



HAL
open science

Artificial intelligence to improve cytology performance in urothelial carcinoma diagnosis: results from validation phase of the French, multicenter, prospective VISIOCYT1 trial

Thierry Lebret, Xavier Paoletti, Géraldine Pignot, Mathieu Roumiguié, Marc C. Colombel, Laurent Savareux, Grégory Verhoest, Laurent Guy, Jérôme Rigaud, Stéphane de Vergie, et al.

► To cite this version:

Thierry Lebret, Xavier Paoletti, Géraldine Pignot, Mathieu Roumiguié, Marc C. Colombel, et al.. Artificial intelligence to improve cytology performance in urothelial carcinoma diagnosis: results from validation phase of the French, multicenter, prospective VISIOCYT1 trial. *World Journal of Urology*, 2023, 41, pp.2381-2388. 10.1007/s00345-023-04519-4 . hal-04195182

HAL Id: hal-04195182

<https://hal.science/hal-04195182>

Submitted on 24 May 2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License



Artificial intelligence to improve cytology performance in urothelial carcinoma diagnosis: results from validation phase of the French, multicenter, prospective VISIOCYT1 trial

Thierry Le Bret¹ · Xavier Paoletti^{2,3} · Geraldine Pignot⁴ · Mathieu Roumiguié⁵ · Marc Colombel⁶ · Laurent Savareux⁷ · Grégory Verhoest⁸ · Laurent Guy⁹ · Jérôme Rigaud¹⁰ · Stéphane De Vergie¹⁰ · Grégoire Poinas¹¹ · Stéphane Droupy¹² · François Kleinclauss¹³ · Monique Courtade-Saïdi¹⁴ · Eric Piaton¹⁵ · Camelia Radulescu¹⁶ · Nathalie Rioux-Leclercq¹⁷ · Christine Kandel-Aznar¹⁸ · Karine Renaudin^{19,20} · Béatrix Cochand-Priollet²¹ · Yves Allory^{22,23} · Sébastien Nivet²⁴ · Morgan Rouprêt²⁵

Received: 22 March 2023 / Accepted: 1 July 2023 / Published online: 22 July 2023
© The Author(s) 2023

Abstract

Purpose Cytology and cystoscopy, the current gold standard for diagnosing urothelial carcinomas, have limits: cytology has high interobserver variability with moderate or not optimal sensitivity (particularly for low-grade tumors); while cystoscopy is expensive, invasive, and operator dependent. The VISIOCYT1 study assessed the benefit of VisioCyt[®] for diagnosing urothelial carcinoma.

Methods VISIOCYT1 was a French prospective clinical trial conducted in 14 centers. The trial enrolled adults undergoing endoscopy for suspected bladder cancer or to explore the lower urinary tract. Participants were allocated either Group 1: with bladder cancer, i.e., with positive cystoscopy or with negative cystoscopy but positive cytology, or Group 2: without bladder cancer. Before cystoscopy and histopathology, slides were prepared for cytology and the VisioCyt[®] test from urine samples. The diagnostic performance of VisioCyt[®] was assessed using sensitivity (primary objective, 70% lower-bound threshold) and specificity (75% lower-bound threshold). Sensitivity was also assessed by tumor grade and T-staging. VisioCyt[®] and cytology performance were evaluated relative to the histopathological assessments.

Results Between October 2017 and December 2019, 391 participants (170 in Group 1 and 149 in Group 2) were enrolled. VisioCyt[®]'s sensitivity was 80.9% (95% CI 73.9–86.4%) and specificity was 61.8% (95% CI 53.4–69.5%). In high-grade tumors, the sensitivity was 93.7% (95% CI 86.0–97.3%) and in low-grade tumors 66.7% (95% CI 55.2–76.5%). Sensitivity by T-staging, compared to the overall sensitivity, was higher in high-grade tumors and lower in low-grade tumors.

Conclusion VisioCyt[®] is a promising diagnostic tool for urothelial cancers with improved sensitivities for high-grade tumors and notably for low-grade tumors.

Keywords Urothelial · Cancer · Bladder · Artificial intelligence · Deep learning · Markers

Introduction

Bladder cancer (BC) is a heterogeneous disease that includes de novo and recurrent low-grade subtypes with indolent disease evolution, and high-grade subtypes with recurrent, progressive, and potentially lethal outcome. Today, the gold standard for diagnosis and surveillance combines urinary cytology with white-light cystoscopy. If a tumor is suspected by cystoscopy, then final diagnosis and pathological staging

are based on cystoscopy with biopsy [1]. However, cytology and cystoscopy are limited. Cytology, although noninvasive with high specificity, has high interobserver variability [2] with moderate sensitivity, particularly for low-grade tumors [3]. Cystoscopy is expensive, invasive, and operator dependent [1]. It detects papillary lesions with high sensitivity but has difficulties differentiating non-papillary and flat lesions in inflammatory lesions. Furthermore, cystoscopy can induce several potential side effects, including urinary tract infections, dysuria, hematuria, and perforation of the bladder wall [4].

Extended author information available on the last page of the article

These limits have driven the development of new non-invasive diagnosis tools [4–7]. In recent years, the use of artificial intelligence (AI) in medicine has expanded, providing opportunities for early detection and management of diseases [8]. AI is a powerful resource for building reliable models from complex and accumulating medical data using machine learning, algorithms, and artificial neuron networks [9]. AI has been successfully applied to urology for diagnosing, staging, treatment, and monitoring of some genitourinary malignancies [10]. AI has for example been used to interpret diagnostic imaging, histopathology, and genomic annotations.

To improve diagnosis and surveillance of patients with BC mainly non-muscle invasive bladder cancer (NMIBC), VitaDX (VitaDX International, France) developed a digital medical device (VisioCyt[®]) that uses deep learning-based automated image processing to analyze urothelial cell morphology [11]. The VisioCyt[®] device detects bladder tumor cell aspects, in both high- and low-grade tumors cells, in smears from voided urine samples, by analyzing the morphological modifications of cell nuclei. Analysis of urine samples using VisioCyt[®] is a practical, noninvasive alternative either alone or combined with cytology and cystoscopy.

The VISIOCYT1 trial investigated this innovative approach for detecting BC. The trial comprised an initial phase to develop and evaluate the algorithm and a second to validate the clinical performances of the VisioCyt[®] diagnostic test. The results of the first stage have been reported [11]. We herein report the results of the second phase.

Methods

The study design, slide preparation, and cell annotation for developing the algorithm, and the automated computed processing have been published [11]. The algorithm was developed from a database of thousands of urothelial cells selected and labelled, by pathologists and cytotechnicians, to identify and classify the various cell types, notably to distinguish normal cells from tumor cells. The global slide annotation used to train the algorithm was based on the results of the ‘gold-standard’ examinations. Patient with a negative cytology (negative for high-grade urothelial carcinoma or atypical urothelial cells) and a negative endoscopy were considered as having negative slides, and those with histologically confirmed urothelial neoplasia as having positive slides. Overall, VisioCyt[®] is an *in vitro* diagnostic medical device that uses whole-slide digitalization and AI algorithms to identify tumor cells. The device performs a morphological analysis of voided urothelial cells, including the shape, size, and color of the nuclei. Below, we highlight important details pertaining to the clinical validation phase of the trial.

Study design and patient selection

VISIOCYT1 was a French, multicenter, prospective, clinical trial conducted in 14 centers.

Patients older than 18 years of age with bladder endoscopy for suspected BC (either *de novo* or relapsed) or for exploring the lower urinary tract for other reasons were eligible. Patients with urinary tract infections, urinary lithiasis, prior pelvic radiotherapy or renal transplants were ineligible. Patients with positive cystoscopy or negative cystoscopy with positive cytology were included in Group 1: patients with BC. Those with negative cystoscopy and negative cytology were included in Group 2: patients without BC.

All study visits were performed according to standard of care. At Visit 1, eligible patients provided urine samples (50 mL) for VisioCyt[®] diagnostic testing and performed cytology and cystoscopy. At Visit 2, histopathological analyses of tissues from cystoscopy with transurethral resection (TUR) or biopsies for patients in Group 1. Follow-up visits with cytology and cystoscopy were performed at 6 months (Visit 3) and 12 months (Visit 4). All assessments and additional visits were at the investigator’s discretion.

The study was approved by an independent Ethics Committee, “CPP Ile de France VIII—Hôpital Ambroise Paré”. All participants provided written informed consent before study participation. The study is registered at ClinicalTrials.gov (NCT02966691).

Urine sample preparation

The natural void urine samples were collected at Visit 1, before cystoscopy. The urine samples were fixed with ThinPrep[®] Cytolite, homogenized, and divided into two samples. One for routine cytology and one for the VisioCyt[®] test (centralized at the Foch Hospital, Suresnes, France).

Standard cytology

At the centers, the slides for cytology were evaluated by trained pathologists. All pathologists attended a 1-day training to obtain consensus on the use of the Paris System 2016 (TPS 2016) criteria [12] for urinary cytology. Urinary cytology was done using transmission microscopy, by standard routine procedures, before the histological results were disclosed. Patients with cytology slides with < 15 urothelial cells were considered as not evaluable and excluded from analyses.

VisioCyt® diagnostic testing

The slides prepared according to the VisioCyt® protocol were scanned on three planes, digitized on brightfield with a dedicated scanner (Hamamatsu S60), and stored on an image sharing web server. Patients with slides containing less than 15 urothelial cells were excluded from the analysis. During the diagnostic validation phase of the VISIO-CYT1 trial, digitized slides were analyzed and interpreted automatically by the VisioCyt® device, without evaluations by pathologists and blinded to the histopathological results of the samples.

Standard histopathological assessment

The histopathological analysis of bladder tissue, from cystoscopy with transurethral resection (TUR) or biopsy, performed at Visit 2, was based on the 2009 TNM classification of BC [13].

Objectives and endpoints

The primary objective of the clinical validation phase of the study was to assess the diagnostic performance of the VisioCyt® test, in terms of sensitivity. The performance of the VisioCyt® test would be considered validated if the sensitivity was above 80% and that the lower bound of the 95% CI was above the 70% threshold. The secondary endpoints included the specificity of VisioCyt® test, with specificity above 85% and that the lower bound of the 95% CI was above the 75% threshold. Furthermore, sensitivity of the VisioCyt® test was also assessed in the following categories: grade of tumor differentiation (low- and high-grade tumors), T-staging within low- and high-grade tumors, and the trial groups. When appropriate, the performance of VisioCyt® was compared with standard cytology. The sensitivity of standard cytology and of the VisioCyt® diagnostic test were calculated relative to the histopathological assessments, performed at Visit 2. The specificity of standard cytology and the VisioCyt® diagnostic test were calculated relative to data from patients in Group 2: those initially with negative cystoscopies and cytologies.

Statistical analysis

Patient demographic and disease-related data were described using mean with standard deviation (SD), median with interquartile range (IQR), and/or numbers with percentages. Sensitivity was calculated from the number of patients positive to the diagnosis test (either cytology or VisioCyt®) among patients with BC (Group 1) and confirmed with an initial positive histology (at Visit 2). The specificity calculated from the number of patients negative to the diagnosis test

among participants without BC (Group 2) confirmed with a negative cytology and cystoscopy (at Visit 2).

To respond to the primary objective, with 80% power for the superiority testing of sensitivity, relative to the theoretical threshold of 70%, and with expected sensitivity of 80%, 155 evaluable patients were required in Group 1. The primary endpoint analysis was planned in individuals allocated Groups 1 and 2. Similarly, to evaluate specificity with 80% power for the superiority testing, with a threshold of 75%, and with expected specificity of 85%, 145 evaluable individuals were required in Group 2.

The sensitivity and specificity were estimated with their confidence interval (CI) at 95% according to the Wilson method. The sensitivity and specificity of the VisioCyt® test were compared using the Wald test with a one-sided alpha level of 2.5%. The performances of the VisioCyt® and standard cytology tests were compared using the McNemar test.

The statistical analyses were performed using SAS (version 9.4 or more recent).

Results

Patient characteristics

Between October 2017 and December 2019, 391 patients were enrolled in the clinical validation phase of the VISIO-CYT1 trial. Among these, 319 were analyzed: 170 in patients with BC (Group 1) and 149 without BC (Group 2). The patient flow diagram for the trial is shown in Fig. 1.

Patients were mainly male, 132 (77.6%) in Group 1 and 91 (61.1%) in Group 2. The median age was 70 years (IQR 63–75) in Group 1 and 67 years (IQR 60–73) in Group 2.

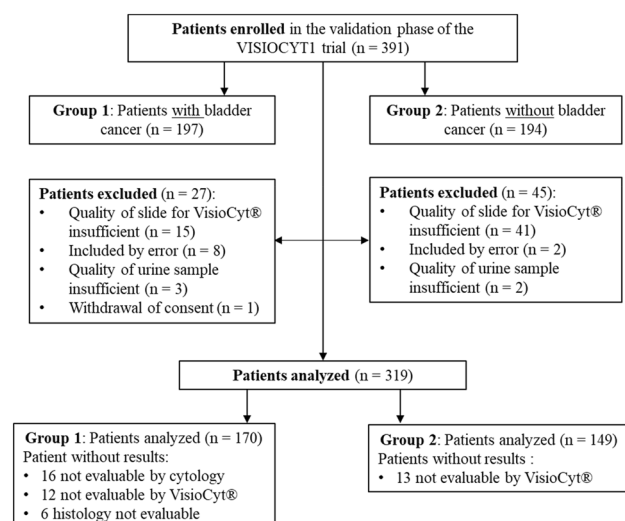


Fig. 1 Patient flow diagram for the validation phase of the VISIO-CYT1 trial

In Group 1, 137 patients (82.5%) had NMIBC (T-stages Ta, Tcis, and T1 [14]). Further baseline characteristics of patients in Group 1 are shown in the Supplementary Information section (Table S1).

Diagnostic performance of the VisioCyt® diagnostic test

The results of the analysis of the diagnostic performance of the VisioCyt® test are summarized in Table 1. The overall sensitivity and specificity were analyzed in patients allocated Group 1 (with BC) and Group 2 (without BC). The overall sensitivity (primary outcome) of the VisioCyt® test was 80.9% (95% CI 73.9–86.4%), and the lower bound of the 95% CI was significantly above the 70% threshold required

for validation ($p=0.002$). The VisioCyt® test specificity was 61.8% (95% CI 53.4–69.5%), below the targeted lower-bound threshold of 75%. The sensitivity of the VisioCyt® test was 93.7% (95% CI 86.0–97.3%) in high-grade tumors and 66.7% (95% CI 55.2–76.5%) in low-grade tumors. The sensitivity of the VisioCyt® test in the subgroups (by tumor grade) according to T-staging were like those reported overall for high-grade tumors (96% for high-grade pTa and 95.8% for pT1) and low-grade tumors (66.2% for low-grade pTa).

Diagnostic performance of standard cytology

The results of the analysis of the diagnostic performance of the standard cytology are summarized in Table 1. The

Table 1 Analysis of the performance (sensitivity and specificity) of the VisioCyt® test and standard cytology in the trial groups and in subgroups of Group 1, according to tumor grade and T-staging

Trial groups/ subgroups analyzed	Histology at Visit 2	VisioCyt® test results		VisioCyt® test performance		Standard cytology results		Standard cytology performance	
		Negative, <i>n</i> (%)	Positive, <i>n</i> (%)	Sensitivity, % (95% CI)	Specificity, % (95% CI)	Negative, <i>n</i> (%)	Positive, <i>n</i> (%)	Sensitivity, % (95% CI)	Specificity, % (95% CI)
Groups 1 and 2 (overall performance evaluation)	Negative	84 (61.8)	52 (38.2)		61.8 (53.4–69.5)	149 (100)			100.0 (97.5–100.0)
	Positive	29 (19.1)	123 (80.9)	80.9 (73.9–86.4)		80 (54.1)	68 (45.9)	45.9 (38.1–54.0)	
Group 1: Patients with bladder cancer (<i>n</i> = 170)	Positive	29 (19.1)	123 (80.9)	80.9 (73.9–86.4)		80 (54.1)	68 (45.9)	45.9 (38.1–54.0)	
High-grade tumors	Positive	5 (6.3)	74 (93.7)	93.7 (86.0–97.3)		29 (37.2)	49 (62.8)	62.8 (51.7–72.7)	
T-staging									
Ta	Positive	1 (4.0)	24 (96.0)	96.0 (80.5–99.3)		12 (50.0)	12 (50.0)	50.0 (31.4–68.6)	
Tcis	Positive	1 (14.3)	6 (85.7)	85.7 (48.7–97.4)		4 (57.1)	3 (42.9)	42.9 (15.8–75.0)	
T1	Positive	1 (4.2)	23 (95.8)	95.8 (79.8–99.3)		6 (26.1)	17 (73.9)	73.9 (53.5–87.5)	
T2	Positive	1 (5.6)	17 (94.4)	94.4 (74.2–99.0)		4 (21.1)	15 (78.9)	78.9 (56.7–91.5)	
Low-grade tumors	Positive	24 (33.3)	48 (66.7)	66.7 (55.2–76.5)		51 (73.9)	18 (26.1)	26.1 (17.2–37.5)	
T-staging									
Ta	Positive	24 (33.8)	47 (66.2)	66.2 (54.6–76.1)		50 (73.5)	18 (26.5)	26.5 (17.4–38.0)	
Group 2: Patients without bladder cancer (<i>n</i> = 149)	Negative	84 (61.8)	52 (38.2)		61.8 (53.4–69.5)	149 (100)			100.0 (97.5–100.0)

CI confidence interval

overall sensitivity of standard cytology was 45.9% (95% CI 38.1–54.0%) and the specificity was 100.0% (95% CI 97.5–100.0%). The sensitivity of standard cytology was 62.8% (95% CI 51.7–72.7%) in high-grade tumors and 26.1% (95% CI 17.2–37.5%) in low-grade tumors. The specificity of standard cytology in Group 2 (patients without BC) was 100.0% (95% CI 97.5–100.0%).

Diagnostic performance of the VisioCyt® test compared to standard cytology

Overall sensitivity was significantly higher with the VisioCyt® test, 80.9% (95% CI 73.9–86.4%), compared to 45.9% (95% CI 38.1–54.0%) with standard cytology, $p < 0.0001$ (McNemar test for paired comparison). Due to the trial's design the specificity with the VisioCyt® could not be compared to the specificity with the standard cytology. The trial was designed to only include patients with negative cytology in Group 2. Therefore, as expected, the specificity of standard cytology in Group 2 was 100.0% (95% CI 97.5–100.0%). Furthermore, the sensitivity of the VisioCyt® test to detecting high-grade tumors was 93.7% compared to 62.8% with standard cytology, $p < 0.0001$. Similarly, the sensitivity of the VisioCyt® test to detect low-grade tumors was 66.7% compared to 26.1% with standard cytology, $p < 0.0001$.

Discussion

The overall sensitivity of the VisioCyt® test was 80.9% (95% CI 73.9–86.4%) with the lower bound of the 95% CI, higher than the 70% threshold established for the trial to validate the VisioCyt® test. In the subgroup analysis, the sensitivity of the VisioCyt® test was 93.7% in high-grade tumors and 66.7% in low-grade tumors. However, the specificity was 61.8%, below the 75% threshold established.

Current standard surveillance comprises cytology and cystoscopy. The overall sensitivity of cytology was reported to be 48%: 84% for high-grade tumors and 16% for low-grade tumors with an overall specificity of 86% [3]. Cystoscopy has an overall sensitivity of about 56–70% and specificity of about 70–85% [15, 16]. In our trial, the VisioCyt® diagnostic test had a sensitivity of 80.9% which was significantly higher than the 45.9% for standard cytology in our trial and the 48% reported [3]. In contrast, the 61.8% specificity of the VisioCyt® test is lower than standard cytology, in our trial (100%) and reported (86%) [3]. In this trial, the sensitivity of standard cytology for high-grade tumors was 62.8%, lower than the 84% reported [3]. However, we used voided urine samples, with a lower concentration of cells, and not on bladder washings following cystoscopy. The specificity of the VisioCyt® test compared to cytology, in our trial, was

biased since all patients with positive standard cytology and those not evaluable were all allocated to Group 1, patients with BC. Thus, overall specificity of standard cytology in our trial was 100%, as expected.

VisioCyt®, compared to cytology from voided urine, has an improved sensitivity, for patients with low- and high-grade tumors. The role of VisioCyt® in clinical practice may be useful for the surveillance of NIMBC and for BC diagnosis. Furthermore, VisioCyt® should be useful for monitoring patients at high and very high risk of recurrence by reducing the number of cystoscopies. Fewer cystoscopies would potentially be more cost effective and reduce the risk of side effects. However, the utility of VisioCyt® still needs to be confirmed in large prospective multicentric studies.

Thus, VisioCyt® that uses voided urinary samples with analysis by a fully automated AI is a valuable addition to the diagnostic tools for detecting urothelial carcinoma. VisioCyt® can be performed whenever a urine sample is available, it may be useful before cystoscopy and when lesions are not visible, and cystoscopy is not conclusive. In these cases, VisioCyt® combined with cytology and complementary examinations, such as computed tomography and magnetic resonance imagery, may avoid repeating of time-consuming cystoscopy.

AI has been extensively applied to several urological conditions [17], including diagnosis of BC by cystoscopy [15, 18–20]. In contrast, few studies have investigated AI applications for diagnosing BC by cytology from voided urine [21–24]. To our knowledge, the VISIOCYT1 trial is the first prospective study to assess an AI-based diagnostic test for BC using urinary cytology. All, including our study, have highlighted the interest in using AI and image morphometry to improve the performance of diagnostic tools for BCs with increased sensitivity: overall and mostly in low-grade tumors [15, 18–20]. Low-grade tumor diagnosis is not reliable by cytology and the low-grade category has been excluded from the Paris System-2 classification [25, 26].

When evaluating diagnostic tools for BCs, it is critical that all suspicious and potentially malignant cells are identified [11, 18]. AI-based diagnostic tools, like VisioCyt®, have several advantages. First, they perform a complete morphological analysis of all urothelial cells present on the slide, thus identifying even the smallest of anomalies. Second, AI-based diagnostic tests ensure reproducibility and reliability of the results. Indeed, intra- and inter-observer variability observed with cytology can often be problematic. Finally, AI-based tools can learn, and it is expected that the performances of these tests will improve as more data are analyzed.

There are limitations to our work. The histological assessments were not centralized, thus there is potential interobserver variability. Furthermore, the standard cytology smears and the biopsy specimens were prepared

according to each center's standard practice. However, morphological cell appearance depends on the preparation technique. Patients included in Group 2 (patients without BC) of the VISIOCYT1 trial do not correspond to patients usually eligible for diagnostic testing.

To assess the benefit of using VisioCyt® during the surveillance of patients with NMIBC at high and very high risk of recurrence, we are currently performing the DMIA study (NCT05176145).

Conclusion

The VisioCyt® test is a promising diagnostic tool for urothelial BCs. The test has improved sensitivity, compared to standard cytology, especially in low-grade tumors, but with lower specificity. VisioCyt® is reproducible, reliable, and practical: requiring only easily accessible voided urine samples. VisioCyt®—performed before cystoscopy—may assist urologists during patient management, particularly during follow-up, to decide on the number and frequency of cystoscopy and other examinations.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s00345-023-04519-4>.

Acknowledgements We are thankful to Yannick Rouach (Antony private Hospital, Antony, France), Jacques Hubert (CHU Brabois, Nancy, France) as co-investigators; Hervé Debaque (Massy, France), Corinne Selle-Patoz (Luxembourg), Jean Marc Desnoyer (Auxerre, France), and Djazia Hmaidi (Nice, France) for processing the cells annotation on the slide; Jacqueline Aziza (IUCT Oncopole, Toulouse, France), André Balaton and Philippe Gros (Massy, France), Samia Gonzalez (Carêmeau Hospital, Nîmes, France), Ruth Borgès-Reis (Labosud, Montpellier, France) for processing the conventional cytologies; Corine Toque (Foch hospital, Suresnes, France), Amanda Simon (CHU Hôtel Dieu Hospital, Nantes, France), Marie-Pierre Valli (Gabriel-Montpied Hospital, Clermont-Ferrand, France), Julie Fouchard (CHU Pontchaillou, Rennes, France), Vasiliki Veloudou (Edouard Herriot Hospital, Lyon, France) for acquisition of data in their role as clinical research associates; Fondation EDDH (Clinical Research Organization) for the management of the study with the investigators centers (Vandoeuvre les Nancy, France). We would like to thank Trevor Stanbury (Pro-Pens) for medical writing assistance.

Author contributions Study concept and design: TL, SN. Acquisition of data: TL, GP, MR, MC, LS, GV, LG, JR, SDV, GP, SD, FK, MC-S, EP, CR, NR-L, CK-A, KR, BC-P, YA. Analysis and interpretation of data: TL, MR, YA, XP, SN. Drafting of the manuscript: TL, MR, YA, XP, SN. Critical revision of the manuscript for important intellectual content: TL, MR, YA, XP, GP, MR, MC, LS, GV, LG, JR, SDV, GP, SD, FK, MC-S, EP, CR, NR-L, CK-A, KR, BC-P. Statistical analysis: XP. Obtaining funding: SN. Administrative, technical, or material support: SN. Supervision: TL, SN. Other: None.

Funding The study was partly funded by VitaDX International. The sponsors played a role in the study design, analyse of the data, and preparation of the publication.

Data availability The data supporting the study results will be provided, upon reason reasonable request, from the corresponding author.

Declarations

Conflict of interests Thierry Lebret declares receiving consulting fees, honoraria, and/or support for attending meetings from Astellas Pharma, Bayer, Bristol Myer Squibb, IPSEN, MSD, Roche, and VitaDX. Grégoire Poinas declares receiving support from VitaDX to conduct this study. Monique Courtade-Saïdi declares being the president of the French Society for Cytology and having received payment for expert testimony from VitaDX. Nathalie Rioux-Leclercq declares having received grants from INCa (the French National Cancer Institute), for a translational study and from the Nominoë Foundation to develop digital pathology at the CHU Rennes. Karine Renaudin declares having received payments from MSD. Béatrix Cochand-Priollet declares receiving support for attending meetings. Nivet declares having shares and being employed by VitaDX. Morgan Rouprêt declares having received consulting fees from Cepheid, OncoDiag, Astellas Pharma, Janssen, IPSEN, and VitaDX. All other authors have nothing to declare.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

1. Flaig TW, Spiess PE, Agarwal N, Bangs R, Boorjian SA, Buyyounouski MK et al (2020) Bladder cancer, version 3.2020, NCCN clinical practice guidelines in oncology. *J Natl Compr Canc Netw* 18:329–354
2. Reid MD, Osunkoya AO, Siddiqui MT, Looney SW (2012) Accuracy of grading of urothelial carcinoma on urine cytology: an analysis of interobserver and intraobserver agreement. *Int J Clin Exp Pathol* 5:882–891
3. Yafi FA, Brimo F, Steinberg J, Aprikian AG, Tanguay S, Kassouf W (2015) Prospective analysis of sensitivity and specificity of urinary cytology and other urinary biomarkers for bladder cancer. *Urol Oncol* 33(66):e25-31
4. Oliveira MC, Caires HR, Oliveira MJ, Fraga A, Vasconcelos MH, Ribeiro R (2020) Urinary biomarkers in bladder cancer: Where do we stand and potential role of extracellular vesicles. *Cancers (Basel)* 12(6):1400
5. Soria F, D'Andrea D, Pohar K, Shariat SF, Lotan Y (2018) Diagnostic, prognostic and surveillance urinary markers in nonmuscle invasive bladder cancer: any role in clinical practice? *Curr Opin Urol* 28:577–583
6. Soubra A, Risk MC (2015) Diagnostics techniques in nonmuscle invasive bladder cancer. *Indian J Urol* 31:283–288
7. Burger M, Kamat AM, McConkey D (2021) Does variant histology change management of non-muscle-invasive bladder cancer? *Eur Urol Oncol* 4:510–514
8. Chen J, Remulla D, Nguyen JH, Aastha D, Liu Y, Dasgupta P et al (2019) Current status of artificial intelligence applications

- in urology and their potential to influence clinical practice. *BJU Int* 124(4):567–577
9. Checcucci E, Autorino R, Cacciamani GE, Amparore D, De Cillis S, Piana A et al (2020) Artificial intelligence and neural networks in urology: current clinical applications. *Minerva Urol Nefrol* 72:49–57
 10. Hosein S, Reitblat CR, Cone EB, Trinh QD (2020) Clinical applications of artificial intelligence in urologic oncology. *Curr Opin Urol* 30:748–753
 11. Leuret T, Pignot G, Colombel M, Guy L, Rebillard X, Savareux L et al (2021) Artificial intelligence to improve cytology performances in bladder carcinoma detection: results of the VisioCyt test. *BJU Int* 129(3):356–363
 12. Barkan GA, Wojcik EM, Nayar R, Savic-Prince S, Quek ML, Kurtycz DF et al (2016) The paris system for reporting urinary cytology: the quest to develop a standardized terminology. *Acta Cytol* 60:185–197
 13. Sobin LH, Gospodarowicz MK, Wittekind C (2009) TNM classification of malignant tumours. John Wiley & Sons
 14. Matulewicz RS, Steinberg GD (2020) Non-muscle-invasive Bladder cancer: overview and contemporary treatment landscape of neoadjuvant chemoablative therapies. *Rev Urol* 22:43–51
 15. Ikeda A, Nosato H, Kochi Y, Kojima T, Kawai K, Sakanashi H et al (2020) Support system of cystoscopic diagnosis for bladder cancer based on artificial intelligence. *J Endourol* 34:352–358
 16. Kufukihara R, Kikuchi E, Shigetani K, Ogihara K, Arita Y, Akita H et al (2022) Diagnostic performance of the vesical imaging-reporting and data system for detecting muscle-invasive bladder cancer in real clinical settings: Comparison with diagnostic cystoscopy. *Urol Oncol* 40:61 (e1–e8)
 17. Shah M, Naik N, Somani BK, Hameed BMZ (2020) Artificial intelligence (AI) in urology-current use and future directions: an iTRUE study. *Turk J Urol* 46:S27–S39
 18. Gosnell ME, Polikarpov DM, Goldys EM, Zvyagin AV, Gillatt DA (2018) Computer-assisted cystoscopy diagnosis of bladder cancer. *Urol Oncol* 36:8.e9–8.e15
 19. Ikeda A, Nosato H, Kochi Y, Negoro H, Kojima T, Sakanashi H et al (2021) Cystoscopic imaging for bladder cancer detection based on stepwise organic transfer learning with a pretrained convolutional neural network. *J Endourol* 35:1030–1035
 20. Lorencin I, Andelic N, Spanjol J, Car Z (2020) Using multi-layer perceptron with Laplacian edge detector for bladder cancer diagnosis. *Artif Intell Med* 102:101746
 21. Sokolov I, Dokukin ME, Kalaparathi V, Miljkovic M, Wang A, Seigne JD et al (2018) Noninvasive diagnostic imaging using machine-learning analysis of nanoresolution images of cell surfaces: detection of bladder cancer. *Proc Natl Acad Sci USA* 115:12920–12925
 22. Muralidaran C, Dey P, Nijhawan R, Kakkar N (2015) Artificial neural network in diagnosis of urothelial cell carcinoma in urine cytology. *Diagn Cytopathol* 43:443–449
 23. Sanghvi AB, Allen EZ, Callenberg KM, Pantanowitz L (2019) Performance of an artificial intelligence algorithm for reporting urine cytopathology. *Cancer Cytopathol* 127:658–666
 24. Vaickus LJ, Suriawinata AA, Wei JW, Liu X (2019) Automating the Paris System for urine cytopathology—a hybrid deep-learning and morphometric approach. *Cancer Cytopathol* 127:98–115
 25. Koss LG, Sherman AB (1984) Image analysis of cells in the sediment of voided urine. *Monogr Clin Cytol* 9:148–162
 26. McCroskey Z, Pambuccian SE, Kleithers S, Antic T, Cohen MB, Barkan GA et al (2015) Accuracy and interobserver variability of the cytologic diagnosis of low-grade urothelial carcinoma in instrumented urinary tract cytology specimens. *Am J Clin Pathol* 144:902–908

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Authors and Affiliations

Thierry Leuret¹ · Xavier Paoletti^{2,3} · Geraldine Pignot⁴ · Mathieu Roumigué⁵ · Marc Colombel⁶ · Laurent Savareux⁷ · Grégory Verhoest⁸ · Laurent Guy⁹ · Jérôme Rigaud¹⁰ · Stéphane De Vergie¹⁰ · Grégoire Poinas¹¹ · Stéphane Droupy¹² · François Kleinclauss¹³ · Monique Courtade-Saïdi¹⁴ · Eric Piaton¹⁵ · Camelia Radulescu¹⁶ · Nathalie Rioux-Leclercq¹⁷ · Christine Kandel-Aznar¹⁸ · Karine Renaudin^{19,20} · Béatrix Cochand-Priollet²¹ · Yves Allory^{22,23} · Sébastien Nivet²⁴ · Morgan Rouprêt²⁵

✉ Sébastien Nivet
sebastien@vitadx.com

¹ Urology Department, Foch Hospital, Suresnes, France

² Institut Curie, Saint Cloud, France

³ Université Versailles Saint-Quentin, Université Paris-Saclay, Saint Cloud, France

⁴ Urology Department, Institut Paoli-Calmettes, Marseille, France

⁵ Urology Department, Centre Hospitalier Universitaire (CHU) Rangueil, IUCT Oncopole, Toulouse, France

⁶ Urology Department, Hôpital Edouard Herriot, Lyon, France

⁷ Urology Auvergne Centre, Clinique de la Chataigneraie, Beaumont, France

⁸ Urology Department, CHU Pontchaillou, Rennes, France

⁹ Urology Department of Urology, CHU Gabriel Montpied, Clermont-Ferrand, France

¹⁰ Urology Department, CHU Hôtel-Dieu, Nantes, France

¹¹ Urology Department, Clinique Beausoleil, Montpellier, France

¹² Urology Department, CHU Caremeau, Nîmes, France

¹³ Urology Department, CHU Besançon, Besançon, France

¹⁴ Pathology and Cytology Department, CHU Toulouse, IUCT Oncopole, Toulouse, France

¹⁵ Centre de Pathologie Est, Hospices Civils de Lyon, Hôpital Femme-Mère-Enfant, Bron, France

¹⁶ Service d'Anatomie et Cytologie Pathologiques, Hôpital Foch, Suresnes, France

¹⁷ Department of Pathology, CHU Pontchaillou, Rennes, France

- ¹⁸ Laboratoire d'Anatomie et Cytologie Pathologique, CHU Hôtel Dieu, Nantes, France
- ¹⁹ Department of Pathology, CHU Hôtel Dieu, Nantes, France
- ²⁰ Centre de Recherche en Transplantation et en Immunologie, UMR 1064, INSERM, Université de Nantes, Nantes, France
- ²¹ Department of Pathology, Hôpital Cochin, Assistance Publique-Hôpitaux de Paris, Paris, France
- ²² Department of Pathology, Institut Curie, Saint-Cloud, France
- ²³ Institut Curie, PSL Research University, CNRS, UMR144, Equipe Labellisée Ligue Contre le Cancer, Paris, France
- ²⁴ VitaDX International, Rennes, France
- ²⁵ Urology Department, GRC n°5, Predictive ONCO-URO, Pitié-Salpêtrière Hospital, AP-HP, Sorbonne University, Paris, France