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Review

# Marine Natural Products from Tunicates and Their Associated Microbes

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**Abstract:** Marine tunicates are identified as a potential source of marine natural products (MNPs), demonstrating a wide range of biological properties, like antimicrobial and anticancer activities. The symbiotic relationship between tunicates and specific microbial groups has revealed the acquisition of microbial compounds by tunicates for defensive purpose. For instance, yellow pigmented compounds, “tambjamines”, produced by the tunicate, *Sigillina signifera* (Sluiter, 1909), primarily originated from their bacterial symbionts, which are involved in their chemical defense function, indicating the ecological role of symbiotic microbial association with tunicates. This review has garnered comprehensive literature on MNPs produced by tunicates and their symbiotic microbionts. Various sections covered in this review include tunicates’ ecological functions, biological activities, such as antimicrobial, antitumor, and anticancer activities, metabolic origins, utilization of invasive tunicates, and research gaps. Apart from the literature content, 20 different chemical databases were explored to identify tunicates-derived MNPs. In addition, the management and exploitation of tunicate resources in the global oceans are detailed for their ecological and biotechnological implications.

**Keywords:** tunicates; symbiotic microbes; pigments; bioactive compounds; alkaloids & peptides



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## 1. Introduction

Tunicates and sea squirts are soft-bodied solitary or colonial (60%) sessile marine organisms belonging to the family Ascidiacea under the subphylum Urochordata, phylum Chordata [1,2]. These organisms are hermaphroditic, filter feeders, and appear in different body colors, such as translucent to blue, green, yellow, red, and brown, with a life span ranging from two months to one year [1–4]. Currently, tunicates are classified into four major clades such as (a) Appendicularia, (b) Thaliacea + Phlebobranchia + Aplousobranchia, (c) Molgulidae, and (d) Styelidae + Pyuridae, on the basis of the phylogenomic transcriptomic approach [5]. Globally, around 2815 tunicate species have been recorded from shallow coastal waters to deep waters [1]. Tunicate larvae resemble tadpole larvae of members of Chordata, but soon after the retrogressive metamorphosis, they lose the notochord and post-anal tail; thus, these organisms are often referred to as the “evolutionary connecting link” between invertebrates and chordates [6,7]. Therefore, tunicates are considered as important model organisms for several research aspects, such as evolution [6], development biology [8,9], invasion success [10], and bioactive compounds.

Tunicates are prolific producers of marine natural products (MNPs), and certain species are also known to release toxins, such as Bistramide A [11,12]. However, a few species, like *Halocynthia roretzi* and *Pyura michaelseni*, are eaten in southeast Asian countries like Korea [13,14]. The strong immune defensive system [15] and their associated symbiotic microbes with bioactive properties [16], makes tunicates highly preferential drug resources in the ocean [15,17]. Since the majority of the tunicate species are known to produce MNP's, attempts are being undertaken in the culturing of these tunicates (e.g., mangrove tunicate *Ecteinascidia turbinata*) in large scale for various applications [18,19]. The process of accumulation of vanadium by vanadocytes of tunicates from seawater is well-known [20]. In contrast, investigations on the acquisition of MNPs by tunicates from their symbiotic bacteria are very limited, except for the antitumor products ecteinascidins [21,22], didemnin [23], and talaropeptides [24]. A recent review highlighted the association of bacteria, actinomycetes, fungi, and cyanobacteria with the tunicates and their bioactive nature [25]. It was also observed that actinomycetes, fungi, and bacteria are the predominant microbes associated with the tunicates, showing cytotoxic and antimicrobial activities [26], with the production of alkaloids as the major source of MNPs [27]. In this context, this review aimed to provide the chemical profiles of various tunicates and their associated microbes for biotechnological and drug development applications.

## 2. Ecological Importance of Tunicates

The tunicates population plays an important role in the marine food web through filter feeding [4]. Earlier studies have suggested that phytoplankton productivity in a shallow fjord is controlled by the tunicates population [28]. Tunicates are known to trap the sinking particulate organic matter and generate mucus rich organic matter and fecal pellets with carbohydrates and minerals [29,30], thereby triggering the downward biogeochemical flux (e.g., carbon flux) patterns from surface to deep waters [29,31,32]. Some obligate photosymbiotic tunicates have been suggested to act as environmental stress indicators [33]. The unknown ecological functions of a few tunicate MNPs [34] in understanding their ecological role is yet to be understood.

## 3. Database Search on Tunicate MNPs

Twenty different public chemical databases such as BIAdb, BindingDB, ChemDB, ChEMBL, ChemSpider, DrugBank, HIT, HMDB, KEGG, NCI, NPACT, PDB-Bind, PDBChem, PharmaGKB, PubChem, SMPDB, SuperDrug, TTD, UNIProt, and ZINC were explored to identify the tunicate-originated MNPs deposited in these databases. The chemical constituents identified from these databases using the search keywords “tunicate and ascidian” are listed in Table 1.

**Table 1.** List of MNPs originated from tunicates available in various public databases. The unknown compound records are excluded from the list.

Database	No. of Known Compounds	No. of Unknown Compounds	Known Chemical Compound	Biological Properties
BIAdb	1	-	Polycarpine	Cytotoxic, antiviral, and antifungal
BindingDB	2	-	Tuberatolides, Sodium 1-(12-hydroxy) octadecanyl sulfate	Farnesoid X receptor antagonists, matrix metalloproteinase 2 inhibitor
ChemDB	2	-	Patellazole B, Patellazole C	Antimicrobial, cytotoxic
ChEMBL	2	-	Ascidiidemin, Trabectedin	Anticancer
ChemSpider	1	-	Trabectedin	Anticancer

Table 1. Cont.

Database	No. of Known Compounds	No. of Unknown Compounds	Known Chemical Compound	Biological Properties
DrugBank	-	-		
HIT	-	-		
HMDB	1	-	Trabectedin	Anticancer
KEGG	1	-	Trabectedin	Anticancer
NCI	-	-		
NPACT	-	-		
PDB-Bind	-	-		
PDBeChem	16	>30	Cystodytin D, cystodytin F, cystodytin E, cystodytin G, cystodytin H, cystodytin I, Diplosoma ylidene 1, Diplosoma ylidene 2, Lejimalide A, lejimalide B, lissoclibadin 1, lissoclibadin 2, lissoclibadin 3, lamellarin alpha 20-sulfate, plitidepsin, trabectedin	Cytotoxic, anticancer
PharmaGKB	1	-	Trabectedin	Anticancer
PubChem	4	2	Patellazole B, Patellazole C, GnRH-II, GnRH-I	Antimicrobial and cytotoxic, induces spawning
SMPDB	-	-		
SuperDrug	1	-	Trabectedin	Anticancer
TTD	-	-		
UniProt	1	1	Retinoic acid	Regeneration of gut
ZINC	1	-	Trabectedin	Anticancer

**Foot note:** Table 1 data are garnered from public chemical databases listed in the main text part 3, but not from the literature. That is why there are no references cited in this table. Readers are asked to refer to Tables 2 and 3 where details are from the literature, and therefore, references are cited.

#### 4. Profile of MNPs from Tunicates and Associated Microbes

Tunicates are known to produce a wide range of MNPs with various bioactive properties (Tables 2 and 3). These organisms are considered as a rich source of cellulose, which varies with different species [35]. Alkaloids and peptides are the major chemical constituents observed in tunicates [36]. Metabolites originated from tunicate hemocytes are also found to be cytotoxic to foreign particles [37] and various cell lines [38]. Microorganisms associated with the invertebrate hosts have also been identified as a source of bioactive metabolites [39]. In fact, bioactive metabolite-producing invertebrate-associated microorganisms have special implications in solving the “supply problem” in the initial steps of drug discovery [40]. Recently, Chen et al. reviewed the biological and chemical diversity of ascidian-associated microorganisms [41].

**Table 2.** Chemical profiles from body parts and fluids of few tunicate species.

Body Component	Chemical Compound	Function	Application	Reference
Tunic ( <i>Ascidia</i> sp., <i>Ciona intestinalis</i> , <i>Halocynthia roretzi</i> , and <i>Styela plicata</i> )	Tunicin (cellulose)	Protection	Material cellulose	[35]
Blood ( <i>Ascidia nigra</i> , <i>Molgula manhattensis</i> )	Vanadium	Oxygen transport		[42]
Blood ( <i>Ascidia nigra</i> )	Tunichromes	Vanadium binding and reduces blood pigments		[42,43]
Hemocytes ( <i>Styela clava</i> )	Clavanins	Multiple functions	Antimicrobial	[44,45]
Hemocytes ( <i>Halocynthia papillosa</i> )	Halocytin and papillosin		Antimicrobial	[46]
Hemocytes ( <i>Halocynthia aurantium</i> )	Halocidin		Antimicrobial	[47]
Gonad (Unknown sp.)	GnRH-2 peptide	Pheromone-like function	Induce spawning	[48]

Microbes associated with tunicates have been found to produce potential metabolites showing antimicrobial and anticancer activities (Figures 1–3 and Table 3). Tunicate-associated bacteria such as *Bacillus*, *Pantoea*, *Pseudoalteromonas*, *Salinicola*, *Streptomyces*, *Vibrio* and *Virgibacillus* have recently been identified with potential antimicrobial activities [16]. The introduced tunicate species are also reported to harbor diverse host-specific microbial populations [49] that produce species-specific metabolites [50]. In general, tunicate associated bacteria and fungi are known to produce a variety of MNPs with various biological properties [41]. The chemistry of yellow pigment-producing parasitic bacteria in the interstitial and blood-filled spaces of planktonic tunicates, *Oikopleura vanhoeffeni* and *Oikopleura dioica*, are yet to be characterized [51].

**Table 3.** Bioactive compounds from various species of tunicates and their associated microbes.

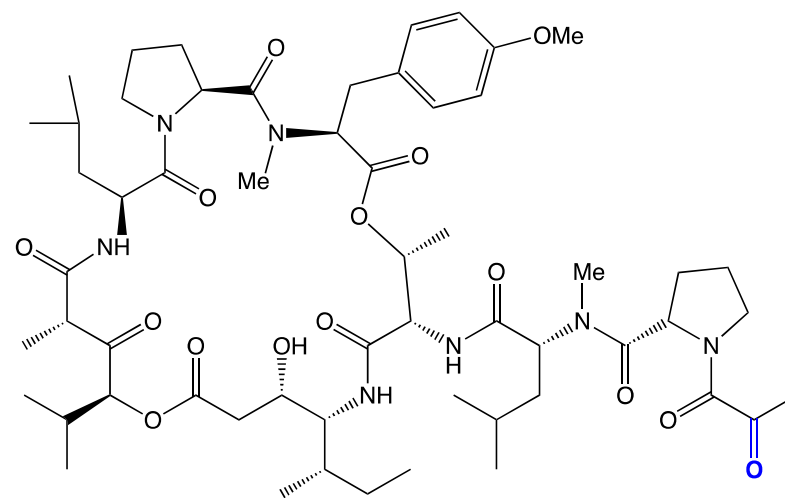
MNPs from Tunicates	Chemical Compound	Function	Application	Reference
<i>Aplidium albicans</i>	Aplidin		Anticancer	[52,53]
<i>Aplidium albicans</i>	Dehydrodidemnin B		Antitumor	[54]
<i>Aplidium glabrum</i>	Quinones		Anticancer, cytotoxic	[55]
<i>Aplidium haouarianum</i>	Haouamine A		Cytotoxic activity	[56]
<i>Aplidium meridianum</i>	Meridianins		Anticancer, antibiofilm	[57]
<i>Aplidium</i> & <i>Synoicum</i>	Meridianins	Feeding deterrents	Antibacterial	[58]
<i>Atapozoa</i> sp.	Tambjamine	Feeding deterrents		[59]
<i>Botryllus tuberatus</i>	Tuberatolides		Farnesoid X receptor antagonists	[60]
<i>Clavelina lepadiformis</i>	Lepadins and villatamines		Antiparasitic, anticancer	[61]
<i>Clavelina picta</i>	Clavepictine A and B		Antimicrobial, cytotoxicity	[62]
<i>Cynthia savignyi</i>	Cynthichlorine		Antifungal, cytotoxicity	[63]
<i>Cystodytes dellechiaiei</i>	Cystodytins A-I		Antitumor, cytotoxic	[64,65]
<i>Cystodytes dellechiaiei</i>	Ascididemin		Antitumor	[66]
<i>Cystodytes</i> sp.	Ascididemin	Feeding deterrents	Antifeedant	[67]

Table 3. Cont.

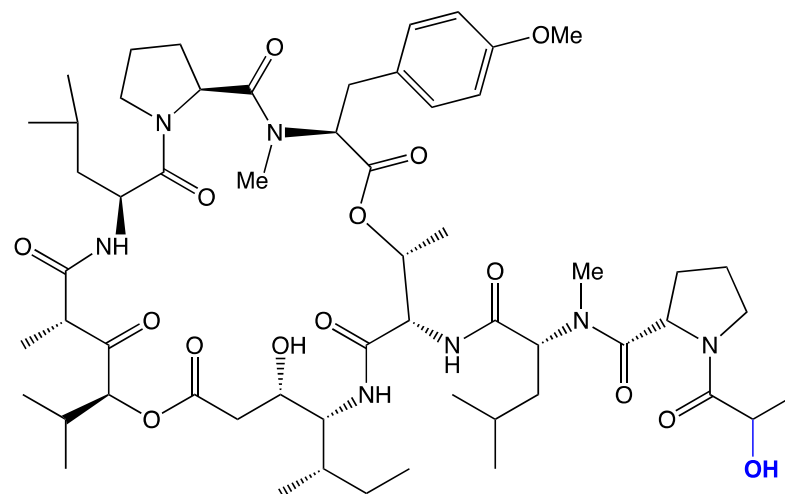
MNPs from Tunicates	Chemical Compound	Function	Application	Reference
Didemnidae	Mellpaladine and dopargimine		Neuroactive	[68]
Didemnidae	Siladenoserinols A and B		Antitumor	[69]
Didemnidae	Sameuramide A		Colony formation	[70]
<i>Didemnum</i> sp.	Lepadins D-F		Antiplasmodial and antitrypanosomal	[71]
<i>Didemnum guttatum</i>	Cyclodidemniserinol trisulfate		Anti-retroviral	[72]
<i>Didemnum granulatum</i>	Granulatamides		Deterrent activity	[73]
<i>Didemnum molle</i>	Lanthipeptide divamide A		anti-HIV drug	[74]
<i>Didemnum molle</i>	Mollamide B		Anticancer	[75]
<i>Didemnum proliferum</i>	Shishijimicins		Antitumor	[76]
<i>Didemnum psammotodes</i>	Methyl esters		Antiproliferative	[77]
<i>Didemnum ternerratum</i>	Lamellarin Sulfates		Anticancer	[78]
<i>Diplosoma</i> sp.	Diplamine		Antibacterial and cytotoxic	[79]
<i>Diplosoma virens</i>	Diplosoma ylidene 1, Diplosoma ylidene 2		Anticancer	[80]
<i>Ecteinascidia turbinata</i>	Ecteinascidin 743 (Trabectedin)		Anticancer	[81]
<i>Eudistoma gilboverde</i>	Methyleudistomins		Antitumor	[82]
<i>Eudistoma olivaceum</i>	Eudistomins G and H	Chemical defense	Antifouling	[34]
<i>Eudistoma olivaceum</i>	Eudistomins A, D, G, H, I, J, M, N, O, P, and Q		Antiviral	[83]
<i>Eudistoma olivaceum</i>	Eudistomins C, E, K, and L		Antiviral	[84]
<i>Eudistoma vancouveri</i>	7-Oxostaurosporine		Anticancer	[85]
<i>Eudistoma viride</i>	Eudistomins H		Anticancer	[86]
<i>Eusynstyela latericius</i>	Eusynstyelamides A, B		Antibacterial	[87]
<i>Eusynstyela tinctoria</i>	Kuanoniamine A	Chemical defense	Antimicrobial, antitumor, antifouling	[88]
<i>Halocynthia aurantium</i>	Halocidin		Antimicrobial	[47]
<i>Halocynthia papillosa</i>	Halocytin and papillosin		Antimicrobial	[46]
<i>Halocynthia roretzi</i>	Lumichrome	Larval metamorphosis		[89]
<i>Halocynthia roretzi</i>	Halocytamine A and B		Antimicrobial, anticancer	[90]
<i>Lissoclinum</i> cf. <i>badium</i>	Lissoclibadins		Anticancer	[91]
<i>Lissoclinum fragile</i>			Antimicrobial, hemolytic, and cytotoxic	[92]
<i>Lissoclinum patella</i>	Patellazole B and C		Antimicrobial, cytotoxic	[93,94]
<i>Phallusia nigra</i>	Vanadium chloride, vanadyl sulfate		Antimicrobial	[95]

Table 3. Cont.

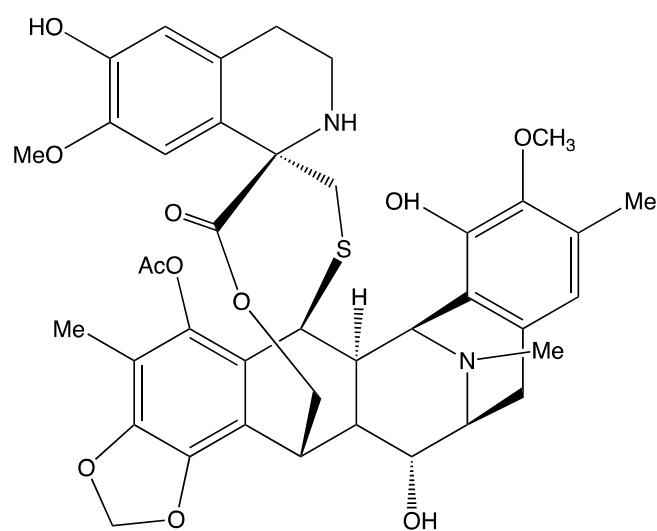
MNPs from Tunicates	Chemical Compound	Function	Application	Reference
<i>Polycarpa aurata</i>	Polyaurines A and B		Antiparasitic	[96]
<i>Polycarpa clavata</i>	Polycarpine dihydrochloride		Cytotoxic	[97]
<i>Polycarpa clavata</i>	Polycarpaurines A and C		Antiviral, antifungal	[98]
Polyclinidae	Sodium 1-(12-hydroxy) octadecanyl sulfate		Matrix metalloproteinase 2 inhibitor	[99]
<i>Polysyncraton lithostrotum</i>	Namenamicin		Cytotoxic, antitumor	[100]
<i>Polyandrocarpa</i> sp.	Polyandrocarpidines		Antimicrobial, cytotoxic, and deterrent activities	[101,102]
<i>Polyandrocarpa misakiensis</i>	Retinoic acid		Regeneration of gut	[103]
<i>Pseudodistoma antinboja</i>	Cadiolides J-M		Antibacterial	[104]
<i>Pycnoclavella kottae</i>	Kottamide D		Cytotoxic, anti-inflammatory, and antimetabolic activities	[105]
<i>Sidnyum turbinatum</i>	Alkyl sulfates		Antiproliferative	[106]
<i>Stolonica</i> sp.	Stolonic acid A and B		Antiproliferative	[107]
<i>Styela clava</i>	Clavanins		Antimicrobial	[108]
<i>Styela plicata</i>	Hemocytetes		Cytotoxic	[109]
<i>Synoicum adareanum</i>	Hyousterones and Abeohyousterone		Cytotoxic and anticancer	[110]
<i>Trididemnum solidum</i>	Didemnins A, B, and C		Antiviral, cytotoxic	[111,112]
<b>MNPs from associated microbes</b>				
<i>Candidatus Endoecteinascidia frumentensis</i>	Tetrahydroisoquinoline			[113]
<i>Microbulbifer</i> sp.	Bulbiferates A and B		Antibacterial	[114]
<i>Penicillium verruculosum</i>	Verruculides A, chrodrimanins A and H		Protein tyrosine phosphatase 1B inhibition	[115]
<i>Pseudoalteromonas rubra</i>	Isatin	Microbial defense	Antibacterial	[116]
<i>Pseudoalteromonas tunicata</i>	Tambjamine	Feeding deterrents		[116]
<i>Pseudoalteromonas tunicata</i>	Tambjamine		Antifungal	[117]
<i>Pseudovibrio denitrificans</i>	Diindol-3-ylmethanes		Antifouling	[118]
<i>Saccharopolyspora</i> sp.	JBIR-66		Cytotoxic	[119]
<i>Serratia marcescens</i>	Tetrapyrrole pigment	Feeding deterrents		[120]
<i>Streptomyces</i> sp.	Granaticin, granatomycin D, and dihydrogranaticin B		Antibacterial	[121]
<i>Talaromyces</i> sp.	Talaropeptides A-D		Plasma stability, Antibacterial, antifungal, cytotoxic	[24]
<i>Tistrella mobilis</i> and <i>Tistrella bauzanensis</i>	Didemnin		Anticancer	[23,122]



Plitidepsin



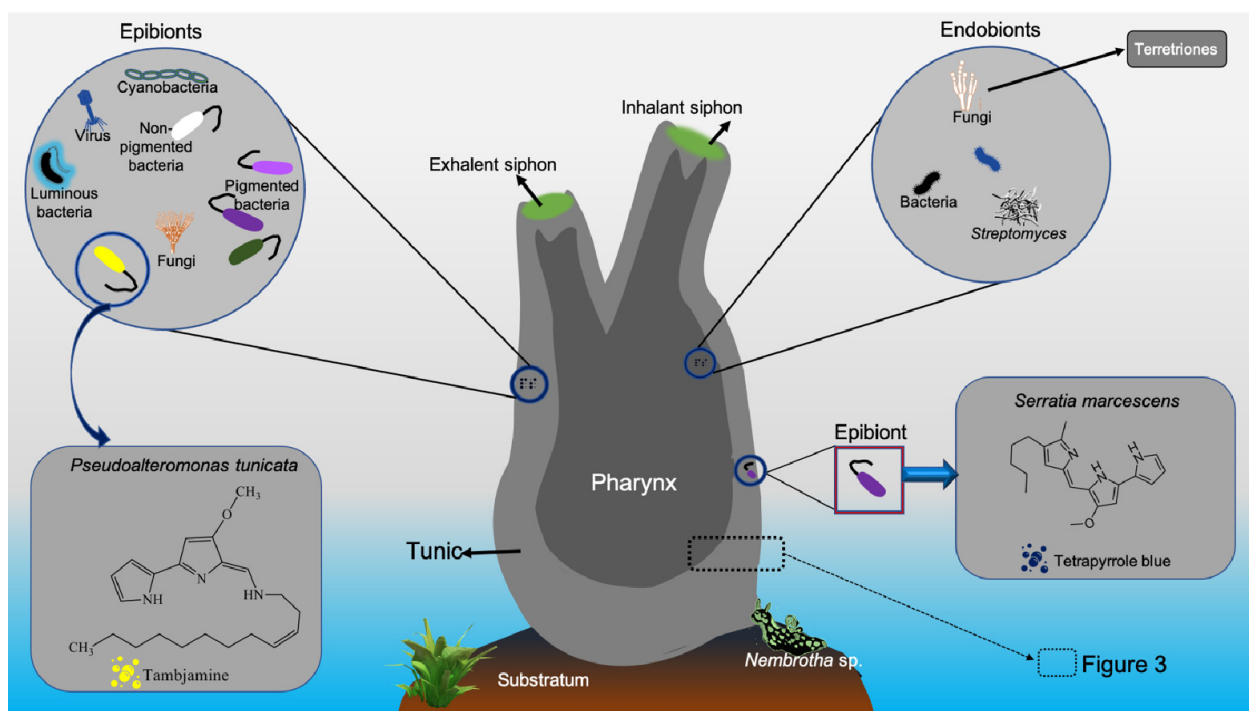
Didemnin B



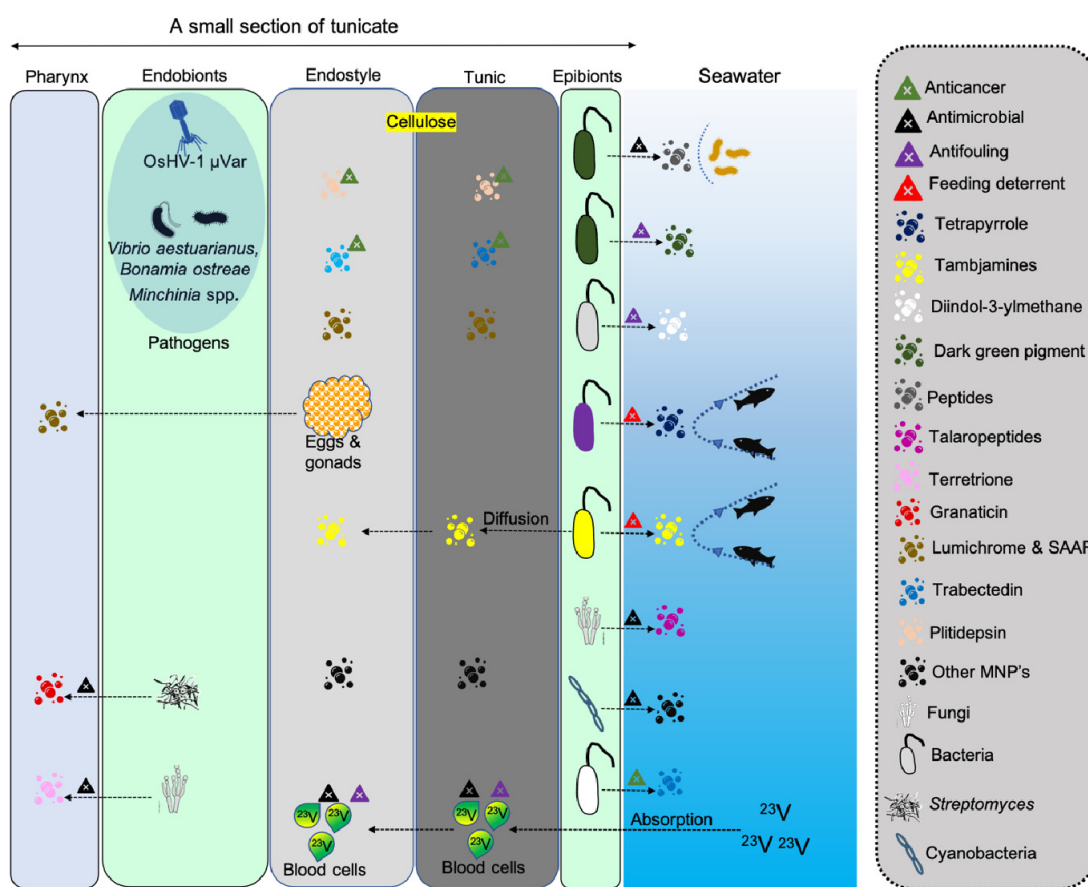
Trabectedin

**Figure 1.** Important anticancer drugs of tunicates and their associated microbes in clinical trials.





**Figure 2.** Tunicate-associated epibiotic and endobiotic symbionts. (the small inserted empty box provides more details in Figure 3).



**Figure 3.** Illustration depicting various MNPs released from endobiotic and epibiotic microbes associated with tunicate's endostyle and tunic.

## 5. Antimicrobial Applications

Tunicates [123], with their associated epi-symbionts [16,124] and endosymbionts [125], are prolific producers of antimicrobial and antifungal compounds inhibiting pathogens. The brominated alkaloids [126] and other compounds from tunicates have been reported to possess several biological activities [25,26]. *Pseudoalteromonas tunicata* produces alkaloid tambjamine (425 nm), an antifungal yellow pigment [127,128], and violacein (575 nm), a purple pigment with antiprotozoal activity [129,130], in addition to a range of bioactive compounds [129,131]. Methanol extraction of *Lissoclinum fragile* displayed antibacterial, antifungal, hemolytic, and cytotoxic activities [92]. The kuanoniamine A metabolite produced by *Eusynstyela tincta* inhibited pathogenic bacteria such as *B. subtilis*, *E. coli*, *S. aureus*, *V. cholerae*, and *V. parahaemolyticus* and fungi *A. fumigatus* and *C. albicans* [88]. A diffusible 190-kDa protein produced by tunicate *Ciona intestinalis* associated bacterium *Pseudoalteromonas tunicata* was found to show antibacterial activity against marine isolates [132]. The four  $\alpha$ -helical peptides “clavanins A, B, C, and D” isolated from the hemocytes of tunicate *Styela clava* showed antibacterial activity against pathogenic *Listeria monocytogenes* strain EGD and antifungal activity against *Candida albicans* [44]. Halocidin, an antimicrobial peptide purified from tunicate *Halocynthia aurantium* showed antibacterial activity against methicillin-resistant *Staphylococcus aureus* and multidrug-resistant *Pseudoalteromonas aeruginosa* [47]. Similarly, halocytin and papillosin peptides isolated from tunicate *Halocynthia papillosa* also displayed antibacterial activity against both Gram-positive and Gram-negative marine bacteria [46]. Halocytamine peptides synthesized by the hemocytes of *Halocynthia roretzi* showed antimicrobial activity against various bacteria and yeasts [90]. Similarly, Halocytamines produced by *Styela clava* also displayed antimicrobial properties [108]. A salt-tolerant peptide isolated from hemocytes of *Ciona intestinalis* showed both antibacterial and antifungal activity [133]. Vanadium chloride and vanadyl sulfate also displayed antibacterial activity against various pathogens [95].

An endobiont, *Streptomyces* sp., isolated from the tunicate, *Styela canopus*, produced antibacterial compounds such as granaticin, granatomycin D, and dihydrogranaticin B [121]. Similarly endosymbiotic fungi associated with the tunicates, *Polycarpa aurata* [134] and *Rhopalaea crassa* [135], showed antimicrobial activity. The fungi *Talaromyces* sp., isolated from an unidentified tunicate, produced talaropeptides A and B, two antibacterial metabolites that inhibited Gram-positive bacteria, *Bacillus subtilis* [24]. The endophytic fungus *Penicillium* sp. isolated from *Didemnum* sp. produced antifungal and cytotoxic compounds, terretrione C and D [136].

Some tunicates produced antiviral molecules, indicating their chemical defense function against environmental viruses. The Caribbean tunicate, *Trididemnum* sp., was found to produce depsipeptides, particularly didemnins A and B, exhibiting antiviral activity against DNA and RNA viruses in vitro [111,137]. Another species of Caribbean tunicate, *Eudistoma olivaceum*, produced prolific MNPs, such as eudistomins A, D, G, H, I, J, M, N, O, P, and Q, which possessed antiviral activity [83]. The ascidian *Didemnum guttatum* was found to produce the cyclodidemniserinol trisulfate compound that showed anti-retroviral activity by inhibiting HIV-1 integrase [72]. The tunicate, *Didemnum molle*, released lanthipeptide divamide A that promised to be a potential anti-HIV drug [74] (Table 4).

## 6. Anticancer and Antitumor Applications

Trabectedin (Ecteinascidin; ET-743; Yondelis<sup>®</sup>), an alkaloid extracted from the orange tunicate, *Ecteinascidia turbinata*, is approved as a first anticancer drug [138] to treat breast cancer [139,140], soft tissue sarcoma [141], and ovarian cancer [142–144]. This molecule is suggested to originate from *E. turbinata* symbiotic bacteria, *Candidatus Endoecteinascidia frumentensis* [145]. However, plitidepsin (Aplidin<sup>®</sup>), a depsipeptide isolated from the Mediterranean tunicate, *Aplidium albicans*, is in phase II clinical trials [138,146] as an anticancer drug against breast cancer [147], human kidney carcinoma cells [52], and multiple myeloma [53]. Didemnins B is also in phase II trials [148], showing anticancer activity against leukaemia P388 cells [111]. Significantly, 60% of the human cervical carcinoma cell lines (HeLa)

were inhibited by Eudistomins H extracts (IC<sub>50</sub> 0.49 µg/mL) obtained from *E. viride* [86]. Clavepictine A and B alkaloids originated from *Clavelina picta* demonstrated potential cytotoxic activity (IC<sub>50</sub> 12 µg/mL) against murine leukemia and human solid tumor cell lines [62]. Lamellarin sulfates originated from *Didemnum ternerratum* [78] and polycarpine dihydrochloride, a disulfide alkaloid extracted from an ascidian *Polycarpa clavata*, were found to inhibit human colon tumor cell lines [97].

Cystodytins A, B, and C, three teracyclic alkaloids isolated from Okinawa tunicate *Cystodytes dellechiaiei*, were reported to show antitumor activities [64]. Macrolides isolated from tunicates *Lissoclinum patella* (Patellazole C) [94] and *Eudistoma cf. rigida* (Lejimalides A, B, C, and D) [149,150] possessed anticancer activity [151]. Diplamine, an orange pigment alkaloid produced by *Diplosoma* sp., demonstrated cytotoxic activity against leukemia cells [79]. Halocyamine A and B peptides extracted from *H. roretzi* showed anticancer activity against various cell lines [90]. A depsipeptide, dehydrodidemnin B, produced by *Aplidium albicans* inhibited Ehrlich carcinoma cells in mice and reduced 80–90% tumor cells [54]. Bryostatins Ecteinascidins products, such as ET-729, 743, 745, 759 A, 759B, and 770, extracted from the Caribbean tunicate *Ecteinascidia turbinata* showed immunomodulator activity and antitumor activity against various leukemia cells [152] and breast, lung, ovary, and melanoma cells [153]. The Brazilian ascidian, *Didemnum granulatum*, produced G2 checkpoint-inhibiting aromatic alkaloids, granulatimide and isogranulatimide [154]. The ascidian *Cystodytes dellechiaiei* produced topoisomerase II-inhibiting ascididemin, which has antitumor activity against various tumor cell lines [66]. This marine alkaloid exhibits marked cytotoxic activities against a range of tumor cells. The kuanoniamine A metabolite extracted from *E. tincta* displayed 100% inhibition of Dalton's lymphoma and Ehrlich ascites tumor cell lines [88]. Cynthichlorine, an alkaloid isolated from the tunicate *Cynthia savignyi*, showed cytotoxicity against *Artemia salina* larva at an LD<sub>50</sub> of 48.5 µg/mL [63]. Siladenoserinols A and B derivatives isolated from didemnid tunicates possessed antitumor activity by inhibiting the interaction of p53-Hdm2 [69] (Table 4).

**Table 4.** Bioactive MNP's from tunicates and associated microbes.

Application	Compound	Activity against	Dose/ Concentration	Growth Inhibition (Diameter/ Percentage)	Assay Method	Reference
<b>Antimicrobial</b>						
	Clavanins	<i>E. coli</i> , <i>L. monocytogenes</i> , <i>C. albicans</i>	1.6–3.5 µg/mL	-	Radial diffusion assay	[44]
	Diplamine	<i>E. coli</i> , <i>S. aureus</i>			-	[79]
	Halocidin	Methicillin-resistant <i>Staphylococcus aureus</i> and multidrug-resistant <i>Pseudomonas aeruginosa</i>	100–200 µg/mL	5–11 mm	Radial diffusion assay	[47]
	Isatin	<i>Bacillus cereus</i> , <i>Bacillus megaterium</i> , <i>Escherichia coli</i> , <i>Micrococcus luteus</i> ,	MIC 200 µg/mL	7–>21 mm	Disk diffusion assay	[16]
	Kuanoniamine A	<i>B. Subtilis</i> , <i>E. coli</i> , <i>S. aureus</i> , <i>V. cholerae</i> , <i>V. parahaemolyticus</i> and fungus <i>A. jumigatus</i> and <i>C. albicans</i>	25 µg/mL	7–13 mm	Disk diffusion assay	[88]
	Cynthichlorine	<i>A. radiobacter</i> , <i>E. coli</i> , <i>P. aeruginosa</i> , <i>Botrytis cinerea</i> , <i>Verticillium albo atrum</i>		6–10 mm	Disc diffusion assay	[63]

Table 4. Cont.

Application	Compound	Activity against	Dose/ Concentration	Growth Inhibition (Diameter/ Percentage)	Assay Method	Reference
	Talaropeptides A and B	<i>Bacillus subtilis</i>	IC <sub>50</sub> 1.5–3.7 µM	50%	Microtiter plate assay	[24]
	Terretrione C and D	<i>Candida albicans</i>	MIC 32 µg/mL	17–19 mm	Disc diffusion assay	[136]
<b>Anticancer &amp; antitumor</b>						
	Aplidin	Multiple myeloma cell lines, MDA-MB-231 breast cancer cells, A-498 and ACHN cell lines	IC <sub>50</sub> 1 to 15 nmol/L		Nuclear Staining Assay; MTT assay	[52,53]
	Clavepictines A and B	Murine leukemia and human solid tumor cell lines	IC <sub>50</sub> 12 µg/mL		Microculture tetrazolium assay	[62]
	Dehydroididemnin B	Ehrlich carcinoma cells	2.5 µg/mouse	70–90%	MTT assay	[54]
	Didemnins A and B	Leukaemia P388 cells	IC <sub>50</sub> 1.5–25 µg/mL		-	[111]
	Diplamine	Leukemia L1210 cells	IC <sub>50</sub> 2 × 10 <sup>-2</sup> µg/mL		-	[79]
	Ecteinascidin 743 (Trabectedin)	Leukemia L1210 cells	IC <sub>50</sub> 0.5 µg/mL		-	[152]
	Eudistomins H	HeLa cell lines	IC <sub>50</sub> 0.49 µg/mL	60%	MTT assay	[86]
	Halocyamine A and B	Rat neuronal cells, mouse neuroblastoma N-18 cells, and human Hep-G2 cells			-	[90]
	Kuanoniamine A	Dalton's lymphoma and Ehrlich ascites tumour cell lines	25 µg/mL	100%	Trypan blue exclusion test	[88]
	Lamellarin Sulfates	HCT-116 human colon tumor cells	IC <sub>50</sub> 9.7 µM		MTS cell proliferation assay	[78]
	Namenamicin	P388 leukemia cells, 3Y1, and HeLa	IC <sub>50</sub> 3.5 nM; IC <sub>50</sub> 3.3–13 pM		Biochemical prophage induction assay	[100]
	Polycarpine dihydrochloride	HCT-116 human colon tumor cells	ED <sub>50</sub> 1.9 µg/mL		-	[97]
	7-oxostaurosporine	HL-60, Molt-4, Jurkat, K562, HCT-8, MDA MB-435, and SF-295 cell lines	IC <sub>50</sub> 10–58 nM	95%	MTT assay	[85]
	Terretrione C and D	Human breast cancer cells	IC <sub>50</sub> 16.5 and 17.6 µM		Sulforhodamine B assay	[136]
<b>Antifouling</b>						
	Diindol-3-ylmethanes	Barnacle, <i>Balanus amphitrite</i> and bryozoan, <i>Bugula neritina</i>	EC <sub>50</sub> 18.57		Microtiter plate assay	[118]
	Eudistomins G and H	Fish and other larvae			Antifeedant assay	[34]

## 7. Antifouling and Anti-Deterrent Activities

The colonial tunicate, *Eudistoma olivaceum*, was found to produce brominated alkaloids, Eudistomins G and H, which acted as antifouling substances and fish antifeedants; thus, the *E. olivaceum* surface was completely free from fouling epibionts [34]. A dark green pigmented bacteria, *Pseudoalteromonas tunicata*, isolated from the surface of *Ciona intestinalis*, collected originally from off the west coast of Sweden, showed antifouling activity against algal spores, invertebrate larvae, and diatoms [131,155,156]. The yellow pigmented *Pseudoalteromonas tunicata* mutants have demonstrated antifouling activity against algal spore germination, bacterial growth, fungal growth, and invertebrate larvae [129]. Diindol-3-ylmethane products isolated from an unidentified ascidian-associated bacteria, *Pseudovibrio denitrificans*, displayed nearly 50% antifouling activity against barnacle *Balanus amphitrite* and bryozoan *Bugula neritina* [118].

Deterring activity of vanadium acidic solutions, such as vanadyl sulfate and sodium vanadate, was observed against *Thalassoma bifasciatum* when incorporated into food pellets [95,157]. Didemnimides C and D from *Didemnum conchyliatum* [158], nordidemnin B [102] and didemnin B [159] from *Trididemnum solidum*, and granulatamides from *Didemnum granulatum* [73] displayed antifeedant effects on various fishes in laboratory experiments. The kuanoniamine A molecule from *E. tinctoria* displayed feeding-deterrent activities against carnivore gold fish, *Carassius auratus* [88]. MNPs isolated from Antarctic tunicates have demonstrated variability in anti-deterrent activities [58]. Both the yellow pigmented tambjamine metabolites and blue tetrapyrrole metabolite released from *Sigillina* sp. (i.e., *Atapozoa* sp.) showed feeding-deterrent activity against various carnivore fishes [59,160]. The blue tetrapyrrole pigment was suggested to originate from the associated bacteria *Serratia marcescens* [120]. Tambjamins and tetrapyrrole chemical constituents from both adult and larvae were reported to function as defensive chemicals against predators [102]. Lipophilic crude extracts from Antarctic tunicate, *Distaplia cylindrica* [161], and polyandrocarpidines from *Polyandrocarpa* sp. [101,102] demonstrated deterrent activity against certain sea-stars, hermit crabs, and snails (Table 4).

## 8. Miscellaneous Applications

The chiton *Mopalia* sp. spawned when injected with 1.0 mg/L of gonadotropin releasing hormone (GnRH2) of a tunicate [48]. Lumichrome, a compound extracted from tunic, gonads, and eggs of ascidian, *Halocynthia roretzi*, was involved in the larval metamorphosis [89]. Similarly, sperm-activating and attracting factors (SAAF) were isolated from eggs of the ascidians *Ciona intestinalis* and *Ascidia sydneiensis* [162]. Lipids extracted from *H. roretzi* have demonstrated the antidiabetic and anti-obese properties in mice models [163]. Two novel alkaloids, mellpaladine and dopargimine, isolated from Palauan tunicate have demonstrated neuroactive behavior in mice [68]. Two new alkaloids, polyaurines A and B, isolated from the tunicate, *Polycarpa aurata*, inhibited blood-dwelling *Schistosoma mansoni* [96]. Lepadins and villatamine alkaloids isolated from *Clavelina lepadiformis* [61] and lepadins from *Didemnum* sp. [71] displayed potential antiparasitic and cytotoxic activities. The ascidian species, *Didemnum psammathodes*, collected from the central west coast of India was extracted in organic solvents. These extracts showed antimicrobial and antifouling properties [164].

## 9. Issues in Extraction & Identification of Tunicate MNPs

Marine organisms have developed diverse secondary metabolic pathways, which produce a vast number of unusual chemical moieties. These compounds belong to a wide variety of chemical classes, including terpenes, shikimates, polyketides, peptides, alkaloids, and many unidentified and uncharacterized structures (Houssen and Jaspars, 2012). There are several technologies in place to isolate and characterize the natural products from even a very small quantity of marine organisms. However, there are still hurdles in the isolation and characterization of bioactive molecules from ascidians. These include 1. taxonomic uncertainty: worldwide, there are very few taxonomists available for proper taxonomic

assignments of tunicates. Sometimes the identification using molecular tools has been complicated by the difficulty in getting pure gDNA from the target species due to complex biotic associations (Houssen and Jaspars, 2012). 2. Quantity of isolated molecules: most of the time, a small quantity of metabolites is available in the organisms, which is not even sufficient for spectroscopic analysis. 3. Instability of molecules: there are extremely labile compounds in the extracts, which decompose during the purification process, and we get artefacts. Of course, these problems are common in other marine invertebrates as well. Research funding has also become a hurdle for many young researchers; thus, many researchers are publishing their works with crude extracts instead of analyzing complete structural elucidation. If we could address these issues, we will be able to isolate and characterize novel bioactive molecules from this unique group of marine invertebrates. The quantity of molecules can be increased if we collect the target tunicate species at the right time (season) from the correct geographic location. This can be achieved by understanding the chemical ecology of the producing species. For this purpose, there should be joint efforts from marine biologists, ecologists, and natural product chemists.

### 10. Metabolic Origin of Some Tunicates and Their Predators

Several bioactive MNPs extracted from tunicates were believed to be originated from tunicates themselves. However, few studies have investigated the original origin of tunicate MNPs from their symbiotic microbes. Tambjamine pigments have been reported to be originated from tunicate-associated symbiotic bacteria like *S. marcescens* [160] and *Pseudoalteromonas tunicata* [116,131]. An identical dark blue pigmented tetrapyrrole compound isolated from an ascidian was observed from a bacterium [165]. The blue tetrapyrrole pigment was reported to have originated from the associated bacteria, *Serratia marcescens* [120]. Didemnins extracted from the tunicate, *T. solidum* [111], are found to be released by associated bacteria, *Tistrella mobilis* and *Tistrella bauzanensis* [23,122]. Similarly, the trabectedin compound identified from the Caribbean tunicate, *E. turbinata* [152,166], has now been observed to be produced by its symbiotic bacteria, *Candidatus Endoecteinascidia frumentensis* [145]. Meridianins isolated from Antarctic tunicates, *Aplidium*, *Synoicum*, and some sponges, are thought to have originated from their symbiotic microbes [58]. Similarly, tetrahydroisoquinoline constituents identified from the tunicate, *Ecteinascidia turbinata*, appeared to be released by the unculturable endosymbiotic bacterium, *Candidatus Endoecteinascidia frumentensis* [113]. Some of the bioactive MNPs identified from Didemnid tunicates also originated from their symbiotic cyanobacterial species, such as *Synechocystis* and *Prochloron* [167,168]. Namenamycin produced by the orange color ascidian, *Polysyncrator lithostrotum*, was suggested to originate from its symbiotic bacterium, *Micromonospora* species [100]. The anti-HIV lanthipeptide, divamide A, isolated from the tunicate, *Didemnum molle*, was found to be produced by uncultivable symbiotic bacteria [74].

Tunicates are known to produce more than 300 alkaloid compounds [126]. The tunicate predatory flatworm *Prostheceraeus villatus* was reported to obtain alkaloids, lepadins, and villatamines by preying (dietary origin) on the tunicate, *Clavelina lepadiformis* [61]. Likewise, tambjamine alkaloids observed in the ascidian *Atapozoa* sp. [160] and associated bacteria [131] were found to be acquired by the predatory nudibranchs, like *Nembrotha* sp., for defense functions [59,169]. Pyridoacridine metabolites observed in ascidians and some sponges indicate a possible microbial origin or convergent evolution of these molecules [170].

### 11. Utilization of Invasive Tunicates Resources

Tunicates usually occur in relatively low abundance in coastal waters. However, some tunicates are reported as invasive species in some coastal waters [171] and are known to cause space competition [172], damage to aquaculture [173,174] by harboring pathogenic viruses and bacteria [175], and ecosystem alteration within the spread area [176]. Few non-invasive tunicate species of the coral reef environment have also been reported to overgrow on massive corals and caused minimal [112] or partial inhibition or delayed

development of coral polyps [177]. A study reported the outbreak of the invasive tunicate, *Diplosoma similis*, that overgrew on corals and macrophytes and resulted in 50% mortality of corals [178] (Table 5).

**Table 5.** Occurrence of invasive tunicate species in the global ocean and their impact on the marine ecosystem.

Invasive Tunicate	Country	Origin Type	Negative Impacts	Reference
<i>Ascidella aspersa</i>	Argentina	Exotic	Space competition	[179]
<i>Botrylloides violaceus</i>	Netherlands	Exotic	Space competition	[172]
<i>Botryllus schlosseri</i>	Netherlands	Indigenous	Space competition	[172]
<i>Botryllus schlosseri</i> , <i>Botrylloides violaceus</i> , <i>Ciona intestinalis</i> , <i>Ciona savignyi</i> , <i>Didemnum vexillum</i> , <i>Molgula manhattensis</i> , <i>Styela clava</i>	USA	Exotic	Competitors for food and space	[180,181]
<i>Ciona intestinalis</i>	Canada	Exotic	Mussel mortality	[176]
<i>Ciona intestinalis</i>	Korea	Exotic	Space competition and damage to aquaculture	[174]
<i>Didemnum psammathodes</i>	India	Indigenous	Space competition	[182]
<i>Didemnum vexillum</i>	USA	Exotic	Threat to eelgrass	[183]
<i>Didemnum vexillum</i>	Wales	Exotic	Space competition	[184]
<i>Diplosoma similis</i>	American Sāmoa	Indigenous	Kill corals	[178]

Therefore, such overwhelming invasive species may be utilized to investigate their biological properties, biotechnological implications, and drug development. The exploitation of antiviral and cytotoxic didemnins from the invasive tunicate, *T. solidum*, has already been investigated [111,112]. Antimicrobial activity of  $\alpha$ -helical peptides “Clavanins” was identified from the hemocytes of the tunicate, *Styela clava* [44]. Thus, other invasive species need to be investigated for their bioactive properties. Seasonal studies on the spread of various invasive tunicates and their biomass estimations are an important research aspect for resource management and coastal conservation. A study suggested that ocean warming is triggering the rise of invasive species in coastal waters [185]. Therefore, identifying the key ocean-warming factors and their mitigation strategies is essential for a sustainable management of the global ocean bioresources.

## 12. Research Gaps and Future Perspective

Tunicates have been an important marine drug reservoir to treat a variety of diseases, including cancer. These resources from the ocean, particularly from the deep-sea, remain untapped for drug discovery. Therefore, exploration and exploitation of tunicate resources from coastal waters to the deep-sea and tropical to polar regions would open new insights in the drug discovery and evolutionary lineages. However, these efforts should be driven by chemical ecology of these organisms. The study of chemical ecology will help in bioprospecting and the efficient production of marine drugs from this unique group of organisms. On the other hand, the mode of colonization and pigment biosynthesis by associated microbes and the acquisition mechanism of pigments (e.g., tambjamins) by tunicates from their associated microbes are yet to be unveiled. Since tunicates have been reported to be colonized by pathogenic bacteria during filter feeding, the pathological implications of tunicates needs to be investigated to understand the possible transfer ways of pathogenic bacteria from tunicates to other biota and aquaculture setups. Therefore, regular biodiversity monitoring and population dynamics of tunicate resources should be performed to understand their distribution patterns and impact on the coastal resources.

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## References

1. Shenkar, N.; Swalla, B.J. Global diversity of Ascidiacea. *PLoS ONE* **2011**, *6*, e20657. [[CrossRef](#)]
2. Holland, L.Z. Tunicates. *Curr. Biol.* **2016**, *26*, R141–R156. [[CrossRef](#)] [[PubMed](#)]
3. Gasparini, F.; Ballarin, L. Reproduction in Tunicates. In *Encyclopedia of Reproduction*, 2nd ed.; Elsevier Inc.: Amsterdam, The Netherlands, 2018; Volume 6, pp. 546–553.
4. Bone, Q.; Carré, C.; Chang, P. Tunicate feeding filters. *J. Mar. Biol. Assoc. UK* **2003**, *83*, 907–919. [[CrossRef](#)]
5. Delsuc, F.; Philippe, H.; Tsagkogeorga, G.; Simion, P.; Tilak, M.K.; Turon, X.; López-Legentil, S.; Piette, J.; Lemaire, P.; Douzery, E.J.P. A phylogenomic framework and timescale for comparative studies of tunicates. *BMC Biol.* **2018**, *16*, 39. [[CrossRef](#)] [[PubMed](#)]
6. Berna, L.; Alvarez-Valin, F. Evolutionary genomics of fast evolving tunicates. *Genome Biol. Evol.* **2014**, *6*, 1724–1738. [[CrossRef](#)]
7. Swalla, B.J. Building divergent body plans with similar genetic pathways. *Heredity* **2006**, *97*, 235–243. [[CrossRef](#)] [[PubMed](#)]
8. Jeffery, W.R. Tunicates: Models for Chordate Evolution and Development at Low Genomic Complexity. In *Comparative Genomics*; Clark, M.S., Ed.; Springer Science + Business Media: New York, NY, USA, 2000; pp. 43–69.
9. Lemaire, P. Evolutionary crossroads in developmental biology: The tunicates. *Development* **2011**, *138*, 2143–2152. [[CrossRef](#)]
10. Zhan, A.; Briski, E.; Bock, D.G.; Ghabooli, S.; MacIsaac, H.J. Ascidiaceans as models for studying invasion success. *Mar. Biol.* **2015**, *162*, 2449–2470. [[CrossRef](#)]
11. Watters, D.J. Ascidian toxins with potential for drug development. *Mar. Drugs* **2018**, *16*, 162. [[CrossRef](#)]
12. Gouiffes, D.; Juge, M.; Grimaud, N.; Welin, L.; Sauviat, M.P.; Barbin, Y.; Laurent, D.; Roussakis, C.; Henichart, J.P.; Verbist, J.F. Bistramide A, a new toxin from the urochordata *Lissoclinum bistratum* Sluiter: Isolation and preliminary characterization. *Toxicol.* **1988**, *26*, 1129–1136. [[CrossRef](#)]
13. Oh, K.-S.; Kim, J.-S.; Heu, M.-S. Food Constituents of Edible Ascidiaceans *Halocynthia roretzi* and *Pyura michaelsoni*. *Korean J. Food Sci. Technol.* **1997**, *29*, 955–962.
14. Ali, A.J.H.; Tamilselvi, M. *Ascidiaceans in Coastal Water: A Comprehensive Inventory of Ascidian Fauna from the Indian Coast*; Springer Nature: Cham, Switzerland, 2016; ISBN 9783319291185.
15. DeFilippo, J.; Beck, G. Tunicate Immunology. In *Reference Module in Life Sciences*; Elsevier: Amsterdam, The Netherlands, 2018; pp. 1–10.
16. Ayuningrum, D.; Liu, Y.; Riyanti; Sibero, M.T.; Kristiana, R.; Asagabaldan, M.A.; Wuisan, Z.G.; Trianto, A.; Radjasa, O.K.; Sabdono, A.; et al. Tunicate-associated bacteria show a great potential for the discovery of antimicrobial compounds. *PLoS ONE* **2019**, *14*, e0213797. [[CrossRef](#)]
17. Franchi, N.; Ballarin, L. Immunity in protochordates: The tunicate perspective. *Front. Immunol.* **2017**, *8*, 674. [[CrossRef](#)] [[PubMed](#)]
18. Walters, T.L.; Gibson, D.M.; Frischer, M.E. Cultivation of the marine pelagic tunicate *Dliolella gegenbauri* (Uljanin 1884) for experimental studies. *J. Vis. Exp.* **2019**, *150*, e59832. [[CrossRef](#)]
19. Fusetani, N. *Drugs from the Sea*; Krager: Basel, Switzerland, 2000.
20. Michibara, H.; Uyama, T.; Ueki, T.; Kanamori, K. The mechanism of accumulation and reduction of vanadium by ascidians. In *The Biology of Ascidiaceans*; Sawada, H., Yokosawa, H., Lambert, C.C., Eds.; Springer: Tokyo, Japan, 2001; pp. 363–373.
21. Shen, G.Q.; Baker, B.J. Biosynthetic studies of the eudistomins in the tunicate *Eudistoma olivaceum*. *Tetrahedron Lett.* **1994**, *35*, 1141–1144. [[CrossRef](#)]
22. Le, V.H.; Inai, M.; Williams, R.M.; Kan, T. Ecteinascidins. A review of the chemistry, biology and clinical utility of potent tetrahydroisoquinoline antitumor antibiotics. *Nat. Prod. Rep.* **2015**, *32*, 328–347. [[CrossRef](#)]
23. Xu, Y.; Kersten, R.D.; Nam, S.J.; Lu, L.; Al-Suwailem, A.M.; Zheng, H.; Fenical, W.; Dorrestein, P.C.; Moore, B.S.; Qian, P.Y. Bacterial biosynthesis and maturation of the didemnin anti-cancer agents. *J. Am. Chem. Soc.* **2012**, *134*, 8625–8632. [[CrossRef](#)]
24. Dewapriya, P.; Khalil, Z.G.; Prasad, P.; Salim, A.A.; Cruz-Morales, P.; Marcellin, E.; Capon, R.J. Talaropeptides A–D: Structure and biosynthesis of extensively N-methylated linear peptides from an Australian marine tunicate-derived *Talaromyces* sp. *Front. Chem.* **2018**, *6*, 394. [[CrossRef](#)]
25. Dou, X.; Dong, B. Origins and bioactivities of natural compounds derived from marine ascidians and their symbionts. *Mar. Drugs* **2019**, *17*, 670. [[CrossRef](#)]
26. Casertano, M.; Menna, M.; Imperatore, C. The ascidian-derived metabolites with antimicrobial properties. *Antibiotics* **2020**, *9*, 510. [[CrossRef](#)]



27. Palanisamy, S.K.; Rajendran, N.M.; Marino, A. Natural products diversity of marine ascidians (Tunicates; Ascidiacea) and successful drugs in clinical development. *Nat. Prod. Bioprospect.* **2017**, *7*, 1–111. [[CrossRef](#)]
28. Petersen, J.K.; Riisgard, H.U. Filtration capacity of the ascidian *Ciona intestinalis* and its grazing impact in a shallow fjord. *Mar. Ecol. Prog. Ser.* **1992**, *88*, 9–17. [[CrossRef](#)]
29. Morris, R.J.; Bone, Q.; Head, R.; Braconnot, J.C.; Nival, P. Role of salps in the flux of organic matter to the bottom of the Ligurian Sea. *Mar. Biol.* **1988**, *97*, 237–241. [[CrossRef](#)]
30. Pomeroy, L.R.; Deibel, D. Aggregation of organic matter by pelagic tunicates. *Limnol. Oceanogr.* **1980**, *25*, 643–652. [[CrossRef](#)]
31. Gorsky, G.; Da Silva, N.L.; Dallot, S.; Laval, P.; Braconnot, J.C.; Prieur, L. Midwater tunicates: Are they related to the permanent front of the Ligurian Sea (NW Mediterranean)? *Mar. Ecol. Prog. Ser.* **1991**, *74*, 195–204. [[CrossRef](#)]
32. Sutherland, K.R.; Madin, L.P.; Stocker, R. Filtration of submicrometer particles by pelagic tunicates. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 15129–15134. [[CrossRef](#)]
33. Hirose, E.; Nozawa, Y. Latitudinal difference in the species richness of photosymbiotic ascidians along the east coast of Taiwan. *Zool. Stud.* **2020**, *59*, e19. [[CrossRef](#)] [[PubMed](#)]
34. Davis, A.R. Alkaloids and ascidian chemical defense: Evidence for the ecological role of natural products from *Eudistoma olivaceum*. *Mar. Biol.* **1991**, *111*, 375–379. [[CrossRef](#)]
35. Zhao, Y.; Li, J. Excellent chemical and material cellulose from tunicates: Diversity in cellulose production yield and chemical and morphological structures from different tunicate species. *Cellulose* **2014**, *21*, 3427–3441. [[CrossRef](#)]
36. Menna, M.; Aiello, A. The Chemistry of Marine Tunicates. In *Handbook of Marine Natural Products*; Fattorusso, E., Gerwick, W.H., Tagliatalata-Scafati, O., Eds.; Springer Science + Business Media B.V.: Berlin/Heidelberg, Germany, 2012; pp. 295–385.
37. Franchi, N.; Ballarin, L. Cytotoxic cells of compound Ascidiaceans. In *Lessons in Immunity: From Single-Cell Organisms to Mammals*; Ballarin, L., Cammarata, M., Eds.; Elsevier Inc.: London, UK, 2016; pp. 193–199.
38. Parrinello, N. Cytotoxic activity of tunicate hemocytes. In *Invertebrate Immunology*; Rinkevich, B., Müller, W.E.G., Eds.; Springer: Berlin/Heidelberg, Germany, 1996; pp. 190–217.
39. Liu, L.; Zheng, Y.-Y.; Shao, C.-L.; Wang, C.-Y. Metabolites from marine invertebrates and their symbiotic microorganisms: Molecular diversity discovery, mining, and application. *Mar. Life Sci. Technol.* **2019**, *1*, 60–94. [[CrossRef](#)]
40. Leal, M.C.; Sheridan, C.; Osinga, R.; Dionísio, G.; Rocha, R.; Silva, B.; Rosa, R.; Calado, C. Marine Microorganism-Invertebrate Assemblages: Perspectives to Solve the “Supply Problem” in the Initial Steps of Drug Discovery. *Mar. Drugs* **2014**, *12*, 3929–3952. [[CrossRef](#)]
41. Chen, L.; Hu, J.S.; Xu, J.L.; Shao, C.L.; Wang, G.Y. Biological and chemical diversity of ascidian-associated microorganisms. *Mar. Drugs* **2018**, *16*, 362. [[CrossRef](#)] [[PubMed](#)]
42. Oltz, E.M.; Smith, M.J.; Nakanishi, K.; Bruening, R.C.; Kustin, K. The tunichromes. A class of reducing blood pigments from sea squirts: Isolation, structures, and vanadium chemistry. *J. Am. Chem. Soc.* **1988**, *110*, 6162–6172. [[CrossRef](#)] [[PubMed](#)]
43. Kustin, K.; Robinson, W.E.; Smith, M.J. Tunichromes, vanadium, and vacuolated blood cells in tunicates. *Invertebr. Reprod. Dev.* **1990**, *17*, 129–139. [[CrossRef](#)]
44. Lee, I.H.; Zhao, C.; Cho, Y.; Harwig, S.S.L.; Cooper, E.L.; Lehrer, R.I. Clavanins,  $\alpha$ -helical antimicrobial peptides from tunicate hemocytes. *FEBS Lett.* **1997**, *400*, 158–162. [[CrossRef](#)]
45. Cima, F.; Franchi, N.; Ballarin, L. Origin and functions of tunicate hemocytes. In *The Evolution of the Immune System: Conservation and Diversification*; Malagoli, D., Ed.; Academic Press: London, UK, 2016; pp. 29–49. ISBN 9780128020135.
46. Galinier, R.; Roger, E.; Sautiere, P.E.; Aumelas, A.; Banaigs, B.; Mitta, G. Halocynthin and papillosin, two new antimicrobial peptides isolated from hemocytes of the solitary tunicate, *Halocynthia papillosa*. *J. Pept. Sci.* **2009**, *15*, 48–55. [[CrossRef](#)] [[PubMed](#)]
47. Jang, W.S.; Kim, K.N.; Lee, Y.S.; Nam, M.H.; Lee, I.H. Halocidin: A new antimicrobial peptide from hemocytes of the solitary tunicate, *Halocynthia aurantium*. *FEBS Lett.* **2002**, *521*, 81–86. [[CrossRef](#)]
48. Gorbman, A.; Whiteley, A.; Kavanaugh, S. Pheromonal stimulation of spawning release of gametes by gonadotropin releasing hormone in the chiton, *Mopalia* sp. *Gen. Comp. Endocrinol.* **2003**, *131*, 62–65. [[CrossRef](#)]
49. Evans, J.S.; Erwin, P.M.; Shenkar, N.; López-Legentil, S. Introduced ascidians harbor highly diverse and host-specific symbiotic microbial assemblages. *Sci. Rep.* **2017**, *7*, 11033. [[CrossRef](#)]
50. Tianero, M.D.B.; Kwan, J.C.; Wyche, T.P.; Presson, A.P.; Koch, M.; Barrows, L.R.; Bugni, T.S.; Schmidt, E.W. Species specificity of symbiosis and secondary metabolism in ascidians. *ISME J.* **2015**, *9*, 615–628. [[CrossRef](#)]
51. Flood, P. Yellow-stained oikopleurid appendicularians are caused by bacterial parasitism. *Mar. Ecol. Prog. Ser.* **1991**, *71*, 291–295. [[CrossRef](#)]
52. Cuadrado, A.; García-Fernández, L.F.; González, L.; Suárez, Y.; Losada, A.; Alcaide, V.; Martínez, T.; Maa Fernández-Sousa, J.; Sánchez-Puelles, J.M.; Muñoz, A. Aplidin<sup>TM</sup> induces apoptosis in human cancer cells via glutathione depletion and sustained activation of the epidermal growth factor receptor, Src, JNK, and p38 MAPK. *J. Biol. Chem.* **2003**, *278*, 241–250. [[CrossRef](#)] [[PubMed](#)]
53. Mitsiades, C.S.; Ocio, E.M.; Pandiella, A.; Maiso, P.; Gajate, C.; Garayoa, M.; Vilanova, D.; Montero, J.C.; Mitsiades, N.; McMullan, C.J.; et al. Aplidin, a marine organism-derived compound with potent antimyeloma activity in vitro and in vivo. *Cancer Res.* **2008**. [[CrossRef](#)]
54. Urdiales, J.L.; Morata, P.; De Castro, I.N.; Sánchez-Jiménez, F. Antiproliferative effect of dehydrodidemnin B (DDB), a depsipeptide isolated from Mediterranean tunicates. *Cancer Lett.* **1996**, *102*, 31–37. [[CrossRef](#)]

55. Fedorov, S.N.; Radchenko, O.S.; Shubina, L.K.; Balaneva, N.N.; Bode, A.M.; Stonik, V.A.; Dong, Z. Evaluation of cancer-preventive activity and structure–activity relationships of 3-demethylubiquinone Q2, isolated from the ascidian *Aplidium glabrum*, and its synthetic analogs. *Pharm. Res.* **2006**, *23*, 70–81. [[CrossRef](#)] [[PubMed](#)]
56. Garrido, L.; Zubía, E.; Ortega, M.J.; Salvá, J. Haouamines A and B: A new class of alkaloids from the ascidian *Aplidium haouarianum*. *J. Org. Chem.* **2003**, *68*, 293–299. [[CrossRef](#)] [[PubMed](#)]
57. Park, N.S.; Park, Y.K.; Ramalingam, M.; Yadav, A.K.; Cho, H.R.; Hong, V.S.; More, K.N.; Bae, J.H.; Bishop-Bailey, D.; Kano, J.; et al. Meridianin C inhibits the growth of YD-10B human tongue cancer cells through macropinocytosis and the down-regulation of Dickkopf-related protein-3. *J. Cell. Mol. Med.* **2018**, *22*, 5833–5846. [[CrossRef](#)]
58. Núñez-Pons, L.; Carbone, M.; Vázquez, J.; Rodríguez, J.; Nieto, R.M.; Varela, M.M.; Gavagnin, M.; Avila, C. Natural products from antarctic colonial ascidians of the genera *Aplidium* and *Syonicum*: Variability and defensive role. *Mar. Drugs* **2012**, *10*, 1741–1764. [[CrossRef](#)]
59. Paul, V.J.; Lindquist, N.; Fenical, W. Chemical defenses of the tropical ascidian *Atapozoa* sp. and its nudibranch predators *Nembrotha* spp. *Mar. Ecol. Prog. Ser.* **1990**, *59*, 109–118. [[CrossRef](#)]
60. Choi, H.; Hwang, H.; Chin, J.; Kim, E.; Lee, J.; Nam, S.J.; Lee, B.C.; Rho, B.J.; Kang, H. Tuberatolides, potent FXR antagonists from the Korean marine tunicate *Botryllus tuberatus*. *J. Nat. Prod.* **2011**, *74*, 90–94. [[CrossRef](#)]
61. Kubanek, J.; Williams, D.E.; de Silva, E.D.; Allen, T.; Andersen, R.J. Cytotoxic alkaloids from the flatworm *Prostheceraeus villatus* and its tunicate prey *Clavelina lepadiformis*. *Tetrahedron Lett.* **1995**, *36*, 6189–6192. [[CrossRef](#)]
62. Raub, M.F.; Cardellina, J.H.; Choudhary, M.I.; Ni, C.Z.; Clardy, J.; Alley, M.C. Clavepictines A and B: Cytotoxic Quinolizidines from the Tunicate *Clavelina picta*. *J. Am. Chem. Soc.* **1991**, *113*, 3178–3180. [[CrossRef](#)]
63. Abourriche, A.; Abboud, Y.; Maoufoud, S.; Mohou, H.; Seffaj, T.; Charrouf, M.; Chaib, N.; Bennamara, A.; Bontemps, N.; Francisco, C. Cynthichlorine: A bioactive alkaloid from the tunicate *Cynthia savignyi*. *Farmaco* **2003**, *58*, 1351–1354. [[CrossRef](#)]
64. Kobayashi, J.; Cheng, J.F.; Nakamura, H.; Ohizumi, Y.; Walchli, M.R.; Hirata, Y.; Sasaki, T. Cystodytins A, B, and C, novel tetracyclic aromatic alkaloids with potent antineoplastic activity from the Okinawan tunicate *Cystodytes dellechiaiei*. *J. Org. Chem.* **1988**, *53*, 1800–1804. [[CrossRef](#)]
65. Kobayashi, J.; Tsuda, M.; Tanabe, A.; Ishibashi, M.; Cheng, J.F.; Yamamura, S.; Sasaki, T. Cystodytins D–I, new cytotoxic tetracyclic aromatic alkaloids from the Okinawan marine tunicate *Cystodytes dellechiaiei*. *J. Nat. Prod.* **1991**, *54*, 1634–1638. [[CrossRef](#)] [[PubMed](#)]
66. Dassonneville, L.; Wattez, N.; Baldeyrou, B.; Mahieu, C.; Lansiaux, A.; Banaigs, B.; Bonnard, I.; Bailly, C. Inhibition of topoisomerase II by the marine alkaloid ascididemin and induction of apoptosis in leukemia cells. *Biochem. Pharmacol.* **2000**, *60*, 527–537. [[CrossRef](#)]
67. López-Legentil, S.; Turon, X.; Schupp, P. Chemical and physical defenses against predators in *Cystodytes* (Asciacea). *J. Exp. Mar. Biol. Ecol.* **2006**, *332*, 27–36. [[CrossRef](#)]
68. Uchimasu, H.; Matsumura, K.; Tsuda, M.; Kumagai, K.; Akakabe, M.; Fujita, M.J.; Sakai, R. Mellpaladines and dopargimine, novel neuroactive guanidine alkaloids from a Palauan Didemnidae tunicate. *Tetrahedron* **2016**, *72*, 7185–7193. [[CrossRef](#)]
69. Torii, M.; Hitora, Y.; Kato, H.; Koyanagi, Y.; Kawahara, T.; Losung, F.; Mangindaan, R.E.P.; Tsukamoto, S. Siladenoserinols M–P, sulfonated serinol derivatives from a tunicate. *Tetrahedron* **2018**, *74*, 7516–7521. [[CrossRef](#)]
70. Machida, K.; Arai, D.; Katsumata, R.; Otsuka, S.; Yamashita, J.K.; Ye, T.; Tang, S.; Fusetani, N.; Nakao, Y. Sameuramide A, a new cyclic depsipeptide isolated from an ascidian of the family Didemnidae. *Bioorg. Med. Chem.* **2018**, *26*, 3852–3857. [[CrossRef](#)] [[PubMed](#)]
71. Wright, A.D.; Goclik, E.; König, G.M.; Kaminsky, R. Lepadins D–F: Antiplasmodial and antitrypanosomal decahydroquinoline derivatives from the tropical marine tunicate *Didemnum* sp. *J. Med. Chem.* **2002**, *45*, 3067–3072. [[CrossRef](#)] [[PubMed](#)]
72. Mitchell, S.S.; Rhodes, D.; Bushman, F.D.; Faulkner, D.J. Cyclodidemniserinol trisulfate, a sulfated serinolipid from the Palauan ascidian *Didemnum guttatum* that inhibits HIV-1 integrase. *Org. Lett.* **2000**, *2*, 1605–1607. [[CrossRef](#)]
73. Selegim, M.H.R.; De Lira, S.P.; Campana, P.T.; Berlinck, R.G.S.; Custódio, M.R. Localization of granulatinamide alkaloids in the tissues of the ascidian *Didemnum granulatum*. *Mar. Biol.* **2007**, *150*, 967–975. [[CrossRef](#)]
74. Smith, T.E.; Pond, C.D.; Pierce, E.; Harmer, Z.P.; Kwan, J.; Zachariah, M.M.; Harper, M.K.; Wyche, T.P.; Maitainaho, T.K.; Bugni, T.S.; et al. Accessing chemical diversity from the uncultivated symbionts of small marine animals. *Nat. Chem. Biol.* **2018**, *14*, 179–185. [[CrossRef](#)]
75. Donia, M.S.; Wang, B.; Dunbar, D.C.; Desai, P.V.; Patny, A.; Avery, M.; Hamann, M.T. Mollamides B and C, cyclic hexapeptides from the Indonesian tunicate *Didemnum molle*. *J. Nat. Prod.* **2008**, *71*, 941–945. [[CrossRef](#)] [[PubMed](#)]
76. Oku, N.; Matsunaga, S.; Fusetani, N. Shishijimicins A–C, novel enediyne antitumor antibiotics from the ascidian *Didemnum proliferum*. *J. Am. Chem. Soc.* **2003**, *125*, 2044–2045. [[CrossRef](#)]
77. Takeara, R.; Jimenez, P.C.; Wilke, D.V.; Moraes, M.O.d.; Pessoa, C.; Lopes, N.P.; Lopes, J.L.C.; Lotufo, T.M.d.C.; Costa-Lotufo, L.V. Antileukemic effects of *Didemnum psammotodes* (Tunicata: Ascidiacea) constituents. *Comp. Biochem. Physiol. A Mol. Integr. Physiol.* **2008**, *151*, 363–369. [[CrossRef](#)]
78. Bracegirdle, J.; Robertson, L.P.; Hume, P.A.; Page, M.J.; Sharrock, A.V.; Ackerley, D.F.; Carroll, A.R.; Keyzers, R.A. Lamellarin Sulfates from the Pacific Tunicate *Didemnum ternerratum*. *J. Nat. Prod.* **2019**, *82*, 2000–2008. [[CrossRef](#)]
79. Charyulu, G.A.; McKee, T.C.; Ireland, C.M. Diplamine, a cytotoxic polyaromatic alkaloid from the tunicate *Diplosoma* sp. *Tetrahedron Lett.* **1989**, *30*, 4201–4202. [[CrossRef](#)]

80. Ogi, T.; Taira, J.; Margiastuti, P.; Ueda, K. Cytotoxic metabolites from the Okinawan ascidian *Diplosoma virens*. *Molecules* **2008**, *13*, 595–602. [[CrossRef](#)]
81. Cvetkovic, R.S.; Figgitt, D.P.; Plosker, G.L. ET-743. *Drugs* **2002**, *62*, 1185–1192. [[CrossRef](#)]
82. Rashid, M.A.; Gustafson, K.R.; Boyd, M.R. New cytotoxic n-methylated  $\beta$ -carboline alkaloids from the marine ascidian *Eudistoma gilboverde*. *J. Nat. Prod.* **2001**, *64*, 1454–1456. [[CrossRef](#)] [[PubMed](#)]
83. Kobayashi, J.; Harbour, G.C.; Gilmore, J.; Rinehart, K.L. Eudistomins A, D, G, H, I, J, M, N, O, P, and Q, Bromo-, Hydroxy-, Pyrrolyl-, and 1-Pyrrolynyl- $\beta$ -carbolines from the antiviral Caribbean tunicate *Eudistoma olivaceum*. *J. Am. Chem. Soc.* **1984**, *106*, 1526–1528. [[CrossRef](#)]
84. Rinehart, K.L.; Kobayashi, J.; Harbour, G.C.; Hughes, R.G.; Mizsak, S.A.; Scahill, T.A. Eudistomins C, E, K, and L, potent antiviral compounds containing a novel oxathiazepine ring from the Caribbean tunicate *Eudistoma olivaceum*. *J. Am. Chem. Soc.* **1984**, *106*, 1524–1526. [[CrossRef](#)]
85. Jimenez, P.C.; Wilke, D.V.; Ferreira, E.G.; Takeara, R.; De Moraes, M.O.; Silveira, E.R.; Lotufo, T.M.D.C.; Lopes, N.P.; Costa-Lotufo, L.V. Structure elucidation and anticancer activity of 7-oxostaurosporine derivatives from the Brazilian endemic tunicate *Eudistoma vannamei*. *Mar. Drugs* **2012**, *10*, 1092–1102. [[CrossRef](#)]
86. Rajesh, R.P.; Annappan, M. Anticancer effects of brominated indole alkaloid eudistomin H from marine ascidian *Eudistoma viride* against cervical cancer cells (HeLa). *Anticancer Res.* **2015**, *35*, 283–294.
87. Tapiolas, D.M.; Bowden, B.F.; Abou-Mansour, E.; Willis, R.H.; Doyle, J.R.; Muirhead, A.N.; Liptrot, C.; Llewellyn, L.E.; Wolff, C.W.W.; Wright, A.D.; et al. Eusynstyelamides A, B, and C, nNOS inhibitors, from the ascidian *Eusynstyela latericius*. *J. Nat. Prod.* **2009**, *72*, 1115–1120. [[CrossRef](#)]
88. Devi, S.; Rajasekharan, K.; Padmakumar, K.; Tanaka, J.; Higa, T. Biological activity and chemistry of the compound ascidian *Eusynstyela tineta*. In *The Biology of Ascidians*; Sawada, H., Yokosawa, H., Lambert, C.C., Eds.; Springer: Tokyo, Japan, 2001; pp. 341–354.
89. Tsukamoto, S.; Kato, H.; Hirota, H.; Fusetane, N. Lumichrome Is a putative intrinsic substance inducing larval metamorphosis in the ascidian *Halocynthia roretzi*. In *The Biology of Ascidians*; Sawada, H., Yokosawa, H., Lambert, C.C., Eds.; Springer: Tokyo, Japan, 2001; pp. 335–340.
90. Azumi, K.; Yokosawa, H.; Ishii, S.I. Halocyanines: Novel antimicrobial tetrapeptide-like substances isolated from the hemocytes of the solitary ascidian *Halocynthia roretzi*. *Biochemistry* **1990**, *29*, 159–165. [[CrossRef](#)]
91. Oda, T.; Fujiwara, T.; Liu, H.; Ukai, K.; Mangindaan, R.E.P.; Mochizuki, M.; Namikoshi, M. Effects of lissoclibadins and lissoclinotoxins, isolated from a tropical ascidian *Lissoclinum cf. badium*, on IL-8 production in a PMA-stimulated promyelocytic leukemia cell line. *Mar. Drugs* **2006**, *4*, 15–21. [[CrossRef](#)]
92. Kumaran, N.S.; Bragadeeswaran, S.; Meenakshi, V.K.; Balasubramanian, T. Bioactivity potential of extracts from ascidian *Lissoclinum fragile*. *Afr. J. Pharm. Pharmacol.* **2012**, *6*, 1854–1859. [[CrossRef](#)]
93. Corley, D.G.; Moore, R.E.; Paul, V.J. Patellazole B: A novel cytotoxic thiazole-containing macrolide from the marine tunicate *Lissoclinum patella*. *J. Am. Chem. Soc.* **1988**, *110*, 7920–7922. [[CrossRef](#)]
94. Zabriskie, T.M.; Mayne, C.L.; Ireland, C.M. Patellazole C: A novel cytotoxic macrolide from *Lissoclinum patella*. *J. Am. Chem. Soc.* **1988**, *110*, 7919–7920. [[CrossRef](#)]
95. Odate, S.; Pawlik, J.R. The role of vanadium in the chemical defense of the solitary tunicate, *Phallusia nigra*. *J. Chem. Ecol.* **2007**, *33*, 643–654. [[CrossRef](#)]
96. Casertano, M.; Imperatore, C.; Luciano, P.; Aiello, A.; Putra, M.Y.; Gimmelli, R.; Ruberti, G.; Menna, M. Chemical investigation of the Indonesian tunicate *Polycarpa aurata* and evaluation of the effects against *Schistosoma mansoni* of the novel alkaloids polyaurines A and B. *Mar. Drugs* **2019**, *17*, 278. [[CrossRef](#)]
97. Kang, H.; Fenical, W. Polycarpine dihydrochloride: A cytotoxic dimeric disulfide alkaloid from the Indian ocean ascidian *Polycarpa clavata*. *Tetrahedron Lett.* **1996**, *37*, 2369–2372. [[CrossRef](#)]
98. Guo, P.; Wang, Z.; Li, G.; Liu, Y.; Xie, Y.; Wang, Q. First discovery of polycarpine, polycarpaurines A and C, and their derivatives as novel antiviral and antiphytopathogenic fungus agents. *J. Agric. Food Chem.* **2016**, *64*, 4264–4272. [[CrossRef](#)]
99. Fujita, M.; Nakao, Y.; Matsunaga, S.; Nishikawa, T.; Fusetani, N. Sodium 1-(12-hydroxy)octadecanyl sulfate, an MMP2 inhibitor, isolated from a tunicate of the family polyclinidae. *J. Nat. Prod.* **2002**, *65*, 1936–1938. [[CrossRef](#)]
100. Nicolaou, K.C.; Li, R.; Lu, Z.; Pitsinos, E.N.; Alemany, L.B. Total synthesis and full structural assignment of namenamicin. *J. Am. Chem. Soc.* **2018**, *140*, 8091–8095. [[CrossRef](#)]
101. Cheng, M.T.; Rinehart, K.L. Polyandrocarpidines: Antimicrobial and Cytotoxic Agents from a Marine Tunicate (*Polyandrocarpa* sp.) from the Gulf of California. *J. Am. Chem. Soc.* **1978**, *100*, 7409–7411. [[CrossRef](#)]
102. Lindquist, N.; Hay, M.E.; Fenical, W. Defense of ascidians and their conspicuous larvae: Adult vs. larval chemical defenses. *Ecol. Monogr.* **1992**, *62*, 547–568. [[CrossRef](#)]
103. Kaneko, N.; Katsuyama, Y.; Kawamura, K.; Fujiwara, S. Regeneration of the gut requires retinoic acid in the budding ascidian *Polyandrocarpa misakiensis*. *Dev. Growth Differ.* **2010**, *52*, 457–468. [[CrossRef](#)]
104. Wang, W.; Kim, H.; Nam, S.J.; Rho, B.J.; Kang, H. Antibacterial butenolides from the Korean tunicate *Pseudodistoma antinboja*. *J. Nat. Prod.* **2012**, *27*, 574–577. [[CrossRef](#)] [[PubMed](#)]
105. Appleton, D.R.; Page, M.J.; Lambert, G.; Berridge, M.V.; Copp, B.R. Kottamides A–D: Novel bioactive imidazolone-containing alkaloids from the New Zealand ascidian *Pycnoclavella kottae*. *J. Org. Chem.* **2002**, *67*, 5402–5404. [[CrossRef](#)]

106. Aiello, A.; Carbonelli, S.; Fattorusso, E.; Iuvone, T.; Menna, M. New bioactive sulfated metabolites from the Mediterranean tunicate *Sidnyum turbinatum*. *J. Nat. Prod.* **2001**, *64*, 219–221. [[CrossRef](#)]
107. Davies-Coleman, M.T.; Cantrell, C.L.; Gustafson, K.R.; Beutler, J.A.; Pannell, L.K.; Boyd, M.R. Stolononic acids A and B, new cytotoxic cyclic peroxides from an Indian Ocean ascidian *Stolonica* species. *J. Nat. Prod.* **2000**, *63*, 1411–1413. [[CrossRef](#)]
108. Menzel, L.P.; Lee, I.H.; Sjostrand, B.; Lehrer, R.I. Immunolocalization of clavanins in *Styela clava* hemocytes. *Dev. Comp. Immunol.* **2002**, *26*, 505–515. [[CrossRef](#)]
109. Raftos, D.A.; Hutchinson, A. Cytotoxicity reactions in the solitary tunicate *Styela plicata*. *Dev. Comp. Immunol.* **1995**, *19*, 463–471. [[CrossRef](#)]
110. Miyata, Y. Ecdysteroids from the Antarctic tunicate *Syonicum adareanum*. *J. Nat. Prod.* **2007**, *70*, 1859–1864. [[CrossRef](#)]
111. Rinehart, K.L.; Gloer, J.B.; Hughes, R.G.; Renis, H.E.; Patrick McGovern, J.; Swynenberg, E.B.; Stringfellow, D.A.; Kuentzel, S.L.; Li, L.H. Didemnins: Antiviral and antitumor depsipeptides from a Caribbean tunicate. *Science* **1981**, *212*, 933–935. [[CrossRef](#)]
112. Rodríguez-Martínez, R.E.; Jordán-Garza, A.G.; Baker, D.M.; Jordán-Dahlgren, E. Competitive interactions between corals and *Trididemnum solidum* on Mexican Caribbean reefs. *Coral Reefs* **2012**, *31*, 571–577. [[CrossRef](#)]
113. Schofield, M.M.; Jain, S.; Porat, D.; Dick, G.J.; Sherman, D.H. Identification and analysis of the bacterial endosymbiont specialized for production of the chemotherapeutic natural product ET-743. *Environ. Microbiol.* **2015**, *17*, 3964–3975. [[CrossRef](#)] [[PubMed](#)]
114. Jayanetti, D.R.; Braun, D.R.; Barns, K.J.; Rajsiki, S.R.; Bugni, T.S. Bulbiferates A and B: Antibacterial acetamidohydroxybenzoates from a marine proteobacterium, *Microbulbifer* sp. *J. Nat. Prod.* **2019**, *82*, 1930–1934. [[CrossRef](#)] [[PubMed](#)]
115. Yamazaki, H.; Nakayama, W.; Takahashi, O.; Kirikoshi, R.; Izumikawa, Y.; Iwasaki, K.; Toraiwa, K.; Ukai, K.; Rotinsulu, H.; Wewengkang, D.S.; et al. Verruculides A and B, two new protein tyrosine phosphatase 1B inhibitors from an Indonesian ascidian-derived *Penicillium verruculosum*. *Bioorganic Med. Chem. Lett.* **2015**, *25*, 3087–3090. [[CrossRef](#)]
116. Pinkerton, D.M.; Banwell, M.G.; Garson, M.J.; Kumar, N.; De Moraes, M.O.; Cavalcanti, B.C.; Barros, F.W.A.; Pessoa, C. Antimicrobial and cytotoxic activities of synthetically derived tambjamins C and E-J, BE-18591, and a related alkaloid from the marine bacterium *Pseudoalteromonas tunicata*. *Chem. Biodivers.* **2010**, *7*, 1311–1324. [[CrossRef](#)]
117. Burke, C.; Thomas, T.; Egan, S.; Kjelleberg, S. The use of functional genomics for the identification of a gene cluster encoding for the biosynthesis of an antifungal tambjamine in the marine bacterium *Pseudoalteromonas tunicata*: Brief report. *Environ. Microbiol.* **2007**, *9*, 814–818. [[CrossRef](#)]
118. Wang, K.L.; Xu, Y.; Lu, L.; Li, Y.; Han, Z.; Zhang, J.; Shao, C.L.; Wang, C.Y.; Qian, P.Y. Low-toxicity diindol-3-ylmethanes as potent antifouling compounds. *Mar. Biotechnol.* **2015**, *17*, 624–632. [[CrossRef](#)] [[PubMed](#)]
119. Takagi, M.; Motohashi, K.; Izumikawa, M.; Khan, S.T.; Hwang, J.-H.; Shin-Ya, K. JBIR-66, a new metabolite isolated from tunicate-derived *Saccharopolyspora* sp. SS081219JE-28. *Biosci. Biotechnol. Biochem.* **2010**, *74*, 2355–2357. [[CrossRef](#)]
120. Wasserman, H.H.; Friedland, D.J.; Morrison, D.A. A novel dipyrrolyldipyrromethene prodigiosin analog from *Serratia marcescens*. *Tetrahedron Lett.* **1968**, *6*, 641–644. [[CrossRef](#)]
121. Sung, A.A.; Gromek, S.M.; Balunas, M.J. Upregulation and identification of antibiotic activity of a marine-derived *Streptomyces* sp. via co-cultures with human pathogens. *Mar. Drugs* **2017**, *15*, 250. [[CrossRef](#)] [[PubMed](#)]
122. Tsukimoto, M.; Nagaoka, M.; Shishido, Y.; Fujimoto, J.; Nishisaka, F.; Matsumoto, S.; Harunari, E.; Imada, C.; Matsuzaki, T. Bacterial production of the tunicate-derived antitumor cyclic depsipeptide didemnin B. *J. Nat. Prod.* **2011**, *74*, 2329–2331. [[CrossRef](#)]
123. Karthikeyan, M.M.; Ananthan, G.; Balasubramanian, T. Antimicrobial activity of crude extracts of some ascidians (Urochordata: Ascidiacea), from Palk Strait, (Southeast Coast of India). *World J. Fish. Mar. Sci.* **2009**, *1*, 262–267.
124. Ayuningrum, D.; Kristiana, R.; Nisa, A.A.; Radjasa, S.K.; Muchlissin, S.I.; Radjasa, O.K.; Sabdon, A.; Trianto, A. Bacteria associated with tunicate, *Polycarpa aurata*, from Lease sea, Maluku, Indonesia exhibiting anti-multidrug resistant bacteria. *Biodiversitas* **2019**, *20*, 956–964. [[CrossRef](#)]
125. Litaay, M.; Christine, G.; Gobel, R.B.; Dwyana, Z. Bioactivity of endo-symbiont bacteria of tunicate *Polycarpa aurata* as antimicrobial. In Proceedings of the 23 National Seminar of Indonesia Biology Society, Jayapura, Indonesia, 18 September 2015.
126. Menna, M.; Fattorusso, E.; Imperatore, C. Alkaloids from marine ascidians. *Molecules* **2011**, *16*, 8694–8732. [[CrossRef](#)]
127. Franks, A.; Haywood, P.; Holmström, C.; Egan, S.; Kjelleberg, S.; Kumar, N. Isolation and structure elucidation of a novel yellow pigment from the marine bacterium *Pseudoalteromonas tunicata*. *Molecules* **2005**, *10*, 1286–1291. [[CrossRef](#)] [[PubMed](#)]
128. Franks, A.; Egan, S.; Holmström, C.; James, S.; Lappin-Scott, H.; Kjelleberg, S. Inhibition of fungal colonization by *Pseudoalteromonas tunicata* provides a competitive advantage during surface colonization. *Appl. Environ. Microbiol.* **2006**, *72*, 6079–6087. [[CrossRef](#)]
129. Egan, S.; James, S.; Holmström, C.; Kjelleberg, S. Correlation between pigmentation and antifouling compounds produced by *Pseudoalteromonas tunicata*. *Environ. Microbiol.* **2002**, *4*, 433–442. [[CrossRef](#)]
130. Matz, C.; Webb, J.S.; Schupp, P.J.; Phang, S.Y.; Penesyan, A.; Egan, S.; Steinberg, P.; Kjelleberg, S. Marine biofilm bacteria evade eukaryotic predation by targeted chemical defense. *PLoS ONE* **2008**, *3*, e2744. [[CrossRef](#)]
131. Holmström, C.; James, S.; Neilan, B.A.; White, D.C.; Kjelleberg, S. *Pseudoalteromonas tunicata* sp. nov., a bacterium that produces antifouling agents. *Int. J. Syst. Bacteriol.* **1998**, *48*, 1205–1212. [[CrossRef](#)]
132. James, S.G.; Holmström, C.; Kjelleberg, S. Purification and characterization of a novel antibacterial protein from the marine bacterium D2. *Appl. Environ. Microbiol.* **1996**, *62*, 2783–2788. [[CrossRef](#)]

133. Fedders, H.; Michalek, M.; Grötzinger, J.; Leippe, M. An exceptional salt-tolerant antimicrobial peptide derived from a novel gene family of haemocytes of the marine invertebrate *Ciona intestinalis*. *Biochem. J.* **2008**, *416*, 65–75. [[CrossRef](#)] [[PubMed](#)]
134. Nurfadillah, A.; Litaay, M.; Gobel, R.B.; Haedar, N. Potency of tunicate *Polycarpa aurata* as inoculum source of sebagai sumber endosymbiotic fungi that produce antimicrobe. *J. Alam Lingkungan.* **2015**, *6*, 10–16.
135. Tahir, E.; Litaay, M.; Gobel, R.B.; Haedar, N.; Al, E. Potency of tunicate *Rhopalaea crassa* as inoculum source of endosymbiont fungi that produce antimicrobe. *Spermonde* **2016**, *2*, 33–37.
136. Shaala, L.A.; Youssef, D.T.A. Identification and bioactivity of compounds from the fungus *Penicillium* sp. CYE-87 isolated from a marine tunicate. *Mar. Drugs* **2015**, *13*, 1698–1709. [[CrossRef](#)] [[PubMed](#)]
137. Canonico, P.G.; Pannier, W.L.; Huggins, J.W.; Rienehart, K.L. Inhibition of RNA viruses in vitro and in Rift Valley fever-infected mice by didemmins A and B. *Antimicrob. Agents Chemother.* **1982**, *22*, 696–697. [[CrossRef](#)] [[PubMed](#)]
138. Mayer, A.M.S.; Glaser, K.B.; Cuevas, C.; Jacobs, R.S.; Kem, W.; Little, R.D.; McIntosh, J.M.; Newman, D.J.; Potts, B.C.; Shuster, D.E. The odyssey of marine pharmaceuticals: A current pipeline perspective. *Trends Pharmacol. Sci.* **2010**, *31*, 255–265. [[CrossRef](#)]
139. Zelek, L.; Yovine, A.; Brain, E.; Turpin, F.; Taamma, A.; Riofrio, M.; Spielmann, M.; Jimeno, J.; Misset, J.L. A phase II study of Yondelis® (trabectedin, ET-743) as a 24-h continuous intravenous infusion in pretreated advanced breast cancer. *Br. J. Cancer* **2006**, *94*, 1610–1614. [[CrossRef](#)]
140. Atmaca, H.; Bozkurt, E.; Uzunoglu, S.; Uslu, R.; Karaca, B. A diverse induction of apoptosis by trabectedin in MCF-7 (HER2−/ER+) and MDA-MB-453 (HER2+/ER−) breast cancer cells. *Toxicol. Lett.* **2013**, *221*, 128–136. [[CrossRef](#)] [[PubMed](#)]
141. Grosso, F.; Jones, R.L.; Demetri, G.D.; Judson, I.R.; Blay, J.-Y.; Cesne, A.L.; Lippo, R.S.; Casieri, P.; Collini, P.; Dileo, P.; et al. Efficacy of trabectedin (ecteinascidin-743) in advanced pretreated myxoid liposarcomas: A retrospective study. *Lancet Oncol.* **2007**, *8*, 595–602. [[CrossRef](#)]
142. Sessa, C.; De Braud, F.; Perotti, A.; Bauer, J.; Curigliano, G.; Noberasco, C.; Zanaboni, F.; Gianni, L.; Marsoni, S.; Jimeno, J.; et al. Trabectedin for women with ovarian carcinoma after treatment with platinum and taxanes fails. *J. Clin. Oncol.* **2005**, *23*, 1867–1874. [[CrossRef](#)]
143. Krasner, C.N.; McMeekin, D.S.; Chan, S.; Braly, P.S.; Renshaw, F.G.; Kaye, S.; Provencher, D.M.; Campos, S.; Gore, M.E. A Phase II study of trabectedin single agent in patients with recurrent ovarian cancer previously treated with platinum-based regimens. *Br. J. Cancer* **2007**, *97*, 1618–1624. [[CrossRef](#)]
144. Monk, B. A randomized phase III study of trabectedin with pegylated liposomal doxorubicin (PLD) versus PLD in relapsed, recurrent ovarian cancer (OC). *Eur. J. Cancer Suppl.* **2008**, *19*, viii1–viii4.
145. Rath, C.M.; Janto, B.; Earl, J.; Ahmed, A.; Hu, F.Z.; Hiller, L.; Dahlgren, M.; Kreft, R.; Yu, F.; Wolff, J.J.; et al. Meta-omic characterization of the marine invertebrate microbial consortium that produces the chemotherapeutic natural product ET-743. *ACS Chem. Biol.* **2011**, *6*, 1244–1255. [[CrossRef](#)] [[PubMed](#)]
146. Tohme, R.; Darwiche, N.; Gali-Muhtasib, H. A journey under the sea: The quest for marine anti-cancer alkaloids. *Molecules* **2011**, *16*, 9665–9696. [[CrossRef](#)]
147. González-Santiago, L.; Suárez, Y.; Zarich, N.; Muñoz-Alonso, M.J.; Cuadrado, A.; Martínez, T.; Goya, L.; Iradi, A.; Sáez-Tormo, G.; Maier, J.V.; et al. Aplidin® induces JNK-dependent apoptosis in human breast cancer cells via alteration of glutathione homeostasis, Rac1 GTPase activation, and MKP-1 phosphatase downregulation. *Cell Death Differ.* **2006**, *13*, 1968–1981. [[CrossRef](#)] [[PubMed](#)]
148. Rinehart, K.L. Antitumor compounds from tunicates. *Med. Res. Rev.* **2000**, *20*, 1–27. [[CrossRef](#)]
149. Kobayashi, J.; Cheng, J.F.; Nakamura, H.; Ohta, T.; Nozoe, S.; Hirata, Y.; Sasaki, T. Lejimalides A and B, novel 24-membered macrolides with potent antileukemic activity from the Okinawan tunicate *Eudistoma* cf. *rigida*. *J. Org. Chem.* **1988**, *53*, 6147–6150. [[CrossRef](#)]
150. Kikuchi, Y.; Ishibashi, M.; Sasaki, T.; Kobayashi, J. Lejimalides C and D, new antineoplastic 24-membered macrolide sulfates from the okinawan marine tunicate *Eudistoma* cf. *rigida*. *Tetrahedron Lett.* **1991**, *32*, 789–797. [[CrossRef](#)]
151. Nguyen, M.H.; Imanishi, M.; Kurogi, T.; Wan, X.; Ishmael, J.E.; McPhail, K.L.; Smith, A.B. Synthetic access to the mandelalide family of macrolides: Development of an anion relay chemistry strategy. *J. Org. Chem.* **2018**, *83*, 4287–4306. [[CrossRef](#)]
152. Rinehart, K.L.; Holt, T.G.; Fregeau, N.L.; Stroh, J.G.; Keifer, P.A.; Sun, F.; Li, L.H.; Martin, D.G. Ecteinascidins 729, 743, 745, 759A, 759B, and 770: Potent antitumor agents from the Caribbean tunicate *Ecteinascidia turbinata*. *J. Org. Chem.* **1990**, *55*, 4512–4515. [[CrossRef](#)]
153. Izbicka, E.; Lawrence, R.; Raymond, E.; Eckhardt, G.; Faircloth, G.; Jimeno, J.; Clark, G.; Von Hoff, D.D. In vitro antitumor activity of the novel marine agent, Ecteinascidin-743 (ET-743, NSC-648766) against human tumors explanted from patients. *Ann. Oncol.* **1998**, *9*, 981–987. [[CrossRef](#)]
154. Berlmeck, R.G.S.; Britton, R.; Piers, E.; Lim, L.; Roberge, M.; Moreira Da Rocha, R.; Andersen, R.J. Granulatimide and isogranulatimide, aromatic alkaloids with G2 checkpoint inhibition activity isolated from the Brazilian ascidian *Didemnum granulatum*: Structure elucidation and synthesis. *J. Org. Chem.* **1998**, *63*, 9850–9856. [[CrossRef](#)]
155. Holmstrom, C.; James, S.; Egan, S.; Kjelleberg, S. Inhibition of common fouling organisms by marine bacterial isolates with special reference to the role of pigmented bacteria. *Biofouling* **1996**, *10*, 251–259. [[CrossRef](#)]
156. Holmstrom, C.; Rittschof, D.; Kjelleberg, S. Inhibition of settlement by larvae of *Balanus amphitrite* and *Ciona intestinalis* by a surface-colonizing marine bacterium. *Appl. Environ. Microbiol.* **1992**, *58*, 2111–2115. [[CrossRef](#)] [[PubMed](#)]
157. Stoecker, D. Resistance of a tunicate to fouling. *Biol. Bull.* **1978**, *155*, 615–626. [[CrossRef](#)]

158. Vervoort, H.C.; Pawlik, J.R.; Fenical, W. Chemical defense of the Caribbean ascidian *Didemnum conchyliatum*. *Mar. Ecol. Prog. Ser.* **1998**, *164*, 221–228. [[CrossRef](#)]
159. Lindquist, N.; Hay, M.E. Can small rare prey be chemically defended? The case for marine larvae. *Ecology* **1995**, *76*, 1347–1358. [[CrossRef](#)]
160. Lindquist, N.; Fenical, W. New tambjamine class alkaloids from the marine ascidian *Atapozoa* sp. and its nudibranch predators. Origin of the tambjamins in *Atapozoa*. *Experientia* **1991**, *47*, 504–506. [[CrossRef](#)]
161. McClintock, J.B.; Amsler, M.O.; Koplovitz, G.; Amsler, C.D.; Baker, B.J. Observations on an association between the dexamimid amphipod *Polycheria antarctica* f. *acanthopoda* and its ascidian host *Distaplia cylindrica*. *J. Crustac. Biol.* **2009**, *29*, 605–608. [[CrossRef](#)]
162. Watanabe, T.; Shibata, H.; Ebine, M.; Tsuchikawa, H.; Matsumori, N.; Murata, M.; Yoshida, M.; Morisawa, M.; Lin, S.; Yamauchi, K.; et al. Synthesis and complete structure determination of a sperm-activating and -attracting factor isolated from the ascidian ascidia *sydnei*ensis. *J. Nat. Prod.* **2018**, *81*, 985–997. [[CrossRef](#)]
163. Mikami, N.; Hosokawa, M.; Miyashita, K. Effects of sea squirt (*Halocynthia roretzi*) lipids on white adipose tissue weight and blood glucose in diabetic/obese KK-Ay mice. *Mol. Med. Rep.* **2010**, *3*, 449–453. [[PubMed](#)]
164. Thakur, N.L. *Studies on Some Bioactive Aspects of Selected Marine Organisms*; Goa University: Goa, India, 2001.
165. Kazlauskas, R.; Marwood, J.F.; Murphy, P.T.; Wells, R.J. A blue pigment from a compound ascidian. *Aust. J. Chem.* **1982**, *35*, 215–217. [[CrossRef](#)]
166. Wright, A.E.; Forleo, D.A.; Gunawardana, G.P.; Gunasekera, S.P.; Koehn, F.E.; McConnell, O.J. Antitumor tetrahydroisoquinoline alkaloids from the colonial ascidian *Ecteinascidia turbinata*. *J. Org. Chem.* **1990**, *55*, 4508–4515. [[CrossRef](#)]
167. Kott, P. Didemnid-algal symbiosis: Host species in the Western Pacific with notes on the symbiosis. *Micronesica* **1982**, *18*, 95–127.
168. Sings, H.L.; Rinehart, K.L. Compounds produced from potential tunicate-blue-green algal symbiosis: A review. *J. Ind. Microbiol. Biotechnol.* **1996**, *17*, 385–396. [[CrossRef](#)]
169. Carté, B.; Faulkner, D.J. Defensive metabolites from three nembrothid nudibranchs. *J. Org. Chem.* **1983**, *48*, 2314–2318. [[CrossRef](#)]
170. Garson, M.J. Marine natural products as antifeedants. In *Comprehensive Natural Products II: Chemistry and Biology*; Elsevier Ltd.: Amsterdam, The Netherlands, 2010; pp. 503–537. ISBN 9780080453828.
171. Lins, D.M.; de Marco, P.; Andrade, A.F.A.; Rocha, R.M. Predicting global ascidian invasions. *Divers. Distrib.* **2018**, *24*, 692–704. [[CrossRef](#)]
172. Gittenberger, A.; Moons, J.J.S. Settlement and possible competition for space between the invasive violet tunicate *Botrylloides violaceus* and the native star tunicate *Botryllus schlosseri* in The Netherlands. *Aquat. Invasions* **2011**, *6*, 435–440. [[CrossRef](#)]
173. Sephton, D.; Vercaemer, B.; Nicolas, J.M.; Keays, J. Monitoring for invasive tunicates in Nova Scotia, Canada (2006–2009). *Aquat. Invasions* **2011**, *6*, 391–403. [[CrossRef](#)]
174. Seo, K.S.; Lee, Y. A first assessment of invasive marine species on Chinese and Korean coasts. In *Biological Invasions in Marine Ecosystems, Ecological Studies 204*; Rilov, G., Crooks, J.A., Eds.; Springer: Berlin/Heidelberg, Germany, 2009; pp. 577–585.
175. Costello, K.E.; Lynch, S.A.; McAllen, R.; O’Riordan, R.M.; Culloty, S.C. The role of invasive tunicates as reservoirs of molluscan pathogens. *Biol. Invasions* **2021**, *23*, 641–655. [[CrossRef](#)]
176. Daigle, R.M.; Herbinger, C.M. Ecological interactions between the vase tunicate (*Ciona intestinalis*) and the farmed blue mussel (*Mytilus edulis*) in Nova Scotia, Canada. *Aquat. Invasions* **2009**, *4*, 177–187. [[CrossRef](#)]
177. Ramesh, C.H.; Koushik, S.; Shunmugaraj, T.; Murthy, M.V.R. Infestation of colonial ascidians on reef biota of Gulf of Mannar Marine Biosphere Reserve, India. *J. New Biol. Rep.* **2019**, *8*, 187–189.
178. Vargas-Ángel, B.; Godwin, L.S.; Asher, J.; Brainard, R.E. Invasive didemnid tunicate spreading across coral reefs at remote Swains Island, American Samoa. *Coral Reefs* **2009**, *28*, 53. [[CrossRef](#)]
179. Tatián, M.; Schwindt, E.; Lagger, C.; Varela, M.M. Colonization of Patagonian harbours (SW Atlantic) by an invasive sea squirt. *Spixiana* **2010**, *33*, 111–117.
180. Pleus, A.; LeClair, L.; Schultz, J.; Lambert, G. *2007–09 Tunicate Management Plan*; In Coordination with the Tunicate Response Advisory Committee; Washington State Department of Fish and Wildlife, Aquatic Invasive Species Unit: Olympia, WA, USA, 2008; pp. 1–64.
181. Cordell, J.R.; Levy, C.; Toft, J.D. Ecological implications of invasive tunicates associated with artificial structures in Puget Sound, Washington, USA. *Biol. Invasions* **2013**, *15*, 1303–1318. [[CrossRef](#)]
182. Abdul Jaffar, H.; Soban Akram, A.; Kaleem Arshan, M.L.; Sivakumar, V.; Tamilselvi, M. Distribution and invasiveness of a colonial ascidian, *Didemnum psammathodes*, along the southern Indian coastal water. *Oceanologia* **2016**, *58*, 212–220. [[CrossRef](#)]
183. Carman, M.R.; Grunden, D.W. First occurrence of the invasive tunicate *Didemnum vexillum* in eelgrass habitat. *Aquat. Invasions* **2010**, *5*, 23–29. [[CrossRef](#)]
184. Griffith, K.; Mowat, S.; Holt, R.H.F.; Ramsay, K.; Bishop, J.D.D.; Lambert, G.; Jenkins, S.R. First records in Great Britain of the invasive colonial ascidian *Didemnum vexillum* Kott, 2002. *Aquat. Invasions* **2009**, *4*, 581–590. [[CrossRef](#)]
185. Sorte, C.J.B.; Williams, S.L.; Zerebecki, R.A. Ocean warming increases threat of invasive species in a marine fouling community. *Ecology* **2010**, *91*, 2198–2204. [[CrossRef](#)]