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NMR Spectroscopy to Study Cyclodextrin-Based Host-Guest Assemblies with Polynuclear Clusters

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10 ABSTRACT. Natural cyclodextrin (CD) macrocycles are known to form diverse inclusion 11 complexes with a wide variety of organic molecules, but recent work has revealed that inorganic 12 clusters also form multicomponent supramolecular complexes and edifices. Such molecular 13 assemblies exhibit a high degree of organization in solution governed by various chemical 14 processes including molecular recognition, host-guest attraction, hydrophobic repulsion, or 15 chaotropic effect. Nuclear magnetic resonance (NMR) spectroscopy is one of the most efficient 16 and practical analytical technique to characterize the nature, the strength and the mechanism of 17 these interactions in solution. This review provides a brief overview on recent examples of the 18 contribution of NMR to the characterization of hybrid systems in solution based on CD with 19 polynuclear clusters, including polyoxometalates (POMs), metallic clusters and hydroborate 20 clusters. The focus will be first on using 1 H (and 13 C) NMR of the host, *i.e.*, CD, to identify the 21 nature of the interactions and measure their strength. Then, 2D NMR methods will be illustrated 22 by DOSY as a means of highlighting the clustering phenomena, and by NOESY/ROESY to 23 evidence the spatial proximity and contact within the supramolecular assemblies. Finally, other 24 NMR nuclei will be selected to probe the inorganic part as a guest molecule. Attention will be paid 25 to classical host-guest complexes Cluster@CD, but also to hierarchical multi-scale, multi-26 component assemblies such as Cluster@CD@Cluster.

27

28 **1. Introduction**

Molecular recognition in biology, which exploits the high affinity of bindings/interactions between specific pairs of host and guest molecules, is an infinite source of inspiration for innovation in materials science.^{1,2} Molecular signal detection is probably one of the most important bioinspired functions, which may be of interest in various fields such as medical engineering, therapeutic treatment, and sensing.³⁻⁵ The processes involved are mainly based on supramolecular chemistry, where self-assembly through weak chemical bonds, *i.e.*, hydrophobic, electrostatic, and ionicdipole interactions, is the driving force behind stable systems.⁶⁻⁹ The spontaneous association of two or more components in solution to produce hybrid systems by coordination or supramolecular chemistry requires favorable physicochemical phenomena with minimal energy cost.^{10,11} The construction of multifunctional assemblies based on supramolecular complexes usually involves the use of macrocyclic molecules to host various guest species.^{5,12,13}

40 Cyclodextrins (CDs) represent a group of macrocyclic oligosaccharides mainly constituted of 6, 7, or 8 D-glucopyranoside units.¹⁴ Thus, the three renowned corresponding forms, *i.e.*, α -, β -, and 41 42 γ -CD, present a hydrophobic cavity of different diameter sizes depending on the number of the 43 repeating glucopyranose group. While CDs are known for a long time to interact with various 44 hydrophobic substances, it is now admitted that CDs are also capable of forming host-guest 45 adducts with a wide variety of hydrophilic polynuclear metallic clusters.¹⁵ Such self-assembly 46 properties of CDs have proven useful for the development of smart materials based on 47 supramolecular hybrid systems.^{16,17} Hybrid associations between CDs and inorganic species can 48 be divided into two categories. The first results from the strong affinity of the CD with a 49 hydrophobic group grafted to the inorganic moieties. Such a mode of interaction fits into the wellestablished classical hydrophobic effect.¹⁸⁻²¹ The second type involves native inorganic species 50 51 such as polyoxometalates (POMs) or ionic cluster compounds which were found to form unusually 52 stable host-guest adducts with CDs. In 2015, both Nau's and Stoddart's groups reported that γ - or 53 β -CD interacts strongly with hydrophilic ions such as $[B_{12}X_{12}]^{2-}$ clusters (with X = H, F, Cl, Br or 54 I) and [PMo₁₂O₄₀]³⁻ anions, as revealed by single-crystal X-ray diffraction (XRD).^{22,23} These solidstate arrangements showed a 2:1 sandwich-type complex where the inorganic ionic species, *i.e.*, 55

56 either the dodecaborate anions or the POM, appears surrounded by two CDs. It is interesting to 57 note that depending on the anionic guest, the interaction of the CD involves its secondary face in 58 the case of the cluster or its primary face in the case of POM. This molecular recognition behavior 59 thus appears as a distinctive feature of CD in the inclusion phenomenon with inorganic clusters. 60 The inclusion phenomenon involving hydrophilic ionic species has been recently well documented 61 and explained as a solvent effect that arises from the chaotropic nature of the ionic guest.^{24,25} Such 62 a chaotropic effect occurring in aqueous solution gives a characteristic thermochemical fingerprint. 63 Low charge density species locally cause the breakdown of the water structure, characterized by a 64 high-enthalpy and a high-entropy solvation shell. Throughout the aggregation process leading to 65 guest encapsulation, the release of water from the hydration shell to the bulk is associated with strong decreases in enthalpy and entropy, *i.e.*, $\Delta_r H < 0$ and $\Delta_r S < 0$. Despite the entropy penalty, 66 the aggregation process remains highly favorable and enthalpically driven ($T\Delta_r S > \Delta_r H$ and $\Delta_r G <$ 67 68 0). Thus, several POM-based chemical systems with low charge density, including the Lindqvist ions $[M_6O_{19}]^{2-2}$, the Preyssler-type anion $[P_5W_{30}O_{110}]^{15-27}$ the Dawson-type anion $[P_2W_{18}O_{62}]^{6-28}$ 69 and even, the giant Mo blue wheel $[Mo_{154}O_{462}H_{14}(H_2O)_{70}]^{14}$, abbreviated $\{Mo_{154}\}$, ^{29,30} have verified 70 71 such affinity. The phenomenon can then be generalized regardless of the structure, composition, size and shape of the POM.^{24,25} Although the chaotropic effect was identified by Hofmeister in 72 73 1888 for small ions, this property has been clearly associated with cluster anions only very recently and opens a new avenue for the design of CD-based smart supramolecular materials.^{7,31-36} 74 75 The characterization of CD complexation by polynuclear clusters is of paramount importance to 76 understand the driving forces of the phenomena involved in solutions such as self-assembly, host-

78 titration,³⁷ ESI-mass spectrometry,^{38,39} X-ray scattering and diffraction,^{1,40} circular

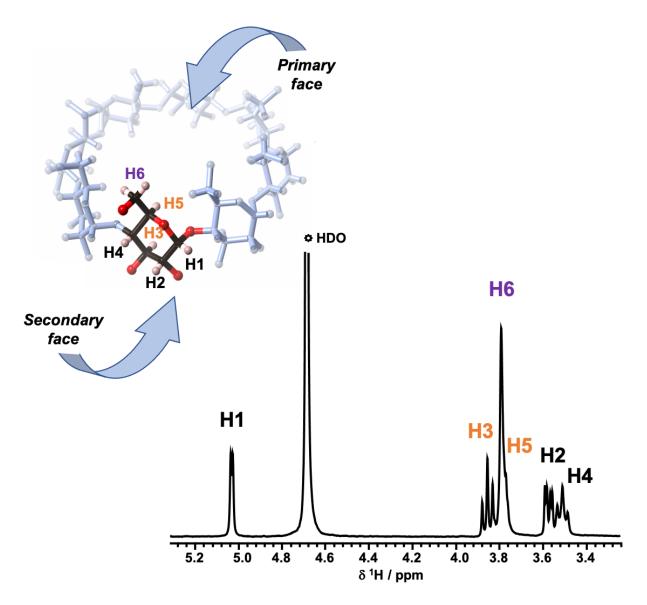
guest inclusion, and aggregation process. Various analytical techniques such as calorimetric

dichroism,^{21,36,41} voltammetry⁴¹ and fluorescence spectroscopy⁴² were applied to study the 79 80 reactivity of CDs with different guest molecules. Furthermore, NMR spectroscopy is one of the most frequently used methods for studying self-assembly processes with CDs,⁴³⁻⁴⁶ allowing to 81 82 determine not only quantitative information, e.g., the stoichiometry of adducts and their binding 83 constants but also structural and dynamic data of the complex and its constituent parts, e.g., the 84 regioselectivity of the binding sites and their equilibria and exchanges. This mini-review focuses 85 on the recent use of NMR methodology to study complexation of CD with inorganic polynuclear 86 species.

- 87
- 88 89

2. Probing host-guest interaction by ¹H and ¹³C NMR of cyclodextrin

90 Cyclodextrins are highly symmetrical molecules consisting of identical D-glucopyranoside 91 motifs. The ¹H NMR in aqueous solution thus shows the signature of this glucopyranoside unit 92 which has six distinctive proton sites, labeled H1 (anomeric position), H2, H3, H4, H5 and H6 93 (See Fig. 1).⁴⁶ The three hydroxyl groups are not distinguishable and merge with the water signal 94 due to their chemical exchange. Because the molecules are cyclic H2 and H4 are exposed to the 95 outside, while H3 and H5 are located inside the central cavity of the tori. The H6 protons 96 correspond to the methanolic arms at the top of the primary face of the macrocycles. Fig. 1 is a 97 representative ¹H NMR spectrum in D₂O of 3 mM solution of γ-CD. The anomeric proton H1 98 appears strongly deshielded at ca. 5 ppm, while the other protons are only partially resolved and 99 divided into two groups, at 3.4-3.6 ppm (outward protons H2 and H4) and 3.7-3.9 ppm (inward 100 protons H3, H5, and H6). The latter are obviously the most sensitive to interaction with guest 101 molecules and are often used as probes for host-guest complexation.



104 Fig. 1. ¹H NMR spectrum of 3 mM γ -CD in D₂O showing the six signals of the macrocycle: the 105 anomeric proton H1, the outward protons H2 and H4, the inward protons H3 and H5, and the 106 methanolic H6 protons on the primary face.

107

108 One of the simplest experiments to identify the host-guest interaction in solution between CD and 109 inorganic species is the ¹H NMR titration of the host molecule by the guest species to observe the 110 effects on the resonances of the most exposed protons, those present on the inner surface of the 111 cavity. Fig. 2 shows an example of titration of 3 mM D₂O solution of γ -CD by two anionic clusters: 112 the selenide octahedral rhenium cluster [Re₆Se₈(CN)₆]⁴ and the Dawson-type POM [P₂W₁₈O₆₂]^{6-113 .^{6,28} Two antagonist behaviors are observed. With the rhenium cluster complex, the most affected} 114 resonance upon addition of 4 eq. corresponds to the H3, down-field shifting from ca. 3.9 to 4.3 115 ppm, whereas in case of the POM, this resonance did not move so much, and instead, H6 116 underwent up to 0.3 ppm downfield shift. This illustrates the difference in supramolecular 117 recognition process in solution. CD reacts with the rhenium cluster to form inclusion complex 118 involving their secondary face, which differs from the sandwich adduct through its primary rim 119 obtained with the Dawson-type POM (see Fig. 2). These observations in aqueous solution are 120 consistent with the molecular organization in the solid-state, but has to consider the dynamics 121 aspect of the host-guest association equilibria. As evidence of molecular recognition, competition 122 occurs in the simultaneous presence of two guests, Dawson-type POM and octahedral rhenium 123 cluster.47

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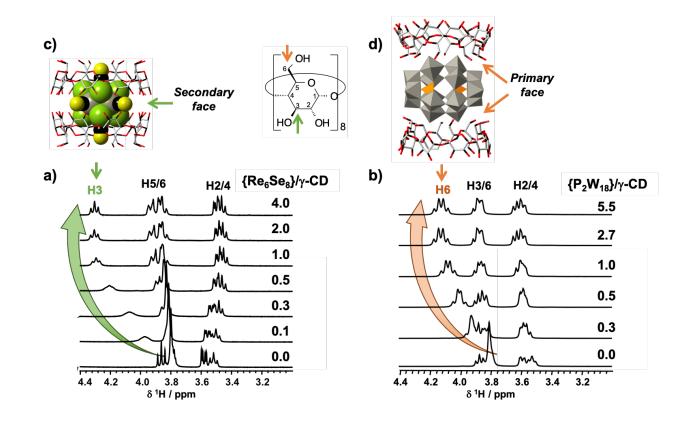


Fig. 2. ¹H NMR titration of 3 mM γ -CD in D₂O by (a) [Re₆Se₈(CN)₆]⁴ and (b) [P₂W₁₈O₆₂]⁶ in the range 3.0-4.4 ppm. (c) and (d) the crystallographic structures of the adducts obtained in each system. Adapted from references.^{6,28}

129

130 The dynamic aspect of the host-guest association could also inform the strength of the interaction 131 between the two components. Generally speaking, a slow dissociative process usually indicates 132 a strong complexation. This can be easily observed in the NMR titration experiment. When the 133 equilibria are fast relative to the NMR time scale, the CD signals appear as a weighted average situation between free and bound species. This is manifested by a continuous shift of some 134 135 specific resonances, as observed in the cases of $[Re_6Se_8(CN)_6]^4$ and $[P_2W_{18}O_{62}]^6$ with γ -CD in Fig. 2. The scenario is different with molybdenum clusters $[Mo_6X_8Cl_6]^{2-}$ (X = Br or I) when the 136 137 NMR signatures of γ -CD exhibit slow exchange regime,⁴⁸ as shown in Fig. 3. Here, the signals 138 of free and complexed CD are observed together, allowing even the stoichiometry of the final 139 adduct to be easily determined. Indeed, consumption of free CD occurs at 0.5 eq. of cluster 140 confirming a 2:1 sandwich complex in agreement with the XRD structures (Fig. 3).

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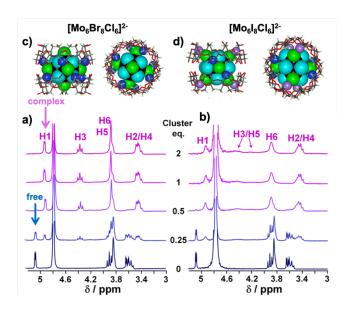


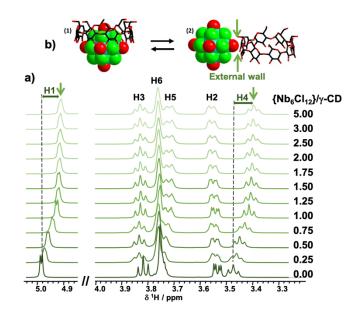
Fig. 3. ¹H NMR titration of 2 mM γ -CD in D₂O by (a) [Mo₆Br₈Cl₆]²⁻ and (b) [Mo₆I₈Cl₆]²⁻. (c) and (d) the crystallographic structures (top and side views) of the adducts obtained in each system. Color code: black = Mo, light blue = X (Br or I), green = Cl. H3 and H5 involved in close host– guest interactions are in dark blue and pink, respectively. Adapted from reference.⁴⁸

147

148 The interaction of octahedral metal clusters with γ -CD often leads to sandwich-like host-guest 149 complexes, as revealed by XRD analysis. However, the behavior in solution may differ 150 dramatically from one cluster to another. Some clusters like $[Re_6X_8(CN)_6]^4$ (X = S or Se) showed 151 labile complexes indicating moderate association affinity ($K_{1:1} < 2000 \text{ M}^{-1}$), while others like 152 $[Mo_6X_8Cl_6]^{2}$ (X = Br or I) are characterized by frozen complexation in solution due to strong 153 attraction between the two components. Such behavior comes from a chaotropic effect induced 154 complexation where the guest dehydration and its association with neutral surfaces such as 155 hydrophobic CD cavities are very favorable processes.^{24,25,49} Here, the overall charge of the 156 polyanion plays a crucial role, and the general trend shows that the lower the charge density, the 157 stronger the host-guest association. In comparison, POMs can have a very strong chaotropic 158 character, but they lead almost exclusively to dynamically labile systems. It seems therefore 159 difficult to reconcile the thermodynamic stability of these supramolecular associations and their 160 dynamics in solution, even if the general tendency shows that frozen complex reflects a strong 161 association.

Internal dynamics can also be detected in a frozen system, as in the case of molybdenum chloride clusters with γ -CD shown in Fig. 3.⁴⁸ The NMR spectra of the iodine derivative are broader than those obtained with Br-based clusters. This would indicate a different rotational dynamic within the host-guest assembly, induced by a reorientation of the cluster in the CD cavities after a 45° tilt from the central axis of the CD (Fig. 3). With iodide compound, all the six apical chlorine ligands are located inside the cavities of the capping CDs, maximizing the weak attractive forces with the host and thus restricting internal molecular movement within the supramolecular framework. Host size may also play a key role in these associations and their dynamics since the volume of cluster $[Mo_6X_8Cl_6]^{2-}$ increases from 395 to 447 Å³ by substituting X with Br and I, respectively.

172 Interaction of cyclodextrins with cationic transition metal octahedral clusters has also been 173 studied, and their behavior in solution has been found mostly to depend on the nature of their 174 inner ligands like with anionic clusters. For instance, very different results were obtained with $[Nb_6Cl_{12}(H_2O)_6]^{2+}$ when compared to $[Ta_6Br_{12}(H_2O)_6]^{2+}$ although both had led to isostructural 175 176 sandwiched inclusion 2:1 complexes with γ -CD in the solid-state.^{1,28} While titration experiment 177 with $[Ta_6Br_{12}(H_2O)_6]^{2+}$ showed frozen complexation in solution, the ¹H NMR spectra in the case 178 of $[Nb_6Cl_{12}(H_2O)_6]^{2+}$, shown in Fig. 4, clearly indicate no host-guest inclusion happened and only 179 weak contacts took place at the external peripheral wall of the CD. Indeed, the signals of H3 and 180 H6 protons at the two different rims did not undergo any significant alteration, while resonances 181 of H1 and H4 protons pointing outward the cavity shifted significantly in the presence of the 182 cluster. In a similar way, no interaction has been recorded with the chloride derivative of $[Mo_6Cl_8Cl_6]^{2-50}$ while the bromide and iodide cluster complexes $[Mo_6X_8Cl_6]^{2-50}$ (X = Br or I) 183 184 interact strongly with γ -CD.⁴⁸



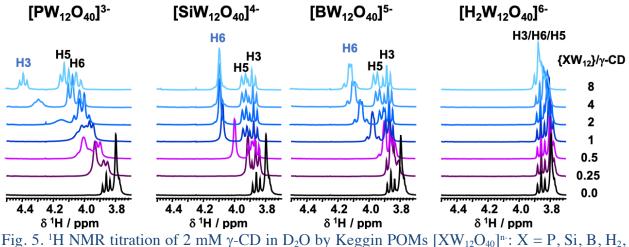
187 Fig. 4. (a) ¹H NMR titration of 1 mM γ -CD in D₂O by [Nb₆Br₁₂(H₂O)₆]²⁺. (b) A schematic 188 illustration of the dynamic dissociation of the host-guest complex observed in the solid state (1) 189 occurring in solution (2). Adapted from reference.¹

190

191 Given their spherical shape and nanoscale size, Keggin-type POMs are expected to react with 192 CDs to readily form host-guest inclusion complexes in a similar way to octahedral metal-atom 193 clusters. Nonetheless, the isolation of well-defined supramolecular assemblies is rather rare with 194 POMs. The interaction in solution was found to be strongly dependent on the charge of the POM 195 as shown in Fig. 5,²⁵ depicting the ¹H NMR titration of 2 mM γ -CD with four different Keggin-196 type POMs $[XW_{12}O_{40}]^{n}$, X = P, Si, B, and H₂, whose overall charge *n* varied from 3- to 6-. 197 Almost no effect can be detected with the most charged POM $[H_2W_{12}O_{40}]^{6}$, indicating very weak 198 interaction between the POM and organic macrocycle. With the boron and silicon derivatives 199 moderate effects on H6 signal can be seen, comparable to that observed with the Dawson-type 200 POM (Fig. 2), indicative of supramolecular contact of the POM with the primary face of the CD. 201 The situation is even more different with the Keggin-type dodecatungstophosphate $[PW_{12}O_{40}]^{3-1}$ 202 where H3 exhibited the strongest effect as clear evidence of embedment of the POM within the

203 central cavity of the CD. Indeed, among the four Keggin-type POMs studied, only $[PW_{12}O_{40}]^{3-1}$ 204 had conducted to crystal structure of host-guest adduct where the POM is deeply embedded 205 within the central cavity through the secondary face of the γ -CD. The X-ray structure with the 206 boron derivative showed co-crystallization of the POM and the CD side-by-side without 207 inclusion complexation. Again, these results are explained by the chaotropic nature of the 208 polyanions. When the hydration network is loosely defined and weakly organized, desolvation 209 can occur easier. Obviously, the charge density of the POM affects the solvation properties and 210 the higher the charge density, the lower the desolvation ability (or the chaotropic power).

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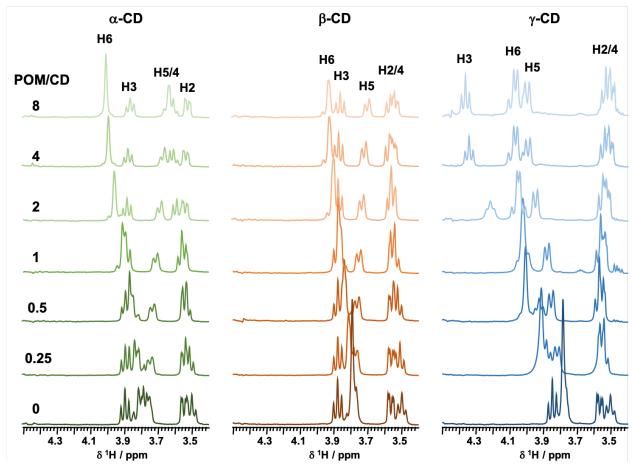


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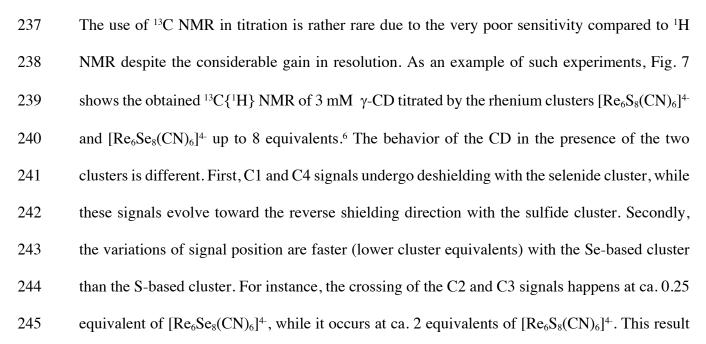
Similar results were obtained with the molybdenum Keggin POMs, confirming that the nature of the metal center, *i.e.*, Mo⁶⁺ or W⁶⁺, does not affect much the interaction with the CD. However, the effects of POM on the ¹H NMR response are different depending on the size of the CD highlighting the importance of the host-guest size matching in the encapsulation process. Fig. 6 displays the obtained ¹H NMR titration of three different solutions of α -, β -, and γ -CD by

222	$[PMo_{12}O_{40}]^{3-}$ (4 mM in D ₂ O). ⁵¹ The evolution of the spectra of γ -CD is very similar to that
223	observed with $[PW_{12}O_{40}]^{3}$ (see Fig. 5), where H2 and H4 show only a negligible effect by
224	comparison to H3, and to a lesser extent to H5 and H6, experiencing all strong deshielding
225	effects. On the other hand, no significant effect can be seen on H3 resonance in α - and β -CD
226	solutions, and only moderate shifts are detected for H6 and H5 signals. These results indicate
227	that interactions occur mainly with the primary face of the CD without deep inclusion within the
228	organic macrocycle since the chemical shift of the internal H3 proton remains unaffected. It
229	appears therefore clear that the α - and β -CD interact mostly with the POM through their outer
230	surface, while γ -CD leads to the usual host-guest inclusion, involving its internal cavity. The α -
231	and β -CD are too small to entirely accommodate Keggin-type POM in their central cavity.
232	



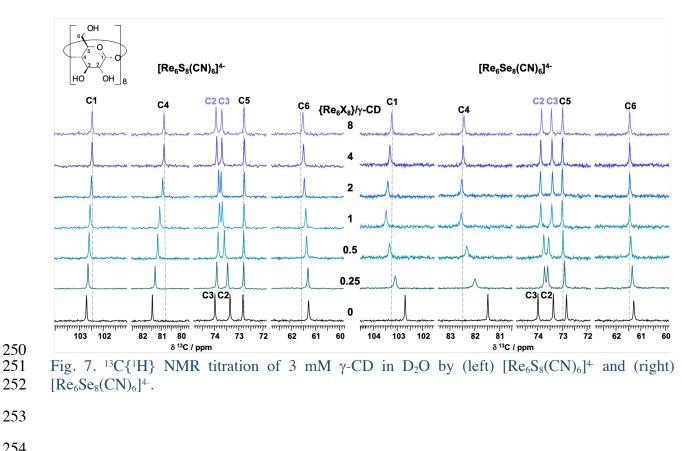
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Fig. 6. ¹H NMR titration of 4 mM α -, β -, and γ -CD in D₂O by Keggin-type POM [PMo₁₂O₄₀]³⁻. Reproduced from reference.⁵¹



246 indicates stronger affinity between the CD and $[Re_6Se_8(CN)_6]^4$ than $[Re_6S_8(CN)_6]^4$ in agreement with the difference in association constants measured for these clusters, $K_{1:1} = 1500 \text{ vs } 900 \text{ M}^{-1}$, 247 248 respectively (see Table 2).

249



254

3. Quantitative analysis: Determination of binding constants 255

NMR is a quantitative analytical tool for studying chemical equilibria and dynamic phenomena.⁵² 256 257 The association constants between the polynuclear cluster and the host CD can be determined from NMR titration by applying mathematical approach.^{53,54} In the case of a fast chemical exchange 258 regime relative to the NMR time scale, when a single NMR signal representative of all the different 259 260 binding states is observed, the variation in the apparent chemical shift (δ_{obs}) of the CD resonances should correspond to a linear combination of the individual chemical shifts of the CD in its 261

different states, *i.e.*, the free CD and the 1:1, 1:2, ... etc. adducts, noted respectively δ_0 , $\delta_{1:1}$, $\delta_{1:2}$, ... etc. This relationship illustrated by equation (1) implies mole fractions x_0 , $x_{1:1}$, $x_{1:2}$, ... etc. of each CD species in free and adduct forms of 1:1, 1:2, ... etc.

265
$$\delta_{obs} = x_0 \delta_0 + x_{1:1} \delta_{1:1} + x_{1:2} \delta_{1:2} + \cdots etc.$$
(1)

In the simplest case of a 1:1 adduct, the association constant *K* is given by equation (2), using the CD molar fractions x_0 and $x_{1:1}$ and expressed as a function of the initial CD concentration C⁰ and the molar ratio R = [Cluster]/[CD].

269
$$K = \frac{x_{1:1}}{x_0 \left(R - \left(\frac{1 - x_0 + x_{1:1}}{2}\right) \right) C^0}$$
(2)

By combining equations (1) and (2), and assuming that $x_0 + x_{1:1} = 1$, we can deduce the expression of the observed chemical shift as a function of binding constant *K* (eqn (3)).

272
$$\delta_{obs} = \delta_0 \left[\frac{KC^0 - KC^0 R - 1 + \sqrt{(KC^0 R - KC^0 + 1)^2 + 4KC^0}}{2KC^0} \right] + \delta_{1:1} \left[\frac{KC^0 + KC^0 R + 1 - \sqrt{(KC^0 R - KC^0 + 1)^2 + 4KC^0}}{2KC^0} \right]$$
(3)

273 Modeling the experimental data ($\delta_{obs} = f(R)$) with equation (3) enables us to estimate the binding constant K for the 1:1 complexation model. The individual chemical shifts (δ_0 and $\delta_{1:1}$) can be 274 275 measured from independent experiments of a free CD solution and a solution with a large excess of Cluster over CD, respectively. For a more complex model, such as a 1:2 adduct, it becomes 276 277 difficult to use this NMR approach to resolve the binding constants $K_{1:1}$ and $K_{1:2}$ due to the 278 overparameterization. Indeed, prior knowledge of the individual chemical shifts $\delta_{1:1}$ and $\delta_{1:2}$, is required, that are difficult to determine in fast exchange spectra. In this case, other experimental 279 280 techniques such as ITC would be more appropriate.

281 Thus, ¹H NMR is a useful internal probe to identify the nature and the strength of the 282 supramolecular interaction between CD and inorganic clusters. In particular, the binding sites 283 (e.g., secondary versus primary face), the location of the interaction (e.g., the interacting proton), 284 and the association constants are the most valuable information determined. As a result, several 285 types of CD-cluster hybrids are obtained from weak surface contact adducts to deeply encapsulated host-guest complexes.^{30,41,55} Tables 1-3 summarize the main CD-based hybrid 286 287 adducts and complexes observed with POM, boron, and octahedral metal-atom clusters studied 288 in solution by NMR.

290	Table 1. Summary	of reported CD-POM addu	cts studied in solution by NMR.

CD	РОМ	CD-POM adduct ^a	binding site ^b	Most affected H	Δδ (ppm) ^c	Binding constant $K_{1:1} (10^3 \text{ M}^{-1})^{\text{d}}$	Ref.
α-CD	$[PW_{12}O_{40}]^{3}$	CD•POM	P. F.	H6	+0.20	0.98	51,56
	$[P_2W_{17}O_{61}\{Sn(C_6H_4I\}]^{7}$	POM@CD	S. F.	Н5	+0.16	0.78	19
	$[P_2Mo_5O_{23}]^{6}$	CD•POM	E. W.	-	0	-	57
	[AlMo ₆ (OH) ₆ O ₁₈] ³ -	3CD•4POM	E. W.	H2	+0.06	-	58
	$[PMo_{12}O_{40}]^{3}$	-	P. F.	H6	+0.20	0.26	51
	$[V_6O_{13}((OCH_2)_3C-CH_2CH_3)_2]^{2-1}$	-	S. F.	Н5	-0.04	0.23	59
	$[V_6O_{13}((OCH_2)_3C-NO_2)_2]^{2-1}$	POM@CD	P. F.	H3	-0.01	0.30	59
	$[V_6O_{13}((OCH_2)_3C\text{-}CH_2OH)_2]^{2\text{-}}$	-	-	-	0	-	59
	$[V_6O_{13}((OCH_2)_3C\text{-}NH(BOC))_2]^{2\text{-}e}$	-	P. F.	H5	+0.12	0.36	59
	$[MPd_{12}O_8(PhAsO_3)_8]^{2\text{-}\mathrm{f}}$	POM@2CD	S. F.	Н3	-0.03	-	35
β-CD	$[PW_{12}O_{40}]^{3}$	-	P. F.	H6	+0.15	0.43	51
	$[P_2W_{17}O_{61}\{Sn(C_6H_4I\}]^{7-}$	POM@CD	P. F.	H5	-0.29	1.0	19
	$[P_2Mo_5O_{23}]^{6}$	CD•POM	E. W.	-	0	-	57
	[MnMo ₉ O ₃₂] ^{6.}	POM@CD	E. W.	H1/H2	-0.07	-	60
	$[PMo_{12}O_{40}]^{3}$	POM@2CD	P. F.	H6	+0.15	0.39	23,51
	$[V_6O_{13}((OCH_2)_3C\text{-}CH_2CH_3)_2]^{2\text{-}}$	-	S. F.	H5	-0.17	0.70	59
	[V ₆ O ₁₃ ((OCH ₂) ₃ C-NO ₂) ₂] ² -	POM@CD	S.F.	H5	-0.15	0.47	59

	[V ₆ O ₁₃ ((OCH ₂) ₃ C-CH ₂ OH) ₂] ²⁻	POM@2CD	S. F.	Н5	-0.10	0.29	59
	[V ₆ O ₁₃ ((OCH ₂) ₃ C-NH(BOC)) ₂] ^{2-e}	POM@2CD	S. F.	Н5	-0.10	1.2	59
γ-CD	[W ₆ O ₁₉] ² -	POM@CD	I. C.	Н5	+0.50	-	26
	$[PW_{12}O_{40}]^{3}$	POM@CD	S. F.	Н3	+0.53	140	25,51,54,61
	[SiW ₁₂ O ₄₀] ⁴⁻	-	P. F.	H6	+0.30	17	25,38
	$[BW_{12}O_{40}]^{5}$	CD•POM	P. F.	H6	+0.33	1.1	25
	$[H_2W_{12}O_{40}]^{6-}$	-	P. F.	H6	+0.07	0.023	25
	[SiW ₁₁ MoO ₄₀] ⁴⁻	-	P. F.	H6	+0.29	11	54
	[SiW ₁₁ MoO ₄₀] ⁵⁻	-	P. F.	H6	+0.10	0.73	54
	[PW ₁₁ VO ₄₀] ⁴⁻	-	P. F.	H6	+0.31	7.8	54
	$[PW_{11}VO_{40}]^{5-}$	-	P. F.	H6	+0.10	0.86	54
	[SiW ₁₁ VO ₄₀] ^{5.}	-	P. F.	H6	+0.28	0.39	54
	[SiW ₁₁ VO ₄₀] ^{6.}	-	-	-	0	0.13	54
	$[P_2W_{18}O_{62}]^{6}$	POM@2CD	P. F.	H6	+0.30	3.2	28
	$[P_5W_{30}O_{110}]^{15}$	CD•POM	S. F.	H6	+0.05	0.43	27
	$[P_2Mo_5O_{23}]^{6}$	CD•POM	E. W.	-	0	-	57
	[Mo ₆ O ₁₉] ²⁻	POM@CD	I. C.	Н5	+0.65	-	26
	[A1Mo ₆ (OH) ₆ O ₁₈] ³ -	CD•POM	E. W.	H2	+0.02	-	58
	[An-AlMo ₆ (OH) ₃ O ₁₈] ^{3-g}	-	I. C.	Н5	-0.40	-	18
	$[An_2 - MnMo_6O_{18}]^{3-g}$	-	I. C.	Н5	-0.40	-	18
	[MnMo ₉ O ₃₂] ^{6.}	POM@2CD	E. W.	H2	-0.11	-	60
	$[PMo_{12}O_{40}]^{3}$	POM@2CD	S. F.	H3	+0.70	56	23,51
	$[Mo_{154}O_{462}H_{14}(H_2O)_{70}]^{14}$	CD@POM	E. W.	H2	+0.50	-	30
	$[V_6O_{13}((OCH_2)_3C-CH_2CH_3)_2]^{2-1}$	-	P. F.	H6	+0.11	0.32	59
	$[V_6O_{13}((OCH_2)_3C-NO_2)_2]^{2}$	POM@2CD	P. F.	Н5	+0.24	1.4	59
	$[V_6O_{13}((OCH_2)_3C-CH_2OH)_2]^{2}$	POM@2CD	P. F.	H6	+0.12	0.090	59
	$[V_6O_{13}((OCH_2)_3C-NH(BOC))_2]^{2-e}$	-	P. F.	H6	+0.22	1.0	59

CD proton between its free and bound states, $\Delta \delta = \delta_{bound} - \delta_{free}$. ^{d)} calculated by NMR or

ITC. ^{e)} BOC = N-tert-butoxycarbonyl. ^{f)} M = Co, Ni, Zn. ^{g)} An = anthracene group.

CD	{M ₆ } cluster	CD-Cluster adduct ^a	binding site ^b	Most affected H	Δδ (ppm) ^c	Binding constant $K_{1:1} (10^3 \text{ M}^{-1})^{\text{d}}$	Ref
α-CD	$[{\rm Re}_6 {\rm S}_8 ({\rm CN})_6]^4$	Cluster@2CD	S. F.	Н3	+0.15	0.27	41
	$[{\rm Re}_6{\rm Se}_8({\rm CN})_6]^4$	CD•POM	S. F.	H3	+0.15	0.030	41
	[Re ₆ Te ₈ (CN) ₆] ⁴	2CD•POM	E. W.	-	0	0	41
β-CD	$[{\rm Re}_6 {\rm S}_8 ({\rm CN})_6]^4$	Cluster@2CD	S. F.	H3	+0.30	0.64	41
	$[{\rm Re}_6{\rm Se}_8({\rm CN})_6]^{4-}$	Cluster@2CD	S. F.	H3	+0.45	0.78	41
	[Re ₆ Te ₈ (CN) ₆] ⁴	Cluster@2CD	S. F.	Н3	+0.15	0.084	41
γ-CD	$[{\rm Re}_6 {\rm S}_8 ({\rm CN})_6]^4$	Cluster@2CD	S. F.	H3	+0.40	0.90	6
	$[{\rm Re}_6{\rm Se}_8({\rm CN})_6]^{3-}$	Cluster@2CD	S. F.	Н3	+0.2	225	49
	$[{\rm Re}_6{\rm Se}_8({\rm CN})_6]^{4-}$	Cluster@2CD	S. F.	Н3	+0.45	1.5	6
	[Re ₆ Te ₈ (CN) ₆] ⁴⁻	Cluster@2CD	S. F.	Н5	+0.35	38	6
	$[{\rm Re}_6 {\rm S}_8 ({\rm H}_2 {\rm O})_6]^{2+}$	Cluster@2CD	S. F.	-	0	-	62
	$[Nb_6Cl_{12}(H_2O)_6]^{2+}$	Cluster@2CD	E. W.	H1/H4	-0.08	2.2	1
	$[W_6Br_8Cl_6]^{2\text{-}}$	Cluster@2CD	S. F.	H3	+0.46	-	48
	$[W_6I_8Cl_6]^{2-}$	Cluster@2CD	S. F.	H3	+0.60	-	48
	$[Mo_6Br_8Cl_6]^{2\text{-}}$	Cluster@2CD	S. F.	H5	+0.40	-	63
	$[Mo_6I_8Cl_6]^{2-}$	Cluster@2CD	S. F.	H5	+0.60	-	63
	$[Mo_6Cl_{14}]^{2}$	Cluster@2CD	S. F.	H3/H5	+0.04	-	50
	$[Ta_6Br_{12}(H_2O)_6]^{2+}$	Cluster@2CD	S. F.	H3	+0.60	150	28

297 Table 2. Summary of reported CD-octaneural metal cluster adducts studied in solution by N	297	Table 2. Summary	of reported CD-octahedral metal cluster adducts studied in solution by N	NMR.
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From crystal structure.^{b)}S.F. = secondary face, E.W. = external wall.^{c)} difference in NMR chemical shift of most affected CD proton between its free and bound states, $\Delta \delta = \delta_{bound} - \delta_{bound}$ 300 δ_{free} . ^{d)} calculated by NMR or ITC.

302 Table 3. Summary of reported CD-boron cluster adducts studied in solution by NMR.

CD	Boron cluster	CD-Cluster adduct ^a	binding site ^b	Most affected H	Δδ (ppm) ^c	Binding constant <i>K</i> _{1:1} (10 ³ M ⁻¹) ^d	Ref.
α-CD	$[B_{10}H_{10}]^{2}$	Cluster@CD	S. F.	H3	+0.05	0.035	55
	$[B_{10}H_9NCCH_3]^-$	-	S.F.	H3	+0.06	0.034	53
	$[\mathbf{B}_{12}\mathbf{H}_{12}]^{2}$	-	-	-	0	-	22
	$[B_{12}H_{11}SH]^{2}$	-	-	-	0	-	22
	$[(1,2-C_2B_9H_{11})_2-3,3'-Co]^-$	-	S. F.	H3	+0.25	-	42

³⁰¹

	$[(1,2-Me_2-1,2-C_2B_9H_9)_2-3,3'-Co]^{-1}$	-	S.F.	Н3	+0.10	-	42
	$[(8,8'-Cl_2-(1,2-C_2B_9H_{10})_2-3,3'-Co]^-$	-	S. F.	Н3	+0.10	-	42
	$[(8-I-1,2-C_2B_9H_{10})(1^{,2}-C_2B_9H_{11})-3,3^{,2}-C_0]$	-	S. F.	Н3	+0.15	-	42
β-CD	$[B_{10}H_{10}]^{2}$	Cluster@CDe	I.C.	Н5	+0.19	0.11	55
	$[B_{10}H_9NCCH_3]^-$	-	P. F.	Н5	+0.15	0.30	53
	$[\mathbf{B}_{12}\mathbf{H}_{12}]^{2}$	-	-	-	0	-	22
	$[B_{12}H_{11}SH]^{2-}$	-	S. F.	Н5	+0.32	-	22
	$[B_{12}I_{12}]^{2}$	-	P. F.	H6	+0.15	-	22
	$[B_{12}I_{11}\text{-}NH_2C_6H_4\text{-}m\text{-}NO_2]^{1-}$	-	-	Н3	+0.10	18	64
	$[B_{12}I_{11}-NH_2C_6H_4-p-NO_2]^{1-2}$	Cluster@CDe	S.F./ I.C.	Н3	-0.15	19	64
	$[B_{12}I_{11}\hbox{-} S_2C_6H2\hbox{-} N_2O\hbox{-} p\hbox{-} NO_2]^{1}$	Cluster@CD ^e	S. F.	H1/H2	-0.10	270	65
	$[B_{12}I_{11}\hbox{-} S_2C_4H_8N\hbox{-} C_6H2\hbox{-} N_2O\hbox{-} p\hbox{-} NO_2]^{1-}$	-	-	H6	-0.15	260	65
	$[B_{12}Br_{12}]^{2}$	-	S.F.	Н3	+0.08	-	22
	$[(1,2-C_2B_9H_{11})_2-3,3'-Co]$	-	S. F.	H3	+0.20	26	42
	$[(1,2-Me_2-1,2-C_2B_9H_9)_2-3,3'-Co]^-$	-	S.F.	H3	+0.30	21	42
	$[(8,8'-Cl_2-(1,2-C_2B_9H_{10})_2-3,3'-Co]^-$	-	S. F.	H3	+0.27	19	42
	$[(8-I-1,2-C_2B_9H_{10})(1^2,2^2-C_2B_9H_{11})-3,3^2-C_0]^-$	-	S. F.	H3	+0.25	24	42
γ-CD	$[\mathbf{B}_{10}\mathbf{H}_{10}]^{2}$	Cluster@CDe	I. C.	Н5	+0.19	0.060	55
	$[B_{10}H_9NCCH_3]$	-	P. F.	Н5	+0.18	0.72	53
	$[B_{12}H_{12}]^{2}$	Cluster@CDe	S.F.	Н5	+0.15	2.0	22
	$[B_{12}H_{11}SH]^{2}$	Cluster@CDe	S.F.	Н5	+0.32	7.8	22
	$[B_{12}Cl_{12}]^{2}$	Cluster@CDe	S. F.	Н3	+0.35	17	22
	$[B_{12}Br_{12}]^{2}$	Cluster@2CD	S. F.	Н5	+0.70	960	22
	$[B_{12}I_{12}]^{2}$	Cluster@CDe	S. F.	H1	+0.10	67	22
	$[B_{12}H_{11}OH]^{2}$	-	S. F.	Н5	+0.10	0.62	22
	$[B_{12}H_{11}NH_3]^{2}$	-	S. F.	Н3	+0.05	1.7	22
	$[B_{12}H_{11}N(CH_2CH_2CH_3)_3]^{2}$	-	P. F.	H6	+0.05	1.1	22
	$[B_{12}H_{11}O(CH_2)_4C(NH_2)CO_2H]^{2-}$	-	S. F.	Н5	+0.20	-	22
	$[B_{12}I_{11}\text{-}NH_2C_6H_4\text{-}m\text{-}NO_2]^{1\text{-}}$	-	-	Н5	+0.20	9.6	64
	$[B_{12}I_{11}\text{-}NH_2C_6H_4\text{-}p\text{-}NO_2]^{1\text{-}}$	-	-	Н5	+0.15	7.6	64
	$[B_{12}I_{11}\text{-}NH_2C_6H_4\text{-}m\text{-}NO_2]^{1\text{-}}$	-	-	H6	+0.20	150	65
	$[B_{12}I_{11}\text{-}NH_2C_6H_4\text{-}p\text{-}NO_2]^{1\text{-}}$	-	-	H6	+0.15	590	65
	$[(1,2-C_2B_9H_{11})_2-3,3'-C_0]^-$	-	P. F.	H6	+0.10	191	42

$[(1,2-Me_2-1,2-C_2B_9H_9)_2-3,3'-Co]^{-1}$	-	S. F.	H3	+0.10	300	42
$[(8,8'-Cl_2-(1,2-C_2B_9H_{10})_2-3,3'-Co]^-$	-	S. F.	H3	+0.15	3600	42
$[(8-I-1,2-C_2B_9H_{10})(1',2'-C_2B_9H_{11})-3,3'-Co]^-$	-	S. F.	H3	+0.15	1300	42

303a)From crystal structure when available. b) P. F. = primary face, S. F. = secondary face, I. C.304= internal cavity. c) difference in NMR chemical shift of most affected CD proton between305its free and bound states, $\Delta \delta = \delta_{bound} - \delta_{free}$. d) calculated by NMR or ITC. e) From DFT306calculation.

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4. Relaxation and line broadening

310 Supramolecular contact may induce line broadening due to dipolar coupling between neighboring 311 nuclear spins, and linewidths could be used as a probe to assess the strength of these interactions 312 and the interatomic distances. In presence of paramagnetic atoms, the effect should even be 313 amplified since the dominant NMR relaxation mechanisms are connected to the coupling of the 314 strong magnetic moment of the unpaired electron with the adjacent nuclear magnetic moment. 315 Examples of NMR studies involving paramagnetic species include reduced POMs and oxidized 316 octahedral clusters.^{49,54} Dramatic line broadening of NMR signals of the CD has been observed 317 upon host-guest complexation, particularly those present in the inner sites (H3 and H5). 318 Furthermore, relaxation studies showed significant decreases of relaxation rates T_1 and T_2 for these 319 specific protons up to two-order of magnitude.⁴⁹ Interestingly, longitudinal relaxation T_1 was found 320 to discriminate interacting protons better than the transverse relaxation T_2 .

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5. DOSY and 2D NMR tools to track aggregation and self-assembly

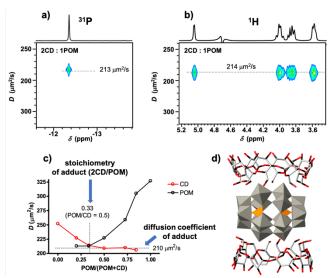
324 Diffusion Ordered SpectroscopY (DOSY) and Nuclear Overhauser Effect SpectroscopY
 325 (NOESY) or Rotating frame Overhauser Effect SpectroscopY (ROESY) are the most used 2D

326 NMR techniques to study the interaction of CD with organic, inorganic and hybrid guests. DOSY 327 is the 2D version of pulsed field gradient (PFG) NMR measurement, used to determine the self-328 diffusivity of dissolved species. Thus, DOSY experiment correlates the chemical shift with the 329 diffusion coefficient in the second dimension. This technique can allow differentiation between 330 free and aggregated CD to other species or between them, and thus provide information about 331 binding and exchange in CD-based supramolecular systems. As an example, the ¹H DOSY 332 experiment revealed the host-guest complex formation between the Lindqvist-type functionalized 333 hexavanadate $[V_6O_{13}((OCH_2)_3C-NO_2)_2]^2$ and γ -CD.⁵⁹ The diffusion coefficient of the free hybrid 334 POM in 2 mM D₂O drops from around 450 to 300 μ m²/s when complexed with γ -CD, as a clear 335 indication of the POM encapsulation process within the macrocycle. The diffusion coefficient of 336 the free CD also decreased in the presence of POM, but only slightly, as the volume/size of the 337 complex is dominated by that of CD. Nonetheless, the diffusion coefficient of hybrid POM is not 338 perfectly aligned with that of the host molecule, indicating dynamic dissociation in solution. 339 DOSY has thus proved an efficient technique for quantitatively studying chemical equilibria and 340 exchange processes in CD-based hybrids with inorganic nanoclusters.^{25,28}

341 For example, the formation of aggregates between γ -CD and Dawson-type POM [P₂W₁₈O₆₂]⁶ 342 has been well demonstrated by ¹H and ³¹P DOSY NMR, providing simultaneous access to the self-343 diffusion coefficient of γ -CD and that of the POM species, respectively.¹⁵ The DOSY 344 investigation, graphically shown in Fig. 8 reveals that the self-diffusion coefficients of the 345 interacting substrates decrease significantly in the POM/y-CD mixture that underline the formation 346 of stable aggregates in solution. Interestingly, the D value of both interacting components reaches 347 together the same minimum value at $D = 210 \ \mu m^2/s$ observed for the composition range close to the 1:2 ratio ($\frac{POM}{POM+CD} = 0.33$). This result highlights that stable aggregates are formed provoking 348

349 large changes of self-diffusion coefficients of each component in the presence of large excess of 350 the corresponding partner (see Fig. 8c). These DOSY NMR data corroborate nicely the analysis 351 given by Stoddart et al. from SAXS/WAXS measurements carried out on aqueous solution containing the Keggin-type anion and γ -CD.¹⁰ However, it is interesting to note that structural 352 353 analysis by XRD (Fig. 8d) reveals that POM and CDs assemble through weak interactions 354 highlighted by minimal variations in NMR chemical shifts (¹H, ¹⁸³W, and ³¹P), while DOSY or 355 SAXS/WAXS studies demonstrate that these hydrophilic species form stable aggregates in 356 aqueous solution. This demonstrates the high sensitivity of the DOSY technique to supramolecular 357 interaction.

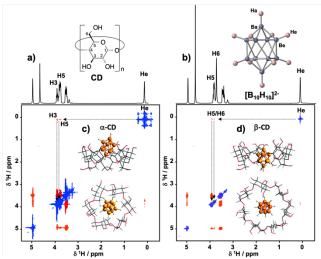
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Fig. 8. 2D DOSY (a) ³¹P and (b) ¹H spectra of a mixture of 2:1 γ -CD (3 mM): Dawson POM [P₂W₁₈O₆₂]⁶⁻ (1.5 mM) revealing common diffusion coefficient at ca. 214 μ m²/s. (c) Diffusion coefficients of the host (CD) and the guest (POM) measured by ¹H and ³¹P DOSY as a function of POM fraction in POM-CD system. The two curves cross at POM/CD ratio of 0.5 corresponding to the stoichiometry of (d) the sandwich adduct evidenced by single crystal XRD. Adapted from reference.¹⁵

367 NOE-based experiments such as NOESY and ROESY are techniques of choice for studying 368 non-covalent interactions in supramolecular systems involving cavitand molecules such as CDs. 369 Such NMR methodologies based on "through space" interactions (or dipolar coupling) allow 370 insight into which parts of a guest molecule are in spatial proximity to the hosting unit. ROESY 371 is generally preferred to NOESY for the study of nanoscopic assemblies built with CD, as it enables to probe longer distances up to 5 Å and provides positive ROE signal for 372 373 macromolecules and large chemical complexes. The sign of the NOE/ROE signals is an 374 important parameter for differentiating other dynamic phenomena such as spin diffusion and 375 exchange process, which lead to negative cross-peaks like the diagonal signals. Fig. 9 shows an 376 example of application of ROESY experiment to the $CD/[B_{10}H_{10}]^{2-}$ systems.⁵⁵ For a 1:1 mixture 377 of α -CD/[B₁₀H₁₀]²⁻, the interactions between equatorial protons of the boron cluster He, with H3 378 and H5 internal protons of the α -CD are revealed through the occurrence of cross NOE peaks. 379 For β -CD, the cross ROE peaks between the unresolved resonances of H5/H6 of the CD and 380 both Ha and He of $[B_{10}H_{10}]^2$ -indicate interactions between hydrogen atoms of the boron cluster 381 and the CD. These results are fully consistent with the DFT-optimized structures of the adducts 382 showing partially embedded cluster through the secondary face of α -CD and fully encapsulated 383 cluster within the β -CD cavity. ROESY NMR technique has been employed successfully to 384 probe the host-guest contacts through space occurring in solution between β-CD and Lindqvisttype hybrid hexavanadates.⁵⁹ Strong correlations were generally observed between the organic 385 386 part of $[V_6O_{13}((OCH_2)_3C-R)_2]^{2-}$ derivatives, $R = CH_2OH$, CH_2CH_3 , NO_2 , and internal protons of 387 the CD (mostly H3 and H5) fairly consistent with the host-guest structures observed in the solid-388 state. Recent examples of zirconium oxoclusters have also been reported.⁶⁶



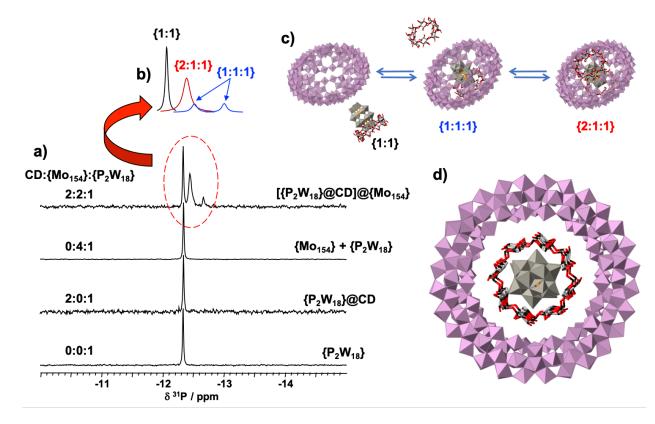
 $\begin{array}{ll} \mbox{391} & \mbox{Fig. 9. 1D and 2D } {}^1\mbox{H-}{}^1\mbox{H} \{ {}^{11}\mbox{B} \} \mbox{ ROESY NMR spectra in } D_2\mbox{O of a 1:1 mixture of (a) } \alpha\mbox{-CD/}[B_{10}\mbox{H}_{10}]^{2-} \\ \mbox{and (b) } \beta\mbox{-CD/}[B_{10}\mbox{H}_{10}]^{2-}.\ \mbox{(c) and (d) the corresponding optimized DFT structures of the CD-cluster} \\ \mbox{adducts. Adapted from reference.}^{55} \end{array}$

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6. Use of guest NMR probes

397 When available other NMR nuclei than ¹H and ¹³C of the host can be used to probe the 398 supramolecular interaction between the CD and the inorganic guest. These NMR nuclei should 399 then be present in the cluster part of the hybrid assembly, providing additional characterization 400 from the guest's point of view. For example, ³¹P NMR is used to follow the Dawson-type POM in 401 the ternary system γ -CD: $[P_2W_{18}O_{62}]^{6}$: $[Mo_{154}O_{462}H_{14}(H_2O)_{70}]^{14}$.³⁰ In the binary system γ -CD: 402 $[P_2W_{18}O_{62}]^{6}$, the characteristic signal of the POM appeared similar to that of the free species at -403 12.3 ppm as an indication of labile dynamic $\{1:1\}$ complex (Fig. 10). Introducing the giant blue 404 wheel {Mo₁₅₄} had led to resolved three additional signals assigned to POM@POM assembly with 405 one and two capping CDs, {1:1:1} and {2:1:1}, respectively (Fig. 10). Such a frozen three-406 component assembly is made possible by the CD binding ability, which enables to bring two 407 anionic species closer together. Indeed, no interaction could be observed in the absence of CD.

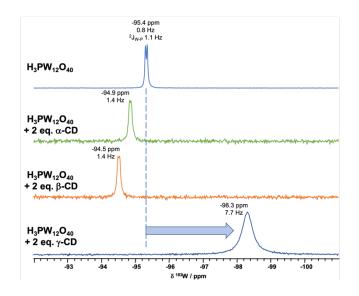


410 Fig. 10. (a) ³¹P NMR spectrum in D₂O of Dawson POM $[P_2W_{18}O_{62}]^{6-}$ in the presence of γ -CD and 411 the molybdenum blue wheel $[Mo_{154}O_{462}H_{14}(H_2O)_{70}]^{14-}$ compared to the spectra in the absence of 412 one of them or both of them. The spectrum in the ternary system can be decomposed into (b) three 413 components corresponding to the labile {1:1} CD:{P_2W_{18}}, and the ternary complexes {1:1:1} and 414 {2:1:1} in the CD:{P_2W_{18}}:{Mo_{154}} system in (c) equilibrium in solution. The crystallographic 415 structure of the {2:1:1} adduct (d) is consistent with a "host in host" hierarchical 416 POM@CD@POM structure. Adapted from reference.³⁰

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In labile dynamically systems, ³¹P NMR could not sense efficiently the host-guest interaction between the POM and the CD when the phosphorus is located in the internal pocket of the POM. The nucleus is in somehow shielded by the surrounding metal atoms, which protect the heteroatom from interaction with the surrounding environment. This is the case of the Dawson-type POM shown in Fig. 10 when exposed to CDs. However, external nuclei at the surface of the POM, such as ¹⁸³W NMR, should exhibit higher sensitivity towards supramolecular interaction at their surface. Fig. 11 shows the effect of presence of α -, β -, and γ -CD on the ¹⁸³W NMR of $[PW_{12}O_{40}]^3$.⁵¹



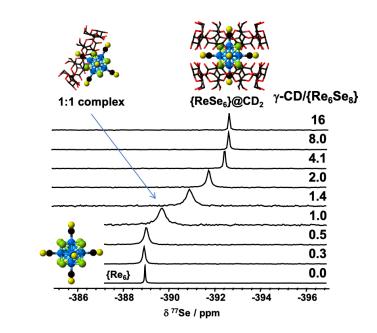


427 Fig. 11. ¹⁸³W NMR spectra in D₂O of 7.5 mM Keggin-type POM $[PW_{12}O_{40}]^{3-}$ in the absence and 428 presence of 2 equivalents of α-, β-, and γ-CD. Adapted from reference.⁵¹

430 The Keggin POM $[PW_{12}O_{40}]^{3-}$ is characterized by a single sharp ¹⁸³W signal at -95.4 ppm 431 corresponding to the 12 W equivalent nuclei of the POM structure (Fig. 11). This signal is actually 432 a doublet due to the scalar coupling with the neighboring P heteroatom (${}^{2}J_{W-P} = 1.1$ Hz). With the 433 addition of two equivalents γ -CD, this signal broadens and shifts toward high fields of about 3 434 ppm. These effects clearly indicate the formation of the inclusion complex with a very strong 435 association, which provides a significant shielding on the W nuclei. The line broadening of the 436 ¹⁸³W signal recalls that of the H3 proton of the CD (see Fig. 5) resulting from a strong host-guest 437 contact. This interaction can therefore be tracked from the host side by ¹H NMR or the guest side 438 by ¹⁸³W NMR. Furthermore, these effects are not observed with the other smaller CDs (Fig. 11), 439 confirming the unique behavior of γ -CD highlighted by the formation of the inclusion complex. 440 These effects are only moderately perceived by ³¹P NMR, with a maximum variation of 0.4 ppm 441 (from -14.3 to -14.7 ppm),⁵¹ while ¹⁸³W NMR recorded a difference of ca. 3 ppm, as the outer metal 442 centers are more exposed to interaction with CD than the inner heteroatom.

444 The metal transition octahedral clusters can also contain interesting active NMR nuclei. For 445 instance, the ½-I nucleus ⁷⁷Se (7.63% natural abundance) has been used to study the interaction of 446 the selenide rhenium cluster $[Re_6Se_8(CN)_6]^4$ with γ -CD.⁶ Fig. 12 shows the NMR titration of 24 447 mM solution by increasing the amount of the CD progressively up to 16 equivalents. A single 448 signal is observed shifting progressively from $\delta = -389$ ppm to the limiting value of -392.6 ppm. 449 This indicates a fast exchange regime between the different cluster-based species, that is, solvated 450 or embedded within γ -CD. The line broadening observed at intermediate CD equivalents (ca. 0.5 451 to 2) is indicative of formation of 1:1 intermediate labile complex. Similar behavior is observed in 452 ¹H NMR titration of the same system shown in Fig. 2a, where the most affected H3 resonance 453 experienced major line broadening at intermediate cluster equivalents. Job plots shown in Fig. 13 454 confirm the formation of the predominant 2:1 CD:Cluster type complex in aqueous solution as in 455 the XRD structure.⁶

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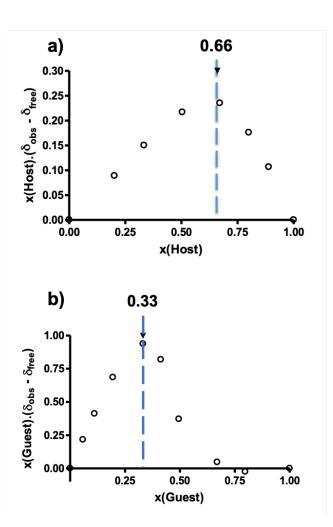
458 Fig. 12. ⁷⁷Se NMR titration of 24 mM [Re₆Se₈(CN)₆]⁴⁻ in D₂O by γ -CD. Adapted from references.³⁶

Job plot allows determining the stoichiometry of the adducts in solution, and can be applied either
on the host or the guest. By plotting the product of the molar fraction of the host (CD) or the guest
(cluster) in the mixture by the difference between observed chemical shift of the host (H3) or the
guest (⁷⁷Se) in presence and absence of the cluster or the CD as a function of the molar fraction in

465 both give consistent results with the formation of 2:1 adduct.

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the host or the guest, respectively, maxima indicate the stoichiometry. Job's plots drawn in Fig. 13

468 Fig. 13. Job plots obtained from NMR titration of (a) the host by the guest, or (b) the guest by the 469 host in the host-guest system γ -CD:[Re₆Se₈(CN)₆]⁴, by using ¹H NMR signal H3 of γ -CD in (a) 470 (spectra shown in Fig. 2a), or ⁷⁷Se NMR signal of the cluster in (b) (spectra shown in Fig. 12).

471 **7. Concluding remarks and outlook**

472 Over the past decade, significant progress has been made in the development of host-guest 473 assemblies based on CDs and inorganic molecules, and this area of research continues to attract 474 growing interest. Understanding the phenomena at play in solution in these systems, and 475 characterizing them in their original environment, is essential for the search of new, more elaborate 476 and complex supramolecular functional materials. NMR has proven to be a versatile, flexible and 477 easy-to-implement technique for probing the weak interactions between these macrocycles and 478 inorganic cluster molecules such as POMs, octahedral metal-atom clusters or boron clusters. 479 Qualitative and quantitative information can be extracted by measuring the strength of mutual 480 interaction, identifying binding sites, and studying chemical equilibria and molecular exchange.

481 Future research should focus on a better understanding of the role of water in host-guest 482 complexation driven by solvent effect. NMR nuclei such as ¹H, ²H, and ¹⁷O would be excellent 483 probes for identifying local interaction sites and monitoring dynamic behavior. Of particular 484 interest, relaxation (T_2 and linewidth) and diffusion (DOSY) experiments could provide valuable 485 information. Variable-temperature NMR experiments can also shed light on molecular motions. 486 Finally, the gradual change in the nature of the medium from water to organic solvents (e.g., 487 DMSO) may also bring insight on the proton exchange process involving the hydroxyl groups in 488 CD.

Nevertheless, with the ever-increasing complexity of these systems, the limits of the technique could be rapidly reached as of the size of the aggregates increases and the viscosity of the native medium decreases, as in hydrogels and supramolecular polymers. The restriction of molecular motion is the main drawback of high-resolution NMR spectroscopy. In this case, other

- 493 techniques more suited to semi-fluids and soft matter systems, such as cryo-TEM, DLS, SAXS,
- 494 SANS, and solid-state NMR, can take over.

495 **Conflicts of interest**

496 The authors declare no competing financial interest.

497

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507	References					
508 509 510 511	1 A. A. Ivanov, T. N. Pozmogova, A. O. Solovieva, T. S. Frolova, O. I. Sinitsyna, O. V. Lundovskaya, A. R. Tsygankova, M. Haouas, D. Landy, E. Benassi, L. V. Shestopalova, C. Falaise, E. Cadot, M. A. Shestopalov, P. A. Abramov and M. N. Sokolov, <i>ChemEur. J.</i> , 2020, 26 , 7479–7485.					
512	2 YM. Zhang, QY. Xu and Y. Liu, Sci. ChinChem., 2019, 62, 549–560.					
513	3 K. Kirakci, M. A. Shestopalov and K. Lang, <i>Coord. Chem. Rev.</i> , 2023, 481 , 215048.					
514	4 J. Lee, SS. Lee, S. Lee and H. B. Oh, <i>Molecules</i> , 2020, 25 , 4048.					

515 5 Y. Li, Y. Su, Z. Li and Y. Chen, *Polymers*, 2022, 14, 4855.

- 516 6 A. A. Ivanov, C. Falaise, P. A. Abramov, M. A. Shestopalov, K. Kirakci, K. Lang, M. A.
- 517 Moussawi, M. N. Sokolov, N. G. Naumov, S. Floquet, D. Landy, M. Haouas, K. A. Brylev, Y. V.
- 518 Mironov, Y. Molard, S. Cordier and E. Cadot, *Chem.-Eur. J.*, 2018, **24**, 13467–13478.
- 519 7 K. Li, K.-L. Zhu, L.-P. Cui and J.-J. Chen, *Dalton Trans.*, DOI:10.1039/d3dt00105a.
- 520 8 L. Ni, H. Li, H. Xu, C. Shen, R. Liu, J. Xie, F. Zhang, C. Chen, H. Zhao, T. Zuo and G.
 521 Diao, ACS Appl. Mater. Interfaces, 2019, 11, 38708–38718.
- 522 9 W. Qi, C. Ma, Y. Yan and J. Huang, *Curr. Opin. Colloid Interface Sci.*, 2021, 56, 101526.
- 523 10 Y. Chen, S. Sun, D. Lu, Y. Shi and Y. Yao, *Chin. Chem. Lett.*, 2019, **30**, 37–43.
- L.-J. Xu, C.-M. Wang, K. Yu, C.-X. Wang and B.-B. Zhou, *Coord. Chem. Rev.*, 2023, 481,
 215044.
- 526 12 A. A. Kuznetsova, V. V. Volchek, V. V. Yanshole, A. D. Fedorenko, N. B. Kompankov,
- 527 V. V. Kokovkin, A. L. Gushchin, P. A. Abramov and M. N. Sokolov, *Inorg. Chem.*, 2022, 61,
 528 14560–14567.
- M.-N. Shen, X.-W. Lin, J. Luo, W.-Z. Li, Y.-Y. Ye and X.-Q. Wang, *Mol. Syst. Des. Eng.*,
 2022, 7, 1570–1587.
- 531 14 Z. Y. Wei, Z. K. Wu, S. Ru, L. B. Ni and W. Wei, *Chem. J. Chin. Univ.-Chin.*, 2022, **43**, 532 20210665.
- 533 15 D. Prochowicz, A. Kornowicz and J. Lewinski, *Chem. Rev.*, 2017, **117**, 13461–13501.
- 534 16 J. M. Cameron, G. Guillemot, T. Galambos, S. S. Amin, E. Hampson, K. Mall Haidaraly,
- 535 G. N. Newton and G. Izzet, *Chem. Soc. Rev.*, 2022, **51**, 293–328.
- 536 17 W. Guan, G. Wang, B. Li and L. Wu, *Coord. Chem. Rev.*, 2023, **481**, 215039.
- 537 18 W. Guan, G. Wang, J. Ding, B. Li and L. Wu, Chem. Commun., 2019, 55, 10788–10791.
- 538 19 G. Izzet, M. Ménand, B. Matt, S. Renaudineau, L.-M. Chamoreau, M. Sollogoub and A.
 539 Proust, *Angew. Chem. Int. Ed.*, 2012, **51**, 487–490.
- 540 20 B. Li, L. Xuan and L. Wu, *Macromol. Rapid Commun.*, 2022, 2200019.
- 541 21 B. Zhang, W. Guan, S. Zhang, B. Li and L. Wu, *Chem. Commun.*, 2016, **52**, 5308–5311.
- 542 22 K. I. Assaf, M. S. Ural, F. Pan, T. Georgiev, S. Simova, K. Rissanen, D. Gabel and W. M.
 543 Nau, *Angew. Chem.-Int. Edit.*, 2015, 54, 6852–6856.
- 544 23 Y. Wu, R. Shi, Y.-L. Wu, J. M. Holcroft, Z. Liu, M. Frasconi, M. R. Wasielewski, H. Li
 545 and J. F. Stoddart, *J. Am. Chem. Soc.*, 2015, 137, 4111–4118.
- 546 24 K. I. Assaf and W. M. Nau, Angew. Chem. Int. Ed., 2018, 57, 13968–13981.
- 547 25 S. Yao, C. Falaise, A. A. Ivanov, N. Leclerc, M. Hohenschutz, M. Haouas, D. Landy, M.
 548 A. Shestopalov, P. Bauduin and E. Cadot, *Inorg. Chem. Front.*, 2021, 8, 12–25.

- 549 26 C. Falaise, M. A. Moussawi, S. Floquet, P. A. Abramov, M. N. Sokolov, M. Haouas and 550 E. Cadot, *J. Am. Chem. Soc.*, 2018, **140**, 11198–11201.
- 551 27 N. Leclerc, M. Haouas, C. Falaise, S. Al Bacha, L. Assaud and E. Cadot, *Molecules*, 2021,
 552 26, 5126.
- M. A. Moussawi, N. Leclerc-Laronze, S. Floquet, P. A. Abramov, M. N. Sokolov, S.
 Cordier, A. Ponchel, E. Monflier, H. Bricout, D. Landy, M. Haouas, J. Marrot and E. Cadot, J. *Am. Chem. Soc.*, 2017, 139, 12793–12803.
- C. Falaise, S. Khlifi, P. Bauduin, P. Schmid, W. Shepard, A. A. Ivanov, M. N. Sokolov,
 M. A. Shestopalov, P. A. Abramov, S. Cordier, J. Marrot, M. Haouas and E. Cadot, *Angew. Chem.*-*Int. Edit.*, 2021, **60**, 14146–14153.
- M. A. Moussawi, M. Haouas, S. Floquet, W. E. Shepard, P. A. Abramov, M. N. Sokolov,
 V. P. Fedin, S. Cordier, A. Ponchel, E. Monflier, J. Marrot and E. Cadot, *J. Am. Chem. Soc.*, 2017, **139**, 14376–14379.
- 562 31 X.-Y. Hu and D.-S. Guo, Angew. Chem. Int. Ed., 2022, e202204979.
- 563 32 D.-L. Long and L. Cronin, in *Recent Highlights I*, eds. C. D. Hubbard and R. VanEldik, 564 2021, vol. 78, pp. 227–267.
- 565 33 J. Luo, X. Sun, J.-F. Yin, P. Yin and T. Liu, *Giant*, 2020, **2**, 100013.
- 566 34 S. Passadis, T. A. Kabanos, Y.-F. Song and H. N. Miras, *Inorganics*, 2018, 6, 71.
- 567 35 M. Stuckart, N. V. Izarova, J. van Leusen, A. Smekhova, C. Schmitz-Antoniak, H. 568 Bamberger, J. van Slageren, B. Santiago-Schübel and P. Kögerler, *Chem. Eur. J.*, 2018, **24**, 569 17767–17778.
- 570 36 Z. Zhu, M. Wei, B. Li and L. Wu, *Dalton Trans.*, 2021, **50**, 5080–5098.
- 571 37 E. Bertaut and D. Landy, *Beilstein J. Org. Chem.*, 2014, **10**, 2630–2641.
- 572 38 Y. Fan, S. Lu and J. Cao, Int. J. Mass Spectrom., 2019, 435, 163–167.
- 573 39 P. Su, A. J. Smith, J. Warneke and J. Laskin, J. Am. Soc. Mass Spectrom., 2019, 30, 1934–
 574 1945.
- 575 40 Y. Yuguchi, *Trend. Glycosci. Glycotechnol.*, 2009, **21**, 1–12.
- A. A. Ivanov, C. Falaise, K. Laouer, F. Hache, P. Changenet, Y. V. Mironov, D. Landy, Y.
 Molard, S. Cordier, M. A. Shestopalov, M. Haouas and E. Cadot, *Inorg. Chem.*, 2019, 58, 13184–
 13194.
- 579 42 K. I. Assaf, B. Begaj, A. Frank, M. Nilam, A. S. Mougharbel, U. Kortz, J. Nekvinda, B.
 580 Grüner, D. Gabel and W. M. Nau, *J. Org. Chem.*, 2019, **84**, 11790–11798.
- 581 43 B. Chankvetadze, *Chem. Soc. Rev.*, 2004, **33**, 337–347.

- 582 44 H. Dodziuk, W. Kozminski and A. Ejchart, *Chirality*, 2004, **16**, 90–105.
- 583 45 M. Haouas, C. Falaise, C. Martineau-Corcos and E. Cadot, *Crystals*, 2018, 8, 457.
- 584 46 H. J. Schneider, F. Hacket, V. Rudiger and H. Ikeda, *Chem. Rev.*, 1998, **98**, 1755–1785.
- A. A. Ivanov, C. Falaise, A. A. Shmakova, N. Leclerc, S. Cordier, Y. Molard, Y. V.
 Mironov, M. A. Shestopalov, P. A. Abramov, M. N. Sokolov, M. Haouas and E. Cadot, *Inorg. Chem.*, 2020, 59, 11396–11406.
- 588 48 C. Falaise, A. A. Ivanov, Y. Molard, M. A. Cortes, M. A. Shestopalov, M. Haouas, E. 589 Cadot and S. Cordier, *Mater. Horizons*, 2020, 7, 2399–2406.
- A. A. Ivanov, C. Falaise, D. Landy, M. Haouas, Y. V. Mironov, M. A. Shestopalov and E.
 Cadot, *Chem. Commun.*, 2019, 55, 9951–9954.
- 50 A. A. Ivanov, P. A. Abramov, M. Haouas, Y. Molard, S. Cordier, C. Falaise, E. Cadot and
 593 M. A. Shestopalov, *Inorganics*, 2023, 11, 77.
- 594 51 S. Yao, C. Falaise, N. Leclerc, C. Roch-Marchal, M. Haouas and E. Cadot, *Inorg. Chem.*,
 595 2022, 61, 4193–4203.
- 596 52 K. Hirose, in *Analytical Methods in Supramolecular Chemistry*, Wiley-VCH, Germany, 597 Ed: C. A. Schalley. Chapter 2, 2012, vol. 1, pp. 27–66.
- 598 53 Z. El Hajj, S. Calancea, M. Haouas, D. Landy, D. Naoufal and S. Floquet, *J. Clus. Sci.*,
 599 2023, 10.1007/s10876-023-02468-x.
- 54 S. Yao, C. Falaise, S. Khlifi, N. Leclerc, M. Haouas, D. Landy and E. Cadot, *Inorg. Chem.*,
 2021, 60, 7433–7441.
- M. Diab, S. Floquet, M. Haouas, P. A. Abramov, X. Lopez, D. Landy, A. Damond, C.
 Falaise, V. Guerineau, D. Touboul, D. Naoufal and E. Cadot, *Eur. J. Inorg. Chem.*, 2019, 3373– 3382.
- 605 56 P. Yang, W. Zhao, A. Shkurenko, Y. Belmabkhout, M. Eddaoudi, X. Dong, H. N.
 606 Alshareef and N. M. Khashab, *J. Am. Chem. Soc.*, 2019, 141, 1847–1851.
- 607 57 P. Yang, B. Atshankiti and N. M. Khashab, *Crystengcomm*, 2020, 22, 2889–2894.
- 58 S. Khlifi, J. Marrot, M. Haouas, W. E. Shepard, C. Falaise and E. Cadot, J. Am. Chem.
 609 Soc., 2022, 144, 4469–4477.
- 610 59 I. F. Bamba, C. Falaise, J. Marrot, P. Atheba, G. Gbassi, D. Landy, W. Shepard, M. Haouas
 611 and E. Cadot, *Chem.-Eur. J.*, 2021, 27, 15516–15527.
- 60 Z.-Q. Qi, M.-Y. Wang, J.-C. Shen, Y.-Z. Lan, Z.-G. Jiang and C.-H. Zhan, *Chem.*613 *Commun.*, 2022, 58, 13616–13619.
- 614 61 Z.-G. Jiang, W.-T. Mao, D.-P. Huang, Y. Wang, X.-J. Wang and C.-H. Zhan, *Nanoscale*,
 615 2020, **12**, 10166–10171.

- 616 62 P. A. Abramov, A. A. Ivanov, M. A. Shestopalov, M. A. Moussawi, E. Cadot, S. Floquet,
 617 M. Haouas and M. N. Sokolov, *J. Clust. Sci.*, 2018, 29, 9–13.
- 63 A. A. Ivanov, M. Haouas, D. V. Evtushok, T. N. Pozmogova, T. S. Golubeva, Y. Molard,
 619 S. Cordier, C. Falaise, E. Cadot and M. A. Shestopalov, *Inorg. Chem.*, 2022, 61, 4462–14469.
- 64 T. Marei, M. K. Al-Joumhawy, M. A. Alnajjar, W. M. Nau, K. Assaf and D. Gabel, *Chem.* 621 *Commun.*, 2022, 58, 2363–2366.
- 65 K. I. Assaf, O. Suckova, N. Al Danaf, V. von Glasenapp, D. Gabel and W. M. Nau, *Org.*623 *Lett.*, 2016, 18, 932–935.
- 624 66 G. Hoyez, J. Rousseau, C. Rousseau, S. Saitzek, J. King, P. A. Szilagyi, C. Volkringer, T.
- 625 Loiseau, F. Hapiot, E. Monflier and A. Ponchel, *Crystengcomm*, 2021, 23, 2764–2772.