

Influence of landscape connectivity on newt's response to a warmer climate

Laurane Winandy, Félix Pellerin, Lucie Di Gesu, Delphine Legrand, Julien

Cote

▶ To cite this version:

Laurane Winandy, Félix Pellerin, Lucie Di Gesu, Delphine Legrand, Julien Cote. Influence of landscape connectivity on newt's response to a warmer climate. Landscape Ecology, In press, 10.1007/s10980-023-01685-z . hal-04135879

HAL Id: hal-04135879 https://hal.science/hal-04135879

Submitted on 21 Jun2023

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 Influence of landscape connectivity on newt's response to a warmer climate

2 Laurane Winandy^{1,2}, Félix Pellerin², Lucie Di Gesu², Delphine Legrand³, Julien Cote².

¹ Laboratory of Ecology and Conservation of Amphibians (LECA), Freshwater and Oceanic Science Unit of
 Research (FOCUS), University of Liège, 4020 Liege, Belgium

 ⁵
 ² Laboratoire Évolution and Diversité Biologique (EDB), UMR5174, CNRS, IRD, Université Toulouse III Paul Sabatier, 118 Route de Narbonne, 31062 Toulouse, France

³ Station d'Écologie Théorique et Expérimentale (SETE), UAR2029, CNRS, 2 route du CNRS, 09200 Moulis,
 France

11

12 Landscape Ecology, 2023, https://doi.org/10.1007/s10980-023-01685-z

- 13 <u>Corresponding author:</u>
- 14 Laurane Winandy, <u>laurane.winandy@uliege.be</u> (0000-0001-6255-2503)
- **15** 0032479486161
- 16 Laboratory of Ecology and Conservation of Amphibians (LECA), Freshwater and Oceanic Science Unit of
- 17 Research (FOCUS), University of Liège, 4020 Liege, Belgium
- 18
- 19 <u>Co-author contacts:</u>
- 20 Felix Pellerin, <u>felix.pellerin23@gmail.com</u> (0000-0002-7062-4957)
- 21 Lucie Di Gesu, l.digesu@ecomail.fr
- 22 Delphine Legrand, <u>Delphine.LEGRAND@sete.cnrs.fr</u>
- 23 Julien Cote, julien.cote@univ-tlse3.fr (0000-0002-4453-5969)
- 24

25 Acknowledgements:

26 We thank the editor and the two anonymous reviewers for their constructive comments and suggestions,

- 27 Dominic Zugai for editing and reviewing this manuscript for English language, Olivier Guillaume, Thomas
- 28 Deruelles, Guillaume Toumi, Elvire Bestion and Audrey Trochet for their help or technical support in the
- 29 achievement of the project.

30 Author contributions:

31 JC, LW and DL designed the study. LW, FP and LDG performed the experiment. JC analyzed the pictures on

32 ImageJ. LW performed the statistical analysis and wrote the first version of the manuscript. All authors

33 contributed substantially to manuscript revisions and gave final approval for publication.

34 Funding:

- 35 This work was supported by funding from the European Research Council (ERC) under the European Union's
- Horizon 2020 research and innovation program (grant agreement no. 817779 to JC), by the French Laboratory of
 Excellence project 'TULIP' (grant nos. ANR-10-LABX-41 and ANR-11-IDEX-0002-02) and by an
- 38 'Investissements d'avenir' program from the Agence Nationale de la Recherche (grant no. ANR-11-INBS-
- 39 0001AnaEE-Services). LW was supported by the Fyssen Foundation Post-Doctoral Fellowship and is currently a
- 40 postdoctoral researcher at the Fonds de la recherche scientifique (F.R.S.-FNRS).
- 41
- 42 Conflict of interest:

43 The authors declare no conflict of interest.

44 Data availability:

- 45 The datasets analyzed during the current study are available from the corresponding author on reasonable
- 46 request.

47 Abstract

48 Context

Climate change and habitat fragmentation exert considerable pressures on biodiversity. The spatial distributionof microclimatic refuges in the landscape can influence species responses to warming climates.

- 51 Objectives
- 52 Using a semi-natural experiment, we investigated the potential synergetic effects of climate warming and habitat53 connectivity on a single amphibian species.
- 54 Methods

55 We monitored populations of the palmate newt, *Lissotriton helveticus*, under two climate treatments, a warmer

56 climate (+~2°C) or a present-day climate, in mesocosms either isolated or connected to the other climatic

57 conditions. We assessed the abundance and phenotype (snout-vent length, body condition and skin coloration:

- 58 darkness and redness) of juvenile and adult newts, and the dispersal propensity of juveniles.
- 59 Results
- 60 Over the 4 years of climatic manipulation, populations tended to increase more in the present-day than in the
- 61 warmer climate. Warmer climates decreased the abundance of adult newts and altered the phenotypic
- 62 composition of populations with darker and less red newts. However, connectivity between the two climates
- 63 cancelled out the effect of a warmer climate on abundance and reversed the effects on phenotype composition.
- 64 We further found that juvenile newts from the present-day climate treatment tended to emigrate more from
- warmer conditions during our laboratory dispersal assay and that population isolation disrupted a commoncovariation between emigration propensity and body size, leg length and skin darkness.
- 67 Conclusions

68 Our results point to a synergetic effect of climate warming and fragmentation on the demography of newt 69 populations and emigration decisions of juveniles, suggesting that microclimate refuges and their accessibility 70 play a key role in buffering the impacts of climate warming, with potential implications for amphibian diversity 71 at a regional scale.

72

73 Keywords: Amphibian, Global warming, Dispersal syndrome, Habitat fragmentation, Microclimate

- 74
- 75

76 Introduction

77 Global warming is one of the largest human-induced disturbances of natural ecosystems affecting biodiversity 78 worldwide (Díaz et al. 2019) and could lead to the extinction of 5 to 37% of all species (Urban 2015). Direct 79 evidence of warming-induced population extinction is scarce (Román-Palacios and Wiens 2020) because 80 organisms may respond to climate change through phenotypic plasticity, selection or relocation (McGaughran et 81 al. 2021). Climate change is indeed already changing the phenotypic composition and life history traits of 82 populations (Merilä and Hendry 2014), with phenological shifts occurring in many species (Parmesan and Yohe 83 2003) and an accelerated pace of life in ectotherm species (Bestion et al. 2015b; Burraco et al. 2020), for 84 example. These changes can alter population dynamics and increase the risk of population extinction, but they 85 can also facilitate species persistence under new conditions (Pellerin et al. 2022). Phenotypic plasticity and 86 genetic selection can result in behavioral, physiological and/or morphological adaptations to prevent extinction 87 (Huey et al. 2012). A notable and widespread example of phenotypic plasticity is thermal acclimation, which 88 allows species to extend their tolerance to higher temperatures (Seebacher et al. 2014).

89 Species can also shift their geographic range through dispersal towards suitable climatic conditions at 90 higher latitudes and/or elevations (Lenoir and Svenning 2015). Highly mobile organisms, such as birds and 91 butterflies, can travel long distances to locate suitable climatic conditions, while other taxa, such as amphibians 92 and reptiles, are more limited in their movement. These less mobile species need much more time to disperse to 93 distant cooler habitats (Sillero 2021), and may instead use locally variable habitats (i.e., microclimate refugia, 94 Suggitt et al. 2018). The survival of these organisms could therefore be more dependent on successful short-95 distance movements across climatically heterogeneous environments. Heterogeneous landscapes can offer a high 96 variety of microclimates (Milling et al. 2018), some of which are similar to conditions at higher latitude or 97 elevation, and these conditions can buffer the consequences of extreme temperatures (i.e., extirpation, Scheffers 98 et al. 2013; Suggitt et al. 2018). Dispersal therefore plays a central role in species' ability to locate habitats that 99 meet their thermal requirements at regional or local scales (Bestion et al. 2015a). Yet dispersal among 100 microclimates may vary by phenotype, and particularly by thermal requirements. Individuals with lower thermal 101 requirements might disperse towards cooler microhabitats, while more thermally tolerant individuals tend to 102 remain in or search for warmer environments (Bestion et al. 2015a; Pellerin et al. 2019). So-called adaptive 103 dispersal syndrome (Clobert et al. 2009) can lead to climate-induced shifts in phenotypic distributions, with 104 consequences for population persistence.

105 Climate-driven dispersal is structured by the spatial distribution of microclimate habitats, which are 106 becoming increasingly fragmented due to human activities (Opdam and Wascher 2004). By reducing landscape 107 connectivity, fragmentation may impede individual movements from temporarily or permanently local 108 detrimental conditions to refuge areas, and may force some species to move longer distances. We therefore 109 expect climate warming and habitat fragmentation to act synergistically to modify population responses to 110 climate change (Opdam and Wascher 2004). However, empirical evidence of this phenomenon is scarce (Urban 111 et al. 2014), necessitating new experimental approaches to test for the combined effects of habitat fragmentation 112 and spatial variability in climate on dispersal.

113 As ectotherms, amphibians are particularly sensitive to climate change because their body temperature, 114 and hence their physiological functions, strongly depend on environmental temperatures (Urban 2015). 115 Moreover, the limited dispersal abilities of many amphibians restrict their options to relocate to optimal climates 116 at regional scales, making them even more threatened by climate change (Urban et al. 2014). Yet thermal 117 acclimation through phenotypic plasticity can potentially reduce the impact of climate change. Previous studies 118 show that newts' locomotor activity, heat tolerance and metabolism rate can be acclimated to higher 119 temperatures (Gvoždík et al. 2007; Winterová & Gvoždík 2022). At a morphological level, changing body size 120 has emerged in many organisms as a response to climate warming but remains underexplored in amphibians (Li 121 et al. 2013). Melanin-based coloration plays an important role in thermoregulation in many animals, with 122 individuals of varying darkness presenting different levels of sensitivity to climate change: while dark skin may 123 provide protection against UV-radiation, it can also increase the risk of overheating (Roulin 2014). In 124 amphibians, skin coloration also results from carotenoid pigments (Mochida et al. 2013), which are acquired 125 through diet and are associated with improved individual fitness and may prevent pro-oxidative effects of 126 enhanced UV (Ogilvy and Preziosi 2012; Mochida et al. 2013). However, predicting the impact of climate 127 change in accordance with color traits depends mostly on the adaptive function of the coloration and how it 128 covaries with physiology and behavior in the studied species (Roulin 2014). For example, darkness is positively 129 correlated to boldness, exploration and dispersal in some species (Mafli et al. 2011; Mateos-Gonzalez and Senar 130 2012; Roulin, 2014; Saino et al., 2014) and would be expected to influence their responses to climate change, 131 particularly in fragmented landscapes.

132 Here, we experimentally assessed the interaction between warmer climate and landscape connectivity 133 on the abundance, phenotypic distribution and movement strategy of the palmate newt (Lissotriton helveticus). 134 We monitored populations of palmate newts inhabiting 24 semi-natural mesocosms undergoing either a warmer 135 (+~2C°) or a present-day climatic treatment across multiple years. Two mesocosms, one for each set of climatic 136 conditions, were paired and either isolated or connected with corridors. We assessed the impact of these 137 conditions on juvenile and adult population abundance and phenotypic traits. We predicted newt abundance 138 would be lower in warm and isolated populations, where newts could not evade the warming temperatures. We 139 also measured morphological traits (body size, body condition), coloration (darkness and redness) and dispersal 140 propensity and their covariation with temperature and phenotype. We predicted phenotypic differences between 141 newts experiencing different climatic conditions, particularly in isolated populations, where the lack of 142 connectivity prevents them from evading warming temperatures. More specifically, we predicted a reduction in 143 body size and body condition at warmer temperatures, as has been shown in previous studies (Reading 2007). 144 We expected that thermal stress would decrease the red coloration, while the expected effects on skin melanism 145 were less clear since darkness of color can have either positive (UV protection) or negative (overheating risk) 146 effects on thermoregulation.

Finally, we measured the emigration propensity of juvenile newts at two temperatures in a laboratory setting and its covariation with phenotypic traits (*i.e.*, dispersal syndrome). We expected newts coming from present-day climate populations to emigrate more from warmer temperatures than newts that were exposed initially to warmer conditions (*i.e.*, from warmer climates), suggesting that newts coming from warmer climates would be acclimated to warmer temperatures. While emigrants are expected to have greater body and leg length and to be darker (*e.g.*, Winandy et al. 2019), we expected challenging environmental conditions experienced in warm and/or isolated populations to disrupt dispersal syndrome (*e.g.*, Cote et al. 2017, 2022), possibly inciting
all individuals to emigrate regardless of phenotype (Legrand et al. 2015).

155 Material and methods

156 *Model species*

157 The palmate newt (Lissotriton helveticus) is a small salamandridae (i.e., tailed amphibian) occurring in a wide range of habitats in Europe, from wooded to more open areas such as meadows, field edges and gardens. The 158 159 palmate newt has an aquatic larval stage before undergoing metamorphosis into terrestrial juveniles. Juveniles 160 usually stay on land for two to three years before reaching sexual maturity and returning to the water to breed. 161 Adult newts have strictly aquatic reproduction and remain in the water during the mating period which lasts several months. As home ranges of newts are quite small (< 50.000 m² (Verell 1987)), aquatic breeding sites 162 163 must be close to the terrestrial habitat (Joly and Miaud 1989). Breeding sites range from small puddles to larger 164 ponds or even lakes and are usually stagnant, permanent or filled with water for at least three months covering 165 the period of reproduction and larval development. On land, newts are usually found sheltering under logs or 166 stones or in small burrows.

167 Study site and experimental design

168 The study took place in the Station d'Ecologie Théorique et Expérimentale in Moulis (France). The Station has a 169 national agreement for use of animals in the laboratory (number B09583), and our experiments were conducted 170 in accordance with French ethics regulations (Ethics permit number APAFIS#30486-2021031815341692v4). 171 The newts were caught in the Metatron under license number 09-2016- 02. At the end of the experiment, they 172 were released exactly where they were caught, following recommendations of the permit.

173 For this study, we used populations of palmate newts naturally occurring in the Metatron, which is a 174 system of 48 semi-natural and interconnected mesocosms in which climatic conditions can be manipulated 175 (Legrand et al. 2012). Mesocosms are large (10 x 10 m), delimited by tarpaulins buried 30 cm into the ground, 176 and fully enclosed with a net allowing the isolation of each enclosure. This system prevents both the escape of 177 the study species and predation from birds and snakes present outside of the Metatron. Each mesocosm acts as a 178 mini-ecosystem of a wet meadow with dense natural vegetation, two permanent pools (30 x 50 x 15cm), shelters 179 (logs and rocks) and large invertebrate communities. They therefore offer all the ecological features required for 180 the different life stages of palmate newts. Temperature, light intensity and hygrometry in each enclosure are 181 monitored every 30 minutes and can be manipulated using motor-driven shutters and a sprinkler system. In 182 addition, mesocosms can be connected by 19 m-long corridors allowing species to move between mesocosms. 183 Newts usually exhibit strong breeding site fidelity, with individuals frequently returning to their natal site (Joly 184 and Miaud 1989) and thus movements between mesocosms and breeding patches could be considered as 185 dispersal (Clobert et al. 2009). While breeding adult newts usually disperse between twenty to a few hundred 186 meters (Cayuela et al. 2018; Denoël et al. 2018), natal dispersal distances are unknown and likely higher as 187 juvenile newts wander on land for several years (Pittman et al. 2014). While the corridor length in the 188 mesocosms does not allow for distinguishing dispersal from migration movements, the harsh conditions in 189 corridors (*i.e.*, a narrow width devoid of shelter and/or puddles) make the crossing challenging for newts and 190 ensures that movements between mesocosm are not routine foraging movements (Fronhofer et al. 2018).

191 In this study, we used 24 mesocosms in which we applied the climatic treatment for four years (2013-192 2017) and the connectivity treatments for two years (2015-2017). The climatic treatment was created with 193 shutters that automatically closed above certain temperature thresholds: 28°C for the present-day climate and 194 38°C for the warm climate. Mesocosms were intrinsically warmer than outside because of the greenhouse effect 195 associated with the nets. The present-day climate treatment created temperatures similar to the mean temperature 196 outside of the Metatron (Bestion et al. 2015a). Since climatic treatments depended on outdoor climatic 197 conditions, they were efficient only during daytime in summer (from mid-June to October), with variation within 198 and between days. This resulted in a 1-2°C difference in mean summer daily temperatures between the present-199 day and warm climates from June 2013 to July 2017 ($\chi^2_1 = 77.12$, P < 0.001, the mean \pm SE are 26.49 \pm 0.10°C 200 and 27.76 \pm 0.11°C for present-day and warm climate treatments respectively). There was also a 2-3°C 201 difference of the summer maximum temperature ($\chi^2_1 = 135.62$, P < 0.001, the mean \pm SE are 29.43 ± 0.13 °C and 202 $31.88 \pm 0.20^{\circ}$ C for present-day and warm climate treatments respectively). The warm climate treatment matched 203 the scenario SSP5-8.5 for 2041–2060 and SSP2-4.5, SSP3-7.0 and SSP5-8.5 for 2081–2100 (Masson-Delmotte 204 et al., 2021). While most global warming studies focus on the effects of a strict increase in mean temperatures, 205 an increase in the magnitude of daily temperature fluctuation (*i.e.*, difference between the maximum and the 206 minimum day temperature) is also predicted (Thompson et al. 2013). Accordingly, in our study, there was also 207 an effect of treatments on the difference between the maximum and the minimum temperature during summer $(\chi^2_1 = 124.96, P < 0.001)$, the mean \pm SE are $15.39 \pm 0.15^{\circ}$ C and $17.94 \pm 0.20^{\circ}$ C for present-day and warm 208 209 climate treatment respectively). During the climate treatment, all mesocosm were watered with soft water every 210 day in the evening to avoid a drying effect of the treatment.

We used 12 pairs of present-day and warm climate mesocosms that were either connected by opening the corridor between them (*i.e.*, for connected treatment) or not to simulate habitat fragmentation (*i.e.*, isolated treatment). The six pairs of each connectivity treatment were randomly distributed across the system.

214 *Population monitoring in the Metatron*

215 After the construction of the Metatron in 2009, all mesocosms remained open for an entire year and, between 216 2010 and 2012, for 2-3 weeks a year to allow colonization by local species (e.g., plants, insects, amphibians, 217 mollusks) present on the site. The palmate newts within habitats around the Metatron rapidly colonized and 218 established themselves inside the mesocosms. In 2012, the 24 mesocosms were closed for a long-term 219 experiment on common lizards (see Bestion et al. 2015b; Pellerin et al. 2022) and migration to and from the 220 mesocosms was thus impossible. The climatic treatment began in 2013 while the connectivity treatment began in 221 2015. Newt abundance in each mesocosm was estimated annually beginning in 2013 with a standardized 222 sampling effort. Our climate treatments were only applied in the summer (from the end of June to the beginning 223 of October) and during the warmest times of the day, while our sampling took place in the early mornings in 224 June, prior to climate treatments. Consequently, we assumed our probability of detection did not differ across 225 climate treatments. Between 2013 and 2016, the sampling consisted of two observers searching for newts in the 226 vegetation during 10 min on the four sides of each mesocosm, where newts were localized due to the presence of 227 moss providing moist conditions.

In June 2017, we estimated newt abundance with a more thorough sampling. To estimate juvenile newt abundance, we did three sessions (on different days) of standardized sampling early in the morning, when there was still morning dew and juvenile newts were more active. Each capture session was conducted by the same two people searching for newts for 10 minutes in each mesocosm. After each session, all newts captured were brought back to the lab. As described above, the climate treatment was not applied during early June and dawn was coolest and wettest part of the day, thus we assumed that there was no difference in detection probability between treatments during sampling. Yet the large size of the mesocosm and the dense vegetation cover likely prevented detection and capture of all juvenile newts. Therefore, the number of captured juveniles in this study represents an estimate of abundance that is comparable between treatments due to equal sampling effort.

With respect to adult newts, we ran two capture sessions on different days by dipnetting for 5 minutes in each of the two pools available to newts in each mesocosm. In June, most adults inhabit water for reproduction purposes. Given the small size of the pools (30 x 50 x 15 cm), we were confident of capturing all adults present in the water at that time. Therefore, we feel that our abundance estimates are likely to match the actual number of adults in each mesocosm.

242 Housing conditions and phenotype assessment

243 From the sampling above, we kept 67 randomly selected juvenile newts from 16 mesocosms in the lab: 31 newts 244 were from seven mesocosms in warm climate (n = 15 in isolated and n = 16 in connected treatments); 36 newts 245 were from nine mesocosms in present-day climate (n = 18 in isolated and n = 18 in connected treatments). The 246 remaining juveniles were released back to the mesocosms after a maximum of three days spent in the lab. The 67 247 juveniles were identified using dorsal and ventral pictures (see below) and housed in individual terraria (25 x 248 15.5 x 15 cm) filled with natural moss, an egg carton as a shelter, and a small pool (3 cm diameter 1 cm depth). 249 Newts were fed five micro-crickets (2-3 mm, Acheta domestica) daily. A natural neon light provided light from 250 9am to 7pm and the temperature of the room was set at 18°C, which is in the range of preferential temperatures 251 for European newts (Balogová & Gvoždík 2015; Gvoždík 2015). All terraria were sprayed with water twice a 252 day to maintain a high humidity level. Juvenile newts were housed in common environmental conditions for 2 253 weeks prior the dispersal test (see below) to reduce the effect of stressful new condition on their emigration rate.

We also randomly selected 86 adult newts: 32 from nine mesocosms with the warm climate (n = 6 in isolated and n = 26 in connected treatments); 54 from 10 mesocosms with the present-day climate (n = 28 in isolated and n = 26 in connected treatments). Adult newts were kept in the lab to measure morphological traits for several days before being released back in the mesocosms. Newts were housed in aquaria (35 cm L × 18 cm l × 20 cm water deep) with a maximum of four individuals from the same mesocosm per aquarium. We provided a shelter and an egg laying substrate (strips of filter paper) and fed individuals 50 mg of thawed *Chironomus* larvae per newt daily to create optimal conditions for palmate newts during aquatic reproduction.

261 Morphological traits

We measured snout-vent length and total length of newts using a ruler (measurement to the mm) and weighed (to 0.001 g) the selected newts (n = 67 juveniles and n = 86 adults) on the day of their capture. The body mass of newts highly depends on their degree of hydration, so we made sure to weigh fully hydrated newts. Adult newts in their aquatic phase, are obviously fully hydrated. For the juveniles, just after capture, they were maintained in boxes filled with soaked paper towels, ensuring their proper hydration. We calculated the body condition of newts using the mass/snout-vent length residuals. We took ventral and dorsal photographs to assess skin 268 coloration of the 67 juveniles. However, due to a technical issue we were only able to photograph 27 adult newts 269 [warm climate: 12 newts from 7 mesocosms (2 in isolated and 10 in connected treatments), present-day climate: 270 15 newts from 7 mesocosms (9 in isolated and 6 in connected treatments)]. For photographs, each newt was 271 carefully positioned on a computer scanner (Canon CadoscanLide110) lined with high density foam to avoid 272 injuries. Photographs were taken at a resolution of 400 dpi and analyzed using ImageJ software. We delimited a 273 zone for the dorsal (from the top of the neck to the posterior legs) and ventral areas (from the throat to the 274 posterior legs). The photographs were transformed in shades of grey and then in black and white according to a 275 fixed grey threshold of 60 chosen to maximize variance in darkness among individuals. A mean grey value and a 276 percentage of black pixels in each zone were computed. We further extracted HSL values (Hue Saturation 277 Lightness, Endler 1990). Hue, often referred to as 'color' (i.e., the difference between green, red or blue), 278 corresponds to the angular position on a color circle where 0° corresponds to red. Saturation refers to the 279 intensity of a hue, from grey tone (scored as 0) to pure and vivid color (scored as 1). Lightness is the luminance 280 (a photometric measure of the luminous intensity per unit area of light travelling in a given direction) ranging 281 from dark (scored as 0) to light (scored as 1). On the ventral images we also measured hindlimb length (to the 282 nearest mm) of juvenile newts, as leg length is a locomotor trait potentially linked to emigration propensity 283 (Denoël et al. 2018; Winandy et al. 2019).

284 Using the FactoMinR package (Lê et al. 2008), adult and juvenile darkness was scored with Principal 285 Component Analyses (PCA) on the mean grey, the percentage of black pixels, and the lightness values of both 286 dorsal and ventral photographs (mean and variance scaled). In adults, the PC1 axis explained 52.6% of variance, 287 had an eigenvalue of 3.16, and was positively correlated to dorsal and ventral lightness and mean grey, and 288 negatively correlated to the percentage of black pixels (Table S1). In juveniles, the PC1 axis explained 57.3% of 289 variance, had an eigenvalue of 3.43, and was positively correlated to dorsal and ventral lightness and mean grey, 290 and negatively correlated to the percentage of black (Table S1). To obtain a darkness score ranging from pale to 291 dark, we multiplied scores by -1.

We scored the red coloration of juvenile newts with PCA using the dorsal and ventral hue and saturation values (variance and mean scaled). The PC1 axis explained 54.4% of variance, had an eigenvalue of 2.17, and was positively correlated to dorsal and ventral hue, and negatively correlated to ventral and dorsal saturation (Table S1). To obtain a redness score (Fig. 1), from grey tone yellow to pure red, we multiplied scores by –1. We did not measure redness in adult newts as they have a beige-brown coloration.



Fig. 1 Coloration of juvenile newts. Redness scores scale from high hue and low saturation (grey-tone yellowcoloration) to low hue and high saturation (pure red), photography by L. Winandy.

300

301 *Emigration propensity*

302 Emigration propensity was assessed only in juvenile newts two weeks after their capture, allowing a period of 303 acclimation to common garden conditions in the laboratory. We used eight experimental systems placed in a 304 greenhouse with controlled temperature and natural light for this experiment. Each system was made of two 130 305 L plastic containers 78 x 56 x 43 cm) connected by a circuitous plastic pipe (diameter: 10 cm, total length: 4.4 306 m). The departure container was filled with soil litter, with several terrestrial shelters and a pool (25 cm 307 diameter, filled with water and natural moss). The corridor (plastic pipe) was directly connected to the departure 308 container and filled with a thin layer of moist soil. To go from the departure to the arrival containers, juvenile 309 newts had to enter the plastic pipe (i.e., the corridor) and fall into the arrival container which was filled with 310 natural moss. Once in the arrival container, newts could not re-enter the corridor. This system has been used in 311 previous studies to assess palmate newt's emigration decisions (Fronhofer et al. 2018; Cote et al. 2022). Here we 312 measured the emigration probability of each individual at two different temperature regimes: a warm 313 temperature (with a daytime temperature of 30.7 ± 0.27 °C and a nighttime temperature of 24.6 ± 0.07 °C), and a 314 cooler temperature (with a daytime temperature of $22.83 \pm 0.18^{\circ}$ C and a nighttime temperature of $20.6 \pm 0.3^{\circ}$ C). 315 These temperatures corresponded to local natural temperatures at this time of the year (*i.e.*, in July).

316 Assays were run on small groups (4-5 newts per system) to prevent high density in departure 317 populations. We carried out two temporal replicates of eight experimental groups each. Juvenile newts from each 318 climatic treatment and mesocosm were homogenously distributed between the temporal replicates. Half of the 319 newts were tested first at the cooler temperature and the other half were tested first at the warmer temperature. 320 Replicates were done several days apart, and the moss and water pools were changed to limit the risk of scent 321 remaining inside the system. Ten days after the first assays, we repeated the assay under the other temperature 322 treatment. For each assay, groups of newts were released in the departure tank in the late afternoon for a 24-h 323 acclimation period to the system and thermal conditions. Food (micro-crickets) was available ad libitum in the 324 departure tank. During the acclimation period, the dispersal pipes were closed to prevent any stress-induced 325 movement from the departure tank. We then opened the pipes for one night as juvenile newts usually move from 326 dusk to dawn. The next morning, we closed all dispersal pipes, captured residents and emigrants (*i.e.*, newts 327 found in departure and arrival tanks, respectively) and identified individuals using the ventral and dorsal 328 photographs. No newts were found inside the dispersal pipes.

- 329 Statistical analysis
- 330 Abundance

Firstly, we ran a generalized linear mixed model (GLMM) using a poisson family (with a log link function) to assess the effect of climatic treatments (warm *vs.* present-day) on the evolution of newt abundance across years (from 2013 to 2017). The mesocosm identity (Mesocosm_ID) was included as a random intercept. Since the connectivity treatment only began in summer 2015, we checked that there was no prior effect of this treatment on newt abundance using a a generalized linear model (GLM), using a quasipoisson family to correct for overdispersion, and testing the effect of connectivity treatment (isolated *vs.* connected) and its interaction withclimatic treatment on newt abundance every year from 2013 to 2015.

Secondly, we analyzed abundance in 2017, in which sampling was more extensive than previous sampling and took place after two years of connectivity treatment. We ran a GLM using a quasipoisson family for adult and juvenile abundance, testing the effect of the climate treatments, the connectivity treatment, and their interaction as fixed effects. Since mesocosms were not seeded with the same number of individuals prior to the start of the experiment, we added the abundance of newts in 2013 as a covariate. For juvenile abundance, we also added the adult abundance as a covariate to prevent multiple testing issues as juvenile and adult abundances should be positively correlated.

345 Phenotype

We ran linear mixed models (LMM) for morphological traits [body size (*i.e.*, snout-vent length), body condition 346 347 (i.e., mass/size residuals) and darkness for adults and juveniles plus leg length (using leg length/body size 348 residuals) and red coloration for juveniles]. All models included the climate treatments, the connectivity 349 treatment, and their interaction as fixed effects. The mesocosm identity (Mesocosm_ID) was included as a 350 random intercept. For adult body size and body condition, the sex and its interaction with treatments were included. Redness and darkness were positively correlated ($F_{1,65} = 18.91$; P < 0.001), so for both variables we 351 352 used the residuals of the regression models (residuals of redness on darkness for the redness variable and 353 darkness/redness residuals for the darkness variable) to obtain an independent analysis of color parameters. For 354 coloration traits, we also added body size (*i.e.*, snout-vent length) as a covariate in the models.

355 *Emigration probability*

356 We used a generalized linear mixed model (GLMM) with a binomial error distribution and a logit link function 357 for the emigration status of juveniles (*i.e.*, 0 = resident, 1 = emigrant), with climate, connectivity, and 358 temperature treatments during the emigration assay and all interactions between these factors as fixed effects. 359 Individual identity (Newt_ID), experimental group (Group_ID) and the mesocosm identity (Mesocosm_ID) were 360 added as random intercepts. The order of passage (warm or cooler temperature first) as well as the temporal replicate (*i.e.*, first or second block of eight experimental groups) did not affect the emigration rate ($\chi^2_1 = 0.355$, 361 P = 0.552; $\chi^2_1 = 0.624$, P = 0.43, respectively) and were therefore removed from the model to avoid over 362 363 parameterization.

364 Dispersal syndromes

We performed a multivariate analysis of variance (MANOVA) to assess the effects of treatments and emigration status on newt phenotypes. The dependent variables were body size, leg length (using leg length/body size residuals), darkness (using darkness/redness residuals), and redness (using redness/darkness residuals). These variables were centered and scaled. Fixed effects included climate treatment, connectivity treatment, and their interactions with the emigration status. In a second step, we ran univariate analysis on the four traits separately.

370 *General procedure*

- Analyses were performed in R 3.5.1 (<u>www.r-project.org</u>). We checked model assumptions by visually screening
- the residuals for normality and homoscedasticity. Body size for adult newts and body condition for juvenilenewts were log-transformed to achieve normality. We used lme4 (Bates et al. 2008) and sjstats (Lüdecke and
- 374 Lüdecke 2019) packages for all models and estimated the significance of factors with Wald χ^2 and likelihood
- 375 ratio tests. We used a backward selection procedure (with *a priori* level of significance of 0.05) to remove non-
- 376 significant interactions and covariates, while keeping the main factors of interest (*i.e.* the climatic and
- 377 connectivity treatments) in the models. After the model selection, we used the performance package (Lüdecke et
- al. 2021) to calculate adjusted R^2 for GLMs and conditional R^2 (R^2_c , effect of the fixed and random effects) and
- marginal R^2 (R^2_m , effect of the fixed effect) for LMMs and GLMMs. For mixed models, when the variance of a
- random effect was equal to 0 (*i.e.*, $R_c^2 = NA$), the random variable of concern was removed from the model and
- the adjusted R^2 was calculated if there were no more random variables included in the model. When we found a significant interaction between climate and connectivity treatments, we investigated the influence of climate in
- each connectivity treatment by separately running the models for each connectivity treatment.

384 **Results**

385 Abundance

During the 4 years of study, we found no effect on abundance from the climate treatment ($\chi^2_1 = 0.201$, P = 0.65). 386 However, we found a strong effect for the specific year ($\chi^2_4 = 330.331$, P < 0.001), showing an increase of newt 387 388 abundance over time and a marginal interaction between year and climate treatment ($\chi^2_4 = 8.834$, P = 0.065), 389 with newt abundance increasing slightly more in the present-day climate than in the warm climate (Fig. 2A). The conditional and marginal R^2 of the model were $R_c^2 = 0.889$ and $R_m^2 = 0.36$, respectively, and the variance of the 390 391 mesocosm identity was 1.532±1.238SD. Before 2017, there was no effect of connectivity treatment or any 392 interaction with climate treatment on newt abundance (P>0.2 for all years from 2013 to 2015, see Table S2 for 393 details).

394 In 2017, over all mesocosms, we captured 187 juvenile newts and 118 adults (52 males and 66 females, 395 see Table S3 for details on the number of newts captured in each mesocosm). We captured 80% of adults in the 396 water during the first session and 13% during the second session. Only 7% of adults were captured on land 397 during the 3 sessions of juvenile sampling (*i.e.* n = 8; 4 newts in each climate treatment), confirming that finding 398 adult newts on land was rare at this time of the year. We found an interaction between the effects of climate and 399 connectivity treatments on the abundance of adult newts (Table 1). In isolated populations, newt abundance was much lower in the warm climate treatment than in the present-day treatment (Fig. 2B; $\chi^2_1 = 7.182$, P = 0.007), 400 401 while there was no difference in the connected populations (Fig. 2B; $\chi^2_1 = 0.13$, P = 0.718). We found no effect 402 of treatment on juvenile abundance, but there was a positive relationship between juvenile and adult abundance 403 (Table 1). The abundance of adult newts in 2013 and 2017 were positively correlated, but the abundance of 404 adults in 2013 and juvenile abundance in 2017 were not related (Table 1).

405

Table 1: Results of GLM (after backward selection procedure) testing the effect of climate treatment, connectivity treatment, and their interaction on the abundance of adult ($R^2_{adj} = 0.903$) and juvenile newts ($R^2_{adj} = 0.227$) in 2017. Estimates for climate and connectivity are given for the present-day climate and the connected population, respectively.



412 Fig. 2 Effect of climate treatments (mean ± SE) on newt abundance across years (A), and of climate and
413 connectivity treatments on adult newt abundance in 2017 (B).

- 414
- 415 Phenotypic differences

416 The climate and connectivity treatments alone or in interaction did not influence body size, leg length or body 417 condition of adult or juvenile newts (Table S4). Adult newts were significantly darker in warm than in present-418 day climates (Table 2, Fig. 3A), while connectivity treatments had no effect (Table 2). There was an interaction 419 between climate and connectivity treatments on juvenile darkness (Table 2, Fig. 3B): in connected populations, 420 newts were darker in the present-day climate than in the warm climate ($\chi^2_1 = 4.564$, P = 0.033). However this 421 was not the case for isolated populations ($\chi^2_1 = 0.740$, P = 0.39). There was an interaction between climate and 422 connectivity treatments on juveniles' redness (Table 2, Fig. 3C): in isolated populations, newts were redder in 423 the present-day climate than in the warm climate ($F_{1,31} = 6.462$, P = 0.016), while in connected populations newts 424 tended to be redder in the warm climate than in the present-day climate ($F_{1,32} = 2.764$, P = 0.106). Body size had 425 no effect on coloration traits in adults or in juveniles (P > 0.1 in all models) and was therefore removed during 426 model selection.

427

Table 2: Results of LMM (after backward selection procedure) testing the effect of climate treatment, connectivity treatment, and their interaction on the darkness of adults (n = 27, $R_c^2 = 0.350$, $R_m^2 = 0.206$) and on darkness and redness of juveniles (n = 67, $R_c^2 = 0.238$, $R_m^2 = 0.083$ and $R_{adj}^2 = 0.12$). Estimates for climate and

431 connectivity are given for the present-day climate and for the connected population, respectively.

Newt coloration	Factors	Estimates	SE	Test value	CI (95%)	Р			
Adult					(-3.07, -0.15)				
Darkness	Climate	-1.609	0.781	$\chi^2_1 = 4.247$	(-1.3, 1.66)	0.039			
	Connectivity	0.180	0.790	$\chi^2_1 = 0.052$		0.820			
	1 Mesocosm_ID		.716SD						
Juvenile									
Darkness	Climate	-0.928	0.742	$\chi^2_1 = 0.0004$	(-2.23, 0.36)	0.985			
	Connectivity	-1.31	0.782	$\chi^2_1 = 0.269$	(-2.68, 0.05)	0.604			
	Climate* Connectivity	1.847	1.042	$\chi^2_1 = 3.968$	(0.04, 3.71)	0.046			
	1 Mesocosm_ID Variance = 1.35±1.16SD								
Juvenile	Climate	1.125	0.430	$F_{1,63} = 0.739$	(0.27, 1.98)	0.393			
Redness	Connectivity	1.445	0.442	$F_{1,63} = 3.105$	(0.56, 2.33)	0.083			
	Climate* Connectivity	-1.703	0.602	$F_{1,63} = 8.002$	(-2.91, -0.50)	0.006			
	1 Mesocosm_ID	Mesocosm_ID Variance = 0*							





434



436 Fig. 3 Effect of treatments on newt coloration (mean \pm SE): Effect of climate treatment (red bar for warm 437 climate and grey bar for present-day climate) on adult newt darkness (A), and of climate and connectivity 438 treatments on juvenile newt darkness (B) and on juvenile newt redness (C).

439

435

440 Emigration probability and dispersal syndromes

While connectivity treatments had no effect on emigration probability and dispersal syndromes, we found a marginal interaction between climate and assay temperature treatments (Table 3, Fig. 4), showing that newts coming from the present-day climates tended to emigrate more often in warm conditions than in cooler conditions, but that the difference was not significant ($\chi^2_1 = 2.300$, P = 0.129). Newts coming from the warm climate emigrated similarly in warm and cooler conditions ($\chi^2_1 = 0.512$, P = 0.474).

446

447 **Table 3:** Results of GLMM (after backward selection procedure) testing the effect climate treatment, 448 connectivity treatment, temperature treatment, and their interaction on the emigration probability of juvenile 449 newts (n = 67, $R_c^2 = 0.990$, $R_m^2 = 0.013$). Estimates for climate, connectivity and temperature are given for the 450 present-day climate, for the connected population, and for cooler temperatures, respectively.

Response	Factors	Estimates	SE	χ^2_1	CI (95%)	Р	
Emigration	Climate	-0.2	1.681	0.014	(-3.5, 3.1)	0.991	
probability	Connectivity	-0.018	1.679	0.0001	(-3.31, 3.27)	0.905	
	Temperature	-1.298	1.003	1.675	(-3.26, 0.67)	0.196	
	Climate*Temperature	-6.342	3.456	3.368	(-13.11, 0.43)	0.066	
	1 newt_ID	Variance = 311.3 ± 17.64 SD					
	1 Group_ID	Variance = 0^*					
	1 Mesocosm_ID	Variance $= 0^*$					

451 *Since the variance of the experimental group (Group_ID) and the mesocosm identity (Mesocosm_ID) were

452 equal to 0, these were removed from the final model.

453





Climate treatment Fig. 4 Interactive effect of climate treatment and temperature treatment (*i.e.*, applied during the movement assay) 455 456 on newt emigration probability (mean \pm SE).

457 The multivariate analysis revealed a significant interaction between emigration status and connectivity 458 treatment (Table 4, Fig. 5). Compared to residents in connected populations, emigrants were bigger (Fig. 5A), 459 had longer legs (Fig. 5B), and were darker (Fig. 5C), while in isolated populations they had shorter legs (Fig. 460 5B) and were redder (Fig. 5D). The univariate analyses showed that this interaction was particularly significant for darkness and leg length (Table 4). Emigrants were darker than residents in connected populations ($F_{(1,65)}$ = 461 462 4.39, P = 0.04, Fig. 5A) but not in isolated populations ($F_{(1,58)} = 0.90$, P = 0.347, Fig. 5A). Emigrants had slightly 463 but non-significantly longer legs than residents in connected populations ($F_{(1,65)} = 2.44$, P = 0.123), Fig. 5B), 464 while in isolated populations emigrants had slightly shorter legs than residents ($F_{(1,55)} = 3.80$, P = 0.056, Fig. 5B). 465 The univariate analysis also revealed that emigrants tended to be redder than residents (mean \pm SE 0.21 \pm 0.14 466 and -0.44 ± 0.29 , respectively, Table 4). Finally, the univariate analysis also revealed a marginal interaction 467 between climate treatment and emigration status on leg length (Table 4). In the warm climate, residents tended to have longer legs than emigrants (mean \pm SE 0.45 \pm 0.010 and 0.42 \pm 0.010, respectively, F_(1.56) = 3.21, P = 468 469 0.079), while residents and emigrants had similar leg length in the present-day climate (mean \pm SE 0.40 \pm 0.01 470 and 0.43 ± 0.01 , respectively; $F_{(1,67)} = 0.41$, P = 0.526).

471

472 Table 4: Results of the multivariate analysis (MANOVA) and univariate analysis (ANOVAs) testing the effect

473 of emigration status and its interaction with the climate and the connectivity treatment on newt traits. Significant

474 and marginal effects are respectively highlighted in bold and underlined.

	Emigration status		Emigration status *		Emigration status *		Emigration status *	
			Connectivity		Climate		Connectivity * Climate	
	F-test	Р	F-test	Р	F-test	Р	F-test	Р
Manova								
All traits	$F_{(4,119)} = 1.72$	0.149	$F_{(4,119)} = 3.63$	0.008	$F_{(4,119)} = 1.62$	0.175	$F_{(4,119)} = 0.44$	0.780
Anovas								
Body size	$F_{(1,119)} = 2.32$	0.130	$F_{(1,119)} = 0.52$	0.473	$F_{(1,119)} = 0.24$	0.624	$F_{(1,119)} = 0.07$	0.797
Leg length	$F_{(1,119)} = 0.10$	0.747	$F_{(1,119)} = 6.62$	0.011	$F_{(1,119)} = 3.74$	<u>0.055</u>	$F_{(1,119)} = 1.05$	0.308
Darkness	$F_{(1,119)} = 0.40$	0.529	$F_{(1,119)} = 5.68$	0.019	$F_{(1,119)} = 0.21$	0.651	$F_{(1,119)} = 0.52$	0.472
Redness	$F_{(1,119)} = 3.64$	<u>0.059</u>	$F_{(1,119)} = 2.35$	0.128	$F_{(1,119)} = 1.83$	0.179	$F_{(1,119)} = 0.27$	0.604
475								



477 Fig. 5 Interactive effects of connectivity treatment (isolated versus connected) and emigration status (resident

478 *versus* disperser) on newt phenotypes (mean ± SE): body size (A), leg length (B), darkness (C) and redness (D).

479

476

480 Discussion

Using experimental mesocosms to simulate natural habitats, we investigated whether population and phenotypic responses of palmate newts to a ~2°C warmer climate depended on the connectivity between warmer and cooler habitats. We found that warmer climates negatively influenced the abundance of newts, altered the phenotype composition of populations, and influenced the emigration propensity in juvenile newts. As predicted, connectivity between thermal habitats negated or reversed these main effects and added a distinct effect on dispersal syndromes. Our results highlight the importance of considering habitat connectivity and microclimates in predicting species responses to warming climates.

488

489 Population abundance and phenotypic composition

490 Newt abundance decreased over four years in mesocosms with warmer climates as has been demonstrated in 491 previous studies where climate warming reduced newt survival and increased the risk of population extinction 492 (Li et al. 2013; Weinbach et al. 2018). The change in newt abundance differed by life-stage, as only adult 493 abundance was reduced in warmer climates. However, our results must be interpreted cautiously as our initial 494 abundance estimates were based on natural colonization of newts rather than establishing standardized 495 population sizes across all mesocosms. Nonetheless this initial abundance was not different across the two 496 climate treatments and our results provide evidence suggesting a negative impact of warm climate on population 497 growing over time. Warmer climates could have impacted adults more than juveniles because adults may not be 498 able to compensate for the energy expenditures induced by warmer conditions. Indeed, unlike juveniles, adult 499 newts may spend part of the summer in the aquatic habitat to avoid hot and dry conditions in the terrestrial 500 habitat (Denoël 2006). However, such habitat shifts might include a cost in depletion of caloric reserves as the 501 metabolic rate of newts is higher in water than on land (Kristín and Gvoždík 2014). Adults may therefore enter 502 hibernation with a lower body condition, in turn impacting their survival. Alternatively, warm climates could 503 provoke an ageing effect that could impact adult survival as observed in other ectotherms (reviewed in Burraco 504 et al. 2020).

505 Previous studies have reported deleterious effects of hot summers on juvenile newt abundance, due 506 mostly to desiccation of breeding sites. Desiccation of breeding sites drastically reduced larval amphibian 507 survival, which had a direct impact on the abundance of post-metamorphic juveniles (Cayuela et al. 2016; 508 Weinbach et al. 2018). In our study, we added water to mesocosms daily, so the aquatic habitats did not dry out 509 even in the warm climate, which could explain the absence of any effect of a warm climate on juvenile 510 abundance. In some species, early life stages are also less impacted by warmer climates (Bestion et al. 2015b) 511 because accelerated energy acquisition in warmer climates are useful for growth. While warmer temperature can 512 accelerate larval development and the timing of metamorphic emergence in amphibians (Gvoždík 2022), it can 513 also result in a smaller size at metamorphosis, which can reduce post metamorphic survival (Urban et al. 2014). 514 At early life stages, amphibians can also show greater thermal tolerance and thermoregulatory behavior, favoring 515 acclimation to warmer temperatures (Fan et al. 2021). Finally, previous studies have showed that compared to 516 cold winters, warm winters more frequently lead to lower survival because they result in depletion of energy 517 reserves in juveniles unable to look for food (Weinbach et al. 2018). We applied climate treatments only during 518 summer, which could again explain the lack of impact on juvenile abundance. However, unlike adult abundance, 519 which we believe was estimated precisely, estimates for juveniles provide relative abundances among 520 mesocosms due to a lower detection probability of juveniles in dense vegetation than adults in pools.

521

522 Climate change can induce multiple phenotypic changes in species. Decreased body size has emerged as 523 a universal response in many organisms to climate warming but seems less pronounced in amphibians (Li et al. 524 2013) and was not observed in our study. In accordance with other taxa, Reading (2007) found that, compared to 525 cold winters, mild winters reduced body size and condition in common toads by depleting more energy reserves 526 during hibernation. Alternatively, in some frog species, body size was increased in warmer habitats (Tryjanowski 527 et al. 2006), which could be explained by either a positive effect of insect prey availability on growth rate (i.e. 528 developmental plasticity), or by an adaptive role of larger body size in desiccation resistance (i.e. selection). In a 529 previous study using the same experimental system, we found that after 3 years of treatment, common lizards inhabiting warm mesocosms were bigger than in the present-day climate, mainly because of accelerated growth
rate at the juvenile stage (Pellerin et al. 2022). In our study, the absence of a climate effect on body size may
result from the summertime climatic manipulation or from opposite effects of energetic saving and prey
availability.

534 Adult newts and, to a smaller extent, juvenile newts were darker in warm climates. Climate warming is 535 predicted to induce a darker skin for UV protection or a paler skin to avoid overheating (Roulin 2014), with the 536 resulting effect depending on the cost-benefit balance between UV protection and overheating. In our 537 experiment, the climatic treatment was created with shutters that automatically closed above certain temperature 538 thresholds, so the warmer treatment implied more exposure to the sun and UV radiation. Amphibian skin is 539 particularly thin and UV radiation can induce strong damage that can directly affect survival (reviewed by 540 Blaustein et al. 2010). Coloration traits have been shown to be highly plastic in amphibians (Nilsson Sköld et al. 541 2013) and the development of darker skin under UV exposure has been observed in several amphibian species 542 (Garcia et al. 2004; Blaustein et al. 2010).

543 Juvenile newts in our study were also less red in warmer than in present-day climates. In amphibians, 544 the red pigments, mostly carotenoids, are acquired through dietary sources (Mochida et al. 2013). In red-eyed 545 tree frogs (Agalychnis callidryas), a diet rich in carotenoids leads to redder skin and has a positive effect on 546 juvenile development and female fecundity (Ogilvy et al. 2012). Warm climates may reduce newt's prey 547 availability and diversity, and hence access to dietary pigments. Carotenoids may also provide protection against the pro-oxidative effects of UV induced by our warming treatment (Ogilvy and Preziosi 2012) and making 548 549 carotenoids less available for skin color. Regardless of the exact determinant, redder individuals are often 550 considered healthier, advantaged against predators (i.e., toxicity signal, (Mochida et al. 2013), and have higher 551 reproductive success (Ogilvy and Preziosi 2012). Overall, the decrease of red pigments in juveniles from our 552 experiment suggests the potential for negative effects from warmer conditions.

553

554 *Connectivity mitigates the effects of climate warming*

555 Interestingly, access to microclimatic refuges negated the effect of warming on newt abundance and suggests a 556 mitigating effect of these refuges for population persistence (Scheffers et al. 2013). Newts in our study may have 557 migrated to the cooler climate habitats during the warmer season to prevent overheating, as found in other 558 ectotherms (Chukwuka et al. 2021). However,, we could not monitor movements between microclimates because 559 newts were not individually marked. Movements could also have allowed a reorganization of individuals 560 according to their thermal optimum, with the phenotypes less adapted to warm climate settling in cooler habitats 561 and vice-versa (Bestion et al. 2015a). Such adaptive dispersal could reduce the negative impacts of warming 562 climate on (meta)population persistence (Pellerin et al. 2019), and is a likely hypothesis given the changes in 563 phenotypic composition in connected populations.

564 Contrary to patterns observed in isolated habitats, newts were paler in the warm climate than in the 565 present-day climate in connected habitats. Darker newts may have moved from the warm climate to the present-566 day climate to prevent overheating (Roulin 2014), and paler individuals may have moved to the warmer climate 567 to meet their thermal needs. In amphibians, smaller sized juveniles are at greater risk of desiccation and their 568 strictly terrestrial lifestyle may lead them to search for terrestrial microclimatic refuges (Enriquez-Urzelai et al. 569 2019). Darker individuals may therefore seek microclimatic refuges to prevent overheating. In contrast to juveniles, darker adults could avoid risk of overheating by remaining in the water during summer, while stillremaining more protected against UV radiation.

572 Connectivity between microclimates further mitigated the effect of warm climate on red coloration. 573 This suggests that the possibility to reach a cooler microclimate can decrease oxidative stress and may leave 574 more carotenoids available for the skin (Ogilvy and Preziosi 2012) or increase the availability of prey that 575 provide the pigments. Information on food availability and diet preference of newts in the two climatic 576 treatments would allow a better understanding of the underlying mechanisms.

577

578 Emigration propensity and dispersal syndrome

579 We found that newts from populations with present-day climate temperatures emigrated slightly more often from 580 warmer than from cooler conditions, while newts from populations with warmer climates emigrated equally 581 under both thermal conditions. This might suggest an acclimation to warmer temperatures by newts from warm 582 climates induced by developmental plasticity. Acclimation to warm temperatures has previously been shown in 583 several newt species through locomotor activity, metabolic rate and critical thermal maximum, and these plastic 584 responses can mitigate the impacts of climate change to some extent (Gvoždík et al. 2007; Winterová & Gvoždík 585 2022). Acclimation to warmer temperatures could also result from natural selection, but given the short duration 586 of this study this explanation is unlikely.

587 While dispersal can facilitate colonization of areas with suitable climatic conditions, this depends on the 588 phenotypic specialization of the dispersers, which modulates their success in travelling or adapting to novel 589 conditions (Clobert et al. 2009). In this study we assessed dispersal syndrome by measuring traits that enhance 590 movement (body size and leg length) on the one hand, and traits related to local adaptation/acclimation 591 (coloration) and movement propensity on the other. In connected populations, we found evidence of dispersal 592 syndrome among almost all the traits measured. Emigrant newts were bigger, had longer legs and were darker 593 than residents. Body size and leg length are the main common enhancing traits of dispersers, and this 594 phenomenon has been well-documented in other amphibians (Denoël et al. 2018; Winandy et al. 2019). The 595 positive association between darkness in coloration and dispersal has been mostly described in birds (Roulin, 596 2014; Saino et al., 2014), and may arise as a by-product of correlation with other physiological, morphological 597 and behavioral traits. For example, darkness is positively correlated to boldness and exploration (Mafli et al. 598 2011; Mateos-Gonzalez and Senar 2012), two behavioral traits often associated to a higher dispersal propensity. 599 Yet habitat isolation largely altered these syndromes in our study. Emigrants had shorter legs than residents but 600 did not differ in body size and darkness. As found in other species (Cote et al. 2022), specific environmental 601 conditions can disrupt the relationships between phenotypic traits and dispersal and may result in additional 602 dispersal costs (Winandy et al. 2019). While the higher abundance of newts in connected populations in our 603 study could suggest adaptive movements, a longer-term follow-up of the fitness of individuals of different 604 phenotypes and emigration decisions is needed to validate this hypothesis. The interaction between the 605 phenotype and environmental conditions is complex, and challenging environments such as warmer and more 606 fragmented habitats can modify dispersal syndrome. This could have consequences for the success of range 607 shifts and, to a greater extent, on phenotypic composition and/or survival of populations (Pellerin et al. 2019).

608

609 Conclusion

610 The spatial configuration of habitats in the landscape can influence the response of organisms to warming 611 climates (Opdam and Wascher 2004). In this study, we found interactive effects of these factors on newt 612 populations: when newts do not have the opportunity to escape warmer temperature, their abundance declines 613 and the phenotypic composition of populations is altered, while microclimate connectivity seems to alleviate 614 these effects. We cannot tease apart the effects of genetic selection and phenotypic plasticity on the observed 615 phenotypic changes. Yet coloration traits have been shown to be highly plastic in amphibians (Nilsson Sköld et 616 al. 2013). By buffering the adverse effects of high temperatures, phenotypic plasticity can be a first step of 617 population persistence (Seebacher et al. 2014). Indeed, in amphibians, thermal acclimation through behavioral 618 thermoregulation or physiological heat tolerance can act as efficient compensatory mechanisms (Gvoždík et al. 619 2007). We did not measure these thermal traits in our study, so we do not know if newts developed thermal 620 compensation in warmer climates. Yet even the most plastic traits may prove insufficient to fully compensate for 621 the consequences of overheating in ectotherms (Gunderson and Stillman 2015). In this case, emigration of less 622 adapted phenotypes to a habitat that better matches their phenotype could be a way of ensuring population 623 persistence, a hypothesis which needs to be validated through long term monitoring of the survival and 624 reproduction of individuals (Merilä and Hendry 2014).

Landscape connectivity to microclimatic refuges seems to be a key factor that could buffer species against regional extirpation (Suggitt et al. 2018). Access to microclimatic refuges can be particularly important in the case of amphibians such as newts that have relatively limited dispersal capacity. Conservation of microclimatic areas like forests and wetlands as well as maintaining landscape connectivity should be a primary consideration to reduce the impact of climate warming on amphibian diversity (Scalercio et al. 2009; Frey et al. 2016).

631

632 References

- Balogová M, Gvoždík L (2015) Can Newts Cope with the Heat? Disparate Thermoregulatory Strategies of Two
 Sympatric Species in Water. PLoS ONE 10(5): e0128155.
- Bates D, Mächler M, Bolker B, Walker S (2008) Fitting mixed-effects models using the lme4 package in R. In:
 International Meeting of the Psychometric SocietyBestion E, Clobert J, Cote J (2015a) Dispersal
 response to climate change: scaling down to intraspecific variation. Ecology Letters 18:1226–1233
- Bestion E, Teyssier A, Richard M, et al (2015b) Live fast, die young: experimental evidence of population
 extinction risk due to climate change. PLoS biology 13:e1002281
- Blaustein AR, Walls SC, Bancroft BA, et al (2010) Direct and indirect effects of climate change on amphibian
 populations. Diversity 2:281–313
- Burraco P, Orizaola G, Monaghan P, Metcalfe NB (2020) Climate change and ageing in ectotherms. Global
 Change Biology 26:5371–5381
- 644 Cayuela H, Arsovski D, Bonnaire E, et al (2016) The impact of severe drought on survival, fecundity, and population persistence in an endangered amphibian. Ecosphere 7:e01246
- 646 Cayuela H, Grolet O, Joly P (2018) Context-dependent dispersal, public information, and heterospecific
 647 attraction in newts. Oecologia 188:1069–1080
- 648 Chukwuka CO, Mello RS, Cree A, Monks JM (2021) Thermal heterogeneity of selected retreats in cool 649 temperate viviparous lizards suggests a potential benefit of future climate warming. Journal of Thermal
 650 Biology 97:102869

- Clobert J, Le Galliard J-F, Cote J, et al (2009) Informed dispersal, heterogeneity in animal dispersal syndromes
 and the dynamics of spatially structured populations. Ecology letters 12:197–209
- 653 Cote J, Bestion E, Jacob S, Travis J, Legrand D, Baguette M (2017). Evolution of dispersal strategies and dispersal syndromes in fragmented landscapes Ecography, 40: 56-73
- 655 Cote J, Dahirel M, Altermatt F, et al (2022) Dispersal syndromes in challenging environments: a cross-species
 656 experiment. Ecology Letters 25: 2675-2687
- 657 Denoël M (2006) Seasonal variation of morph ratio in facultatively paedomorphic populations of the palmate
 658 newt Triturus helveticus. Acta oecologica 29:165–170
- Denoël M, Dalleur S, Langrand E, et al (2018) Dispersal and alternative breeding site fidelity strategies in an amphibian. Ecography 41:1543–1555
- Díaz SM, Settele J, Brondízio E, et al (2019) The global assessment report on biodiversity and ecosystem
 services: Summary for policy makers
- Enriquez-Urzelai U, Bernardo N, Moreno-Rueda G, et al (2019) Are amphibians tracking their climatic niches in response to climate warming? A test with Iberian amphibians. Climatic Change 154:289–301
- Fan XL, Lin ZH, Scheffers BR (2021) Physiological, developmental, and behavioral plasticity in response to thermal acclimation. Journal of Thermal Biology 97:102866
- Frey SJ, Hadley AS, Johnson SL, et al (2016) Spatial models reveal the microclimatic buffering capacity of old growth forests. Science advances 2:e1501392
- Fronhofer EA, Legrand D, Altermatt F, et al (2018) Bottom-up and top-down control of dispersal across major
 organismal groups. Nature Ecology & Evolution 2:1859–1863
- Garcia TS, Stacy J, Sih A (2004) Larval salamander response to UV radiation and predation risk: color change
 and microhabitat use. Ecological Applications 14:1055–1064
- 673 Gunderson AR, Stillman JH (2015) Plasticity in thermal tolerance has limited potential to buffer ectotherms
 674 from global warming. Proceedings of the Royal Society B: Biological Sciences 282:20150401
- Gvoždík, L. (2015) Mismatch Between Ectotherm Thermal Preferenda and Optima for Swimming: A Test of the
 Evolutionary Pace Hypothesis. Evolutionary Biology 42: 137–145
- Gvoždík L, Puky M, ŠUGERKOVÁ M (2007) Acclimation is beneficial at extreme test temperatures in the
 Danube crested newt, Triturus dobrogicus (Caudata, Salamandridae). Biological Journal of the Linnean
 Society 90:627–636
- Huey RB, Kearney MR, Krockenberger A, et al (2012) Predicting organismal vulnerability to climate warming:
 roles of behaviour, physiology and adaptation. Philosophical Transactions of the Royal Society B:
 Biological Sciences 367:1665–1679
- Joly P, Miaud C (1989) Fidelity to the breeding site in the alpine newt Triturus alpestris. Behavioural Processes
 19:47–56
- Kristín P, Gvoždík L (2014) Aquatic-to-terrestrial habitat shift reduces energy expenditure in newts. Journal of
 Experimental Zoology Part A: Ecological Genetics and Physiology 321:183–188
- Lê S, Josse J, Husson F (2008) FactoMineR: an R package for multivariate analysis. Journal of statistical software 25:1–18
- Legrand D, Guillaume O, Baguette M, et al (2012) The Metatron: an experimental system to study dispersal and metaecosystems for terrestrial organisms. Nature methods 9:828–833

- Legrand D, Trochet A, Moulherat S, et al (2015) Ranking the ecological causes of dispersal in a butterfly.
 Ecography 38:822–831
- Lenoir J, Svenning J-C (2015) Climate-related range shifts-a global multidimensional synthesis and new research directions. Ecography 38:15-28
- Li Y, Cohen JM, Rohr JR (2013) Review and synthesis of the effects of climate change on amphibians.
 Integrative Zoology 8:145–161
- 697 Lüdecke D, Ben-Shachar MS, Patil I, et al (2021) performance: An R package for assessment, comparison and
 698 testing of statistical models. Journal of Open Source Software 6:
- 699 Lüdecke D, Lüdecke MD (2019) Package 'sjstats.' Statistical functions for Regression Models, Version 017 3:
- Mafli A, Wakamatsu K, Roulin A (2011) Melanin-based coloration predicts aggressiveness and boldness in captive eastern Hermann's tortoises. Animal Behaviour 81:859–863
- Masson-Delmotte V, Zhai P, Pirani A, Connors SL, Péan C, Berger S et al. (2021). Climate change 2021: the physical science basis. Contribution of working group I to the sixth assessment report of the intergovernmental panel on climate change, 2.
- Mateos-Gonzalez F, Senar JC (2012) Melanin-based trait predicts individual exploratory behaviour in siskins,
 Carduelis spinus. Animal Behaviour 83:229–232
- 707 McGaughran A, Laver R, Fraser C (2021) Evolutionary responses to warming. Trends in Ecology & Evolution
 708 36:591–600
- Merilä J, Hendry AP (2014) Climate change, adaptation, and phenotypic plasticity: the problem and the
 evidence. Evolutionary applications 7:1–14
- Milling CR, Rachlow JL, Olsoy PJ, et al (2018) Habitat structure modifies microclimate: An approach for
 mapping fine-scale thermal refuge. Methods in Ecology and Evolution, 9: 1648-1657
- Mochida K, Kitada M, Ikeda K, et al (2013) Spatial and temporal instability of local biotic community mediate a
 form of aposematic defense in newts, consisting of carotenoid-based coloration and tetrodotoxin.
 Journal of chemical ecology 39:1186–1192
- Nilsson Sköld H, Aspengren S, Wallin M (2013) Rapid color change in fish and amphibians–function,
 regulation, and emerging applications. Pigment cell & melanoma research 26:29–38
- Ogilvy V, Preziosi RF (2012) Can carotenoids mediate the potentially harmful effects of ultraviolet light in
 Silurana (Xenopus) tropicalis larvae? Journal of animal physiology and animal nutrition 96:693–699
- Ogilvy V, Preziosi RF, Fidgett AL (2012) A brighter future for frogs? The influence of carotenoids on the health,
 development and reproductive success of the red-eye tree frog. Animal Conservation 15:480–488
- 722 Opdam P, Wascher D (2004) Climate change meets habitat fragmentation: linking landscape and biogeographical scale levels in research and conservation. Biological conservation 117:285–297
- Parmesan C, Yohe G (2003) A globally coherent fingerprint of climate change impacts across natural systems.
 nature 421:37–42
- Pellerin F, Bestion E, Winandy L, et al (2022) Connectivity among thermal habitats buffers the effects of warm climate on life-history traits and population dynamics. Journal of Animal Ecology 91: 2301–2313
- Pellerin F, Cote J, Bestion E, Aguilée R (2019) Matching habitat choice promotes species persistence under
 climate change. Oikos 128:221–234
- Pittman SE, Osbourn MS, Semlitsch RD. (2014). Movement ecology of amphibians: a missing component for
 understanding population declines. Biological Conservation, 169, 44-53.

- Reading CJ (2007) Linking global warming to amphibian declines through its effects on female body condition
 and survivorship. Oecologia 151:125–131
- Román-Palacios C, Wiens JJ (2020) Recent responses to climate change reveal the drivers of species extinction
 and survival. Proceedings of the National Academy of Sciences 117:4211–4217
- Roulin A (2014) Melanin-based colour polymorphism responding to climate change. Global Change Biology 20:3344–3350
- Saino N, Romano M, Scandolara C, et al (2014) Brownish, small and lousy barn swallows have greater natal dispersal propensity. Animal Behaviour 87:137–146
- Scalercio S, Russo M, Dapporto L (2009) Wetlands are refuge areas that delay global warming-induced range
 shift of Lepidoptera. Handbook of nature conservation: Global, Environmental and Economic Issues,
 Nova Publishers, New York 393–406
- Scheffers BR, Phillips BL, Laurance WF, et al (2013) Increasing arboreality with altitude: a novel biogeographic
 dimension. Proceedings of the Royal Society B: Biological Sciences 280:20131581
- Seebacher F, Beaman J, Little AG (2014) Regulation of thermal acclimation varies between generations of the
 short-lived mosquitofish that developed in different environmental conditions. Functional Ecology 28:137–148
- Sillero N (2021) Climate change in action: local elevational shifts on Iberian amphibians and reptiles. Regional
 Environmental Change 21:1–13
- Suggitt AJ, Wilson RJ, Isaac NJ, et al (2018) Extinction risk from climate change is reduced by microclimatic
 buffering. Nature Climate Change 8:713–717
- Thompson RM, Beardall J, Beringer J, et al (2013) Means and extremes: building variability into community level climate change experiments. Ecology Letters 16:799–806
- Tryjanowski P, Sparks T, Rybacki M, Berger L (2006) Is body size of the water frog Rana esculenta complex
 responding to climate change? Naturwissenschaften 93:110–113
- 756 Urban MC (2015) Accelerating extinction risk from climate change. Science 348:571–573
- 757 Urban MC, Richardson JL, Freidenfelds NA (2014) Plasticity and genetic adaptation mediate amphibian and
 758 reptile responses to climate change. Evolutionary Applications 7:88–103
- van den Brink V, Dreiss AN, Roulin A (2012) Melanin-based coloration predicts natal dispersal in the barn owl,
 Tyto alba. Animal Behaviour 84:805–812
- Verell PA (1987). The directionality of migrations of amphibians to and from a pond in southern England, with
 particular reference to the smooth newt, Triturus vulgaris. Amphibia-Reptilia 8: 93-100
- Weinbach A, Cayuela H, Grolet O, et al (2018) Resilience to climate variation in a spatially structured
 amphibian population. Scientific Reports 8:1–9
- Winandy L, Cote J, Di Gesu L, et al (2019) Local predation risk and matrix permeability interact to shape
 movement strategy. Oikos 128:1402–1412