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## RESEARCH ARTICLE

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## Environmental Controls of Size Distribution of Modern Planktonic Foraminifera in the Tropical Indian Ocean

### Key Points:

- Optimum size-hypothesis holds true in planktonic foraminifera if one considers the main parameters driving each species' size distribution
- Size variations in planktonic foraminifera are linked to species' niches and diversity does not increase with productivity
- Within-species size is driven by  $\text{CO}_3^{2-}$  concentration, temperature, and salinity; assemblage size by  $\text{CO}_3^{2-}$  concentration and temperature

### Supporting Information:

Supporting Information may be found in the online version of this article.

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**Abstract** Paleooceanographic studies often rely on abundance changes in microfossil species, with little consideration for characteristics such as organism size, which may also be related to environmental changes. Using a tropical Indian Ocean (TIO) core-top data set, we test the Optimum size-hypothesis (OSH), investigating whether relative abundance or environmental variables are better descriptors of planktonic foraminifera species' optimum conditions. We also investigate the environmental drivers of whole-assemblage planktonic foraminiferal test size variation in the TIO. We use an automated imaging and sorting system (MiSo) to identify planktonic foraminiferal species, analyze their morphology, and quantify fragmentation rate using machine learning techniques. Machine model accuracy is confirmed by comparison with human classifiers (97% accuracy). Data for 33 environmental parameters were extracted from modern databases and, through exploratory factor analysis and regression models, we explore relationships between planktonic foraminiferal size and oceanographic parameters in the TIO. Results show that the size frequency distribution of most planktonic foraminifera species is unimodal, with some larger species showing multimodal distributions. Assemblage size<sub>95/5</sub> (95th percentile size) increases with increasing species diversity, and this is attributed to vertical niche separation induced by thermal stratification. Our test for the OSH reveals that relative abundance is not a good predictor of species' optima and within-species size<sub>95/5</sub> response to environmental parameters is species-specific, with parameters related to carbonate ion concentration, temperature, and salinity being primary drivers. At the species and assemblage levels, our analyses indicate that carbonate ion concentration and temperature play important roles in determining size trends in TIO planktonic foraminifera.

**Plain Language Summary** In core-top samples from the tropical Indian Ocean (TIO), we investigate the optimum size-hypothesis, testing whether species' relative abundance or environmental parameter(s) are better descriptors of planktonic foraminifera species' optimum conditions. Further, we investigate the main environmental drivers of size variations in planktonic foraminifera at the assemblage-level, given that temperature has been reported to primarily drive assemblage size trends. We use a state-of-the-art machine (MiSo) to automatically identify planktonic foraminiferal species, analyze their size, and quantify fragmentation using machine learning techniques. When compared to identification carried out by human experts across 21 species, the machine classified the species accurately 97% of the time. The MiSo-generated size data was similar to that by other researchers. The frequency distributions of the species' size spectra show that most species have distributions that form bell-shaped curves. As species diversity increased, so did the assemblage size (95th percentile size); we attribute this observation to the effect of temperature-dependent niche separation. We find that, in the TIO, environmental parameters are better descriptors of optimum conditions in planktonic foraminifera than relative abundance. Our results also reveal that size variation at the species and assemblage levels is mostly driven by ambient carbonate chemistry and temperature.

## 1. Introduction

Increasing anthropogenic emissions of carbon dioxide ( $\text{CO}_2$ ) and other greenhouse gases are drastically affecting the physico-chemical characteristics of the ocean, resulting in higher temperatures, lower pH, stronger stratification, and reduced biological productivity (Bindoff et al., 2019; Caldeira & Wickett, 2003; Gao et al., 2012; Stips et al., 2016; Zeebe & Wolf-Gladrow, 2001). Over the last century, the surface ocean has warmed by on average  $0.9^\circ\text{C}$  while upper-ocean stratification has increased by  $4.9\% \pm 1.5\%$  between 1970 and 2018 (Fox-Kemper

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et al., 2021). Under more stratified conditions, upper ocean vertical mixing decreases, leading to reduced nutrient inputs and primary productivity (PP) (Gruber, 2011; He & Mahadevan, 2021). These surface ocean changes impact calcifying marine plankton, including planktonic foraminifera, which are major contributors to the global carbonate pump driven by the production of  $\text{CaCO}_3$  in surface waters and its subsequent export to deep waters and, ultimately, sediments (Berner & Raiswell, 1983; Fabry et al., 2008; Henehan et al., 2017; Manno et al., 2012; Moy et al., 2009; Orr et al., 2005; Schiebel, 2002). Planktonic foraminifera are unicellular marine protozoans that precipitate calcite tests, accounting for ~25% of the total calcite exported to the deep ocean (Buitenhuis et al., 2019; Schiebel, 2002; Schiebel et al., 2007). Their growth, calcification, and reproduction processes are directly linked to ambient environmental conditions (Schiebel & Hemleben, 2017; Weinkauff et al., 2013, 2022). Fossil planktonic foraminiferal assemblages and the geochemical signatures of tests are routinely used to reconstruct past upper ocean conditions, including temperature and productivity (e.g., Cayre et al., 1999; Kucera, 2007; Kucera et al., 2005; Tachikawa et al., 2008). Another key variable that we can extract from the planktonic foraminiferal fossil record is test size, yet comparatively little is known about how it varies in response to the environment, despite existing studies showing that test size distribution can provide valuable insight into present and past oceanographic conditions (Moller et al., 2013; Schmidt, Renaud, et al., 2004).

The size of an organism is a functional trait that scales with ecological parameters such as population growth rate, competition, and reproduction (Arendt, 2007; Millien et al., 2006), and influences its metabolic rate, its ability to coexist with other organisms, and its response to changes in ambient conditions (Brown et al., 2007; Hart et al., 2016; Violle et al., 2012). Different rules have been proposed to explain the relationship between an organism's size and temperature. First, Bergmann's rule, originally proposed for endotherms in the context of heat conservation, states that larger organisms tend to inhabit colder environments (Bergmann, 1847). This rule has since been recast for groups including marine crustaceans, birds, and some ectotherms (Ashton & Feldman, 2003; Fan et al., 2019; Ollala-Tàrraga et al., 2006). Second, the temperature–size rule states that the body size of ectothermic organisms (including protozoans, copepods, and fish) generally decreases with increasing temperature (Atkinson, 1994). Whilst neither of these rules have been applied specifically to planktonic foraminifera, a positive temperature–size relationship on global biogeographical and evolutionary scales has been documented (Schmidt, Renaud, et al., 2004; Schmidt, Thierstein, et al., 2004); that is, modern planktonic foraminifera are larger in warm tropical waters than in colder high latitude waters, and Paleocene (greenhouse climate) planktonic foraminifera are larger than Pleistocene (icehouse climate) planktonic foraminifera.

Another theory suggests that size variations in planktonic foraminifera follow the optimum size-hypothesis (OSH), which states that populations are largest at their ecological optimum (Hecht, 1976). Testing the OSH by examining the relationship between a species' size and a measure of its optimum conditions is challenging, and multiple factors can contribute to the diversity of results obtained (e.g., Beer et al., 2010; Hecht, 1976; Kahn, 1981; Kennett, 1976; Malmgren & Kennett, 1976, 1977; Rillo et al., 2020; Schmidt, Renaud, et al., 2004; Schmidt, Thierstein, et al., 2004; Weinkauff et al., 2016; Zarkogiannis et al., 2020). These factors include differences in the population unit investigated (single or multiple populations), in the methodology or statistics adopted, in the choice of metric used to define size in a population (e.g., mean, maximum,  $\text{size}_{95/5}$  (95th percentile size: the size separating the top 5% from the remaining 95% size within a population or assemblage), shell weight, size normalized weight), and lastly how optimum growth conditions are defined (maximum absolute or relative abundance, environmental preferences). In addition, the range of environmental gradients covered within the samples of a study may also influence the outcome with respect to the OSH (Kontakiotis et al., 2021; Weinkauff et al., 2016). Aside from the global study of Schmidt, Renaud, et al. (2004), most studies that found support for the OSH investigated single or very few species, and defined optimum conditions based on species' absolute or relative abundance (Hecht, 1976; Malmgren & Kennett, 1976; Moller et al., 2013). Schmidt, Renaud, et al. (2004) investigated the OSH using the correlation between temperature at maximum  $\text{size}_{95/5}$  and temperature at maximum relative abundance, under the assumption that convergence between the two temperatures indicated an ecological optimum. While this is plausible, this approach uses only a small proportion of the available data, and because different species have different thermal niches, a 1:1 temperature relationship as observed in their multispecies study will still occur even if there is no relationship between size and abundance in individual species (Rillo et al., 2020). In Weinkauff et al. (2016) and Rillo et al. (2020), correlations between size (mean size and  $\text{size}_{95/5}$ ) and environmental parameters such as sea surface temperature (SST), net PP, and chlorophyll a showed no support for the OSH. Therefore, there is a need to evaluate whether optimum conditions in planktonic foraminifera are better inferred from species' relative abundances or from their preferred environmental conditions using robust statistical methods.

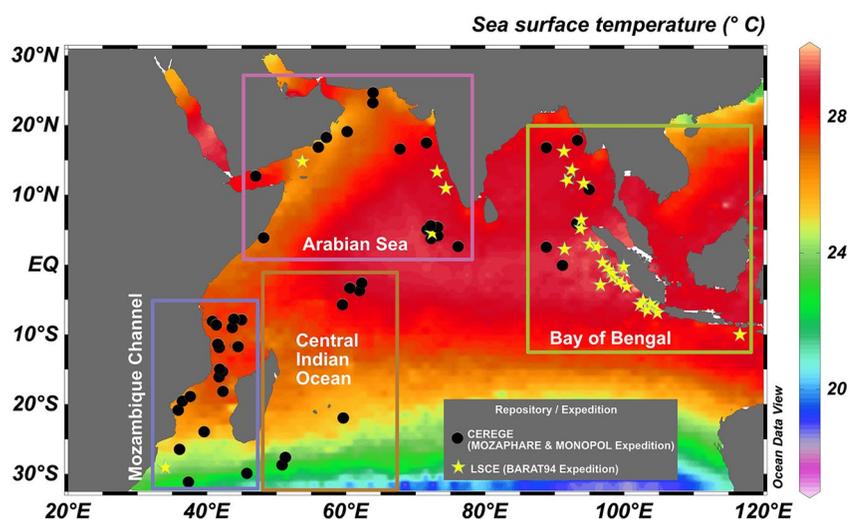
Within the context of planktonic foraminifera size response to abiotic factors, it can be difficult to define which parameter serves as the good measure of optimum conditions for each species, since different species have varying ecological preferences (Schiebel & Hemleben, 2017). Whilst SST has been implicated as a primary driver of size variation in planktonic foraminifera (Schmidt, Renaud, et al., 2004), laboratory experiments have shown that size variations can also be linked to salinity, carbonate saturation, nutrients, and oxygen availability (Bé et al., 1981; Bijma et al., 1990; Caron et al., 1982; Hemleben et al., 1987; Kuroyanagi et al., 2013), making the SST–size relationship difficult to discern at the species level (Rillo et al., 2020). The lack of consensus on which physicochemical parameter(s) exert the strongest controls on planktonic foraminiferal size may in part result from: (a) differential size responses to ambient conditions at the assemblage level (Schmidt, Renaud, et al., 2004) and the species level (Colombo & Cita, 1980; Hecht, 1974; Moller et al., 2013; Rillo et al., 2020) due to species-specific ecological adaptation and resource requirements; (b) investigation of few parameters ( $n < 8$ ) rather than a broader range of variables that more closely represent species' living conditions (e.g., Rillo et al., 2020); (c) methodological differences arising from the use of different size metrics such as diameter, area, aspect ratio, and roundness (Brombacher et al., 2017); (d) differences in sample material, that is, whether data from modern samples (laboratory culture experiments, plankton tow, or sediment trap) or fossil materials (core-tops and down-core sediment samples) are used; and (e) local or regional effects (Moller et al., 2013). This lack of consensus warrants further study to understand which environmental variable(s) exert major controls on planktonic foraminiferal size distribution at the species and assemblage levels.

In this study, we quantified planktonic foraminiferal species compositions and analyzed their size variations in a suite of 82 core-top samples from the tropical Indian Ocean (TIO). We use state-of-the-art automated morphometric and identification methods to collect a large and robust data set, and our new high-throughput workflow based on machine-learning techniques is validated by comparison with human counts. Using statistical modeling, we test the OSH, investigating whether species' optimum conditions are best inferred from their relative abundances or from the environmental parameters that mostly drive variations in their sizes (33 parameters are tested). We investigated the environmental drivers of size variations in TIO planktonic foraminifera at the species and assemblage levels. We focus on the TIO because a significant amount of data on planktonic foraminifera distribution in this region already exists, and because this region contains highly contrasted oceanic environments in terms of physico-chemical characteristics (oxygenation, stratification, PP, carbonate chemistry, salinity). In the TIO region encompassing our samples, mean annual sea surface temperatures range from 19 to 30°C and average mixed layer (0–200 m) temperatures range from 12 to 22°C (Locarnini et al., 2018), thus the influence of temperature on size is unlikely to override the influence of other environmental factors.

## 2. Materials and Methods

### 2.1. Oceanographic Setting

The TIO, with its unique seasonally-reversing monsoons and transient cyclonic storms, is landlocked in the north, and represents an area of intense air–sea interaction (R. R. Rao et al., 1991; Schott et al., 2009). The northern TIO is divided into two parts by the Indian Peninsula: the Arabian Sea and the Bay of Bengal. Due to its proximity to the Persian Gulf and Red Sea, the Arabian Sea has an upper layer of highly saline waters and experiences more evaporation than precipitation (Levitus, 1982; L. V. G. Rao & Ram, 2005). The Bay of Bengal receives high freshwater inputs via river discharge from abundant precipitation during the southwest (summer) monsoon, resulting in a low-salinity upper layer and a stratification dominated by salinity gradients rather than temperature gradients (L. V. G. Rao & Ram, 2005; Wyrтки, 1971). In the Arabian Sea, wind-driven coastal upwelling in the west is strong during the southwest monsoon and vertical mixing is also enhanced during the northeast monsoon, promoting higher productivity (Barber et al., 2001). In the southwestern TIO, the Mozambique Channel is important for water exchanges between the Indian and Atlantic Oceans via the Agulhas Current (Schott et al., 2009; Ternon et al., 2014). During the summer monsoon from April to October, southeasterly winds from Australia induce Ekman pumping in the southeastern TIO, causing upwelling along the coasts of Sumatra and Java (Susanto et al., 2001). Within the TIO (31°N, 31°S, 20–120°E), mean annual SST ranges from 18°C (southern part; July–October) to >30°C (northern part; April–May) (Locarnini et al., 2018; L. V. G. Rao & Ram, 2005). Meanwhile, sea surface salinity ranges from 30 to 36 psu, dissolved oxygen concentration ranges from 189 to 255  $\mu\text{mol kg}^{-1}$ , surface Chl-*a* concentration ranges from 0.1 to 0.5  $\text{mg m}^{-3}$ , and carbonate ion content of surface waters ranges from 200 to 280  $\mu\text{mol kg}^{-1}$ , with an average of >220  $\mu\text{mol kg}^{-1}$  (Broecker & Sutherland, 2000; Garcia et al., 2018a; Lovenduski et al., 2015; L. V. G. Rao & Ram, 2005; Wyrтки, 1971; Zweng et al., 2018).



**Figure 1.** Map showing the location of the core-top samples used in this study as well as the laboratories where they are archived. Underlaid is the zonal sea surface temperature gradient of the tropical Indian Ocean. Shown also are the categories of the regions denoted in this study: Arabian Sea (pink box), Bay of Bengal (yellow box), central Indian Ocean (brown box), and Mozambique Channel (blue box).

## 2.2. Sample Material

We analyzed 82 core-top samples collected in the TIO (Figure 1), stored in core repositories at the Center Européen de Recherche et d'Enseignement de Géosciences de l'Environnement (CEREGE, France) and at the Laboratoire des Sciences du Climat et de l'Environnement (LSCE, France). Samples were taken during expeditions carried out on the RV *Marion Dufresne* and RV *Baruna Jaya I* research vessels. The sample set covers the Arabian Sea, Bay of Bengal (including offshore Java), Mozambique Channel, and central Indian Ocean (31° 30'S, 36° 45'E to 24° 50'N, 60° 36'E) and core-tops are from water depths between 364 and 4,425 m (mean depth 2,346 m). Surface sediment samples (top 2–3 cm) were dried at 50°C then washed over a 150 μm sieve, and the coarse fractions were oven-dried at 50°C. Using a SIO microsplitter, two splits were prepared per sample; one split served as an archive half while the other was used as the working half. The coarse fraction (>150 μm) for the working half of each sample was used for automated imaging on the Microfossil Sorter (MiSo, patent pending) at CEREGE. This automation takes multiple (generally 12) images of single foraminifera tests at several focal depths (vertical resolution of 11 μm). Processed, Z-stacked foraminifera images have a resolution of 1.1594 pixels per micron.

## 2.3. Environmental Data

The environmental data for the ocean overlying each site were retrieved from oceanographic databases using mean annual values from different sources as reported in Table 1. Environmental parameters were selected to capture a wide variety of variables including (but not limited to) parameters that have been suggested to influence planktonic foraminifera size and assemblage distribution: temperature, nutrient concentrations, salinity, oxygen concentrations, carbonate system parameters, and thermal gradients (Cayre et al., 1999; Clemens et al., 1996; Kuroyanagi et al., 2013; Moller et al., 2013; Schmidt, Renaud, et al., 2004). Since the average living depth of most planktonic foraminifera is within the upper 200 m of the water column (Lessa et al., 2020), most of the environmental data used in this work come from 0 to 200 m, so as to capture both surface and sub-surface conditions. The environmental data sets that we used are archived in the French Open Access database, SEANOE, and can be accessed at: <https://doi.org/10.17882/86211> (Adebayo, Bolton, Marchant, Bassinot, Sandrine, et al., 2022). The spatial variability in surface ocean parameters in the TIO was analyzed using a covariance matrix principal component analysis (PCA) to identify the surface hydrographic settings of our study area. The paleontological statistical software Past (version 4.01, Hammer et al., 2001) was used for this analysis.

**Table 1**  
*List of Environmental Parameters Included in the Analysis for This Study*

	Variable name/resolution	Unit	Description	Var_code	Source
1	Sea surface temperature/1°	°C	1955–2017 annual mean temperature at 0 m	Sst	World Ocean Atlas (WOA 18) (Locarnini et al., 2018)
2	Sea temperature/1°	°C	1955–2017 annual mean temperature at 10 m	T10	World Ocean Atlas (WOA 18) (Locarnini et al., 2018)
3	Sea temperature/1°	°C	1955–2017 annual mean at 50 m	T50	World Ocean Atlas (WOA 18) (Locarnini et al., 2018)
4	Sea temperature/1°	°C	1955–2017 annual mean temperature at 100 m	T100	World Ocean Atlas (WOA 18) (Locarnini et al., 2018)
5	Sea temperature/1°	°C	1955–2017 annual mean temperature at 200 m	T200	World Ocean Atlas (WOA 18) (Locarnini et al., 2018)
	Sea temperature/1°	°C	Δ Temperature between 0 and 200 m	ΔT	World Ocean Atlas (WOA 18) (Locarnini et al., 2018)
6	Sea temperature/1°	°C	1955–2017 Summer SST (July–September)	Sumsst	World Ocean Atlas (WOA 18) (Locarnini et al., 2018)
7	Sea Temperature/1°	°C	1955–2017 Winter SST (January–March)	Winsst	World Ocean Atlas (WOA 18) (Locarnini et al., 2018)
8	Sea temperature/1°	°C	Δ Temperature between Winter and Summer Temperatures	Winsum	World Ocean Atlas (WOA 18) (Locarnini et al., 2018)
9	Phosphate/1°	μmol/kg	1955–2017 annual mean phosphate concentration at 0 m	Phos	World Ocean Atlas (WOA 18) (Garcia et al., 2018a)
10	Phosphate/1°	μmol/kg	1955–2017 annual mean phosphate concentration at 200 m	Phos200	World Ocean Atlas (WOA 18) (Garcia et al., 2018a)
11	Phosphate/1°	μmol/kg	Δ Phosphate between 0 and 200 m	ΔPhos	World Ocean Atlas (WOA 18) (Garcia et al., 2018a)
12	Nitrate/1°	(μmol/kg)	1955–2017 annual mean nitrate at 0 surface	Nit	World Ocean Atlas (WOA 18) (Garcia et al., 2018a)
13	Nitrate/1°	μmol/kg	1955–2017 annual mean nitrate at 200 m	Nit200	World Ocean Atlas (WOA 18) (Garcia et al., 2018a)
14	Nitrate/1°	μmol/kg	Δ Nitrate between 0 and 200 m	ΔNit	World Ocean Atlas (WOA 18) (Garcia et al., 2018a)
15	Silicate/1°	μmol/kg	1955–2017 annual mean silicate concentration at 0 m	Sil	World Ocean Atlas (WOA 18) (Garcia et al., 2018a)
16	Salinity/1°		1955–2017 annual mean salinity at 0 m	Sal	World Ocean Atlas (WOA 18) (Zweng et al., 2018)
17	Dissolved oxygen/1°	μmol/kg	1955–2017 annual mean oxygen at 0 m	Ox	World Ocean Atlas (WOA 18) (Garcia et al., 2018b)
18	Dissolved oxygen/1°	μmol/kg	1955–2017 annual mean oxygen at 200 m	Ox200	World Ocean Atlas (WOA 18) (Garcia et al., 2018b)
19	Dissolved oxygen/1°	μmol/kg	Δ oxygen between 0 and 200 m	ΔOx	World Ocean Atlas (WOA 18) (Garcia et al., 2018b)
20	Apparent oxygen saturation/1°	μmol/kg	1955–2017 annual mean apparent oxygen saturation at 0 m	aos	World Ocean Atlas (WOA 18) (Garcia et al., 2018b)
21	Primary productivity/2°	g C/m <sup>2</sup> /yr	Annual Mean Primary Productivity	pp	Ocean Productivity (National Center for Atmospheric Research Staff) (Eds)/ <a href="https://climatedataguide.ucar.edu/climate-data/ocean-productivity-phytoplankton-size-estimates-satellite">https://climatedataguide.ucar.edu/climate-data/ocean-productivity-phytoplankton-size-estimates-satellite</a>

**Table 1**  
*Continued*

	Variable name/resolution	Unit	Description	Var_code	Source
22	Carbonate ion concentration/1°	[CO <sub>3</sub> <sup>2-</sup> ]	Calculated annual mean [CO <sub>3</sub> <sup>2-</sup> ] at 0 m	Scarb	Calculated in CO2SYS using temperature, phosphate, depth, silicate, and salinity data from WOA 18, and alkalinity and carbon dioxide data from Global Ocean Data Analysis (GLODAP) as input data (Key et al., 2015; Olsen et al., 2016; Pierrot et al., 2006)
23	Carbonate ion concentration/1°	[CO <sub>3</sub> <sup>2-</sup> ]	Calculated annual mean [CO <sub>3</sub> <sup>2-</sup> ] at core depth	Carbcd	Same as source for Scarb
24	Carbonate ion concentration/1°	[CO <sub>3</sub> <sup>2-</sup> ]	Calculated annual mean [CO <sub>3</sub> <sup>2-</sup> ] at 200 m	Carb200	Same as source for Scarb
25	Carbonate ion concentration/1°	[CO <sub>3</sub> <sup>2-</sup> ]	Δ [CO <sub>3</sub> <sup>2-</sup> ] between 0 and 200 m	Carb0-200	Same as source for Scarb
26	Carbonate ion concentration/1°	[CO <sub>3</sub> <sup>2-</sup> ]	Calculated annual mean [CO <sub>3</sub> <sup>2-</sup> ] at 500 m	Carb500	Same as source for Scarb
27	Carbonate ion concentration/1°	[CO <sub>3</sub> <sup>2-</sup> ]	Δ [CO <sub>3</sub> <sup>2-</sup> ] between 0 and 200 m	ΔCarb	Same as source for Scarb
28	Carbonate ion concentration/1°	[CO <sub>3</sub> <sup>2-</sup> ]	Δ [CO <sub>3</sub> <sup>2-</sup> ] between 0 and 500 m	Carb0-500	Same as source for Scarb
29	Thermocline depth/2°	m	Defined as the depth above which the temperature is superior to the surface temperature (at 10 m depth) minus 0.2°C	Thermdepth	IFREMER/LOS Mixed Layer Depth Climatology website ( <a href="http://www.ifremer.fr/cerweb/deboyer/mld">www.ifremer.fr/cerweb/deboyer/mld</a> )/Mignot et al. (2007)
30	Pycnocline depth/2°	m	Values obtained for each core location	pycdepth	NOAA( <a href="https://data.noaa.gov/dataset/dataset/mixed-layer-depth-isothermal-layer-depth-barrier-layer-depth-and-other-upper-ocean-thermohaline">https://data.noaa.gov/dataset/dataset/mixed-layer-depth-isothermal-layer-depth-barrier-layer-depth-and-other-upper-ocean-thermohaline</a> )
31	Mixed layer depth/1°	m	Surface mixed layer depth	Mld_ifremer	IFREMER/LOS Mixed Layer Depth Climatology website ( <a href="http://www.ifremer.fr/cerweb/deboyer/mld/surface_mixed_layer_depth">www.ifremer.fr/cerweb/deboyer/mld/surface_mixed_layer_depth</a> )/de Boyer Montégut et al. (2004)
32	Mixed layer depth/1°	m	Surface mixed layer depth	Mld_NOAA	NOAA( <a href="https://data.noaa.gov/dataset/dataset/mixed-layer-depth-isothermal-layer-depth-barrier-layer-depth-and-other-upper-ocean-thermohaline">https://data.noaa.gov/dataset/dataset/mixed-layer-depth-isothermal-layer-depth-barrier-layer-depth-and-other-upper-ocean-thermohaline</a> )
33	Core depth/m	m	Values retrieved from cruise reports	Depth	Cruise Reports

#### 2.4. Automated Imaging, Analysis, and Convolutional Neural Network (CNN) Training

Following Marchant et al. (2020), splits of the >150 μm size fraction were transferred to MiSo and images of each particle (primarily/exclusively whole foraminifera or foraminiferal fragments) were acquired (total  $N = 311,380$ , average  $N$  per sample = 3,797). With the aid of ParticleTrieur, a software developed at CEREGE (Marchant et al., 2020), 20,734 images were then manually labeled. Six convolutional neural networks (“Base Cyclic 16,” “ResNet Cyclic 4,” “ResNet Cyclic 8,” “ResNet50 Cyclic Gain TL,” “ResNet Cyclic TL,” and “VGG19”) with different topologies were then trained by random selection of 80% of the manually labeled images which served

as the training image set, while the remaining 20% served as the validation set that was used to evaluate CNN performance on three metrics:

1. accuracy, expressed as  $\left(\frac{P_{\text{true}} + N_{\text{true}}}{P_{\text{true}} + P_{\text{false}} + N_{\text{true}} + N_{\text{false}}}\right)$ ;
2. recall, expressed as  $\left(\frac{P_{\text{true}}}{P_{\text{true}} + N_{\text{false}}}\right)$ ; and
3. precision expressed as  $\left(\frac{P_{\text{true}}}{P_{\text{true}} + P_{\text{false}}}\right)$ .

In the three above expressions,  $P_{\text{true}}$  = true positives,  $P_{\text{false}}$  = false positives,  $N_{\text{true}}$  = true negatives, and  $N_{\text{false}}$  = false negatives. The best performing CNN was then adopted in the automatic identification of the remaining images. To ascertain the reliability of our model, all 311,380 images were validated manually, and comparison was made between the model classification and that collectively performed by two human classifiers (a graduate student and a senior researcher). Morphometric parameters (area, perimeter, convex area, convex perimeter, major axis, minor axis, minimum enclosing circle area, minimum enclosing circle radius, eccentricity, solidity, roundness, circularity, mean, standard deviation, standard deviation invariant, skewness, kurtosis, fifth moment, and sixth moment) as well as the abundance of each species within each assemblage were extracted from ParticleTrier. For more details on ParticleTrier workflows, see Marchant et al. (2020). The image data sets generated in this study using MiSo are archived in the SEANOE database and can be accessed at: <https://doi.org/10.17882/86411> (Adebayo, Bolton, Marchant, Bassinot, Conrod, et al., 2022).

### 2.5. Size and Relative Abundance Data

We measured individual foraminiferal size based on the test maximum diameter. The test maximum diameter was calculated from the area using the formula:

$$\text{MD} = \left( \sqrt{\frac{\text{Area (mm}^2\text{)}}{\pi}} \right) * 2.$$

We chose to measure test size using the maximum diameter because it is less affected by random object orientation (Schmidt, Renaud, et al., 2004) and has been reported to be a consistent size proxy for two-dimensional images (Brombacher et al., 2018). The 95th percentile size of each assemblage, hereafter referred to as size<sub>95/5</sub>, was selected to investigate the size response of planktonic foraminifera assemblages to environmental variables. We chose size<sub>95/5</sub> because, compared to mean and maximum size, it is less sensitive to single outliers (Rillo et al., 2020; Schmidt, Renaud, et al., 2004). To validate our size spectra, we compared our size<sub>95/5</sub> data with published measurements reported for the TIO from core-top samples (Rillo et al., 2019, 2020). We also compared the relative abundances of the species recorded in this study with those reported for the closest sites to our core locations in the ForCenS Database, which compiles planktonic foraminifera assemblages from core-top sediments (Siccha & Kucera, 2017).

### 2.6. Dissolution

To estimate the degree of dissolution in our samples based on fragmentation intensity, we used the index of Suárez-Ibarra et al. (2021):

$$\text{FI} = \frac{\frac{(fb)}{(fb+pf)}}{\frac{(\text{area})}{(\text{perimeter})}}$$

where *fb* represents the total number of broken tests and fragments, *pf* is the number of whole planktonic foraminifera tests, and area and perimeter are the averages for the broken and fragmented specimens in each sample. Values closer to zero indicate well preserved samples while values closer to one indicate that tests are more affected by dissolution. In addition, we explored other dissolution indicators by investigating the relationship between size<sub>95/5</sub> versus depth, size<sub>95/5</sub> versus carbonate saturation state at core depth ( $\Delta[\text{CO}_3^{2-}]$ ), and size<sub>95/5</sub> versus fragmentation rate.

## 2.7. Statistical Analyses

Before analyzing the planktonic foraminiferal size and assemblage data, we excluded rare species (i.e., species with <50 individuals per sample in all samples). We also excluded samples with low whole specimen counts (<300 whole planktonic foraminifera in a sample); this brought the total number of core-top samples analyzed from 82 to 62. We used the Jarque–Bera normality test (Jarque, 2011) in the R package “tseries” (version 0.10–51) to evaluate whether our size data set showed a normal distribution (based on skewness and kurtosis). This enabled us to decide whether parametric or non-parametric correlation and regression analyses would be preferable to explore relationships between variables. Results of the Jarque–Bera normality test confirmed that the distribution of our size data set is non-parametric ( $X^2 = 478,591$ ,  $p < 2.2e-16$ ); therefore, only non-parametric correlations and regression analyses were adopted going forward.

For comparison purposes, we restricted the Rillo et al. (2020) data set to the TIO (30°S–30°N, 54–112°E) and utilized shell diameter data instead of shell area. Using the Spearman's rank-order correlation (“ $\rho$ ”), we compared our size<sub>95/5</sub> data with that contained in the Rillo et al. (2019) data set. Likewise, we compared the species' relative abundances recorded in this study with those reported for the closest sites to our core locations in the ForCenS Database (REF). Aside from making comparisons based on size measurements, through robust regression analysis, we also assessed the relationship between the size<sub>95/5</sub> of nine planktonic foraminifera species (*Pulleniatina obliquiloculata*, *Globorotalia menardii*, *Neogloboquadrina dutertei*, *Globigerinoides ruber albus*, *Trilobatus sacculifer*, *Globigerinella siphonifera*, *Globigerinoides conglobatus*, *Globorotalia truncatulinoides*, and *Globococconeella inflata*) and mean annual SST as contained in the Rillo et al. (2019) data set for the TIO. We used the “robustbase” package (version 0.95-0) in R for this and subsequent robust regression analyses.

A size frequency distribution (SFD) was used to show the size ranges, the number of observations within a given size, as well as the number of species represented across the study sites. We determined the modal distribution of each species' SFD using the “LaplacesDemon” package (version 16.1.6) in R which tests for unimodality, bimodality, and multimodality in a given distribution. Through robust regression analyses, we tested the OSH by: (a) examining the relationship between individual species' size<sub>95/5</sub> and their relative abundance, assuming that relative abundance is a measure of optimum growth conditions, and (b) investigating the relationship between individual species' size<sub>95/5</sub> and environmental parameters. For these tests,  $p$ -values were adjusted using the Holm-Bonferroni correction method (Holm, 1979) to control the family-wise error rate and reduce the chances of obtaining false positive significance levels. In visualizing the species-specific environmental controls on the individual species size<sub>95/5</sub> (hypothesis test (b) above), because the number of variables tested were many, we binned the variables under umbrella parameters as follows: *carbonate* (includes surface carbonate concentration and carbonate concentration at core depth); *temperature* (includes SST, temperature at 10 m, temperature at 50 m, temperature at 100 m, temperature at 200 m, and difference between winter and summer temperatures); *nutrient* (surface phosphate concentration, surface nitrate concentration, nitrate concentration at 200 m); *oxygen* (includes  $\Delta$ oxygen concentration between 200 and 0 m and oxygen concentration at 200 m); and *salinity* (surface salinity).

We further explored the relationship between size<sub>95/5</sub>, species richness (defined as the number of species in each sample), and species diversity through robust regression analyses. Species diversity was calculated using the classical Shannon–Weiner diversity index (Shannon & Weaver, 1949):

$$H = \sum_{i=1}^S Pi(\ln pi)$$

where,

$H$  = Shannon–Weiner diversity index

$S$  = Total number of species encountered

$Pi$  = proportion of  $i$ th species in the population

$\ln$  = natural logarithm

$\sum$  = sum from species 1 to species  $S$

We assessed the influence of assemblage composition on size<sub>95/5</sub> in two ways: first, in each sample, we investigated the size spectrum of the 13 most abundant species relative to the sample size<sub>95/5</sub> using the density function in the R ggplot2 (version 3.3.6) package (Wickham, 2016); second, through robust linear regression analysis, we evaluated the relationship between size<sub>95/5</sub>, the total number of foraminifera, and the percentage contribution of species with sizes greater than the regional size<sub>95/5</sub> in our data set. The regional size<sub>95/5</sub> was calculated by binning

**Table 2**  
Principal Component Analysis Factor Loading of Surface Hydrographic Parameters in the Tropical Indian Ocean

	PC 1	PC 2	PC 3	PC 4	PC 5	PC 6	PC 7
SST	-0.158	<b>0.735</b>	0.384	-0.455	0.278	0.046	0.021
NIT	0.063	-0.161	-0.114	0.202	<b>0.941</b>	0.112	-0.135
PHOS	-0.094	0.127	0.636	<b>0.748</b>	-0.059	0.071	0.039
PP	-0.060	-0.015	-0.055	-0.049	-0.141	<b>0.959</b>	-0.225
CO <sub>3</sub> <sup>2-</sup>	<b>0.978</b>	0.163	0.107	-0.003	-0.036	0.056	-0.037
OXY	-0.034	0.625	<b>-0.644</b>	0.435	-0.063	-0.021	-0.021
SAL	0.039	-0.027	-0.073	0.006	0.092	0.237	<b>0.963</b>

Note. Keys—SST (sea surface temperature); NIT (surface nitrate concentration), PHOS (surface phosphate concentration); PP (primary productivity); CO<sub>3</sub><sup>2-</sup> (surface carbonate ion concentration); OXY (surface oxygen concentration); and SAL (salinity). The values in bold indicate the parameter with the highest weighting on each PC axis.

the individual sizes of planktonic foraminifera from all core-tops and computing the 95th percentile size. Species with sizes higher than the regional size<sub>95/5</sub> are regarded as “large” because we assume that the size of adult-sized species differ and the maximum size they can attain is fixed. Hence, the percentage contribution of species with sizes greater than the regional size<sub>95/5</sub> represents the ratio of the total number of species where at least one specimen in a sample is larger than the regional size<sub>95/5</sub> to the total number of species in the sample.

To investigate the environmental controls on planktonic foraminiferal assemblage size<sub>95/5</sub> in the TIO, we first characterized planktonic foraminiferal size<sub>95/5</sub> distribution by computing a factor analysis to identify the discernible ecological patterns in the assemblage-wide planktonic foraminifera size<sub>95/5</sub> records. The factor analysis was carried out only on the species' size<sub>95/5</sub> data, with the species name on the horizontal axis and the core IDs on the vertical axis. Each value in the matrix represents the size<sub>95/5</sub> of each species within the core-top sample they were found. The output of the factor analysis are the factor scores and factor loadings. Thereafter, we regressed the factor scores against environmental data to identify the primary and secondary drivers of size at the assemblage level using a robust regression model. We assessed the reliability of our analytical procedure by carrying out factor analysis on the species' relative abundance data, identifying their latent variables, and comparing our results to a similar work by Cayre et al. (1999), who reconstructed paleoproductivity of the TIO from planktonic foraminifera assemblage distribution. Our goal was to check whether the same parameters found by Cayre et al. (1999) to primarily drive planktonic foraminifera assemblage distribution within the TIO were also identified using our machine learning approach. All analyses were carried out in R version 4.0.3 (R Core Team, 2020) and all packages used are listed in Table S1 of Supporting Information S1.

### 3. Results

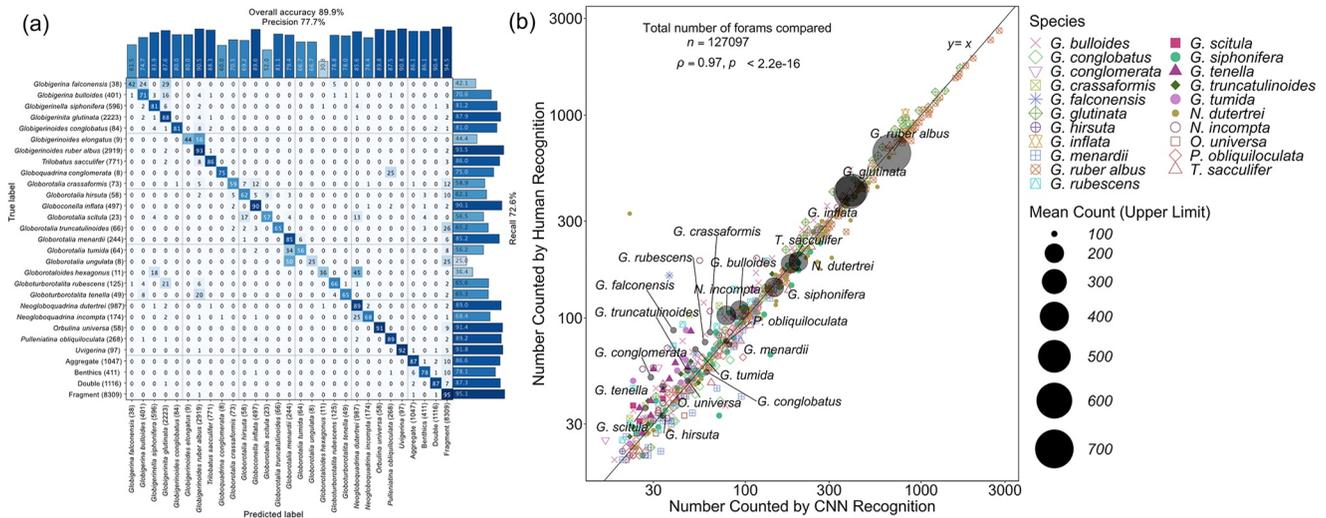
#### 3.1. PCA of Surface Oceanographic Parameters

Results of PCA run on TIO surface hydrographic data show that the first two principal components explain 63% of the total variance (Figure S1 in Supporting Information S1). PC1 alone explains 41% of the variance and is likely associated with seawater carbonate chemistry, indicated by its large component loading on carbonate ion concentration (97%). PC2 explains 22% of the variation and mainly relates to variations in SST as shown by its large component loading on SST (74%; Table 2). We present the full component loadings of the surface parameters analyzed in Table 2.

#### 3.2. Ground-Truthing Our High Throughput Workflow

##### 3.2.1. CNN Performance and Comparison With Human Classifier

Of the six CNNs trained, the “Base Cyclic 16” network gives the best results, reaching an accuracy of 89.9%; a precision of 77.7%; and a recall of 72.6% over five iterations (Figure 2a). “Base Cyclic 16” is robust in identifying classes such as whole planktonic foraminifera, fragments (broken foraminifera), *Uvigerina peregrina* (a benthic foraminiferal species), aggregates (including radiolarians, acritarchs, and other organic materials such as broken non-foraminiferal shells), and doubles (i.e., more than one particle in an image). This is shown by



**Figure 2.** (a) Confusion matrix showing the performance of “Base Cyclic 16” convolutional neural network on the validation data set. We observed that poorly represented samples in the validation set were more frequently misidentified (more false positives) compared to well-represented species. (b) Count comparability between human and machine classifiers for the most abundant species. Although identified, rare species such as *Globigerinella calida* are excluded from this analysis. *Globigerinoides ruber albus* and *Globigerinita glutinata* were the most abundant species in the tropical Indian Ocean data set. Values on both  $x$ - and  $y$ -axis are represented as log-scales. The mean count (upper limit) represents the average count per sample within a range binning of 100.

its high accuracy scores ranging between 86% and 95%, and a recall of 78%–95%. Other CNNs trained typically ranged between 81% and 88% in accuracy, 64%–76% in precision, 60%–70% in recall. The results of the performance of the “Base Cyclic 16” network tested over five iterations is presented in Table S2 of Supporting Information S1.

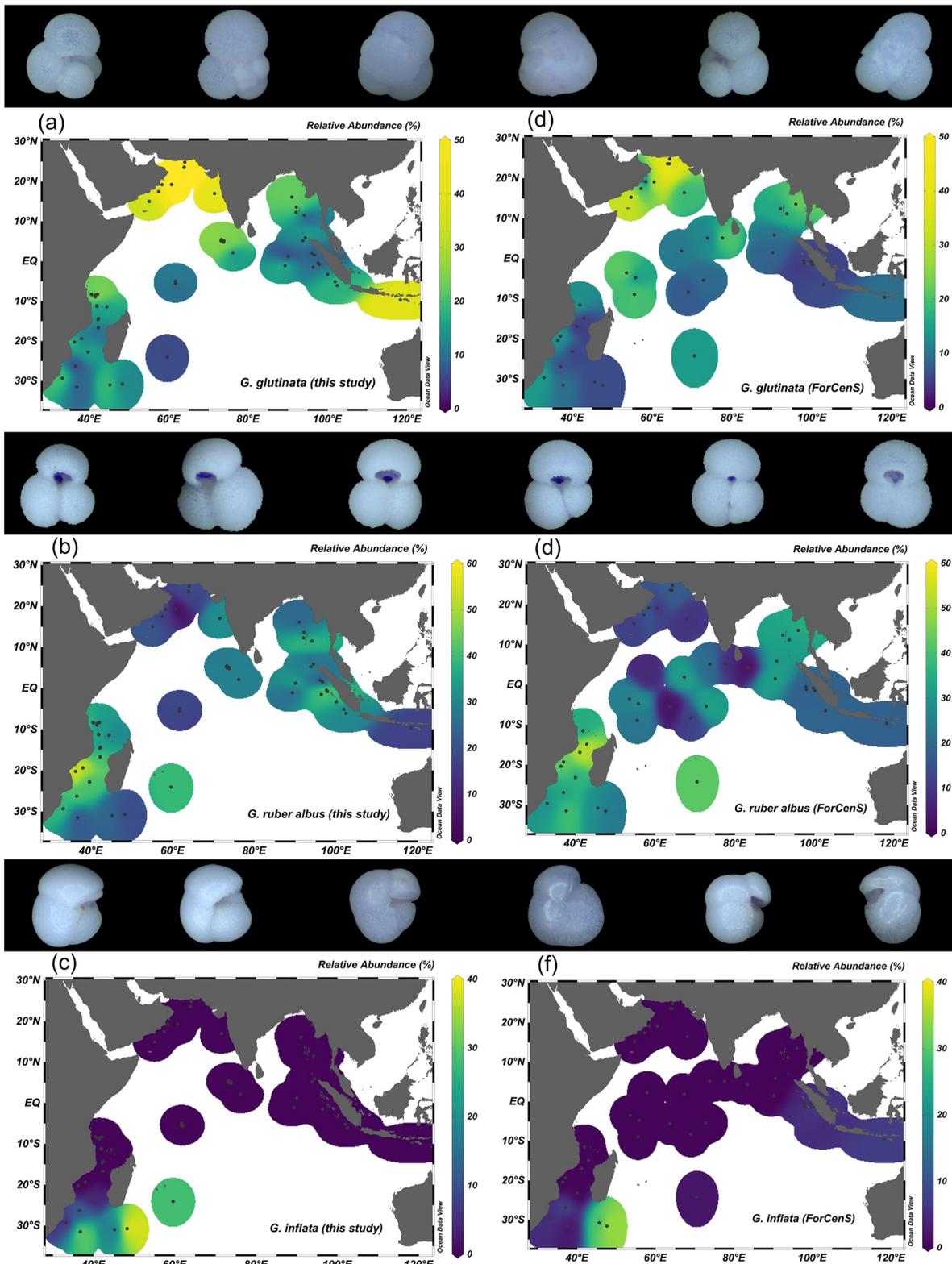
After deploying the “Base Cyclic 16” CNN model in the automatic classification of all images, comparison with the classification by a human classifier shows that the machine successfully labeled most of the species and achieved near full accuracy compared to a human classifier ( $\rho = 0.97, p < 2.2e-16$ , Figure 2b). Using this automated classification scheme, we plot the spatial distribution of planktonic foraminifera from our core-top data set (Figures 3a–3c). While *G. ruber albus*, *G. inflata*, and *Globigerinita glutinata* are the most abundant species in the TIO, we observe a dominance of *G. glutinata* in the northern TIO reaching 85%, while the southern TIO is dominated by *G. ruber* up to 69%, as shown in Figure S2 of Supporting Information S1.

### 3.2.2. Relative Abundance Comparison With ForCenS Database

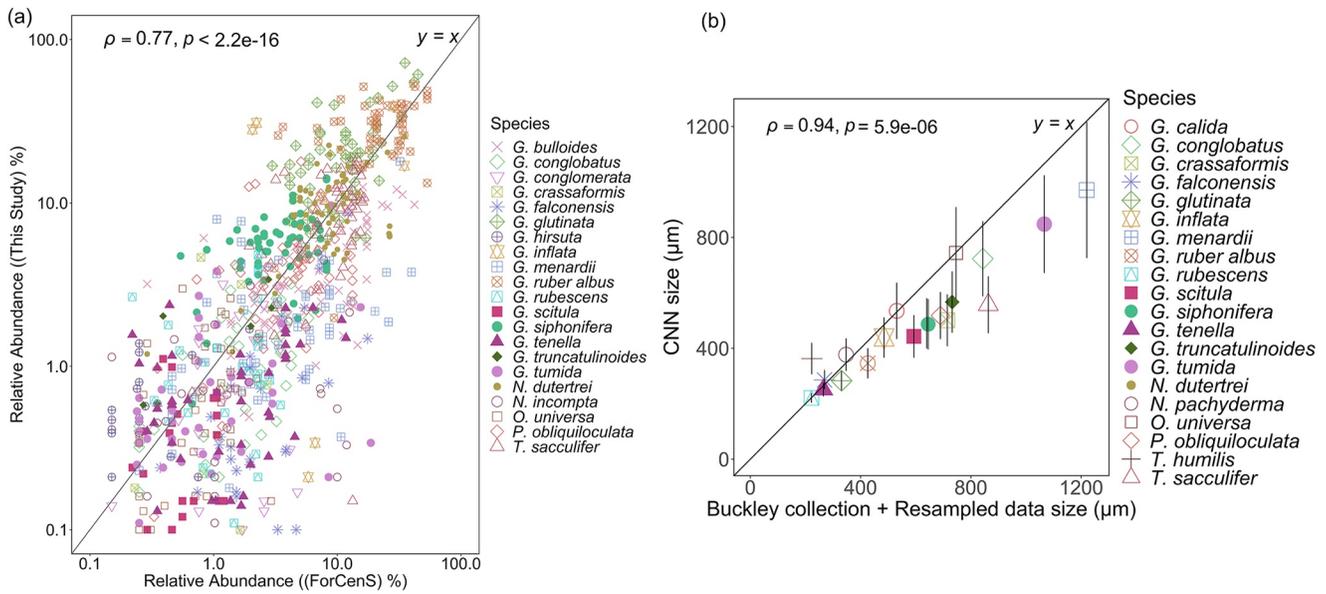
Comparison of the relative abundance of the 21 most abundant species from our study sites with the corresponding closest sites (pairing was based on a single site) in the *ForCenS Database* (Siccha & Kucera, 2017) shows a significant correlation (Figure 4a). While the data fall near the  $y = x$  line and the correlation between both data sets is strong ( $\rho = 0.77, p < 2.2e-16$ ), some differences in the relative abundances of some species occur. These could result from differences in the total number of individuals counted in this study (2,000 individuals per sample on average) compared to that of the *ForCenS* database (300 individuals per sample on average). Nevertheless, the similarity between the two data sets becomes more evident when we observe the regions of highest relative abundance of some of the species. For example, in both studies, the relative abundance of *G. glutinata* is highest in the Arabian Sea while the highest relative abundance of *G. ruber albus* is seen around the Mozambique Channel. Likewise, *G. inflata* relative abundance is highest around the Southern Mozambique Basin (Figure 3 and Figure S3 in Supporting Information S1).

### 3.2.3. Morphometry Data Comparison With Rillo et al. (2020) Data Set

There is a strong correlation ( $\rho = 0.94, p = 5.9e-06$ ) between the  $size_{95/5}$  of the TIO planktonic foraminiferal species measured in the present study and those recorded in the Rillo et al. (2020) data set, confirming the reliability of the morphometric data generated using our workflow (Figure 4b). However, the size of many of the species fall below the  $y = x$  line, suggesting that the Rillo et al. (2020) data set is biased toward larger sizes. This bias, noted by Rillo et al. (2020), comes from a collector bias who predominantly picked, identified, and curated the largest foraminifera in their samples.



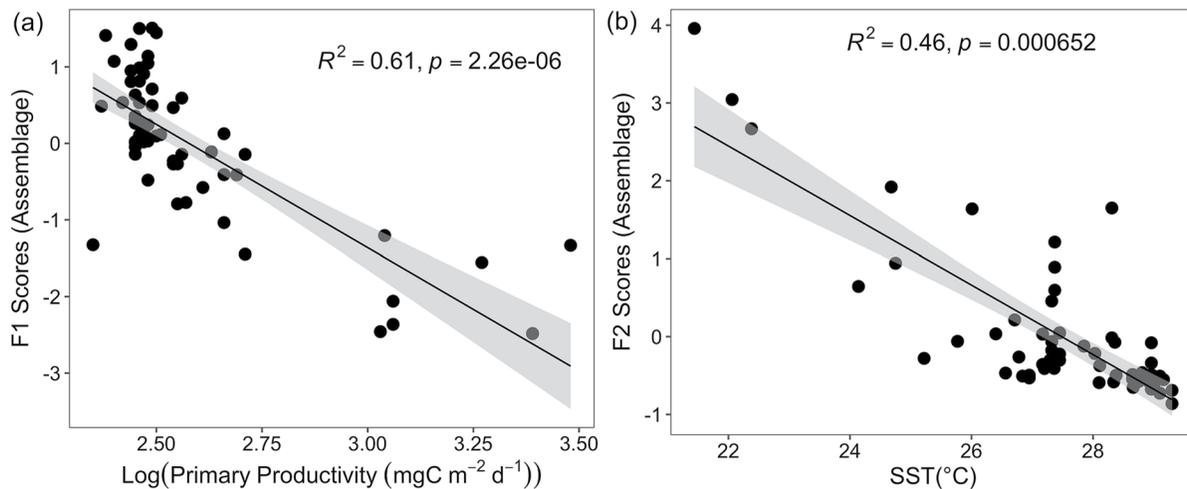
**Figure 3.** Similarity in the areas of highest relative abundance of *G. glutinata*, *G. ruber albus*, and *G. inflata* in this study versus ForCenS database. Purple color represents areas of low relative abundance while yellow color represents areas of high relative abundance. Gradient colors between purple and yellow colors represent intermediate relative abundance (See Figure S3 in Supporting Information S1 for other species). Representative images of each species are shown above their distribution map.



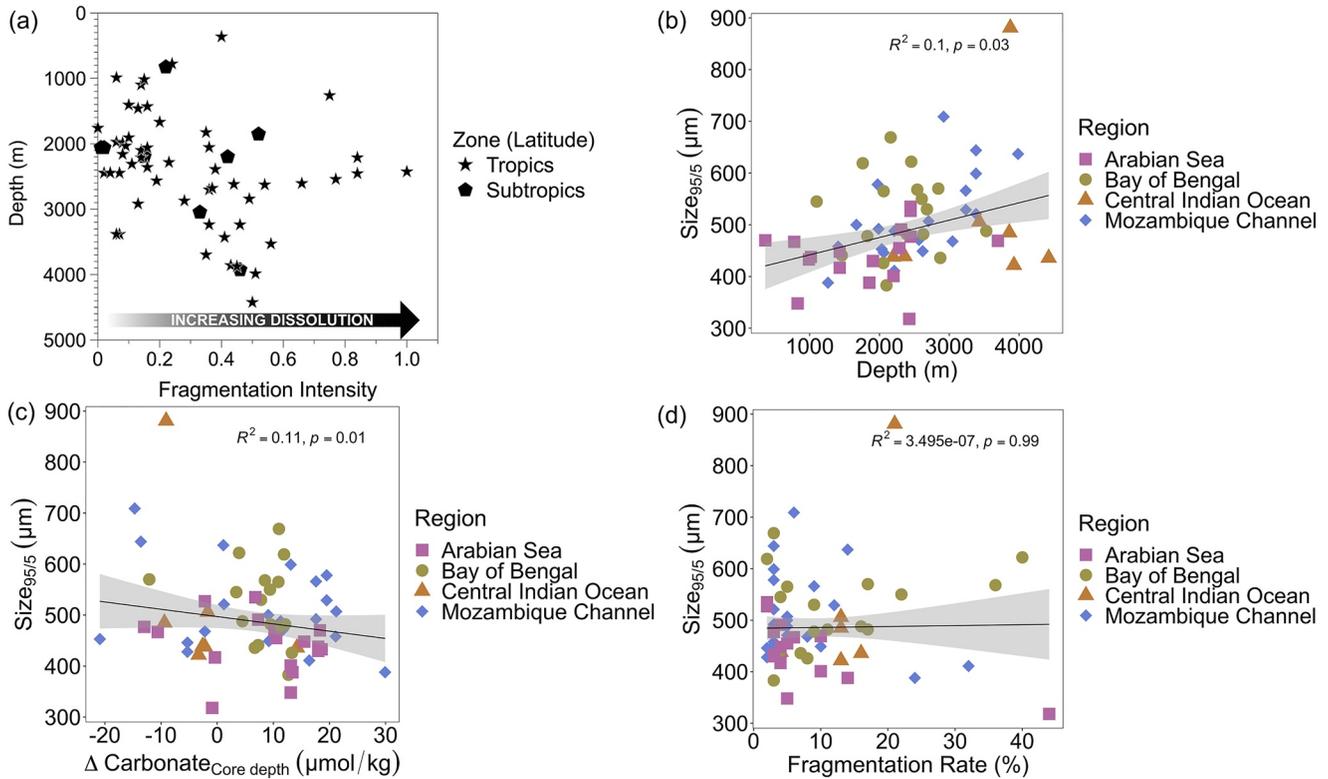
**Figure 4.** (a) Comparison between the relative abundance of planktonic foraminifera species (this study) versus the ForCenS database. Site selection was based on nearest-neighbor that is, the site closest to our core locations in the ForCenS Database (Siccha & Kucera, 2017). (b) Comparison between planktonic foraminifera size<sub>95/5</sub> recorded in this study and those from the tropical Indian Ocean samples in the Rillo et al. (2020) data set. Line is identity function. Error bars represent 1 standard deviation in our measurements. The Buckley collection represents samples from the Henry Buckley Collection of Planktonic Foraminifera stored at The Natural History Museum in London, UK while the “resampled data” refers to the data from the bulk sediments from which Rillo et al. (2020) validated the Buckley collection. The resampling was done on the Ocean Bottom Deposits collection which were bulk sediments from which Buckley built his collections.

### 3.2.4. Assemblage Distribution Factor Analysis Comparison With Cayre et al. (1999)

Factor analysis on species' relative abundance data from our study sites reveal that 10 axes explain 72% of the total variance in the assemblage distribution of the species studied, with the first two axes accounting for 32% of the total variance. Thus, we make interpretations based on the first two axes. High negative factor scores for the first factorial axis are related to species such as *Globigerina falconensis*, *Globigerina bulloides*, *G. glutinata*, and *T. sacculifer*. In the second factorial axis, high positive factor scores are represented by species such as *Globorotalia crassaformis*, *G. truncatulinoides*, and *G. inflata* (see Table S3 in Supporting Information S1); interestingly, these same species are highly weighted in the second axis of the size factor loading. Regression between



**Figure 5.** Robust regression analysis between the *F1*-Axis factor scores computed from the factor analysis of 26 planktonic foraminifera species' relative abundance data and environmental parameters showed best fit with log of primary productivity (PP) (a), while the *F2*-Axis factor scores showed best fit with sea surface temperature (b). Note that the PP data was log-transformed because of the non-normal distribution of the data and the larger data range relative to other variables.



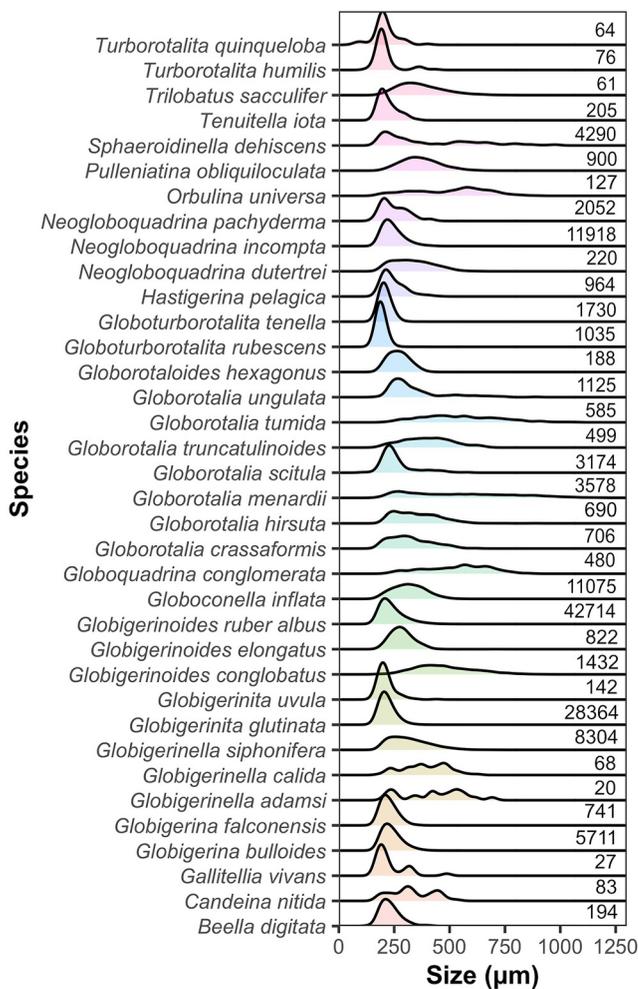
**Figure 6.** Dissolution proxies: (a) Fragmentation intensity according to Suárez-Ibarra et al. (2021) versus depth; (b) size<sub>95/5</sub> versus depth; (c) size<sub>95/5</sub> versus carbonate saturation state at core depth; (d) size<sub>95/5</sub> versus fragmentation rate. Shaded intervals represent 95% confidence interval.

the  $F1$ -Axis and environmental parameters show highest  $R^2$  values with (log transformed) PP ( $\text{mgC m}^{-2} \text{d}^{-1}$ ) ( $R^2 = 0.61$ ,  $p < 0.05$ , Figure 5a) while the  $F2$ -Axis show highest  $R^2$  values with SST ( $R^2 = 0.46$ ,  $p < 0.05$ , Figure 5b). This result is similar to that of Cayre et al. (1999) whose first factorial axis after PCA on planktonic foraminifera assemblage distribution in core-top samples from the TIO showed best fit with PP. This result gave us confidence in our subsequent analysis on the assemblage size data set.

### 3.3. Size Distribution of Planktonic Foraminifera in the TIO

#### 3.3.1. Dissolution

The fragmentation intensity dissolution index indicates good preservation in most of the samples (Figure 6a). However, it suggests relatively high levels of dissolution (index  $>0.6$ ) in six out of 62 samples, representing about 10% of the samples. A weak relationship is observed between size<sub>95/5</sub> and depth ( $R^2 = 0.10$ ,  $p = 0.03$ ) (Figure 6b). Similarly, changes in carbonate saturation state at core depth ( $\Delta[\text{CO}_3^{2-}]$ ) show a weak relationship with size<sub>95/5</sub> ( $R^2 = 0.11$ ,  $p = 0.01$ ) (Figure 6c). No relationship is found between size<sub>95/5</sub> and fragmentation rate ( $R^2 = 3.50\text{e}-07$ ,  $p > 0.99$ ) (Figure 6d). Given that changes in seafloor depth and bottom water carbonate saturation state respectively explain 10% and 11% in the variation of our size metric (size<sub>95/5</sub>), and only 10% of the samples were identified by the fragmentation intensity dissolution index as being significantly affected by dissolution, we conclude that the influence of dissolution on our species' size records is small and that our results in relation to environmental parameters are likely robust. This is supported by the fact that the size<sub>95/5</sub> values in some deeper sites are smaller than those in shallower waters, whereas preferential dissolution of smaller individuals might be expected at deeper sites with lower carbonate saturation. Thus, we did not exclude any of the sites identified by the fragmentation intensity index from our analysis because their species size spectra and size<sub>95/5</sub> do not indicate any significant bias toward larger individuals or more robust species, and the total whole foraminifera counts from these samples were quite high.



**Figure 7.** An overview of the size frequency distribution (SFD) of the planktonic foraminifera species recorded in this study. About 56% of the species showed unimodal distribution for adult sized species ( $>150 \mu\text{m}$ ), while 44% of the species showed multimodal distributions. The numbers to the right of each species' SFD represent the total number of individuals represented in their distribution.

### 3.3.2. Species Size Frequency Distribution

Of the 36 species recorded, the size-frequency distribution (SFD) of 23 species appear to possess unimodal distributions, while the remaining 13 species show multimodal distributions (Figure 7). Bimodal distribution is observed in species such as *N. pachyderma* and *Candeina nitida*. Meanwhile, the SFD of the largest species such as *G. menardii*, *Globoquadrina conglomerata*, and *Globorotalia tumida* are widely spread, showing no clear peaks. Statistical confirmation of the modal distribution types observed for each species is presented in Table S4 of Supporting Information S1.

### 3.4. Test for the OSH

#### 3.4.1. Size Versus Relative Abundance

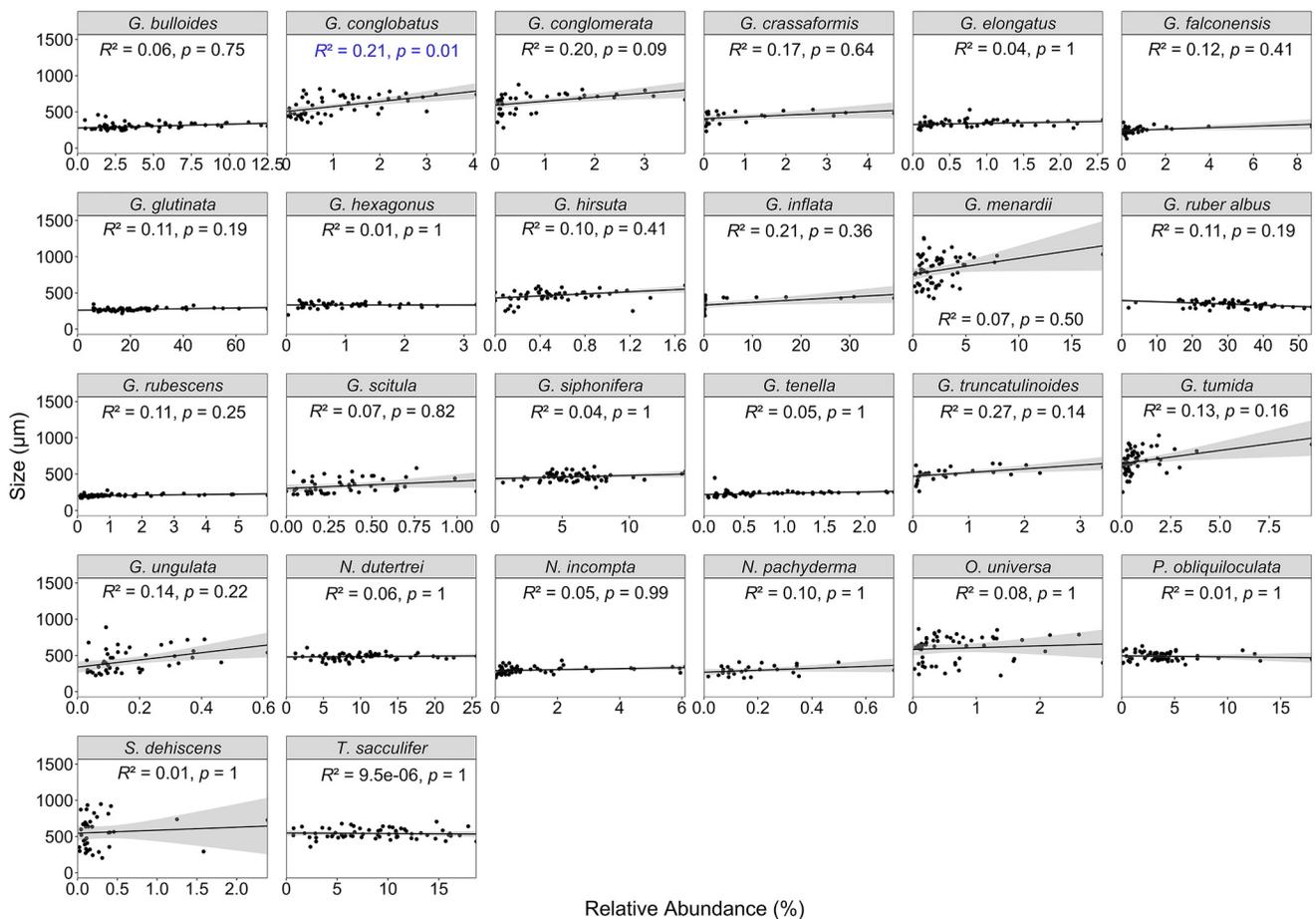
Out of the 26 species tested, only *G. conglobatus* increased in  $\text{size}_{95/5}$  with increasing relative abundance ( $R^2 = 0.21$ ,  $p = 0.01$ ) (Figure 8). No significant relationship is observed between the  $\text{size}_{95/5}$  and relative abundance of most species, including the abundant species such as *G. glutinata*, *G. elongatus*, *N. dutertrei*, and *T. sacculifer*.

#### 3.4.2. Size Versus Abiotic Forcing (Environmental Parameters)

Evaluated based on the parameter that showed the best coefficient of determination ( $R^2$ ), the response of individual species'  $\text{size}_{95/5}$  to environmental parameters investigated in this study reveals that 25% of the species show a low to moderate significant response ( $R^2$  values range from 0.10 to 0.35,  $p < 0.025$ ) to temperature-related parameters excluding SST (Figure 9). On the other hand, parameters related to carbonate ion concentration ( $R^2$  values range from 0.10 to 0.29,  $p < 0.025$ ) explain  $\text{size}_{95/5}$  variation in 35% of the species, especially in deep-dwelling and large-sized species (e.g., *G. menardii*, *G. tumida*, *G. truncatulinoides*, and *P. obliquiloculata*). Factors such as salinity ( $R^2$  values range from 0.22 to 0.51,  $p < 0.025$ ) and oxygen ( $R^2$  values range from 0.18 to 0.23,  $p < 0.025$ ) explain  $\text{size}_{95/5}$  variation in 20% and 10% of the species respectively. Meanwhile, nutrient availability explains  $\text{size}_{95/5}$  variation in 10% of species ( $R^2$  values range from 0.19 to 0.31,  $p < 0.025$ ). Only *Neogloboquadrina incompta*  $\text{size}_{95/5}$  shows no significant relationship with any environmental parameter (see Table S5 in Supporting Information S1 for details of the exact parameters that are found to drive the variation in individual species'  $\text{size}_{95/5}$ ).

With a focus on the impact of SST on the size variations of the species examined, nine out of 20 species show weak but significant relationships with SST including *G. bulloides* ( $R^2 = 0.07$ ,  $p = 0.01$ ), *G. siphonifera* ( $R^2 = 0.07$ ,  $p = 0.002$ ), *T. sacculifer* ( $R^2 = 0.05$ ,  $p = 0.03$ ), *G. conglomerata* ( $R^2 = 0.13$ ,  $p = 0.01$ ), *G. crassaformis* ( $R^2 = 0.18$ ,  $p = 0.01$ ), *Globorotalia hirsuta* ( $R^2 = 0.19$ ,  $p = 0.001$ ), *G. inflata* ( $R^2 = 0.30$ ,  $p = 2.79 \times 10^{-5}$ ), *G. menardii* ( $R^2 = 0.05$ ,  $p = 0.03$ ), and *N. dutertrei* ( $R^2 = 0.04$ ,  $p = 0.05$ ). However, none of these  $R^2$  values are greater than the parameters that we find to primarily drive size changes in all these species. See Table S6 in Supporting Information S1 for details of the  $R^2$  and  $p$ -values describing the relationship between individual planktonic foraminifera species  $\text{size}_{95/5}$  and mean annual SST.

Our re-analyses of TIO planktonic foraminifera in the Rillo et al. (2020) data set reveal that, out of the nine species investigated, only three show significant relationships between their  $\text{size}_{95/5}$  and SST. The three species are *T. sacculifer* ( $R^2 = 0.22$ ,  $p = 0.03$ ), *G. truncatulinoides* ( $R^2 = 0.58$ ,  $p = 0.02$ ), and *G. inflata* ( $R^2 = 0.69$ ,  $p = 0.03$ ), although we note that the number of samples per species are significantly lower than in our study (62 vs. 14 for *T. sacculifer*, 27 vs. 9 for *G. truncatulinoides*, and 25 vs. 4 for *G. inflata*) (Figure S4 in Supporting Information S1).



**Figure 8.** Single-species robust regression analysis showing the relationship between individual species' size<sub>95/5</sub> and their relative abundances. We assume that their relative abundances indicate optimum growth conditions. *P*-values were corrected using the Holm–Bonferroni correction method.

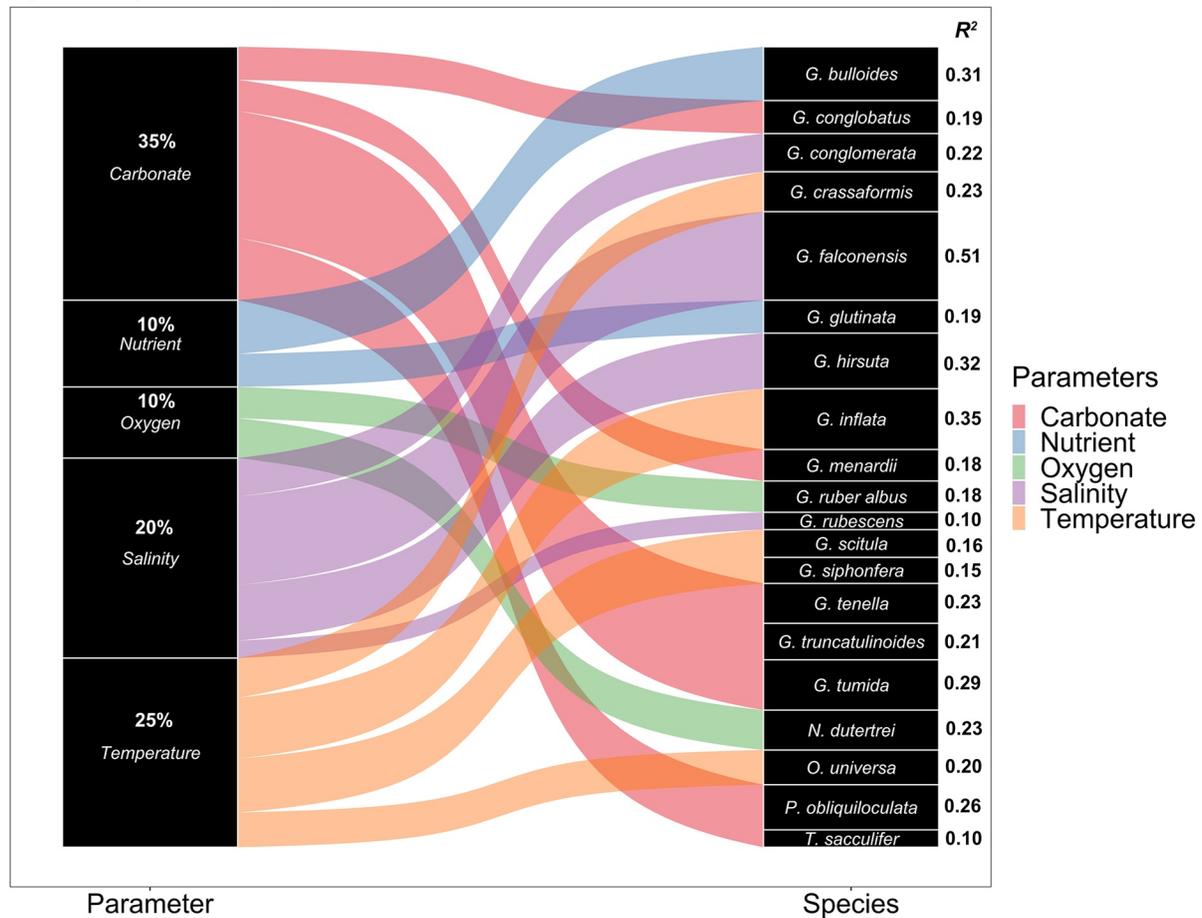
### 3.5. Size Versus Species Diversity and Species Richness

Species richness shows no relationship with assemblage size<sub>95/5</sub> ( $R^2 = 0.013$ ,  $p = 0.546$ ) (Figure 10a). Conversely, the Shannon–Weiner diversity index increases with increasing assemblage size<sub>95/5</sub> ( $R^2 = 0.49$ ,  $p = 6.43 \times 10^{-9}$ ) (Figure 10b). Highest Shannon diversity (2.34) is found in a core-top located near the Madingley Rise (central Indian Ocean) at a water depth of 3,875 m while minimum Shannon diversity (1.21) is recorded in a core located in the Arabian Sea at a water depth of 2,427 m.

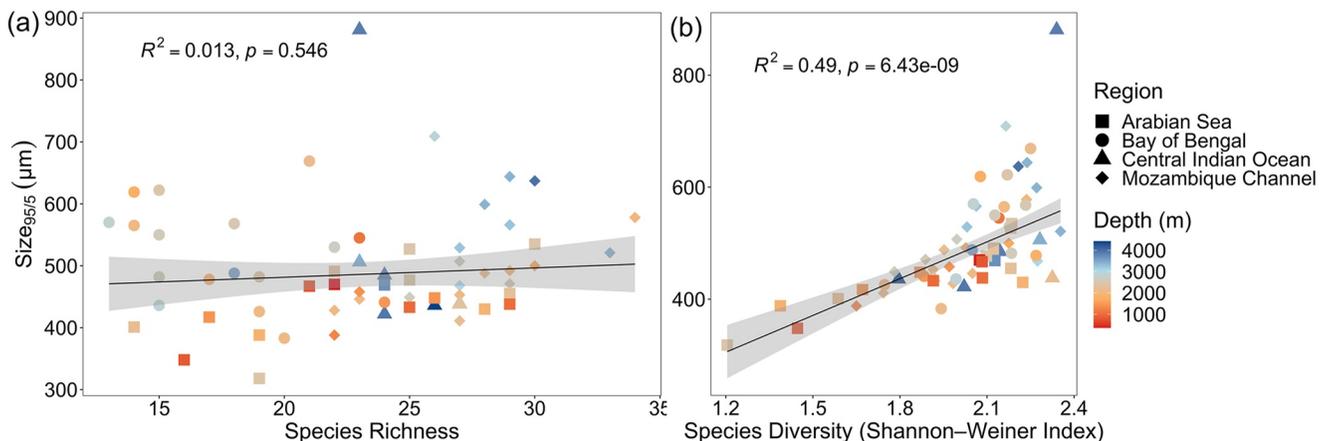
### 3.6. Influence of Assemblage Composition on Assemblage Size<sub>95/5</sub>

Generally, the percentage contributions of species with sizes greater than the regional size<sub>95/5</sub> in our samples are around 20%–40%, with the largest contributions (50%) observed predominantly in core-tops from the central Indian Ocean and Bay of Bengal (including offshore Java) (Figure 11). Above a size<sub>95/5</sub> of 550 µm, assemblage size<sub>95/5</sub> seems to be impacted by the increasing percentage contribution of species with sizes greater than the regional size<sub>95/5</sub>. Within the central Indian Ocean, irrespective of the total number of foraminifera, size<sub>95/5</sub> values do not exceed ~500 µm except in one core-top. Thus, this observation suggests that foraminifera growing in this region are smaller. Finally, we observe that maximum size<sub>95/5</sub> values are attained at around a count of 2,000 individuals. We suggest that this might be a minimum count limit for foraminiferal size analysis using automated systems. Further, we find no clear regional character in the modal size and size<sub>95/5</sub> in the studied sites based on assemblage composition. In all cores, the modal size is lower than the size<sub>95/5</sub>, thus indicating that the size distribution in all the cores is not skewed by large sized species and that small sized species contribute significantly to the assemblages (see Figure S5 in Supporting Information S1).

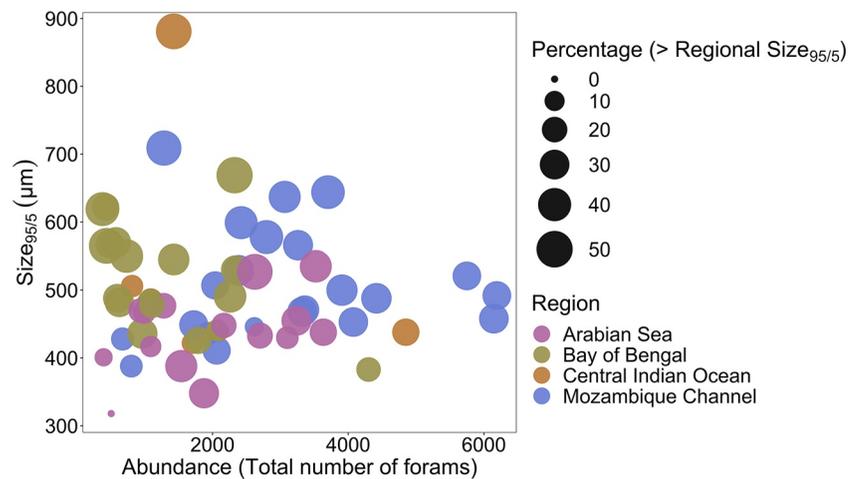
Species-specific Response to Environmental Variables



**Figure 9.** Robust regression analysis between individual planktonic foraminifera species' size<sub>95/5</sub> and environmental parameters. Values contained in each parameter box represent the percentage proportion explained by that parameter among the five broad parameters tested. Values to the right of each species represent the coefficient of determination between that species' size<sub>95/5</sub> and the parameter it showed the best fit with. Because the number of variables tested were many, we binned the variables under umbrella parameters as follows: **Carbonate** (includes surface carbonate concentration, carbonate concentration at 200 m, and  $\Delta$ Carbonate concentration between 0 and 500 m); **Temperature** (includes temperature at 100 m, temperature at 200 m, winter sea surface temperature (SST), and summer SST); **Nutrient** (includes surface phosphate concentration and phosphate concentration at 200 m); **Oxygen** (includes  $\Delta$ oxygen concentration between 200 and 0 m); and **Salinity** (surface). See Table S6 in Supporting Information S1 for details about the exact parameters that were found to drive individual species' size<sub>95/5</sub> variation.



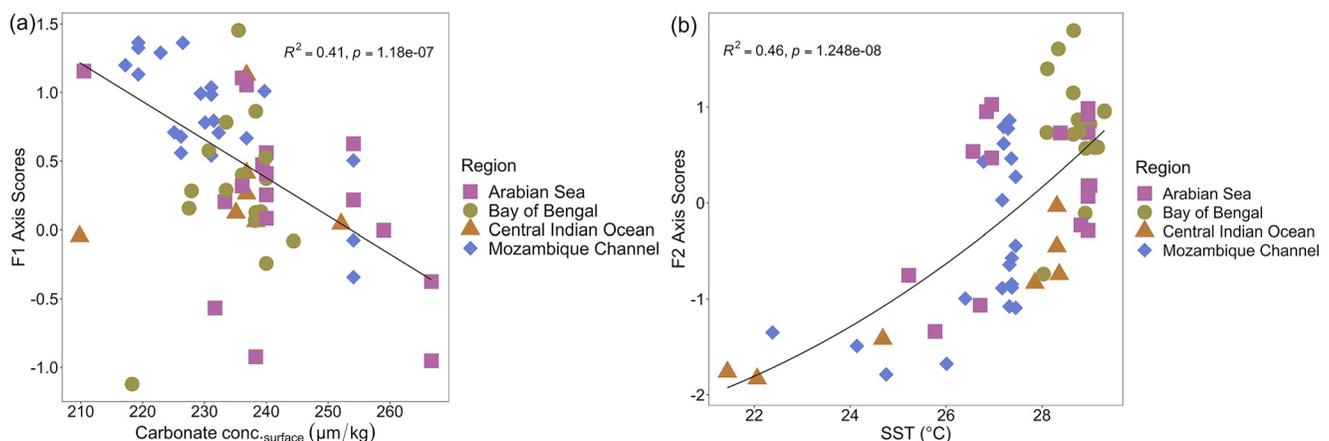
**Figure 10.** (a) Relationship between size<sub>95/5</sub> and species richness; (b) relationship between size<sub>95/5</sub> and species diversity. Shaded intervals represent the 95% confidence interval.



**Figure 11.** Size<sub>95/5</sub> versus total numbers of planktonic foraminifera per sample. The size of the circles corresponds to the percentage contribution of species with sizes greater than the regional size<sub>95/5</sub> in the Indian Ocean. The regional size<sub>95/5</sub> here is taking as the arbitrary size cut-off that defines the size above which species are regarded as “large” in the tropical Indian Ocean. Increasing percentage representation show the effect of assemblage composition on local size<sub>95/5</sub> values.

### 3.7. Assemblage Size Response (Factor Analysis)

The size spectra of 26 planktonic foraminiferal species were used for this analysis. Ten factors explain 70% of the total size variance. The first and second axes explain 45% of the variance while axes three to 10 explain only 25% and were often monospecific, thus, we only used the first and second axes. The first factorial coordinate of the *F*-Matrix derived from the factor analysis of planktonic foraminifera assemblage size attributes high positive factor scores mostly to species that either bear calcite crusts or are large-sized. These include *Globigerinoides conglobatus*, *G. menardii*, *G. tumida*, *Pulleniatina obliquiloculata*, *Neogloboquadrina dutertrei*, and *Orbulina universa*. On the other hand, high negative factor scores are mainly attributed to deep-dwelling species such as *G. inflata*, *G. crassaformis*, and *G. truncatulinoides* on the second factorial coordinate (see Table S7 in Supporting Information S1). Among the environmental parameters explored, the first factorial coordinate shows the best fit with surface carbonate ion concentration (Figure 12a), while the second factorial coordinate shows strongest fit with SST (Figure 12b).



**Figure 12.** (a) Robust regression analysis between *F1*-Axis factor scores computed from the Factor Analysis of 26 planktonic foraminifera species morphometric data versus surface carbonate concentration; (b) second order polynomial relationship between *F2*-Axis scores versus sea surface temperature.

## 4. Discussion

### 4.1. CNN Classifies Species Images With High Accuracy

A review of our neural network “Base Cyclic 16” CNN confusion matrix (Figure 2a) reveals that despite its high accuracy, it misclassified some planktonic foraminifera species. The major identified biases by the CNN were inductive (i.e., when a model incorrectly predicts a species' class due to the non-inclusion of the species in the training data set or favoring of highly sampled classes over less-represented classes), numerical, and taxonomic. For instance, *G. elongatus* was often misidentified as *G. ruber albus*, *G. tumida* as *G. menardii*, *Globorotalia ungulata* as either *G. tumida* or *G. menardii*, and *Globorotaloides hexagonus* as *N. dutertrei*.

In comparison to the other models tested, our “Base Cyclic 16” model was less susceptible to biases resulting from orientation, over-representation of a single species, and pigmentation (a situation where two morphologically similar species are distinguishable by color). Due to misclassification of species with low abundance, the CNN classifier underestimates the population of minor species, but this bias is noticed by the human classifier. Similar to human classifiers, the CNN classifier accuracy improves as the average species count increases. The strong correlation ( $\rho = 0.97$ ,  $p < 2.2e-16$ ) between the number of individuals counted by the CNN versus human classifiers across 21 species highlights the reliability of the data produced using our “Base Cyclic 16” neural network (Figure 2b). This is further supported by our results correlating the relative abundance data generated by the model with the data retrieved from the closest stations to the cores used in this study from the modern surface sediment in the ForCenS database (Figure 4a); this also showed significant positive correlation ( $\rho = 0.77$ ,  $p < 2.2e-16$ ). Species such as *G. ruber albus*, *G. glutinata*, and *G. inflata*, recorded the highest relative abundance in the ForCenS database at similar locations in our study (Figure 3). Although skewed toward large sizes, the relatively comparable size spectra of the Rillo et al. (2019) and the data set produced by MiSo provides confidence in the use of automated workflows such as MiSo/ParticleTrieur in the identification, classification, and measurement of planktonic foraminiferal species (Figure 4b) (Marchant et al., 2020). Our workflow and CNN model show a better accuracy (89.9%, Figure 3a) than other workflows utilizing supervised learning models in the automatic identification of planktonic foraminifera, for example, the “VGG 16” model test (27,737 training images, 6,903 validation data set) by Hsiang et al. (2019) achieved an accuracy of 87.4% when used to classify modern planktonic foraminifera images. Likewise, the “ResNet50 + VGG 16” model test (1,258 training images, 180 validation data set) by Mitra et al. (2019) achieved an accuracy of ~86% when used to classify images of modern planktonic foraminifera.

### 4.2. TIO Planktonic Foraminifera Mostly Exhibit Unimodal Size Frequency Distribution at the Species Level

The SFD of a species is a measure of the density of the species' size along a size spectrum within a sample (Peeters et al., 1999). In this study, about half of the species in our data set exhibit a unimodal distribution while others show flat to multimodal distributions (Figure 7, Table S4 in Supporting Information S1). The species *Neogloboquadrina pachyderma* and *Candeina nitida* show bimodality in their SFDs. SFDs in this study compare favorably with those reported in the literature, though existing data sets are sparse. For example, consistent with our findings, Peeters et al. (1999) reported unimodal SFDs for *G. bulloides*, *G. ruber albus*, *T. sacculifer*, and *G. glutinata* in Arabian Sea surface sediments. The modal sizes of these species were found to be very similar to those in our study, with values of 210  $\mu\text{m}$  (*G. bulloides*), 206  $\mu\text{m}$  (*G. ruber*), 299  $\mu\text{m}$  (*T. sacculifer*), and 189  $\mu\text{m}$  (*G. glutinata*) reported for the Arabian Sea (Peeters et al., 1999), compared to 226  $\mu\text{m}$  (*G. bulloides*), 198  $\mu\text{m}$  (*G. ruber*), 290  $\mu\text{m}$  (*T. sacculifer*), and 193  $\mu\text{m}$  (*G. glutinata*) reported in this study.

*N. pachyderma* and *C. nitida* showed bimodal SFDs, however, we note that these species have the lowest counts (127 and 83 specimens respectively) in the entire data set. Hence, although we have confirmed the bimodality of both species via a distribution test, we think that the observed bimodal SFDs may have also resulted from the low representation of these species in our data set, and larger populations need to be analyzed to confirm this result. Nevertheless, in a study investigating the distribution of planktonic foraminifera in the Arctic Ocean, Carstens et al. (1997) reported a weak bimodal SFD in *N. pachyderma* populations ( $n = 6,768$ ). Planktonic foraminiferal SFD can be controlled by multiple processes including surface water physical and chemical properties (Naidu & Malmgren, 1995), reproductive cycle (Bijma et al., 1994), size-dependent weight, morphology (spinose vs. non-spinose) and sinking speed (Bijma et al., 1994; Takahashi & Be, 1984), dissolution (Peeters et al., 1999),

cryptic diversity (Huber et al., 1997), winnowing after settling on the sea floor (Peeters et al., 1999), and the tendency for the morphological characteristics of a species to vary depending on how the species reproduces (that is, sexually or asexually) (Meilland et al., 2022). Since no published water column size spectra are available to the best of our knowledge to compare our data set with living communities, we infer that the SFDs of the well-represented species reported here are representative of their regional SFDs.

#### 4.3. Vertical Niche Partitioning May Provide a Link Between Assemblage Size and Diversity

In this study restricted to the TIO, increasing species richness does not influence assemblage size<sub>95/5</sub> ( $R^2 = 0.013$ ,  $p = 0.546$ ) (Figure 10a). This result is supported by Zarkogiannis et al. (2020), who also observed no relationship between size and species richness in core-top samples from the Eastern Mediterranean Sea. Meanwhile, planktonic foraminiferal size<sub>95/5</sub> has been reported to positively correlate with species richness on a global scale (Schmidt, Renaud, et al., 2004). This suggests that the size versus species richness relationship is only observable on a global scale and that at the regional to sub-regional scale, the signal of this relationship is lost because of the lower variability in prevailing hydrographic conditions.

Conversely, we found a positive relationship between assemblage size<sub>95/5</sub> and species diversity ( $R^2 = 0.49$ ,  $p = 6.43e-09$ ) (Figure 10b). Two models could explain this relationship. First, the diversity–productivity model suggests that diversity in planktonic foraminifera is a unimodal function of biomass and this in turn affects species' size distribution as communities try to achieve balance between food availability and selective predation (Irigoin et al., 2004). In this case, a positive relationship between species diversity and PP is expected. Secondly, the thermal stratification model suggests that vertical heterogeneity supports niche and depth-parapatric speciation, resulting in the gradual addition of new niches and larger sized species (Whittaker et al., 2001). In this case, the relationship between species diversity and thermal stratification (here we use the temperature difference between 0 and 200 m,  $\Delta T$ ) should be positive. Our results from the TIO lend support to the thermal stratification model rather than the diversity–productivity model because the correlation between species diversity and PP yielded a negative relationship ( $\rho = -0.27$ ,  $p = 0.03$ ), while its relationship with  $\Delta T$  is positive ( $\rho = 0.31$ ,  $p = 0.02$ ). In our study, the association of higher planktonic foraminiferal diversities and larger size<sub>95/5</sub> at sites in oligotrophic areas (and lower diversities and smaller size<sub>95/5</sub> in highly productive upwelling areas), contradicts the diversity–productivity model. This is consistent with the findings of Schmidt, Renaud, et al. (2004) on a global scale. Al-Sabouni et al. (2007) also found a similar negative relationship between species diversity and PP in the Atlantic, whilst Rutherford et al. (1999) reported that changes in upper-ocean thermal structure strongly drive planktonic foraminiferal diversity. Seasonality may also play a role in explaining the size–diversity relationship in the TIO. Although the correlation between species diversity and temperature seasonality ( $\Delta T$  summer–winter) reveals no relationship ( $\rho = 0.08$ ,  $p = 0.56$ ), PP in the TIO is highly seasonal (Longhurst, 2007), increasing the range of ecological niches over the year that species can adapt to. This warrants further investigation.

#### 4.4. Abundance and Ecological Optima

The OSH postulates that planktonic foraminiferal species attain largest size at their ecological optimum (Hecht, 1976). Thus, if a species' abundance or relative abundance is a biotic measure of stress (Drake & Griffen, 2010), and by extension ecological optimum, then its largest size should be attained where it is most abundant and least stressed. In this study, among the 26 species where the relationship between size<sub>95/5</sub> and relative abundance was investigated, only *G. conglobatus* ( $R^2 = 0.21$ ,  $p = 0.013$ ) significantly increased in size<sub>95/5</sub> with increasing abundance (Figure 8). This implies that relative abundance is not a good measure of optimum growth conditions in planktonic foraminifera, or that size does not increase under optimum conditions. This result is consistent with Rillo et al. (2020), who found no relationship between size<sub>95/5</sub> and relative abundance in eight out of nine planktonic foraminifera species in core-top samples. Similarly, Weinkauff et al. (2019) found no relationship between the test size of large populations of *O. universa* and their abundance. Although previous studies found support for the OSH based on the relationship between species' size and abundance (Hecht, 1976; Kahn, 1981; Malmgren & Kennett, 1976; Moller et al., 2013), our results do not support the use of relative abundance as a determinant of optimum conditions, if one assumes that size<sub>95/5</sub> indeed does increase under optimum conditions.

The lack of relationship between size<sub>95/5</sub> and abundance in nearly all species could be related to adaptation strategies, or the multitude of factors influencing the (relative) abundance of a species. Under high stress conditions,

a species may adapt by adopting a stabilizing selection strategy that allows only the most tolerant phenotype (in this case, size ranges) to thrive (van Valen, 1965; Weinkauf et al., 2019). As a result, total abundance of the species will be reduced, but its “survival size ranges” will persist and attain high relative abundances within the species even under sub-optimal conditions and at the expense of less-tolerant phenotypes (Hecht, 1976; Rillo et al., 2020). Furthermore, a species' abundance can be influenced by many biotic (competition, predation) and abiotic (temperature, salinity, oxygen, nutrients) factors within its living environment as well as by post-depositional dissolution that could impact the representation of a species in a sample (Hecht, 1976; Schiebel & Hemleben, 2017).

#### 4.5. Environmental Forcing of Planktonic Foraminiferal Size at the Species Level

Based on regression of temperature at maximum size with temperature at maximum relative abundance, Schmidt, Renaud, et al. (2004) proposed that mean annual SST primarily drives planktonic foraminifera size at the species level. Our results from the TIO show that SST did not primarily drive size changes in any of the species studied (see Table S6 in Supporting Information S1). Although the  $size_{95/5}$  of some species showed weak but significant relationships with SST, in all cases, the coefficients of determination were markedly lower than the parameters with which they showed the best fit (Table S5 in Supporting Information S1). In the global core-top study of Rillo et al. (2020), the size (log of shell area) of nine planktonic foraminifera species showed little to no response to SST and PP. This led to the conclusion that environmental parameters cannot consistently predict planktonic foraminifera shell size. Whilst results from our study support this general conclusion, they also reveal species-specific environmental controls on the size of some of the most abundant species, including *O. universa*, *G. bulloides*, *G. ruber albus*, *G. inflata*, and *G. glutinata* (Figure 9). In Figure 9, we binned the parameters (e.g., all temperature parameters, all carbonate chemistry parameters) to identify broad environmental controls on species' size distribution. Parameters related to carbonate ion concentration explain size variation in 35% of the species while parameters related to other factors such as temperature (25%), salinity (20%), nutrient concentration (10%), and oxygenation (10%) also explain some of the size variations in individual planktonic foraminiferal species (see Table S5 in Supporting Information S1).

Results from re-analysis of the Rillo et al. (2020) data set show that three of the nine species studied (*T. sacculifer*,  $R^2 = 0.22$ ,  $p = 0.03$ ; *G. truncatulinoides*,  $R^2 = 0.58$ ,  $p = 0.02$ ; and *G. inflata*,  $R^2 = 0.69$ ,  $p = 0.03$ ) showed significant relationships with SST in the TIO (see Figure S4 in Supporting Information S1). In this study, *T. sacculifer* ( $R^2 = 0.05$ ,  $p = 0.03$ ) and *G. inflata* ( $R^2 = 0.30$ ,  $p = 0.01$ ) were also found to show significant relationships with SST. Whether the high coefficients of determinations of the three species reported for the Rillo et al. (2020) data set would be realized when a higher number of samples are investigated remain in doubt because the number of samples per species in our study were significantly higher than theirs (62 vs. 14 for *T. sacculifer*, 27 vs. 9 for *G. truncatulinoides*, and 25 vs. 4 for *G. inflata*). Nevertheless, we argue that the relationships observed between the  $size_{95/5}$  of these three species (and the species observed to show similar relationships in our study) and SST indicate a possible secondary effect of SST on their size variations. Thus, two inferences can be drawn from the results of our study combined with those of Rillo et al. (2020). Regionally and globally, size variation in individual species is linked to multiple processes such as calcification (initial test growth), respiration, trophic strategy, and resource availability, and no single parameter can predict intra-specific size variation (Kontakiotis et al., 2021; Kuroyanagi et al., 2013; Rillo et al., 2020). In addition, while SST may affect the rates of the aforementioned processes differently among species (e.g., Lombard et al., 2009; Weinkauf et al., 2016), in tropical environments with predominantly warm SSTs, the net impact of SST on intra-specific size variation does not outweigh the impact of other physiochemical parameters. By inference, SST is not a suitable parameter for testing the OSH at the species level.

In our data set, the identified drivers of size in some species agree with described ecological and environmental preferences (Figure 9). For instance, phosphate concentration (an indicator of the nutritional landscape or “water fertility”) was found to be a significant driver of size in *G. bulloides* ( $R^2 = 0.31$ ,  $p = 2.4e-07$ ) and *G. glutinata* ( $R^2 = 0.19$ ,  $p = 2.94e-05$ ), and these species are known to thrive in nutrient-rich and upwelling environments (Bé & Hutson, 1977; Conan & Brummer, 2000; Ivanova et al., 2003; Kroon & Ganssen, 1988; Schiebel & Hemleben, 2017; Seears et al., 2012). Interestingly, calcification in *G. bulloides* in the North Atlantic has also been reported to be linked to phosphate concentration (Aldridge et al., 2012). Therefore, phosphate concentration may be key to providing optimum conditions for these species. Carbonate ion concentration was found to

significantly drive size variation in sub-surface species which bear gametogenic calcite such as *G. conglobatus* ( $R^2 = 0.19$ ,  $p = 0.01$ ), *G. tumida* ( $R^2 = 0.29$ ,  $p = 0.001$ ), and *P. obliquiloculata* ( $R^2 = 0.26$ ,  $p = 0.047$ ). On the other hand, tropical to sub-tropical species such as *G. ruber albus* and *T. sacculifer*, which are surface dwellers with a preference for warm water masses (Bé, 1977), showed no relationship with any temperature-related parameter. Using a small data set from the Caribbean Sea, Stone (1956) reported that test size in *O. universa* showed no relationship with SST while test size in *G. ruber albus* is weakly sensitive to SST compared to *T. sacculifer* which showed a higher sensitivity. In this study, similarly, size<sub>95/5</sub> variation in *O. universa* showed no sensitivity to SST ( $R^2 = 0.06$ ,  $p = 0.06$ ) and *T. sacculifer* showed more sensitivity to SST ( $R^2 = 0.05$ ,  $p = 0.03$ ), albeit weak. Meanwhile, *G. ruber albus* size<sub>95/5</sub> is not impacted by SST changes ( $R^2 = 0.02$ ,  $p = 0.15$ ) in this study, and its size related more to oxygenation. Our results suggest that, in the TIO, size<sub>95/5</sub> variation in *O. universa* is in part driven by sub-surface temperatures (at 100 m water depth;  $R^2 = 0.20$ ,  $p = 1.2e-05$ ) while size<sub>95/5</sub> in *T. sacculifer* is influenced by carbonate concentration ( $R^2 = 0.10$ ,  $p = 0.01$ ).

Culture experiments on two subtropical–transitional planktonic foraminifer species (*O. universa* and *G. bulloides*) show increased shell length under increasing dissolved oxygen conditions (Kuroyanagi et al., 2013). Symbiont-bearing *O. universa* not only increased in length but also in pore diameter and size under varying dissolved oxygen concentrations and low temperature variability. This may explain the positive correlation we found between size and oxygenation in *G. ruber albus* (also a symbiont-bearing species like *O. universa*) and we speculate that it may imply that under low temperature variability, growth in *G. ruber albus* could be linked to increased gas exchange even though some of its nutrients are derived from organic carbon supply due to enhanced photosynthetic activity of its symbionts (Jørgensen et al., 1985; LeKieffre et al., 2018).

Broadly, our results suggest that environmental parameters have limited but significant impacts on species-specific geographical size distribution in TIO planktonic foraminifera. Building upon the conclusions of Rillo et al. (2020), we infer that no single environmental parameter can consistently predict intra-specific size variations in planktonic foraminifera, but that specific environmental parameters might define each species' ecological optimum, supporting the OSH.

#### 4.6. Primary and Secondary Environmental Controls on Assemblage Size Trends

##### 4.6.1. Surface Carbonate Ion Concentration Exerts Primary Controls on Assemblage Size Trends

In a previous global study, SST was reported to primarily drive size changes in planktonic foraminifera assemblages (Schmidt, Renaud, et al., 2004). Our investigation of this hypothesis suggests that SST does not primarily drive assemblage size changes regionally in the TIO. The *F*-matrix loadings from the factor analysis on planktonic foraminifera assemblage size shows characteristic faunal aggregation on the first factor axis. The high weighting and aggregation of tropical and sub-tropical species such as *T. sacculifer*, *P. obliquiloculata*, *G. menardii*, *G. tumida*, *G. elongatus*, *N. dutertrei*, and *O. universa*, along Factor axis 1 of the assemblage size data makes it difficult to decipher a clear temperature-related size distribution (see Table S7 in Supporting Information S1). Likewise, the low weighting on species with preferences for nutrient rich environments (upwelling zones) such as *G. glutinata*, *G. falconensis*, and *G. bulloides* on this axis also rules out the possibility that this axis is related to resource availability. However, notably, large-sized and relatively more resistant species as defined by Berger (1970), have higher loadings on this axis. We note that the Berger (1970) solubility ranking included most species but not all. In addition, some of the species with high positive factor scores such as *P. obliquiloculata*, *G. conglobatus*, *G. menardii*, and *G. tumida* bear gametogenic calcite. Robust regression analysis between this first factor axis and hydrographic parameters revealed a best fit with surface carbonate ion concentration ( $R^2 = 0.41$ ,  $p = 1.18e-07$ ) (Figure 12a), consistent with the importance of surface carbonate ion concentration in the TIO revealed by our PCA of surface hydrographic parameters (Figure S1 in Supporting Information S1, Table 2). The weighting of different species on this axis might reflect the different sensitivities of thermocline/sub-thermocline species versus mixed layer species to carbonate ion concentration.

Our results imply that, at the assemblage level, upper water column carbonate ion concentration is significantly inversely correlated with assemblage size<sub>95/5</sub> in TIO planktonic foraminifera. While this result may suggest a possible influence of carbonate ion in driving growth and indirectly calcification at the assemblage level, the use of assemblage size as a proxy of calcification is not straightforward because it integrates all single-species responses, and their full life histories. More so, carbonate ion has only so far been reported to influence calcification at the

species level (Barker and Elderfield, 2002; de Moel et al., 2009; Henehan et al., 2017; Naik et al., 2011; Weinkauff et al., 2013), and it remains to be tested whether the same observation will apply at the assemblage level. Nevertheless, if one posits that the assemblage size<sub>95/5</sub> signal were linked to community calcification, our result yields a negative relationship compared to the positive relationship observed in foraminifera at the species level (Barker & Elderfield, 2002; Bijma et al., 1999, 2002). We propose two possible explanations for this observation. First, it might be that in assemblages in more supersaturated waters in the TIO, calcification carries less energetic costs to the foraminifera (i.e., reduced energetic costs of elevating pH at the calcification site) such that species (a) produce their full test more quickly and (b) have more energy to spend on other metabolic processes (e.g., reproduction); by reserving energy for such an activity, species complete their life or reproductive cycle more efficiently and quickly, but do so within the lower limits of their survival size ranges. This hypothesis cannot be tested using our data set, as further data including size-normalized shell weight or CT-scans of multiple species would be needed to fully understand the link between size, growth and calcification in each assemblage.

Second, the negative size to carbonate ion relationship observed may be linked to differing carbonate production patterns between upwelling and more oligotrophic environments. Upwelling environments have been reported to be characterized by high carbonate fluxes, lower surface carbonate ion concentration, and a dominance of small-sized foraminiferal species (e.g., *G. glutinata*, *G. bulloides*, and *N. incompta*) (Peeters & Brummer, 2002; Ramaswamy & Nair, 1994; Thunell et al., 2007). Conversely, oligotrophic environments are generally characterized by lower carbonate fluxes, higher surface carbonate ion concentration, and high relative abundance of larger-sized species (e.g., *O. universa*, *T. sacculifer*, and *G. siphonifera*) (Peeters & Brummer, 2002; Ramaswamy & Nair, 1994; Schiebel & Hemleben, 2017). We test this hypothesis through correlation and “correlation cluster” analyses to examine the relationship between the relative abundances of small-sized upwelling species and large-sized oligotrophic species versus surface carbonate ion concentration in the TIO (Figure S6 in Supporting Information S1). In our full data set, we find that higher relative abundances of small-sized upwelling species occur under higher surface carbonate ion concentration and that higher relative abundances of large-sized oligotrophic species occur under lower surface carbonate condition (Figures S6a and S6b in Supporting Information S1). This leads to an overall lower assemblage size<sub>95/5</sub> under high surface carbonate condition and a higher assemblage size<sub>95/5</sub> under low surface carbonate ion conditions. However, this result is sensitive to the inclusion of the Arabian Sea data set, and its exclusion leads to opposite results (i.e., high relative abundances of small-sized species under low surface carbonate ion conditions and vice versa) (Figures S6c–S6d and S7 in Supporting Information S1). Overall, in the TIO, our results show that surface carbonate ion concentration exerts primary control on assemblage size<sub>95/5</sub> variations, and we suggest that this response might be due to energetic optimization and/or changes in the dominance of the main species linked to nutrient regimes, though this pattern remains to be confirmed in other ocean basins.

#### 4.6.2. SST Exerts Secondary Controls on Assemblage Size Trends

A temperature effect on assemblage size might be expected for different reasons. First, previous studies have shown that test size decreases from the tropics toward the poles (de Villiers, 2004; Renaud & Schmidt, 2003). This implies that planktonic foraminifera assemblages are sensitive to temperature changes and as communities shift from the poles toward the equator, assemblage size<sub>95/5</sub> increases. Support for this is found in Schmidt, Renaud, et al. (2004) who suggested that SST primarily drives assemblage size changes in planktonic foraminifera on a global scale (Schmidt, Renaud, et al., 2004). Second, in fossil sediments and plankton net samples, temperature has been reported to influence shell calcification (shell weight and SNW) in planktonic foraminifera (Aldridge et al., 2012; Gonzalez-Mora et al., 2008). Third, the second axis of the PCA analysis on surface hydrographic parameters show that SST is an important hydrographic parameter in the TIO after surface carbonate ion concentration. We test this assumption by correlating our  $F2$  scores with hydrographic parameters.

On the second factor axis of the  $F$ -matrix loading of the assemblage size factor analysis, unlike the first factor axis, a clear niche partitioning is observed. Factor 2 is characterized by deep-dwellers including *G. inflata*, *G. crassaformis*, and *G. truncatulinoides* (see Table S7 in Supporting Information S1), suggesting a possible relationship with a temperature-related parameter based on the common thermal niche preference of these species. Assemblage size  $F2$  scores indeed showed the best relationship with SST ( $R^2 = 0.46$ ,  $p = 1.24e-08$ ) among the environmental variables tested (Figure 12b). In agreement with previous studies, our result shows that assemblage size increased with temperature (Schmidt, Renaud, et al., 2004). The influence of SST may be more pronounced on assemblage size<sub>95/5</sub> and not species' size<sub>95/5</sub> because the aggregate net effect of SST on assemblage calcification

processes becomes more evident when species composition is heterogenous. Therefore, based on our results, we suggest that ambient carbonate ion concentration primarily influences assemblage size<sub>95/5</sub>, but temperature plays a significant role in modulating the ontogenic calcification process. The influence of both carbonate ion concentration and temperature on assemblage size<sub>95/5</sub> (and not species' size<sub>95/5</sub>) has not been reported previously, thus our results constitute the first in this regard. Yet, while most species-specific studies on the factors that influence shell calcification have shown that carbonate ion concentration drives shell calcification (Barker & Elderfield, 2002; Bijma et al., 2002; Lombard et al., 2010; Spero et al., 1997), other studies have shown that shell calcification can be attributed to the influence of carbonate ion concentration and temperature (Aldridge et al., 2012; Gonzalez-Mora et al., 2008); our results support the latter. Therefore, we conclude that SST plays a secondary but significant role in driving planktonic foraminifera assemblage size<sub>95/5</sub> trends in the TIO.

## 5. Summary and Conclusions

In this study, through automated foraminiferal analysis, we test the OSH to determine whether relative abundance or environmental parameters are better indicators of optimum conditions in TIO planktonic foraminiferal species. We also investigate the dominant environmental controls on TIO planktonic foraminifera assemblage size<sub>95/5</sub> distribution. Combined, our high-throughput imaging machine as well as our CNN-trained model achieved classification and measurement accuracies similar to human experts. Although most species show unimodal SFD, we report flat to multimodal SFDs in larger species such as *G. menardii*, *G. conglomerata*, and *G. tumida*. We infer that vertical niche separation explains the relationship between planktonic foraminiferal species' size and diversity. Using environmental parameters rather than relative abundances as indicators of optimum conditions in planktonic foraminiferal species, we found support for the OSH. However, we report that the environmental drivers of size in planktonic foraminifera are species-specific, with parameters related to carbonate ion concentration and temperature exerting significant controls on species-level size<sub>95/5</sub> variations. Therefore, no single parameter can predict intra-specific size variation in planktonic foraminifera, as suggested by Rillo et al. (2020). Further, in contrast to previous reports that planktonic foraminifera assemblage size is primarily driven by temperature, results from this study suggest that, within tropical oceans where surface ocean temperature variability is restricted, carbonate ion concentration exerts the strongest control on planktonic foraminifera size variations, although temperature was found to exert a secondary control on assemblage size<sub>95/5</sub> variation. In support of previous work (Cayre et al., 1999) and as a validation of our automated method, we found that PP exerts primary control on TIO species assemblage distribution, but temperature was found to be a secondary driver of species assemblage distribution in the TIO. Although it was found to be a secondary driver, temperature influence on planktonic foraminiferal assemblage size variations and assemblage distribution is significant. Because of multicollinearity and complexity in relationships between environmental variables, we conclude that the isolated drivers of size<sub>95/5</sub> variation identified in this study act as reasonable proxies for a larger suite of drivers. Overall, if our findings apply to other tropical ocean planktonic foraminifera populations and communities, changes in planktonic foraminiferal test size variations are mostly linked to the effects of parameters related to carbonate ion and temperature.

## Data Availability Statement

Data sets associated with this paper are available in the SEANOE open-access database at <https://doi.org/10.17882/86211> (Adebayo, Bolton, Marchant, Bassinot, Sandrine, et al., 2022) and <https://doi.org/10.17882/86411> (Adebayo, Bolton, Marchant, Bassinot, Conrod, et al., 2022), published under a CC-BY-NC license. The scripts for the code used in this work can be accessed at <https://github.com/mikolinton>. The *ParticleTrieur* software used in this work is available with a tutorial from <http://particle-classification.readthedocs.io>, and the Python scripts to train the CNNs in this paper are available from <http://www.github.com/microfossil/particle-classification> and can be installed using the pip Python package.

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