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## Biomimetic photocatalysts for the transformation of CO<sub>2</sub>: Design, properties and mechanistic insights

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#### **Abstract**

The increase in CO<sub>2</sub> concentration in the atmosphere due to the intensive burning of fossil fuels, for the needs in energy of our modern society, has strongly contributed to the global warming. One of the promising strategies to fight this menace is to capture atmospheric CO<sub>2</sub> and convert it into value added products, including fuels and chemicals. Among the methods developed for the transformation of CO<sub>2</sub>, the sunlight-activated photocatalytic conversion appears as the renewable and easy to use method. For this purpose, it is required to have at disposal proper materials to act as catalysts and light absorbing materials to harvest the sun energy, and then to combine these components with an electron carrier, in an efficient way in order to access the desired fuels by a photocatalytic sustainable CO<sub>2</sub> conversion. Among the efforts in research dedicated to the production of solar fuels, studies focused on the design and development of bio-based photocatalytic systems as alternative to conventional photocatalytic systems have emerged in the literature in the last decade. The objective of this review is to show the readers the recent advances in the fabrication of bio-based photocatalysts for the transformation of CO<sub>2</sub> into value added products. This review specifically focuses on the conversion of CO<sub>2</sub> into carbon monoxide, methanol, formic acid/formate and acetic acid which are the main target products described. Methods adopted to enhance the selectivity and efficiency of the bio-derived photocatalysts developed are also discussed and a section is dedicated to computational

investigations. Finally, the current challenges and perspectives in the development and use of these novel and greener photocatalysts are presented.

Keywords: CO<sub>2</sub> transformation; solar fuels; biomimetic photocatalysts; enzymatic catalysis

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#### 1. Introduction

Up to recent period, the burning of fossil fuels has been a major source of energy of our modern society. The inordinate consumption of fossil resources has led to a severe global warming (with new records in the last three years) due to the excessive discharge of greenhouse gasses into the atmosphere, and most particularly of carbon dioxide (O=C=O), and this has become an international crisis that calls for an urgent solution. CO2 is the primary driver of the global warming [1]. In addition, it is a long-lived greenhouse gas, with one-third still present in the atmosphere after a century of release [2]. Scientists believe that if purposeful action is not taken with immediate alacrity to lower CO<sub>2</sub> concentration in the atmosphere, global warming and impact of climate change could inevitably lead to the death of more than 250,000 people per year, put 100 million people into abject poverty by 2030 and displace 150 million people or more by 2050 [3]. The transformation of atmospheric CO<sub>2</sub> is a herculean task because of its high stability. However, it may have significant benefits in terms of access to carbon based platform molecules and fuels [4]. Among the various strategies available for the reduction of CO<sub>2</sub>, photocatalytic transformation of CO<sub>2</sub> is unarguably a promising and green strategy that could solve the global challenge [5]. The reduction of CO<sub>2</sub> by photocatalysis is similar in mechanism to the natural photosynthesis, which can be explained using three separate but linked steps: (i) the

absorption of solar energy (ii) the separation of charges and (iii) surface redox reaction. Utilization of the ubiquitous solar energy in order to reduce CO<sub>2</sub> into renewable value-added products would in turn simultaneously improve the lingering global energy shortage, climate change and take a step towards the circular economy [6]. However, the major bottlenecks of existing photocatalysts (i.e. monocatalysts, liquid state Z-scheme, all solid states Z-scheme, direct Z-scheme and type II heterojunctions) are (1) fast recombination of photogenerated charge carriers, (2) poor redox power and (3) Inefficient charge separation thereby hindering the practical application of the technique [7-9]. According to general agreement, a good photocatalytic system must have quick charge transition, efficient charge separation, and good absorption in the visible spectrum, be composed of non-toxic elements and display high stability [10]. Among the photocatalysts reported in the literature, bio-based photocatalysts have gained popularity in the recent decade. This is mainly due to their high reaction efficiency and their fascinating catalytic mechanism inspired by nature [11]. In comparison to their photosynthetic counterparts, non-photosynthetic organisms, especially those created in contemporary synthetic biology, have more desirable metabolic pathways, providing a tunable CO<sub>2</sub> metabolism for a variety of products [12]. Other interests of the bio-based photocatalytic systems for the transformation of CO2 into hydrocarbon fuels emerge from the abundance of the biomaterials used, environmental friendliness, and energy-saving if the process is activated by sunlight, to mention but a few. In the reviewing exercises that cover materials for application as catalysts in the transformation of CO<sub>2</sub>, the recent advancements in the use of bio-based photocatalysts is only little highlighted. For example, Yau and co-workers [13] recently published an updated review which gives precedence to microbial hybrid photosystems for CO<sub>2</sub> fixation. Notwithstanding, the review work did not provide elaborate survey on other bio-based photocatalysts and it fails to address the theoretical investigation in order to rationalize the mechanism of CO<sub>2</sub> reduction.

In order to sensitize the scientific community to the advantages of using bio-derived photocatalysts (i.e. a natural light driven catalytic reaction using enzyme, bacterial and polymers) for the conversion of CO<sub>2</sub>, this review intends to gather the recent research (i.e. covering the last decade) on the design of such photocatalytic systems for the transformation of CO<sub>2</sub> into fuels. The examples reported are classified into five sections in line with the target hydrocarbon fuel, namely carbon monoxide, methanol, formic acid/formate and acetic acid (see sections 2, 3, 4 and 5, respectively). Furthermore, in section 6, an in-depth analysis on the interest of computational

investigations for the development of bio-derived photocatalysts for CO<sub>2</sub> reduction is given. Computational data may either complement or to give prior understanding into experimental investigations. Finally, current challenges and future outlook are discussed.

#### 2. Biomimetic photocatalysts for the transformation of CO<sub>2</sub> to carbon monoxide

Carbon monoxide (CO) is one of the attractive value-added products that can be generated from the reduction of CO<sub>2</sub>. It is commonly used as an intermediate in some chemical processes including the production of drugs, fragrances, formaldehyde or benzaldehyde and methanol [14]. Some chemical industries also use CO in the preparation of inorganic chemicals such as metal carbonyls like nickel tetracarbonyl (**Eq. 1**) and iron pentacarbonyl (**Eq. 2**).

$$Ni + 4CO \rightarrow Ni(CO)_4$$
 (1)

$$Fe + 5CO \rightarrow Fe(CO)_5$$
 (2)

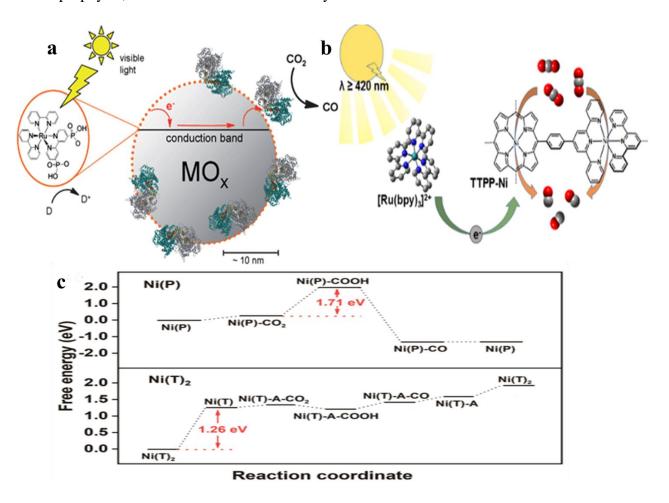
Furthermore, CO plays an important role alongside with hydrogen gas, forming together the syngas that is used in the Fischer-Tropsch process towards superior hydrocarbons [15].

From thermodynamic and kinetic points of view, the generation of CO from  $CO_2$  is one of the easiest compared to that of other hydrocarbon fuels. The reduction of  $CO_2$  into CO and  $H_2O$  requires two electrons and two protons with a reduction potential of 0.53V (vs. NHE at pH = 7) (**Eq. 3**).

$$CO_2 + 2H^+ + 2e^- \rightarrow CO + H_2O E = 0.53V$$
 (3)

Different bio-based photocatalytic materials have been employed for the conversion of CO<sub>2</sub> to CO. In one of the earliest studies, Woolerton et al. developed a photocatalytic system that comprises metal-oxide nanoparticles functionalized by the enzyme carbon monoxide dehydrogenase (CODH) as catalyst and sensitized to visible light by a ruthenium bipyridyl photosensitizer, both components being supported on an anatase/rutile TiO<sub>2</sub> mixture as the semiconductor (electron donor). Using this photocatalyst activated by visible light they could effectively reduce CO<sub>2</sub> to CO [16]. The authors examined the CO production rates and stability as a function of each component (photosensitizer, enzyme and TiO<sub>2</sub>). Tolerance to O<sub>2</sub> and effects of different electron donors were also investigated, together with strategies to control enzyme binding at the surface of TiO<sub>2</sub> in order to enhance overall activity (**Fig. 1a**). Ding and co-authors [17] reported a functional porous coordination hybrid system composed of nickel porphyrin and

nickel bis(terpyridine) dual active sites as efficient bio-based photocatalyst for  $CO_2$  reduction into CO under the illumination of visible light ( $\lambda \ge 420$  nm) (**Fig. 1b**). The biomaterials exhibited an extraordinary photocatalytic activity towards the CO formation with an outstanding generation rate of 3900  $\mu$ molh<sup>-1</sup>g<sup>-1</sup> and selectivity of 98%. Also, they investigated the mechanism of action associated with dual active sites of the system, composed of metal porphyrin and metal bis(terpyridine), using theoretical simulations which further supported an excellent photocatalytic performance of these bio-hybrids as observed in the experimental investigation (**Fig. 1c**). The theoretical investigation demonstrated that the binding between  $CO_2$  and Ni(II) in nickel bis(terpyridine) is much easier than that with nickel porphyrin. The Ni–C Mayer bond order of \* $CO_2$  intermediate in nickel bis(terpyridine) was 0.533, much higher than that (0.009) in nickel porphyrin, which accounts for the stability of the intermediate of the former.



**Figure 1.** Mechanism of CO<sub>2</sub> reduction to CO by (a) CODH/ruthenium bipyridyl/anatase or rutile-TiO<sub>2</sub>. Reproduced with the permision from [16]. Copyright 2011 Royal Society of Chemistry. (b) Ni Porphyrin and Ni bis(terpyridine) coordinated polymer (TPP-Ni) (c) Schematic mechanism of CO<sub>2</sub> reduction catalyzed by Ni Porphyrin and Ni bis(terpyridine). Adapted with the permission from [17]. Copyright 2022 American Chemical Society.

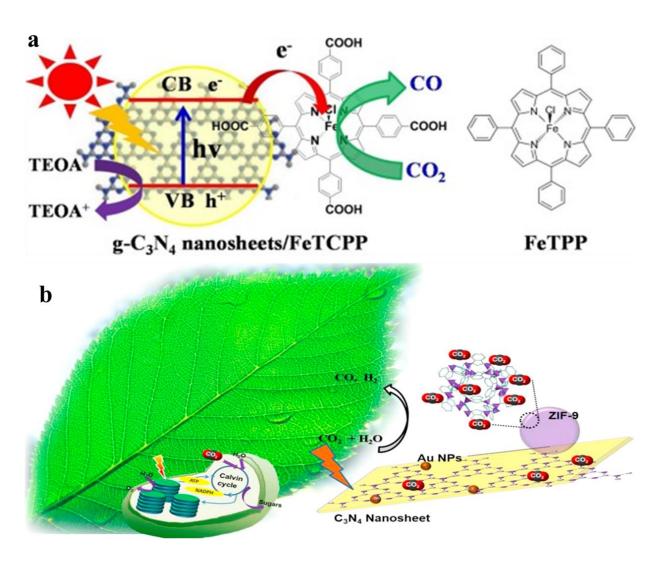
In a related study, Liang and co-workers [11] developed a biomimetic system based on metalloporphyrin and  $ZrO_2$  nanoframes that showed an evolution CO yield of 35.3  $\mu$ mol within 3 h of reaction and high selectivity rate of 93.1% as well as an apparent quantum efficiency of 1.84% which is 60.8 greater than pristine porphyrin-Ni with evolution rate of 0.58  $\mu$ mol.

The group of Zhao [18] constructed a covalently bonded reaction centre\_bio-hybrids comprised of Co-based porphyrin and a low molecular mass graphitic C<sub>3</sub>N<sub>4</sub> for the transformation of CO<sub>2</sub> to CO under the irradiation of visible light. They obtained a photocatalytic activity 13 times greater than that of pure C<sub>3</sub>N<sub>4</sub> (CO generation rate of 17 μmolg<sup>-1</sup>h<sup>-1</sup>), thus evidencing the positive role of the Co porphyrin. The improved activity was attributed to the efficient electron transfer and trapping by CO active sites, improved electron-hole separation and strong affinity of Coporphyrin for CO<sub>2</sub> adsorption.

Lin and colleagues [19] designed a highly efficacious g-C<sub>3</sub>N<sub>4</sub>/tetra(4-carboxyphenyl)porphyrin (III) chloride heterogeneous catalyst for selective photoreduction of CO<sub>2</sub> into CO under visible light (**Fig. 2a**). In this catalytic system they used abundant, environmentally friendly FeTCPP as the catalytic center and inexpensive g-C<sub>3</sub>N<sub>4</sub> nanosheets as the light absorber, maintaining a simple mechanical mix under constant stirring. A maximum rate of 6.52 mmol g<sup>-1</sup> of CO in 6 h, with up to 98% selectivity was obtained. The catalytic performance was attributed to an effective electron transfer from g-C<sub>3</sub>N<sub>4</sub> nanosheets to FeTCPP

Apart from the enzymitic/bacterial based photosystems reviewed so far, Zhou and co-authors [20] reported biomimetic polymeric semiconductor based hybrid nanosystems that combined a typical CO<sub>2</sub>/proton reduction catalyst, a cocatalyst, an electron mediator, and a CO<sub>2</sub> activator for artificial photosynthesis with water and CO<sub>2</sub>. g-C<sub>3</sub>N<sub>4</sub> nanosheets, which mimic the nanolayered thylakoids stacks, are shown to be promising photocatalytic elements with a planar configuration and a high surface area, and they serve as an excellent platform for the assembly of other

analogous elements (**Fig. 2b**). Au NPs are used as a suitable cocatalyst. ZIF-9 is used as a CO<sub>2</sub> concentrator and an electron mediator to promote the redox reaction. The highest production of CO and H<sub>2</sub> was achieved by ZIF-9(1%)Au-CNS at the evolution rates of 0.3 and 7.5 μmolh<sup>-1</sup>g<sup>-1</sup>, respectively. The study revealed that the incorporation of polymeric based g-C<sub>3</sub>N<sub>4</sub> and ZIF-9 significantly enhanced the CO evolution rates which is 122 folds higher compared to non-ZIF-9 modified systems.



**Figure 2**. Plausible mechanism of CO<sub>2</sub> photoreduction by (a) g-C<sub>3</sub>N<sub>4</sub>/FeTCPP hybrids (b) Photoconversion activity of C<sub>3</sub>N<sub>4</sub>/AuNPs/ZIF-9. Reproduced with the permision from [19] and [20] . Copyright 2018, 2016 Elsevier, respectively.

**Table 1** below outlines biomimetic photocatalysts that have been explored for CO production from CO<sub>2</sub> reduction. The synergistic incorporation of biomimetic photocatalysts has shown to be promising for the reduction of CO<sub>2</sub> to CO with high selectvity. However, the present bio based photocatalytic systems for CO production have not provided satisfactory activity and stability, key points that necessitate further research.

**Table 1**. Carbon monoxide production using bio-based photocatalysts

Photocatalytic system	Reaction conditions	CO	Comments	Reference
		yield/TON		
TTPP-Ni/metal based	Photocatalytic activity	3900 <b>μmolh</b> -	The	[17]
bis(terpyridine) with	under $\lambda \geq 420$ nm,	<sup>1</sup> g <sup>-1</sup>	composites	
dual active sites	trisopropanolamine		exhibited high	
	employed as		CO yield.	
	sacrificial agent, 15mg		However,	
	catslyst		there is partial	
			dissolution of	
			photocatalysts	
Zn-porphyrin sensitised	Aqeuous medium, red	Low yield	The slow	[21]
Mn (I)	light irradiation ( $\lambda =$		conversion rate	
	620 nm)		of CO <sub>2</sub> to CO	
			was atributed	
			to inadequate	
			e- migration	
			from porphyrin	
			to the catalyst	
Carbon monoxide	Carried out under	Good yield	The size and	[22]
dehydrogenase	viisible light		morphologgy	
functionalized with	irradiation		of the	
CdS nanocrystals			nanocrystals	
			greatly	

			enhanced the
Nafion assisted Re(I)-RuL	The catalytic activity was done under visibile light for 20h,	454 μmolh <sup>-1</sup> g <sup>-</sup>	of nation improved the photocatalytic
Polymeric based g- C <sub>3</sub> N <sub>4</sub> /AuNPs/ZIF-9	80 kPa of pure CO <sub>2</sub> gas was purged, 300W Xe arc lamp as light source	0.4 μmolh <sup>-1</sup> g <sup>-1</sup>	activity  The dominant [20] evoluted gas is  H <sub>2</sub> which is not favourble for the reaction
g-C <sub>3</sub> N <sub>4</sub> /FeTCPP	Under visible light greater than 420 nm. TEOA was used as sacrificial agent	6.52 mmolg <sup>-1</sup>	The findings [19] indicates that the electrons migrated from $g-C_3N_4$ to FeTCPP
ZrO <sub>2</sub> nanoframes/metallopor phyrins	Visible light	35.3 μmol	The bio based [11] material efficiently transfer electrons to the Ni catalytic sites which in turn increased the CO generation

TON: Turn over frequency, FeTCPP: tetra(4-carboxyphenyl)porphyrin (III) chloride

#### 3. Biomimetic photocatalysts for the transformation of CO<sub>2</sub> to methanol

Methanol or methyl alcohol (CH<sub>3</sub>OH) is an organic compound primarily used as a solvent, cosolvent, or feedstock in the chemical industry. Methanol has an octane number of 113 and a density that is half that of the gasoline, as thus, can be mixed with gasoline with no technical modifications and be used in methanol-fuelled vehicles [24]. Although methanol is considered an environmentally friendly fuel (no oxides of sulphur or nitrogen), the production of CO during its combustion limits its use as a pure fuel (100%). Hence it is mixed with gasoline on a ratio of 85% (methanol) - 15% (unleaded gasoline) [25]. More usages of methanol include hydrogen storage and transportation as an energy carrier as well as an easily transportable fuel, and also production of intermediates and synthetic hydrocarbons as a C1 building block [26]. Methanol is therefore regarded as a transition molecule from fossil fuels to renewable energies.

Methanol can be synthesized from carbon-containing materials such as natural gas, coal, biomass, and carbon dioxide (CO<sub>2</sub>). Natural gas is currently the primary source of methanol contributing approximately 90% to the world 111 million metric ton production [27]. Unfortunately, natural gas and coal are fossil fuel resources, thus depleting non-renewable resources and significantly contributing to greenhouse gas emissions. The burning of fossil fuels (natural gas, coal, and oil) leads to CO<sub>2</sub> emissions which contribute significantly to global warming. Therefore, green production of methanol has gained a great attention of academic and industrial researchers.

Catalytic hydrogenation of CO<sub>2</sub> has been proposed as a green methanol production process. This process falls under the carbon capture and utilization (CCU) strategy which uses catalysts in the reaction of CO<sub>2</sub> and hydrogen (Eq. 4). The transformation of CO<sub>2</sub> to methanol is considered a green process when using captured CO<sub>2</sub> and/or hydrogen from renewable energy resources. The study by González-Garay et al. [28] shows how much CO<sub>2</sub> emissions can be avoided when using the green methanol synthesis process. However, the green methanol production *via* CO<sub>2</sub> hydrogenation is not economically feasible compared to the fossil-based process due to high costs of renewable hydrogen [28]. Therefore, there is an urgent need for developing cost effective CO<sub>2</sub> conversion technologies to produce methanol in a green way. Amongst many CO<sub>2</sub> conversion technologies such as biological, thermochemical, and electrochemical routes, the photochemical process has gained extensive attention [29]. If using solar light as its source of

energy, the photocatalytic reduction of CO<sub>2</sub> into valuable chemicals like methanol may be a sustainable approach [30–34].

$$CO_2 + 6H^+ + 6e^- \longrightarrow CH_3OH + H_2O$$
 (4)

Enzymatic conversion of CO<sub>2</sub> requires energy source like for the photochemical processes. The energy sources can be diverse, including ultraviolet, electrical potential, or energy carrying molecules (adenosine triphosphate (ATP), guanosine triphosphate (GTP), nicotinamide adenine dinucleotide (NADH)). While in a photochemical process, an electron donor, a photosensitizer, and catalysts (e.g. TiO<sub>2</sub>, ZnO, graphene, etc.) are mainly required to drive selective production to methanol [35,36] [37–40]. These photocatalysts may be incorporated into biomaterials such as biochars and biopolymers with conductive properties (e.g. cellulose, chitosan, etc.) to improve their structural and electronic properties [32,41–43]. There have been few case studies on biomimetic photocatalysts for the reduction of CO<sub>2</sub> to methanol since 2015 due to cost of enzyme production, regeneration, nonetheless, the research area is slightly increasing (as seen in **Table 2**).

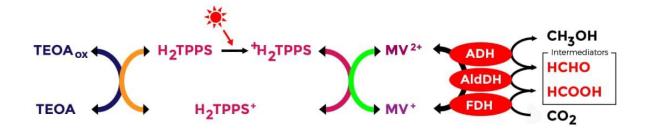
Recently, Tseng and co-workers [44] utilised sugarcane bagasse (SB) to support graphitic carbon nitride (gCN) for photocatalytic reduction of CO<sub>2</sub> to CO and methanol using a 14-watt LED light bulb. The presence of lignin in SB resulted in the semiconductor-like absorption edge and provided the interaction with gCN. The use of SB alone for photocatalytic study resulted in the formation of small amounts of MeOH. This is linked to the electronic transition of conjugated species within lignin (from SB) thus providing photoinduced charge carriers on SB. The introduction of SB (containing lignin) influenced the semiconductor-like absorption edge properties of the photocatalyst. Catalyst synthesis, g-SB-HT, using water (g-SB-HT-W) and ethanol (g-SB-HT-E) influenced MeOH yields, as the later solvent may be seen to have acted as hole scavenger thus increasing the yield of methanol (see Table 2).

Tseng and co-workers [45] also developed titanium oxide-chitosan film hybrid material (TiO<sub>2</sub>-chitosan) for the photocatalytic CO<sub>2</sub> reduction to methanol. This TiO<sub>2</sub>-chitosan material was able to selectively convert CO<sub>2</sub> in methanol at a production rate of 0.37 nmol/g (**Table 2**), showing a good stability. Similarly, Kavil and co-workers [46] synthesized a TiO<sub>2</sub>-based hybrid photocatalyst (Cu-C(glucose)/TiO<sub>2</sub>) by co-doping TiO<sub>2</sub> with copper (from copper sulfate) and carbon (from glucose) to photocatalytically reduce CO<sub>2</sub> to methanol. The photocatalytic experiment was performed in a Pyrex glass photoreactor filled with the catalyst and a NaOH

suspension and then purged with certified super–critical fluid grade carbon dioxide for 30 min. The photoreactor was irradiated with UV lamps (365 nm wavelength) for 5 h. In these conditions the Cu-C/TiO<sub>2</sub> catalyst allowed to achieve the highest methanol production ( $\sim$ 2600  $\mu$ M) with compared to counterparts C/TiO<sub>2</sub> (TiO<sub>2</sub> doped with only carbon) and TiO<sub>2</sub> P25, thus evidencing the synergy effect between the three components, Cu, C and TiO<sub>2</sub>.

Studies have shown that enzymes catalysed photoreduction of CO<sub>2</sub> to methanol involves using a three-step enzyme cascade including formate dehydrogenase (FDH), formaldehyde dehydrogenase (AldDH or FDH), and alcohol dehydrogenase (ADH) [46]. Co-enzymes and co-factors are usually sacrificial for photoredox reactions and are depleted in the process, thus, requiring enzymes that may regenerate the reduced forms. To further improve methanol yield, sequential and co-immobilization of dehydrogenases with polymeric membranes, metal complexes and oxides have been proposed [47].

Biocatalysts such as enzymes, namely formate (FDH), aldehyde (AldDH), formaldehyde (FalDH), lactate (LDH) and alcohol (ADH) dehydrogenases was tested for  $CO_2$  reduction to methanol [47,48]. Co-enzymes (also known as co-factors) such as nicotinamide adenine dinucleotide (NAD<sup>+</sup>) or its reduced form (NADH) and methylviologen (MV<sup>2+</sup>) are often used with FDH, NADH, AldDH and ADH. [48][49–51]. Amao and co-workers [36] developed a photochemical enzyme-based conversion system for the  $CO_2$  photoreduction to methanol, using FDH, AldDH, and ADH combined with  $MV^{2+}$  under the visible-light photosensitization of water-soluble porphyrin, tetraphenylporphyrin tetrasulfonate (H<sub>2</sub>TPPS) in the presence of triethanolamione (TEOA) (**Fig. 3**). This complex photoredox system allowed to achieve a methanol production concentration of 6.8  $\mu$ M. Study of the enzymatic kinetics evidenced that the rate-limiting step is the formic acid to formaldehyde reaction with AldDH and reduced form of  $MV^{2+}$  (Methyl viologen).



**Figure 3.** Methanol production from CO<sub>2</sub> catalysed with H<sub>2</sub>TPPS/MV<sup>2+</sup>/TEOA/FDH, AldDH and ADH photoredox system. Reproduced with permission from reference [36]. Copyright 2018 Elsevier.

The use of co-enzyme nicotinamide adenine dinucleotide reduced form (NADH), FDH catalyses both the reduction of CO<sub>2</sub> to formic acid, ADH catalyses the reduction of formic acid to formaldehyde, and ADH catalyses both the reduction of formaldehyde to methanol (Fig 4). The resulting co-enzyme nicotinamide adenine dinucleotide (NAD<sup>+</sup>) catalyses the oxidation of formic acid to CO<sub>2</sub>, formaldehyde to formic acid and oxidation of methanol to formaldehyde, respectively, thus inhibiting methanol production due to the redox nature of NAD<sup>+</sup>/NADH (Fig. 4). The oxidation processes were suppressed using oxidized form of MV<sup>2+</sup>, an electron carrier molecule, to drive the photochemical production of methanol. A reaction scheme for methanol production from CO<sub>2</sub> with FDH, AldDH and ADH using natural co-enzyme NAD<sup>+</sup>/NADH and artificial co-enzyme MV<sup>2+</sup>/MV<sup>+</sup> is presented in **Figure 4**.

For the system using NADH as a co-enzyme

$$CO_2 + NADH + H^+ \stackrel{FDH}{\longleftrightarrow} HCOOH + NAD^+$$

$$AIdDH$$

$$HCOOH + NADH + H^+ \stackrel{ADH}{\longleftrightarrow} HCHO + NAD^+ + H_2O$$

$$HCHO + NADH + H^+ \stackrel{ADH}{\longleftrightarrow} CH_3OH + NAD^+$$

$$Total reaction$$

$$CO_2 + 3 NADH + 3 H^+ \stackrel{CH_3OH}{\longleftrightarrow} CH_3OH + 3 NAD^+ + H_2O$$

For the system using reduced form of MV<sup>2+</sup> as a co-enzyme

$$CO_2 + 2 \text{ MV}^+ + 2 \text{ H}^+ \frac{\text{FDH}}{\text{AldDH}} + \text{COOH} + 2 \text{ MV}^{2+}$$

$$HCOOH + 2 \text{ MV}^+ + 2 \text{ H}^+ \xrightarrow{\text{ADH}} + \text{CHO} + 2 \text{ MV}^{2+} + \text{H}_2O$$

$$HCHO + 2 \text{ MV}^+ + 2 \text{ H}^+ \xrightarrow{\text{ADH}} + \text{CH}_3OH + 2 \text{ MV}^{2+}$$

$$Total \ reaction$$

$$CO_2 + 6 \text{ MV}^+ + 6 \text{ H}^+ \longrightarrow + \text{CH}_3OH + 6 \text{ MV}^{2+} + \text{H}_2O$$

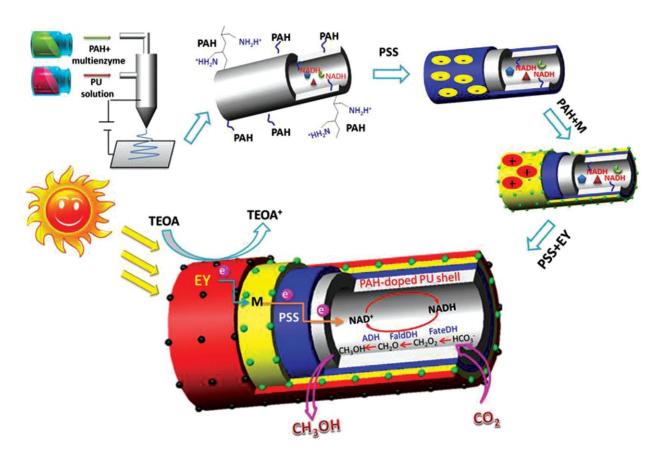
**Figure 4.** Scheme for the methanol production from CO<sub>2</sub> with FDH, AldDH and ADH using natural co-enzyme NAD+/NADH and artificial co-enzyme MV<sup>2+</sup>/MV<sup>+</sup>.

Zezzi Do Valle Gomes and co-workers [48] reported the biocatalytic cascade conversion of CO<sub>2</sub> to methanol using three dehydrogenases (FDH, ADH and FalDH) co-immobilized in tailored siliceous mesostructured cellular foams (MCF) functionalized with mercaptopropyl groups (MP) with no cofactor. With this catalytic system they obtained a methanol yield of 0.35 mmol/g<sub>enzyme</sub>/min, a value which might be improven by adding NADH regeneration.

Yadav et al. [52] prepared a CCG-IP type photocatalyst by attaching isatin-porphyrin (IP) chromophores with chemically converted graphene (CCG). A methanol production concentration of 11.21 µM was achieved using CCG-IP under visible light with an NADH photoregeneration of 39%.

Ji et al. [53] developed a polyelectrolyte-doped hollow nanofibers/microcapsules-based integrated photosynthesis system. Basically, the system was designed by assembling the components involved in the electron-transfer chain in a specific order, such that the photoexcited electrons can be transported efficiently in the opposite direction. This system was integrated in a device in the following order: three dehydrogenases and a cofactor (FDH, FalDH,

ADH and NAD(H)) in the innermost layer, electron mediators (M) in the middle layer, a photosensitizer (EY $^-$ Eosin Y) in the outermost layer and an electron donor (TEOA) for the CO $_2$  conversion to methanol under solar energy (depicted in **Fig. 5**). They achieved a methanol production of 60.37  $\mu$ M (hollow mixture of poly(allylamine hydrochloride) (PAH) and polyurethane (PU) nanofiber-based system) and 61.98  $\mu$ M (microcapsules-based system) with 86.04% NADH photo-regeneration.



**Figure 5.** Schematic illustration of preparation of polyelectrolyte-doped hollow nanofibers/microcapsules-based integrated photosynthesis system for the solar-driven CO<sub>2</sub> transformation to methanol. Reproduced with permission from reference [53]. Copyright 2016 John Wiley & Sons.

Note that in different works, the co-immobilization of NADH with multi-enzymatic catalytic systems allowed to attend a higher methanol production efficiency over a single enzyme-based system. Furthermore, addition of photosensitisers, inorganic metal oxides and electron mediators as well as electron donors improved the photoreduction of CO<sub>2</sub> to methanol [53] [36].

Table 2. Methanol production from CO<sub>2</sub> using biomimetic photocatalysts

Photocatalytic system	Reaction conditions	CH <sub>3</sub> OH yield	Reference
Sugarcane bagasse (SB)	CO <sub>2</sub> was purged	110 μmol/g (g-	[44]
supported on graphitic carbon	through water bubbler	SB-HT-E)	
nitride (gCN)	and photocatalyst and	15-20 μmol/g (g-	
(SB-modified gCN)	irradiated for 12 h	SB-HT-W)	
	with 14 W LED		
Chitosan-TiO <sub>2</sub>	CO <sub>2</sub> was purged	0.37  nmol/g	[45]
	through water bubbler		
	and photocatalyst and		
	irradiated for 12 h		
	with UV light		
Cuprous Oxide Nanoparticles	CO <sub>2</sub> was purged	1656.7 µmol/g	[54]
Coated with Aminated Cellulose	through NaOH (120		
(Cu <sub>2</sub> O/NCC-EDA)	mL of $0.5 \text{ mol } L^{-1}$ )		
	and photocatalyst (0.1		
	g) suspension and		
	irradiated for 12 h		
	with 300 W Xenon		
	lamp		
CuO/ ZnO-embedded	Catalyst Films were	0.030 mmol/g	[55]
carbohydrate polymer films	irradiated for 6 h with	0.037 mmol/g	
(CuO/ZnO-Den-TOCNF)	UV light at 5cm		
(CuO/ZnO-CS)	distance from the		
	closed reactor bottle		
	where water was		
	dropped and dry ice		
	(440–880 mg) was		
	held		
Cu-loaded carbon modified	NaOH (0.2 M) and	2600 μΜ	[46]
titanium oxide (Cu-C/TiO <sub>2</sub> ).	catalyst suspension		

Carbon from glucose	was purged with CO <sub>2</sub> for 30 min and irradiated for 5 h with		
	UV lamps		
Tetraphenylporphyrin	Suspension of	6.8 μΜ	[36]
tetrasulfonate with	H <sub>2</sub> TPPS (100 μM),		
methylviologen, triethanolamine	$MV^{2+}$ (2.0 mM),		
and dehydrogenases (H <sub>2</sub> TPPS/	TEOA (0.3 M), FDH		
MV <sup>2+</sup> /TEOA/FDH/AldDH/ADH)	(2.0 μM), AldDH (2.0		
	$\mu M$ ) and ADH (2.0		
	μM) in CO <sub>2</sub> saturated		
	50mM sodium		
	pyrophosphate buffer		
	was irradiated for 100		
	min with 250W		
	halogen lamp		
Dehydrogenases	10 mL buffer solution	60.37 μΜ	[53]
(FDH/FalDH/ADH) encapsulated	(50 mM PBS, pH	(hollow	
in hollow nanofibers and	7.0), 15% (w/v)	nanofibers) and	
microcapsules	TEOA, 25 mg hollow	61.98 μM	
	nanofibers or	(microcapsules)	
	microcapsules, 0.1 mg	86.04% NADH	
	mL <sup>-1</sup> FateDH, 0.1 mg	photoregeneration	
	mL <sup>-1</sup> FaldDH, 0.1 mg		
	$mL^{-1}$ ADH, 0.2 $mM$		
	NADH, 0.25 mM M,		
	and 50 $\mu M$ EY		
	irradiated for 10 h		
	with 450 W Xenon		
	lamp		
Chemically converted graphene	CO <sub>2</sub> (0.5 mL/min), β-	11.21 μΜ	[52]

attached to isatin-porphyrin and NAD<sup>+</sup> (1.24 µmol), 38.99% NADH coupled with dehydrogenases Rh (0.62)µmol), photoregenration (CCG-IP/ FalDH/FDH/ADH) TEOA (1.24 mmol), photocatalyst (0.5)mg), 9 units of each enzyme (FDH, FalDH, and ADH) in 3.1 mL of sodium phosphate buffer (100 mM, pН 7.0)irradiated for 30 min with visible light Siliceous mesostructured cellular CO<sub>2</sub>, MFC-MP, 1.35 mmol/g/min [48] foams (MCF) functionalized with FateDH (0.05)mg mercaptopropyl groups (MP) and ml<sup>-1</sup>), FaldDH (0.75) with  $mg ml^{-1}$ ), ADH (3.75) co-immobilized  $ml^{-1}$ ), NADH dehydrogenases mg (MFC-MP/FDH/ADH/FaldDH) (100)mM), and KHCO<sub>3</sub> (200 mM) in 100 mM phosphate buffer (pH 6.5)

#### 4. Biomimetic photocatalysts for the transformation of CO<sub>2</sub> to formic acid/formate

Formic acid (FA) also known as methanoic acid is a colourless, sour, pungent-smelling, and fuming liquid, with as chemical structure HCOOH. If this weak acid is toxic in case of skin and oral exposure, it is non-toxic and stable at room temperature conditions [56,57]. A variety of wood ants and bees produce large quantities of FA naturally, to protect their nest by spraying at prey [58]. FA is one of the most important chemicals as it has found application in a wide range of industries. It has been used for example as an anti-bacterial reagent, food additive, textile dyeing, leather tanning, rubber, finishing, drilling fluids, anti-icing, fragrance ingredient,

cleaning products, pH modifier, and corrosion inhibitor. It is also used as organic synthesis reagent in various industrial chemical processes [59–61].

Due to the high industrial demand for FA, its production at high scale crucial. Formerly, FA was produced as a by-product of acetic acid production through hydrocarbon oxidation which has been reduced after the implementation of the carbonylation of methanol method for acetic acid production. FA production was then fulfilled via hydrolysis of formamide, this method however was economically unfeasible due to the consumption of sulfuric acid and ammonia as well as ammonium sulfate formation [56]. Finally, the carbonylation of methanol followed by hydrolysis of methyl formate (**Eq. 5 and 6**) has been the main industrial process applied for FA production [62][63].

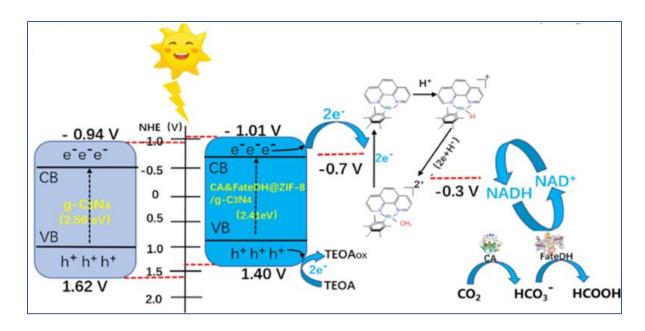
$$CH_3OH + CO \rightarrow HCO_2CH_3$$
 (5)  
 $HCO_2CH_3 + H_2O \rightarrow HCOOH (FA) + CH_3OH$  (6)

However, this conventional production process has encountered predicaments, being the work-up of the hydrolysis mixture the main issue. Also, in the course of separating unreacted methyl formate, re-esterification of FA and methanol to methyl formate happens very quickly due to the unfavourable equilibrium [64,65]. Finally, this conventional method has negative environmental impacts, and high energy and economic costs [66,67]. As a result, alternative ways of producing FA from other sources such as the reductive conversion of CO<sub>2</sub> are extensively researched [68– 70]. Transformation of CO<sub>2</sub> to FA can significantly reduce carbon emissions and assist in fighting climate change whilst finding applications in hydrogen storage, chemical feedstock, and fuel in direct formic acid fuel cells (DFAFCs) [71][72]. Several methods allow the reduction of CO<sub>2</sub> to FA among which photochemical [73][74], electrochemical (ECR) [75][68], hydrogenation [69][76], and biological processes [77][78], stand a good position. The attractivity of the biomimetic photocatalytic transformation of CO<sub>2</sub> for FA production derives the abundance of biomaterials, environmental friendliness, and energy-saving to mention a few advantages. The mostly used biocatalyst is an enzyme, mainly the p-2-hydroxy acid dehydrogenase (FDH) that is found on methylotrophic microorganisms like bacteria and yeast that can use methanol. Tishkov and Popov reported that the FDH from bacterial sources exhibit 1.7 times higher activity than yeast [52]. The FDH enzyme catalyses the oxidation of formate to CO<sub>2</sub> and converts NAD<sup>+</sup> to NADH [79][80]. The cofactor β-nicotinamide adenine dinucleotide in its reduced form (NADH)

is normally needed by FDH enzymes to attain the reduction of CO<sub>2</sub> to HCOOH according to reaction (**Eq. 7**) below [81]. Around twenty studies that deal with the reduction of CO<sub>2</sub> into FA utilizing bio-based photocatalytic systems have been reported between 2012 and 2022 (see Table 3). These studies increased sharply from 2018 and exhibited promising conversion systems and high FA production yields.

$$CO_2 + NADH \leftrightarrow ^{CbFDH}HCOO^- + NAD^+$$
 (7)

Yu et al. reported an effective and environmentally friendly light-driven enzymatic nanosystem [82]. This catalytic system consists of a combination between graphitic carbon nitride (g-C<sub>3</sub>N<sub>4</sub>) photocatalyst and two enzymes (carbon anhydrase (CA) and formate dehydrogenase (FateDH/FDH)), encapsulated in ZIF-8 MOF (CA&FateDH@ZIF-8/g-C<sub>3</sub>N<sub>4</sub>). Fig. 6 illustrates the possible mechanism where photocatalytic NADH regeneration and enzymatic hydrolysis reduced CO<sub>2</sub> as described by the authors. Upon light irradiation, CA&FateDH@ZIF-8/g-C<sub>3</sub>N<sub>4</sub> is excited to generate electron-hole pairs which then receive two electrons from the oxidation of one molecule of TEOA which are then excited from the valence band (VB) to the conduction band (CB) by light. The electron transport complex (M) was used to produce more 1,4-NADH, the biological entity needed by the enzymes. Thus, the highly selective M transports two electrons and one photon to NAD<sup>+</sup> for NADH regeneration. With the two enzymes, CA quickly converted CO<sub>2</sub> into HCO<sub>3</sub> which is converted into HCOOH using FateDH which, at the same time, consumes NADH as shown in Fig. 6. Reduction of CO<sub>2</sub> with CA&FateDH@ZIF-8/g-C<sub>3</sub>N<sub>4</sub> system resulted in 198 µM of FA whilst the system without CA and free FateDH produced only 75 and 38 µM, respectively. When 2 mg.mL<sup>-1</sup> of CA&FateDH@ZIF-8/g-C<sub>3</sub>N<sub>4</sub> was utilized, the optimized yield could reach 243 µM of FA. The activity of the enzymatic activity was retained above 80% after 10 runs, thus indicating the reusability of this catalytic system, that was also reported as able to shorten the distance between photocatalyst and enzyme entities, that is beneficial to the diffusion of substances and electrons.

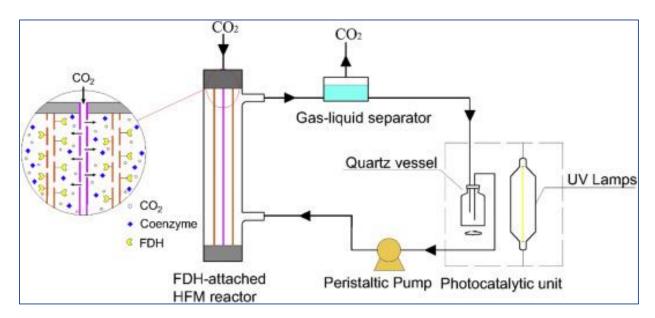


**Figure 6.** Illustration of electron and energy transfer processes for light-driven enzymatic CA&FateDH@ZIF-8/g-C<sub>3</sub>N<sub>4</sub> nanosystem. Reproduced with permission from Reference [82]. Copyright 2021 Elsevier.

Wang et al. [83] reported a CO<sub>2</sub> reduction system consisting in two enzymes as well i.e. CA and FDH. Using a catalyst that combines two enzymes has been found to regulate the reaction rate and increase the loading amount of FDH which allows to decrease the ratio of of CA/FDH. This is one of the routes enhancing FA production. Sustaining a stable pH environment for this system in the course of CO<sub>2</sub> hydration was mandatory as the process releases hydrogen ions. The increase in the hydration rate of CO<sub>2</sub> increased the yield of FA by aspect of 4.2-fold. However, the costs of cofactor and enzymes are economically unfeasible and enzyme immobilization and cofactor regeneration appeared as a probable solution for low costs, recyclability, and enhanced biocatalyst stability.

A FDH-bearing hollow fiber membrane (HFM) reactor (**Fig. 7**) has been developed by Gu et al. [70] for CO<sub>2</sub> reduction which led to high-efficient and sustainable FA production. They used a UV-TiO<sub>2</sub> photocatalytic coenzyme regeneration system with a FDH-bearing HFM reactor which exhibited a maximum initial reaction rate of 1.04 mM.h<sup>-1</sup> and 81.7% yield. The optimization of operating conditions and NADH concentration provided a turnover number of 125 after 4.5 h. Considering that one mole of NADH is consumed to generate one mole of FA, NADH regeneration for continuous enzymatic production is important. Thus, the system regenerated

NADH when the reaction mixture was transported into a photocatalytic cell. This resulted in an instant regeneration of NADH which eventually showed that even at a low loading amount of NADH high FA production could be achieved.

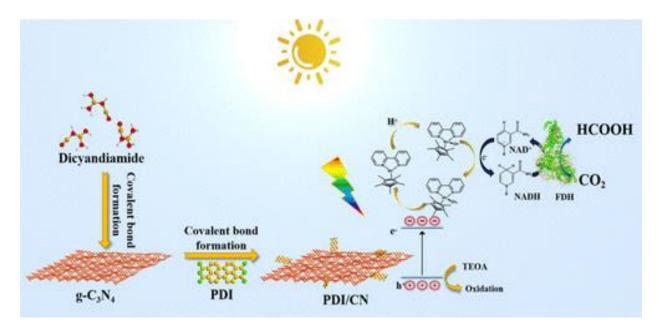


**Figure 7.** Illustration of a photocatalytic/enzymatic HFM system. Reproduced with permission from Reference [70]. Elsevier 2020.

Zhang et al. built an artificial thylakoid consisting of protamine-titania (PTi) micro-capsules with cadmium sulfide quantum dots (CdS QDs) and enzymes (FDH extracted from *Candida boidinii* (CbFDH)) for FA production from CO<sub>2</sub> [84]. The improvement of NADH regeneration activity (4226±121 μmolg<sup>-1</sup>h<sup>-1</sup>) and enzyme compatibility of CdS/PTi microcapsules contributed to high FA production. Compartmentalization of photogenerated holes and reactive oxygen species (ROS) by the structure of CdS/PTi microcapsules protected FDH from deactivation, resulting in high compatibility between bio- and photo-catalyst. This catalytic system exhibited a quantum yield of 0.66±0.13% of FA over multiple light-dark cycles.

Zhang et al. reported a highly selective reduction system with a metal-free heterojunction of pyromellitic diimide/g-C<sub>3</sub>N<sub>4</sub> (PDI/CN) coupled with FDH enzyme [85]. **Fig. 8** shows the possible mechanism of the photoenzymatic catalytic cascade system on the PDI/CN heterojunction. In this cyclic process, the photocatalyst could continuously reduce NAD<sup>+</sup> to NADH while FDH could oxidize NADH to NAD<sup>+</sup>, concurrently, in order to accelerate CO<sub>2</sub> transformation to FA. The process can successfully advance continual FA production. Another

study reported the utilization of [Cp\*Rh(bpy)H<sub>2</sub>O]<sup>2+</sup> as a cocatalyst since the biologically inactive 1.6-NADH and the dimers of NAD<sup>+</sup> may be formed under the action of the intermediate radical. Thus, it was proven that [Cp\*Rh(bpy)H<sub>2</sub>O]<sup>2+</sup> was effective for the selective production of biologically active 1,4-NADH [70]. Zhang et al. [85] have also shown herein the promoting effect of [Cp\*Rh(bpy)H<sub>2</sub>O]<sup>2+</sup> in their system. The NADH yield increases linearly with the increase in sunlight irradiation time, and the system with [Cp\*Rh(bpy)H<sub>2</sub>O]<sup>2+</sup> showed better performance signifying that this cocatalyst assists NAD<sup>+</sup> to accept photogenerated electrons thereby accelerating electron transfer to generate NADH. The outstanding performance of this photocatalytic system was ascribed to good conductivity, heterointerface effect, broad sunlight response range of the catalyst as well as a distinctive photoenzymatic catalytic cascade system. The system achieved 75% NADH yield and 114.8 µmol for FA production.



**Figure 8.** Possible mechanism of the photoenzymatic catalytic cascade system on the PDI/CN heterojunction. Reproduced with permission from Reference [85]. ACS Publications 2021.

Several studies concern the use of carbon-based materials such as carbon nanodots-silica hybrid, graphene film, graphene oxide modified with cobalt metallated aminoporphyrin, and  $C_{60}$  polymer film as photocatalysts [86][33][87][34]. The interest in carbon materials emanates from their wide availability, environmental friendliness, and good light harvesting properties. Carbon nanodots were reported to have excellent biocompatibility, high solubility, and low toxicity whilst GO properties include biological function, thermal, mechanical, and biomedical [88].

Carbon nanodots-silica hybrid/FDH and graphene film/FDH systems showed NADH regeneration and FA yield of 74.10±0.17%, 203.33±1.9 μmol [86], and 71.8%, 228.6 μmol [33], respectively. For the FDH@GO-Co-ATPP system, NADH regeneration and FA yield of 48.53% and 96.49 μmol, respectively were reported [87] whilst the C<sub>60</sub>-based polymeric/FDH system exhibited 91.07% of NADH regeneration and 239.46 μmol [34].

Ji et al. reported chloroplast-mimicking TCPP/SiO<sub>2</sub>/Rh HNP-based wholly integrated APS and AM/M/BP HNS-based integrated Z-scheme APS models in 2018 [89] and 2020 [90]. These systems decreased the transfer distance of the energy and electron among the enzymatic redox and light-harvest system as well as enabling sufficient charge separation and photogenerated excitons transportation. In both studies, the effect of the TCPP/SiO<sub>2</sub>/Rh HNPs and AM/M/BP HNS-based Z-scheme APS was confirmed by detecting photochemical, optical, and electrochemical properties. The TCPP/SiO<sub>2</sub>/Rh HNPs system showed NADH regeneration which enhanced from 11 – 75% and FA production which increased from 10 – 100 μmol. AM/M/BP HNS-based Z-scheme APS, on the other hand, exhibited significant enhancement of NADH regeneration and FA production which reached 89% and 365 μmol, respectively [89][90].

The use of graphitic carbon nitride has been on the rise as well. This nitrogen-containing material possesses exceptional solar light-harvesting photocatalytic ability due to its slow recombination ability which increases photo-responsive energy transfer from the dye to a g-C<sub>3</sub>N<sub>4</sub> moiety [91]. It has high stability, a tuneable band structure, strong visible light response, and astounding biocompatibility [92]. The systems reported by Cheng et al. [93], Zhang et al. [92], Singh et al. [91], and Kumar et al. [94] polymeric carbon nitride (PCN) mediated by a tannic acid/polyethyleneamine (TA/PEI), Rh complex NH<sub>2</sub>-functionalized polymeric carbon nitride, 1-4,-diaminoanthaquinone monomer using a modified CN, and Se-doped CN nanosheets, respectively, showed FA production of 2.1 mM, 5.0 mM, 344.12 μmol, and 200.26 μmol.

Tian et al. [51] assembled a functionally compartmental inorganic photocatalyst-enzyme system that imitated chloroplast. Photoexcitation and enzymatic  $CO_2$  reduction were compartmentalized by the thylakoid membrane with the purpose of protecting enzyme from photodamage whilst the system enables electron transfer thereby fostering photocatalyst. They built a tightly integrated artificial photosystem comprising of Rh complex cocatalyst onto thiophene-modified  $C_3N_4$  (TPE- $C_3N_4$ ) and FDH encapsulated with MAF-7 which is a subclass of zeolitic imidazolate-

based MOF. The improved NADH regeneration of the system was attributed to the tightly integrated structure which promoted fast electron transfer from TPE- $C_3N_4$  to the Rh complex. MAF-7 not only protected FDH from photoinduced deactivation but also improved enzymatic stability in photosynthesis conditions. This photocatalytic system resulted in 16.75 mM FA after 9 h of illumination. **Table 3** shows the summary of the studies done on the transformation of  $CO_2$  to FA using bio-inspired photocatalysts and the progress is promising.

**Table 3.** Biomimetic photocatalysts transformation of FA from CO<sub>2</sub>.

Catalyst	Reaction conditions	Yield	Reference
Bromophenol-Bakelite (BPB)/Rh	NAD <sup>+</sup> – 1.24 μM, BPB –	NADH	[95]
complex/FHD	$0.031~\mu M,~AsA~-~1.24$	regeneration –	
	μM, Rh complex - 0.62	82.52%	
	μM, sodium phosphate		
	buffer – 3.1 mL	FA yield – 220	
		μmol	
Bamboo-shape magnesium	Photocatalyst $-15 \mu L$ in	NADH	[50]
tetraphenyl porphyrin-based	3.1 mL of sodium	regeneration –	
framework (Mg-TPPBF)/FDH	phosphate buffer (100	73.9%	
	mM, pH 7.0), $\beta$ -NAD <sup>+</sup> –		
	124 μL, rhodium complex	FA yield – 197.2	
	$-62~\mu L,~FDH-3~units,$	µmol in 2 h	
	$AsA-310\;\mu L$		
1,4-diaminoanthraquinone (AQ)	NAD <sup>+</sup> /NADP <sup>+</sup> – 1.24	NADH	[91]
monomer using a modified carbon	μmol, CpM – 0.62 μmol,	regeneration –	
nitride (CN) (AQBCN)/FDH	AsA - 0.1  mmol, FDH - 3	97.21/95%	
	units and the AQBCN		
	photocatalyst - 0.5 mg in	FA - 344.12	
	3.1 mL of SPB (100 mM;	μmol	
	pH, ~7.0)		
Rh complex covalently grafted onto	Photocatalyst – 12 mg,	NADH	[92]
NH <sub>2</sub> -functionalized polymeric	NAD – 120 μL, PBS –	regeneration –	

```
carbon nitride (NH<sub>2</sub>-PCN) (Rh<sub>m3</sub>- 2.88 mL, FDH – 1 unit
                                                                62.4%
N-PCN)/FDH
                                                                FA yield - 5.0
                                                                mM
CA&FateDH@ZIF-8/g-C<sub>3</sub>N<sub>4</sub>
                                    CA&FateDH@ZIF-8/g-
                                                                FA yield – 243 [82]
                                    C_3N_4 - 2 \text{ mg.mL}^{-1} in a \mu M
                                    0.1 M PBS, TEOA - 10
                                    w/v\%, mediator – 1
                                    mg.mL^{-1}, β-NAD<sup>+</sup> – 0.8 m
Pyromellitic
                   diimide/g-C<sub>3</sub>N<sub>4</sub> FDH – 3 units, catalyst – NADH
                                                                                   [85]
(PDI/CN)/FDH
                                    10 mg, TEOA – 15 wt.%, regeneration
                                    phosphate buffer solution 75%
                                    (pH 7.4, 100 mM) - 10
                                    mL, \beta-NAD<sup>+</sup> - 0.1 mM, FA yield - 114.8
                                    [Cp*Rh(bpy)-(H_2O)]^{2+} – µmol
                                    0.25 mM
Polymeric carbon nitride (PCN) Photocatalyst
                                                                                   [93]
                                                           0.5 NADH
                                   mgmL^{-1}, FDH – 2 regeneration
mediated
              by
                      a
                            tannic
                         (TA/PEI) mgmL<sup>-1</sup>, NAD<sup>+</sup> – 1 mM, 37.8%
acid/polyethyleneimine
(PCN@TA/PEI-Rh
                                    PBS buffer (pH 7.0, 100
                                    mM), free Rh -0.5 FA yield -2.1
core@shell)/FDH
                                    mgmL^{-1}, and CO_2 - 0.3 mM
                                    MPa.
Covalent-organic framework/2,4,6- Photocatalyst – 0.5 mg, β- NADH
                                                                                   [96]
                                    NAD<sup>+</sup> – 1.24 mmol, AsA regeneration
triformylresorcinol (COF-4)/FDH
                                    - 0.1 mmol, Rh complex 94.6%
                                    -0.62 mmol, and 3 units
                                    of FDH in 3.1 mL of 100 FA yield - 226.3
                                    mM buffer (pH 7.0). CO<sub>2</sub> μmol
                                    flow rate -0.5 \text{ mLmin}^{-1}
                                    for 1 h in the absence of
```

```
light
                                     NADH – 2mM, liquid FA yield
FDH@HFM/UV/TiO<sub>2</sub>
                                                                                - [70]
                                     velocity – 40 mL.min<sup>-1</sup>, 81.7%
                                     gas velocity – 3 mL.min<sup>-1</sup>
Antimonene/Cp*Rh(phen)Cl/black
                                     TEOA – 15 w/v%, AM NADH
                                                                                    [90]
                                    NSs or BP NSs - 0.2 regeneration
phosphorus hybrid nanosheet-based
Z-scheme (AM/M/BP HNS)/FDH
                                     mgmL^{-1}, M - 0.1 mM, 35±4%
                                     FDH – 0.3 units, NAD<sup>+</sup> –
                                     0.1 \, \mathrm{mM}
                                                                 FA
                                                                        yield
                                                                 160±24 µmol
FDH@MAF-7/TPE-C<sub>3</sub>N<sub>4</sub>/PEI/Rh
                                     NAD^{+} g - 1 mM, FA yield - 16.75 [51]
                                     phosphate buffer – 100 mM after 9 h
                                     mM (pH 7), TEOA – 15
                                     w/v%, MAF@FDH - 7.5 5th cycle FA -
                                     mg, TPE-C<sub>3</sub>N<sub>4</sub>/PEI/Rh -3 ~76%
                                     mg
FDH@Rh-NU-1006
                                     FDH@Rh-NU-1006
                                                             - FA yield - 0.144 [97]
                                     1.8 \times 10^{-3} M in Tris buffer \pm 0.003 in 24 h
                                     (2 \text{ mL}), TEOA -0.5 \text{ mM},
                                     NAD^+ - 1 mM
C<sub>60</sub>-based
             polymeric
                            system CO<sub>2</sub> flow rate - 0.5 NADH
                                                                                    [34]
                                     mLmin<sup>-1</sup>, \beta-NAD<sup>+</sup> – 1.24 regeneration
coupled with FDH enzyme
                                     μmol, TEOA – 1.24 91.07%
                                     mmol, 3 units of FDA in
                                     3.1
                                           mL
                                                  of
                                                        sodium FA
                                                                        vield
                                     phosphate buffer (pH 7.0, 239.46 µmol
                                     100 mM)
                                     [CO<sub>2</sub>] – 0.3 MPa, 37 °C, NADH
CdS/TPi microcapsule-CbFDH
                                                                                    [84]
                                     visible light illumination regeneration
                                     (\lambda = 405\pm 5 \text{ nm})
                                                                 93.03±3.84%
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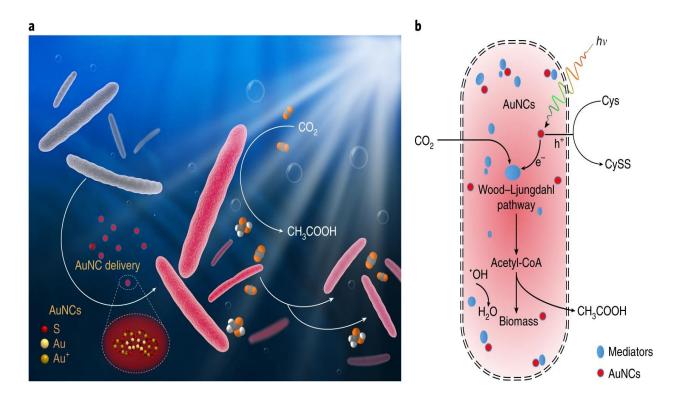
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NAD<sup>+</sup> - 5 nm, TEOA -
                                     400 nm, CdS/PTi – 1 gL<sup>-1</sup>, FA yield – 1500
                                     PBS buffer – pH 7.0, 100 μMh<sup>-1</sup>
                                     nm, CbFDH - 1 gL^{-1}
FDH@GO-Co-ATPP
                                     GO-Co-ATTP - 0.7 mg, NADH
                                                                                    [87]
                                     NAD<sup>+</sup> – 1.24 mol, TEOA regeneration
                                     - 1.24 mmol, 3 units of 48.53%
                                     FDH in 3 ml of (100 mM,
                                     pH 7) sodium phosphate FA yield – 96.49
                                     buffer
                                                                 μmol
Porphyrin/SiO<sub>2</sub>/Cp*Rh(bpy)Cl
                                     Phosphate buffer solution NADH
                                                                                    [89]
hybrid
                      nanoparticles
                                    (100 \text{ mM}, \text{ pH } 7.0) - 10 \text{ regeneration}
                                     mL, NAD^+ - 1 mM, 75\%
(TCPP/SiO<sub>2</sub>/Rh-HNPs)/FDH
                                     TCPP - 0.5 \text{ mM}, FDH -
                                     0.3 units mL<sup>-1</sup>, CO_2 - 0.3 FA yield -100
                                     MPa
                                                                 µmol after 4 h.
Se-doped carbon nitride (C<sub>3</sub>N<sub>4</sub>) Photocatalyst – 0.5 mg,
                                                                                    [94]
                                                                NADH
nanosheets/FDH
                                     Rh – 0.62 \mumol, \beta-NAD<sup>+</sup> regeneration
                                     - 1.24 μmol, FDH - 3 82.84%
                                     units, AsA - 0.1 mmol in
                                                        sodium FA
                                     3.1
                                            mL
                                                  of
                                                                        yield
                                     phosphate buffer (100 200.26 µmol in
                                     mM, pH 7.0)
                                                                 1 h
                                     CO_2 flow rate -0.5
                                     mLmin<sup>-1</sup>
Graphene
             film
                      photocatalyst β–NAD<sup>+</sup> – 1.24 μmol, Rh NADH
                                                                                    [33]
(GFPC)/FDH
                                     complex – 0.62 µmol, regeneration
                                     FDH - 3 units in 3.1 mL 71.8%
                                     of
                                           sodium
                                                     phosphate
                                     buffer (100 mM, pH 7.0), FA yield – 228.6
                                     AsA - 0.1 mmol, CO<sub>2</sub> µmol in 2 h
```

flow rate  $-0.5 \text{ mLmin}^{-1}$ Carbon Photocatalyst – 0.5 mg,  $\beta$ - NADH [86] nanodots-silica hybrid (CNDSH)/FDH - 1.24 mmol, regeneration rhodium complex – 0.62 74.10±0.17% mmol, ascorbic acid -0.1mmol, FDH – 3 units in FA vield 3.1 mL of Na<sub>2</sub>HPO<sub>4</sub>-203.33±1.9 NaH<sub>2</sub>PO<sub>4</sub> buffer (100)umol mM, pH 7.0) Cobaloxime/eosin/REOA/FDH Eosin Y - 0.10 mM, FA yield - 36% [98]  $NAD^+ - 0.50$  mM, TEOA - 0.20 M, 5 units of FDH in 0.15 M phosphate buffer (pH 7.0)

#### 5. Biomimetic photocatalyts for the transformation of CO2 to acetic acid

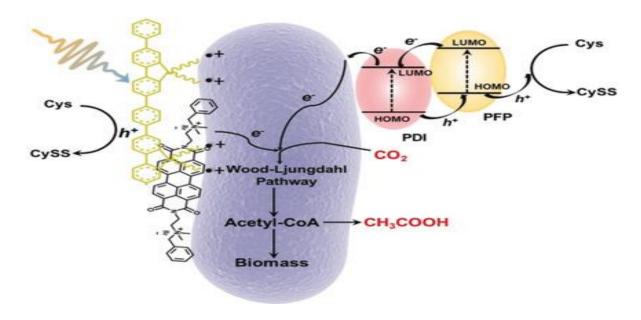
Acetic acid otherwise known as vinegar acid, ethanoic acid and methane carboxylic acid is a clear, colourless liquid, of pungent smell and with the chemical formula CH<sub>3</sub>COOH. It is the second simplest carboxylic acid after formic acid and is of high importance also due to a plethora of industrial and pharmaceutical applications. It is produced synthetically and it can also be prepared via bacterial fermentation. It is a crucial chemical reagent being primarily used for the production of cellulose acetate for photographic films and that of acetic anhydride. Acetic acid also possesses antiseptic properties, so it can be employed as antimicrobial agent. The current approach for the production of acetic acid is not environmental benign, with high energy cost and the production scale cannot meet the demand for both industrial and pharmaceutical needs [99,100]. Development of alternative production routes that will cover the current demand and be environmentally friendly is thus needed. One of the promising and cost-effective methods to produce acetic acid is the photocatalytic transformation of CO<sub>2</sub>. Recent advancements in this domain are described below but note that only a few of case studies were found in the literature of the last decade.

Zhang and co-authors [12] integrated ultra-small gold nanoclusters (AuNCs) and cadmium sulfide (CdS) into M. thermoacetica bacteria and compared their performance in the photosynthesis of acetic acid from CO<sub>2</sub> (Fig. 9a). These bio-based photocatalytic systems were found efficient for CO<sub>2</sub> fixation into acetic acid under low light irradiation and dark (Fig. 9b). The authors observed that the yield in acetic acid by the gold-based biophotocatalyst after four days is conspicuously higher than that obtained with cadmium sulfide-based bio-photocatalyst under the irradiation of low light. Moreover, the authors found that the activity of ordinary bacterium (M. thermoacetica) and gold-based bio-photocatalyst in darkness are relatively low. A monitoring of the electron transfer mechanism via photoluminescence confirmed that the photoexcited electrons from the gold nanoclusters are migrated to distributed redox mediators in the bacteria cytoplasm such as flavoproteins. Evaluation of the effect of cysteine on the yield of acetic acid production revealed that the gold-based bio-photocatalyst produced the highest yield (6.01 mmolg<sup>-1</sup>) at 0.3 % wt Cys. It is worth to mention that the ligands at the gold nanoclusters surface and the gold core size can be changed in order to tune their biochemical and biophysical characteristics, including cell uptake, cytotoxicity, biocompatibility, and molecular electronic structures. This study demonstrated that integration of gold nanoclusters as intercellular photosensitizer inside the cytoplasm of M. thermoacetica bacterium allowed this nonphotosynthetic latter to conduct effective photosynthesis of acetic acid from CO<sub>2</sub>.



**Figure 9** (a) Schematic illustration of *M. thermoacetica*/AuNCs hybrid system (b) Schematic bacteria electron transfer pathway for acetic acid generation under illumination of light

In another study, Gai et al. [101] developed an organic semiconductor – bacteria biohybrid photosynthetic system that efficiently conversed  $CO_2$  into acetic acid. The bacterium involved is Moorella thermoacetica whose surface was coated with perylene diimide derivative (PDI) and poly(fluorene-co-phenylene) (PFP) as photosensitizers. This led to the formation of a p-n heterojunction (PFP/PDI) layer with high hole/electron separation efficiency (**Fig. 10**). The  $\pi$ -conjugated semiconductors display excellent light-harvesting ability and biocompatibility. In addition, the cationic side chains of the two organic semiconductors could intercalate into cell membranes, thus allowing an efficient transfer of electrons to the bacterium. In this photocatalytic system, the bacterium could harvest photoexcited electrons from the PFP/PDI heterojunction and synthesize acetic acid from  $CO_2$  under illumination. This bio-photocatalytic system showed an efficiency rate of 1.6% which is comparable to those of reported inorganic biohybrid systems.



**Figure 10.** Wood–Ljungdahl pathway for CO<sub>2</sub> reduction in the presence of PFP/PDI/M. thermoacetica. Reproduced with the permission from [101]. Copyright 2020 Willey online

Tharak and colleagues [102] reported microbial-mediated CO<sub>2</sub> in gas and gas-electro fermentation systems with and without applied potential by selectively enriched biocatalysts (Fig. 11). In the Heat + CO<sub>2</sub> condition during the gas fermentation (GF) stage, the maximum production was 0.84 g/L acetic acid with 31 mg.L<sup>-1</sup>.h<sup>-1</sup> inorganic carbon fixation rate (ICFR). The GF was tested in a single-chambered microbial electrochemical system (MES; Gas-electro fermentation) at different applied potentials (-0.4 and -0.6 V vs Ag/AgCl [3.5 M KCl]). With -0.6 V applied potential, a higher acetic acid production of 1.02 g/L was obtained. The CO<sub>2</sub> fermentation efficiencies were improved by higher charge transfer rates (52% faradaic efficiency) and lower corrosion and resistances (Rct -0.4 ohm). On continuous conditions, an abundance of Proteobacteria, Firmicutes, and Actinobacteria in the enriched biocatalyst represented a significant shift of the microbiome towards CO<sub>2</sub> feeding microbes.

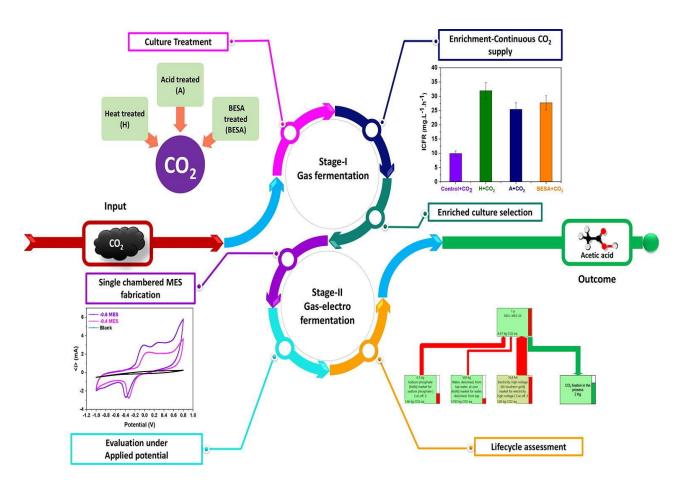


Fig. 11. Schematic diagram of acetic acid production from CO<sub>2</sub> via lignin/ionic based catalytic system. Reproduced with the permission from [102]. Copyright 2023 Elsevier

Wang and co-workers [103] reported a new strategy to produce renewably sourced acetic acid from lignin and CO<sub>2</sub>/H<sub>2</sub>. The catalytic system is based on an ionic liquid (that is 1- butyl-3-methylimidazolium chloride, [BMIm][Cl]) that contains a Ru-Rh bimetal catalyst (Ru<sub>3</sub>(CO)<sub>12</sub> and RhI<sub>3</sub>) and Li salt additives (LiI and LiBF<sub>4</sub>). This system was effective for the production of acetic acid in the temperature range of 120-180 °C under low pressure of CO<sub>2</sub> (1 MPa), affording an acetic acid yield of 94% at 180 °C. In addition, taking lignin as the feedstock, 0.17 g of acetic acid was produced from 1 g of lignin. Finally, the catalytic system could be recycled 5 times without decrease in performance. Investigation of the reaction mechanism evidenced that lignin provided methyl group to form CH<sub>3</sub>I, while the [BMIm]<sup>+</sup> -stabilized RhI<sub>3</sub> and I <sup>-</sup>-complexed Ru(CO)<sub>n(n=1~4)</sub> cooperated and catalyzed the formation of acetic acid from CH<sub>3</sub>I and CO<sub>2</sub>/H<sub>2</sub>.

**Table 4** below summarizes the biomimetic materials explored for the production of acetic acid. While there is appreciable development towards the production of acetic acid from CO<sub>2</sub> using

biomimetic materials, the yields reported are very low and cannot satisfy the commercial purpose. This calls for more research efforts in this hot topic.

Table 4. Biomimetic photocatalysts for acetic acid production from CO<sub>2</sub>

Photocatalystic system	Reaction conditions	Yield	Reference
M.	Low-intensity	6.01 mmolg <sup>-1</sup> after six	[12]
thermoacetica/AuNCs	simulated sunlight	days in the presence of	
hybrid system	(calibrated by	cysteine as sacrificial	
	photodiode, air mass	agent	
	1.5 global spectrum		
	$(AM 1.5), 2Wm^{-2})$		
M.	Air mass 1.5 global	85±12% quantum yield	[104]
thermoacetica/CdSNPs	spectrum, 2 W m <sup>-2</sup> ,	after four days	
hybrid	photons; 5 x 10 <sup>13</sup> cm <sup>-2</sup>		
	s <sup>-1</sup> under light and		
	dark flux at $\lambda = 435$ to		
	485 nm		
PFP/PDI/M.	5.0 Mw/cm <sup>2</sup> power	0.1 mM in dark, 0.6	[101]
thermoacetica p-n	density, under of light-	mM under light	
heterojunction	dark cycle for 12h		
Different bacteria	Heat-shock, Acid-	1.02 g/L at 52% FE	[102]
mediated	shock and 2-		
biocomposites	Bromoethano		
	sulphonate (BESA),		
	31 mg L <sup>-1</sup> h <sup>-1</sup> of		
	ICFR, (-0.4 and -0.6 V		
	vs Ag/AgCl [3.5 M		
	KCl]), $R_{ct} = 0.4$ ohms		
Lignin/ionic based	180 °C, 1 MPa	94% yield, 0.17g acetic	[103]
catalytic system		acid produced from 1g	
		of lignin	

PFP: poly(fluorene-co-phenylene), PDI: perylene diimide derivative, FE: Faradaic efficiency, ICFR: Inorganic carbon fixation rate

#### 6. Mechanistic insights of biomimetic photocatalysts for CO<sub>2</sub> Reduction

There is no definite mechanism for the reduction of CO<sub>2</sub> as it is a complex reaction involving radicals, electrons, and protons [105]. Generally, after the reduction of CO<sub>2</sub> by a one-electron transfer and the formation of surface-bound CO<sub>2</sub>, the process proceeds through a sequence of elementary steps involving the transfer of a proton, electron or radical (H·), as well as breaking C-O bonds and creating C-H bonds. Several intermediates are radical species, partially accounting for the number of possible pathways and final products at different stages of recombination. The exact sequence and mechanistic flow of each subsequent step have not been thoroughly elucidated, but some literature suggests that photocatalytic reduction of CO<sub>2</sub> *via* biocatalysis has a similar pathway than that described for transition metal photocatalysis [105].

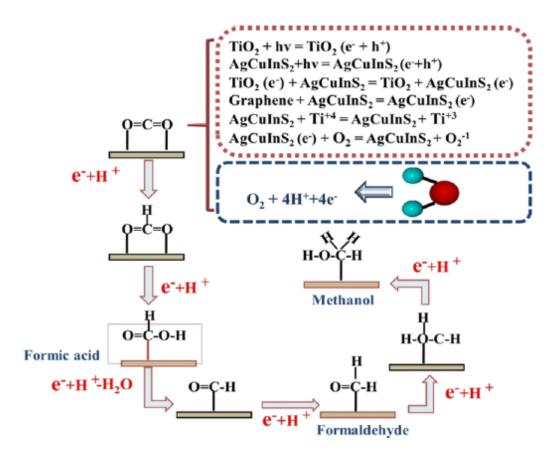
Hydrogen radicals (H·) bind to the oxygen atom of the adsorbed carboxyl radicals (CO<sub>2</sub> <sup>-</sup>), resulting in an immediate cleavage of the C-O bond, subsequently leading to the formation of carbon monoxide. In a separate mechanism, the carboxyl radicals recombine with hydrogen radical to form formic acid. In a next step, formic acid accepts another hydrogen radical to form a dihydroxymethyl radical, which is dehydrated when another hydrogen radical is attached to the formaldehyde. Two additional steps of reduction lead to CH<sub>3</sub>OH formation, which is further reduced to methane in two more steps of reduction. A proposed mechanistic pathway (by formaldehyde, carbene, glycol pathways) for the reduction of CO<sub>2</sub> to CO, formic acid, and methanol as described in the literature is presented in **Fig. 12** [106].

#### Formaldehyde pathway

# Carbene pathway

**Figure 12.** Two reduction routes for CO<sub>2</sub>; top, formaldehyde pathway and bottom carbene pathway [106].

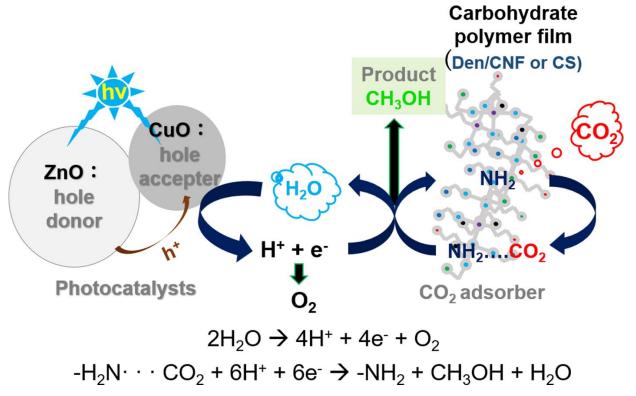
The mechanism of CO<sub>2</sub> conversion involves two-electron reduction mediated by metal-based catalysts or enzymes. Most photocatalysts are metal-based [107][108][109][110][111]. For a commonly described metal photocatalyst, the semiconductor part absorbs photon energy, which leads to the excitation of electrons from valence band (VB) to conduction band (CB), creating electron-hole pair. The holes in VB have strong oxidizing ability, especially for water oxidation and electrons in CB reduce CO<sub>2</sub>. Water oxidation produces hydrogen ions and hydroxyl radicals, and the activated CO<sub>2</sub> reacts with excited electrons and hydrogen ions, which finally result in the formation of formic acid, formaldehyde, and methanol [112]. When AgCuInS<sub>2</sub> supported on graphene-TiO<sub>2</sub> are parallelly activated, electrons transfer through the graphene to reach the VB of TiO<sub>2</sub> to form the donor level, thus, resulting in the formation of photogenerated electrons (e<sup>-</sup>) and holes (h<sup>+</sup>) at the active sites of AgCuInS<sub>2</sub> photocatalyst. The holes (h<sup>+</sup>) react with the adsorbed 2H<sub>2</sub>O to produce oxygen (O<sub>2</sub>), electron (e<sup>-</sup>) and protons (H<sup>+</sup>). Surface-adsorbed CO<sub>2</sub>\*radical generated by the injection of a single electron reacts with e<sup>-</sup> and H<sup>+</sup>, producing CO, which undergoes a further reduction process forming the surface-adsorbed C. CO<sub>2</sub> further reacts with excited electrons (2e<sup>-</sup>) and hydrogen ions (2H<sup>+</sup>) to yield HCOOH. The further reaction of HCOOH with excited electrons (2e<sup>-</sup>) and hydrogen ions (2H<sup>+</sup>) led to a loss of water (H<sub>2</sub>O) to produce an intermediate, formaldehyde, (HCOH) (typical of formaldehyde pathway, see Fig 11). Surface adsorbed formaldehyde reacts with 2e<sup>-</sup> and 2H<sup>+</sup> to yield methanol as a main product. A schematic representation of photocatalytic CO<sub>2</sub> reduction process with AgCuInS<sub>2</sub> is illustrated in Fig. 13.



**Figure 13.** Schematic diagram of the photocatalytic CO<sub>2</sub> reduction process. Re-used with permission from. Copyright (2021) American Chemical Society. [112]

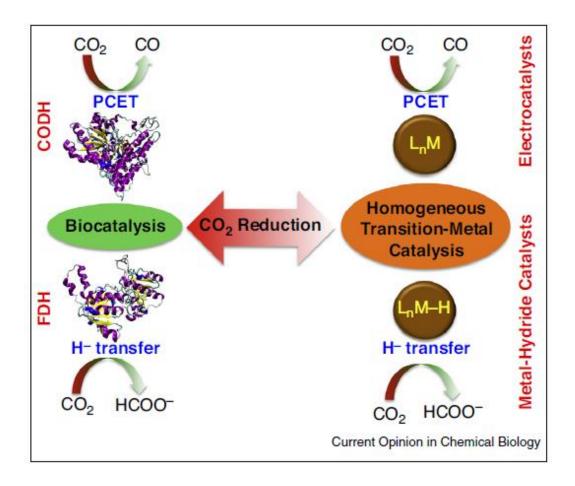
Getahun et al. [55] concerned the use of metal oxides (ZnO and CuO) embedded with bio-based materials (such as chitosan) as photocatalysts. Both ZnO and CuO absorb UV light, making energy separation to occur, and resulting in hydrogen ion (H<sup>+</sup>) and electron (e<sup>-</sup>) generation. The schematic representation of photocatalytic  $CO_2$  reduction using ZnO and CuO under UV light is illustrated in **Fig. 14**. The valence-band energy level of CuO (n-type) is higher than that of ZnO (p-type), the holes of ZnO generated by absorbing light energy transfers to the valence-band level of CuO, leading to the holes (h<sup>+</sup>) on CuO/ZnO composite oxidizing water (H<sub>2</sub>O) to oxygen (O<sub>2</sub>), hydrogen ion (H<sup>+</sup>) and an electron (e<sup>-</sup>).  $CO_2$  (trapped within amine groups of chitosan and dendrimer, -NH<sub>2</sub>--- $CO_2$ ) reacts with e<sup>-</sup> to form anion radical  $CO_2$ <sup>-</sup> (a single electron reduction of  $CO_2$ ) followed by a reaction with protons H<sup>+</sup>, a process known as "proton-assisted multielectron reduction". Surface-adsorbed  $CO_2$ <sup>-</sup> radical reacted with 6e<sup>-</sup> and 6H<sup>+</sup> ( $CO_2$  + 6H<sup>+</sup> + 6e<sup>-</sup>  $\rightarrow$   $CH_3OH + H_2O$ ), producing methanol. Therefore, the photoredox pathway could be considered a

proton-assisted multielectron reduction via the formaldehyde route. Pathway for CO and formic acid production are similar to those reported by Otgonbayar et al [110].



**Figure 14.** Illustration of CO<sub>2</sub> photocatalytic reduction to methanol with H<sub>2</sub>O oxidation on films of CuO/ZnO catalytic-embedded carbohydrate polymer. Reproduced with permission from [55]. Copy rights (2022) Elsevier.

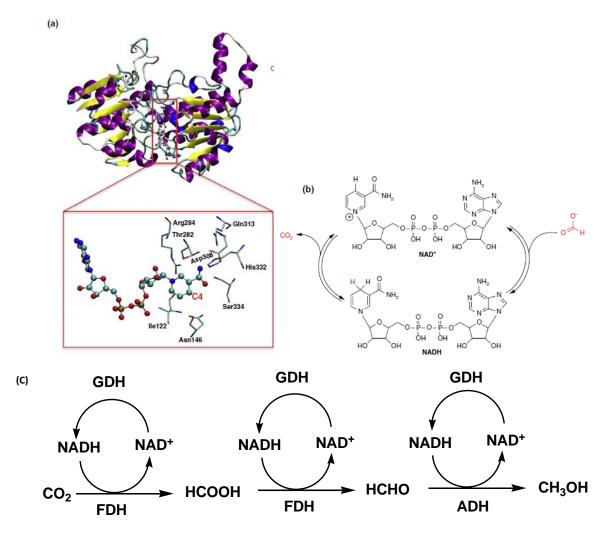
The photocatalytic reaction mechanism of metal-based systems are similar to those followed by enzymes such as carbon monoxide dehydrogenases (CODHs) [113] and formate dehydrogenases (FDHs) [114][115], in which two to four electron reduction of  $CO_2$  is required for conversion to carbon monoxide, formic acid/formate, and methanol. The bio-inspired mechanistic insight for two-electron reduction of  $CO_2$  to CO, as described by Mondal et al, 2014 [105] goes through proton-coupled electron transfer (PCET,  $H^+ + e^-$ ), from a molecule that acts as "electron and proton" donor (reductant) to another one that acts as an acceptor (oxidant). Both biocatalysis and homogeneous transition-metal catalysis usually proceed *via* redox reaction where hydride transfer takes place for formate ion production (**Fig. 15**).



**Figure 15.** Overview of the bio-inspired mechanistic insight into the two-electron reduction of CO<sub>2</sub> to CO and FA (PCET = proton-coupled electron transfer). Copyright [105].

FDHs are known to catalyze the reduction of  $CO_2$  to formic acid through a proton (H<sup>+</sup>) transfer mechanism ( $CO_2 + NADH + H^+ \leftrightarrow HCOOH + NAD^+$ ) as well as reversible oxidation of formate to  $CO_2$  (HCOO<sup>-</sup>  $\leftrightarrow$   $CO_2 + H^+ + 2e^-$ ) [114][116][117]. β-Nicotinamide adenine dinucleotide hydrate reduced salt (NADH) is an essential co-factor for FDH, its efficiency is highly dependent on the NADH concentration as a hydrogen donor. NAD<sup>+</sup> is the oxidized form of β-Nicotinamide adenine dinucleotide hydrate (NAD). The active site of the NAD<sup>+</sup>-based FDH showing the C4-atom position on pyridine ring (**Fig. 16a**). Mondal et al., [105] reported the direct hydride transfer, occurs from the C-atom of the formate to the C4-atom of pyridine ring of NAD<sup>+</sup>, leading to NADH and  $CO_2$  (**Fig. 16b**). The neighbouring residues on FDH such as Thr (Thr282), Asp (Asp308), Ser (Ser334) and Gly (Gly335) interact with the positively charged NAD<sup>+</sup> ring and stabilize it, thereby promoting the hydride transfer from formate to C4 [118][119][120].

Generally, enzymatic reduction of CO<sub>2</sub> usually proceeds to HCO<sub>2</sub><sup>-</sup>, and the oxidized form of nicotinamide cofactor (NAD<sup>+</sup>) is reduced to NADH by glutamate dehydrogenase (GDH).



**Figure 16.** Two-electron reduction of CO<sub>2</sub> to HCOOH; (a) the active site of the NAD<sup>+</sup>-based FDH, (b) the catalytic mechanism of FDH Copyright [105], (c) a simplified reaction pathway of CO<sub>2</sub> to methanol (FDH, formaldehyde dehydrogenase (FDH), alcohol dehydrogenase (ADH)). The regeneration of NADH was catalyzed by glutamate dehydrogenase (GDH) [114].

A complex system of enzymes such as adenosine triphosphate (ATP), guanosine triphosphate (GTP), nicotinamide adenine dinucleotide (NAD or NADH for the reduced form), or nicotinamide adenine dinucleotide phosphate (NADP or NADPH for the reduced form), working in tandem is vital when considering biological conversion of CO<sub>2</sub>. There is a need to further

research in this area especially around the regeneration of cofactor NADH, which in turn, improves enzyme efficiency during complex CO<sub>2</sub> bioprocessing applications.

In the formation of acetic acid, methanol carbonylation is required, and the reaction comes after the formation of Methanol (photocatalytic reduction of CO<sub>2</sub>) [101] by using metals catalysts such as gold, platinum, palladium, osmium, copper-nickel, copper-Mordenite, palladium-cerium, ruthenium, rhodium, and lanthanum. Metal complexes play a huge role in achieving the carbonylation reaction through formation. In a typical Rh/CH<sub>3</sub>I catalysed system, a series of activated anionic species such as cis-[Rh(CO)<sub>2</sub>I<sub>2</sub>]<sup>-</sup>, [(CH<sub>3</sub>CO)-Rh(CO)I<sub>3</sub>]<sup>-</sup> and [(CH<sub>3</sub>CO)-Rh(CO)<sub>2</sub>I<sub>3</sub>]<sup>-</sup> to assist with product formation. Halogen additives is required to promote the reaction by activating C-O bond in methanol and on transition metal complexes to form methyl iodide (Eq. 1) which reacts with carbon monoxide to form acetyl-iodide (Eq. 2) and can quickly reach acetic acid final product when hydrolyzed in water (Eq. 3). The metal catalyst is recycled at the end of the reaction.

$$CH_3OH + HI \rightarrow CH_3I + H_2O$$
 (1)

$$CH_3I + CO \rightarrow CH_3COOI$$
 (2)

$$CH_3COOH + H_2O \rightarrow CH_3COOH + HI$$
 (3)

# **Conclusion and perspectives**

The potential of using bio-based photocatalysts for the conversion of CO<sub>2</sub> into value-added products, has been emphasized by describing the main advances in this domain from the literature in the last decade. The investigation of such catalytic systems is fundamental as this may afford the development of environmentally friendly, and sustainable processes as alternatives to replace the use of fossil fuels and consequently, contribute to the decrease global warming. The reported bio-inspired photocatalytic systems are presented following the target chemicals, namely carbon monoxide, formic acid, methanol and acetic acid (via carbonylation of methanol) all being key chemicals for a direct use in various applications or as intermediate synthons. Several case studies concern the use of metal catalysts combined with biomaterials which, under photocatalytic conditions, allow the formation of CO<sub>2</sub>-derived carbon molecules. The reduction of CO<sub>2</sub> using bioinspired catalysts is generally accompanied by proton-coupled

electron transfer (PCET) resulting in C–O bond cleavage to yield CO, an initial carbon-based product. CO<sub>2</sub> conversion to formic acid is usually promoted by formate dehydrogenase (FDH) and its co-factor NADH, responsible for donating hydrogen [117][118]. Carbonic anhydrase one of the fastest enzymes that catalyses CO<sub>2</sub>-hydration reaction has been combined with formate dehydrogenase (FDH) with co-factor NADH to form formate/formic acid [119][122][121][119]. One of the major drawbacks is the depletion of NADH during catalytic reduction reaction to NAD+, thus requiring continuous regeneration to NADH to have sustained formic acid production. The enzymes used for conversion of CO<sub>2</sub> to CH<sub>3</sub>OH are Formate Dehydrogenase (FDH) that utilizes NADH as a cofactor, and alcohol dehydrogenase (ADH). Enzymes usually exhibit much reduced activities over time due to impurities, slight change in reaction condition, (such as pH and temperature). The presence of transition metal/metal oxides incorporated unto biopolymers and enzymes facilitate the reduction of CO<sub>2</sub> to various products by promoting metal-CO<sub>2</sub> interaction. An example is [NiFe] CO Dehydrogenase ([NiFe] CODH), an enzyme used for conversion of CO<sub>2</sub> to CO. Zinc complexed Carbonic Anhydrase (CA), Zn[CA] is used for the conversion of CO<sub>2</sub> to bicarbonate (HCO<sub>3</sub>) [119].

Most of the enzymes being investigated in CO<sub>2</sub> reduction are naturally occurring and can denature easily under varying pH and temperature. Some key issues still remain that need to be considered when looking for further advancement in the biological catalysed CO<sub>2</sub> reduction. Firstly, the use of transition metals/metal ions incorporated unto enzymes protein ligands having high CO<sub>2</sub> binding affinity is vital for converting CO<sub>2</sub> molecules to desired products. Secondly, the depletion of NADH during catalytic CO<sub>2</sub> reduction reaction to NAD<sup>+</sup>, as well as the higher stability of CO<sub>2</sub> molecule, resulting in its lower reactivity. To avoid this problem, it is necessary to regenerate NADH during CO<sub>2</sub> reduction. A separate NADH regeneration step which involve the use of combined chemical, photocatalytic, biocatalytic, and/or electrochemical reactions within a single set-up must be considered when engineering efficient bioinspired catalyst for CO<sub>2</sub> reduction.

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### **Conflicts of interest**

The authors declare no conflict of interest

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