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# *In situ* Bioconjugation of a Maleimide-Functionalized Ruthenium-based Photosensitizer to Albumin for Photodynamic Therapy

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### **Abstract**

Maleimide-containing prodrugs can quickly and selectively react with circulating serum albumin following their injection in the blood stream. The drug-albumin complex then benefits from longer blood circulation times and better tumor accumulation. Herein, we have applied this strategy to a previously reported highly phototoxic Ru polypyridyl complex-based photosensitizer to increase its accumulation at the tumor, reduce off-target cytotoxicity and therefore improve its pharmacological profile. Specifically, two complexes were synthesized bearing a maleimide group: one complex with the maleimide directly incorporated onto the bipyridyl ligand, the other has a hydrophilic linker between the ligand and the maleimide group. Their interaction with albumin was studied in-depth, revealing their ability to efficiently bind both covalently and non-covalently to the plasma protein. A crucial finding is that the maleimide-functionalized complexes exhibited significantly lower cytotoxicity in non-cancerous cells under dark conditions compared to the non-functionalized complex, which is a highly desirable property for a photosensitizer. The binding to albumin also led to a decrease in the phototoxicity of the Ru bioconjugates in comparison to the non-functionalized complex, probably due to a decreased cellular uptake. Unfortunately, this decrease in phototoxicity was not compensated by a dramatic increase in tumor accumulation, as was

demonstrated in a tumor-bearing mouse model using ICP-MS studies. Consequently, this study provides valuable insight into the future design of *in situ* albumin-binding complexes for photodynamic therapy, in order to maximize their effectiveness and realize their full potential.

## Introduction

In the past few years, photodynamic therapy (PDT) has gained a growing interest for the fight against cancer. This approved medical technique involves the use of a photosensitizer (PS) and light in order to induce cellular damages in tumors in a spatially and temporally controlled manner.<sup>1-4</sup>

Typical PDT procedures start with the local or systemic administration of the PS, followed after a certain time by local light irradiation of the targeted tissues. Light-mediated excitation of the PS leads to the generation of reactive oxygen species (ROS) and/or singlet oxygen (<sup>1</sup>O<sub>2</sub>), following a type I or type II mechanism, respectively, which are responsible for local cellular damages.<sup>5</sup>

Most clinically applied PSs are based on tetrapyrrolic scaffolds. First-generation PSs (*i.e.*, Photofrin® and hematoporphyrin derivatives) are a mixture of several chemical entities. They have often been associated with slow elimination rates, leading to extended photosensitivity in patients, low aqueous solubility, photodegradation, tedious synthetic procedures and lack of selectivity towards diseased tissues. Some of these drawbacks have been addressed to yield second-generation PSs (*e.g.*, Photosens®<sup>®</sup>, Foscan®<sup>®</sup>, Laserphyrin®<sup>®</sup>, Redaporphin®<sup>®</sup>, Photogem®<sup>®</sup>, Radachlorin®<sup>®</sup>, Puryltin®<sup>®</sup>, Lutrin®<sup>®</sup>, TOOKAD®<sup>®</sup> soluble) with better-defined structures, improved aqueous solubility, longer absorption wavelengths and reduced skin sensitization. However, these second-generation porphyrin-based compounds are still far from the ideal of a perfect PS, since only few combine all of the aforementioned advantages.<sup>2,4,6-8</sup>

Among non-porphyrin PSs, the use of ruthenium(II) (Ru(II)) polypyridyl complexes has recently received much interest thanks to their simple synthesis, chemical stability, good <sup>1</sup>O<sub>2</sub> production yields and generally good aqueous solubility.<sup>9-18</sup> Notably, TLD-1433 is currently involved in a phase II clinical trial on patients with bladder cancer.<sup>19,20</sup> However, these Ru(II)-based PSs are known to poorly absorb light in the biological spectral window (600-900 nm).<sup>8,21</sup> This feature prevents the treatment of tumors located deeply in the tissues or of large tumors.<sup>22</sup>

To address this drawback, we recently reported the DFT-guided design and *in vitro* study of new Ru(II) polypyridyl complexes absorbing at longer wavelengths.<sup>23</sup> One of these compounds, namely **[Ru(bphen)<sub>2</sub>dmbipy][PF<sub>6</sub>]<sub>2</sub>** (bphen = 4,7-diphenyl-1,10-phenantroline, dmbipy = 4,4'-dimethyl-2,2'-bipyridine, Figure 1), which has previously been described by others for its cytotoxic properties<sup>24</sup>, showed a great PDT potential on various cancer cell lines in a 2D cell monolayer model as well as in a 3D HeLa multicellular tumor spheroid (MCTS)

model. In contrast to the majority of published Ru(II) polypyridyl complexes, this compound was able to exert a phototoxic effect upon irradiation at up to 595 nm, which lies at the frontier of the biological spectral window. In addition, **[Ru(bphen)<sub>2</sub>dmbipy][PF<sub>6</sub>]<sub>2</sub>** displays little photobleaching and a good stability in human plasma.

PDT features a spatial and temporal control of the therapeutic effect, which intrinsically provides PDT with a first level of selectivity towards diseased tissues. However to improve its therapeutic potential, PDT could benefit from a second level of selectivity by directly targeting the tumor.<sup>25</sup> Over the years, many strategies have been developed to increase chemotherapeutics selectivity towards cancer cells either through active or passive targeting.<sup>26–29</sup> Some active targeting strategies have already been applied to Ru(II)-based PSs, including conjugation of the complex to aptamers,<sup>30</sup> antibodies and nanobodies,<sup>29,31,32</sup> oligonucleotides,<sup>33,34</sup> bombesin,<sup>35–37</sup> biotin,<sup>33,38–41</sup> folic acid,<sup>42</sup> somatostatin,<sup>43</sup> glucose,<sup>44</sup> mannose,<sup>45</sup> tamoxifen,<sup>46</sup> cobalamin,<sup>47</sup> taurine<sup>48</sup> or transferrin.<sup>34,49,50</sup> Passive targeting strategies exploiting the enhanced permeation and retention (EPR) effect have also been explored through encapsulation or conjugation methods using polymers,<sup>38,51–58</sup> liposomes,<sup>59,60</sup> carbon nanotubes and graphene oxide,<sup>61–63</sup> inorganic materials,<sup>34,42,64–72</sup> or proteins,<sup>71,73,74</sup>. It is worth noting that a substantial proportion of these studies combine both active and passive targeting to further enhance PSs tumor selectivity.

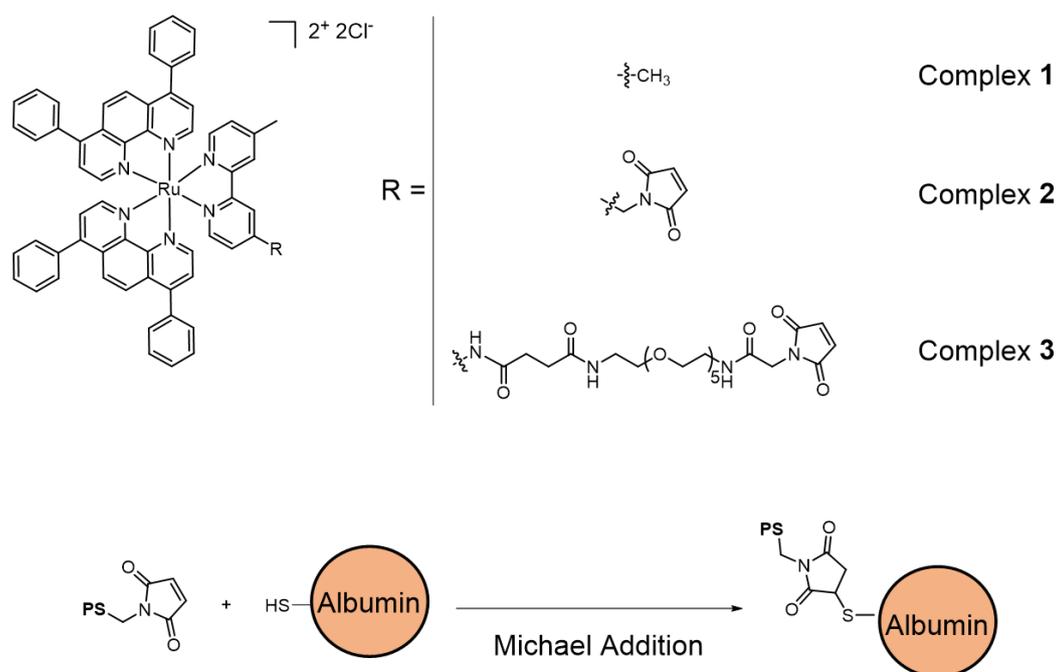
Among these targeting approaches, the conjugation or binding of the PS to endogenous blood plasma proteins, and more particularly albumin, seems especially attractive to increase its blood residence time and promote its accumulation specifically within tumors.<sup>75,76</sup> Albumin is the most abundant plasma protein with a concentration of 35 – 50 g/L of human plasma.<sup>77</sup> This protein has been described to accumulate in tumors through the EPR effect<sup>78–80</sup> but also through a gp60 and caveolin-1-mediated transcytosis, promoting its extravasation from the blood circulation to the diseased tissues.<sup>81,82</sup> Several attempts to conjugate chemotherapeutic drugs to albumin in order to increase their tumor accumulation resulted in the clinical success of Abraxane®, a now marketed paclitaxel formulation with enhanced efficacy and safety compared to the classical formulation.<sup>83–85</sup> While albumin was predominantly investigated as a therapeutic carrier for purely organic drugs, there is a growing interest in exploring its potential for anticancer metal complexes.<sup>86–90</sup>

In particular, this strategy has already been successfully employed in the case of Ru(II)-based PSs. Shi *et al.* reported the elaboration of a nanocarrier based on lanthanide-doped upconversion nanoparticles coated with human serum albumin (HSA) and a Ru(II)

polypyridyl complex for both imaging and PDT purposes. This nanocarrier showed a good cellular uptake as well as an enhanced *in vitro* phototoxicity on HeLa and HepG2 cell lines compared to the Ru(II) complex alone.<sup>71</sup> In an alternative approach, Chakraborty *et al.* reported the design and *in vitro* evaluation of an engineered HSA molecule functionalized with mitochondria-targeting groups, solubilizing polyethylene glycol chains and Ru(II) polypyridyl complexes. This macromolecule demonstrated an impressive phototoxic effect in HeLa cells which was attributed to an enhanced <sup>1</sup>O<sub>2</sub> production yield and a better photostability of the complex associated to a favorable mitochondrial localization.<sup>73</sup> Unfortunately, none of these studies investigated the *in vivo* potential of their systems. Moreover, these studies involved the *ex vivo* conjugation of human serum albumin (HSA) to the PS, which requires the use of commercially available HSA. An alternative strategy involves the design of a prodrug incorporating a maleimide (Mal) moiety. Upon intravenous (IV) injection of the prodrug, the Mal group reacts quickly and selectively with the circulating albumin's cysteine-34 (Cys-34) sulfhydryl group, leading to the *in situ* formation of a drug-albumin conjugate. This method allows therefore the development of a simple, well characterized low-molecular weight drug, able to benefit from the advantages of albumin-bound drugs.<sup>91,92</sup>

To the best of our knowledge, while this strategy has already been successfully applied to chemotherapeutic complexes of platinum, Ru and osmium, it has never been applied to vectorize a metal-based PS for PDT applications.<sup>93-100</sup> We believe Ru(II)-based PSs, and particularly **[Ru(bphen)<sub>2</sub>dmbipy][PF<sub>6</sub>]<sub>2</sub>** or its analog with chlorides as counter ions **[Ru(bphen)<sub>2</sub>dmbipy][Cl]<sub>2</sub>** (**1**), could greatly benefit from *in situ* albumin conjugation to improve its tumor accumulation. Therefore, in this article, we describe the synthesis, *in vitro* and *in vivo* characterization of the first Ru(II) polypyridyl complexes containing a Mal function for *in situ* albumin conjugation. The Mal function was either directly added onto the complex, or spaced using an hydrophilic polyethylene glycol (PEG) linker to further improve its aqueous solubility and blood circulation time yielding respectively **[Ru(bphen)<sub>2</sub>dmbipyMal][Cl]<sub>2</sub>** (**2**) (dmbipyMal = 4-maleimidomethyl-4'-methyl-2,2'-bipyridine) and **[Ru(bphen)<sub>2</sub>dmbipyPEGMal][Cl]<sub>2</sub>** (**3**) (PEGMal = N-(1-(2,5-dioxo-2,5-dihydro-1H-pyrrol-1-yl)-2-oxo-6,9,12,15,18-pentaoxa-3-azaicosan-20-yl)-4-oxopentanamidyl). Of note, we have already described complex **2** in another study, albeit it was used only as an intermediate to the synthesis of a peptide-conjugated PDT PS.<sup>37</sup> We show

that all three complexes can bind non-covalently to albumin, and only **2** and **3** exhibit the capability of covalently binding to the protein. The three complexes were then characterized *in vitro* and in a preliminary *in vivo* experiment in tumor-bearing mice, to evaluate the potential of this targeting strategy.



**Figure 1.** Chemical structures of the Ru(II) complexes studied in this work and putative scheme of reaction of a maleimide functionalized PS with the thiol group of the Cys-34 of albumin.

## Results and discussion

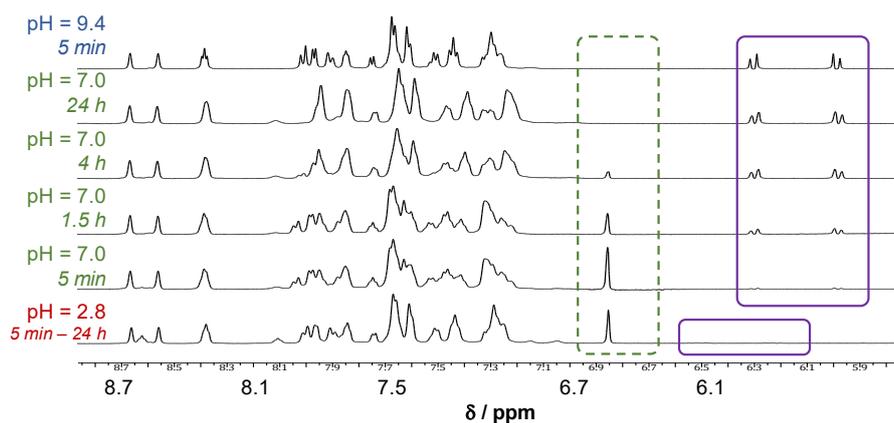
The ligand 4-aminomethyl-4'-methyl-2,2'-bipyridine was synthesized as previously reported from 4-bromomethyl-4'-methyl-2,2'-bipyridine with hexamethylenetetramine.<sup>101</sup> The precursor [Ru(bphen)<sub>2</sub>]Cl<sub>2</sub> was prepared by reduction of Ru(III)Cl<sub>3</sub> and substitution with bphen in the presence of LiCl. Following this, the [Ru(bphen)<sub>2</sub>dmbipyNH<sub>2</sub>][PF<sub>6</sub>]<sub>2</sub> complex was obtained by a coordination reaction of the precursor with the above mentioned ligand. The desired [Ru(bphen)<sub>2</sub>dmbipyMal][PF<sub>6</sub>]<sub>2</sub> complex was then obtained using maleic anhydride condensation on the amine (Scheme S1). In parallel, [Ru(bphen)<sub>2</sub>dmbipyNH<sub>2</sub>][PF<sub>6</sub>]<sub>2</sub> was conjugated to a PEG<sub>5</sub> linker generated as previously described,<sup>102–105</sup> which was activated as a NHS-ester using standard conditions. The Mal moiety was finally added onto the linker using 2,5-dioxopyrrolidin-1-yl 2-(2,5-dioxo-2,5-

dihydro-1H-pyrrol-1-yl)acetate (Scheme S1). The compounds were then converted into their chloride salts using a counter ion exchange resin to yield complexes **2** and **3**. The identity of the compounds was confirmed by <sup>1</sup>H-, <sup>13</sup>C-NMR and HRMS (Figure S1-S10) and the purity confirmed by elemental analysis. The control complex [Ru(bphen)<sub>2</sub>dmbipy][PF<sub>6</sub>]<sub>2</sub> was obtained in a previous study and also converted to the dichloride salt using a counter ion exchange resin to give complex **1**. Complexes were used as racemic mixtures of the Δ and Λ isomers.

### Solubility and stability

Complexes **1** and **2** showed a good aqueous solubility (≥1 mM), which decreases significantly in buffered media like 15 mM phosphate or PBS at pH 7.40 or even in the presence of 0.1 M KCl. Moderate (*ca.* 7%) precipitation takes place on the basis of the observed decrease in the UV–Vis absorption spectra (see Figure S11A for **2**) even at complex concentrations of 20 μM after 24 h in PBS buffered samples. We have already observed this behavior with analogous complexes in previous studies. We hypothesized that a high anion concentration decreases the repulsive interaction between the positively charged complexes, leading to their aggregation.<sup>37,40</sup> Additionally, as we previously observed, the presence of HSA in buffered media could prevent the precipitation of complexes **1** and **2** for at least two days (see UV–Vis spectra in Figure S11B).<sup>37,40</sup> In contrast, complex **3**, which includes a hydrophilic PEG spacer, only suffers from a rather moderate decrease of its aqueous solubility in the presence of PBS components. A concentration of 100 μM of complex **3** was attainable in this medium. Overall, all studied complexes displayed an acceptable solubility at a 20 μM concentration in PBS over a 3 h time span (96% of **1** and **2**, and 100% of **3** remained in solution). At the same time, pure aqueous stock solutions of **1** and **2** were proven to be stable for at least 3 days based on the <sup>1</sup>H NMR measurements (spectra not shown). However, when the pH was set to 7.4, then hydrolysis of the maleimide group in complex **2** took place as signals corresponding to a maleamate group formed (Figure S12). Analogously, aqueous stock solutions of **3** were stable in slightly acidic solutions for at least 24 h according to the <sup>1</sup>H NMR spectra depicted in Figure 2. However, at pH 7.4 and 7.0, complex **3** behaved as complex **2** and maleamate formed in a somewhat slower process (Figure 2). A negligible amount of maleamate could be detected at neutral pH for the freshly prepared (5 min) sample, while the hydrolysis progressed to 37%, 62% and 100 % at time points 1.5 h, 4 h and 24 h, respectively. The

hydrolysis appeared even faster at a higher pH of 9.4, which suggests that the hydrolysis rate is pH dependent as it was proven for alkyl-maleimides.<sup>106</sup> Noteworthy, the hydrolysis of the present compounds is faster (*ca.* 52%/1h for **2** and 62%/4h for **3**) compared to that of ethylmaleimide reported at pH 7.16 and 30 °C ( $k_{\text{obs}} \approx 10^{-5}$  1/s,  $t_{1/2} \approx 19$  h).<sup>106,107</sup> Therefore, stock solutions of **3** used for *in vitro* experiments were prepared in slightly acidic solution and used within 12 h. Additionally, stability studies made on complex **2** in Eagle's minimal essential (EMEM) cell culture medium at pH 7.4 showed interaction of **2** with EMEM components within 30 min and hydrolysis to maleamate was initially suppressed (see Figure S13). This results however should be treated with caution because some precipitate also formed in the NMR tube. Overall, complexes **2** and **3** appear to slowly hydrolyze in aqueous medium in physiologic conditions. However, as we expect the Mal group to react quickly and selectively with the albumin contained both in complete culture medium and in blood, this instability might not dramatically affect the performances of the complexes.



**Figure 2.** <sup>1</sup>H NMR spectra recorded for complex **3** at pH 2.8, 9.4 and at pH 7.0 in function of time; dashed frame: maleimide CH, solid frames: maleamate and at pH 2.8 maleamic acid CH. No differences were observed when the complex was incubated for 5 min or 24 h at pH 2.8. {*c* = 0.5 mM; 10% (v/v) D<sub>2</sub>O}.

### Lipophilicity

The lipophilic and/or hydrophilic character is an important property influencing the ability of a drug candidate to penetrate biological membranes. The shake flask method was used for the determination of the *n*-octanol/water partition of the Ru(II) complexes. The complexes [Ru(phen)<sub>3</sub>]Cl<sub>2</sub> and [Ru(bpy)<sub>3</sub>]Cl<sub>2</sub> were used as references. Table 1 exhibits the logarithm of

the distribution coefficient of the complexes determined at pH 7.40 ( $\log D_{7.40}$ ). Interestingly the lipophilicity of complexes **1-3** appears to be highly dependent on the salt content of the aqueous medium as previously observed.<sup>40</sup> **1** shows a slight preference for the aqueous phase over the non-polar solvent ( $\log D = -0.03$ ) when no extra salt is added in the aqueous fraction, while it becomes extremely lipophilic in PBS buffer ( $\log D > +3.7$ ). A  $\log D = +0.48$  was reported by Mazuryk *et al.* for **1** dissolved in water.<sup>108</sup> Experiments were repeated with the two main components of PBS separately, *i.e.* with 0.1 M NaCl and 15 mM phosphate. The results showed the more pronounced role of the chloride ions over phosphate (see Table 1). The same tendency was observed for complexes **2** and **3**. Ion pair formation most probably takes place between the positively charged complexes and anions like  $\text{Cl}^-$  and  $\text{H}_2\text{PO}_4^-/\text{HPO}_4^{2-}$  leading to a reduced aqueous partition. Phenyl substitution of the coordinating 1,10-phenantrolines at positions 4 and 7 might be responsible for this phenomenon through  $\pi$ -stacking, since the lipophilicity of the non-substituted complexes  $[\text{Ru}(\text{phen})_3]\text{Cl}_2$  and  $[\text{Ru}(\text{bpy})_3]\text{Cl}_2$  do not display a significant dependency on the electrolyte concentration of the aqueous phase. Both appear to be rather hydrophilic even in the presence of 1 M NaCl and 15 mM phosphate ( $\log D_{7.40} = -1.09$  and  $< -1.7$ , respectively).

**Table 1.** *n*-Octanol/aqueous phase distribution coefficients at pH 7.40 (expressed as  $\log D_{7.40}$ ) determined for **1-3**,  $[\text{Ru}(\text{phen})_3]\text{Cl}_2$  (Ru-phen) and  $[\text{Ru}(\text{bpy})_3]\text{Cl}_2$  (Ru-bpy) at 25 °C.

aqueous medium	<b>1</b>	<b>2</b>	<b>3</b>	<b>Ru-phen</b> <sup>a</sup>	<b>Ru-bpy</b> <sup>b</sup>
H <sub>2</sub> O <sup>c</sup>	-0.03±0.02 <sup>d</sup>	+0.81±0.05	+0.82±0.05	-1.6±0.1 <sup>e</sup>	< -1.7
0.1 M NaCl <sup>c</sup>	> +3.7	> +3.7	+2.4±0.1	-	-
PBS	> +3.7	> +3.7	+2.3±0.1	-1.30±0.05 <sup>f</sup>	< -1.7 <sup>g</sup>
15 mM phosphate	+1.5±0.1	+2.3±0.1	+1.1±0.1	-1.5±0.1	< -1.7

<sup>a</sup>  $\log D_{7.40} = -1.09$  in 15 mM phosphate and 1 M NaCl. <sup>b</sup>  $\log D_{7.40} < -1.7$  in 15 mM phosphate and 1 M NaCl.

<sup>c</sup> pH between 6.5–8.0. <sup>d</sup> Reported partition coefficient:  $\log P = +0.48$  in water.<sup>24</sup> <sup>e</sup> Reported partition coefficient:  $\log P = -1.5$  in water.<sup>109</sup> <sup>f</sup> Reported partition coefficients:  $\log P = -1.1$  in PBS,<sup>110</sup>  $\log P = -0.33$  solvent not reported.<sup>109</sup> <sup>g</sup> Reported partition coefficients:  $\log P = -1.21$  in PBS,<sup>111</sup>  $\log P = -0.41$  solvent not reported.<sup>109</sup>

Since the  $\log D$  of a drug is related to the putative partitioning between lipid membrane and blood plasma the  $\log D$  values determined in *n*-octanol – PBS buffer system can be considered

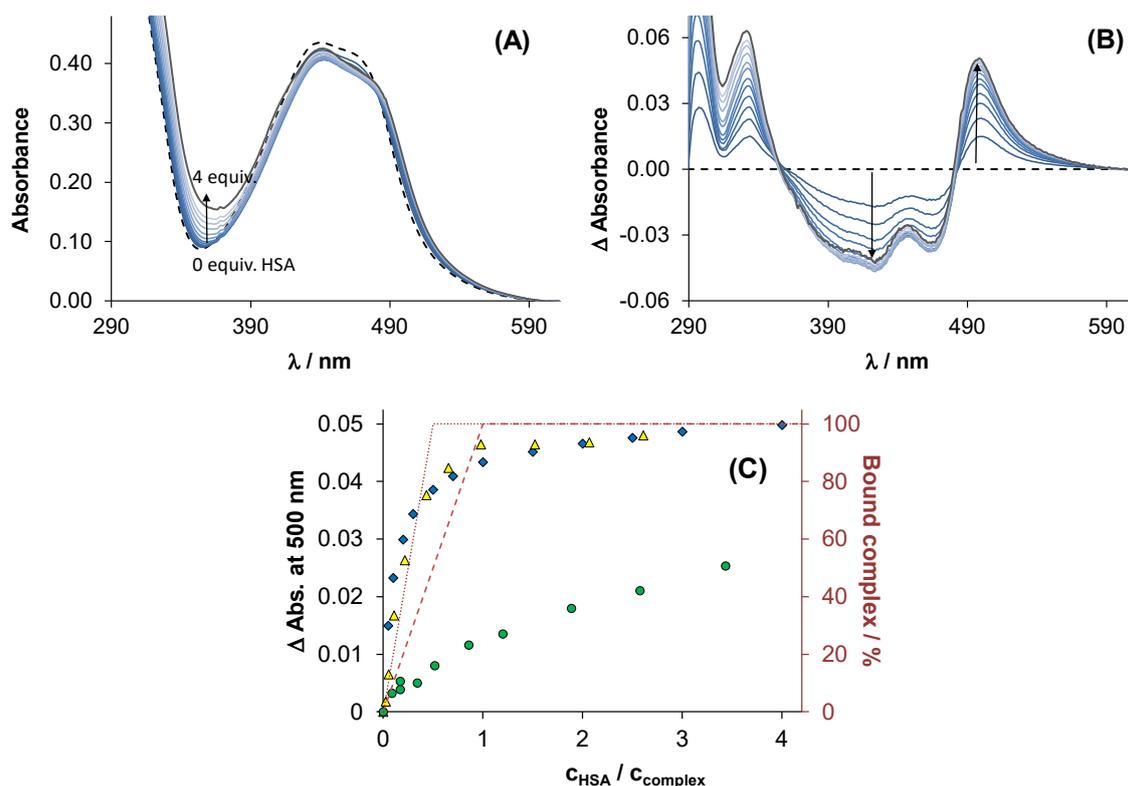
as more realistic. The results of lipophilicity measurements for complexes **1-3** show that they are highly lipophilic, which suggest that their cellular uptake via passive transport is possible.

### **Albumin binding studies**

In order to evaluate the ability of the complexes to bind albumin, complexes **1-3** were incubated with bovine serum albumin (BSA) in a 2:1 molar excess. BSA was initially used as a model for HSA, since it bears the same cysteine at position 34. The mixtures were then dialyzed against distilled water, to remove any unbound complex, and subjected to MALDI-TOF mass spectrometry, using BSA as a control. While BSA can be observed as a single peak in the 60-70 kDa region (mean mass of 66332 Da), an additional peak could be observed when complexes **2** and **3** were incubated with BSA (Figure S14). This would suggest that the complexes were successfully covalently bound to BSA. However, the same phenomenon was observed for complex **1**. The additional peak observed could therefore represent a non-covalent complex between BSA and the respective compounds, although it is surprising that such a complex would resist the ionization process. We therefore cannot exclude that ion pairing between BSA and free complexes occurred in the plasma during the ionization. Thus, a more in-depth study was performed using spectroscopic techniques. Of note, HSA was used in the following experiments to better represent what could happen in human patients.

To study the interaction of complexes **1-3** with albumin, the UV-Vis spectra of the complexes in the presence of HSA were recorded to follow protein adduct formation processes. Figure 3 shows the minor but significant spectral changes of **1** upon addition of increasing amounts of HSA in a wavelength range where the protein does not absorb light. The binding process takes rapidly place as the equilibrium was reached within 1-2 min. The spectrum profile of the albumin-bound complex is practically identical with the one recorded in *n*-octanol (see Figure S15A). This observation suggests that the metal complex is accommodated in one or more of the hydrophobic binding pockets of HSA. The calculated difference between the UV-Vis spectra (Figure 3B) indicates two isosbestic points at 356 nm and 482 nm. The changes of the absorbance difference values at 500 nm (Figure 3C) suggest the existence of more than two binding sites on HSA since quantitative binding curves assuming one (dashed line) or two (dotted line) binding sites plotted in the figure are crossed by the collected absorbance values. Complexes **2** and **3** displayed somewhat different behaviour (Figures S16 and S17). Although, spectral changes are similar to those observed for complex **1**, no isosbestic points were observed. This observation, and the slight differences

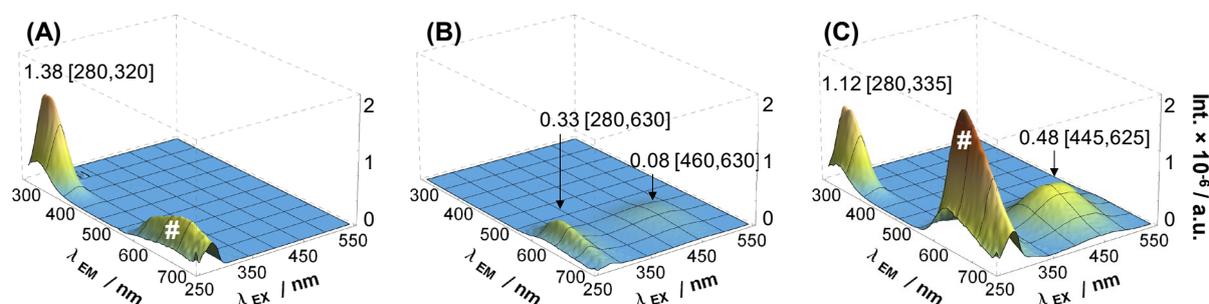
between the octanolic and HSA-bound spectra (see Figure S15B for **2**), strongly support an additional binding mode for complexes **2** and **3** via the maleimide moiety (*vide infra*). In the case of complex **2**, a biphasic binding character is clearly seen (Figures S16), while **3** shows a lower binding affinity towards HSA, since the same amount of HSA affects much less the absorbance spectrum of **3** in comparison to **1** and **2** (Figure 3C).



**Figure 3.** UV–Vis absorbance spectra of **1** in the presence of increasing amounts of HSA (A); derived difference spectra of the same system (B) and absorbance changes of **1** ( $\blacklozenge$ ), **2** ( $\blacktriangle$ ) and **3** ( $\bullet$ ) at 500 nm plotted together with quantitative binding curves corresponding to one (red dashed line) and two (red dotted line) binding sites on the protein.  $\{c_{\text{complex}} = 20 \mu\text{M}; c_{\text{HSA}} = 0\text{--}80 \mu\text{M}; \text{pH} = 7.4 \text{ (PBS)}; 25 \text{ }^\circ\text{C}\}$ . Difference spectra are calculated as follows:  $\text{Absorbance}_{(\text{complex} - \text{HSA mixture})} - \text{Absorbance}_{(\text{complex alone})} - \text{Absorbance}_{(\text{HSA alone})}$ .

To further study the binding mode between **1-3** and albumin, we exploited the intrinsic luminescence of these Ru(II) polypyridyl complexes.<sup>112</sup> As the three-dimensional luminescence spectra in Figure 4 show, complex **2** has two excitation maxima at 280 nm and 460 nm and emits light between 550 and 750 nm. A *ca.* 6-fold increase of phosphorescence was observed in the presence of 1 equivalent of albumin. On the contrary, the emission peak

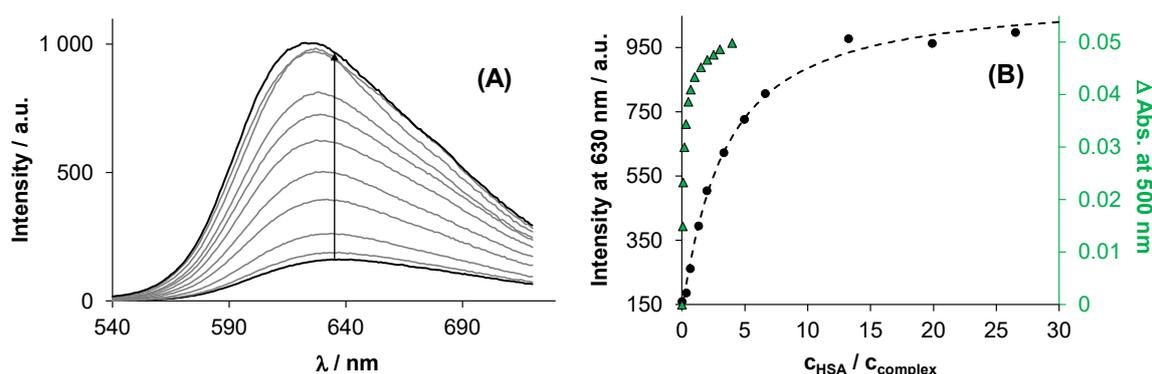
of HSA at  $\lambda_{EM} = 320$  nm (that originates mainly from the tryptophan at position 214 (Trp-214) situated near site I in subdomain IIA) is significantly reduced, which is a result of the binding of complex **2** at site I. The very same behaviour was obtained for **1**, while **3** quenched only slightly the intrinsic fluorescence of HSA (Figure S18).



**Figure 4.** Three-dimensional luminescence spectra of HSA (A), **2** (B) and the HSA–**2** (1:1) (C) system. Peak intensities (Int.) and wavelength coordinates ( $[\lambda_{EX}, \lambda_{EM}]$ ) are indicated in the figure; peaks with symbol # originate (partly) from the second order diffraction effect of the monochromator.  $\{c_{HSA} = c_{complex} = 5 \mu\text{M}; \text{pH} = 7.40$  (PBS);  $25 \text{ }^\circ\text{C}$ ; spectra are corrected by self-absorbance and inner filter effect $\}$ .

When the Ru(II) complexes are titrated by HSA, the emission intensity gradually increases as seen for complex **1** on Figure 5. The emission maximum is gradually shifted towards shorter wavelengths and the measured intensities reached saturation after the addition of *ca.* 15 equivalents of HSA. In Figure 5/B it can be seen that the fluorometric titration curve is far less steep compared to the absorbance-ratio curve, namely the intrinsic fluorescence of complex **1** is not sensitive to all the binding events. The titration data could be fitted well with the simple 1:1 binding model; therefore, no more complex binding model was not applied. The binding constant of  $\log K' = 5.1 \pm 0.1$  was calculated with the computer program HypSpec.<sup>100</sup> Complex **2** showed a rather similar behaviour in the presence of HSA, although a slightly higher binding constant was calculated on the basis of **2**–HSA titrations (Table 2). Complex **3** behaved again somewhat differently since no shifting of the emission maximum was observed (Figure S19), and a remarkably lower binding constant was calculated ( $\log K' = 4.2 \pm 0.1$ ) in comparison to those of **1** and **2**. The measured and calculated data points in Figures 5 and S19 show a strong fit, although it should be noted that binding of a second (and

third) complex on HSA cannot be ruled out. Of note these results could also be confirmed by following the changes in lifetime decay of the metal complexes in presence and absence of HSA (Figure S20). Interestingly, the lifetime of the albumin-bound complexes increased in absence of molecular oxygen (Table S1). This suggests that the complexes are still able to photosensitize oxygen, and thus maintain their photodynamic potential even when bound to albumin.



**Figure 5.** Phosphorescence spectra of **1** in the presence of increasing amounts of HSA (A). Measured and fitted intensities at 630 nm plotted together with the absorbance changes of **1** titrated by HSA (B). { $C_{\text{complex}} = 3 \mu\text{M}$  (fluorometry),  $20 \mu\text{M}$  (UV-Vis);  $\lambda_{\text{EX}} = 445 \text{ nm}$ ;  $\text{pH} = 7.40$  (PBS);  $25^\circ\text{C}$ }.

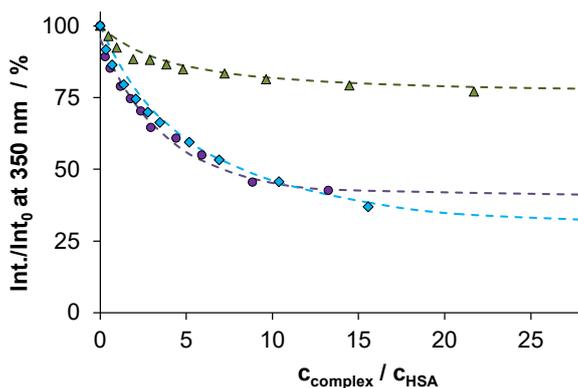
**Table 2.** Conditional binding constants ( $\log K^*$ ) of the complexes at binding sites I and II of HSA and binding constant calculated based on the intrinsic phosphorescence of the complexes determined by spectrofluorometric measurements. { $\text{pH} = 7.40$  (PBS);  $25^\circ\text{C}$ }.

$\log K^*$ <sup>a</sup>	<b>1</b>	<b>2</b>	<b>3</b>
Trp quenching	$5.2 \pm 0.1$ <sup>b</sup>	$5.2 \pm 0.1$	-
WF displacement	$5.5 \pm 0.1$	$5.5 \pm 0.1$	-
DG displacement	$5.4 \pm 0.1$	$5.4 \pm 0.1$	$4.5 \pm 0.1$
intrinsic phosphorescence of the complex	$5.1 \pm 0.1$	$5.4 \pm 0.1$	$4.2 \pm 0.1$

<sup>a</sup> Data presented are the mean value  $\pm$  standard deviation of at least two independent assays. <sup>b</sup>  $\log K^* = 5.09$  reported by Mazuryk *et al.*<sup>24</sup> WF: warfarin, DG: dansylglycine.

## Binding location study

The binding location of the metal complexes in HSA was investigated as a next step. The binding at site I (subdomain IIA) and site II (subdomain IIIA) was studied *via* Trp quenching and site marker displacement experiments. Conditional binding constants computed by HypSpec based on the titration data are listed in Table 2 and fitted quenching curves are depicted in Figure 6. Calculated constants for complexes **1** and **2** are fairly similar at both sites. The Trp-214 quenching constant for **1** is in good agreement with the data reported by Mazuryk *et al.* ( $\log K' = 5.09$ ).<sup>24</sup> The binding site preference and affinity of **3** to HSA were generally different from that of **1** and **2**. Although a Trp-214 quenching constant similar to that of **1** could be calculated for **3** ( $\log K' = 5.4 \pm 0.1$ ), the fluorescence of Trp-214 was quenched in only 27% (Figure 6). The warfarin (WF) displacement experiment showed even less alteration in the fluorescence signal of the HSA–WF system upon addition of **3** (see Figure S21). The results of this latter finding and the Trp-214 quenching experiments strongly suggest that the binding of **3** takes place on other place(s) than site I. The calculated quenching constant ( $\log K' = 5.4 \pm 0.1$ ) cannot be handled as a binding constant at site I, therefore it is not listed in Table 2. Most probably the allosteric effect of binding of complex **3** can be registered at site I. This phenomenon can be also observed in the reverse experiment, namely addition of WF to the **3**–HSA system increases the measured emission intensity of the complex (Figure S22). Lifetime measurements (data in Table S1) reveal that this increase originates mainly from the growing lifetime of bound **3**, *i.e.*, the close environment of bound metal complex changes but not primarily the bound fraction. The other two complexes behaved differently, both **1** and **2** could displace WF from site I and similar binding constants could be calculated from the displacement data as it was obtained in quenching experiments (Table 2). At the same time WF addition did not affect the fluorescence of **1**–HSA or **2**–HSA systems (Fig S23).



**Figure 6.** Fluorometric quenching curves recorded for the HSA–**1** (●), HSA–**2** (◆) and HSA–**3** (▲) systems, dashed lines are the fitted curves calculated on the basis of quenching constants ( $\log K'$ ) listed in Table 2 ( $C_{\text{HSA}} = 1 \mu\text{M}$ ;  $\lambda_{\text{EX}} = 295 \text{ nm}$ ;  $\text{pH} = 7.40$  (PBS);  $25 \text{ }^\circ\text{C}$ ).

The binding at site II was investigated in the HSA – dansylglycine (DG) – **complex 1-3** ternary systems (see three-dimensional fluorescence spectra and lifetime data collected for HSA – DG – **3** in Figure S24 and Table S2). A relatively low binding constant of **3** at site II could be computed, which is about one order of magnitude lower compared to those of **1** and **2** (Table 2). The decrease of HSA-bound **3** fluorescence in the presence of DG seems to originate both from changes of the environment of the bound form ( $\tau_2$  decreased from 3060 ns to 2870 ns) and the slight increase of the bound fraction ( $\alpha_2$ : 47%  $\rightarrow$  53%). In contrast, **1** and **2** could displace DG, but phosphorescence of these complexes did not change in the reversed type titrations.

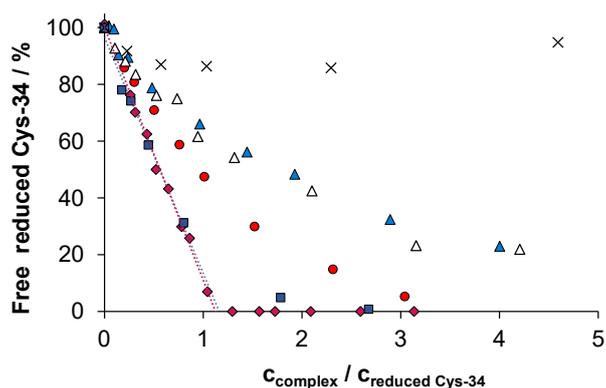
To summarize, complexes **1** and **2** can effectively displace WF and DG, apparently occupying sites I and II. On the other hand, **3** can displace slightly DG but not WF. Most probably, its binding at another additional site could be responsible for the elevated phosphorescence of the complex. This particular site seems to be insensitive to any events occurring at sites I or II in the case of complexes **1** and **2**, while the binding site of complex **3** appears to be allosterically connected to site I and II.

### Covalent binding with albumin study

The possible covalent interaction of **2** and **3** with Cys-34 thiol moiety in albumin was investigated as well. Molecules containing sulfhydryl group can form a stable thioether

conjugate with maleimide at pH between 6.5 and 7.5. HSA, as the most abundant plasma protein having one accessible cysteine (Cys-34), is the most likely target of Mal-linked agents in blood. *Ca.* 20–30% of the Cys-34 thiol groups of HSA are oxidized in the human blood (disulphide bridges are formed with small sulfhydryl compounds);<sup>113</sup> in contrast, commercially available lyophilized HSA is usually oxidized at higher extent (depending on the recovering and storage conditions). The concentration of the free thiol group of Cys-34 of the protein was determined via reaction with 4,4'-dithiodipyridine (DTDP) by spectrophotometry according to our previously reported protocol.<sup>100</sup>  $18\pm 2\%$  of Cys-34 of the albumin stock was determined to be present in its non-oxidized form. Since reaction of the Mal moiety of **2** and **3** with the thiol group of Cys-34 decreases the free thiol content of the protein, DTDP protocol can be used in order to quantify the sulfhydryl groups of HSA in the presence of the metal complexes (see Figure S25 for UV–Vis spectra). Figure 7 shows the changes of free thiol content in HSA as a function of the added equivalents of the complexes. Complex **1** applied as negative control, affects barely the quantity of free thiol groups. On the other hand, the Mal-functionalized complex **2** and **3** interact in a significant extent with Cys-34. The incubation of **2** with HSA for 3 h ( $\Delta$ ) or 30 min ( $\blacktriangle$ ) did not result in remarkable differences. In addition, the interaction with the native protein did not result in a quantitative covalent binding at this site. A similar behaviour was observed for **3**. Measurements on the sodium dodecyl sulphate (SDS)-denatured protein revealed nearly quantitative interaction of **2** and **3** with the Cys-34 thiol group of HSA. Two scenarios are possible regarding the interaction with the native protein: (i) concurrent binding at other sites in HSA reduces the effective concentration of **2** and **3**, or (ii) the hydrolysis of the Mal moiety competes with the covalent binding of the complexes with the Cys-34, which takes relatively longer on the native protein. Complex **3** seems to react more efficiently with the native protein than complex **2**, which could be due to its lower non-covalent binding.

Complexes **2** and **3** were found to react with the Cys-34 thiol group of HSA. Although the interaction is not quantitative, a significant binding *via* the maleimide moiety is highly probable in blood as well.



**Figure 7.** Free Cys-34 SH content of HSA at various **complex**-to-HSA ratios; symbols denote the following complexes and conditions  $\Delta$ : **2**, 3 h incubation,  $\blacktriangle$ : **2**, 30 min incubation;  $\blacksquare$ : **2**, 0.5% SDS, 30 min;  $\bullet$ : **3**, 30 min;  $\blacklozenge$ : **3**, 0.5 % SDS, 30 min and  $\times$ : **1** as negative control, 30 min  $\{c_{\text{HSA}} = 130 \mu\text{M}; c_{\text{Cys-34}} = 23 \mu\text{M}$  (18% of HSA); pH = 7.00 (100 mM phosphate buffer)}.

Overall, these data suggest that complexes **1** and **2** strongly bind non-covalently to sites I and II of albumin, while this interaction appears significantly weaker for **3**. This could be due to the relatively higher hydrophilicity and steric hindrance provided by the PEG spacer. While both **2** and **3** are able to bind covalently *via* their Mal moiety to the Cys-34 of albumin, the relatively lower non-covalent binding of **3** is counterbalanced with a relatively higher covalent binding efficiency.

### Cell studies

Complex **1** has been described previously as cytotoxic even when cells were incubated in the absence of light exposure, which is an undesired property for a PDT PS.<sup>23,24</sup> We therefore compared the cytotoxic potential of complexes **1-3** in the dark on non-cancerous immortalized human retinal pigment epithelial cells (RPE-1) following 48 h of incubation. As shown in Table 3, complex **1** induced a significant cytotoxic effect, with an  $\text{IC}_{50}$  of about  $2 \mu\text{M}$ , which represents the concentration needed to kill 50% of the cells. Of note, the  $\text{IC}_{50}$  of complex **1** was in the same range than the cytotoxicity measured by Mazuryk *et al.* on 4T1 cells following 24 h of incubation.<sup>24</sup> In contrast, complexes **2** and **3** were found to be significantly less toxic, with  $\text{IC}_{50}$  values of 74 and  $100 \mu\text{M}$ , respectively. However, following their irradiation at 595 nm, complexes **2** and **3** remained highly phototoxic, with  $\text{IC}_{50}$  values in the range of 4-8  $\mu\text{M}$  on CT26 and RPE-1 cells. Their phototoxicity index (PI), which represents

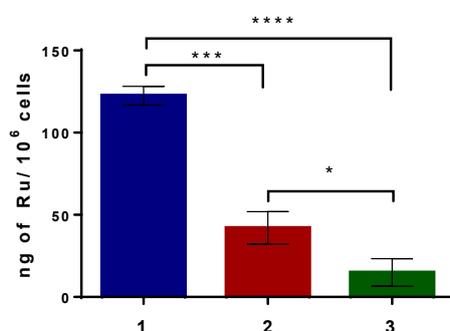
the ratio between their toxicity in the dark vs. their toxicity following irradiation, was found to be in the same range than that of complex **1**, if not slightly better. Additional experiments using higher concentrations of the complexes would be required to precisely determine their PI. However, while all three complexes appeared to be soluble at the highest concentration tested in culture medium (100  $\mu\text{M}$ ), solubility issues will occur at higher concentrations and significantly alter the results. Of note, much higher PIs were previously obtained for Ru(II)-based PSs. For instance, the PI of TLD-1433 was reported to be above 9'000 on CT26 cells.<sup>20</sup> However, many other parameters (*e.g.* tumor accumulation, irradiation wavelength, light source) are to be taken into account to evaluate the efficiency of a PS.

**Table 3.** IC<sub>50</sub> values of complexes **1-3** in various conditions. Column A: cells were incubated continuously with the complexes for 48 h in the dark. Column B: cells were incubated with the complexes for 4 h in the dark, then the medium was exchanged, and cells were incubated for an additional 44 h in the dark. Column C: cells were incubated with the complexes for 4 h in the dark, then the medium was exchanged, cells were irradiated for 2 h at 595 nm (22.47 J/cm<sup>2</sup>) and then incubated in the dark for an additional 44 h in the dark. The cell viability was determined using a resazurin assay. Results are presented as means  $\pm$  SD of three replicate experiments. PI: phototoxicity index, N.D.: not determined.

Cell Line	Compound	A - 48 h cytotoxicity ( $\mu\text{M}$ )	B - Dark ( $\mu\text{M}$ )	C - 595 nm ( $\mu\text{M}$ )	PI
CT26	<b>1</b>		5.35 $\pm$ 0.08	0.43 $\pm$ 0.07	12
	<b>2</b>	N.D.	>100	8.4 $\pm$ 0.6	> 12
	<b>3</b>		>100	4.43 $\pm$ 0.02	>23
RPE-1	<b>1</b>	2.2 $\pm$ 0.1	10.1 $\pm$ 0.5	0.8 $\pm$ 0.3	13
	<b>2</b>	74 $\pm$ 7	>100	4.1 $\pm$ 0.1	> 24
	<b>3</b>	100 $\pm$ 2	>100	4.4 $\pm$ 0.2	> 23

To rationalize these results, a cellular internalization assay was performed on CT26 cells. Cells were incubated with 5  $\mu\text{M}$  of the complexes **1-3** for 4 h. Following a thorough washing procedure, the cells were harvested and digested in nitric acid. Subsequently, the digests were analyzed by ICP-MS to quantify the amount of Ru internalized by the cells. As can be seen in

Figure 8, complexes **2** and **3** were significantly less internalized than complex **1** (3 and 8 times less, respectively). It is well known that the lipophilicity of a compound can have a significant effect on its cellular internalization.<sup>114</sup> While this could explain that the more hydrophilic complex **3** is less internalized than **1** and **2**, it does not seem to apply to complex **2**, which is highly hydrophobic. However, as complex **2** and **3** can covalently bind to albumin, which is present in the culture medium, their reduced internalization could be due to their covalent binding to the bulky protein, which would in turn prevent their uptake *via* passive diffusion. This could be an issue for future *in vivo* applications. If the complexes are not released and subsequently internalized by cancer cells once vectorized to the tumor, they will not be able to reach cellular organelles, such as the nucleus, the mitochondria, or the lysosomes, where they could exert the highest effect.<sup>115</sup>



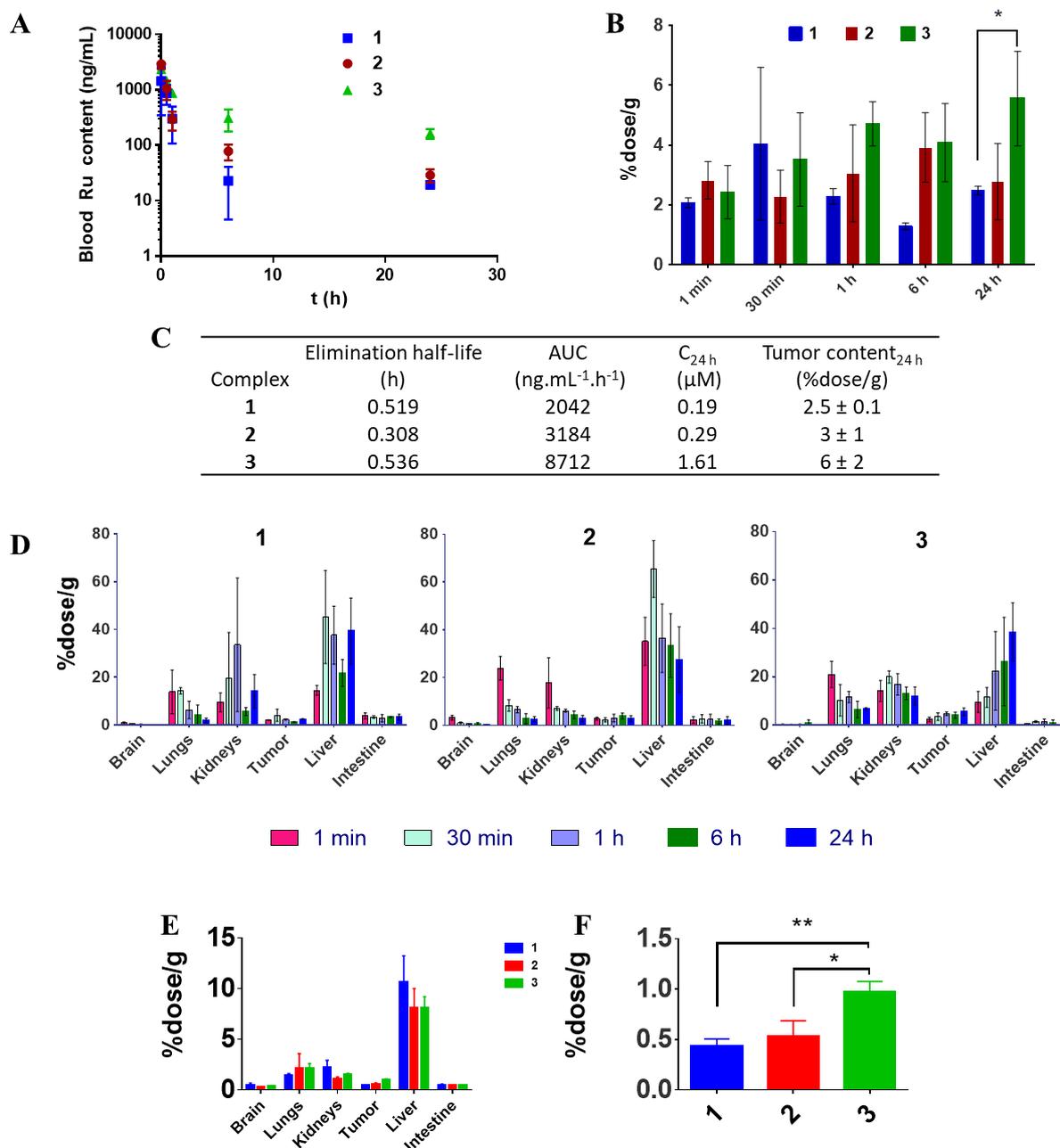
**Figure 8.** Cellular uptake of complexes **1-3** in CT26 cells. The results are presented as mean and SD of three replicates. (\*  $P < 0.05$ , \*\*\*  $P < 0.0003$ , \*\*\*\*  $P < 0.0001$ , t-test).

Nevertheless, the thiol-Mal bond can be reversible through thiol-exchange reactions.<sup>116</sup> If the albumin-complexes are efficient at accumulating and residing at the tumor site, one could hope that the PS will eventually be released from the protein. On the other hand, this phenomenon could also lead to the premature release of the PS in the blood circulation or in sensitive organs, thus increasing off-target toxicity. Such complex processes are however hard to predict using solely *in vitro* experiment, and *in vivo* data could help understanding the fate of complexes **1-3** following their administration.

### Animal studies

We therefore sought out to evaluate the biodistribution of the complexes **1-3** in a CT26 tumor-bearing mouse model. Complexes **1-3** were injected into the tail vein at a dose of 2  $\mu\text{mol/kg}$

(corresponding to 2.04, 2.27 and 3.25 mg/kg, respectively). After a given time interval (ranging from 1 min to 24 h), the mice were euthanized and their blood and organs were harvested, digested in nitric acid, and the resulting digests were then analyzed by ICP-MS (3 mice/group). It is important to mention that this method only provides the amount of Ru in tissues but does not give any information about its form (initial complex, degradation product, metabolites, etc.). Of note, due to solubility issues, complexes **1** and **2** required the addition of polysorbate 80 (1%) to prevent aggregation, while complex **3** appeared completely soluble at this concentration. The formulation of **1** and **2** with polysorbate was performed using the film rehydration technique, described more in detail in the experimental part, while complex **3** was directly solubilized in 5% glucose. While polysorbate might slightly alter the biodistribution of the compounds, we first made sure that it does not prevent the binding of complex **2** with albumin (see Figure S26). As shown on Figure 9A, complexes **1** and **2** were quickly eliminated from the blood stream within the first 6 hours following injection and only 0.19 and 0.29  $\mu\text{M}$  of Ru were detected in the blood 24 h following injection (Figure 9C). In contrast, complex **3** appeared to be eliminated more slowly in the first hours following injection, and 1.61  $\mu\text{M}$  of Ru were still detected in the blood at 24 h following injection. The comparable elimination half-lives for the three complexes suggest a similar elimination mechanism. Consequently, the area under the curve (AUC) for complex **3**, which represents the overall exposition of the animals to the complex, is significantly higher than that of complex **1** and **2**. While we cannot exclude that the PEG spacer in complex **3**, or the different formulation used might be fully responsible for this difference in the blood Ru profiles, its slightly better covalent binding efficiency to albumin (see Figure 7) might also participate in the extension of its blood circulation. Complexes **1-3** distributed in the organs in a similar way, and accumulated mainly in the liver and the kidney, suggesting that the elimination of the complexes occurs both by biliary and renal excretion (Figure 9D). All three complexes also appeared to accumulate in the tumors in a comparable manner (Figure 9B). The concentration of complex **3** appeared slightly but significantly higher in the tumor 24 h following injection in comparison to complex **1**. Of note, these results are to be taken with caution considering the small number of animals per group and the high variability of the results.



**Figure 9.** Complexes 1-3 were injected IV at a 2 μmol/kg dose of the complexes formulated in 1% polysorbate 80 in PBS (complexes 1 and 2) or in 5% glucose (complex 3) (n = 3 mice/time group). In (E) and (F), all three complexes were formulated in 5% glucose. (A) Blood Ru content as a function of time following the administration of complexes 1-3 determined by ICP-MS. (B) Time-dependent Ru tumour content determined by ICP-MS. Data are presented as mean and SD of the % of the total injected dose per g of tumour (\* P < 0.05, t-test). (C) Blood Ru kinetic parameters using a two-phase exponential decay fit. AUC: Area Under the Curve. (D) Time-dependent biodistribution of complexes 1-3 in CT26 tumour-

bearing mice following the IV injection complexes **1-3**. (n = 3 mice/time group). Results are presented as the mean and SD of the % of the Ru dose injected per g of organ. (E) Biodistribution of complexes **1-3** in CT26 tumour-bearing mice 48 h following the injection of complexes **1-3**. Results are presented as the mean and SD of the % of the total Ru injected dose per g of the corresponding organ. (F) Ru tumour content determined by ICP-MS 48 h following the injection of complexes **1-3**. Data are presented as mean and SD of the % of the total injected dose per g of tumour (\* P < 0.05, \*\* P < 0.005, t-test).

In order to confirm the trend observed previously, the experiment was repeated on a reduced number of animals to evaluate the biodistribution at a single timepoint, 48 h following administration of complexes **1-3**. As we realized in the meantime that the low solubility of complexes **1** and **2** in saline and PBS was not due to their low solubility in water, but to the aggregating effect induced by high salt concentrations, we formulated the three complexes in 5% glucose. All three complexes appeared soluble in this vehicle at the concentration required for IV administration (about 400  $\mu$ M). As shown in Figure 9E and 9F, complex **3** demonstrated a modestly (*ca.* 2-fold) higher accumulation in the tumour in comparison to complex **1**. Complex **2**, however, did not accumulate significantly more in the tumour in comparison to complex **1**. These results confirm the trend initially observed and show that replacing the polysorbate formulation by 5% glucose did not reverse the superiority of complex **3** over complex **2**.

Overall, these results suggest that adding a Mal moiety on complex **1** had a minor effect on its tumor accumulation. Since this strategy has already been proven as efficient in other metal complexes,<sup>93-95</sup> one could hypothesize that the high hydrophobicity of the three compounds lead to their fast capture in the liver, before they had a chance to react with albumin *in vivo*, which gives a slight advantage to the more hydrophilic complex **3**. Given the overall low tumor accumulation and inferior phototoxicity of complexes **2** and **3** in comparison to **1**, we decided not to evaluate their PDT efficiency *in vivo*.

## Conclusion

The *in situ* conjugation of drug-maleimide prodrugs to albumin has been described as an efficient strategy to improve the tumor accumulation and efficiency of antitumor drugs. In this

study, we showed that this strategy is not applicable to every compound. *In vitro*, the Ru(II) complexes bearing a Mal moiety described in this work were able to strongly bind non-covalently in several hydrophobic pockets of albumin, as well as covalently to its Cys-34. This property appeared to have a significant and negative impact on their ability to penetrate the cells, which led to a decrease in their phototoxicity. Unfortunately, this decrease was not compensated by a dramatic increase in their tumor accumulation *in vivo*, although the addition of a hydrophilic linker slightly improved the blood circulation of the complex. One improvement to this system would be to add a cleavable linker between the maleimide group and the PS, which would enable its release in the tumor microenvironment. Interestingly, the ability of the complexes **2** and **3** to react with the Cys-34 of albumin appeared to be inversely correlated with their propensity to be bound in the hydrophobic pockets of the protein. This phenomenon may be responsible for a hampered thiol-Mal reaction, which consequently increase the chances for the Mal moiety to be hydrolyzed before it could react with the Cys-34. Thus, Mal-prodrugs that strongly bind non-covalently to albumin are to be considered with caution. In addition, incorporating a hydrophilic spacer between the Mal group and the complex appears to be beneficial to both its blood circulation time and its albumin conjugation. Applying the *in situ* Mal-mediated albumin bioconjugation to photosensitizers therefore appears as promising strategy to improve the efficiency and safety of PDT. Nevertheless, a strong investigation into the interaction between Ru(II) complexes and plasma proteins is needed to design the appropriate system.

## Experimental section

### *Materials*

All chemicals were obtained from commercial sources and were used without further purification. The Ru(II) complexes *cis*-dichlorotetrakis(dimethylsulfoxide) Ru(II)<sup>117</sup> and dichlorobis(4,7-diphenyl-1,10-phenanthroline)Ru(II) (Ru(bphen)<sub>2</sub>Cl<sub>2</sub>)<sup>102</sup> and the complexes [Ru(phen)<sub>3</sub>]Cl<sub>2</sub> and [Ru(bpy)<sub>3</sub>]Cl<sub>2</sub><sup>118</sup> were synthesized as previously reported. The ligand 5-(aminomethyl)-2,2'-bipyridine was prepared as previously reported.<sup>101</sup> Spectral data were in accordance with the literature. The complex [Ru(bphen)<sub>2</sub>dmbipy][PF<sub>6</sub>]<sub>2</sub> was obtained in a previous study<sup>23</sup> and converted to the dichloride salt by elution with MeOH from the ion exchange resin Amberlite IRA-410 to yield [Ru(bphen)<sub>2</sub>dmbipy][PF<sub>6</sub>]<sub>2</sub> (**1**). The hydrophilic BOC-protected linker 2,2-dimethyl-4,24-dioxo-3,8,11,14,17,20-hexaoxa-5,23-diazaheptacosan-27-oic acid was prepared as previously described.<sup>102-105</sup> Cells were purchased from the ATCC®. Sinapic acid (SA, used as the matrix for MALDI-TOF experiments, was of the highest grade available and used without further purification) was purchased from Sigma Aldrich Co. Human serum albumin (HSA) containing fatty acids (A8763, essentially globulin free), KCl, NaCl, NaH<sub>2</sub>PO<sub>4</sub> and KH<sub>2</sub>PO<sub>4</sub>, Na<sub>2</sub>HPO<sub>4</sub>, warfarin (WF), dansylglycine (DG), 2,2'-dithiodipyridine (DTDP), sodium dodecyl sulfate (SDS), trimethylsilylpropanesulfonate (DSS) were purchased from Sigma-Aldrich. Milli-Q water was used for sample preparation. Samples for albumin binding studies were prepared in phosphate buffered saline (PBS) at pH 7.40. Stock solutions of HSA, WF and DG were prepared as described previously.<sup>119</sup> Pure aqueous stock solutions (*c* = 1 mM) of the Ru(II) complex were freshly prepared every day and their concentrations and molar absorptivities were determined based on a weight-in-volume basis. In the HSA binding experiments these stock solutions were diluted with PBS to get the working solutions (*c* = 50–200 μM, pH = 7.40). All measurements were carried out at 25.0 ± 0.2 °C.

### *Instrumentation and methods*

<sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on a Bruker 400 MHz NMR spectrometer. Chemical shifts (δ) are reported in parts per million (ppm) referenced to tetramethylsilane (δ 0.00) ppm using the residual proton solvent peaks as internal standards. Coupling constants (J) are reported in Hertz (Hz) and the multiplicity is abbreviated as follows: s (singlet), d (doublet) and m (multiplet). ESI-MS experiments were carried out using a LTQ-Orbitrap XL from

Thermo Scientific (Thermo Fisher Scientific, Courtaboeuf, France) and operated in positive ionization mode, with a spray voltage at 3.6 kV. No Sheath and auxiliary gas were used. Applied voltages were 40 and 100 V for the ion transfer capillary and the tube lens, respectively. The ion transfer capillary was held at 275 °C. Detection was achieved in the Orbitrap with a resolution set to 100,000 (at  $m/z$  400) and a  $m/z$  range between 150-2000 in profile mode. Spectrum was analysed using the acquisition software XCalibur 2.1 (Thermo Fisher Scientific, Courtaboeuf, France). The automatic gain control (AGC) allowed accumulation of up to  $2 \cdot 10^5$  ions for FTMS scans, Maximum injection time was set to 300 ms and 1  $\mu$ scan was acquired. 10  $\mu$ L was injected using a Thermo Finnigan Surveyor HPLC system (Thermo Fisher Scientific, Courtaboeuf, France) with a continuous infusion of methanol at 100  $\mu$ L $\cdot$ min $^{-1}$ . Elemental microanalyses were performed on a Thermo Flash 2000 elemental analyser. MALDI-TOF experiments were performed on a MALDI-TOF/TOF UltrafleXtreme mass spectrometer (Bruker Daltonics, Bremen). Mass spectra were obtained in linear positive ion mode. The laser intensity was set just above the ion generation threshold to obtain peaks with the highest possible signal-to-noise (S/N) ratio without significant peak broadening. All data were processed using the FlexAnalysis software package (Bruker Daltonics). Fluorescence and phosphorescence lifetime measurements were carried out on a Fluoromax (Horiba Jobin Yvon) spectrofluorometer equipped with a DeltaHub time correlated single photon counting (TCSPC) system using nanoLED light sources N-295, N-350 and N-460 (Horiba Jobin Yvon). ICP-MS studies were performed using an Element II HR-ICP-MS (Thermo Fisher Scientific).

## ***Synthesis***

### **[Ru(bphen)<sub>2</sub>dmbipyNH<sub>2</sub>][PF<sub>6</sub>]<sub>2</sub>**

Ru(bphen)<sub>2</sub>Cl<sub>2</sub> (200 mg, 0.24 mmol, 1.0 equiv.) and 5-(aminomethyl)-2,2'-bipyridine (57 mg, 0.29 mmol, 1.2 equiv.) were dissolved in a 1:3 mixture of H<sub>2</sub>O/ethanol (50 mL) and refluxed overnight under N<sub>2</sub> atmosphere. The solvent was evaporated, and the residue dissolved in 10 mL of H<sub>2</sub>O. A saturated, aq. NH<sub>4</sub>PF<sub>6</sub> solution was added, and the resulting precipitate was collected by vacuum filtration. The solid was washed with H<sub>2</sub>O (50 mL) and Et<sub>2</sub>O (50 mL). The product was dried in high vacuum. Yield: 89%. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>CN)  $\delta$  = 8.59 (s, 1H), 8.44 (s, 1H), 8.29 (d,  $J$  = 5.5 Hz, 1H), 8.26 (d,  $J$  = 5.5 Hz, 1H), 8.15-8.07 (m, 6H), 7.93 (d,  $J$  = 5.9 Hz, 1H), 7.74-7.70 (m, 3H), 7.62-7.47 (m, 22H), 7.39 (dd,  $J$  = 5.9, 1.7 Hz, 1H),

7.24 (d,  $J = 5.7$  Hz, 1H), 4.38 (s, 2H), 2.53 (s, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CD}_3\text{CN}$ )  $\delta = 158.4$ , 156.6, 152.9, 152.6, 152.4, 152.3, 151.8, 151.3, 149.5, 149.4, 149.4, 148.9, 148.7, 148.6, 142.9, 136.2, 136.2, 130.3, 130.2, 130.0, 130.0, 129.6, 129.5, 129.4, 129.4, 129.3, 127.4, 126.8, 126.7, 126.6, 126.4, 125.6, 124.5, 42.8, 20.8.

### **[Ru(bphen)<sub>2</sub>dmbipyMal][Cl]<sub>2</sub> (2)**

**[Ru(bphen)<sub>2</sub>dmbipyMal][PF<sub>6</sub>]<sub>2</sub>** was synthesized as previously described.  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra were in accordance with the reported data.<sup>37</sup>

The counter ion  $\text{PF}_6^-$  was exchanged to  $\text{Cl}^-$  by elution with MeOH from the ion exchange resin Amberlite IRA-410. Elemental analysis calcd. for  $\text{C}_{64}\text{H}_{45}\text{Cl}_2\text{N}_7\text{O}_2\text{Ru}+\text{H}_2\text{O}$  (%): C 67.78, H 4.18, N 8.65; found: C 67.73, H 3.94, N 8.36.

### **[Ru(bphen)<sub>2</sub>dmbipyPEGNH<sub>2</sub>][PF<sub>6</sub>]<sub>2</sub>**

To a solution of 2,2-dimethyl-4,24-dioxo-3,8,11,14,17,20-hexaoxa-5,23-diazaheptacosan-27-oic acid (0.150 g, 0.31 mmol, 1.00 equiv.) in anhydrous dichloromethane (DCM) (5 mL) was added N-hydroxysuccinimide (0.043 g, 0.37 mmol, 1.2 equiv.) and dicyclohexylcarbodiimide (0.076 g, 0.37 mmol, 1.2 equiv.). The mixture was agitated for 15 h at room temperature. The solid was filtered off and the filtrate evaporated to yield a transparent oil (0.185 g). To a solution of this oil in anhydrous DCM (2 mL) was added a solution of **[Ru(bphen)<sub>2</sub>dmbipyNH<sub>2</sub>][PF<sub>6</sub>]<sub>2</sub>** (0.197 mg, 0.16 mmol, 1.00 equiv.) and N,N-diisopropylethylamine (DIPEA) (0.08 mL, 0.48 mmol, 3.0 equiv.) in DCM (3 mL). The mixture was agitated in the dark at room temperature under nitrogen for 15 h. The solvent was evaporated, and the residue was dissolved in methanol (10 mL). A saturated solution of  $\text{NH}_4\text{PF}_6$  was added and the orange solid was isolated by filtration, washed with cold water and diethyl ether ( $\text{Et}_2\text{O}$ ), and finally dried under vacuum. Under nitrogen, the solid was dissolved in DCM (1 mL), and trifluoroacetic acid (0.1 mL) was added. The mixture was agitated at room temperature for 15 h. The solvent was evaporated to dryness, the residue was dissolved in a small amount of methanol, and a saturated solution of  $\text{NH}_4\text{PF}_6$  was added. The orange solid was isolated by filtration, washed with cold water and  $\text{Et}_2\text{O}$ . The crude product was purified by column chromatography on silica gel with an  $\text{CH}_3\text{CN}$  /aq.  $\text{KNO}_3$  (0.4 M) solution (10:1). The fractions containing the product were united and the solvent was removed. The residue was dissolved in  $\text{CH}_3\text{CN}$  and undissolved  $\text{KNO}_3$  was removed by filtration. The solvent was evaporated, and the product was dissolved in  $\text{H}_2\text{O}$  (20 mL). Upon addition of

$\text{NH}_4\text{PF}_6$  the product precipitated as a  $\text{PF}_6$  salt. The solid was obtained by filtration and was washed with  $\text{H}_2\text{O}$  (50 mL) and  $\text{Et}_2\text{O}$  (50 mL) to yield **[Ru(bphen)<sub>2</sub>dmbipyPEGNH<sub>2</sub>][PF<sub>6</sub>]<sub>2</sub>** as an orange powder (85 mg, 0.05 mmol, 58%). <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>CN)  $\delta$  = 8.64 (d,  $J$  = 1.7 Hz, 1H), 8.48 (d,  $J$  = 1.8 Hz, 1H), 8.28 (t,  $J$  = 5.5 Hz, 2H), 8.24 – 8.15 (m, 4H), 8.11 (t,  $J$  = 5.7 Hz, 2H), 7.76 (dd,  $J$  = 5.5, 1.4 Hz, 2H), 7.74 (d,  $J$  = 6.0 Hz, 1H), 7.71 – 7.53 (m, 21H), 7.26 – 7.21 (m, 2H), 7.17 (t,  $J$  = 6.1 Hz, 1H), 6.83 (s, 2H), 6.69 (t,  $J$  = 5.9 Hz, 1H), 4.56 (d,  $J$  = 6.1 Hz, 2H), 3.68 – 3.61 (m, 2H), 3.59 – 3.39 (m, 18H), 3.35 – 3.26 (m, 2H), 3.01 (dd,  $J$  = 6.1, 4.3 Hz, 2H), 2.61 – 2.54 (m, 4H), 2.51 – 2.46 (m, 2H); <sup>13</sup>C NMR (100 MHz, CD<sub>3</sub>CN)  $\delta$  = 174.0, 158.2, 157.6, 153.2, 153.1, 152.9, 152.6, 152.5, 152.2, 151.6, 149.94, 149.87, 149.85, 149.5, 149.31, 149.30, 136.71, 136.68, 136.6, 130.8, 130.72, 130.66, 130.62, 130.57, 130.11, 130.06, 129.9, 129.3, 127.09, 127.05, 127.0, 126.3, 126.2, 122.7, 70.84, 70.82, 70.80, 70.6, 70.5, 70.3, 70.2, 70.0, 67.3, 42.4, 40.6, 39.6, 31.3, 21.3.

### **[Ru(bphen)<sub>2</sub>dmbipyPEGMal][Cl]<sub>2</sub> (3)**

Under nitrogen, 2,5-dioxopyrrolidin-1-yl 2-(2,5-dioxo-2,5-dihydro-1H-pyrrol-1-yl)acetate (23 mg, 0.09 mmol, 2.0 equiv.) was added to a solution of **[Ru(bphen)<sub>2</sub>dmbipyPEGNH<sub>2</sub>][PF<sub>6</sub>]<sub>2</sub>** (73 mg, 0.045 mmol, 1.0 equiv.) and DIPEA (80 mg, 0.068 mmol, 1.5 equiv.) in DCM (5 mL). The mixture was agitated at room temperature in the dark for 15 h. The solvent was removed, and the residue was dissolved in a small amount of methanol, and a saturated solution of  $\text{NH}_4\text{PF}_6$  was added. The orange solid was isolated by filtration, washed with cold water and  $\text{Et}_2\text{O}$ . The counter ion  $\text{PF}_6^-$  was exchanged to  $\text{Cl}^-$  by elution with MeOH from the ion exchange resin Amberlite IRA-4101 to yield **[Ru(bphen)<sub>2</sub>dmbipyPEGMal][PF<sub>6</sub>]<sub>2</sub> (3)** as an orange powder (61 mg, 0.040 mmol, 89%). <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>CN)  $\delta$  = 8.79 (s, 1H), 8.53 (s, 1H), 8.30 (dd,  $J$  = 5.5, 2.8 Hz, 2H), 8.23 – 8.14 (m, 4H), 8.10 (dd,  $J$  = 5.5, 3.6 Hz, 2H), 7.76 (dd,  $J$  = 5.5, 3.8 Hz, 2H), 7.73 (d,  $J$  = 5.9 Hz, 1H), 7.70 – 7.52 (m, 23H), 7.28 – 7.17 (m, 3H), 6.78 (s, 2H), 6.68 – 6.60 (m, 1H), 6.00 (s, 1H), 4.58 (dd,  $J$  = 17.6, 6.1 Hz, 1H), 4.51 (dd,  $J$  = 17.6, 6.1 Hz, 1H), 4.03 (s, 2H), 3.53 – 3.34 (m, 20H), 3.31 – 3.19 (m, 4H), 2.58 (s, 3H), 2.52 (s, 4H). <sup>13</sup>C NMR (100 MHz, CD<sub>3</sub>OD)  $\delta$  = The counter ion  $\text{PF}_6^-$  was exchanged to  $\text{Cl}^-$  by elution with MeOH from the ion exchange resin Amberlite IRA-4101. <sup>13</sup>C NMR (100 MHz, CD<sub>3</sub>CN)  $\delta$  = 175.6, 174.6, 171.9, 169.2, 158.8, 158.3, 153.3, 153.0, 152.5, 152.2, 150.74, 150.68, 149.9, 149.8, 137.1, 137.0, 135.7, 131.1, 131.02, 130.94, 130.88, 130.4, 130.3, 130.0, 127.7, 127.4, 127.0, 126.8, 71.5, 71.3, 71.2, 70.4, 70.3, 58.3, 43.0, 40.64, 40.55, 40.4, 31.7, 31.4, 21.4, 18.4. ESI-HRMS (pos. detection mode): calcd. for C<sub>82</sub>H<sub>78</sub>N<sub>10</sub>O<sub>10</sub>Ru

$m/z$   $[M]^{2+}$  732.2467; found: 732.2481. The counter ion  $PF_6^-$  was exchanged to  $Cl^-$  by elution with MeOH from the ion exchange resin Amberlite IRA-4101. Elemental analysis calcd. for  $C_{64}H_{45}Cl_2N_7O_2Ru+5H_2O$  (%): C 60.59, H 5.46, N 8.62; found: C 60.68, H 5.46, N 8.75.

## ***Physicochemical properties***

### ***$^1H$ NMR spectroscopy for stability studies***

$^1H$  NMR spectroscopic studies were carried out on a Bruker Avance III HD Ascend 500 Plus instrument. Complexes were dissolved in 0.5 mM concentration in water, in PBS buffer, in 5 mM phosphate (pH 7.4) or in EMEM, 10% (v/v)  $D_2O$  was added to the samples. Spectra were recorded with the WATERGATE water suppression pulse scheme using DSS as internal standard.

### ***Lipophilicity and membrane permeability***

Distribution constants ( $D$ ) of **1**, **2** and **3** were determined by the traditional shake-flask method in *n*-octanol/buffered aqueous solution at pH 7.40 (PBS, or 15 mM phosphate buffer) and in water containing 0 M or 0.1 M KCl as described previously.<sup>90,120</sup> The complexes were dissolved in *n*-octanol pre-saturated aqueous solution at *ca.* 20  $\mu$ M concentrations. Aqueous phases and water pre-saturated *n*-octanolic phases were gently mixed in 1:1, or in 100:1 volume ratios with Heidolph Reax 2 overhead shaker (~20 rpm) for 2 h. After separation, UV–Vis spectra of the compounds in the aqueous phase were compared to those of the original aqueous solutions solution and  $D$  values of the compounds were calculated according to the following equation:

$$D = \left[ \frac{\text{Abs}_{(stock\ sol.)}}{\text{Abs}_{(aqueous\ phase\ after\ separation)}} - 1 \right] \times \frac{V_{(aqueous\ phase)}}{V_{(n\text{-}octanol\ phase)}} \quad (3)$$

An Agilent Cary 8454 diode array spectrophotometer was used to measure the UV–Vis spectra in the interval 200–800 nm.

### ***Albumin interaction***

### **MALDI Sample Preparation**

Complexes **1-3** from stock solution in ethanol were added to a 100  $\mu\text{M}$  solution of BSA in PBS at a final complex concentration of 200  $\mu\text{M}$  (final ethanol concentration: 5%). The mixtures were agitated at 450 rpm, 37  $^{\circ}\text{C}$  for 2 h in a Thermomixer (Eppendorf). The mixtures were then dialyzed against 2 L distilled water for 24 h (3 buffer exchanges) using dialysis cassettes (Slide-a-lyzer MWCO 10 kDa, Thermo Fisher).

The matrix solution was prepared at a concentration of 45 mM in  $\text{H}_2\text{O}/\text{CH}_3\text{CN}/\text{trifluoroacetic acid}$  1/1/0.1.

The samples were prepared by mixing the BSA-complex mixture solution with matrix solution at a volume ratio of 1:9.

### **UV-visible spectrophotometric and circular dichroism spectroscopic measurements**

UV-visible (UV-Vis) spectrophotometry was utilized to follow stability of the complexes in various aqueous solutions and monitor their interaction with HSA on an Agilent Cary 8454 diode array spectrophotometer and an Agilent Cary 3500 spectrophotometer in the wavelength range between 200 and 800 nm. Samples contained *ca.* 20  $\mu\text{M}$  metal complex dissolved in various buffer systems (PBS, phosphate) or in water. In protein binding studies a constant amount of complex was titrated by HSA (0–80  $\mu\text{M}$ ).

Interaction of the metal complexes at the Cys-34 residue of HSA was investigated via the DTDP method described in our former work.<sup>100</sup> The available Cys-thiol content in HSA was determined to be 18%. Complex binding was tested in the following setup: 130  $\mu\text{M}$  HSA (23  $\mu\text{M}$  free thiol) and various amounts of complex (0–120  $\mu\text{M}$ ) were incubated for 3 h or 30 min at pH 7.00 (100 mM phosphate), and the UV-Vis spectra were recorded (a), then 110  $\mu\text{M}$  DTDP was added, and the UV-Vis spectra were measured again after another 40 min waiting time (b). Spectrum of the colored reaction product 2-thiopyridone (2-TP) was derived by the subtraction of spectrum (a) from the spectrum (b). Blank experiment with **1** was carried out as well as a control experiment. The effect of protein unfolding on the thiol binding of **2** and **3** was studied by the addition of 0.5% (w/w) SDS to the protein prior to its reaction with the complex.

### **Spectrofluorometric measurements**

Samples were measured in 1×1 cm cells. PBS buffer was used for sample preparation; and emission spectra were recorded after 5 min incubation. Four kinds of experiments were carried out: (i) 1 μM HSA and various amounts of complex (from 0 to 20 equiv.) were used for quenching experiments; and (ii) fluorescence of site markers WF and DG was measured in samples containing 1 μM HSA, 1 μM marker and 0–20 μM compound; intrinsic fluorescence of the complexes was followed as well applying (iii) 5 μM complex and various HSA concentrations (0–110 μM) or (iv) 5 μM complex, 5 μM HSA and various site marker (WF, DG) concentrations (0–160 μM). Instrumental parameters are listed in Table S3. Computer program HypSpec<sup>121</sup> was utilized for calculation of formation constants for HSA–compound adducts as described in our former work.<sup>119,122</sup> Corrections for self-absorbance and inner filter effect were done as described in our former work using the formula suggested by Lakowicz.<sup>112</sup>

### Time-resolved fluorescence measurements

Ludox® (from Sigma-Aldrich) was used as scatter to obtain the instrument response function. The background (obtained with blank samples) was subtracted from the decay. The program DAS6 (version 6.6.; Horiba, Jobin Yvon) was used for the analysis of the experimental fluorescence decays. The fluorescence intensity decay over time is described by a sum of exponentials as the following equation shows:

$$I(t) = \sum_{i=1}^n \alpha_i \exp\left(\frac{-t}{\tau_i}\right) \quad (1)$$

where  $\alpha_i$  and  $\tau_i$  are the normalized amplitude and lifetime of component  $i$ , respectively. From these parameters, the fraction of emitted light by each component  $i$  can be calculated through the equation:

$$f_i = \frac{\alpha_i \tau_i}{\sum(\alpha_i \tau_i)} \quad (2)$$

The goodness of the fit was judged from a  $\chi^2_R$  value close to 1.0 and a random distribution of weighted residuals. See details on the instrument settings for different kinds of fluorophores in Table S3.

## ***In cellulo experiments***

### **Cell culture**

CT-26 cells were cultured in DMEM medium (Gibco, Life Technologies, USA) supplemented with 10 % of fetal calf serum (Gibco) and 100 U/mL penicillin streptomycin mixture (Gibco) and maintained in humidified atmosphere at 37 °C and 5% CO<sub>2</sub>. RPE-1 cells were cultured in DMEM-F12 medium (Gibco, Life Technologies, USA) supplemented with 10 % of fetal calf serum (Gibco) and 100 U/mL penicillin streptomycin mixture (Gibco) and maintained in humidified atmosphere at 37 °C and 5% CO<sub>2</sub>.

### **48 h cytotoxicity assay**

Cells were seeded in triplicate plates in 96-well plates at a 4,000 cells/well density in 100 µL and were incubated at 37 °C, 5% CO<sub>2</sub> for 24 h. The medium was replaced with increasing concentrations of test compounds in 100 µL of fresh medium and cells were incubated at 37 °C, 5% CO<sub>2</sub> for 48 h. The medium was replaced with 100 µL of fresh medium containing 0.2 mg/mL of resazurin. After 4 h of incubation at 37 °C, 5% CO<sub>2</sub>, the fluorescence intensity of resorufin was read at 590 nm with an excitation wavelength of 540 nm using a SpectraMaxM5 microplate reader (Molecular Devices). Data were fitted using Graphpad Prism V6.07 Software. Fluorescence intensities (Y values) were plotted against compounds concentrations (X values). X values were transformed in log(X) values. Y values were normalized by setting 100% cell viability for the highest fluorescence intensity and 0% cell viability for the lowest fluorescence intensity in each data set. IC<sub>50</sub> were calculated by non-linear regression using the algorithm “log(inhibitor) vs. normalized response”.

### **Phototoxicity assay**

Cells were seeded in triplicate plates in 96-well plates at a 4,000 cells/well density in 100 µL and were incubated at 37 °C, 5% CO<sub>2</sub> for 24 h. The medium was replaced with increasing concentrations of test compounds in 100 µL of fresh medium and cells were incubated at 37 °C, 5% CO<sub>2</sub> for 4 h. The medium was then replaced with fresh medium and wells were individually irradiated at 595 nm for 2 h (22,47 J/cm<sup>2</sup>) using a LUMOS-BIO photoreactor (Atlas Photonics). Control plates were kept in the dark in a non-CO<sub>2</sub> incubator for 10 min or 2h. Cells were then incubated for an additional 44 h at 37 °C, 5% CO<sub>2</sub>. The medium was replaced with 100 µL of fresh medium containing 0.2 mg/mL of resazurin. After 4 h of

incubation at 37 °C, 5% CO<sub>2</sub>, the fluorescence intensity of resorufin was read at 590 nm with an excitation wavelength of 540 nm using a SpectraMaxM5 microplate reader (Molecular Devices). Data were fitted using Graphpad Prism V6.07 Software. Fluorescence intensities (Y values) were plotted against compounds concentrations (X values). X values were transformed in log(X) values. Y values were normalized by setting 100% cell viability for the highest fluorescence intensity and 0% cell viability for the lowest fluorescence intensity in each data set. IC<sub>50</sub> were calculated by non-linear regression using the algorithm “log(inhibitor) vs. normalized response”.

### **Cellular uptake**

5×10<sup>6</sup> CT26 cells were seeded in 10-cm petri dishes (10 mL/dish) and were incubated at 37 °C, 5% CO<sub>2</sub>. The next day, the medium was replaced with 5 μM of complexes dilution in 10 mL of culture medium and the dishes were incubated for 4 hours at 37 °C, 5% CO<sub>2</sub>. Cells were washed three times with cold PBS, trypsinized, harvested, and a 10 μL aliquot of each cell suspension was sampled for accurate counting. The cell suspensions were centrifuged, and the supernatant was discarded. The pellets were digested in 100 μL of 70% HNO<sub>3</sub> at 65 °C for 24 h and then diluted in 5 mL of MilliQ water (final HNO<sub>3</sub> concentration: 1.4 %). The Ru content in each sample was determined by ICP-MS. The amount of Ru detected in the digests was transformed from ppb into ng and normalized using the total number of cells digested. Experiments were performed in triplicate.

### ***In vivo* studies**

#### **Animals**

This study was carried out in accordance with EU regulations and approved by the Ethical Commission of the faculty of Pharmaceutical and Biological Sciences Paris-Descartes (agreement number: E-75-06-02).

8-weeks old Balb/c mice were provided by Janvier Lab and housed with food and water supplied *ad libitum* in a 12-hour day/night cycle.

#### **Formulation in polysorbate 80**

1.74 mmol of the test compound and 50 mg of polysorbate 80 were dissolved in 5 mL of anhydrous ethanol. The solution was concentrated to dryness under vacuum in a round-bottom flask to yield an orange viscous film that was further dried at 40 °C under vacuum for 10 min. The film was then rehydrated in 5 mL of PBS (Gibco). The orange solution was finally sterile-filtered on a 0.2 µm cellulose-acetate membrane (Corning®). Test compound's concentration was determined by measuring the absorbance of the solution after dilution in acetonitrile at 450 nm in quartz cuvettes using a SpectraMaxM5 (Molecular Devices) spectrophotometer.

### **[Ru(bphen)<sub>2</sub>dmbipyMal]Cl<sub>2</sub> conjugation to albumin kinetic**

A solution of complex **2** (0.2 mg/mL) formulated either in 5% ethanol in PBS (Gibco) or 1% polysorbate 80 in PBS (Gibco) was added to a solution of bovine serum albumin (BSA) (120 mg/mL) in PBS (Gibco). The mixture was shaken at 450 rpm at 37 °C in a Thermomixer (Eppendorf). At each time point, 300 µL of the mixture was added to 800 µL of CH<sub>3</sub>CN. The mixture was vortexed for 5 s and then centrifugated for 5 min at 10,000 g. Absorbance of the supernatants were recorded at 450 nm in quartz cuvette using a SpectraMaxM5 (Molecular Devices) spectrophotometer. The experiment was performed in triplicates. %conversion was calculated using the following formula:

$$\%conversion = \frac{Abs_{t_0} - Abs_t}{Abs_{t_0}}$$

### **Biodistribution study**

15 10-week-old BALB/c mice were implanted subcutaneously in both flanks with two CT-26 tumor fragments with a diameter of 1 mm, previously extracted from donor mice.

After 8 days, mice were randomly divided into 5 groups and injected intravenously in the caudal vein with 2 µmol/kg of test compound formulated in 1% polysorbate 80 in PBS (complex **1** and **2**) or 5% glucose in water (complex **3**). After 1 min, 30 min, 1 h, 6 h and 24 h, mice were sacrificed and relevant organs including blood, tumors, liver, kidneys, intestine, lungs, and brain were harvested and weighed. Organs were then digested in 70% nitric acid at 65 °C for 24 h. Digests were diluted 100 times in 1% HCl and Ru contents were determined by ICP-MS. The amount of Ru detected in organ digests was transformed from ppb into ng of Ru and expressed as a % of the injected dose/g of organs. Tumor Ru contents are presented as an average of left and right tumors Ru content. Kinetic parameters were calculated using

Graphpad Prism V6.07 Software by fitting the blood Ru content in function of time a two-phase exponential decay equation. The 48 h biodistribution experiment was performed similarly, but complexes **1-3** were all formulated in 5% glucose in water for comparison purposes. Three mice were used for each compound.

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### Supporting Information

Synthesis scheme (Scheme S1). <sup>1</sup>H, <sup>13</sup>C NMR spectra and HRMS (Fig. S1-S10). Complex-albumin additional figures (Fig. S11-S22). Binding kinetic of complex **2** formulated in polysorbate 80 or ethanol (Fig. S23). Tables containing fluorescence lifetime parameters (Tables S1, S2, S3).

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Table of Contents Graphic: a maleimide-functionalized photosensitizer binds covalently and non-covalently with the circulating albumin following intravenous injection.

