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# Flower development in the Solanaceae

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**Running head:** Molecular mechanisms of flower development in the nightshades (Solanaceae)  
family

**Keywords:** flower / development / Solanaceae / nightshades / ABC model / floral organ identity /  
petunia / tomato / Physalis

## 17 **Introduction**

18 Flower development is the process leading from a reproductive meristem to a mature flower with  
19 fully developed floral organs. This multi-step process is complex and involves thousands of genes  
20 in intertwined regulatory pathways; navigating through the FLOR-ID website will give an  
21 impression of this complexity and of the astonishing amount of work that has been carried on the  
22 topic [1]. Our understanding of flower development mostly comes from the model species  
23 *Arabidopsis thaliana*, but numerous other studies outside of Brassicaceae have helped apprehend  
24 the conservation of these mechanisms in a large evolutionary context [2–4]. Integrating additional  
25 species and families to the research on this topic can only advance our understanding of flower  
26 development and its evolution.

27 In this chapter, we will review the particular contribution that the Solanaceae family (Fig. 1)  
28 has made to the comprehension of flower development. While many of the general features of  
29 flower development (i.e. the key molecular players involved in flower meristem identity,  
30 inflorescence architecture or floral organ development) are similar to *Arabidopsis*, our main  
31 objective in this chapter is to highlight the points of divergence, and emphasize specificities of the  
32 Solanaceae. We will not discuss the large topics of flowering time regulation, inflorescence  
33 architecture and fruit development, and we will restrict ourselves to the mechanisms included in a  
34 time window after the floral transition and before the fertilization. Moreover, this review will not be  
35 exhaustive of the large amount of work carried on the topic, and the choices that we made to  
36 describe in large details some stories from the literature are based on the soundness of the functional  
37 work performed, and surely as well on our own preferences and expertise.

38 First, we will give a brief overview of the Solanaceae family and some of its specificities.  
39 Then, our main focus will be on the molecular mechanisms controlling floral organ identity, for  
40 which extended functional work in petunia led to substantial revisions to the famous ABC model.  
41 Finally, after reviewing some studies on floral organ initiation and growth, we will discuss floral  
42 organ maturation, using the examples of the inflated calyx of the Chinese lantern *Physalis* and  
43 petunia petal pigmentation.

#### 44 **The Solanaceae family (nightshades): crops and model species**

45 The Solanaceae, also called nightshades, belong to the Asterids clade in the eudicots and gather  
46 more than 2,500 species and 115 genera [5]. The family has a world-wide distribution but the  
47 greatest diversity in species is found in south and central america, probably reflecting the origin of  
48 the family. The Solanaceae include many agronomically important crops, such as tomato (*Solanum*  
49 *lycopersicum*), tobacco (*Nicotiana tabacum*), potato (*Solanum tuberosum*), petunia (*Petunia x*  
50 *hybrida*), pepper (*Capsicum annuum*) or eggplant (*Solanum melongena*) (Fig. 1). Several members  
51 of the family produce alkaloid compounds with an agronomical interest, such as nicotin in tobacco,  
52 or psychoactive or poisonous substances such as those found in belladonna, stramonium or  
53 mandrake. Indeed the name Solanaceae might come from the latin *solare*, meaning "to soothe", in  
54 reference to the pharmaceutical properties of many members of the family. The Solanaceae display  
55 a large variety of inflorescence structures but the family is typified by the characteristic cymose  
56 inflorescence, where the terminal flower dies out and new flowers grow from lateral buds. The  
57 flowers generally have a type-5 symmetry with 5 petals fused at different degrees and 5 stamens  
58 partly fused to the corolla, and the ovary can develop into either a fleshy (e.g. tomato) or dry (e.g.  
59 petunia) fruit.

60 The Solanaceae contain model species such as petunia, tomato, potato and tobacco, that can  
61 be easily transformed with *Agrobacterium tumefaciens*, and whose genomes have been sequenced  
62 (<https://solgenomics.net/>) [6]. These species of agronomical interest have been bred to improve  
63 specific traits, and therefore constitute good model systems to understand the genetic processes of  
64 domestication [7]. Petunia is a famous bedding plant with high agronomic value, and hundreds of  
65 floral varieties selected over the years provide a large repertoire for variation in flower morphology.  
66 In particular, petunia has been instrumental to decipher the anthocyanin biosynthesis pathway  
67 involved in petal pigmentation [8] and evolution of flower colour in relation to changes in  
68 pollinators [9]. Moreover, a large transposon-insertion database allows for reverse genetics to be  
69 carried out easily, which distinguishes it from the other model Solanaceae species [10, 11]. In  
70 tobacco, the famous Maryland Mammoth short-days flowering mutant plant has revealed the role of  
71 photoperiodism on the floral transition [12, 13], but afterwards tobacco has been little used for  
72 research on flower development [6]. The same stands true for potato, although this species has  
73 revealed the particular parallel between flowering and tuberization, that uses a similar molecular  
74 toolkit with day-length-dependent traveling proteins [14]. Tomato was a major model species for  
75 classical genetics before the rise of Arabidopsis and is still widely used as a model fruit-bearing  
76 crop [6, 15]. In the next paragraph, we will review how members of the Solanaceae family, and in  
77 particular petunia, have contributed to a better understanding of how flowers are built.

## 78 **Specification of floral organ identity: revisions to the ABC model**

79 Environmental and endogenous signals inform the plant when to flower, and at that point, the  
80 vegetative meristem that produces leaves will turn into an inflorescence meristem that produces  
81 flowers. At an early stage, groups of cells in the flower will acquire a specific identity depending on  
82 their position, by the apposition of a precise molecular identity. In Arabidopsis, this floral organ  
83 identity is specified by the combinatorial action of A-, B- and C-class proteins, as summarized in  
84 the classical ABC model and in the floral quartet model that derives from it [16–18] (Fig. 2).  
85 Although the ABC model is generally considered valid across a wide variety of flowering plants,  
86 this often relies on expression data only and the tedious genetic work that is needed to validate the  
87 function of ABC genes has only been done in a few species. For this reason, extended work in  
88 petunia, where forward and reverse genetics can be carried at a large scale [11], has been key to  
89 extend the ABC model to species outside of Brassicaceae. In the following paragraphs, we will  
90 review this work, together with pieces of evidence from other nightshades, to illustrate how a  
91 detailed floral organ identity patterning model, quite distant to the idealized ABC model found in  
92 textbooks, has been built over the years (Fig. 2). To help the reader mostly familiar with  
93 Arabidopsis genes, we have included a phylogeny of the ABC MADS-box transcription factors  
94 discussed in this chapter, showing orthologs in petunia and tomato (Fig. 3).

95

### 96 *Redefining the A function*

97 In contrast to the B- and C-functions that are generally well conserved in flowering plants, the  
98 molecular identity of the players encoding the A-function, and even the existence of the A-function  
99 itself, have been debated [19–21]. A-class genes, as formulated in the classical ABC model, have a  
100 dual role: on one hand they antagonize the expression of C-class genes in the two first whorls of the  
101 flower, and on the other hand they specify alone the identity of sepals, and together with B-class  
102 genes the identity of petals [16, 17]. As a result, A-class mutants are expected to develop carpels in  
103 the first whorl and stamens in the second whorl of the flower. In Arabidopsis, *APETALA1* (*AP1*) and  
104 *AP2* are classified as A-class genes. The *ap1* mutant flowers lack petals and have sepals that display  
105 bract features [22, 23], whereas the *ap2* mutant sepals are converted into carpel- or leaf-like organs,  
106 and its petals are absent or transformed into stamen-like structures [24, 25], suggesting indeed that  
107 *AP2* (and *AP1* to some extent) is necessary for sepals and petals to form correctly and to repress C-  
108 class gene expression from these organs. However, such expected A-class mutants were never  
109 clearly found outside of Arabidopsis, suggesting that the A-function as initially defined is not  
110 universal [19]. Instead, in *Antirrhinum* only a gain-of-function mutant of the C-class gene *PLE*  
111 produces the expected A-class mutant phenotype [26], showing that the wild-type sepal and petal

112 identity is rather due to the repression of B- and C-class gene expression from the outer whorls of  
113 the flower, than by an "added value" of A-class genes [20, 27]. For these reasons, variations to the  
114 ABC model have been proposed, and in particular the (A)BC model [16, 20, 28]. In this model, the  
115 (A) function merely provides a floral context in which the B- and C-class genes are active to specify  
116 petal, stamen and carpel identity, whereas sepals represent the ground floral organ identity. In other  
117 words, the A-function plays a cadastral role, setting the outside boundaries of B- and C-genes  
118 expression, rather than specifying a precise floral organ identity.

119 In petunia, the cadastral function of A-class genes on C-class gene expression is clearly  
120 apparent in the double mutant for *BLIND* (*BL*) and *BLIND ENHANCER* (*BEN*), in which petals are  
121 absent or stamenoid, and sepals are converted into carpel-like structures [29]. *BL* encodes a  
122 microRNA from the miR169 family [30] while *BEN* is a member of the *euAP2* lineage [29], to  
123 which the Arabidopsis A-class gene *AP2* also belong. Therefore in petunia, the C-repression role of  
124 the A-function is fulfilled by the combinatorial action of two distinct molecular players: a  
125 microRNA and an AP2-type transcription factor. Although *BEN* encodes the functional equivalent  
126 of Arabidopsis *AP2*, they are not orthologs since *BEN* belongs to the *TOE*-type genes of the *euAP2*  
127 lineage (Fig. 3), members of which in Arabidopsis redundantly act as floral repressors [31–34].  
128 Combining the *ben* mutant with mutations in the *euAP2*-type genes *ROB1*, *ROB2* and *ROB3*  
129 (orthologous to Arabidopsis *AP2*) causes a complete homeotic conversion of sepals into petals,  
130 witnessing the full derepression of B-class genes in the first whorl [29]. In addition, mutations in the  
131 four members of the *AP1/SQUA* subfamily (*euAP1*, *PGF*, *FLORAL BINDING PROTEIN 26*  
132 (*FBP26*) and *FBP29*, see Fig. 3) results in terminal flowers with normal petals and sepals with  
133 petaloid sectors, this last feature being strongly enhanced when combined with mutations in *ROBs*  
134 genes [35]. This demonstrates the existence of a cadastral function to restrict B-class gene  
135 expression to its correct domain, fulfilled by *BEN*, *ROBs* and *AP1/SQUA* genes in petunia. In  
136 contrast, in Arabidopsis only indirect evidence for a repressive role of *AP2* on B-class genes had  
137 been reported [36]. Therefore, in petunia a combination of different molecular players act to restrict  
138 B- and C-class gene expression to their respective domains, and fulfilling altogether the cadastral  
139 part of the A-function (Fig. 2).

140 It is now clear in the Solanaceae family that *AP1*- and *AP2*-like genes are not needed to  
141 specify petal identity. In tomato, a knock-out insertion mutant in the gene *MACROCALYX* (*MC*),  
142 orthologous to the A-function gene *AP1*, shows no defects in petal identity, with sepals being  
143 enlarged like bracts [37, 38]. However, this could be masked by redundancy with other *AP1*-like  
144 genes. But as shown previously, in petunia a quadruple mutant of the four *AP1/SQUA* genes leads to  
145 terminal flowers with perfectly normal petals, and with enlarged sepals with petaloid sectors, as a

146 result of B-class gene derepression [35]. Similarly, the *rob1 rob2 rob3* mutant (*ROBs* being  
147 orthologous to Arabidopsis *AP2*) forms petals, although with some growth and pigmentation defects  
148 [29]. To conclude, neither *AP1*-like nor *AP2*-like genes are necessary for basic specification of petal  
149 identity in petunia flowers, in sharp contrast to what is known in Arabidopsis and generalized in the  
150 textbook ABC model.

151

#### 152 *Variations on the B function: Specializing for a single floral organ*

153 The classical ABC model states that B-genes specify petals and stamens, in combination with A-  
154 and C-genes respectively. Indeed, B-class mutants in eudicots usually show a phenotype affecting  
155 both the petal and stamen whorls and converting them into sepals and carpels respectively; this is  
156 for instance the case in tomato when the *APETALA3* (*AP3*) ortholog *STAMENLESS* is knocked-out  
157 [39, 40]. However, the situation is different in petunia: mutant in the B-class gene *PhDEFICIENS*  
158 (*PhDEF*), also known as *green petals*, has a full conversion of petals into sepals, but stamens  
159 remain unaffected [41]. *PhDEF* is expressed in petal and stamen primordia, suggesting that another  
160 gene redundantly controls stamen identity [42]. This gene was found to be its paralog *PhTM6*,  
161 which resembles the ancestral *paleoAP3* type of B-class genes, rather than the classical *euAP3* type  
162 B-class genes to which *PhDEF* belongs (Fig. 3) [43]. Surprisingly, *PhTM6* appeared to be  
163 expressed rather as a C-class gene in whorls 3 and 4 [44]. This unconventional expression pattern  
164 for a B-class gene is caused by the fact that C-class genes activate *PhTM6* expression [21]. The  
165 *phtm6* mutant has no visible phenotype, while the *phdef phtm6* double mutant displays full  
166 homeotic conversion of petals into sepals and stamens into carpels, as would be expected for a B-  
167 class mutant [44]. Thus in petunia, B-class genes from the *AP3/DEF* clade have specialized into  
168 controlling petal and stamen, or only stamen, identity in a partially redundant fashion (Fig. 4).

169 Furthermore, it was shown that the *AP3/DEF* proteins act in heterocomplexes with  
170 *PISTILLATA/GLOBOSA* (*PI/GLO*) proteins, also belonging to the B-class family (Fig. 3), and that  
171 this complex activates its own expression [45, 46]. For instance, Arabidopsis *AP3* interacts with *PI*,  
172 and Antirrhinum *DEF* interacts with *GLO* [47, 48]. In petunia, there are two *PI/GLO* proteins, both  
173 expressed in petals and stamens, and their interaction pattern with the two *AP3/DEF* proteins is  
174 logically more complex than in Arabidopsis or Antirrhinum: *PhDEF* interacts with both *PhGLO1*  
175 and *PhGLO2*, while *PhTM6* only interacts with *PhGLO2*, which was confirmed genetically in the  
176 corresponding mutants [42]. In addition, fusion of the stamen filaments with the inner petal tube is  
177 specifically regulated by the *PhDEF/PhGLO1* heterodimer [42]. In summary, the increased  
178 complexity in gene number and protein interaction pattern in petunia led to subtle  
179 subfunctionalization of these genes in specifying petal and stamen identity and development (Fig.

180 4). But how the different protein complexes divide up tasks to regulate target genes and specify  
181 correct organ identity at the molecular level remains to be understood.

182

### 183 *Multiple functions for C-class genes*

184 The C-function, i.e. the specification of stamen and carpel identity, generally coupled with the  
185 control of floral determinacy, is controlled by members of the *AGAMOUS* (*AG*) family. This family  
186 is subdivided into the *euAG* and *PLENA* (*PLE*) clades (Fig. 3), whose names come from the rosid  
187 species *Arabidopsis* and the asterid species *Antirrhinum*, where *AG* and *PLE* respectively specify  
188 the C-function [16, 17]. In contrast, the *Arabidopsis* *PLE*-like genes *SHATTERPROOF1* and 2  
189 (*SHP1/2*) play a late role in fruit shattering but are not essential for the C-function [49], while the  
190 *Antirrhinum* *euAG*-like gene *FARINELLI* (*FAR*) only has a little contribution to stamen  
191 development [50]. These observations led to build a model where, after the gene duplication that  
192 generated the *euAG*- and *PLE*-clades, control of the C-function has been randomly allocated to  
193 either member of the two clades, with the other member adopting a distinct function after changes  
194 in gene expression pattern or protein biochemical properties [50].

195 In Solanaceae, several evidences suggest that the C-function is largely redundantly  
196 controlled by both members of the *euAG* and *PLE* clades, which might reflect the ancestral state  
197 just after the *euAG/PLE* duplication [50, 51]. Indeed, in petunia single and double knock-out  
198 mutants in the *euAG*-like gene *PETUNIA MADS BOX GENE3* (*PMADS3*) and the *PLE*-like gene  
199 *FBP6* demonstrate that the two genes redundantly control stamen and carpel identity and floral  
200 determinacy [21]. In tomato, different RNAi lines against the *euAG*-like gene *TOMATO*  
201 *AGAMOUS1* (*TAG1*) and the *PLE*-like gene *ARLEQUIN/TOMATO AGAMOUS-LIKE1* (*TAGL1*)  
202 generated slightly conflicting results, likely due to different degrees of gene down-regulation and  
203 possible co-silencing of paralogous genes [52, 53]. Still, these studies suggest a partially redundant  
204 function of *TAG1* and *TAGL1* in the control of the C-function [52, 53]. Consistently, the beautiful  
205 arlequin tomato (Fig. 1G), a semi-dominant mutant of *TAGL1*, forms carpelloid sepals appearing  
206 fleshy and bright red when the fruit is mature [54]. Both genes also appear to play a role in fruit  
207 development and ripening [52, 53]. In tobacco, VIGS lines suggest that the *euAG*- and *PLE*-like  
208 genes *NbAG* and *NbSHP* redundantly control the C-function, while *NbSHP* has an additional role in  
209 fruit dehiscence (Fourquin and Ferrándiz, 2012). Similarly in *Physalis*, downregulation of the  
210 *euAG*- and *PLE*-like genes *PFAG1* and *PFAG2* by VIGS revealed their partially redundant role in  
211 regulating the C-function [55]. Although work in tomato, tobacco and *Physalis* is not fully clear  
212 since gene function was assessed by down-regulation and not by complete knock-out, overall it  
213 appears that in Solanaceae control of the C-function is largely shared between *euAG*- and *PLE*-like



214 genes, and that *PLE*-like genes can have an additional role in fruit development or ripening  
215 (similarly to the *SHP* genes in Arabidopsis). Together with previous studies in Arabidopsis and  
216 Antirrhinum, this supports an evolutionary model with several independent subfunctionalization  
217 events between *AG*-family members for the control of the C-function [50, 51]. B-class genes have  
218 also been reported to participate in the determination of carpel identity in Solanaceae: as discussed  
219 previously, in petunia the *paleoAP3*-type gene *PhTM6* is expressed in the carpel [44], suggesting it  
220 plays a role in its establishment, although this has not been formally demonstrated yet. In *Physalis*,  
221 it was recently shown that the *GLO*-like gene *DOLL1* is expressed in the carpel whorl where it  
222 activates the expression of the ortholog of the carpel regulator *CRABS CLAW (CRC)*, which ensures  
223 proper carpel development and fertility [56, 57].

224 C-class genes can have other roles in addition to the C-function: these genes also trigger  
225 nectary development [58]. Interestingly, this newly discovered function is true both in petunia  
226 where nectaries are found at the base of the ovary, and in Arabidopsis where nectaries are found at  
227 the base of the stamens. In both species, the *AG*-family genes (i.e. *AG* and *SHP1/2* in Arabidopsis,  
228 and *FBP6* and *PMADS3* in petunia) redundantly activate expression of the YABBY transcription  
229 factor *CRC* ( or its orthologs *PhCRC1/2* in petunia), that are essential for nectary development [57,  
230 58]. This work revealed that the same *AG-CRC* genetic module is involved in nectary development  
231 in two distantly-related species with different nectary positioning, suggesting the possibility of an  
232 ancestral mechanism for nectary specification before the asterids-rosids divergence [58].

233 Finally, the petunia C-class genes *FBP6* and *PMADS3* also participate to the D-function,  
234 specifying ovule identity [21]. This function has been first identified in petunia, where co-silencing  
235 of the D-class genes *FBP7* and *FBP11* converts ovules into carpelloid organs [59]. However, full  
236 knock-out lines revealed later that both genes were dispensable for ovule identity (suggesting that  
237 other genes had been silenced in the cosuppression lines), but that combining the *fbp7 fbp11*  
238 mutations with additional mutation or silencing in *FBP6* or *PMADS3* (therefore creating a partial-  
239 C/full-D class mutant) leads to a strong loss of ovule identity [21]. Indeed, this is in accordance  
240 with what is observed in Arabidopsis, where the C- and D-class genes *AG*, *SHP1/2* and  
241 *SEEDSTICK* also participate in defining ovule identity [60]. In tomato, the D-class genes *Sl-AGL11*  
242 and *Sl-MBP3* are expressed both in the carpel, the seeds and the fruit, and overexpression of *Sl-*  
243 *AGL11* leads to early fruit ripening together with a dramatic conversion of sepals into fleshy fruit  
244 tissue [61]. This suggests that at least one of the D-class genes in tomato has maintained the  
245 capacity to specify carpel identity when ectopically expressed, and may have also developed an  
246 additional role during fruit development.

247

248 *Uncovering the E function*

249 The Solanaceae flowers brought the first evidence for the existence of an extra floral function in  
250 addition to the ABC(D) functions. Indeed, although clearly not gene-specific, co-suppression lines  
251 of the petunia *FBP2* or the tomato *TM5* genes (both MADS-box genes orthologous to Arabidopsis  
252 *SEP3*) exhibited flowers with homeotic conversion of petals, stamens and carpels into sepaloid  
253 organs, together with floral indeterminacy [62, 63]. At that time, this was interpreted as the  
254 involvement of *FBP2* and *TM5* in floral meristem identity or in the repression of sepaloid identity  
255 [62, 63]. Both of these interpretations still stand true today, although we do not think now of  
256 sepaloid identity as being repressed but rather that petal, stamen and carpel identities are activated  
257 on a sepaloid background, and that this activation of the B- and C-functions requires genes from the  
258 E function, such as petunia *FBP2*, tomato *TM5* or Arabidopsis *SEP3*. Indeed, it was later found in  
259 Arabidopsis that petals, stamens and carpels in the *sep1 sep2 sep3* triple mutant were transformed  
260 into sepals [64], while all floral organs in a *sep1 sep2 sep3 sep4* mutant developed as leaf-like  
261 organs [65], leading to the idea that the *SEP* genes are required for the identity of all floral organs in  
262 a largely redundant fashion, thereby incarnating the E function. Molecular support for this  
263 additional function came from the identification of "floral quartets", where E-class proteins bridge  
264 other MADS-box proteins together in a floral organ specific manner, providing a physical  
265 explanation for the implication of *SEP* proteins in the identity of all floral organs [18, 66, 67]. In  
266 addition, work in petunia and Arabidopsis further showed that ovule identity also requires *SEP*  
267 activity, as illustrated by the homeotic conversions of all ovules into leaf-like organs in the petunia  
268 *fbp2 fbp5* mutant [68] and of a part of the ovules into leaf-like or carpel-like organs in the  
269 *SEP1/sep1 sep2 sep3* mutant (Favaro et al., 2003). Furthermore, it was later shown that Petunia  
270 *AGL6*, a member of the MADS-box *AGL6* subfamily closely related to the *SEP* subfamily (Fig. 3),  
271 also performs *SEP-like* functions, redundantly with some of the petunia *SEP* genes [70], adding  
272 further genetic complexity to the *SEP* function in these Solanaceous species. More recently, a large  
273 genetic study was performed aimed at revealing all individual and redundant functions of Petunia  
274 *AGL6* and its six *SEP-like* genes, based on the analysis of single and higher order mutants [35]. This  
275 study revealed that the petunia *SEP1/2/3* orthologs (Fig. 3) together with *AGL6* encode the classical  
276 *SEP* floral organ identity and floral termination functions, with a master role for the petunia *SEP3*  
277 ortholog *FBP2*. Remarkably however, it was found that the *FBP9* subclade members *FBP9* and  
278 *FBP23*, for which no clear ortholog is present in Arabidopsis, play a major role in determining  
279 floral meristem identity together with *FBP4*, while contributing only very moderately to floral  
280 organ identity. Indeed, triple *fbp4 fbp9 fbp23* mutants completely lack flowers and exhibit a highly  
281 branched inflorescence structure due to the homeotic transformation of its floral meristems into

282 inflorescence meristems. This shows that in contrast to Arabidopsis, a subset of the Petunia *SEP*  
283 genes (*FBP4*, -9 and -23) have evolved a specific function as floral meristem identity genes rather  
284 than encoding the classical organ identity *SEP* function. This is remarkably similar to the earlier  
285 proposed roles for three orthologous tomato genes *LIN* (*LONG INFLORESCENCE*), *J2*  
286 (*JOINTLESS 2*) and *EJ2* (*ENHANCER OF JOINTLESS 2*) which are responsible for the transition  
287 from the inflorescence to the floral meristem identity [71], suggesting that this is conserved within  
288 the Solanaceae. Interestingly, mutations in *J2* and *EJ2* were individually selected during tomato  
289 breeding as they both caused beneficial traits (elimination of the fruit abscission zone and increased  
290 calyx size), but when combined together affect inflorescence branching and fertility [71]. Finely  
291 controlling the expression levels of *J2* and *EJ2* by genetic engineering allows to generate tomato  
292 varieties with a better combination of beneficial traits [71, 72]. This reveals the wide and complex  
293 roles of the tomato *SEP*-like genes in the control of inflorescence branching, calyx size, flower  
294 abscission and flower fertility. Finally, silencing approaches suggest that the two tomato *SEP1/2/3*  
295 members *TM5* and *TM29* and the tomato *AGL6* gene exhibit classical *SEP* organ identity functions  
296 (Pnueli et al., 1994; Ampomah-Dwamena et al., 2002; Yu et al., 2017) similar to what we found in  
297 petunia.

298 In Arabidopsis, the floral meristem identity function is attributed to members of the  
299 *AP1/SQUA* MADS-box subfamily (*AP1*, *CAL* and *FUL* genes in Arabidopsis) [73, 74] rather than to  
300 a specific subclass of *SEP* genes as described above for petunia and tomato. Interestingly, *AP1*, *SEP*  
301 and *AGL6* genes all form a superclade with shared ancestry (Fig. 3). This suggests that the floral  
302 meristem identity function might have been ancestral to the duplication event that generated those  
303 three different gene families, and has been distributed primarily to *AP1*-like genes in Arabidopsis  
304 and to a subclass of *SEP*-like genes in petunia and tomato.

305

### 306 *Divergence to the textbook ABC model*

307 Research in the Solanaceae, and in particular in petunia, has led to the construction of a floral organ  
308 identity patterning model quite distinct to the textbook ABC model (Fig. 2). Why so many  
309 differences? Is this a specificity of the Solanaceae? The ABC model is by essence, a simplification  
310 of the reality, and even the founder species of the model, Arabidopsis and Antirrhinum, do not fit  
311 perfectly in the frame. For instance, no complete A-function mutant was ever found in Antirrhinum,  
312 and the A-class gene *AP1* is fully dispensable for petal identity in Arabidopsis [19, 20]. Therefore it  
313 is only normal that, as we discover more and more details about the molecular players of floral  
314 organ identity, the model that was proposed more than 30 years ago becomes less adequate and  
315 needs to be complexified [75]. Moreover, the number of ABC genes within a species necessarily

316 complexifies the model, as each gene copy can evolve new functions (neofunctionalization) or  
317 retain part of the ancestral function (subfunctionalization). [76] Therefore, the increased complexity  
318 of the model in Fig. 3, as compared to the textbook ABC model, is due to the large number of  
319 (A)BC genes in petunia, the high power of functional analysis that can be performed in this species  
320 and the large amount of work that was carried on the topic over the years. This only advocates for  
321 the need of many more model species where such a fine analysis can be performed, in order to find  
322 more general principles and specificities of floral organ identity patterning.

### 323 **Initiation and fusion of floral organs**

324 While floral organ identity is continuously specified by homeotic genes throughout organ  
325 development, other molecular players direct the pattern of floral organ initiation and the hormone  
326 auxin appears as a key player in this process (Smyth, 2018 and references therein). The synthetic  
327 promoter DR5, containing auxin-responsive elements and fused to a reporter gene like GFP, is often  
328 used as a late reporter for auxin signalling [77]. During floral development, *DR5:GFP* expression  
329 peaks successively at incipient floral organ primordia, first marking the sepals initiation domain,  
330 followed by petals, stamens and carpels [78, 79].

331 So far, Solanaceae have brought relatively little contribution to the understanding of floral  
332 organ initiation and growth. Yet, some specific features from the family could strongly enrich the  
333 field of research. For instance, flowers have a type-5 symmetry, with 5 sepals, 5 petals and 5  
334 stamens initiating successively, and almost jointly within each whorl, in contrast to the type-4  
335 symmetry of Arabidopsis flowers. Models, based on periodic auxin accumulation and increasing  
336 space in the floral meristem, generate self-organizing patterns of primordia initiation [80, 81].  
337 Researchers have attempted to explain the emergence of different floral organ numbers per whorl  
338 [82], but these models would strongly benefit from additional data in species with a type-5  
339 symmetry, such as the detailed spatio-temporal pattern of DR5 expression that was characterized  
340 during tomato flower and fruit development [83].

341 Petal fusion (sympetaly) is a trait of major evolutionary importance, that led to new floral  
342 structures and possibly accelerated speciation rates. Some molecular players involved in petal  
343 fusion have been found in Arabidopsis, based on mutants with fused petals [84]. Using a model  
344 species with fused petals, such as *e.g.* petunia, should allow to identify other actors of petal fusion,  
345 and in particular those involved in the evolution of the trait. The petunia mutants *maewest* (*maw*)  
346 and *choripetala suzanne* (*chsu*) were isolated in a genetic screen for petal fusion defects [85]. These  
347 mutants form partly fused and narrow petals, together with narrow leaves, suggesting general  
348 defects in organ laminar lateral growth. Petal fusion is congenital in petunia (i.e. petals are fused by  
349 the confluence of their individual primordia), and the defects observed in *maw* or *chsu* appear to be  
350 due to defects in petal primordia lateral expansion, suggesting that fusion and lateral growth are  
351 inherently coupled during petunia petal development. *MAW* is a WUS-like homeobox (WOX) gene,  
352 to which the famous gene *WUSCHEL* belongs, that controls stem cell maintenance in the meristem  
353 [86, 87]. In tomato flowers, whose petals are partly fused at the base, mutation in the *MAW* ortholog  
354 *UN-FUSED FLOWER* (*UF*) also causes narrow and unfused petals, and narrow leaves [88]. *MAW*  
355 and *UF* are homologous to Arabidopsis *WOX1*, whose mutation causes no obvious phenotypic  
356 defects in the flower [85]. However, combining mutations in *wox1* and in another WOX gene,

357 *PRESSED FLOWER (PRS)*, results in plants with very narrow leaves and petals, reminiscent of the  
358 *petunia maw* phenotype [85, 89]. This indicates that petal lateral growth (and fusion in *petunia*) is  
359 controlled by similar classes of genes from the *WOX* subfamily in *Arabidopsis* and *petunia*, but  
360 different members have been recruited to fulfill this function in the two species [85, 86]. Although  
361 *Arabidopsis WOX1* and *PRS* genes appear to fulfill together the same function as *MAW* in *petunia*,  
362 *Arabidopsis* petals are not fused, suggesting that other differences in the regulatory network caused  
363 divergence in this trait during evolution. Sympetaly likely has a single origin in asterids [84], hence  
364 identifying the key event leading to the evolution of this trait might require comparative work  
365 involving a sister group to the asterids, such as Caryophyllaceae family where most species form  
366 flowers with unfused petals.

## 367 **Growth and maturation of floral organs : Chinese lanterns and petal colours**

368 The elegant Chinese lanterns from *Physalis* and *Whitania* form an encapsulated fruit after  
369 pollination, named the inflated calyx syndrome (ICS), due to late sepal growth (Fig. 1H). Indeed  
370 sepals of Chinese lanterns resume growth after pollination, and this trait was shown to be an  
371 advantageous morphological novelty, because the inflated calyx has photosynthetic capability and  
372 provides a micro-environment improving fruit maturation [90]. It had been proposed that the  
373 MADS-box gene *MPF2*, orthologous to Arabidopsis *AGAMOUS-LIKE 24*, and genes from the  
374 *MPF2-like-A* family were involved in ICS: down-regulating *MPF2* expression by RNA interference  
375 in *Physalis* resulted in small sepals, and ectopic expression of *MPF2* in tomato enlarged its calyx by  
376 increasing cell division rates [91, 92]. In tomato, the *MPF2* ortholog *STMADS16* is only expressed  
377 in leaves, whereas in *Physalis* *MPF2* is expressed both in leaves and sepals, suggesting that during  
378 evolution *MPF2* was recruited in the calyx, by heterotopic expression, to form the Chinese lantern  
379 phenotype [91]. However, these results were recently reevaluated, because knocking-out *MPF2* by  
380 CRISPR was not sufficient to disrupt the ICS, and neither was the individual knock-out of ten other  
381 MADS-box genes from the *AP1*, *SEP4*, *AG* or *DEF/GLO* clades [93]. RNA interference notoriously  
382 silences both on- and off-targets with partial efficiencies, which probably explains the inconsistency  
383 of phenotypes observed between the RNA interference and CRISPR lines. This suggests that  
384 multiple MADS-box genes may play a role in the formation of the inflated calyx, but the exact  
385 genetic change that caused the emergence of this novelty has not been pinpointed yet [94].

386

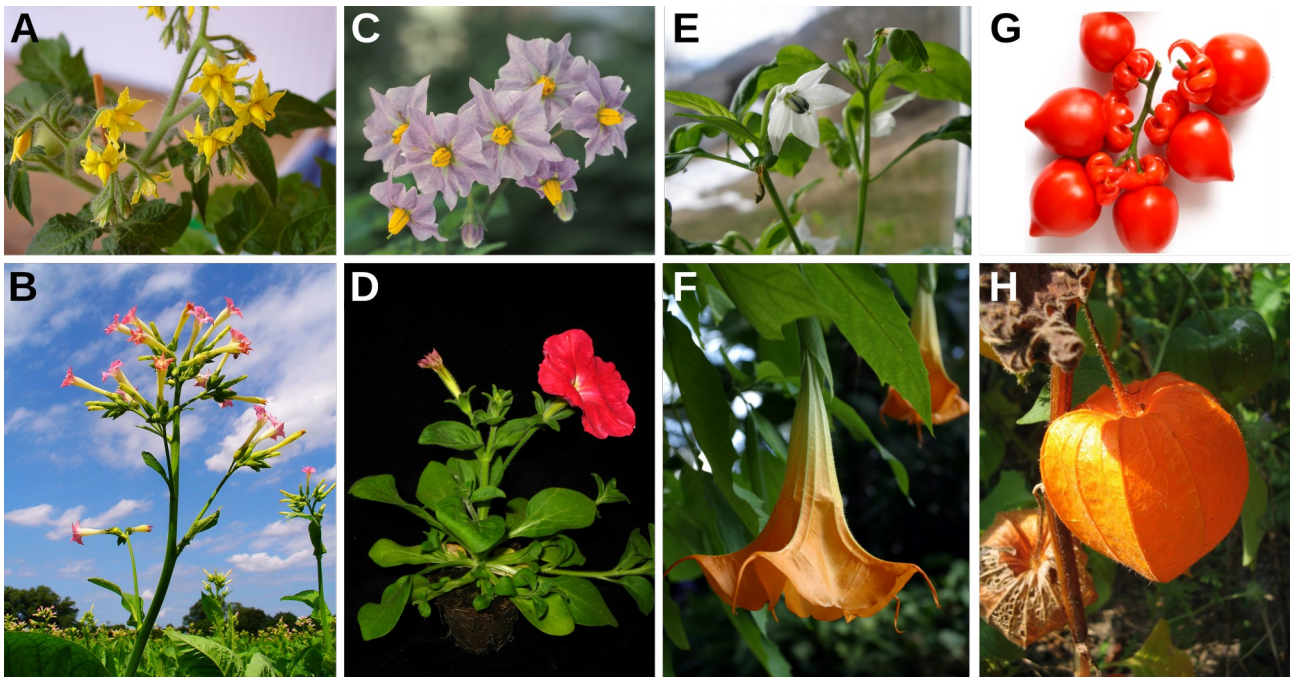
387 Mature floral organs acquire a whole set of tissue and cell properties, and in particular petals  
388 display a specific pigmentation, crucial for their interaction with pollinators. Since petunia petals  
389 are big and showy, and because insertion mutants frequently arise in the line with an active  
390 transposase, a mutagenesis screen for the production of anthocyanins (the main pigments  
391 accumulating in petunia petals) entails a beautiful and easy phenotyping process. Consequently,  
392 petunia has been instrumental in identifying the molecular players of flower colour, with the  
393 majority of enzymes and regulators involved in anthocyanin production cloned and characterized in  
394 this species [8]. Production of anthocyanins first relies on the production of flavonols, later  
395 modified into anthocyanins through the action of specific enzymes and regulators [8]. Interestingly,  
396 the mutant phenotypes of these different regulators are distinct: the *an1* mutant loses pigmentation  
397 in all tissues, while the *an2* and *an8* mutations only affect pigmentation of the limbs (the upper part  
398 of the corolla) and the tube respectively [8, 96, 97]. This suggests that the wild-type petal  
399 pigmentation is the result of the global and local action of a combination of regulatory genes. The  
400 action mechanism of the regulator AN1 has been elucidated in more details: additional to the

401 regulation of anthocyanin biosynthesis, AN1 regulates the pH of the vacuole of petal epidermal  
402 cells, which directly affects the absorption spectra of anthocyanins and hence the resulting petal  
403 colour [96]. In particular, AN1 regulates the expression of *PH1* and *PH5*, two vacuolar P-ATPases  
404 that pump protons into the vacuole and therefore acidify it [98, 99]. Surprisingly, it was recently  
405 found that fruit acidity in lemon and oranges is caused by the same mechanism: the genes *CitPH1*  
406 and *CitPH5* acidify the vacuoles in the juice vesicle cells of the fruit and can thereby generate a pH  
407 as low as 2 [100]. The regulator of anthocyanin production *AN2* was also found to be involved in  
408 the evolution of petal colour between wild petunia species. Indeed, in the white-coloured flowers of  
409 *Petunia axillaris*, five independent losses of function in *AN2*, some of them by frame-shift  
410 mutations, were found in the wild, affecting flower colour and pollinator preferences [101].  
411 Moreover, the surprising "resurrection" of the *AN2* gene after pseudogenization, by a secondary  
412 mutation that restored the original reading frame of the gene, has been reported in the purple-  
413 coloured petals of *P. secreta* [102]. The study of petal pigmentation has proven to be a rich field of  
414 research in the Solanaceae, bridging gaps between the subcellular and the microevolutionary scale.



## 415 **Conclusion**

416 Flower development is a massive field of research that can be apprehended under different angles,  
417 at different scales and in different species. For the last 30 years, research on Arabidopsis has led to  
418 the characterization of most key concepts and molecular players of flower development. However,  
419 each species has its specificities and Arabidopsis is no exception; therefore some of the mechanisms  
420 uncovered in Arabidopsis are highly divergent in comparison to the common ancestor of eudicots.  
421 Moreover, due to the random nature of gene duplication and subfunctionalization in different  
422 species, gene functions can appear hidden in redundancy in Arabidopsis but be revealed by gene  
423 subfunctionalization in other species (or vice versa). Finally, some important features of flower  
424 development simply do not exist in Arabidopsis, for instance petal fusion or petal pigmentation. For  
425 these reasons, including several model species in the research on flower development is now crucial  
426 to further improve our understanding of this process [11]. Solanaceae have brought their share of  
427 results on this topic and surely this family will continue bringing new insights on flower  
428 development, also with potential application for agronomical purposes.

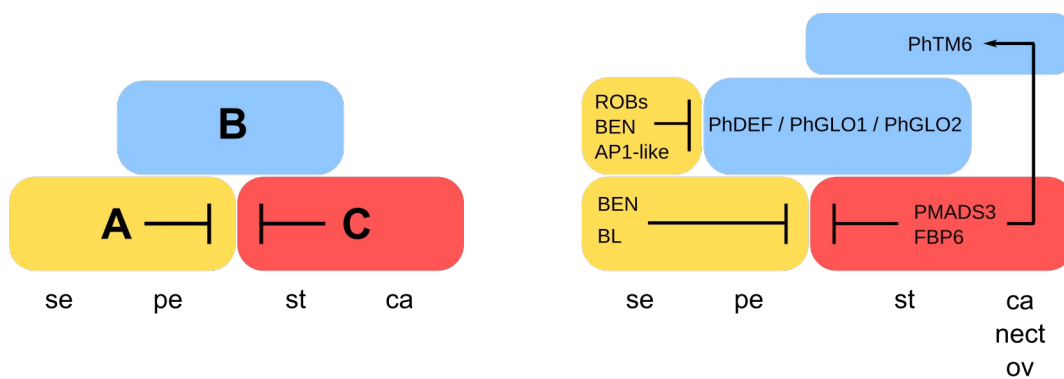


429

430 **Figure 1.** Flowers and inflorescences from various Solanaceae members: *Solanum lycopersicum*  
 431 (*tomato*, A), *Nicotiana tabacum* (tobacco, B), *Solanum tuberosum* (potato, C), *Petunia x hybrida*  
 432 (petunia, D), *Capsicum annuum* (pepper, E), *Brugmansia sp.* (F), fruits from the arlequin tomato  
 433 mutant (G), fruit from *Physalis alkekengi* (H). Picture credits respectively, from A to H: Niek  
 434 Willems, H. Zell, Keith Weller, Michiel Vandenbussche, Simone Stibbe, Tom Morphy, arlequin  
 435 tomato picture reproduced with authorization from Rafael Lozano [54], and Michael Gasperl.  
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437

438



### The textbook ABC model

### Floral organ identity patterning in petunia

439

440 **Figure 2.** The textbook ABC model and its variation in petunia, as we currently understand it. In the

441 textbook ABC model, A-class genes and C-class genes mutually repress their expression domains.

442 A-class genes alone and in combination with B-class genes specify the identities of sepals (se) and

443 petals (pe) respectively, C-class genes alone and in combination with B-class genes specify the

444 identities of carpels (ca) and stamens (st) respectively. In the petunia model, the 2 C-class genes

445 *PMADS3* and *FBP6* redundantly share the C-function and specify carpel identity alone, and stamen

446 identity together with the 4 B-class genes *PhDEF*, *PhTM6*, *PhGLO1* and *PhGLO2*. These 4 genes

447 show additional patterns of subfunctionalization detailed in Figure 4. C-class genes additionally

448 specify nectary (nect) and ovule (ov) development. *PhTM6* is expressed in the stamens and carpels

449 and its expression is activated by C-class genes; however the function of *PhTM6* in the carpels is

450 unknown so far. The (A)-function is ensured by the genes *BL*, *BEN*, *ROBs* and *AP1-like*: *BL* and

451 *BEN* repress C-class genes expression in sepals and petals, and *BEN*, *ROBs* and *AP1-like* genes

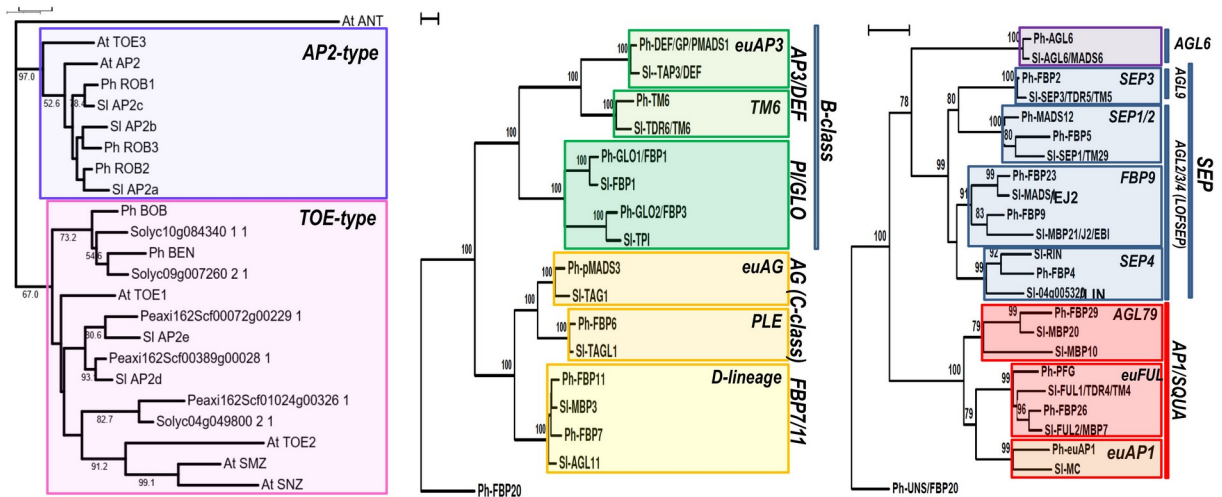
452 repress B-class genes expression in the sepal whorl. In this model, the coloured boxes represent the

453 region of action of the genes and not necessarily their domain of expression (*BL*, *BEN* and the *ROB*

454 genes are for instance expressed in all floral organs).

455

456

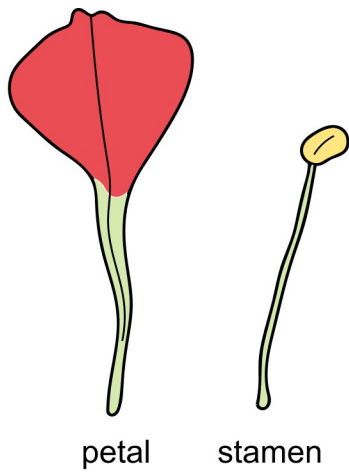


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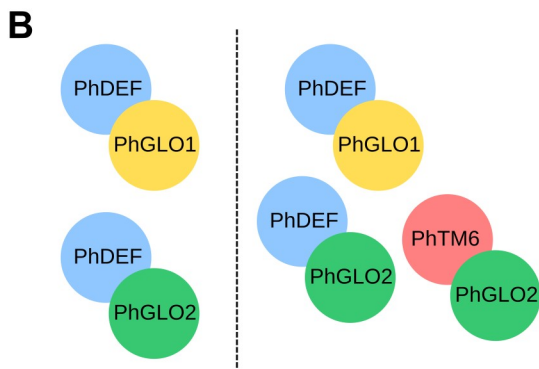
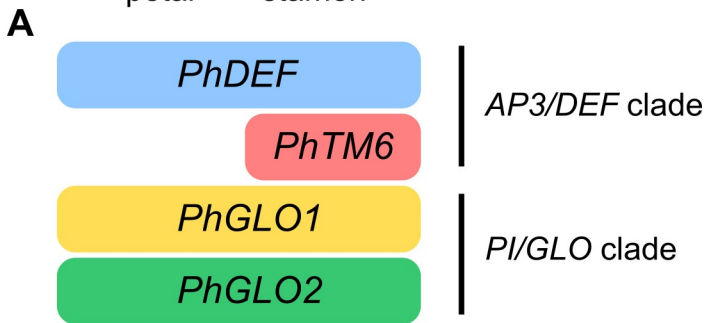
458 **Figure 3.** Neighbor joining trees of euAP2 transcription factors (left), B/C/D class (middle) and  
 459 AP1/SEP/AGL6 superclade (right) MADS-box transcription factors. Prefixes At, Ph and Sl  
 460 represent *Arabidopsis thaliana*, *Petunia hybrida* and *Solanum lycopersicum* respectively. The  
 461 euAP2 phylogenetic analysis was obtained using the pipeline offered by <https://ngphylogeny.fr>, and  
 462 the tree was rooted with Arabidopsis ANT, an AP2 transcription factor not belonging to the euAP2  
 463 lineage. Both MADS-box trees were rooted with Ph-UNS/FBP20, a SOC1 subfamily member.  
 464 Bootstrap values (based on 1000 replicates) supporting tree branching above 70% are indicated near  
 465 the branching points. Scalebars correspond with 0.1 substitutions per site. The MADS trees were  
 466 computed with Treecon software [103] using the Tajima & Nei Distance Calculation method and  
 467 further default settings.

468

469



petal      stamen



470

471 **Figure 4.** Subfunctionalization of the B-class genes for specification of petal and stamen identity  
 472 in petunia. **A:** Expression domains of the *PhDEF*, *PhTM6*, *PhGLO1* and *PhGLO2* genes in petal  
 473 and stamen initiation domains. **B:** Protein complexes formed between AP3/DEF- and PI/GLO-type  
 474 proteins in each floral organ, based on [42, 44].

475

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