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Biomimetic photocatalysts for the transformation of CO₂: Design, properties and mechanistic insights

Yakubu Adekunle Alli^{a,b*}, Nokuthula E. Magida^c, Funeka Matebese^c, Nuria Romero^a, Adeniyi Sunday Ogunlaja^c, Karine Philippot^a

^a CNRS, LCC (Laboratoire de Chimie de Coordination), UPR8241, Université de Toulouse, UPS, INPT, Toulouse cedex 4 F-31077, France.

^b Department of Chemical Sciences, Faculty of Science and Computing, Ahman Pategi University, Patigi-Kpada Road, Patigi, Kwara State, Nigeria

^c Department of Chemistry, Nelson Mandela University, Gqeberha (Port Elizabeth), South Africa

*Corresponding authors' e-mail address: yakubu.alli@lcc-toulouse.fr

Abstract

The increase in CO₂ concentration in the atmosphere due to the intensive burning of fossil fuels, for the needs in energy of our modern society, has strongly contributed to the global warming. One of the promising strategies to fight this menace is to capture atmospheric CO₂ and convert it into value added products, including fuels and chemicals. Among the methods developed for the transformation of CO₂, the sunlight-activated photocatalytic conversion appears as the renewable and easy to use method. For this purpose, it is required to have at disposal proper materials to act as catalysts and light absorbing materials to harvest the sun energy, and then to combine these components with an electron carrier, in an efficient way in order to access the desired fuels by a photocatalytic sustainable CO₂ conversion. Among the efforts in research dedicated to the production of solar fuels, studies focused on the design and development of bio-based photocatalytic systems as alternative to conventional photocatalytic systems have emerged in the literature in the last decade. The objective of this review is to show the readers the recent advances in the fabrication of bio-based photocatalysts for the transformation of CO₂ into value added products. This review specifically focuses on the conversion of CO₂ into carbon monoxide, methanol, formic acid/formate and acetic acid which are the main target products described. Methods adopted to enhance the selectivity and efficiency of the bio-derived photocatalysts developed are also discussed and a section is dedicated to computational

investigations. Finally, the current challenges and perspectives in the development and use of these novel and greener photocatalysts are presented.

Keywords: CO₂ transformation; solar fuels; biomimetic photocatalysts; enzymatic catalysis

Table of contents

1. Introduction
2. Biomimetic-photocatalysts for the transformation of CO₂ to carbon monoxide
3. Biomimetic photocatalysts for the transformation of CO₂ to methanol
4. Biomimetic photocatalysts for the transformation of CO₂ to formic acid/formate
5. Biomimetic photocatalysts for the transformation of CO₂ to acetic acid
6. Mechanistic insights of biomimetic photocatalysts for CO₂ Reduction
7. Conclusions and perspectives

1. Introduction

Up to recent period, the burning of fossil fuels has been a major source of energy of our modern society. The inordinate consumption of fossil resources has led to a severe global warming (with new records in the last three years) due to the excessive discharge of greenhouse gasses into the atmosphere, and most particularly of carbon dioxide (O=C=O), and this has become an international crisis that calls for an urgent solution. CO₂ is the primary driver of the global warming [1]. In addition, it is a long-lived greenhouse gas, with one-third still present in the atmosphere after a century of release [2]. Scientists believe that if purposeful action is not taken with immediate alacrity to lower CO₂ concentration in the atmosphere, global warming and impact of climate change could inevitably lead to the death of more than 250,000 people per year, put 100 million people into abject poverty by 2030 and displace 150 million people or more by 2050 [3]. The transformation of atmospheric CO₂ is a herculean task because of its high stability. However, it may have significant benefits in terms of access to carbon based platform molecules and fuels [4]. Among the various strategies available for the reduction of CO₂, photocatalytic transformation of CO₂ is unarguably a promising and green strategy that could solve the global challenge [5]. The reduction of CO₂ by photocatalysis is similar in mechanism to the natural photosynthesis, which can be explained using three separate but linked steps: (i) the

absorption of solar energy (ii) the separation of charges and (iii) surface redox reaction. Utilization of the ubiquitous solar energy in order to reduce CO₂ into renewable value-added products would in turn simultaneously improve the lingering global energy shortage, climate change and take a step towards the circular economy [6]. However, the major bottlenecks of existing photocatalysts (i.e. monocatalysts, liquid state Z-scheme, all solid states Z-scheme, direct Z-scheme and type II heterojunctions) are (1) fast recombination of photogenerated charge carriers, (2) poor redox power and (3) Inefficient charge separation thereby hindering the practical application of the technique [7–9]. According to general agreement, a good photocatalytic system must have quick charge transition, efficient charge separation, and good absorption in the visible spectrum, be composed of non-toxic elements and display high stability [10]. Among the photocatalysts reported in the literature, bio-based photocatalysts have gained popularity in the recent decade. This is mainly due to their high reaction efficiency and their fascinating catalytic mechanism inspired by nature [11]. In comparison to their photosynthetic counterparts, non-photosynthetic organisms, especially those created in contemporary synthetic biology, have more desirable metabolic pathways, providing a tunable CO₂ metabolism for a variety of products [12]. Other interests of the bio-based photocatalytic systems for the transformation of CO₂ into hydrocarbon fuels emerge from the abundance of the biomaterials used, environmental friendliness, and energy-saving if the process is activated by sunlight, to mention but a few. In the reviewing exercises that cover materials for application as catalysts in the transformation of CO₂, the recent advancements in the use of bio-based photocatalysts is only little highlighted. For example, Yau and co-workers [13] recently published an updated review which gives precedence to microbial hybrid photosystems for CO₂ fixation. Notwithstanding, the review work did not provide elaborate survey on other bio-based photocatalysts and it fails to address the theoretical investigation in order to rationalize the mechanism of CO₂ reduction.

In order to sensitize the scientific community to the advantages of using bio-derived photocatalysts (i.e. a natural light driven catalytic reaction using enzyme, bacterial and polymers) for the conversion of CO₂, this review intends to gather the recent research (i.e. covering the last decade) on the design of such photocatalytic systems for the transformation of CO₂ into fuels. The examples reported are classified into five sections in line with the target hydrocarbon fuel, namely carbon monoxide, methanol, formic acid/formate and acetic acid (see sections 2, 3, 4 and 5, respectively). Furthermore, in section 6, an in-depth analysis on the interest of computational

investigations for the development of bio-derived photocatalysts for CO₂ reduction is given. Computational data may either complement or to give prior understanding into experimental investigations. Finally, current challenges and future outlook are discussed.

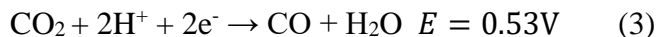
2. Biomimetic photocatalysts for the transformation of CO₂ to carbon monoxide

Carbon monoxide (CO) is one of the attractive value-added products that can be generated from the reduction of CO₂. It is commonly used as an intermediate in some chemical processes including the production of drugs, fragrances, formaldehyde or benzaldehyde and methanol [14]. Some chemical industries also use CO in the preparation of inorganic chemicals such as metal carbonyls like nickel tetracarbonyl (**Eq. 1**) and iron pentacarbonyl (**Eq. 2**).



Furthermore, CO plays an important role alongside with hydrogen gas, forming together the syngas that is used in the Fischer-Tropsch process towards superior hydrocarbons [15].

From thermodynamic and kinetic points of view, the generation of CO from CO₂ is one of the easiest compared to that of other hydrocarbon fuels. The reduction of CO₂ into CO and H₂O requires two electrons and two protons with a reduction potential of 0.53V (vs. NHE at pH = 7) (**Eq. 3**).



Different bio-based photocatalytic materials have been employed for the conversion of CO₂ to CO. In one of the earliest studies, Woolerton et al. developed a photocatalytic system that comprises metal-oxide nanoparticles functionalized by the enzyme carbon monoxide dehydrogenase (CODH) as catalyst and sensitized to visible light by a ruthenium bipyridyl photosensitizer, both components being supported on an anatase/rutile TiO₂ mixture as the semiconductor (electron donor). Using this photocatalyst activated by visible light they could effectively reduce CO₂ to CO [16]. The authors examined the CO production rates and stability as a function of each component (photosensitizer, enzyme and TiO₂). Tolerance to O₂ and effects of different electron donors were also investigated, together with strategies to control enzyme binding at the surface of TiO₂ in order to enhance overall activity (**Fig. 1a**). Ding and co-authors [17] reported a functional porous coordination hybrid system composed of nickel porphyrin and

nickel bis(terpyridine) dual active sites as efficient bio-based photocatalyst for CO₂ reduction into CO under the illumination of visible light ($\lambda \geq 420$ nm) (**Fig. 1b**). The biomaterials exhibited an extraordinary photocatalytic activity towards the CO formation with an outstanding generation rate of 3900 $\mu\text{molh}^{-1}\text{g}^{-1}$ and selectivity of 98%. Also, they investigated the mechanism of action associated with dual active sites of the system, composed of metal porphyrin and metal bis(terpyridine), using theoretical simulations which further supported an excellent photocatalytic performance of these bio-hybrids as observed in the experimental investigation (**Fig. 1c**). The theoretical investigation demonstrated that the binding between CO₂ and Ni(II) in nickel bis(terpyridine) is much easier than that with nickel porphyrin. The Ni–C Mayer bond order of *CO₂ intermediate in nickel bis(terpyridine) was 0.533, much higher than that (0.009) in nickel porphyrin, which accounts for the stability of the intermediate of the former.

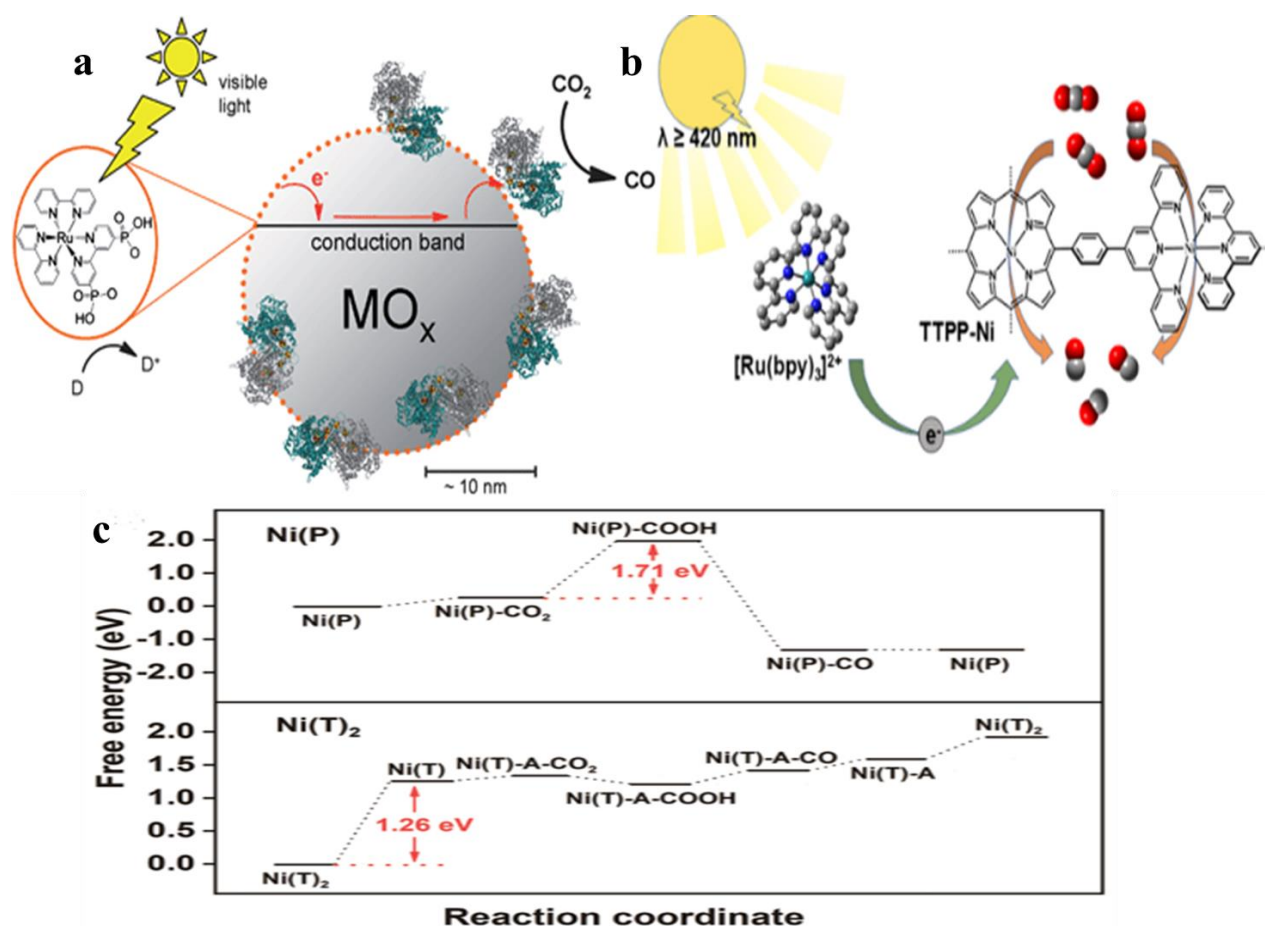


Figure 1. Mechanism of CO₂ reduction to CO by (a) CODH/ruthenium bipyridyl/anatase or rutile-TiO₂. Reproduced with the permission from [16]. Copyright 2011 Royal Society of Chemistry. (b) Ni Porphyrin and Ni bis(terpyridine) coordinated polymer (TPP-Ni) (c) Schematic mechanism of CO₂ reduction catalyzed by Ni Porphyrin and Ni bis(terpyridine) . Adapted with the permission from [17]. Copyright 2022 American Chemical Society.

In a related study, Liang and co-workers [11] developed a biomimetic system based on metalloporphyrin and ZrO₂ nanoframes that showed an evolution CO yield of 35.3 μmol within 3 h of reaction and high selectivity rate of 93.1% as well as an apparent quantum efficiency of 1.84% which is 60.8 greater than pristine porphyrin-Ni with evolution rate of 0.58 μmol.

The group of Zhao [18] constructed a covalently bonded reaction centre bio-hybrids comprised of Co-based porphyrin and a low molecular mass graphitic C₃N₄ for the transformation of CO₂ to CO under the irradiation of visible light. They obtained a photocatalytic activity 13 times greater than that of pure C₃N₄ (CO generation rate of 17 μmolg⁻¹h⁻¹), thus evidencing the positive role of the Co porphyrin. The improved activity was attributed to the efficient electron transfer and trapping by CO active sites, improved electron-hole separation and strong affinity of Co-porphyrin for CO₂ adsorption.

Lin and colleagues [19] designed a highly efficacious g-C₃N₄/tetra(4-carboxyphenyl)porphyrin (III) chloride heterogeneous catalyst for selective photoreduction of CO₂ into CO under visible light (**Fig. 2a**). In this catalytic system they used abundant, environmentally friendly FeTCPP as the catalytic center and inexpensive g-C₃N₄ nanosheets as the light absorber, maintaining a simple mechanical mix under constant stirring. A maximum rate of 6.52 mmol g⁻¹ of CO in 6 h, with up to 98% selectivity was obtained. The catalytic performance was attributed to an effective electron transfer from g-C₃N₄ nanosheets to FeTCPP

Apart from the enzymatic/bacterial based photosystems reviewed so far, Zhou and co-authors [20] reported biomimetic polymeric semiconductor based hybrid nanosystems that combined a typical CO₂/proton reduction catalyst, a cocatalyst, an electron mediator, and a CO₂ activator for artificial photosynthesis with water and CO₂. g-C₃N₄ nanosheets, which mimic the nanolayered thylakoids stacks, are shown to be promising photocatalytic elements with a planar configuration and a high surface area, and they serve as an excellent platform for the assembly of other

analogous elements (**Fig. 2b**). Au NPs are used as a suitable cocatalyst. ZIF-9 is used as a CO₂ concentrator and an electron mediator to promote the redox reaction. The highest production of CO and H₂ was achieved by ZIF-9(1%)Au-CNS at the evolution rates of 0.3 and 7.5 μmolh⁻¹g⁻¹, respectively. The study revealed that the incorporation of polymeric based g-C₃N₄ and ZIF-9 significantly enhanced the CO evolution rates which is 122 folds higher compared to non-ZIF-9 modified systems.

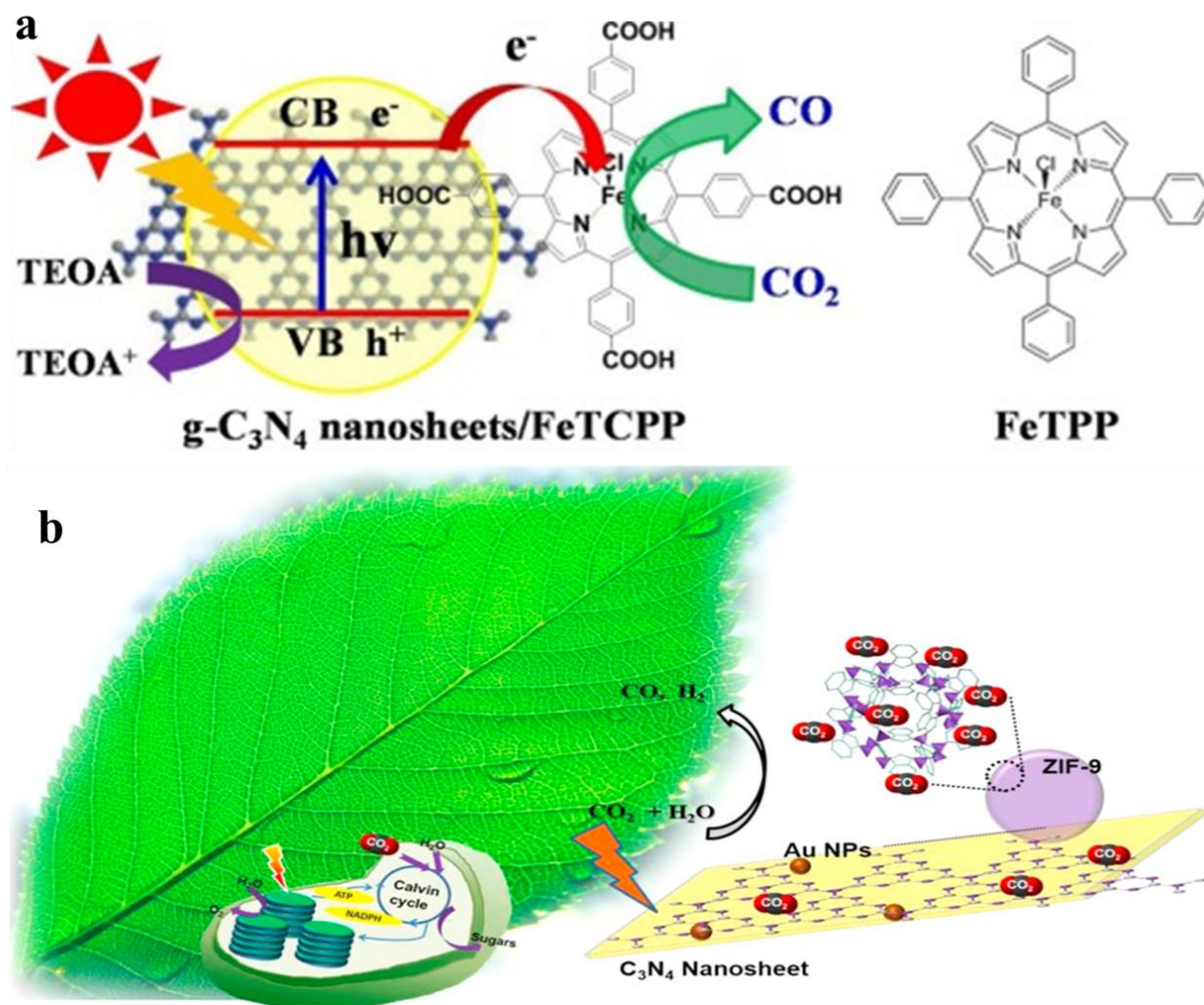


Figure 2. Plausible mechanism of CO₂ photoreduction by (a) g-C₃N₄/FeTCPP hybrids (b) Photoconversion activity of C₃N₄/AuNPs/ZIF-9. Reproduced with the permission from [19] and [20] . Copyright 2018, 2016 Elsevier, respectively.

Table 1 below outlines biomimetic photocatalysts that have been explored for CO production from CO₂ reduction. The synergistic incorporation of biomimetic photocatalysts has shown to be promising for the reduction of CO₂ to CO with high selectivity. However, the present bio based photocatalytic systems for CO production have not provided satisfactory activity and stability, key points that necessitate further research.

Table 1. Carbon monoxide production using bio-based photocatalysts

Photocatalytic system	Reaction conditions	CO yield/TON	Comments	Reference
TTPP-Ni/metal based bis(terpyridine) dual active sites	Photocatalytic activity under $\lambda \geq 420$ nm, trisopropanolamine employed as sacrificial agent, 15mg catslyst	3900 $\mu\text{molh}^{-1}\text{g}^{-1}$	The composites exhibited high CO yield. However, there is partial dissolution of photocatalysts	[17]
Zn-porphyrin sensitised Mn (I)	Aqueous medium, red light irradiation ($\lambda = 620$ nm)	Low yield	The slow conversion rate of CO ₂ to CO was attributed to inadequate e- migration from porphyrin to the catalyst	[21]
Carbon monoxide dehydrogenase functionalized CdS nanocrystals	Carried out under visible light irradiation with	Good yield	The size and morphology of the nanocrystals greatly	[22]

Nafion assisted Re(I)-RuL	The catalytic activity was done under visible light for 20h,	454 $\mu\text{molh}^{-1}\text{g}^{-1}$	enhanced the activity The presence of nafion improved the photocatalytic activity	[23]
Polymeric based $\text{C}_3\text{N}_4/\text{AuNPs}/\text{ZIF-9}$	g- 80 kPa of pure CO_2 gas was purged, 300W Xe arc lamp as light source	0.4 $\mu\text{molh}^{-1}\text{g}^{-1}$	The dominant evolved gas is H_2 which is not favourable for the reaction	[20]
g- $\text{C}_3\text{N}_4/\text{FeTCPP}$	Under visible light greater than 420 nm. TEOA was used as sacrificial agent	6.52 mmolg^{-1}	The findings indicates that the electrons migrated from g- C_3N_4 to FeTCPP	[19]
ZrO_2 nanoframes/metalloporphyrins	Visible light	35.3 μmol	The bio based material efficiently transfer electrons to the Ni catalytic sites which in turn increased the CO generation	[11]

TON: Turn over frequency, FeTCPP: tetra(4-carboxyphenyl)porphyrin (III) chloride

3. Biomimetic photocatalysts for the transformation of CO₂ to methanol

Methanol or methyl alcohol (CH₃OH) is an organic compound primarily used as a solvent, co-solvent, or feedstock in the chemical industry. Methanol has an octane number of 113 and a density that is half that of the gasoline, as thus, can be mixed with gasoline with no technical modifications and be used in methanol-fuelled vehicles [24]. Although methanol is considered an environmentally friendly fuel (no oxides of sulphur or nitrogen), the production of CO during its combustion limits its use as a pure fuel (100%). Hence it is mixed with gasoline on a ratio of 85% (methanol) - 15% (unleaded gasoline) [25]. More usages of methanol include hydrogen storage and transportation as an energy carrier as well as an easily transportable fuel, and also production of intermediates and synthetic hydrocarbons as a C1 building block [26]. Methanol is therefore regarded as a transition molecule from fossil fuels to renewable energies.

Methanol can be synthesized from carbon-containing materials such as natural gas, coal, biomass, and carbon dioxide (CO₂). Natural gas is currently the primary source of methanol contributing approximately 90% to the world 111 million metric ton production [27]. Unfortunately, natural gas and coal are fossil fuel resources, thus depleting non-renewable resources and significantly contributing to greenhouse gas emissions. The burning of fossil fuels (natural gas, coal, and oil) leads to CO₂ emissions which contribute significantly to global warming. Therefore, green production of methanol has gained a great attention of academic and industrial researchers.

Catalytic hydrogenation of CO₂ has been proposed as a green methanol production process. This process falls under the carbon capture and utilization (CCU) strategy which uses catalysts in the reaction of CO₂ and hydrogen (**Eq. 4**). The transformation of CO₂ to methanol is considered a green process when using captured CO₂ and/or hydrogen from renewable energy resources. The study by González-Garay et al. [28] shows how much CO₂ emissions can be avoided when using the green methanol synthesis process. However, the green methanol production *via* CO₂ hydrogenation is not economically feasible compared to the fossil-based process due to high costs of renewable hydrogen [28]. Therefore, there is an urgent need for developing cost effective CO₂ conversion technologies to produce methanol in a green way. Amongst many CO₂ conversion technologies such as biological, thermochemical, and electrochemical routes, the photochemical process has gained extensive attention [29]. If using solar light as its source of

energy, the photocatalytic reduction of CO₂ into valuable chemicals like methanol may be a sustainable approach [30–34].



Enzymatic conversion of CO₂ requires energy source like for the photochemical processes. The energy sources can be diverse, including ultraviolet, electrical potential, or energy carrying molecules (adenosine triphosphate (ATP), guanosine triphosphate (GTP), nicotinamide adenine dinucleotide (NADH)). While in a photochemical process, an electron donor, a photosensitizer, and catalysts (e.g. TiO₂, ZnO, graphene, etc.) are mainly required to drive selective production to methanol [35,36] [37–40]. These photocatalysts may be incorporated into biomaterials such as biochars and biopolymers with conductive properties (e.g. cellulose, chitosan, etc.) to improve their structural and electronic properties [32,41–43]. There have been few case studies on biomimetic photocatalysts for the reduction of CO₂ to methanol since 2015 due to cost of enzyme production, regeneration, nonetheless, the research area is slightly increasing (as seen in **Table 2**).

Recently, Tseng and co-workers [44] utilised sugarcane bagasse (SB) to support graphitic carbon nitride (gCN) for photocatalytic reduction of CO₂ to CO and methanol using a 14-watt LED light bulb. The presence of lignin in SB resulted in the semiconductor-like absorption edge and provided the interaction with gCN. The use of SB alone for photocatalytic study resulted in the formation of small amounts of MeOH. This is linked to the electronic transition of conjugated species within lignin (from SB) thus providing photoinduced charge carriers on SB. The introduction of SB (containing lignin) influenced the semiconductor-like absorption edge properties of the photocatalyst. Catalyst synthesis, g-SB-HT, using water (g-SB-HT-W) and ethanol (g-SB-HT-E) influenced MeOH yields, as the later solvent may be seen to have acted as hole scavenger thus increasing the yield of methanol (see Table 2).

Tseng and co-workers [45] also developed titanium oxide-chitosan film hybrid material (TiO₂-chitosan) for the photocatalytic CO₂ reduction to methanol. This TiO₂-chitosan material was able to selectively convert CO₂ in methanol at a production rate of 0.37 nmol/g (**Table 2**), showing a good stability. Similarly, Kavil and co-workers [46] synthesized a TiO₂-based hybrid photocatalyst (Cu-C(glucose)/TiO₂) by co-doping TiO₂ with copper (from copper sulfate) and carbon (from glucose) to photocatalytically reduce CO₂ to methanol. The photocatalytic experiment was performed in a Pyrex glass photoreactor filled with the catalyst and a NaOH

suspension and then purged with certified super-critical fluid grade carbon dioxide for 30 min. The photoreactor was irradiated with UV lamps (365 nm wavelength) for 5 h. In these conditions the Cu-C/TiO₂ catalyst allowed to achieve the highest methanol production (~2600 μM) with compared to counterparts C/TiO₂ (TiO₂ doped with only carbon) and TiO₂ P25, thus evidencing the synergy effect between the three components, Cu, C and TiO₂.

Studies have shown that enzymes catalysed photoreduction of CO₂ to methanol involves using a three-step enzyme cascade including formate dehydrogenase (FDH), formaldehyde dehydrogenase (AldDH or FDH), and alcohol dehydrogenase (ADH) [46]. Co-enzymes and co-factors are usually sacrificial for photoredox reactions and are depleted in the process, thus, requiring enzymes that may regenerate the reduced forms. To further improve methanol yield, sequential and co-immobilization of dehydrogenases with polymeric membranes, metal complexes and oxides have been proposed [47].

Biocatalysts such as enzymes, namely formate (FDH), aldehyde (AldDH), formaldehyde (FalDH), lactate (LDH) and alcohol (ADH) dehydrogenases was tested for CO₂ reduction to methanol [47,48]. Co-enzymes (also known as co-factors) such as nicotinamide adenine dinucleotide (NAD⁺) or its reduced form (NADH) and methylviologen (MV²⁺) are often used with FDH, NADH, AldDH and ADH. [48][49–51]. Amao and co-workers [36] developed a photochemical enzyme-based conversion system for the CO₂ photoreduction to methanol, using FDH, AldDH, and ADH combined with MV²⁺ under the visible-light photosensitization of water-soluble porphyrin, tetraphenylporphyrin tetrasulfonate (H₂TPPS) in the presence of triethanolamine (TEOA) (**Fig. 3**). This complex photoredox system allowed to achieve a methanol production concentration of 6.8 μM. Study of the enzymatic kinetics evidenced that the rate-limiting step is the formic acid to formaldehyde reaction with AldDH and reduced form of MV²⁺ (Methyl viologen).

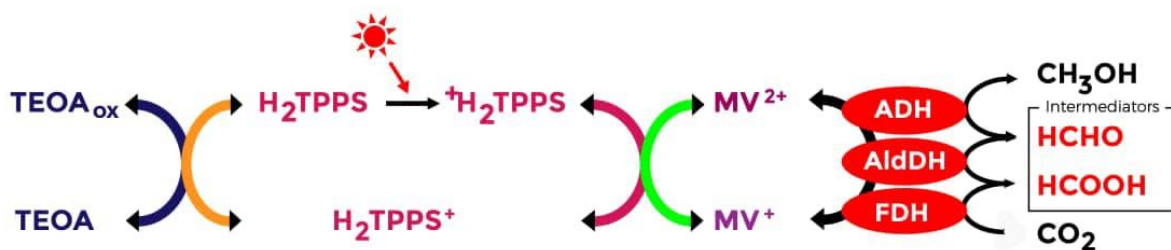


Figure 3. Methanol production from CO₂ catalysed with H₂TPPS/MV²⁺/TEOA/FDH, AldDH and ADH photoredox system. Reproduced with permission from reference [36]. Copyright 2018 Elsevier.

The use of co-enzyme nicotinamide adenine dinucleotide reduced form (NADH), FDH catalyses both the reduction of CO₂ to formic acid, ADH catalyses the reduction of formic acid to formaldehyde, and ADH catalyses both the reduction of formaldehyde to methanol (Fig 4). The resulting co-enzyme nicotinamide adenine dinucleotide (NAD⁺) catalyses the oxidation of formic acid to CO₂, formaldehyde to formic acid and oxidation of methanol to formaldehyde, respectively, thus inhibiting methanol production due to the redox nature of NAD⁺/NADH (Fig. 4). The oxidation processes were suppressed using oxidized form of MV²⁺, an electron carrier molecule, to drive the photochemical production of methanol. A reaction scheme for methanol production from CO₂ with FDH, AldDH and ADH using natural co-enzyme NAD⁺/NADH and artificial co-enzyme MV²⁺/MV⁺ is presented in **Figure 4**.

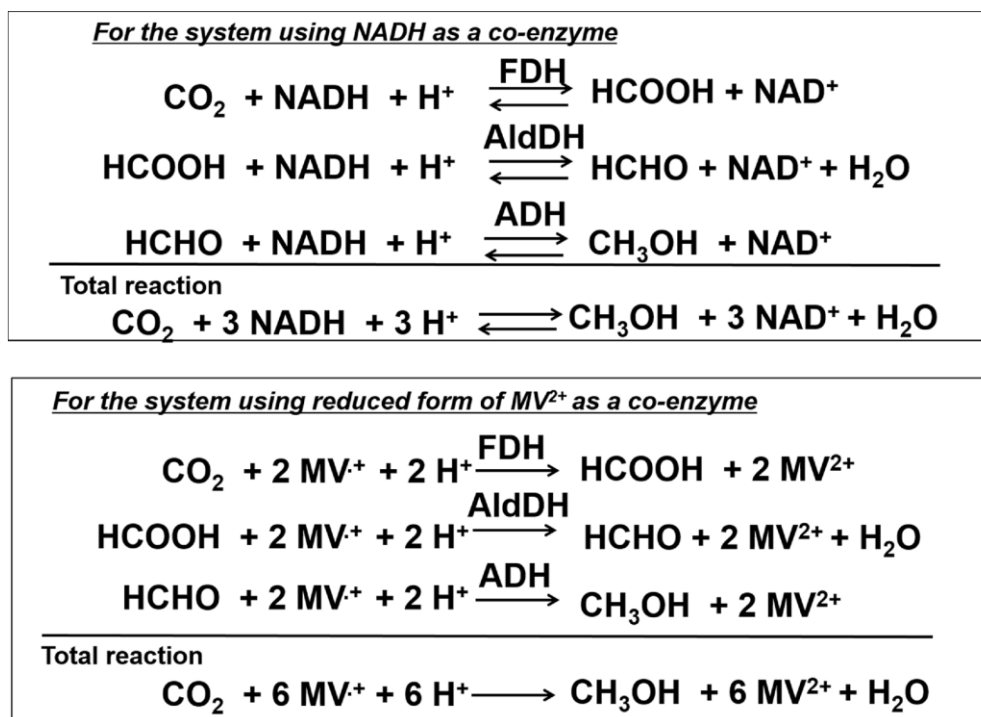


Figure 4. Scheme for the methanol production from CO₂ with FDH, AldDH and ADH using natural co-enzyme NAD⁺/NADH and artificial co-enzyme MV²⁺/MV⁺.

Zezi Do Valle Gomes and co-workers [48] reported the biocatalytic cascade conversion of CO₂ to methanol using three dehydrogenases (FDH, ADH and FalDH) co-immobilized in tailored siliceous mesostructured cellular foams (MCF) functionalized with mercaptopropyl groups (MP) with no cofactor. With this catalytic system they obtained a methanol yield of 0.35 mmol/g_{enzyme}/min, a value which might be improved by adding NADH regeneration.

Yadav et al. [52] prepared a CCG-IP type photocatalyst by attaching isatin-porphyrin (IP) chromophores with chemically converted graphene (CCG). A methanol production concentration of 11.21 μM was achieved using CCG-IP under visible light with an NADH photoregeneration of 39%.

Ji et al. [53] developed a polyelectrolyte-doped hollow nanofibers/microcapsules-based integrated photosynthesis system. Basically, the system was designed by assembling the components involved in the electron-transfer chain in a specific order, such that the photo-excited electrons can be transported efficiently in the opposite direction. This system was integrated in a device in the following order: three dehydrogenases and a cofactor (FDH, FalDH,

ADH and NAD(H)) in the innermost layer, electron mediators (M) in the middle layer, a photosensitizer (EY⁻ Eosin Y) in the outermost layer and an electron donor (TEOA) for the CO₂ conversion to methanol under solar energy (depicted in **Fig. 5**). They achieved a methanol production of 60.37 μM (hollow mixture of poly(allylamine hydrochloride) (PAH) and polyurethane (PU) nanofiber-based system) and 61.98 μM (microcapsules-based system) with 86.04% NADH photo-regeneration.

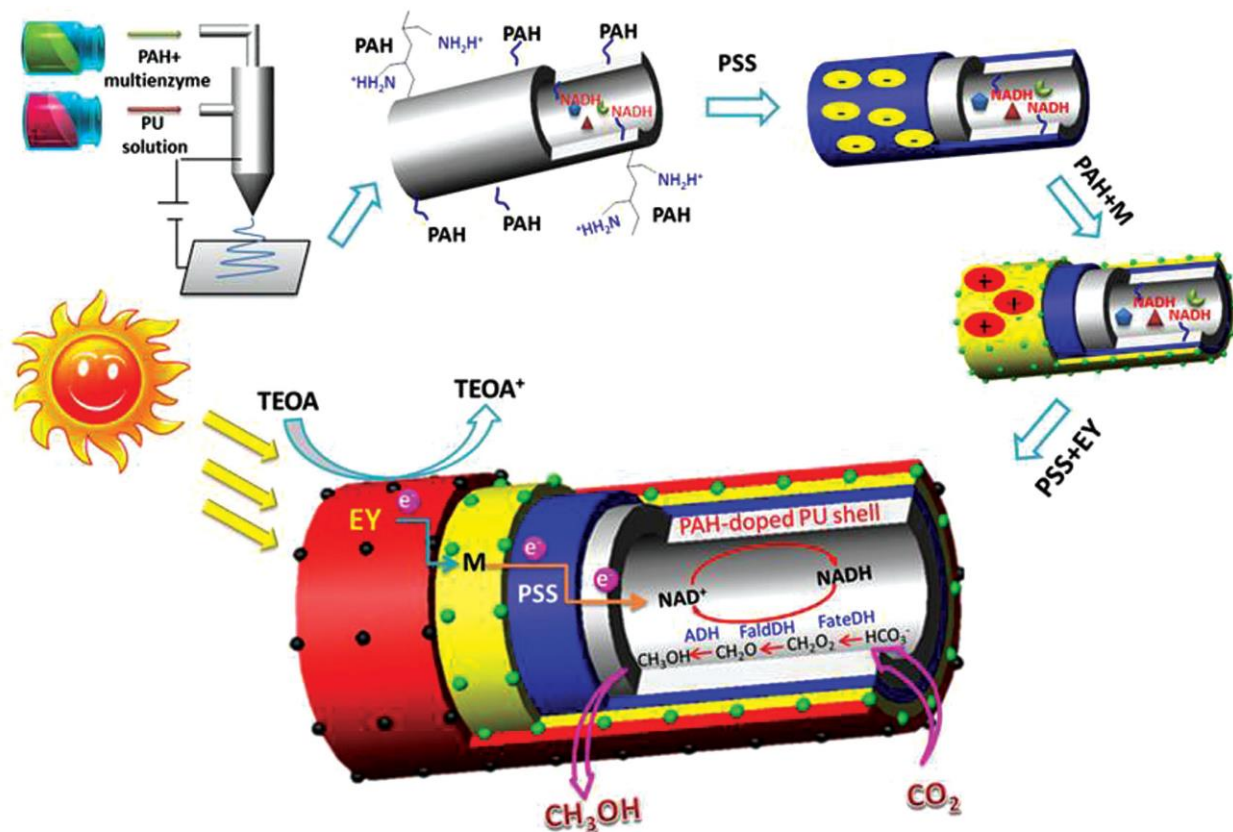


Figure 5. Schematic illustration of preparation of polyelectrolyte-doped hollow nanofibers/microcapsules-based integrated photosynthesis system for the solar-driven CO₂ transformation to methanol. Reproduced with permission from reference [53]. Copyright 2016 John Wiley & Sons.

Note that in different works, the co-immobilization of NADH with multi-enzymatic catalytic systems allowed to attend a higher methanol production efficiency over a single enzyme-based system. Furthermore, addition of photosensitisers, inorganic metal oxides and electron mediators as well as electron donors improved the photoreduction of CO₂ to methanol [53] [36].

Table 2. Methanol production from CO₂ using biomimetic photocatalysts

Photocatalytic system	Reaction conditions	CH ₃ OH yield	Reference
Sugarcane bagasse (SB) supported on graphitic carbon nitride (gCN) (SB-modified gCN)	CO ₂ was purged through water bubbler and photocatalyst and irradiated for 12 h with 14 W LED	110 μmol/g (g-SB-HT-E) 15-20 μmol/g (g-SB-HT-W)	[44]
Chitosan-TiO ₂	CO ₂ was purged through water bubbler and photocatalyst and irradiated for 12 h with UV light	0.37 nmol/g	[45]
Cuprous Oxide Nanoparticles Coated with Aminated Cellulose (Cu ₂ O/NCC-EDA)	CO ₂ was purged through NaOH (120 mL of 0.5 mol L ⁻¹) and photocatalyst (0.1 g) suspension and irradiated for 12 h with 300 W Xenon lamp	1656.7 μmol/g	[54]
CuO/ ZnO-embedded carbohydrate polymer films (CuO/ZnO-Den-TOCNF) (CuO/ZnO-CS)	Catalyst Films were irradiated for 6 h with UV light at 5cm distance from the closed reactor bottle where water was dropped and dry ice (440–880 mg) was held	0.030 mmol/g 0.037 mmol/g	[55]
Cu-loaded carbon modified titanium oxide (Cu-C/TiO ₂).	NaOH (0.2 M) and catalyst suspension	2600 μM	[46]

Carbon from glucose	was purged with CO ₂ for 30 min and irradiated for 5 h with UV lamps		
Tetraphenylporphyrin tetrasulfonate with methylviologen, triethanolamine and dehydrogenases (H ₂ TPPS/MV ²⁺ /TEOA/FDH/AldDH/ADH)	Suspension of H ₂ TPPS (100 μM), MV ²⁺ (2.0 mM), TEOA (0.3 M), FDH (2.0 μM), AldDH (2.0 μM) and ADH (2.0 μM) in CO ₂ saturated 50mM sodium pyrophosphate buffer was irradiated for 100 min with 250W halogen lamp	6.8 μM	[36]
Dehydrogenases (FDH/FaldH/ADH) encapsulated in hollow nanofibers and microcapsules	10 mL buffer solution (50 mM PBS, pH 7.0), 15% (w/v) TEOA, 25 mg hollow nanofibers or microcapsules, 0.1 mg mL ⁻¹ FateDH, 0.1 mg mL ⁻¹ FaldDH, 0.1 mg mL ⁻¹ ADH, 0.2 mM NADH, 0.25 mM M, and 50 μM EY irradiated for 10 h with 450 W Xenon lamp	60.37 μM 61.98 μM 86.04%	[53]
Chemically converted graphene	CO ₂ (0.5 mL/min), β-	11.21 μM	[52]

attached to isatin-porphyrin and coupled with dehydrogenases (CCG-IP/ FalDH/FDH/ADH) NAD⁺ (1.24 μmol), 38.99% NADH photoregeneration Rh (0.62 μmol), TEOA (1.24 mmol), photocatalyst (0.5 mg), 9 units of each enzyme (FDH, FalDH, and ADH) in 3.1 mL of sodium phosphate buffer (100 mM, pH 7.0) irradiated for 30 min with visible light

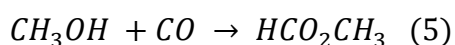
Siliceous mesostructured cellular foams (MCF) functionalized with mercaptopropyl groups (MP) and co-immobilized with dehydrogenases (MFC-MP/FDH/ADH/FalDDH) CO₂, MFC-MP, 1.35 mmol/g/min [48] FateDH (0.05 mg ml⁻¹), FalDDH (0.75 mg ml⁻¹), ADH (3.75 mg ml⁻¹), NADH (100 mM), and KHCO₃ (200 mM) in 100 mM phosphate buffer (pH 6.5)

4. Biomimetic photocatalysts for the transformation of CO₂ to formic acid/formate

Formic acid (FA) also known as methanoic acid is a colourless, sour, pungent-smelling, and fuming liquid, with as chemical structure HCOOH. If this weak acid is toxic in case of skin and oral exposure, it is non-toxic and stable at room temperature conditions [56,57]. A variety of wood ants and bees produce large quantities of FA naturally, to protect their nest by spraying at prey [58]. FA is one of the most important chemicals as it has found application in a wide range of industries. It has been used for example as an anti-bacterial reagent, food additive, textile dyeing, leather tanning, rubber, finishing, drilling fluids, anti-icing, fragrance ingredient,

cleaning products, pH modifier, and corrosion inhibitor. It is also used as organic synthesis reagent in various industrial chemical processes [59–61].

Due to the high industrial demand for FA, its production at high scale crucial. Formerly, FA was produced as a by-product of acetic acid production through hydrocarbon oxidation which has been reduced after the implementation of the carbonylation of methanol method for acetic acid production. FA production was then fulfilled via hydrolysis of formamide, this method however was economically unfeasible due to the consumption of sulfuric acid and ammonia as well as ammonium sulfate formation [56]. Finally, the carbonylation of methanol followed by hydrolysis of methyl formate (**Eq. 5 and 6**) has been the main industrial process applied for FA production [62][63].



However, this conventional production process has encountered predicaments, being the work-up of the hydrolysis mixture the main issue. Also, in the course of separating unreacted methyl formate, re-esterification of FA and methanol to methyl formate happens very quickly due to the unfavourable equilibrium [64,65]. Finally, this conventional method has negative environmental impacts, and high energy and economic costs [66,67]. As a result, alternative ways of producing FA from other sources such as the reductive conversion of CO₂ are extensively researched [68–70]. Transformation of CO₂ to FA can significantly reduce carbon emissions and assist in fighting climate change whilst finding applications in hydrogen storage, chemical feedstock, and fuel in direct formic acid fuel cells (DFAFCs) [71][72]. Several methods allow the reduction of CO₂ to FA among which photochemical [73][74], electrochemical (ECR) [75][68], hydrogenation [69][76], and biological processes [77][78], stand a good position. The attractiveness of the biomimetic photocatalytic transformation of CO₂ for FA production derives the abundance of biomaterials, environmental friendliness, and energy-saving to mention a few advantages. The mostly used biocatalyst is an enzyme, mainly the D-2-hydroxy acid dehydrogenase (FDH) that is found on methylotrophic microorganisms like bacteria and yeast that can use methanol. Tishkov and Popov reported that the FDH from bacterial sources exhibit 1.7 times higher activity than yeast [52]. The FDH enzyme catalyses the oxidation of formate to CO₂ and converts NAD⁺ to NADH [79][80]. The cofactor β-nicotinamide adenine dinucleotide in its reduced form (NADH)

is normally needed by FDH enzymes to attain the reduction of CO₂ to HCOOH according to reaction (Eq. 7) below [81]. Around twenty studies that deal with the reduction of CO₂ into FA utilizing bio-based photocatalytic systems have been reported between 2012 and 2022 (see Table 3). These studies increased sharply from 2018 and exhibited promising conversion systems and high FA production yields.



Yu et al. reported an effective and environmentally friendly light-driven enzymatic nanosystem [82]. This catalytic system consists of a combination between graphitic carbon nitride (g-C₃N₄) photocatalyst and two enzymes (carbon anhydrase (CA) and formate dehydrogenase (FateDH/FDH)), encapsulated in ZIF-8 MOF (CA&FateDH@ZIF-8/g-C₃N₄). **Fig. 6** illustrates the possible mechanism where photocatalytic NADH regeneration and enzymatic hydrolysis reduced CO₂ as described by the authors. Upon light irradiation, CA&FateDH@ZIF-8/g-C₃N₄ is excited to generate electron-hole pairs which then receive two electrons from the oxidation of one molecule of TEOA which are then excited from the valence band (VB) to the conduction band (CB) by light. The electron transport complex (M) was used to produce more 1,4-NADH, the biological entity needed by the enzymes. Thus, the highly selective M transports two electrons and one photon to NAD⁺ for NADH regeneration. With the two enzymes, CA quickly converted CO₂ into HCO₃⁻ which is converted into HCOOH using FateDH which, at the same time, consumes NADH as shown in Fig. 6. Reduction of CO₂ with CA&FateDH@ZIF-8/g-C₃N₄ system resulted in 198 μM of FA whilst the system without CA and free FateDH produced only 75 and 38 μM, respectively. When 2 mg.mL⁻¹ of CA&FateDH@ZIF-8/g-C₃N₄ was utilized, the optimized yield could reach 243 μM of FA. The activity of the enzymatic activity was retained above 80% after 10 runs, thus indicating the reusability of this catalytic system, that was also reported as able to shorten the distance between photocatalyst and enzyme entities, that is beneficial to the diffusion of substances and electrons.

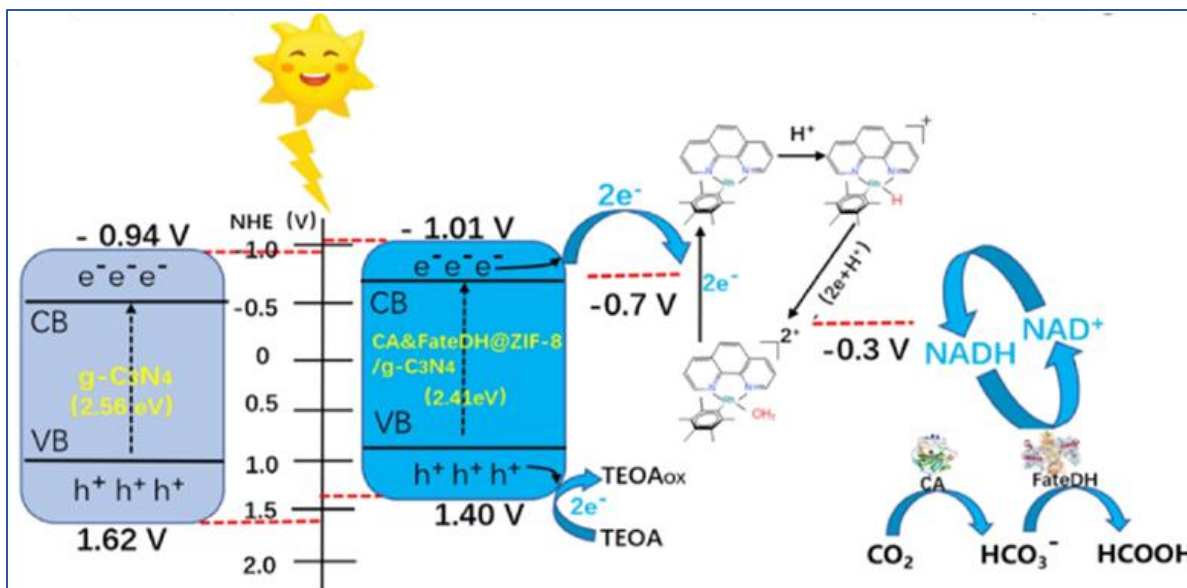


Figure 6. Illustration of electron and energy transfer processes for light-driven enzymatic CA&FateDH@ZIF-8/g-C₃N₄ nanosystem. Reproduced with permission from Reference [82]. Copyright 2021 Elsevier.

Wang et al. [83] reported a CO₂ reduction system consisting in two enzymes as well i.e. CA and FDH. Using a catalyst that combines two enzymes has been found to regulate the reaction rate and increase the loading amount of FDH which allows to decrease the ratio of CA/FDH. This is one of the routes enhancing FA production. Sustaining a stable pH environment for this system in the course of CO₂ hydration was mandatory as the process releases hydrogen ions. The increase in the hydration rate of CO₂ increased the yield of FA by aspect of 4.2-fold. However, the costs of cofactor and enzymes are economically unfeasible and enzyme immobilization and cofactor regeneration appeared as a probable solution for low costs, recyclability, and enhanced biocatalyst stability.

A FDH-bearing hollow fiber membrane (HFM) reactor (**Fig. 7**) has been developed by Gu et al. [70] for CO₂ reduction which led to high-efficient and sustainable FA production. They used a UV-TiO₂ photocatalytic coenzyme regeneration system with a FDH-bearing HFM reactor which exhibited a maximum initial reaction rate of 1.04 mM.h⁻¹ and 81.7% yield. The optimization of operating conditions and NADH concentration provided a turnover number of 125 after 4.5 h. Considering that one mole of NADH is consumed to generate one mole of FA, NADH regeneration for continuous enzymatic production is important. Thus, the system regenerated

NADH when the reaction mixture was transported into a photocatalytic cell. This resulted in an instant regeneration of NADH which eventually showed that even at a low loading amount of NADH high FA production could be achieved.

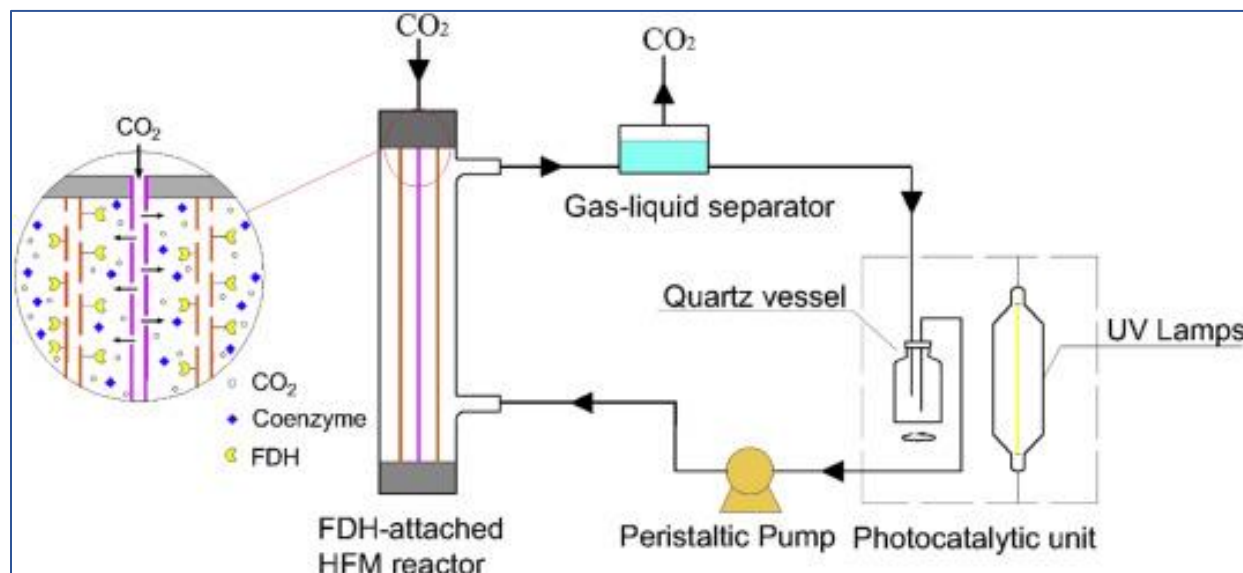


Figure 7. Illustration of a photocatalytic/enzymatic HFM system. Reproduced with permission from Reference [70]. Elsevier 2020.

Zhang et al. built an artificial thylakoid consisting of protamine-titania (PTi) micro-capsules with cadmium sulfide quantum dots (CdS QDs) and enzymes (FDH extracted from *Candida boidinii* (CbFDH)) for FA production from CO₂ [84]. The improvement of NADH regeneration activity ($4226 \pm 121 \mu\text{mol g}^{-1} \text{h}^{-1}$) and enzyme compatibility of CdS/PTi microcapsules contributed to high FA production. Compartmentalization of photogenerated holes and reactive oxygen species (ROS) by the structure of CdS/PTi microcapsules protected FDH from deactivation, resulting in high compatibility between bio- and photo-catalyst. This catalytic system exhibited a quantum yield of $0.66 \pm 0.13\%$ of FA over multiple light-dark cycles.

Zhang et al. reported a highly selective reduction system with a metal-free heterojunction of pyromellitic diimide/g-C₃N₄ (PDI/CN) coupled with FDH enzyme [85]. **Fig. 8** shows the possible mechanism of the photoenzymatic catalytic cascade system on the PDI/CN heterojunction. In this cyclic process, the photocatalyst could continuously reduce NAD⁺ to NADH while FDH could oxidize NADH to NAD⁺, concurrently, in order to accelerate CO₂ transformation to FA. The process can successfully advance continual FA production. Another

study reported the utilization of $[\text{Cp}^*\text{Rh}(\text{bpy})\text{H}_2\text{O}]^{2+}$ as a cocatalyst since the biologically inactive 1,6-NADH and the dimers of NAD^+ may be formed under the action of the intermediate radical. Thus, it was proven that $[\text{Cp}^*\text{Rh}(\text{bpy})\text{H}_2\text{O}]^{2+}$ was effective for the selective production of biologically active 1,4-NADH [70]. Zhang et al. [85] have also shown herein the promoting effect of $[\text{Cp}^*\text{Rh}(\text{bpy})\text{H}_2\text{O}]^{2+}$ in their system. The NADH yield increases linearly with the increase in sunlight irradiation time, and the system with $[\text{Cp}^*\text{Rh}(\text{bpy})\text{H}_2\text{O}]^{2+}$ showed better performance signifying that this cocatalyst assists NAD^+ to accept photogenerated electrons thereby accelerating electron transfer to generate NADH. The outstanding performance of this photocatalytic system was ascribed to good conductivity, heterointerface effect, broad sunlight response range of the catalyst as well as a distinctive photoenzymatic catalytic cascade system. The system achieved 75% NADH yield and 114.8 μmol for FA production.

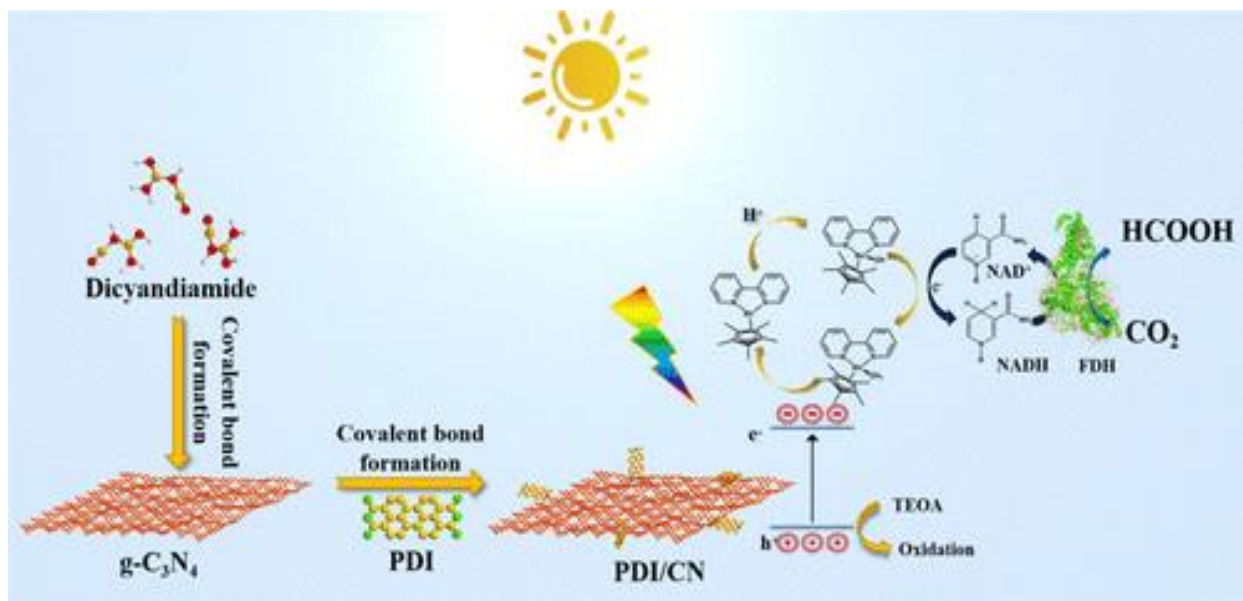


Figure 8. Possible mechanism of the photoenzymatic catalytic cascade system on the PDI/CN heterojunction. Reproduced with permission from Reference [85]. ACS Publications 2021.

Several studies concern the use of carbon-based materials such as carbon nanodots-silica hybrid, graphene film, graphene oxide modified with cobalt metallated aminoporphyrin, and C₆₀ polymer film as photocatalysts [86][33][87][34]. The interest in carbon materials emanates from their wide availability, environmental friendliness, and good light harvesting properties. Carbon nanodots were reported to have excellent biocompatibility, high solubility, and low toxicity whilst GO properties include biological function, thermal, mechanical, and biomedical [88].

Carbon nanodots-silica hybrid/FDH and graphene film/FDH systems showed NADH regeneration and FA yield of $74.10 \pm 0.17\%$, $203.33 \pm 1.9 \mu\text{mol}$ [86], and 71.8% , $228.6 \mu\text{mol}$ [33], respectively. For the FDH@GO-Co-ATPP system, NADH regeneration and FA yield of 48.53% and $96.49 \mu\text{mol}$, respectively were reported [87] whilst the C₆₀-based polymeric/FDH system exhibited 91.07% of NADH regeneration and $239.46 \mu\text{mol}$ [34].

Ji et al. reported chloroplast-mimicking TCPP/SiO₂/Rh HNP-based wholly integrated APS and AM/M/BP HNS-based integrated Z-scheme APS models in 2018 [89] and 2020 [90]. These systems decreased the transfer distance of the energy and electron among the enzymatic redox and light-harvest system as well as enabling sufficient charge separation and photogenerated excitons transportation. In both studies, the effect of the TCPP/SiO₂/Rh HNPs and AM/M/BP HNS-based Z-scheme APS was confirmed by detecting photochemical, optical, and electrochemical properties. The TCPP/SiO₂/Rh HNPs system showed NADH regeneration which enhanced from $11 - 75\%$ and FA production which increased from $10 - 100 \mu\text{mol}$. AM/M/BP HNS-based Z-scheme APS, on the other hand, exhibited significant enhancement of NADH regeneration and FA production which reached 89% and $365 \mu\text{mol}$, respectively [89][90].

The use of graphitic carbon nitride has been on the rise as well. This nitrogen-containing material possesses exceptional solar light-harvesting photocatalytic ability due to its slow recombination ability which increases photo-responsive energy transfer from the dye to a g-C₃N₄ moiety [91]. It has high stability, a tuneable band structure, strong visible light response, and astounding biocompatibility [92]. The systems reported by Cheng et al. [93], Zhang et al. [92], Singh et al. [91], and Kumar et al. [94] polymeric carbon nitride (PCN) mediated by a tannic acid/polyethyleneamine (TA/PEI), Rh complex NH₂-functionalized polymeric carbon nitride, 1-4,-diaminoanthraquinone monomer using a modified CN, and Se-doped CN nanosheets, respectively, showed FA production of 2.1 mM , 5.0 mM , $344.12 \mu\text{mol}$, and $200.26 \mu\text{mol}$.

Tian et al. [51] assembled a functionally compartmental inorganic photocatalyst-enzyme system that imitated chloroplast. Photoexcitation and enzymatic CO₂ reduction were compartmentalized by the thylakoid membrane with the purpose of protecting enzyme from photodamage whilst the system enables electron transfer thereby fostering photocatalyst. They built a tightly integrated artificial photosystem comprising of Rh complex cocatalyst onto thiophene-modified C₃N₄ (TPE-C₃N₄) and FDH encapsulated with MAF-7 which is a subclass of zeolitic imidazolate-

based MOF. The improved NADH regeneration of the system was attributed to the tightly integrated structure which promoted fast electron transfer from TPE-C₃N₄ to the Rh complex. MAF-7 not only protected FDH from photoinduced deactivation but also improved enzymatic stability in photosynthesis conditions. This photocatalytic system resulted in 16.75 mM FA after 9 h of illumination. **Table 3** shows the summary of the studies done on the transformation of CO₂ to FA using bio-inspired photocatalysts and the progress is promising.

Table 3. Biomimetic photocatalysts transformation of FA from CO₂.

Catalyst	Reaction conditions	Yield	Reference
Bromophenol-Bakelite complex/FHD	(BPB)/Rh NAD ⁺ – 1.24 μM, BPB – 0.031 μM, AsA – 1.24 μM, Rh complex – 0.62 μM, sodium phosphate buffer – 3.1 mL	NADH regeneration – 82.52% FA yield – 220 μmol	[95]
Bamboo-shape tetraphenyl framework (Mg-TPPBF)/FDH	magnesium porphyrin-based Photocatalyst – 15 μL in 3.1 mL of sodium phosphate buffer (100 mM, pH 7.0), β-NAD ⁺ – 124 μL, rhodium complex – 62 μL, FDH – 3 units, AsA – 310 μL	NADH regeneration – 73.9% FA yield – 197.2 μmol in 2 h	[50]
1,4-diaminoanthraquinone monomer using a modified carbon nitride (CN) (AQBCN)/FDH	(AQ) NAD ⁺ /NADP ⁺ – 1.24 μmol, CpM – 0.62 μmol, AsA – 0.1 mmol, FDH – 3 units and the AQBCN photocatalyst – 0.5 mg in 3.1 mL of SPB (100 mM; pH, ~7.0)	NADH regeneration – 97.21/95% FA – 344.12 μmol	[91]
Rh complex covalently grafted onto NH ₂ -functionalized polymeric	Photocatalyst – 12 mg, NAD – 120 μL, PBS –	NADH regeneration –	[92]

carbon nitride (NH ₂ -PCN) (Rh _{m3} -N-PCN)/FDH	2.88 mL, FDH – 1 unit	62.4%	
			FA yield – 5.0 mM
CA&FateDH@ZIF-8/g-C ₃ N ₄	CA&FateDH@ZIF-8/g-C ₃ N ₄ – 2 mg.mL ⁻¹ in a 0.1 M PBS, TEOA – 10 w/v%, mediator – 1 mg.mL ⁻¹ , β-NAD ⁺ – 0.8 m	FA yield – 243	[82]
Pyromellitic diimide/g-C ₃ N ₄ (PDI/CN)/FDH	FDH – 3 units, catalyst – 10 mg, TEOA – 15 wt.%, phosphate buffer solution (pH 7.4, 100 mM) – 10 mL, β-NAD ⁺ – 0.1 mM, [Cp*Rh(bpy)-(H ₂ O)] ²⁺ – 0.25 mM	NADH regeneration – 75%	[85]
Polymeric carbon nitride (PCN) mediated by a tannic acid/polyethyleneimine (TA/PEI) (PCN@TA/PEI-Rh core@shell)/FDH	Photocatalyst – 0.5 mgmL ⁻¹ , FDH – 2 mgmL ⁻¹ , NAD ⁺ – 1 mM, PBS buffer (pH 7.0, 100 mM), free Rh – 0.5 mgmL ⁻¹ , and CO ₂ – 0.3 MPa.	NADH regeneration – 37.8%	[93]
Covalent-organic framework/2,4,6-triformylresorcinol (COF-4)/FDH	Photocatalyst – 0.5 mg, β-NAD ⁺ – 1.24 mmol, AsA – 0.1 mmol, Rh complex – 0.62 mmol, and 3 units of FDH in 3.1 mL of 100 mM buffer (pH 7.0). CO ₂ flow rate – 0.5 mLmin ⁻¹ for 1 h in the absence of	NADH regeneration – 94.6%	[96]
			FA yield – 226.3 μmol

	light			
FDH@HFM/UV/TiO ₂	NADH – 2mM, liquid velocity – 40 mL.min ⁻¹ , gas velocity – 3 mL.min ⁻¹	FA yield – 81.7%		[70]
Antimonene/Cp*Rh(phen)Cl/black phosphorus hybrid nanosheet-based Z-scheme (AM/M/BP HNS)/FDH	TEOA – 15 w/v%, AM NSs or BP NSs – 0.2 mgmL ⁻¹ , M – 0.1 mM, FDH – 0.3 units, NAD ⁺ – 0.1 mM	NADH regeneration – 35±4% FA yield – 160±24 μmol		[90]
FDH@MAF-7/TPE-C ₃ N ₄ /PEI/Rh	NAD ⁺ g – 1 mM, phosphate buffer – 100 mM (pH 7), TEOA – 15 w/v%, MAF@FDH – 7.5 mg, TPE-C ₃ N ₄ /PEI/Rh – 3 mg	FA yield – 16.75 mM after 9 h 5 th cycle FA – ~76%		[51]
FDH@Rh-NU-1006	FDH@Rh-NU-1006 – 1.8×10 ⁻³ M in Tris buffer (2 mL), TEOA – 0.5 mM, NAD ⁺ – 1 mM	FA yield – 0.144 ± 0.003 in 24 h		[97]
C ₆₀ -based polymeric system coupled with FDH enzyme	CO ₂ flow rate – 0.5 mLmin ⁻¹ , β-NAD ⁺ – 1.24 μmol, TEOA – 1.24 mmol, 3 units of FDA in 3.1 mL of sodium phosphate buffer (pH 7.0, 100 mM)	NADH regeneration – 91.07% FA yield – 239.46 μmol		[34]
CdS/TPi microcapsule-CbFDH	[CO ₂] – 0.3 MPa, 37 °C, visible light illumination (λ = 405±5 nm)	NADH regeneration – 93.03±3.84%		[84]

	NAD ⁺ – 5 mM, TEOA – 400 nm, CdS/PTi – 1 gL ⁻¹ , PBS buffer – pH 7.0, 100 μMh ⁻¹ nm, CbFDH – 1 gL ⁻¹	FA yield – 1500	
FDH@GO-Co-ATPP	GO-Co-ATTP – 0.7 mg, NAD ⁺ – 1.24 mol, TEOA – 1.24 mmol, 3 units of FDH in 3 ml of (100 mM, pH 7) sodium phosphate buffer	NADH regeneration – 48.53% FA yield – 96.49 μmol	[87]
Porphyrin/SiO ₂ /Cp*Rh(bpy)Cl hybrid nanoparticles (TCPP/SiO ₂ /Rh-HNPs)/FDH	Phosphate buffer solution (100 mM, pH 7.0) – 10 mL, NAD ⁺ – 1 mM, TCPP – 0.5 mM, FDH – 0.3 units mL ⁻¹ , CO ₂ – 0.3 MPa	NADH regeneration – 75% FA yield – 100 μmol after 4 h.	[89]
Se-doped carbon nitride (C ₃ N ₄) nanosheets/FDH	Photocatalyst – 0.5 mg, Rh – 0.62 μmol, β-NAD ⁺ – 1.24 μmol, FDH – 3 units, AsA – 0.1 mmol in 3.1 mL of sodium phosphate buffer (100 mM, pH 7.0) CO ₂ flow rate – 0.5 mLmin ⁻¹	NADH regeneration – 82.84% FA yield – 200.26 μmol in 1 h	[94]
Graphene film photocatalyst (GFPC)/FDH	β-NAD ⁺ – 1.24 μmol, Rh complex – 0.62 μmol, FDH – 3 units in 3.1 mL of sodium phosphate buffer (100 mM, pH 7.0), AsA – 0.1 mmol, CO ₂	NADH regeneration – 71.8% FA yield – 228.6 μmol in 2 h	[33]

		flow rate – 0.5 mLmin ⁻¹		
Carbon nanodots-silica hybrid (CNDSH)/FDH		Photocatalyst – 0.5 mg, β-NAD ⁺ – 1.24 mmol, rhodium complex – 0.62 mmol, ascorbic acid – 0.1 mmol, FDH – 3 units in 3.1 mL of Na ₂ HPO ₄ -NaH ₂ PO ₄ buffer (100 mM, pH 7.0)	NADH regeneration – 74.10±0.17% FA yield – 203.33±1.9	[86]
Cobaloxime/eosin/REOA/FDH		Eosin Y – 0.10 mM, NAD ⁺ – 0.50 mM, TEOA – 0.20 M, 5 units of FDH in 0.15 M phosphate buffer (pH 7.0)	FA yield – 36%	[98]

5. Biomimetic photocatalysts for the transformation of CO₂ to acetic acid

Acetic acid otherwise known as vinegar acid, ethanoic acid and methane carboxylic acid is a clear, colourless liquid, of pungent smell and with the chemical formula CH₃COOH. It is the second simplest carboxylic acid after formic acid and is of high importance also due to a plethora of industrial and pharmaceutical applications. It is produced synthetically and it can also be prepared via bacterial fermentation. It is a crucial chemical reagent being primarily used for the production of cellulose acetate for photographic films and that of acetic anhydride. Acetic acid also possesses antiseptic properties, so it can be employed as antimicrobial agent. The current approach for the production of acetic acid is not environmental benign, with high energy cost and the production scale cannot meet the demand for both industrial and pharmaceutical needs [99,100]. Development of alternative production routes that will cover the current demand and be environmentally friendly is thus needed. One of the promising and cost-effective methods to produce acetic acid is the photocatalytic transformation of CO₂. Recent advancements in this domain are described below but note that only a few of case studies were found in the literature of the last decade.

Zhang and co-authors [12] integrated ultra-small gold nanoclusters (AuNCs) and cadmium sulfide (CdS) into *M. thermoacetica* bacteria and compared their performance in the photosynthesis of acetic acid from CO₂ (**Fig. 9a**). These bio-based photocatalytic systems were found efficient for CO₂ fixation into acetic acid under low light irradiation and dark (**Fig. 9b**). The authors observed that the yield in acetic acid by the gold-based biophotocatalyst after four days is conspicuously higher than that obtained with cadmium sulfide-based bio-photocatalyst under the irradiation of low light. Moreover, the authors found that the activity of ordinary bacterium (*M. thermoacetica*) and gold-based bio-photocatalyst in darkness are relatively low. A monitoring of the electron transfer mechanism via photoluminescence confirmed that the photo-excited electrons from the gold nanoclusters are migrated to distributed redox mediators in the bacteria cytoplasm such as flavoproteins. Evaluation of the effect of cysteine on the yield of acetic acid production revealed that the gold-based bio-photocatalyst produced the highest yield (6.01 mmolg⁻¹) at 0.3 % wt Cys. It is worth to mention that the ligands at the gold nanoclusters surface and the gold core size can be changed in order to tune their biochemical and biophysical characteristics, including cell uptake, cytotoxicity, biocompatibility, and molecular electronic structures. This study demonstrated that integration of gold nanoclusters as intercellular photosensitizer inside the cytoplasm of *M. thermoacetica* bacterium allowed this non-photosynthetic latter to conduct effective photosynthesis of acetic acid from CO₂.

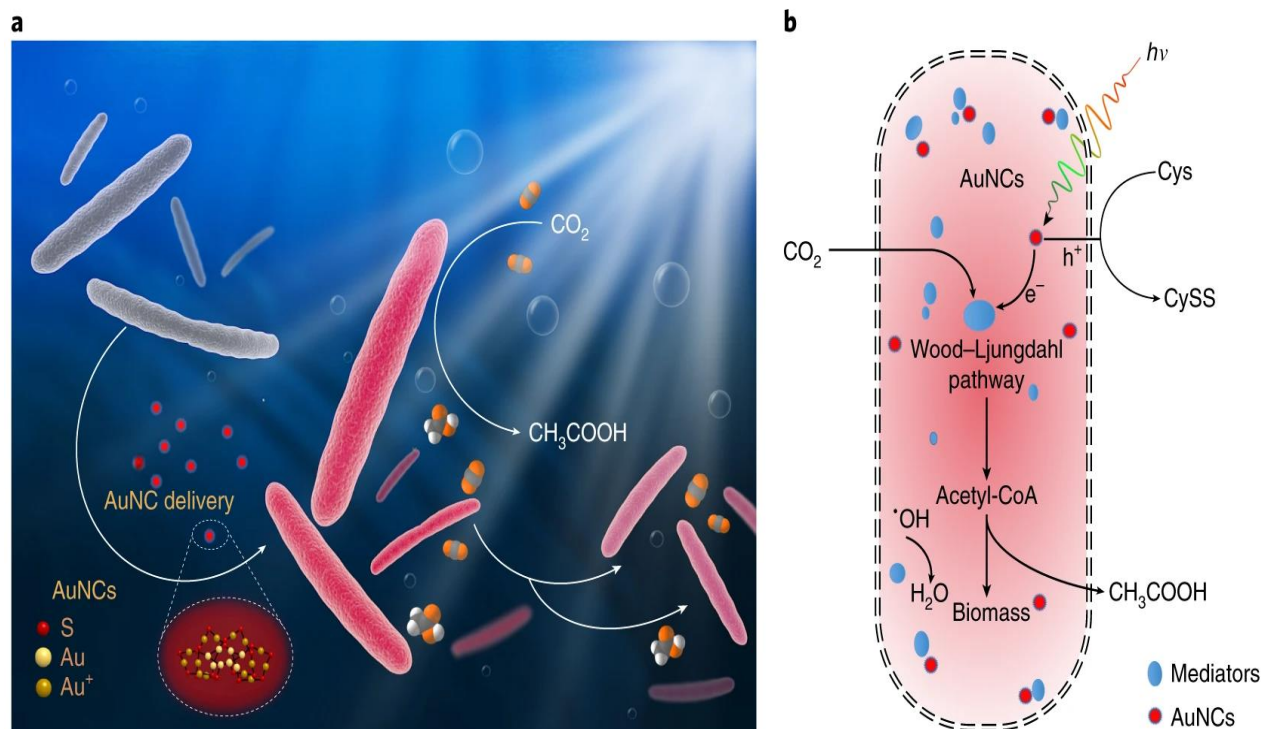


Figure 9 (a) Schematic illustration of *M. thermoacetica*/AuNCs hybrid system (b) Schematic bacteria electron transfer pathway for acetic acid generation under illumination of light

In another study, Gai et al. [101] developed an organic semiconductor – bacteria biohybrid photosynthetic system that efficiently converted CO₂ into acetic acid. The bacterium involved is *Moorella thermoacetica* whose surface was coated with perylene diimide derivative (PDI) and poly(fluorene-co-phenylene) (PFP) as photosensitizers. This led to the formation of a p-n heterojunction (PFP/PDI) layer with high hole/electron separation efficiency (**Fig. 10**). The π -conjugated semiconductors display excellent light-harvesting ability and biocompatibility. In addition, the cationic side chains of the two organic semiconductors could intercalate into cell membranes, thus allowing an efficient transfer of electrons to the bacterium. In this photocatalytic system, the bacterium could harvest photoexcited electrons from the PFP/PDI heterojunction and synthesize acetic acid from CO₂ under illumination. This bio-photocatalytic system showed an efficiency rate of 1.6% which is comparable to those of reported inorganic biohybrid systems.

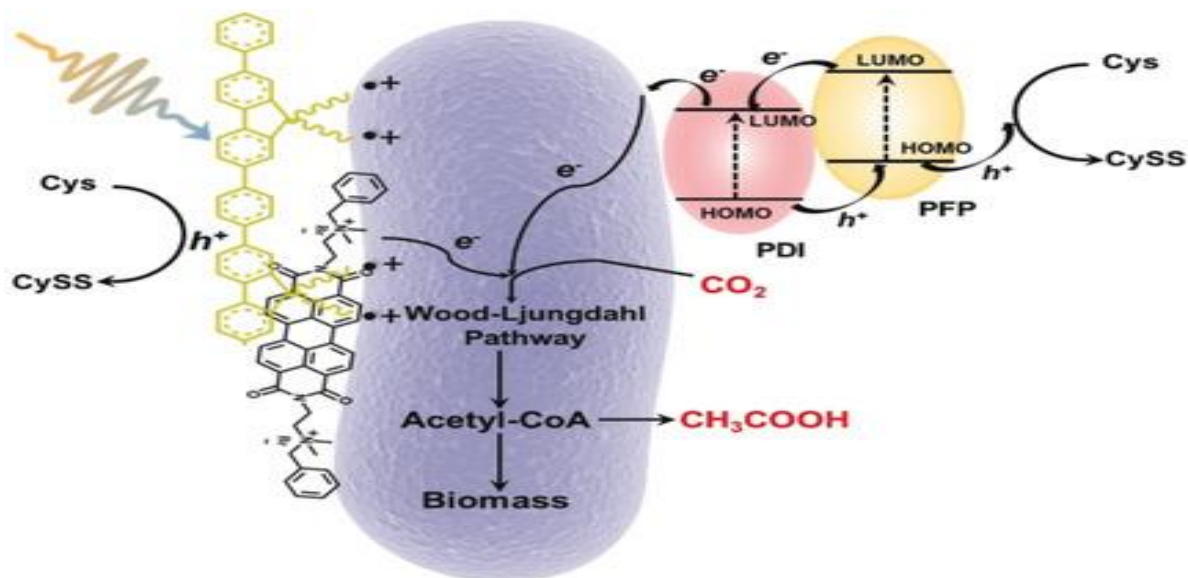


Figure 10. Wood–Ljungdahl pathway for CO₂ reduction in the presence of PFP/PDI/M. thermoacetica. Reproduced with the permission from [101]. Copyright 2020 Wiley online

Tharak and colleagues [102] reported microbial-mediated CO₂ in gas and gas-electro fermentation systems with and without applied potential by selectively enriched biocatalysts (**Fig. 11**). In the Heat + CO₂ condition during the gas fermentation (GF) stage, the maximum production was 0.84 g/L acetic acid with 31 mg.L⁻¹.h⁻¹ inorganic carbon fixation rate (ICFR). The GF was tested in a single-chambered microbial electrochemical system (MES; Gas-electro fermentation) at different applied potentials (-0.4 and -0.6 V vs Ag/AgCl [3.5 M KCl]). With -0.6 V applied potential, a higher acetic acid production of 1.02 g/L was obtained. The CO₂ fermentation efficiencies were improved by higher charge transfer rates (52% faradaic efficiency) and lower corrosion and resistances (R_{ct} -0.4 ohm). On continuous conditions, an abundance of Proteobacteria, Firmicutes, and Actinobacteria in the enriched biocatalyst represented a significant shift of the microbiome towards CO₂ feeding microbes.

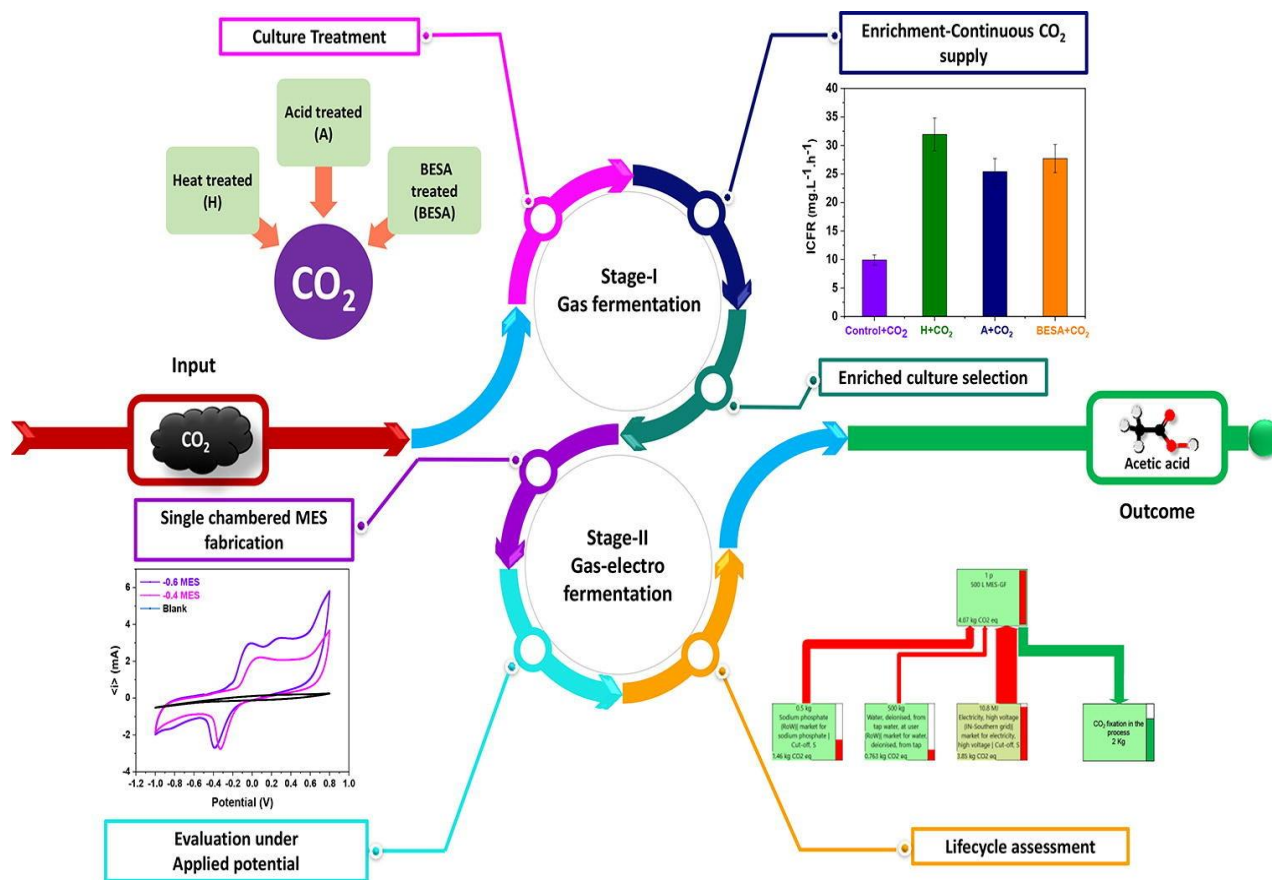


Fig. 11. Schematic diagram of acetic acid production from CO₂ via lignin/ionic based catalytic system. Reproduced with the permission from [102]. Copyright 2023 Elsevier

Wang and co-workers [103] reported a new strategy to produce renewably sourced acetic acid from lignin and CO₂/H₂. The catalytic system is based on an ionic liquid (that is 1-butyl-3-methylimidazolium chloride, [BMIm][Cl]) that contains a Ru-Rh bimetal catalyst (Ru₃(CO)₁₂ and RhI₃) and Li salt additives (LiI and LiBF₄). This system was effective for the production of acetic acid in the temperature range of 120-180 °C under low pressure of CO₂ (1 MPa), affording an acetic acid yield of 94% at 180 °C. In addition, taking lignin as the feedstock, 0.17 g of acetic acid was produced from 1 g of lignin. Finally, the catalytic system could be recycled 5 times without decrease in performance. Investigation of the reaction mechanism evidenced that lignin provided methyl group to form CH₃I, while the [BMIm]⁺-stabilized RhI₃ and I⁻-complexed Ru(CO)_n(n=1-4) cooperated and catalyzed the formation of acetic acid from CH₃I and CO₂/H₂.

Table 4 below summarizes the biomimetic materials explored for the production of acetic acid. While there is appreciable development towards the production of acetic acid from CO₂ using

biomimetic materials, the yields reported are very low and cannot satisfy the commercial purpose. This calls for more research efforts in this hot topic.

Table 4. Biomimetic photocatalysts for acetic acid production from CO₂

Photocatalytic system	Reaction conditions	Yield	Reference
M. thermoacetica/AuNCs hybrid system	Low-intensity simulated sunlight (calibrated by photodiode, air mass 1.5 global spectrum (AM 1.5), 2Wm ⁻²)	6.01 mmolg ⁻¹ after six days in the presence of cysteine as sacrificial agent	[12]
M. thermoacetica/CdSNPs hybrid	Air mass 1.5 global spectrum, 2 W m ⁻² , photons; 5 x 10 ¹³ cm ⁻² s ⁻¹ under light and dark flux at λ = 435 to 485 nm	85±12% quantum yield after four days	[104]
PFP/PDI/M. thermoacetica heterojunction	p-n 5.0 Mw/cm ² power density, under of light-dark cycle for 12h	0.1 mM in dark, 0.6 mM under light	[101]
Different mediated biocomposites	bacteria Heat-shock, Acid-shock and 2-Bromoethano sulphonate (BESA), 31 mg L ⁻¹ h ⁻¹ of ICFR, (-0.4 and -0.6 V vs Ag/AgCl [3.5 M KCl]), R _{ct} = 0.4 ohms	1.02 g/L at 52% FE	[102]
Lignin/ionic catalytic system	based 180 °C, 1 MPa	94% yield, 0.17g acetic acid produced from 1g of lignin	[103]

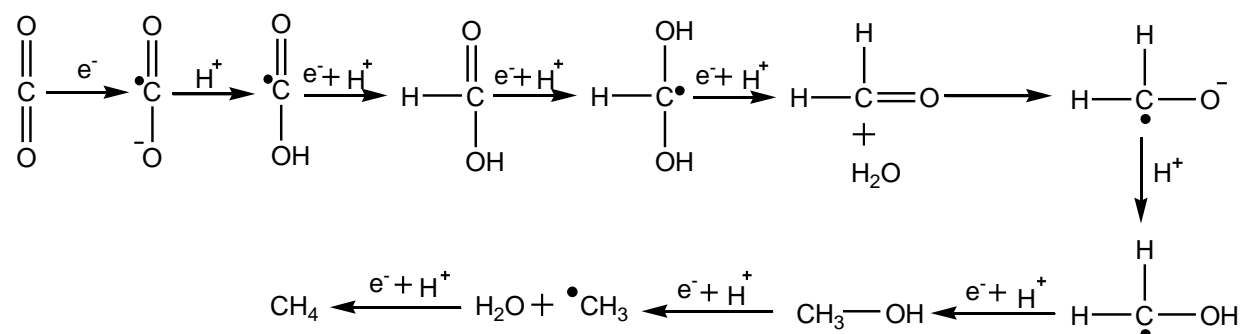
PPF: poly(fluorene-co-phenylene), PDI: perylene diimide derivative, FE: Faradaic efficiency, ICFR: Inorganic carbon fixation rate

6. Mechanistic insights of biomimetic photocatalysts for CO₂ Reduction

There is no definite mechanism for the reduction of CO₂ as it is a complex reaction involving radicals, electrons, and protons [105]. Generally, after the reduction of CO₂ by a one-electron transfer and the formation of surface-bound CO₂⁻, the process proceeds through a sequence of elementary steps involving the transfer of a proton, electron or radical (H[•]), as well as breaking C-O bonds and creating C-H bonds. Several intermediates are radical species, partially accounting for the number of possible pathways and final products at different stages of recombination. The exact sequence and mechanistic flow of each subsequent step have not been thoroughly elucidated, but some literature suggests that photocatalytic reduction of CO₂ *via* biocatalysis has a similar pathway than that described for transition metal photocatalysis [105].

Hydrogen radicals (H[•]) bind to the oxygen atom of the adsorbed carboxyl radicals (CO₂⁻), resulting in an immediate cleavage of the C-O bond, subsequently leading to the formation of carbon monoxide. In a separate mechanism, the carboxyl radicals recombine with hydrogen radical to form formic acid. In a next step, formic acid accepts another hydrogen radical to form a dihydroxymethyl radical, which is dehydrated when another hydrogen radical is attached to the formaldehyde. Two additional steps of reduction lead to CH₃OH formation, which is further reduced to methane in two more steps of reduction. A proposed mechanistic pathway (by formaldehyde, carbene, glycol pathways) for the reduction of CO₂ to CO, formic acid, and methanol as described in the literature is presented in **Fig. 12** [106].

Formaldehyde pathway



Carbene pathway

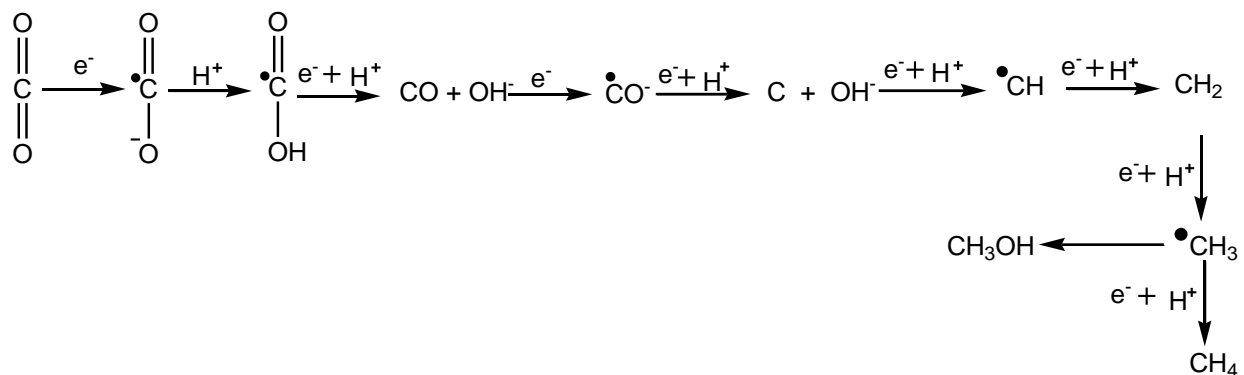


Figure 12. Two reduction routes for CO₂; top, formaldehyde pathway and bottom carbene pathway [106].

The mechanism of CO₂ conversion involves two-electron reduction mediated by metal-based catalysts or enzymes. Most photocatalysts are metal-based [107][108][109][110][111]. For a commonly described metal photocatalyst, the semiconductor part absorbs photon energy, which leads to the excitation of electrons from valence band (VB) to conduction band (CB), creating electron-hole pair. The holes in VB have strong oxidizing ability, especially for water oxidation and electrons in CB reduce CO₂. Water oxidation produces hydrogen ions and hydroxyl radicals, and the activated CO₂ reacts with excited electrons and hydrogen ions, which finally result in the formation of formic acid, formaldehyde, and methanol [112]. When AgCuInS₂ supported on graphene-TiO₂ are parallelly activated, electrons transfer through the graphene to reach the VB of TiO₂ to form the donor level, thus, resulting in the formation of photogenerated electrons (e⁻) and holes (h⁺) at the active sites of AgCuInS₂ photocatalyst. The holes (h⁺) react with the adsorbed 2H₂O to produce oxygen (O₂), electron (e⁻) and protons (H⁺). Surface-adsorbed CO₂⁻ radical generated by the injection of a single electron reacts with e⁻ and H⁺, producing CO, which undergoes a further reduction process forming the surface-adsorbed C. CO₂ further reacts with excited electrons (2e⁻) and hydrogen ions (2H⁺) to yield HCOOH. The further reaction of HCOOH with excited electrons (2e⁻) and hydrogen ions (2H⁺) led to a loss of water (H₂O) to produce an intermediate, formaldehyde, (HCOH) (typical of formaldehyde pathway, see Fig 11). Surface adsorbed formaldehyde reacts with 2e⁻ and 2H⁺ to yield methanol as a main product. A schematic representation of photocatalytic CO₂ reduction process with AgCuInS₂ is illustrated in **Fig. 13**.

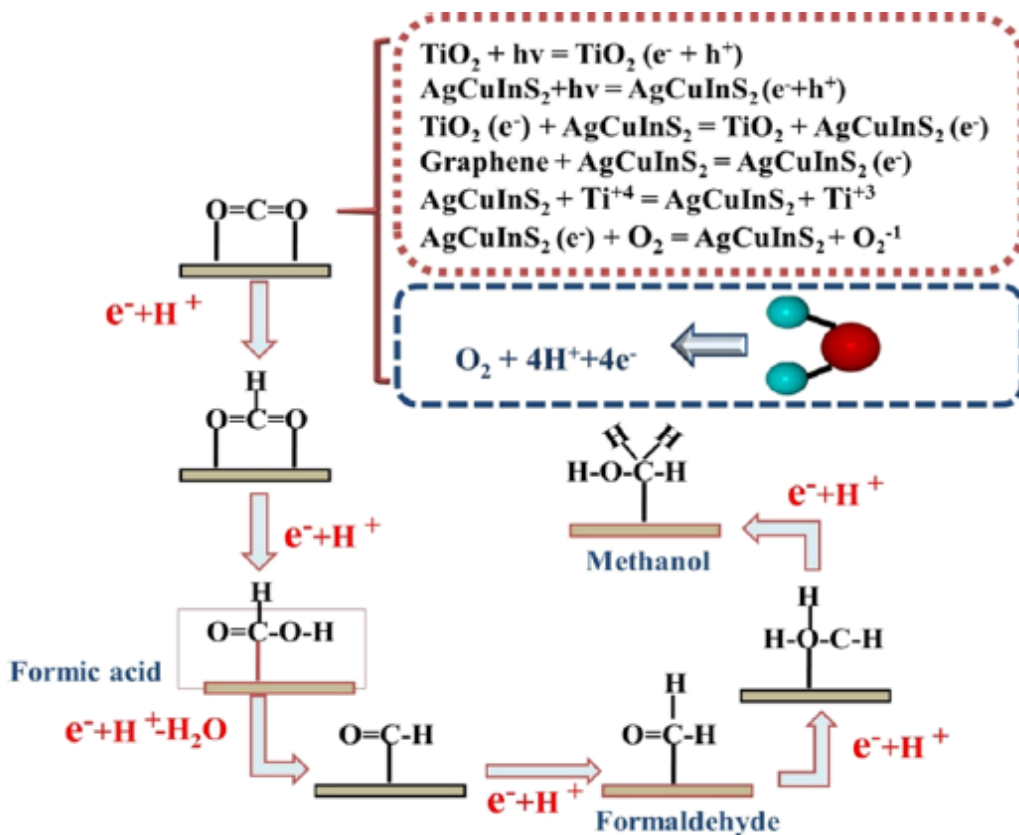


Figure 13. Schematic diagram of the photocatalytic CO₂ reduction process. Re-used with permission from. Copyright (2021) American Chemical Society. [112]

Getahun et al. [55] concerned the use of metal oxides (ZnO and CuO) embedded with bio-based materials (such as chitosan) as photocatalysts. Both ZnO and CuO absorb UV light, making energy separation to occur, and resulting in hydrogen ion (H⁺) and electron (e⁻) generation. The schematic representation of photocatalytic CO₂ reduction using ZnO and CuO under UV light is illustrated in **Fig. 14**. The valence-band energy level of CuO (n-type) is higher than that of ZnO (p-type), the holes of ZnO generated by absorbing light energy transfers to the valence-band level of CuO, leading to the holes (h⁺) on CuO/ZnO composite oxidizing water (H₂O) to oxygen (O₂), hydrogen ion (H⁺) and an electron (e⁻). CO₂ (trapped within amine groups of chitosan and dendrimer, -NH₂---CO₂) reacts with e⁻ to form anion radical CO₂^{•-} (a single electron reduction of CO₂) followed by a reaction with protons H⁺, a process known as “proton-assisted multielectron reduction”. Surface-adsorbed CO₂^{•-} radical reacted with 6e⁻ and 6H⁺ (CO₂ + 6H⁺ + 6e⁻ → CH₃OH + H₂O), producing methanol. Therefore, the photoredox pathway could be considered a

proton-assisted multielectron reduction via the formaldehyde route. Pathway for CO and formic acid production are similar to those reported by Otgonbayar et al [110].

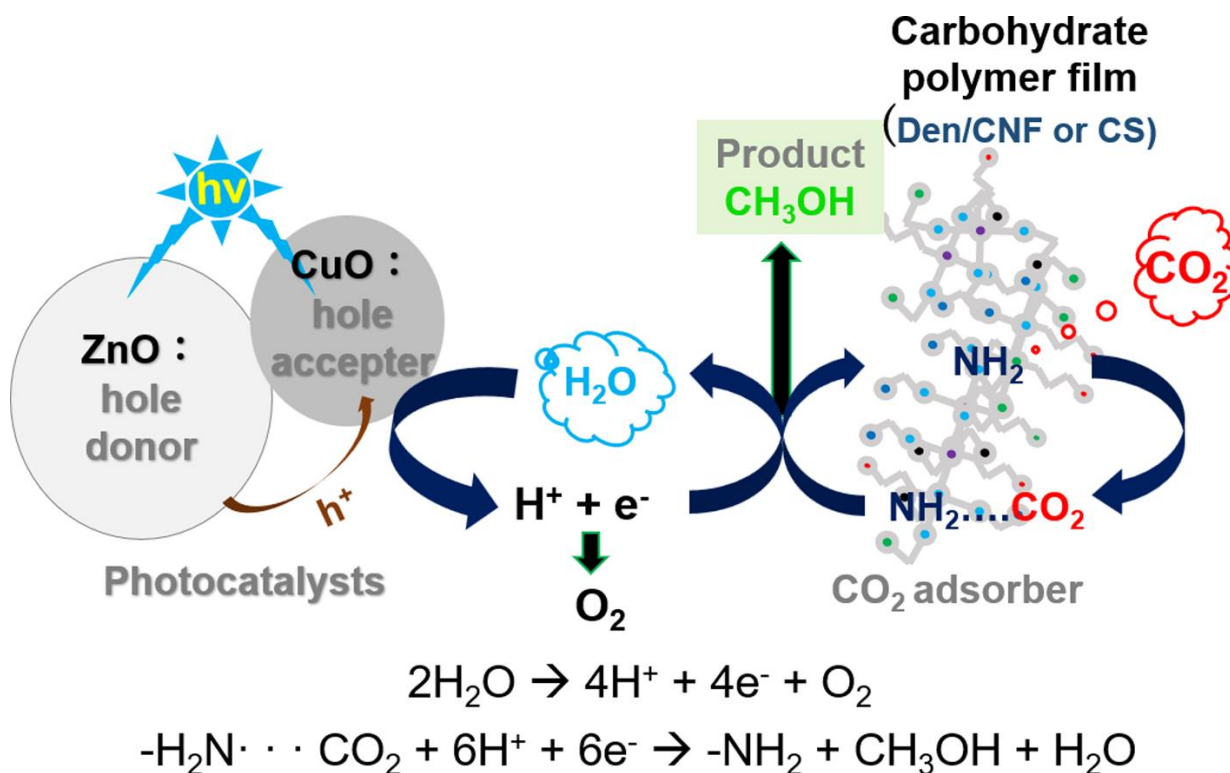


Figure 14. Illustration of CO₂ photocatalytic reduction to methanol with H₂O oxidation on films of CuO/ZnO catalytic-embedded carbohydrate polymer. Reproduced with permission from [55]. Copy rights (2022) Elsevier.

The photocatalytic reaction mechanism of metal-based systems are similar to those followed by enzymes such as carbon monoxide dehydrogenases (CODHs) [113] and formate dehydrogenases (FDHs) [114][115], in which two to four electron reduction of CO₂ is required for conversion to carbon monoxide, formic acid/formate, and methanol. The bio-inspired mechanistic insight for two-electron reduction of CO₂ to CO, as described by Mondal et al, 2014 [105] goes through proton-coupled electron transfer (PCET, H⁺ + e⁻), from a molecule that acts as “electron and proton” donor (reductant) to another one that acts as an acceptor (oxidant). Both biocatalysis and homogeneous transition-metal catalysis usually proceed *via* redox reaction where hydride transfer takes place for formate ion production (**Fig. 15**).

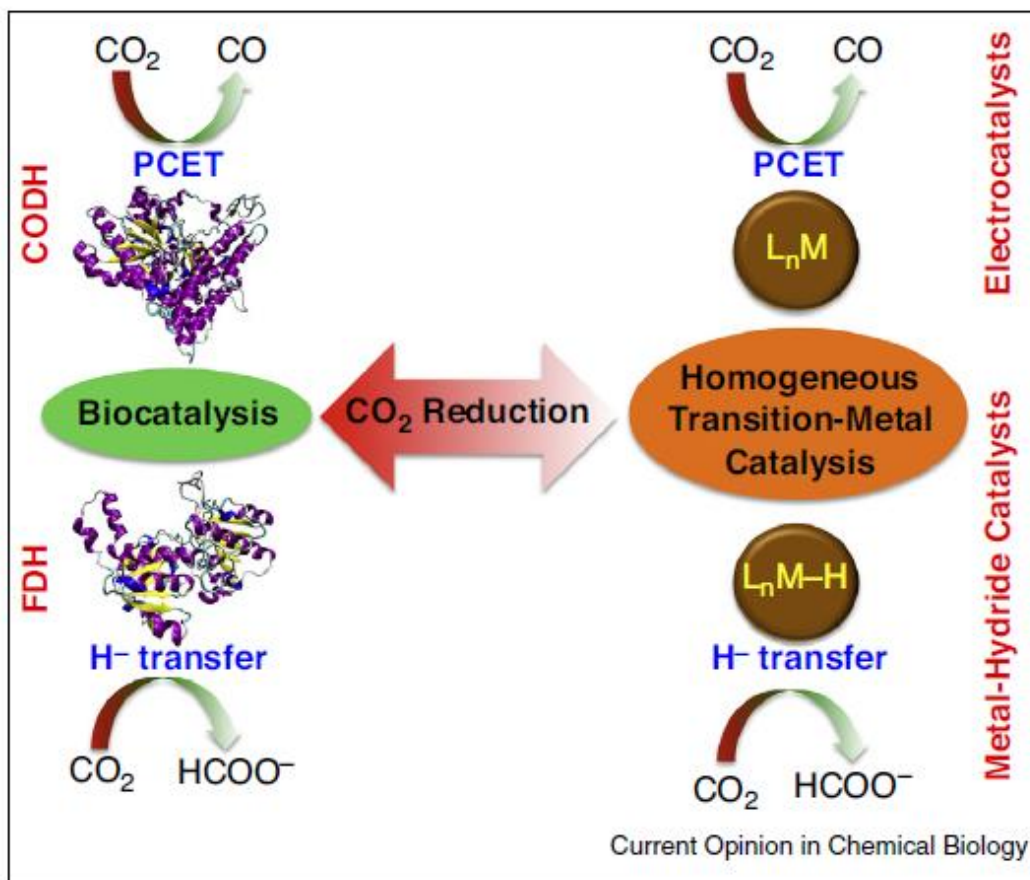


Figure 15. Overview of the bio-inspired mechanistic insight into the two-electron reduction of CO_2 to CO and FA (PCET = proton-coupled electron transfer). Copyright [105].

FDHs are known to catalyze the reduction of CO_2 to formic acid through a proton (H^+) transfer mechanism ($\text{CO}_2 + \text{NADH} + \text{H}^+ \leftrightarrow \text{HCOOH} + \text{NAD}^+$) as well as reversible oxidation of formate to CO_2 ($\text{HCOO}^- \leftrightarrow \text{CO}_2 + \text{H}^+ + 2\text{e}^-$) [114][116][117]. β -Nicotinamide adenine dinucleotide hydrate reduced salt (NADH) is an essential co-factor for FDH, its efficiency is highly dependent on the NADH concentration as a hydrogen donor. NAD^+ is the oxidized form of β -Nicotinamide adenine dinucleotide hydrate (NAD). The active site of the NAD^+ -based FDH showing the C4-atom position on pyridine ring (**Fig. 16a**). Mondal et al., [105] reported the direct hydride transfer, occurs from the C-atom of the formate to the C4-atom of pyridine ring of NAD^+ , leading to NADH and CO_2 (**Fig. 16b**). The neighbouring residues on FDH such as Thr (Thr282), Asp (Asp308), Ser (Ser334) and Gly (Gly335) interact with the positively charged NAD^+ ring and stabilize it, thereby promoting the hydride transfer from formate to C4 [118][119][120].

Generally, enzymatic reduction of CO_2 usually proceeds to HCO_2^- , and the oxidized form of nicotinamide cofactor (NAD^+) is reduced to NADH by glutamate dehydrogenase (GDH).

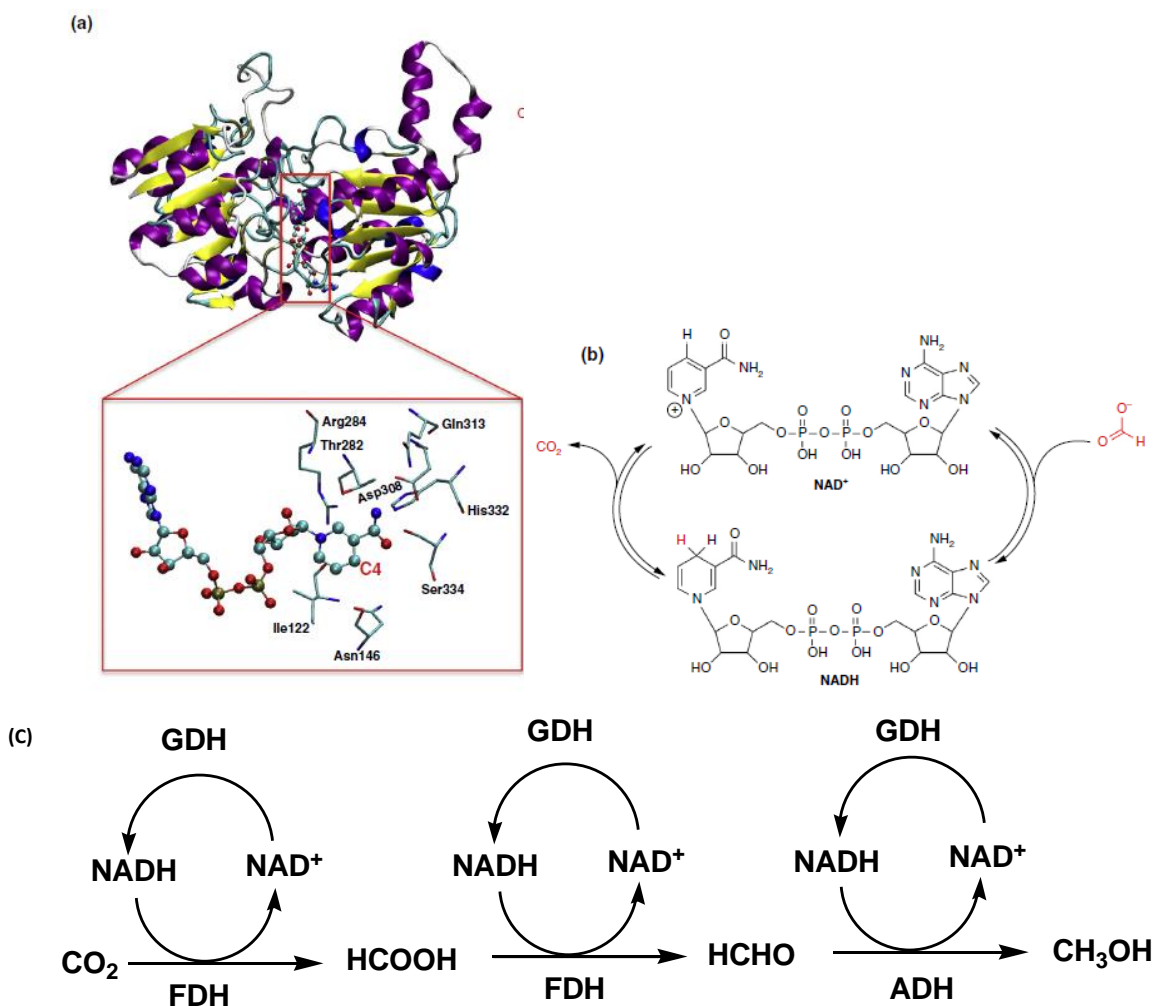
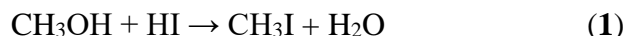


Figure 16. Two-electron reduction of CO_2 to HCOOH ; (a) the active site of the NAD^+ -based FDH , (b) the catalytic mechanism of FDH Copyright [105], (c) a simplified reaction pathway of CO_2 to methanol (FDH , formaldehyde dehydrogenase (FDH), alcohol dehydrogenase (ADH)). The regeneration of NADH was catalyzed by glutamate dehydrogenase (GDH) [114].

A complex system of enzymes such as adenosine triphosphate (ATP), guanosine triphosphate (GTP), nicotinamide adenine dinucleotide (NAD or NADH for the reduced form), or nicotinamide adenine dinucleotide phosphate (NADP or NADPH for the reduced form), working in tandem is vital when considering biological conversion of CO_2 . There is a need to further

research in this area especially around the regeneration of cofactor NADH, which in turn, improves enzyme efficiency during complex CO₂ bioprocessing applications.

In the formation of acetic acid, methanol carbonylation is required, and the reaction comes after the formation of Methanol (photocatalytic reduction of CO₂) [101] by using metals catalysts such as gold, platinum, palladium, osmium, copper-nickel, copper-Mordenite, palladium-cerium, ruthenium, rhodium, and lanthanum. Metal complexes play a huge role in achieving the carbonylation reaction through formation. In a typical Rh/CH₃I catalysed system, a series of activated anionic species such as cis-[Rh(CO)₂I₂]⁻, [(CH₃CO)-Rh(CO)I₃]⁻ and [(CH₃CO)-Rh(CO)₂I₃]⁻ to assist with product formation. Halogen additives is required to promote the reaction by activating C-O bond in methanol and on transition metal complexes to form methyl iodide (**Eq. 1**) which reacts with carbon monoxide to form acetyl-iodide (**Eq. 2**) and can quickly reach acetic acid final product when hydrolyzed in water (**Eq. 3**). The metal catalyst is recycled at the end of the reaction.



Conclusion and perspectives

The potential of using bio-based photocatalysts for the conversion of CO₂ into value-added products, has been emphasized by describing the main advances in this domain from the literature in the last decade. The investigation of such catalytic systems is fundamental as this may afford the development of environmentally friendly, and sustainable processes as alternatives to replace the use of fossil fuels and consequently, contribute to the decrease global warming. The reported bio-inspired photocatalytic systems are presented following the target chemicals, namely carbon monoxide, formic acid, methanol and acetic acid (via carbonylation of methanol) all being key chemicals for a direct use in various applications or as intermediate synthons. Several case studies concern the use of metal catalysts combined with biomaterials which, under photocatalytic conditions, allow the formation of CO₂-derived carbon molecules. The reduction of CO₂ using bioinspired catalysts is generally accompanied by proton-coupled

electron transfer (PCET) resulting in C–O bond cleavage to yield CO, an initial carbon-based product. CO₂ conversion to formic acid is usually promoted by formate dehydrogenase (FDH) and its co-factor NADH, responsible for donating hydrogen [117][118]. Carbonic anhydrase one of the fastest enzymes that catalyses CO₂-hydration reaction has been combined with formate dehydrogenase (FDH) with co-factor NADH to form formate/formic acid [119][122][121][119]. One of the major drawbacks is the depletion of NADH during catalytic reduction reaction to NAD⁺, thus requiring continuous regeneration to NADH to have sustained formic acid production. The enzymes used for conversion of CO₂ to CH₃OH are Formate Dehydrogenase (FDH) that utilizes NADH as a cofactor, and alcohol dehydrogenase (ADH). Enzymes usually exhibit much reduced activities over time due to impurities, slight change in reaction condition, (such as pH and temperature). The presence of transition metal/metal oxides incorporated unto biopolymers and enzymes facilitate the reduction of CO₂ to various products by promoting metal-CO₂ interaction. An example is [NiFe] CO Dehydrogenase ([NiFe] CODH), an enzyme used for conversion of CO₂ to CO. Zinc complexed Carbonic Anhydrase (CA), Zn[CA] is used for the conversion of CO₂ to bicarbonate (HCO₃⁻) [119].

Most of the enzymes being investigated in CO₂ reduction are naturally occurring and can denature easily under varying pH and temperature. Some key issues still remain that need to be considered when looking for further advancement in the biological catalysed CO₂ reduction. Firstly, the use of transition metals/metal ions incorporated unto enzymes protein ligands having high CO₂ binding affinity is vital for converting CO₂ molecules to desired products. Secondly, the depletion of NADH during catalytic CO₂ reduction reaction to NAD⁺, as well as the higher stability of CO₂ molecule, resulting in its lower reactivity. To avoid this problem, it is necessary to regenerate NADH during CO₂ reduction. A separate NADH regeneration step which involve the use of combined chemical, photocatalytic, biocatalytic, and/or electrochemical reactions within a single set-up must be considered when engineering efficient bioinspired catalyst for CO₂ reduction.

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Conflicts of interest

The authors declare no conflict of interest

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