

Undissociable chemically cross-linked and single-chain gonadotropins

Thi Mong Diep Nguyen, Danièle Klett, Yves Combarnous

To cite this version:

Thi Mong Diep Nguyen, Danièle Klett, Yves Combarnous. Undissociable chemically cross-linked and single-chain gonadotropins. Theriogenology, 2023, 198, pp.250-255. $10.1016/j.theriogenology.2022.12.034$. $\,$ hal-04206790 $\,$

HAL Id: hal-04206790 <https://hal.science/hal-04206790v1>

Submitted on 8 Jan 2025

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

[Distributed under a Creative Commons Attribution - NonCommercial 4.0 International License](http://creativecommons.org/licenses/by-nc/4.0/)

Clean 1

Undissociable chemically cross-linked and single-chain gonadotropins

- 2 Thi Mong Diep NGUYEN^{1,2}, Danièle KLETT¹ and Yves COMBARNOUS¹
- 1. INRAe, CNRS Unit of Reproductive Physiology and Behaviour, 37380 Nouzilly (France)
- 2. Faculty of Natural Sciences, Quy Nhon University, Quy Nhon 820000 (Vietnam)
-

Abstract

- Undissociable gonadotropins can be obtained either by chemical cross-linking of the natural heterodimeric
- hormones or by expressing recombinant single-chain molecules through the fusion of their α and β polypeptide
- sequences. These undissociable hormones are not more active than their natural heterodimeric counterparts
- indicating that the β-subunit seatbelt embracing the α-subunit ensures the αβ heterodimer stability in
- physiological conditions. The main interests of single-chain gonadotropins are that 1/ only one single plasmid is
- required to produce an active recombinant hormone, 2/ the two subunits' domains are constantly present in equal
- amounts and 3/ they remain in close proximity even at low concentration for forming the hormone bioactive 3D
- structure. These undissociable gonadotropins have been shown to exhibit excellent stability and activity but they
- have not yet been commercialized probably because of immunogenicity risks and cost of production.
- Nevertheless, they might be used as a basis for the development of chemically simpler and cheaper ligands of
- LH and FSH receptors.
-

Introduction

- It has been now known for almost sixty years that ovine Luteinizing Hormone (oLH), also previously named
- 21 Interstitial Cell Stimulating Hormone (ICSH) in some old papers, is a ~30kD dimer consisting of two non-
- covalently associated 15kD « monomers » [1]. These monomers were later found to be two dissimilar subunits
- 23 [2], initially named C1 and C2 and later α and β . Both α and β subunits are devoid of biological activity when
- alone but recover it when recombined [3]. It was then demonstrated that one of the subunits was common to
- Thyroid-Stimulating Hormone (TSH) and LH in the bovine species [4, 5]. However, the common human α-
- 26 subunit from human Chorionic Gonadotropin (hCG α) was found to be able to recombine with either the bovine
- TSHβ-subunit or the bovine LHβ-subunit to give active TSH or LH heterodimers respectively [6]. Subsequent
- studies in 1970-72 showed that the pituitary and chorionic glycoprotein hormone α and β subunits from different
- mammalian species can recombine into potent chimeric heterodimers [7-11].
- In the present short review, we will present the quaternary structure of gonadotropins, and address the
- consequences of the chemical linkage of their subunits or, more recently, the covalent association of their
- subunits in single-chain recombinant molecules. Finally, we will consider the present and potential advantages
- and drawbacks of these gonadotropins for clinical or veterinary usage.

Quaternary structure of glycoprotein hormones (GPHs)

- The quaternary structure of glycoprotein hormones is accompanied by changes in the secondary and tertiary
- structures of their constitutive subunits. The changes in their secondary structures have been initially followed by
- circular dichroism (CD) [12-18]. Only ~50% of the sequences in glycoprotein hormone were found by CD to
- adopt a secondary periodic structure (essentially β-sheet and a mere 4% as α-helix), the rest of it being in a non-
- periodic structure. Their tertiary structures were studied essentially by specific group chemical modifications (for
- example [19-24]) or by analyzing their epitopes at their surface (for example [25-29]).
- The dynamic of the three-dimensional structure of glycoprotein hormones has been studied using nuclear
- magnetic resonance (NMR) [30-33] and isotopic hydrogen exchange (HX) [34]. Only much later, the crystal 3D
- structure of hCG was disclosed by two independent groups [35, 36]. The most unexpected feature in this
- 44 structure was the β-subunit peptide (β CTP) embracing its α -subunit counterpart in the heterodimer, forming a
- « seatbelt » that is fastened by a single intra-β disulfide bridge between Cysβ26 and Cysβ110. Another
- 46 outstanding feature of these crystal structures is the presence of one three-bridge cystine knot in the α and β
- 47 subunits [37]. Interestingly, these knots already exist in the α and β-subunit's glycoprotein ancestors,
- respectively, named GPA2 and GPB5 [38]. Moreover, the GPB5 possesses a sequence shorter than the
- β-subunits and, most importantly, does not possess the seatbelt sequence, so it does not naturally form a stable
- heterodimer with GPA2 [39, 40].
- The αβ heterodimeric non-covalent association is mandatory for allowing the expression of the bioactivity
- specified by the identity of the β-subunit [3, 6, 8-10, 41, 42]. Nevertheless, the specific β-subunits do not express
- any biological activity when isolated, and their association with the α-subunit, common to all glycoprotein
- hormones in each species, is indispensable for expressing gonadotropic (LH, FSH) or thyrotropic (TSH) activity.
- Likewise, the isolated α-subunit which is synthesized in excess relative to the β-subunit does not exhibit any
- gonadotropic or thyrotropic activity. Nevertheless, it has been shown to exert lactogen activity during embryonic
- pituitary development [43, 44] as well as in the human placenta [45]. In contrast to the α-subunits combined with
- β-subunits, the free α-subunit has been shown to bear an O-glycan chain at Thr43 in the bovine pituitary [46]. It
- is unclear whether this modification is the cause or the consequence of its non-association with a β-subunit [47].
- Moreover, it is still unclear whether this O-glycan plays a role in the free alpha lactogenic activity [44].

Consequences of chemical linkage of glycoprotein hormone subunits

- Since glycoprotein hormones are heterodimeric, it is interesting to question the functional importance of the non-
- covalent association of their subunits. Our first guess in the 1970s' was that these hormones could act like the
- cholera toxin, with one subunit allowing membrane binding and the other alone being transferred into the target
- cells [48]. In such a case, the subunits must dissociate for acting. It was thus interesting to test whether an
- undissociable glycoprotein hormone lost its bioactivity. Alternatively, considering that the hormone dissociation
- could play a role in its elimination from blood, it was interesting to test whether an undissociable glycoprotein
- hormone exhibited enhanced *in vivo* bioactivity. We cross-linked the porcine LH subunits to answer these
- questions using water-soluble carbodiimide EDC treatment [49]. A porcine LH derivative that was undissociable
- under urea, guanidinium, low pH, or high pH treatments was obtained and analyzed. Its in vivo bioactivity was
- unchanged, indicating that it does not act through a cholera toxin-like mechanism and does not exhibit a longer
- half-life in the blood. The same EDC treatment applied to ovine LH led to multiple cross-links and a substantial
- 73 loss of bioactivity, probably because of the derivatization of multiple NH₃⁺ and COO⁻ groups and/or
- rigidification of the hormone structure [50]. The difference between porcine and ovine LH is due to the absence
- of Lys residues in pLHβ so that only α-LysεNH-CO-Glu/Asp-β covalent amide bridge(s) can be formed and not
- the reverse as in ovine LH. This procedure was also used for bLH, bTSH, and hCG with 0 to 80% activity loss.
- 77 These losses were due to COO–and/or NH₃⁺ group derivatization and not to the undissociability of hormone
- structures [51].
- Are heterodimeric quaternary structures already present in subunits' ancestors GPA2 and GPB5? Recombinant
- GPA2 and GPB5 form heterodimers only upon reaction with a bifunctional crosslinker and not in its absence
- [38, 40]. Nevertheless, the recombinant GPB5 expressed alone in a baculovirus/insect cell system has been
- shown to form homodimers [40]. Thus, it is likely that these molecules do not form stable heterodimers in
- contrast to glycoprotein hormone subunits because of the absence of the seatbelt sequence in GPB5.
- Artificial cross-linked GPA2/GPB5 heterodimers have been shown to exhibit TSH-like activity, and the
- molecule has been named « thyrostimulin » [39]. The reality of GPA2/GPB5 heterodimer formation *in vivo* is
- 86 still a matter of debate in all animal species. Considering its weak stability, it is likely that GPA2 and GPB exert
- their still-unknown functions mainly as monomers or maybe as a heterodimer at a short range, i.e., as a paracrine
- factor [52].
-

The emergence of recombinant single-chain glycoprotein hormones

- Although chemically cross-linked glycoprotein hormones provided interesting information, they were not easy to prepare. Most importantly, they were found not to be more or less active than the unmodified natural hormones.
- The idea of synthesizing recombinant single-chain glycoprotein hormones emerged from studies dealing with the
- enhancement of the in vivo FSH half-life through the fusion of the hCGβ CTP at the FSHβ C-terminus [53-55].
- It was later shown that fusion of the hCGβ CTP at the hα subunit N-terminus also increased the in vivo
- bioactivity of heterodimers comprising various β-subunits but was deleterious when fused at the hα subunit
- C-terminus [56]. On this basis, it appeared reasonable to fuse the hα sequence downstream of the glycoprotein
- hormone β-subunits to get single-chain βα hormones. This was first done using the hCG subunits [57-59] and the
- hCGβ subunit CTP extension could act as a hinge between the two. Indeed, with hFSH subunits, introducing a
- hCGβ CTP linker between the β- and α-subunit led to a 3-fold increase in its expression with retention of its *in*
- *vitro* biological activity [60]. Likewise, with hLH or hTSH subunits, the presence of the hCGβ CTP as a linker
- between the β and α subunits had a favorable effect on their expression and did not alter their *in vitro*
- bioactivities [61, 62]. Various single-chain glycoprotein hormones (LH, FSH, TSH, CG) have now been
- obtained in many species and characterized [63-71]. It has been shown that the hCGβ CTP linker in hFSH can be
- replaced by a different sequence including two N-glycan chains. This single-chain « hFSH2N » exhibited in vivo
- activity thanks to its prolonged half-life in blood circulation [72]. Nevertheless, in this paper, the authors have
- not tested a similar single-chain hFSH with no N-glycans in the linker. It is therefore not possible to distinguish
- the importance of covalent subunits linkage relative to that of the supplementary N-glycans.
- Multidomain single-chain molecules comprising several β-subunit sequences fused with one α sequence were
- also produced and characterized [63, 73, 74]. In this case, the two activities determined by the two β-subunits are
- expressed by this complex molecule. This dual activity can be due to either a static or dynamic sharing of the
- sole α-subunit by two fused β-subunits (figure 1). A dynamic interconversion would be dependent on the redox
- 113 potential which affects the CysS-SCys \leftrightarrow 2 CysSH equilibrium controlling the seatbelt opening/fastening [75].
- Moreover, single-chain molecules comprising the βα sequence fused to that of the corresponding receptor were
- also expressed [76-78], leading to the receptor's sustained stimulation. Further studies in this direction allowed
- new insights into the structure-function relationships of glycoprotein hormones [79-83] but are outside the scope
- of the present review.
-

Scaling-up of single-chain glycoprotein hormones production

- *In vitro* cell culture production. Different CTPs have been used to optimize the heterodimeric and single-chain
- 121 hormone secretions [84-86]. The human hCGβ CTP has been the most frequently used linker, but the equine
- LH/CGβ CTP was also used in some instances [87]. It is interesting to point out that the pseudo-CTP sequence,
- downstream of the bovine LHβ stop codon also enhanced by about 3-fold the rate and level of secretion from the
- transfected cells [84]. This bovine pseudo-CTPβ or cryptic-CTPβ did so despite it contains no O-glycosylated
- Thr and/or Ser in contrast to the genuine hCGβ or eLH/CGβ CTPs. Such a linker without glycosylation might be
- modified to introduce N-glycosylation sites since N-glycosylations are more manageable to synthesize in
- classical CHO or HEK cells than O-glycosylations. The choice of cells should also be primordial to get the
- highest possible quantities of secreted glycoprotein hormones. Cell lines such as expiCHO and expiHEK
- harboring an episomal expression system for recombinant protein production are well suited for this purpose [88, 89].
- Production in rabbit milk. Recombinant single-chain eLH/CG could be expressed at a concentration of 21.7 mg/L in rabbit milk [83] and exhibited both *in vitro* LH and FSH bioactivities as expected. However, it showed 133 an extremely rapid clearance $(t_{1/2} \sim 10 \text{ min})$, which explains its lack of *in vivo* bioactivities. Likely, the glycosyl 134 sidechains introduced by the mammary gland in the $\beta\alpha$ polypeptide chain are recognized by the recipients' macrophages, which rapidly eliminate the hormone from blood circulation [90]. Another difficulty is that if the hormone could leak toward blood, it would alter reproduction and consequently impede milk production in the rabbit. At present, this approach does not seem the most promising for the production of single-chain glycoprotein hormones.
-

Production in *Pichia*/*Spodoptera* system Several groups have produced single-chain glycoprotein hormones [64, 65, 70, 71] in insect cells such as the *Spodoptera frugiperda* cell line using recombinant *Pichia pastoris* baculovirus for infection. Although these recombinant single-chain hormones were active *in vitro*, they were not *in vivo,* most probably because of their rapid removal from blood.

In vivo **biological properties of recombinant single-chain glycoprotein hormones**

Although sc-gonadotropins have been shown to be biologically active *in vitro*, it remains a great challenge to use them *in vivo*.

- Clinical uses: Recombinant FSHs with hCG CTP at their β-subunit C-terminus or as a linker between their β and
- α have been tested in humans and animals [91-95]. Nevertheless, single-chain gonadotropins have not yet been
- commercialized for clinical use in humans [96] probably because of potential immunogenicity of these
- molecules. Indeed, although the hCG CTP is of human origin, it cannot be excluded that the junctions
- respectively between hFSHβ and hCGβ-CTP and/or between hCGβ-CTP and hα can be immunogenic as they do
- not exist in the natural hormones (figure 2). Alternative flexible, non-immunogenic sequences [97, 98] might be
- used in the future to link the two glycoprotein hormone subunits.
- Another limitation of the development of single-chain gonadotropins in Humans is that they do not offer superior
- biological properties relative to their heterodimeric counterparts. Their advantage lies in their production: The
- 157 use of a single plasmid allows 1/ strictly balanced 1 :1 synthesis of the α and β domains (avoiding the useless
- synthesis of a free subunit in excess) and 2/ their close proximity in the single-chain molecule (allowing their
- highly efficient association).
- Veterinary uses. For the time being, there are only a few available commercial single-chain glycoprotein
- hormones for animals [93, 99, 100], as it is very expensive to develop recombinant gonadotropins specifically
- for the veterinary market. Nevertheless, a patent for single-chain equine choriogonadotropin derivatives has been
- recently published [101]. These molecules comprise either one or several additional eCGβ carboxy-terminal
- peptide(s) (CTP) including 12 O-glycosylation sites and/or one or several amino-terminal peptide(s) (NTP) each
- including four N-glycosylation sites. One of these recombinant single-chain eCG derivatives is expected to be
- commercialized soon in Argentina and other Latin American countries.
- The human and animal undissociable gonadotropins have thus been shown to exhibit excellent stability and
- activity but their commercialization is still very limited probably because of immunogenicity risks and cost of production.
-

Conclusion

- Recombinant single-chain gonadotropins or chemically-linked undissociable gonadotropins are highly active but
- not more than their natural heterodimeric counterparts. This indicates that the β-subunit seatbelt embracing the α-
- 174 subunit in natural gonadotropins ensures the $\alpha\beta$ heterodimer stability in physiological conditions. The main
- interests of single-chain gonadotropins are that 1/ only one single plasmid is required to produce an active
- recombinant hormone, 2/ the two subunits' domains are constantly present in equal amounts and 3/ they remain
- in close proximity even at low concentration for forming the hormone bioactive 3D structure.

Fundings

Research is supported by Vingroup Innovation Foundation (VINIF) in project code VINIF.2020.DA05. It is also 181 supported by INRAe and CNRS (France).

Declaration of interest

- The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of
- the research reported.

References

- [1] Li CH, Starman B. Molecular Weight of Sheep Pituitary Interstitial Cell-Stimulating Hormone. Nature. 1964;202:291-2.
- [2] Papkoff H, Samy TS. Isolation and partial characterization of the polypeptide chains of ovine interstitial cell-stimulating hormone. Biochim Biophys Acta. 1967;147:175-7.
- [3] De la Llosa P, Jutisz M. Reversible dissociation into subunits and biological activity of ovine luteinizing hormone. Biochim Biophys Acta. 1969;181:426-36.
- 195 [4] Liao TH, Hennen G, Howard SM, Shome B, Pierce JG. Bovine thyrotropin. Countercurrent distribution and a
196 comparison with the isolated subunits of luteinizing hormone. J Biol Chem. 1969:244:6458-67. comparison with the isolated subunits of luteinizing hormone. J Biol Chem. 1969;244:6458-67.
- [5] Liao TH, Pierce JG. The presence of a common type of subunit in bovine thyroid-stimulating and luteinizing hormones. J Biol Chem. 1970;245:3275-81.
- [6] Pierce JG, Bahl OP, Cornell JS, Swaminathan N. Biologically active hormones prepared by recombination of the alpha chain of human chorionic gonadotropin and the hormone-specific chain of bovine thyrotropin or of bovine luteinizing hormone. J Biol Chem. 1971;246:2321-4.
- [7] Reichert LE, Jr., Midgley AR, Jr., Niswender GD, Ward DN. Formation of a hybrid molecule from subunits of human bovine luteinizing hormone. Endocrinology. 1970;87:534-41.
- [8] Reichert LE, Jr. Biological evidence for the subunit structure of human follicle-stimulating hormone. Endocrinology. 1971;89:925-8.
- [9] Ward DN, Reichart LE, Jr., Fitak BA, Nahm HS, Sweeney CM, Neill JD. Isolation and properties of subunits of rat pituitary luteinizing hormone. Biochemistry. 1971;10:1796-802.
- [10] Closset J, Hennen G, Lequin RM. Isolation and properties of human luteinizing hormone subunits. FEBS Lett. 1972;21:325-9.
- [11] Bousfield GR, Liu WK, Ward DN. Hybrids from equine LH: alpha enhances, beta diminishes activity. Mol Cell Endocrinol. 1985;40:69-77.
- [12] Jirgensons B, Ward DN. Circular dichroism of ovine luteinizing hormone and its subunits. Tex Rep Biol Med. 1970;28:553-9.
- [13] Puett D, Nureddin A, Holladay LA. Circular dichroism of human pituitary luteinizing hormone and its
- glycopeptides. Curve resolution and band assignments to the peptide chromophore, aromatic residues, disulfides, and N-acetylated amino sugars. Int J Pept Protein Res. 1976;8:183-91.
- [14] Puett D, Holladay LA. The effects of denaturant and temperature on the circular dichroic spectrum of ovine lutropin-alpha. Int J Pept Protein Res. 1977;10:102-6.
- [15] Strickland TW, Puett D. Circular dichroism of gonadotropin recombinants. Int J Pept Protein Res. 1983;21:374-80.
- [16] Burova T, Lecompte F, Galet C, Monsallier F, Delpech S, Haertle T, et al. Conformational stability and in vitro bioactivity of porcine luteinizing hormone. Mol Cell Endocrinol. 2001;176:129-34.
- [17] Salesse R, Combarnous Y, Brahms S, Garnier J. Conformational study of native and acid-dissociated states of porcine lutropin (pLH) by vacuum circular dichroism and ultraviolet absorption spectroscopy. Arch Biochem Biophys. 1981;209:480-5.
- [18] Combarnous Y, Maghuin-Rogister G. Luteinizing hormone. I. Circular dichroism and spectrophotometric titration of porcine and bovine hormones and of their alpha and beta subunits. Eur J Biochem. 1974;42:7-12.
- [19] Ward DN, Cofey JA. Analysis of the Acetyl Groups in Ovine Luteinizing Hormone by Gas Chromatography. Biochemistry. 1964;3:1575-7.
- [20] Liu WK, Yang KP, Nakagawa Y, Ward DN. The role of the amino group in subunit association and
- receptor site interaction for ovine luteinizing hormone as studied by acylation. J Biol Chem. 1974;249:5544-50.
- 232 [21] Liu WK, Esfahani M, Ward DN. Guanidination of ovine luteinizing hormone and effects on activity.
233 Endocr Res Commun. 1975:2:47-63. Endocr Res Commun. 1975;2:47-63.
- [22] Liu WK, Ward DN. Preparation of lutropin with acetyl or acetimidinyl substituents on the amino groups of the beta-subunit. Biochim Biophys Acta. 1975;405:522-6.
- [23] Burleigh BD, Liu WK, Ward DN. Reaction of tetranitromethane with lutropin, oxytocin, and vasopressin. J Biol Chem. 1976;251:308-15.
- 238 [24] Combarnous Y, Maghuin-Rogister G. Luteinizing hormone. 2. Relative reactivities of tyrosyl residues of the porcine hormone towards iodination. Eur J Biochem. 1974:42:13-9. the porcine hormone towards iodination. Eur J Biochem. 1974;42:13-9.
- [25] Chopineau M, Maurel MC, Combarnous Y, Durand P. Topography of equine chorionic gonadotropin 241 epitopes relative to the luteinizing hormone and follicle-stimulating hormone receptor interaction sites. Mol Cell
242 Endocrinol. 1993:92:229-39.
- Endocrinol. 1993;92:229-39.
- [26] Dirnhofer S, Lechner O, Madersbacher S, Klieber R, de Leeuw R, Wick G, et al. Free alpha subunit of human chorionic gonadotrophin: molecular basis of immunologically and biologically active domains. J
- Endocrinol. 1994;140:145-54.
- [27] Moyle WR, Ehrlich PH, Canfield RE. Use of monoclonal antibodies to subunits of human chorionic 247 gonadotropin to examine the orientation of the hormone in its complex with receptor. Proc Natl Acad Sci U S A.
248 1982:79:2245-9. 1982;79:2245-9.
- [28] Dighe RR, Moudgal NR. Use of alpha- and beta-subunit specific antibodies in studying interaction of hCG with Leydig cell receptors. Arch Biochem Biophys. 1983;225:490-9.
- [29] Bidart JM, Troalen F, Salesse R, Bousfield GR, Bohuon CJ, Bellet DH. Topographic antigenic determinants recognized by monoclonal antibodies on human choriogonadotropin beta-subunit. J Biol Chem. 1987;262:8551- 6.
- [30] Frankenne F, Maghuin-Rogister G, Birdsall B, Roberts GC. H-NMR studies of the histidine residues of human choriogonadotropin and its alpha- and beta-subunits. FEBS Lett. 1983;151:197-200.
- 256 [31] De Beer T, Van Zuylen CW, Leeflang BR, Hard K, Boelens R, Kaptein R, et al. NMR studies of the free
257 alpha subunit of human chorionic gonadotropin. Structural influences of N-glycosylation and the beta subunit alpha subunit of human chorionic gonadotropin. Structural influences of N-glycosylation and the beta subunit on the conformation of the alpha subunit. Eur J Biochem. 1996;241:229-42.
- [32] Erbel PJ, Karimi-Nejad Y, De Beer T, Boelens R, Kamerling JP, Vliegenthart JF. Solution structure of the alpha-subunit of human chorionic gonadotropin. Eur J Biochem. 1999;260:490-8.
- [33] Parsons TF, Pierce JG. Studies of the histidine residues of human and bovine glycoprotein hormones by nuclear magnetic resonance. Int J Pept Protein Res. 1983;21:522-35.
- [34] Combarnous Y, Nabedryk-Viala E. Porcine lutropin: a study of the association of its subunits by hydrogen-deuterium exchange. Biochem Biophys Res Commun. 1978;84:1119-24.
- [35] Lapthorn AJ, Harris DC, Littlejohn A, Lustbader JW, Canfield RE, Machin KJ, et al. Crystal structure of human chorionic gonadotropin. Nature. 1994;369:455-61.
- [36] Wu H, Lustbader JW, Liu Y, Canfield RE, Hendrickson WA. Structure of human chorionic gonadotropin at 2.6 A resolution from MAD analysis of the selenomethionyl protein. Structure. 1994;2:545-58.
- [37] Alvarez E, Cahoreau C, Combarnous Y. Comparative structure analyses of cystine knot-containing
-
- 270 molecules with eight aminoacyl ring including glycoprotein hormones (GPH) alpha and beta subunits and GPH-
271 related A2 (GPA2) and B5 (GPB5) molecules. Reprod Biol Endocrinol. 2009;7:90. related A2 (GPA2) and B5 (GPB5) molecules. Reprod Biol Endocrinol. 2009;7:90.
- [38] Hsu SY, Nakabayashi K, Bhalla A. Evolution of glycoprotein hormone subunit genes in bilateral metazoa: identification of two novel human glycoprotein hormone subunit family genes, GPA2 and GPB5. Mol
- Endocrinol. 2002;16:1538-51.
- [39] Nakabayashi K, Matsumi H, Bhalla A, Bae J, Mosselman S, Hsu SY, et al. Thyrostimulin, a heterodimer of
- 276 two new human glycoprotein hormone subunits, activates the thyroid-stimulating hormone receptor. J Clin 277 Invest. 2002;109:1445-52.
- Invest. $2002;109:1445-52$.
- [40] Cahoreau C, Klett D, Combarnous Y. Structure-function relationships of glycoprotein hormones and their subunits' ancestors. Front Endocrinol (Lausanne). 2015;6:26.
- [41] Gospodarowicz D. Purification and physicochemical properties of the pregnant mare serum gonadotropin (PMSG). Endocrinology. 1972;91:101-6.
- [42] Combarnous Y. Molecular basis of the specificity of binding of glycoprotein hormones to their receptors. Endocr Rev. 1992;13:670-91.
- [43] Begeot M, Hemming FJ, Dubois PM, Combarnous Y, Dubois MP, Aubert ML. Induction of pituitary lactotrope differentiation by luteinizing hormone alpha subunit. Science. 1984;226:566-8.
- [44] Chabot V, Magallon T, Taragnat C, Combarnous Y. Two free isoforms of ovine glycoprotein hormone alpha-subunit strongly differ in their ability to stimulate prolactin release from foetal pituitaries. J Endocrinol. 2000;164:287-97.
- [45] Blithe DL, Richards RG, Skarulis MC. Free alpha molecules from pregnancy stimulate secretion of
- prolactin from human decidual cells: a novel function for free alpha in pregnancy. Endocrinology. 1991;129:2257-9.
- [46] Parsons TF, Bloomfield GA, Pierce JG. Purification of an alternate form of the alpha subunit of the glycoprotein hormones from bovine pituitaries and identification of its O-linked oligosaccharide. J Biol Chem. 1983;258:240-4.
- [47] Blithe DL, Iles RK. The role of glycosylation in regulating the glycoprotein hormone free alpha-subunit and free beta-subunit combination in the extraembryonic coelomic fluid of early pregnancy. Endocrinology. 1995;136:903-10.
- [48] Van Heyningen WE, Van Heyningen S, King CA. The nature and action of cholera toxin. Ciba Found Symp. 1976:73-88.
- [49] Combarnous Y, Hennen G. Luteinizing hormone derivatives with covalently-linked subunits. FEBS Lett. 1974;44:224-8.
- [50] van Dijk S, Ward DN. Chemical cross-linking of porcine luteinizing hormone: location of the cross-link and consequences for stability and biological activity. Endocrinology. 1993;132:534-8.
- [51] Parsons TF, Pierce JG. Biologically active covalently cross-linked glycoprotein hormones and the effects of modification of the COOH-terminal region of their alpha subunits. J Biol Chem. 1979;254:6010-5.
- [52] Querat B. Unconventional Actions of Glycoprotein Hormone Subunits: A Comprehensive Review. Front Endocrinol (Lausanne). 2021;12:731966.
- [53] Fares FA, Suganuma N, Nishimori K, LaPolt PS, Hsueh AJ, Boime I. Design of a long-acting follitropin
- 309 agonist by fusing the C-terminal sequence of the chorionic gonadotropin beta subunit to the follitropin beta
310 subunit. Proc Natl Acad Sci U S A. 1992:89:4304-8. subunit. Proc Natl Acad Sci U S A. 1992;89:4304-8.
- [54] LaPolt PS, Nishimori K, Fares FA, Perlas E, Boime I, Hsueh AJ. Enhanced stimulation of follicle
- 312 maturation and ovulatory potential by long acting follicle-stimulating hormone agonists with extended carboxyl-
313 terminal peptides. Endocrinology. 1992;131:2514-20. terminal peptides. Endocrinology. 1992;131:2514-20.
- [55] Fauser BC, Mannaerts BM, Devroey P, Leader A, Boime I, Baird DT. Advances in recombinant DNA
- technology: corifollitropin alfa, a hybrid molecule with sustained follicle-stimulating activity and reduced
- injection frequency. Hum Reprod Update. 2009;15:309-21.
- [56] Furuhashi M, Shikone T, Fares FA, Sugahara T, Hsueh AJ, Boime I. Fusing the carboxy-terminal peptide of
- 318 the chorionic gonadotropin (CG) beta-subunit to the common alpha-subunit: retention of O-linked glycosylation
319 and enhanced in vivo bioactivity of chimeric human CG. Mol Endocrinol. 1995:9:54-63. and enhanced in vivo bioactivity of chimeric human CG. Mol Endocrinol. 1995;9:54-63.
- [57] Narayan P, Wu C, Puett D. Functional expression of yoked human chorionic gonadotropin in baculovirus-infected insect cells. Mol Endocrinol. 1995;9:1720-6.
- [58] Sugahara T, Pixley MR, Minami S, Perlas E, Ben-Menahem D, Hsueh AJ, et al. Biosynthesis of a
- biologically active single peptide chain containing the human common alpha and chorionic gonadotropin beta subunits in tandem. Proc Natl Acad Sci U S A. 1995;92:2041-5.
- [59] Ben-Menahem D, Boime I. Converting heterodimeric gonadotropins to genetically linked single chains: new approaches to structure activity relationships and analogue design. Trends Endocrinol Metab. 1996;7:100-5.
-
- 327 [60] Sugahara T, Sato A, Kudo M, Ben-Menahem D, Pixley MR, Hsueh AJ, et al. Expression of biologically
328 active fusion genes encoding the common alpha subunit and the follicle-stimulating hormone beta subunit. Re active fusion genes encoding the common alpha subunit and the follicle-stimulating hormone beta subunit. Role of a linker sequence. J Biol Chem. 1996;271:10445-8.
- 330 [61] Garcia-Campayo V, Sato A, Hirsch B, Sugahara T, Muyan M, Hsueh AJ, et al. Design of stable biologically active recombinant lutropin analogs. Nat Biotechnol. 1997;15:663-7. active recombinant lutropin analogs. Nat Biotechnol. 1997;15:663-7.
- [62] Fares FA, Yamabe S, Ben-Menahem D, Pixley M, Hsueh AJ, Boime I. Conversion of thyrotropin heterodimer to a biologically active single-chain. Endocrinology. 1998;139:2459-64.
- [63] Garcia-Campayo V, Kumar TR, Boime I. Thyrotropin, follitropin, and chorionic gonadotropin expressed as a single multifunctional unit reveal remarkable permissiveness in receptor-ligand interactions. Endocrinology. 2002;143:3773-8.
- [64] Fidler AE, Lin JS, Lun S, Ng Chie W, Western A, Stent V, et al. Production of biologically active tethered ovine FSHbetaalpha by the methylotrophic yeast Pichia pastoris. J Mol Endocrinol. 2003;30:213-25.
- 339 [65] Kasuto H, Levavi-Sivan B. Production of biologically active tethered tilapia LHbetaalpha by the methylotrophic yeast Pichia pastoris. Gen Comp Endocrinol. 2005;140:222-32. methylotrophic yeast Pichia pastoris. Gen Comp Endocrinol. 2005;140:222-32.
- [66] Rayalam S, Eizenstat LD, Hoenig M, Ferguson DC. Cloning and sequencing of feline thyrotropin (fTSH): heterodimeric and yoked constructs. Domest Anim Endocrinol. 2006;30:203-17.
- [67] Jablonka-Shariff A, Roser JF, Bousfield GR, Wolfe MW, Sibley LE, Colgin M, et al. Expression and bioactivity of a single chain recombinant equine luteinizing hormone (reLH). Theriogenology. 2007;67:311-20.
-
- [68] Hollander-Cohen L, Golan M, Aizen J, Shpilman M, Levavi-Sivan B. Characterization of carp gonadotropins: Structure, annual profile, and carp and zebrafish pituitary topographic organization. Gen Comp Endocrinol. 2018;264:28-38.
- [69] Burow S, Fontaine R, von Krogh K, Mayer I, Nourizadeh-Lillabadi R, Hollander-Cohen L, et al. Medaka
- follicle-stimulating hormone (Fsh) and luteinizing hormone (Lh): Developmental profiles of pituitary protein and gene expression levels. Gen Comp Endocrinol. 2019;272:93-108.
- 351 [70] Blanchard V, Gadkari RA, George AV, Roy S, Gerwig GJ, Leeflang BR, et al. High-level expression of
352 biologically active glycoprotein hormones in Pichia pastoris strains-selection of strain GS115, and not X-33. biologically active glycoprotein hormones in Pichia pastoris strains--selection of strain GS115, and not X-33, for
- the production of biologically active N-glycosylated 15N-labeled phCG. Glycoconj J. 2008;25:245-57.
- [71] Legardinier S, Duonor-Cerutti M, Devauchelle G, Combarnous Y, Cahoreau C. Biological activities of
- 355 recombinant equine luteinizing hormone/chorionic gonadotropin (eLH/CG) expressed in Sf9 and Mimic insect cell lines. J Mol Endocrinol. 2005;34:47-60. cell lines. J Mol Endocrinol. 2005;34:47-60.
- [72] Klein J, Lobel L, Pollak S, Lustbader B, Ogden RT, Sauer MV, et al. Development and characterization of a long-acting recombinant hFSH agonist. Hum Reprod. 2003;18:50-6.
- [73] Ben-Menahem D, Hyde R, Pixley M, Berger P, Boime I. Synthesis of multi-subunit domain gonadotropin complexes: a model for alpha/beta heterodimer formation. Biochemistry. 1999;38:15070-7.
- [74] Garcia-Campayo V, Jablonka-Shariff A, Boime I. A single-chain bifunctional gonadotropin analog is secreted from Chinese hamster ovary cells as two distinct bioactive species. J Biol Chem. 2004;279:44286-93.
- [75] Galet C, Lecompte F, Combarnous Y. Association/dissociation of gonadotropin subunits involves disulfide
- bridge disruption which is influenced by carbohydrate moiety. Biochem Biophys Res Commun. 2004;324:868- 73.
- [76] Narayan P, Wu C, Puett D. Genetic engineering of single-chain gonadotropins and hormone-receptor fusion proteins. Methods. 2000;21:59-66.
- [77] Fralish GB, Narayan P, Puett D. High-level expression of a functional single-chain human chorionic
- gonadotropin-luteinizing hormone receptor ectodomain complex in insect cells. Endocrinology. 2001;142:1517- 24.
- [78] Narayan P, Gray J, Puett D. Yoked complexes of human choriogonadotropin and the lutropin receptor: evidence that monomeric individual subunits are inactive. Mol Endocrinol. 2002;16:2733-45.
- [79] Schubert RL, Narayan P, Puett D. Specificity of cognate ligand-receptor interactions: fusion proteins of
374 human chorionic gonadotropin and the heptahelical receptors for human luteinizing hormone, thyroid-
- human chorionic gonadotropin and the heptahelical receptors for human luteinizing hormone, thyroid-
- stimulating hormone, and follicle-stimulating hormone. Endocrinology. 2003;144:129-37.
- [80] Schubert RL, Puett D. Single-chain human chorionic gonadotropin analogs containing the determinant loop of the beta-subunit linked to the alpha-subunit. J Mol Endocrinol. 2003;31:157-68.
- [81] Garcia-Campayo V, Boime I, Ma X, Daphna-Iken D, Kumar TR. A single-chain tetradomain glycoprotein hormone analog elicits multiple hormone activities in vivo. Biol Reprod. 2005;72:301-8.
- [82] Jablonka-Shariff A, Kumar TR, Eklund J, Comstock A, Boime I. Single-chain, triple-domain gonadotropin 381 analogs with disulfide bond mutations in the alpha-subunit elicit dual follitropin and lutropin activities in vivo.
382 Mol Endocrinol. 2006;20:1437-46. Mol Endocrinol. 2006;20:1437-46.
- [83] Galet C, Le Bourhis CM, Chopineau M, Le Griec G, Perrin A, Magallon T, et al. Expression of a single βα chain protein of equine LH/CG in milk of transgenic rabbits and its biological activity. Molecular and Cellular Endocrinology. 2001;174:31-40.
- [84] Nakav S, Dantes A, Pen S, Chadna-Mohanty P, Braw-Tal R, Amsterdam A, et al. Homologous and heterologous carboxyl terminal peptide (CTP) linker sequences enhance the secretion of bioactive single-chain bovine LH analogs. Exp Clin Endocrinol Diabetes. 2006;114:95-104.
- [85] Galet C, Guillou F, Foulon-Gauze F, Combarnous Y, Chopineau M. The beta104-109 sequence is essential for the secretion of correctly folded single-chain beta alpha horse LH/CG and for its FSH activity. J Endocrinol. 2009;203:167-74.
- [86] Jablonka-Shariff A, Boime I. Secretory trafficking signal encoded in the carboxyl-terminal region of the CGbeta-subunit. Mol Endocrinol. 2009;23:316-23.
- 394 [87] Legardinier S, Poirier JC, Klett D, Combarnous Y, Cahoreau C. Stability and biological activities of heterodimeric and single-chain equine LH/chorionic gonadotropin variants. J Mol Endocrinol. 2008:40:18 heterodimeric and single-chain equine LH/chorionic gonadotropin variants. J Mol Endocrinol. 2008;40:185-98.
- [88] Zhong X, Schwab A, Ma W, Meade CL, Zhou J, D'Antona AM, et al. Large-Scale Transient
-
- 397 ProductionTransient productionin ExpiCHO-S™ with Enhanced N-Galactosylation-SialylationN-
398 Galactosylation-Sialylationand PEI-Based TransfectionTransfection. In: Houen G, editor. Therape Galactosylation-Sialylationand PEI-Based TransfectionTransfection. In: Houen G, editor. Therapeutic
- Antibodies: Methods and Protocols. New York, NY: Springer US; 2022. p. 143-50.
- [89] Jain NK, Barkowski-Clark S, Altman R, Johnson K, Sun F, Zmuda J, et al. A high density CHO-S transient transfection system: Comparison of ExpiCHO and Expi293. Protein Expr Purif. 2017;134:38-46.
- [90] O'Riordan N, Kane M, Joshi L, Hickey RM. Structural and functional characteristics of bovine milk protein glycosylation. Glycobiology. 2014;24:220-36.
- [91] Balen AH, Mulders AG, Fauser BC, Schoot BC, Renier MA, Devroey P, et al. Pharmacodynamics of a
- single low dose of long-acting recombinant follicle-stimulating hormone (FSH-carboxy terminal peptide,
- 406 corifollitropin alfa) in women with World Health Organization group II anovulatory infertility. J Clin Endocrinol
407 Metab. 2004:89:6297-304. Metab. 2004:89:6297-304.
- [92] Devroey P, Fauser BC, Platteau P, Beckers NG, Dhont M, Mannaerts BM. Induction of multiple follicular
- development by a single dose of long-acting recombinant follicle-Stimulating hormone (FSH-CTP,
- corifollitropin alfa) for controlled ovarian stimulation before in vitro fertilization. J Clin Endocrinol Metab.
- 2004;89:2062-70.
- [93] Lemke EP, Adams BM, Jablonka-Shariff A, Boime I, Adams TE. Single-chain human gonadotropin analogs induce follicle development in sheep. J Endocrinol. 2008;196:593-600.
- [94] Loutradis D, Drakakis P, Vlismas A, Antsaklis A. Corifollitropin alfa, a long-acting follicle-stimulating hormone agonist for the treatment of infertility. Curr Opin Investig Drugs. 2009;10:372-80.
- [95] Loutradis D, Vlismas A, Drakakis P. Corifollitropin alfa: a novel long-acting recombinant follicle-stimulating hormone agonist for controlled ovarian stimulation. Womens Health (Lond Engl). 2010;6:655-64.
-
- [96] Lunenfeld B, Bilger W, Longobardi S, Alam V, D'Hooghe T, Sunkara SK. The Development of Gonadotropins for Clinical Use in the Treatment of Infertility. Front Endocrinol (Lausanne). 2019;10:429.
- [97] Van Rosmalen M, Krom M, Merkx M. Tuning the Flexibility of Glycine-Serine Linkers To Allow Rational Design of Multidomain Proteins. Biochemistry. 2017;56:6565-74.
- [98] Arslan M, Karadag M, Onal E, Gelinci E, Cakan-Akdogan G, Kalyoncu S. Effect of non-repetitive linker on in vitro and in vivo properties of an anti-VEGF scFv. Scientific Reports. 2022;12.
-
- 424 [99] Carvalho PD, Hackbart KS, Bender RW, Baez GM, Dresch AR, Guenther JN, et al. Use of a single
425 injection of long-acting recombinant bovine FSH to superovulate Holstein heifers: a preliminary study. injection of long-acting recombinant bovine FSH to superovulate Holstein heifers: a preliminary study. Theriogenology. 2014;82:481-9.
- [100] Rutigliano HM, Adams BM, Jablonka-Shariff A, Boime I, Adams TE. Effect of single-chain ovine gonadotropins with dual activity on ovarian function in sheep. Reproduction. 2014;148:129-36.
- [101] Perez-Saez JM, Busmann LE. Recombinant Chorionic Gonadotropin, procedure for preparation,
- pharmaceutical compositions and use of the same. WO 2021/037384 A1 Patent. Syntex SA Argentina. 2021.
-

Figures

Figure 1 : **Hypothetical structure of multidomain LH + FSH single-chain glycoprotein**

Either the βFSH or the βLH domain embraces the α-domain with its « seatbelt » (sb) sequence to form the structures with

FSH or LH activity, respectively. Depending on the physicochemical conditions, particularly the redox potential (allowing

opening and fastening of the « seatbelt latch » formed by the disulfide bridge between Cysβ26 and Cysβ110). The two

bioactive conformations are expected to be either static or dynamically inter-convertible.

Figure 2: **Schematic structure of single-chain hFSH and potential immunogenic sites (red arrows)**

The fusion sequences between the βFSH and CTP and between the CTP and hα (red arrows) do not exist naturally and are

thus potentially immunogenic. The β-subunit's « seatbelt » (sb) sequence embracing the ^α-subunit, present in the natural

- hormone, is conserved in the single-chain gonadotropin structures.
- sb = seatbelt.