Plant Health Workshop: Advances in plant disease and pest management: a holistic approach February 28th, 2023 IRTA, Spain

Breeding for resistance/tolerance in fruit crops: New trends

Bénédicte QUILOT-TURION

INRAE GAFL







Major stakes for stone fruit production today



Need for low input cultivation

Treatment frequency [15-30] High dependance to irrigation and fertilization

Varieties with low adaptation to agro-ecological transition:

Few resilient varieties No incentive in registration procedure

Perennial crops

Health management over years ~20 years to create a new variety

Major solutions:

→Genetic diagnostics

→Screening plant material faster and more accurately

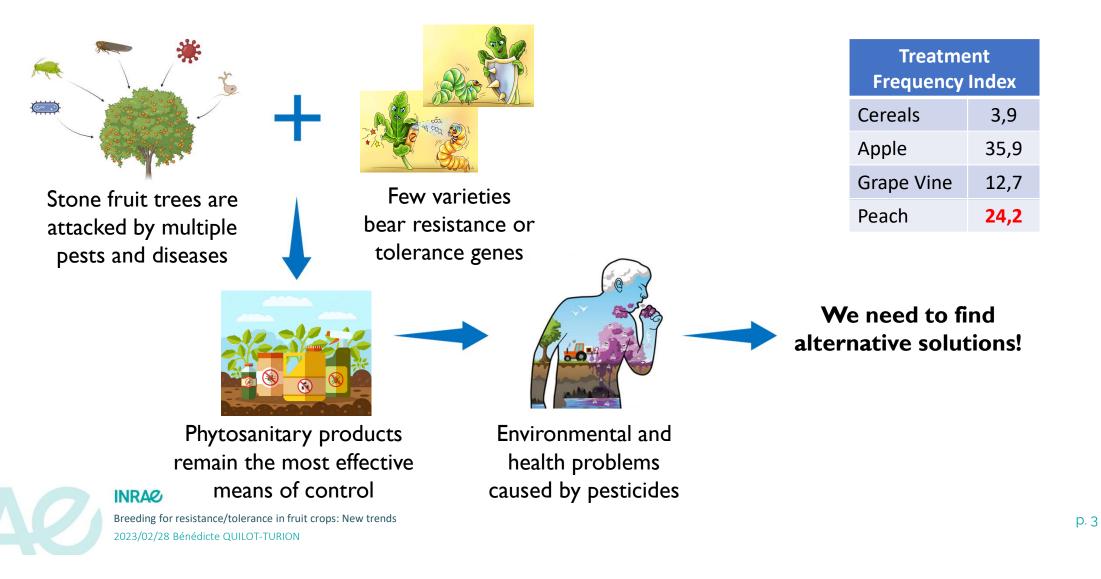


Potential for agro-ecology

Large diversity within stone fruit Development of agroforestry Carbon storage in soil Demand for organic fruits New « eco-labels » on market

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> The use of pesticides: a real problem in fruit production



Makes genetic an efficient lever for agroecology

Combining genetics and agrotechnical levers to develop agroecological, sustainable systems

- 1. Mobilize the arsenal of modern tools
 - Sequencing, genotyping, genetic engineering
 - Digital phenotyping, NIRS
- 2. Ensure the durability of the resistance genes
- 3. Enlarge genetic diversity <- use wild relatives
- 4. Target new traits : eg resilience

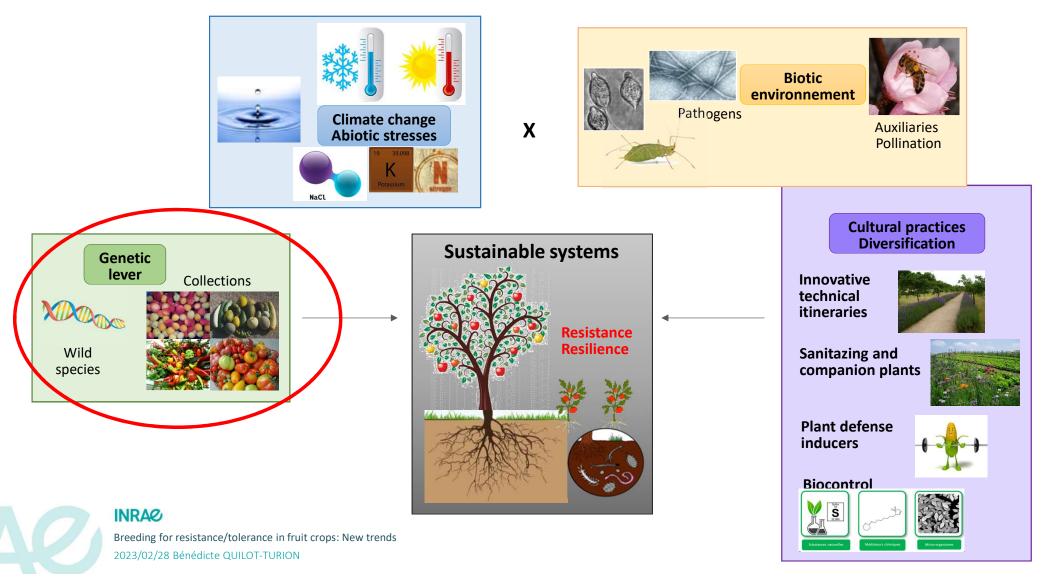
Methodologies

Marker Assisted Selection Marker Assisted Introgression Genomic prediction Gene editing

Phenotype characterization Phenomic prediction

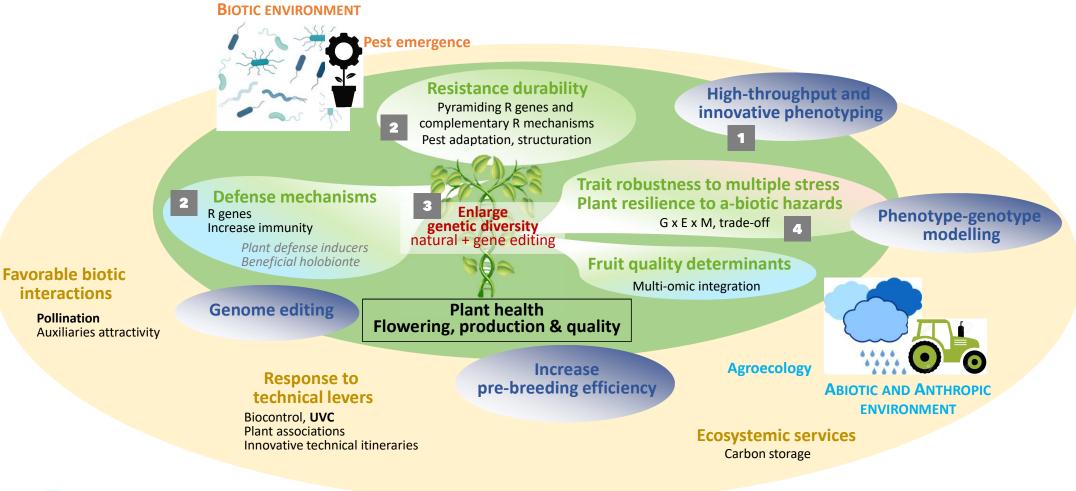


Makes genetic an efficient lever for agroecology



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Makes genetic an efficient lever for agroecology



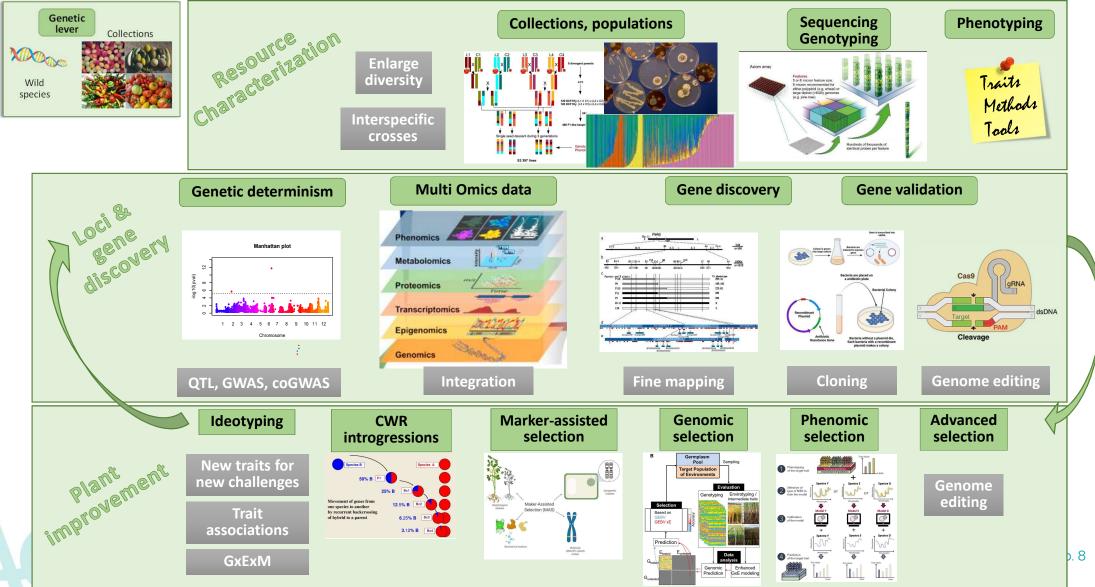
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> Summary

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> The genetician pipeline



> Phenotyping

Controlled experiments



Emphasis platform network

Phenotyping tools

Highthrouput phenotyping

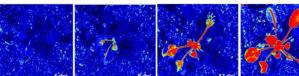


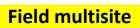
Captors for ecophysiological traits

Images for symptoms / architecture







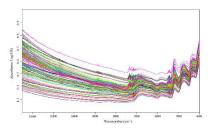




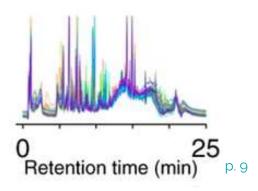
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NIRS for phenomics



Metabolomics



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Durable management of resistance

- Resistance genes are a limited resource
- Resistance gene breakdowns have already been observed
- A breeding program is a long-term and costly process
- The question of the management of resistance durability is crucial, particularly for perennial species

Quantitative resistance (QR) : the foundation of breeding for disease resistance in crops to achieve durable resistance

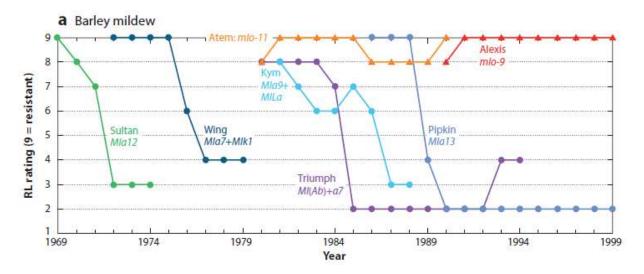
-> it remains poorly understood in comparison to the well studied gene-for-gene recognition process

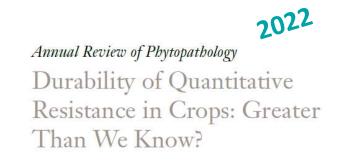
Pyramiding resistance genes is an efficient strategy to increase resistance durability

Appropriate cultural practices and monitoring of variety deployment also contribute to the durability of resistance

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Christina Cowger¹ and James K.M. Brown²

Durable Resistance of Crops to Disease: A Darwinian Perspective James K.M. Brown Annual Review of Phytopathology 2015 53:1, 513-539

> **Durable resistance** is defined as resistance that has been effective for a relatively long time over a relatively large area where conditions are generally conducive to the disease

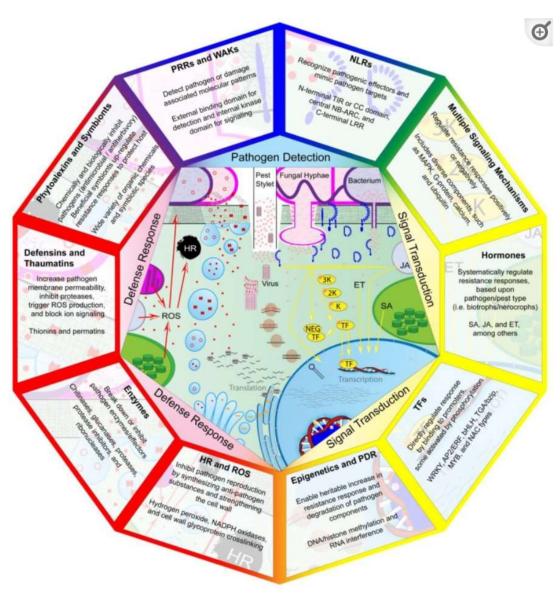
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Plant defense mechanisms



Review Disease Resistance Mechanisms in Plants

Ethan J. Andersen ¹, Shaukat Ali ², Emmanuel Byamukama ², Yang Yen ¹⁽⁰⁾ and Madhav P. Nepal ^{1,*}⁽⁰⁾





> Durable management of resistance

Many Effector-Triggered Resistance genes with large effects have not been durable because of their specificity to pathogen genotypes

Specific resistances	Absolute, total, qualitative resistances	Monogenic Major gene	Prevents pathogen establishment	Triggers a strong plant response to neutralize the pathogen; not very robust Selection pressure on the pathogen selects more virulent strains: circumvention
	Partial, incomplete, quantitative resistances	Polygenic Several minor genes	Limits colonization and the impact of the presence of the pathogen	Causes disease reduction Longevity, durability of resistance
Non- specific resistances				to multiple diseases allowing plants to defend themselves against a broad spectrum of pathogens

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> INRA-ResDur for durable resistance in grape vine

Resistance breakdown of

- Rpv3 : resistance to Plasmopara viticola (downy mildew)
- Run1 : resistance to Uncinula necator (powdery mildew)

Marker-assisted selection to stack resistance factors derived from multiple sources.

- -> development of candidate cultivars two or three genes to control each disease
- Rpv1, Rpv3 and Rpv10 for downy mildew
- Run1, Ren3 and Ren3.2 for powdery mildew

INRA-ResDur: the French grapevine breeding programme for durable resistance to downy and powdery mildew

C. Schneider^a, C. Onimus, E. Prado, V. Dumas, S. Wiedemann-Merdinoglu, M.A. Dorne, M.C. Lacombe, M.C. Piron, A. Umar-Faruk, E. Duchêne, P. Mestre and D. Merdinoglu



Breeding for durable resistance to downy and powdery mildew in grapevine

D. Merdinoglu*, C. Schneider, E. Prado, S. Wiedemann-Merdinoglu and P. Mestre

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Enlarge genetic diversity : use wild relatives



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MDP

Trends in

> INRAE peach resistance program : green aphid and powdery mildew

Theor Appl Genet (2003) 107:227–238 DOI 10.1007/s00122-003-1238-8

2003

M. Foulongne · T. Pascal · P. Arús · J. Kervella

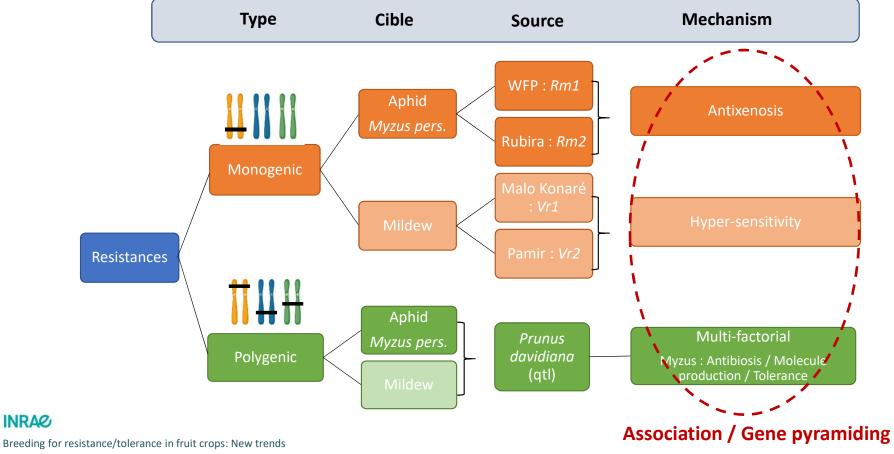
The potential of *Prunus davidiana* for introgression into peach [*Prunus persica* (L.) Batsch] assessed by comparative mapping

Trait/Resistance	Organism	Pathogen	Inoculation	Resistance/ tolerance	Inheritance	Gene name	LG	Reference
P. davidiana								
Powdery mildew	Fungi	P. pannosa	Natural	Tolerance	Quantitative	QTLs	1, 2, 4, 6, 8	Foulongne et al. (2003)
Sharka	Virus	Plum pox virus	Controlled	Tolerance	Quantitative	QTLs	1, 2, 4, 5, 6,7	Rubio et al. (2013)
Green peach aphid	Insect	Myzus persicae	Natural	Tolerance	Quantitative	QTLs	1, 2, 3, 4, 5,6, 8	Sauge et al. (2012)
Root-knot nematode	Nematode	Meloidogyne spp.	Natural	Tolerance	Major gene		Not-mapped	Reighard and Loreti (2008)
Brown rot	Fungi	Monilinia spp.	Controlled	Tolerance	Quantitative		Not-mapped	Pascal et al. (1998)

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INRAE peach resistance program : green aphid and powdery mildew



Major genes associated to minor genes for resistance durability

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Incorporation of resistance genes from wild species : a promising option

identify the valuable germplasm

very little information about resistance sources in non-crop *Prunus* relatives

-> systematic survey of *Prunus* wild relatives around the world

-> characterization of their phenotype for various resistance traits

Aranzana et al. Horticulture Research (2019)6:58 https://doi.org/10.1038/s41438-019-0140-8 2019

Horticulture Research www.nature.com/hortres

REVIEW ARTICLE

Open Access

Prunus genetics and applications after de novo genome sequencing: achievements and prospects

Maria José Aranzana¹, Véronique Decroocq², Elisabeth Dirlewanger², Iban Eduardo¹, Zhong Shan Gao ³, Ksenija Gasic⁴, Amy lezzoni⁵, Sook Jung⁶, Cameron Peace⁶, Humberto Prieto⁷, Ryutaro Tao⁸, Ignazio Verde⁹, Albert G. Abbott¹⁰ and Pere Arús¹

-> introgression of the genetic factors linked to resistance into elite *Prunus* cultivars through interspecific crosses

The construction of a *Prunus* pan-genome would promote and accelerate the development of appropriate genomic tools for all *Prunus* species, including wild relatives



Development of interspecific lines and molecular tool

Tree Genetics & Genomes (2016) 12: 96 DOI 10.1007/s11295-016-1056-1

2016

ORIGINAL ARTICLE

Marker-assisted introgression (MAI) of almond genes into the peach background: a fast method to mine and integrate novel variation from exotic sources in long intergeneration species

Octávio Serra¹ · José Manuel Donoso^{1,2} · Roger Picañol^{1,3} · Ignasi Batlle⁴ · Werner Howad¹ · Iban Eduardo¹ · Pere Arús¹

Article

Research

Construction of a collection of introgression lines of "Texas" almond DNA fragments in the "Earlygold" peach genetic background

Naveen Kalluri¹, Octávio Serra², José Manuel Donoso³, Roger Picañol⁴, Werner Howad^{1,5}, Iban Eduardo^{1,5,*} and Pere Arús^{1,5,*}

HORIZON-CL6-2023-BIODIV-01-13 / Crop wild relatives for sustainable agriculture

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CWR inventory and knowledge WP1 WP2 CWR genotypic characterisation WP3 Phenotypic characterisation HTP genotyping of CWR core-WP4 collections WP5 Genomics-based methods to integrate CWR in pre-breeding programs Promotion and plans for sustainable WP6 conservation of CWR in situ populations

Horticulture 2022 (E) CrossMark

> An endless work, long and laborious

	# pathogens	# studied	included in breeding programs
Peach	70	9	Powdery mildew, brown rot, leaf curl, Bacterial spot, PPV, green peach aphid, root-knot nematodes
Cherry	61	2	Monilinia, bacterial canker
Apricot	59	5	Monilina, Bacterial canker, PPV, Apricot chlorotic leafroll phytoplasma

source WOS

Few resistance genes known Old and emerging diseases Too many traits to target one by one



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> Why is resilience so relevant ?

Convergent selection targets in animals and plants

Farming challenges

- Resistances et tolerances to a large number of diseases and parasites
- Reduction of antibiotic/parasitic molecules **Examples of biomarkers of resilience**:
- Milk production (ex. Elgersma et al. 2018)
- Blood tests (ex. Bai et al. 2020)
- Diet (ex. Putz et al. 2019)



Crop challenges

- Resistances et tolerances to a large number of diseases and parasites
- Reduction of pesticides

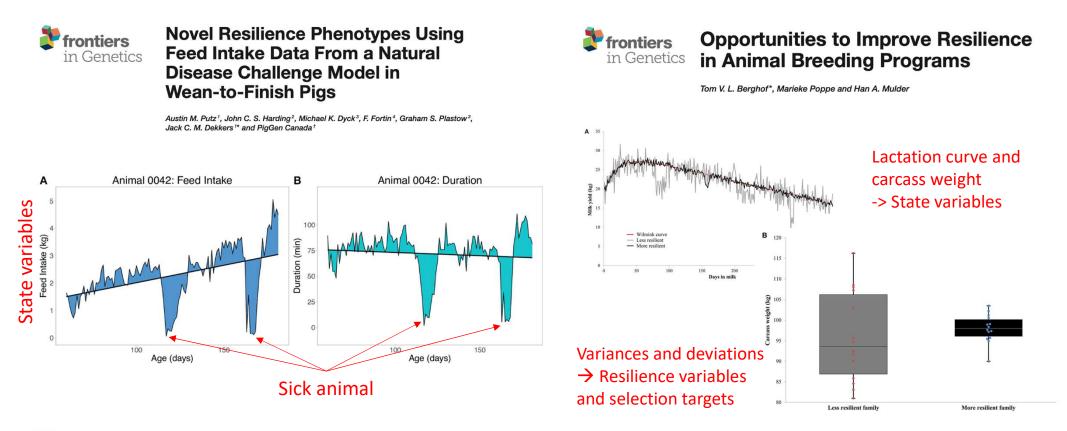
Biomarkers of resilience not defined yet:

- Aerial or root biomass?
- Photosynthetic activity?
- Evapo-transpiration?



Disease-resilience in animals : one step ahead?

« Polymicrobial natural disease model » versus « disease by disease »

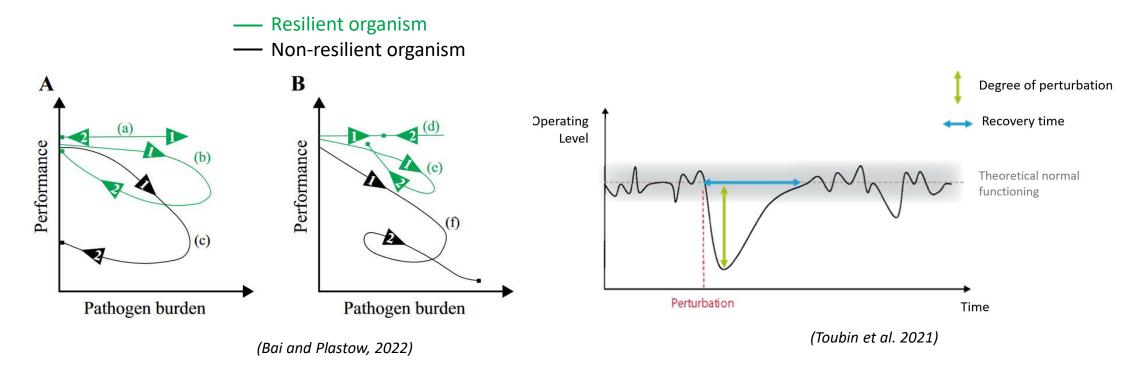


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A source of inspiration

> Breeding for resilience : a durable way to manage the biotic pressures



RESILIENCE : the capacity of an organism to be minimally affected by disturbances or to rapidly return to the state pertained before exposure to a disturbance

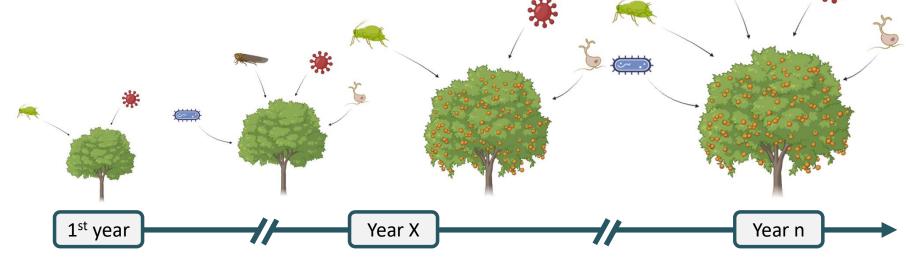
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1st PhD committee 2022/09/28 Marie Serrie it crops: New trends

How to quantify resilience: multi-year and integrative monitoring of orchards

Multi-year monitoring of orchards

Pests/diseases damages



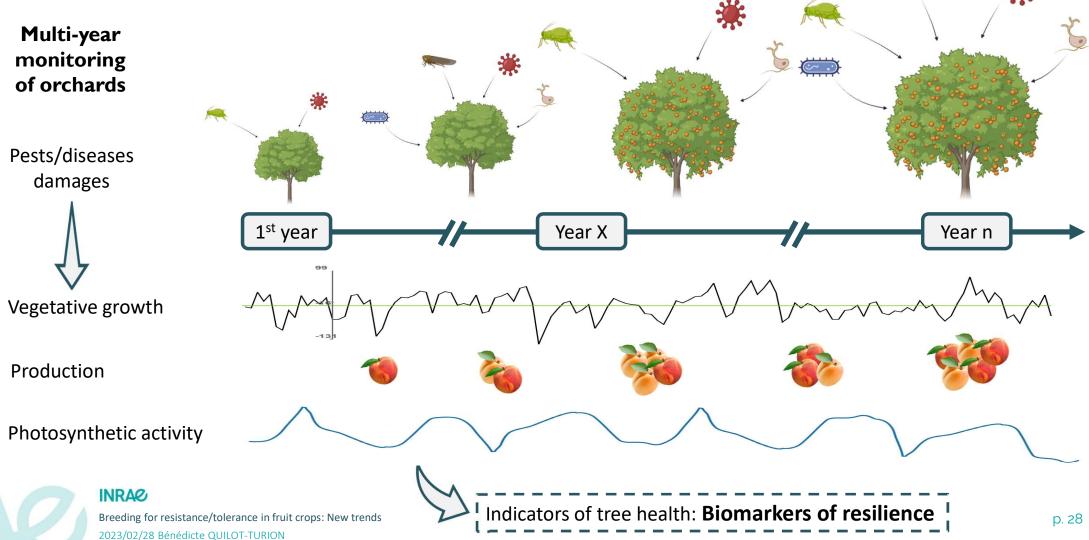


What are the impacts of these biotic stresses on tree health? How to take these data into account in an integrative way?



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How to quantify resilience: multi-year and integrative monitoring of orchards



> Approach

Uncover the fundamental principles of resilience in stone fruit trees

- Multi-trait observations among diversity panels
- Interactions between pest/diseases
- Quantify the impact of biotic stresses on tree health/vigor over the years
- Quantify risk factors with epidemiological covariables
- Summarizing multiple observations of symptoms into resilience metrics

Find genetic markers associated to disease resilience and/or resilience components (resistance/tolerance)

- Identify inheritance modes of resilience sub-trait and integrative traits
- Impact of the genetic, environment, G x E interactions effects
- Identify best-suited GWAS for multi-trait and multi-environment data



Explore the applications of the methods developed in the project for future disease-resilience breeding

- Defining "resilient ideotypes" as a new target for pre-breeding activities
- Discuss the feasibility of MAS for resilience traits
- Compare resilience characteristics between peach and apricot

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> Plant material: multi-species core collections with a large genetic diversity



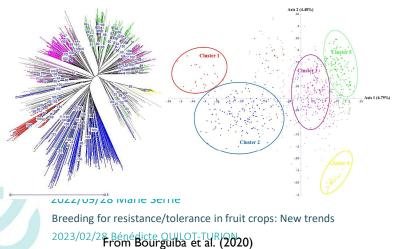
Apricot cultivated

(P. armeniaca)

I 50 accessions Resequenced with Illumina HiSeq 2000 NGS

Selecting accessions

- ✓ Genetic variability of 25 Microsatellites (Bourguiba et al., 2012)
- ✓ Within the five known genetic groups (Bourguiba et al., 2020)
- ✓ Incl. of modern accessions and hybrids from recombinant lines between phylogroups





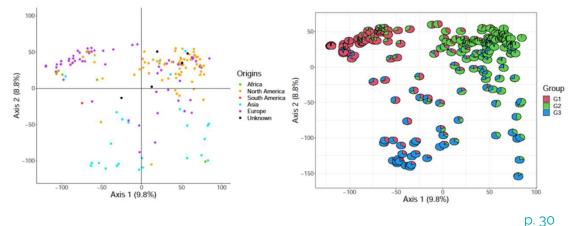
Peach cultivated and wild

(P. persica + relatives)

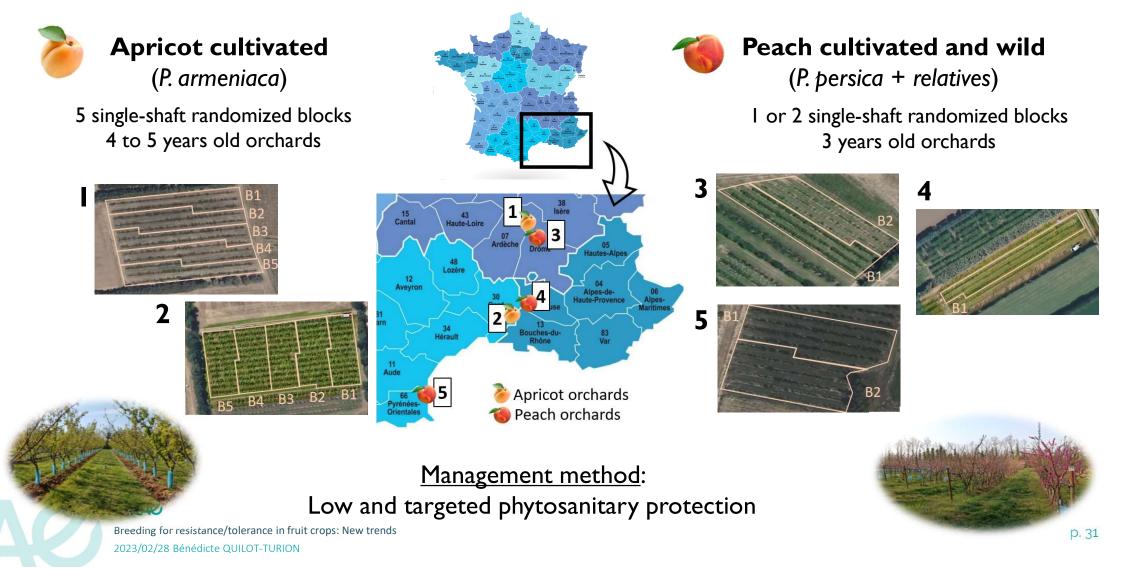
206 accessions Genotyped with 16k SNP chip

Selecting accessions

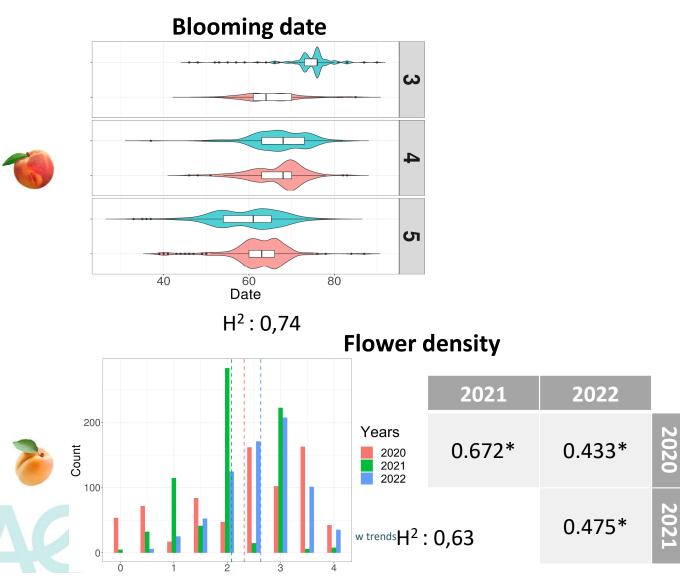
- ✓ SNP polymorphism with 9K chip (Micheletti et al 2015)
- ✓ Incl. of related species and interspecific
- ✓ 3 genetics groups
- ✓ Several geographical origin

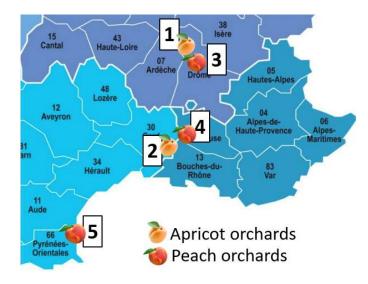


Experimental design: multi-site with low pesticide conditions

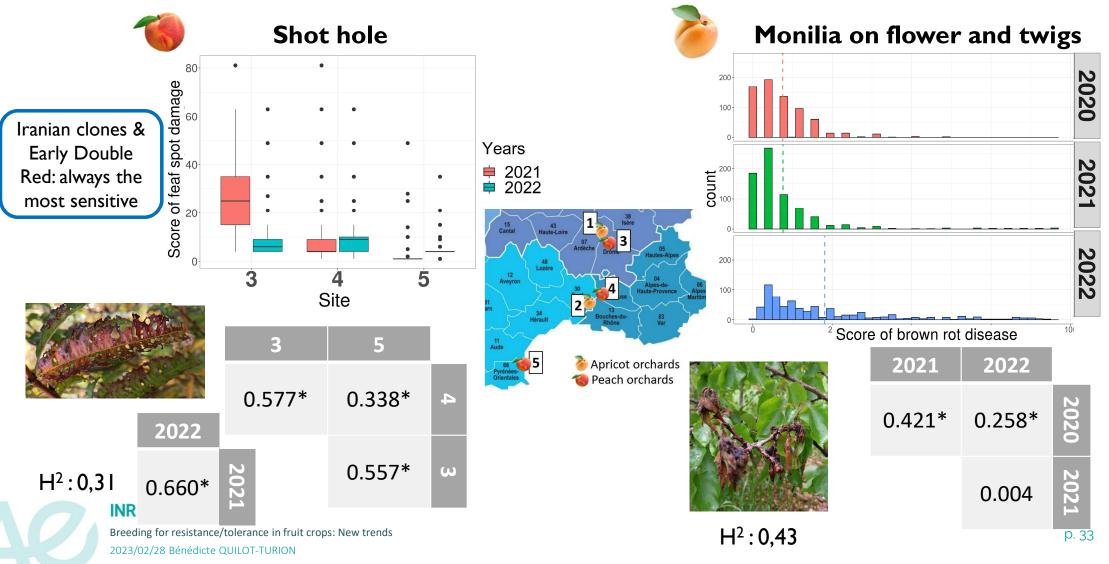


A broad phenological diversity in the core collections





Strong effect of the environment: Necessity of multi-site and years trials



First genome-wide association results on peach >

	SNP detected Trait		Chromosome	P-value	Effect size (β)		
Peach_AO_0175780		Date of vegetative bud break	2	5.24e-08	-3.99	✓ MLMM method	
Peach_AO_0459115		Date of vegetative bud break	4	3.11e-07	-3.22	 (Segura et al. 2012) ✓ Bonferroni correction ✓ 13,975 markers after filtering 	
Pe	each_AO_0060498	Blooming date	1	1.1e-06	3.67		
S	SNP_IGA_283581	Blooming date	2	7.3e-07	4.22		
S	SNP_IGA_116812	Shot hole	1	2.79e-06	-0.27		
[Date of vegetativ	e bud break	Bloomi	ng date		Shot hole	
9			ω - • •		- a	•	
$-\log_{10}(p)$					p. 34		
	1 2 3 4 chromosom	5678 PE	1 2 3 chrom	4 5 6 7 8 OSOME	1 2 3 4 5 6 7 8 chromosome		

> A new breeding approach : a better integration of the information

CLASSICAL SELECTION OF FRUIT TREES	OBJECTIVES FOR TOMORROW'S SELECTION			
"Trait-by-trait" phenotypic selection of varieties with high yield and quality potential	Selection of resilient ideotypes assisted by high-throughput phenotyping and genetic information			
Elite varieties with a wide geographical spectrum that are greedy in pesticides	Varieties able to cope with biotic stresses in a given environment			
Exclusively visual phenotyping	Gain in precision and throughput and new descriptive variables by digital phenotyping			
Sorting candidates for selection on adult plants	Early genetic diagnostics			
Pyramiding of traits of interest, cross after cross within a restricted gene pool	Multi-trait modeling and optimization of the use of diversity			
Resistance to rare diseases and only based on major genes	Consideration of so-called qualitative resistances and tolerances by quantitative genetics			

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> Acknowledgments

Team working



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