

## Corticothalamic network abnormalities underlying absence seizures

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1	<b>Corticothalamic network abnormalities</b>
2	underlying absence seizures
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#### 34 Abstract

Absence seizures (ASs) are genetic generalized seizures and one of the critical neurological 35 36 phenotypes that originates from abnormalities in corticothalamic (CT) networks. Genetic 37 analyses of cohorts with only ASs have identified 2p16.1 and 2q22.3 as significant loci and a 38 few genes related to intrinsic and synaptic membrane channels. In normal animals, the genetic 39 mutation of a single voltage- or ligand-gated channel in a single cortical or thalamic neuronal 40 population can lead to spontaneous ASs. The electrographic activity of ASs invariably initiates from a localized cortical area but the full expression of these seizures' clinical 41 42 symptoms requires interactions between cortical and thalamic networks. In contrast to focal and other genetic generalized seizures, the ictal activity of cortical and thalamic neurons 43 44 during ASs is predominantly characterized by a decreased, but highly synchronized, firing 45 with individual neurons often showing a different firing pattern from one seizure to the next. 46 The high remittance rate of children with ASs indicates that CT networks, and/or their 47 modulatory inputs from the basal ganglia and brain stem, have the inherent capacity to 48 compensate for the developmental abnormalities that lead to ASs, but the mechanisms of 49 remittance have not been investigated since all available models lack this feature of human 50 ASs.

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#### 53 **1. Introduction**

54 As early as the first half of the 20th century, the German neurologist Hans Berger 55 hypothesized that the 3Hz oscillations that he observed in the EEG of a patient during an absence seizure (AS) resulted from an abrupt withdrawal of tonic inhibitory influences 56 57 exerted by the thalamus on the cortex (see Avoli, 2012). Thus, although at that time the 58 mechanism of these electroencephalographic (EEG) oscillations (now known as spike-and-59 wave discharges, SWDs) was completely unknown, the cortex and the thalamus were already considered major players in the generation of this paroxysmal activity. Nowadays, there is a 60 61 general consensus that ASs are a disease of corticothalamic (CT) networks with a critical 62 modulation by the basal ganglia and brain stem (Crunelli et al., 2020).

63 Following a short summary of the clinical presentation, therapy and comorbidities of ASs, we 64 first discuss their genetics and the mechanisms involved in their initiation and maintenance in humans and in models of ASs. The main body of this chapter is a focused review of the 65 66 current state of knowledge of the molecular and cellular abnormalities in different neuronal 67 populations of CT networks that can bring about ASs, using evidence both from 68 investigations that have used selective genetic manipulations in normal non-epileptic animals 69 and from studies in five genetic models with spontaneous ASs that have been validated for 70 their strong face-, construct- and content-validity: the polygenic GAERS and WAG/Rij rats, 71 and the monogenic stargazer (STG), tottering (TG) and lethargic (LH) mice (Pitkänen et al., 72 2017). Due to space limitations, studies using only pharmacological manipulations are not 73 discussed.

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#### 75 **2.** Clinical presentation of ASs

76 An AS is a sudden and relatively brief lapse of consciousness associated with lack of 77 voluntary movements and distinctive SWDs at 2.5-4Hz in the EEG (Crunelli et al., 2020). 78 ASs are present, together with other seizure types, in many genetic generalised epilepsies 79 (GGEs), here we focus on ASs observed in childhood absence epilepsy (CAE) since in this 80 epilepsy type ASs are the only clinical symptom present and not compromised by other 81 epileptic phenotypes. In CAE, ASs begin between 3 and 8 years of age and can occur up to 82 ~200 per day. Approximately 70% of CAE patients go into spontaneous remission around 83 adolescence (Crunelli and Leresche, 2002), though the mechanisms underlying remission 84 have not been elucidated, since currently researched ASs models lack spontaneous remission. 85 Pharmacological treatment of ASs relies primarily on ethosuximide (ETX), valproic acid,

lamotrigine or their combinations, but ~30% of children are pharmaco-resistant and ~60%
show neuropsychiatric comorbidities (see Crunelli et al., 2020).

88

#### 89 **3. Genetics of ASs**

90 Strong evidence exists for a genetic component in CAE, possibly of a polygenic nature 91 (Perucca et al., 2020) and concordances of 70%–85% and 33% have been reported in 92 monozygotic twins and first-degree relatives, respectively (Gruenbaum et al., 2021). 93 However, most CAE patients do not have family history of epilepsy and siblings of affected 94 individuals have an 8% risk of ASs, which is lower than the expected risk for a recessive 95 (25%) or dominant (50%) inherited trait (Perucca et al., 2020).

96 Most AS genetic studies have used cohorts with a complex phenotype that included various 97 epilepsies and other neurological deficits, rendering it difficult to demonstrate an association 98 between gene abnormalities and ASs. Further, these studies focused on a few families and 99 reported diverse mutations mainly in  $\gamma$ -aminobutyric acid receptor (GABA<sub>A</sub>R) subunits and voltage-gated Ca<sup>2+</sup> channels (reviewed in Crunelli and Leresche, 2002). More recently, in a 100 101 large study of GGEs (that included CAE as well as juvenile absence epilepsy), no single gene 102 reached exome-wide significance, though lead associations were found for CACNAIG, 103 EEF1A2 and GABRG2, and, notably, GABAAR genes, but not excitatory receptor genes, were 104 enriched for missense variants (Feng et al., 2019).

105 The first genome-wide association study of a large, pure CAE cohort (778 Caucasian, 15 106 African) has identified two significant loci: 2p16.1 and 2q22.3 (International League Against 107 Epilepsy Consortium on Complex Epilepsies, 2018). Though the precise genes in these loci 108 that contribute to the CAE phenotype are still unknown, prioritized genes include FANCL, 109 BCL11A and ZEB2, i.e., genes not linked to membrane channels or transmitter pathways that 110 are generally considered key proteins for AS expression. Notwithstanding, mutations in 111 SCN1A, SCN8A, CACNA1H and CLCN2 have been identified in different Chinese CAE 112 cohorts (Xie et al., 2019).

A large number of spontaneous single gene mutations bring about spontaneous ASs in mice (Frankel, 2009), some of which have been validated as ASs models, including *CACNG2* (in STG mice), *CACNA1A* (in TG mice) and *CACNB4*, (in LH mice). Whole-genome sequencing of the polygenic GAERS rats has identified *CACNA1H* (R1584P) as the only gene-variant that selectively co-segregate with high-seizuring F2 progenies (Casillas-Espinosa et al., 2017). Notably, a combined proteomic and metabolomic analysis of GAERS rats has found an increase in pathways involved in synaptic transmission and metabolism of neurotransmitters and the down-regulation of the lysine degradation pathway in both cortex and thalamus (Harutyunyan et al., 2022). However, adult rats were used in this study, raising the issue of whether the observed changes in proteins and metabolites are the cause or a consequence of ASs.

In summary, single gene variants are causally linked to the expression of spontaneous ASs in mice models. However, the human homologs of these genes as well as the mutant genes found in the Chinese CAE cohorts are not located in the significant loci (2p16.1 and 2q22) identified in a large CAE cohort.

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#### 129 4. Cortical initiation networks of ASs

130 Although ASs are classified as generalized seizures, many studies in humans report that 131 SWDs originate from cortical regions before spreading to the entire cortical surface and other 132 brain regions, though the cohorts of these studies had a mixed phenotype including different 133 GGEs (reviewed in Crunelli et al., 2020). The first study on a pure CAE cohort reported 134 cortical-selective increases in the amplitude of blood-oxygen-level-dependent (BOLD) signals 135 occurring as early as 14s before the clinical and electrographic signatures of an AS could be 136 observed (Bai et al., 2010). The cortical area(s) showing these pre-ictal increases in BOLD 137 amplitude are mostly in frontal regions and can differ from child to child, but are generally 138 consistent across seizures for a given child (Bai et al., 2010). Importantly, the same cortical 139 areas show a post-ictal decrease in BOLD amplitude that can persist for up to 10s after 140 seizure-offset (Bai et al., 2010).

Changes in BOLD signals also occur in the thalamus during ASs (particularly, in the anterior,
centromedian, parafasicular and reticular nuclei), supporting its critical involvement in the
expression of these seizures (Bai et al., 2010; Guo et al., 2016).

144 The lack of generalization at the onset of an AS, i.e., the existence of a cortical initiation 145 network (CIN), had been originally reported in two rat models of ASs. It was found that the 146 peri-oral region of the primary somatosensory cortex (S1) was the cortical initiation network 147 in GAERS and WAG/Rij rats (Figure 1A) and that S1 Layer 5/6 excitatory pyramidal neurons 148 were the first to fire at the start of SWDs (Figure 1B) (Meeren et al., 2002; Polack et al., 149 2007). Moreover, BOLD signals in awake GAERS rats recapitulate human BOLD results 150 with an AS-linked decrease and increase in signal in S1 and ventrobasal (VB) thalamus, 151 respectively (McCafferty et al., 2023).

152 All these pre-seizure changes observed with non-invasive imaging and EEG in humans and 153 animal models may be the substrate for impaired consciousness, including loss of responses to 154 external stimuli and a pause in ongoing interactive behaviours, which occur not only during, 155 but even before the development of, SWDs (Guo et al., 2016). Notably, the magnitude of 156 BOLD and EEG changes in the cortex and thalamus that precede an AS are positively linked 157 to the severity of altered behavioural responses and consciousness during a seizure and may 158 thus be pre-determined by the state of CT networks before seizure-onset (Bai et al., 2010; 159 Guo et al., 2016). Notably, in GAERS rats, the expression of the AS-related behavioural 160 phenotype requires the mutual interactions of cortical and thalamic networks, as shown by the 161 lack of any overt behavioural sign when short (1-2sec) small amplitude SWDs are present in 162 the cortex but not in the thalamus (Figure 1C) (Polack et al., 2007), indicating that the 163 recruitment of the thalamus is a necessary step for the expression and maintenance of ASs 164 (see Part 6 below for intralaminar thalamic nuclei).

In summary, it is now well established that 1) SWDs are not generalized from the start but originate from selective CINs, with S1 Layer 5/6 pyramidal neurons being the first to fire in animal models; 2) the cortex and thalamus are critical elements for the full expression of ASs; and 3) the severity of the clinical symptoms of ASs may be predicted by the degree of preseizure EEG and BOLD changes and predetermined by persistently abnormal CT networks.

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#### 171 5. Mechanisms of altered excitability in deep layers of the CIN

172 The presence of a CIN in animal models has allowed to study the mechanism(s) underlying 173 the AS initiation. Strong evidence obtained in vivo and in vitro in four AS models (GAERS 174 and WAG/Rij rats, STG and TG mice) has shown enhanced excitability of Layer 5/6 175 pyramidal neurons (Kole et al., 2007; Polack et al., 2007) that is evident from the presence of 176 spontaneous giant depolarizing potentials, an increased number of neurons with intrinsic 177 strong bursting and N-methyl-D-aspartate (NMDA)-mediated EPSPs as well as an enhanced 178 firing rate, response to weak stimuli and gain of input-output function of Layer 5/6 neurons 179 (D'Antuono et al., 2005; Polack et al., 2007). Moreover, in WAG/Rij rats, it has elegantly 180 been shown that a selective decrease in HCN1, but not HCN2 channels in the distal apical 181 dendrites of Layer 5 pyramidal neurons leads to a decreased dendritic I<sub>h</sub> current, larger and 182 faster backpropagating action potentials and increased somato-dendritic coupling, so that the 183 summation of synaptic potentials is enhanced (Figure 1D) (Kole et al., 2007). These intrinsic 184 HCN channel-dependent changes in excitability are strengthened by a decreased peak

- 185 conductance and paired-pulse depression (PPD) of evoked GABA<sub>A</sub> inhibitory post-synaptic
- 186 potentials (eIPSPs) in S1 Layer 5 (and Layer 2/3) pyramidal neurons and evoked excitatory
- 187 post-synaptic potentials (eEPSPs) with a reduced PPD and sensitivity to baclofen, a GABA<sub>B</sub>R
- agonist. The net result of all these changes in synaptic activity in S1 Layer 5/6 pyramidal
- 189 neurons of WAG/Rij rats is an increased EPSP/IPSP ratio since i) the PPD of both EPSPs and
- 190 IPSP is decreased, ii) the number of NMDA EPSPs is increased, and iii) the peak conductance
- 191 of IPSCs is decreased.
- 192 Notably, a similar decrease in cortical inhibition is present in S1 of the TG mice, where a 193 comprehensive study of EPSPs and IPSPs evoked either by thalamic stimulation or locally 194 (either in Layer 4 or 5), has compellingly shown that the feed-forward inhibition from the 195 thalamus to Layer 4 is markedly compromised and correlated to the developmental onset of 196 ASs (Sasaki et al., 2006).
- Notably, there is a lower expression of PV+ interneurons in all S1 layers of WAG/Rij rats
  (Papp et al., 2018). Functionally, chemogenetic silencing of S1 PV+ interneurons in normal
  mice is sufficient to generate ETX-sensitive ASs (Panthi and Leitch, 2019), whereas
  DREADD-mediated activation of S1 PV+ interneurons, i.e., activation of local feed-forward
  inhibition, protects against pentylenetetrazol-induced SWDs (Panthi and Leitch, 2019) (see
  Part 7, below).
- 203 In summary, in WAG/Rij rats, we have a clear mechanistic explanation of the aberrant 204 behaviour of S1 Layer 5/6 pyramidal neurons where SWDs originate: intrinsic HCN1 loss-of-205 function (leading to enhanced synaptic potential summation) combines with an increased 206 EPSPs/IPSPs ratio to bring about the enhanced excitability observed in this neuronal 207 population. A decreased cortical inhibition is also present in TG mice, but similar detailed 208 mechanistic studies of S1 are not available for GAERS rats and STG and LH mice. PV+ 209 interneurons have an anti-absence role since their selective silencing in normal non-epileptic 210 animals leads to AS expression.
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#### 212 6. Cellular activity in the neocortex and thalamus during ASs

Using electrophysiological and imaging approaches in non-anesthetized AS models, the ictal neuronal activities that occur in the cortex and thalamus have recently been studied with single-cell resolution. Using  $Ca^{2+}$ -sensitive dyes in the primary visual cortex in STG mice, Meyer et al. (2018) reported that ~80% of neurons in all cortical layers reduce their activity before a SWD is detected in the EEG, while smaller neuronal pools show an increased, or no

218 change in, activity (Figure 2A). Interestingly, the ictal involvement of each neuron is not 219 fixed but fluctuates from seizure to seizure. An overall decrease in neuronal activity during 220 ASs has been confirmed in S1 cortical neurons of GAERS rats (McCafferty et al., 2023). 221 However, on closer examination, four patterns of ictal activity could be identified. While 37% 222 of neurons experienced a sustained ictal decrease in firing, 28% showed a transient firing 223 peak at seizure-onset. Importantly, regardless of the type of firing, all cortical neurons show a 224 marked increase in rhythmicity during SWDs compared to interictal periods and all changes 225 precede seizure-onset by up 80 seconds.

226 Simultaneous ensemble recordings in cortical neurons and VB thalamocortical (TC) and 227 nucleus reticularis thalami (NRT) neurons during spontaneous ASs in GAERS rats and y-228 hydroxybutyric acid (GHB)-induced ASs in Wistar rats under freely moving conditions show 229 that cortical afferent activity drives thalamic firing and temporally frames thalamic output via 230 feed-forward inhibition of TC neurons by NRT neurons (Figure 2D) (McCafferty et al., 231 2018). This results in a marked decrease in the overall firing of TC neurons (with only rare T-232 type  $Ca^{2+}$  channel-dependent burst firing being present ictally) (Figure 2B), which is causally 233 linked to the constitutively enhanced tonic GABA<sub>A</sub> current of TC neurons that is present in 234 this AS model as well as in STG and LH mice (Cope et al., 2009). Nevertheless, the total 235 firing of TC (and NRT) neurons is strictly in phase with each SWD cycle (Figure 2C), 236 indicating that the synchrony and strength of the ictal output of TC neurons to the neocortex is 237 not a feature of single, rhythmically bursting TC neurons, as previously suggested (Sorokin et 238 al., 2017), but an emergent property of this neuronal population. Notably, an altered firing 239 that precedes the start of an AS is also present in WAG/Rij rats and STG mice (Sorokin et al., 240 2017).

241 A clear limitation in our understanding of thalamic activity during ASs is that the vast 242 majority of available data have been obtained in first-order somatosensory thalamic nuclei 243 (see Part 7, below), whereas higher-order and intralaminar thalamic nuclei have been 244 relatively neglected. Notwithstanding, the rodent thalamic posterior nucleus may be one of the 245 key thalamic hubs in view of its critical involvement in pre- and post-ictal activity 246 (Lüttjohann and Pape, 2019). Notably, TC neurons in intralaminar thalamic nuclei show a 247 similar decrease in total firing as that of first-order thalamic nuclei during ASs but present 248 two patterns of ictal burst firing: one with a transient increase in bursts at seizure-onset and 249 one with a sustained increase in burst firing that lasts for the duration of the seizure (Atherton 250 et al., 2023). Moreover, it is interesting to note that extracellular alkalosis elicits ASs by activating intralaminar thalamic, but not cortical and first-order thalamic neurons of WAG/Rij
rats (Salvati et al., 2022b), a finding of key clinical significance since hyperventilation is a
trigger of ASs in many CAE patients.

Finally, although ASs are generated by CT network, both the direct and indirect basal ganglia pathways provide a fine control of the expression of these seizures (Deransart and Depaulis, 2002). Ictal changes in MRI BOLD signals in the basal ganglia of CAE cohorts have been consistently reported (Bai et al., 2010). Moreover, genetic deletion of the Munc-18-1 protein in cortico-striatal Layer 5 neurons elicits ETX-sensitive ASs, and activation of DREADDtransfected striatal fast-spiking interneurons blocks the spontaneous ASs of mice with wholebrain KO of Munc-18-1 (Miyamoto et al., 2019).

In summary, most cortical neurons decrease their firing during ASs but there are significant neuronal pools that either increase or do not change their firing rate. In the thalamus, the total and burst firing rate of TC neurons are reduced but that of the GABAergic NRT neurons is increased. Each cortical and thalamic neuron may show different activity changes form one seizure to the next. Intrathalamic thalamic nuclei are critical for AS synchronization and maintenance.

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#### 268 7. Induced genetic alterations in CT networks that lead to ASs

269 To understand how alterations in different neuronal populations of CT networks lead to the 270 expression of ASs, i.e., to an abnormal function of these networks, many studies have used 271 genetic tools to manipulate voltage- and ligand-gated membrane channels in normal animals. 272 In the following, we provide a detailed summary of all the studies that genetically 273 manipulated voltage- and ligand-gated membrane channels in normal non-epileptic animals 274 and compare these findings with studies that have investigated abnormalities in voltage- and 275 ligand-gated membrane channels of five well-validated AS models: GAERS and WAG/Rij 276 rats, and STG, TG and LH mice. Due to space limitations, we report changes in the 277 expression levels of intrinsic and synaptic membrane channels only if their functional 278 consequences have been investigated. Moreover, the abnormalities are described in 279 comparison with the respective non-epileptic control animals of each model.

#### 280 7.1 Ligand-gated channels

GABA<sub>A</sub>R  $\alpha$ 3 subunit knock-out (KO) mice do not express spontaneous ASs but show a decrease in GHB-induced SWDs (Winsky-Sommerer et al., 2008), whereas GABA<sub>A</sub>R β3 subunit KO mice show a complex epileptic phenotype that only at times includes ASs 284 (DeLorey et al., 1998). Spontaneous ASs are observed in mice knock-in (KI) for the human 285  $\gamma 2$  subunit R43Q mutation (Tan et al., 2007) (which had been identified in probands with 286 febrile seizures and ASs (see Crunelli and Leresche, 2002), whereas whole-brain KO of 287 GABA<sub>B</sub>R1 or GABA<sub>B</sub>R2 lead only to rare or no spontaneous SWDs, respectively (Gassmann 288 et al., 2004; Schuler et al., 2001).

Paradoxically, whole brain ablation of one of the subunits of the  $\alpha$ -amino-3-hydroxy-5methyl-4-isoxazolepropionic acid (AMPA) receptors (*GRIA4* KO mice) results in spontaneous ASs with decreased EPSCs in NRT but not TC neurons (Paz et al., 2011). Notably, decreased eEPSCs are present in TC neurons of TG mice and reduced NMDA and non-NMDA eEPSCs in LH mice (Caddick et al., 1999).

294 A common finding in GAERS rats and STG and LH mice is the constitutively enhanced tonic 295 GABA<sub>A</sub> current of TC neurons that results from a non-genetic loss-of-function of GAT1 (one 296 of the GABA transporters) that is necessary and sufficient for AS expression (Cope et al., 297 2009). In TC neurons of LH mice, GABA<sub>B</sub> eIPSCs are decreased but there is no change in 298 GABA<sub>A</sub> eIPSCs and baclofen-evoked currents (Caddick et al., 1999). The lack of changes in 299 GABAA IPSCs is shared by other models since no difference was found in GABAA miniature 300 IPSCs (mIPSCs) and eIPSCs in TC neurons of GAERS rats (Bessaïh et al., 2006; Cope et al., 301 2009) and TG mice (Caddick et al., 1999), respectively. An increased amplitude, and 302 decreased frequency and duration of mIPSCs but no change in PPD was reported in GAERS 303 NRT neurons (Bessaïh et al., 2006).

As far as excitatory transmitter function is concerned, a detailed analysis of synaptic potentials in NRT neurons has only been carried out in STG mice. Using a combination of electrical stimulation and photolysis of caged-glutamate, it was elegantly shown that a trafficking defect of AMPA receptors (i.e., decreased synaptic and increased perisynaptic receptors) leads to a reduction of AMPA EPSCs and a compensatory increase of NMDA EPSCs efficacy, leading to an increased NMDA/AMPA charge of EPSCs of both thalamic and cortical origin (Lacey et al., 2012).

311 7.2 Voltage-gated Ca<sup>+2</sup> channels

Selective knock-down (KD) of  $Ca_V 2.1$  channels in Layer 6 pyramidal neurons of normal mice leads to spontaneous ETX-sensitive ASs accompanied by increased T-type  $Ca^{2+}$  currents in the TC and NRT neurons and increased burst firing in the latter but not the former thalamic neural population (Bomben et al., 2016) (Figure 3). Since no changes in cortical Cav2.1

channels expression were observed with light microscopy, T-type Ca2+ currents and 316 317 associated burst firing was not investigated in cortical excitatory and inhibitory neurons in 318 these Layer 6 pyramidal neuron-selective KD mice, making it difficult to accept the authors' 319 conclusion that changes in thalamic neuron excitability were the only ones causally linked to 320 AS expression. Indeed, ablation of Cav2.1 channels in adult mice brings about ASs that are 321 not associated with changes in T-type  $Ca^{2+}$  currents or burst firing in TC neurons (Miao et al., 322 2019). Similarly, selective KO of Ca<sub>V</sub>2.1 channels in PV+ and somatostatin-positive (SST+) cortical interneurons leads to ASs with no change in T-type Ca<sup>2+</sup> currents or burst firing of TC 323 neurons (Figure 3) (Rossignol et al., 2013), demonstrating that a constitutive or induced 324 325 increase in these TC neuron parameters, as observed in TG mice and mice with selective 326 Layer 6 pyramidal neurons KD of Cav2.1 channels, is not necessary for the expression of 327 ASs.

328  $Ca_V 2.3$  channels are expressed in cortical and NRT, but not TC neurons, and their whole-329 brain KO leads to a decrease in GHB-induced ASs (Zaman et al., 2011). However, whether 330 this pathological phenotype is due to NRT or cortical Ca<sub>V</sub>2.3 channels remains to be 331 investigated.

332 Whole-brain overexpression of Ca<sub>v</sub>3.1 channels lead to spontaneous ETX-sensitive ASs 333 (Ernst et al., 2009), whereas mice with KO of these channels show a decrease in baclofen and 334 GHB-induced SWDs (Kim et al., 2001). Notably, the spontaneous ASs observed in Cav2.1 KO mice (Bomben et al., 2016; Miao et al., 2019; Song et al., 2004) are abolished by the 335 336 simultaneous KO of Ca<sub>V</sub>3.1 channels. However, despite the substantial expression of Ca<sub>V</sub>3.1 337 channels in cortical territories (Talley et al., 1999), the analysis of the cellular/network 338 abnormalities in these single and double KO mice was only carried out in the thalamus. So, 339 though these studies strongly point to a key role of Ca<sub>V</sub>3.1 channels in the genesis of ASs, 340 they do not clarify the brain area(s) and neuronal population(s) where these channels are 341 critical for AS expression (see below).

Notably, elevated T-type  $Ca^{2+}$  currents (mainly due to  $Ca_V 3.1$  channels) are present in TC neurons of different thalamic nuclei of four AS models (WAG/Rij rat, and STG, TG and LG mice) (Broicher et al., 2008; Zhang et al., 2002). The block of ASs by whole-brain genetic ablation of  $Ca_V 3.1$  channels in STG, TG and LH mice has been causally linked to the loss of function of these channels in TC neurons (Song et al., 2004). However, such conclusion is flawed since the block of ASs in these mice could also be due to the simultaneous ablation of  $Ca_V 3.1$  channels in the cortex where they are highly expressed (Talley et al., 1999). Indeed,

- ASs in GAERS rats are not affected by selectively blocking Ca<sub>v</sub>3.1 channels and related burst
  firing of TC neurons with a potent T-type Ca<sup>2+</sup> channels antagonist (McCafferty et al., 2018).
  Notwithstanding, TC neuron firing is necessary for AS expression since brief optogenetic
  manipulation of TC neurons abort ASs in WAG/Rij rats and STG mice (Sorokin et al., 2017).
- 353 Mice with whole-brain KO or selective KD of Cav3.3 channels in the NRT do not show 354 spontaneous ASs, but an unexpected increase in GHB-induced ASs (Lee et al., 2014) (Figure 355 3). The same phenotype is observed in double KO mice of  $Ca_V 3.3$  and  $Ca_V 3.2$  channels (Lee et al., 2014) where the T-type Ca<sup>2+</sup> current of NRT neurons is completely abolished. Thus, 356 notwithstanding the key role of the NRT neurons T-type  $Ca^{2+}$  current (and the burst firing that 357 358 it generates) in physiological activities expressed by CT networks (Crunelli et al., 2018), full 359 ablation of these NRT channels leads to potentiation of pharmacologically induced ASs. 360 Whether this is due to the increased tonic firing of NRT neurons that is present in these single 361 and double KO mice (Lee et al., 2014) or results from alterations in the activity of cortical 362 neuronal populations remains to be elucidated since the analysis of the cellular and network 363 abnormalities of these mice was only carried out in the thalamus.
- Elevated T-type Ca<sup>2+</sup> currents are also present in the NRT neurons of GAERS and WAG rats (Broicher et al., 2008; Cain et al., 2018) but have not been studied in the NRT of STG, TG and LH mice. In GAERS rats, this increased current is due to the R158P missense variant of *CACNA1H* (the Cav3.2 channel gene) and selective KD of these channels in NRT neurons markedly reduces ASs and normalizes their elevated burst firing (Cain et al., 2018). Moreover, KI of the human *CACNA1H* variant C456S in S1 of normal non-epileptic rats elicits spontaneous ASs in >60% of the injected animals (Wang et al., 2015).
- 371 7.3 Voltage-gated  $Na^+$  and  $Na^+/K^+$  channels
- Ablation of Nav1.2 channels in the whole-brain or selectively in telencephalic and 372 373 diencephalic neurons leads to spontaneous ETX-sensitive ASs with smaller SWDs being 374 present in the thalamus compared to those in S1, medial prefrontal cortex, and hippocampus 375 (Ogiwara et al., 2018). Ablation of Nav1.6 channels either in the whole-brain, only in 376 inhibitory neurons or in the NRT (Figure 3) brings about ETX-sensitive ASs, whereas no 377 seizures are observed when these channels are ablated in either all cortical neurons, in 378 glutamatergic excitatory neurons, or only in inhibitory neurons of the cortex, hippocampus, 379 and striatum (Makinson et al., 2017).
- 380 Mice with whole-brain KO of hyperpolarization-activated, cyclic nucleotide-gated  $Na^+/K^+$ 381 channels subtype 2 (HCN2) show spontaneous ASs (Ludwig et al., 2003) but HCN4 channel

382 KO mice do not (Zobeiri et al., 2019). However, extensive analysis of thalamic neuron 383 abnormalities but no data on cortical neurons were provided in these two studies, leaving a 384 knowledge gap on the relative contribution of HCN2 and HCN4 channels in the cortical and 385 thalamic sectors of the CT network. In TC neurons of different first-order thalamic nuclei of 386 both GAERS and WAG/Rij rats, there is an increased I<sub>h</sub> (Budde et al., 2005; Cain et al., 2015; 387 Kanyshkova et al., 2012), that most likely results from an increased HCN1 channel density 388 with no change in HCN2 and HCN4 expression (Cain et al., 2015). Notably, the increased 389 density of HCN1 channels in thalamic nuclei of WAG/Rij rats (Kuisle et al., 2006) is in 390 contrast to their decreased expression in S1 (Kole et al., 2007) (see Part 5, above), indicating 391 that in a given AS model, and potentially in humans with ASs, the same channels can be 392 differentially expressed in diverse sectors of the CT network. Notably, pharmacological block 393 of all HCN subtypes systemically, or locally in S1 or VB, blocks ASs (Iacone et al., 2021).

394 In summary, the following significant conclusions arise from these studies:

395 1. genetic ablations of diverse single channel abnormalities in single neuronal populations of 396 the CT network lead to spontaneous ETX-sensitive ASs (Figure 3); Cav2.1 channels in 397 cortical layer 6 neurons (Bomben et al., 2016), Cav2.1 channels in cortical PV+ and SST+ 398 interneurons (Rossignol et al., 2013), Nav1.6 channels in NRT neurons (Makinson et al., 399 2017), phospholipase C β1 subunit in NRT neurons (Chang et al., 2022) and phospholipase 400 C β4 subunit in TC neurons (Cheong et al., 2009), whereas selective KD of Cav3.3 401 channels in NRT neurons does not lead to spontaneous ASs but increases GHB-induced 402 SWDs (Lee et al., 2014);

- 403 2. secondary (compensatory) changes in a CT sector other than the one where the primary
  404 genetic manipulation had been made may be the critical element for ASs expression
  405 indicated by the increased T-type Ca<sup>2+</sup> currents in TC neurons of mice with selective KD
  406 of Ca<sub>v</sub>2.1 channels in cortical Layer 6 neurons (Bomben et al., 2016);
- different changes of membrane proteins expression may occur in diverse areas of CT
  network, as indicated by the decreased and increased expression of HCN1 channels in the
  cortex and thalamus, respectively, of WAG/Rij rats (Cain et al., 2015; Kole et al., 2007);
- 410 4. in view of the cortical location of SWDs initiation in humans and animal models (see Parts
  411 4 and 5, above), it is unfortunate that almost all investigations that used whole-brain
  412 genetic manipulations have studied abnormalities only in thalamic but not cortical neurons,
  413 even when the manipulated channels are (highly) expressed in both the cortical and
  414 thalamic sectors of CT networks.

415

#### 416 8. Conclusions

417 Abnormal genes found either in small Chinese CAE cohorts or in mice AS models are not 418 located in the significant loci (2p16.1 and 2q22) identified in the only large genome-wide 419 CAE study, though CACNA1H variants are present in Chinese probands and the GAERS rat 420 model of ASs. Selective ablation of Cav2.1, Cav3.3, and Nav1.6 channels as well as 421 phospholipase C  $\beta$ 1 and  $\beta$ 4 subunits in either Layer 6 pyramidal neurons, cortical PV+ and 422 SST+ interneurons, TC or NRT neurons of normal non-epileptic animals leads to ASs. In 423 contrast to focal seizures and other GGEs, the ictal activity of cortical and thalamic neurons is 424 predominantly characterized by a decreased, but highly synchronized, firing with all 425 component members of a neuronal population often changing firing pattern from one seizure 426 to the next. Selective chemogenetic and optogenetic manipulation of S1 PV+ interneurons and 427 TC neurons, respectively, conclusively demonstrates that these neuronal populations are 428 necessary for AS expression. The evidence in normal non-epileptic animals that a single 429 manipulation of different voltage- and ligand-gated membrane channels does lead to ASs 430 indicates the liability of CT networks towards the generation of these seizures. On the other 431 hand, the high remittance rate of children with CAE indicates that CT networks, and/or their 432 modulatory inputs from the basal ganglia and brain stem, have the inherent capacity to 433 compensate for the developmental abnormalities that lead to ASs. However, the mechanism 434 of remittance has not been investigated since all available AS models lack this feature of the 435 human disease.

436

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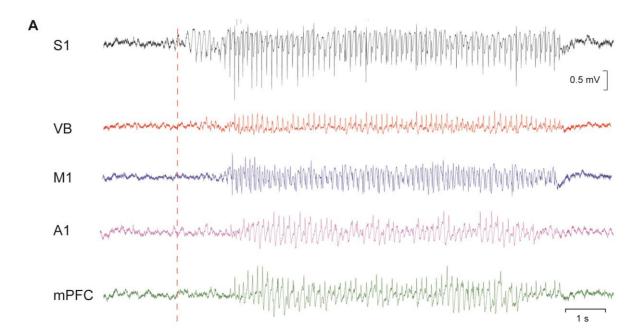
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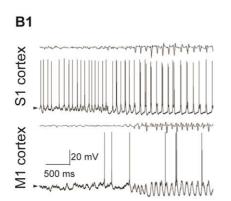
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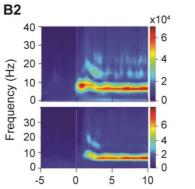
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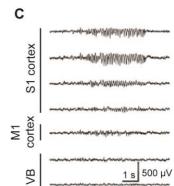
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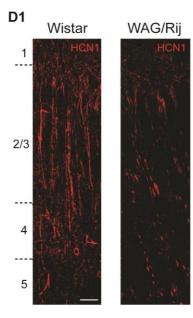
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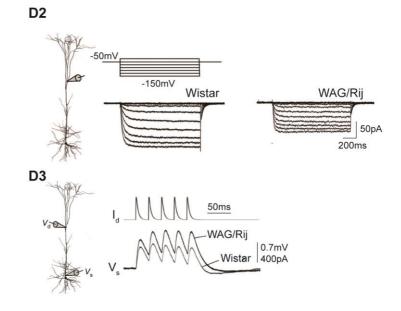








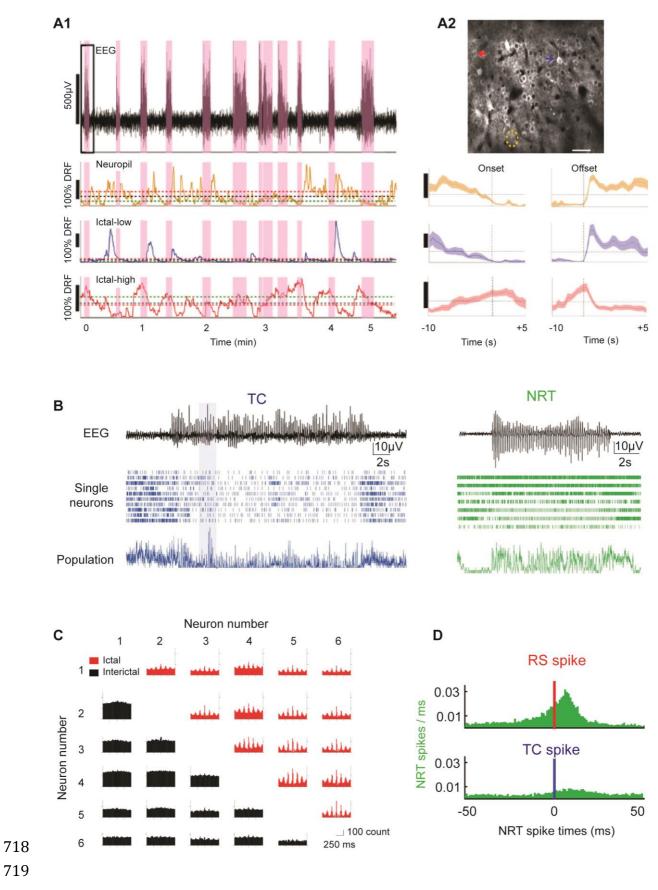




# Figure 1. Cortical initiation of absence seizures (ASs) and cellular abnormalities in Layer 5 neurons of AS models.

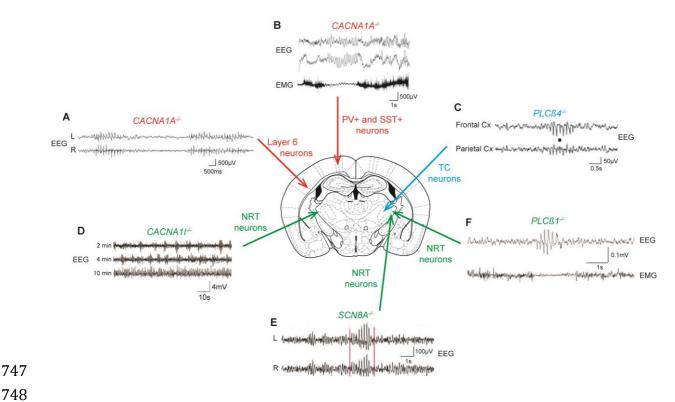
696 (A) Electroencephalographic (EEG) traces from the indicated cortical and thalamic areas in a 697 freely moving GAERS rat show the initiation of SWDs in the primary somatosensory cortex 698 (S1); VB: ventrobasal thalamic nucleus, M1: primary motor cortex, A1: primary auditory 699 cortex, mPFC: medial prefrontal cortex. (B1) Interictal and ictal intracellular activity (bottom 700 trace in each pair) of Layer 5 neurons, recorded in the same GAERS rat simultaneously with 701 the corresponding EEG (top trace in each pair). Note the hyperactivity of the neuron located 702 in the primary somatosensory cortex (S1 cortex), the cortical initiation network in this animal 703 model, compared to the neuron in the primary motor cortex (M1 cortex). (B2) Averaged time-704 frequency maps showing the occurrence of 6-8Hz SWDs in S1 before M1. (C) EEG traces 705 from different sites in the CT network of a GAERS rat show large amplitude SWDs in S1 706 (and very small SWDs in M1) but not in the VB, the somatotopic thalamic nucleus of S1, 707 which were not accompanied by any behavioural sign. (D1) HCN1 immunofluorescence in S1 708 shows weaker labeling in a WAG/Rij rat compared to a control Wistar rat (scale bar, 100µm). 709 (D2) Dendritic cell-attached patch-clamp recordings in the apical dendrites of an S1 Layer 5 710 pyramidal neuron show significantly lower amplitude of I<sub>h</sub> in a WAG/Rij rat compared to a 711 control Wistar rat. (D3) Simultaneous dendritic and somatic whole-cell recordings in an S1 712 Layer 5 pyramidal neuron. EPSPs were simulated by current injection in the dendrite (Id, top 713 trace) and their summation at the soma (Vs, bottom traces) was increased in a WAG/Rij rat 714 neuron compared to a control Wistar rat neuron. Adapted from McCafferty et al. (2018) (A), 715 Polack et al. (2007) (B, C) and Kole et al. (2007) (D1-D3).

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720 Figure 2. Activity of cortical and thalamic neurons during absence seizures (ASs).

(A1) Concomitant electroencephalographic (EEG) (top trace) and intracellular Ca<sup>2+</sup> changes 721 722 from Layer 2/3 GCaMP6-filled neurons in the visual cortex of a head-restrained STG mouse. The Ca<sup>2+</sup> changes of each neuron/neuropil patch over time are shown with the seizure epochs 723 724 highlighted in pink. Mean ictal activity (green dashed line), mean interictal activity (red 725 dashed line), and overall mean activity (black dashed line). (A2) The average activity (vertical 726 bar = 20%  $\Delta$ F/F) time-locked to the seizure onset and offset is shown for exemplary neuropil 727 (top), a neuron with low activity during ASs (ictal-low, middle) and a neuron with high 728 activity during ASs (ictal-high, bottom). The ictal-low and ictal-high neuron are marked by a 729 blue and red arrow, respectively in the 2-photon microscopy image shown at the top, with the 730 neuropil area highlighted by a yellow dotted circle. Scale bar =  $50\mu m$ . (B) EEG (top trace) 731 and spike-time raster plots from 10 thalamocortical (TC) neurons in the ventrobasal thalamic 732 nucleus and 7 nucleus reticularis thalami (NRT) neurons simultaneously recorded with a 733 silicon probe in a freely moving GAERS rats. Most TC neurons show a decrease in firing rate during the ASs, which is also evident from the population activity (bottom trace). NRT 734 735 neurons show two different types of ictal firing: one group increases their firing ictally (top 736 three cells in raster-plot) and another group shows a decreased activity (bottom four cells). 737 (C) Cross-correlograms of the total firing of 6 simultaneously recorded GAERS TC neurons 738 during interictal periods (black plots) and ASs (red plots). Despite the overall decrease in 739 firing during ASs (red plots), there is a marked increase in neuronal synchrony. (D) 740 Histograms of the firing of an NRT neuron recorded simultaneously with a cortical regular 741 spiking (RS) neuron (top) or a TC neuron (bottom) in a freely moving GAERS rat. Note that 742 the ictal NRT neuron firing (green bars) is mainly driven by the cortical RS neuron spikes 743 rather than the TC neuron spikes. Time zero marks the firing of the RS and TC neurons (red 744 and blue lines, respectively). Adapted from Meyer et al. (2018) (A) and McCafferty et al. 745 (2018) (B-D).





#### 749 Figure 3. Single genetic ablations that lead to absence seizures (ASs).

750 Schematic brain drawing (center panel) showing the location of the different single mutations 751 induced in either Layer 6 neurons, cortical parvalbumin- and somatostatin-positive (PV+ and 752 SST+, respectively) interneurons, thalamocortical (TC) and nucleus reticularis thalami (NRT) 753 neurons that have been shown to elicit spontaneous ASs (A, B, C, E, and F) or increase  $\gamma$ -754 hydroxybutyric acid-induced ASs (D). Panels show electroencephalographic (EEG) and 755 electromyographic (EMG) activity in mice with the illustrated genetic ablation. Minutes on 756 the left-hand side of the traces in D indicate the time from the  $\gamma$ -hydroxybutyric acid injection 757 (Cx: cortex; L: left; R: right). Adapted from Bomben et al. (2016) (A), Rossignol et al. (2013) 758 (B), Cheong et al. (2009) (C), Lee et al. (2014) (D), Makinson et al. (2017) (E), and Chang et 759 al. (2022) (F).