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## Transamidation vitrimers enabled by neighboring fluorine atom activation

Dimitri Berne<sup>a</sup>, Gwendal Tanguy<sup>a</sup>, Sylvain Caillol<sup>a</sup>, Rinaldo Poli<sup>b,c</sup>, Vincent Ladmiral\*,<sup>a</sup>, Eric Leclerc\*,<sup>a</sup>

The activation of associative transamidation via neighboring fluorine atom activation has been unveiled, quantified and applied for the construction of catalyst-free polyamide vitrimers. The inherent thermal and chemical stability of the amide functionality usually renders this linkage not suited for chemical exchange. The present study demonstrated that the positioning of a  $CF_2$  group  $\alpha$  to the amide group strongly activates this functionality, effectively turning on the dynamic transamidation. This effect was initially studied through combined DFT and kinetic investigations on the transamidation of small model molecules, prior to its application to polyamides networks. A range of fluorinated transamidation vitrimers were easily and rapidly synthesized by reaction of  $\alpha$ , $\alpha$ -difluoroesters with amines. Different amide/free amine ratios were employed, leading to materials that feature a range of relaxation and mechanical properties. Hence, flow activation energies going from 88.5 to 126 kJ.mol<sup>-1</sup> were obtained, allowing to easily modulate the vitrimer dynamic properties. Recycling tests of these additive-free transamidation vitrimers were performed at temperatures ranging from 150 to 200 °C and the mechanical and thermal properties of the reshaped materials were compared to those of the initial ones. Thanks to this approach, transamidation vitrimers with tuneable reshaping temperature, Young modulus and glass transition temperature have become accessible, paving the way to further applications of these materials.

#### **Keywords**

Vitrimer, transamidation, fluorine activation, neighboring group participation, polyamide

#### Introduction

Polyamides (PAs) feature high chemical resistance and excellent mechanical properties due to the high cohesive energy linked to the strong hydrogen bonds formed between amide functions of different chains. 1 A wide range of monomers are available to prepare PAs, providing access to a large structural diversity.<sup>2,3</sup> PAs have thus been used for a broad range of applications including automotive, clothing and electronics.<sup>4,5</sup> Since the commercialization of nylon 6,6 in 1938, numerous other thermoplastic PAs have been developed.<sup>6-9</sup> The synthesis of biobased thermoplastic PAs obtained from renewable resources such as plant oils enables reducing the environmental impact of the PA family. 10-13 The PA networks synthesis is generally performed in two steps. First, PA pre-polymers are prepared by polycondensation between a diacyl chloride and a diamine or by ring-opening polymerization. These conditions are considered "harsh" as they require either the synthesis of moisture-sensitive chloride compounds, or high temperatures when using diacid synthons. In the second step, the prepolymers are cross-linked by thermal- or photo-curing of double or triple-bonds. 14-21 In any case, PA networks often are not crystalline and thus do not feature the remarkable properties of PA thermoplastics. 17,22,23 In the current context of global waste reduction, the concept of Covalent Adaptable Network (CAN) has emerged as a new manufacturing process for thermosets and as a promising solution for the recycling of cross-linked polymers. CANs refer to polymer networks composed of covalent bonds capable of exchanging under a specific stimulus (often the elevation of temperature), enabling reshaping while still characteristic properties of thermosets such

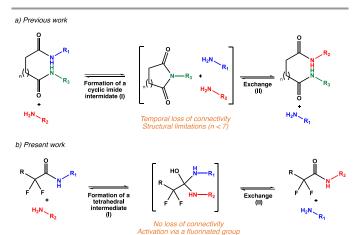
insolubility. 24-26 Numerous exchange reactions 27-38 have been developed since the pioneering work on disulfide exchange, 39,40 allowing the preparation of a broad range of CANs and vitrimers. Nevertheless, there are only a few examples of CANs based on a PA matrix. 23,41-43

Based on the previous research work, it appears of great interest to improve on one hand the efficiency of the PA synthesis and, on the other hand, to endow crosslinked PAs with dynamic properties to allow their reprocessing. A few examples of PA networks showcasing CAN properties that rely on the use of other linkages such as disulfide<sup>41</sup> or imine bonds<sup>42</sup> as exchangeable functions have been reported. Using the amide bond itself as the dynamic linkage is difficult due to the thermal stability and the chemical inertness of this function. Indeed, even if an amide group may theoretically exchange by reaction with an amine, with a carboxylic acid, or with another amide (amide metathesis), the reported methods for such reactions on model molecules generally require long reaction times, high temperatures (above 250 °C) or highly airand moisture-sensitive catalysts. 44-47 Most of the developed catalysts to achieve fast transamidation at low temperature are metal complexes (e.g. Fe(III) salts, Ni(cod)2, zirconocene dichloride, scandium triflate, AlCl3, Ti(NMe2)4, lanthanides).48-<sup>57</sup> Only few reports have presented metal-free transamidation of amides with amines, but they all relied on the use of coreactants or catalysts such as boric acid, hydroxylamine hydrochloride, selenium dioxide or potassium persulfate.  $^{58-61}$ Transamidation is however unavoidable during melt processing, resulting in the formation of segmented block copolyamides when two chemically different homopolyamides are processed as a blend. 62,63 The main mechanism for this transamidation is thought to involve hydrolysis-recombination and is thus largely affected by moisture and acidity. 64

Starting from these observations, the concept of neighboring group participation (NGP)<sup>65,66</sup> seems to be particularly suitable to promote transamidation. Indeed, NGP enables the activation of covalent bonds towards exchange reactions by the presence of specific functional groups in direct proximity

to the targeted bonds. 65,66 So far, NGP has mostly been used for the activation of transesterification. Hence, additional hydroxyl<sup>67,68</sup> or carboxylic acid<sup>69,70</sup> functions placed in epoxy networks accelerate the exchange process via the carbonyl group activation by H-bonding. Moreover, phthalate compounds were used to activate transesterification and transthioesterification via the formation cyclic intermediates.<sup>71–75</sup> Finally, of an increase the transesterification rate in epoxy-acid networks induced by neighboring fluorine atoms was recently demonstrated by our group. 76,77 This ester function activation is due to the strong electron-withdrawing effect of the fluorine atoms, which dramatically increases the electrophilicity of the ester carbonyl function.<sup>78</sup> It is important to underline that the fluorine activation maintained the associative character of the exchange reaction, whereas the formation of cyclic intermediate observed with other neighboring groups led to a dissociative mechanism. 71,79

Following the development of the NGP concept in CANs, Du Prez et al. recently highlighted that amide bonds could exchange via formation of an imide intermediate (Scheme 1a). 23 This exchange was made possible by the proximity of the amide groups (six or fewer carbon atoms between the two amide groups), enabling cyclization with the release of a free exchangeable amine under a thermal stimulus. This specific exchange was later used by Sijbesma et al. in a polymer network produced via addition of diamines onto polyimide.<sup>43</sup> In contrast to these dissociative CANs, transamidation vitrimers prepared from renewable resources were recently reported and presumed to involve an exchange between an amide and a free amine promoted by a boric acid catalyst.80 Mechanistic considerations were not discussed in this study but the exchange seemed to occur through an associative pathway, in opposition to the previously reported dissociative CANs proceeding through the imide formation.



Scheme 1. Transamidation by a) the formation of an intermediate imide  $^{23}$  and b) the formation of a tetrahedral intermediate

In the present study, our aim was to develop catalyst-free transamidation vitrimers by activation of the associative amine exchange reaction with the use of neighboring fluorine atoms (Scheme 1b). The fluorine effect on the amide formation as well as on the transamidation exchange was first highlighted on model compounds and rationalized by DFT calculations.

These preliminary studies prompted us to prepare additive-free transamidation vitrimers based on this fluorine activation. The use of different amide/free amine ratios indeed led to crosslinked materials that feature a range of relaxation and mechanical properties. Rheological experiments and reprocessability tests highlighted the impact of this ratio on the dynamic properties of the resulting vitrimers and provided insights into the transamidation key parameters.

#### Results and discussion

#### Model molecules

In order to compare their transamidation reactivity and quantify the reaction rates, difluorinated amides and their non-fluorinated counterparts were synthesized in two steps (Scheme 2). First, difluorinated (1, Figures S1-3) and nonfluorinated (2, Figures S4-5) esters were obtained by the substitutionof nucleophilic bromide bromodifluoroacetate and bromoacetate by *n*-octanethiol (Scheme 2A), respectively. This reaction was more efficient for the non-fluorinated bromoacetate, providing a cleaner reaction mixture and a higher isolated yield. One rationalizing hypothesis is that, in terms of the classical S<sub>N</sub>2 mechanism, the C-Br bond of ethyl bromoacetate is more polarized than that of ethyl bromodifluoroacetate, due to the powerful electronwithdrawing effect of the two fluorine atoms. However, it is also well-known that substitution reactions of fluoroalkyl bromides with thiolates may proceed through alternative mechanisms involving halophilic substitution or single electron-transfer steps. 81 Therefore, care should be taken when comparing halogen-substitution in the fluorinated and non-fluorinated series. In contrast, the subsequent solventfree amidation with benzylamine (Scheme 2B) occurred at 25 °C in 10 min on the  $\alpha,\alpha$ -difluorinated ester **1** to yield amide **3** (conversion > 99 %), whereas 48 h at 100 °C were required to achieve a 91 % conversion with the non-fluorinated analogue 2 to yield amide 4. The resulting benzylamides 3 and 4 were characterized by <sup>1</sup>H, <sup>19</sup>F and <sup>13</sup>C-NMR (Figures S6-8 for **3** and Figures S9-10 for 4). These observations clearly show that the two fluorine atoms dramatically increased the electrophilicity of the carbonyl group and enhanced the reaction rate.

Scheme 2. A) Syntheses of fluorinated and non-fluorinated esters. B) Syntheses of fluorinated and non-fluorinated benzylamides with their respective yields.

In order to examine the effect of the fluorine atoms on transamidation, the fluorinated and non-fluorinated amides (3 and 4) were reacted with an excess (10 eq.) of octylamine (5)

at 150 °C under neat conditions. After 72 h, 85 % (determined by <sup>1</sup>H-NMR) of **3** had been converted into fluorinated octylamide (**6**, Scheme 3A), while no conversion was observed for the non-fluorinated substrate **4** (Scheme 3). Note that **6** was not isolated nor purified.

(A) 
$$C_8H_{17}$$
  $S$   $F$   $F$   $H$   $+$   $C_7H_{15}$   $NH_2$   $\frac{\text{neat, 150°C}}{72 \text{ h}}$   $C_8H_{17}$   $S$   $F$   $H$   $C_7H_{15}$   $+$   $\frac{\text{BnNH}_2}{7}$   $\frac{\text{neat, 150°C}}{72 \text{ h}}$   $\frac{\text{Neath 150°C}}{72 \text{ h}}$   $\frac{\text{Neath 150°C}}{72 \text{ h}}$   $C_8H_{17}$   $\frac{\text{Neath 150°C}}{10 \text{ eq.}}$   $\frac{\text{Neath 150°C}}{10 \text{ eq.}}$ 

Scheme 3. Transamidation of A) 3 and B) 4 with 5 under neat conditions at 150  $^{\circ}\mathrm{C.}$ 

DFT These preliminary results were corroborated bv calculations. The computations used the same methodology (functional, basis sets and various corrections, see details in the Experimental section) as our previously published study of the F substitution effect on the transesterification rate. 18 The literature reports only few DFT investigations of the transamidation reaction mechanism, mostly restricted to metal-catalyzed<sup>53,82,83</sup> or organocatayzed<sup>84–86</sup> processes. It seems that only one report has addressed the non-catalyzed transamidation, for a special amide (RCO-NMeOMe, a "Weinreb" amide), in comparison with the CO<sub>2</sub>-catalyzed process.<sup>86</sup> As in this literature precedent, a second amine molecule was introduced in our calculations to serve as proton shuttle, leading to a tetrahedral intermediate (Scheme 4). Our investigation has addressed three model systems, as shown in Scheme 4. The first two (A and B) serve to assess the effect of the F substitution at the amide  $\alpha\text{-}C$  atom on the activation barrier, while comparison of systems A and C provides information on the additional effect of the sulfur atom. The alkyl chain was simplified to either a CH<sub>3</sub>S group for A and B or an ethyl group for C, while the attacking and leaving amines were simplified to methylamine, yielding a symmetric tetrahedral intermediate. The formation of the new amide product by amine elimination is thus the microscopic reverse of the initial amine addition.

Scheme 4. Non-catalyzed transamidation mechanism and model systems used in the present DFT investigation.

The results of these calculations are summarized in Figure 1. Starting with the separate reagents  $CH_3ZCX_2CONHCH_3$  and  $CH_3NH_2...NH_2CH_3$  (system I at the zero Gibbs energy reference point), the process starts with the formation of an H-bonded adduct (II), in which the amide NH is a proton donor and one N atom of the  $CH_3NH_2...NH_2CH_3$  dimer is the proton acceptor. This interaction is exoergic, the enthalpic gain dominating over

the entropic penalty. The stabilization is greater in the order A > B > C, correlating with the N-H bond polarity, which is regulated by the inductive effects of the X and Z groups (see optimized geometries in Figure S19). Along the path leading to the transition state, another local minimum (III) was identified, in which the interaction between the reactants involves an Hbond between the amine dimer as a proton donor and the amide carbonyl function as proton acceptor. For each system, this intermediate was located and optimized using, as a guess geometry, the optimized TS structure modified along the imaginary frequency normal mode. The higher G of III relative to II is essentially of enthalpic nature (no significant entropic contribution), due to the weaker NH···O bond (lower acidity of the amine vs. amide NH bond; lower basicity of C=O vs. amine). The optimized geometries of these intermediates (Figure S20) also reveal an electrostatic interaction between the incoming amine N atom and activated carbonyl C atom in IIIA and IIIC, whereas this N atom prefers to accept the amide NH proton in H-bonding in the structure of IIIB. The TS energies (optimized geometries in Figure S21) increase in the order A < C << B, highlighting the highly beneficial effect of the two F substituents (A << B), but also a beneficial effect of the  $\beta$ -S atom (A < C). The overall barriers from the lowest G hydrogen-bonded intermediate II to the TS are A (20.0 kcal  $mol^{-1}$ ) < C (21.0 kcal  $mol^{-1}$ ) < B (26.4 kcal  $mol^{-1}$ ). Finally, the tetrahedral intermediates (geometries shown in Figure S22) follow the same relative G trend as the transition states. The Gibbs energies of all molecules were also recalculated at 403 K (150 °C), yielding greater barriers because of the contribution of a negative activation entropy. The major entropic penalty is in the formation of the reactant H-bonded adduct, which is endoergic relative to the separate reagents at 150 °C. The activation barriers remain in the same relative order (A, 23.6; < C, 26.6 < B,  $33.2 \text{ kcal mol}^{-1}$ ), see profiles in Figure S23. The Cartesian coordinates of all optimized molecules are available in Table S1.

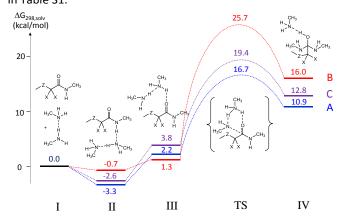


Figure 1. Gibbs energy profile (T = 298 K) for the transamidation process of the model molecules defined in Scheme 4.

In addition to the DFT calculations, a kinetic study was carried out to determine the activation enthalpy and entropy of the transamidation of **3** using dodecylamine (**8**) (Scheme 5). The reaction was monitored by GC-MS, which allowed the accurate measurement of the product concentrations, especially at low conversions. Dodecylamine **8** was chosen as the competing amine, over octylamine **5** used for the preliminary tests, as the

resulting amide **9** was well-separated from **3** by GC-MS, whereas **3** and **6** had similar retention times. The isolated fluorinated amide based on dodecylamine (**9**) was characterized by  ${}^{1}H$ ,  ${}^{19}F$  and  ${}^{13}C$ -NMR (Figures S11-13).

$$C_8H_{17}$$
  $\xrightarrow{S}$   $\xrightarrow{F}$   $\xrightarrow{N}$   $\xrightarrow{H}$   $\xrightarrow{$ 

Scheme 5. Transamidation of 3 with 8 in mesitylene at temperature ranging from 110 to 140  $^{\circ}$  C, x varied from 10 to 20.

The kinetic investigation started with the determination of the rate law for this transamidation reaction, which was anticipated to have a positive reaction order for both reagents, i.e. the amide (first-order) and the competing amine (firstorder or more). Mesitylene (MST, bp = 165 °C) was used as the solvent due to its high boiling point. Using different [8]<sub>0</sub>/[3]<sub>0</sub> ratios (10, 15 and 20) at 130°C, all kinetic profiles were in agreement with a pseudo-first-order rate law in amide, exhibiting very good linear fits when plotting  $ln([8]_0/([8]_0-[9]) =$ f(t) (Figure S24). The rate constants  $(k_{obs})$  deduced from the three reactions showed a good linear dependence vs. [8], as expected for a first order in amine (Table S2 and Figure S25). Hence, these first kinetic investigations confirmed the expected rate law for an associative exchange mechanism. Three additional kinetic measurements were carried out at 110, 120 and 140 °C using a  $[8]_0/[3]_0$  ratio of 10. The k values obtained for these reactions are reported in Table S3. These experiments allowed the determination of the experimental activation parameters of the transamidation reaction through the Eyring-Polanyi relationship (Figure 2). A good linear fit was obtained and the following activation parameters were extracted:  $\Delta H^{\dagger} = 7.78 \pm 0.63 \text{ kcal.mol}^{-1} \text{ and } \Delta S^{\dagger} = -65.7 \pm 1.6$ cal.mol<sup>-1</sup>.K<sup>-1</sup> (Figure 2B). The negative activation entropy is in agreement with the associative character of the reaction (ordered transition state).

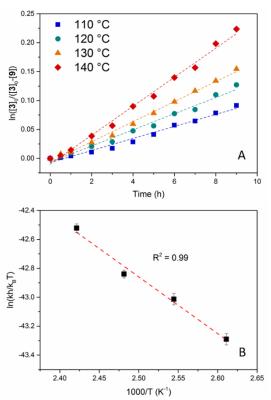
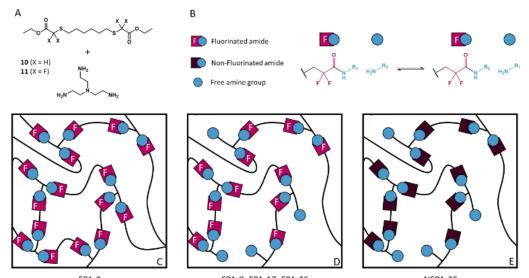


Figure 2. A) Monitoring of the transamidation of 3 with 8 in MST at 110, 120, 130 and 140  $^{\circ}\text{C}$  B) Eyring plot of the rate constants in the 110-140  $^{\circ}\text{C}$  range.

#### Polyamide networks synthesis

Based on the promising results obtained with model molecules, dynamic fluorinated polyamide (**FPA**) networks constituted of  $\alpha,\alpha$ -difluoroamide exchangeable groups were synthesized. A new bis- $(\alpha,\alpha$ -difluoro)ester (**10**) and its nonfluorinated analogue (**11**) were synthesized by substitution of, respectively, ethyl bromodifluoroacetate and bromoacetate with 1,6-hexanedithiol in DMF in the presence of  $K_2CO_3$  (Scheme S1). The desired products **10** and **11** were purified by column chromatography in order to remove the monoaddition product and the unreacted dithiol and their structures were confirmed by  $^1H$ ,  $^{19}F$  and  $^{13}C$ -NMR (Figures S14-16 for **10** and Figures S17-18 for **11**). It is important to note that these monomers were specifically designed so that the formation of an imide from the two resulting amides is strongly disfavoured as this would involve the formation of a 13-membered ring.



FPA-0 FPA-8, FPA-17, FPA-25

Figure 3. A) Di-esters and tri-amine used for the synthesis of the PA networks. Schematic representation of B) the associative transamidation, C) the fluorinated PA network without free amine groups (FPA-0), D) the fluorinated PA networks containing free amine groups (FPA-7, FPA-8 and FPA-25), and E) the non-fluorinated PA network containing free amine groups (NFPA-25).

Previous studies demonstrated that the degrees of crosslinking and of reactive functions (represented by free amine group in FPA) have an influence on the relaxation time and on the activation energy of the associated CANs. 66 Hence, to highlight the influence of the fraction of free amine in the network, a series of FPAs with different degrees of free amine functionalities were synthesized. It should be noted that a 3D structure was maintained, even at the highest free amine/amide ratio. 10 was thus mixed with tris(2aminoethyl)amine (TREN) as the cross-linker at different ratios (10/TREN =3/2, 2.75/2, 2.5/2 and 2.25/2) leading to materials containing 0, 8, 17 and 25 mol% of free amine groups, respectively and named FPA-0, FPA-8, FPA-17 and FPA-25 accordingly. The non-fluorinated equivalent of FPA-25 (named NFPA-25) was also synthesized by reaction of 11 and TREN in a 2.25/2 ratio (Figure 3).

As already observed in the synthesis of the model molecules, the fluorine atoms clearly accelerated the amidation reaction leading to the network formation. In the case of the FPA materials, gelation readily occurred 2 min after mixing the two monomers and a fully cured material was obtained after only 2 hours at 100 °C. In contrast, 30 hours at 100 °C were required to complete the synthesis of NFPA-25. Subsequently, preliminary reshaping tests using a hot press were carried out on all materials to quickly assess the influence of the free amine group fraction and of the fluorine activation. The temperature required for reshaping decreased as the percentage of free amine groups increased in the FPA series (Table 1). The transamidation exchange rate thus seems to

correlate with the percentage of free amine groups. **FPA-0** failed to (re)shape even after a thermal treatment at 220 °C under a 3-ton load for two hours. This suggests that the mechanism of transamidation involved in these FPA materials is associative (as it requires free amine groups). Finally, the fluorine effect on transamidation was evidenced by the required temperature for shaping **NFPA-25** (200 °C) which was 50 °C above that of **FPA-25**.

The formation of a cross-linked network for all the PA materials was confirmed by several analyses (Table 1). First, all the synthesized materials demonstrated a high gel content (GC), above 93 %. The percentage of free amine groups did not considerably impact the solubility properties of the PA networks, which all showed similar gel contents and swelling indexes (SI) in THF. ATR-FTIR analyses also confirmed the high conversion of the reactive functions as the initial absorption band of the ester bond of 10 at 1681 cm<sup>-1</sup> was completely replaced by the amide peak at 1760 cm<sup>-1</sup> in the **FPA** networks (Figure S26). These analyses did not enable differentiating the N-H bands of the amine from that of the amide, as their characteristic wavenumbers are too close. Finally, no residual exothermic peak was observed during the DSC analyses, confirming the complete curing of the materials (Figure S27). The glass transition temperature (Tg) determined from these DSC analyses increased as the percentage of free amine groups decreases, in agreement with the corresponding lower crosslinking density.

Table 1. Properties of fluorinated polyamide (FPA) and non-fluorinated polyamide (NFPA) networks

Material	GC (%) <sup>a</sup>	SI (%) <sup>a</sup>	T <sub>d</sub> <sup>5%</sup> (°C) <sup>b</sup>	T <sub>g</sub> (°C) <sup>c</sup>	T <sub>α</sub> (°C) <sup>d</sup>	E'glassy e (GPa)	E' <sub>rubbery</sub> f (MPa)	T <sub>(re)shape</sub> (°C) <sup>g</sup>
FPA-0	93 ± 2	62 ± 5	301	-	-	-	-	_h _
FPA-8	95 ± 1	60 ± 7	298	28	15	2.1	1.6	200
FPA-17	99 ± 1	69 ± 2	287	20	7	1.3	1.3	180
FPA-25	96 ± 1	75 ± 7	284	12	3	1.2	0.8	150
NFPA-25	97 ± 2	105 ± 2	292	8	-	-	-	200

 $^{a}$ Gel content and swelling indexes measured after 24 h immersion in THF;  $^{b}$ Temperature of degradation corresponding to 5 % weight loss determined by TGA analysis;  $^{c}$ Glass transition temperature determined by DMA analysis at E'' maxima;  $^{e}$ Determined at T<sub>α</sub> – 50  $^{e}$ C;  $^{f}$ Determined at T<sub>α</sub> + 50  $^{e}$ C;  $^{g}$ (Re)shaping was carried out at T<sub>(re)shape</sub> under 3 tons in 1 h;  $^{h}$ Shaping failed at 220  $^{e}$ C.

All the **PA** networks demonstrated high thermal stabilities with  $T_d^{5\%}$  (temperatures corresponding to a 5 % degradation) above 280 °C (Figure S28, and Table 1). Higher free amine fractions slightly decreased the thermal resistances as this chemical function is known to undergo degradation reactions at high temperatures. The **NFPA-25** network also demonstrated high cross-linking and high thermal stability. Its  $T_g$  was slightly lower than that of its fluorinated analogue, which may be rationalized by the higher steric hindrance induced by the presence of the fluorine atoms.  $^{87}$ 

**FPA-8, FPA-17** and **FPA-25** were evaluated by dynamic mechanical analyses (Figure 4). As expected, the  $T_{\alpha}$  measured by DMA followed the same trend as the  $T_{g}$  measured by DSC: as the free amine groups content increases, the chain mobility increases and the glassy/rubbery transition temperature decreases. Moreover, the storage modulus value slightly decreases with increase of free amine groups content, as expected.

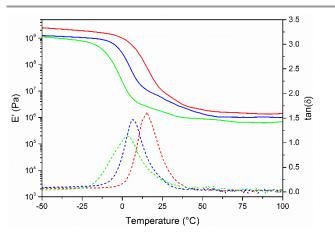


Figure 4. Storage modulus (plain lines) and  $tan(\delta)$  (dashed lines) of **FPA-8** (red), **FPA-17** (blue) and **FPA-25** (green).

#### **Dynamic properties**

The dynamic nature of the **FPA** networks and the influence of the free amine group content were evaluated *via* frequency sweep, relaxation tests and creep/recovery experiments. Frequency sweep analyses were performed on the **FPA**s at temperatures ranging between 120 °C and 180 °C (Figures S29-31) and showed that the storage moduli did not vary significantly with temperature. This observation is an additional indication that an associative transamidation exchange takes place in these FPA networks.<sup>79</sup> Indeed, for dissociative CANs, a decrease of the storage modulus with

increasing T caused by a loss of connectivity is generally observed. <sup>79,88</sup> Moreover, the increase of the loss modulus observed at low frequencies and high temperatures is related to the stress relaxation of the material. <sup>89</sup> This increase, particularly visible for **FPA-17** and **FPA-25**, indicates that these networks likely relax stress faster than **FPA-8**.

Stress-relaxation experiments were then performed on the FPA-X networks in the [140-180 °C] temperature window for X = 17 or 25 and in the  $[160-200 \, ^{\circ}C]$  window for X =8 (Figure 5). The initial relaxation modulus did not decrease with increasing temperature (Figure S32-34), confirming the frequency sweep observations that the exchange reaction does not induce a loss of connectivity and thus likely does not proceed via a dissociative mechanism. The normalized stress relaxation curves were fitted with a stretched exponential,  $G(t) = G_0 \exp(-\frac{t}{2})$  $(t/\tau)$ ) (Figure 5). These stress relaxation experiments show that the relaxation time of the FPA networks at any given temperature is shortened by an increase of the free amine group content. This result can be rationalized as follows. As previously demonstrated for transesterification vitrimers, 90 the stress relaxation time decreases with increasing content of hydroxyl groups in the network. Hence, the transamidation rate increased from FPA-8 to FPA-25 as the free amine group content in the network increased. In addition, the material cross-link density plays a role on the relative mobility, diffusion and availability of the exchangeable functions within the polymer network. Several studies (dealing with dioxaborolane chemistry, 91 transesterification 92,93 or vinylogous urethane exchange<sup>94</sup>) have demonstrated that an increase of the crosslink density is associated to a slower stress relaxation. Accordingly, FPA-25 relaxes the stress faster than FPA-17, which in turn relaxes stress faster than FPA-8.

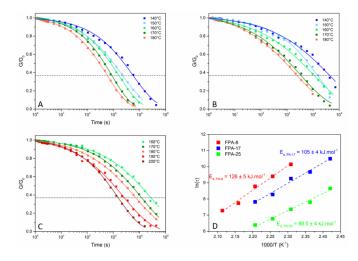


Figure 5. Normalized stress relaxation for A) FPA-25, B) FPA-17 and C) FPA-8. D) Arrhenius plot of FPA-X networks (X = 8, 17 or 25).

The Arrhenius plots of the **FPA** relaxation times (Figure 5D) are also consistent with an associative exchange. Indeed, even though a few dissociative CANs were shown to follow an Arrhenius behaviour within a certain T range, this property is always observed in associative CANs. The flow activation energies of 88.5, 105 and 126 kJ.mol<sup>-1</sup> obtained for **FPA-25**, **FPA-17** and **FPA-8** (Figure 5D), respectively, can be correlated to the increasing cross-link density, as described in previous studies. <sup>91–93</sup> It is also interesting to note that the activation energy is inversely proportional to the percentage of free amine groups, even though this trend was observed within a limited range of material composition (Figure S35). In conclusion, an increase of free amine group content in the presented **FPA** networks translates into a faster relaxation and a lower sensitivity of the material viscosity to temperature.

The preliminary kinetics on model molecules and the DFT study already demonstrated that the transamidation reaction rate is considerably accelerated by the neighboring fluorine atoms. This effect was further demonstrated by the comparison of the relaxation curves at 180 °C of NFPA-25 and FPA-25 (Figure S36). Both systems were able to relax stress at this temperature, but the relaxation time of FPA-25 was three times shorter than that of NFPA-25 (586 vs. 1703 s). The relatively fast relaxation of the non-fluorinated system is somewhat surprising. Indeed, according to previous studies,  $^{\rm 43,88}$  only materials in which the formation of an imide was possible allowed transamidation via dissociative exchange and were therefore able to relax stress. Here, the formation of imide is highly unlikely, compared to the associative exchange with a free amine, due to the number of carbon between the amide and amine functions. In addition, FTIR measurements at the reprocessing temperature failed to detect the imide characteristic bands in any of the tested materials. This further supports the hypothesis of the transamidation reaction occurring through an associative mechanism strongly accelerated by the high electron-withdrawing effect of the fluorine atoms, which increase the electrophilicity of the amide carbonyl functions. The transamidation reaction occurring in NFPA-25, though not activated by fluorine, is probably slightly favoured by the presence of the sulfur atom on the  $\alpha$ -carbon of the amide. This moderate influence was highlighted in the DFT study. Another possible way to rationalize this unexpected result is the high concentration of tertiary amines (from the TREN monomer) in this polyamide networks. Indeed, tertiary amines have been shown to promote transesterification in epoxy-acid vitrimers and may have the same effect on transamidation.

In order to assess the flow properties of the **FPA** systems, creep/recovery measurements were also performed at 50, 100 and 150 °C (Figure 6). At 50 °C, none of the **FPA** networks demonstrated any critical deformation, allowing their use at service temperatures under 50 °C. The higher temperature creep results were in good agreement with the stress-relaxation properties: **FPA-25** demonstrated higher creep at 150 °C than **FPA-17** or **FPA-8**. The 100 °C data show that **FPA-8** and **FPA-17** have almost no dynamic behaviour (no deformation), suggesting that these materials could be used up to 100 °C without significant flow.

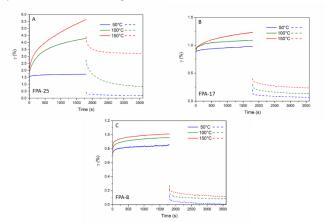


Figure 6. Creep and recovery behaviour at 50, 100 and 150 °C for an applied stress of 3 kPa for A) **FPA-25**, B) **FPA-17** and C) **FPA-8**.

#### Recyclability

One of the main advantages of vitrimers compared to thermosets is their capability to be reprocessed/reshaped. To test this property, the **FPA** networks were cut into small pieces (ca. 1 mm<sup>2</sup>) and placed in a hot press for 1 h under a 3-ton load. The temperature of reshaping was first optimized for each sample by visual observation, before assessing the reprocessing quality by TGA, DSC, FTIR, insolubility test, frequency sweep and tensile analyses. Slower-relaxing materials require higher reprocessing temperatures, hence **FPA-25**, **FPA-17** and **FPA-8** were respectively reprocessed at 150, 180 and 200 °C.

The properties of the reprocessed materials were evaluated after each processing up to three cycles for **FPA-17** and **FPA-25**. For **FPA-25**. No significant difference was observed between the DSC traces of the initial and reshaped materials (Figure S37), whereas a slight increase of T<sub>g</sub> was observed for **FPA-17** and **FPA-8**, indicating an increase of cross-link density, probably related to a slight material degradation (Figures S38-39). The retention of high thermal stability by the reprocessed samples was also indicated by thermogravimetric analyses for **FPA-25** and **FPA-17**, whereas a slight thermal stability decrease was observed for **FPA-8** (Figure S40-42). The FTIR spectra were nearly identical before and after reshaping for all CANs,

(Figures S43-45), further indicating that secondary reactions did not significantly occur during the hot press treatment and that the chemical integrity of the networks was maintained. Despite this relative material alteration with temperature, a reuse is still possible, depending on the targeted secondary application.

Frequency sweep measurements of **FPA-25** and **FPA-17** at 180 °C (Figure S46-47) also confirmed that the cross-link density

was not significantly impacted by the reprocessing step, as the storage modulus did not increase significantly for the recycled materials compared to that of the initial one. Moreover, a loss modulus increase at low frequencies could still be observed at 180 °C, demonstrating that the vitrimer dynamic behaviour was not lost in the reshaped materials.

Table 2. Physical properties of the initial and reshaped FPA networks

				FPA-25			
Material	T <sub>d5%</sub> (°C) <sup>a</sup>	Tg (°C) <sup>b</sup>	SI (%)°	GC (%) <sup>c</sup>	Young modulus (MPa) <sup>d</sup>	Stress at break (MPa) <sup>d</sup>	Strain at break (%) <sup>d</sup>
Initial	284	12	75 ± 7	96 ± 1	0.38 ± 0.03	0.38 ± 0.05	218 ± 21
Reshape 1	283	13	74 ± 1	98 ± 2	0.41 ± 0.05	0.40 ± 0.04	202 ± 30
Reshape 2	286	18	64 ± 7	94 ± 1	0.33 ± 0.03	0.24 ± 0.05	158 ± 29
Reshape 3	284	17	74 ± 1	99 ± 1	0.40 ± 0.06	0.38 ± 0.01	219 ± 25
				FPA-17			
Initial	287	20	69 ± 2	99 ± 1	2.19 ± 0.14	1.34 ± 0.41	86.7 ± 23.1
Reshape 1	291	25	65 ± 1	99 ± 1	1.71 ± 0.16	0.78 ± 1.40	56.6 ± 17.2
Reshape 2	291	30	61 ± 2	99 ± 2	7.10 ± 3.12	4.76 ± 2.22	10.5 ± 2.3
Reshape 3	288	33	61 ± 1	99 ± 2	216 ± 25	8.26 ± 3.01	6.8 ± 1.9
				FPA-8			
Initial	298	28	60 ± 7	95 ± 1	3.37 ± 0.63	1.68 ± 0.85	55.1 ± 12.7
Reshape 1	271	33	56 ± 1	97 ± 1	106 ± 24	7.78 ± 4.03	13.4 ± 5.1

<sup>a</sup>Temperature of degradation corresponding to 5 % weight loss determined by TGA analysis; <sup>b</sup>Glass transition temperature determined by DSC analysis; <sup>c</sup>Gel content and swelling indexes measured after 24 h immersion in THF; <sup>d</sup>Tensile tests were performed on dog bones of 1 mm thickness with a tensile speed of 0.1 mm s<sup>-1</sup>.

The reshaping efficiency was also evaluated by performing tensile tests on the initial and the reshaped samples. The Young moduli and stress/strains at break are reported in Table 2. The Young moduli of the initial FPAs are comparable to those obtained by the DMA analyses. Focusing on the FPA-25 results, no significant change of the characteristic properties was observed over three reshaping cycles. For **FPA-17**, on the other hand, the Young modulus as well as stress and strain at break were slightly decreased after one reshaping cycle. The evolution of these properties with the number of reshaping cycles was expected. Indeed, even though not evolving significantly, the glass transition temperature of FPA-X was raised above the analysis temperature (25 °C) by the reshaping process. Hence, after 2 or 3 cycles, FPA-17 is in a state closer to the glassy state rather than in the rubbery state of the initial and once-reshaped materials. The same phenomenon justifies the difference between the pristine and once-reshaped FPA-8. Nevertheless, these results are consistent with the other analyses and confirmed an efficient reshaping protocol for each material.

#### Conclusion

In this study, additive-free transamidation vitrimers could be developed thanks to neighboring fluorine atom activation. The

potential of  $\alpha$ , $\alpha$ -difluorination on the transamidation rate was highlighted by combined DFT and kinetic investigations on model molecules. The use of  $\alpha$ , $\alpha$ -difluoesters enables the rapid and facile synthesis of **FPA** bulk materials. The modulation of the free amine fraction appears as a facile way to control the dynamic behaviour of these **FPA** networks, as the reprocessing temperature of these materials could be tuned from 150 °C to 200 °C. The mechanical and thermal properties of these materials are also dependent on the free amine content, hence the Young modulus ranged from 0.38 to 3.4 MPa and the glass transition temperature from 12 to 28 °C. Considering the large amount of available aminefunctionalised molecules, the synthetic simplicity and the structural flexibility, these **FPAs** may find many practical applications, such as composites or reversible adhesives.

The polyamide vitrimers presented in this study did not demonstrate crystallinity as classical polyamides do. Hence, these materials should rather be considered as an extension of dynamic materials to polyamide networks and transamidation exchange reactions. Further works may focus on combining the crystallinity of polyamides and such dynamic properties in order to obtain ultra-resistant recyclable materials.

#### **Conflicts of interest**

The authors declare no conflicts of interest.

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#### **Materials**

1,6-Hexanedithiol (purity > 96%), 1-octanethiol (98.5%), ethyl bromodifluoroacetate (98%), 2-bromoethyl acetate (97%), tris(2-aminoethyl)amine (96%), benzylamine (99%), dodecylamine (99%), octylamine (98%) and mesytylene (MST, 98%) were purchased from Sigma-Aldrich (Darmstadt Germany). Pentane, Ethyl actetae (EtAc), DMF and CDCl $_3$  (99.5% D) were provided by Eurisotop (Saint- Aubin, France). All materials were used as received.

#### Characterizations

#### **Nuclear Magnetic Resonance**

The  $^1\text{H},~^{13}\text{C}$  and  $^{19}\text{F}$  NMR analyses were carried out in CDCl $_3$  using a Bruker Avance III 400 MHz NMR spectrometer at 25 °C. For  $^1\text{H}$  NMR (400 MHz),  $^{13}\text{C}$  NMR (100 MHz) and  $^{19}\text{F}$  NMR (377 MHz), CHCl $_3$  served as internal standard ( $\delta=7.26$  ppm) and data are reported as follows: chemical shift (in ppm), multiplicity (s = singlet, d = doublet, t = triplet, m = multiplet), coupling constant (in Hz), and number of nuclei from the integration. For  $^{13}\text{C}$  and  $^{19}\text{F}$  NMR (100 MHz), spectra were obtained with complete proton decoupling.

#### Gas Chromatography - Mass spectrometry

Gas chromatography—mass spectra (GC-MS) were recorded on a Shimadzu QP2012-SE with a Zebron ZB-5MS (20 m  $\times$  0.18 mm), capillary apolar column (stationary phase: 0.18  $\mu m$  film). GC-MS method: Initial temperature: 50 °C; initial time: 2 min; ramp: 22 °C/min; final temperature: 280 °C; final time: 15 min. MS spectra were recorded from 2.5 min to 4.4 min and 5.3 min to 15 min to avoid saturation with mesytylene (MST, 4.5 min) and octylamine (5.2 min).

#### Mass spectrometry (MS)

Mass spectrometry (MS) analyses were recorded on a Bruker Daltonics micrOTOF-Q with an ESI source and a positive ion polarity.

#### **Fourier Transform Infrared Spectroscopy**

The IR spectra were recorded on a Nicolet 210 spectrometer, equipped with an ATR accessory. The characteristic absorptions mentioned in the text are reported in cm<sup>-1</sup>.

#### **Thermogravimetric Analyses**

The thermogravimetric analyses (TGA) were carried out using a TG 209F1 apparatus (Netzch). Approximately 10 mg of sample

were placed in an aluminum crucible and heated from room temperature to 580 °C at a heating rate of 20 °C/min under a nitrogen atmosphere (60 mL/min).

#### **Differential Scanning Calorimetry**

The differential scanning calorimetry (DSC) analyses were carried out using a Netzsch DSC200F3 calorimeter, which was calibrated using indium, n-octadecane and n-octane standards. Nitrogen was used as purge gas. Approximately 10 mg of sample were placed in a perforated aluminum pan and the thermal properties were recorded between -100 °C and 150 °C at 20 °C/min heating/cooling rates to observe the glass transition temperature. The  $T_g$  values were measured on the second heating ramp to erase the thermal history of the polymer.

#### Dynamic mechanical analyses

The dynamic mechanical analyses (DMA) were carried out on Metravib DMA 25 with Dynatest 6.8 software. The samples were tested in the uniaxial tension mode at a frequency of 1 Hz with a fixed strain of  $10^{-5}$  m (0.1 % of material height), while applying a temperature ramp at a rate of 3 °C/min from -100 °C to +150 °C. The T $\alpha$  was determined as the maximum of the loss modulus E".

#### Rheology

The rheology measurements were carried out on a ThermoScientific Haake Mars 60 rheometer equipped with a lower electrical temperature module and an active upper heating system, with a textured 8-mm plane-plane geometry. For all rheology experiments, the applied stress was comprised in the linear viscoelastic region. A 1 N axial force was applied to ensure proper contact between the plates and the samples for all experiments. For the stress-relaxation experiments, a 0.5 % torsional strain was applied on the samples, and the rubbery modulus evolution with time was monitored at different temperatures. The obtained characteristic relaxation time ( $\tau$ ) was used to calculate the activation energy. Creep recovery experiments were carried out at 50, 100 and 150°C by applying a 3 kPa shear stress for a duration of 1800 s, followed by a recovery period of 1800 s.

#### **Swelling index**

Three samples from the same batch of the material, of ca. 20 mg each, were separately immersed in THF for 24 h. The swelling index (SI) was calculated using Equation 1, where  $m_2$  is the mass of the swellen material and  $m_1$  is the initial mass. Reported swelling index are average values of the three samples.

Equation 1 
$$SI = \frac{m_2 - m_1}{m_1} \times 100$$

#### **Gel content**

Three samples from the same material, of around 20 mg each, were separately immersed in THF for 24 h. The samples were then filtrated and dried in a ventilated oven at 70 °C for 24 h. The gel content (GC) was calculated using Equation 2, where

 $\rm m_2$  is the mass of the dried material and  $\rm m_1$  is the initial mass. Reported gel content are average values of the three samples.

Equation 2 
$$GC = \frac{m_2}{m_1} \times 100$$

#### **Tensile tests**

The material tensile tests were executed on an Instron 3366L5885 mechanical tester. Dog-bone sample of  $10 \times 2 \times 1$  mm were used for these analyses. The tensile speed was set at  $0.1 \text{ mm.s}^{-1}$  with a 100 ms sampling time. The Young modulus and the stress and strain at break were measured on five samples.

#### Procedure for the kinetic experiments

A flame-dried 10 mL scintillation vial was charged with the appropriate excess molar amount of dodecylamine, the amine was stirred at the appropriate temperature for 2 h to ensure amine decarboxylation. **3** (0.25 mmol, 1 eq), dissolved in mesytylene (MST, the volume of which was adjusted so that the total reaction volume is 2.5 mL), was then added quickly and the reaction was monitored by GC of periodically withdrawn aliquots (ca. 100  $\mu$  of reaction mixture diluted in approximately 5 mL of chloroform). The reaction temperature was varied from 110 °C to 140 °C. The reactions were carried out in the presence of TMB (trimethoxybenzene, 0.3 eq.) as an internal standard for the GC quantification.

#### Preparation of model molecules and monomers

Synthetic protocols used for the preparation of fluorinated molecules are followed by the ones used for the preparation of their non-fluorinated counterparts.

#### Ethyl 2,2-difluoro-2-(n-octylthio)acetate 1

$$SH + Br \downarrow 0$$
  $M_2CO_3$   $M_2CO_3$   $M_2CO_3$   $M_2CO_3$ 

In a 250 mL round-bottom flask were placed n-octanethiol (8.4 g, 57.6 mmol, 1 eq) and ethyl 2-bromo-2,2-difluoroacetate (17.5 g, 86.4 mmol,1.5 eq) dissolved in 100 mL of DMF. Potassium carbonate (7.9 g, 57.6 mmol, 1 eq) was then added slowly at 0°C. The mixture was then stirred and allowed to warm to room temperature. After 2 h, the reaction was quenched with water (100 mL). The mixture was extracted with diethyl ether (3 x 100 mL) and the organic layers were washed with a saturated sodium bicarbonate solution (3 x 100 mL), then with water (3 x 100 mL) and finally dried over MgSO<sub>4</sub>. The solvent was removed under reduced pressure. The crude product was purified by column chromatography on silica gel (from 100/0 up to 80/20 pentane/AcOEt). The product was obtained as a colourless oil (9.57 g, 62%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) 4.38 (q,  ${}^3J$  = 7.2 Hz, 2H, CH<sub>3</sub>-C<u>H</u><sub>2</sub>-O-CO) 2.89 (t,  ${}^3J$  = 7.4 Hz, 2H, S-C<u>H</u><sub>2</sub>-CH<sub>2</sub>), 1.75 – 1.62 (m, 2H, S-CH<sub>2</sub>-CH<sub>2</sub>-(CH<sub>2</sub>)<sub>5</sub>), 1.48 – 1.20 (m, 13H, C<u>H</u><sub>3</sub>-CH<sub>2</sub>-O-CO-CF<sub>2</sub>-S-CH<sub>2</sub>-CH<sub>2</sub>-(C<u>H</u><sub>2</sub>)<sub>5</sub>-CH<sub>3</sub>), 0.97 – 0.81 (m, 3H, S-CH<sub>2</sub>-CH<sub>2</sub>-(CH<sub>2</sub>)<sub>5</sub>-C<u>H<sub>3</sub></u>).

 $^{19}$ F NMR (377 MHz, CDCl<sub>3</sub>): δ (ppm) -82.9 (s).

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ (ppm) 162.0 (t,  $^2J$  = 33.0 Hz, O-CO-CF<sub>2</sub>-S), 120.7 (t,  $^1J$  = 285 Hz, O-CO-CF<sub>2</sub>-S), 63.6 (s, CH<sub>3</sub>-CH<sub>2</sub>-O-CO), 28.8 (t,  $^3J$  = 3.0 Hz, S-CH<sub>2</sub>-CH<sub>2</sub>), 28.7, 29.0, 29.1, 29.6 and 31.8 (s, S-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>3</sub>), 22.6 (s, S-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>3</sub>), 14.1 (s, CH<sub>3</sub>-CH<sub>2</sub>-O-CO), 13.9 (s, S-CH<sub>2</sub>-CH<sub>2</sub>-(CH<sub>2</sub>)<sub>5</sub>-CH<sub>3</sub>).

#### N-Benzyl-2,2-difluoro-2-(n-octylthio)acetamide 3

In a test tube were added compound **1** (0.50 g, 1.86 mmol, 1eq) and benzylamine (0.60 g, 5.58 mmol, 3 eq). The mixture was stirred for 10 minutes at 25 °C. The crude product was purified by column chromatography on silica gel (from 95/5 up to 70/30 pentane/AcOEt). The product was obtained as a colourless oil (0.50 g, 82%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) 7.42 – 7.30 (m, 5H, aromatics), 4.53 (d,  ${}^3J$  = 5.8 Hz, 2H, CO-NH-C $\underline{H}_2$ -Ar), 2.92 (t,  ${}^3J$  = 7.4 Hz, 2H, CH<sub>3</sub>-(CH<sub>2</sub>)<sub>5</sub>-CH<sub>2</sub>-C $\underline{H}_2$ -S), 1.78 – 1.61 (m, 2H, CH<sub>3</sub>-(CH<sub>2</sub>)<sub>5</sub>-C $\underline{H}_2$ -CH<sub>2</sub>-S), 1.49 – 1.20 (m, 10H, CH<sub>3</sub>-(C $\underline{H}_2$ )<sub>5</sub>-CH<sub>2</sub>-CH<sub>2</sub>-S), 0.96 – 0.87 (m, 3H, C $\underline{H}_3$ -(CH<sub>2</sub>)<sub>5</sub>-CH<sub>2</sub>-CH<sub>2</sub>-S).

<sup>19</sup>F NMR (377 MHz, CDCl<sub>3</sub>): δ (ppm) -82.5 (s).

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ (ppm) 162.1 (t,  $^2J$  = 28.9 Hz, S-CF<sub>2</sub>-CO-NH), 136.6 (s, aromatic carbon), 128.92 (s, 2 aromatic carbons), 128.02 (s, aromatic carbon), 127.86 (s, 2 aromatic carbons), 123.29 (t,  $^2J$  = 286.4 Hz, S-CF<sub>2</sub>-CO-NH), 43.78 (s, CO-NH-CH<sub>2</sub>-Ar), 28.7, 29.0, 29.1, 29.6 and 31.8 (s, S-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>

HRMS (ESI+): calc. m/z for  $[M+H]^+$ : 330.1698, measured: 330.1694.

#### N-Dodecyl-2,2-difluoro-2-(n-octylthio)acetamide 9

In a test tube were added compound **1** (0.5 g, 1.86 mmol, 1 eq) and dodecylamine (1.03 g, 5.58 mmol, 3 eq). The mixture was stirred for 1 h. The crude product was purified by column chromatography on silica gel (from 95/5 up to 70/30 pentane/AcOEt). The product was obtained as a colourless oil (0.61 g, 80%).

 $^{1}$ H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) 3.35 (q,  $^{3}J$  = 6.9 Hz, 2H, CO-NH-C $_{12}$ -CH<sub>2</sub>), 2.91 (t,  $^{3}J$  = 7.4 Hz, 2H, CH<sub>2</sub>-C $_{12}$ -S-CO-NH), 1.79 – 1.52 (m, 4H, CH<sub>2</sub> on alkyl chains), 1.47 – 1.22 (m, 28H, CH<sub>2</sub> on alkyl chains), 1.01 – 0.84 (m, 6H, CH<sub>3</sub> at the end of both alkyl chains).

<sup>19</sup>F NMR (377 MHz, CDCl<sub>3</sub>): δ (ppm) -82.6 (s).

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ (ppm) 162.0 (t,  ${}^{3}J$  = 28.4 Hz, S-CF<sub>2</sub>-CO-NH), 123.3 (t,  ${}^{1}J$  = 286 Hz, S-CF<sub>2</sub>-CO-NH), 28.7, 29.0, 29.1, 29.2, 29.2, 29.4, 29.5, 29.6, 29.6, 29.6, 31.8, 31.9 and 39.9 (s, C in alkyl chains), 28.6 (t,  ${}^{3}J$  = 2.9 Hz, CH<sub>3</sub>-(CH<sub>2</sub>)<sub>5</sub>-CH<sub>2</sub>-CH<sub>2</sub>-S), 14.1, 14.1, 22.6, 22.7, 26.7 (s, C in alkyl chains).

HRMS (ESI+): calc. m/z for  $[M+H]^+$ : 408.3182, measured: 408.3189.

#### Synthesis of ethyl 2-(n-octylthio)acetate 2

In a 50 mL round-bottom flask were placed octanethiol (3.3 g, 23 mmol, 1 eq) and ethyl 2-bromoacetate (5.8 g, 34.5 mmol, 1.5 eq), dissolved in 25 mL of DMF. Then, potassium carbonate (3.2 g, 23 mmol, 1 eq) was added slowly at 0°C. The mixture was then stirred and allowed to warm to room temperature. After 2 h, the reaction was quenched with water (50 mL). The mixture was extracted with diethyl ether (3 x 50 mL) and the organic layers were washed with a saturated sodium bicarbonate solution (3 x 50 mL), then with water (3 x 50 mL) and finally dried over MgSO<sub>4</sub>. The solvent was removed under reduced pressure. The resulting product (4.97 g, 93%) was used without further purification.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) 4.20 (q,  ${}^3J$  = 7.1 Hz, 2H, CH<sub>3</sub>-CH<sub>2</sub>-O-CO), 3.21 (s, 2H, O-CO-CH<sub>2</sub>-S), 2.64 (t,  ${}^3J$  = 7.3 Hz, 2H, S-CH<sub>2</sub>-CH<sub>2</sub>-(CH<sub>2</sub>)<sub>5</sub>-CH<sub>3</sub>), 1.67 – 1.55 (m, 2H, S-CH<sub>2</sub>-CH<sub>2</sub>-(CH<sub>2</sub>)<sub>5</sub>-CH<sub>3</sub>), 1.43 – 1.22 (m, 13H, CH<sub>3</sub>-CH<sub>2</sub>-O-CO-CH<sub>2</sub>-S-CH<sub>2</sub>-CH<sub>2</sub>-(CH<sub>2</sub>)<sub>5</sub>-CH<sub>3</sub>), 0.92 – 0.86 (m, 3H, S-CH<sub>2</sub>-CH<sub>2</sub>-(CH<sub>2</sub>)<sub>5</sub>-CH<sub>3</sub>).

 $^{13}\text{C NMR}$  (101 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) 170.6 (s, O-CO-CH<sub>2</sub>-S), 61.3 (s, CH<sub>3</sub>-CH<sub>2</sub>-O-CO), 33.7 (s, O-CO-CH<sub>2</sub>-S), 28.8, 29.0, 29.1, 29.2, 31.8 and 32.7 (s, S-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>3</sub>), 22.6 (s, S-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>

#### N-Benzyl-2-(n-octylthio)acetamide 4

In a test tube were added compound  $\bf 2$  (1 g, 4.3 mmol, 1 eq) and benzylamine (1.38 g, 12.9 mmol, 3 eq). The mixture was stirred for 48 h at 100 °C. The crude product was purified by column chromatography on silica gel (from 95/5 Pentane up to 70/30 pentane/AcOEt) to yield a colourless oil (0.93 g, 74%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) 7.41 – 7.30 (m, 5H, aromatics), 4.52 (d,  ${}^3J$  = 5.9 Hz, 2H, CO-NH-C $\underline{H}_2$ -Ar), 3.30 (s, 2H, S-C $\underline{H}_2$ -CO-NH), 2.53 (t,  ${}^3J$  = 7.4 Hz, 2H, CH<sub>3</sub>-(CH<sub>2</sub>)<sub>5</sub>-CH<sub>2</sub>-C $\underline{H}_2$ -S), 1.61 – 1.51 (m, 2H, CH<sub>3</sub>-(CH<sub>2</sub>)<sub>5</sub>-C $\underline{H}_2$ -CH<sub>2</sub>-C), 1.44 – 1.19 (m, 10H, CH<sub>3</sub>-(C $\underline{H}_2$ )<sub>5</sub> -CH<sub>2</sub>-CH<sub>2</sub>-S), 0.91 (t,  ${}^3J$  = 7.0 Hz, 3H, C $\underline{H}_3$ -(CH<sub>2</sub>)<sub>5</sub>-CH<sub>2</sub>-CH<sub>2</sub>-S).

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ (ppm) 168.8 (s, S-CH<sub>2</sub>-<u>C</u>O-NH), 138.0 (s, aromatic carbon), 128.8 (s, 2 aromatic carbons), 127.7 (s, 2 aromatic carbons), 127.6 (s, aromatic carbon), 43.8 (s, CO-NH-<u>C</u>H<sub>2</sub>-Ar), 36.3 (s, S-<u>C</u>H<sub>2</sub>-CO-NH), 29.1, 29.2, 29.2, 31.8 and 33.3 (s, S-CH<sub>2</sub>-<u>C</u>H<sub>2</sub>-<u>C</u>H<sub>2</sub>-<u>C</u>H<sub>2</sub>-<u>C</u>H<sub>2</sub>-<u>C</u>H<sub>2</sub>-CH<sub>2</sub>-CH<sub>3</sub>), 28.8 (s, CH<sub>3</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-

### Diethyl 2,2'-(hexane-1,6-diylbis(sulfanediyl))bis(2,2-difluoroacetate) 10

In a 250 mL round-bottom flask were placed hexane-1,6-dithiol (10 g, 66.5 mmol, 1 eq) and ethyl 2-bromo-2,2-difluoroacetate

(40.5 g, 200 mmol, 3 eq), dissolved in 100 mL of DMF. Then, potassium carbonate (18.4 g, 133 mmol, 2 eq) was added slowly at 0°C. The mixture was then stirred and allowed to warm to room temperature. After 2 h, the reaction was quenched with water (100 mL). The mixture was extracted with diethyl ether (3 x 100 mL) and the organic layers were washed with a saturated sodium bicarbonate solution (3 x100 mL), then with water (3 x 100 mL) and finally dried over MgSO<sub>4</sub>. The solvent was removed under reduced pressure. The crude product was purified by column chromatography on silica gel (from 99/1 up to 80/20 pentane/AcOEt) to yield a colourless oil (15.4 g, 58%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) 4.38 (q,  ${}^{3}J$  = 7.1 Hz, 4H, CH<sub>3</sub>-CH<sub>2</sub>-O-CO), 2.89 (t,  ${}^{3}J$  = 7.4 Hz, 4H, S-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>), 1.76 – 1.66 (m, 4H, S-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>), 1.49 – 1.43 (m, 4H, S-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>), 1.40 (t,  ${}^{3}J$  = 7.1 Hz, 6H, CH<sub>3</sub>-CH<sub>2</sub>-O-CO).

<sup>19</sup>F NMR (377 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) -82.8 (s).

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ (ppm) 161.9 (t,  $^2J$  = 32.9 Hz, CH<sub>3</sub>-CH<sub>2</sub>-O- $\underline{\text{CO}}$ ), 120.7 (t,  $^1J$  = 286 Hz, O-CO- $\underline{\text{CF}}_2$ -S), 63.6 (s, CH<sub>3</sub>- $\underline{\text{CH}}_2$ -O-CO), 29.4 (s, S-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>), 28.6 (t,  $^3J$  = 3.1 Hz, S- $\underline{\text{CH}}_2$ -CH<sub>2</sub>-CH<sub>2</sub>), 28.0 (s, S-CH<sub>2</sub>-CH<sub>2</sub>- $\underline{\text{CH}}_2$ ), 13.9 (s,  $\underline{\text{CH}}_3$ -CH<sub>2</sub>-O-CO).

HRMS (ESI+): calc. m/z for  $[M+NH_4]^{+}$ : 412.1234, measured: 412.1229.

#### Diethyl 2,2'-(hexane-1,6-diylbis(sulfanediyl))diacetate 11

HS 
$$\sim$$
 SH + 2 Br  $\sim$  DMF, 0°C, 2h

In a 100 mL round-bottom flask were placed hexane-1,6-dithiol (4 g, 26.6 mmol, 1 eq) and ethyl 2-bromoacetate (13.3 g, 79.8 mmol, 3 eq), dissolved in 25 mL of DMF. Then, potassium carbonate (7.4 g, 53.2 mmol, 2 eq) was added slowly at 0°C. The mixture was then stirred and allowed to warm to room temperature. After 2 h, the reaction was quenched with water (50 mL). The mixture was extracted with diethyl ether (3 x 50 mL) and the organic layers were washed with a saturated sodium bicarbonate solution (3 x 50 mL), then with water (3 x 50 mL) and finally dried over MgSO<sub>4</sub>. The solvent was removed under reduced pressure. The resulting product (8.16 g, 95%) was used without further purification.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) 4.18 (q,  ${}^{3}J$  = 7.1 Hz, 4H, CH<sub>3</sub>-CH<sub>2</sub>-O-CO), 3.19 (s, 4H, CO-CH<sub>2</sub>-S), 2.62 (t,  ${}^{3}J$  = 7.6 Hz, 4H, S-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>), 1.66 – 1.53 (m, 4H, S-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>), 1.45 – 1.35 (m, 4H, S-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>), 1.28 (t,  ${}^{3}J$  = 7.7 Hz, 6H, CH<sub>3</sub>-CH<sub>2</sub>-O-CO)

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ (ppm) 170.5 (s, O-<u>C</u>O-CH<sub>2</sub>-S), 61.2 (s, CH<sub>3</sub>-<u>C</u>H<sub>2</sub>-O-CO), 33.7 (s, O-CO-<u>C</u>H<sub>2</sub>-S), 32.5 (s, S-CH<sub>2</sub>-<u>C</u>H<sub>2</sub>-CH<sub>2</sub>), 28.8 (s, S-<u>C</u>H<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>), 28.2 (s, S-CH<sub>2</sub>-CH<sub>2</sub>-<u>C</u>H<sub>2</sub>), 14.2 (s, <u>C</u>H<sub>3</sub>-CH<sub>2</sub>-O-CO).

HRMS (ESI+): calc. m/z for [M+H]<sup>+</sup>: 323.1345, measured: 323.1344.

### Synthesis of fluorinated and non-fluorinated polyamide (FPA and NFPA)

For the synthesis of the polyamide networks, TREN and compounds **10** or **11** were mixed in a SpeedMixer for 3 minutes and then cured at 100 °C for 2 hours for the fluorinated materials (**FPA**) and 30 hours for the non-fluorinated material (**NFPA**). The resulting samples were cut into small pieces (ca. 1 cm²) and then shaped under a hot press for 1 h under 3 tonnes of pressure at different temperatures, depending on the initial material formulation.

#### **Computational details**

The computational work was carried out using the Gaussian09 suite of programs. The geometry optimizations were performed without any symmetry constraint using the BP86 functional and the 6-311G(d,p) basis functions for all atoms. The effect of dispersion forces (Grimme's D3 empirical method and solvation (SMD, were included during the optimization. The ZPVE, PV, and TS corrections at 298 K were obtained with Gaussian09 from the solution of the nuclear equation using the standard ideal gas and harmonic approximations at T = 298.15 K, which also verified the nature of all optimized geometries as local minima or first-order saddle points. A correction of 1.95 kcal/mol was applied to all G values to change the standard state from the gas phase (1 atm) to solution (1 M).

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