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Ex Situ **Generation of ¹⁸O² and ¹⁷O² from Endoperoxides for *O-Labeling and Mechanistic Studies of Oxidations by Dioxygen**

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ABSTRACT: Near-stoichiometric amounts of ¹⁸O₂ & ¹⁷O₂ were generated *ex situ* from endoperoxides in a twochamber glassware to oxidize various substrates. This strategy gave [*O_2]endoperoxides, $[^*O_1]$ quinones, $[^*O_1]$ phenols and [^{*}O_x]artemisin in moderate to good yields and high isotopic enrichments (up to 83%) at affordable costs. Moreover, mass spectrometry and ¹⁷O NMR of the [^{*}O]products provided valuable information on the chemical mechanisms involved

Among the various elements, oxygen plays a key role in many functional groups, and its isotopic labeling often proves determinant for mechanistic insights.¹ Indeed, ¹⁸O can be easily differentiated by mass analysis from the predominant ¹⁶O (99.759%), and recent advances in NMR instrumentation allows efficient detection of the chemical shifts of ^{17}O (-30 to +1000 ppm).² Due to the extremely low natural abundance of ^{18}O and ^{17}O (0.204% and 0.037%, respectively), the use of isotopically enriched compounds is essential, and synthetic methodologies for the incorporation of labeled oxygen (*O) have been extensively studied.^{3,4,5} They generally rely on one of the cheapest isotope precursors: $[^*O|H_2O$, but often require harsh conditions limiting their use to simple synthons, and/or involve reversible isotopic exchange yielding moderate isotopic enrichments. Advanced isotopic oxygen sources ($[^*O]CO_2$, $[^*O]CH_3OH,...$) are compatible with more specific reactions, but are extremely costly due to their low availability. Interestingly, some examples also reported the use of gaseous $*O_2$,^{3,6} for which the molar cost of the *O-labeled atoms is comparable to $[^*O]H_2O$.⁷ However, the need to employ large excesses of this gas and the difficulty to manipulate it precisely greatly increase the overall cost of these procedures, which made them under-used. All isotopically labeled gases share these drawbacks, but as demonstrated in the literature for some examples such as $[{}^*C]CO^8$, ${}^*H_2{}^9$ or $[{}^*C_2{}^*H_4]$ ethylene,¹⁰ they might be solved using a two-chamber glassware and a stable precursor able to release "*ex situ*" stoichiometric amounts of the desired gas. Thus, we decided to investigate the *ex situ* generation of $*O₂$ from easy-to-access solid [$*O$]compounds to enable cost-efficient $*O$ -labeling (Scheme 1, a).

Scheme 1: Strategy envisaged for efficient *O-labeling with *O² and syntheses of EPO [¹⁸O2]2 and [¹⁷O2]2.

Notably, endoperoxides scaffolds (EPO) are known to be precursors of singlet oxygen,^{11,12,13} and among the various possibilities tested, endoperoxide **2** was selected as the most promising structure. Indeed, it possessed both the ability to generate dioxygen quantitatively when heated over $100^{\circ}C^{14}$ and a high stability at room temperature, which avoided specific storage conditions and ensured practical manipulations. Moreover, multi-gram quantities of its $*$ O-labeled versions $[$ ¹⁸O₂ $]$ 2 and $[$ ¹⁷O₂ $]$ 2 were obtained in almost quantitative yields by reacting the commercially available 9,10-diphenylanthracene 1 with photosensitized singlet [*O₂]dioxygen coming from a lowpressure gas bottle (1L, 1.02 equiv., Scheme 1, b). Interestingly, the isotopic loss in this step was moderate, and starting from 90-99% *O-labeled dioxygen provided the desired EPOs with high isotopic enrichments of 84% and 82% for **[¹⁸O2]2** and **[¹⁷O2]2**, respectively.¹⁵ Then, the use of these compounds for *O-labeling on small scales (0.2- 0.5 mmol) was explored in combination with a specific two-chamber glassware designed to allow independent heating/light irradiation of each chamber.¹⁶ Firstly, the efficiency of the $*O_2$ generation & transfer was investigated by adding 1.5 equiv. of **[¹⁸O2]2** in chamber A and 9,10-diphenylanthracene **1** (1 equiv.) in chamber B in dichloromethane in presence of a catalytic amount of methylene blue (Scheme 2, a). After flushing the system under N_2 by freeze-and-pump cycles, chamber A was heated over 150°C for 3 min.¹⁷ Then, chamber B was irradiated at r.t. for 66 h hours under red light (660 nm LEDs), and **[¹⁸O2]2** could be isolated from this chamber in quantitative yield after a simple filtration on an alumina pad. Moreover, the excellent isotopic enrichment of 81% determined for **[¹⁸O2]2** indicated minimal isotopic losses (Scheme 2, a).¹⁸ Switching to a limiting amount of starting **[¹⁸O2]2** in chamber A also gave in chamber B **[¹⁸O2]2** in quantitative yield (based on the initial EPO) with 79% *O-labeling, which demonstrated the excellent efficiency of this set-up. Then, the scope of these oxidations by *O-labeled singlet dioxygen was explored, and various substrates were engaged with only a slight excess of $[^*O_2]$ endoperoxides **[¹⁸O2]2** or **[¹⁷O2]2** (1.5 to 2 equiv.). When 9,10-dimethylanthracene **3** was employed as starting material in conditions A at -10°C, the desired **[¹⁸O2]4** and **[¹⁷O2]4** were also obtained in very good yields of 87% and 96%, respectively (Scheme 2, b). These results demonstrated the reproducibility of the procedure, despite observing slightly lowered *O-contents in this case (63%-78%). Then, the synthesis $[^*O_2]$ ascaridole (an anthelmintic natural product) was investigated by 1,4-cycloaddition of α-terpinene **5** with *O-labeled singlet oxygen.¹⁹ In this case, **[¹⁸O2]6** & **[¹⁷O2]6** were produced easily in 96% and 67% yields with excellent isotopic contents of ¹⁸O/¹⁷O ranging from 78% to 82% (Scheme 2, c). The oxidation of furfural **7** was also attempted using 1.5 equiv. of **[¹⁸O2]2** or **[¹⁷O2]2**, and 5 hydroxy-2(5*H*)-furanone²⁰ was obtained with excellent yields (93% and 100%, Scheme 2, d). Mass analyses determined that the main isotopic molecular formula was $C_4H_4O_2*O_1$ for both (83% & 77%), which indicated minimal

isotopic losses. However, ¹⁷O NMR spectrum of **[¹⁷O1]8**, revealed the presence of two signals with identical integrations. As previously described in the literature,²¹ this confirmed that either the carbonyl- or the intracyclic-*Oatom was labeled, the two isotopomers being in equilibrium *via* the transient opening of the lactone.¹⁶ Oxidation of naphthoquinol **9** was also explored, and the mono-*O-labeled naphtoquinones **[¹⁸O1]10** and **[¹⁷O1]10** were synthesized in one step using conditions A. Yields were moderate (32-51%), but high isotopic enrichments were again measured (68%-83%, Scheme 2, e). Applying directly these conditions to substituted naphthoquinols **11** and **13** also gave the desired quinones, but yields were only ranging from 11% to 17%. However, this method still provided a straightforward access to the isotopically labeled versions of menadione (vitamin K3)²² [*O₁]12 and juglone²³ **[*O1]14** with high *O-enrichments (68-76%, Scheme 2, f & g). Moreover, ¹³C NMR demonstrated that the desired carbonyl possessed the sole ¹⁸O-labeled atom in **[¹⁸O1]12** and **[¹⁸O1]14**. 16

The photooxidation of substituted phenols was also investigated by irradiating substrate **15** under green light in presence of catalytic amounts of rose bengal $\&$ cesium carbonate,²⁴ while being exposed to *O-labeled dioxygen generated from two equivalents of **[*O2]2**. A mixture of quinol **[*O1]16a** and epoxyquinol **[*O2]16b** was obtained in both cases, with overall yields of 32% to 53% (Scheme 2, h). Isotopic enrichments of 84% and 77%, were determined for **[¹⁸O1]16a**and **[¹⁷O1]16a**, respectively. These values indicated very limited isotopic losses from the parent EPO $[{}^{*}O_2]2$, and ¹⁷O NMR of $[{}^{17}O_1]16a$ confirmed the unique labeling at the hydroxyl group. On the other hand, despite an overall ¹⁸O-content of 77% for the alcohol and epoxide functions of **[¹⁸Ox]16b**, the distribution of the isotopic molecular formulas was determined as 72% of $C_8H_{10}{}^{16}O_1{}^{18}O_2$, 24% of $C_8H_{10}{}^{16}O_2{}^{18}O_1$ and 4% of $C_8H_{10}^{16}O_3$.¹⁶ Taking in account the isotopic distribution of the initial endoperoxide of 84% of $[^{18}O_2]$ and 16% of **[¹⁶O2]2**, ¹⁶ the high ratio of mono-*O-labeled compound **[¹⁸O1]16b** observed was more consistent with an independent labeling of the two oxygen atoms rather than with an intramolecular double-oxidation pathway. Similar ratios were also measured for **[¹⁷Ox]16b**, and the ¹⁷O NMR experiment revealed that the two signals of the hydroxy and epoxy groups had equivalent integrations.¹⁶ Taken together, these results demonstrated that the epoxidation of the double bond mainly occurred by an intermolecular reaction with the hydroperoxide intermediate generated after the first step of phenol oxidation.^{24c} Finally, the photooxidation of boronic acids into phenols²⁵ was investigated using this two-chamber set-up with 1.5 equivalent of *O-labeled endoperoxide **[*O2]2**. Interestingly, irradiation of arylboronic acids **17**, **19** and **21** under green light in presence of 40 mol% of triethylamine and 5 mol% of rose bengal in ethanol at r.t. for 36 h provided smoothly the desired *O-phenols **[*O1]18**, **[*O1]20** and **[*O1]22**. Especially, good yields ranging from 51% to 90% and excellent isotopic enrichments of 78-83% were obtained (Scheme 2, i, j & k). These results confirmed that the labeled *O-atom introduced originated from of O_2 even at a low partial pressure. Moreover, these conditions were also applied to prepare in one-step $[\eta$ -¹⁷O₁]-(S)-*N*-Boc-tyrosine 24 in 40% yield and 72% of ¹⁷O-labeling at the phenol position (Scheme 2, l), which, after *N*-Boc-deprotection, can be used to study protein structures by ¹⁷O NMR.²⁶

Scheme 2: Oxidations of various substrates by near-stoichiometric amounts of *O² produced *ex situ* **from EPO [¹⁸O2]2 or [¹⁷O2]2.**

*General protocol: [*O2]2 (x equiv.) was placed in chamber A, substrate (0.2 to 0.4 mmol) & photosensitizer were solubilized in the solvent in chamber B. The system was flushed by N² using freeze-and-pump cycles. Chamber A was heated over 150°C for 3 min, then chamber B was irradiated by a suitable light. Conditions A: methylene blue (5 mol%), CH₂Cl₂ or CH₃OH, r.t. or -10°C, 660 nm LEDs (150W), 2 d. Conditions B: Cs2CO³ (15 mol%), rose bengal (1 mol%), MeOH, r.t., 525 nm LEDs (40W), 36-48h. Conditions C: NEt³ (40 mol%), rose bengal (5 mol%), EtOH, r.t., 525 nm LEDs (40W), 36h. Isotopic enrichments determined by mass analyses. δ(¹⁷O) in orange. See SI for detailed conditions. ^a Isolated in mixture with 1. b isolated in mixture with 16% of p-cymene. ^c 5-hydroxy-2(5H)-furanone [*O]8 converted partially converted into [*O]-trans-βformylacrylic acid after isolation ^dCalculated on the *O-atoms introduced.*

Finally, the possibility to perform complex and/or multiple oxidations was investigated, and artemisin **27** was identified as a suitable molecular target. Indeed, this well-studied anti-malarial drug is a natural endoperoxide compound²⁷ that can be obtain from dihydroartemisinic acid **25** by two consecutive dioxygen oxidations involving a postulated intermediary Hock cleavage.²⁸ In this context, compound 25 was placed in CH_2Cl_2 under N₂ in the chamber B of the two-chamber glassware with 5 mol% of methylene blue, while chamber A was previously loaded with three equivalents of $[^*O_2]$ (Scheme 3). After thermal release of ¹⁸O₂ or ¹⁷O₂, chamber B was irradiated under red light at r.t. for three days to enable the desired ene-reaction that formed *in situ* endoperoxide **[*O2]26**. In a second step, trifluoroacetic acid was added to the reaction mixture via a gas tight syringe to favor the one-pot Hock cleavage leading to the enol. Interestingly, it could reacted directly with the residual *O-labeled triplet oxygen in the two-chamber glassware, and spontaneous cyclization occurred to give the desired *O-labeled versions of artemisinin **[¹⁸Ox]27** and **[¹⁷Ox]27** in decent isolated yields of 18% and 20%, respectively.

Scheme 3: Synthesis of [*Ox]Artemisinin 27 by one-pot double oxidation of dihydroartemisinic acid 25 using $*$ **O**₂ produced *ex situ* from 3 equivalents of $[$ ¹⁸ O ₂ $]$ 2 or $[$ ¹⁷ O ₂ $]$ 2.

MB: methylene blue. Isotopic distribution determined by mass analyses. δ(¹⁷O) in orange. See SI for exact experimental conditions.

In both cases, mass analyses revealed a mixture of isotopic formulas ranging from zero to five oxygen atoms labeled, yet the expected **[*O4]27** was the main specie detected in 43% to 45%. Detailed examination of the isotopic mass distribution for the ¹⁸O-labeled artemisinin showed that summing the proportions for **[¹⁸O4]27** & $[{}^{18}O_3]$ 27 (= 73%) and $[{}^{18}O_2]$ 27 & $[{}^{18}O_1]$ 27 (= 22%) gave values close to the expected results (i.e. 71% of $[{}^{18}O_4]$ 27, 26% of **[¹⁸O2]27** and 3% of **[¹⁸O0]27**). This suggested that one *O-labeled atom was lost partially during the double oxidation process. These results confirmed the Hock mechanism (Scheme 4), which involved the *O-*O bond cleavage of $[^*O_2]26$ *via* protonation of the terminal oxygen of the peroxide and consecutive $[^*O]H_2O$ departure. Indeed, the water molecule is recaptured in these case by the carbocation issued from the vinyl migration. Thus, as unlabeled H_2O is in the final cyclization cascade, competition occurred at this stage between [*O]H₂O and [¹⁶O]H₂O to trap the carbocation, which decreased partially from one the number of $*O$ -labeled atoms in the final $[*O_x]$ artemisin **27**. Moreover, the unreacted *O-labeled water generated was able to freely exchange with the *O-atoms of the carboxylic acid until the cyclization step, which explains the small amounts of **[¹⁸O5]27** detected (3.8%, Scheme 3). Finally, the ¹⁷O NMR spectra of \lceil ¹⁷O_x]artemisin **27** showed the presence of three main signals (endoperoxide at 268 ppm, O-ester at 200 ppm and O-acetal at 88 ppm) and a small signal at 367 ppm, confirming a minimal amount of *O-atoms in the C=O bond.¹⁶

To conclude, easy-to-handle EPOs **[¹⁸O2]2** and **[¹⁷O2]2** were synthesized and used to produce near-stoichiometric amounts of $*O_2$ *ex situ* in a two-chamber glassware. This strategy allowed the ¹⁸O- and ¹⁷O-labeling of various compounds on small scales with very high isotopic enrichments and at affordable costs. Particularly, analyzing the [*O]products by mass spectrometry and 17O NMR provided valuable information on the chemical mechanisms involved. Thus, we believe that this method will bring significant benefits for studies on oxidations with $O₂$.

Scheme 4: Proposed mechanism for the formation of [*Ox]artemisinin from peroxide [*O2]26

ASSOCIATED CONTENT

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Supporting Information

General experimental procedures and analysis/spectroscopic data for new compounds $(^1H \text{ NMR}$, $^{13}C \text{ NMR}$, ^{17}O NMR and mass spectra). The Supporting Information is available free of charge on the ACS Publications website as PDF file [\(https://pubs.acs.org/doi/suppl/10.1021/acs.orglett.3c01487/suppl_file/ol3c01487_si_001.pdf\).](https://pubs.acs.org/doi/suppl/10.1021/acs.orglett.3c01487/suppl_file/ol3c01487_si_001.pdf)

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Author Contributions

P. H. conceptualized the project and acquired the funding. Investigation and methodology were performed by A. D. & M.-F. B., and P. H. & E. F. supervised the study. P. H. wrote the original draft and A. D., M.-F. B. and E. F. reviewed and edited the draft.

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- (16) See SI for details.
- (17) **EPO** 2 was never described as potentially explosive and it possesses a very low "Active oxygen content" (\leq 5), which should discard any safety issues of this protocol on millimolar scales using appropriate safety equipment. For general safety regards concerning peroxides, see: Dussault, P., "Working with organic peroxides in the academic lab", Organic Peroxides: Safety Issues. 2**, 2018**, doi:10.13014/K2GT5KCJ.
- (18) No reaction was observed in chamber B without irradiation, indicating that singlet dioxygen generated from **[¹⁸O2]2** in chamber A was fully de-excited to triplet dioxygen upon diffusion to the other chamber.
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