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▶ To cite this version:

Jonathan Rolland, L. Francisco Henao-Diaz, Michael Doebeli, Rachel Germain, Luke J Harmon, et al.. Conceptual and empirical bridges between micro and macroevolution. Nature Ecology & Evolution, 2023, 7 (8), pp.1181-1193. 10.1038/s41559-023-02116-7. hal-04190499

HAL Id: hal-04190499

https://hal.science/hal-04190499

Submitted on 31 Aug 2023

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1 Conceptual and empirical bridges between micro and macroevolution

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- 36 Keywords: adaptive landscape, speciation, extinction, demography, molecular evolution,
- 37 phenotypic evolution
- 38 Abstract
- 39 Explaining broad molecular, phenotypic, and species biodiversity patterns necessitates a
- 40 unifying framework spanning multiple evolutionary scales. We argue here that, although
- 41 substantial effort has been made to reconcile microevolution and macroevolution, much work
- remains to identify the links between biological processes at play. We highlight four major
- 43 questions of evolutionary biology whose solutions require conceptual bridges between micro
- and macroevolution. We review potential avenues for future research to establish how
- 45 mechanisms at one scale (drift, mutation, migration, selection) translate to processes at the
- other (speciation, extinction, biogeographic dispersal) and vice versa. We propose ways in
- 47 which current comparative methods to infer molecular evolution, phenotypic evolution and
- 48 species diversification could be improved to specifically address those questions. We conclude
- 49 that researchers are in a better position than ever before to build a synthesis to understand how
- 50 microevolutionary dynamics unfold over millions of years.

- 52 Main Text
- Reconciling scales of evolution

Since the modern synthesis¹, many evolutionary biologists have focused their attention on evolution at one of two different time scales: microevolution, i.e., the evolution of populations below the species level (in fields such as population genetics, phylogeography, and quantitative genetics), or macroevolution, i.e., the evolution of species or higher taxonomic levels (e.g., phylogenetics, paleobiology, and biogeography). Patterns at the two scales often seem to contradict one another. For example, accumulating evidence suggests that rates of evolution seem to be faster at shorter time scales than at longer timescales². This discrepancy in rates has been detected in molecular evolution, phenotypic evolution and even lineage diversification rates², leading to several paradoxes and exposing conceptual gaps in current theory (see Figure 1). In the past few decades, attempts have been made to bridge these gaps and reconcile these two scales³⁻¹². Nevertheless, the conceptual framework, terminology, and mathematical models remain largely separate and opportunities exist to unite them. For example, speciation is often described as a lengthy divergence between two populations at the microevolutionary scale and as an instantaneous process in birth-death models at the macroevolutionary scale, but recent efforts have sought to improve this ^{e.g.13}. Additionally, a known pitfall of comparative methods is their inability to identify which microevolutionary mechanisms underlie associations between phenotypic evolution and speciation-extinction dynamics. We argue here that solving many long-standing questions in evolutionary biology will benefit from tighter conceptual linkages between micro- and macroevolutionary approaches. We focus on four as yet unsolved questions in evolutionary biology and propose avenues to tackle them.

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74 *1-Why does the rate of evolution appear to accelerate close to present time?*

Pattern The paradox of stasis is the apparent discrepancy between the slow rate of phenotypic evolution measured in the fossil record, over geological time scales and the fast rates of evolution observed in populations at the present time¹⁴⁻¹⁸. The pattern can be decomposed in two distinct components. The first is the acceleration of evolutionary rates close to the present (Figure 1), which has been observed in molecular¹⁹⁻²¹, phenotypic^{7,22}, phylogenetic²³⁻²⁶ and fossil data²⁷. The second component is the phenotypic stasis observed through long spans of geological time, which we address in the next section (see section 2).

Explanations The perception of accelerated rates of molecular and phenotypic evolution, as well as diversification, near the present might be due to methodological biases and/or biological processes². Methodological bias can result from model misspecification, and from sampling or

parameter estimation errors^{2,22}. Errors in the estimation of a time interval biases rates upwards, particularly when the time interval (the denominator of a rate) is small. An example of model misspecification occurs when rate heterogeneity is ignored in a model, because more variability is expected over shorter time frames, resulting in younger clades exhibiting the highest rates of evolution^{2,28}. Similarly, phenotypic evolution over short time periods measures instantaneous rate, whereas measures over long time scales provide net rate, which need not be the same. Biological reasons include bounds on evolution, which become evident only at the macroevolutionary scale (e.g., limits on the number of combinations of nucleotides for a genome sequence of a given length or bounded phenotypic traits) or simply, if unrealistically, evolution might be accelerating towards the present day². Another potential factor is fluctuating selection, which yields measurable evolution in the short term, is recorded in the fossils only as time-averaged population variation. The fact that all rates tend to accelerate toward the present seems to suggest a common biological explanation and/or a sampling/methodological bias shared between subdisciplines of evolutionary biology.

One methodological bias that could contribute to the three patterns is our inability to fully account for extinction and the incompleteness of the data in the past (Figure 2). Because much of the past evolutionary history of a lineage is lost over time, there is much more data sampled at the present time and more evolutionary change can be detected, which likely leads to a perceived acceleration of the rates of evolution and diversification²⁹. An excess of lineages sampled at present time has already been described as the "pull of the present" in the phylogenetic birth-death model literature³⁰ or "the pull of the recent" in the paleontology literature³¹, which is related to the probability that a (fossil) lineage is lost over time³². The reconstructed birth-death process described in Nee et al.²⁹ is intended to explicitly account for the pull of the present but only if rates are homogenous across time and among clades. While a range of solutions have been derived to account for heterogeneous rates (e.g., ³³), estimating extinction rates accurately, especially in the absence of fossil data, remains a challenging task³⁴.

Similarly, most past genetic polymorphisms are not recorded through time. This results from both neutral processes, whereby many polymorphisms are lost through genetic drift and selective processes that purge deleterious polymorphisms. Regardless of the cause, only a fraction of polymorphisms will be fixed. The fact that many alleles are deleterious and short-lived helps to explain why short-term rates of molecular evolution are much higher than longer-term rates of substitution¹⁹ (Figure 1).

Finally, we can follow the same reasoning with phenotypic differences between populations. Futuyma³⁵⁻³⁶ proposed that a large part of the variation in phenotypes observed at present — 'ephemeral divergences' — will likely be lost over long time scales when populations go extinct or merge. Reconstructing ancestral values based on a small fraction of the phenotypic variance preserved in the fossil record from only persisting populations could lead to biased phenotypic values on deeper timescales and lower estimated rates compared to the present².

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Future Future research should systematically assess the relative contribution of specific methodological and biological biases to the perceived increase in evolutionary rates toward the present. We identify two complementary avenues: (1) develop new approaches and (2) gather more high-quality data. The development of new approaches will ideally span different time scales, and include more complex models for both micro and macroevolution (see Box 1 for some examples), using phylogenetic information, fossils or both. Current phylogenetic methods are already correcting, to some extent, for the pull of the present³⁷, intraspecific phenotypic variance³⁸⁻³⁹, the temporal sampling heterogeneity of the fossil record⁴⁰, and for the fact that speciation is not instantaneous (protracted speciation models⁴¹). More complex macroevolutionary models could be developed to account explicitly for phenotypic 'ephemeral divergences' and the evolution of intraspecific genetic polymorphisms over long time scales (i.e., at the clade level). Some attempts have already been made with the multispecies coalescent and other recent genomic approaches⁴². To better account for extinction, phylogenies can potentially be combined with the fossil record (see attempts⁴³⁻⁴⁷), at least for the few clades for which both neontological and paleontological data are available (such as mammalian lineages, Cetacea³⁷, Carnivora⁴³ or Rodentia⁴⁸). Importantly, these models of molecular, phenotypic evolution, and diversification should be tested with individual-based simulations on geological time scales (such as^{7,49}) to assess if they can tell apart the relative contribution of methodological and biological biases in the increase of evolutionary rates near the present.

Improving comparative methods may not be enough, given that they can only estimate a low number of parameters, so we also need better data. Those data should allow a more precise and unbiased estimation of rates from past populations that are comparable with modern populations. Reanalysis of fine-scale fossil records, such as Bryozoa⁵⁰, Foraminifera⁵¹, sticklebacks⁵² or diatoms⁵³ (reviewed in⁵⁴) or long-term lab experiments⁵⁵ may also help to

determine whether time-scaling patterns are artifactual and why. Fossil databases are also important resources⁵⁶⁻⁵⁸. Finally, ancient genomes reconstructed for multiple individuals in past populations may be used to estimate past polymorphism. Comparing these ancient with present-day genomes could potentially make it possible to estimate how polymorphisms have been lost and estimate variation in the rate of molecular evolution through time with higher accuracy⁵⁹⁻⁶¹.

2- Why is there stasis over long evolutionary time scales?

Pattern The second part of the paradox of stasis consists of the observation that phenotypes in the fossil record are not evolving gradually but rather seem to show long periods of stasis (i.e., a slow rate of evolution) punctuated by short bursts of rapid evolution. Although this observation has received some support from fossil data ("stasis is data" from S. J. Gould¹⁶), recent studies show results consistent with a large variety of scenarios, ranging from a slow rate of phenotypic evolution on short time-scales (<1 Myrs) and cumulative evolutionary changes over longer time-scales with bursts of evolution (e.g., every ~25 Myrs on average⁶², or every 1-100 Myrs depending on the size of the shift⁶³), or gradual evolution on both short and long time-scales^{50,64}.

No paradox of stasis has been described for lineage diversification, as speciation and extinction rates have been shown to be highly heterogeneous through time and among clades^{33,37}. Only some small clades have been shown to fit a model of constant diversification rates, while larger clades are generally highly heterogeneous^{33,37,65}. Nonetheless, in some cases, net diversification may slow down when speciation decreases or extinction increases over time, e.g. due to diversity-dependence or other reasons⁶⁶.

Similarly, stasis is not generally expected at the molecular level. Even 'living fossil' clades, such as coelacanths and tuataras, show slow, but non-zero long-term average rates of molecular substitution and phenotypic evolution⁶⁷⁻⁶⁹. Genome composition also changes at heterogenous rates among clades, via changing rates of inversion, translocation, chromosome fusion, and polyploidization^{e.g. 70}.

Explanations Several biological processes may explain why we detect stasis in phenotypes. These include oscillating directional selection around an optimum or stabilizing selection¹⁸, which can be caused by genetic interactions between traits (epistasis, pleiotropy) or the complexity of selective pressures acting on several traits simultaneously. Signals of adaptive

evolution may often be missed when measuring selection, for statistical and/or biological reasons, such as phenotypic plasticity or age-specific response⁷⁰⁻⁷³. This may lead us to infer stasis despite abundant genetic variation for functional traits and strong selection⁷⁰⁻⁷³. Conversely, selection on skewed trait distributions can lead to signals of directional selection when in fact it is stabilizing⁷³.

The tempo and mode of evolution over long time scales will be dictated by the shape of the adaptive landscape (the mean fitness surface plotted in the phenotypic and genetic space, e.g.⁷⁵, see Box 2, Figure 3 & 4). We can formulate the hypothesis that a lineage following a 'stasis' dynamic is often stuck at one peak of the adaptive landscape but is still evolving around this peak at the same rate as other lineages. This hypothesis is consistent with previous results showing that the distance traveled by the phenotype in morphospace was similar for all fossil time series, with or without 'stasis'⁷⁶. Therefore, macroevolutionary dynamics are thought to be mainly related to changes in this landscape through time ("seascape"⁷⁷⁻⁷⁹), whereas short-term evolution depends primarily on the current landscape.

Future To better describe adaptive phenotypic evolution, we have to understand adaptive landscapes and how they change over time. Quantifying the topography of the adaptive landscape and how it changes through time might help to explain why some traits are evolving fast or slow in the long-term (Figure 3 & 4). A fast-evolving landscape or a flat landscape would lead to relatively unconstrained evolution and high lability of the trait, such as ovoviviparity in amphibians or migratory behavior in birds⁸⁰. Other traits may also have landscapes with fitness valleys corresponding to "hard" boundaries — general limits caused by physical properties of the organism (related to biomechanical constraints, maximum metabolic rate, frost tolerance, e.g.⁸¹), biological interactions (traits that would make species vulnerable to parasites or predators, such as maximum running speed for a given body mass⁸²) or simply the properties of the phenotypes studied (proportion bounded between 0 and 1, latitude bounded at 90°). For instance, the maximum critical temperature is thought to be very stable through time, with the body temperature above 40-45°C representing a deep fitness valley leading to a conserved maximum across most organisms⁸³.

Reconciling the multiple types of evolutionary landscapes that have been described and their relation to each other would also be a fruitful bridge between evolution on shorter and longer time scales (Figure 3). At least four types of "landscapes" have been envisioned: (1) The fitness function describes fitness of individuals varying in phenotype⁸⁴. (2) The adaptive

landscape is defined as the mean fitness of a population over different combinations of trait means⁸⁵ — or genotype frequencies⁷⁵ and so is closely connected to the fitness function. (3) The macroevolutionary surface is a function representing a phylogenetic model of phenotypic evolution at the species level (e.g., Ornstein Uhlenbeck model⁸⁶, Bounded Brownian Motion⁸⁷, Surface⁸⁸). It describes the frequency distribution of species means over the long run. (4) The species-selection surface describes how speciation and extinction rates depend on species values in trait space (e.g., QuaSSE⁸⁹). The connections between those different representations of selection are not yet well understood, but they might be key to predicting evolutionary trajectories from one time scale to another. In theory, one could plot all the individuals of all populations of all species in the same individual fitness landscape, and therefore all species under the same adaptive landscape. However, in practice this is difficult because a fitness peak for one species can be a valley for another, and because a peak at one scale might be a valley at another scale⁹⁰⁻⁹¹(Box 3). For example, an increase in body size leading to a higher fitness at the individual level could lead to a higher probability of extinction for the species (see also⁹² for another example on the short and long-term effects of costly traits on fitness).

Finally, as a side note, we propose that the terminologies "stasis" and "punctuated equilibrium" should be avoided when possible, as rates vary all the time with phases of bursts and slow-downs. We encourage researchers to discuss and estimate variation in rates of evolution and to investigate the causes for shifts in rates. For example, more factors potentially contributing to slow rates of evolution should be investigated, such as covariance between traits (G matrices⁸⁵ in a comparative context⁹³⁻⁹⁴), complex developmental processes (e.g., hourglass model⁹⁵), traits with no intraspecific variation or complex adaptations to multiple correlated selective pressures (e.g., humidity and temperature).

3- Do bursts of phenotypic evolution and speciation occur at the same time?

Pattern There is strong evidence that rates of evolution vary over time and along the branches of the tree of life. Punctuated equilibrium theory proposed that bursts in phenotypic evolution mainly occur with speciation events^{16,96}, an idea with little support from empirical fossil data^{50,97}. Indirect evidence has been provided at the clade level from adaptive radiations (e.g., African cichlids, Anolis lizards, Galápagos finches⁹⁸), where speciation rate and phenotypic evolution have been shown to increase concomitantly⁹⁹, but the exact synchrony between phenotypic change and speciation has proved hard to test. At the lineage level, shifts in the rate

of phenotypic evolution and of molecular evolution in adaptive genes can occur during ecological speciation: when lineages in a clade adapt to different niches (e.g., as in adaptive radiations⁹⁸), respond to a changing environment⁶⁵, or fill ecological opportunities, e.g., after a mass extinction¹⁰⁰. Interestingly, recent studies have also found evidence for an association between major genetic changes: such as major genomic rearrangements (i.e., gene duplication) associated with increased phenotypic innovation¹⁰¹ and potential speciation¹⁰². Contrastingly, other modes of speciation do not require major changes in the molecular rate nor the phenotypic rate of evolution. For instance, the evolution of Bateson-Dobzhansky-Muller incompatibilities via genetic conflict might only involve a few genes and do not necessarily affect phenotypic differences¹⁰³.

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Explanations On the one hand, there are many reasons why phenotypic evolution should be concentrated around speciation events. For example, sympatric speciation requires some form of character displacement for incipient species to coexist¹⁰⁴⁻¹⁰⁵. During allopatric speciation, geographic barriers to gene flow should lead to the accumulation of divergent mutations, which should lead to faster phenotypic divergence than the average long-term rate. Futuyma³⁵⁻³⁶ proposed that most phenotypic divergence observed at the present time is ephemeral and might not be recorded in long-term evolution, except when speciation events partition variation between daughter species. This is because gene flow, which homogenizes phenotypic and genetic differences between populations, diminishes during the emergence of reproductive isolation. The process of partitioning of variation itself may be associated with an increase in the rate of phenotypic evolution close to speciation events. In the case of asymmetric segregation of traits at speciation, individuals of the two daughter species represent a nonrandom draw of individuals from the parent population, driven by geography or traits involved in the speciation process, which can lead to an apparent jump in trait evolution⁵⁰. The hypothesis of ephemeral divergence is also consistent with the empirical observation that the accumulation of phenotypic differences speeds up after a period, about one million years⁶² in vertebrates, that is on the same order of magnitude as the expected waiting time between speciation events. In terms of molecular evolution, a reduction in population size during speciation may increase drift, or increased selective pressure on adaptive genes may lead to a higher substitution rate associated with speciation events¹⁰⁶. Finally, speciation could also be largely a by-product of genetic divergence¹⁰⁷, and consequently, speciation could be associated with bursts of molecular genetic change. Polyploidization, for example, shows signs of clustering around

speciation events⁷¹, as expected from polyploid speciation and hybrid speciation. In those cases, we may expect a correlation between genetic change, phenotypic change and speciation.

On the other hand, phenotypic change, substitution and speciation rates can be disconnected in many cases. There are many examples of putatively non-adaptive radiations, which unfold without any clear sign of ecological divergence (reported for some clades of damselflies, snails, and salamanders 108). In a similar vein, key evolutionary innovations can spread without changes in speciation rate (examples reviewed in¹¹⁰, such as the Australian from arboreal/nectarivorous honeyeaters adapting an foraging strategy terrestrial/insectivorous strategy). Additionally, it remains unclear whether substitution rates accelerate at speciation events¹¹¹⁻¹¹². The association between mutation and speciation is at the heart of classic theories, such as the metabolic theory of ecology¹¹³, which predicts that higher rates of mutation at high temperature (such as in the tropics) should foster faster molecular evolution and speciation. However, this theory has received relatively weak support, at least in the context of the latitudinal diversity gradient¹¹⁴.

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Future Overall, whether bursts of speciation and phenotypic evolution take place concurrently, or whether one typically precedes the other, remains to be definitively demonstrated. To this end, we first need to understand when and how rate shifts are happening at large macroevolutionary scales. Data on molecular and phenotypic traits, as well as speciation/extinction data, need to be compiled through time using fossil, ancient DNA and genomes in order to identify the causes and the consequence of each speciation event, measured over a long enough time scale to capture shifts. Fine-scale analysis of planktonic fossil sequences¹¹⁵, long-term lab experiments⁵⁵ and resurrection experiments¹¹⁶ can help to test the causes of shifts and speciation events. For example, comparison between ancient DNA in sediments and present-day genomes of sticklebacks has led to the identification of genes under selection during the transition from marine to freshwater habitat ~12,000 years ago⁶¹. Traitdependent models of diversification (e.g., ClaSSE¹¹⁷ distinguishing cladogenetic and anagenetic trait evolution) and the development of models that combine fossil observations and phylogenies¹¹⁸ allow for probabilistic tests of the correlation between speciation and trait evolution at nodes and along the branches of the phylogenetic tree¹¹⁹. Finally, in order to better understand how phenotypic variation is distributed between the two daughter lineages during speciation and if jumps occur at speciation, current comparative models (such as BM, OU or Levy flight⁶³) could be improved to account for asymmetrical inheritance of the intraspecific variation during speciation for quantitative traits⁴⁹.

We are currently unsure under what circumstances macroevolution can predict microevolution⁷⁻⁸ and vice versa (Box 3). We lack a framework to predict how adaptive landscapes shift and the resulting impact on phenotypic evolution and diversification. A striking example was the inability of researchers to predict the speed and the direction of change leading to the emergence of new variants of the SARS-CoV-2 virus (but see¹²⁰). An important factor complicating the prediction is that bursts of evolution strongly depend on the environmental and the ecological context. For example, in the context of adaptive radiation, phenotypic jumps to an alternative adaptive peak depends on whether the peaks (niches) are already occupied^{85,88,98}. More generally, macroevolution could potentially be used to understand when an adaptive jump will likely occur, by comparing mutations affecting fitness in other related lineages at the clade level, and by estimating the probability of a beneficial mutation to evolve given the sequence of the protein at a given time. Microevolution could also be helpful to predict the rate of evolution and trajectory of evolution during the jump, using population-based quantities such as intraspecific genetic variance (G matrix) for adaptive traits⁷³. This gives hope that microevolution and macroevolution may soon be modeled jointly to improve the predictive power of evolutionary biology concerning the probability of speciation, extinction, and the adaptation of species.

4- Do ecological interactions leave a predictable signature on macroevolution?

Pattern Detecting the imprint of species interactions on macroevolution remains a difficult task. Although evolution caused by ecological interactions is often thought to happen over short time scales, interactions can influence evolutionary dynamics below and above the species level¹²¹⁻¹²⁵. Although a number of potential biotic interactions (i.e., mutualism, symbiosis, commensalism, antagonism) may impact both micro and macroevolution¹²⁶, previous studies focused mostly on competition and predation. At the macroevolutionary scale, slowdowns in diversification rates through time, are often inferred from small and medium-size phylogenetic trees⁶⁵ and have often been interpreted as a potential signature of increased competition accompanying the buildup of biodiversity reducing ecological opportunity. Speciation rates should decline and extinction rates increase as the number of species increases over time, owing to the increasing competition between the species for the limited resources. Under this process, called diversity dependence^{25,127-128}, we expect that the rate at which new species accumulate

slows near the species "carrying capacity" of the environment. This process can be mitigated when the resource limits are far from being reached or when evolutionary innovation continually affords access to new resources. But the cause of observed slowdowns in diversification is uncertain⁶⁸. Many other factors could cause slowdowns in diversification, such as the temperature decline through the Phanerozoic⁶⁵, the effect of extinction and/or protracted speciation at the tips of phylogenetic trees⁶⁶. Clade interactions could also influence diversification dynamics on macroevolutionary scales, such as competition between Bivalves and Brachiopods¹²⁹, Canids¹³⁰, or between Angiosperms and Gymnosperms¹³¹.

Explanations Many studies aimed at detecting the signal of biotic interactions on macroevolution focus on the effect of interspecific competition. For instance, diversity-dependent models assume that every environment can only carry a given number of species, which is thought to be limited by the number of niches (a challenging concept to quantify¹⁰⁰) or resource availability (see the discussion in¹³² and¹³³). Diversity-dependent diversification models have been developed to estimate carrying capacity at the clade-level¹³⁴⁻¹³⁶, but these models have important limitations.

First, these models do not account for all lineages potentially competing for resources. They should account for interactions with lineages outside of the clade within the same geographical region. Second, they do not explicitly include phenotypic data or pre-existing data on species interactions. Species competing for the same niche are expected to show signs of character displacement, which should be detectable with coexistence experiments ¹⁰⁵, disparity through time analysis of phenotypes (DTT¹³⁷) or other models of phenotypic evolution (diversity-dependent, matching competition and early burst models ¹¹⁹). These types of analyses permit the detection of interspecific phenotypic interaction during species coexistence ¹³⁸ and across the latitudinal gradient ¹³⁹. Interactions beyond temporal co-occurrence, such as predation and spatial competition can be observed directly in the fossil record ¹⁴⁰ and could potentially be incorporated in modeling frameworks.

Finally, diversity-dependent macroevolutionary models do not account for the demographic consequences of species interactions. At the microevolutionary scale, a slowdown in population growth rate due to density dependence is often observed when individuals compete for limited resources, so detecting this slowdown, i.e. using fossil density estimates¹⁴¹, may help to validate the hypothesis that species are indeed interacting.

Future To better test whether biotic interactions leave a detectable imprint on macroevolution, we propose that several lines of evidence from population trends, ecological experiments, phenotypic data and birth-death models should be used to connect data at microevolutionary and macroevolution scales. First, in order to understand the impact of direct ecological competition on diversification, there is a need to improve our fundamental understanding of the links between diversity-dependent and density-dependent processes¹⁴². In theory, if one species is outcompeting another, it should experience an increase in effective population size, while the other should decline. Correlations in abundance data through time are thus helpful to understanding how species interact and if the fitness of the individuals from one species is affected by other species. Fossil community data through time (such as pollen and macrofossil database, e.g., Neotoma) could help to identify more such interactions between clades that were coexisting in the past^{48,143}, given that species abundance can be obtained and compared through time (from fossils¹⁴¹ or quantification of environmental DNA¹⁴⁴). Effective population size can also be reconstructed, extending back for millions of years, either using ancient DNA or current genomes with Pairwise Sequentially Markovian Coalescent¹⁴⁵. Interlinked patterns among species could reveal the nature of interactions among them, keeping in mind that estimates of effective population size can be biased and do not directly correspond to census population size.

Although there are many microevolutionary models of species interactions, future research should also investigate the demographic consequences of species interactions and complex feedback loops between ecology and evolution across long time scales^{121,124,129-131}. For instance, at the microevolutionary scale, the evolution of a predator can change the shape of the adaptive landscape at a lower trophic level, which can, in turn, impact the evolution of the predator¹⁴⁶. Such feedback can then affect speciation and extinction probabilities of all interacting lineages. Similarly, as species alter their environment, they can create or destroy niches of other species¹⁴⁷, altering diversification rates in the community. Interestingly, each peak in the adaptive landscape defined by a resource can be stable over long time scales, leading to the specialization of phenotypes, which in some extreme cases can lead to decreased speciation and increased extinction risk (e.g., *X. morganii praedicta* feeding on *A. sesquipedale*¹⁴⁸), or be more labile and related to the coexistence of species over short time scales. A specific example is the coexistence of incipient species during speciation¹⁰⁵. The strength of resource competition between two new species impacts both the probability that a new isolated population forms and the persistence of isolated populations, two key parameters

rarely estimated in the literature^{11,149}. The appearance and persistence of populations should in turn be linked to the probability of speciation and extinction at larger time scales^{9,129}. Recently developed comparative models¹³⁹ have the potential to be used to study a large variety of positive and negative species interactions (such as competition, mutualisms, symbiosis)¹²⁶.

Finally, through longer time scales, comparing similar environments at different places on earth may also help to identify empty and occupied peaks in the adaptive landscape, in order to better understand biotic constraints¹⁵⁰ and ecological opportunities for phenotypic evolution and diversification.

Concluding remarks

The four questions we address highlight many of the challenges but also the advantages to be gained by reconciling microevolution within species and macroevolution above the species level. Success is not assured, and we have listed reasons why in some cases it may not be possible to bridge across evolutionary scales. Yet, many of the previous attempts have already been illuminating. Seeming discrepancies between rates of evolution over long and short time scales have indicated the existence of potentially key understudied processes, such as bounded evolution and ephemeral evolution. These examples illustrate how insights about microevolutionary processes may sometimes be revealed only after considering macroevolutionary data.

Bridging scales of evolution will require using transdisciplinary approaches in order to explain how combinations of microevolutionary processes produce macroevolutionary patterns and to solve long-standing questions. In this review, we propose that future research should move away from framing that ignores the dynamic interplay among processes. Quantifying more precisely the tempo and mode of evolution among clades and through time promises to unveil times when evolutionary rates change at the phenotypic and species level. We propose that future research should (i) assess the relative contribution of methodological biases and biological processes to understand why estimated molecular, phenotypic and diversification rates appear to increase through time, (ii) attempt to measure adaptive landscapes across space and time to explain changes in the tempo of evolution, (iii) measure more precisely variation in rates and the causes of large shifts, especially near speciation events, and (iv) conduct further tests of the impact of species interactions on macroevolution, combining abundance and phenotypic data. Unification across scales in evolutionary biology seems more possible than ever before. We anticipate that, in the next decades, researchers will likely revisit old questions

with new models and data in genomics, computational biology and paleontology in order to better describe and understand evolution in all its complexity.

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Acknowledgments and Funding

- 453 This article was designed during the working group "Linking micro and macroevolution"
- organized by J.R. with advices from D.Sc. at the University of British Columbia (UBC).
- 455 Funding was provided to J.R. through a UBC grant for Catalyzing Biodiversity research. We
- 456 thank K. Beall and L. Rieseberg for their support of this meeting. We thank three anonymous
- reviewers for their very constructive comments. J.R. also received funding from the European
- 458 Union's Horizon 2020 Research and Innovation Program under the Marie Skłodowska-Curie
- 459 (grant no. 785910) and from Investissement d'Avenir grants managed by the Agence Nationale
- de la Recherche (CEBA: ANR-10-LABX-25-01; TULIP: ANR-10-LABX-0041). M.D., R.G.,
- 461 J.E.M., S.P.O., M.P., and D.Sc. received grant support from the Natural Sciences and
- Engineering Research Council of Canada (Discovery Grant). D.Si. received funding from the
- Swiss National Science Foundation (PCEFP3 187012), from the Swedish Research Council
- 464 (VR: 2019-04739), and from the Foundation for Environmental Strategic Research, Sweden
- 465 (BIOPATH). A.M. was funded by the Charles University Research Centre programme (no.
- 466 204069); StGACR 23-05977S Czech Science Foundation; European Union's Horizon 2020
- Research and Innovation Programme under the Marie Sklodowska-Curie Grant (agreement no.
- 468 785799). L.H.L. has received funding from the European Research Council (ERC) under the
- 469 European Union's Horizon 2020 research and innovation programme (grant agreement No
- 470 724324). C.E.W. was partially supported by the US National Science Foundation grant DEB-
- 471 1556963.

Contributions

- 473 J.R., L.F.H-D., M.D., R.G., L.J.H., L.L.K., L.H.L., J.E.M., A.M., S.P.O., M.P., N.S., D.Si.,
- 474 M.S., J.U., C.E.W. and D.Sc. participated at the workshop and designed the research. J.R. wrote
- 475 the first draft. All co-authors participated in commenting and editing the manuscript, with a
- 476 substantial contribution of J.R. and D.Sc.

Competing interest 478 479 The authors declare no competing interest. 480 References 481 482 483 1. Huxley, J. Evolution. The modern synthesis. Evolution. The Modern Synthesis. (1942). 484 485 2. Harmon, L. J., Pennell, M. W., Henao-Diaz, L. F., Rolland, J., Sipley, B. N. & Uyeda, J. C. 486 Causes and consequences of apparent timescaling across all estimated evolutionary rates. Annu. Rev. Ecol. Evol. Syst. 52, 587-609 (2021). 487 488 489 3. Charlesworth, B., Lande, R. & Slatkin, M. A neo-Darwinian commentary on 490 macroevolution. Evolution 36, 474-498 (1982). 491 492 4. Estes, S. & Arnold, S. J. Resolving the paradox of stasis: models with stabilizing selection explain evolutionary divergence on all timescales. Am. Nat. 169, 227-244. (2007). 493 494 495 5. Hansen, T. F. & Martins, E. P. Translating between microevolutionary process and 496 macroevolutionary patterns: the correlation structure of interspecific data. Evolution 50, 497 1404-1417 (1996). 498 499 6. Reznick, D. N. & Ricklefs, R. E. Darwin's bridge between microevolution and 500 macroevolution. Nature 457, 837-842 (2009). 501 502 7. Rolland, J., Silvestro, D., Litsios, G., Faye, L. & Salamin, N. Clownfishes evolution below 503 and above the species level. *Proc. Roy. Soc. B* **285**, 20171796 (2018). 504 505 8. Arnold, S. J., Pfrender, M. E., & Jones, A. G. The adaptive landscape as a conceptual 506 bridge between micro-and macroevolution. Microevolution rate, pattern, process, 9-32 507 (2001).508 509 9. Singhal, S., Colli, G. R., Grundler, M. R., Costa, G. C., Prates, I. & Rabosky, D. L. No link 510 between population isolation and speciation rate in squamate reptiles. *Proc. Natl. Acad.* 511 Sci. USA 119, e2113388119 (2022). 512 513 10. Rabosky, D. L., & Matute, D. R. Macroevolutionary speciation rates are decoupled from 514 the evolution of intrinsic reproductive isolation in Drosophila and birds. Proc. Natl.

517 11. Dynesius, M. & Jansson, R. Persistence of within-species lineages: a neglected control of speciation rates. *Evolution* **68**, 923-934 (2014).

Acad. Sci. USA 110, 15354-15359 (2013).

515

- 520 12. Alencar, L. R. V. D., & Quental, T. B. Linking population-level and microevolutionary
- 521 processes to understand speciation dynamics at the macroevolutionary scale. Ecology
- 522 and Evolution, **11**, 5828-5843 (2021).
- 523 13. Hua, X., Herdha, T. & Burden, C.J. Protracted speciation under the state-dependent
- speciation and extinction approach. Syst. Biol. 71, 1362-1377 (2022).
- 525 14. Simpson, G. G. Tempo and mode in evolution. Columbia University Press (1944).

526

- 527 15. Lewontin, R. C. *The genetic basis of evolutionary change*. New York: Columbia
- 528 University Press (1974).

529

530 16. Gould, S. J. & Eldredge, N. Punctuated equilibria: the tempo and mode of evolution 531 reconsidered. *Paleobiology* **3**, 115-151 (1977).

532

- 17. Hansen, T. F. & Houle, D. Evolvability, stabilizing selection and the problem of stasis.
- Phenotypic integration: Studying the ecology and evolution of complex phenotypes. eds
- 535 Pigliucci M. and Preston K. pp130–150 (2004).

536

- 18. Haller, B. C. & Hendry, A. P. Solving the paradox of stasis: squashed stabilizing selection
- and the limits of detection. *Evolution* **68**, 483-500 (2014).

539

- 19. Ho, S. Y., Phillips, M. J., Cooper, A. & Drummond, A. J. Time dependency of molecular rate estimates and systematic overestimation of recent divergence times. *Mol. Biol.*
- 542 Evol. **22**, 1561-1568 (2005).

543

- 544 20. Ho, S. Y., Lanfear, R., Bromham, L., Phillips, M. J., Soubrier, J., Rodrigo, A. G. &
- Cooper, A. Time-dependent rates of molecular evolution. *Mol. Ecol.* **20**, 3087-3101
- 546 (2011).

547

548 21. Ho, S. Y., Duchêne, S., Molak, M. & Shapiro, B. Time-dependent estimates of molecular 549 evolutionary rates: evidence and causes. *Mol. Ecol.* **24**, 6007-6012 (2015).

550

551 22. Gingerich, P. D. *Rates of evolution: A quantitative synthesis*. Cambridge University Press (2019).

553

- 23. Magallon, S., & Sanderson, M. J. Absolute diversification rates in angiosperm clades.
- 555 Evolution **55**, 1762-1780 (2001).

556

24. Ricklefs, R. E. Evolutionary diversification and the origin of the diversity–environment relationship. *Ecology* **87**, S3-S13 (2006).

- 25. McPeek, M. A. & Brown, J. M. Clade age and not diversification rate explains species richness among animal taxa. *Am. Nat.* **169**, E97-E106 (2007).
- 26. Louca, S., Henao-Diaz, L. F. & Pennell, M. The scaling of diversification rates with age is likely explained by sampling bias. *Evolution* **76**, 1625–1637 (2022).
- 564 27. Henao Diaz, L. F., Harmon, L. J., Sugawara, M. T., Miller, E. T. & Pennell, M. W.
- Macroevolutionary diversification rates show time dependency. *Proc. Natl. Acad. Sci.*
- 566 *USA* **116**, 7403-7408 (2019).

28. Yang Z. Computational Molecular Evolution. Oxford, UK: Oxford Univ. Press (2006).

569

570 29. Budd, G. E., & Mann, R. P. History is written by the victors: the effect of the push of the past on the fossil record. *Evolution* **72**, 2276-2291 (2018).

572

- 573 30. Nee, S., R. Mccredie May, & Harvey P. H. The reconstructed evolutionary process.
- 574 Philos. Trans. R. Soc. **344**, 305-311 (1994).

575

31. Jablonski, D., Roy, K., Valentine, J. W., Price, R. M., & Anderson, P. S. The impact of the pull of the recent on the history of marine diversity. *Science* **300**, 1133-1135 (2003).

578

32. Raup, D. M. Size of the Permo-Triassic bottleneck and its evolutionary implications.
 Science 206, 217-218 (1979).

581

33. Stadler, T. Mammalian phylogeny reveals recent diversification rate shifts. *Proc. Natl.* Acad. Sci. USA 108, 6187-6192 (2011).

584

34. Brian C. O'Meara & Beaulieu J. M. Potential survival of some, but not all, diversification methods. EcoEvoRxiv https://doi.org/10.32942/osf.io/w5nvd (2022).

587

588 35. Futuyma, D. J. On the role of species in anagenesis. *Am. Nat.*, **130**, 465-473 (1987).

589

590 36. Futuyma, D. J. Evolutionary constraint and ecological consequences. *Evolution*, **64**, 1865-591 1884 (2010).

592

593 37. Morlon, H., Parsons, T. L. & Plotkin, J. B. Reconciling molecular phylogenies with the fossil record. *Proc. Natl. Acad. Sci. USA* **108**, 16327-16332 (2011).

595

38. Kostikova, A., Silvestro, D., Pearman, P. B., & Salamin, N. Bridging inter- and
 intraspecific trait evolution with a hierarchical Bayesian approach. *Syst. Biol.*, 65, 417 431 (2016).

- 39. Gaboriau, T., Mendes, F. K., Joly, S., Silvestro, D., & Salamin, N. A multi-platform
 package for the analysis of intra- and interspecific trait evolution. *Methods Ecol. Evol.*,
 11, 1439-1447 (2020).
- 40. Silvestro, D., Salamin, N., Antonelli, A., & Meyer, X. Improved estimation of
 macroevolutionary rates from fossil data using a Bayesian framework. *Paleobiology*,
 45, 546-570 (2019).

607

611

614

617

621

625

628

632

635

- 41. Etienne, R. S. & Rosindell, J. Prolonging the past counteracts the pull of the present:
 protracted speciation can explain observed slowdowns in diversification. *Syst. biol.* 61,
 204 (2012).
- 42. De Maio, N., Schrempf, D., & Kosiol, C. PoMo: an allele frequency-based approach for species tree estimation. *Syst. Biol.*, **64**, 1018-1031 (2015).
- 43. Slater, G. J., Harmon, L. J. & Alfaro, M. E. Integrating fossils with molecular
 phylogenies improves inference of trait evolution. *Evolution* 66, 3931-3944 (2012).
- 44. Rolland, J., Silvestro, D., Schluter, D., Guisan, A., Broennimann, O. & Salamin, N. The
 impact of endothermy on the climatic niche evolution and the distribution of vertebrate
 diversity. *Nat. Ecol. Evol.* 2, 459-464 (2018).
- 45. Silvestro, D., Warnock, R., Gavryushkina, A., & Stadler, T. Closing the gap between
 palaeontological and neontological speciation and extinction rate estimates. *Nature Comm.*, 9, 1-14 (2018).
- 46. Mitchell, J. S., Etienne, R. S. & Rabosky, D. L. Inferring diversification rate variation from phylogenies with fossils. *Syst. Biol.*, **68**, 1-18 (2019).
- 47. Černý, D., Madzia, D., & Slater, G. J. Empirical and methodological challenges to the
 model-based inference of diversification rates in extinct clades. *Syst. Biol.* 71, 153-171
 (2022).
- 48. Blois, J. L., Zarnetske, P. L., Fitzpatrick, M. C., & Finnegan, S. Climate change and the past, present, and future of biotic interactions. *Science* **341**, 499-504 (2013).
- 49. Duchen, P., Alfaro, M., Rolland, J., Salamin, N. & Silvestro D. On the effect of
 asymmetrical trait inheritance on models of trait evolution. *Syst. Biol.* 70, 376-388
 (2021).
- 50. Voje, K. L., Di Martino, E. & Porto, A. Revisiting a landmark study system: no evidence for a punctuated mode of evolution in Metrarabdotos. *Am. Nat.* **195**, 899-917 (2020).

- 51. Brombacher, A., Wilson, P. A., Bailey I. & Ezard T. H. G. The breakdown of static and evolutionary allometries during climatic upheaval. *Evolution* **190**, 299-450 (2017).
- 52. Hunt, G. The relative importance of directional change, random walks, and stasis in the evolution of fossil lineages. *Proc. Natl. Acad. Sci. USA* **104**, 18404-18408 (2007).
- 53. Voje, K. L. Testing eco-evolutionary predictions using fossil data: Phyletic evolution following ecological opportunity. *Evolution* **74**, 188-200 (2020).
- 54. Webster, M. Morphological homeostasis in the fossil record. In Seminars in cell &
 developmental biology 88, 91-104 (2019).
- 55. Fox, J. W. & Lenski, R. E. From here to eternity—the theory and practice of a really long experiment. *PLoS biol.* **13**, e1002185 (2015).
- 56. Fenton, I. S., et al. Triton, a new species-level database of Cenozoic planktonic
 foraminiferal occurrences. *Sci. Data*, 8, 160 (2021).
- 57. https://paleobiodb.org/

648

651

654

657

660

662

666

670

673

677

680

- 58. Fan, J.X., Shen, S.Z., Erwin, D.H., Sadler, P.M., MacLeod, N., Cheng, Q.M., Hou, X.D.,
 Yang, J., Wang, X.D., Wang, Y. & Zhang, H. A high-resolution summary of Cambrian
 to Early Triassic marine invertebrate biodiversity. *Science* 367, 272-277 (2020).
- 59. Lambert, D. M., Ritchie, P. A., Millar, C. D., Holland, B., Drummond, A. J., & Baroni, C.
 Rates of evolution in ancient DNA from Adelie penguins. *Science* 295, 2270-2273
 (2002).
- 671 60. Hay, J. M. et al. Rapid molecular evolution in a living fossil. *Trends Genet.* **24**, 106-109 (2008).
- 674 61. Kirch, M., Romundset, A., Gilbert, M. T. P., Jones, F. C. & Foote, A. D. Ancient and 675 modern stickleback genomes reveal the demographic constraints on adaptation. *Curr*. 676 *Biol.* 31, 2027-2036. (2021).
- 62. Uyeda, J. C., Hansen, T. F., Arnold, S. J. & Pienaar, J. The million-year wait for 679 macroevolutionary bursts. *Proc. Natl. Acad. Sci. USA* **108**, 15908-15913 (2011).
- 63. Landis, M. J. & Schraiber, J. G. Pulsed evolution shaped modern vertebrate body sizes.

 82. *Proc. Natl. Acad. Sci. USA* **114,** 13224-13229 (2017).

- 684 64. Pagel, M., O'Donovan, C. & Meade, A. General statistical model shows that
- 685 macroevolutionary patterns and processes are consistent with Darwinian gradualism.
- 686 *Nat. commun.* **13**, 1-12 (2022).

688 65. Condamine, F. L., Rolland, J. & Morlon, H. Assessing the causes of diversification 689 slowdowns: temperature-dependent and diversity-dependent models receive equivalent 690 support. *Ecol. lett.* **22**, 1900-1912 (2019).

691

692 66. Moen, D. & Morlon, H. Why does diversification slow down? *Trends Ecol. Evol.* **29**, 190-197 (2014).

694

695 67. Amemiya, C. T., Alföldi, J., Lee, A. P., Fan, S., Philippe, H., MacCallum, I. *et al.* The 696 African coelacanth genome provides insights into tetrapod evolution. *Nature* **496**, 311-697 316 (2013).

698

68. Gemmell, N. J., et al. The tuatara genome reveals ancient features of amniote evolution. *Nature*, **584**, 403-409. (2020).

701

702 69. Zhan, S. H., Otto, S. P., & Barker, M. S. Broad variation in rates of polyploidy and
 703 dysploidy across flowering plants is correlated with lineage diversification. bioRxiv
 704 2021-03 (2021).

705

70. Merilä, J. & Crnokrak, P. Comparison of genetic differentiation at marker loci and quantitative traits. *J. Evol. Biol.* **14**, 892-903 (2001).

708

71. Merilä, J., & Hendry, A. P. Climate change, adaptation, and phenotypic plasticity: the problem and the evidence. *Evol. Appl.* **7**, 1–14 (2014).

711

72. Pujol, B., Blanchet, S., Charmantier, A., Danchin, E., Facon, B., Marrot, P., *et al.* The missing response to selection in the wild. *Trends Ecol. Evol.* **33**, 337-346 (2018).

714

73. Bonnet, T., Morrissey, M. B., de Villemereuil, P., Alberts, S. C., Arcese, P., Bailey, L. D. *et al.* Genetic variance in fitness indicates rapid contemporary adaptive evolution in wild animals. *Science* **376**, 1012-1016 (2022).

718

74. Bonamour, S., Teplitsky, C., Charmantier, A., Crochet, P. A. & Chevin, L. M. Selection on skewed characters and the paradox of stasis. *Evolution* **71**, 2703-2713 (2017).

721

722 75. Schluter, D. & Grant, P. R. Determinants of morphological patterns in communities of Darwin's finches. *Am. Nat.* **123**, 175-196 (1984).

724

725 76. Voje, K. L. Tempo does not correlate with mode in the fossil record. *Evolution*, **70**, 2678-2689 (2016).

728 77. Mustonen, V. & Lässig, M. From fitness landscapes to seascapes: non-equilibrium dynamics of selection and adaptation. *Trends genet.* **25**, 111-119 (2009).

730

731 78. Payne, J. L. & Wagner, A. The causes of evolvability and their evolution. *Nature Rev.*732 *Genet.* **20**, 24-38 (2019).

733

734 79. Hansen, T. F. & Svensson, E. Adaptive landscapes and macroevolutionary dynamics. *The adaptive landscape in evolutionary biology* **205**, 26 (2012).

736

737 80. Rolland, J., Jiguet, F., Jønsson, K. A., Condamine, F. L., & Morlon, H. Settling down of 738 seasonal migrants promotes bird diversification. *Proc. Roy. Soc. B* **281**, 20140473 739 (2014).

740

741 81. Zanne, A. E., Tank, D. C., Cornwell, W. K., Eastman, J. M., Smith, S. A., FitzJohn, R. G.
 742 et al. Three keys to the radiation of angiosperms into freezing environments. *Nature* 743 506, 89-92 (2014).

744

745 82. Hirt, M. R., Jetz, W., Rall, B. C. & Brose, U. A general scaling law reveals why the largest animals are not the fastest. *Nature Ecol. Evol.* **1**, 1116-1122 (2017).

747

83. Araújo, M. B., Ferri-Yáñez, F., Bozinovic, F., Marquet, P. A., Valladares, F. & Chown, S.
L. Heat freezes niche evolution. *Ecol. lett.* 16, 1206-1219 (2013).

750

751 84. Martin, C. H. & Wainwright, P. C. Multiple fitness peaks on the adaptive landscape drive adaptive radiation in the wild. *Science* **339**, 208-211 (2013).

753

754 85. Schluter, D. Adaptive radiation along genetic lines of least resistance. *Evolution* **50**, 1766-755 1774 (1996).

756

757 86. Butler, M. A. & King, A. A. Phylogenetic comparative analysis: a modeling approach for adaptive evolution. *Am. Nat.* **164**, 683-695 (2004).

759

87. Boucher, F. C. & Démery, V. Inferring bounded evolution in phenotypic characters from
 phylogenetic comparative data. *Syst. biol.* 65, 651-661 (2016).

762

763 88. Mahler, D. L., Ingram, T., Revell, L. J. & Losos, J. B. Exceptional convergence on the macroevolutionary landscape in island lizard radiations. *Science* **341**, 292-295 (2013).

765

766 89. FitzJohn, R. G. Quantitative traits and diversification. *Syst. biol.* **59**, 619-633 (2010).

767

90. Gould, S. J. Gulliver's further travels: the necessity and difficulty of a hierarchical theory of selection. *Phil. Trans. R. Soc. Lond.* **353**, 307-314 (1998).

91. Rabosky, D. L. & McCune, A. R. Reinventing species selection with molecular phylogenies. *Trends Ecol. Evol.* 25, 68-74 (2010).

773

92. Martins, M. J. F., Puckett, T. M., Lockwood, R., Swaddle, J. P. & Hunt, G. High male sexual investment as a driver of extinction in fossil ostracods. *Nature* **556**, 366–369 (2018).

777

93. McGlothlin, J. W., Kobiela, M. E., Wright, H. V., Mahler, D. L., Kolbe, J. J., Losos, J. B.,
& Brodie III, E. D. Adaptive radiation along a deeply conserved genetic line of least
resistance in Anolis lizards. *Evol. Lett.* 2, 310-322 (2018).

781

94. Houle, D., Bolstad, G. H., van der Linde, K., & Hansen, T. F. Mutation predicts 40
 million years of fly wing evolution. *Nature*, 548, 447-450 (2017).

784

95. Zalts, H., & Yanai, I. Developmental constraints shape the evolution of the nematode mid-developmental transition. *Nat. Ecol. Evol.* **1**, 1-7. (2017).

787

96. Pennell, M. W., Harmon, L. J. & Uyeda, J. C. Is there room for punctuated equilibrium in macroevolution? *Trends Ecol. Evol.* **29**, 23-32 (2014).

790

791 97. Hunt, G. Testing the link between phenotypic evolution and speciation: an integrated palaeontological and phylogenetic analysis. *Methods Ecol. Evol.*, **4**, 714-723 (2013).

793

98. Schluter, D. The ecology of adaptive radiation. OUP Oxford. (2000).

795

99. Rabosky, D. L., Santini, F., Eastman, J., Smith, S. A., Sidlauskas, B., Chang, J. & Alfaro,
 M. E. Rates of speciation and morphological evolution are correlated across the largest
 vertebrate radiation. *Nat. Comm.* 4, 1-8 (2013).

799

800 100. Erwin, D. H., Valentine, J. W. & Sepkoski, J. J. A comparative study of diversification events: the early Paleozoic versus the Mesozoic. *Evolution* **41**, 1177-1186 (1987).

802

101. Parins-Fukuchi, C., Stull, G. W. & Smith, S. A. Phylogenomic conflict coincides with rapid morphological innovation. *Proc. Natl. Acad. Sci. USA* **118**, e2023058118 (2021).

805

Stull, G. W., Qu, X. J., Parins-Fukuchi, C., Yang, Y. Y., Yang, J. B., Yang, Z. Y. *et al.* Gene duplications and phylogenomic conflict underlie major pulses of phenotypic
 evolution in gymnosperms. *Nat. Plants.* 7, 1015-1025 (2021).

809

810 103. Schluter, D., & Rieseberg, L. H. (2022). Three problems in the genetics of speciation by 811 selection. *Proc. Natl. Acad. Sci. USA* **119**, e2122153119.

813 104. Anderson, S. A., & Weir, J. T. Character displacement drives trait divergence in a continental fauna. *Proc. Natl. Acad. Sci. USA*, **118**, e2021209118 (2021).

815

816 105. Germain, R. M., Hart, S. P., Turcotte, M. M., Otto, S. P., Sakarchi, J., Rolland, J., *et al.* 817 On the origin of coexisting species. *Trends Ecol. Evol.* **36**, 284-293 (2021).

818

819 106. Venditti, C., & Pagel, M. Speciation as an active force in promoting genetic evolution.
 820 Trends Ecol. Evol. 25, 14-20 (2010).

821

822 107. Coyne, J. A. & Orr, H. A. Speciation. Sinauer, Sunderland, MA. (2004).

823

824 108. Rundell, R. J., & Price, T. D. Adaptive radiation, nonadaptive radiation, ecological speciation and nonecological speciation. *Trends Ecol. Evol.*, *24*, 394-399 (2009).

826

110. Rabosky, D. L. Phylogenetic tests for evolutionary innovation: the problematic link
between key innovations and exceptional diversification. *Phil. Trans. R. Soc. Lond.*,
372, 20160417 (2017).

830

111. Lanfear, R., Ho, S. Y., Love, D., & Bromham, L. Mutation rate is linked to diversification in birds. *Proc. Natl. Acad. Sci. USA* **107**, 20423-20428 (2010).

833

112. Dowle, E. J., Morgan-Richards, M. & Trewick, S. A. Molecular evolution and the latitudinal biodiversity gradient. *Heredity* **110**, 501-510 (2013).

836

113. Brown, J. H., Gillooly, J. F., Allen, A. P., Savage, V. M., & West, G. B. Toward a
 metabolic theory of ecology. *Ecology* 85, 1771-1789 (2004).

839

114. Orton, M. G., J. A. May, W. Ly, D. J. Lee & S. J. Adamowicz. Is molecular evolution faster in the tropics? *Heredity* **122**, 513-524 (2019).

842

115. Hull, P. M. & Norris, R. D. Evidence for abrupt speciation in a classic case of gradual evolution. *Proc. Natl. Acad. Sci. USA* **106**, 21224–21229 (2009).

845

116. Franks, S. J., Hamann, E., & Weis, A. E. Using the resurrection approach to understand
 contemporary evolution in changing environments. *Evol. Appl.* 11, 17-28 (2018).

848

849 117. Goldberg, E. E. & Igić, B. Tempo and mode in plant breeding system evolution.
850 Evolution 66, 3701-3709 (2012).

851

Wright, A. M., Bapst, D. W., Barido-Sottani, J., & Warnock, R. C. Integrating fossil
 observations into phylogenetics using the fossilized birth–death model. *Annu. Rev. Ecol. Evol. Syst.* 53, 251–73 (2022).

119. Manceau, M., Marin, J., Morlon, H. & Lambert, A. Model-based inference of punctuated molecular evolution. *Mol. Biol. Evol.* **37**, 3308-3323 (2020).

858

120. Obermeyer, F., et al. Analysis of 6.4 million SARS-CoV-2 genomes identifies mutations associated with fitness. *Science* **376**, 1327-1332 (2022).

861

121. Weber, M. G., Wagner, C. E., Best, R. J., Harmon, L. J. & Matthews, B. Evolution in a community context: on integrating ecological interactions and macroevolution. *Trends Ecol. Evol.* **32**, 291-304 (2017).

865

122. Aristide, L. & Morlon, H. Understanding the effect of competition during evolutionary radiations: an integrated model of phenotypic and species diversification. *Ecol. lett.* **22**, 2006-2017 (2019).

869

870 123. Nuismer, S. L., & Harmon, L. J. Predicting rates of interspecific interaction from phylogenetic trees. *Ecol. Lett.* **18**, 17-27 (2015).

872

124. Harmon, L. J., Andreazzi, C. S., Débarre, F., Drury, J., Goldberg, E. E., Martins, A. *et al.* Detecting the macroevolutionary signal of species interactions. *J. Evol. Biol.* 32, 769-782 (2019).

876

125. Blasco-Costa, I., Hayward, A., Poulin, R., & Balbuena, J. A. Next-generation
 cophylogeny: unravelling eco-evolutionary processes. *Trends Ecol. Evol.* 36, 907-918
 (2021).

880

126. Hembry, D. H., & Weber, M. G. Ecological interactions and macroevolution: a new field with old roots. *Annu. Rev. Ecol. Evol. Syst.* **51**, 215-243 (2020).

883

884 127. Phillimore, A. B. & Price, T. D. Density-dependent cladogenesis in birds. *PLoS Biol* **6**, 885 e71 (2008).

886

128. Morlon, H., Potts, M. D. & Plotkin, J. B. Inferring the dynamics of diversification: a coalescent approach. *PLoS Biol.* **8**, e1000493 (2010).

889

Liow, L. H., Reitan, T. & Harnik, P. G. Ecological interactions on macroevolutionary time scales: clams and brachiopods are more than ships that pass in the night. *Ecol. Lett.* 18, 1030-1039 (2015).

893

130. Silvestro, D., Antonelli, A., Salamin, N., & Quental, T. B. The role of clade competition
in the diversification of North American canids. *Proc. Natl. Acad. Sci. USA* 112, 8684-896
8689 (2015).

131. Condamine, F. L., Silvestro, D., Koppelhus, E. B., & Antonelli, A. The rise of
angiosperms pushed conifers to decline during global cooling. *Proc. Natl. Acad. Sci. USA* 117, 28867-28875 (2020).

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935

- 902 132. Harmon, L. J. & Harrison, S. Species diversity is dynamic and unbounded at local and continental scales. *Am. Nat.* **185**, 584-593 (2015).
- 905 133. Rabosky, D. L., & Hurlbert, A. H. Species richness at continental scales is dominated by ecological limits. *Am. Nat.* **185**, 572-583 (2015).
- 908 134. Etienne, R. S., Haegeman, B., Stadler, T., Aze, T., Pearson, P. N., Purvis, A., & Phillimore, A. B. Diversity-dependence brings molecular phylogenies closer to agreement with the fossil record. *Proc. Roy. Soc. B* **279**, 1300-1309 (2012).
- 912 135. Etienne, R. S. & Haegeman, B. A conceptual and statistical framework for adaptive 913 radiations with a key role for diversity dependence. *Am. Nat.* **180**, E75-E89 (2012). 914
- 915 136. Rabosky, D. L. Diversity-dependence, ecological speciation, and the role of competition 916 in macroevolution. *Annu. Rev. Ecol. Evol. Syst.* **44**, 481-502 (2013).
- 918 137. Harmon, L. J., Schulte, J. A., Larson, A., & Losos, J. B.Tempo and mode of 919 evolutionary radiation in iguanian lizards. *Science* **301**, 961-964 (2003).
- 138. Tobias, J. A., Cornwallis, C. K., Derryberry, E. P., Claramunt, S., Brumfield, R. T., &
 Seddon, N. Species coexistence and the dynamics of phenotypic evolution in adaptive
 radiation. *Nature* 506, 359-363 (2014).
- 925 139. Drury, J. P., Clavel, J., Tobias, J. A., Rolland, J., Sheard, C. & Morlon, H. Tempo and mode of morphological evolution are decoupled from latitude in birds. *PLoS Biol.* **19**, e3001270 (2021).
- 929 140. Carvalho, M. R., Jaramillo, C., de la Parra, F., Caballero-Rodríguez, D., Herrera, F., 930 Wing, S., *et al.* Extinction at the end-Cretaceous and the origin of modern Neotropical 931 rainforests. *Science* **372**, 63-68 (2021).
- 933 141. Reitan, T., Schweder, T. & Henderiks J. Phenotypic evolution studied by layered stochastic differential equations *Ann. Appl. Stat.* **6**, 1531–1551 (2012).
- 936 142. Xu, L., Van Doorn, S., Hildenbrandt, H. & Etienne, R. S. Inferring the effect of species interactions on trait evolution. *Syst. Biol.* **70**, 463-479 (2021).
- 143. Tóth, A. B., Lyons, S. K., Barr, W. A., Behrensmeyer, A. K., Blois, J. L., Bobe, R., *et al.* Reorganization of surviving mammal communities after the end-Pleistocene
 megafaunal extinction. *Science* 365, 1305-1308. (2019).

943 144. Luo, M., Ji, Y., Warton, D., & Yu, D. W. Extracting abundance information from DNA-944 based data. *Mol. Ecol. Resour.* **23**, 174-189. (2023).

945

946 145. Li, H. & Durbin, R. Inference of human population history from individual whole-947 genome sequences. *Nature* **475**, 493-496 (2011).

948

949 146. Ferriere, R. & Legendre, S. Eco-evolutionary feedbacks, adaptive dynamics and evolutionary rescue theory. *Phil. Trans. R. Soc. Lond* **368**, 20120081 (2013).

951

952 147. Odling-Smee, F. J., Laland, K. N., & Feldman, M. W. Niche construction. In *Niche Construction*. Princeton university press (2013).

954

148. Ardetti, J., Elliott, J., Kitching, I.J. & Wasserthal, L.T. 'Good Heavens what insect can
 suck it' – Charles Darwin, Angraecum sesquipedale and Xanthopan morganii praedicta.
 Bot. J. Linn. Soc. 169, 403-432 (2012).

958

959 149. Harvey, M. G., Seeholzer, G. F., Smith, B. T., Rabosky, D. L., Cuervo, A. M. & Brumfield, R. T. Positive association between population genetic differentiation and speciation rates in New World birds. *Proc. Natl. Acad. Sci. USA* **114**, 6328-6333 (2017).

962

150. Freeman, B. G., Strimas-Mackey, M., & Miller, E. T. Interspecific competition limits bird species' ranges in tropical mountains. *Science* **377**, 416-420. (2022).

965

151. FitzJohn, R. G. Diversitree: comparative phylogenetic analyses of diversification in R.
 Methods Ecol. Evol. 3, 1084-1092 (2012).

968

152. Freeman, B. G., Rolland, J., Montgomery, G. A. & Schluter, D. Faster evolution of a
 premating reproductive barrier is not associated with faster speciation rates in New
 World passerine birds. *Proc. Roy. Soc. B* 289, 20211514 (2022).

972

973 153. Rolland, J., Schluter, D., & Romiguier, J. Vulnerability to fishing and life history traits 974 correlate with the load of deleterious mutations in teleosts. *Mol. Biol. Evol.* **37**, 2192-975 2196 (2020).

976

977 154. Ogbunugafor, C. B., Wylie, C. S., Diakite, I., Weinreich, D. M. & Hartl, D. L. Adaptive 978 landscape by environment interactions dictate evolutionary dynamics in models of drug 979 resistance. *PLoS Comput. Biol.* **12**, e1004710 (2016).

980

981 155. Lewontin, R. C. The units of selection. Annu. Rev. Ecol. Evol. Syst. 1, 1-18 (1970).

- 983 156. Pimiento, C., Bacon, C. D., Silvestro, D., Hendy, A., Jaramillo, C., Zizka, A. et al.
- Selective extinction against redundant species buffers functional diversity. *Proc. Roy.*
- *Soc. B* **287**, 20201162 (2020).

987 157. Vrba, E. S., & Gould, S. J. The hierarchical expansion of sorting and selection: sorting and selection cannot be equated. *Paleobiology* **12**, 217-228. (1986).

990 158. Gould, S. J. Gulliver's further travels: the necessity and difficulty of a hierarchical theory of selection. *Philos. Trans. R. Soc. Lond.* **353**, 307-314. (1998).

993 159. Erwin, D. H. Macroevolution is more than repeated rounds of microevolution. *Evol.* 994 *Dev.* **2**, 78–84 (2000).

160. Gould, S. J. (1989). Wonderful life: the Burgess Shale and the nature of history. WW
 Norton & Company.

999 161. Blount, Z. D., Lenski, R. E., & Losos, J. B. Contingency and determinism in evolution: 1000 Replaying life's tape. *Science* **362**, eaam5979 (2018).

1002 162. Grant, P. R. & Grant, B. R. Unpredictable evolution in a 30-year study of Darwin's finches. *Science* **296**, 707-711 (2002).

1005 163. Uyeda, J. C., Zenil-Ferguson, R., & Pennell, M. W. Rethinking phylogenetic comparative methods. *Syst. Biol.* **67**, 1091-1109. (2018).

1008 164. Li, J., Huang, J. P., Sukumaran, J., & Knowles, L. L. Microevolutionary processes impact macroevolutionary patterns. *BMC evol. biol.* **18**, 1-8 (2018).

Figure 1. Time-scale dependent rates of molecular evolution, phenotypic evolution, and speciation using phylogenetic or fossil data. Molecular evolution is shown in panels A and E, phenotypic evolution in panels B and F, speciation rate using phylogenetic data in panels C and G and speciation rate using fossil data in panels D and H. All rates appear faster nearer the present and slower over longer timescales. Both axes in E, F, G, H have been log-transformed in order to show the long-term trend. This figure has been recreated from the data provided in^{19,27,62}. Darwin values of zero were removed in the panel F to avoid infinite values. Note that the timescales are different for each panel with an acceleration in the last 100kyr for genes, the last million of years for morphology, and the last tens of Myrs for speciation rates, while all relationships are linear when time is log-transformed. Why this pattern is shared between different measures of evolution is not well understood². In the literature, the "paradox of stasis" refers to morphological changes (panels B and F). The x-axis represents time intervals between calibration points (A and E), time intervals between measurements (B and F), clade ages (C and G), or clade durations (D and H).

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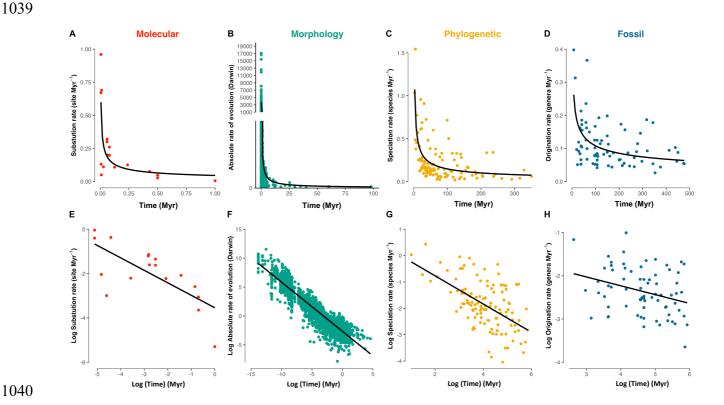


Figure 2. Current and fossil data represent past evolutionary processes incompletely. Reconstructions of past molecular evolution, phenotypic evolution and diversification are biased because most of the information about past evolutionary changes is lost from molecular, phenotypic, phylogenetic and fossil history. Panel A shows how mutations in red cannot be reconstructed when several mutations affect the same position in the genome, a phenomenon called saturation. The comparison between a present-day genome and the genome of a sister species (or a sample of ancient DNA) only allows reconstructing mutations shown in green, biasing rate estimates if the method fails to take saturation into account, as shown in panel C, compared to the true rate in panel B. There may be other reasons why polymorphisms are not recorded over the long term, such as deleterious mutations not persisting or lost polymorphisms in extinct populations (not represented here). Similarly, phenotypic changes through geological time can only be accurately reconstructed when the fossil record is exceptionally well preserved. Reconstructed phenotypes based on phylogenies and fossils (panel E) will lead either to wrongly assigning a fossil to a branch when it belonged to another extinct lineage (e.g., fossil 1), or to missing shifts in phenotype that are not recorded in the fossil information. Phenotypes reconstructed only using phylogenetic data (F) will also largely miss past evolutionary changes. Finally, reconstructed phylogenies based on present-day data (H) are also likely missing most of the past speciation and extinction events (G). The observed lineagethrough-time plot (I) obtained with the phylogenetic tree (black) is usually very far from the truth (blue).

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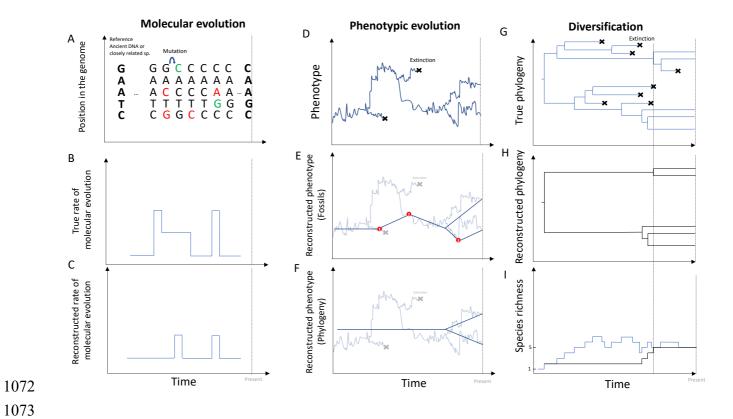
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Box 1. How to connect macroevolutionary trends to underlying population parameters?

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The structure and the dynamics of evolving populations within species affect speciation and extinction rates. The probability of speciation is influenced by the probability of population establishment or splitting, the probability that populations persist long enough for speciation to occur, as well as the rate of evolution of reproductive barriers. The probability of species extinction is influenced by the number of populations and their size and genetic diversity. With the increasing availability of population parameters estimated for a large number of species (e.g. from population genomic datasets), there are growing opportunities to build more 'mechanistic' and potentially more informative comparative methods that include such details. Several comparative methods have already been developed to model trait evolution while accounting for intraspecific variance (jive model³⁸⁻³⁹) or to model diversification accounting for incipient lineages (protracted speciation model⁴¹), but they do not directly test the effects of population parameters on speciation and extinction rates. Trait-dependent diversification models (SSE models¹⁵¹) could be used for this exact purpose. SSE models were originally developed to test the effect of species traits on diversification¹⁵¹ with the probability of speciation and the probability of extinction at each point in time being a function of the species' trait value. They generally estimate the likelihood of trait-dependent speciation and extinction rates, given a phylogenetic tree and species' traits measured at the present time. Transition rates between character states reflect both mutation and selection, although those two components are not explicitly modeled. One could thus treat microevolutionary parameters as species' traits to estimate their impact on diversification rates. For instance, the probability of speciation is expected to depend on the rate of establishment of new populations and the probability of population splitting¹¹. Traits that could be used as microevolutionary proxies in diversification analyses include the number of populations within a species, the rate at which populations differentiate 149 (e.g., using the slope of isolation by distance, estimated from F_{ST})9, or the rate at which reproductive isolation builds (e.g., via song evolution in birds¹⁵²). Similarly, the probability of species extinction is expected to be a function of the long-term population size¹⁵³ and the probability that populations persist¹¹. Although estimates of persistence are rare, they are much needed and could be potentially estimated from high-quality fossil records or approximated from other traits implicated in extinction risk, such as effective population size, genetic diversity or range size. We thus need more studies to understand how those proxies relate to persistence/extinction and differentiation/speciation. Importantly, these comparative models provide insights on how those difficult-to-collect but crucial microevolutionary proxies could relate to differentiation/speciation and persistence/extinction.

Box 2. Four evolutionary landscapes. In the literature, four evolutionary landscapes have been described at different scales (Figure 3). At the microevolutionary level, one can either map the fitness of each individual onto trait values (the fitness function; e.g., 84) or the mean fitness of a population as a function of gene frequencies or mean phenotype (adaptive landscape 85). The fitness function and adaptive landscape are usually measured from experiments or more rarely in natural ecosystems 84,77. Peaks in the adaptive landscape represent genotypes/trait values where mean fitness is maximized, and valleys represent genotypes/phenotypes that are detrimental. The adaptive landscape concept is more complicated when selection is frequency-dependent, as when species interact.

Macroevolutionary surfaces are functions that represent probabilities of phenotypic change over time. They lack a fitness interpretation but can be a useful summary of the outcome of the adaptive evolution of component species. The Brownian motion model, whereby instantaneous change is equiprobable in all directions, is represented by a flat macroevolutionary surface. It fits cases in which the squared difference between species increases linearly with time. The Ornstein-Uhlenbeck process fits data in which differences between species over time increasingly behave as though randomly sampled from a fixed underlying gaussian frequency distribution. The model can be represented by a concave downward surface with a single "peak" that is essentially the long-term mean of the underlying frequency distribution of species phenotype values. More complex surfaces can have multiple "peaks", which fit data for which the long-term frequency distribution of species differences is best described by a mixture of probability distributions rather than a single distribution. For instance, Mahler et al. ⁸⁸ showed that phenotypes of species in different lineages of the *Anolis* phylogeny clustered around the same set of "peaks" inferred from the distribution of species trait means, indicating convergence.

"Species selection" surfaces describe the rates of speciation and extinction as a function of species mean trait values. The trait-dependent speciation and extinction can be estimated by fitting a model to data on trait means and branching rates in reconstructed phylogenetic trees^{89,151}. The concept can be applied to model speciation/extinction rates as a function of discrete or quantitative traits. The speciation and extinction rates are not interpretable on a fitness scale, but a lineage is more likely to persist if it splits often into daughter lineages and/or

has a low extinction rate (see previous debates on species selection^{89,155}). A multi-trait framework still requires further development for quantitative characters (but see¹⁵⁶ for fossil data, and MuSSE for discrete traits¹⁵¹). Additionally, some methods based on the analysis of the fossil record also allow modeling a changing "species selection" surface (*evoTS* and *layeranalyzer* packages).

Box 3. Why microevolution may not be coupled with macroevolution. There are many reasons why all the details of microevolution would not predict macroevolution. The first reason is that large macroevolutionary patterns may emerge unpredictably from microevolutionary processes (e.g., the extinction risk of lineages may reflect unusual bouts of strong selection, rather than the average rate of selection). Classic philosophical discussions about challenges to the predictability of macroevolution from microevolution center on the hierarchical nature of biological organization, and the potential for properties to emerge at higher levels of organization (e.g. species) that cannot be explained at a lower organizational level 156-159. A key element is that selection can change direction at different scales (section 2). Although the intensity of these debates has lessened, these issues remain and there is no consensus as to the relative frequency by which microevolutionary processes can predict macroevolution, and when they cannot.

In his book *Wonderful life*¹⁶⁰, S.J. Gould proposed to "replay the tape of life" to know whether long-term evolution would follow the same trajectories over and over. Decades after this book, most evolutionary biologists would still argue that it is very unlikely that life will evolve following the same exact trajectory twice¹⁶¹. First, there is a large amount of stochasticity in any (complex) biological system, with survival, reproduction, and mutation all subject to chance¹⁶². For example, the COVID-19 pandemic hinged upon the chance of transmission of the virus in its first human hosts. Similarly, the time at which adaptations arise may be unpredictable, particularly for mutations that have arisen only once in the history of life¹⁶³. Second, there is a great deal of stochasticity and contingency in the abiotic environment, which affects both microevolutionary processes (e.g., lightning striking an individual) and macroevolutionary processes (e.g., a meteorite leading to a mass extinction¹⁵⁹). Another important problem is that several microevolutionary processes may lead to the same macroevolutionary pattern¹⁶⁴, which may lessen the identifiability of the models. Accounting for the microevolutionary processes underlying macroevolutionary patterns may also be challenging when several evolutionary processes operate simultaneously (e.g., population

bottlenecks and periods of strong selection). Even if individual and population-level processes may never totally explain broad-scale biodiversity patterns, we should strive to understand which general rules at macroevolutionary scales can inform microevolution and vice versa.

Figure 3. Four representations of evolutionary landscapes and surfaces at different scales related to phenotype. Panel A and B represent the fitness function and the adaptive landscape at the microevolutionary scale. Panel A represents the fitness of all individuals in a population as a function of their trait values (e.g., 84). Panel B represents the fitness averaged at the level of populations as a function of the mean trait value in a population (the so-called adaptive landscape 85). Panel C and D represent surfaces at the macroevolutionary scale. Panel C represents the likelihood of the parameter Θ, representing the attraction point of an OU process estimated using the frequency distribution of species trait values and a phylogeny or a time series. The peak is often interpreted as an "optimum" or "adaptive peak," but it is better thought of as the mean of the underlying long-run frequency distribution of species' trait values. Panel D represents the rates of speciation and extinction as a function of lineage trait values, which are sometimes considered as parameters describing "species selection" on traits 89. The locations of "peaks" need not correspond between different landscapes.

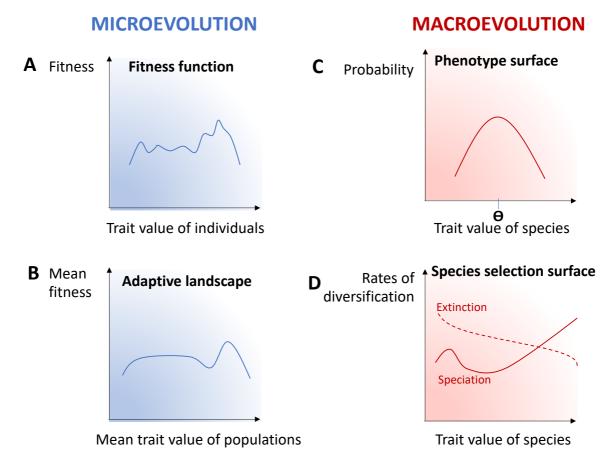


Figure 4. A changing microevolutionary adaptive landscape integrated over a macroevolutionary time scale. The relationship between the adaptive landscape, governing the evolution of populations, to the macroevolutionary surface, representing probabilities of change in species means over time, is not well understood. Panel A shows a hypothetical adaptive landscape for a single trait at different slices of time. Each time slice describes mean fitness of a single population (color goes from blue/low fitness to red/high fitness). Panel C shows the ensuing changes in mean phenotype of the population over time, as it roughly tracks the changing adaptive landscape (e.g., as recorded in a fossil sequence). Panel B represents the average of the adaptive landscape through time, for each row of panel A over the entire time period. The value of Θ , the attractor of the Ornstein-Uhlenbeck (OU) process is estimated by fitting the model to the distribution of species values over time (not shown here). In this case, Θ can be regarded as a summary of the outcome of adaptive evolution (e.g., as described across a fossil sequence). In phylogenetics, Θ is usually estimated from a clade of species, each of which might have its own unique adaptive landscape with a trajectory through time (one panel A per lineage). Here too, Θ can be regarded as a summary of the outcome of lower-level processes giving rise to the distribution of species values over the long run. This figure shows how a macroevolutionary model of phenotypic evolution may be summarizing complex microevolutionary processes.

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