

Towards microbiota-based disease management: analysis of grapevine microbiota in plots with contrasted levels of downy mildew infection

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1. Context and hypotheses

Vineyards are colonized by a myriad of microorganisms that interact with each other and with grapevine plants^{1,2}. Some microorganisms are plant pathogens, such as the oomycete Plasmopara viticola that causes grapevine downy mildew. Others have a positive influence on vine health, such as plant growth promoting bacteria and disease biocontrol agents. These beneficial interactions have gained more attention in recent years because they could represent an alternative to the use of fungicides in viticulture³.

To develop microbiota-based disease management, we aim to cultivate the myriad of microorganisms that colonize grapevine leaves and then assemble multi-strain microbial consortia that could protect vines against *P. viticola*. To increase the probability of isolating microorganisms of interest, we developed a plantbased culture medium. We tested the hypothesis that it increases the cultivable microbial diversity in comparison to standard culture media.

2. Materials and methods

We combined metabarcoding (Fig.1(2.a)) and culture-dependent (Fig.1(2.b)) approaches to analyze the microbiota of soil and leaf samples collected in plots with contrasted levels of downy mildew infection.



Bibliography

- 1. Bettenfeld et al., 2021. J. Adv. Res.
- 2. Fournier, Pellan et al., 2022. Adv. Ecol. Res.
- 3. Busby *et al.*, 2017. *PLOS Biol*.

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Downy mildew

Plasmopara viticola

Oomycete

Fig.3: Bacterial genera identified using MALDI-TOF MS in grapevine leaf samples, depending on the culture media (GLA, LBA and TSA). Abundance is the number of bacterial isolates belonging to each genus. Bacterial genera identified in only one, two or three culture media are circled in red, green and orange, respectively.

We were able to cultivate and isolate >1000 bacterial strains from grapevine leaves. Diversifying the culture media allowed us to isolate more strains. After completing the Sanger sequencing of all microbial isolates (including yeasts and filamentous fungi; Fig. 1 (2.b)), we will analyze the complementarities between the grapevine leaf-based culture medium and standard culture media. We will test the hypothesis that the vine-based medium selects for microbial taxa that are different from those selected on standard media and/or poorly referenced in the current version of the MALDI-TOF MS database. Statistical analysis of metabarcoding data (Fig. 1 (2.a)) will provide an estimate of the % of cultivable taxa and will allow us to identify microbial taxa that are significantly more abundant in vineyard plots with low levels of downy mildew infection. Those microorganisms will be retrieved from our microbial collection. Their effect on P. viticola will be tested in vitro and in planta, as single strains and as consortia. Modes of action will be elucidated for the most promising microbial consortia (those that thrive on grapevine leaves, are resilient to disturbances and have a biocontrol activity against *P. viticola*).

MALDI-TOF MS Identification of bacteria

3. Results and discussion

A total of 1262 bacterial isolates were analyzed by MALDI-TOF, of which 60% could be identified at least to genus level (Fig.2).

Strains isolated from GLA and LBA were less well assigned than those isolated from TSA (higher % of non-identification and lower % of assignment to the species level) (Fig.2).

• A total of 32 bacterial genera were identified on the three culture media. Eight genera were isolated on all three media (Fig.3).

The genera Gordonia and Rahnella were isolated only on GLA (Fig.3).



4. Conclusion and perspectives



Fig.2: Number of bacterial strains isolated on each culture media and percentage of taxonomic assignments by MALDI-TOF MS.