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1 Distribution and trophic functioning of planktonic communities in coastal marshes in Atlantic Coast of
2 France

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14

15 Abstract:

16 Coastal marshes are submitted to huge management due to anthropogenic pressure and thus, it is
17 essential to preserve their biodiversity, their ecological functions and the ecosystem services they can
18 provide. This study investigates the diversity and abundance of planktonic communities (heterotrophic
19 prokaryotes, heterotrophic protists, microphytoplankton and metazooplankton). The aim of this study is
20 to provide a first quantitative inventory on the plankton communities present in two marshes and to
21 construct the different typologies of planktonic food webs in these marshes. Seasonal samplings (4) for
22 environmental variables, nutrient concentrations and planktonic communities were conducted at 2-3
23 stations on each marsh during a year. A total of five different types of planktonic food web were
24 determined, three of them were found in both marshes. The saltmarsh phytoplanktonic communities
25 were dominated by Cryptophyta, nanoflagellates and Cyanobacteria. The freshwater marsh was
26 dominated by Cryptophyta in autumn and winter and by a diverse phytoplankton community in spring.
27 Marine copepods (Calanoida and Harpacticoida) characterized the saltwater metazooplanktonic
28 communities, Cladocerans and Rotifera the freshwater ones. Overall, the planktonic diversity was higher
29 in the freshwater marsh (102 taxa) than in the saltwater marsh (54 taxa).

30 Phytoplankton blooms represented nutrient and CO₂ uptake in both marshes and this reification function
31 seemed most efficient in autumn in the saltmarsh. Considerable zooplankton communities represented
32 a potential for nursery. Of the three management actions performed, only periodic water renewals might
33 affect the seasonal dynamics of planktonic communities.

34

35 Keywords: plankton, purification function, nursery function, salt- and freshwater marshes, food web

36

37 Graphical abstract

38

39 Highlights

- 40 - Planktonic diversity differs highly between the salt- and freshwater marsh.
- 41 - Similar planktonic ecological functions can be found in both marshes.
- 42 - Seasonal variations might be affected by management operation.
- 43 - Monitoring planktonic communities is essential to adapt coastal marsh management.

44

45 1. Introduction

46 Coastal marshes are habitats with high biological productivity, sustaining many ecological functions
47 (Vernberg, 1993; Więski et al., 2010). According to their uses and management, these environments
48 can provide several ecosystem services. Some of the most common services are directly linked to the
49 supply of human food resources, such as salt production, shellfish farming (Guillou, 1997; Leloup et al.,
50 2008), and fisheries (Maltby, 1991). Moreover, coastal marshes can provide regulation services by
51 forming a buffer zone against flooding of nearby inhabited areas (Sheng et al., 2022). They can also
52 provide cultural services as recreational spaces for the human population (Ghermandi and Nunes, 2013)
53 or provide refuge areas for migratory birds (Bradshaw et al., 2020). Overall, these different services are
54 often valued by their economic value (Woodward and Wui, 2001), which drives the type of management
55 applied to these environments. For example, in North America, dikes were constructed on some
56 marshes to directly control flooding to favor the development of specific marsh grass for salt hay farming
57 (Hinkle and Mitsch, 2005).

58 The various ecological functions and ecosystem services provided depend to a high extent on the
59 biodiversity present in coastal marshes. Halophyte vegetations and macrophytes provide shelter for

60 breeding and nesting for a wide diversity of animals from invertebrates to migratory birds (Pétillon et al.,
61 2014; Jerabek et al., 2017; Joyeux et al., 2017). In addition, coastal marshes constitute crucial nurseries
62 for juveniles of mobile species (*i.e.*, fish) that find requested trophic resources before migrating to a new
63 habitat (Cattrijsse and Hampel, 2006). One of the main resources of fish juveniles using these habitats
64 is metazooplankton (Stephenson, 1990; Kaneko et al., 2018; Talanda et al., 2022).

65 Planktonic communities are one of the actors of many ecological functions (Harris, 2012; D'Alelio et al.,
66 2016; Hébert et al., 2016). For instance, in coastal marshes, they are known to take part in the water
67 purification function by retaining suspended matter and nutrients and pollutants in the water column,
68 thus preventing eutrophication (Azim et al., 2005; Verhoeven et al., 2006; Nyman, 2011). Bradley et al.
69 (2010), showed that phytoplanktonic communities, especially picophytoplankton, were able to consume
70 both organic and inorganic nitrogen sources. While many papers consider the importance of benthic
71 communities on marsh ecology (Craft, 2000; Sullivan and Currin, 2002; Nordström et al., 2014), few
72 studies focus on the planktonic communities and their ecological associated functions in coastal fresh
73 and saltwater marshes (Quintana et al., 1998; Kobayashi et al., 2009). Even less consider the effect of
74 management measures on plankton communities and associated functions and ecosystem services.

75 David et al. (2020) studied the effect of human activities on phyto- and zooplankton diversity in
76 specifically drained marshes; they demonstrated that water replenishment and urbanization have a
77 strong effect on planktonic diversity and eutrophication. Koch and Gobler (2009) showed that draining
78 in salt marshes directly affects water quality and can increase development of diatoms and
79 dinoflagellates (*i.e.*, bloom). The introduction of species such as fish stocking for recreational fisheries
80 can reshape planktonic diversity (Reissig et al., 2006) and might also bring non-native planktonic
81 species. These invasive species might take over on native species if they are suited to the environmental
82 conditions in the marshes (Dexter et al., 2020). The aim of this study is to provide a first quantitative
83 inventory on the plankton communities present in the Fiers d'Ars and Tadon marsh, located on the west
84 coast of France, and to construct the different typologies of planktonic food webs in these marches. Both
85 marches being submitted to management practices, some considerations are formulated concerning
86 the potential impact of these on the plankton community composition, abundance and associated
87 ecological functions, especially the water purification function and nursery function.

88 In this study, the term metazooplankton is used to cover meso- and microzooplankton. Management
89 actions applied on the marshes were: (1) an annual drought, (2) the management of water inlet and

90 renewal in the marshes and (3) the introduction of non-native species. The potential effect of these
91 actions was compared to seasonal changes as observed in the planktonic communities during the study
92 period.

93 Based on the collected data and the study of Masclaux et al., (2014), who quantified both abundances
94 and flux between different planktonic compartments in a similar marsh also situated in Charente -
95 Maritime (France) and supported by literature references, we deduce the most likely trophic pathways
96 present in the studied marshes.

97 In short, microbial loops are found when both picophytoplankton and heterotrophic prokaryote
98 abundances are high, (Legendre and Rassoulzadegan, 1995). Herbivorous food webs can be found
99 when the temperature rises enough and nutrient concentration in the water column is sufficient to sustain
100 phytoplankton blooms (Huppert et al., 2002; Masclaux et al., 2014; Trombetta et al., 2019). Multivorous
101 food webs are likely when all planktonic communities are present and active in the water column
102 (Masclaux et al., 2014). Biological winters reflect situations where abundance and activity of planktonic
103 communities are low despite considerable concentrations of nutrients in the water column (Masclaux et
104 al., 2014). More details on these characterizations are given in the discussion of the specific situations
105 found in this study.

106

107 2. Methods

108 2.1. Study areas

109 The two marshes are located in Charente-Maritime, on the west coast of France (Fig 1). This area
110 contains several coastal marshes (salt and freshwater type) with variable uses and management types
111 and is considered as one of the biggest wetland zones in France (almost 100 000 ha) (David et al.,
112 2020).

113 The saltwater marsh (salinity between 22.9 and 37.6 throughout the year) studied is the Fier d'Ars
114 marsh, located at the North of the Ré-Island. This area is protected by a dike and is a highly touristic
115 site where traditional shellfish and salt farming activities are carried out (Fig 1a). The vegetation around
116 the marsh is mainly composed of *Halimione* sp., *Salsola soda* and *Tamarix* sp.. A part of this marsh is
117 included in a natural reserve managed by the "Ligue pour la Protection des Oiseaux" (LPO). Sampling
118 stations were selected in two areas. The first station FA (N 46° 14' 5.964"; W 1° 30' 20.707") is a 40 097
119 m² polyhaline basin with an average depth of 0.6 m located in the natural reserve. It is located about 0.1

120 km from the sea and connected to it by a channel. The management of this basin does not include
121 dredging or cleaning and no species have been introduced in addition to the resident species in the
122 marsh. The second station FB (N 46° 13' 26.4"; W 1° 27' 21.599") is an 8 483 m² polyhaline basin with
123 an average depth of 0.75 m. It is located about 2 km from the sea and connected to it by a channel;
124 when the tidal range reaches about 5.5 m, the water crossing the channels can enter the marshes. Thus,
125 juvenile fish can enter the marsh. The management of this basin includes dredging once a year in March.
126 As for FA, no species have been introduced in FB. The water inlets are regulated by a weir followed by
127 a floodgate to control water renewals depending on the season and the tidal range.

128 The freshwater marsh (salinity between 0.1 and 0.5 through the year) studied is the urban Tasdon marsh
129 located between La Rochelle and Aytré (Fig 1b). The vegetation around the marsh is mainly composed
130 of *Arrhenatheretea elatioris*. Sampling stations were selected in three distinct areas located between
131 1.5 and 2.5 km from the sea. The first station TA (N 46° 8' 56.4"; W 1° 7' 26.4") is a 72 000 m² freshwater
132 lake with a maximum depth of 1.35 m. The second station TB (N 46° 9' 3.6"; W 1° 8' 9.599") is a 21 400
133 m² freshwater basin with a maximum depth of 0.8 m. The third station labelled as TC (N 46° 8' 49.2"; W
134 1° 8' 13.2") is a 23 700 m² elongated freshwater basin with a maximum depth of 1 m. The management
135 of these three stations does not include dredging nor cleaning. However, freshwater fish such as black-
136 bass and sunfish have been introduced in these areas (early 2019) and tend to develop faster than the
137 native fishes present in the marsh, such as *Carassius* spp. and *Gambusia* spp.. Water inlets are coming
138 from an adjacent river connected to the stations: TB and TC located further downstream than at TA.

139

140 2.2. Sampling methods

141 Four seasonal samplings, corresponding to a season cycle (winter, spring, summer, autumn) were
142 conducted in 2019. All samplings were made in triplicate at each station at 0.5 m depth (sub-surface;
143 measured with a measuring rod). Temperature, salinity, pH, and dissolved oxygen concentration were
144 measured using a multiparametric probe (VWR™). A total of 5 L of water was sampled in polythene
145 bottles (mid water column) for quantification of Suspended Particulate Matter (SPM: Suspended
146 Particulate Inorganic Matter (PIM) and Suspended Particulate Organic Matter (POM)), dissolved nutrient
147 analysis (NO₂, NO₃, NH₄, PO₄, Si), chlorophyll *a* concentration and abundances of different planktonic
148 compartments (see after). These samples were preserved from light in opaque black plastic bags. To
149 estimate the abundance of heterotrophic prokaryotes and pico-nanophytoplankton, 1.5 mL of water was

150 directly transferred in cryotubes containing a mixture of glutaraldehyde (final concentration 0.2%) and
151 poloxamer (final concentration 0.001%). These fixed samples were frozen in liquid nitrogen and then
152 stored at -80°C in the lab. To study the diversity and abundance of microphytoplankton, 50 mL of water
153 sample were directly fixed with alkaline lugol iodine (final concentration of 4%). For heterotrophic
154 nanoflagellates (HNF), a 60 mL water sample was fixed with paraformaldehyde filtered at 0.22 µm (final
155 concentration of 1%). Metazoan microzooplankton was sampled by filtering 6 L of water through a 63
156 µm mesh stainless-steel sieve and then fixed with buffered formaldehyde (final concentration 4%).
157 Horizontal hauls were done at each station with a standard WP2 (200 µm mesh net) with an attached
158 flow meter (Mechanical Flow Meter with back-run stop, Model 438 115) to sample mesozooplanktonic
159 communities. The content of the net collector was fixed with buffered formaldehyde (final concentration
160 4%). Since the maximum depth was 1.35 m (TA), the effect of planktonic vertical migration was
161 considered as low (Gliwicz, 1986; Semyalo et al., 2009; Das et al., 2020). Horizontal hauls were then
162 sufficient to sample planktonic communities in the water column; thus, no additional vertical hauls were
163 done. Except for samples contained in cryotubes, all the samples were kept at 4°C in a cold-storage
164 room until analysis.

165

166 2.3. Sample analyses

167 The concentration of nitrate (NO₃), nitrite (NO₂), ammonium (NH₄), phosphate (PO₄) and silicate (Si)
168 was determined on filtered water (0.7 µm GF/F glass fiber membrane, Whatman) using a SEAL AA3
169 autoanalyzer with the detection limit of: 0.02 µmol.L⁻¹ (NO₃), 0.003 µmol.L⁻¹ (NO₂), 0.06 µmol.L⁻¹ (NH₄),
170 0.01 µmol.L⁻¹ (PO₄), 0.04 µmol.L⁻¹ (Si) according to the protocol of Aminot and K erouel (2004). SPM
171 were measured by weighing filters according to the protocol of Aminot and K erouel (2004). PIM was
172 also measured by weighing filters after acidification. POM was calculated as the difference between
173 SPM and PIM weight.

174 The concentration of chlorophyll *a* and phaeopigments was measured for each replicate for three size
175 classes: microphytoplankton (> 20 µm), nanophytoplankton (between 3 and 20 µm) and
176 picophytoplankton (< 3 µm). To do this, a triplicate filtration for each sampling was first performed on a
177 20 µm nylon fiber membrane (Millipore). Then, a fraction of the resulting filtrate was filtered onto a 3 µm
178 nucleopore membrane (Whatman) and the rest of the filtrate onto a 0.7 µm GF/F glass fiber membrane
179 (Whatman). Chlorophyll *a* and phaeopigments (*i.e.*, concentration of degraded chlorophyll pigment)

180 concentrations were quantified by fluorimetry according to the protocol of Lorenzen (1966). The
181 proportion of active chlorophyll *a* was measured by calculating the ratio of concentration of
182 phaeopigments to phaeopigment plus chlorophyll *a* initially measured.

183 FlowCam analyses were conducted rapidly after field work at the laboratory on living microplanktonic
184 organisms for each replicate using water from the polythene bottles. The abundance of microplanktonic
185 groups (phytoplankton and heterotrophic protists) were determined by FlowCam analyses following the
186 protocol of Buskey and Hyatt (2006). Microphytoplanktonic organisms (samples fixed with lugol) were
187 identified at the highest taxonomic level possible and counted under an inverted microscope (IX70
188 Olympus) coupled with a camera (DFC 450C Leica) (mainly at x400) according the Utermöhl protocol
189 (1958). One sample for each station-season were processed.

190 The abundance of heterotrophic prokaryotes, pico- and nanophytoplankton were determined by flow
191 cytometry analyses performed with a FACS Cantoll flow cytometer according to the method described
192 in Marie et al. (1999). The abundance of HNF from 2 to 20 μm was assessed by counting cells by
193 epifluorescence microscopy using the method of Bloem et al. (1986). Micro- and mesozooplanktonic
194 organisms were identified and counted with binocular microscope observations using the protocol of
195 Postel et al. (2000). Identification of micro- and mesozooplankton was performed to the species level
196 whenever possible (genus or order if damaged). To achieve species level identification, crustaceans
197 were dissected and observed with an optical microscope (x400) when necessary. All five copepodite
198 stages were regrouped for each copepod order. Planktonic larvae were identified to order directly under
199 a stereomicroscope (x120) without prior dissection.

200

201 *2.4. Statistical analyses*

202 All statistical analyses were conducted with R software (version 4.0.3.). First, heatmaps including
203 dendrograms based on euclidean distances were done to evaluate phytoplanktonic and
204 metazooplanktonic diversities on both marshes for each sampling station on the four seasons. In
205 addition, the data set obtained in this study was analyzed using a Multiple Factor Analysis (MFA) (Pagès,
206 2002), with FactoMineR and factoextra packages, to determine key variables guiding the seasonal
207 dynamics of planktonic communities on both marshes. Quantitative variables were divided in three
208 groups: a group including all abiotic parameters, a group including metazooplanktonic taxa and a group
209 including phytoplanktonic taxa. The management actions, including annual drought, water inlet and

210 renewal and introduction of non-native species, were used as descriptive qualitative variables (A). Then,
211 boxplots (package ggplot2) with associated Permanova and Fisher test (package vegan) were done to
212 determine the different types of planktonic food web (FW) per group, as described by Masclaux et al.
213 (2014). These analyses allow to compare, for each station-season couple, the concentration of nutrients
214 (NO₂, NO₃, NH₄), PIM, the proportion of active chlorophyll *a* and the concentration of chlorophyll *a* for
215 the three size classes (larger than 20 µm, between 20 and 3 µm, less than 3 µm), and the abundance
216 of micro- and mesozooplankton, ciliates, diatoms and heterotrophic prokaryotes.

217

218 3. Results

219 3.1. Diversity of microphytoplankton and metazooplankton

220 A total of 54 taxa for the Fier d'Ars marsh and 102 taxa for the Tasdon marsh were determined for the
221 microphytoplanktonic compartment. Taxa which were only present at one season for a single sampling
222 station were removed to retain the most representative taxa of the total abundance corresponding to
223 98% of the total abundance. Therefore, respectively 33 and 64 taxa for the Fier d'Ars marsh and the
224 Tasdon marsh were kept to study relative abundances of microphytoplanktonic communities per
225 season-station with a heatmap. For the Fier d'Ars marsh (Fig 2a), the dominant phytoplanktonic species
226 was *Teleaulax amphioxeia* (Cryptophyta), which was most abundant in autumn at both FA and FB and
227 in winter in FA. In summer, flagellates sp2 (not determined) were most abundant at FB whereas
228 *Planktolyngbya limnetica* (Cyanobacteria) was the most abundant at FA for the same season.
229 Considering the distribution of microphytoplankton communities between stations and by season
230 (results of clustering on heatmap), three groups could be distinguished: a first group including FB station
231 in spring and summer, a second group including both FA and FB in autumn and FA in winter, and finally
232 a third group including FA in summer and spring and also FB in winter.

233 For the Tasdon marsh (Fig 2b), the dominant phytoplanktonic species was *Plagioselmis nannoplanctica*
234 (Cryptophyta) which was most abundant in autumn and winter at TA and in autumn at TB. In summer,
235 microalgae of the genus *Scenedesmus* and *Willea rectangularis* (Chlorophyta) were most abundant at
236 TA and TB while *Lepocinclis acus* (Euglenozoa), *Haslea* sp., *Aulacoseira granulata* (diatoms), and
237 cyanobacteria of the genus *Aphanocapsa* were most abundant at TC. In spring, *Dinobryon* sp.
238 (Chrysophyceae) and flagellates were the most present at TA while green algae of the genus
239 *Monoraphidium* were dominant at TB. In the same season, *Monoraphidium* algae were also found with

240 a high abundance at TC with also *Haslea* sp., *Cryptomonas curvata* and flagellates sp6. Again
241 considering, only the distribution of phytoplankton communities (results of clustering on heatmap), four
242 groups could be distinguished: a first group including TA and TB in summer, a second group including
243 TA and TC in spring and TC in summer, a third group including all stations in autumn, and TA and TB
244 in winter, and a fourth group including TB in spring and TC in winter.

245 For the metazooplanktonic compartment, 30 and 42 taxa were determined for the Fier d'Ars marsh and
246 the Tasdon marsh, respectively. As for the phytoplanktonic compartment, relative abundances for each
247 taxon were studied per season-station. For the Fier d'Ars marsh (Fig 2c), copepods were most abundant
248 with the dominant genus *Acartia* at both FA and FB, especially in autumn. Copepod nauplii were
249 dominant in spring at both FA and FB, as well as in summer at FB. A high abundance of Zoe larvae (*i.e.*,
250 larvae of crab) was also noticed in spring at FA. Considering the distribution of metazooplankton
251 communities between stations and by season, two groups seemed to stand out: a group including both
252 FA and FB in autumn and winter and FA in summer, and a group including both FA and FB in spring
253 and FB in summer.

254 For the Tasdon marsh (Fig 2d), Cladocerans were the most abundant, especially *Bosmina longirostris*
255 with highest relative abundances observed in spring at TA and TB as well as in autumn at TA and TC.
256 Cyclopoid copepodite abundances were also high in summer at TA and TB. Rotifers of the genus
257 *Asplanchna* were the most abundant in winter at the three stations, and TC in spring. Nauplii of copepods
258 were the most abundant in summer in TC and in autumn in TB. In addition, two exotic and tropical
259 zooplankton species were found in Tasdon marsh: *Keratella tropica* and *Brachionus falcatus*. Looking
260 at the distribution of mesozooplankton communities between stations and by season, three groups could
261 be distinguished: a first group including the three stations in winter and TC in spring, a second group
262 including TA in spring and autumn, TB in spring and TC in autumn, and finally a third group including
263 the three stations in summer and TB in autumn gathering the juveniles' forms of copepods (nauplii and
264 copepodites).

265 Comparing both planktonic communities, the heatmaps showed different station-season's clustering
266 according to the type of planktonic community studied. First, for both study areas, the number of groups
267 resulting from clustering with the relative abundances of microphytoplankton or metazooplankton was
268 different. For the Fier d'Ars data, the majority of station-seasons were grouped in the same way for
269 phyto- and metazooplankton with the exception of FA in spring and winter. For the Tasdon data, more

270 differences were observed: whereas all three stations were grouped together for respectively summer
271 and winter when looking at metazooplanktonic relative abundances, they were only all grouped together
272 for autumn when looking at phytoplanktonic relative abundances. TA and TC in spring were grouped
273 together with TC in summer when looking at phytoplanktonic relative abundances. In contrast, when
274 looking at zooplankton relative abundances, TA and TB in spring were grouped together with TA and
275 TC in autumn, and TC in spring was grouped with all three stations in winter.

276

277 *3.2. Spatio-temporal variations of planktonic communities*

278 For the Fier d'Ars marsh, the three axes of the MFA explained around 72% of the variance of data (Fig
279 3). The abundance of heterotrophic prokaryotes (HPT), euglena (EUG), rotifera (RTF),
280 metazooplanktonic larvae (LRV), copepoda (COP), other metazooplanktonic organisms (OTH) and
281 pennate diatoms (DTMp) defined the first axis of the MFA with abiotic variables such as the water
282 temperature (T.w) and salinity (Sal). It was noted that all these variables were in opposition with the
283 dissolved oxygen concentration (O₂). The abundance of cryptophyceae, cladocerans (CLA) and the four
284 other metazooplanktonic groups which already defined the first axis (COP, OTH, LRV, RTF), euglena,
285 as well as the concentration of particulate organic matter (POM) and dissolved silicate (SI), hydrogen
286 potential (pH) and the proportion of active chlorophyll a for organisms between 3 and 20 µm defined the
287 second axis. The third axis was explained by the abundance of naked ciliates (CILn) and the
288 concentration of nitrate (NO₃), nitrite (NO₂), phosphate (PO₄) and particulate inorganic matter (PIM). The
289 importance of environmental parameters (especially temperature, salinity, and dissolved oxygen
290 concentration) in the distribution of the data indicated a strong seasonal effect. This observation was
291 confirmed by the representation of station-season individuals with ellipses of confidence via the MFA
292 (Fig 4), where especially summer and winter seemed to stand out from the other seasons. Contrary to
293 the seasonal effect, the three management operations included in the MFA as descriptive qualitative
294 variables, were not well represented in the three axes of the MFA.

295 For the Tasdon marsh, around 70% of the variance in the data was explained by the three axes of the
296 MFA (Fig 5). The first axis was explained by the abundance of copepoda (COP), cladocera (CLA),
297 metazooplanktonic larvae, heterotrophic prokaryotes, euglena, picoeukaryotes, nanoeukaryotes (NEC1
298 and NEC2), pennate diatoms and cyanobacteria, as well as the concentration of particular organic
299 matter, silicate. This group was in opposition with nitrates (NO₃) and nitrites (NO₂). Water temperature

300 also positively explained the first axis. The second axis was explained by the abundance of other
301 metazooplanktonic organisms, centric and chain diatoms, as well as the proportion of total active
302 chlorophyll *a* (total and for the three studied size classes), the concentration of particulate inorganic
303 matter, dissolved O₂, and negatively by salinity and pH. The third axis was mainly determined by the
304 proportion of total active chlorophyll *a* (total and for the three studied size classes), the concentration of
305 phosphate and negatively by the abundance of Rotifera. The abundance of photosynthetic communities
306 on the three size classes was an important factor characterizing the station-month pairs. Furthermore,
307 and similarly to the Fier d'Ars, the importance of environmental parameters (especially temperature and
308 nutrients) in the data distribution indicated a strong seasonal effect. This seasonal effect was revealed
309 by the representation of station-season individuals with ellipses of confidence via the MFA (Fig 6), where
310 especially summer seemed to stand out from the other seasons. Water inlet and renewal was
311 represented on the second axis.

312

313 *3.3. Planktonic food web typologies*

314 For the Fier d'Ars marsh, four different food web types were determined during the year of the survey
315 (Fig 7). An herbivorous food web (noted FW1, see Fig 9), characterized by high abundances of
316 phytoplanktonic communities and high proportion of active chlorophyll *a*, was observed in spring on both
317 FA and FB. A multivorous food web (noted FW2, see Fig 9), characterized by high abundances of
318 heterotrophic prokaryotes, micro- and mesozooplanktonic communities, and a medium abundance of
319 phytoplanktonic communities, was observed in summer on both FA and FB. In the case of FA, this food
320 web (FW2, see Fig 9) was developing in summer, with an increase in abundance of heterotrophic
321 prokaryotes and phytoplanktonic communities compared to spring, although the abundance of
322 metazooplanktonic communities did not yet increase. The third type of food web, defined as microbial
323 loop (FW3, see Fig 9) was observed in autumn on FA, characterized by high concentrations of NH₄, with
324 average abundance of heterotrophic prokaryotes. The food web "biological winter" (noted FW4, see Fig
325 9) was observed in autumn on FB and in winter on both FA and FB, and was characterized by low
326 abundances of phyto- and zooplanktonic communities, lower abundances of heterotrophic prokaryotes
327 than in the previous season (autumn for FA, summer for FB), as well as an increase of the concentration
328 of NO₂, NO₃ and PIM.

329 For the Tasdon marsh, four different food web types were determined, three of which are identical to
330 those identified at the Fier d'Ars marsh (Fig 8). As for FA and FB, the herbivorous food web FW1 was
331 observed in spring on TA and TB, and also in winter on TC. The multivorous food web FW2 was
332 observed in summer on all three stations (TA, TB, TC) and in spring on TC. However, this last station
333 was more characterized by FW2 in summer than in spring because the abundances of micro- and
334 mesozooplanktonic communities, phytoplanktonic organisms larger than 20 μm and smaller than 3 μm ,
335 diatoms and heterotrophic prokaryotes were higher in summer than in spring. The food web FW4 was
336 observed at all three stations in autumn. Another food web identified as "biological winter" (noted FW5,
337 see Fig 9) was observed in winter on TA and TB. The main difference between this "biological winter"
338 and FW4 was the higher concentration of NO_2 and NO_3 measured in winter compared to the
339 concentration measured in autumn for these two stations.

340

341 4. Discussion

342 4.1. Diversity and trophic functions of microphytoplankton and metazooplankton

343 Overall, the planktonic diversity was higher in the freshwater marsh of Tasdon (102 taxa) than in the
344 saltwater marsh of the Fier d'Ars (54 taxa). The saltmarsh phytoplanktonic communities were dominated
345 by Cryptophyta in autumn and winter, and by nanoflagellates and Cyanobacteria in summer. Marine
346 copepods (Calanoida and Harpacticoida) characterized the metazooplanktonic communities. The
347 freshwater marsh (Tasdon) was also dominated by Cryptophyta in autumn and winter. However, in
348 summer, Chlorophyta dominated at TA and TB while Euglenozoa, some diatoms and cyanobacteria
349 dominated at TC. In spring, phytoplanktonic communities were dominated by Chrysophyceae,
350 Chlorophyta and undetermined flagellates dominated the and metazooplanktonic communities by
351 Cladocerans and Rotifera.

352 These differences of planktonic taxa are consistent with planktonic diversity described in different
353 habitats (salt or freshwater) in the literature (*e.g.*, Walkusz et al., 2010; Badsı et al., 2012; Thakur et al.,
354 2013; Grujcic et al., 2018). Focusing on phytoplanktonic species, the data showed similarities in the taxa
355 found in both salt- and freshwater marsh, such as, for example, the presence of Cryptophyta. However,
356 high abundances of cyanobacteria and Euglenozoa were found in addition in the freshwater marsh in
357 summer. This observation is often correlated with high turbidity (up to an average of $223.84 \pm 13.41 \mu\text{g}$
358 L^{-1} of PIM at TC in summer) and can cause the seasonal succession of food webs in the marsh (Abrantes

359 et al., 2006). However, Ersanli and Gönülol (2006) concluded that such environmental conditions are
360 an indicator of low water quality.

361 It has been demonstrated in many aquatic ecosystems that a higher diversity might generate a better
362 stability and resilience (e.g., Ives and Carpenter, 2007; Pennekamp et al., 2018). Although diversity can
363 induce functional redundancy and hence not necessarily additional ecological functions, it can also
364 induce more diversified ecological functions. Indeed, the more different species there are, the higher the
365 probability of having species with different functional traits, and thus of having actors of different
366 ecological functions. It is therefore important to keep a high planktonic diversity in these environments
367 in order to maintain a maximum of ecological functions provided by these communities (Norlin et al.,
368 2006; Chaparro et al., 2019). In this study, the higher diversity of phytoplanktonic communities in the
369 freshwater marsh might produce a higher primary production than in the saltwater marsh, which has
370 lower phytoplanktonic diversity. Some protists (Euglenozoa, Cryptophyta, some of the ciliates) are
371 mixotrophic and can change their trophic regime according to the environmental conditions (Esteban et
372 al., 2010; Cordoba et al., 2021), thus their trophic functions are more difficult to determine. Nevertheless,
373 these communities contribute, with the other autotrophic phytoplanktonic communities, to the fixation of
374 atmospheric carbon through photosynthesis (Yan et al., 2021; Jia et al., 2022; Cohen, 2022). The
375 resulting phytoplankton biomass, which might then be transferred along food web to higher trophic levels
376 via the protists and mesozooplankton in these marshes and as such fuel the supply function (Atkinson
377 et al., 1996).

378 Focusing on the trophic diets of metazooplanktonic species identified in this study, there was a high
379 proportion of omnivorous species in the Fier d'Ars marsh, such as Harpacticoid copepods, *Acartia* sp.,
380 *Temora longicornis*, *Eurytemora pacifica*, *Evadne* spp., *Brachionus angularis* (Rieper, 1982; Kleppel,
381 1993; Berggreen et al., 1988). Different meroplanktonic larvae were also observed (Polychaeta, Zoe,
382 Echinoderma, veligers), confirming the importance of omnivory in the Fier d'Ars marsh (McEdward,
383 2020). Furthermore, detritivore species, such as *Mytilina* sp., Ostracoda and Amphipoda (Navarro-
384 Barranco et al., 2013; Galir Balki et al., 2017) were also present.

385 For the Tasdon marsh, *Asplanchna* sp., *Ceriodaphnia pulchella*, *Acanthocyclops trajani*, *Brachionus*
386 *calyciflorus*, *Brachionus falcatus*, *Polyarthra major*, *Keratella tropica*, *Lecane bulla*, described as
387 omnivorous species (Amoros, 1984; Abrantes et al., 2009; Santos-medrano et al., 2016; Rahmati et al.,
388 2020), were present. Non-selective filter feeder species as *Bosmina longirostris* and *Sida cristallina*

389 (Amoros, 1984; Bleiwas and Stokes, 1985) were also present. Detritivore species (Amoros, 1984;
390 Robertson, 1988) in the Tasdon marsh, were *Disparalona rostrata* and *Alonella excisa*.

391 Since omnivorous and non-selective filter feeder species can feed indifferently on plant or animal
392 organisms, they are more likely to find resources, which explains their high abundance in the marshes.
393 High abundances of these metazooplanktonic communities can provide a significant food source for
394 higher trophic level, and thus support the nursery function (Milstein et al., 2006; Rahmati et al., 2018;
395 Didenko et al., 2020). Detritivore species feed on suspended detrital matter by filtration and are therefore
396 actors of the purification function (Stanachkova et al., 2017). Overall, in both marshes, similar trophic
397 functions were found, which means that similar types of planktonic food web probably exist in both of
398 these marshes.

399

400 4.2. Seasonal changes of planktonic communities

401 On both marshes, a strong seasonal effect was observed in the planktonic communities. The
402 environmental parameter most strongly associated with the variability of these communities was
403 temperature, which explains the difference between the confidence ellipses of the stations in summer
404 compared to other seasons (Fig 4 and 6). Phytoplankton blooms take up nutrients and as such purify
405 the water. At FA station, the NO₂ and NO₃ concentrations were low in all the seasons except in winter,
406 and in parallel, phytoplankton abundance or biomass of chlorophyll were high in spring and summer.
407 Phytoplankton biomass provides resources for herbivorous (e.g., *Paracartia grani* for the Fier d'Ars
408 marsh and *Daphnia ambigua* for the Tasdon marsh), omnivorous and non-selective filter-feeder
409 zooplankton (Timms and Moss, 1984; Landry et Calbet, 2004), thus supporting the entire food web
410 (Costalgo et al., 2020). In addition, for the Fier d'Ars marsh, there was a strong effect of nutrients (NO₂,
411 NO₃, PO₄) observed with the MFA analysis, possibly linked to the run-off. For the Tasdon marsh, the
412 MFA analysis showed that phytoplanktonic communities were key actors with the strong correlation of
413 the proportion of active chlorophyll *a* for all three size classes. Apart from TA which can receive
414 freshwater from a connected river according to its seasonal flow, TB and TC are stagnant marshes,
415 without water renewal. It is known that planktonic communities tend to develop better with higher
416 abundances in lentic and stagnant environments than in lotic environments because their residence
417 time in the environment is longer (Baker and Baker, 1979; Koslow et al., 2008). So here also, the
418 considerable phytoplankton presence represents a purification function

419

420

421 *4.3. Food web and ecological functions*

422 While the present study does not include flux measurements, the deduction of food web typologies made
423 from biomass data is justified by the fact that solid theoretical basis exists for this approach (Volterra,
424 1928; Tortajada, 2011; Barbier and Loreau, 2019). Also, several of the food web typologies reported in
425 the marches studied, such as an herbivorous, microbial and mixed loop types have also been found
426 experimentally in other coastal, SPM rich environments, such as brackish and freshwater estuarine
427 reaches (Tackx et al., 2003; David et al., 2006; Van de Meersche et al., 2009; Masclaux et al. 2014;
428 Kimmel et al., 2015; David et al., 2016).

429 A total of five different types of food web were determined between the two marshes studied (Fig 9).
430 The herbivorous food web (FW1) was defined by high abundances of phytoplanktonic communities and
431 sufficiently high abundance of herbivorous zooplanktonic communities to regulate them by grazing
432 (Dupuy et al., 2011; Chenillat et al., 2021). Landry et al. (1995) demonstrated that pigment analyses of
433 zooplankton stomach contents showed a higher concentration of chlorophyll pigments in autumn and
434 spring, and a higher grazing rate especially for microzooplankton. These results are consistent with
435 those published by Masclaux et al. (2014) on the regulation of phytoplankton communities by
436 zooplankton in the herbivorous food-web (FW1). In the Tasdon marsh, the high proportion of active
437 chlorophyll *a* indicated a strong photosynthetic activity from autotrophic organisms during this period,
438 thus a strong contribution to atmospheric carbon fixation in the water column (Copping and Lorenzen,
439 1980). The data indicated two orders of magnitude higher nutrient concentrations in the Tasdon marsh
440 (see Fig 7 and 8) than in the Fier d'Ars marsh. This could be explained by the fact that the Tasdon
441 marsh, unlike the Fier d'Ars marsh, is located on an urbanized watershed, and therefore receives more
442 nutrients by leaching during rainfall (Rudy et al., 1994). Thus, the intake of nutrients might have a higher
443 effect on the saltwater marsh studied since their concentrations of NO₃ and NO₂ were initially lower. The
444 efficiency of the purification function in both marshes might be equivalent, since the concentrations of
445 NO₃ and NO₂ tended to decrease significantly between seasons in FA, TA, TB and TC. The difference
446 in the initial concentrations of nutrients in this case is important to consider. Since the decrease of NO₃
447 and NO₂ concentrations between winter and spring, was significant (p-value = 0.009) at FA but not at
448 FB the water purification function might be more efficient at FA than at FB. It is however clear that more

449 information is needed on nutrient and organic matter inputs to be able to fully assess the importance of
450 purification activities by plankton and prokaryote communities.

451 The multivorous food web (FW2) was defined by high abundances of heterotrophic prokaryotes and
452 protists, phyto- and metazooplanktonic communities as well as high turbidity linked to a high
453 concentration of particulate organic matter resulting from bacterial degradation and mineralization
454 processes (Kirchman, 1994; Jia et al., 2017), and a low concentration of nutrients (Masclaux et al.,
455 2014). This food web appeared directly after the herbivorous food web: the increase of the abundance
456 of phytoplankton during the previous period induced the increase of available prey for zooplankton which
457 can thus grow in the habitat (Chenillat et al., 2021). A multivorous food web can be qualified as weak or
458 strong depending on the abundances of all planktonic compartments (Masclaux et al., 2014). Therefore,
459 there was a weak multivorous food web in spring at TC compared to all three stations of Tasdon marsh
460 in summer because HPT and mesozooplanktonic abundances were not as high. Moreover, at FA in
461 summer, the multivorous food web identified was beginning because the abundance of micro- and
462 mesozooplankton were just starting to increase compared to their higher abundance in spring. Overall,
463 these high planktonic abundances represented a large pool of resources and food for planktivorous
464 species and thus might indicate a substantial nursery function if they are simultaneously present in these
465 habitats, the case in both marshes during summer, and at TC in spring.

466 The microbial loop (FW3) was identified only in autumn at FA and defined by high concentration of NH_4 ,
467 which is a product of particulate matter recycling in the water column, low concentrations of NO_2 and
468 NO_3 , and high abundances of heterotrophic prokaryotes (Legendre and Rassoulzadegan, 1995;
469 Almuktar et al., 2018). Since the concentrations of nitrite and nitrate were low despite the nutrient input
470 coming from water inlet, the purification function was well shown at FA. The first type of “biological
471 winter” (FW4) was defined by low phytoplanktonic, metazooplanktonic and heterotrophic prokaryote
472 abundances. The second type of “biological winter” (FW5) was similar as FW4 except for the higher
473 concentration of nitrite and nitrate, which may be linked to inlets of water bringing nutrients in the
474 freshwater marsh (Nalubega, 1999). FW5 might correspond to a late state of biological winter close to
475 transition with an herbivorous food web when the temperature increases with phenology (Trombetta et
476 al., 2019).

477 The herbivorous and multivorous food webs were found in both marshes, as well as the first type of
478 “biological winter” FW4. The microbial loop was only noted in the Fier d’Ars marsh, whereas the second

479 type of “biological winter” FW5 was only revealed in the Tasdon marsh. According to the statistical
480 analyses, the effect of nutrient concentrations in the water column seemed to have a stronger impact in
481 the saltmarsh than in the freshwater marsh. Again, knowing that the intake of nutrients by water inlets
482 might be more important at Fier d’Ars than Tasdon; sampling the water directly in the channel leading
483 to the marshes and measuring nutrients concentrations during each the season would be necessary to
484 solidify this statement.

485 It is highly likely that a season is composed of a succession of different food webs; unfortunately, with
486 only one sampling per season, it is possible that we missed some situations. The second type of
487 “biological winter” might have been identified in the saltmarsh if the sampling had been done later in
488 winter, closer to the beginning of spring. It should be noted that the presence and dynamics of some
489 phyto- and zooplanktonic species seem to be directly linked to the season, which correlates with other
490 published studies (Ogbuagu and Ayoade, 2012; Ratnam et al., 2022), therefore the efficiency of their
491 associated ecological functions might be affected as well.

492 Overall, the seasonal succession of food web types seemed to follow the patterns generally described
493 in the literature (Legendre and Rassoulzadegan, 1995; Masclaux et al., 2014).

494 The results of the MFA indicated that, during the year studied, the three management operations studied
495 did not impact planktonic communities more than the natural seasonal effect. Angeler and Moreno, 2007
496 also state that planktonic communities are resilient this type of one-time and infrequently repeated
497 methods applied on coastal marshes. It would be interesting to study whether similar results are found
498 if these same methods are used at higher frequencies, to assess the limit of resilience of planktonic
499 communities. Also, the potential combined effects of different management operations, such as the three
500 actions studied here and sediment resuspension caused by landscapes modelling, should be studied
501 over consecutive years to determine how resilient the communities involved in these ecological
502 functions.

503

504 5. Conclusion

505 On the whole, this study showed that, although planktonic communities composition, abundance and
506 diversity are substantially different, similar planktonic food web types can be found on salt and
507 freshwater coastal marshes. This one-year survey, with a sampling each season, provides a basis of
508 information on the composition and abundance of the planktonic communities in the Fier d’Ars and

509 Tasdon marshes and allows to deduce their most likely trophic pathways and associated ecological
510 functions. Integration into the context of the interdisciplinary project PAMPAS will allow more detailed
511 analysis of the role of planktonic communities within the total marsh ecosystems.

512

513 CRediT authorship contribution statement

514 Lauriane Bergeon: Conceptualization, Formal analysis, Investigation, Writing – Original Draft, Writing –
515 Review & Editing, Visualization. Frédéric Azémar: Conceptualization, Resources, Writing – Review &
516 Editing, Supervision. Claire Carré: Resources, Data Curation, Writing – Review & Editing. Bénédicte
517 Dubillot: Methodology, Validation, Investigation, Resources. Claire Emery: Investigation, Resources,
518 Data Curation. Hélène Agogué: Investigation, Resources, Data Curation, Writing – Review & Editing.
519 Philippe Pineau: Investigation, Resources, Data Curation. Thomas Lacoue-Labarthe: Project
520 administration, Funding acquisition, Writing – Review & Editing. Marc Bouvy: Resources, Data Curation,
521 Writing – Review & Editing. Michèle Tackx: Conceptualization, Validation, Writing – Review & Editing,
522 Supervision. Christine Dupuy: Conceptualization, Methodology, Validation, Investigation, Resources,
523 Data Curation, Writing – Review & Editing, Supervision, Project administration.

524

525 Declaration of Competing Interest

526 The authors declare that they have no known competing financial interests or personal relationships that
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528

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543

544 Data availability statement

545 Data will be made available upon reasonable request.

546

547 References

548 Abrantes, N., Antunes, S.C., Pereira, M.J., Gonçalves, F., 2006. Seasonal succession of cladocerans
549 and phytoplankton and their interactions in a shallow eutrophic lake (Lake Vela, Portugal). *Acta*
550 *Oecologica* 29, 54–64. <https://doi.org/10.1016/j.actao.2005.07.006>

551 Abrantes, N., Nogueira, A., Gonçalves, F., 2009. Short-term dynamics of Cladocerans in a eutrophic
552 shallow lake during a shift in the phytoplankton dominance. *Ann. Limnol– - Int. J. Lim.* 45, 237–245.
553 <https://doi.org/10.1051/limn/2009027>

554 Almuktar, S.A.A.N., Abed, S.N., Scholz, M., 2018. Wetlands for wastewater treatment and subsequent
555 recycling of treated effluent: a review. *Environ Sci Pollut Res* 25, 23595–23623.
556 <https://doi.org/10.1007/s11356-018-2629-3>

557 Aminot, A., Kérouel, R., 2004. *Hydrologie des écosystèmes marins : paramètres et analyses*. Editions
558 Quae.

559 Amoros, C., 1984. *Introduction pratique à la systématique des organismes des eaux continentales*
560 *françaises-5. Crustacés Cladocères*. Publications de la Société Linnéenne de Lyon 53, 72–107.

561 Angeler, D.G., Moreno, J.M., 2007. Zooplankton community resilience after press-type anthropogenic
562 stress in temporary ponds. *Ecological Applications* 17, 1105–1115. <https://doi.org/10.1890/06-1040>

563 Arbach Leloup, F., Desroy, N., Le Mao, P., Pauly, D., Le Pape, O., 2008. Interactions between a natural
564 food web, shellfish farming and exotic species: The case of the Bay of Mont Saint Michel (France).
565 *Estuarine, Coastal and Shelf Science* 76, 111–120. <https://doi.org/10.1016/j.ecss.2007.06.011>

566 Atkinson, A., Shreeve, R., Pakhomov, E., Priddle, J., Blight, S., Ward, P., 1996. Zooplankton response
567 to a phytoplankton bloom near South Georgia, Antarctica. *Mar. Ecol. Prog. Ser.* 144, 195–210.
568 <https://doi.org/10.3354/meps144195>

569 Azim, M.E., Verdegem, M.C.J., Dam, A.A. van, Beveridge, M.C.M., 2005. Periphyton: Ecology,
570 Exploitation and Management. CABI. ISBN: 978-0-85199-097-2

571 Badsı, H., Ali, H.O., Loudiki, M., Aamiri, A., 2012. Phytoplankton Diversity and Community Composition
572 along the Salinity Gradient of the Massa Estuary. *American Journal of Human Ecology* 1, 58–64.

573 Baker, A.L., Baker, K.K., 1979. Effects of temperature and current discharge on the concentration and
574 photosynthetic activity of the phytoplankton in the upper Mississippi River. *Freshwater Biol* 9, 191–198.
575 <https://doi.org/10.1111/j.1365-2427.1979.tb01502.x>

576 Barbier M., Loreau, M. (2019) Pyramids and cascades: a synthesis of food chain functioning and
577 stability. *Ecology Letters*, 22 : 405-419. <https://doi.org/10.1111/ele.13196>

578 Berggreen, U., Hansen, B., Kiørboe, T., 1988. Food size spectra, ingestion and growth of the copepod
579 *Acartia tonsa* during development: Implications for determination of copepod production. *Mar. Biol.* 99,
580 341–352. <https://doi.org/10.1007/BF02112126>

581 Bleiwas, A.H., Stokes, P.M., 1985. Collection of large and small food particles by *Bosmina*. *Limnol.*
582 *Oceanogr.* 30, 1090–1092. <https://doi.org/10.4319/lo.1985.30.5.1090>

583 Bradley, P.B., Sanderson, M.P., Frischer, M.E., Brofft, J., Booth, M.G., Kerkhof, L.J., Bronk, D.A., 2010.
584 Inorganic and organic nitrogen uptake by phytoplankton and heterotrophic bacteria in the stratified Mid-
585 Atlantic Bight. *Estuarine, Coastal and Shelf Science* 88, 429–441.
586 <https://doi.org/10.1016/j.ecss.2010.02.001>

587 Bradshaw, T.M., Blake-Bradshaw, A.G., Fournier, A.M.V., Lancaster, J.D., O'Connell, J., Jacques, C.N.,
588 Eichholz, M.W., Hagy, H.M., 2020. Marsh bird occupancy of wetlands managed for waterfowl in the
589 Midwestern USA. *PLoS ONE* 15, 1–19. <https://doi.org/10.1371/journal.pone.0228980>

590 Buskey, E.J., Hyatt, C.J., 2006. Use of the FlowCAM for semi-automated recognition and enumeration
591 of red tide cells (*Karenia brevis*) in natural plankton samples. *Harmful Algae* 5, 685–692.
592 <https://doi.org/10.1016/j.hal.2006.02.003>

593 Cattrijsse, A., Hampel, H., 2006. *Marine Ecology Progress Series* 324:293. *Mar Ecol Prog Ser* 324, 293–
594 307.

595 Chaparro, G., O'Farrell, I., Hein, T., 2019. Multi-scale analysis of functional plankton diversity in
596 floodplain wetlands: Effects of river regulation. *Science of The Total Environment* 667, 338–347.
597 <https://doi.org/10.1016/j.scitotenv.2019.02.147>

598 Chenillat, F., Rivière, P., Ohman, M.D., 2021. On the sensitivity of plankton ecosystem models to the
599 formulation of zooplankton grazing. *PLoS ONE* 16, e0252033.
600 <https://doi.org/10.1371/journal.pone.0252033>

601 Cohen, N.R., 2022. Mixotrophic plankton foraging behaviour linked to carbon export. *Nat Commun* 13,
602 1–3. <https://doi.org/10.1038/s41467-022-28868-7>

603 Copping, A.E., Lorenzen, C.J., 1980. Carbon budget of a marine phytoplankton-herbivore system with
604 carbon-14 as a tracer. *Limnol. Oceanogr.* 25, 873–882. <https://doi.org/10.4319/lo.1980.25.5.0873>

605 Cordoba, J., Perez, E., Van Vlierberghe, M., Bertrand, A.R., Lupo, V., Cardol, P., Baurain, D., 2021. De
606 Novo Transcriptome Meta-Assembly of the Mixotrophic Freshwater Microalga *Euglena gracilis*. *Genes*
607 12, 842. <https://doi.org/10.3390/genes12060842>

608 Costalago, D., Forster, I., Nemcek, N., Neville, C., Perry, R.I., Young, K., Hunt, B.P.V., 2020. Seasonal
609 and spatial dynamics of the planktonic trophic biomarkers in the Strait of Georgia (northeast Pacific) and
610 implications for fish. *Sci Rep* 10, 8517. <https://doi.org/10.1038/s41598-020-65557-1>

611 Craft, C., 2000. Co-development of wetland soils and benthic invertebrate communities following salt
612 marsh creation. *Wetlands Ecology and Management* 8, 197–207.
613 <https://doi.org/10.1023/A:1008448620605>

614 D’Alelio, D., Libralato, S., Wyatt, T., Ribera d’Alcalà, M., 2016. Ecological-network models link diversity,
615 structure and function in the plankton food-web. *Sci Rep* 6, 21806. <https://doi.org/10.1038/srep21806>

616 Das, P., Kar, S., Das, U., Bimola, M., Kar, D., Aditya, G., 2020. Day time variations of zooplankton
617 species composition: observations from the wetlands of Assam, India. *Acta Limnol. Bras.* 32, e10.
618 <https://doi.org/10.1590/s2179-975x1418>

619 David, V., Tortajada, S., Savoye, N., Breret, M., Lachaussée, N., Philippine, O., Robin, F.-X., Dupuy,
620 C., 2020. Impact of human activities on the spatio-seasonal dynamics of plankton diversity in drained
621 marshes and consequences on eutrophication. *Water Research* 170, 115287.
622 <https://doi.org/10.1016/j.watres.2019.115287>

623 David, V., Selleslagh, J., Nowaczyk, A., Dubois, S., Bachelet, G., Blanchet, H., Gouillieux, B., Lavesque,
624 N., Leconte, M., Savoye, N., Sautour, B., Lobry, J., 2016. Estuarine habitats structure zooplankton
625 communities: Implications for the pelagic trophic pathways. *Estuarine, Coastal and Shelf Science* 179,
626 99–111. <https://doi.org/10.1016/j.ecss.2016.01.022>

627 David, V., Sautour, B., Galois, R., Chardy, P., 2006. The paradox high zooplankton biomass–low vegetal
628 particulate organic matter in high turbidity zones: What way for energy transfer? *Journal of Experimental*
629 *Marine Biology and Ecology* 333, 202–218. <https://doi.org/10.1016/j.jembe.2005.12.045>

630 Dexter, E., Katz, S.L., Bollens, S.M., Rollwagen-Bollens, G., Hampton, S.E., 2020. Modeling the trophic
631 impacts of invasive zooplankton in a highly invaded river. *PLoS ONE* 15, e0243002.
632 <https://doi.org/10.1371/journal.pone.0243002>

633 Didenko, A., Buzevych, I., Volikov, Y., Kruzhylina, S., Gurbyk, A., 2020. Population dynamics and
634 feeding ecology of the invasive Caucasian dwarf goby, *Knipowitschia caucasica*, in a freshwater habitat
635 in Ukraine. *Knowl. Manag. Aquat. Ecosyst.* 421, 1–12. <https://doi.org/10.1051/kmae/2020018>

636 Dupuy, C., Talarmin, A., Hartmann, H.J., Delmas, D., Courties, C., Marquis, E., 2011. Community
637 structure and grazing of the nano-microzooplankton on the continental shelf of the Bay of Biscay.
638 *Estuarine, Coastal and Shelf Science* 95, 1–13. <https://doi.org/10.1016/j.ecss.2011.05.002>

639 Ersanli, E., Gönülol, A., 2006. A study on the phytoplankton of Lake Simenit, Turkey. *Cryptogamie, Algol.*
640 23, 289–305.

641 Esteban, G.F., Fenchel, T., Finlay, B.J., 2010. Mixotrophy in Ciliates. *Protist* 161, 621–641.
642 <https://doi.org/10.1016/j.protis.2010.08.002>

643 Galir Balkić, A., Ternjej, I., Špoljar, M., 2018. Hydrology driven changes in the rotifer trophic structure
644 and implications for food web interactions. *Ecohydrology* 11, e1917. <https://doi.org/10.1002/eco.1917>

645 Ghermandi, A., Nunes, P.A.L.D., 2013. A global map of coastal recreation values: Results from a
646 spatially explicit meta-analysis. *Ecological Economics* 86, 1–15.
647 <https://doi.org/10.1016/j.ecolecon.2012.11.006>

648 Gliwicz, M.Z., 1986. Predation and the evolution of vertical migration in zooplankton. *Nature* 320, 746–
649 748. <https://doi.org/10.1038/320746a0>

650 Grujicic, V., Nuy, J.K., Salcher, M.M., Shabarova, T., Kasalicky, V., Boenigk, J., Jensen, M., Simek, K.,
651 2018. Cryptophyta as major bacterivores in freshwater summer plankton. *ISME J* 12, 1668–1681.
652 <https://doi.org/10.1038/s41396-018-0057-5>

653 Guillou, P., 1997. La communauté salicole du bas-pays guérandais ou les interrelations entre l'homme
654 et son environnement naturel. Université Marc Bloch (Strasbourg) (1971-2008).

655 Harris, G., 2012. *Phytoplankton ecology: structure, function and fluctuation*. Springer Science &
656 Business Media.

657 Hébert, M.-P., Beisner, B.E., Maranger, R., 2017. Linking zooplankton communities to ecosystem
658 functioning: toward an effect-trait framework. *J. Plankton Res.* 39, 3–12.
659 <https://doi.org/10.1093/plankt/fbw068>

660 Henry Ogbuagu, D., Abeke Ayoade, A., 2012. Seasonal Dynamics in Plankton Abundance and Diversity
661 of a Freshwater Body in Etche, Nigeria. *ENRR* 2, p48. <https://doi.org/10.5539/enrr.v2n2p48>

662 Hinkle, R.L., Mitsch, W.J., 2005. Salt marsh vegetation recovery at salt hay farm wetland restoration
663 sites on Delaware Bay. *Ecological Engineering* 25, 240–251.

664 Huppert, A., Blasius, B., Stone, L., 2002. A Model of Phytoplankton Blooms. *The American Naturalist*
665 159, 156–171. <https://doi.org/10.1086/324789>

666 Ives, A.R., Carpenter, S.R., 2007. Stability and Diversity of Ecosystems. *Science* 317, 58–62.
667 <https://doi.org/10.1126/science.1133258>

668 Jerabek, A., Darnell, K.M., Pellerin, C., Carruthers, T.J.B., 2017. Use of Marsh Edge and Submerged
669 Aquatic Vegetation as Habitat by Fish and Crustaceans in Degrading Southern Louisiana Coastal
670 Marshes. *Southeastern Geographer* 57, 212–230. <https://doi.org/10.1353/sgo.2017.0022>

671 Jia, J., Bai, J., Gao, H., Wen, X., Zhang, G., Cui, B., Liu, X., 2017. In situ soil net nitrogen mineralization
672 in coastal salt marshes (*Suaeda salsa*) with different flooding periods in a Chinese estuary. *Ecological*
673 *Indicators* 73, 559–565. <https://doi.org/10.1016/j.ecolind.2016.10.012>

674 Jia, J., Gao, Y., Sun, K., Lu, Y., Wang, J., Shi, K., 2022. Phytoplankton community composition, carbon
675 sequestration, and associated regulatory mechanisms in a floodplain lake system. *Environmental*
676 *Pollution* 306, 119411. <https://doi.org/10.1016/j.envpol.2022.119411>

677 Joyeux, E., Carpentier, A., Corre, F., Haie, S., Pétilion, J., 2017. Impact of salt-marsh management on
678 fish nursery function in the bay of Aiguillon (French Atlantic coast), with a focus on European sea bass
679 diet. *J Coast Conserv* 21, 435–444. [https://doi.org/DOI 10.1007/s11852-017-0501-0](https://doi.org/DOI%2010.1007/s11852-017-0501-0)

680 Kaneko, S., Kanou, K., Sano, M., 2019. Comparison of fish assemblage structures among microhabitats
681 in a salt marsh in Lake Hinuma, eastern Japan. *Fish Sci* 85, 113–125. [https://doi.org/10.1007/s12562-](https://doi.org/10.1007/s12562-018-1269-3)
682 [018-1269-3](https://doi.org/10.1007/s12562-018-1269-3)

683 Kimmel, D.G., McGlaughon, B.D., Leonard, J., Paerl, H.W., Taylor, J.C., Cira, E.K., Wetz, M.S., 2015.
684 Mesozooplankton abundance in relation to the chlorophyll maximum in the Neuse River Estuary, North
685 Carolina, USA: Implications for trophic dynamics. *Estuarine, Coastal and Shelf Science* 157, 59–68.
686 <https://doi.org/10.1016/j.ecss.2015.02.014>

687 Kirchman, D.L., 1994. The uptake of inorganic nutrients by heterotrophic bacteria. *Microb Ecol* 28, 255–
688 271. <https://doi.org/10.1007/BF00166816>

689 Kleppel, G., 1993. On the diets of calanoid copepods. *Mar. Ecol. Prog. Ser.* 99, 183–195.
690 <https://doi.org/10.3354/meps099183>

691 Kobayashi, T., Ryder, D.S., Gordon, G., Shannon, I., Ingleton, T., Carpenter, M., Jacobs, S.J., 2009.
692 Short-term response of nutrients, carbon and planktonic microbial communities to floodplain wetland
693 inundation. *Aquat Ecol* 43, 843–858. <https://doi.org/10.1007/s10452-008-9219-2>

694 Koch, F., Gobler, C.J., 2009. The Effects of Tidal Export from Salt Marsh Ditches on Estuarine Water
695 Quality and Plankton Communities. *Estuaries and Coasts* 32, 261–275. [https://doi.org/10.1007/s12237-
696 008-9123-y](https://doi.org/10.1007/s12237-008-9123-y)

697 Koslow, J.A., Pesant, S., Feng, M., Pearce, A., Fearn, P., Moore, T., Matear, R., Waite, A., 2008. The
698 effect of the Leeuwin Current on phytoplankton biomass and production off Southwestern Australia. *J.*
699 *Geophys. Res.* 113, C07050. <https://doi.org/10.1029/2007JC004102>

700 Landry, M.R., Peterson, W.K., Lorenzen, C.J., 1995. Zooplankton grazing, phytoplankton growth, and
701 export flux: inferences from chlorophyll tracer methods. *ICES Journal of Marine Science* 52, 337–345.
702 [https://doi.org/10.1016/1054-3139\(95\)80049-2](https://doi.org/10.1016/1054-3139(95)80049-2)

703 Landry, M.R., Calbet, A., 2004. Microzooplankton production in the oceans. *ICES Journal of Marine*
704 *Science* 61, 501–507. <https://doi.org/10.1016/j.icesjms.2004.03.011>

705 Legendre, L., Rassoulzadegan, F., 1995. Plankton and nutrient dynamics in marine waters. *Ophelia* 41,
706 153–172. <https://doi.org/10.1080/00785236.1995.10422042>

707 Lorenzen, C.J., 1966. A method for the continuous measurement of in vivo chlorophyll concentration.
708 *Deep Sea Research and Oceanographic Abstracts* 13, 223–227. [https://doi.org/10.1016/0011-
709 7471\(66\)91102-8](https://doi.org/10.1016/0011-7471(66)91102-8)

710 Maltby, E., 1991. Wetland management goals: wise use and conservation. *Landscape and Urban*
711 *Planning* 20, 9–18. [https://doi.org/10.1016/0169-2046\(91\)90085-Z](https://doi.org/10.1016/0169-2046(91)90085-Z)

712 Marie, D., Brussaard, C., Partensky, F., Vaulot, D., Wiley, J., 1999. Flow cytometric analysis of
713 phytoplankton, bacteria and viruses. *Current protocols in cytometry* 11, 1–15.

714 Masclaux, H., Tortajada, S., Philippine, O., Robin, F.-X., Dupuy, C., 2015. Planktonic food web structure
715 and dynamic in freshwater marshes after a lock closing in early spring. *Aquat Sci* 77, 115–128.
716 <https://doi.org/10.1007/s00027-014-0376-1>

717 McEdward, L., 2020. Ecology of Marine Invertebrate Larvae. CRC Press.

718 Milstein, A., Valdenberg, A., Harpaz, S., 2006. Fish larvae – zooplankton relationships in microcosm
719 simulations of earthen nursery ponds. I. Freshwater system. *Aquacult Int* 14, 231–246.
720 <https://doi.org/10.1007/s10499-005-9006-1>

721 Morrissey, E.M., Gillespie, J.L., Morina, J.C., Franklin, R.B., 2014. Salinity affects microbial activity and
722 soil organic matter content in tidal wetlands. *Glob Change Biol* 20, 1351–1362.
723 <https://doi.org/10.1111/gcb.12431>

724 Nalubega, M., 1999. Water Balance and Hydrodynamics of the Nakivubo Swamp, in: *Wastewater
725 Treatment by a Natural Wetland: The Nakivubo Swamp, Uganda*. CRC Press.

726 Navarro-Barranco, C., Tierno-de-Figueroa, J.M., Guerra-García, J.M., Sánchez-Tocino, L., García-
727 Gómez, J.C., 2013. Feeding habits of amphipods (Crustacea: Malacostraca) from shallow soft bottom
728 communities: Comparison between marine caves and open habitats. *Journal of Sea Research* 78, 1–7.
729 <https://doi.org/10.1016/j.seares.2012.12.011>

730 Nordström, M., Currin, C., Talley, T., Whitcraft, C., Levin, L., 2014. Benthic food-web succession in a
731 developing salt marsh. *Mar. Ecol. Prog. Ser.* 500, 43–55. <https://doi.org/10.3354/meps10686>

732 Norlin, J.I., Bayley, S.E., Ross, L.C.M., 2006. Zooplankton Composition and Ecology in Western Boreal
733 Shallow-water Wetlands. *Hydrobiologia* 560, 197–215. <https://doi.org/10.1007/s10750-005-1185-2>

734 Nyman, J.A., 2011. Ecological Functions of Wetlands, in: LePage, B.A. (Ed.), *Wetlands: Integrating
735 Multidisciplinary Concepts*. Springer Netherlands, Dordrecht, pp. 115–128. [https://doi.org/10.1007/978-
736 94-007-0551-7_6](https://doi.org/10.1007/978-94-007-0551-7_6)

737 Pagès, J., 2002. Analyse factorielle multiple appliquée aux variables qualitatives et aux données mixtes.
738 *Revue de statistique appliquée* 50, 5–37.

739 Pennekamp, F., Pontarp, M., Tabi, A., Altermatt, F., Alther, R., Choffat, Y., Fronhofer, E.A.,
740 Ganesanandamoorthy, P., Garnier, A., Griffiths, J.I., Greene, S., Horgan, K., Massie, T.M., Mächler, E.,
741 Palamara, G.M., Seymour, M., Petchey, O.L., 2018. Biodiversity increases and decreases ecosystem
742 stability. *Nature* 563, 109–112. <https://doi.org/10.1038/s41586-018-0627-8>

743 Peter Sheng, Y., Paramygin, V.A., Rivera-Nieves, A.A., Zou, R., Fernald, S., Hall, T., Jacob, K., 2022.
744 Coastal marshes provide valuable protection for coastal communities from storm-induced wave, flood,
745 and structural loss in a changing climate. *Sci Rep* 12, 3051. [https://doi.org/10.1038/s41598-022-06850-
746 z](https://doi.org/10.1038/s41598-022-06850-)

747 Pétillon, J., Potier, S., Carpentier, A., Garbutt, A., 2014. Evaluating the success of managed realignment
748 for the restoration of salt marshes: Lessons from invertebrate communities. *Ecological Engineering* 69,
749 70–75. <https://doi.org/10.1016/j.ecoleng.2014.03.085>

750 Postel, L., Fock, H., Hagen, W., 2000. Biomass and abundance, in: *ICES Zooplankton Methodology*
751 *Manual*. Elsevier, pp. 83–192. <https://doi.org/10.1016/B978-012327645-2/50005-0>

752 Quintana, X.D., Comián, F.A., Moreno-Amich, R., 1998. Nutrient and plankton dynamics in a
753 Mediterranean salt marsh dominated by incidents of flooding. Part 2: Response of the zooplankton
754 community to disturbances. *J Plankton Res* 20, 2109–2127. <https://doi.org/10.1093/plankt/20.11.2109>

755 Rahmati, R., Fereidouni, A.E., Rouhi, A., Agh, N., 2020. Effects of different diets on population growth
756 and fatty acids composition in cyclopoid copepod, *Acanthocyclops trajani* (Mirabdullayev and Defaye,
757 2002): A potential supplementary live food for freshwater fish larvae. *Iranian Journal of Fisheries*
758 *Sciences* 19, 1447–1462. DOI: 10.22092/ijfs.2019.120729

759 Ratnam, K., Limna Mol, V.P., Venkatnarayanan, S., Jha, D.K., Dharani, G., Devi, P., 2022. Seasonal
760 variations influencing the abundance and diversity of plankton in the Swarnamukhi River Estuary,
761 Nellore, India. *J. Threat. Taxa* 14, 20615–20624. <https://doi.org/10.11609/jott.7663.14.2.20615-20624>

762 Reissig, M., Trochine, C., Queimaliños, C., Balseiro, E., Modenutti, B., 2006. Impact of fish introduction
763 on planktonic food webs in lakes of the Patagonian Plateau. *Biological Conservation* 132, 437–447.
764 <https://doi.org/10.1016/j.biocon.2006.04.036>

765 Rieper, M., 1982. Feeding Preferences of Marine Harpacticoid Copepods for Various Species of
766 Bacteria. *Mar. Ecol. Prog. Ser.* 7, 303–307. <https://doi.org/10.3354/meps007303>

767 Robertson, A.L., 1988. Life histories of some species of Chydoridae (Cladocera: Crustacea). *Freshwater*
768 *Biol* 20, 75–84. <https://doi.org/10.1111/j.1365-2427.1988.tb01719.x>

769 Rudy, M., McDonnell, K., Valiela, I., Foreman, K., 1994. Dissolved Organic Nitrogen in Groundwater
770 Bordering Estuaries of Waquoit Bay, Massachusetts: Relations with Watershed Landscape Mosaics.
771 *The Biological Bulletin* 187, 278–279. <https://doi.org/10.1086/BBLv187n2p278>

772 Santos-Medrano, G.E., Robles-Vargas, D., Hernández-Flores, S., Rico-Martínez, R., 2017. Life table
773 demography of *Asplanchna brightwellii* Gosse, 1850 fed with five different prey items. *Hydrobiologia*
774 796, 169–179. <https://doi.org/10.1007/s10750-016-3069-z>

775 Semyalo, R., Nattabi, J.K., Larsson, P., 2009. Diel Vertical Migration of zooplankton in a eutrophic bay
776 of Lake Victoria. *Hydrobiologia* 635, 383–394. <https://doi.org/10.1007/s10750-009-9931-5>

777 Stanachkova, M., Stefanova, M., Kozuharov, D., Raikova-Petrova, G., Fikovska, E., 2017. Community
778 structure of zooplankton as key factor for self-purification capacity of iskar reservoir. *Ecological*
779 *Engineering and Environment Protection* 9, 39–46.

780 Stephenson, T.D., 1990. Fish Reproductive Utilization of Coastal Marshes of Lake Ontario Near Toronto.
781 *J. Great Lakes Res.* 16, 71–81.

782 Sullivan, M.J., Currin, C.A. (2002). Community Structure and Functional Dynamics of Benthic
783 Microalgae in Salt Marshes. In: Weinstein, M.P., Kreeger, D.A. (eds) *Concepts and Controversies in*
784 *Tidal Marsh Ecology*. Springer, Dordrecht. https://doi.org/10.1007/0-306-47534-0_6

785 Tackx, M.L.M., Herman, P.J.M., Gasparini, S., Irigoien, X., Billiones, R., Daro, M.H., 2003. Selective
786 feeding of *Eurytemora affinis* (Copepoda, Calanoida) in temperate estuaries: model and field
787 observations. *Estuarine, Coastal and Shelf Science* 56, 305–311. [https://doi.org/10.1016/S0272-](https://doi.org/10.1016/S0272-7714(02)00182-8)
788 [7714\(02\)00182-8](https://doi.org/10.1016/S0272-7714(02)00182-8)

789 Tałanda, J., Maszczyk, P., Babkiewicz, E., Rutkowska, K., Ślusarczyk, M., 2022. The short-term effects
790 of planktivorous fish foraging in the presence of artificial light at night on lake zooplankton. *Journal of*
791 *Plankton Research* 00, 1–5. <https://doi.org/10.1093/plankt/fbac046>

792 Thakur, R.K., Jindal, R., Singh, U.B., Ahluwalia, A.S., 2013. Plankton diversity and water quality
793 assessment of three freshwater lakes of Mandi (Himachal Pradesh, India) with special reference to
794 planktonic indicators. *Environ Monit Assess* 185, 8355–8373. [https://doi.org/10.1007/s10661-013-3178-](https://doi.org/10.1007/s10661-013-3178-3)
795 [3](https://doi.org/10.1007/s10661-013-3178-3)

796 Timms, R.M., Moss, B., 1984. Prevention of growth of potentially dense phytoplankton populations by
797 zooplankton grazing, in the presence of zooplanktivorous fish, in a shallow wetland ecosystem: Plankton
798 grazing and predation. *Limnol. Oceanogr.* 29, 472–486. <https://doi.org/10.4319/lo.1984.29.3.0472>

799 Trombetta, T., Vidussi, F., Mas, S., Parin, D., Simier, M., Mostajir, B., 2019. Water temperature drives
800 phytoplankton blooms in coastal waters. *PLoS ONE* 14, e0214933.
801 <https://doi.org/10.1371/journal.pone.0214933>

802 Utermöhl H., 1958. Zur Vervollkommung der quantitativen Phytoplankton Methodik. *Mitteilung*
803 *Internationale Vereinigung für Theoretische und Angewandte Limnologie* 9, 1-38

804 Van den Meersche, K.V., Rijswijk, P.V., Soetaert, K., Middelburg, J.J., 2009. Autochthonous and
805 allochthonous contributions to mesozooplankton diet in a tidal river and estuary: Integrating carbon

806 isotope and fatty acid constraints. *Limnol. Oceanogr.* 54, 62–74.
807 <https://doi.org/10.4319/lo.2009.54.1.0062>

808 Verhoeven, J., Arheimer, B., Yin, C., Hefting, M., 2006. Regional and global concerns over wetlands
809 and water quality. *Trends in Ecology & Evolution* 21, 96–103. <https://doi.org/10.1016/j.tree.2005.11.015>

810 Vernberg, F.J., 1993. Salt-marsh processes: A Review. *Environ Toxicol Chem* 12, 2167–2195.
811 <https://doi.org/10.1002/etc.5620121203>

812 Volterra V. (1928) Variations and fluctuations of the number of individuals in animal species living
813 together. *ICES J. Marine Sci.*, 3: 3-51

814 Walkusz, W., Paulić, J.E., Kwaśniewski, S., Williams, W.J., Wong, S., Papst, M.H., 2010. Distribution,
815 diversity and biomass of summer zooplankton from the coastal Canadian Beaufort Sea. *Polar Biol* 33,
816 321–335. <https://doi.org/10.1007/s00300-009-0708-0>

817 Więski, K., Guo, H., Craft, C.B., Pennings, S.C., 2010. Ecosystem Functions of Tidal Fresh, Brackish,
818 and Salt Marshes on the Georgia Coast. *Estuaries and Coasts* 33, 161–169.
819 <https://doi.org/10.1007/s12237-009-9230-4>

820 Woodward, R.T., Wui, Y.-S., 2001. The economic value of wetland services: a meta-analysis. *Ecological*
821 *Economics* 37, 257–270. [https://doi.org/10.1016/S0921-8009\(00\)00276-7](https://doi.org/10.1016/S0921-8009(00)00276-7)

822 Yan, Z., Shen, T., Li, W., Cheng, W., Wang, X., Zhu, M., Yu, Q., Xiao, Y., Yu, L., 2021. Contribution of
823 microalgae to carbon sequestration in a natural karst wetland aquatic ecosystem: An in-situ mesocosm
824 study. *Science of The Total Environment* 768, 144387. <https://doi.org/10.1016/j.scitotenv.2020.144387>