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The phylogenetic limits to diversity-dependent diversification

RAMPAL S. ETIENNE^{1,*}, BART HAEGEMAN², ÁLVARO DUGO-COTA³, CARLES VILÀ³,

Alejandro Gonzalez-Voyer
4 & Luis Valente 1,5

¹ Groningen Institute for Evolutionary Life Sciences, University of Groningen, Box 11103, 9700 CC

- ² Groningen, The Netherlands.
- ³ ² Theoretical and Experimental Ecology Station, CNRS and Paul Sabatier University, Moulis, France.
- ⁴ ³ Conservation and Evolutionary Genetics Group, Doñana Biological Station (EBD-CSIC), Seville, Spain.
- ⁵ ⁴ Department of Evolutionary Ecology. Instituto de Ecología, Universidad Nacional Autónoma de
- 6 México, Mexico city, Mexico.

 $_{7}$ ⁵ Naturalis Biodiversity Center, Darwinweg 2, 2333 CR Leiden, The Netherlands

^{*} Corresponding author. Email: r.s.etienne@rug.nl. Phone: +31 50 363 2230

⁹ Running head: Phylogenetic limits to diversity-dependence

Author contribution: RSE and LV conceived the idea for this paper, RSE and BH worked out the IW model mathematically, RSE implemented the likelihood calculation in the DAISIE package, LV and RSE conducted the bootstrap analyses, ADC, AGV and CV provided the data and expertise on the

¹³ Eleutherodactylus system, ADC prepared the Eleutherodactylus phylogeny and distribution maps, and

¹⁴ RSE, BH and LV each wrote parts of the manuscript. All authors commented on the manuscript.

Abstract

While the theory of micro-evolution by natural selection assigns a crucial role to competition, its role in 15 macroevolution is less clear. Phylogenetic evidence for a decelerating accumulation of lineages suggests a 16 feedback of lineage diversity on diversification. However, does this feedback only occur between close 17 relatives, or do distant relatives also influence their diversification? In other words: are there phylogenetic limits to this diversity-dependence? Islands form ideal systems to answer these questions, because their boundedness facilitates an overview of all potential competitors. The DAISIE (Dynamic Assembly of 20 Island biota through Speciation Immigration and Extinction) framework allows for testing the presence 21 of diversity-dependence on islands given phylogenetic data on colonization and branching times. The 22 current inference models in DAISIE assume that this diversity-dependence only applies within a 23

colonizing clade, i.e. all mainland species can colonize and diversify independently from one another. We term this clade-specific (CS) diversity-dependence. Here we introduce a new DAISIE model that assumes that diversity-dependence applies to all island species of a taxonomic group regardless of their mainland 26 ancestry, i.e. diversity-dependence applies both to species within the same clade and between different 27 clades established by different mainland species. We call this island-wide (IW) diversity-dependence. We present a method to compute a likelihood for this model given phylogenetic data on colonization and branching events and use likelihood ratio bootstrapping to compare it to the likelihood of the CS model in order to overcome biases known for standard model selection. We apply it to the diversification of 31 *Eleutherodactylus* frogs on Hispaniola. Across the Greater Antilles archipelago, this radiation shows 32 repeated patterns of diversification in ecotypes which are similar across clades. This could be suggestive 33 of overlapping niche space and hence between-clade interactions, i.e. IW diversity-dependence. But it 3/ could also be suggestive of only within-clade interactions, because between-clade interactions would have 35 blocked the same ecotype re-appearing. We find that the CS model fits the data much better than the IW model, indicating that different colonizations, while resulting in similar ecotypes, are sufficiently distinct 37 to avoid interacting strongly. We argue that non-overlapping distributions between clades (both spatially and in terms of ecotypes) cannot be used as evidence of CS diversity-dependence, because this pattern 30 may be a consequence of IW diversity-dependence. By contrast, by using phylogenetic data rather than 40 distributional data our method does allow for inferring the phylogenetic limits to diversity-dependent diversification. We discuss possibilities for future extensions and applications of our modelling approach.

Key words: birth-death model, diversity-dependence, *Eleutherodactylus*, Caribbean, adaptive radiation,
 island biogeography

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INTRODUCTION

"As species of the same genus have usually, though by no means invariably, some similarity in habits and
constitution, and always in structure, the struggle will generally be more severe between species of the
same genus, when they come into competition with each other, than between species of distinct genera".
This statement by Darwin in the Origin of Species (Darwin, 1859), known as the competition-relatedness
hypothesis (Cahill et al., 2008) or the phylogenetic limiting similarity hypothesis (Violle et al., 2011), or
Darwin's naturalization hypothesis in the field of invasion biology (Proches et al., 2008), has been the
subject of debate over the past decades, particularly in the field of phylogenetic community ecology
(Mayfield and Levine, 2010; HilleRisLambers et al., 2012; Pigot and Etienne, 2015; Gerhold et al., 2015;

Narwani et al., 2015; Germain et al., 2016; Cadotte et al., 2017; Wilcox et al., 2018). The consequences of the competition-relatedness hypothesis for macroevolution have received much less attention. Darwin (1859) formulated these consequences himself as "each new variety or species, during the progress of its formation, will generally press hardest on its nearest kindred, and tend to exterminate them." This 57 implies that with increasing diversity, speciation rates decline or extinction rates increase. This phenomenon has been referred to as diversity-dependent diversification (also somewhat confusingly called density-dependent diversification) since the 1970s (Raup et al., 1973; Walker and Valentine, 1984). Rabosky (2013) distinguishes Darwinian diversity-dependence, which does not imply an upper bound, 61 from asymptotic diversity-dependence, which by definition does impose an upper bound on diversity. We 62 leave the question aside whether an upper bound exists, and rather focus on the commonality of these 63 types of diversity-dependence: that diversity levels affect diversification, and in particular colonization and speciation rates decline with increasing diversity. (In other words: even though in this paper we model this negative diversity-dependence with a finite upper bound, we do not believe that whether the diversity limit is finite or infinite is crucial for our findings.) 67

There have been many suggestions of how such diversity limits come about (Rabosky, 2013; 68 Rabosky and Hurlbert, 2015). Here we do not enter this discussion, but we are interested whether there 60 is a phylogenetic limit to the effect of diversity, i.e. whether diversity-dependence only acts between 70 closely related species or (also) between distantly related species. There is considerable support for diversity-dependence in clades of phylogenetically closely related species (Foote and Miller, 2006; Phillimore and Price, 2008; Rabosky and Glor, 2010; Etienne et al., 2012; Jønsson et al., 2012; Foote et al., 2018), but there is also some evidence that phylogenetically distantly related (but ecologically similar) taxa reduce each other's diversification rates (Stanley, 1973; Sepkoski, 1996; Valkenburgh, 1999; 75 Jablonski, 2008; Silvestro et al., 2015; Pires et al., 2017). However, the latter evidence is relatively scarce 76 and comes mostly from fossil data. The question then presents itself whether molecular phylogenies can 77 also inform us about the phylogenetic limits to diversity-dependent diversification.

We propose that islands are the ideal arena to study these questions, because they are clearly defined systems where (exceptional) radiations have occurred. Moreover, as islands tend to be depauperate, we see cases where species released from competition have radiated to fill niches usually occupied by a different clade, e.g., woodpecker finches in the Galápagos. In MacArthur & Wilson's original work on island biogeography (MacArhur and Wilson, 1967) speciation receives little attention and therefore also diversity-dependent speciation, but colonization and extinction are diversity-dependent in their theory, as per capita colonization rates decrease and per capita extinction

rates are assumed to increase with increasing island diversity. The General Dynamic Model of island biogeography (Whittaker et al., 2008) explicitly assumes that the island's carrying capacity influences the diversification rates. However, neither of these classic works discusses the phylogenetic nature of the limits to diversification. Here, we consider two types of diversity-dependence, differing in the phylogenetic extent of diversity-dependence: the clade-specific (CS) level, where only species that descend from the same mainland (extinct or extant) species (possibly through multiple colonizations) reduce each other's speciation rate and colonization rates, and the island-wide (IW) level, where all island species of a predefined taxonomic group, that may descend from very different mainland ancestors, inhibit each other's speciation and colonization (see Fig. 1). Our focus is on species that occur on the island, and therefore by 'clade' we refer to a lineage of island species descending from the same mainland ancestor species. These island lineages are evidently embedded in a wider lineage containing both insular and mainland species, but the phylogeny of the species outside of the island is not considered here. The 97 CS scenario can be modelled by assuming a carrying capacity or upper limit to the number of species for each clade, while the IW model can be modelled by assuming an island-wide carrying capacity or upper limit to the total number of species. The CS and IW models are thus two extremes of a continuum of 100 diversity effects on colonization and speciation. In practice, these effects will not stop directly at the 101 clade established by a colonizing species as in the CS model, but they will also not generally extend to 102 all island species of the considered taxon as in the IW model. Modelling the continuum between the 103 extremes taking into account the phylogenetic (or phenotypic) distances between the mainland species 104 would be ideal, but the IW model is already difficult to handle mathematically and computationally (see 105 below), and thus modelling an intermediate case is currently unfeasible. However, we believe that with 106 these two extremes we are still able to gain more insight into the phylogenetic limits to 107 diversity-dependent colonization and speciation. 108

Diversity-dependence in speciation rates and colonization rates has been incorporated in the 109 DAISIE framework (Dynamic Assembly of Island biota through Speciation, Extinction and Immigration, 110 Valente et al. (2015)) that allows maximum likelihood estimation of rates of colonization, speciation and 111 extinction from phylogenetic data of the clades that colonized an island (or archipelago). In the first 112 simulations in this framework, diversity-dependence was of the IW-type (Valente et al., 2014). For 113 inference (i.e. parameter estimation), only the CS model was implemented (Valente et al., 2015), using 114 insight from analyses on single clades of closely-related species (Rabosky and Lovette, 2008; Etienne 115 et al., 2012), because the IW model presented technical difficulties. Here we overcome (some of) these technical difficulties by presenting a method to compute the likelihood of colonization and branching 117



Figure 1. Schematic representation of the two types of diversity-dependence. Phylogenetic trees A to C represent clades of island species descending from different colonization events. In the IW model (a) diversity-dependence extends to all island lineages in the data, whereas in the CS model (b) diversity-dependence extends only to species descending from the same mainland species. In this example there is a recolonization of species B. In the CS model diversification in clade B_1 depends on diversity in clade B_1 as well as clade B_2 , because these two clades descend from the same mainland ancestor (in a phylogeny that includes exclusively the island species, clades B_1 and B_2 would form a single monophyletic lineage). This is evidently also true for the IW model. For the CS model the same clade-level carrying capacity K applies to each clade separately (represented by the different colours of the shaded areas), while for the IW model it applies to all clades together.

¹¹⁸ events under the IW model.

We illustrate our method with an application to the colonization of Hispaniola by five lineages of 119 *Eleutherodactylus* frogs (genus *Eleutherodactylus*; Dugo-Cota et al., 2019), for which both CS and IW models can be verbally argued to apply. On the one hand, these lineages show, across the Greater 121 Antillean archipelago, repeated patterns of diversification into a similar set of ecotypes (Dugo-Cota 122 et al., 2019), suggesting a limited set of niches is available, which in turn implies that 123 diversity-dependence acts, but no further than within each clade (CS). On the other hand, the relatively 124 low geographic overlap in ecotypes between clades on Hispaniola suggests that diversity-dependence 125 extends to all *Eleutherodactylus* species on the island (IW) because species may have blocked 126 colonization of the same ecotype regardless of their phylogenetic relatedness. Our analysis, using only 127 phylogenetic data, shows that the CS model fits the data much better than the IW model. We discuss 128 this result and provide suggestions for further research avenues. 129

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Methods

¹³¹ Under the original DAISIE inference model (Valente et al., 2015) and its subsequent extensions (Valente ¹³² et al., 2017a,b, 2019) species can colonize an island at a rate γ , go extinct at a rate μ , and speciate via ¹³³ cladogenesis (when one island species splits into two, forming two new endemic species) at a rate λ^c or ¹³⁴ via anagenesis (when one island species diverges from its mainland ancestor becoming a new endemic ¹³⁵ species, without leading to an increase in diversity on the island) at a rate λ^a . CS-type ¹³⁶ diversity-dependence is implemented by allowing for rates of cladogenesis and colonization to decline ¹³⁷ with increasing diversity within a clade, with the number of species within each clade being limited by a ¹³⁸ CS carrying capacity, K. The maximum-likelihood implementation of DAISIE allows γ , μ , λ^c , λ^a and K ¹³⁹ to be estimated based on the distribution of times of island colonization and branching times within an ¹⁴⁰ island, extracted from divergence-dated molecular phylogenies. A diversity-independent model (DI) is ¹⁴¹ also implemented, i.e. by fixing K to infinity so that λ^c and γ do not decline with diversity.

A logical alternative model to CS in the island context is the IW model, where instead of a K per 142 clade there is an island-wide K that determines the maximum number of species that can coexist on an 143 island across all clades. This model was implemented in the first version of DAISIE, but only in 144 simulations (Valente et al., 2014). Until now, estimating parameters of an IW model has not been 145 attempted, because a) the model equations are rather cumbersome to write down and implement, and b) 146 parameter estimation is computationally demanding in terms of memory requirements and runtimes, 147 even for small data sets. Here we take on these hurdles. We develop a method of estimating parameters 148 of an IW model from phylogenetic data. The data requirements, parameters and simulation approach of 149 the DAISIE IW model are the same as for CS, except that diversity-dependence in λ^c and γ is 150 determined by an island-wide K, so that these rates decline with diversity of all island species rather 151 than simply diversity of the colonist clade they belong to. 152

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Likelihood of colonization and branching data for the IW model

We compute the likelihood of the data, consisting of colonization and branching events, for the IW model 154 using the Q-approach (Etienne et al., 2012; Laudanno et al., 2019). This approach is named after the 155 quantity Q(t), which is the probability that a random realisation of the model is consistent with the data 156 up to an arbitrary time t. In the supplementary material we construct the differential equations 157 governing the dynamics of Q(t), and explain how these equations, which apply to the dynamics between 158 colonization and branching events, are connected to one another across the colonization and branching 159 events. By solving these equations from the island emergence time to the present, we obtain $Q(t_p)$, the 160 quantity Q(t) evaluated at the present time t_p , from which the likelihood can be extracted (see 161 supplementary material for details). 162

Our computational procedure is based on the assumption that we have full information about the extant species. That is, we assume that the island phylogenies of the full set of extant species are known, together with the corresponding colonization times. This assumption simplifies not only the likelihood ¹⁶⁶ computation, but also the comparability with the CS likelihood. Indeed, in case of partial sampling from ¹⁶⁷ the phylogeny, the CS model distinguishes to what clade the missing species belong, while the IW model ¹⁶⁸ does not, making their likelihood incomparable. To guarantee full comparability we have also treated the ¹⁶⁹ likelihood of the CS model as a product of IW likelihoods with mainland pool size of 1 across the M¹⁷⁰ mainland species. That is, CS and IW only differ in whether clades established by mainland ancestors ¹⁷¹ are independent (CS) or are connected through each other's diversity (IW).

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Model fitting

We fitted five DAISIE models: a model without diversity dependence (DI, 4 free parameters), a model with clade-specific diversity dependence (CS-DD, 5 free parameters), a model identical to CS-DD but without anagenesis (CS-DD-noA, 4 free parameters); a model with island-wide diversity dependence (IW-DD, 5 free parameters), and a model identical to IW-DD but without anagenesis (IW-DD-noA, 4 free parameters). In all DD models the per capita rates of cladogenesis λ_N^c and colonization γ_N were assumed to linearly decline with diversity:

$$\lambda_N^c = \lambda_0^c (1 - N/K)$$
$$\gamma_N = \gamma_0 (1 - N/K)$$

where N is the total number of species in a clade in the CS model, and the total number of species on the entire island in the IW model. K is the carrying capacity per clade for the CS model (hence the same for each clade) and for the entire island for the IW model. We follow the original DAISIE model (Valente et al., 2015) by assuming no diversity-dependence in extinction or anagenesis, and we solely focus on diversity-dependence in rates of colonization and cladogenesis.

Phylogenetic data

¹⁷⁹ We used the dated phylogeny of *Eleutherodactylus* frogs by Dugo-Cota et al. (2019), which is based on ¹⁸⁰ four mitochondrial and three nuclear genes. The data set comprises 152 species of the genus, including ¹⁸¹ 148 Caribbean species, i.e. 89% of the Caribbean diversity, as well as four continental species. The ¹⁸² divergence dated phylogeny was reconstructed in BEAST v1.8.2, using secondary time calibration points ¹⁸³ extracted from the wider eleutherodactyline phylogeny of Heinicke et al. (2007). Dugo-Cota et al. (2019) ¹⁸⁴ reconstructed the biogeographical history of Caribbean *Eleutherodactylus* using BioGEOBEARS ¹⁸⁵ (Matzke, 2013, 2014) with a time-stratified analysis and nine geographical regions. They inferred five ¹⁸⁶ colonizations of Hispaniola from the mainland and surrounding islands (which are collectively referred to 8

as the mainland hereafter), each of which radiated on the island, to a great or lesser degree, producing
five in situ radiations of 28, 21, 8, 5 and 3 species (Table 1).

The Dugo-Cota et al. (2019) phylogeny includes 57 of the 66 Hispaniola species. Because fitting 180 the IW model assumes complete sampling of the extant species of the focal island, we inserted the 190 missing Hispaniola species by assigning them at random locations within the Hispaniola subclades that 191 they have been hypothesized to belong to. Information on the nine missing species and the detailed 192 rationale for including them in a given subclade are given in supplementary Table S1. There is no genetic 193 data available on GenBank for these missing species, because they have been recently described, are 194 known from a single specimen or are possibly extinct. We used a set of functions from the phytools R 195 package to assign missing species to clades (Revell, 2012). Four of the missing species were previously 196 considered subspecies, and have recently been elevated to species, and we thus randomly inserted them 197 at any height along the tip branch of the species they were previously assigned to. Four other species 198 have been proposed to belong to well-defined terminal clades based on morphology, and we randomly 199 inserted them at any position and at any height within those clades. We repeated this procedure 100 200 times on the five clades from the maximum clade credibility tree from BEAST, producing 100 sets of five 201 clades with complete sampling. The exact procedure is detailed in Table S1. One of the missing species, 202 E. neiba was not added to the tree because there is no previous hypothesis regarding its phylogenetic 203 position. We ran a sensitivity analysis including this species as a separate colonization, to assess whether in the unlikely case it formed a separate clade this would affect the results. These analyses showed that even if E. neiba formed an independent colonization, the same model would still be preferred. We 206 therefore did not include it in the analyses, as it is unlikely to modify the main findings. 207

We extracted colonization and branching times for each of the five Hispaniola radiations from these data. Colonization times were assumed to be the stem ages of the Hispaniola clades, as the stem age marks the divergence from the mainland sister clade (which is assumed to be do to the colonization event). Information on each of the Hispaniola clades is given in Table 1 and the phylogeny is shown in Fig. 2.

As the downstream DAISIE analyses are computationally demanding, we wanted to use only one data set for subsequent analyses. In order to perform an informed selection of the tree, we fitted the CS and the IW model with no anagenesis to each of the 100 sets of trees. The results of the analysis on the 100 sets of trees are shown in Table S2. The preferred model in all trees was CS. We thus used only tree set 52, which was the one with the highest likelihood for CS, for all subsequent analyses (hereafter 'empirical data set'). This may seem to introduce a bias in favor of the CS model, but we note that all



Figure 2. Phylogeny of the Hispaniola *Eleutherodactylus* frogs, and the ecotypes and spatial distribution of the tip species. A separate time-calibrated phylogenetic tree is shown for each of the five clades. Gray islands show the five inferred independent colonization events of Hispaniola. Colors at the tips of the phylogeny represent the species ecotypes (see legend). The same colors are used to show the species distributions for each independent colonization on the Hispaniola map (for visual clarity some transparency has been applied). The asterisk indicates where missing species have been added to the phylogeny according to taxonomic information, see Table S1. We note that our inference method only uses the phylogenetic information in the data, i.e. only the colonization and branching events.

loglikelihood differences (and all parameter estimates) were very similar across the 100 sets of trees: the
loglikelihood differences between CS and IW were between 5.2 and 6.6 with a median of 5.9 and set 52
had a loglikelihood difference of 6.4. All 100 data sets would have led to the same conclusions in our
model comparison (see Results). All sets of trees and corresponding DAISIE colonization/branching time

Clade	Number of species	Colonization time (Ma)	Geographical distribution
1	28	22.09(18.21 - 26.36)	Mixed
2	21	13.75(10.73 - 16.97)	100% South
3	3	11.03 (8.31 - 13.89)	100% North
4	5	8.85(6.91 - 10.8)	100% South
5	8	8.43(6.3 - 10.73)	87.5% North

Table 1. Characteristics of the five clades on Hispaniola: diversity, colonization times and geographical distribution within the island of Hispaniola.

R objects are provided in the supplementary material. The simulation code, functions to compute the likelihood under the two models, and a tutorial on how to run simulations and perform model-fitting are available in the R package DAISIE on CRAN and on Github (https://github.com/rsetienne/DAISIE).

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Likelihood optimization on the empirical data

We fitted each of the five DAISIE models 5 times to the empirical data set using different random sets of 227 starting parameters to avoid being trapped in local likelihood optima. We assumed an island age of 30 228 million years, consistent with the paleogeographical reconstruction of Iturralde-Vinent (2006) for when 229 Hispaniola was isolated from other landmasses. The mainland pool size M was set to 1000 frog species. 230 We note that this value is not crucial, because mainland pool size affects only the rate of colonization; 231 the product of mainland pool size and the rate of colonization, i.e. the total rate of colonization, is 232 practically constant (Valente et al., 2019). Indeed, parameter estimates were very similar for 233 optimizations with M = 300. 234

Maximum likelihood optimizations were run on the high-performance (Peregrine) cluster of the University of Groningen. Optimization of DI and CS-type models generally converged in a few hours. IW-DD model optimizations took between a few hours up to 10 days to complete.

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Goodness-of-fit

We simulated 5,000 data sets using the maximum likelihood parameters of the preferred CS-type model (CS-DD no anagenesis) and preferred IW-type model (IW-DD no anagenesis), hereafter the CS and IW models. We then plotted relevant statistics from the simulated data sets and compared them to those in the empirical data to study how well the models fit the data.

Bootstrap analysis

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We computed the AIC and BIC values and weights for model comparison, but because model selection 244 involving diversity-dependent models is known to be troublesome (Etienne et al., 2016), we used a 245 parametric bootstrap likelihood ratio test similar to Boettiger et al. (2012). This bootstrap analysis 246 additionally allowed us to assess bias and precision of parameter estimates. We chose the first 1000 out of 247 the 5000 data sets from each of the CS and IW simulations. Not all 5000 simulated data sets were used 248 for the bootstrap likelihood ratio test, because the subsequent analyses on these data sets were 240 computationally demanding. For each of the chosen 2000 data sets we fitted both CS and IW models, 250 resulting in a total of 4000 maximum likelihood optimizations. As starting values of the optimizations we 251 used the maximum likelihood parameters of the given model obtained for the empirical data, to make it 252 as likely as possible that we will find the global likelihood optimum, as it is expected to be around these 253 empirical maximum likelihood parameters. To really ensure local optima are avoided, the optimizations would need to be run many times from many different starting values, but this was computationally 255 unfeasible. In some cases it was not possible to use the parameters obtained in the optimization analyses 256 as initial parameters for the optimization. For instance, if a clade in data sets generated under the IW 257 model had more species than the value of K estimated for the CS model fitted to the empirical data, 258 using that K as a starting value to fit the CS model to the IW-simulated data would give a likelihood of 259 0. Therefore we calculated the starting K for each data set using the largest value of either the K260 estimated from the empirical data for the given model being fitted, or the maximum number of species in 261 a clade (CS model) or the total number of species on the island (IW model) in the simulated data set. 262

For the bootstrap likelihood ratio test we compared the logarithm of the likelihood ratio of CS 263 and IW in the empirical data (i.e. loglikelihood difference, loglikelihood of the CS model – loglikelihood 264 of the IW model) with the distribution of the logarithm of likelihood ratios from the data sets simulated 265 under CS and under IW (1000 data sets each). We computed the 95th percentile of the distribution under the IW model. If the loglikelihood difference of the empirical data falls to the right of this value, 267 then they are unlikely to be produced by the IW model, and if it is well within the distribution of the 268 data generated under the CS model, the CS model is selected. We also computed the 5th percentile of the 269 distribution under the CS model. If the loglikelihood difference of the empirical data falls to the left of 270 this value, then they are unlikely to be produced by the CS model, and if it is well within the distribution 271 for the data generated under the IW model, the IW model is selected. If the loglikelihood difference of 272 the empirical data falls between the two percentiles, then no model can be selected decisively.

RESULTS

Convergence of the five independent optimizations per model to the empirical data set was very good, 276 with all five runs finding the same maximum likelihood parameter set for each model. The preferred model using both AIC or BIC was CS-DD with no anagenesis (four free parameters) (Table 2). The 278 loglikelihood difference between the best CS model and the best IW model (both diversity-dependent) 279 was 6.43. This value points to the CS model as the best model also in the likelihood ratio bootstrap test 280 (see below). The models without anagenesis had virtually the same parameter values as their 281 counterparts allowing anagenesis to be different from 0. That is, the latter models had estimated rate of 282 anagenesis that were very close to 0. This is to be expected because all five frog clades radiated and 283 hence there is no evidence for anagenesis. The only signal of anagenesis in such a case could come from the observation of recolonizations of the same mainland species that established the clade(s). This is 285 because the model assumes that recolonizations can only occur after speciation has taken place (if it 286 happens before speciation takes place, the recolonization is assumed to reset the colonization time and is 287 then not observed). As the data did not contain recolonizations, the maximum likelihood estimate of 288 anagenesis is expected to be 0. Across the 100 sets of empirical trees the loglikelihood difference between the diversity-dependent CS and IW models ranged from 5.16 to 6.63, all of which suggest the CS model is highly preferred.

Table 2. Maximum likelihood parameter estimates and corresponding loglikelihood (LL) for five fitted models. DI: diversity-independent rates; DD-CS: clade-specific diversity-dependence in colonization and cladogenesis; DD-CS-noA: same, but with anagenesis rate fixed to 0; DD-IW: island-wide diversity-dependence in colonization and cladogenesis; DD-IW-noA: same, but with anagenesis rate fixed to 0. An asterisk indicates that the estimated value is practically 0. df: degrees of freedom, i.e. number of free parameters; AIC: Akaike information criterion; BIC: Bayesian information criterion.

model	λ_0^c	μ	K	γ	λ^a	LL	df	AIC	AIC weight	BIC	BIC weight
DI	0.18	0.03	∞	0.0002	0*	-215.87	4	439.75	0.00	445.53	0.00
DD-CS	0.44	0.11	36.44	0.0002	0^{*}	-208.67	5	427.34	0.27	434.58	0.15
DD-CS-noA	0.44	0.11	36.45	0.0002	0	-208.67	4	425.34	0.73	431.13	0.85
DD-IW	0.40	0.17	131.89	0.0003	0^{*}	-215.10	5	440.20	0.00	447.43	0.00
DD-IW-noA	0.40	0.17	131.96	0.0003	0	-215.10	4	438.20	0.00	443.98	0.00

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Goodness-of-fit

²⁹³ Using the estimated parameters for the diversity-dependent CS and IW models we generated simulated ²⁹⁴ data for which we computed several summary statistics. The distributions of the summary statistics ²⁹⁵ across these simulations fitted well with the empirical data for both models, but somewhat better for the

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²⁹⁶ CS model. (Fig. 3, Fig. S1, Fig. S2), as the empirical statistics and the medians across the simulations ²⁹⁷ are slightly more similar for this model.



Figure 3. Goodness-of-fit plots. Distributions of relevant metrics (number of species, number of colonizations leading to extant clades, size of the largest clade, and rank of the largest clade when clades are ordered according to their colonization time, rank 1 corresponding to the first colonization) obtained from 5,000 data sets simulated with the maximum likelihood parameters of the CS (top row) and IW (bottom row) models. Black line: median value, blue arrow and blue bar: value in the empirical data.

Bootstraps

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The analyses fitting the CS model to each of 1000 CS and 1000 IW simulated data sets all completed 299 successfully. For the analyses fitting the IW model to the same data sets, some runs runs could not be 300 completed within the limit we set (10 days). For the CS simulated data sets this was 0.8% of the 301 simulations and for the IW simulated data sets this was 1.6%. These were all data sets with little 302 information (only a single clade) where the estimation procedure went to very high values of the rate of 303 cladogenesis and colonization. We used the loglikelihood that was obtained after 10 days which is thus an 304 underestimate of the maximum IW loglikelihood, but the ML may not be much higher than this value 305 after 10 days. However, even if we make the unlikely assumption that in all of these aberrant simulated data sets the IW model is a better fit, they are so rare that our qualitative conclusion that the CS model is a better fit does not change. 308

Parameters were estimated with high precision and little bias under both models: the median and means of the distribution of parameters estimated under the CS and IW models for data sets simulated under those models closely matched the simulated values (Figs. 4 and 5). When fitting the CS model to IW simulations (Fig. S3) we observe that the K is estimated to be much higher than in the CS simulations (Fig. 4). This is because the IW simulations show more variability in clade sizes that can 14

only be accommodated by the CS model by assuming a larger clade-level K. When fitting the IW model to CS simulations (Fig. S4), we do not observe such a discrepancy. Indeed, in this case the total number of species matters rather than the number of species per clade. All parameter estimates and corresponding loglikelihoods are available in Table S3.



Figure 4. Bootstrap precision estimates of the parameters of the CS model. In a parametric bootstrap analysis the CS model was fitted to 1000 data sets simulated with the maximum likelihood parameters of the CS model for the empirical data. The panels show density histograms of the estimated parameters. The black lines indicate the median estimated values across all simulations and the blue arrows point to the values used in the simulations.



Figure 5. Bootstrap precision estimates of the parameters of the IW model. In a parametric bootstrap analysis the IW model was fitted to 1000 data sets simulated with the maximum likelihood parameters of the IW model for the empirical data. The panels show density histograms of the estimated parameters. The black lines indicate the median estimated values across all simulations and the blue arrows point to the values used in the simulations.

The simulated data can be used to check the reliability of model selection (because we know the

generating process). When performing model selection by simply selecting the model with the highest 319 likelihood, the IW model was incorrectly preferred over CS in 7.7% of data sets simulated under CS. The 320 CS model was incorrectly preferred over IW in 19% of data sets simulated under IW. When imposing at 321 least two log-units of difference before selecting a model, these numbers become 0.9% and 1.6%, 322 respectively. The CS and IW model were then correctly selected in 77% and 40.6% of the corresponding 323 data sets respectively, leaving 22.1% and 57.8% undecided between the two models. Because using 324 highest likelihood or higher by at least two log-units is quite arbitrary, and still leads to either high type 325 I error (highest likelihood) or low power (two log-units difference), we used the bootstrap likelihood ratio 326 (or loglikelihood difference) distribution to set the permissible type I error to 5% (two left-most arrows in 327 Fig. 6). This distribution of differences in loglikelihood between the CS and the IW model revealed that 328 it was highly unlikely (p < 0.001) that the empirical loglikelihood difference (6.43) would have been 329 found if the underlying model was IW, because the loglikelihood difference found in the empirical data 330 (black arrow in Fig. 6) falls clearly beyond the tail of the distribution of loglikelihood differences 331 obtained from data simulated under IW (higher than the largest likelihood), but falls right in the middle 332 of the distribution of differences for data simulated under CS (at the 49.6th percentile). This all suggests 333 that the CS model is strongly supported as the best model for the empirical data. 334

The power to select the generating model is relatively high. The power to detect CS is 85% (part of distribution generated under CS model that is larger than the middle arrow in Fig. 6) whereas the power to detect IW is 72% (part of distribution under IW model that is smaller than the left-most arrow in Fig. 6). If the empirical data had had a loglikelihood ratio between -0.29 (the leftmost arrow in Fig. 6) and 1.11 (middle arrow in Fig. 6), model selection would have been indecisive.

DISCUSSION

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We have developed a method to determine, using phylogenetic data on island colonization and branching 341 times, whether diversity-dependence in rates of colonization and speciation is limited to species within a 342 clade, or extends to species from different clades, or whether the information in the data is too limited to 343 make a clear call. In Hispaniolan *Eleutherodactylus* frogs we find that models including 344 diversity-dependence outperform models without a negative feedback of diversity on colonization and 345 speciation rates, suggesting that diversity limits play an important role. Diversity limits operating at the 346 clade-specific level (i.e. species from different colonizing clades do not interact) predominate over limits 347 at the island-wide level (i.e. species from different clades reduce each other's rate of colonization and speciation), because the model with clade-specific diversity-dependence clearly outperformed the model 349



Figure 6. Likelihood ratio bootstrap test. Distribution of differences between the loglikelihood of the CS model and the loglikelihood of the IW model when fitting both models to data sets simulated under CS and IW. The rightmost (black) arrow shows the difference in the empirical data, while the leftmost and middle arrows indicate the 5th percentile and the 95th percentile of the distributions generated under the CS model and the IW model respectively. The black arrow falls well inside the distribution for data simulated under CS, and to the right of the 95th percentile (in fact even the maximum!) of the distribution for data generated under IW. Therefore, the CS model is strongly preferred.

³⁵⁰ with island-wide diversity-dependence.

Although *Eleutherodactylus* frogs show repeated patterns of evolution into the same set of 351 ecotypes (Dugo-Cota et al., 2019), these results suggest that these ecotypes do not interfere with each 352 other across clades. One could argue that Fig. 2 already tells us this because the overlap in ecotypes and 353 in ranges between the clades on Hispaniola is limited, and hence species do not seem to interact across 354 clades. However, one can also explain this pattern as a *consequence* of interaction across clades, because 355 under IW earlier clades block later ones from radiating into the same habitats (both ecologically and 356 spatially). Our results do not support this explanation and lead us to conjecture that there has been 357 sufficient (niche) space that IW diversity-dependence does not occur. Of course, this may change in time: 358 if we wait millions of years, (niche) space may eventually become saturated, but currently there is no 350 signal of IW diversity-dependence. In summary, present-day spatial distributions and ecological 360 distributions into ecotypes cannot be taken as evidence that species from different clades do not interact, as these patterns may be a consequence of such interactions in the past. The approach we have taken in this paper is to infer such diversity-dependence from the phylogenetic branching pattern. We have shown 363 that if IW diversity-dependence operates, we would often pick up its signal from the phylogenetic data. 364 In our *Eleutherodactylus* frog example, we did not, as there is only 1% chance that the pattern we 365 observed would be generated by an IW model (i.e. only 1 in 100 simulations of an IW model we would 366 obtain a loglikelihood ratio between CS and IW models which is equal or higher than observed for the empirical data).

Our simulations were limited to the parameters estimated from the *Eleutherodactylus* frog data. To assess the more general ability of our approach to identify CS and IW when they are operating would 370 require analyzing many more simulated data sets for a wide range of parameters sets. This is currently 371 computationally unfeasible, because the likelihood maximizations, although performed with highly 372 optimized code, take quite a bit of time (at least a few hours per data set), which bars extensive 373 simulation studies across a sizeable number of replicates. Instead we suggest that researchers wishing to 374 compare CS and IW models for their study system should fit these models to their data and take the 375 estimated parameters to run simulations, just like we did here. This allows one to establish whether CS 376 and IW models can be distinguished by plotting figures such as Fig. 6. We have shown that it is 377 important to do so, because model selection based solely on AIC may be biased (Etienne et al., 2016). 378

The CS model assumes the same carrying capacity K for each clade, which is a constraint to each clade's size, and hence our model selection may be somewhat biased towards IW, which only limits the overall number of species by its K. Because the CS model nevertheless outperforms the IW model, this is not an issue for this study, and it may be indicative of a similar K among clades, which is in line with ecotype space limiting the number of species equally in each clade. Still, models with different K values for each clade could in principle be fitted to the data to confirm this. In practice, however, this is not really feasible, because we are already estimating four or five parameters, and there may not be enough information in a data set of this size to allow for more parameters to be reliably estimated.

We have only considered two models of diversity-dependence: one where diversity-dependence only applies to species within the same clade, and one where it also applies to species of other 388 *Eleutherodactylus* clades establishing on an island. Various other models can be conceived. First, 389 diversity-dependence might apply to Anurans species beyond Eleutherodactulus or include other 390 amphibians or even non-amphibians. We have chosen the level of textitEleutherodactylus species as it 391 seems, arguably, the largest group where the assumptions of equal rates of colonization, speciation and 392 extinction are not too strongly violated. Our results indicate that diversity-dependence does not extend 393 to this scale. Second, diversity-dependence could also occur at a higher taxonomic level, i.e. the number 394 of clades, rather than the number of species within them may be limiting further colonization or 395 diversification. Third, the effect of phylogenetic relatedness may also differ for speciation, extinction and 396 colonization. For instance, Pires et al. (2017) found that speciation is mostly affected by within-clade 397 diversity-dependence, whereas extinction is mostly affected by between-clade diversity-dependence. Fourth, one could define phylogenetic limits in terms of actual phylogenetic distances so that we move from a within- and between-clade dichotomy to a more continuous spectrum where some phylogenetically 400

related clades may interact, but more distantly related clades do not. There are no likelihood methods
for such models yet. One may have to resort to simulation-based approaches such as Approximate
Bayesian Computation (Janzen et al., 2015). These methods need to integrate over all possible
trajectories of the clades through time which is not trivial, because the space of these trajectories is
extremely high-dimensional.

We assumed no diversity-dependence in extinction and anagenesis to focus on the effect on colonization and cladogenesis, and for model and computational simplicity. However, 407 diversity-dependence in extinction and anagenesis is conceivable. For extinction one may assume higher 408 extinction rates for higher diversity. We note that this causes a stronger pull-of-the-present in 409 lineages-through-time plots contrary to what is commonly observed (Phillimore and Price, 2008; Etienne 410 et al., 2012). However, such a pull-of-the-present may not be visible in empirical data, because we fail to 411 account for incipient species (Etienne and Rosindell, 2012). Likelihood methods that incorporate both 412 diversity-dependence and protracted speciation do not yet exist, however. Diversity-dependence in 413 anagenesis is also conceivable, but it may be both negative and positive. High diversity can inhibit 414 anagenesis bt limiting the ecological space to evolve into. However, higher diversity might also mean that 415 there is more selective pressure for a species to evolve away from the mainland sister (sub)species. 416 Anagenesis can also occur through drift alone simply due to long-term isolation from the mainland, in 417 which case a diversity effect on anagenesis seems unlikely. One might also argue that if local adaptation 418 is the primary cause of anagenesis, competition with other species (and thus diversity-dependence in 419 anagenesis) is unlikely to prevent anagenesis. In such a scenario the species will probably not establish at 420 all (some adaptation seems necessary to survive in a new environment) which would be accounted for by 421 diversity-dependence in the colonization rate. 422

One may wonder what it is in the branching pattern that allows for selecting one model over the 423 other. An intuitively obvious candidate may be the rank of the largest clade. The IW model can be 424 expected to have the first clade as the largest, because later clades will be suffering from 425 diversity-dependence and hence not be able to grow very large. However, we noticed that the first clade 426 is also almost equally often the largest clade under the CS model (Fig. 3). Hence, a pattern we may put 427 down to incumbency and interclade competition (Silvertown, 2004; Schenk et al., 2013) arises equally 428 prominently under a model without interclade competition. Apparently, in our empirical example time 429 since colonization is a more important determinant of the size of a clade than diversity-dependence. The 430 IW or CS nature of the colonization and diversification process, and thus the presence or absence of 431 priority effects at the macroevolutionary scale, is hidden in a more complex way in the phylogenetic 432

⁴³³ branching pattern that is not easily picked up by simple summary statistics, but is detected by our ⁴³⁴ likelihood ratio test. We do note that the estimate of the island-wide carrying capacity K (132) is quite a ⁴³⁵ bit larger than the number of species present on the island (66), suggesting that the island is still far ⁴³⁶ from saturation under the IW model. Other systems may have a lower K and the effect of priority effects ⁴³⁷ may be relatively stronger. It is an interesting avenue to study whether there is a relationship between ⁴³⁸ the magnitude of these priority effects and invasibility of islands, which may contribute to our ⁴³⁹ understanding of biological invasions (Fraser et al., 2015).

The new IW model may also be applicable in other fields, such as epidemiology where it may 440 serve as a tool in understanding spread of an infectious disease, e.g. a virus, in island-like systems such as 441 schools or hospitals. In such systems there may be multiple sources of infections that spread through the 442 local population and can be modelled as colonizations. The carrying capacity is the number of children 443 or patients. The IW model would be the appropriate model if once the host is infected, it builds up 444 immunity against all strains, thus hindering further colonization and diversification. This scenario is 445 most likely if the colonizing strains are phylogenetically related. The CS model would be a better 446 description if a host can be infected by multiple strains, but within each strain there is viral interference 447 (see e.g. Ojosnegros et al. (2010)). This scenario is most likely if the colonizing strains are 448 phylogenetically (and hence functionally) distinct. 449

The CS model implementation allows incomplete phylogenetic information: if the island species 450 are recognized (including their endemicity status), but their colonization or branching times are not 451 known, the model integrates over the possible colonization and branching times. The same principle can 452 be applied to the IW model, but this is currently computationally unfeasible. Sampling colonization and 453 branching times to obtain a complete data set and then repeating the analysis for each sampled complete 454 phylogeny, as we have done, is probably the most straightforward way to get an idea of the impact of 455 phylogenetic uncertainty. Incomplete knowledge due to failure to recognize incipient or cryptic species is 456 a more fundamental problem that the field has not been able to address completely satisfactorily. There 457 are models that can account for this, e.g. the protracted speciation model (Etienne and Rosindell, 2012; 458 Lambert et al., 2014; Etienne et al., 2014), but the mathematical approach to compute the likelihood 459 under this model seems incompatible with the approach used for diversity-dependence models (Etienne 460 et al., 2012; Laudanno et al., 2019). Simulation-based approaches may be the only (long) way to a 461 resolution (Richter et al., 2020).

We have provided a new model for island biogeography with diversity-dependent feedback on colonization and diversification occurring between all island species. Our implementation of this IW

REFERENCES

model has some computational limitations for islands with large numbers of colonizations, particularly if these are non-endemic, but typical insular data sets with a moderate number of colonizations or a high 466 level of endemism (such as our *Eleventherodactylus* frogs) are perfectly feasible. Our single empirical 467 example serves as an illustration to the empiricist how to explore the phylogenetic limits of diversity 468 limits to diversification from phylogenetic data alone despite the limitations of phylogenetic data (Losos, 460 2011). Although this example showed a clearly better fit of the CS model, future applications may reveal 470 different and more nuanced impacts of clade competition on diversification at a wider range of 471 phylogenetic scales. Further extension of our approach to allow integrating ecological data with 472 phylogenetic data (Harmon et al., 2019), for instance to examine simultaneous diversity-dependence and 473 trait-dependence of diversification, is an exciting but challenging next direction. 474

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References

- Boettiger, C., G. Coop, and P. Ralph. 2012. Is your phylogeny informative? measuring the power of
 comparative methods. Evolution 66:2240–2251.
- Cadotte, M. W., T. J. Davies, and P. Peres-Neto. 2017. Why phylogenies do not always predict ecological
 differences. Ecological Monographs 87:535–551.

⁴⁹² Cahill, J. F., S. W. Kembel, E. G. Lamb, and P. A. Keddy. 2008. Does phylogenetic relatedness influence

⁴⁹³ the strength of competition among vascular plants? Perspectives in Plant Ecology, Evolution and

- 494 Systematics 10:41-50.
- ⁴⁹⁵ Darwin, C. 1859. On the origin of species by means of natural selection, or preservation of favoured races
 ⁴⁹⁶ in the struggle for life. John Murray, London.
- ⁴⁹⁷ Dugo-Cota, A., C. Vilá, A. Rodríguez, and A. Gonzalez-Voyer. 2019. Ecomorphological convergence in ⁴⁹⁸ *Eleutherodactylus* frogs: a case of replicate radiations in the Caribbean. Ecology Letters 22:884–893.
- Etienne, R. S., B. Haegeman, T. Stadler, T. Aze, P. N. Pearson, A. Purvis, and A. B. Phillimore. 2012.
 Diversity-dependence brings molecular phylogenies closer to agreement with the fossil record.
- ⁵⁰¹ Proceedings of the Royal Society of London B 279:1300–1309.
- Etienne, R. S., H. Morlon, and A. Lambert. 2014. Estimating the duration of speciation from
 phylogenies. Evolution 68:2430–2440.
- Etienne, R. S., A. L. Pigot, and A. B. Phillimore. 2016. How reliably can we infer diversity-dependent
 diversification from phylogenies? Methods in Ecology and Evolution 7:1092–1099.
- Etienne, R. S. and J. Rosindell. 2012. Prolonging the past counteracts the pull of the present: protracted
 speciation can explain observed slowdowns in diversification. Systematic Biology 61:204–213.
- Foote, M., R. A. Cooper, J. S. Crampton, and P. M. Sadler. 2018. Diversity-dependent evolutionary rates
 in early Palaeozoic zooplankton. Proceedings of the Royal Society of London B: Biological sciences
 285:20180122.
- ⁵¹¹ Foote, M. and A. I. Miller. 2006. Principles of psaleontology. W.H. Freeman, New York.
- Fraser, C. I., S. C. Banks, and J. M. Waters. 2015. Priority effects can lead to underestimation of
 dispersal and invasion potential. Biological Invasions 17:1–8.
- Gerhold, P., J. F. C. Jr, M. Winter, I. V. Bartish, and A. Prinzing. 2015. Phylogenetic patterns are not proxies of community assembly mechanisms (they are far better). Functional Ecology 29:600–614.
- ⁵¹⁶ Germain, R. M., J. T. Weir, and G. Benjamin. 2016. Species coexistence: macroevolutionary
 ⁵¹⁷ relationships and the contingency of historical interactions. Proceedings of the Royal Society B:
 ⁵¹⁸ Biological Sciences 283:20160047.
- ⁵¹⁹ Harmon, L. J., C. S. Andreazzi, F. Dbarre, J. Drury, E. E. Goldberg, A. B. Martins, C. J. Melin,
- A. Narwani, S. L. Nuismer, M. W. Pennell, S. M. Rudman, O. Seehausen, D. Silvestro, M. Weber, and
- B. Matthews. 2019. Detecting the macroevolutionary signal of species interactions. Journal of
- 522 Evolutionary Biology 32:769–782.

- Heinicke, M. P., W. E. Duellman, and S. B. Hedges. 2007. Major Caribbean and Central American frog
 faunas originated by ancient oceanic dispersal. Proceedings of the National Academy of Sciences
 104:10092–10097.
- ⁵²⁶ HilleRisLambers, J., P. B. Adler, W. S. Harpole, J. M. Levine, and M. M. Mayfield. 2012. Rethinking
 ⁵²⁷ community assembly through the lens of coexistence theory. Annual Review of Ecology, Evolution, and
 ⁵²⁸ Systematics 43:227–248.
- Iturralde-Vinent, M. A. 2006. Meso-cenozoic Caribbean paleogeography: Implications for the historical
 biogeography of the region. International Geology Review 48:791–827.
- Jablonski, D. 2008. Biotic interactions and macroevolution: Extensions and mismatches across scales and levels. Evolution 62:715–739.
- Janzen, T., S. Höhna, and R. S. Etienne. 2015. Approximate bayesian computation of diversification rates from molecular phylogenies: introducing a new efficient summary statistic, the nltt. Methods in Ecology and Evolution 6:566–575.
- Jønsson, K. A., P.-H. Fabre, S. A. Fritz, R. S. Etienne, R. E. Ricklefs, T. B. Jørgensen, J. Fjeldså,
 C. Rahbek, P. G. P. Ericson, F. Woog, E. Pasquet, and M. Irestedt. 2012. Ecological and evolutionary
 determinants for the adaptive radiation of the Madagascan vangas. Proceedings of the National
 Academy of Sciences 109:6620–6625.
- Lambert, A., H. Morlon, and R. S. Etienne. 2014. The reconstructed tree in the lineage-based model of protracted speciation. Journal of Mathematical Biology. In press. doi: 10.1007/s00285-014-0767-x.
- Laudanno, G., B. Haegeman, and R. S. Etienne. 2019. Additional analytical support for a new method to compute the likelihood of diversification models. bioRxiv.
- Losos, J. B. 2011. Seeing the forest for the trees: The limitations of phylogenies in comparative biology. The American Naturalist 177:709–727.
- MacArhur, R. H. and E. O. Wilson. 1967. Island Biogeography. Princeton University Press, Princeton,
 NJ.
- ⁵⁴⁸ Matzke, N. J. 2013. Probabilistic historical biogeography: new models for founder-event speciation,
- ⁵⁴⁹ imperfect detection, and fossils allow improved accuracy and model-testing. Frontiers of Biogeography
- ⁵⁵⁰ 5:242–248.

- Matzke, N. J. 2014. Model Selection in Historical Biogeography Reveals that Founder-Event Speciation
 Is a Crucial Process in Island Clades. Systematic Biology 63:951–970.
- ⁵⁵³ Mayfield, M. M. and J. M. Levine. 2010. Opposing effects of competitive exclusion on the phylogenetic ⁵⁵⁴ structure of communities. Ecology Letters 13:1085–1093.
- ⁵⁵⁵ Narwani, A., B. Matthews, J. Fox, and P. Venail. 2015. Using phylogenetics in community assembly and
 ⁵⁵⁶ ecosystem functioning research. Functional Ecology 29:589–591.
- ⁵⁵⁷ Ojosnegros, S., N. Beerenwinkel, T. Antal, M. A. Nowak, C. Escarmís, and E. Domingo. 2010.
 ⁵⁵⁸ Competition-colonization dynamics in an rna virus. Proceedings of the National Academy of Sciences
 ⁵⁵⁹ 107:2108–2112.
- Phillimore, A. B. and T. D. Price. 2008. Density-dependent cladogenesis in birds. PLoS Biology
 6:0483–0489.
- ⁵⁶² Pigot, A. L. and R. S. Etienne. 2015. A new dynamic null model for phylogenetic community structure.
 ⁵⁶³ Ecology Letters 18:153–163.
- ⁵⁶⁴ Pires, M. M., D. Silvestro, and T. B. Quental. 2017. Interactions within and between clades shaped the
 ⁵⁶⁵ diversification of terrestrial carnivores. Evolution 71:1855–1864.
- ⁵⁶⁶ Proches, S., J. R. U. Wilson, D. M. Richardson, and M. Rejmnek. 2008. Searching for phylogenetic
 ⁵⁶⁷ pattern in biological invasions. Global Ecology and Biogeography 17:5–10.
- Rabosky, D. L. 2013. Diversity-dependence, ecological speciation, and the role of competition in
 macroevolution. Annual Review of Ecology, Evolution, and Systematics 44:481–502.
- Rabosky, D. L. and R. E. Glor. 2010. Equilibrium speciation dynamics in a model adaptive radiation of
 island lizards. Proceedings of the National Academy of Sciences 107:22178–22183.
- Rabosky, D. L. and A. H. Hurlbert. 2015. Species richness at continental scales is dominated by
 ecological limits. The American Naturalist 185:572–583.
- Rabosky, D. L. and I. J. Lovette. 2008. Density-dependent diversification in North American wood
 warblers. Proceedings of the Royal Society of London B 275:2363–2371.
- Raup, D. M., S. J. Gould, T. J. M. Schopf, and D. S. Simberloff. 1973. Stochastic models of phylogeny
 and the evolution of diversity. Journal of Geology 81:525–542.

- Revell, L. J. 2012. phytools: an R package for phylogenetic comparative biology (and other things).
 Methods in Ecology and Evolution 3:217–223.
- Richter, F., B. Haegeman, R. S. Etienne, and E. C. Wit. 2020. Introducing a general class of species
 diversification models for phylogenetic trees. Statistica Neerlandica 74:261–274.
- Schenk, J. J., K. C. Rowe, and S. J. Steppan. 2013. Ecological Opportunity and Incumbency in the
 Diversification of Repeated Continental Colonizations by Muroid Rodents. Systematic Biology
 62:837–864.
- Sepkoski, J. 1996. Competition in macroevolution: the double wedge revisited. Pages 211–255 in
 Evolutionary Paleobiology (J. L. D. Jablosinki, D.H. Erwin, ed.). University of Chicago Press.
- Silvertown, J. 2004. The ghost of competition past in the phylogeny of island endemic plants. Journal of
 Ecology 92:168–173.
- Silvestro, D., A. Antonelli, N. Salamin, and T. B. Quental. 2015. The role of clade competition in the
 diversification of North American canids. Proceedings of the National Academy of Sciences
 112:8684–8689.
- ⁵⁹² Stanley, S. M. 1973. An explanation for cope's rule. Evolution 27:1–26.
- Valente, L., R. S. Etienne, and L. M. Dávalos. 2017a. Recent extinctions disturb path to equilibrium
 diversity in Caribbean bats. Nature Ecology & Evolution 1:0026.
- Valente, L., R. S. Etienne, and J. C. Garcia-R. 2019. Deep macroevolutionary impact of humans on New
 Zealand's unique avifauna. Current Biology 29:2563–2569.e4.
- ⁵⁹⁷ Valente, L., J. C. Illera, K. Havenstein, T. Pallien, R. S. Etienne, and R. Tiedemann. 2017b. Equilibrium
 ⁵⁹⁸ bird species diversity in Atlantic islands. Current Biology 27:1660–1666.e5.
- ⁵⁹⁹ Valente, L. M., R. S. Etienne, and A. B. Phillimore. 2014. The effects of island ontogeny on species
 diversity and phylogeny. Proceedings of the Royal Society B: Biological Sciences 281:20133227.
- Valente, L. M., A. B. Phillimore, and R. S. Etienne. 2015. Equilibrium and non-equilibrium dynamics
 simultaneously operate in the Galápagos islands. Ecology Letters 18:844–852.
- Valkenburgh, B. V. 1999. Major patterns in the history of carnivorous mammals. Annual Review of
 Earth and Planetary Sciences 27:463–493.

24

- ⁶⁰⁵ Violle, C., D. R. Nemergut, Z. Pu, and L. Jiang. 2011. Phylogenetic limiting similarity and competitive
 ⁶⁰⁶ exclusion. Ecology Letters 14:782–787.
- ⁶⁰⁷ Walker, T. D. and J. W. Valentine. 1984. Equilibrium models of evolutionary species diversity and the ⁶⁰⁸ number of empty niches. American Naturalist 124:887–899.
- ⁶⁰⁹ Whittaker, R. J., K. A. Triantis, and R. J. Ladle. 2008. A general dynamic theory of oceanic island ⁶¹⁰ biogeography. Journal of Biogeography 35:977–994.
- ⁶¹¹ Wilcox, T. M., M. K. Schwartz, and W. H. Lowe. 2018. Evolutionary community ecology: Time to think
- outside the (taxonomic) box. Trends in Ecology & Evolution 33:240 250.