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1 **Limits of calcium isotopes diagenesis in fossil bone and enamel**

2

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21

22 **Abstract**

23 Diagenesis has been recognized for decades to significantly alter the trace elements biogenic
24 signatures in fossil tooth enamel and bone that are routinely used for paleobiological and
25 paleoenvironmental reconstructions. This signature is modified during diagenesis according
26 to a complex continuum between two main processes, addition, and substitution. For an
27 additive-like, or early diagenesis, the trace elements biogenic profiles can be restored by
28 leaching secondary minerals, but this technique is inefficient for a substitutive-like, or
29 extensive diagenesis for which secondary trace elements are incorporated into the biogenic
30 mineral. This scheme is however unclear for Ca, the major cation in tooth enamel and bone
31 hydroxylapatite, which stable isotope composition ($\delta^{44/42}\text{Ca}$) also conveys biological and
32 environmental information. We present a suite of leaching experiments for monitoring
33 $\delta^{44/42}\text{Ca}$ values in artificial and natural fossil enamel and bone from different settings. The
34 results show that enamel $\delta^{44/42}\text{Ca}$ values are insensitive to an additive-like diagenesis that
35 involves the formation of secondary Ca-carbonate mineral phases, while bone shows a
36 consistent offset toward ^{44}Ca -enriched values, that can be restored to the biogenic baseline
37 by a leaching procedure. In the context of a substitutive-like diagenesis, bone exhibits
38 constant $\delta^{44/42}\text{Ca}$ values, insensitive to leaching, and shows a REE pattern symptomatic of
39 extensive diagenesis. Such a REE pattern can be observed in fossil enamel for which $\delta^{44/42}\text{Ca}$
40 values are still fluctuating and follow a trophic pattern. We conclude that Ca isotopes in fossil
41 enamel are probably not prone to extensive diagenesis and argue that this immunity is due
42 to the very low porosity of enamel that cannot accommodate enough secondary minerals to
43 significantly modify the isotopic composition of the enamel Ca pool.

44

45 **1. Introduction**

46 Bone, dentin, and enamel are valuable archives for recording information about the
47 life history and environment of fossil and extant vertebrates. The information is embedded
48 as trace elements or isotopic ratios signature during the genesis of the mineralized tissue, a
49 signature that is further obfuscated by diagenesis during fossilization. Regarding trace
50 elements, the comprehension of diagenetic mechanisms has greatly benefited from in-situ
51 laser ablation measurements (Kohn and Moses, 2013; Kral et al., 2022), highlighting the
52 complex interplay of the diffusion, adsorption, and transport reaction processes. However,
53 while sophisticated techniques are now deployed to fully integrate all the hallmarks of
54 diagenesis (Suarez and Kohn, 2020; Weber et al., 2021; Kral et al., 2022), the results still
55 recognize the central importance of porosity which was already suspected in the 2000's
56 (Hedges, 2002; Trueman and Tuross, 2002; Kral et al., 2021). Porosity mediates fluid
57 circulation and secondary mineral deposition, and increases as a result of collagen
58 degradation, augmenting the exposure of carbonated hydroxylapatite (HAp) crystallites in the
59 first stages of bone diagenesis. Because porosity is forty times higher in bone than in enamel
60 (Wang and Cerling, 1994), the latter is widely accepted to be far more resistant to diagenesis
61 than bone and is nowadays the tissue of reference for measuring trace elements
62 concentration (Le Houedec et al., 2013; Joannes-Boyau et al., 2019; Nava et al., 2020; Funston
63 et al., 2022) and/or isotope composition (Martin et al., 2014, 2015; Jaouen et al., 2017;
64 McCormack et al., 2022).

65 In all types of tissue, some diagenetic overprints can be reverted using leaching
66 pretreatment (Sillen and LeGeros, 1991; Koch et al., 1997; Nielsen-Marsh and Hedges, 2000;
67 Balter et al., 2002a; Wathen et al., 2022). When formation of secondary mineral phases is
68 thought to have occurred (additive-like diagenesis), leaching is potentially efficient at
69 removing these minerals. One issue is that leaching is often performed in batch, leading
70 samples to be leached more than necessary, therefore dissolving the most soluble HAp
71 fraction that is the most pristine (Sillen and LeGeros, 1991; Balter et al., 2002a). The efficiency
72 of the leaching method is even more questionable when diffusion of diagenetic elements
73 (substitutive-like diagenesis) into the HAp has occurred.

74 All told, these considerations hold for trace elements, but the situation is unclear for
75 major elements. Traditionally, the $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ values of CO_3 present in HAp (~5 wt%) are
76 measured in leached samples due to the widespread presence of calcite in the porosity, but

77 the $\delta^{18}\text{O}$ value of HAp PO_4 (~55 wt%) is always measured in raw samples. The isotope
78 composition of Ca of HAp (~40 wt%) varies according to trophic position (Skulan and DePaolo,
79 1999; Heuser et al., 2011; Martin et al., 2018; Hu et al., 2022) and physiology (Tacaïl et al.,
80 2017; Hassler et al., 2021; Koutamanis et al., 2021; Li et al., 2022), but deserves an assessment
81 of potential diagenetic effects on $\delta^{44/42}\text{Ca}$ values in bone and tooth enamel. Simple mass
82 balance calculations already show that changes of the original $\delta^{44/42}\text{Ca}$ values in bone and
83 tooth enamel by secondary Ca minerals are either small or would require an uncommonly
84 fractionated Ca isotopes diagenetic pool (Heuser et al., 2011; Martin et al., 2017).

85 Here, we attempt to resolve this issue by studying how Ca isotopes behave during
86 controlled experiments of artificial fossil bone and enamel leaching and in different diagenetic
87 case studies. The Ca isotope compositions are measured along with Ca concentrations and
88 Ca/P ratios to monitor the origin of Ca, and Rare Earth Elements (REE) to decipher the type
89 (additive or substitutive) and extrapolate the extend of diagenetic alteration (Reynard et al.,
90 1999; Trueman and Tuross, 2002).

91

92 **2. Material and methods**

93 First, we analyzed the Ca isotope composition in leached and unleached artificial
94 samples mimicking fossil bone and enamel that have undergone an additive-like diagenesis.
95 These samples consist of mixtures of HAp and calcite powder which $\delta^{44/42}\text{Ca}$ value are distinct
96 and constrained. An artificial fossil bone was prepared using the bone ash certified reference
97 material (SRM-1400, NIST; $\delta^{44/42}\text{Ca} = -1.21 \pm 0.04\text{‰}$, $\pm 2\text{SD}$, $n = 3$) to which was added various
98 proportions of the calcite certified reference material SRM-915b (NIST, $\delta^{44/42}\text{Ca} = -0.28 \pm$
99 0.04‰ , $\pm 2\text{SD}$, $n = 21$). The SRM-1400 certified reference material has been ashed and is
100 devoid of organic matter and has a high crystallinity, both being characteristics of fossil bone.
101 Because enamel certified reference material does not exist, we prepared artificial fossil
102 enamel by mixing a sample of extant enamel from an isolated and partial molar tooth of
103 modern elephant (*Elephas maximus*; $\delta^{44/42}\text{Ca} = -1.43 \pm 0.07\text{‰}$, $\pm 2\text{SD}$, $n = 10$) with various
104 proportions of SRM-915b. Second, we analyzed a suite of natural samples from different
105 localities and ages that were selected according to their potential degree of diagenetic
106 alteration. Pleistocene bone and tooth enamel samples were chosen to be representative of
107 weak diagenesis, with preserved trace elements compositions, and Devonian bone of more

108 pronounced diagenesis. A first set of material is composed by a suite of fossil bone samples
109 (n = 13) from the Camiac cave (Gironde, France). The site is a hyena dens from the upper
110 Pleistocene (Mousterian period, 35.1 ± 2 ka BP; Balter et al., 2002b). Taxonomic information
111 is given in [Table S2](#). This material is characterized by trace elements (Sr/Ca and Ba/Ca)
112 distributions that reflect a trophic pattern (Balter et al., 2002b). A suite of fossil enamel
113 samples (n = 14) from the Plio-Pleistocene of the Shungura Formation in the Lower Omo
114 Valley (Ethiopia, sample age bracketed between 3.6 and 1.76 Ma; Boisserie et al., 2008)
115 represents the second set of material. Here, the material is composed by only two extinct
116 genera of large African suids (*Metridiochoerus* sp. and *Notochoerus* sp., [Table S2](#)). Leached
117 fossil bone at Shungura exhibit Sr/Ca ratios characteristic of a trophic pattern (Sillen, 1986).
118 A third set of samples gather vertebrate (fish and tetrapod) fossil bone (n = 25, [Table S2](#)) from
119 the Devonian locality of Strud (360 Ma, Belgium; Olive et al., 2015). To our knowledge, no
120 geochemical studies has already been performed on this material.

121 The samples were processed in the clean room and analyzed on the facilities at the
122 Laboratoire de Géologie de Lyon: Terre, Planètes, Environnements (LGL-TPE, ENS de Lyon).
123 All the samples (ca. ~ 5 mg) were leached using 0.5 ml of 0.1 M acetic acid per mg of sample
124 during 30 min in an ultrasonic bath and rinsed with 18.2 MΩ/cm grade MilliQ water. This
125 represents 2.5-fold the quantity of acid necessary to dissolve an equivalent amount of pure
126 calcite. The samples were dissolved in distilled concentrated HNO₃ and an aliquot measured
127 for major elements (P and Ca) by ICP-OES (Thermo Scientific, iCap 6000 Radial) and trace
128 elements (REE) by ICP-MS (Thermo Scientific, iCap-Q) according to Balter and Lécuyer (2004).
129 Calcium from the remaining solution was purified by ion-exchange chromatography and Ca
130 isotope composition measured by MC-ICP-MS (Thermo Scientific, Neptune Plus) according to
131 Tacail et al. (2014). Briefly, three ion-exchange chromatography steps are necessary, first
132 allowing the recovery of Ca, Fe and Sr only, the second to purify Ca from Fe and the third to
133 remove Sr ([Table 1](#)). Blanks for the procedure did not exceed 100 ng of Ca.

134

135 **Table 1:** Detailed procedure for Ca extraction and purification.

1. Matrix elimination		
AG50W-X12 resin (200-400 mesh) ~ 2 mL		
Step	Eluent	Vol. (mL)
Condition	1 M HCl	10
Load	1 M HCl	2+1
Elution (matrix)	1 M HCl	55
Ca elution (Ca, Sr, Fe)	6 M HCl	10
2. Fe elimination if necessary		
AG1-X8 resin (100-200 mesh) ~ 2 mL		
Step	Eluent	Vol. (mL)
Condition	6 M HCl	10
Load	6 M HCl	1+1
Elution (Ca, Sr)	6 M HCl	6
3. Sr elimination		
Sr-Specific resin (Eichrom) ~ 0.7 mL		
Step	Eluent	Vol. (mL)
Condition	3 M HNO ₃	5
Load	3 M HNO ₃	0.5+0.5
Elution (Ca)	3 M HNO₃	6

136

137 Calcium isotope abundance ratios (⁴⁴Ca/⁴²Ca and ⁴³Ca/⁴²Ca) were measured using a
 138 Neptune Plus multi-collector ICP-MS using the standard-sample-standard bracketing method.
 139 All Ca isotope compositions are expressed using the delta notation, calculated as follows:

140

$$141 \quad \delta^{44/42}\text{Ca} = \left(\frac{(^{44}\text{Ca}/^{42}\text{Ca})_{\text{Sample}}}{0.5 \times (^{44}\text{Ca}/^{42}\text{Ca})_{\text{ICP-Ca-Lyon}}^{n-1} + 0.5 \times (^{44}\text{Ca}/^{42}\text{Ca})_{\text{ICP-Ca-Lyon}}^{n+1}} - 1 \right) \times 1000$$

142

143 The ICP Ca Lyon standard, used routinely in Lyon, was used as the bracketing standard.
 144 The results can be converted relative to the SRM915a with an offset of -0.52‰.

145

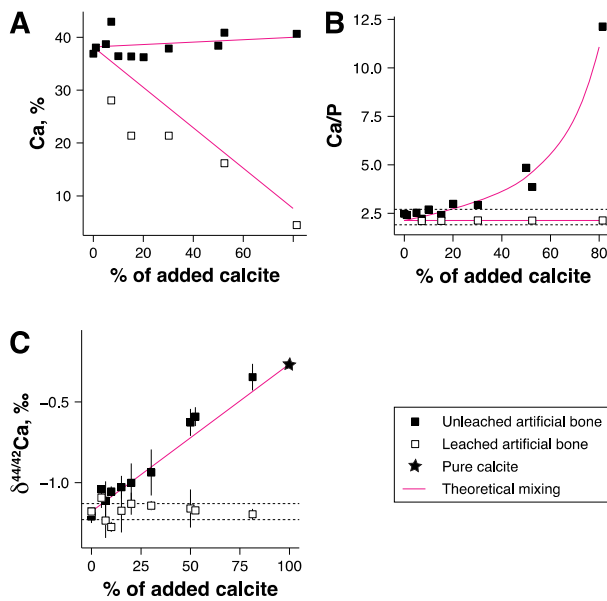
146 **3. Data presentation and interpretation**

147 Results are given in **Table S1** for artificial fossil bone and enamel and in **Table S2** for
 148 natural fossil bone and enamel. The mass fractionation measured in this study agrees with
 149 the 0.5 slope predicted by the linear approximation of mass-dependent fractionation (**Fig. S1**).

150 We first test the effect of leaching on the $\delta^{44/42}\text{Ca}$ value of the certified reference
 151 material SRM-1400 and found no difference after leaching ($\delta^{44/42}\text{Ca} = -1.21 \pm 0.03\text{‰}$, $\pm 2\text{SD}$,
 152 $n = 3$) compared to unleached material ($\delta^{44/42}\text{Ca} = -1.21 \pm 0.04\text{‰}$, $\pm 2\text{SD}$, $n = 3$). Absence of
 153 difference between the $\delta^{44/42}\text{Ca}$ values of unleached and leached enamel is also observed as

154 we measured a constant $\delta^{44/42}\text{Ca}$ value of -1.43‰ (Table S1). Because of the very small
155 amount of processed material (~ 5 mg), the remaining bone or enamel residue after leaching
156 is so small that it is impossible to weigh it, so Ca concentration for leached material is
157 underestimated. Nevertheless, using in 0.1 M acetic acid for 30 min generally leads to a loss
158 of about 30% of initial material. The absence of Ca isotope fractionation during leaching
159 therefore indicates that the dissolution of HAp is quantitative and that leaching *per se* does
160 not affect fossil bone and enamel $\delta^{44/42}\text{Ca}$ values.

161 In the mixture of bone HAp with increasing proportion of added calcite, the slight
162 theoretical increase of Ca concentration is barely detected in unleached mixtures, probably
163 due to the ~5% error inherent to ICPMS measurements (Fig. 1A), but the decrease of Ca
164 content is inambiguous in leached mixture (Fig. 1A). In leached mixtures, the Ca content is
165 generally lower than predicted particularly for low calcite content (Fig. 1A). This suggests
166 partial dissolution of HAp that further occurs after complete dissolution of calcite (Balter et
167 al., 2002b). The leaching restores the Ca/P ratio of the mixture to the pure bone value (Fig.
168 1B). The leaching also restores the original bone $\delta^{44/42}\text{Ca}$ value even up to 80% of added calcite
169 (Fig. 1C). The deviation of the bulk $\delta^{44/42}\text{Ca}$ value can be detectable with $\geq 10\%$ added calcite
170 and generally follows the theoretical balance between the HAp and the calcite end-members.
171



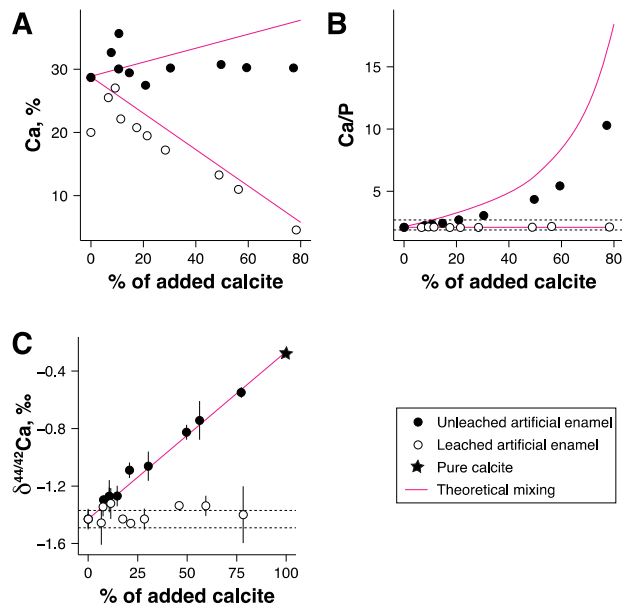
172

173 **Fig. 1:** Effect of leaching on the Ca concentration (A), Ca/P ratio (B), and the $\delta^{44/42}\text{Ca}$ (C) value of artificial fossil
 174 bone material (SRM-1400 + SRM-915b). Percentage of added calcite is relative to the initial total mass of the
 175 mixture. Error bars are typically < 5% in panel A. Dotted lines in panel B represent the natural range of the Ca/P
 176 ratio from 1.9 to 2.7 in biological HAP. Dotted lines in panel C represent the 2 standard deviations of the mean
 177 $\delta^{44/42}\text{Ca}$ value of raw material. The $\delta^{44/42}\text{Ca}$ value is reported relative to ICP-Ca Lyon.

178

179 In the mixture of enamel with increasing proportion of calcite, the Ca concentration
 180 and Ca/P ratio between unleached and leached samples follow the same pattern than for
 181 bone (Fig. 2A, 2B). Dissolution of enamel is also observed for low calcite content as measured
 182 Ca concentrations stand below theoretical values (Fig. 2A). Similar to bone, deviation of the
 183 bulk $\delta^{44/42}\text{Ca}$ value is detected with $\geq 10\%$ added calcite, which follows the theoretical balance
 184 between the HAP and the calcite end-members, and leaching restores the initial enamel
 185 $\delta^{44/42}\text{Ca}$ value even up to 80% of added calcite (Fig. 1C). Porosity in bone and enamel is
 186 variable (see Kral et al., 2021 for a discussion) but is grossly 40% in bone and one order of
 187 magnitude less ($\sim 1\%$) in enamel (Wang and Cerling, 1994). Given that bone and enamel can
 188 accommodate calcite to a magnitude equal to porosity, the results from artificial fossil bone
 189 and enamel suggest that the incorporation of calcite in natural fossil materials can be
 190 detected in the bone $\delta^{44/42}\text{Ca}$ value but not for that enamel, except for unrealistic calcite
 191 $\delta^{44/42}\text{Ca}$ value or important secondary porosity (e.g., post burial enamel surface cracks). This
 192 assumption is confirmed with results obtained on natural fossil bone and enamel.

193



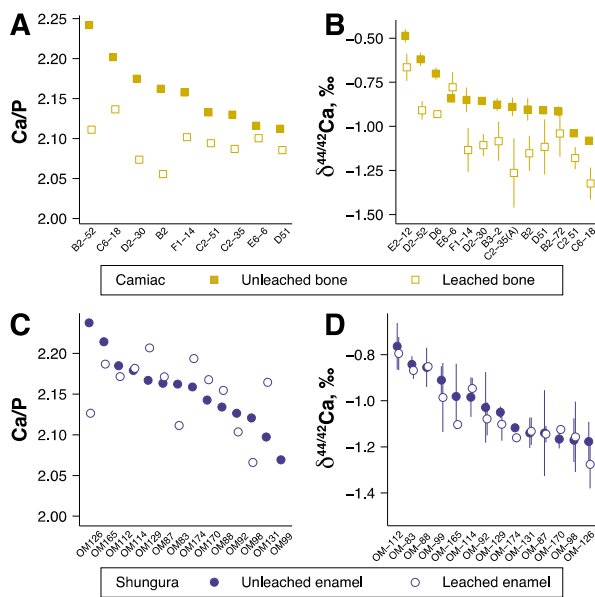
194

195 **Fig. 2:** Effect of leaching on the Ca concentration (A), Ca/P ratio (B), and the $\delta^{44/42}\text{Ca}$ (C) value of artificial fossil
 196 bone material (enamel + SRM-915b). Percentage of added calcite is relative to the initial total mass of the
 197 mixture. Error bars are typically < 5% in panel A. Dotted lines in panel B represent the natural range of the Ca/P
 198 ratio from 1.9 to 2.7 in biological HAp. Dotted lines in panel C represent the 2 standard deviations of the mean
 199 $\delta^{44/42}\text{Ca}$ value of raw material. The $\delta^{44/42}\text{Ca}$ value is reported relative to ICP-Ca Lyon.

200

201 Fossil bone from Camiac (Table S2) shows a Ca/P decrease indicative of efficient
 202 removal of additive Ca of calcitic origin (Fig. 3A) along with a consistent decrease (-0.21 ± 0.11 ,
 203 $n = 12$) of the $\delta^{44/42}\text{Ca}$ values during the leaching (Fig. 3B), validating the assumption that the
 204 added diagenetic Ca from calcite with a higher $\delta^{44/42}\text{Ca}$ value than bone has been removed
 205 accordingly. Fossil enamel unleached and leached samples from Shungura (Table S2) show a
 206 Ca/P ratio that remains constant (Fig. 3C) and similar $\delta^{44/42}\text{Ca}$ values (Fig. 3D), suggesting that
 207 non-significant amount of calcite was present in raw samples and that a quantitative
 208 fractionation of Ca isotopes occurred during leaching.

209



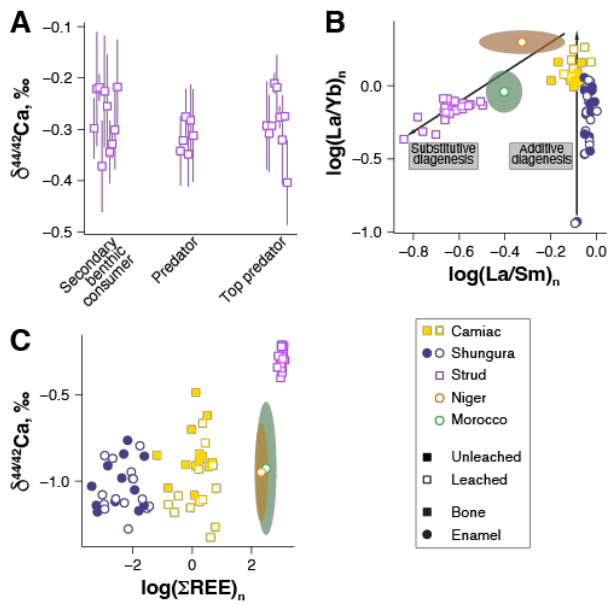
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211

212 **Fig. 3:** Effect of leaching on the Ca/P ratio and $\delta^{44/42}\text{Ca}$ value of natural fossil bone from Camiac (A and B) and
 213 fossil enamel from Shungura (C and D). The variability of the Ca/P ratio is in the natural range of the Ca/P ratio
 214 in biological Hap. The $\delta^{44/42}\text{Ca}$ value is reported relative to ICP-Ca Lyon.

215

216 Finally, the Devonian bone from Strud (Table S2) show invariable $\delta^{44/42}\text{Ca}$ values with
 217 no preserved trophic information (Fig. 4) despite the leaching procedure. Here, the range of
 218 variation of the $\delta^{44/42}\text{Ca}$ values (0.19‰, n = 25) is much more contracted than at Camiac
 219 (0.84‰, n = 26) or Shungura (0.51‰, n = 28). Note that Shungura samples are represented
 220 by two taxa of suids only (*Metridiochoerus* sp. and *Notochoerus* sp.), thus are not
 221 representative of the possible full trophic variability. The Ca/P range of variation at Strud (2.12
 222 \pm 0.07, \pm 2SD, n = 25) is in the variability of biological HAP and does not indicate any Ca excess.



223

224 **Fig. 4:** (A) $\delta^{44/42}\text{Ca}$ values of leached bone at Strud. Trophic levels are assessed based on ecomorphological
 225 features. (B) $(\text{La}/\text{Sm})_n$ vs $(\text{La}/\text{Yb})_n$ distribution of fossil samples analyzed in the study. Morocco and Niger samples
 226 are from Hassler et al. (2018). REE have been normalized to PAAS. (C) $(\Sigma\text{REE})_n$ vs $\delta^{44/42}\text{Ca}$ distribution of fossil
 227 samples analyzed in the study. Morocco and Niger samples are from Hassler et al. (2018). REE have been
 228 normalized to PAAS. The $\delta^{44/42}\text{Ca}$ value is reported relative to ICP-Ca Lyon.

229

230 Taken together, the results of leaching experiments suggest that Ca isotopes in bone,
 231 but not in enamel, are sensitive to an additive-like diagenesis probably because bone porosity
 232 is high enough to accommodate significant amount of calcite to modify the original HAp
 233 $\delta^{44/42}\text{Ca}$ value. In case of clear additive-like diagenesis (artificial fossil bone and enamel,
 234 Camiac fossil bone) the original HAp $\delta^{44/42}\text{Ca}$ value is restored by leaching. However, in the
 235 case of the Strud fossil bones, the original HAp $\delta^{44/42}\text{Ca}$ value cannot be restored by leaching
 236 as suggested by the absence of trophic pattern. At Strud, fossil bone probably experienced a
 237 substitutive-like diagenesis that was sufficiently pervasive to reset original HAp $\delta^{44/42}\text{Ca}$
 238 values.

239

240 4. Discussion

241 The type of fossil bone and enamel diagenesis, additive or substitutive, can be
 242 depicted by the REE pattern, and therefore help to detect whether the Ca isotope
 243 composition is potentially reset or not. REE trapping in fossil bone or enamel is a post-mortem
 244 phenomenon that occurs through two main processes, namely adsorption and substitution.

245 The suite of REE differs by ionic radius, middle REE (Sm) having an ionic radius close to that of
246 Ca (~ 100 pm) and thus more liable to incorporate the HAp crystal lattice compared to light
247 (La) or heavy (Yb) REE. The REE pattern can thus typify the nature of diagenetic processes
248 because the relative partitioning among REE will be characteristic of the incorporation
249 process. A quantitative incorporation without relative REE partitioning first occurs during
250 early diagenesis, followed during protracted diagenesis by a non-quantitative adsorption
251 mechanism controlled by surface crystal-chemical properties. Those mechanisms are
252 considered additive in that the REE content increases during diagenesis, constituting an extra
253 burden of trace elements that can be theoretically removed by leaching. While this
254 assumption can be easily accepted concerning fossils that have undergone early diagenesis
255 only, the efficiency of leaching is questionable in case of protracted diagenesis during which
256 recrystallization inevitably occurred (Trueman and Tuross, 2002). Note that Hap
257 recrystallisation, as witnessed by the increase of crystallite size can occur within a few years
258 post-mortem (Trueman et al., 2004), but is linked to the exposure of HAp crystallite surfaces
259 during the breakdown of collagen (Trueman et al., 2008). Finally, a substitution mechanism,
260 which is controlled by bulk crystal-chemical properties, can occur during extensive diagenesis,
261 leading to strong REE partitioning (Reynard et al., 1999), along with progressive
262 transformation of HAp into francolite (Trueman and Tuross, 2002). The effects of this late-
263 stage substitutive-like diagenesis are anticipated to be insensitive to leaching.

264 As expected, the Late Pleistocene Camiac bone and Early Pleistocene Shungura
265 enamel samples exhibit $(La/Sm)_n$ and $(La/Yb)_n$ ratios characteristic of early diagenesis,
266 supporting the idea that well preserved $\delta^{44/42}Ca$ values (Fig. 4B) can be obtained in leached
267 samples, which is necessary for bone (Fig. 3B) but not for enamel (Fig. 3D) due to the presence
268 of secondary Ca-carbonate in bone but not in enamel. Leaching has no influence on the
269 $(La/Sm)_n$ and $(La/Yb)_n$ ratios for both bone from Camiac and enamel from Shungura (Table
270 S2). The REE pattern at Strud is indicative of a substitutive-like diagenesis (Fig. 4B), and despite
271 leaching, the associated bone $\delta^{44/42}Ca$, which are contracted and invariable among trophic
272 positions, are reset by diagenesis. The picture is however different for fossil enamel. For
273 instance, Cretaceous dinosaur enamel samples from Hassler et al. (2018), which have not
274 been leached, still exhibit highly variable $\delta^{44/42}Ca$ values (from -1.6‰ to -0.4‰) with
275 preserved trophic systematics, despite obvious signs of substitutive-like diagenesis, i.e., hat-
276 shaped REE patterns (Hassler et al., 2018) and low $(La/Sm)_n$ ratios (Fig. 4B). Fossil bone or

277 enamel are open systems and accumulate REE during diagenesis, being through the
278 adsorption and/or the substitution processes. The REE concentration (ΣREE)_n thus increases
279 during diagenesis, spanning four orders of magnitude between Shungura enamel samples
280 ($0.009 \pm 0.020 \mu\text{g/g}$, $\pm 2\text{SD}$, $n = 28$, Table S2) and Strud bone samples ($1028 \pm 724 \mu\text{g/g}$, $\pm 2\text{SD}$,
281 $n = 25$, Table S2, Fig. 4C). Dinosaurs enamel samples from Morocco ($417 \pm 581 \mu\text{g/g}$, $\pm 2\text{SD}$, n
282 $= 23$) and from Niger ($237 \pm 282 \mu\text{g/g}$, $\pm 2\text{SD}$, $n = 49$) display high (ΣREE)_n concentrations, but
283 still variable $\delta^{44/42}\text{Ca}$ values despite the absence of leaching.

284 The substitutive-like REE pattern is thus not a good proxy of Ca isotope extended
285 diagenesis in fossil enamel. An explanation is that the enamel one percent of porosity is
286 insufficient to accommodate enough Ca in diagenetic fluids and/or secondary minerals that
287 will further substitute with HAp to significantly modify the isotopic composition of the enamel
288 bulk Ca. This assumption holds providing a realistic $\delta^{44/42}\text{Ca}$ value of the diagenetic pool (fluid
289 or mineral), which must stand close to the value of river water, i.e., $\sim 0\text{‰}$ (Heuser et al., 2011;
290 Martin et al., 2017). A simple mass balance calculation indicates that a 1% porosity full of Ca
291 necessitates the diagenetic pool to be $\pm 4\text{‰}$ different from the HAp to modify the $\delta^{44/42}\text{Ca}$
292 value by $\pm 0.1\text{‰}$. In bone, the situation is clearly different because the porosity ($\sim 40\%$) grossly
293 equals the HAp Ca content, so the diagenetic pool can have realistic $\delta^{44/42}\text{Ca}$ value, i.e., close
294 to that of HAp. Taking all the above results into account, we conclude that Ca isotopes are
295 largely immune to diagenesis in fossil enamel, but not in fossil bone.

296

297 5. Conclusion

298 We demonstrate that an addition of a diagenetic phase containing Ca in the form of
299 Ca-carbonate significantly modifies the bulk $\delta^{44/42}\text{Ca}$ value of artificial fossil bone and enamel,
300 whose initial composition can be restored using leaching. For fossils that have undergone
301 weak diagenesis, this process is reproduced in fossil bone, but not in fossil enamel, which
302 cannot accommodate enough diagenetic Ca in the porosity to modify the bulk $\delta^{44/42}\text{Ca}$ value.
303 For fossils that have undergone extensive diagenesis, as typified by the REE patterns, leaching
304 is unable to restore the initial bone $\delta^{44/42}\text{Ca}$ value, while enamel can still contain non-
305 significantly modified $\delta^{44/42}\text{Ca}$ values, even in the absence of leaching. The diagenetic Ca
306 content that can be added in the porosity during early diagenesis of enamel is too low and is
307 further unable to modify its bulk $\delta^{44/42}\text{Ca}$ value by substitution. The REE pattern of fossil

308 enamel is thus not a reliable proxy for extensive diagenetic alteration of Ca isotopes, which
309 remains to be potentially uncovered by crystal-chemical means.

310

311 **Declaration of competing interest**

312 The authors declare that they have no known competing financial interests or per-
313 sonal relationships that could have appeared to influence the work reported in this paper.

314

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329

330 **Appendix A. Supplementary material**

331 Supplementary figure 1: Mass fractionation in three-isotope space $\delta^{43/42}\text{Ca}$ vs $\delta^{44/42}\text{Ca}$
332 calculated from the measurements of the samples in this study.

333 Supplementary table 1: Table of results for artificial fossil bone and enamel samples.

334 Supplementary table 2: Table of results for natural fossil bone and enamel samples.

335

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- 457

1 **Limits of calcium isotopes diagenesis in fossil bone and enamel**

2

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21

22 **Abstract**

23 Diagenesis has been recognized for decades to significantly alter the trace elements biogenic
24 **signatures** in fossil tooth enamel and bone that are routinely used for paleobiological and
25 paleoenvironmental reconstructions. **This signature is modified during diagenesis according**
26 **to a complex continuum between two main processes, addition, and substitution.** For an
27 additive-like, or early diagenesis, the trace elements biogenic profiles can be restored by
28 leaching secondary minerals, but this technique is inefficient for a substitutive-like, or
29 extensive diagenesis for which secondary trace elements are incorporated into the biogenic
30 mineral. This scheme is however unclear for Ca, the major cation in tooth enamel and bone
31 hydroxylapatite, which stable isotope composition ($\delta^{44/42}\text{Ca}$) also conveys biological and
32 environmental information. We present a suite of leaching experiments for monitoring
33 $\delta^{44/42}\text{Ca}$ values in artificial and natural fossil enamel and bone from different settings. The
34 results show that enamel $\delta^{44/42}\text{Ca}$ values are insensitive to an additive-like diagenesis that
35 involves the formation of secondary Ca-carbonate mineral phases, while bone shows a
36 consistent offset toward ^{44}Ca -enriched values, that can be restored to the biogenic baseline
37 by a leaching procedure. In the context of a substitutive-like diagenesis, bone exhibits
38 constant $\delta^{44/42}\text{Ca}$ values, insensitive to leaching, and shows a REE pattern symptomatic of
39 extensive diagenesis. Such a REE pattern can be observed in fossil enamel **for which** $\delta^{44/42}\text{Ca}$
40 values are still fluctuating and follow a trophic pattern. We conclude that Ca isotopes in fossil
41 enamel are probably not prone to extensive diagenesis and argue that this immunity is due
42 to the very low porosity of enamel that cannot accommodate enough secondary minerals to
43 significantly modify the isotopic composition of the enamel Ca pool.

44

45 **1. Introduction**

46 Bone, dentin, and enamel are valuable archives for recording information about the
47 life history and environment of fossil and extant vertebrates. The information is embedded
48 as trace elements or isotopic ratios signature during the genesis of the mineralized tissue, a
49 signature that is further obfuscated by diagenesis during fossilization. Regarding trace
50 elements, the comprehension of diagenetic mechanisms has greatly benefited from in-situ
51 laser ablation measurements (Kohn and Moses, 2013; Kral et al., 2022), highlighting the
52 complex interplay of the diffusion, adsorption, and transport reaction processes. However,
53 while sophisticated techniques are now deployed to fully integrate all the hallmarks of
54 diagenesis (Suarez and Kohn, 2020; Weber et al., 2021; Kral et al., 2022), the results still
55 recognize the central importance of porosity which was already suspected in the 2000's
56 (Hedges, 2002; Trueman and Tuross, 2002; Kral et al., 2021). Porosity mediates fluid
57 circulation and secondary mineral deposition, and increases as a result of collagen
58 degradation, augmenting the exposure of carbonated hydroxylapatite (HAp) crystallites in the
59 first stages of bone diagenesis. Because porosity is forty times higher in bone than in enamel
60 (Wang and Cerling, 1994), the latter is widely accepted to be far more resistant to diagenesis
61 than bone and is nowadays the tissue of reference for measuring trace elements
62 concentration (Le Houedec et al., 2013; Joannes-Boyau et al., 2019; Nava et al., 2020; Funston
63 et al., 2022) and/or isotope composition (Martin et al., 2014, 2015; Jaouen et al., 2017;
64 McCormack et al., 2022).

65 In all types of tissue, some diagenetic overprints can be reverted using leaching
66 pretreatment (Sillen and LeGeros, 1991; Koch et al., 1997; Nielsen-Marsh and Hedges, 2000;
67 Balter et al., 2002a; Wathen et al., 2022). When formation of secondary mineral phases is
68 thought to have occurred (additive-like diagenesis), leaching is potentially efficient at
69 removing these minerals. One issue is that leaching is often performed in batch, leading
70 samples to be leached more than necessary, therefore dissolving the most soluble HAp
71 fraction that is the most pristine (Sillen and LeGeros, 1991; Balter et al., 2002a). The efficiency
72 of the leaching method is even more questionable when diffusion of diagenetic elements
73 (substitutive-like diagenesis) into the HAp has occurred.

74 All told, these considerations hold for trace elements, but the situation is unclear for
75 major elements. Traditionally, the $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ values of CO_3 present in HAp (~5 wt%) are
76 measured in leached samples due to the widespread presence of calcite in the porosity, but

77 the $\delta^{18}\text{O}$ value of HAp PO_4 (~55 wt%) is always measured in raw samples. The isotope
78 composition of Ca of HAp (~40 wt%) varies according to trophic position (Skulan and DePaolo,
79 1999; Heuser et al., 2011; Martin et al., 2018; Hu et al., 2022) and physiology (Tacaïl et al.,
80 2017; Hassler et al., 2021; Koutamanis et al., 2021; Li et al., 2022), but deserves an assessment
81 of potential diagenetic effects on $\delta^{44/42}\text{Ca}$ values in bone and tooth enamel. Simple mass
82 balance calculations already show that changes of the original $\delta^{44/42}\text{Ca}$ values in bone and
83 tooth enamel by secondary Ca minerals are either small or **would require an uncommonly**
84 **fractionated Ca isotopes diagenetic pool** (Heuser et al., 2011; Martin et al., 2017).

85 Here, we attempt to resolve this issue by studying how Ca isotopes behave during
86 controlled experiments of artificial fossil bone and enamel leaching and in different diagenetic
87 case studies. **The Ca isotope compositions are measured along with Ca concentrations and**
88 **Ca/P ratios to monitor the origin of Ca**, and Rare Earth Elements (REE) to decipher the type
89 (additive or substitutive) and extrapolate the extend of diagenetic alteration (Reynard et al.,
90 1999; Trueman and Tuross, 2002).

91

92 **2. Material and methods**

93 First, we analyzed the Ca isotope composition in leached and unleached artificial
94 samples mimicking fossil bone and enamel that have undergone an additive-like diagenesis.
95 These samples consist of mixtures of HAp and calcite powder which $\delta^{44/42}\text{Ca}$ value are distinct
96 and constrained. An artificial fossil bone was prepared using the bone ash certified reference
97 material (SRM-1400, NIST; $\delta^{44/42}\text{Ca} = -1.21 \pm 0.04\text{‰}$, $\pm 2\text{SD}$, $n = 3$) to which was added various
98 proportions of the calcite certified reference material SRM-915b (NIST, $\delta^{44/42}\text{Ca} = -0.28 \pm$
99 0.04‰ , $\pm 2\text{SD}$, $n = 21$). The SRM-1400 certified reference material has been ashed and is
100 devoid of organic matter and has a high crystallinity, both being characteristics of fossil bone.
101 Because enamel certified reference material does not exist, we prepared artificial fossil
102 enamel by mixing a sample of extant enamel from an isolated and partial molar tooth of
103 modern elephant (*Elephas maximus*; $\delta^{44/42}\text{Ca} = -1.43 \pm 0.07\text{‰}$, $\pm 2\text{SD}$, $n = 10$) with various
104 proportions of SRM-915b. Second, we analyzed a suite of natural samples from different
105 localities and ages that were selected according to their potential degree of diagenetic
106 alteration. Pleistocene bone and tooth enamel samples were chosen to be representative of
107 weak diagenesis, with preserved trace elements compositions, and Devonian bone of more

108 pronounced diagenesis. A first set of material is composed by a suite of fossil bone samples
109 (n = 13) from the Camiac cave (Gironde, France). The site is a hyena dens from the upper
110 Pleistocene (Mousterian period, 35.1 ± 2 ka BP; Balter et al., 2002b). Taxonomic information
111 is given in [Table S2](#). This material is characterized by trace elements (Sr/Ca and Ba/Ca)
112 distributions that reflect a trophic pattern (Balter et al., 2002b). A suite of fossil enamel
113 samples (n = 14) from the Plio-Pleistocene of the Shungura Formation in the Lower Omo
114 Valley (Ethiopia, sample age bracketed between 3.6 and 1.76 Ma; Boisserie et al., 2008)
115 represents the second set of material. Here, the material is composed by only two extinct
116 genera of large African suids (*Metridiochoerus* sp. and *Notochoerus* sp., [Table S2](#)). Leached
117 fossil bone at Shungura exhibit Sr/Ca ratios characteristic of a trophic pattern (Sillen, 1986).
118 A third set of samples gather vertebrate (fish and tetrapod) fossil bone (n = 25, [Table S2](#)) from
119 the Devonian locality of Strud (360 Ma, Belgium; Olive et al., 2015). To our knowledge, no
120 geochemical studies has already been performed on this material.

121 The samples were processed in the clean room and analyzed on the facilities at the
122 Laboratoire de Géologie de Lyon: Terre, Planètes, Environnements (LGL-TPE, ENS de Lyon).
123 All the samples (ca. ~ 5 mg) were leached using 0.5 ml of 0.1 M acetic acid per mg of sample
124 during 30 min in an ultrasonic bath and rinsed with 18.2 M Ω /cm grade MilliQ water. This
125 represents 2.5-fold the quantity of acid necessary to dissolve an equivalent amount of pure
126 calcite. The samples were dissolved in distilled concentrated HNO₃ and an aliquot measured
127 for major elements (P and Ca) by ICP-OES (Thermo Scientific, iCap 6000 Radial) and trace
128 elements (REE) by ICP-MS (Thermo Scientific, iCap-Q) according to Balter and Lécuyer (2004).
129 Calcium from the remaining solution was purified by ion-exchange chromatography and Ca
130 isotope composition measured by MC-ICP-MS (Thermo Scientific, Neptune Plus) according to
131 Tacail et al. (2014). Briefly, three ion-exchange chromatography steps are necessary, first
132 allowing the recovery of Ca, Fe and Sr only, the second to purify Ca from Fe and the third to
133 remove Sr ([Table 1](#)). Blanks for the procedure did not exceed 100 ng of Ca.

134

135 **Table 1:** Detailed procedure for Ca extraction and purification.

1. Matrix elimination		
AG50W-X12 resin (200-400 mesh) ~ 2 mL		
Step	Eluent	Vol. (mL)
Condition	1 M HCl	10
Load	1 M HCl	2+1
Elution (matrix)	1 M HCl	55
Ca elution (Ca, Sr, Fe)	6 M HCl	10
2. Fe elimination if necessary		
AG1-X8 resin (100-200 mesh) ~ 2 mL		
Step	Eluent	Vol. (mL)
Condition	6 M HCl	10
Load	6 M HCl	1+1
Elution (Ca, Sr)	6 M HCl	6
3. Sr elimination		
Sr-Specific resin (Eichrom) ~ 0.7 mL		
Step	Eluent	Vol. (mL)
Condition	3 M HNO ₃	5
Load	3 M HNO ₃	0.5+0.5
Elution (Ca)	3 M HNO₃	6

136

137 Calcium isotope abundance ratios (⁴⁴Ca/⁴²Ca and ⁴³Ca/⁴²Ca) were measured using a
 138 Neptune Plus multi-collector ICP-MS using the standard-sample-standard bracketing method.
 139 All Ca isotope compositions are expressed using the delta notation, calculated as follows:

140

$$141 \quad \delta^{44/42}\text{Ca} = \left(\frac{(^{44}\text{Ca}/^{42}\text{Ca})_{\text{Sample}}}{0.5 \times (^{44}\text{Ca}/^{42}\text{Ca})_{\text{ICP-Ca-Lyon}}^{n-1} + 0.5 \times (^{44}\text{Ca}/^{42}\text{Ca})_{\text{ICP-Ca-Lyon}}^{n+1}} - 1 \right) \times 1000$$

142

143 The ICP Ca Lyon standard, used routinely in Lyon, was used as the bracketing standard.
 144 The results can be converted relative to the SRM915a with an offset of -0.52‰.

145

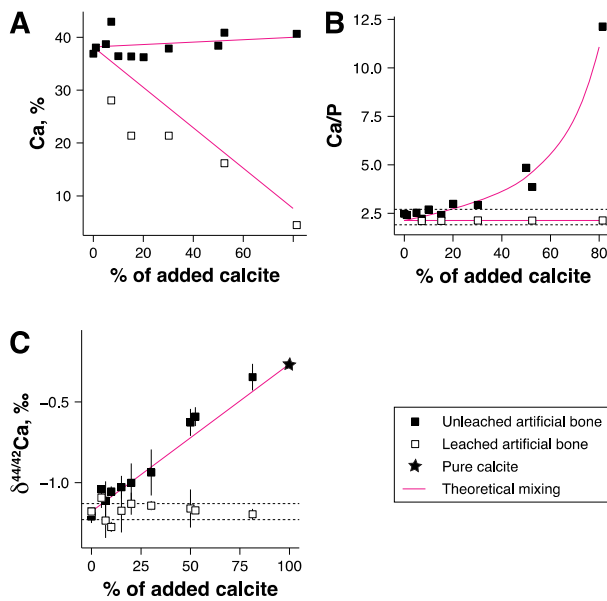
146 3. Data presentation and interpretation

147 Results are given in **Table S1** for artificial fossil bone and enamel and in **Table S2** for
 148 natural fossil bone and enamel. The mass fractionation measured in this study agrees with
 149 the 0.5 slope predicted by the linear approximation of mass-dependent fractionation (**Fig. S1**).

150 We first test the effect of leaching on the $\delta^{44/42}\text{Ca}$ value of the certified reference
 151 material SRM-1400 and found no difference after leaching ($\delta^{44/42}\text{Ca} = -1.21 \pm 0.03\text{‰}$, $\pm 2\text{SD}$,
 152 $n = 3$) compared to unleached material ($\delta^{44/42}\text{Ca} = -1.21 \pm 0.04\text{‰}$, $\pm 2\text{SD}$, $n = 3$). Absence of
 153 difference between the $\delta^{44/42}\text{Ca}$ values of unleached and leached enamel is also observed as

154 we measured a constant $\delta^{44/42}\text{Ca}$ value of -1.43‰ (Table S1). Because of the very small
155 amount of processed material (~ 5 mg), the remaining bone or enamel residue after leaching
156 is so small that it is impossible to weigh it, so Ca concentration for leached material is
157 underestimated. Nevertheless, using in 0.1 M acetic acid for 30 min generally leads to a loss
158 of about 30% of initial material. The absence of Ca isotope fractionation during leaching
159 therefore indicates that the dissolution of HAp is quantitative and that leaching *per se* does
160 not affect fossil bone and enamel $\delta^{44/42}\text{Ca}$ values.

161 In the mixture of bone HAp with increasing proportion of added calcite, the slight
162 theoretical increase of Ca concentration is barely detected in unleached mixtures, probably
163 due to the ~5% error inherent to ICPMS measurements (Fig. 1A), but the decrease of Ca
164 content is inambiguous in leached mixture (Fig. 1A). In leached mixtures, the Ca content is
165 generally lower than predicted particularly for low calcite content (Fig. 1A). This suggests
166 partial dissolution of HAp that further occurs after complete dissolution of calcite (Balter et
167 al., 2002b). The leaching restores the Ca/P ratio of the mixture to the pure bone value (Fig.
168 1B). The leaching also restores the original bone $\delta^{44/42}\text{Ca}$ value even up to 80% of added calcite
169 (Fig. 1C). The deviation of the bulk $\delta^{44/42}\text{Ca}$ value can be detectable with $\geq 10\%$ added calcite
170 and generally follows the theoretical balance between the HAp and the calcite end-members.
171



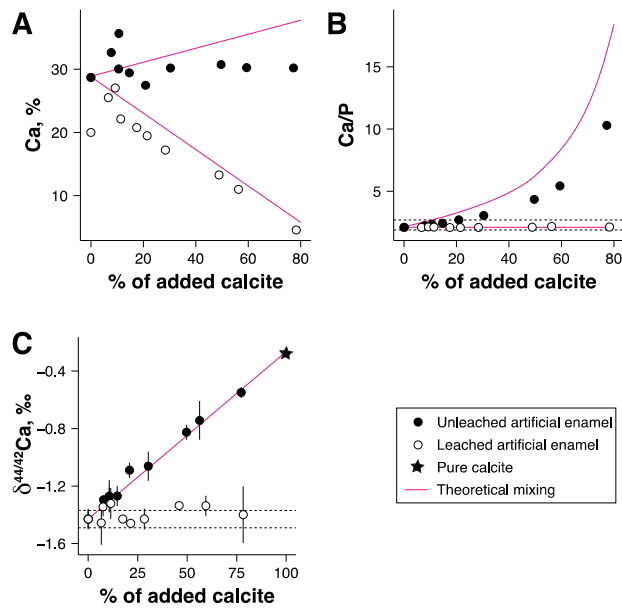
172

173 **Fig. 1:** Effect of leaching on the Ca concentration (A), Ca/P ratio (B), and the $\delta^{44/42}\text{Ca}$ (C) value of artificial fossil
 174 bone material (SRM-1400 + SRM-915b). Percentage of added calcite is relative to the initial total mass of the
 175 mixture. Error bars are typically < 5% in panel A. Dotted lines in panel B represent the natural range of the Ca/P
 176 ratio from 1.9 to 2.7 in biological HAP. Dotted lines in panel C represent the 2 standard deviations of the mean
 177 $\delta^{44/42}\text{Ca}$ value of raw material. The $\delta^{44/42}\text{Ca}$ value is reported relative to ICP-Ca Lyon.

178

179 In the mixture of enamel with increasing proportion of calcite, the Ca concentration
 180 and Ca/P ratio between unleached and leached samples follow the same pattern than for
 181 bone (Fig. 2A, 2B). Dissolution of enamel is also observed for low calcite content as measured
 182 Ca concentrations stand below theoretical values (Fig. 2A). Similar to bone, deviation of the
 183 bulk $\delta^{44/42}\text{Ca}$ value is detected with $\geq 10\%$ added calcite, which follows the theoretical balance
 184 between the HAP and the calcite end-members, and leaching restores the initial enamel
 185 $\delta^{44/42}\text{Ca}$ value even up to 80% of added calcite (Fig. 1C). Porosity in bone and enamel is
 186 variable (see Kral et al., 2021 for a discussion) but is grossly 40% in bone and one order of
 187 magnitude less ($\sim 1\%$) in enamel (Wang and Cerling, 1994). Given that bone and enamel can
 188 accommodate calcite to a magnitude equal to porosity, the results from artificial fossil bone
 189 and enamel suggest that the incorporation of calcite in natural fossil materials can be
 190 detected in the bone $\delta^{44/42}\text{Ca}$ value but not for that enamel, except for unrealistic calcite
 191 $\delta^{44/42}\text{Ca}$ value or important secondary porosity (e.g., post burial enamel surface cracks). This
 192 assumption is confirmed with results obtained on natural fossil bone and enamel.

193



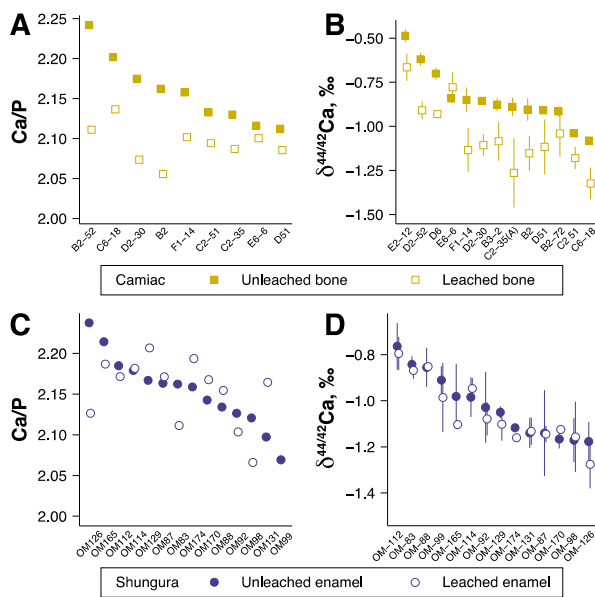
194

195 **Fig. 2:** Effect of leaching on the Ca concentration (A), Ca/P ratio (B), and the $\delta^{44/42}\text{Ca}$ (C) value of artificial fossil
 196 bone material (enamel + SRM-915b). Percentage of added calcite is relative to the initial total mass of the
 197 mixture. Error bars are typically < 5% in panel A. Dotted lines in panel B represent the natural range of the Ca/P
 198 ratio from 1.9 to 2.7 in biological HAp. Dotted lines in panel C represent the 2 standard deviations of the mean
 199 $\delta^{44/42}\text{Ca}$ value of raw material. The $\delta^{44/42}\text{Ca}$ value is reported relative to ICP-Ca Lyon.

200

201 Fossil bone from Camiac (Table S2) shows a Ca/P decrease indicative of efficient
 202 removal of additive Ca of calcitic origin (Fig. 3A) along with a consistent decrease (-0.21 ± 0.11 ,
 203 $n = 12$) of the $\delta^{44/42}\text{Ca}$ values during the leaching (Fig. 3B), validating the assumption that the
 204 added diagenetic Ca from calcite with a higher $\delta^{44/42}\text{Ca}$ value than bone has been removed
 205 accordingly. Fossil enamel unleached and leached samples from Shungura (Table S2) show a
 206 Ca/P ratio that remains constant (Fig. 3C) and similar $\delta^{44/42}\text{Ca}$ values (Fig. 3D), suggesting that
 207 non-significant amount of calcite was present in raw samples and that a quantitative
 208 fractionation of Ca isotopes occurred during leaching.

209



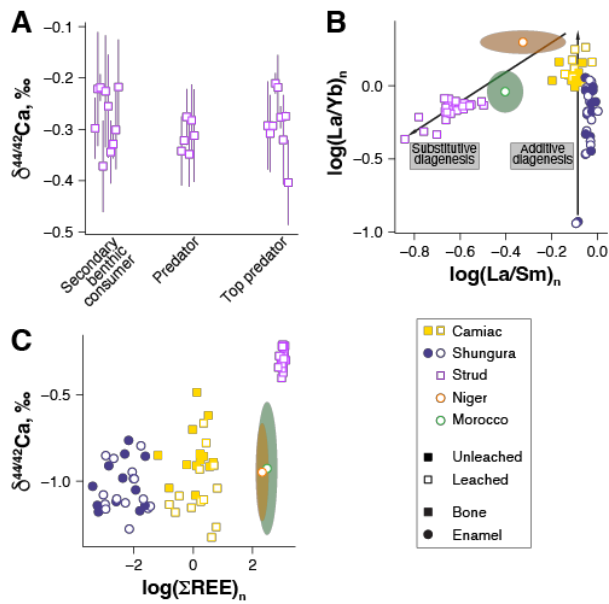
210

211

212 **Fig. 3:** Effect of leaching on the Ca/P ratio and $\delta^{44/42}\text{Ca}$ value of natural fossil bone from Camiac (A and B) and
 213 fossil enamel from Shungura (C and D). The variability of the Ca/P ratio is in the natural range of the Ca/P ratio
 214 in biological Hap. The $\delta^{44/42}\text{Ca}$ value is reported relative to ICP-Ca Lyon.

215

216 Finally, the Devonian bone from Strud (Table S2) show invariable $\delta^{44/42}\text{Ca}$ values with
 217 no preserved trophic information (Fig. 4) despite the leaching procedure. Here, the range of
 218 variation of the $\delta^{44/42}\text{Ca}$ values (0.19‰, n = 25) is much more contracted than at Camiac
 219 (0.84‰, n = 26) or Shungura (0.51‰, n = 28). Note that Shungura samples are represented
 220 by two taxa of suids only (*Metridiochoerus* sp. and *Notochoerus* sp.), thus are not
 221 representative of the possible full trophic variability. The Ca/P range of variation at Strud (2.12
 222 ± 0.07 , $\pm 2\text{SD}$, n = 25) is in the variability of biological HAP and does not indicate any Ca excess.



223

224 **Fig. 4:** (A) $\delta^{44/42}\text{Ca}$ values of leached bone at Strud. Trophic levels are assessed based on ecomorphological
 225 features. (B) $(\text{La}/\text{Sm})_n$ vs $(\text{La}/\text{Yb})_n$ distribution of fossil samples analyzed in the study. Morocco and Niger samples
 226 are from Hassler et al. (2018). REE have been normalized to PAAS. (C) $(\Sigma\text{REE})_n$ vs $\delta^{44/42}\text{Ca}$ distribution of fossil
 227 samples analyzed in the study. Morocco and Niger samples are from Hassler et al. (2018). REE have been
 228 normalized to PAAS. The $\delta^{44/42}\text{Ca}$ value is reported relative to ICP-Ca Lyon.

229

230 Taken together, the results of leaching experiments suggest that Ca isotopes in bone,
 231 but not in enamel, are sensitive to an additive-like diagenesis probably because bone porosity
 232 is high enough to accommodate significant amount of calcite to modify the original HAp
 233 $\delta^{44/42}\text{Ca}$ value. In case of clear additive-like diagenesis (artificial fossil bone and enamel,
 234 Camiac fossil bone) the original HAp $\delta^{44/42}\text{Ca}$ value is restored by leaching. However, in the
 235 case of the Strud fossil bones, the original HAp $\delta^{44/42}\text{Ca}$ value cannot be restored by leaching
 236 as suggested by the absence of trophic pattern. At Strud, fossil bone probably experienced a
 237 substitutive-like diagenesis that was sufficiently pervasive to reset original HAp $\delta^{44/42}\text{Ca}$
 238 values.

239

240 4. Discussion

241 The type of fossil bone and enamel diagenesis, additive or substitutive, can be
 242 depicted by the REE pattern, and therefore help to detect whether the Ca isotope
 243 composition is potentially reset or not. REE trapping in fossil bone or enamel is a post-mortem
 244 phenomenon that occurs through two main processes, namely adsorption and substitution.

245 The suite of REE differs by ionic radius, middle REE (Sm) having an ionic radius close to that of
246 Ca (~ 100 pm) and thus more liable to incorporate the HAp crystal lattice compared to light
247 (La) or heavy (Yb) REE. The REE pattern can thus typify the nature of diagenetic processes
248 because the relative partitioning among REE will be characteristic of the incorporation
249 process. A quantitative incorporation without relative REE partitioning first occurs during
250 early diagenesis, followed during protracted diagenesis by a non-quantitative adsorption
251 mechanism controlled by surface crystal-chemical properties. Those mechanisms are
252 considered additive in that the REE content increases during diagenesis, constituting an extra
253 burden of trace elements that can be theoretically removed by leaching. While this
254 assumption can be easily accepted concerning fossils that have undergone early diagenesis
255 only, the efficiency of leaching is questionable in case of protracted diagenesis during which
256 recrystallization inevitably occurred (Trueman and Tuross, 2002). Note that Hap
257 recrystallisation, as witnessed by the increase of crystallite size can occur within a few years
258 post-mortem (Trueman et al., 2004), but is linked to the exposure of HAp crystallite surfaces
259 during the breakdown of collagen (Trueman et al., 2008). Finally, a substitution mechanism,
260 which is controlled by bulk crystal-chemical properties, can occur during extensive diagenesis,
261 leading to strong REE partitioning (Reynard et al., 1999), along with progressive
262 transformation of HAp into francolite (Trueman and Tuross, 2002). The effects of this late-
263 stage substitutive-like diagenesis are anticipated to be insensitive to leaching.

264 As expected, the Late Pleistocene Camiac bone and Early Pleistocene Shungura
265 enamel samples exhibit $(La/Sm)_n$ and $(La/Yb)_n$ ratios characteristic of early diagenesis,
266 supporting the idea that well preserved $\delta^{44/42}Ca$ values (Fig. 4B) can be obtained in leached
267 samples, which is necessary for bone (Fig. 3B) but not for enamel (Fig. 3D) due to the presence
268 of secondary Ca-carbonate in bone but not in enamel. Leaching has no influence on the
269 $(La/Sm)_n$ and $(La/Yb)_n$ ratios for both bone from Camiac and enamel from Shungura (Table
270 S2). The REE pattern at Strud is indicative of a substitutive-like diagenesis (Fig. 4B), and despite
271 leaching, the associated bone $\delta^{44/42}Ca$, which are contracted and invariable among trophic
272 positions, are reset by diagenesis. The picture is however different for fossil enamel. For
273 instance, Cretaceous dinosaur enamel samples from Hassler et al. (2018), which have not
274 been leached, still exhibit highly variable $\delta^{44/42}Ca$ values (from -1.6‰ to -0.4‰) with
275 preserved trophic systematics, despite obvious signs of substitutive-like diagenesis, i.e., hat-
276 shaped REE patterns (Hassler et al., 2018) and low $(La/Sm)_n$ ratios (Fig. 4B). Fossil bone or

277 enamel are open systems and accumulate REE during diagenesis, being through the
278 adsorption and/or the substitution processes. The REE concentration (ΣREE)_n thus increases
279 during diagenesis, spanning four orders of magnitude between Shungura enamel samples
280 ($0.009 \pm 0.020 \mu\text{g/g}$, $\pm 2\text{SD}$, $n = 28$, Table S2) and Strud bone samples ($1028 \pm 724 \mu\text{g/g}$, $\pm 2\text{SD}$,
281 $n = 25$, Table S2, Fig. 4C). Dinosaurs enamel samples from Morocco ($417 \pm 581 \mu\text{g/g}$, $\pm 2\text{SD}$, n
282 $= 23$) and from Niger ($237 \pm 282 \mu\text{g/g}$, $\pm 2\text{SD}$, $n = 49$) display high (ΣREE)_n concentrations, but
283 still variable $\delta^{44/42}\text{Ca}$ values despite the absence of leaching.

284 The substitutive-like REE pattern is thus not a good proxy of Ca isotope extended
285 diagenesis in fossil enamel. An explanation is that the enamel one percent of porosity is
286 insufficient to accommodate enough Ca in diagenetic fluids and/or secondary minerals that
287 will further substitute with HAp to significantly modify the isotopic composition of the enamel
288 bulk Ca. This assumption holds providing a realistic $\delta^{44/42}\text{Ca}$ value of the diagenetic pool (fluid
289 or mineral), which must stand close to the value of river water, i.e., $\sim 0\text{‰}$ (Heuser et al., 2011;
290 Martin et al., 2017). A simple mass balance calculation indicates that a 1% porosity full of Ca
291 necessitates the diagenetic pool to be $\pm 4\text{‰}$ different from the HAp to modify the $\delta^{44/42}\text{Ca}$
292 value by $\pm 0.1\text{‰}$. In bone, the situation is clearly different because the porosity ($\sim 40\%$) grossly
293 equals the HAp Ca content, so the diagenetic pool can have realistic $\delta^{44/42}\text{Ca}$ value, i.e., close
294 to that of HAp. Taking all the above results into account, we conclude that Ca isotopes are
295 largely immune to diagenesis in fossil enamel, but not in fossil bone.

296

297 5. Conclusion

298 We demonstrate that an addition of a diagenetic phase containing Ca in the form of
299 Ca-carbonate significantly modifies the bulk $\delta^{44/42}\text{Ca}$ value of artificial fossil bone and enamel,
300 whose initial composition can be restored using leaching. For fossils that have undergone
301 weak diagenesis, this process is reproduced in fossil bone, but not in fossil enamel, which
302 cannot accommodate enough diagenetic Ca in the porosity to modify the bulk $\delta^{44/42}\text{Ca}$ value.
303 For fossils that have undergone extensive diagenesis, as typified by the REE patterns, leaching
304 is unable to restore the initial bone $\delta^{44/42}\text{Ca}$ value, while enamel can still contain non-
305 significantly modified $\delta^{44/42}\text{Ca}$ values, even in the absence of leaching. The diagenetic Ca
306 content that can be added in the porosity during early diagenesis of enamel is too low and is
307 further unable to modify its bulk $\delta^{44/42}\text{Ca}$ value by substitution. The REE pattern of fossil

308 enamel is thus not a reliable proxy for extensive diagenetic alteration of Ca isotopes, which
309 remains to be potentially uncovered by crystal-chemical means.

310

311 **Declaration of competing interest**

312 The authors declare that they have no known competing financial interests or per-
313 sonal relationships that could have appeared to influence the work reported in this paper.

314

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329

330 **Appendix A. Supplementary material**

331 Supplementary figure 1: Mass fractionation in three-isotope space $\delta^{43/42}\text{Ca}$ vs $\delta^{44/42}\text{Ca}$
332 calculated from the measurements of the samples in this study.

333 Supplementary table 1: Table of results for artificial fossil bone and enamel samples.

334 Supplementary table 2: Table of results for natural fossil bone and enamel samples.

335

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