) we probed the wettability properties of carbonate rocks with different amounts of organic material. The adhesion properties of two types of limestones were determined in liquid environments at different length scales (nm to mm) using functionalized tips with different chemical groups to determine the extent of surface hydrophobic and hydrophilic organo-mineral interactions.

Findings: We observed homogeneous hydrophobic areas at length scales below $<5 \mu\text{m}$. The origin of this hydrophobicity is linked to the presence of organics, whose amount and spatial distribution depend on the rock composition. Specifically, our results reveal that the biogenic vs. non-biogenic origin of the mineral grains is the main rock property controlling the wettability of the solid surface. Overall, our methodology offers a multi-scale approach to unravel the role that organic moieties and biominerals play in controlling the wettability of rock-water interfaces.

Keywords: wettability, carbonates, biominerals, adhesion force, AFM/AFS

35 1. Introduction

36 Wettability is one of the key properties controlling fundamental physicochemical
37 interfacial processes in fields such as the environmental sciences, geology, biology,
38 material science and civil engineering. This includes the fluid transport of organic
39 pollutants in the subsurface,[1] radwaste geological storage,[2] nucleation and growth of
40 biominerals,[3] uptakes of nutrients by plant roots,[4] development of novel filtration
41 systems with high physical stability[5], geological CO₂ sequestration,[6,7] enhanced oil
42 recovery,[8] proton exchange membrane in fuel cells,[9] water desalination
43 systems,[10,11] or conversion of CO₂ in the cement and concrete industry.[12]

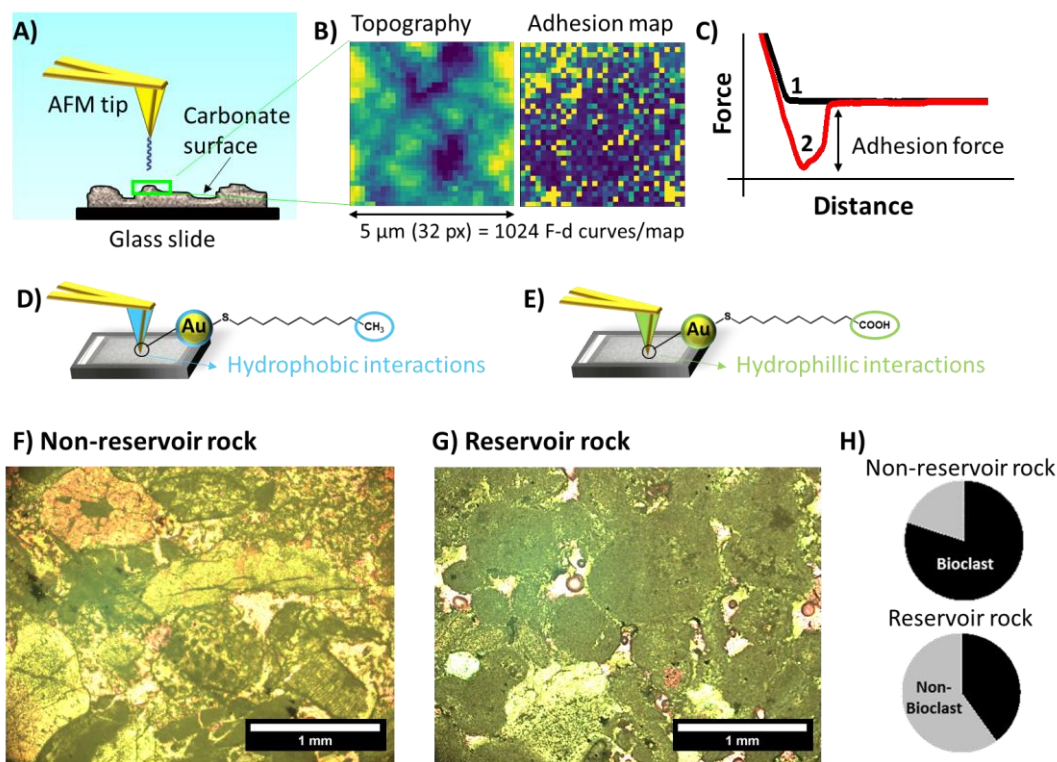
44 The term wettability describes the preference of a solid surface to be in contact with
45 one type of liquid rather than another one, which is essentially determined by
46 intermolecular attractive interactions (adhesion forces).[13] Although wettability is very
47 often reduced to hydrophilic (water-wet) and hydrophobic (oil-wet) behaviors,[14] this
48 simplification is masking the complexity present at real surfaces, especially when
49 investigating heterogeneous systems such as natural rocks. Most commonly, rock
50 wettability is determined macroscopically using contact angle measurements.[15–20]
51 However, the question arises if this type of measurement is representative of the
52 wettability of complex systems where the surface heterogeneities are smaller than the
53 droplet size (~0.05 mL) that is deposited on the rock surface. Probing molecular
54 interactions at the rock-water interface could shed light on this matter, providing not
55 only fundamental insights on the influence that such heterogeneities have on the rock
56 wettability at the nanoscale but also the physical and chemical mechanisms behind the
57 complex concept of surface wettability.

58 Surface imaging and molecular interactions can be determined in liquid media with
59 Atomic Force Microscopy (AFM) and Spectroscopy (AFS). In AFS, an array of force-
60 distance measurements between the AFM tip and the sample surface are recorded, from
61 which adhesion forces can be extracted (**Fig. 1**). In addition, the functionalization of the
62 AFM tips with organic moieties allows discerning mechanistic insights into the
63 interactions between specific organic functionalities and the scanned surfaces.[21–23]
64 This approach has led to significant progress in the exploration of rock wettability at the
65 atomic scale during the last few years.[24–28] For example, various AFS studies on
66 minerals have demonstrated the different wetting behavior induced by adsorbed or

67 dissolved ions (e.g. Mg^{2+} or SO_4^{2-}) or the presence of adsorbed organic matter.[29–35]
68 Nevertheless, little attention has been paid to understanding the physical and chemical
69 properties of the natural rock components, which will ultimately control the wettability
70 in natural environments.

71 The ubiquitous presence in the nature of carbonate rocks and their abundant
72 industrial uses have resulted in extensive research on the comprehension of their
73 wettability.[36–38] However, carbonate rocks are highly heterogeneous in terms of
74 surface roughness, porosity and chemical composition,[39,40] and despite many years
75 of study, there is still not a general description of their wetting behavior. Importantly,
76 carbonate rocks are usually formed through sedimentation processes, via the
77 accumulation and lithification of fragments of micro-organism remains, which results in
78 a large variety of structures, compositions – potentially including organic matter – and
79 textures made of fossilized biominerals.[41] Biominerals are natural nanocomposite
80 materials, composed of a hierarchical organization of organic molecules and a mineral
81 matrix. This organization leads to the inherent heterogeneity of carbonate rocks down to
82 the nanoscale.[42–44] Elucidating the role of the organic-inorganic nanostructures of
83 the biominerals is paramount to understand the wettability of the rock surfaces, a task
84 up to now never studied.

85 Here, we present a detailed investigation of molecular interactions at the carbonate
86 rock-water interface using AFM/AFS (**Fig. 1A**). We have simultaneously measured
87 topography and adhesion force maps (**Fig. 1B**) of the carbonate rock surfaces where
88 each pixel corresponds to a single force-distance curve (**Fig. 1C**). From these curves, we
89 determine the intermolecular – attractive or repulsive – forces between the sample and
90 the tip either during the approach (position 1 in **Fig. 1C**) or retract step of the tip
91 (position 2 in **Fig. 1C**). To measure hydrophobic/hydrophilic forces at the molecular
92 level, that is, to evaluate the solid wettability properties, we used AFM tips coated with
93 a monolayer of alkyl chains with non-polar ($-\text{CH}_3$, **Fig. 1D**) and polar ($-\text{COOH}$, **Fig.**
94 **1E**) end functional groups, that is, hydrophobic and hydrophilic nature, respectively.
95 We performed this study from the microscale —where bioclasts can be identified— to
96 the nanoscale and evaluated the relevant spatial scale where areas with heterogeneous
97 surface wettability can be observed.



98

99 **Fig. 1.** A) Schematic representation of the functionalization strategy used in this work by using
 100 AFM tips to measure intermolecular forces (adhesion forces) at the rock-water interface. We
 101 used non-polished thin sections of the carbonate rocks and performed AFS experiments where
 102 B) topographical and adhesion force maps are obtained. Each pixel of those maps corresponds
 103 to force-distance curves as the one represented in C). We used AFM tip coated with alkyl chains
 104 with D) methyl (-CH₃) and E) carboxyl acid (-COOH) end groups to study hydrophobic and
 105 hydrophilic interactions, respectively. F) Optical image of the non-reservoir and G) reservoir
 106 rock took using plane-polarized light (PPL) and H) pie chart containing the percentage of
 107 bioclasts (in black) for each rock which shows a higher content of bioclasts in the non-reservoir
 108 rock which are quite well-preserved.

109 The samples under study are two limestone rocks, one from southern France – a non-
 110 reservoir rock (**Fig. 1F**) – and one from the Guinea gulf – an oil reservoir rock (**Fig.**
 111 **1G**). The non-reservoir rock is a limestone sample with a rudstone-packstone texture,
 112 with different types of porosity, and mainly composed of bioclast-type grains
 113 (echinoderms, bryozoans, red algae, mollusks, small benthic foraminifera). The
 114 reservoir rock is a limestone sample with a grainstone-packstone texture, with different
 115 types of porosity, and is mainly composed of peloid-type grains and a few bioclast-type
 116 grains (echinoderms, mollusks). The use of these two contrasting rocks with significant
 117 differences in the bioclastic abundance (**Fig. 1H**) as well as in the adsorbed organic
 118 matter (potentially higher in the oil reservoir rock), allows determining the effect on
 119 wetting properties of foreign organic matter over the organic remains inherent from the
 120 bioclasts. Importantly, our observations at the nanoscale are carefully performed in

121 selected regions previously identified from optical microscopy and classified as bioclast
122 (fragments from identified biomineral remains) vs. non-bioclast (fragments non-
123 recognized as having a direct biomineral origin, likely peloids or intraclasts from
124 diagenetic recrystallization) grains. This approach allows for unraveling the role of
125 organic matter and the role of biominerals in the wettability of carbonate rocks.

126 **2. Materials and methods**

127 *2.1. Materials*

128 Blocs of limestone samples were provided by TotalEnergies collected from South
129 East France (a non-reservoir limestone) and central Africa (a reservoir limestone). The
130 non-reservoir limestone was formed during the Upper Tertiary period, Miocene epoch,
131 Burdigalian stage. It comprises mostly calcite (>99%) with traces of quartz. This
132 limestone is a bioclastic carbonate rock with a dual porosity system with macro and
133 micropores (inter- and intragranular) and dual pore throats distribution with a maximum
134 of ~6.8 and ~0.2 microns. Its specific area is ~0.46 m²/g, the total porosity is around
135 32% and permeability is ~100-300 mD. The reservoir limestone is a medium to coarse
136 oncoidal-bioclastic grainstone of mostly calcite (>95%) with dual porosity (inter- and
137 intragranular porosity) and dual pore throats distribution with a maximum at ~10 and
138 ~0.1 microns. Its total porosity is around 20% and permeability is ~100-200 mD. The
139 core of both limestones was washed with toluene and isopropanol before their use to
140 avoid any possible organic contaminant. Thin sections (40 µm thick) were used for all
141 the experiments shown in this work. These sections were non-covered and non-polished
142 to modify as less as possible the topographical and surface features of the different
143 components of the carbonate rocks.

144 *2.2. Characterization of materials*

145 A polarizing light microscope (Olympus) was employed at 5-10X magnification for
146 petrographic observations and paleontological identification of the constituents of the
147 carbonate rock. Phase analysis was performed using X-Ray Diffraction (XRD), Bruker
148 D 8 Advance v1 on the carbonate thin sections using Cu K α radiation. Raman spectra
149 were obtained with a LabRAM Soleil Horiba Raman microspectrometer with 532 nm
150 wavelength laser-excitation and 1 µm spot size. Peaks were fitted using mixed
151 Lorentzian functions using Quasar software.[45] A laser power of 15 mW was used,

152 which we confirmed to be sufficiently low to avoid damage in the sample by the laser.
153 Fluorescence spectra were obtained with an experimental device for stationary and
154 time-resolved fluorescence. Both fluorescence spectra were acquired by a nanosecond
155 pulsed YAG laser beam excitation (266 and 355 nm, 1 μ J). The sample was set on two
156 axes translation stage to achieve fluorescence maps. With a confocal optical set-up, the
157 15 μ m focused beam excited the sample orthogonally and the fluorescence scattering is
158 analyzed by a monochromator (Jobin, Yvon Micro HR) and detected with a back-
159 illuminated CCD (Syncerity S10420). Spectra were simulated by a linear combination
160 of lognormal curves with a MATLAB script (least square fit). With the same set-up for
161 excitation, the detection of fluorescence decay was measured by the way of a filter
162 wheel and a Photomultiplier device (Hamamatsu H9305-01). The electric signal was
163 recorded with an oscilloscope after pre-amplification (200 mHz of bandwidth). This
164 detection setup allows the detection of decay longer than 5 ns. After correction, decays
165 were simulated by way of an exponential fit and a power-like law (Jakub Włodarczyk
166 and Borys Kierdaszuk, 2003) (MATLAB script).

167 2.3. *Contact angle measurements*

168 Static contact angles were measured on the untreated thin sections of the reservoir
169 rock and of the non-reservoir rock. The measurements were performed with a Tracker
170 Tensiometer from Teclis at room temperature. 2 μ L-drops of brine (composition:
171 NaCl=32.3 g/L; MgCl₂.6H₂O=12.6 g/L; CaCl₂.2H₂O=1.7 g/L) were deposited on the
172 samples using an automatic procedure. 12 and 7 drops were deposited in line for the
173 non-reservoir rock and the reservoir rock respectively, to have statistical results. For all
174 drops, the contact angle was measured after 120s.

175 2.4. *Atomic Force Microscopy and Spectroscopy*

176 The topographical features of the limestone sample were evaluated using Atomic
177 Force Microscopy (AFM) operating in contact and dynamic mode.[46] All
178 measurements were performed using a MFP-3D microscope from Asylum Research
179 (Santa Barbara, USA) with a maximum planar range of the piezo scanner of 120 μ m in
180 the planar direction (x,y) and 15 μ m in the vertical direction (z). The microscope is
181 isolated inside a chamber and further, it contains a vibration isolation control unit from
182 Herzan. Complementary, the AFM contains an inverted optical microscope mounted on

183 the AFM head that enables to record optical images of the cantilever and the sample. All
184 the AFM images were acquired by using triangular silicon nitride (PNP-TR from
185 NanoWorld) with a nominal length of 200 μm , a width of 28 μm , and a thickness of 500
186 nm. The employed AFM probes had a nominal spring constant of $0.08 \text{ N}\cdot\text{m}^{-1}$ and were
187 routinely calibrated using the thermal method. All the obtained images were processed
188 using the AR and WSxM software.[47]

189 Owing to the ability of thiols to form self-assembled monolayers (SAMs) on certain
190 gold substrates, we functionalized gold-coated AFM tips (PNP-TR-Au from
191 NanoWorld) with 35 nm of gold coating. The functionalization protocol starts with a
192 UV/O₃ plasma treatment to clean gold coated-AFM tips for 20 minutes and after that,
193 the tips are immersed into a 2mM ethanolic solution of the desired thiol for at least 24
194 hours. In this work, we have used undecanethiol and 11-mercaptoundecanoic acid to
195 obtain hydrophobic and hydrophilic AFM tips, respectively. We used these
196 functionalized tips to measure adhesion force maps of 32 x 32 px (a pixel corresponds
197 to the region where a force-distance curve is measured and from where adhesion force is
198 determined), containing thus 1024 force-distance curves for each adhesion force map.
199 We further covered different spatial scales from 60 μm to 1 μm which corresponds to
200 spectral resolutions from 2 $\mu\text{m}/\text{px}$ to 30 nm/px.

201 All the force spectroscopy measurements were performed in low salinity conditions
202 (1 mM NaCl solution) using a loading force of 2 nN to avoid damage to the
203 hydrocarbon monolayer. We also checked if we unintentionally modified the rock
204 surface by depositing the organic monolayer of the functionalized tip during the AFS
205 measurements across the scales which were performed keeping the same center and just
206 reducing the image size. Thus, we routinely measured again the largest area (60 μm x
207 60 μm) and confirmed that neither the adhesion force nor the surface roughness of the
208 carbonate surface was not altered by the force spectroscopy measurements (**Fig. S1**).

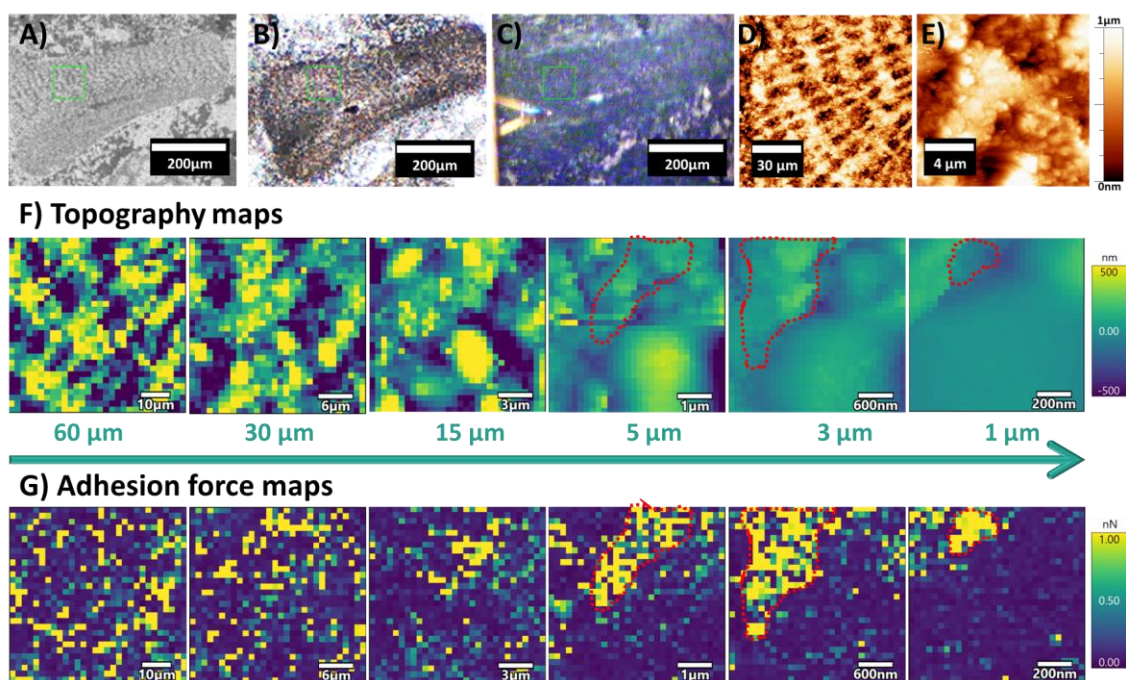
209 **3. Results and discussion**

210 *3.1. Does the scale of observation play a role in determining the wettability of*
211 *carbonate rocks?*

212 When studying the wettability properties of heterogeneous surfaces, such as
213 carbonate rocks, the scale of observation can play an important role. At the macroscale,

214 we measured the contact angle of our carbonate rocks (**Fig. S2**). For the non-reservoir
215 and reservoir rocks, we obtained average values of $63.8 \pm 3.4^\circ$ and $73.4 \pm 3.9^\circ$ (mean \pm
216 SD) respectively, which indicates a more hydrophobic behavior of the reservoir rock,
217 although information about any potential heterogeneities is unavailable at this point.
218 Observations of both limestone rocks using SEM and polarizing optical microscopy
219 enabled the identification of the biologic constituents, as exemplified by the red algae
220 fragment of the non-reservoir rock sample (**Fig. 2A-C**). Such identification is
221 fundamental for further measurements of the wettability properties of individual
222 components of the carbonate rock from the micro- to the nano-scale.

223 To study the molecular interactions at the rock-water interface, we performed AFS
224 experiments on the limestone surfaces over a spatial range from 60 μm down to 1 μm ,
225 covering thus spatial resolutions from 2 $\mu\text{m}/\text{px}$ to 30 nm/px . Making use of an optical
226 microscope integrated into the AFM unit, we intentionally placed the tip in the region of
227 interest for the wettability measurements (**Fig. 2C**) and imaged the carbonate surface at
228 different scales (**Fig. 2D,E**). We used CH_3 -functionalized tips to directly probe
229 hydrophobic interactions by simultaneously measuring the surface topography and
230 adhesion force maps of the surface of carbonate bioclots (i.e. red algae in **Fig. 2F,G**
231 and for other examples **Fig. S3**). From the topography maps, we determined the surface
232 roughness of the rock, which decreased across the scales from 500 to 80 nm, while from
233 adhesion force maps we observed minor differences across the studied spatial scales
234 (**Fig. S4**). Only below 5 μm image size, we observe homogeneous hydrophobic regions
235 which are highlighted with a contoured red dotted line in **Fig. 2G**. Although the
236 influence of surface roughness on the macroscopic wettability properties has been
237 reported previously,[48,49] we did not observe such a roughness effect on the adhesion
238 data in the studied spatial resolution (**Fig. S5**). This suggests that nano-wettability is
239 rather influenced by differences in the chemical composition of the rock surface.
240 Overall, these results confirm that the scale of observation is an important parameter
241 when studying rock surface wettability.



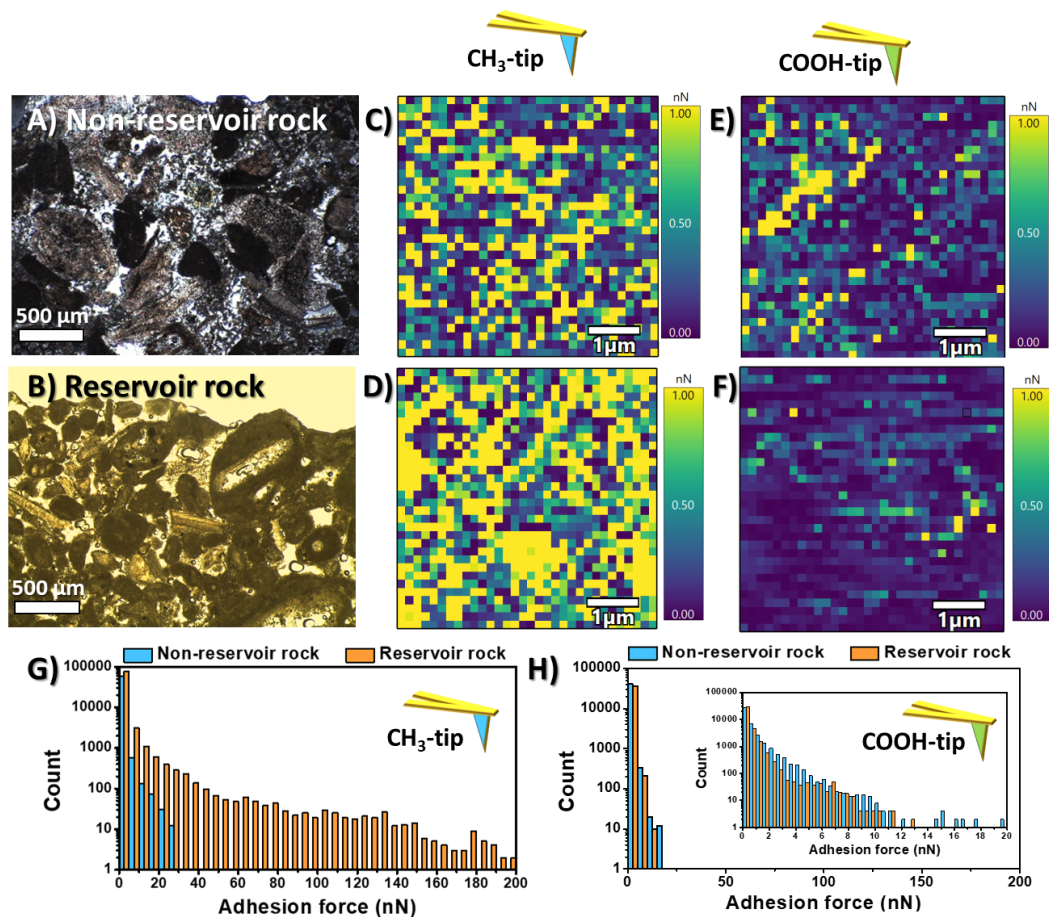
242 **Fig. 2.** Images of a bioclast of the non-reservoir rock that contains a red algae fragment obtained
 243 by: A) SEM, B) PPL optical microscope and C) inverse optical microscope coupled with AFM
 244 microscope and where the AFM tip is also presented. D,E) AFM images of the red algae area
 245 highlighted with a green square in images A-C at 120 and 20 μm image size. E) Topography
 246 and F) adhesion force maps obtained with a hydrophobic tip in a range of image sizes from 60
 247 to 1 μm , covering resolutions of 2 $\mu\text{m}/\text{px}$ to 30 nm/px . Only below an image size of 5 μm , we
 248 observe homogeneous hydrophobic regions (highlighted with a contoured dotted line in red).
 249

250 3.2. Does adsorbed organic matter play a role in the wettability of carbonate rocks?

251 Once the relevant scale of observation is identified, the nano-wettability of the
 252 limestone rocks was explored using image sizes of 5 μm (where we observed
 253 homogeneous hydrophobic regions) and acquiring force maps with a resolution of 150
 254 nm/px . The adhesion properties of the non-reservoir (**Fig. 3A**) and a reservoir (**Fig. 3B**)
 255 rock samples were studied using CH_3 - and COOH -functionalized tips as non-polar and
 256 polar probes to mimic hydrophobic and hydrophilic organic-carbonate interactions,
 257 respectively. Representative adhesion force maps obtained with the CH_3 -tip (**Fig. 3C,D**)
 258 show higher adhesive-type interactions than those obtained with COOH -tip (**Fig. 3E,F**).
 259 This is due to the higher ligand density of methyl groups enabling a better surface
 260 coverage of the AFM tip that leads to larger adhesion values, expressed as mean \pm SE.
 261 The hydrophobic interactions are especially pronounced in the reservoir rock as
 262 evidenced by the larger adhesion force data (1.73 ± 0.29 nN) found in comparison to
 263 that of the non-reservoir rock (0.68 ± 0.09 nN). In addition to the preferential interaction
 264 of the reservoir rock surface with non-polar molecules, this rock type presents a highly
 265 homogeneous surface in terms of wetting behavior. In contrast, the non-reservoir rock

266 presents distinct homogeneous sizes of hydrophobic and hydrophilic properties.
267 Furthermore, our measurements with COOH-tips led to slightly higher adhesion force in
268 the non-reservoir rock (0.74 ± 0.1 nN) than in the reservoir rock (0.68 ± 0.09 nN).
269 Looking at representative force-distance curves of both carbonate rocks using CH₃- and
270 COOH-tips (**Fig. S6**), we observed that the approaching curves usually present
271 electrostatic repulsion with the exception when the CH₃-tip is approaching to reservoir
272 rock surface, which usually presents Van der Waals interactions. The adhesion peaks in
273 the retracting curves correspond to the hydrophobic or hydrophilic interactions
274 depending on the use of CH₃- or COOH-tips. The reservoir rock surface presents the
275 largest adhesion peak when interacting with the CH₃-tip (7.4 nN), whereas a very small
276 adhesion peak is observed when using COOH-tip (0.1 nN), confirming its predominant
277 hydrophobic character. However, similar (low) adhesion peaks are observed in the
278 interactions of the hydrophobic and hydrophilic tips with the non-reservoir rock (1.2
279 and 0.7 nN, respectively), ascribed to a lower content of organic matter.

280 Statistical analysis of our AFS results (about 250.000 force-distance curves for both
281 rocks and both functionalized tips) was done to obtain representative wettability data of
282 the rock surface from nanoscale interactions. The statistical results are presented as
283 adhesion histograms in **Fig. 3G,H** and confirm the stronger hydrophobic behavior of the
284 reservoir rock in comparison with that of the non-reservoir rock. In agreement with the
285 macroscopic contact angle measurements (**Fig. S2**), these nanoscale measurements
286 reveal the critical role that foreign organic matter (i.e. oil) plays in controlling the
287 surface wettability.



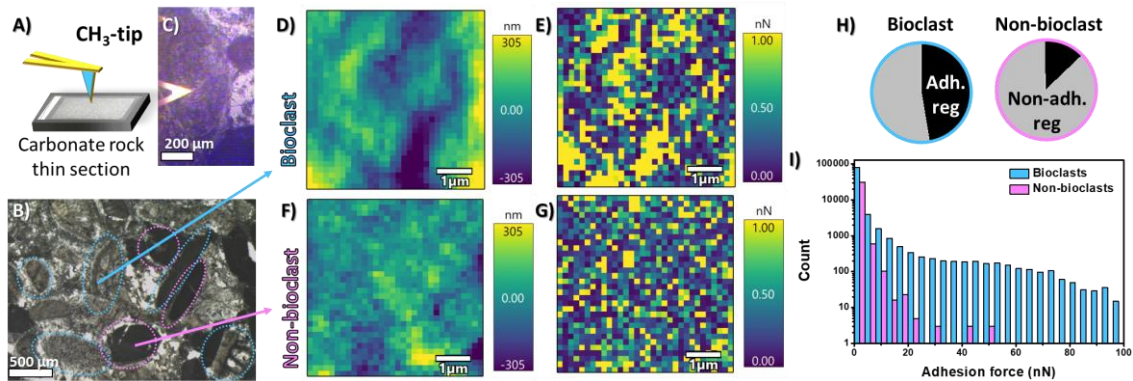
288

289 **Fig. 3.** Thin section optical images of A) non-reservoir and B) reservoir carbonate rocks where
 290 AFS measurements were performed. Representative adhesion force maps were obtained in both
 291 rocks using the hydrophobic (C,D) and hydrophilic (E,F) tips. Adhesion force histograms
 292 corresponding to 75000 force-curve data G) CH₃-tip and H) COOH-tip.

293 *3.3. Is the wettability linked to the biogenicity of a rock fragment?*

294 Hydrophobic interactions of both carbonate rocks were further explored and related
 295 to their biological origin, classifying them into different types of bioclusters. Before AFS
 296 measurements (**Fig. 4A**), the carbonate thin sections were observed with a polarizing
 297 optical microscope to identify the components of the rocks (**Fig. 4B**). We classified the
 298 rock components as bioclusters or skeletal grains with biological origin and non-biocluster
 299 including non-skeletal grains and types of cement. Then, we explored if the bioclusters
 300 influence the rock wettability of the non-reservoir and reservoir rocks using AFM and
 301 an integrated optical microscope (**Fig. 4C**). Although trivial, the identification of rock
 302 constituents from the micro- to the nano-scale has been fundamental to understand the
 303 wettability properties of the rocks. The topography and adhesion force maps of **Fig. 4D-**
 304 **G** show larger hydrophobic regions in the biocluster regions (**Fig. 4D-E**) than in the non-
 305 bioclusters ones (**Fig. 4F-G**). In fact, we calculated from multiple force maps that the

306 adhesive region in bioclusters corresponds to 47% of the imaged area, while for non-
 307 bioclusters it only represents 13% (**Fig. 4H**). Furthermore, adhesion histograms,
 308 corresponding to the analysis of 150.000 force-distance curves (**Fig. 4I**), confirm that
 309 biocluster regions interact more strongly with the hydrophobic tip than the non-biocluster
 310 ones. However, we did not observe any relevant inter-biomineral differences, i.e.
 311 echinoderm, mollusk, foraminifera, bryozoan and red algae all displayed comparable
 312 values of adhesion (**Fig. S7**).



313 **Fig. 4.** A) Schematic representation of the AFS measurements using CH₃-tips and carbonate
 314 thin sections. B) Optical micrograph of the non-reservoir carbonate rock thin section. D)
 315 Topography and E) adhesion force maps of bioclusters in comparison with F) topography and G)
 316 adhesion force maps of non-bioclusters. H) Pie-chart representing the percentage of adhesion
 317 region measured over the total map area for bioclusters and non-bioclusters. I) Adhesion force
 318 histograms of the AFS measurements.

320 The significant difference in the wettability (**Fig. S8**) between bioclusters (0.80 and
 321 2.89 nN for non-reservoir and reservoir rock) and non-bioclusters (0.56 and 1.35 nN for
 322 non-reservoir and reservoir rock) can be ascribed to the presence of organic remains
 323 within the former (indeed, biominerals are composite organo-mineral materials). The
 324 organic remainders favor interactions with non-polar organic molecules. Fossilized
 325 biominerals can therefore induce hydrophobicity, also facilitating the adsorption of
 326 organic matter (i.e. in oil reservoirs) through non-polar interactions.

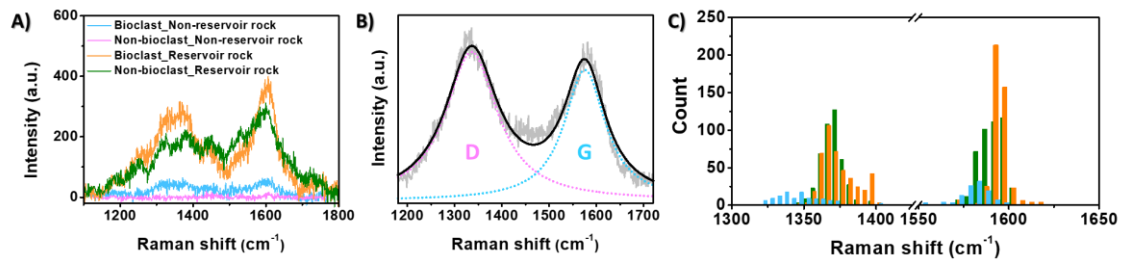
327 3.4. Evaluating the role of organic matter on the wettability properties of carbonate 328 rocks.

329 Based on our nanoscale observations, we have established that a more hydrophobic
 330 behavior is present in the reservoir rock. Furthermore, we have observed a significant
 331 difference between the wetting properties of biocluster and non-biocluster grains. These
 332 results highlight the key role that organic matter, either adsorbed hydrocarbons from

333 crude oil or gas or the breakdown products of biomolecules included in biominerals,
334 plays on the wettability (hydrophobicity) of rock surfaces. To identify the presence of
335 organic matter in the carbonate rock surface, we performed Raman microspectroscopy
336 on thin sections of both limestone rocks. In addition to the calcite peaks in the low
337 wavenumber region (calcite is the dominant calcium carbonate polymorph –**Fig. S9**),
338 we also detected the fingerprints of organic matter in the higher wavenumber region
339 (**Fig. 5A**). In particular, we observed the characteristic D and G bands of graphitic
340 structures, which are attributed to in-plane defects and sp^2 C-C stretching vibrations
341 within the aromatic rings of the graphitic layer, respectively.[50] The presence of wide
342 D and G bands with similar intensities indicates the presence of disordered organic
343 matter in all the samples. However, their distribution on the surface differs: the
344 bioclasts, non-bioclasts and cement components of the reservoir rock present a
345 homogeneous distribution on the rock surface (**Fig. S10**). But, for the non-reservoir
346 rock, the signal from organic matter was only presented in the bioclastic grains. These
347 results support the idea that the presence of organic matter from a natural origin is
348 responsible for the hydrophobic behavior. Also, these maps with a resolution of 1
349 $\mu\text{m}/\text{px}$ agree well with the spatial heterogeneity observed in the wetting properties at
350 small spatial scales.

351 The position and broadening of D and G bands were determined by simultaneous
352 peak fitting to Lorentzian profiles after baseline correction to remove the fluorescence
353 contribution of the organic matter using Quasar software[45] (**Fig. 5B**). Based on the
354 Raman observations in the bioclasts of the non-reservoir rock (**Fig. 5C**), the position of
355 the G band at 1580 cm^{-1} corresponds to that of crystalline graphitic structure and the D
356 band at 1347 cm^{-1} of high intensity indicates the presence of heteroatoms such as
357 hydrogen, nitrogen or oxygen in the aromatic rings of the organic matter.[51] For the
358 reservoir rock, we observed a significant blue-shift in the D and the G bands, being
359 larger for the bioclasts (1375 and 1595 cm^{-1}) than for the non-bioclasts, including grains
360 and cements (1370 and 1591 cm^{-1}). The shift in the G peak has been attributed to the
361 presence of hydrocarbons adsorbed on the rock surface from the crude oil.[52] The G
362 peaks of the bioclasts are also narrower than those of the non-bioclust grains and
363 cements (**Fig. S11**), which reveals a higher degree of graphitization of the organic
364 molecules in the bioclast grains. This effect is also reflected in the shift of the D peak of

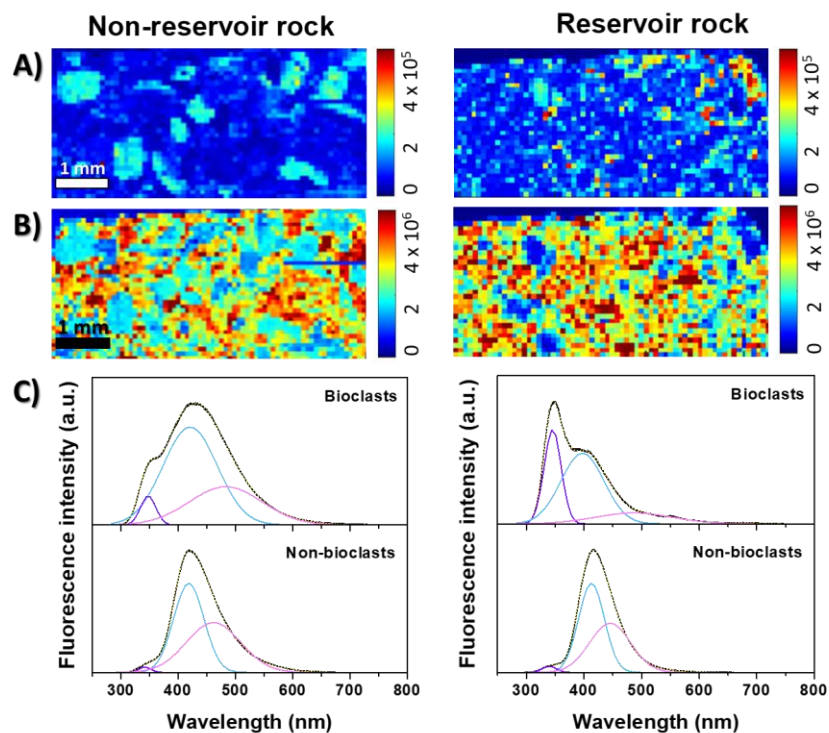
365 the reservoir rock, which has been previously attributed to a higher degree of maturation
366 of the organic matter in rocks.[53]



367

368 **Fig. 5.** A) Raman spectra of C-signal region for the bioclasts and non-bioclasts of (including
369 grains and cements) non-reservoir and reservoir rocks. B) Example of a double Lorentzian fit
370 ascribed to the D (pink) and G (blue) bands of graphitic structures. C) Distributions of D and G
371 band position obtained from Raman mapping of 10 x 10 μm x μm spatial areas with a resolution
372 of 0.5 μm for the bioclasts of the non-reservoir rock (blue) and the bioclasts (orange) and non-
373 bioclasts (green) of the reservoir rock.

374 Additionally, we explored the fluorescence of organic matter contained in our
375 carbonate rocks in the wavelength range of 300-800 nm using an excitation wavelength
376 of 266 nm. We explored a large area of 18 mm² on thin sections of both carbonate rocks
377 to observe the emission of the different components. Fluorescence intensity maps at 340
378 nm (**Fig. 6A**) and 420 nm (**Fig. 6B**) of non-reservoir and reservoir rocks reveal that the
379 former contribution is characteristic of the bioclast grains. Here, the spatial resolution of
380 the fluorescence maps does not allow for the observation of heterogeneities in the
381 nanoscale but still gives relevant insights into the composition of the organic matter of
382 the carbonate rocks. **Fig. 6C** shows representative spectra of bioclast and non-bioclast
383 grains in both rocks and their fits. The fit results clearly show the presence of the peak
384 around 340 nm (purple line) in the bioclasts – especially that of the reservoir rock –
385 whereas it is negligible for the non-bioclasts. The fluorescence around 420 nm (blue
386 line) is the main component of all the rock constituents. This peak is considerably wider
387 in the bioclasts, attributed to the presence of multiple fluorophores although less intense
388 than in the non-bioclasts. Finally, we observed a third contribution (magenta line) with
389 an emission center around 500 nm in the bioclasts and shifted to 450 nm in the case of
390 the non-bioclasts grains.



391

392 **Fig. 6.** Stationary fluorescence intensity maps for wavelengths of A) 340 and B) 450 nm for
 393 non-reservoir (left) and reservoir rocks (right) with a resolution of 100 μm . C) Representative
 394 fluorescence spectra of bioclasts and non-bioclasts for non-reservoir (left) and reservoir rocks
 395 (right) together to their fit using 3 gaussian peaks around 340 (purple), 420 (blue) and 500 nm
 396 (magenta).

397 We have further measured decay spectra in the bioclasts and non-bioclaster grains of
 398 both rocks for the three contributions observed in the stationary fluorescence. The decay
 399 times were simulated with Powerlike function estimations[54] for each of the center
 400 wavelengths using and passband filter with 10 nm wavelength width. For the 340 and
 401 the 420 nm bands, decay times are low and close to the detection limit of our setup,
 402 typically between 5 and 6 ns. These results suggest that the fluorescence at 340 and 420
 403 nm could correspond to small organic molecules rather than polyaromatic molecules
 404 which are known to exhibit a long-time decay fluorescence.[55,56] The 500 nm
 405 centered band shows a longer fluorescence decay time on the bioclast, longer than 8 ns
 406 whereas, for non-bioclaster, fluorescence decay time remains between 5 and 6 ns.
 407 Furthermore, we observed a high sensitivity of the contribution around 340 and 420 nm
 408 to photodegradation (**Fig. S12**) whereas the contribution around 500 nm remains
 409 constant. This result together with the longer time decay on bioclast indicates that some
 410 hydrophobic compounds such as protein-like or humic-like (also referred to as
 411 unidentified natural organic compounds – UNOC) are present in the carbonate rocks
 412 and partially trapped by the bioclaster.[57,58] Overall, we found that bioclaster show a

413 different fluorescence spectrum than that of the non-bioclats with a contribution around
414 340 nm that furthermore, becomes the main component in the reservoir rock. Since
415 these results are highly in concordance with the wettability of the bioclats, we ascribe
416 the emission at 340 nm to organic matter of organic behavior with a plausible biological
417 origin.

418 **4. Conclusion**

419 There is a common use to describe the wettability of a surface sample by using
420 contact angle measurements. However, the macro-scale approach of such measurements
421 only provides an average bulk value of the solid surface, missing thus valuable
422 information from the nanoscale. Our approach to probing molecular interactions within
423 carbonate rocks allows elucidating the complexity of rock surface wettability, especially
424 in heterogeneous natural systems, by exploring surface heterogeneities from the micro-
425 to the nano-scale. We demonstrate that the scale of observation is an important
426 parameter when describing the wettability properties of natural rocks, being necessary
427 to explore areas below $25 \mu\text{m}^2$ to study heterogeneities of the carbonate rock in terms of
428 wetting behavior. At this length scale, preferential interaction (stronger adhesion force)
429 with hydrophobic molecules showed a homogeneous wetting behavior of the reservoir
430 rock. However, at the same length scale, the non-reservoir rock presented heterogeneous
431 hydrophobic regions only at the surface of biominerals. Furthermore, our methodology
432 with the identification of individual components of the carbonate rock from the micro-
433 to the nano-scale has been fundamental to explore the wettability of specific bioclast
434 and non-bioclast grains resulting that the former presenting higher hydrophobic
435 behavior. Such hydrophobicity can be induced in the bioclast grains by facilitating the
436 adsorption of organic matter through non-polar interactions. The combination of
437 adhesion force maps with micro-Raman and fluorescence spectroscopy maps have
438 confirmed the hypothesis that organic remains within the biominerals are the main
439 responsible for the hydrophobic character of bioclast grain surfaces. The fluorescence
440 emission of the carbonate rocks corresponds mainly to organic matter which
441 experiments photobleaching at long time exposition. The main signals correspond to
442 protein-like or humic-like compounds and their short decay times reveal that small
443 molecules with certain aromaticity are present in the rock. A high degree of maturation,
444 observed in the position of the C signal in the Raman spectra, elucidates that small
445 molecules with natural origin (i.e. amino acids) can be present within the rock which

446 leads to homogeneous hydrophobic regions. All results together highlight the critical
447 role that the biogenic origin of the minerals plays in the wetting behavior of the studied
448 limestones at the nanoscale, which had never been studied up to now. Furthermore, this
449 could be used as a potential parameter to control the wettability properties of carbonate-
450 water interfaces.

451 **CRedit authorship contribution statement**

452 **Alicia Moya:** Investigation, Data curation, Visualization, Formal analysis,
453 Methodology, Validation, Writing – original draft, Writing – review & editing.
454 **Fabienne Giraud:** Investigation, Validation, Writing – review & editing. **Valerie**
455 **Molinier:** Investigation, Resources, Writing – review & editing. **Yves Perrette:**
456 Investigation, Data curation, Formal analysis, Methodology, Writing – review &
457 editing. **Laurent Charlet:** Conceptualization, Resources, Supervision, Funding
458 acquisition, Writing – review & editing. **Alexander Van Driessche:** Conceptualization,
459 Resources, Supervision, Funding acquisition, Writing – review & editing. **Alejandro**
460 **Fernandez-Martinez:** Conceptualization, Resources, Supervision, Funding acquisition,
461 Writing – review & editing.

462 **Data availability**

463 The data that support the plots within this paper and other findings of this study are
464 available from the corresponding author upon reasonable request.

465 **Declaration of Competing Interest**

466 The authors declare no competing interests.

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471 **Appendix A. Supporting material**

472 Supplementary data to this article can be found online at

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