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Biomimetic photocatalysts for the transformation of CO₂: Design, properties and mechanistic insights

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Abstract

The increase in CO_2 concentration in the atmosphere due to the intensive burning of fossil fuels, for the needs in energy of our modern society, has strongly contributed to the global warming. One of the promising strategies to fight this menace is to capture atmospheric CO₂ and convert it into value added products, including fuels and chemicals. Among the methods developed for the transformation of CO₂, the sunlight-activated photocatalytic conversion appears as the renewable and easy to use method. For this purpose, it is required to have at disposal proper materials to act as catalysts and light absorbing materials to harvest the sun energy, and then to combine these components with an electron carrier, in an efficient way in order to access the desired fuels by a photocatalytic sustainable CO₂ conversion. Among the efforts in research dedicated to the production of solar fuels, studies focused on the design and development of bio-based photocatalytic systems as alternative to conventional photocatalytic systems have emerged in the literature in the last decade. The objective of this review is to show the readers the recent advances in the fabrication of bio-based photocatalysts for the transformation of CO₂ into value added products. This review specifically focuses on the conversion of CO₂ into carbon monoxide, methanol, formic acid/formate and acetic acid which are the main target products described. Methods adopted to enhance the selectivity and efficiency of the bio-derived photocatalysts developed are also discussed and a section is dedicated to computational

investigations. Finally, the current challenges and perspectives in the development and use of these novel and greener photocatalysts are presented.

Keywords: CO₂ transformation; solar fuels; biomimetic photocatalysts; enzymatic catalysis

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1. Introduction

Up to recent period, the burning of fossil fuels has been a major source of energy of our modern society. The inordinate consumption of fossil resources has led to a severe global warming (with new records in the last three years) due to the excessive discharge of greenhouse gasses into the atmosphere, and most particularly of carbon dioxide (O=C=O), and this has become an international crisis that calls for an urgent solution. CO₂ is the primary driver of the global warming [1]. In addition, it is a long-lived greenhouse gas, with one-third still present in the atmosphere after a century of release [2]. Scientists believe that if purposeful action is not taken with immediate alacrity to lower CO_2 concentration in the atmosphere, global warming and impact of climate change could inevitably lead to the death of more than 250,000 people per year, put 100 million people into abject poverty by 2030 and displace 150 million people or more by 2050 [3]. The transformation of atmospheric CO_2 is a herculean task because of its high stability. However, it may have significant benefits in terms of access to carbon based platform molecules and fuels [4]. Among the various strategies available for the reduction of CO₂, photocatalytic transformation of CO₂ is unarguably a promising and green strategy that could solve the global challenge [5]. The reduction of CO₂ by photocatalysis is similar in mechanism to the natural photosynthesis, which can be explained using three separate but linked steps: (i) the

absorption of solar energy (ii) the separation of charges and (iii) surface redox reaction. Utilization of the ubiquitous solar energy in order to reduce CO₂ into renewable value-added products would in turn simultaneously improve the lingering global energy shortage, climate change and take a step towards the circular economy [6]. However, the major bottlenecks of existing photocatalysts (i.e. monocatalysts, liquid state Z-scheme, all solid states Z-scheme, direct Z-scheme and type II heterojunctions) are (1) fast recombination of photogenerated charge carriers, (2) poor redox power and (3) Inefficient charge separation thereby hindering the practical application of the technique [7-9]. According to general agreement, a good photocatalytic system must have quick charge transition, efficient charge separation, and good absorption in the visible spectrum, be composed of non-toxic elements and display high stability [10]. Among the photocatalysts reported in the literature, bio-based photocatalysts have gained popularity in the recent decade. This is mainly due to their high reaction efficiency and their fascinating catalytic mechanism inspired by nature [11]. In comparison to their photosynthetic counterparts, non-photosynthetic organisms, especially those created in contemporary synthetic biology, have more desirable metabolic pathways, providing a tunable CO₂ metabolism for a variety of products [12]. Other interests of the bio-based photocatalytic systems for the transformation of CO₂ into hydrocarbon fuels emerge from the abundance of the biomaterials used, environmental friendliness, and energy-saving if the process is activated by sunlight, to mention but a few. In the reviewing exercises that cover materials for application as catalysts in the transformation of CO₂, the recent advancements in the use of bio-based photocatalysts is only little highlighted. For example, Yau and co-workers [13] recently published an updated review which gives precedence to microbial hybrid photosystems for CO₂ fixation. Notwithstanding, the review work did not provide elaborate survey on other bio-based photocatalysts and it fails to address the theoretical investigation in order to rationalize the mechanism of CO₂ reduction.

In order to sensitize the scientific community to the advantages of using bio-derived photocatalysts (i.e. a natural light driven catalytic reaction using enzyme, bacterial and polymers) for the conversion of CO_2 , this review intends to gather the recent research (i.e. covering the last decade) on the design of such photocatalytic systems for the transformation of CO_2 into fuels. The examples reported are classified into five sections in line with the target hydrocarbon fuel, namely carbon monoxide, methanol, formic acid/formate and acetic acid (see sections 2, 3, 4 and 5, respectively). Furthermore, in section 6, an in-depth analysis on the interest of computational

investigations for the development of bio-derived photocatalysts for CO_2 reduction is given. Computational data may either complement or to give prior understanding into experimental investigations. Finally, current challenges and future outlook are discussed.

2. Biomimetic photocatalysts for the transformation of CO₂ to carbon monoxide

Carbon monoxide (CO) is one of the attractive value-added products that can be generated from the reduction of CO_2 . It is commonly used as an intermediate in some chemical processes including the production of drugs, fragrances, formaldehyde or benzaldehyde and methanol [14]. Some chemical industries also use CO in the preparation of inorganic chemicals such as metal carbonyls like nickel tetracarbonyl (**Eq. 1**) and iron pentacarbonyl (**Eq. 2**).

$Ni + 4CO \rightarrow Ni(CO)_4$	(1)
$Fe + 5CO \rightarrow Fe(CO)_5$	(2)

Furthermore, CO plays an important role alongside with hydrogen gas, forming together the syngas that is used in the Fischer-Tropsch process towards superior hydrocarbons [15].

From thermodynamic and kinetic points of view, the generation of CO from CO₂ is one of the easiest compared to that of other hydrocarbon fuels. The reduction of CO₂ into CO and H₂O requires two electrons and two protons with a reduction potential of 0.53V (vs. NHE at pH = 7) (**Eq. 3**).

$$CO_2 + 2H^+ + 2e^- \rightarrow CO + H_2O \ E = 0.53V$$
 (3)

Different bio-based photocatalytic materials have been employed for the conversion of CO_2 to CO. In one of the earliest studies, Woolerton et al. developed a photocatalytic system that comprises metal-oxide nanoparticles functionalized by the enzyme carbon monoxide dehydrogenase (CODH) as catalyst and sensitized to visible light by a ruthenium bipyridyl photosensitizer, both components being supported on an anatase/rutile TiO₂ mixture as the semiconductor (electron donor). Using this photocatalyst activated by visible light they could effectively reduce CO₂ to CO [16]. The authors examined the CO production rates and stability as a function of each component (photosensitizer, enzyme and TiO₂). Tolerance to O₂ and effects of different electron donors were also investigated, together with strategies to control enzyme binding at the surface of TiO₂ in order to enhance overall activity (**Fig. 1a**). Ding and co-authors [17] reported a functional porous coordination hybrid system composed of nickel porphyrin and

nickel bis(terpyridine) dual active sites as efficient bio-based photocatalyst for CO₂ reduction into CO under the illumination of visible light ($\lambda \ge 420$ nm) (**Fig. 1b**). The biomaterials exhibited an extraordinary photocatalytic activity towards the CO formation with an outstanding generation rate of 3900 µmolh⁻¹g⁻¹ and selectivity of 98%. Also, they investigated the mechanism of action associated with dual active sites of the system, composed of metal porphyrin and metal bis(terpyridine), using theoretical simulations which further supported an excellent photocatalytic performance of these bio-hybrids as observed in the experimental investigation (**Fig. 1c**). The theoretical investigation demonstrated that the binding between CO₂ and Ni(II) in nickel bis(terpyridine) is much easier than that with nickel porphyrin. The Ni–C Mayer bond order of *CO₂ intermediate in nickel bis(terpyridine) was 0.533, much higher than that (0.009) in nickel porphyrin, which accounts for the stability of the intermediate of the former.



Figure 1. Mechanism of CO_2 reduction to CO by (a) CODH/ruthenium bipyridyl/anatase or rutile-TiO₂. Reproduced with the permission from [16]. Copyright 2011 Royal Society of Chemistry. (b) Ni Porphyrin and Ni bis(terpyridine) coordinated polymer (TPP-Ni) (c) Schematic mechanism of CO_2 reduction catalyzed by Ni Porphyrin and Ni bis(terpyridine) . Adapted with the permission from [17]. Copyright 2022 American Chemical Society.

In a related study, Liang and co-workers [11] developed a biomimetic system based on metalloporphyrin and ZrO_2 nanoframes that showed an evolution CO yield of 35.3 µmol within 3 h of reaction and high selectivity rate of 93.1% as well as an apparent quantum efficiency of 1.84% which is 60.8 greater than pristine porphyrin-Ni with evolution rate of 0.58 µmol.

The group of Zhao [18] constructed a covalently bonded reaction centre_bio-hybrids comprised of Co-based porphyrin and a low molecular mass graphitic C_3N_4 for the transformation of CO_2 to CO under the irradiation of visible light. They obtained a photocatalytic activity 13 times greater than that of pure C_3N_4 (CO generation rate of 17 µmolg⁻¹h⁻¹), thus evidencing the positive role of the Co porphyrin. The improved activity was attributed to the efficient electron transfer and trapping by CO active sites, improved electron-hole separation and strong affinity of Coporphyrin for CO₂ adsorption.

Lin and colleagues [19] designed a highly efficacious $g-C_3N_4$ /tetra(4-carboxyphenyl)porphyrin (III) chloride heterogeneous catalyst for selective photoreduction of CO₂ into CO under visible light (**Fig. 2a**). In this catalytic system they used abundant, environmentally friendly FeTCPP as the catalytic center and inexpensive $g-C_3N_4$ nanosheets as the light absorber, maintaining a simple mechanical mix under constant stirring. A maximum rate of 6.52 mmol g^{-1} of CO in 6 h, with up to 98% selectivity was obtained. The catalytic performance was attributed to an effective electron transfer from $g-C_3N_4$ nanosheets to FeTCPP

Apart from the enzymitic/bacterial based photosystems reviewed so far, Zhou and co-authors [20] reported biomimetic polymeric semiconductor based hybrid nanosystems that combined a typical CO₂/proton reduction catalyst, a cocatalyst, an electron mediator, and a CO₂ activator for artificial photosynthesis with water and CO₂. g-C₃N₄ nanosheets, which mimic the nanolayered thylakoids stacks, are shown to be promising photocatalytic elements with a planar configuration and a high surface area, and they serve as an excellent platform for the assembly of other

analogous elements (**Fig. 2b**). Au NPs are used as a suitable cocatalyst. ZIF-9 is used as a CO₂ concentrator and an electron mediator to promote the redox reaction. The highest production of CO and H₂ was achieved by ZIF-9(1%)Au-CNS at the evolution rates of 0.3 and 7.5 μ molh⁻¹g⁻¹, respectively. The study revealed that the incorporation of polymeric based g-C₃N₄ and ZIF-9 significantly enhanced the CO evolution rates which is 122 folds higher compared to non-ZIF-9 modified systems.



Figure 2. Plausible mechanism of CO_2 photoreduction by (a) g-C₃N₄/FeTCPP hybrids (b) Photoconversion activity of C₃N₄/AuNPs/ZIF-9. Reproduced with the permission from [19] and [20]. Copyright 2018, 2016 Elsevier, respectively.

Table 1 below outlines biomimetic photocatalysts that have been explored for CO production from CO_2 reduction. The synergistic incorporation of biomimetic photocatalysts has shown to be promising for the reduction of CO_2 to CO with high selectvity. However, the present bio based photocatalytic systems for CO production have not provided satisfactory activity and stability, key points that necessitate further research.

 Table 1. Carbon monoxide production using bio-based photocatalysts

Photocatalytic system	Reaction conditions CO		Comments	Reference
		yield/TON		
TTPP-Ni/metal based	Photocatalytic activity	3900 μmolh ⁻	The	[17]
bis(terpyridine) with	under $\lambda \geq 420$ nm,	¹ g-1	composites	
dual active sites	trisopropanolamine		exhibited high	
	employed as		CO yield.	
	sacrificial agent, 15mg		However,	
	catslyst		there is partial	
			dissolution of	
			photocatalysts	
Zn-porphyrin sensitised	Aqeuous medium, red	Low yield	The slow	[21]
Mn (I)	light irradiation (λ =		conversion rate	
	620 nm)		of CO_2 to CO	
			was atributed	
			to inadequate	
			e- migration	
			from porphyrin	
			to the catalyst	
Carbon monoxide	Carried out under	Good yield	The size and	[22]
dehydrogenase	viisible light		morphologgy	
functionalized with	irradiation		of the	
CdS nanocrystals			nanocrystals	
			greatly	

			enhanced the
			actvity
Nafion assisted Re(I)-	The catalytic activity	454 µmolh ⁻¹ g ⁻	The presence [23]
RuL	was done under	1	of nafion
	visibile light for 20h,		improved the
			photocatalytic
			activity
Polymeric based g-	80 kPa of pure CO_2	$0.4 \ \mu molh^{-1}g^{-1}$	The dominant [20]
C ₃ N ₄ /AuNPs/ZIF-9	gas was purged, 300W		evoluted gas is
	Xe arc lamp as light		H ₂ which is
	source		not favourble
			for the reaction
g-C ₃ N ₄ /FeTCPP	Under visible light	6.52 mmolg ⁻¹	The findings [19]
	greater than 420 nm.		indicates that
	TEOA was used as		the electrons
	sacrificial agent		migrated from
			$g-C_3N_4$ to
			FeTCPP
ZrO ₂	Visible light	35.3 µmol	The bio based [11]
nanoframes/metallopor			material
phyrins			efficiently
			transfer
			electrons to the
			Ni catalytic
			sites which in
			turn increased
			the CO
			generation

TON: Turn over frequency, FeTCPP: tetra(4-carboxyphenyl)porphyrin (III) chloride

3. Biomimetic photocatalysts for the transformation of CO₂ to methanol

Methanol or methyl alcohol (CH₃OH) is an organic compound primarily used as a solvent, cosolvent, or feedstock in the chemical industry. Methanol has an octane number of 113 and a density that is half that of the gasoline, as thus, can be mixed with gasoline with no technical modifications and be used in methanol-fuelled vehicles [24]. Although methanol is considered an environmentally friendly fuel (no oxides of sulphur or nitrogen), the production of CO during its combustion limits its use as a pure fuel (100%). Hence it is mixed with gasoline on a ratio of 85% (methanol) - 15% (unleaded gasoline) [25]. More usages of methanol include hydrogen storage and transportation as an energy carrier as well as an easily transportable fuel, and also production of intermediates and synthetic hydrocarbons as a C1 building block [26]. Methanol is therefore regarded as a transition molecule from fossil fuels to renewable energies.

Methanol can be synthesized from carbon-containing materials such as natural gas, coal, biomass, and carbon dioxide (CO₂). Natural gas is currently the primary source of methanol contributing approximately 90% to the world 111 million metric ton production [27]. Unfortunately, natural gas and coal are fossil fuel resources, thus depleting non-renewable resources and significantly contributing to greenhouse gas emissions. The burning of fossil fuels (natural gas, coal, and oil) leads to CO_2 emissions which contribute significantly to global warming. Therefore, green production of methanol has gained a great attention of academic and industrial researchers.

Catalytic hydrogenation of CO_2 has been proposed as a green methanol production process. This process falls under the carbon capture and utilization (CCU) strategy which uses catalysts in the reaction of CO_2 and hydrogen (**Eq. 4**). The transformation of CO_2 to methanol is considered a green process when using captured CO_2 and/or hydrogen from renewable energy resources. The study by González-Garay et al. [28] shows how much CO_2 emissions can be avoided when using the green methanol synthesis process. However, the green methanol production *via* CO_2 hydrogenation is not economically feasible compared to the fossil-based process due to high costs of renewable hydrogen [28]. Therefore, there is an urgent need for developing cost effective CO_2 conversion technologies to produce methanol in a green way. Amongst many CO_2 conversion technologies such as biological, thermochemical, and electrochemical routes, the photochemical process has gained extensive attention [29]. If using solar light as its source of energy, the photocatalytic reduction of CO_2 into valuable chemicals like methanol may be a sustainable approach [30–34].

 $CO_2 + 6H^+ + 6e^- \longrightarrow CH_3OH + H_2O$ (4)

Enzymatic conversion of CO₂ requires energy source like for the photochemical processes. The energy sources can be diverse, including ultraviolet, electrical potential, or energy carrying molecules (adenosine triphosphate (ATP), guanosine triphosphate (GTP), nicotinamide adenine dinucleotide (NADH)). While in a photochemical process, an electron donor, a photosensitizer, and catalysts (e.g. TiO₂, ZnO, graphene, etc.) are mainly required to drive selective production to methanol [35,36] [37–40]. These photocatalysts may be incorporated into biomaterials such as biochars and biopolymers with conductive properties (e.g. cellulose, chitosan, etc.) to improve their structural and electronic properties [32,41–43]. There have been few case studies on biomimetic photocatalysts for the reduction of CO₂ to methanol since 2015 due to cost of enzyme production, regeneration, nonetheless, the research area is slightly increasing (as seen in **Table 2**).

Recently, Tseng and co-workers [44] utilised sugarcane bagasse (SB) to support graphitic carbon nitride (gCN) for photocatalytic reduction of CO₂ to CO and methanol using a 14-watt LED light bulb. The presence of lignin in SB resulted in the semiconductor-like absorption edge and provided the interaction with gCN. The use of SB alone for photocatalytic study resulted in the formation of small amounts of MeOH. This is linked to the electronic transition of conjugated species within lignin (from SB) thus providing photoinduced charge carriers on SB. The introduction of SB (containing lignin) influenced the semiconductor-like absorption edge properties of the photocatalyst. Catalyst synthesis, g-SB-HT, using water (g-SB-HT-W) and ethanol (g-SB-HT-E) influenced MeOH yields, as the later solvent may be seen to have acted as hole scavenger thus increasing the yield of methanol (see Table 2).

Tseng and co-workers [45] also developed titanium oxide-chitosan film hybrid material (TiO₂chitosan) for the photocatalytic CO₂ reduction to methanol. This TiO₂-chitosan material was able to selectively convert CO₂ in methanol at a production rate of 0.37 nmol/g (**Table 2**), showing a good stability. Similarly, Kavil and co-workers [46] synthesized a TiO₂-based hybrid photocatalyst (Cu-C(glucose)/TiO₂) by co-doping TiO₂ with copper (from copper sulfate) and carbon (from glucose) to photocatalytically reduce CO₂ to methanol. The photocatalytic experiment was performed in a Pyrex glass photoreactor filled with the catalyst and a NaOH suspension and then purged with certified super–critical fluid grade carbon dioxide for 30 min. The photoreactor was irradiated with UV lamps (365 nm wavelength) for 5 h. In these conditions the Cu-C/TiO₂ catalyst allowed to achieve the highest methanol production (~2600 μ M) with compared to counterparts C/TiO₂ (TiO₂ doped with only carbon) and TiO₂ P25, thus evidencing the synergy effect between the three components, Cu, C and TiO₂.

Studies have shown that enzymes catalysed photoreduction of CO₂ to methanol involves using a three-step enzyme cascade including formate dehydrogenase (FDH), formaldehyde dehydrogenase (AldDH or FDH), and alcohol dehydrogenase (ADH) [46]. Co-enzymes and co-factors are usually sacrificial for photoredox reactions and are depleted in the process, thus, requiring enzymes that may regenerate the reduced forms. To further improve methanol yield, sequential and co-immobilization of dehydrogenases with polymeric membranes, metal complexes and oxides have been proposed [47].

Biocatalysts such as enzymes, namely formate (FDH), aldehyde (AldDH), formaldehyde (FalDH), lactate (LDH) and alcohol (ADH) dehydrogenases was tested for CO₂ reduction to methanol [47,48]. Co-enzymes (also known as co-factors) such as nicotinamide adenine dinucleotide (NAD⁺) or its reduced form (NADH) and methylviologen (MV²⁺) are often used with FDH, NADH, AldDH and ADH. [48][49–51]. Amao and co-workers [36] developed a photochemical enzyme-based conversion system for the CO₂ photoreduction to methanol, using FDH, AldDH, and ADH combined with MV²⁺ under the visible-light photosensitization of water-soluble porphyrin, tetraphenylporphyrin tetrasulfonate (H₂TPPS) in the presence of triethanolamione (TEOA) (**Fig. 3**). This complex photoredox system allowed to achieve a methanol production concentration of 6.8 μ M. Study of the enzymatic kinetics evidenced that the rate-limiting step is the formic acid to formaldehyde reaction with AldDH and reduced form of MV²⁺ (Methyl viologen).



Figure 3. Methanol production from CO_2 catalysed with H₂TPPS/MV²⁺/TEOA/FDH, AldDH and ADH photoredox system. Reproduced with permission from reference [36]. Copyright 2018 Elsevier.

The use of co-enzyme nicotinamide adenine dinucleotide reduced form (NADH), FDH catalyses both the reduction of CO_2 to formic acid, ADH catalyses the reduction of formic acid to formaldehyde, and ADH catalyses both the reduction of formaldehyde to methanol (Fig 4). The resulting co-enzyme nicotinamide adenine dinucleotide (NAD⁺) catalyses the oxidation of formic acid to CO_2 , formaldehyde to formic acid and oxidation of methanol to formaldehyde, respectively, thus inhibiting methanol production due to the redox nature of NAD⁺/NADH (Fig. 4). The oxidation processes were suppressed using oxidized form of MV²⁺, an electron carrier molecule, to drive the photochemical production of methanol. A reaction scheme for methanol production from CO_2 with FDH, AldDH and ADH using natural co-enzyme NAD⁺/NADH and artificial co-enzyme MV²⁺/MV⁺ is presented in **Figure 4**.



Figure 4. Scheme for the methanol production from CO_2 with FDH, AldDH and ADH using natural co-enzyme NAD+/NADH and artificial co-enzyme MV^{2+}/MV^+ .

Zezzi Do Valle Gomes and co-workers [48] reported the biocatalytic cascade conversion of CO_2 to methanol using three dehydrogenases (FDH, ADH and FalDH) co-immobilized in tailored siliceous mesostructured cellular foams (MCF) functionalized with mercaptopropyl groups (MP) with no cofactor. With this catalytic system they obtained a methanol yield of 0.35 mmol/g_{enzyme}/min, a value which might be improven by adding NADH regeneration.

Yadav et al. [52] prepared a CCG-IP type photocatalyst by attaching isatin-porphyrin (IP) chromophores with chemically converted graphene (CCG). A methanol production concentration of 11.21 μ M was achieved using CCG-IP under visible light with an NADH photoregeneration of 39%.

Ji et al. [53] developed a polyelectrolyte-doped hollow nanofibers/microcapsules-based integrated photosynthesis system. Basically, the system was designed by assembling the components involved in the electron-transfer chain in a specific order, such that the photo-excited electrons can be transported efficiently in the opposite direction. This system was integrated in a device in the following order: three dehydrogenases and a cofactor (FDH, FalDH,

ADH and NAD(H)) in the innermost layer, electron mediators (M) in the middle layer, a photosensitizer (EY⁻ Eosin Y) in the outermost layer and an electron donor (TEOA) for the CO₂ conversion to methanol under solar energy (depicted in **Fig. 5**). They achieved a methanol production of 60.37 μ M (hollow mixture of poly(allylamine hydrochloride) (PAH) and polyurethane (PU) nanofiber-based system) and 61.98 μ M (microcapsules-based system) with 86.04% NADH photo-regeneration.



Figure 5. Schematic illustration of preparation of polyelectrolyte-doped hollow nanofibers/microcapsules-based integrated photosynthesis system for the solar-driven CO_2 transformation to methanol. Reproduced with permission from reference [53]. Copyright 2016 John Wiley & Sons.

Note that in different works, the co-immobilization of NADH with multi-enzymatic catalytic systems allowed to attend a higher methanol production efficiency over a single enzyme-based system. Furthermore, addition of photosensitisers, inorganic metal oxides and electron mediators as well as electron donors improved the photoreduction of CO_2 to methanol [53] [36].

Photocatalytic system	Reaction conditions	CH ₃ OH yield	Reference
Sugarcane bagasse (SB)	CO ₂ was purged	110 µmol/g (g-	[44]
supported on graphitic carbon	through water bubbler	SB-HT-E)	
nitride (gCN)	and photocatalyst and	15-20 µmol/g (g-	
(SB-modified gCN)	irradiated for 12 h	SB-HT-W)	
	with 14 W LED		
Chitosan-TiO ₂	CO ₂ was purged	0.37 nmol/g	[45]
	through water bubbler		
	and photocatalyst and		
	irradiated for 12 h		
	with UV light		
Cuprous Oxide Nanoparticles	CO ₂ was purged	1656.7 µmol/g	[54]
Coated with Aminated Cellulose	through NaOH (120		
(Cu ₂ O/NCC-EDA)	mL of 0.5 mol L^{-1})		
	and photocatalyst (0.1		
	g) suspension and		
	irradiated for 12 h		
	with 300 W Xenon		
	lamp		
CuO/ ZnO-embedded	Catalyst Films were	0.030 mmol/g	[55]
carbohydrate polymer films	irradiated for 6 h with	0.037 mmol/g	
(CuO/ZnO-Den-TOCNF)	UV light at 5cm		
(CuO/ZnO-CS)	distance from the		
	closed reactor bottle		
	where water was		
	dropped and dry ice		
	(440–880 mg) was		
	held		
Cu-loaded carbon modified	NaOH (0.2 M) and	2600 µM	[46]
titanium oxide (Cu-C/TiO ₂).	catalyst suspension		

Table 2. Methanol production from CO₂ using biomimetic photocatalysts

Carbon from glucose	was purged with CO_2		
	for 30 min and		
	irradiated for 5 h with		
	UV lamps		
Tetraphenylporphyrin	Suspension of	6.8 µM	[36]
tetrasulfonate with	H ₂ TPPS (100 μM),		
methylviologen, triethanolamine	MV ²⁺ (2.0 mM),		
and dehydrogenases (H ₂ TPPS/	TEOA (0.3 M), FDH		
MV ²⁺ /TEOA/FDH/AldDH/ADH)	(2.0 µM), AldDH (2.0		
	$\mu M)$ and ADH (2.0		
	μ M) in CO ₂ saturated		
	50mM sodium		
	pyrophosphate buffer		
	was irradiated for 100		
	min with 250W		
	halogen lamp		
Dehydrogenases	10 mL buffer solution	60.37 μM	[53]
(FDH/FalDH/ADH) encapsulated	(50 mM PBS, pH	(hollow	
in hollow nanofibers and	7.0), 15% (w/v)	nanofibers) and	
microcapsules	TEOA, 25 mg hollow	61.98 μM	
	nanofibers or	(microcapsules)	
	microcapsules, 0.1 mg	86.04% NADH	
	mL ⁻¹ FateDH, 0.1 mg	photoregeneration	
	mL ⁻¹ FaldDH, 0.1 mg		
	mL^{-1} ADH, 0.2 mM		
	NADH, 0.25 mM M,		
	and 50 μM EY		
	irradiated for 10 h		
	with 450 W Xenon		
	lamp		
Chemically converted graphene	CO ₂ (0.5 mL/min), β-	11.21 µM	[52]

attached to isatin-porphyrin and	NAD ⁺ (1.24 μ mol),	38.99% NADH
coupled with dehydrogenases	Rh (0.62 µmol),	photoregenration
(CCG-IP/ FalDH/FDH/ADH)	TEOA (1.24 mmol),	
	photocatalyst (0.5	
	mg), 9 units of each	
	enzyme (FDH,	
	FalDH, and ADH) in	
	3.1 mL of sodium	
	phosphate buffer (100	
	mM, pH 7.0)	
	irradiated for 30 min	
	with visible light	
Siliceous mesostructured cellular	CO ₂ , MFC-MP,	1.35 mmol/g/min [48]
foams (MCF) functionalized with	FateDH (0.05 mg	
mercaptopropyl groups (MP) and	ml^{-1}), FaldDH (0.75	
co-immobilized with	mg ml $^{-1}$), ADH (3.75	
dehydrogenases	mg ml ^{-1}), NADH	
(MFC-MP/FDH/ADH/FaldDH)	(100 mM), and	
	$KHCO_3\ (200\ mM)$ in	
	100 mM phosphate	
	buffer (pH 6.5)	

4. Biomimetic photocatalysts for the transformation of CO₂ to formic acid/formate

Formic acid (FA) also known as methanoic acid is a colourless, sour, pungent-smelling, and fuming liquid, with as chemical structure HCOOH. If this weak acid is toxic in case of skin and oral exposure, it is non-toxic and stable at room temperature conditions [56,57]. A variety of wood ants and bees produce large quantities of FA naturally, to protect their nest by spraying at prey [58]. FA is one of the most important chemicals as it has found application in a wide range of industries. It has been used for example as an anti-bacterial reagent, food additive, textile dyeing, leather tanning, rubber, finishing, drilling fluids, anti-icing, fragrance ingredient,

cleaning products, pH modifier, and corrosion inhibitor. It is also used as organic synthesis reagent in various industrial chemical processes [59–61].

Due to the high industrial demand for FA, its production at high scale crucial. Formerly, FA was produced as a by-product of acetic acid production through hydrocarbon oxidation which has been reduced after the implementation of the carbonylation of methanol method for acetic acid production. FA production was then fulfilled via hydrolysis of formamide, this method however was economically unfeasible due to the consumption of sulfuric acid and ammonia as well as ammonium sulfate formation [56]. Finally, the carbonylation of methanol followed by hydrolysis of methyl formate (**Eq. 5 and 6**) has been the main industrial process applied for FA production [62][63].

$$CH_3OH + CO \rightarrow HCO_2CH_3$$
 (5)
 $HCO_2CH_3 + H_2O \rightarrow HCOOH (FA) + CH_3OH$ (6)

However, this conventional production process has encountered predicaments, being the work-up of the hydrolysis mixture the main issue. Also, in the course of separating unreacted methyl formate, re-esterification of FA and methanol to methyl formate happens very quickly due to the unfavourable equilibrium [64,65]. Finally, this conventional method has negative environmental impacts, and high energy and economic costs [66,67]. As a result, alternative ways of producing FA from other sources such as the reductive conversion of CO₂ are extensively researched [68– 70]. Transformation of CO₂ to FA can significantly reduce carbon emissions and assist in fighting climate change whilst finding applications in hydrogen storage, chemical feedstock, and fuel in direct formic acid fuel cells (DFAFCs) [71][72]. Several methods allow the reduction of CO₂ to FA among which photochemical [73][74], electrochemical (ECR) [75][68], hydrogenation [69][76], and biological processes [77][78], stand a good position. The attractivity of the biomimetic photocatalytic transformation of CO₂ for FA production derives the abundance of biomaterials, environmental friendliness, and energy-saving to mention a few advantages. The mostly used biocatalyst is an enzyme, mainly the D-2-hydroxy acid dehydrogenase (FDH) that is found on methylotrophic microorganisms like bacteria and yeast that can use methanol. Tishkov and Popov reported that the FDH from bacterial sources exhibit 1.7 times higher activity than yeast [52]. The FDH enzyme catalyses the oxidation of formate to CO_2 and converts NAD⁺ to NADH [79][80]. The cofactor β -nicotinamide adenine dinucleotide in its reduced form (NADH)

is normally needed by FDH enzymes to attain the reduction of CO_2 to HCOOH according to reaction (**Eq. 7**) below [81]. Around twenty studies that deal with the reduction of CO_2 into FA utilizing bio-based photocatalytic systems have been reported between 2012 and 2022 (see Table 3). These studies increased sharply from 2018 and exhibited promising conversion systems and high FA production yields.

$$CO_2 + NADH \leftrightarrow ^{CbFDH}HCOO^- + NAD^+$$
 (7)

Yu et al. reported an effective and environmentally friendly light-driven enzymatic nanosystem [82]. This catalytic system consists of a combination between graphitic carbon nitride $(g-C_3N_4)$ photocatalyst and two enzymes (carbon anhydrase (CA) and formate dehydrogenase (FateDH/FDH)), encapsulated in ZIF-8 MOF (CA&FateDH@ZIF-8/g-C₃N₄). Fig. 6 illustrates the possible mechanism where photocatalytic NADH regeneration and enzymatic hydrolysis reduced CO₂ as described by the authors. Upon light irradiation, CA&FateDH@ZIF-8/g-C₃N₄ is excited to generate electron-hole pairs which then receive two electrons from the oxidation of one molecule of TEOA which are then excited from the valence band (VB) to the conduction band (CB) by light. The electron transport complex (M) was used to produce more 1,4-NADH, the biological entity needed by the enzymes. Thus, the highly selective M transports two electrons and one photon to NAD⁺ for NADH regeneration. With the two enzymes, CA quickly converted CO₂ into HCO₃ which is converted into HCOOH using FateDH which, at the same time, consumes NADH as shown in Fig. 6. Reduction of CO2 with CA&FateDH@ZIF-8/g-C3N4 system resulted in 198 µM of FA whilst the system without CA and free FateDH produced only 75 and 38 µM, respectively. When 2 mg.mL⁻¹ of CA&FateDH@ZIF-8/g-C₃N₄ was utilized, the optimized yield could reach 243 µM of FA. The activity of the enzymatic activity was retained above 80% after 10 runs, thus indicating the reusability of this catalytic system, that was also reported as able to shorten the distance between photocatalyst and enzyme entities, that is beneficial to the diffusion of substances and electrons.



Figure 6. Illustration of electron and energy transfer processes for light-driven enzymatic CA&FateDH@ZIF-8/g-C₃N₄ nanosystem. Reproduced with permission from Reference [82]. Copyright 2021 Elsevier.

Wang et al. [83] reported a CO_2 reduction system consisting in two enzymes as well i.e. CA and FDH. Using a catalyst that combines two enzymes has been found to regulate the reaction rate and increase the loading amount of FDH which allows to decrease the ratio of of CA/FDH. This is one of the routes enhancing FA production. Sustaining a stable pH environment for this system in the course of CO_2 hydration was mandatory as the process releases hydrogen ions. The increase in the hydration rate of CO_2 increased the yield of FA by aspect of 4.2-fold. However, the costs of cofactor and enzymes are economically unfeasible and enzyme immobilization and cofactor regeneration appeared as a probable solution for low costs, recyclability, and enhanced biocatalyst stability.

A FDH-bearing hollow fiber membrane (HFM) reactor (**Fig. 7**) has been developed by Gu et al. [70] for CO₂ reduction which led to high-efficient and sustainable FA production. They used a UV-TiO₂ photocatalytic coenzyme regeneration system with a FDH-bearing HFM reactor which exhibited a maximum initial reaction rate of 1.04 mM.h⁻¹ and 81.7% yield. The optimization of operating conditions and NADH concentration provided a turnover number of 125 after 4.5 h. Considering that one mole of NADH is consumed to generate one mole of FA, NADH regeneration for continuous enzymatic production is important. Thus, the system regenerated

NADH when the reaction mixture was transported into a photocatalytic cell. This resulted in an instant regeneration of NADH which eventually showed that even at a low loading amount of NADH high FA production could be achieved.



Figure 7. Illustration of a photocatalytic/enzymatic HFM system. Reproduced with permission from Reference [70]. Elsevier 2020.

Zhang et al. built an artificial thylakoid consisting of protamine-titania (PTi) micro-capsules with cadmium sulfide quantum dots (CdS QDs) and enzymes (FDH extracted from *Candida boidinii* (CbFDH)) for FA production from CO₂ [84]. The improvement of NADH regeneration activity $(4226\pm121 \ \mu molg^{-1}h^{-1})$ and enzyme compatibility of CdS/PTi microcapsules contributed to high FA production. Compartmentalization of photogenerated holes and reactive oxygen species (ROS) by the structure of CdS/PTi microcapsules protected FDH from deactivation, resulting in high compatibility between bio- and photo-catalyst. This catalytic system exhibited a quantum yield of $0.66\pm0.13\%$ of FA over multiple light-dark cycles.

Zhang et al. reported a highly selective reduction system with a metal-free heterojunction of pyromellitic diimide/g-C₃N₄ (PDI/CN) coupled with FDH enzyme [85]. **Fig. 8** shows the possible mechanism of the photoenzymatic catalytic cascade system on the PDI/CN heterojunction. In this cyclic process, the photocatalyst could continuously reduce NAD⁺ to NADH while FDH could oxidize NADH to NAD⁺, concurrently, in order to accelerate CO_2 transformation to FA. The process can successfully advance continual FA production. Another

study reported the utilization of $[Cp*Rh(bpy)H_2O]^{2+}$ as a cocatalyst since the biologically inactive 1.6-NADH and the dimers of NAD⁺ may be formed under the action of the intermediate radical. Thus, it was proven that $[Cp*Rh(bpy)H_2O]^{2+}$ was effective for the selective production of biologically active 1,4-NADH [70]. Zhang et al. [85] have also shown herein the promoting effect of $[Cp*Rh(bpy)H_2O]^{2+}$ in their system. The NADH yield increases linearly with the increase in sunlight irradiation time, and the system with $[Cp*Rh(bpy)H_2O]^{2+}$ showed better performance signifying that this cocatalyst assists NAD⁺ to accept photogenerated electrons thereby accelerating electron transfer to generate NADH. The outstanding performance of this photocatalytic system was ascribed to good conductivity, heterointerface effect, broad sunlight response range of the catalyst as well as a distinctive photoenzymatic catalytic cascade system. The system achieved 75% NADH yield and 114.8 µmol for FA production.



Figure 8. Possible mechanism of the photoenzymatic catalytic cascade system on the PDI/CN heterojunction. Reproduced with permission from Reference [85]. ACS Publications 2021. Several studies concern the use of carbon-based materials such as carbon nanodots-silica hybrid, graphene film, graphene oxide modified with cobalt metallated aminoporphyrin, and C_{60} polymer film as photocatalysts [86][33][87][34]. The interest in carbon materials emanates from their wide availability, environmental friendliness, and good light harvesting properties. Carbon nanodots were reported to have excellent biocompatibility, high solubility, and low toxicity whilst GO properties include biological function, thermal, mechanical, and biomedical [88].

Carbon nanodots-silica hybrid/FDH and graphene film/FDH systems showed NADH regeneration and FA yield of 74.10 \pm 0.17%, 203.33 \pm 1.9 µmol [86], and 71.8%, 228.6 µmol [33], respectively. For the FDH@GO-Co-ATPP system, NADH regeneration and FA yield of 48.53% and 96.49 µmol, respectively were reported [87] whilst the C₆₀-based polymeric/FDH system exhibited 91.07% of NADH regeneration and 239.46 µmol [34].

Ji et al. reported chloroplast-mimicking TCPP/SiO₂/Rh HNP-based wholly integrated APS and AM/M/BP HNS-based integrated Z-scheme APS models in 2018 [89] and 2020 [90]. These systems decreased the transfer distance of the energy and electron among the enzymatic redox and light-harvest system as well as enabling sufficient charge separation and photogenerated excitons transportation. In both studies, the effect of the TCPP/SiO₂/Rh HNPs and AM/M/BP HNS-based Z-scheme APS was confirmed by detecting photochemical, optical, and electrochemical properties. The TCPP/SiO₂/Rh HNPs system showed NADH regeneration which enhanced from 11 - 75% and FA production which increased from $10 - 100 \mu$ mol. AM/M/BP HNS-based Z-scheme APS, on the other hand, exhibited significant enhancement of NADH regeneration and FA production which reached 89% and 365 μ mol, respectively [89][90].

The use of graphitic carbon nitride has been on the rise as well. This nitrogen-containing material possesses exceptional solar light-harvesting photocatalytic ability due to its slow recombination ability which increases photo-responsive energy transfer from the dye to a $g-C_3N_4$ moiety [91]. It has high stability, a tuneable band structure, strong visible light response, and astounding biocompatibility [92]. The systems reported by Cheng et al. [93], Zhang et al. [92], Singh et al. [91], and Kumar et al. [94] polymeric carbon nitride (PCN) mediated by a tannic acid/polyethyleneamine (TA/PEI), Rh complex NH₂-functionalized polymeric carbon nitride, 1-4,-diaminoanthaquinone monomer using a modified CN, and Se-doped CN nanosheets, respectively, showed FA production of 2.1 mM, 5.0 mM, 344.12 µmol, and 200.26 µmol.

Tian et al. [51] assembled a functionally compartmental inorganic photocatalyst-enzyme system that imitated chloroplast. Photoexcitation and enzymatic CO_2 reduction were compartmentalized by the thylakoid membrane with the purpose of protecting enzyme from photodamage whilst the system enables electron transfer thereby fostering photocatalyst. They built a tightly integrated artificial photosystem comprising of Rh complex cocatalyst onto thiophene-modified C_3N_4 (TPE- C_3N_4) and FDH encapsulated with MAF-7 which is a subclass of zeolitic imidazolatebased MOF. The improved NADH regeneration of the system was attributed to the tightly integrated structure which promoted fast electron transfer from TPE-C₃N₄ to the Rh complex. MAF-7 not only protected FDH from photoinduced deactivation but also improved enzymatic stability in photosynthesis conditions. This photocatalytic system resulted in 16.75 mM FA after 9 h of illumination. **Table 3** shows the summary of the studies done on the transformation of CO₂ to FA using bio-inspired photocatalysts and the progress is promising.

Catalyst	Reaction conditions	Yield	Reference
Bromophenol-Bakelite (BPB)/Rh	$NAD^+ - 1.24 \ \mu M, \ BPB \ -$	NADH	[95]
complex/FHD	$0.031 \ \mu M$, AsA – 1.24	regeneration –	
	μM , Rh complex – 0.62	82.52%	
	µM, sodium phosphate		
	buffer – 3.1 mL	FA yield – 220	
		μmol	
Bamboo-shape magnesium	Photocatalyst – 15 μL in	NADH	[50]
tetraphenyl porphyrin-based	3.1 mL of sodium	regeneration –	
framework (Mg-TPPBF)/FDH	phosphate buffer (100	73.9%	
	mM, pH 7.0), β -NAD ⁺ –		
	124 µL, rhodium complex	FA yield – 197.2	
	$-$ 62 μ L, FDH $-$ 3 units,	µmol in 2 h	
	$AsA - 310 \ \mu L$		
1,4-diaminoanthraquinone (AQ)	$NAD^+/NADP^+ - 1.24$	NADH	[91]
monomer using a modified carbon	µmol, CpM – 0.62 µmol,	regeneration –	
nitride (CN) (AQBCN)/FDH	AsA – 0.1 mmol, FDH – 3	97.21/95%	
	units and the AQBCN		
	photocatalyst - 0.5 mg in	FA – 344.12	
	3.1 mL of SPB (100 mM;	μmol	
	pH, ~7.0)		
Rh complex covalently grafted onto	Photocatalyst – 12 mg,	NADH	[92]
NH ₂ -functionalized polymeric	NAD – 120 $\mu L,\ PBS$ –	regeneration –	

Table 3. Biomimetic photocatalysts transformation of FA from CO₂.

carbon nitride (NH ₂ -PCN) (Rh _{m3} -	2.88 mL, FDH – 1 unit	62.4%	
N-PCN)/FDH			
		FA yield – 5.0 mM	
CA&FateDH@ZIF-8/g-C ₃ N ₄	CA&FateDH@ZIF-8/g-	FA yield – 243	[82]
	$C_3N_4-2\ mg.mL^{\text{-1}} \text{in a}$	μM	
	0.1 M PBS, TEOA – 10		
	w/v%, mediator – 1		
	mg.mL ⁻¹ , β -NAD ⁺ – 0.8 m		
Pyromellitic diimide/g-C ₃ N ₄	FDH – 3 units, catalyst –	NADH	[85]
(PDI/CN)/FDH	10 mg, TEOA - 15 wt.%,	regeneration –	
	phosphate buffer solution	75%	
	(pH 7.4, 100 mM) - 10		
	mL, β -NAD ⁺ – 0.1 mM,	FA yield – 114.8	
	$[Cp*Rh(bpy)-(H_2O)]^{2+}$ –	μmol	
	0.25 mM		
Polymeric carbon nitride (PCN)	Photocatalyst – 0.5	NADH	[93]
mediated by a tannic	$mgmL^{-1}$, FDH – 2	regeneration –	
acid/polyethyleneimine (TA/PEI)	$mgmL^{-1}$, $NAD^+ - 1 mM$,	37.8%	
(PCN@TA/PEI-Rh	PBS buffer (pH 7.0, 100		
core@shell)/FDH	mM), free Rh – 0.5	FA yield – 2.1	
	mgmL ⁻¹ , and CO ₂ – 0.3	mM	
	MPa.		
Covalent-organic framework/2,4,6-	Photocatalyst – 0.5 mg, β -	NADH	[96]
triformylresorcinol (COF-4)/FDH	NAD ⁺ - 1.24 mmol, AsA	regeneration –	
	– 0.1 mmol, Rh complex	94.6%	
	- 0.62 mmol, and 3 units		
	of FDH in 3.1 mL of 100	FA yield – 226.3	
	mM buffer (pH 7.0). CO ₂	μmol	
	flow rate -0.5 mLmin^{-1}		
	for 1 h in the absence of		

light		
NADH – 2mM, liquid	FA yield –	[70]
velocity -40 mL.min ⁻¹ ,	81.7%	
gas velocity – 3 mL.min ⁻¹		
TEOA – 15 w/v%, AM	NADH	[90]
NSs or BP NSs $-$ 0.2	regeneration –	
$mgmL^{-1}$, M – 0.1 mM,	35±4%	
FDH $-$ 0.3 units, NAD $^{\scriptscriptstyle +}$ $-$		
0.1 mM	FA yield –	
	$160\pm24\ \mu mol$	
NAD^+ g $-$ 1 mM,	FA yield – 16.75	[51]
phosphate buffer - 100	mM after 9 h	
mM (pH 7), TEOA – 15		
w/v%, MAF@FDH - 7.5	5^{th} cycle FA –	
mg, TPE-C ₃ N ₄ /PEI/Rh $- 3$	~76%	
mg		
FDH@Rh-NU-1006 –	FA yield – 0.144	[97]
1.8×10^{-3} M in Tris buffer	±0.003 in 24 h	
(2 mL), TEOA – 0.5 mM,		
$NAD^+ - 1 \ mM$		
CO_2 flow rate – 0.5	NADH	[34]
mLmin ⁻¹ , β -NAD ⁺ – 1.24	regeneration –	
µmol, TEOA – 1.24	91.07%	
mmol, 3 units of FDA in		
3.1 mL of sodium	FA yield –	
phosphate buffer (pH 7.0,	239.46 µmol	
100 mM)		
[CO ₂] – 0.3 MPa, 37 °C,	NADH	[84]
visible light illumination	regeneration –	
$(\lambda = 405 \pm 5 \text{ nm})$	93.03±3.84%	
	light NADH – 2mM, liquid velocity – 40 mL.min ⁻¹ , gas velocity – 3 mL.min ⁻¹ TEOA – 15 w/v%, AM NSs or BP NSs – 0.2 mgmL ⁻¹ , M – 0.1 mM, FDH – 0.3 units, NAD ⁺ – 0.1 mM NAD ⁺ g – 1 mM, phosphate buffer – 100 mM (pH 7), TEOA – 15 w/v%, MAF@FDH – 7.5 mg, TPE-C ₃ N ₄ /PEI/Rh – 3 mg FDH@Rh-NU-1006 – 1.8×10 ⁻³ M in Tris buffer (2 mL), TEOA – 0.5 mM, NAD ⁺ – 1 mM CO ₂ flow rate – 0.5 mLmin ⁻¹ , β-NAD ⁺ – 1.24 µmol, TEOA – 1.24 µmol, 3 units of FDA in 3.1 mL of sodium phosphate buffer (pH 7.0, 100 mM) [CO ₂] – 0.3 MPa, 37 °C, visible light illumination (λ = 405±5 nm)	light NADH - 2mM, liquid FA yield - velocity - 40 mL.min ⁻¹ , 81.7% gas velocity - 3 mL.min ⁻¹ TEOA - 15 w/v%, AM NADH NSs or BP NSs - 0.2 regeneration - mgmL ⁻¹ , M - 0.1 mM, $35\pm4\%$ FDH - 0.3 units, NAD ⁺ - 0.1 mM FA yield - 0.1 mM FA yield - 160±24 µmol NAD ⁺ g - 1 mM, FA yield - 16.75 phosphate buffer - 100 mM after 9 h mM (pH 7), TEOA - 15 w/v%, MAF@FDH - 7.5 w/v%, MAF@FDH - 7.5 5 th cycle FA - mg, TPE-C ₃ N ₄ /PEI/Rh - 3 ~76% mg FDH@Rh-NU-1006 FA yield - 0.144 1.8×10 ⁻³ M in Tris buffer ± 0.003 in 24 h (2 mL), TEOA - 0.5 mM, NADH NAD ⁺ - 1 mM CO2 flow rate - 0.5 NADH mLmin ⁻¹ , β-NAD ⁺ - 1.24 regeneration - - µmol, TEOA - 1.24 91.07% mmol, 3 units of FDA in 3.1 mL of sodium FA yield - - µmol, 100 mM) ICO2] - 0.3 MPa, 37 °C, NADH NaDH visible light illumination regeneration - -

	NAD^+ – 5 nm, TEOA –		
	400 nm, CdS/PTi – 1 gL ⁻¹ ,	FA yield - 1500	
	PBS buffer - pH 7.0, 100	μMh^{-1}	
	nm, CbFDH – 1 gL ⁻¹		
FDH@GO-Co-ATPP	GO-Co-ATTP - 0.7 mg,	NADH	[87]
	NAD ⁺ – 1.24 mol, TEOA	regeneration –	
	- 1.24 mmol, 3 units of	48.53%	
	FDH in 3 ml of (100 mM,		
	pH 7) sodium phosphate	FA yield – 96.49	
	buffer	μmol	
Porphyrin/SiO ₂ /Cp*Rh(bpy)Cl	Phosphate buffer solution	NADH	[89]
hybrid nanoparticles	(100 mM, pH 7.0) – 10	regeneration –	
(TCPP/SiO ₂ /Rh-HNPs)/FDH	mL, $NAD^+ - 1$ mM,	75%	
	TCPP $-$ 0.5 mM, FDH $-$		
	0.3 units mL ⁻¹ , $CO_2 - 0.3$	FA yield – 100	
	MPa	µmol after 4 h.	
Se-doped carbon nitride (C ₃ N ₄)	Photocatalyst – 0.5 mg,	NADH	[94]
nanosheets/FDH	$Rh - 0.62 \ \mu mol, \ \beta\text{-NAD}^+$	regeneration –	
	$-$ 1.24 μ mol, FDH $-$ 3	82.84%	
	units, AsA - 0.1 mmol in		
	3.1 mL of sodium	FA yield –	
	phosphate buffer (100	200.26 μ mol in	
	mM, pH 7.0)	1 h	
	CO_2 flow rate – 0.5		
	mLmin ⁻¹		
Graphene film photocatalyst	β –NAD ⁺ – 1.24 µmol, Rh	NADH	[33]
(GFPC)/FDH	complex $-$ 0.62 µmol,	regeneration –	
	FDH – 3 units in 3.1 mL	71.8%	
	of sodium phosphate		
	buffer (100 mM, pH 7.0),	FA yield – 228.6	
	AsA $- 0.1$ mmol, CO ₂	µmol in 2 h	

			flow rate -0.5 mLmin^{-1}	
Carbon	nanodots-silica	hybrid	Photocatalyst $-0.5 \text{ mg}, \beta$	- NADH [86]
(CNDSH)/FDH		NAD^+ – 1.24 mmol	, regeneration –
			rhodium complex - 0.62	2 74.10±0.17%
			mmol, ascorbic acid -0.1	
			mmol, FDH - 3 units in	n FA yield –
			3.1 mL of Na ₂ HPO ₄	- 203.33±1.9
			NaH ₂ PO ₄ buffer (100) µmol
			mM, pH 7.0)	
Cobaloxi	me/eosin/REOA/F	DH	Eosin Y – 0.10 mM	, FA yield – 36% [98]
			$NAD^+ - 0.50 \text{ mM}, \text{TEOA}$	
			-0.20 M, 5 units of FDH	I
			in 0.15 M phosphate	
			buffer (pH 7.0)	

5. Biomimetic photocatalyts for the transformation of CO₂ to acetic acid

Acetic acid otherwise known as vinegar acid, ethanoic acid and methane carboxylic acid is a clear, colourless liquid, of pungent smell and with the chemical formula CH₃COOH. It is the second simplest carboxylic acid after formic acid and is of high importance also due to a plethora of industrial and pharmaceutical applications. It is produced synthetically and it can also be prepared via bacterial fermentation. It is a crucial chemical reagent being primarily used for the production of cellulose acetate for photographic films and that of acetic anhydride. Acetic acid also possesses antiseptic properties, so it can be employed as antimicrobial agent. The current approach for the production of acetic acid is not environmental benign, with high energy cost and the production scale cannot meet the demand for both industrial and pharmaceutical needs [99,100]. Development of alternative production routes that will cover the current demand and be environmentally friendly is thus needed. One of the promising and cost-effective methods to produce acetic acid is the photocatalytic transformation of CO₂. Recent advancements in this domain are described below but note that only a few of case studies were found in the literature of the last decade.

Zhang and co-authors [12] integrated ultra-small gold nanoclusters (AuNCs) and cadmium sulfide (CdS) into M. thermoacetica bacteria and compared their performance in the photosynthesis of acetic acid from CO_2 (Fig. 9a). These bio-based photocatalytic systems were found efficient for CO₂ fixation into acetic acid under low light irradiation and dark (Fig. 9b). The authors observed that the yield in acetic acid by the gold-based biophotocatalyst after four days is conspicuously higher than that obtained with cadmium sulfide-based bio-photocatalyst under the irradiation of low light. Moreover, the authors found that the activity of ordinary bacterium (M. thermoacetica) and gold-based bio-photocatalyst in darkness are relatively low. A monitoring of the electron transfer mechanism via photoluminescence confirmed that the photoexcited electrons from the gold nanoclusters are migrated to distributed redox mediators in the bacteria cytoplasm such as flavoproteins. Evaluation of the effect of cysteine on the yield of acetic acid production revealed that the gold-based bio-photocatalyst produced the highest yield (6.01 mmolg⁻¹) at 0.3 % wt Cys. It is worth to mention that the ligands at the gold nanoclusters surface and the gold core size can be changed in order to tune their biochemical and biophysical characteristics, including cell uptake, cytotoxicity, biocompatibility, and molecular electronic structures. This study demonstrated that integration of gold nanoclusters as intercellular photosensitizer inside the cytoplasm of M. thermoacetica bacterium allowed this nonphotosynthetic latter to conduct effective photosynthesis of acetic acid from CO₂.



Figure 9 (a) Schematic illustration of *M. thermoacetica*/AuNCs hybrid system (b) Schematic bacteria electron transfer pathway for acetic acid generation under illumination of light

In another study, Gai et al. [101] developed an organic semiconductor – bacteria biohybrid photosynthetic system that efficiently conversed CO₂ into acetic acid. The bacterium involved is Moorella thermoacetica whose surface was coated with perylene diimide derivative (PDI) and poly(fluorene-co-phenylene) (PFP) as photosensitizers. This led to the formation of a p-n heterojunction (PFP/PDI) layer with high hole/electron separation efficiency (**Fig. 10**). The π conjugated semiconductors display excellent light-harvesting ability and biocompatibility. In addition, the cationic side chains of the two organic semiconductors could intercalate into cell membranes, thus allowing an efficient transfer of electrons to the bacterium. In this photocatalytic system, the bacterium could harvest photoexcited electrons from the PFP/PDI heterojunction and synthesize acetic acid from CO₂ under illumination. This bio-photocatalytic system showed an efficiency rate of 1.6% which is comparable to those of reported inorganic biohybrid systems.



Figure 10. Wood–Ljungdahl pathway for CO₂ reduction in the presence of PFP/PDI/M. thermoacetica. Reproduced with the permission from [101].Copyright 2020 Willey online

Tharak and colleagues [102] reported microbial-mediated CO_2 in gas and gas-electro fermentation systems with and without applied potential by selectively enriched biocatalysts (**Fig. 11**). In the Heat + CO_2 condition during the gas fermentation (GF) stage, the maximum production was 0.84 g/L acetic acid with 31 mg.L⁻¹.h⁻¹ inorganic carbon fixation rate (ICFR). The GF was tested in a single-chambered microbial electrochemical system (MES; Gas-electro fermentation) at different applied potentials (-0.4 and -0.6 V vs Ag/AgCl [3.5 M KCl]). With -0.6 V applied potential, a higher acetic acid production of 1.02 g/L was obtained. The CO₂ fermentation efficiencies were improved by higher charge transfer rates (52% faradaic efficiency) and lower corrosion and resistances (Rct -0.4 ohm). On continuous conditions, an abundance of Proteobacteria, Firmicutes, and Actinobacteria in the enriched biocatalyst represented a significant shift of the microbiome towards CO₂ feeding microbes.



Fig. 11. Schematic diagram of acetic acid production from CO₂ via lignin/ionic based catalytic system. Reproduced with the permission from [102]. Copyright 2023 Elsevier

Wang and co-workers [103] reported a new strategy to produce renewably sourced acetic acid from lignin and CO_2/H_2 . The catalytic system is based on an ionic liquid (that is 1- butyl-3methylimidazolium chloride, [BMIm][Cl]) that contains a Ru-Rh bimetal catalyst (Ru₃(CO)₁₂ and RhI₃) and Li salt additives (LiI and LiBF₄). This system was effective for the production of acetic acid in the temperature range of 120-180 °C under low pressure of CO_2 (1 MPa), affording an acetic acid yield of 94% at 180 °C. In addition, taking lignin as the feedstock, 0.17 g of acetic acid was produced from 1 g of lignin. Finally, the catalytic system could be recycled 5 times without decrease in performance. Investigation of the reaction mechanism evidenced that lignin provided methyl group to form CH₃I, while the [BMIm^{]+} -stabilized RhI₃ and I ⁻-complexed Ru(CO)_{n(n=1~4)} cooperated and catalyzed the formation of acetic acid from CH₃I and CO₂/H₂.

Table 4 below summarizes the biomimetic materials explored for the production of acetic acid. While there is appreciable development towards the production of acetic acid from CO_2 using

biomimetic materials, the yields reported are very low and cannot satisfy the commercial purpose. This calls for more research efforts in this hot topic.

Photocatalystic system	Reaction conditions	Yield	Reference
М.	Low-intensity	6.01 mmolg ⁻¹ after six	[12]
thermoacetica/AuNCs	simulated sunlight	days in the presence of	
hybrid system	(calibrated by	cysteine as sacrificial	
	photodiode, air mass	agent	
	1.5 global spectrum		
	$(AM 1.5), 2Wm^{-2})$		
М.	Air mass 1.5 global	85±12% quantum yield	[104]
thermoacetica/CdSNPs	spectrum, 2 W m^{-2} ,	after four days	
hybrid	photons; 5 x 10^{13} cm ⁻²		
	s ⁻¹ under light and		
	dark flux at $\lambda = 435$ to		
	485 nm		
PFP/PDI/M.	5.0 Mw/cm ² power	0.1 mM in dark, 0.6	[101]
thermoacetica p-n	density, under of light-	mM under light	
heterojunction	dark cycle for 12h		
Different bacteria	Heat-shock, Acid-	1.02 g/L at 52% FE	[102]
mediated	shock and 2-		
biocomposites	Bromoethano		
	sulphonate (BESA),		
	31 mg L ⁻¹ h ⁻¹ of		
	ICFR, (-0.4 and -0.6 V		
	vs Ag/AgCl [3.5 M		
	KCl]), $R_{ct} = 0.4$ ohms		
Lignin/ionic based	180 °C, 1 MPa	94% yield, 0.17g acetic	[103]
catalytic system		acid produced from 1g	
		of lignin	

Table 4. Biomimetic photocatalysts for acetic acid production from CO₂

PFP: poly(fluorene-co-phenylene), PDI: perylene diimide derivative, FE: Faradaic efficiency, ICFR: Inorganic carbon fixation rate

6. Mechanistic insights of biomimetic photocatalysts for CO₂ Reduction

There is no definite mechanism for the reduction of CO_2 as it is a complex reaction involving radicals, electrons, and protons [105]. Generally, after the reduction of CO_2 by a one-electron transfer and the formation of surface-bound CO_2^- , the process proceeds through a sequence of elementary steps involving the transfer of a proton, electron or radical (H·), as well as breaking C-O bonds and creating C-H bonds. Several intermediates are radical species, partially accounting for the number of possible pathways and final products at different stages of recombination. The exact sequence and mechanistic flow of each subsequent step have not been thoroughly elucidated, but some literature suggests that photocatalytic reduction of CO_2 *via* biocatalysis has a similar pathway than that described for transition metal photocatalysis [105].

Hydrogen radicals (H[·]) bind to the oxygen atom of the adsorbed carboxyl radicals (CO₂ ⁻), resulting in an immediate cleavage of the C-O bond, subsequently leading to the formation of carbon monoxide. In a separate mechanism, the carboxyl radicals recombine with hydrogen radical to form formic acid. In a next step, formic acid accepts another hydrogen radical to form a dihydroxymethyl radical, which is dehydrated when another hydrogen radical is attached to the formaldehyde. Two additional steps of reduction lead to CH₃OH formation, which is further reduced to methane in two more steps of reduction. A proposed mechanistic pathway (by formaldehyde, carbene, glycol pathways) for the reduction of CO₂ to CO, formic acid, and methanol as described in the literature is presented in **Fig. 12** [106].

Formaldehyde pathway



Carbene pathway

$$\bigcup_{O} \bigoplus_{O} \bigoplus_{$$

Figure 12. Two reduction routes for CO₂; top, formaldehyde pathway and bottom carbene pathway [106].

The mechanism of CO_2 conversion involves two-electron reduction mediated by metal-based catalysts or enzymes. Most photocatalysts are metal-based [107][108][109][110][111]. For a commonly described metal photocatalyst, the semiconductor part absorbs photon energy, which leads to the excitation of electrons from valence band (VB) to conduction band (CB), creating electron-hole pair. The holes in VB have strong oxidizing ability, especially for water oxidation and electrons in CB reduce CO_2 . Water oxidation produces hydrogen ions and hydroxyl radicals, and the activated CO₂ reacts with excited electrons and hydrogen ions, which finally result in the formation of formic acid, formaldehyde, and methanol [112]. When $AgCuInS_2$ supported on graphene-TiO₂ are parallelly activated, electrons transfer through the graphene to reach the VB of TiO₂ to form the donor level, thus, resulting in the formation of photogenerated electrons (e^{-}) and holes (h⁺) at the active sites of AgCuInS₂ photocatalyst. The holes (h⁺) react with the adsorbed $2H_2O$ to produce oxygen (O₂), electron (e⁻) and protons (H⁺). Surface-adsorbed CO₂^{•-} radical generated by the injection of a single electron reacts with e⁻ and H⁺, producing CO, which undergoes a further reduction process forming the surface-adsorbed C. CO₂ further reacts with excited electrons (2e⁻) and hydrogen ions (2H⁺) to yield HCOOH. The further reaction of HCOOH with excited electrons $(2e^{-})$ and hydrogen ions $(2H^{+})$ led to a loss of water (H₂O) to produce an intermediate, formaldehyde, (HCOH) (typical of formaldehyde pathway, see Fig 11). Surface adsorbed formaldehyde reacts with 2e⁻ and 2H⁺ to yield methanol as a main product. A schematic representation of photocatalytic CO₂ reduction process with AgCuInS₂ is illustrated in Fig. 13.



Figure 13. Schematic diagram of the photocatalytic CO₂ reduction process. Re-used with permission from. Copyright (2021) American Chemical Society. [112]

Getahun et al. [55] concerned the use of metal oxides (ZnO and CuO) embedded with bio-based materials (such as chitosan) as photocatalysts. Both ZnO and CuO absorb UV light, making energy separation to occur, and resulting in hydrogen ion (H⁺) and electron (e⁻) generation. The schematic representation of photocatalytic CO₂ reduction using ZnO and CuO under UV light is illustrated in **Fig. 14**. The valence-band energy level of CuO (n-type) is higher than that of ZnO (p-type), the holes of ZnO generated by absorbing light energy transfers to the valence-band level of CuO, leading to the holes (h⁺) on CuO/ZnO composite oxidizing water (H₂O) to oxygen (O₂), hydrogen ion (H⁺) and an electron (e⁻). CO₂ (trapped within amine groups of chitosan and dendrimer, -NH₂---CO₂) reacts with e⁻ to form anion radical CO₂⁺⁻ (a single electron reduction of CO₂) followed by a reaction with protons H⁺, a process known as "proton-assisted multielectron reduction," Surface-adsorbed CO₂⁺⁻ radical reacted with 6e⁻ and 6H⁺ (CO₂ + 6H⁺ + 6e⁻ \rightarrow CH₃OH + H₂O), producing methanol. Therefore, the photoredox pathway could be considered a

proton-assisted multielectron reduction via the formaldehyde route. Pathway for CO and formic acid production are similar to those reported by Otgonbayar et al [110].



Figure 14. Illustration of CO₂ photocatalytic reduction to methanol with H₂O oxidation on films of CuO/ZnO catalytic-embedded carbohydrate polymer. Reproduced with permission from [55]. Copy rights (2022) Elsevier.

The photocatalytic reaction mechanism of metal-based systems are similar to those followed by enzymes such as carbon monoxide dehydrogenases (CODHs) [113] and formate dehydrogenases (FDHs) [114][115], in which two to four electron reduction of CO₂ is required for conversion to carbon monoxide, formic acid/formate, and methanol. The bio-inspired mechanistic insight for two-electron reduction of CO₂ to CO, as described by Mondal et al, 2014 [105] goes through proton-coupled electron transfer (PCET, $H^+ + e^-$), from a molecule that acts as "electron and proton" donor (reductant) to another one that acts as an acceptor (oxidant). Both biocatalysis and homogeneous transition-metal catalysis usually proceed *via* redox reaction where hydride transfer takes place for formate ion production (**Fig. 15**).



Figure 15. Overview of the bio-inspired mechanistic insight into the two-electron reduction of CO_2 to CO and FA (PCET = proton-coupled electron transfer). Copyright [105].

FDHs are known to catalyze the reduction of CO₂ to formic acid through a proton (H⁺) transfer mechanism (CO₂ + NADH + H⁺ \leftrightarrow HCOOH + NAD⁺) as well as reversible oxidation of formate to CO₂ (HCOO⁻ \leftrightarrow CO₂ + H⁺ + 2e⁻) [114][116][117]. β-Nicotinamide adenine dinucleotide hydrate reduced salt (NADH) is an essential co-factor for FDH, its efficiency is highly dependent on the NADH concentration as a hydrogen donor. NAD⁺ is the oxidized form of β-Nicotinamide adenine dinucleotide hydrate (NAD). The active site of the NAD⁺-based FDH showing the C4atom position on pyridine ring (**Fig. 16a**). Mondal et al., [105] reported the direct hydride transfer, occurs from the C-atom of the formate to the C4-atom of pyridine ring of NAD⁺, leading to NADH and CO₂ (**Fig. 16b**). The neighbouring residues on FDH such as Thr (Thr282), Asp (Asp308), Ser (Ser334) and Gly (Gly335) interact with the positively charged NAD⁺ ring and stabilize it, thereby promoting the hydride transfer from formate to C4 [118][119][120]. Generally, enzymatic reduction of CO_2 usually proceeds to HCO_2^- , and the oxidized form of nicotinamide cofactor (NAD⁺) is reduced to NADH by glutamate dehydrogenase (GDH).



Figure 16. Two-electron reduction of CO_2 to HCOOH; (a) the active site of the NAD⁺-based FDH, (b) the catalytic mechanism of FDH Copyright [105], (c) a simplified reaction pathway of CO_2 to methanol (FDH, formaldehyde dehydrogenase (FDH), alcohol dehydrogenase (ADH)). The regeneration of NADH was catalyzed by glutamate dehydrogenase (GDH) [114].

A complex system of enzymes such as adenosine triphosphate (ATP), guanosine triphosphate (GTP), nicotinamide adenine dinucleotide (NAD or NADH for the reduced form), or nicotinamide adenine dinucleotide phosphate (NADP or NADPH for the reduced form), working in tandem is vital when considering biological conversion of CO₂. There is a need to further

research in this area especially around the regeneration of cofactor NADH, which in turn, improves enzyme efficiency during complex CO₂ bioprocessing applications.

In the formation of acetic acid, methanol carbonylation is required, and the reaction comes after the formation of Methanol (photocatalytic reduction of CO₂) [101] by using metals catalysts such as gold, platinum, palladium, osmium, copper-nickel, copper-Mordenite, palladium-cerium, ruthenium, rhodium, and lanthanum. Metal complexes play a huge role in achieving the carbonylation reaction through formation. In a typical Rh/CH₃I catalysed system, a series of activated anionic species such as cis-[Rh(CO)₂I₂]⁻, [(CH₃CO)-Rh(CO)I₃]⁻ and [(CH₃CO)-Rh(CO)₂I₃]⁻ to assist with product formation. Halogen additives is required to promote the reaction by activating C-O bond in methanol and on transition metal complexes to form methyl iodide (**Eq. 1**) which reacts with carbon monoxide to form acetyl-iodide (**Eq. 2**) and can quickly reach acetic acid final product when hydrolyzed in water (**Eq. 3**). The metal catalyst is recycled at the end of the reaction.

 $CH_3OH + HI \rightarrow CH_3I + H_2O \tag{1}$

$$CH_3I + CO \rightarrow CH_3COOI$$
 (2)

 $CH_3COOH + H_2O \rightarrow CH_3COOH + HI$ (3)

Conclusion and perspectives

The potential of using bio-based photocatalysts for the conversion of CO_2 into value-added products, has been emphasized by describing the main advances in this domain from the literature in the last decade. The investigation of such catalytic systems is fundamental as this may afford the development of environmentally friendly, and sustainable processes as alternatives to replace the use of fossil fuels and consequently, contribute to the decrease global warming. The reported bio-inspired photocatalytic systems are presented following the target chemicals, namely carbon monoxide, formic acid, methanol and acetic acid (via carbonylation of methanol) all being key chemicals for a direct use in various applications or as intermediate synthons. Several case studies concern the use of metal catalysts combined with biomaterials which, under photocatalytic conditions, allow the formation of CO_2 -derived carbon molecules. The reduction of CO_2 using bioinspired catalysts is generally accompanied by proton-coupled electron transfer (PCET) resulting in C–O bond cleavage to yield CO, an initial carbon-based product. CO₂ conversion to formic acid is usually promoted by formate dehydrogenase (FDH) and its co-factor NADH, responsible for donating hydrogen [117][118]. Carbonic anhydrase one of the fastest enzymes that catalyses CO₂-hydration reaction has been combined with formate dehydrogenase (FDH) with co-factor NADH to form formate/formic acid [119][122][121][119]. One of the major drawbacks is the depletion of NADH during catalytic reduction reaction to NAD⁺, thus requiring continuous regeneration to NADH to have sustained formic acid production. The enzymes used for conversion of CO₂ to CH₃OH are Formate Dehydrogenase (FDH) that utilizes NADH as a cofactor, and alcohol dehydrogenase (ADH). Enzymes usually exhibit much reduced activities over time due to impurities, slight change in reaction condition, (such as pH and temperature). The presence of transition metal/metal oxides incorporated unto biopolymers and enzymes facilitate the reduction of CO₂ to various products by promoting metal-CO₂ interaction. An example is [NiFe] CO Dehydrogenase ([NiFe] CODH), an enzyme used for conversion of CO₂ to CO. Zinc complexed Carbonic Anhydrase (CA), Zn[CA] is used for the conversion of CO₂ to bicarbonate (HCO₃) [119].

Most of the enzymes being investigated in CO_2 reduction are naturally occurring and can denature easily under varying pH and temperature. Some key issues still remain that need to be considered when looking for further advancement in the biological catalysed CO_2 reduction. Firstly, the use of transition metals/metal ions incorporated unto enzymes protein ligands having high CO_2 binding affinity is vital for converting CO_2 molecules to desired products. Secondly, the depletion of NADH during catalytic CO_2 reduction reaction to NAD⁺, as well as the higher stability of CO_2 molecule, resulting in its lower reactivity. To avoid this problem, it is necessary to regenerate NADH during CO_2 reduction. A separate NADH regeneration step which involve the use of combined chemical, photocatalytic, biocatalytic, and/or electrochemical reactions within a single set-up must be considered when engineering efficient bioinspired catalyst for CO_2 reduction.

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Conflicts of interest

The authors declare no conflict of interest

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