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1	Pilot-scale direct UV-C photodegradation of pesticides in groundwater and recycled
2	wastewater for agricultural use.
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- 27 Abstract
- 28

Pesticides widely used for intensive agriculture may leach to groundwater and pose problems 29 to drinking water and irrigation. UV-C disinfection systems (UV-DS) for water disinfection 30 can be used also for the abatement of organic micropollutants. A pilot-scale continuous flow-31 32 through UV-DS system was evaluated for its degradation efficiency of atrazine (ATR), malathion (MAL) and glyphosate (GLY) from 40 L water. Groundwater used to irrigate 33 potato fields and recycled wastewater used to wash potatoes were treated without catalysts to 34 avoid any toxicity effect on potatoes. Chromatographic methods were used to quantify very 35 low pesticide levels before and after UV-C treatments ($<10 \ \mu g \ L^{-1}$), while a specific method 36 was adapted to analyse traces of GLY (0.0008-10 µg L⁻¹) in recycled wastewater containing 37 suspended particulate matter (SPM). ATR was completely eliminated from groundwater after 38 15 min photodegradation while 80% was removed from the turbid wastewater after 25 min. 39 For MAL, 70-80% was removed in 25 minutes from the groundwater. For wastewater, the 40 initial concentration was important for the performance of the photolytic process. An amount 41 of 75% of GLY was eliminated after 10 minutes irradiation at concentrations higher than 42 those found in natural groundwater. In wastewater, the UV-C treatment was less efficient 43 because GLY was mainly adsorbed to SPM which obstruct the photodegradation process. 44 45 Therefore, the pilot-scale UV-DS using a turbulent flow and a multiple-lamp system was performant to remove quantitatively traces of pesticides from large volumes of water, by 46 direct photolytic oxidation, when the turbidity of the treated water was limited. 47

48

49 Key words

50 UV-C photolysis; recycled water; pesticides; glyphosate; pilot-scale treatment; ultra-trace
51 analysis

53 Abbreviations

- 54
- 55 AOP: Advanced oxidation processes
- 56 AMPA: aminomethylphosphonic acid
- 57 ATR: Atrazine
- 58 CIP: Cleaning in place
- 59 DEA: Desethylatrazine
- 60 DI: Deionized
- 61 DIA: Deisopropyl-atrazine
- 62 DOM: Dissolved organic matter
- 63 FLD: Fluorescence detection
- 64 FMOC: Fluorenylmethoxycarbonyl
- 65 GC-MS: Gas chromatography coupled to mass spectrometry
- 66 GLY: Glyphosate
- 67 HA: Humic acids
- 68 IARC: International Agency for Research on Cancer
- 69 LC: Liquid chromatography
- 70 LOD: Limit of detection
- 71 LOQ: Limit of quantification
- 72 MAL: Malathion
- 73 SIP: Sterilization in place
- 74 S/N: Signal to noise
- 75 SPE: Solid phase extraction
- 76 SPM: Suspended particulate matter
- 77 UV-DS: UV disinfection systems
- 78

79 **1. Introduction**

The extension of intensive agriculture worldwide has led to an increase in the use of 80 pesticides for plant and crop protection in the past decades and they have become among the 81 most frequently occurring organic pollutants in surface and groundwater [1, 2]. A majority of 82 pesticides are persistent and resist to natural degradation processes and so they remain for 83 extended periods in soils (after field applications) where they can accumulate. Some 84 pesticides may leach to groundwater and pose problems for drinking water and irrigation. 85 Because of possible adverse effects on the environment and human health caused by a 86 majority of pesticides, the European Union established a maximum permissible concentration 87 of 0.1 μ g L⁻¹ in drinking water for one target pesticide and a total concentration of 0.5 μ g L⁻¹ 88 for the sum of them [3]. 89

Each year in France, more than 5 billion m³ of water is collected for agricultural use, of which 60% is used for irrigation. In potato production, 0.6 m³ water is required to produce 1 kg of potato tubers. In a context of sustainable development and to save drinking water, wastewater is expected to be recycled and reused, but the presence of pesticides above regulated concentrations can, as for irrigation water, pose a serious problem.

Atrazine (ATR) is one of the most commonly used herbicides in the world [4]. It is not 95 96 volatile, not very polar and not very soluble in water, and it can accumulate onto soil organic matter (Table 1). Before its ban in France in 2003, the organochlorine ATR was commonly 97 used in corn and wheat cultivation, but also in potato fields. ATR and two of its degradation 98 products (desethylatrazine (DEA) and hydroxyatrazine) were among the most detected 99 100 substances in the French rivers between 2009 and 2013 [5]. ATR is classified in group 3 of the International Agency for Research on Cancer (IARC), with an inadequate evidence for the 101 102 carcinogenicity in humans but a sufficient evidence in experimental animals (Table 1).

After 2003, ATR was largely replaced by glyphosate (GLY) in France, which is now the most 103 104 widely used herbicide and became (with its main transformation product aminomethylphosphonic acid (AMPA)) the most measured in French rivers [6]. GLY is a 105 non-selective broad-spectrum organophosphate herbicide, not volatile, very polar and very 106 soluble into water (Table 1). However, it can also accumulate into soils through the formation 107 of complexes with soil cations, depending on soil pH [7]. In 2015, IARC classified GLY as 108 "probably carcinogenic to humans" (Group 2A) (Table 1). GLY drifts can also cause leaf 109 deformation of potato plants and can consequently lead to serious economic losses for 110 impacted producers after lot rejections. Just like GLY, malathion (MAL) is an 111 112 organophosphorous compound but is generally used as an insecticide. As ATR, MAL is neither volatile, nor being very polar and not very soluble into water, so it can accumulate 113 onto soil organic matter (Table 1). It has been forbidden in France since 2007, but it was 114 115 previously used to eliminate fungi from seed potato lots. Even if it is no more a major pesticide found in French rivers, it is a dangerous persistent compound, which has been 116 117 classified as "probably carcinogenic to humans" (Group 2A) by IARC (Table 1), as one of its transformation product malaoxon. 118

119 Table 1: Physico-chemical and carcinogenic characteristics of the studied pesticides

	Atrazine	Malathion	Glyphosate	
Chemical structure		H ₃ C O CH ₃ H ₃ C O CH ₃		
рКа	1.64	-	2.34; 5.73; 10.2	
Vapour Pressure (25°C) (Pa)	3.85×10^{-5}	4.5×10^{-4}	1.31 × 10 ⁻⁵	
Solubility (20°C) (mg L ⁻¹)	30	145	10,500 (pH=2)	
Log K _{ow}	2.5	2.8	-3.2 (pH=5-9)	
$K_{oc} (L kg^{-1})$	86	151-308	81-7564	
Maximal absorption wavelength (nm)	225 [8]	210 [8]	< 200 (pH=7) [9]	
Carcinogenicity (IARC)	Group 3	Group 2A	Group 2A	

As pesticides can adversely impact aquatic environment at low levels and must be analysed at 123 124 trace levels in groundwater as well as in drinking water, sensitive analytical methods must be developed. ATR and MAL can be analysed using liquid chromatography (LC) or gas 125 126 chromatography coupled to mass spectrometry detection (GC-MS) but a pre-enrichment procedure (using solid phase extraction (SPE) for example) is generally required to reach 127 concentrations lower than 0.1 μ g L⁻¹ [10-12]. GLY and AMPA are generally analysed using 128 129 LC coupled to MS or to fluorescence detection (FLD), but they can also be analysed using GC requiring previous derivatization [13-15]. As for ATR and MAL, various preparation 130 steps must be developed before analysis (SPE, lyophilisation, large volume injections...) to 131 132 analyse GLY and AMPA at trace levels [16-19].

Ultraviolet (UV) radiation-based processes are feasible treatment solutions to eliminate or 133 drastically decrease a broad range of persistent organic pollutants such as pesticides in 134 135 aqueous effluents. UV-C direct photolysis has been reported to achieve better degradation of persistent organic compounds than UV-A due to its higher energy [20, 21]. Photolysis 136 processes include direct photolysis of compounds that are excited by absorbing energy from 137 UV-C photons (180-280 nm) resulting in bond cleavage or rearrangement. ATR, MAL and 138 GLY can potentially absorb such energies and could be possibly degraded (Table 1). But 139 140 indirect photolysis can also occur when photons are absorbed by photoactive compounds producing reactive oxygen species (ROS) or dissolved organic matter (DOM) in excited state 141 [22]. The application of a photocatalyst in advanced oxidation processes (AOPs) can enhance 142 the degradation of too stable or photo-inactive compounds by a significant increase of the 143 generation of ROS that oxidize and degrade them [23, 24]. UV-C disinfection systems (UV-144 DS) can be simultaneously used for water disinfection (eliminating pathogens) [25] but also 145 for the abatement of organic micropollutants in water. The lower generation of toxic 146 byproducts during the process compared to AOPs using catalysts can be an advantage of UV-147

DS to treat aqueous effluents in contact with food or vegetables [26]. For example, the UV-C treatment of MAL in presence of TiO₂ catalyst produces more phosphate byproduct, which can be toxic [27].

151 In some agricultural practices, cooperatives ensure the reception of tons of potatoes that must be washed before their conditioning and delivery. Potatoes tubers are washed with natural 152 groundwater and then wastewater is intended to be continuously collected and recycled for 153 154 subsequent potatoes tubers cleaning, in order to save thousands cubic meters of water and to develop a circular economic model. In such current process, leached persistent pesticides can 155 accumulate after few washing cycles, leading to significant exceedance of tolerated levels. 156 157 UV-DS can be a promising technology to eliminate leached pesticides from wastewater in order to reuse it after each washing cycle. But continuous flow-through UV-DS reported 158 experiments are rare and their degradation efficiency are not well known [28]. Moreover, the 159 160 scaling up of photolysis or photocatalytic reactors is limited and studies are required for testing large-scale photodegradation processes and determining kinetic models for a variety of 161 aqueous effluents, particularly natural water containing mixtures of persistent pollutants [29]. 162

In this study, a pilot-scale disinfection process using UV-C irradiation for treating water in a 163 continuous flow-through system was evaluated for the first time for its degradation efficiency 164 of three pesticides, ATR, MAL and GLY (and some of their transformation products). 165 Groundwater used to irrigate potato fields and recycled wastewater used to wash potatoes 166 were treated. No chemical catalyst was introduced to avoid any interaction with potatoes used 167 for animal or human feed. The disappearance kinetics of the three targeted compounds were 168 established. In the case of GLY/AMPA a new SPE-HPLC-Fluorimetry analytical method had 169 to be developed to reach ultra-trace levels in water containing solid particulate matter. 170

171

172 **2.** Material and methods

173 2.1 Materials

174 2.1.1 Chemicals

Standards of ATR, MAL, deisopropyl-atrazine (DIA), diethyl-atrazine (DEA), malaoxon,
isomalathion and fluorenylmethoxycarbonyl chloride (FMOC-Cl) were obtained from Sigma
Aldrich (Saint Quentin Fallavier, France). GLY, AMPA, derivatized GLY-FMOC and
AMPA-FMOC were from Dr Ehrenstorfer GmbH (Augsburg, Germany). Deuterated labelled
analytical standards atrazine-d5 (ATR-d5) and malathion-d10 (MAL-d10) were from Sigma
Aldrich.

The LC grade solvents acetonitrile, dichloromethane, methanol and ethylacetate were 181 182 purchased from Fischer Scientific (Illkirsh, France) while octanol was from Sigma Aldrich. Boric acid, sodium bicarbonate and formate ammonium were from Fisher Scientific. 183 Potassium chloride and sodium hydroxide were from Sigma Aldrich, while disodium EDTA 184 185 was from VWR (Strasbourg, France). Formate ammonium buffer 10 mM (pH 8) was prepared by mixing 10 mL of formate ammonium 1 M and 6.19 mL of sodium hydroxide 1 M in 1 L 186 pure deionized (DI) water. Borate buffer 0.3 M (pH 8.7) was prepared by mixing 25 mL of a 187 solution of boric acid and potassium chloride (0.3 M each) with 10.7 mL of sodium hydroxide 188 0.3 M in 100 mL DI water. 189

Oxy-Anios 5 (Laboratoires ANIOS, France), the solution used to clean the UV-C pilot, was a
solution of peracetic acid (48 mg g⁻¹) and hydrogen peroxide (255.9 mg g⁻¹).

192

193 2.1.2 Water sampling

Pure DI water was supplied by a Milli-Q water system (Fisher Scientific). A large volume of drilling water as well as wastewater from potato tubers cleaning was necessary to conduct the pilot study. Drilled water used for irrigation of potato crops was sampled from a potato field area near Evreux, Normandy France. Direct access to the drilling point was possible and

sampling was done with the aid of a lance. A description of the drilling point is presented in 198 199 Figure S1 (supplementary materials). Groundwater is commonly used for crop irrigation due to its high degree of purity as a result of natural filtration through the soil. However, in recent 200 201 years pure groundwater has become increasingly scarce due to drought and/or otherwise, contaminated by pesticide residues. Groundwater had a pH of 8.2, a mean conductivity of 202 0.453 ± 0.006 mS cm⁻¹ (n=9) and a mean turbidity of 21.6 ± 3.0 FTU (n=9). Compared with 203 typical French tap water (turbidity ≤ 2 FTU and 0.180 \leq conductivity ≥ 1 mS cm⁻¹, quality 204 reference values from 2001-1220 French decree), the collected groundwater was slightly 205 206 turbid and contained few ionized mineral salts.

207 Wastewater from potato tuber cleaning was sampled with a pump, which was inserted into a clean settled reservoir that collects wastewater from a potato packaging company. The 208 reservoir is made up of four water pools which are inter-connected. As such, movement of the 209 wastewater from the potatoes cleaning point crosses the four water pools, with debris settling 210 211 in each of the pools, thus water arriving at the last pool is clearer and is intended be reused for cleaning potato tubers. Apart from this pre-treatment step, there was no additional treatment 212 applied to the wastewater. As such, it is possible that all potato crop-related treatments and 213 214 microbial contaminants or pathogens of the potato tubers maybe leached into the reservoirs, leading to an accumulation of pesticides and proliferation of pathogens. It is why the 215 SurePure TurbulatorTM device was expected to be installed after the last reservoir for an 216 217 additional treatment of the washing water (Figure S2). Wastewater had a pH of 7.9, a mean conductivity of 0.477±0.020 mS cm⁻¹ (n=9) and a mean turbidity of 492±91 FTU (n=9). As 218 219 compared with typical French tap water, the recycled water was very turbid and contained few 220 ionized mineral salts.

After sampling, ground- or wastewater was stored at room temperature in 10 L plastic containers, previously rinsed to eliminate plastic debris and avoid any possible contamination of the samples before analyses.

After water collection at the outlet of the disinfection setup, ground- or waste-water was centrifuged at 10,000 g for 5 min to eliminate a maximum of suspended particulate matter (SPM).

227

228 2.2 Experimental disinfection setup and procedure

229 2.2.1 Experimental setup

230 Fig. 1 shows the pilot-scale disinfection set-up (SurePure Europe). The patented SurePure TurbulatorTM device exploits swirling turbulent flows and is designed for continuous flow of 231 turbid fluids, so it is well adapted for treating complex solutions such as wine, blood, milk 232 233 [30]. The turbulent flow and the multiple-lamp system (primarily used as germicide lamps) increase the liquid exposure to UV-C, limiting biofouling, enabling greater homogeneity and 234 235 efficiency in purification. The disinfection pilot consists of a reservoir tank that can contain 100 L water, of a liquid pump (flow-rate 4500 L h⁻¹), of 10 quartz pipes of 80 cm long and 78 236 mm diameter in a series circuit, each containing one mercury lamp (254 nm wavelength) (Fig. 237 1). Each lamp is protected by a quartz sleeve and can deliver 36 W (which corresponds to 29 238 239 W in the UV-C range), but with 15% loss due the quartz tubing material (measured and specified by the manufacturer). 240

- Figure 1: (a) Pilot-scale disinfection UV-C pilot: "1" and "2" indicate the entrance water flow, "3" and "4" indicate the outlet flow; (b) 3D-modelling of the stainless steel tube; (c) the arrows indicate how the solution moves around the UV-C lamp.



248 2.2.2 Photolytic experiments procedure

249 *Cleaning and sterilization in place (CIP and SIP steps):*

This procedure aimed at preparing the installation for the reception of the water to be treated. 250 251 It consisted of a chemical cleaning and decontamination using Oxy-Anios 5. In the starting tank, 200 mL of Oxy-Anios 5 (oxidizing agent) in 39.8 L water (0.5% mixture) was disposed. 252 253 This mixture flowed through the installation for 10 min with no light, passing through the 254 pump and the pipes, then returning in the starting tank in a close circuit. After 10 min, the installation was entirely rinsed with 100 L of tap water to eliminate any remaining Oxy-Anios 255 5. Then the starting tank was again charged with 40 L of tap water which flowed for 10 min 256 257 through the machine but with lights switched on, in order to eliminate residual Oxy-Anios 5. 258 After rinsing, the water was eliminated and the machine was purged. At the end of each water treatment, the CIP and SIP steps were undertaken. 259

260 *Treatment step:*

To prevent some damages of the machine, especially of the quartz tubes, the water to be 261 treated was previously filtered through sieves of 450 and 250 µm. The coarsest fractions of 262 particulate matter were eliminated. The 40 L water samples (to be treated) were charged in the 263 starting tank, then the connection between the tank and the pump was opened and the water 264 265 circulation was switched on. During the first 5 min, the lamps were not switched on allowing the water to homogenize correctly in all the installation. As 32 seconds are required to flush 266 40 L of sample once in the tubes, 5 min was enough to reach a steady concentration after few 267 circulation cycles [25]. As a control sample, a non-treated water sub-sample was collected 268 269 before the UV-C lamps were switched on. After each exposure time (1, 3, 5, 10, 15 and 25 min), the lamps and the pump were stopped while 200 mL of sub-sample of treated water was 270 271 collected at the end-point of the pipes circuit. Then, the collecting point was closed and the lamp and the pump were restarted again until the next collection. 272

273 **2.3 Analytical methods**

274 2.3.1 Sample preparation and GC-MS analysis of ATR, MAL and some transformation275 products

After water centrifugation, a preconcentration step was performed on the supernatant using 276 solid phase extraction (SPE) with Oasis HLB 200 mg cartridges (Waters, Saint-Quentin-en-277 Yvelines, France). Solid phase conditioning was done with 6 mL dichloromethane, 6 mL 278 acetonitrile followed by 6 mL DI water. The prepared water sample (V=200 to 1000 mL) was 279 passed through the SPE cartridge by a vacuum pump. The cartridge was finally dried by a 280 nitrogen flow (1 h). The analytes were eluted with 2×4 mL dichloromethane. An amount of 281 10 µL octanol was added as solvent keeper and the eluate was evaporated to dryness 45 min 282 at 45°C and 200 mBar (Mivac concentrator, Genevac, Fisher Scientific). At the end, 170 µL 283 ethyl acetate was added alongside 10 µL of internal standards of deuterated ATR-d5 and 284 MAL-d10 (at 100 mg L⁻¹ in methanol). An aliquot of 1 µL was injected (splitless mode, 285 280°C) into a gas chromatographer GC (7890B, Agilent, Santa Clara, USA), coupled with a 286 287 mass spectrometer MS (7000C). Separation was performed using a 60 m×0.25 mm Zebron ZB- SemiVolatiles capillary column (0.25 µm film thickness) obtained from Phenomenex, 288 with helium at 1.4 mL min⁻¹. The oven temperature was programmed to begin at 100°C (1 289 min), increased to 190°C (30°C min⁻¹), then 200°C (0.8°C min⁻¹), 230°C (5°C min⁻¹) and 290 291 finally 240°C (4°C min⁻¹). The MS detector operated at 70 eV. Quantification was based on selected ion monitoring for better sensitivity. The linearity of the detection was in the range 292 0.1-8 mg L⁻¹ for ATR, MAL, DEA, DIA, malaoxon, isomalathion, with $r^2 > 0.995$ for 293 294 calibration curves. Limits of detection (LOD) (signal to noise S/N=3) and quantification (LOQ) (S/N=10) of these compounds were evaluated on real samples of recycled wastewater 295 (V=1000 mL) spiked with 2 μ g L⁻¹ of ATR, MAL and some transformation products (Table 296 2). Their recoveries were in the range 75-106% (Table 2). 297

Table 2: Limits of detection, of quantification and recovery yields for the different pesticides
and some transformation products spiked in the turbid wastewater and analysed after SPE
concentration.

	LOD (ng L ⁻¹)	LOQ (ng L ⁻¹)	Recoveries (%) (n=5)	
ATR	0.71	2.6	78.0±9.6	
DIA	1.3	4.1	106.0±8.7	
DEA	0.3	1.9	75.0±9.3	
MAL	0.1	2.3	95.0±16.5	
Malaoxon	0.5	2.5	98.0±13.7	
Isomalathion	2.8	8.6	90.0±18.7	
GLY *	0.25	0.84	58.0 (n= 3)	

** Results obtained from spiked groundwater, only for GLY.*

304 2.3.2. Sample preparation and LC-FLD analysis of GLY and AMPA

305 After water centrifugation and before the preconcentration step, 8 mL of EDTA 0.1 M (pH=10, adjusted with NaOH 5M) was added to 200 mL of the aqueous supernatant and was 306 307 homogenized prior to filtration through a cellulose acetate filter (5-8 µm, Fisher Scientific) under vacuum. The preconcentration step was performed using a PSOH Chromabond (500 308 309 mg) SPE cartridges (Macherey Nagel, Hoerdt, France). Solid phase conditioning was done 310 with 2×2 mL DI water, then 2×1 mL sodium bicarbonate (1 M) followed by 2×2 mL DI water. Then the water sample mixed with EDTA was passed through the SPE cartridge with 311 the aid of a vacuum pump. After drying the cartridge under N₂ flow, analytes were eluted with 312 2×1 mL KCl 1 M. 313

For the derivatization step, 0.4 mL borate buffer 0.3 M (pH 8.7) was added to the eluate and 314 shaken for 20 s. Then 0.4 mL of FMOC-Cl (6 mM into acetonitrile) was added and the 315 316 mixture was shaken in darkness for 20 min. Thereafter, 3×1 mL ethyl acetate were added and after shaking, the lower aqueous phase was recovered and analysed using LC (Beckman Gold 317 126, Villepinte, France) coupled with a fluorimetric (FLD) detector Prostar 363 Varian 318 (Agilent). An amount of 20 µL of sample was injected into a Gemini C₁₈ column (150×4.6 319 mm, $d_p=5 \mu m$) (Phenomenex), at 30°C and with a flow-rate of 1 mL min⁻¹. The mobile phase 320 was composed of solutions (A) (ammonium formate 10 mM, pH 9.5) and (B) (acetonitrile). 321 The analysis began with 15% (B) for 5 min, followed by a linear gradient to 98% (B) over 15 322 min before stabilizing at 98% (B) over 5 min. Optimal fluorescence excitation/emission 323 wavelengths were, respectively, 250/310 nm to obtain a sensitive detection. The linearity of 324 the detection was in the range 1-30 μ g L⁻¹ for GLY and AMPA, with r²>0.997 for calibration 325 curves. The LOD, LOQ and recoveries are discussed in the section 3.1 (Table 2). 326

328 **3. Results and discussion**

329 3.1. Adaptation of the analytical methodology for the analysis of GLY at ultra-trace 330 levels in wastewater.

GLY is often analysed at trace levels in drinking, surface water or groundwater, which do not 331 contain high amounts of SPM or DOM and rarely in raw turbid water, where it has been 332 demonstrated that GLY can be associated to particulate matter [17, 18, 31]. However, in the 333 334 present study, although filtration and centrifugation were performed before analysis of the turbid recycled wastewater, organic colloids (DOM) and nano- or micro-particles from SPM 335 could not be fully removed. Thus, there was a requirement for the development of new 336 337 strategies in the sample preparation to analyse GLY at ultra-trace levels in turbid water. It was particularly critical to quantify GLY at concentrations in the $\mu g L^{-1}$ range as direct 338 photodegradation processes in these low concentration ranges, encountered in environmental 339 340 water effluents, are not well known.

As GLY and AMPA were detected through a fluorimetric detector, allowing higher sensitivity 341 342 than UV or MS detectors, they were derivatized using an excess of FMOC-Cl, because it can also react with other amines or amino acids which may be present in recycled water. FMOC-343 Cl was prepared into acetonitrile to obtain better derivatization yields [32]. It is known that 344 345 the reaction must be done in alkaline conditions and that derivatization yields increase when pH increase from 7.0 to 9.1, but here, a pH=8.7 was selected to avoid high hydrolysis of 346 FMOC-Cl into FMOC-OH at more alkaline pH [33]. As GLY and AMPA were obtained after 347 concentration of 200 mL water in 2 mL aqueous solutions containing concentrated KCl, 348 several volumes (300-600 µL) and concentrations (0.125-0.3 M) of FMOC-Cl were tested. 349 Even if higher amounts of FMOC-Cl favor the derivatization process, a medium volume (400 350 μ L) of the highest concentrated solution was selected to avoid sample dilution. At last, the 351 best reaction time was determined allowing the formation of derivatized analytes without the 352

formation of secondary by-products. The reaction time was tested between 0-24 h [34, 35]. 353 354 No significant difference on derivatization yields was observed after 20 min reaction (p>0.05, Student test, n=3) and after 1h, higher amounts of secondary by-products could be observed. 355 Therefore, 20 min derivatization was selected, obtaining mean derivatized glyphosate (GLY-356 FMOC) yields of 100.8±9.7% (n=3). Using DI water spiked with GLY and AMPA, the limit 357 of detection (LOD) (S/N=3) and of quantification (LOQ) (S/N=10) using LC-FLD after 358 derivatization were 0.013 μ g L⁻¹ and 0.040 μ g L⁻¹, respectively, for GLY-FMOC and 0.09 μ g 359 L⁻¹ and 0.30 µg L⁻¹, respectively, for AMPA-FMOC. A review on analytical methods used for 360 361 GLY/AMPA analyses in water using LC-FLD mentioned current LODs between 0.01-0.1 µg L^{-1} for both chemicals [36]. GLY had to be quantified into water at levels as low as 0.1 µg L^{-1} , 362 so the attained LOQ could be considered satisfactory. 363

GLY spiked in turbid water can be lost, due to various sorption phenomena on solid materials. 364 Therefore, a concentration process through SPE, allowing a concentration factor of 100, was 365 366 developed to accurately determine ultra-traces of GLY. Derivatized GLY-FMOC and AMPA-FMOC are anionic species in a large alkaline pH range, so various anion exchange SPE 367 cartridges were tested, such as Chromabond PSOH (strong anion exchanger in the OH⁻ form), 368 369 Chromabond NH₂ (weak anion exchanger with aminopropyl bonded material) and Oasis MAX (containing a mixed-mode polymeric sorbent with quaternary ammonium groups). 370 Various eluting solvents were tested to desorb GLY from the SPE cartridges. With Oasis 371 MAX, aqueous solutions containing HCl (0.5 M) or sodium citrate (0.6 M) were tested, and 372 low or inconsistent recoveries (2±2% and 140±24% (n=3 replicates), respectively) for GLY-373 FMOC were obtained. The HCl acid was not a good eluting solvent for the further 374 derivatization step whereas sodium citrate yielded a more favorable alkaline pH=8. With 375 376 Chromabond NH₂, aqueous solutions containing sodium citrate (0.6 M) and a phosphate buffer (0.1 M, pH=8) gave 43±9% and 108±5% recoveries, respectively (n=3). Unfortunately, 377

the chromatographic resolution of AMPA-FMOC was deteriorated when phosphate buffer 378 379 was used. With Chromabond PSOH, an aqueous phase containing KCl (1 M) was used as eluting solvent and gave a mean recovery of 102±6% (n=3). This latter combination for the 380 SPE step was chosen rather than the combination involving Oasis MAX/sodium citrate 381 because the relative standard deviation was lower. The reproducibility of the recovery yields 382 383 for quantifying GLY at very low levels in spiked DI water after SPE, evaluated on 3 tests on different days, were 91.9 \pm 20.4% and 98.7 \pm 8.7% for GLY spiked at 0.1 µg L⁻¹ and 0.6 µg L⁻¹, 384 respectively. 385

As previously mentioned, quantifying GLY in drilling or recycled wastewater appeared not so 386 387 easy, as they contain more or less SPM/DOM which required an elimination before LC analysis. Several membrane filters were tested, with cellulose acetate, nylon or nitrocellulose 388 materials which demonstrated 11%, 24% and 45% GLY loss, respectively. As such as, the 389 390 filter had to be carefully chosen to accurately quantify GLY in raw water containing particles, and cellulose acetate material seemed appropriate. Moreover, even if ground- and wastewater 391 were centrifuged after their collection at the outlet of the disinfection setup in order to 392 eliminate SPM, mineral or organic colloids, they could not be totally eliminated. Mineral 393 colloids could contain iron or aluminium oxides that can sorb GLY [37]. Also, it has been 394 395 demonstrated that metal cations could interfere on the GLY and AMPA derivatization [17, 38]. Therefore, EDTA was introduced after centrifugation to precipitate mineral colloids and 396 associated bivalent metallic species in order to eliminate them with further filtration. In the 397 best analytical conditions, GLY spiked at 10 μ g L⁻¹ into groundwater and placed into the 398 399 disinfection pilot without irradiation, was quantified with 58% recovery (Table 2). The 42% loss could be attributed to the sorption of GLY onto SPM/colloids (which were eliminated 400 401 through centrifugation, precipitation and filtration before analysis), but also to the sorption onto the surface of the inox and quartz material of the pilot. 402

Concerning wastewater which was highly turbid and contained much more SPM, initial 403 404 concentrations of GLY were not negligible. From one collected in October 2017 and another collected in March 2018, mean concentrations of 4.1 \pm 3.2 µg L⁻¹ (n=5) and 2.6 \pm 0.9 µg L⁻¹ 405 (n=6), respectively, were measured. The main transformation product AMPA could not be 406 detected in wastewater. In comparison, GLY was initially found in groundwater at 0.3±0.2 µg 407 L^{-1} (n=3) which is significantly lower (p<0.05, Student test). Then GLY was spiked at 10 µg 408 L⁻¹ into wastewater and placed into the disinfection pilot without irradiation: it was quantified 409 with only 19% recovery. Compared with the results from groundwater, it is clear that an 410 important quantity was sorbed onto the pilot surfaces and coatings, but also on the high 411 amount of SPM/colloids and so could not be quantified in the dissolved aqueous fraction. 412

413 Considering the moderate GLY losses related to the presence of SPM/DOM into 414 groundwater, procedural LODs and LOQ using SPE and LC-FLD could be measured only 415 from spiked groundwater and were 0.84 ng L^{-1} and 0.25 ng L^{-1} , respectively (Table 2). Thus, 416 the very low LOQ values obtained for ATR, MAL and GLY (Table 2) allowed for the 417 quantification of those pesticides at ultra-trace levels into water of various turbidity, even 418 after their degradation following UV-C treatment.

419

420 **3.2.** Degradation of pesticides using the pilot-scale disinfection process

421 3.2.1. Energy of UV-C exposure per liter of treated water

422 The transmitted UV-C energy (or its power $P_{transmitted}$ in J s⁻¹) to the aqueous fluid through the 423 quartz tubing could be calculated using equation (1), knowing the power of each UV lamp 424 ($P_{UV lamp}$):

425
$$P_{\text{transmitted}} = (P_{\text{UV lamp}} \times \%_{\text{Loss}}) \times \text{number of lamps} = (29 \times 0.85) \times 10 = 246.5 \text{ J s}^{-1}$$
 (1)

426 The time (t_{pass} in s) needed for the passage of the volume of water to be treated in the machine
427 could be calculated using equation (2):

428
$$t_{\text{pass}} = \frac{(V_{\text{T}} \times 3600)}{4500} = \frac{(40 \times 3600)}{4500} = 32 \text{ s}$$
 (2)

Where V_T was the volume of water to treat (40 L) and V=4500 L was the total volume of water which could be delivered by the pump in 1 h. The energy applied to the volume of water to treat (E_{pass} in J L⁻¹) in one passage could be calculated using equation (3):

433
$$E_{\text{pass}} = \frac{(t_{\text{pass}} \times P_{\text{transmitted}})}{V_{\text{T}}} = \frac{(32 \times 246.5)}{40} = 197.2 \text{ J L}^{-1}$$
 (3)

We could then calculate the total energy of UV-C exposure (E_{exposure} in J L⁻¹ or kJ m⁻³)
(corresponding to the fluence) for each treatment time using equation (4):

436
$$E_{exposure} = \frac{time \times E_{pass}}{t_{pass}}$$
 (4)

437 Where time is the treatment time (in min). Thus, the fluence could be calculated using a 438 simple equation (5): $E_{exposure} = 369.75 \times time$ (5)

439

440 3.2.2 Dissipation of pesticides

441 *Dissipation of atrazine and malathion*

The kinetic curves of ATR and MAL degradation were compared according to the water 442 treated. The measured relative concentrations (C_t/C_0) are shown as a function of the 443 444 irradiation time on Fig. 2. The mean values of C_t/C_0 were obtained through duplicate experiments for a particular concentration of spiked pesticide, carried out on two different 445 446 days and using two different 40 L water sample cans, which explains the variability of the results. As illustrated on equation (5), the longer the residence time into the UV-DS tubes, the 447 higher the energy delivered from the UV-C lamps. Both MAL and ATR were quantitatively 448 photodegraded by UV-C irradiation but a limit appeared after a few dozen minutes. 449



Figure 2: Plots of C_t/C_0 versus photoreaction time for MAL and ATR in groundwater (a, b) and wastewater (c, d). Co is the initial concentration measured without irradiation (t=0) and Ct is the measured concentration after t (min) irradiation (n=2 experiments).

MAL and ATR could be dissipated through direct photolysis, but also through oxidation 453 454 reaction with ROS. Indeed, in the case of UV-C irradiation, the water molecules can decompose in photoactive hydroxyl radicals that absorb the energy of irradiation [39]. The 455 yield of this reactive intermediate can reach 0.280 µmol per J of absorbed radiation energy 456 [40]. In view of eq. (5), it was possible that 1760 μ g L⁻¹ could be generated per min 457 irradiation in the pilot, which is significantly above the concentrations of the dissolved 458 459 pesticides to remove. Morevover, other oxidants such as superoxide radical or singlet oxygen could be generated: the big section of the turbulator device could help to oxygenize the 460 aqueous solution during the process and promote these ROS formation [38]. 461

462 More than 80% ATR could be eliminated in less than 25 minutes irradiation (corresponding to a fluence of 9244 kJ m⁻³, eq. (5)) irrespective of the quality of water treated and for all the 463 tested concentrations, from 0.5 to 10 μ g L⁻¹ (Fig. 2b and 2d). Only 15 minutes were necessary 464 to completely eliminate ATR from the less turbid groundwater (fluence = 5546 kJ m^{-3}) (Fig. 465 2.b). For wastewater the dissipation was limited to 80%. The larger amount of interfering 466 compounds probably interacted with the ROS, limiting the efficiency of the oxidation reaction 467 in the liquid phase. Also, the persistence of residual ATR which could not be eliminated after 468 25 min irradiation was possibly due to their sorption onto SPM, where they were not 469 accessible to direct irradiation. 470

It has been reported that DEA, DIA and 2-hydroxyatrazine are the main transformation products obtained by photolysis of ATR [41, 42]. These transformation products are obtained by excitation of the triazine, the dechlorination and the insertion of the hydroxyl radical generated by the water photolysis. As discussed, the GC-MS method was adapted to analyse ultra-traces of ATR and some of its current transformation products. The analysis of groundwater and wastewater before UV-C treatment demonstrated that ATR was not detectable (<0.00071 μ g L⁻¹), contrary to some of its transformation products. DEA was

detected (<0.002 μ g L⁻¹), and DIA could be quantified at 0.15±0.04 μ g L⁻¹ (n=4) and 478 0.90±0.20 µg L⁻¹ (n=12) in ground- and wastewater, respectively. Such contamination of 479 natural water by ATR metabolites, produced from the metabolic activity of bacteria, has 480 already been reported by Moreira et al [42]. After all the UV-C treatments, even the longer 481 482 ones, DEA and DIA levels were not significantly changed in treated water compared with 483 their initial concentrations. It is possible that the hydroxylation of the chlorinated carbon of ATR increased the stability of the chemical structure of these compounds [42]. Thus, DEA 484 and DIA are difficult to remove by photolysis and it contributes to their prevalence and so to 485 their wide detection in various water environments, even groundwater, and for decades after 486 487 the phasing out of the parent molecule [43]. Also, Khan et al. [44] demonstrated that the degradation rate of DEA and DIA increased with the increase in their initial concentration 488 when they were submitted to UV-C irradiation (254 nm). Therefore, their very low initial 489 490 concentrations in ground- or wastewater might also explain why no DEA or DIA concentration change could be measured during the exposure to UV-C irradiation. 491

For MAL, the UV-C degradation was limited to 50-80% depending on water quality, for 492 concentrations ranging from 0.5 to 10 μ g L⁻¹ and after 25 min treatment (corresponding to a 493 fluence of 9244 kJ m⁻³) (Fig. 2a and 2c). It has been reported that three oxidation by-products 494 495 could be generated during photolysis [45]. The degradation of MAL could be initiated by the cleavage of the P-S bond to obtain the diethyl-2-mercaptosuccinate, and then the mercapto 496 group was substituted by an hydroxyl group with the help of hydroxyl radicals, to obtain the 497 diethyl malate and finally, the internal hydroxyl group could react with an hydrogen to give 498 499 diethyl maleate [27]. The GC-MS analysis confirmed that malaoxon and isomalathion were not present, even at ultra-trace levels in water, before and after UV-C treatment, possibly 500 501 because the P-S bond was rapidly cleaved by photolysis.

The results presented in Fig. 2 indicates that the degradation was more efficient in 502 503 groundwater than in wastewater for both ATR and MAL. We could emphasize that SPM present in the unclear wastewater limits the interaction with photons involved in the 504 505 photolysis and sorption protects pesticides from photolysis by competitive light attenuation [46]. Moreover, the largest amount of interfering compounds (DOM and inorganic ions) in 506 wastewater should compete with ATR and MAL for photochemical reactions with ROS. 507 508 Particularly, humic substances can reduce the pesticide photodegradation by a strong quenching effect and it can also quickly react with ROS [46, 47]. 509

In both the tested water, ATR was more sensitive to UV-C degradation than MAL. Residual concentrations remaining in the solution after 25 min irradiation were always greater for MAL than for ATR. ATR presents a maximum absorption between 220-230 nm, so with a sensitization effect more favorable than for MAL, which presents a maximum absorption wavelength at the limit of the UV-C region (Table 1) [8, 41].

515 In order to better understand the photodegradation process, the kinetic of degradation was 516 evaluated for both ATR and MAL. The pseudo-first order kinetic modeling was applied in 517 waste- and groundwater according to equation (6):

518
$$\ln\left(\frac{c_t}{c_0}\right) = -k t \tag{6}$$

where C_0 and C_t represent the concentration at time 0 and t, respectively, and k (min⁻¹) is the pseudo-first order rate constant.

The values of the rate constants k and the coefficients of determination (r^2) corresponding to the mathematical modeling (eq. 6) are summarized in Table 3. The pseudo-first order model fitted well with the experimental values for ATR and MAL and it was relevant for the different concentrations tested and whatever the water quality. Fig. 3 shows the plot obtained from equation (6) which is in accordance with the mean experimental values (obtained from duplicates).

- 527 Table 3: Pseudo-first order kinetic parameters for the photodegradation of MAL and ATR in528 ground- and wastewater.

		Groundwater		Wastewater			
		k (min ⁻¹)	t _{1/2} (min)	r ²	k (min ⁻¹)	$t_{1/2}(min)$	r ²
	0.5 μg L ⁻¹	0.1852	3.7	0.996	0.0817	8.5	0.945
Atrazine	$2 \ \mu g \ L^{-1}$	0.2847	2.4	0.970	0.0594	11.7	0.980
	10 µg L ⁻¹	0.2237	3.1	0.993	0.0622	11.9	0.964
	0.5 μg L ⁻¹	0.0512	13.5	0.980	0.0471	14.7	0.980
Malathion	$2 \ \mu g \ L^{-1}$	0.0669	10.4	0.990	0.0257	27.0	0.870
	10 µg L ⁻¹	0.0456	15.2	0.992	0.0183	37.9	0.970



Although the kinetics of the photochemical reactions can be complex because of the number of steps and the reactive species involved, the OH radical is usually regarded as the sole and rate limiting reactive species [46], which explains why the pseudo-first order kinetic could properly model the photodegradation kinetic. According to first-order kinetics, the time required for dissipating 50% of the initial concentration ($t_{1/2}$) of MAL or ATR could be calculated by the equation (7):

541
$$t_{1/2} = \frac{\ln 2}{k}$$
 (7)

542 The values of $t_{1/2}$ are given in Table 3. It shows that the first stage of ATR dissipation (for 543 degrading at least one half of the molecules) was always faster than that of MAL.

In groundwater, the photodegradation process was less sensitive to the initial concentration of 544 ATR or MAL than in wastewater, in the range 0.5-10 μ g L⁻¹. The constant rates k were only 545 slightly higher at 2 μ g L⁻¹ than at 0.5 or 10 μ g L⁻¹ for both the molecules in groundwater. So 546 the fluence corresponding to the dissipation of 50% of these pesticides in groundwater was 547 between 887 and 1368 kJ per m³ of water for ATR and between 3845 and 5620 kJ per m³ for 548 MAL, in the 0.5-10 μ g L⁻¹ concentrations range. It is important to note here that continuous 549 flow-through reactor studies for organic contaminants removal by UV-C photolysis are rare at 550 the pilot scale, and that the results obtained for these pesticides are promising not only for the 551 removal capacity but also for the energy expenditure [23]. For comparison, direct UV-C 552 photolysis of a persistent pharmaceutical compound in drinking water showed degradation 553 less than 6% at 15 min, in a bench-scale continuous flow-through reactor, corresponding to a 554 fluence of 5000 kJ m⁻³ [28]. 555

For wastewater, the photodegradation of ATR and MAL was significantly slower than for groundwater. The time required to dissipate 50% of ATR was between 2.3 to 4.9 time longer in wastewater and between 2.6 to 6 times longer for MAL (Table 3). The exception was for MAL at 0.5 μ g L⁻¹ whose t_{1/2} was quite the same for the two water qualities. So at the lowest

MAL concentration, the amount of ROS seemed to be sufficient to react with MAL even in 560 the turbid water, but at MAL concentrations > 2 μ g L⁻¹, t_{1/2} increased drastically in wastewater 561 which shows that generated active species were no longer sufficient (Table 3). The fluence 562 corresponding to the dissipation of 50% ATR was between 3143 and 4400 kJ m⁻³ of 563 wastewater and increased from 5435 to 14014 kJ m⁻³ of wastewater for MAL, depending on 564 its initial concentration. Once again, the results obtained with the SurePure device are 565 promising, even for a very turbid water. For comparison, the removal of ten pesticides (of 566 which ATR) from leaching water containing DOM at pilot plant scale, using solar irradiation 567 (UV-A to UV-C), was less than 20% after 60 min [39]. 568

569

570 *Dissipation of Glyphosate*

The photolytic dissipation of GLY was investigated for 30 minutes (Fig. 4). Some authors 571 572 noted that no sign of direct photolysis was observed using UV-C irradiation for GLY at 50 mg L⁻¹ in pure water, which was justified by the absorption spectrum of GLY which is at the limit 573 574 of the UV-C region [48]. However, the sample pH is important to take into account. McConnell et al. (1993) demonstrated that the absorption spectra of GLY could shift to 575 higher values with a pH increase. Thus, the wavelength of the maximum absorption could 576 shift to 214 nm for pHs higher than 7.0 [49]. In our case, treated waters were alkaline (7.9-577 8.2) which could explain a part of the GLY photosensitivity. 578

Figure 4: Dissipation of free GLY (initial spiking: $10 \ \mu g \ L^{-1}$) in groundwater (n=3 tests) and wastewater (n=1 test) after UV-C irradiation (measured in the dissolved aqueous fraction from t=0).

583



584

As already mentioned (section 3.1), the direct photolysis of GLY in the very low 586 concentration ranges, found in environmental water (µg L⁻¹), is not well known, as 587 photodegradation of GLY is more often studied using spiked GLY in the mg L⁻¹ range and is 588 usually assisted by photocatalysts in AOPs [50]. Here, the dissipation of GLY (spiked at 10 589 µg L⁻¹) seemed very fast in groundwater because only one minute of treatment was enough to 590 remove 57% of GLY (fluence = 370 kJ m⁻³) (Fig. 4). It is important to note that the reported 591 Ct/Co values were calculated from the initial Co concentration measured in the dissolved 592 aqueous fraction at t₀, just before the irradiation began, so taking into account the GLY losses 593 due to sorption (approximately 42% GLY was sorbed at t₀ (section 3.1) and could not be 594 595 measured in the dissolved fraction). The dissipation of free GLY was more than 75% after 10 minutes of irradiation, corresponding to a fluence of 3698 J m⁻³ of treated groundwater (eq. 596 597 (5)).

This fast and efficient dissipation of free GLY at low concentration (< 10 μ g L⁻¹) could not be 598 599 fully explained by direct photolysis, but mainly by reactions with ROS such as hydroxyl or 600 superoxide radicals, for instance, and also possibly by the presence of trace of metals and humic acids (HA) in groundwater. Groundwater may contain a small fraction of soluble 601 organic colloids (DOM) composed of HA complexed to iron, manganese or aluminum, 602 coming from soil lixiviation. The various aromatic compounds of HA, as polyphenols or 603 604 organic acids, can form charge transfer complexes with some herbicides, further sensitising them for the photodegradation [51]. Also, it is known that inorganic Fe or Mn elements can 605 act as catalysts for GLY degradation by oxidation in photodegradation or electro-Fenton like 606 processes [52, 53]. In particular, HA and Fe³⁺ complexes can sensitize aquatic 607 photodegradation of GLY, with HA interacting with Fe³⁺ to produce more hydroxyl radicals, 608 609 causing the C-N and C-P bonds cleavage to form phosphate [53]. It should be noted here that AMPA apparition could not be measured during the exposure of GLY to UV-C. 610

It must be underlined that the results obtained for GLY removal with the SurePure device are 611 612 very encouraging. According to the review from Diez et al. (2019), there was just few publications on agrochemicals degradation by UV-C light at the pilot scale without catalyst 613 [23]. The cited authors reached a degradation of 30% in 100 minutes for a pesticide and 5% in 614 100 minutes for a fungicide for 19 L of treated water, demonstrating that 75% GLY removal 615 616 after 10 min irradiation, obtained in this work, is promising. Moreover, it is important to note 617 that these cited works were investigated in pure water and not in groundwater (or wastewater) like in this study. At last, experiments on GLY using UV-C irradiation at the pilot scale are 618 even more rare and Jönsson et al. mentioned that the removal of GLY, spiked in drinking 619 water at 3 µg L⁻¹, was not exceeding 36% after 7 days of treatment with UV-C irradiation in a 620 flow through system [54]. 621

For wastewater, the performance of photolysis was also interesting with more than 50% of the 622 623 initial concentration of free GLY removed from the water after 10 minutes exposure and a fluence of 3698 kJ m⁻³ (Fig. 4). However, after 10 minutes, the concentration of soluble GLY 624 increased in the pilot, indicating that a part of GLY was again released in the aqueous phase. 625 Compared with groundwater, an important part of GLY was sorbed to the pilot surfaces 626 (stainless steel and quartz tubes) as well as to the high amount of SPM. Approximately 80% 627 628 GLY was sorbed to all the solid surfaces (known initial spiked amounts were compared with those measured after equilibrium of wastewater into the pilot and the water collected at t_o, just 629 before the irradiation began). It has been reported that GLY and AMPA were more frequently 630 found associated with SPM than in the soluble phase despite their high solubility, with 631 partition coefficients in the range of 81-7564 L kg⁻¹ which is significantly higher than those of 632 ATR and MAL (Table 1) [9, 31, 55]. Therefore, in the case of the turbid wastewater, a 633 photodegradation of the soluble GLY fraction may occur rapidly, although SPM limit the 634 interaction with photons and the generation of hydroxyl radicals with the aid of HA from 635

DOM. However, particulate matter tended to release sorbed GLY, possibly at each water cycle and because the solid/liquid partition was continuously modified in this dynamic system. Unfortunately, GLY disappearance was lower than GLY release. It means that UV-C irradiation must be significantly longer than 30 min to eliminate all the dissolved GLY molecules from the turbid water.

641

642 Conclusions

This study investigated the degradation of ATR, MAL and GLY pesticides by direct UV-C 643 photolysis in order to develop a circular economic model to clean water used in agriculture. 644 645 The direct photolysis, applied in a continuous flow-through system which could treat 40 L water, was investigated on natural spiked ground- and wastewater. For diluted (0.5 μ g L⁻¹) 646 647 and more concentrated (10 μ g L⁻¹) solutions, ATR was completely eliminated after 15 minutes from groundwater while 80% was removed from the more turbid wastewater after 25 648 minutes treatment. The GC-MS analysis demonstrated that ATR was not quantitatively 649 converted to DIA or DIE and that trace levels of these transformation products were not 650 eliminated by UV-C treatment in both water samples. For MAL, 70 to 80% (as a function of 651 the initial concentration) was removed in 25 minutes from the groundwater by photolytic 652 oxidation. For wastewater, the initial concentration of MAL was important on the 653 performance of the photolytic process. The dissipation of GLY at concentrations (10 μ g L⁻¹) 654 higher than those found in natural groundwater was promising because it was quantitatively 655 eliminated after 10 minutes of irradiation (>75%), possibly with the aid of HA. In wastewater 656 containing high amount of SPM, the UV-C treatment was less efficient because GLY was 657 mainly adsorbed to particles which obstruct the photodegradation process. It seems that only 658 dissolved GLY could be photodegraded at low concentration and the continuous release of 659 GLY from particles in the dynamic UV-DS system was an obstacle to its rapid quantitative 660 elimination. This study therefore, demonstrated that the UV-C treatment applied to large 661

volumes of water, using a turbulent flow and the multiple-lamp system, was effective to
rapidly and quantitatively remove some herbicides and pesticides by direct photolytic
oxidation, without added catalysts, when the turbidity of the treated water is limited.

- As transformation products can sometimes be more toxic than parent pesticides, the ability of
- the UV-C treatment to remove quantitatively problematic pesticides in water will need, in the
- 667 future, to be supported by toxicity assays and researches on potential ecological effects.
- 668
- 669

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674 **References**

- 675 [1] E. Villanneau, N.P.A. Saby, D. Arrouays, C.C. Jolivet, L. Boulonne, G. Caria, E. Barriuso, A. Bispo, O.
- Briand, Spatial distribution of lindane in topsoil of Northern France, Chemosphere, 77 (2009) 1249-
- 677 1255. https://doi.org/10.1016/j.chemosphere.2009.08.060
- [2] V.C. Schreiner, E. Szöcs, A.K. Bhowmik, M.G. Vijver, R.B. Schäfer, Pesticide mixtures in streams of
 several European countries and the USA, Science of The Total Environment, 573 (2016) 680-689.
 https://doi.org/10.1016/j.scitotenv.2016.08.163
- [3] F.J. Benitez, F.J. Real, J.L. Acero, C. Garcia, Photochemical oxidation processes for the elimination
 of phenyl-urea herbicides in waters, Journal of hazardous materials, 138 (2006) 278-287.
 https://doi.org/10.1016/j.jhazmat.2006.05.077
- [4] M.P. Silva, A.P. dos Santos Batista, S.I. Borrely, V.H.O. Silva, A.C.S.C. Teixeira, Photolysis of atrazine
 in aqueous solution: role of process variables and reactive oxygen species, Environmental Science
 and Pollution Research, 21 (2014) 12135-12142. https://doi.org/10.1007/s11356-014-2881-0
- [5] N. Chen, D. Valdes, C. Marlin, P. Ribstein, F. Alliot, E. Aubry, H. Blanchoud, Transfer and
 degradation of the common pesticide atrazine through the unsaturated zone of the Chalk aquifer
 (Northern France), Environmental Pollution, 255 (2019) 113125.
 https://doi.org/10.1016/j.envpol.2019.113125
- [6] L. Carles, H. Gardon, L. Joseph, J. Sanchís, M. Farré, J. Artigas, Meta-analysis of glyphosate
 contamination in surface waters and dissipation by biofilms, Environment International, 124 (2019)
 284-293. https://doi.org/10.1016/j.envint.2018.12.064
- [7] J. Dollinger, C. Dagès, M. Voltz, Glyphosate sorption to soils and sediments predicted by
 pedotransfer functions, Environmental Chemistry Letters, 13 (2015) 293-307.
 https://doi.org/10.1007/s10311-015-0515-5
- [8] L. Velkoska-Markovska, B. Petanovska-Ilievska, A. Markovski, Application of High Performance
 Liquid Chromatography to the Analysis of Pesticide Residues in Apple Juice, Contemporary
 Agriculture, 67 (2018) 93-102. doi:10.2478/contagri-2018-0014
 - 34

- [9] R.L. Glass, Adsorption of glyphosate by soils and clay minerals, Journal of Agricultural and Food
 Chemistry, 35 (1987) 497-500. https://doi.org/10.1021/jf00076a013
- [10] G.-M. Momplaisir, C.G. Rosal, E.M. Heithmar, K.E. Varner, L.A. Riddick, D.F. Bradford, N.G.
 Tallent-Halsell, Development of a solid phase extraction method for agricultural pesticides in large-
- volume water samples, Talanta, 81 (2010) 1380-1386. https://doi.org/10.1016/j.talanta.2010.02.038

705 [11] S. Barrek, C. Cren-Olivé, L. Wiest, R. Baudot, C. Arnaudguilhem, M.-F. Grenier-Loustalot, Multi-

residue analysis and ultra-trace quantification of 36 priority substances from the European Water Framework Directive by GC–MS and LC-FLD-MS/MS in surface waters, Talanta, 79 (2009) 712-722.

708 https://doi.org/10.1016/j.talanta.2009.04.058

- R. Carabias-Martínez, E. Rodríguez-Gonzalo, E. Herrero-Hernández, F.J. Sánchez-San Román,
 M.G. Prado Flores, Determination of herbicides and metabolites by solid-phase extraction and liquid
 chromatography: Evaluation of pollution due to herbicides in surface and groundwaters, Journal of
 Chromatography: A 050 (2002) 457 455, https://doi.org/10.1016/00024.0573(01)01512.2
- 712 Chromatography A, 950 (2002) 157-166. https://doi.org/10.1016/S0021-9673(01)01613-2
- [13] C. Hao, D. Morse, F. Morra, X. Zhao, P. Yang, B. Nunn, Direct aqueous determination of
 glyphosate and related compounds by liquid chromatography/tandem mass spectrometry using
 reversed-phase and weak anion-exchange mixed-mode column, Journal of Chromatography A, 1218
 (2011) 5638-5643. https://doi.org/10.1016/j.chroma.2011.06.070
- [14] B. Le Bot, K. Colliaux, D. Pelle, C. Briens, R. Seux, M. Clément, Optimization and performance
 evaluation of the analysis of glyphosate and AMPA in water by HPLC with fluorescence detection,
 Chromatographia, 56 (2002) 161-164. https://doi.org/10.1007/BF02493205
- T. Arkan, A. Csámpai, I. Molnár-Perl, Alkylsilyl derivatization of glyphosate and
 aminomethylphosphonic acid followed by gas chromatography mass spectrometry, Microchemical
 Journal, 125 (2016) 219-223. https://doi.org/10.1016/j.microc.2015.11.027
- [16] J. Jiang, C.A. Lucy, Determination of glyphosate using off-line ion exchange preconcentration and
 capillary electrophoresis-laser induced fluorescence detection, Talanta, 72 (2007) 113-118.
 https://doi.org/10.1016/j.talanta.2006.10.001
- 726 [17] C.E. Ramirez, S. Bellmund, P.R. Gardinali, A simple method for routine monitoring of glyphosate 727 and its main metabolite in surface waters using lyophilization and LC-FLD+MS/MS. Case study: canals
- with influence on Biscayne National Park, Science of The Total Environment, 496 (2014) 389-401.
- 729 https://doi.org/10.1016/j.scitotenv.2014.06.118
- 730 [18] C. Hidalgo, C. Rios, M. Hidalgo, V. Salvadó, J.V. Sancho, F. Hernández, Improved coupled-column
- liquid chromatographic method for the determination of glyphosate and aminomethylphosphonic
 acid residues in environmental waters, Journal of Chromatography A, 1035 (2004) 153-157.
- 733 https://doi.org/10.1016/j.chroma.2004.02.044
- [19] J. Patsias, A. Papadopoulou, E. Papadopoulou-Mourkidou, Automated trace level determination
 of glyphosate and aminomethyl phosphonic acid in water by on-line anion-exchange solid-phase
 extraction followed by cation-exchange liquid chromatography and post-column derivatization,
 Journal of Chromatography A, 932 (2001) 83-90. https://doi.org/10.1016/S0021-9673(01)01253-5
- [20] A. Tufail, W.E. Price, M. Mohseni, B.K. Pramanik, F.I. Hai, A critical review of advanced oxidation
 processes for emerging trace organic contaminant degradation: Mechanisms, factors, degradation
 products, and effluent toxicity, Journal of Water Process Engineering, (2020) 101778.
 https://doi.org/10.1016/j.jwpe.2020.101778
- [21] J.-L. Shie, C.-H. Lee, C.-S. Chiou, C.-T. Chang, C.-C. Chang, C.-Y. Chang, Photodegradation kinetics
 of formaldehyde using light sources of UVA, UVC and UVLED in the presence of composed silver
 titanium oxide photocatalyst, Journal of Hazardous Materials, 155 (2008) 164-172.
 https://doi.org/10.1016/j.jhazmat.2007.11.043
- 746 [22] C. Bertoldi, A.G. Rodrigues, A.N. Fernandes, Removal of endocrine disrupters in water under 747 artificial light: the effect of organic matter, Journal of Water Process Engineering, 27 (2019) 126-133.
- 748 https://doi.org/10.1016/j.jwpe.2018.11.016
- [23] A.M. Díez, M.A. Sanromán, M. Pazos, New approaches on the agrochemicals degradation by UV
 oxidation processes, Chemical Engineering Journal, 376 (2019) 120026.
 https://doi.org/10.1016/j.cej.2018.09.187

752 [24] R.O. Ramos, M.V.C. Albuquerque, W.S. Lopes, J.T. Sousa, V.D. Leite, Degradation of indigo 753 carmine by photo-Fenton, Fenton, H2O2/UV-C and direct UV-C: Comparison of pathways, products 754 and kinetics, Journal of Water Process Engineering, 37 (2020) 101535. 755 https://doi.org/10.1016/j.jwpe.2020.101535

756 [25] E. Blázquez, C. Rodríguez, J. Ródenas, N. Navarro, C. Riquelme, R. Rosell, J. Campbell, J. 757 Crenshaw, J. Segalés, J. Pujols, J. Polo, Evaluation of the effectiveness of the SurePure Turbulator

viltraviolet-C irradiation equipment on inactivation of different enveloped and non-enveloped viruses
inoculated in commercially collected liquid animal plasma, PLOS ONE, 14 (2019) e0212332.
10.1371/journal.pone.0212332

- [26] N. Bustos, A. Cruz-Alcalde, A. Iriel, A. Fernández Cirelli, C. Sans, Sunlight and UVC-254 irradiation
 induced photodegradation of organophosphorus pesticide dichlorvos in aqueous matrices, Sci Total
 Environ, 649 (2019) 592-600. https://doi.org/10.1016/j.scitotenv.2018.08.254
- [27] M. Bavcon Kralj, U. Černigoj, M. Franko, P. Trebše, Comparison of photocatalysis and photolysis
 of malathion, isomalathion, malaoxon, and commercial malathion—Products and toxicity studies,
 Water Research, 41 (2007) 4504-4514. https://doi.org/10.1016/j.watres.2007.06.016
- 767 [28] P. Somathilake, J.A. Dominic, G. Achari, C.H. Langford, J.-H. Tay, Use of low pressure mercury
 768 lamps with H2O2 and TiO2 for treating carbamazepine in drinking water: Batch and continuous flow
 769 through experiments, Journal of Water Process Engineering, 26 (2018) 230-236.
 770 https://doi.org/10.1016/j.jwpe.2018.10.015
- [29] R. Ahmad, Z. Ahmad, A.U. Khan, N.R. Mastoi, M. Aslam, J. Kim, Photocatalytic systems as an
 advanced environmental remediation: Recent developments, limitations and new avenues for
 applications, Journal of Environmental Chemical Engineering, 4 (2016) 4143-4164.
 https://doi.org/10.1016/j.jece.2016.09.009
- [30] F. Alberini, M.J.H. Simmons, D.J. Parker, T. Koutchma, Validation of hydrodynamic and microbial
 inactivation models for UV-C treatment of milk in a swirl-tube 'SurePure Turbulator™', Journal of
 Food Engineering, 162 (2015) 63-69. https://doi.org/10.1016/j.jfoodeng.2015.04.009
- 778 [31] T.M. Mac Loughlin, M.L. Peluso, V.C. Aparicio, D.J.G. Marino, Contribution of soluble and 779 particulate-matter fractions to the total glyphosate and AMPA load in water bodies associated with 780 horticulture, Science of The Total Environment, 703 (2020) 134717. https://doi.org/10.1016/j.scitotenv.2019.134717 781
- [32] I. Hanke, H. Singer, J. Hollender, Ultratrace-level determination of glyphosate,
 aminomethylphosphonic acid and glufosinate in natural waters by solid-phase extraction followed by
 liquid chromatography-tandem mass spectrometry: performance tuning of derivatization,
 enrichment and detection, Analytical and Bioanalytical Chemistry, 391 (2008) 2265-2276.
 https://doi.org/10.1007/s00216-008-2134-5
- [33] S. Wang, B. Liu, D. Yuan, J. Ma, A simple method for the determination of glyphosate and
 aminomethylphosphonic acid in seawater matrix with high performance liquid chromatography and
 fluorescence detection, Talanta, 161 (2016) 700-706. https://doi.org/10.1016/j.talanta.2016.09.023
- [34] T.V. Nedelkoska, G.K.C. Low, High-performance liquid chromatographic determination of
 glyphosate in water and plant material after pre-column derivatisation with 9-fluorenylmethyl
 chloroformate, Analytica Chimica Acta, 511 (2004) 145-153.
 https://doi.org/10.1016/j.aca.2004.01.027
- [35] R.J. Vreeken, P. Speksnijder, I. Bobeldijk-Pastorova, T.H.M. Noij, Selective analysis of the
 herbicides glyphosate and aminomethylphosphonic acid in water by on-line solid-phase extraction–
 high-performance liquid chromatography–electrospray ionization mass spectrometry, Journal of
 Chromatography A, 794 (1998) 187-199. https://doi.org/10.1016/S0021-9673(97)01129-1
- [36] W.C. Koskinen, L.J. Marek, K.E. Hall, Analysis of glyphosate and aminomethylphosphonic acid in
 water, plant materials and soil, Pest Management Science, 72 (2016) 423-432.
 https://doi.org/10.1002/ps.4172
- [37] Y.S. Hu, Y.Q. Zhao, B. Sorohan, Removal of glyphosate from aqueous environment by adsorption
 using water industrial residual, Desalination, 271 (2011) 150-156.
 https://doi.org/10.1016/j.desal.2010.12.014

- [38] I. Freuze, A. Jadas-Hecart, A. Royer, P.-Y. Communal, Influence of complexation phenomena with
 multivalent cations on the analysis of glyphosate and aminomethyl phosphonic acid in water, Journal
 of Chromatography A, 1175 (2007) 197-206. https://doi.org/10.1016/j.chroma.2007.10.092
- 807 [39] M.P. Silva, A.P. dos Santos Batista, S.I. Borrely, V.H.O. Silva, A.C.S.C. Teixeira, Photolysis of 808 atrazine in aqueous solution: role of process variables and reactive oxygen species.
- [40] G.V. Buxton, C.L. Greenstock, W.P. Helman, A.B. Ross, Critical Review of rate constants for
 reactions of hydrated electrons, hydrogen atoms and hydroxyl radicals (·OH/·O– in Aqueous Solution,
 Journal of Physical and Chemical Reference Data, 17 (1988) 513-886. 10.1063/1.555805
- [41] H. Prosen, L. Zupančič-Kralj, Evaluation of photolysis and hydrolysis of atrazine and its first
 degradation products in the presence of humic acids, Environmental Pollution, 133 (2005) 517-529.
 https://doi.org/10.1016/j.envpol.2004.06.015
- [42] A.J. Moreira, A.C. Borges, L.F.C. Gouvea, T.C.O. MacLeod, G.P.G. Freschi, The process of atrazine
 degradation, its mechanism, and the formation of metabolites using UV and UV/MW photolysis,
 Journal of Photochemistry and Photobiology A: Chemistry, 347 (2017) 160-167.
 https://doi.org/10.1016/j.jphotochem.2017.07.022
- 819 [43] D.W. Kolpin, E.M. Thurman, S.M. Linhart, Finding minimal herbicide concentrations in ground
- water? Try looking for their degradates, Science of The Total Environment, 248 (2000) 115-122.
 https://doi.org/10.1016/S0048-9697(99)00535-5
- [44] J.A. Khan, X. He, N.S. Shah, M. Sayed, H.M. Khan, D.D. Dionysiou, Degradation kinetics and
 mechanism of desethyl-atrazine and desisopropyl-atrazine in water with OH and SO4– based-AOPs,
 Chemical Engineering Journal, 325 (2017) 485-494. https://doi.org/10.1016/j.cej.2017.05.011
- [45] W. Li, Y. Zhao, X. Yan, J. Duan, C.P. Saint, S. Beecham, Transformation pathway and toxicity
 assessment of malathion in aqueous solution during UV photolysis and photocatalysis, Chemosphere,
 234 (2019) 204-214. https://doi.org/10.1016/j.chemosphere.2019.06.058
- [46] S. Navarro, J. Fenoll, N. Vela, E. Ruiz, G. Navarro, Removal of ten pesticides from leaching water
 at pilot plant scale by photo-Fenton treatment, Chemical Engineering Journal, 167 (2011) 42-49.
 https://doi.org/10.1016/j.cej.2010.11.105
- 831 [47] L. Varanasi, E. Coscarelli, M. Khaksari, L.R. Mazzoleni, D. Minakata.
- [48] A. Manassero, C. Passalia, A.C. Negro, A.E. Cassano, C.S. Zalazar, Glyphosate degradation in
 water employing the H2O2/UVC process, Water Research, 44 (2010) 3875-3882.
 https://doi.org/10.1016/j.watres.2010.05.004
- [49] J.S. McConnell, R.M. McConnell, Ultraviolet Spectra of Acetic Acid, Glycine, and Glyphosate,
 Journal of the Arkansas Academy of Science, 47 (1993) 73-76.
- [50] D. Feng, A. Soric, O. Boutin, Treatment technologies and degradation pathways of glyphosate: A
 critical review, Science of The Total Environment, 742 (2020) 140559.
 https://doi.org/10.1016/j.scitotenv.2020.140559
- K. Lányi, P. Laczay, J. Lehel, Effects of some naturally occurring substances on the
 photodegradation of herbicide methabenzthiazuron, Journal of Environmental Chemical Engineering,
 4 (2016) 123-129. https://doi.org/10.1016/j.jece.2015.11.002
- 4 (2010) 123-123. https://doi.org/10.1010/j.jece.2013.11.002
 843 [52] B. Balci, M.A. Oturan, N. Oturan, I. Sirés, Decontamination of Aqueous Glyphosate,
 844 (Aminomethyl)phosphonic Acid, and Glufosinate Solutions by Electro-Fenton-like Process with Mn2+
 845 as the Catalyst, Journal of Agricultural and Food Chemistry, 57 (2009) 4888-4894.
- 846 https://doi.org/10.1021/jf900876x
- [53] J. Liu, J. Fan, T. He, X. Xu, Y. Ai, H. Tang, H. Gu, T. Lu, Y. Liu, G. Liu, The mechanism of aquatic
 photodegradation of organophosphorus sensitized by humic acid-Fe3+ complexes, Journal of
 Hazardous Materials, 384 (2020) 121466. https://doi.org/10.1016/j.jhazmat.2019.121466
- [54] J. Jönsson, R. Camm, T. Hall, Removal and degradation of glyphosate in water treatment: a
 review, Journal of Water Supply: Research and Technology-Aqua, 62 (2013) 395-408.
 10.2166/aqua.2013.080
- 853 [55] R. Kanissery, B. Gairhe, D. Kadyampakeni, O. Batuman, F. Alferez, Glyphosate: Its Environmental 854 Persistence and Impact on Crop Health and Nutrition, Plants, 8 (2019). 855 https://doi.org/10.3390/plants8110499

