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1 **Ligand fishing as a tool to screen natural products with anticancer potential**

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17 18 **ABSTRACT**

19 Cancer is the second leading cause of death in the world and its incidence is expected to
20 increase with the aging of the world's population and globalization of risk factors. Natural
21 products and their derivatives have provided a significant number of approved anticancer
22 drugs and the development of robust and selective screening assays for the identification of
23 lead anticancer natural products is essential in the challenge of developing personalized
24 targeted therapies tailored to the genetic and molecular characteristics of tumors. To this end,
25 the ligand fishing assay is a remarkable tool to rapidly and rigorously screen complex
26 matrices, such as plant extracts, for the isolation and identification of specific ligands that
27 bind to relevant pharmacological targets. In this paper, we review the application of ligand
28 fishing with cancer-related targets to screen natural product extracts for the isolation and
29 identification of selective ligands. We provide critical analysis of the system configurations,
30 targets, and key phytochemical classes related to the field of anticancer research. Based on
31 the data collected, ligand fishing emerges as a robust and powerful screening system for the
32 rapid discovery of new anticancer drugs from natural resources. It is currently an
33 underexplored strategy according to its considerable potential.

34 **Keywords:** Affinity chromatography, cancer, drug discovery, ligand fishing, natural
35 products

36
37 **List of abbreviations:** 2-AG, 2-arachidonylglycerol; AA, arachidonic acid; ACS, American
38 Chemical Society BDNF, brain-derived neurotrophic factor; CM, cell membrane; COXs,
39 cyclooxygenases; cPLA₂, cytosolic phospholipase 2; CTAC, hexadecyltrimethylammonium

chloride; CYP450, cytochrome P450; DAG, diacylglycerol; ECM, extracellular matrix; EGFR, epidermal growth factor receptor; FDA, Food and Drug Administration; FGF, fibroblast growth factor; FGFR4, fibroblast growth factor receptor 4; GO, graphene oxide; GSK-3 β , glycogen synthase kinase 3 β ; GST, glutathione-S-transferase; HF, hollow fiber; HSPs, heat shock proteins; IOMN, iron oxide magnetic nanoparticle; IP₃, inositol triphosphate; iPLA₂, calcium-independent phospholipase A₂; JAK2, Janus kinase 2; LDH, lactate dehydrogenase; LOXs, lipoxygenases; MAGL, monoacylglycerol lipase; MAPK, mitogen-activated protein kinase; MAPKAPK2, MAPK-activated protein kinase 2; MKK3, mitogen-activated protein kinase kinase 3; MMPs, matrix metalloproteinases; MPs, magnetic particles; NAD, nicotinamide adenine dinucleotide; NGF, nerve growth factor; OGO, oriented layered graphene oxide pad; PAF, platelet-activating factor; PAI-1, plasminogen activator inhibitor; PI3K, phosphatidylinositol 3 kinase; PGs, prostaglandins; PGE₂, prostaglandin E₂; PLA₂s, phospholipases A₂; PKC, protein kinase C; PLC, phospholipase C; QD, quantum dots; QR2, quinone reductase 2; SIRT, sirtuin; sPLA₂, secreted phospholipase A₂; STAT3, signal transducer and activator of transcription 3; TCMs, traditional Chinese medicines; TrkB receptor, tropomyosin-related kinase B receptor; UF, ultrafiltration; uPA, urokinase plasminogen activator; uPAR, urokinase plasminogen activator receptor; UHPLC-LTQ, ultra-high-performance liquid chromatography-linear trap quadrupole; UPLC-Q-TOF/MS, ultra-high performance liquid chromatography-quadrupole-time-of-flight mass spectrometry

1. Introduction

With 10 million deaths by 2020, corresponding to nearly one in six, cancer is the second leading cause of death in the world [1]. The most common cancers are breast, lung, colon, and prostate. The incidence of cancer is rapidly increasing worldwide because of the aging population, spread of viral and bacterial infections, pollution, and globalization of behavioral and dietary risk factors, including obesity, lack of exercise, tobacco and alcohol use, as well as low fruit and vegetable consumption [2].

Although many cancers can be cured if detected early and treated effectively, many patients still die for a variety of reasons, including lack of accessible treatments in poor countries, inability to complete treatment in patients at risk of heart or kidney failure, late diagnosis of metastatic tumors, natural and acquired resistance to treatments, anticancer drugs toxicity, and insensitivity of cancer stem cells to treatment with surgery, chemotherapy, radiation or immunotherapy [2–7]. Cancer cells develop multiple adaptative mechanisms to survive, become persistent against metabolic, hypoxic, nutritional, and therapeutic pressure, and escape detection and destruction by the immune system. Understanding these characteristics has led to major advances in the development of targeted drugs acting on the deregulation of cell metabolism, proliferative signaling, epigenetic reprogramming, cell cycle deregulation, tumor angiogenesis, and resistance to apoptosis and antitumor immunity [8]. Thus, cancer treatments are currently moving from standardized and validated protocols for each type of cancer to personalized protocols based on the genetic and molecular characteristics of tumors. From this perspective, the discovery of molecules with new modes

of action and the identification of key cellular pharmacological targets for the survival of cancer cells have opened the way to important therapeutic innovations [9].

Natural products have provided considerable value to the development of anticancer drugs because of their unique chemical scaffolds and original mechanisms of action. A variety of naturally derived agents are used in cancer chemotherapy, whether as active, preventive, or chemosensitizing drugs and the identification and evaluation of natural anticancer molecules is one of the most active areas of research in medicinal chemistry and biotechnology [10]. However, the screening and purification of natural products are often laborious, time-consuming, and expensive. In this sense, rapid and efficient methods need to be developed, especially for purifying bioactive compounds from complex matrices. In exploring these gaps, ligand fishing has emerged as an innovative, efficient, and specific high-throughput screening method designed to save research groups time and resources in the search for new chemical hits. This strategy consists in purifying a molecule with a selective affinity for a biological molecular target of therapeutic interest from a complex mixture of natural products obtained by extraction from biomass. To date, a large number of ligand fishing techniques have been developed with various protocols and efficiently optimized to identify ligands from plant, animal, or bacterial extracts. Combined with highly sensitive high-resolution analytical methods (e.g., LC-HRMS and NMR), ligand fishing allows for the rapid identification and purification of fished ligands, accelerating the pharmacological evaluation and characterization of bioactive natural products [11–13].

In this paper, we aimed to propose an exhaustive updated review and critical analysis of the ligand fishing technique as an effective screening strategy to identify natural products with anticancer potential. We reviewed the contribution of this technique to the identification of anticancer natural products in various biological extracts, discussed the different technical approaches developed and explored its gaps and possible improvements.

2. Materials and Methods

2.1 Search strategy

The present systematic review was performed based on original reports published in the scientific literature from all years until the 1st of October 2022. The scientific databases chosen for this work were Science Direct, PubMed, Google Scholar, Scopus, and Web of Science, using the following descriptors: “ligand fishing”, “natural products” and “cancer”. The obtained results were filtered to remain with research articles only. Moreover, we did not contact authors to retrieve unpublished data or unavailable full texts.

2.2 Inclusion/exclusion criteria

Papers were included in this study if the authors used ligand fishing to screen natural products, if the authors developed a proof of concept with known ligands to rationalize the method, and if the target was directly related to cancer pathophysiology. Only articles published in English and that met the inclusion criteria described above were selected for further evaluation. Manuscripts that did not satisfy the inclusion criteria, reviews, meta-

122 analysis, abstracts, conferences, editorials, letters, case reports, thesis, conferences, and
123 manuscripts with full text not available were excluded from the present review.

124 Two independent investigators (L.A.M.S. Duarte-Filho and P.C.O. Oliveira) were in
125 charge to select the articles evaluating the title, abstract and full text. All retrieved articles
126 were thoroughly examined in an attempt to identify the ones that did not fit into the inclusion
127 criteria properly. In case of divergence in the selection of manuscripts, two additional expert
128 reviewers were consulted (L. Picot and M.C. Moraes). Manually added papers were also
129 retrieved from the references of the articles selected through our systematic search.

130 *2.3 Data Collection*

131 All data were submitted to standardized forms to comprehensively collect information
132 about the country and year of publication, diversity of ligand fishing techniques and
133 experimental conditions, cancer targets and their respective cancer types, natural products,
134 identified ligands, and their chemical classes, as well as pre-clinical assessments (when
135 applicable). Furthermore, the selected data here presented was verified based on the expertise
136 of the authors and summarized into figures, tables, and diagrams for facilitated
137 comprehension of the numerical results.

138 **3. Results and Discussion**

139 **3.1. Bibliometric data**

140 The initial systematic search produced 569 results, of which 175 were from Science Direct,
141 19 from PubMed, 242 from Google Scholar, 118 from Scopus, and 15 from Web of Science.
142 One hundred and eleven articles were excluded because of duplicity. The remaining 458
143 articles were screened and 403 were excluded after reading of the title and abstract.
144 Afterward, we remained with 55 articles for a full examination, of which 3 could not be
145 retrieved and 17 were excluded due to divergencies with the inclusion criteria previously set.
146 In addition, 32 studies from reference lists of selected articles were manually added, of which
147 1 could not be retrieved and 8 were excluded after full-text analysis. At last, the total number
148 of articles selected for the present review was 58. The works included in the present review
149 were sorted according to the country where the research was developed, year of publication,
150 ligand fishing technique, anticancer targets, and respective cancer types. Additionally, details
151 concerning the natural product (family, species, parts used, ligands fished, and chemical
152 class) as well as the type of extraction, solvents, and buffers employed were also gathered
153 (**Tables S1** and **S2** respectively).

154 According to our data, approximately 83% of the selected works (n=48) were from
155 China, 5.1% from the USA (n=3), 3.4% from a cooperation China-USA (n=2), 3.4% from
156 China-South Korea (n=2), 1.7% from USA-Turkey (n=1), 1.7% from USA-Taiwan (n=1)
157 and 1.7% from Brazil (n=1), as shown in **Figure 1**. Considering the year of publication, the
158 first seven years (2011-2017) account for 36.2% (n=21) of the works published. On the other
159 hand, publications from the last four years (2018-2022) account for 63.8% of the included
160 articles (n=37) (**Figure 2**).

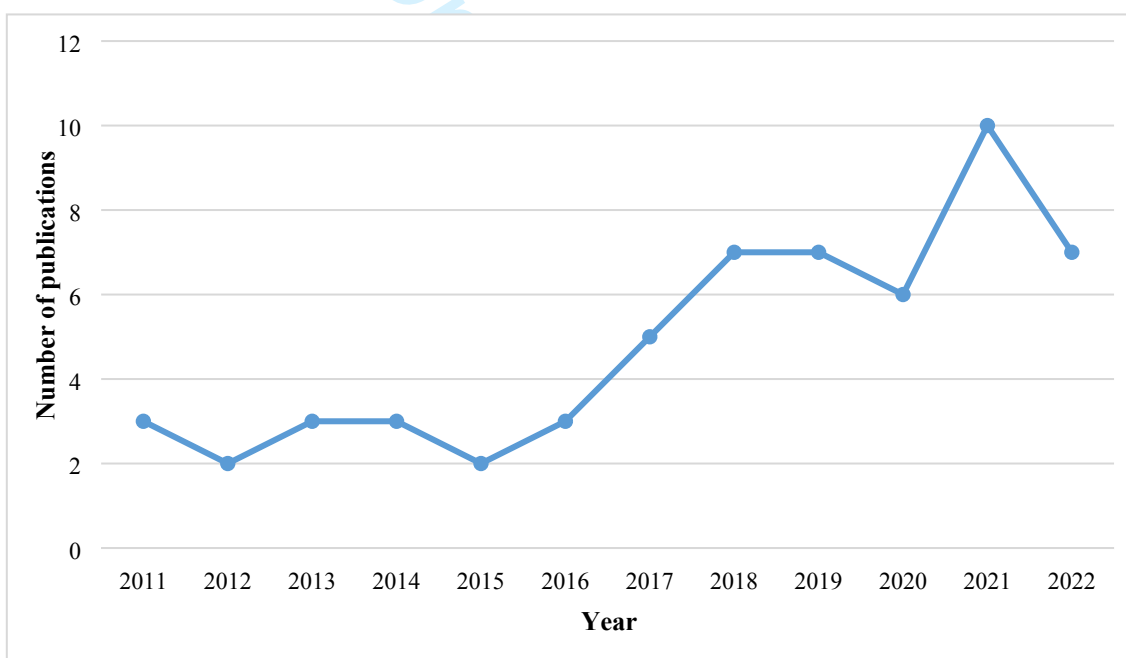
161

162 Figure 1 – Geographical distribution of selected studies



163

164 Additional assessments of the fished ligands were also part of the investigations. In
 165 this regard, more than two-thirds (n=40 – 69%) of the selected articles performed *in vitro*
 166 assays in order to investigate the enzymatic inhibition and/or *in vitro* anticancer potential of
 167 the fished-out ligands.



168

169 Figure 2 – Evolution of the annual number of world publications dealing with the
 170 identification of anticancer natural products using a ligand fishing strategy.

171 3.2. Diversity of ligand fishing techniques developed to isolate anticancer natural 172 products

173 According to the numbers shown in Figure 2, ligand fishing is a technique that has
 174 raised increasing interest in the last years and is gaining more attention from the scientific
 175 community as a versatile and powerful purification assay. The possibility of using a complex
 176 matrix with minimal sample preparation represents one of its major advantages, as it results
 177 in an economy of time and resources. Various techniques to bind ligands and separate bound
 178 ligands interacting with the molecular target from unbound ligands have been developed. The
 179 macromolecular target may be free in solution or immobilized on solid supports, and bound
 180 ligands may be finally dissociated from the complex matrix using a variety of techniques

181 such as ultrafiltration, centrifugation, or magnetic separation. Although immobilized targets
 182 usually demand more preparation time and reagents, they also provide better chemical and
 183 thermal stability and, especially, reusability for the system [14,15]. The use of solid support
 184 opens many possibilities and the diversity of immobilization chemistry and methodology
 185 results in a wide range of protocols and system configurations [16]. However, despite the
 186 benefits, immobilization is not mandatory for the ligand fishing assay.

187

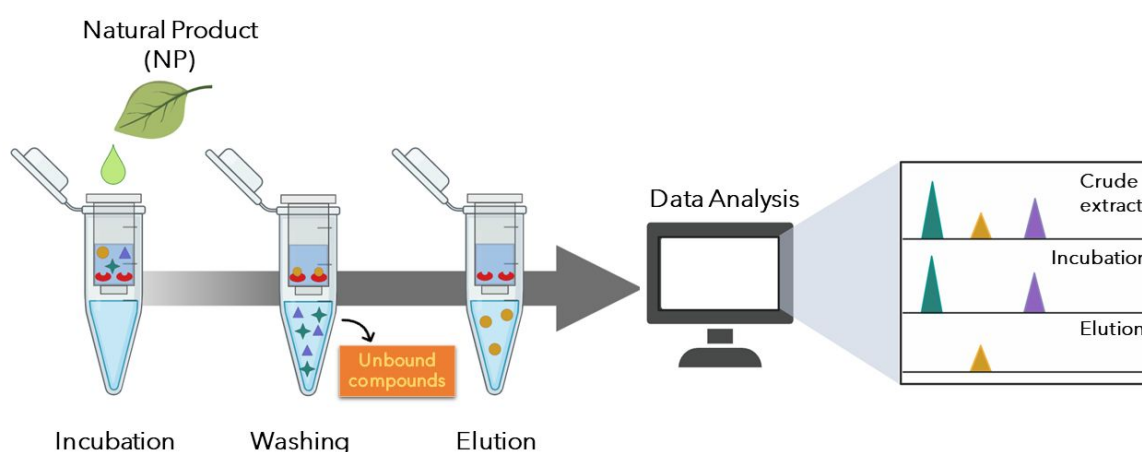
188 3.2.1. Free biological target

189 Ligand fishing methodologies that use the free biological target in solution require a
 190 step to recover the ligand-target complex from the solution. The isolation of the complex is
 191 based on the difference in size or mass of the complex with the remaining elements of the
 192 complex matrix. In the literature, techniques such as ultrafiltration and centrifugation are
 193 often used to separate the components by differences in size and mass.

194

195 *3.2.1.1. Ultrafiltration*

196 Ultrafiltration (UF) membranes can selectively retain macromolecules and
 197 consequently any ligand-target complex with a molecular weight higher than the pore size,
 198 but not small substances (although non-specific interactions between the UF membrane and
 199 small compounds may occur through electrostatic or low energy non-covalent forces) as
 200 shown by Figure 3. In this sense, it is possible to use the target in its free form, eliminating
 201 all the steps required for biomolecules immobilization, but more importantly, the UF
 202 membranes usually present weak interaction with the components of a mixture, so the ligands
 203 with no affinity for the target will readily cross this physical barrier [13,17] The necessity to
 204 perform several washes to completely remove the unbound components is considered a
 205 drawback of this technique, as well as the need for high pressure to eliminate non-targeted
 206 molecules. Therefore, the washing steps are important to avoid retention by non-specific
 207 interactions.



208 Figure 3 – Schematic illustration of ligand fishing assay using ultrafiltration membrane. In
 209 this configuration, it is used a physical barrier (membrane) to retain the target. As a result,
 210 the ligand-target complex can be isolated, and the washing of weakly bound ligands and
 211 elution of ligands with strong affinity can be promptly executed.

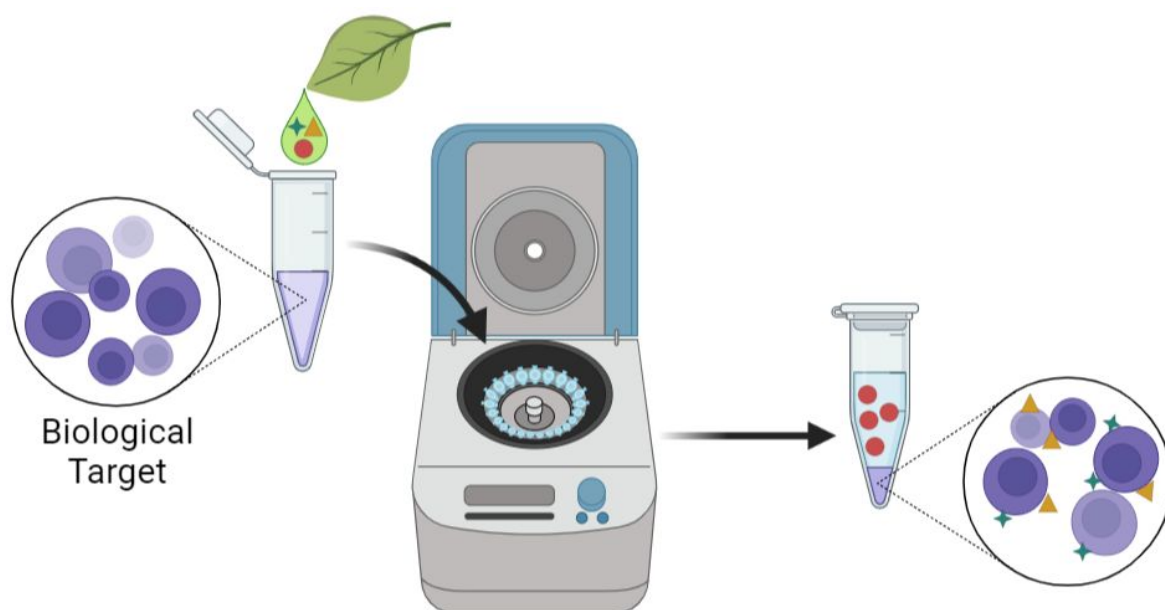
1
2
3 212 In two different works, *Glycyrrhiza uralensis* Fisch. root and mango (*Mangifera*
4 213 *indica* L.) leaves extracts were screened for tyrosinase inhibitors using the ligand fishing
5 214 assay associated with the UF technique [18,19]. Liu et al. [18] compared the performance of
6 215 UF and magnetic support to identify ligands in the *G. uralensis* Fisch. extract. Even though
7 216 both techniques were able to recognize the same ligands, it was observed different responses
8 217 for each assay. Comparing methodologies, the two presented easy operation, however, the
9 218 UF process demanded more time (more than 1 hour) while the usage of magnetic particles
10 219 resulted in a 20-min assay. UF is usually presented as a fast technique, however, this time
11 220 can be shortened when considering other techniques. The authors argued that this method has
12 221 easy operation and fast response with few parameters to adjust [18]. Ma and collaborators
13 222 [12] applied UF to identify DNA-binding substances from several herbal extracts. After
14 223 evaluating different membranes and conditions, they chose a modified poly (ether sulfone)
15 224 membrane that was able to retain the target and operated under a variety of eluting solvents.
16 225 Once established, this UF affinity assay was evaluated and it was able to fish out 10 ligands
17 226 (berberine, coptisine, palmitine, daurisolone, dauricine, fangchinoline, tetrandrine,
18 227 actinomycin D, actinomycin V, and actinomycin X0 β) proving to be a reliable method for
19 228 the recognition of potential ligands in a complex matrix.

229 3.2.1.2. Centrifugation

230 Centrifugation is particularly useful for suspensions that contain components with
231 large differences in density and sedimentation time, such as macromolecules (cells, DNA,
232 etc.) versus low molecular weight molecules. This property is thus particularly tailored to the
233 separation of ligands bound on target macromolecules from unbound molecules (Figure 4).

234 To explore *Bufo melanostictus* Schneider venom as a source of anti-cancer
235 components, Ren et al. [11] used a simple procedure to perform the ligand fishing assay. In
236 this work, the biological targets were HepG2 and MCF-7 cells, and the affinity assay was
237 conducted in the cell culture flask, where the growth medium was replaced by a toad venom
238 solution. After incubation, the cells were washed and centrifugated and the venom molecules
239 bound to the cells were eluted using methanol as organic solvent. Comparison of the raw toad
240 venom composition with the eluted compounds, using UPLC-MS analysis, allowed them to
241 identify the toxins that interacted with the cellular targets [11]. Similarly, Wei and
242 collaborators [20] performed this cell affinity-based assay to screen bioactive components in
243 a *Curcuma longa* L. extract. Using centrifugation, PC3 cells incubated with the extract were
244 decanted and separated from unbound molecules and washed to elute the ligands. After
245 concentration and analysis by UHPLC-LTQ Orbitrap MS, three curcuminoids (curcumin,
246 demethoxycurcumin, and bisdemethoxycurcumin) with significant anticancer activity were
247 identified [20].

248



249

250 Figure 4 – Ligand fishing assay using centrifugation as a method to isolate the
 251 macromolecule and unbound ligands (low-mass fraction) through the action of centripetal
 252 force. Therefore, the supernatant can be easily retrieved, and the ligand-target complex can
 253 proceed to the next steps (washing and elution).

254 3.2.2. Grafting of the biological target on a solid support

255 Solid supports are usually used for enzyme immobilization, and a large panel of
 256 materials and strategies can be used. Amino acids present in the macromolecule can be used
 257 for the formation of covalent bonds between the support and the target. Electrostatic forces
 258 and weak interactions, such as Van der Waals, can also promote immobilization. The solid
 259 supports can be classified as inorganic, such as silica, metals, metallic oxides, and carbon, or
 260 organic like natural and synthetic polymers [15,21].

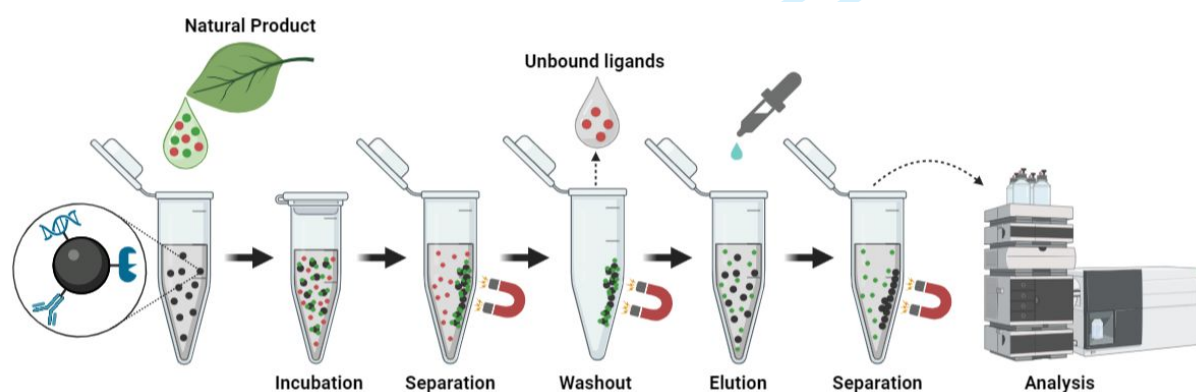
261 *3.2.2.1. Magnetic particles*

262 The convenience of this technique resides in the superparamagnetic properties of the
 263 particles, which allow the practical and fast isolation of interesting ligands using an external
 264 electromagnetic field applied by a simple magnet. These characteristic conserves the
 265 colloidal stability of nano-sized magnetic particles since they do not present remaining
 266 magnetization after the removal of the external field, theoretically avoiding agglomeration
 267 [22]. Beyond superparamagnetic properties, the surface functionalization of magnetic
 268 particles confers versatile chemical features to this material that are responsible for its wide
 269 range of applications [23–26].

270 Iron oxide magnetic nanoparticles (IOMN) have multiple applications in biomedical
 271 sciences due to their unique physicochemical properties. However, their chemical reactivity
 272 and high surface energy also represent two important issues able to reduce the
 273 superparamagnetic properties of this material. In this sense, IOMN silica-coating is an
 274 interesting procedure to enhance the chemical stability and reduce agglomeration and
 275 eventual cytotoxicity of this material [27]. In a sophisticated experiment, Yi et al. [28]
 276 fabricated a polydopamine-functionalized silica-coated iron oxide mesoporous magnetic
 277 particle meant to immobilize glutathione-S-transferase (GST) on its surface. The authors
 278 used a hexadecyltrimethylammonium chloride (CTAC) template to create mesopores in the

279 silica-coated IOMN, aiming to screen different plant extracts (*P. frutescens* (L.) Britt leaves,
 280 *A. orientale* (Sam.) Juz. roots, and *C. sinensis* (L.) Kuntze extracts). As a result, they
 281 successfully fished and identified six compounds, of which two ((-)-Epigallocatechin-3-O-
 282 gallate and (-)-Epicatechin-3-gallate) presented IC₅₀ values for (GST) lower than 100µM
 283 [28]. It is important to note that mesoporous silica offers a larger surface area in comparison
 284 to ordinary silica coating, allowing the attachment of more functional groups to the
 285 nanocomposite, hence increasing fishing yields. [29].

286 Although several techniques are available for protein immobilization on the surface
 287 of magnetic particles, most of these approaches are not adequate to immobilize
 288 transmembrane receptors, which in turn are the targets of more than one-third of all approved
 289 drugs [30,31]. Indeed, to bind relevant bioactive ligands, these receptors must keep their
 290 physiological tridimensional structure to expose functional binding domains, and their
 291 purification and grafting on magnetic particles may lead to denaturation. Exploring this
 292 limitation, three works developed cell membrane-coated magnetic nanoparticles
 293 overexpressing certain types of receptors involved in cancer pathophysiology. Arituluk and
 294 colleagues [32] engineered SH-SY5Y neuroblastoma cells overexpressing tropomyosin-
 295 related kinase B (TrkB) receptors to encapsulate IOMN and screen Gotu kola (*Centella*
 296 *asiatica* (L.) Urban) plant extract seeking ligands. Since the TrkB receptor is a
 297 transmembrane integral protein and its functionality depends greatly on the integrity of the
 298 phospholipids in its vicinity, using TrkB-enriched membrane fragments instead of purified
 299 TrkB receptors represents a very interesting strategy to simulate more realistic conditions.
 300 However, some validated tyrosine kinase receptor inhibitors, such as sunitinib and
 301 nintedanib, act through the receptor's intracellular domains, which in turn are unlikely to be
 302 accessible in this fishing system since most of the intracellular space is occupied by the
 303 magnetic particles [33,34].

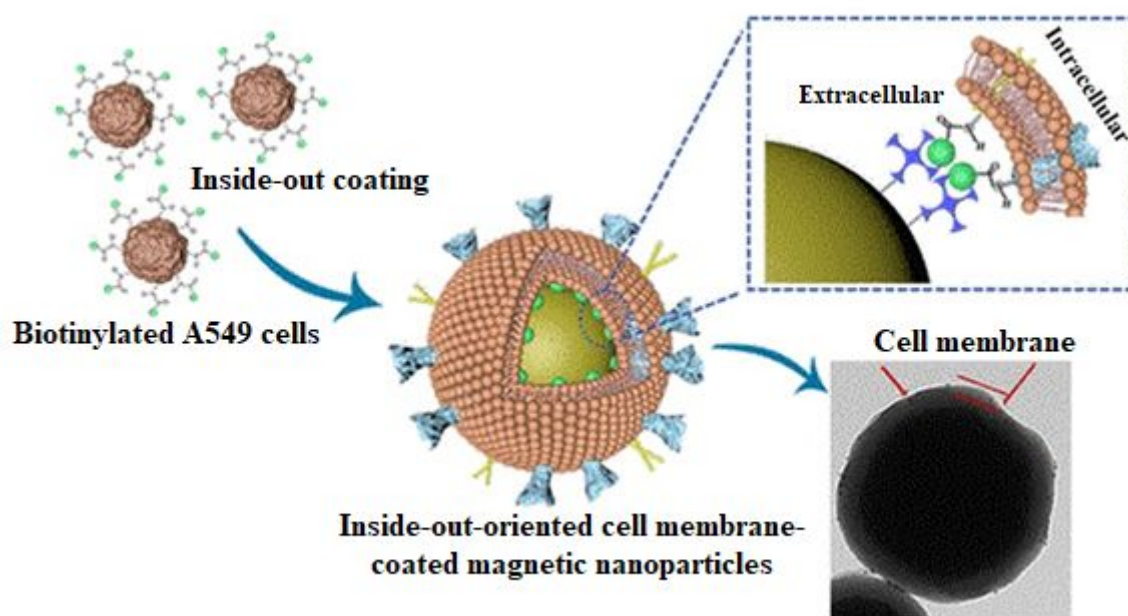


304 Figure 5 – Ligand fishing using magnetic particles (MPs). The MPs can be used to
 305 immobilize a variety of targets such as DNA, antibodies, and other proteins. After successful
 306 immobilization, MPs are incubated with the natural product. By affinity, the green compound
 307 is adsorbed by MPs whereas the red compound remains unbound. With the aid of an external
 308 magnetic field, the separation step occurs. In it, the complex MPs-green compound is retained
 309 inside the tube whereas the unbound red compound is washed out. The washout step is
 310 repeated multiple times to mitigate unspecific binding. Then, the complex MPs-green

311 compound is eluted using an organic solvent. Finally, a magnetic field is applied once again
 312 to recover MPs whereas the green compound-containing supernatant is brought to analysis.

313 In another study, Bu and collaborators [35] addressed this limitation. They
 314 functionalized carboxy-terminated iron oxide nanoparticles with streptavidin and
 315 biotinylated human lung adenocarcinoma epithelial (A549) cell fragments overexpressing
 316 the epidermal growth factor receptor (EGFR) to obtain inside-out-oriented cell membrane
 317 magnetic nanoparticles. With this system, the authors intended to fish out ligands from
 318 *Strychnos nux-vomica* L. extract that act in the intracellular domains of this receptor. As one
 319 of the negative controls, they used outside-out-oriented magnetic nanoparticles and observed
 320 that erlotinib and gefitinib, two known EGFR inhibitors, were more efficiently fished out by
 321 the inside-out-oriented magnetic nanoparticles in comparison to the outside-side-out one,
 322 highlighting the importance of transmembrane receptors binding site accessibility. Figure 6
 323 shows the orientation of the cell membrane concerning the encapsulated magnetic
 324 nanoparticle.

325 The fabrication and functionalization of magnetic particles can be diversified and
 326 adapted to confer desired physicochemical properties to this material. However, controlling
 327 parameters such as particle size, shape, stability, and dispersibility in specific solvents is
 328 often a challenging task [36] Furthermore, the characterization of nanomaterials involves
 329 expensive analytical techniques and equipment, which in turn represent a bottleneck to the
 330 study of such materials [37]. In this regard, commercially available functionalized magnetic
 331 beads represent a feasible alternative, as explored by Yasuda et al. and Singh et al. [38,39]



332
 333 Figure 6 – Representation of the inside-out-oriented cell membrane-coated magnetic
 334 nanoparticle. Reprinted (adapted) with permission from [35]. Copyright 2021 American
 335 Chemical Society (ACS).

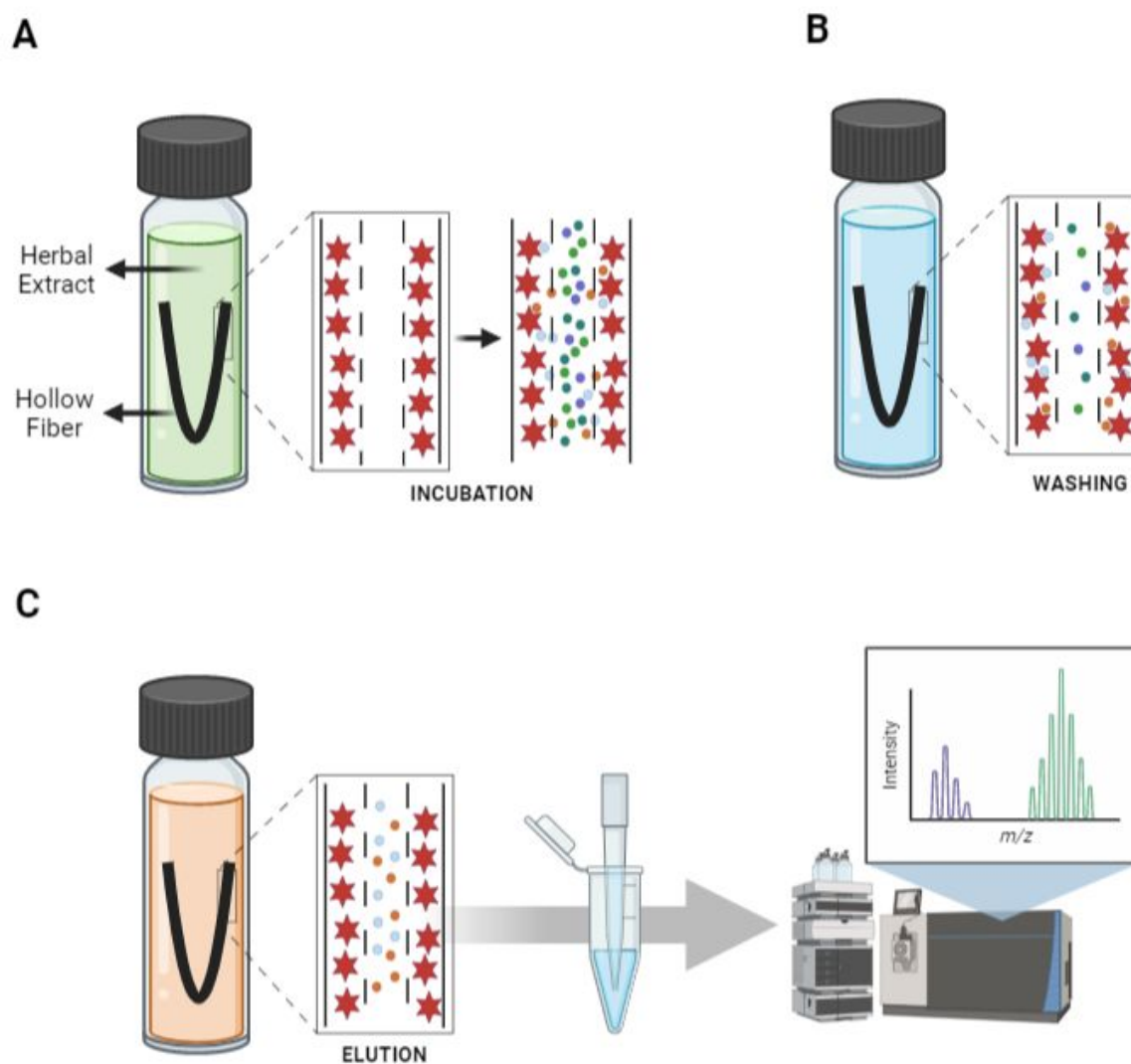
336 Based on the above-mentioned, magnetic particles represent versatile and very
 337 efficient supports for ligand fishing assays, able to be modified, adapted, and associated with
 338 different approaches in order to meet realistic physiological conditions depending on the
 339 target and its location inside the body.

340

341 *3.2.2.2. Hollow Fiber*

342 The hollow fibers (HF) are organic polymeric membranes with pore diameter and
343 inner cavities that operate as physical barriers capable to entrap the biological target, in a
344 similar way to ultrafiltration membranes. Hollow fibers have been used as a carrier for
345 proteins, cells, and liposomes [40,41]. The main characteristics of HF include its
346 biocompatibility, large surface area, high preconcentration capacity, and good recovery.
347 Additionally, this approach has an easy operation and fast response. The ligand fishing assay
348 that uses HF (Figure 7) requires usual laboratory instrumentation, however, as in other
349 techniques, false positives are problematic. For this reason, the washing solvent is a critical
350 part of the assay.

351 Zhao et al. [42] developed an efficient approach based on adsorbed hollow fiber to
352 screen and identify potential tyrosinase inhibitors in *Pueraria lobata* (Willd.) Ohwi extract.
353 Seven tyrosinase ligands were fished out in this study, and three compounds had their
354 inhibitory effect confirmed by *in vitro* assays. In 2019, Li and collaborators [43] used hollow
355 fiber to screen a series of coumarins and *Angelicae dahurica* Fisch. extract against human
356 hepatoma and renal tubular cells. The same system was applied to different sample matrices.
357 The first application was to screen the bioactive components directly from the plant extract,
358 as an *in vitro* approach. In the second study, the same herbal extract was orally administered
359 to rats. After five hours, abdominal aortal blood and rats' kidney and liver tissues were
360 collected and submitted to the HF fishing assay. In this *in vivo* assay, the authors aimed to
361 track which substances were able to reach these target organs. For all applications the hollow
362 fiber affinity-based assay was successful in the recognition and isolation of binders,
363 indicating the versatility of this method [43].



364
365 Figure 7 – Ligand fishing using hollow fiber (A: Incubation step - the biotarget is trapped
366 inside de fiber allowing diffusion and interaction of ligands; B: Washing step: the hollow
367 fiber was removed from extract solution and cleaned to remove unbound substances; C:
368 Elution step: the ligand-target complex is disrupted, the bound ligands are eluted and
369 proceeded to analysis)

370 371 3.2.2.3 Other immobilization techniques

372 This topic presents distinct systems for screening bioactive components from natural
373 products with anticancer potential, which from now on will be referred to as miscellaneous.
374 The application of different approaches allows the study of new characteristics in the
375 development of ligand fishing assays. In this regard, the majority of ligand fishing studies
376 here discussed used innovative configurations, supports, or immobilization approaches. In
377 three insightful studies, the Heat shock protein 90 (Hsp 90) was grafted in mesoporous silica
378 beads associated with InP/ZnS [44,45] and CdTe [46] quantum dots (QD). The support used
379 combines the feature of *in-situ* imaging through the fluorescence response of QD with the
380 ligand fishing technique. The QD was used as fluorescent probes in an imaging assay, this
381 way the ligand-target complex can be directly tested in a cell-based assay. Using this support
382 the authors propose not only to screen and identify potential binders by ligand fishing assay
383 but also to assess the biological activity of fished ligands on HeLa and MCF-7 cells. The
384 application of these silica nanocomposites was successful in the screening of potential
385 inhibitors of Hsp 90 on *Alisma plantago-aquatica* L., *Tripterygium Wilfordii* Hook. f., and

1
2
3 386 *Curcuma longa* L., in which, for the first time, four ligands (zingiberenol, (6R,7R)-
4 387 bisabolone, (*E*)-Atlantone and 6-(1-Hydro-xymethylvinyl)-4,8a-dimethyl-3,5,6,7,8,8a-
5 388 hexahydro-1H-naphthalen-2-one) were associated to this extract.

8 389 Another example includes the use of biotinylated triplex DNA on agarose beads. In
9 this approach, Xu and collaborators [47] used affinity immobilization of the target. They
10 390 proposed a screening of a known collection of small molecules to validate the methodology
11 391 and applied it to search for ligands in the *Phellodendron chinense* Schneid cortexes. The
12 392 developed screening assay was able to fish two components (berberine and palmatine) from
13 393 the complex mixture. Even though carbon-based materials have numerous applications, Zhou
14 394 et al. [48] state that this is the first time that graphene oxide (GO)/oriented layered graphene
15 395 oxide pad (OGO) is used as solid support for the immobilization of cell membranes (CM) in
16 396 ligand fishing assay. In the search for EGFR inhibitors, CM from H1819 and A549 cells were
17 397 selected to coat GO/OGO. This system was used to screen potential ligands from the Comfrey
18 398 rhizome extract (*Lithospermum erythrorhizon* Siebold & Zucc.). During the evaluation, OGO
19 399 presented a better performance than GO with a higher target load and better stability. The
20 400 H1819-CM@OGO proved to be a better target for fishing EGFR inhibitors. The developed
21 401 platform successfully fished three components from the extract (salvianolic acid B,
22 402 deoxyshikonin, and shikonin) that showed a strong biological response against H1819 cells
23 403 [48].
24 404

25 405 In order to better understand the well-known anticancer activity of a Traditional
26 406 Chinese Medicine (*Tripterygium wilfordii* Hook. f.), Tang and colleagues [51] used a
27 407 singular approach to uncover more information regarding bioactive components and
28 408 mechanism of action. In this work, the authors synthesized nanoparticles of CaCO₃ that were
29 409 covered by MCF-7 cell membranes. The exposed receptors in the cell membrane were
30 410 available for interaction with potential ligands in the plant extract. The differential of this
31 411 solid support resides in its capacity to hydrolase in an acidic medium. This feature permits
32 412 the support's removal preserving the target and the release of the bound ligands. This affinity
33 413 assay fished out seven ligands from the *T. wilfordii* extract. The most potent fished ligand
34 414 (celastrol) was used in the second application of the CaCO₃ nanoparticles - a target protein
35 415 fishing assay. For this assay, the celastrol was fixed into the support surface and it was used
36 416 to screen the protein collection from the MCF-7 cells digestion. The novelty in this
37 417 bioanalytical method is the use of the same support for both ligand and protein fishing, which
38 418 the authors named as a none-loss target release. Therefore, besides the screening of bioactive
39 419 substances in complex mixtures this strategy could be used to fish potential protein targets
40 420 [51]

As a summary of the works here discussed, we observed that most of them (n=20 – 34.5%) used miscellaneous approaches to screen natural products. Additionally, magnetic particles (n=17 – 29.3%), ultrafiltration (n=12 – 20.7%), hollow fiber (n=8 – 13.8%), and dialysis (n=1 – 1.7%) were also performed in an attempt to find anticancer leads from natural products (**figure 8**).

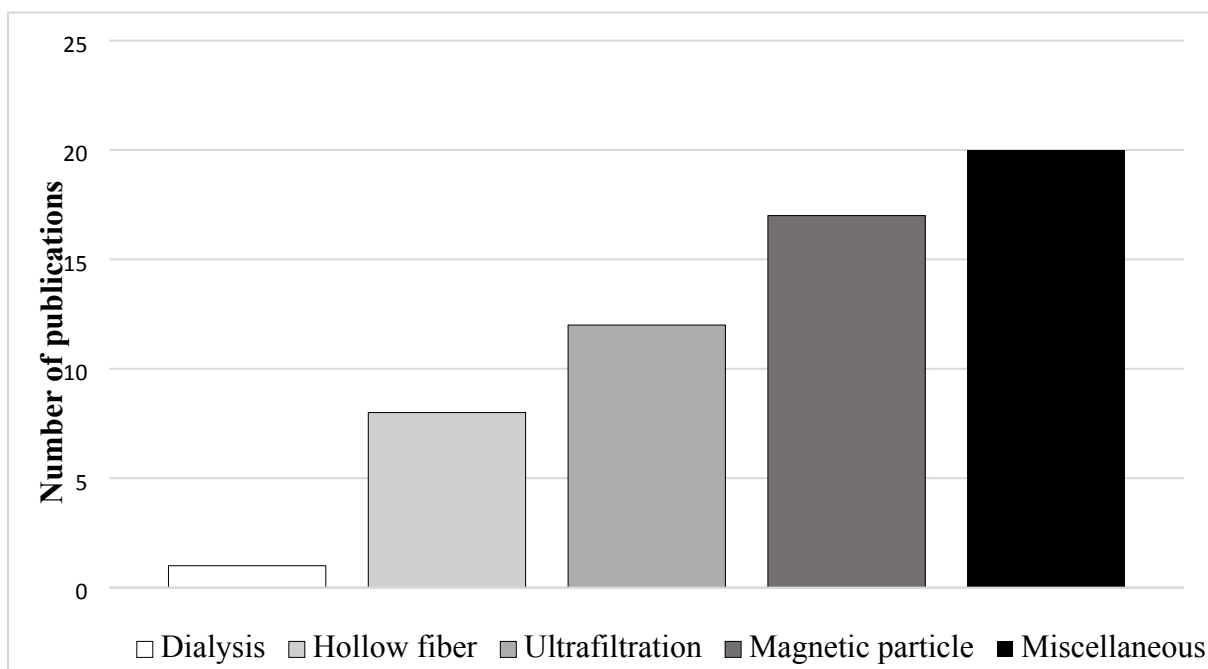


Figure 8 – Number of articles sorted by ligand fishing technique. Each bar represents the number of works included in each category.

As discussed in this section, the ligand fishing assays can be presented in different system configurations. Most of the works analyzed in this review fit the offline definition. This means that the isolation and identification of potential ligands are performed in two independent and physically disconnected steps [52]. Even though this configuration has proven to be useful, it is important to mention that there are different approaches to perform this affinity assay. Online assays are the ones that automatically insert the samples into the system, where the analyst has minimal interference. In more detail, this technique uses a bypass to transfer aliquots from one line to another [52,53]. Besides avoiding errors from preparation due to the automated transferring of samples and aliquots, this continuous flow can represent an economy of time and resources. These advantages are especially relevant in the screening of small biologically active compounds. To the best of our knowledge, no articles were found applying online ligand fishing using cancer-related targets to screen bioactive substances. Certainly, each assay has benefits and limitations but the fact that there is a gap in the literature for this type of system configuration indicates that there is still a lot to explore.

3.3. Anticancer natural products identified by ligand fishing

Table S1 presents the list of natural products identified from various natural sources as binders of specific molecular or cellular targets dysregulated in different cancers, using the ligand fishing strategy. For each molecule identified, the chemical class is indicated. Analysis of the selected works showed that flavonoids (n=23), alkaloids (n=17), and phenolic

1
2
3 449 acids (n=13) were the most fished secondary metabolites, followed by terpenoids (n=11),
4
5 450 coumarins (n=5), anthraquinones (n=5), and steroids (n=4). Ligands from other chemical
6
7 451 classes (i.e., saponins, tannins, lignans, etc.) were fished in 16 works (**figure 9A**).
8
9 452 Unsurprisingly, these classes have been extensively studied for their anticancer potential,
10
11 453 which allowed the discovery of remarkable anticancer compounds such as vinca alkaloids,
12
13 454 camptothecin, and paclitaxel [54–61]

14 455 Flavonoids are one of the largest classes of secondary metabolites, corresponding to
15
16 456 about 60% of all natural polyphenols, with approximately 8000 molecules [62,63]. Their
17
18 457 biological activity is usually attributed to a high antioxidant power [64], however, it is also
19
20 458 well known that flavonoids can act on several pathways, including some involved with cancer
21
22 459 pathophysiology. For example, *in vitro* studies have demonstrated that flavonoids can
23
24 460 modulate several protein kinases, epidermal growth factor receptors, vascular endothelial
25
26 461 growth factor receptors, and cytoskeleton dynamics, besides inhibiting some metabolizing
27
28 462 enzymes that activate a large number of pro-carcinogens [61,65-67]. In total, 96 flavonoids
29
30 463 from 23 works were identified as binders of molecular targets related to cancer,
31
32 464 demonstrating the potency of this technique to rapidly screen and identify flavonoids with
33
34 465 anticancer potential. Among them, polymethoxylated flavones are interesting molecules that
35
36 466 have been intensively studied for their anticancer potential. These natural products are
37
38 467 versatile pharmacological agents that can modulate different pathways involved with cell
39
40 468 proliferation, apoptosis, and cytoskeleton dynamics [68-70]. Moreover, Cheng and
41
42 469 collaborators [71] for instance have identified for the first time eight polymethoxylated
43
44 470 flavones in the fruit peels of *Citrus reticulata* Blanco that act as inhibitors of the matrix
45
46 471 metalloproteinase 2. It is important to note that although *C. reticulata* is a well-studied plant
47
48 472 species, these ligands were only detected after the implementation of a ligand fishing
49
50 473 approach, which unveils the power of the technique in identifying undiscovered compounds.

51
52 474 The fact that flavonoids are abundant in nature and act by several mechanisms may
53
54 475 explain why this class has been the most fished one among the selected articles. Quercetin
55
56 476 and kaempferol, for example, are known to exhibit a pleiotropic enzymatic inhibition activity.
57
58 477 It should also be noted that flavonoids can aggregate and form colloidal suspensions in *in*
59
60 478 *vitro* assays, with a high potential to unspecifically bind and inhibit enzymes, which could
479
480 479 produce false-positive results, misleading drug discovery research [72-74]. In this sense,
481
482 480 none of the studies dealing with ligand fishing and identifying flavonoids have addressed the
483
484 481 possible formation of colloidal aggregates, although this process in physiological conditions
485
486 482 is unconfirmed [75,76].

487
488 483 The most successful class of natural products being used to treat cancer is probably
489
490 484 alkaloids. They represent a highly diverse group of compounds containing cyclic structures
485
486 485 with at least one basic nitrogen atom incorporated within [77,78]. There are approximately
487
488 486 12,000 different alkaloids identified and classified according to their chemical structures,
489
490 487 biosynthetic pathways, and biological activities. Since the 1960s, alkaloids have been
488
489 488 recognized as an important class in the treatment of cancer, with the vinca alkaloids being
490
491 489 the first plant-derived antimitotic agents to be introduced into the drug market [60]. Alkaloids
492
493 490 are considered important sources of anticancer drugs, with several preclinical and clinical

1
2
3 491 studies being conducted [79]. They act through complex mechanisms, affecting multiple
4 492 targets, and therefore can be useful to treat various forms of cancers [78]. Camptothecin, for
5 493 instance, is a monoterpene pentacyclic quinoline alkaloid first extracted from the leaves of
6 494 *Camptotheca acuminata* that originated irinotecan and topotecan, two FDA-approved
7 495 topoisomerase-1 inhibitors able to treat metastatic ovarian, colorectal, and primary colon
8 496 cancers. Another successful example is taxol, a β -tubulin-binding alkaloid extracted from the
9 497 barks of *Taxus brevifolia* Nutt. that was approved in the 1980s for the treatment of Kaposi
10 498 sarcoma, ovarian, breast, colon lung, bladder, prostate, melanoma, esophageal, and neck
11 499 cancers [80].

12 500 Given the importance of alkaloids to the discovery of new anticancer agents,
13 501 seventeen studies have successfully fished and characterized 37 alkaloids. However, some
14 502 minor constituents like the ones that were reported for the first time in the complex venom
15 503 of *Bufo melanostictus* Schneider, such as compound 18 ($C_{14}H_{17}O_4N_2$), were referred to as
16 504 unknown due to their novelty [11]. This highlights the importance of sensitive chemical
17 505 screening methods, such as ligand fishing, in the discovery of new alkaloids with unexplored
18 506 pharmacological potential.

19 507

20 508 **3.4. Natural sources screened for anticancer compounds using the Ligand Fishing** 21 509 **strategy**

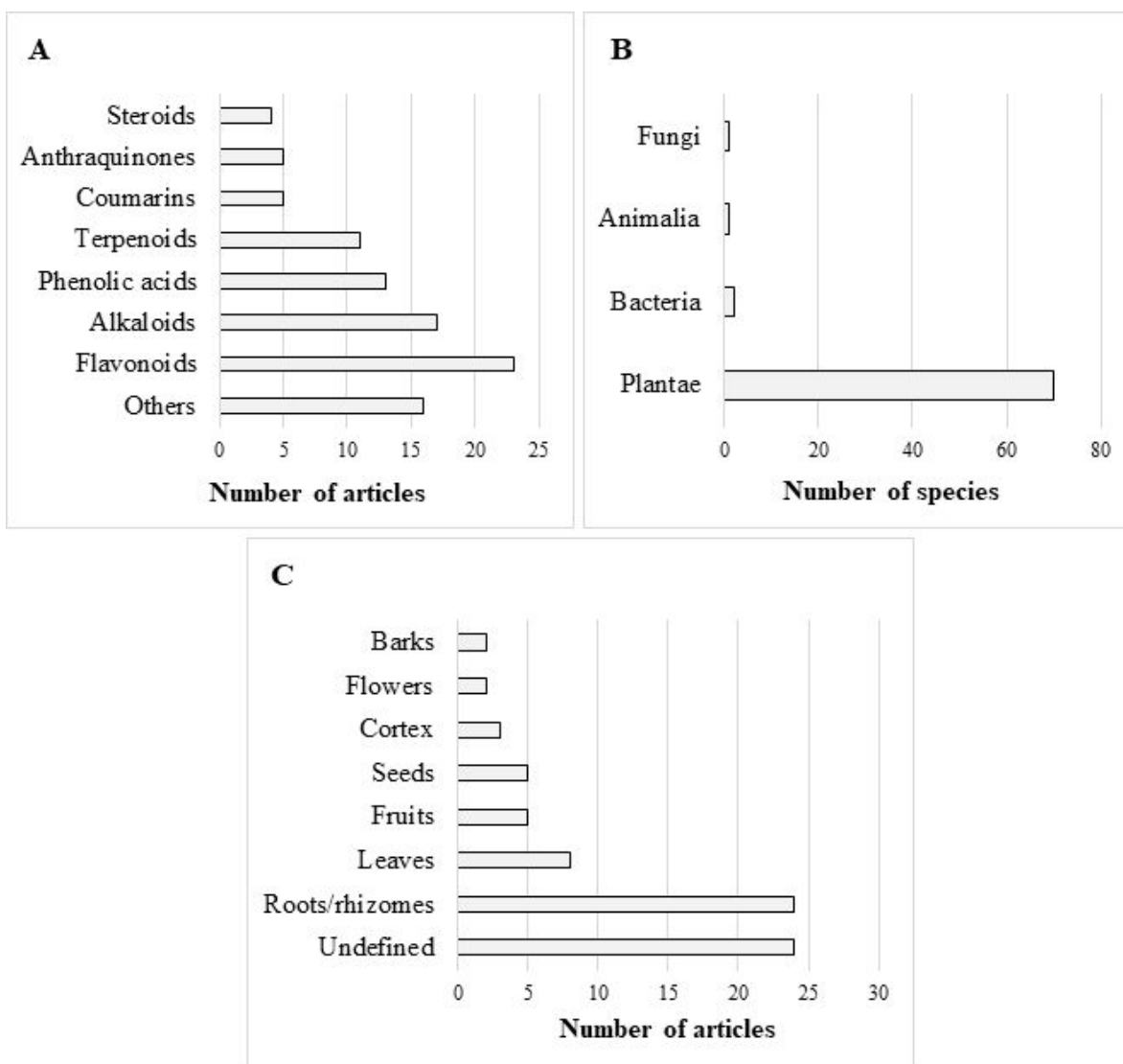
22 510 Among the taxonomic kingdoms of the species searched (n=74), the Plantae kingdom
23 511 stands out as the most studied one, with 94,6% of all species in the selected articles (n=70)
24 512 (**figure 9B and Table S1**). Different parts of plants were investigated in these studies (**table**
25 513 **S1**), the roots/rhizomes being the most studied ones (n=24), followed by the leaves (n=8),
26 514 fruits (n=5), seeds (n=5), cortex (n=3), flowers (n=2) and barks (n=2). Interestingly, there
27 515 was a considerable number of studies (n=24) that did not specify the part used, being referred
28 516 to as undefined (**figure 9C and table S1**).

29 517 It is also worth mentioning that most ligand fishing experiments performed to identify
30 518 anticancer natural products were made with terrestrial plant extracts (**Figure 9B**). However,
31 519 there is a huge and undiscovered potential in marine organisms [87][89], which were barely
32 520 screened by ligand fishing studies for anticancer leads. The group of Choi and collaborators
33 521 [82] was the only one that screened a marine-derived natural product. They fished out
34 522 tetrangulol methyl ether, a potent quinone reductase-2 inhibitor ($IC_{50} = 0.16 \mu M$), from an
35 523 extract of *Actinomyces* sp. derived from marine sediment. Indeed, marine-derived natural
36 524 products from seaweeds, microalgae, and plants have shown great potential, not only for
37 525 cancerology but also for human health [83–85]. In this sense, they represent an immense
38 526 reservoir of interesting natural molecules, and as such, should be better explored using the
39 527 ligand fishing strategy.

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41 529

42 530



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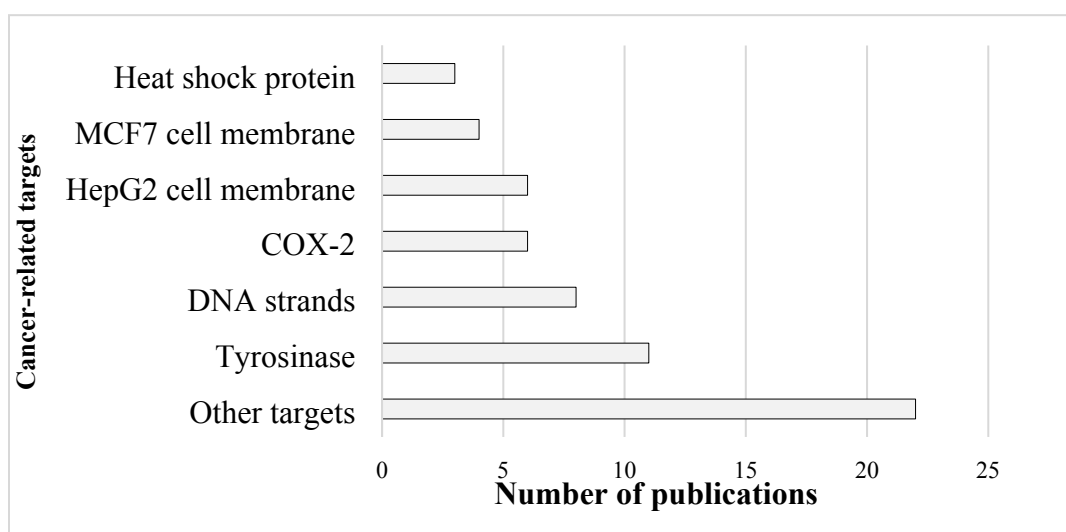
532 **Figure 9** – Graphical summary of ligands fished (A), the taxonomic kingdom of the screened
 533 natural product (B), and the parts used to make the extraction (C).

534

535 3.5. Cancer-related targets

536 Most newly approved anticancer drugs explore specific oncoproteins encoded by
 537 mutated genes. However, the number of identified genes is extremely small compared to the
 538 diversity of cancer targets described in the literature, which makes the oncogene
 539 determination and search for new target inhibitors two complementary approaches in
 540 anticancer drug discovery [86]. Hereupon, in this section, we focus on the latter aspect of
 541 anticancer drug discovery, discussing closely related cancer targets, general pathways, and
 542 other technologies involved in the ligand fishing works selected for this review. Among
 543 cancer molecular targets, tyrosinase stands out as the most studied one (n=11 – 18.3%),
 544 followed by DNA (n=8 – 13.3%) the cell membrane of HepG2 cells (n=6 – 10.4%),
 545 cyclooxygenase-2 (n=6 – 10.4%), cell membrane of MCF7 cells (n=4 – 6.6%) and heat shock
 546 protein 90 (n=3 – 6.3%). Other targets were also employed, such as lactate dehydrogenase,
 547 glutathione-S-transferase, fibroblast growth factor receptor 4 (FGFR4), and tropomyosin-
 548 related kinase B (TrkB) receptor among others (**figure 10** and **table S2**).

549



550 **Figure 10** – Main anticancer targets exploited in natural products ligand fishing studies.

551 Each bar represents the number of studies included

552

553 3.5.1. Proteins

554 Proteins are the most explored targets for disease treatment and their modulating
 555 agents represent the vast majority of all anticancer drugs approved by the FDA [87]. Not
 556 surprisingly, proteins were the most used targets to construct ligand fishing systems able to
 557 capture anticancer compounds from natural products.

558

559 *3.5.1.1 Tyrosinase*

560 Tyrosinase is an enzyme involved in the production of melanin in different organisms
 561 such as animals and plants. In humans, tyrosinase is closely related to melanogenesis and its
 562 inhibitors may represent an important clinical strategy to treat melanoma [88]. In this sense,
 563 tyrosinase inhibitors were sought by different ligand fishing approaches such as hollow fiber
 564 [42], ultrafiltration [19,88–94], and magnetic particles [95,96], in an attempt to efficiently
 565 capture ligands with affinity for this target.

566 Yang and colleagues [89], for example, successfully fished out two derivatives from
 567 quercetin and kaempferol (quercetin-3-O-(6-O-malonyl)- β -D-glucopyranoside and
 568 kaempferol-3-O-(6-O-malonyl)- β -D-glucopyranoside, respectively) from the *Morus alba* L.
 569 leaf extract as promising tyrosinase inhibitors. However, it is important to note that the
 570 oxidation of phenolic compounds by the active tyrosinase contributes to the formation of
 571 reactive species, stimulating apoptosis via the activation of p53. Therefore, the indirect
 572 cytotoxic effect of tyrosinase depends on its activations, instead of inhibition [96]. Indeed,
 573 the prodrug-activating feature of tyrosinase has been exploited to improve melanoma
 574 treatment since melanoma cells distinguish from normal cells, among other characteristics,
 575 by the overexpression of tyrosinase, which in turn favors cytotoxicity towards cancer over
 576 normal cells [97].

577 On the other hand, the exaggerated production of melanin in melanoma cells confers
 578 radioprotection and resistance to photodynamic therapy. Additionally, melanin can chelate
 579 chemotherapeutic agents like cyclophosphamide whereas cell whitening promotes

1
2
3 580 sensitization, indicating that tyrosinase inhibitors may represent a feasible alternative for
4
5 581 melanoma chemosensitization [98].

6 582

7
8 583 *3.5.1.2. Cyclooxygenase-2 (COX-2)*

9
10 584 Cyclooxygenase-2 (COX-2) is one of the most studied targets involved with
11
12 585 inflammation. COX-2 inhibitors are widely used to contain inflammation since they present
13
14 586 fewer side effects than COX-1 inhibitors. These enzymes catalyze the oxidation of fatty acids
15
16 587 to produce eicosanoids such as prostaglandins (PG), which in turn are closely related to the
17
18 588 transformation of normal hyperplastic into neoplastic cells. In this regard, several levels of
19
20 589 evidence indicate that the inhibition of cyclooxygenases, particularly COX-2, is intimately
21
22 590 linked to a significant risk reduction for cancer development. Furthermore, COX-2 is
23
24 591 overexpressed in various cancer types such as breast [99], colon [100], cervical [101], and
25
26 592 non-small cell lung cancer [102]. Additionally, COX-2 overexpression is associated with
27
28 593 poor prognosis in the brain, ovarian, uterine, hepatocarcinoma, and melanoma, among other
29
30 594 cancers, which makes this enzyme an important target for anticancer therapy research [103].

31 595 The prostaglandin E₂ (PGE₂) is one of the most important eicosanoids produced by
32
33 596 COXs, having the ability to stimulate several intracellular pathways involved in cancer
34
35 597 pathophysiology. For instance, PGE₂ can promote apoptosis resistance via upregulation of
36
37 598 Bcl-2, stimulate angiogenesis via upregulation of vascular endothelial growth factor (VEGF),
38
39 599 metastasis through increase in metalloproteinases types 2 and 9, tumor growth via
40
41 600 upregulation of epidermal growth factor receptor (EGFR) and invasion via PI3K-AKT
42
43 601 signaling pathway [104].

44 602 Based on the importance of COX-2 for cancerology, six works employed this target
45
46 603 in the development of ligand fishing assays to screen natural products, aiming to find
47
48 604 innovative inhibitors. Four of the articles focused on the role of COX-2 inhibitors in cancer
49
50 605 therapy [105-108], whereas two works focused on the anti-inflammatory features of the
51
52 606 fished inhibitors [109, 110]. In any case, COX-2 inhibitors can be extremely valuable in both
53
54 607 scenarios. Celecoxib, for instance, is a widely used selective COX-2 inhibitor that promotes
55
56 608 local anti-inflammatory response as well as acts as a chemopreventive agent, reducing
57
58 609 recurrences of adenoma and breast cancer by more than 40% [111].

59 610

60 611 *3.5.1.3 Phospholipases A₂ (PLA₂s)*

612 Phospholipases A₂ (PLA₂s) are enzymes belonging to the esterase superfamily that
613
614 613 catalyze the hydrolyzation of the acyl bond of glycerophospholipids to release
615
616 614 lysophospholipids and free fatty acids, being subdivided in cytosolic (cPLA₂), calcium-
617
618 615 independent (iPLA₂), secreted (sPLA₂), lysosomal, adipose-specific PLA₂s and platelet-
619
620 616 activating factor (PAF) [112]. Among its different isoforms, cPLA₂α, iPLA₂β, sPLA₂-IIA,
617
618 617 and sPLA₂-III have demonstrated involvement with human carcinogenesis, therefore being
619
620 618 able to be considered for anticancer therapies. Arachidonic acid (AA) is, undoubtedly, one
619
620 619 of the most important products of PLA₂s' action. It is a glycerophospholipid derivative that
620
620 620 is transformed into eicosanoids closely related to inflammation and tumorigenesis by the

1
2
3 621 action of cyclooxygenases (COXs), lipoxygenases (LOXs), and cytochrome p450 (CYP450)
4
5 622 enzymes [113].

6
7 623 Based on this, numerous studies have demonstrated the dual role of PLA₂s isoforms
8
9 624 in tumorigenesis. The sPLA₂, for instance, acts as a tumor suppressor in gastric and intestinal
10
11 625 cancers and as a tumor promoter in breast, lung, prostate, ovarian, and esophageal cancers
12
13 626 [113], whereas cPLA₂ is mostly tumorigenic, except for colon cancer where it acts as a tumor
14
15 627 suppressor [114]. This highlights the importance of targeting the appropriate isoforms of
16
17 628 PLA₂s depending on the type of cancer.

18
19 629 Given the above-mentioned, PLA₂ inhibitors may be promising pharmacological
20
21 630 tools to treat cancer. In this regard, Wei and collaborators [115] elaborated a magnetic
22
23 631 nanoparticle-based ligand fishing system with immobilized porcine PLA₂ in an attempt to
24
25 632 find inhibitors from *Pseudopithomyces* sp. extract. In it, they fished six inhibitors and tested
26
27 633 them against human lung carcinoma (A549), bone marrow neuroblastoma (SH-SY5Y), and
28
29 634 cervical cancer (HeLa) cell lines. Interestingly, the authors found good potency of fusaristatin
30
31 635 C (IC₅₀ = 10.10 μM) against A549 cells, but not against SH-SY5Y and HeLa cells, which
32
33 636 may correlate with the alleged role of PLA₂s in specific types of cancers.

32 33 637 34 638 *3.5.1.4 Lactate dehydrogenase (LDH)*

35
36 639 Lactate dehydrogenase (LDH) is a tetrameric enzyme formed by two subunits (A and
37
38 640 B) with different body distributions. While subunit A is more present in the liver and
39
40 641 muscles, subunit B predominates in the heart, being both upregulated in different types of
41
42 642 cancer [116]. Differently from normal cells, cancer cells obtain most of their energy from
43
44 643 glycolysis and subsequent conversion of pyruvate into lactate by LDH, even under normal
45
46 644 aerobic conditions [117]. This distinguished activity pattern turns LDH into an interesting
47
48 645 target in the search for inhibitors with anticancer potential since diminished levels of LDH
49
50 646 activity reduce tumorigenicity causing minor side effects [116,118]. Exploring this
51
52 647 potentiality, Cheng and collaborators [119] fabricated a bovine LDH-grafted magnetic
53
54 648 particle capable of rapidly screening natural inhibitors from two complex matrices. In it, the
55
56 649 authors successfully fished out twelve different ligands of which at least six (emodin, rhein,
57
58 650 aloe-emodin, chrysophanic acid, lunatin, and (-)-epicatechin-3-O-gallate) have
59
60 651 demonstrated promising activity against cancer cells by other research groups [120-125].
652 However, in this work, the authors did not perform enzymatic assays to check inhibition in
653 the presence of the fished-out ligands, not being possible to confirm their alleged LDH
654 inhibiting potential.

655 656 *3.5.1.5 Glutathione S-transferase (GST)*

657
658 659 Glutathione S-transferase (GST) is a family of enzymes that catalyze the conjugation
660
661 661 of glutathione into different types of electrophilic substances, being responsible for phase II
detoxification. Additionally, GST (μ and π classes) participates in other cellular processes
including apoptosis, carcinogenesis, tumor formation, and multidrug resistance, in which it
undermines the chemotherapy drug efficacy by extrusion and inhibits the MAPK pathways

1
2
3 662 to avoid tumor cell apoptosis [126]. Indeed, the GST isozyme GSTP1 ($\pi 1$) is overexpressed
4
5 663 in different types of cancer (glioma, gastric, prostate, and breast cancer) and its
6
7 664 inhibition/knockdown increases the response to anticancer-approved drugs [127]. For these
8
9 665 reasons, GST inhibitors have been considered for anticancer drug discovery and treatment,
10
11 666 being used as an immobilized target in the ligand fishing system developed by Yi and
12
13 667 colleagues [28]. In it, the authors engineered a mesoporous silica magnetic particle with GST
14
15 668 immobilized on its surface intending to screen natural products with inhibitory potential
16
17 669 against GST. With it, they could efficiently fish seven ligands, of which three presented high
18
19 670 binding affinity for GST, being them rosmarinic acid, (-)-epigallocatechin-3-O-gallate, and
20
21 671 (-)-epicatechin-3-gallate [28].

22
23 672 GST inhibitors are classified according to their activity in specific binding sites,
24
25 673 namely G- and H-sites. G-site inhibitors are glutathione-based molecules since this specific
26
27 674 site only accepts glutathione as a substrate. On the other hand, H-site inhibitors are more
28
29 675 structurally diversified, making the development of inhibitors quite challenging [127]. One
30
31 676 way or the other, natural products are interesting sources of GST inhibitors and when
32
33 677 associated with ligand fishing approaches, this enables fast screening and discovery of new
34
35 678 chemosensitizers to aid current cancer chemotherapy.

36 37 679 38 39 680 *3.5.1.6 Quinone reductase-2 (QR2)*

40
41 681 Like GST, quinone reductase-2 (QR2) is a phase II enzyme that protects the body
42
43 682 against toxic and reactive chemical species, although its role in cancer appears to be less
44
45 683 evident. Yet, QR2 inhibitors have been hypothesized as cancer chemopreventive agents,
46
47 684 resveratrol being one of the most studied natural products for this purpose [128,129]. Indeed,
48
49 685 resveratrol has been associated with all three steps of carcinogenesis and as a QR2 inhibitor,
50
51 686 it seems to regulate the degradation of cyclin D1 by the proteasome via the AKT-GSK-3 β
52
53 687 pathway, limiting cell growth. Moreover, QR2 knockdown results in slower proliferation and
54
55 688 cell cycle arrest at the G1 phase, indicating that this enzyme plays a key role in cell division
56
57 689 [130].

58
59 690 From this perspective, Choi and collaborators [82] screened different natural products
60
61 691 seeking QR2 inhibitors by a ligand fishing assay. They incubated human recombinant QR2
62
63 692 with marine sediment bacteria (*Actinomyces* sp.) and hop (*Humulus lupulus* L.) extracts,
64
65 693 being able to fish out and identify three different inhibitors, one from bacteria (tetrangulol
66
67 694 methyl ether) and two from hops (xanthohumol and xanthohumol D). As compared to
68
69 695 resveratrol ($IC_{50} = 5.1 \mu M$), xanthohumol and xanthohumol D showed lower activity ($IC_{50} >$
70
71 696 $100 \mu M$) whereas tetrangulol methyl ether was quite potent ($IC_{50} = 0.16 \mu M$) [131]. In fact,
72
73 697 the latter compound has shown interesting cytotoxicity against different types of cancer cells
74
75 698 (KB, MCF-7, and NCI-H187) [132], which may also correlate with its potent inhibitory
76
77 699 activity against QR2.

78 79 700 80 81 701 *3.5.1.7 Heat-shock proteins (HSPs)*

1
2
3 702 The heat-shock proteins (HSP) are mammalian chaperones that regulate protein
4
5 703 folding and refolding in the face of stressful conditions such as abrupt temperature changes
6
7 704 and exposure to high concentrations of reactive oxygen species (ROS). In this sense, HSPs
8
9 705 represent the primary line of defense against stress-related cellular events, therefore being
10
11 706 related to various stress-originated diseases. For these reasons, HSP has been widely
12
13 707 considered in the search for new drugs, especially in cancerology. In this regard, HSPs play
14
15 708 a key role in carcinogenesis surveillance since cancer cells require distinct and often mutated
16
17 709 biochemical machinery to favor proliferation, invasion, metastasis, evasion from apoptosis,
18
19 710 and drug resistance [133].

20
21 711 Among the families of HSPs (HSP27, HSP40, HSP60, HSP70, and HSP90), HSP90
22
23 712 is the most studied one. This protein is overexpressed in several types of cancer and is related
24
25 713 to a bad prognosis in cholangiocarcinoma, glioblastoma, gastric, lung, and breast cancers.
26
27 714 The body of evidence correlating HSP90 to cancer is vast. The increased expression of
28
29 715 HSP90, for example, is responsible for the activation and stabilization of oncogenic protein
30
31 716 kinases, such as STAT3, JAK2, AKT, PI3K, and MAPK, which ultimately stimulate cancer
32
33 717 cell growth and tumor progression [134].

34
35 718 In the field of ligand fishing, HSP90 has been explored as a target in the search for
36
37 719 new anticancer agents. Three works from the same research group used quantum dots as a
38
39 720 support to immobilize HSP90 and screen natural products for drug leads [44-46]. The group
40
41 721 found multiple HSP90 inhibitors, some with characterized cytotoxicity *in vitro* and antitumor
42
43 722 effect *in vivo* [135-137]. However, developing HSP90 inhibitors is quite challenging as they
44
45 723 frequently fail in clinical trials either due to high toxicity or lack of beneficial effects
46
47 724 [138,139].

48
49 725

50 51 726 3.5.1.8 Sirtuin 6 (SIRT6)

52
53 727 Sirtuins are nicotinamide adenine dinucleotide-dependent (NAD⁺) proteins belonging
54
55 728 to the class III histone deacetylase family, being involved in DNA damage repair, stress
56
57 729 response, ribosome biogenesis, cell proliferation, and cell death [140]. There are seven
58
59 730 isoforms of sirtuins (SIRT1-7), classified according to their function and location. Sirtuin 6
60
731 (SIRT6), in particular, is a nuclear deacetylase involved in the modulation of various
732
733 732 biological processes such as inflammation, metabolism, aging, tumor suppression, and
734
735 733 carcinogenesis. The reduction of SIRT6 levels is associated with the human premature aging
736
737 734 disorder and restoration of its activity reduces cell senescence. Additionally, SIRT6
738
739 735 possesses both tumor suppressor and oncogenic properties that will emerge depending on the
740
741 736 type of cell. In colorectal, ovarian, and pancreatic cancers, for example, functional SIRT6
742
743 737 inhibits progression whereas its upregulation is related to hepatocarcinoma, skin, lung,
744
745 738 breast, and prostate cancers [141].

746
747 739 In this context, two similar works used immobilized SIRT6 onto the surface of
748
749 740 magnetic particles to screen *Trigonella foenum-graecum* seed extract in an attempt to find
750
751 741 interesting ligands [38,39] In the first, the authors develop the fishing system using catalytic
752
753 742 mutant SIRT6 and heat-inactivated SIRT6 as negative controls, being able to identify

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2
3 743 quercetin and vitexin as inhibitors. In the second work, the authors used the same fishing
4 744 technique but with a different identification approach, being able to identify orientin and
5 745 fourteen other SIRT6 modulators for the first time.
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9 747 3.5.1.9 $\alpha_v\beta_3$ Integrin

10 748 Cancer invasion and metastasis strongly depend on the partial degradation of the
11 749 extracellular matrix and the formation of new blood vessels [142]. Integrins are a protein
12 750 family responsible for cell-cell and cell-extracellular matrix adhesion as well as angiogenesis,
13 751 being the $\alpha_v\beta_3$ integrin (also known as vitronectin receptor) the most important one [143]. In
14 752 fact, this protein is highly expressed in newborn vessels whereas is lacking in resting
15 753 endothelial cells [144]. In this sense, $\alpha_v\beta_3$ integrin has been explored as an anticancer target
16 754 since the inhibition of its activity has been associated with endothelial apoptosis and
17 755 inhibition of angiogenesis, which is a crucial step in tumor formation and growth [145,146].
18 756 Cilengitide, for example, is one of the most studied $\alpha_v\beta_3$ antagonists, being able to potentiate
19 757 the radiotherapy outcomes in patients with glioma [147].

20 758 Having this challenge in mind, Uliana and colleagues [148] developed a ligand
21 759 fishing system with immobilized $\alpha_v\beta_3$ integrin able to efficiently fish out [10]-gingerol from
22 760 the ginger extract. The [10]-gingerol has been tested as an adjuvant to the treatment with
23 761 doxorubicin using *in vitro* and *in vivo* models of triple-negative breast cancer where it was
24 762 able to reduce metastasis, chemotherapy-related weight loss, and hepatotoxicity [149].
25 763

26 764 3.5.1.10 Tropomyosin-related kinase B (TrkB) receptor

27 765 The tropomyosin-related kinase is a family of tyrosine kinase receptors involved in
28 766 neural development and cell functioning, survival, and proliferation. As tyrosine kinase
29 767 receptors, after neurotrophin stimulus, they undergo a dynamic homodimerization with
30 768 subsequent transactivation of the tyrosine kinase domains which in turn allows the
31 769 recruitment of cytoplasmic proteins and activation of intracellular pathways [150].
32 770 Alterations in these receptors, either via mutations or rearrangements, have been related to
33 771 oncogenesis, therefore making these proteins potential anticancer targets [151]. They are
34 772 divided into three types depending on their activating neurotrophin: the TrkA, stimulated by
35 773 the nerve growth factor (NGF), the TrkB, which is essentially activated by the brain-derived
36 774 neurotrophic factor (BDNF) and the TrkC which in turn is stimulated in the presence of the
37 775 neurotrophin-3 [152]. Preclinical studies have demonstrated that TrkA and TrkB inhibitors
38 776 are capable of attenuating the proliferation and aggressiveness of pancreatic, neuroblastoma,
39 777 and neuroendocrine tumors via apoptosis induction [153]. Additionally, the pan-Trk inhibitor
40 778 (TrkA, B, and C inhibitor) entrectinib (Rozlytrek®), recently approved in Japan, the USA,
41 779 and the European Union has demonstrated great results in the treatment of solid tumors in
42 780 patients with specific genetic conditions [154].

43 781 Aware of the potential of Trk inhibitors, Arituluk and colleagues [32] developed a
44 782 ligand fishing assay using magnetic nanoparticles coated with TrkB-overexpressing SH-
45 783 SY5Y cell membranes to screen a Chinese natural product in the search for new inhibitors.

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3 784 Despite some limitations, this model proved to be a fast-track in the screen of TrkB inhibitors
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5 785 from a very complex natural matrix, with a great degree of specificity as it was able to
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7 786 discriminate between TrkB binders and non-binders mixed in equimolar concentration [32].
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9 787

10 788 *3.5.1.11 Fibroblast growth factor receptor 4 (FGFR4)*

11
12 789 Fibroblast growth factor receptor 4 (FGFR4) is a highly conserved tyrosine kinase
13
14 790 receptor involved in various physiological processes such as tissue growth and repair,
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16 791 production of bile acids, metabolism, and muscle differentiation. Upon activation by the
17
18 792 fibroblast growth factor (FGF) in the extracellular domains, the FGFR4 undergoes
19
20 793 dimerization and transphosphorylation (intracellular domains), which in turn allows the
21
22 794 direct activation of the PLC γ -IP $_3$ -DAG-PKC pathway and indirect (via FGFR substrate 2 –
23
24 795 FRS2) activation of RAS-RAF-MAPK and PI3K-AKT pathways [155]. Moreover, FGFR4
25
26 796 overexpression and mutation as well as upregulation of its endogenous agonists are closely
27
28 797 related to the development of cancer, especially breast cancer [156]. Despite that, the
29
30 798 oncogenic functions of FGFR4 seem to vary greatly depending on the type of cancer as it is
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32 799 related to poor prognosis in colorectal, esophageal, liver, rhabdomyosarcoma, and lung
33
34 800 cancers and with better survival in cervical, cholangiocarcinoma, pancreatic and head and
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36 801 neck cancers [157].

37 802 Based on the close relationship between FGFR4 receptors and cancer, Bu and
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39 803 colleagues [158] used a previously engineered HEK293 cell transfected with the Y367C
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41 804 mutation [159] to produce a mutant FGFR4-expressing cell membrane-coated silica particle
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43 805 to screen a TCM product. It is important to note that the FGFR4 Y367C mutation is a
44
45 806 dominant oncogene in MDA-MB453 breast cancer cells [160] and as such, it is a good model
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47 807 for the screening of anticancer compounds through cell membrane ligand fishing assay. As a
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49 808 result, the authors fished out four alkaloids of which one (napellonine) had inhibitory action
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51 809 on FGFR4, with IC $_{50}$ of 133 μ g/mL. Yet according to the authors, in comparison to the
52
53 810 standard FGFR4 inhibitor PD173074 (IC $_{50}$ = 71.3 μ g/mL), napellonine would need structural
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55 811 modifications to increase its potency towards FGFR4 [158].
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57 812

58 813 *3.5.1.12 Urokinase plasminogen activator (uPA)*

59
60 814 The urokinase plasminogen activator (uPA) is an essential protease involved in the
815
816 815 mechanism of conversion of plasminogen into plasmin. The extracellular matrix (ECM) is
817
818 816 particularly affected by the activation of the uPA receptor (uPAR) by uPA as it is degraded
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820 817 by plasmin-activated metalloproteinases. The activation of this system is regulated by the
821
822 818 plasminogen activator inhibitor 1 and 2 (PAI-1/2), which in turn signals for internalization
823
824 819 of the complex uPA-uPAR. This phenomenon is crucial for various biological processes like
820
821 820 embryogenesis and wound healing as well as in pathological states such as cancer invasion
822
823 821 and metastasis [161]. Furthermore, the uPA system can facilitate cancer metastasis by
824
825 822 different mechanisms other than ECM degradation. The binding of uPA to the uPAR can
826
827 823 activate MAPK pathways (Ras-Raf-MEK-ERK and Rac1-MKK3-p38-MAPKAPK2) which
828
829 824 are involved with cell growth, differentiation, survival, migration, and apoptosis [162]. The

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3 825 invasion of breast cancer cells, for example, is thought to be mediated by uPA via the p38-
4 826 MAPK pathway with the aid of transmembrane integrins since the uPAR lacks
5 827 transmembrane and cytosolic domains. Additionally, PAI-1 has a dual role in cancer since it
6 828 stimulates the expression and internalization of the uPA-uPAR via low-density lipoproteins.
7 829 Interestingly, this endogenous inhibitor has been associated with a poor cancer prognosis.
8 830 Indeed, PAI-1's ability to limit excessive degradation of the ECM seems to help cancer cells
9 831 to invade other tissues and spread through the bloodstream. Furthermore, PAI-1 also
10 832 stimulates angiogenesis, apparently by exerting the same control over the ECM [163].
11 833 Previous reports showed that the uPA antagonist amiloride could modulate uPA gene
12 834 expression at transcription and post-transcriptional levels, proving to be effective against
13 835 cancer in preclinical models [164].

14 836 From this panorama, Li and collaborators [165] developed a fishing system designed
15 837 to screen natural uPA ligands from traditional Chinese medicines (TCMs). With it, they
16 838 screened forty-two TCMs to find seven uPA ligands, of which pentagalloylglucose and 28-
17 839 *O*- β -D-glucopyranosyl pomolic acid presented good inhibitory potential against uPA (IC_{50} =
18 840 1.639 μ M and 37.82 μ M respectively). The high throughput nature of this approach allowed
19 841 the authors to screen dozens of complex matrices, with thousands of chemical entities each,
20 842 in a very short time as compared to classical natural product screenings, which makes this
21 843 technique suitable for accelerated natural product screen and drug discovery.

22 844

23 845 3.5.1.13 Matrix Metalloproteinase-2 (MMP-2)

24 846 Matrix metalloproteinases (MMPs) are a large family of zinc-dependent proteases
25 847 that act by cleaving and rebuilding the extracellular matrix in connective tissues. These
26 848 enzymes participate in physiological processes such as apoptosis, ovulation, inflammation,
27 849 angiogenesis, neural growth, bone remodeling, embryonic development, and organ
28 850 morphogenesis. On the other hand, pathological processes such as cancer invasion and
29 851 metastasis occur with the aid of MMPs, highlighting the importance of these enzymes to
30 852 cancerology [166]. There are more than 20 isoforms of MMPs, each having specific
31 853 substrates, tissue distribution, molecular mass, and gene coding. Among them, gelatinases
32 854 (MMP-2 and 9) are particularly involved in cancer invasion and metastasis as their increased
33 855 expression has been related to the invasiveness of gliomas [167], malignant transformations
34 856 after human papilloma virus infections [168], reduced survival in patients with primary breast
35 857 carcinoma [169] and esophageal cancer [170].

36 858 In this context, Tao and colleagues [171] constructed a very interesting ligand fishing
37 859 system that combines immobilization on magnetic particles with a microfluidic chip system.
38 860 In it, the authors prepared and characterized a quantum dot magnetic particle with
39 861 immobilized MMP-2 before submitting it to a microfluidic chip. Through two different
40 862 channels (b and c), they injected the MMP-2-functionalized magnetic particles and the fruit
41 863 extract of *Rosa roxburghii* Tratt. After incubation, a small ring magnet trapped the magnetic
42 864 particles, and a wash solvent (phosphate buffer solution) was pumped through two other
43 865 channels (d and f) to remove unspecifically bound ligands that were collected through two

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3 866 additional channels (e and g). In the end, the dissociating solvent (30% acetonitrile in water)
4
5 867 was pumped through another channel (h) to separate the ligands from the MMP-2-
6
7 868 functionalized magnetic particles. Finally, the eluted ligands were pumped into a UPLC-Q-
8
9 869 TOF/MS system to be analyzed. Among the twelve ligands fished and identified, ellagic acid
10
11 870 stands out as it is a well-known MMP-2 inhibitor [171,172]. This polyphenolic compound
12
13 871 possesses antiproliferative activity in various types of cancer cells, such as glioblastoma (U87
14
15 872 and U118), human osteogenic sarcoma (CRL1343), human colorectal adenocarcinoma
16
17 873 (Caco-2), human prostate cancer (DU 145) and breast cancer (Hs 578T and MCF-7)
18
19 874 [173-175]. Furthermore, the other eleven ligands fished (niga-ichigoside F₁, rosamultin,
20
21 875 kajiichigoside F₁, zebirioside O, peduncloside, potentillanoside A, quadranoside VIII,
22
23 876 corilagin, oreganol A, medicagenic acid, and valerenic acid) were described as MMP-2
24
25 877 inhibitors for the first time, evidencing the high throughput potential of this approach to
26
27 878 isolate new molecules that could be useful for cancer treatment [171].
28
29 879

30 880 3.5.1.14 Monoacylglycerol lipase (MAGL)

31 881 The monoacylglycerol lipase (MAGL) is a soluble enzyme that catalyzes the
32
33 882 hydrolyzation of monoacylglycerols to glycerol and fatty acids. Moreover, this enzyme is
34
35 883 responsible for the hydrolysis of the endocannabinoid 2-arachidonoylglycerol (2-AG),
36
37 884 releasing arachidonic acid, which in turn is a major precursor of pro-inflammatory agents
38
39 885 that are involved in cancer progression, such as prostaglandins. In aggressive types of cancer,
40
41 886 MAGL provides the free fatty acid supply that triggers downstream pro-tumorigenic
42
43 887 pathways [176]. Nevertheless, MAGL is highly active in melanoma, ovarian, breast,
44
45 888 prostate, and liver cancer cells [177].

46 889 In this context, Mei and collaborators [178] engineered yeast cells (KM71H *Pichia*
47
48 890 *pastoris*) to expose MAGL on their surface. In a preliminary study, the authors screened
49
50 891 twelve TCMs (Corydalis Rhizoma, Salvia Miltiorrhiza Radix et Rhizoma, Chuanxiong
51
52 892 Rhizoma, Carthami Flos, Andrographis Herba, Eucommiae Cortex, Asari Radix et Rhizoma,
53
54 893 Ginkgo Folium, Cinnamomum Ramulus, Rhei Radix et Rhizoma, Acorus Tatarinowii
55
56 894 Rhizoma, and Paridis Rhizoma) to verify their inhibitory potential against MAGL. From it,
57
58 895 three TCMs had inhibitory activity greater than 50%, Corydalis Rhizoma (*Corydalis*
59
60 896 *yanhusuo* W. T. Wang) being the most active one (66.6% of inhibition). After the
897 characterization of the ligand fishing system with fluorescent antibodies, the authors
898 incubated the modified yeast with the extract of *Corydalis yanhusuo* to search for MAGL
899 inhibitors. Among the seven ligands fished and characterized, the dehydrocorydaline
900 (presented the highest relative binding rate in comparison with the control (JZL184) [178].
901 In another study, dehydrocorydaline demonstrated anticancer activity *in vitro* against MCF-
902 7 cells, promoting apoptosis via the Bax-Bcl-2 signaling pathway [179]. On the other hand,
903 it was not demonstrated yet if the anticancer effect of DHC is also mediated through the
904 inhibition of MAGL, which can be an important research question for subsequent studies.

905 906 3.5.2 DNA

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3 907 DNA structural characterization has advanced tremendously in the last decades.
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5 908 Various possible arrangements differ significantly from the B-form of DNA proposed by
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7 909 Watson and Crick in the 1950s. While the DNA B-form is relatively conserved in most cells,
8
9 910 other forms such as slipped, cruciform, triplex, and quadruplex are much more susceptible to
10
11 911 mutations, arousing great interest in the field of cancer-related drug discovery [180,181].
12
13 912 DNA G-quadruplexes, for instance, are guanine-rich DNA strands that can inhibit telomerase
14
15 913 under physiological conditions, being this enzyme overexpressed in various types of cancer.
16
17 914 Telomerase overexpression is associated with apoptosis suppression and the formation of
18
19 915 spontaneous tumors in animal models [189][191]. In this sense, small molecules that act as
20
21 916 G-quadruplex stabilizers represent an interesting alternative to limit telomerase activity and
22
23 917 therefore its oncogenic effects. Telomestatin, for example, is a potent telomerase inhibitor
24
25 918 derived from *Streptomyces anulatus* that has shown promising anticancer activity both *in*
26
27 919 *vivo* and *in vitro* [183,184].

28
29 920 Based on our survey, eight works used DNA strands (double, triple, and quadruple)
30
31 921 as smart baits to fish out ligands from natural products with anticancer potential. Different
32
33 922 approaches were used to apply DNA strands into ligand fishing settings, including dialysis
34
35 923 [185], ultrafiltration [12], magnetic particles [186], and microfluid chips [187,188] as well
36
37 924 as other less trivial supports such as streptavidin-coated 96-well plates [49,50] and agarose
38
39 925 beads [47]. The DNA-bound ligands were tested in preclinical settings in two specific reports.
40
41 926 Ma and colleagues [12] found seven DNA binders from 62 plant samples, of which two
42
43 927 (daurisolone and tetrandrine) were active against various triple-negative breast cancer and
44
45 928 pediatric cancer cell lines ($GI_{50} < 20\mu\text{M}$). In another report, Gao and colleagues [194][196]
46
47 929 found four DNA-bound alkaloids (sanguinarine, chelerythrine, protopine, and
48
49 930 allocryptopine). They found no cytotoxic activity for protopine and allocryptopine whereas
50
51 931 chelerythrine and sanguinarine presented cytotoxicity against both normal (LO2 cell line)
52
53 932 and liver cancer cells (HepG2 cell line) [187]. In fact, DNA binders are known for their
54
55 933 undistinguishable toxicity against cancer and normal cells, which dramatically limits their
56
57 934 utilization in clinical settings [189].

58 935 3.5.3 Cancer cell membranes

59
60 936 In the late 1990s, He and colleagues [190] developed a technique that used cell
937 membranes for the fabrication of stationary phases applied in chromatographic studies. This
938 innovative approach reproduces *in vivo* drug-receptor interactions and adsorbs ligands on the
939 cell membrane. Coupled with it, high-performance liquid chromatography (HPLC) systems
940 separated the retained ligands for further analysis, giving rise to cell membrane
941 chromatography (CMC). From then on, various adaptations of the technique were performed
942 to adequate to different research topics, including ligand fishing with a focus on anticancer
943 drug discovery.

944 In this review, several papers used a great variety of cancer cell lines to perform ligand
945 fishing assays such as PC3 (human androgen-refractory prostate cancer) [20], MCF-7 (breast
946 cancer) [11,51,191], HepG2 (hepatoma) [11,192], HCT116 (colorectal cancer) [193], A549
947 (lung carcinoma) [35], SKOV-3 (human ovarian cancer) [190,194], ACHN (renal

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3 948 adenocarcinoma) [190,195,196], HEK293 cells (immortalized human embryonic kidney
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5 949 cells) [197] among others. The rationale of the techniques resides in the slight differences
6
7 950 between normal and cancer cells as well as the preservation of membrane receptors' vicinity.
8
9 951 In this sense, cancer cells are used as smart baits to fish out ligands according to their affinity
10
11 952 for cell membrane surface structures, mainly receptors. Some cell membrane ligand fishing
12
13 953 studies have induced the overexpression of membrane receptors to potentialize the adsorption
14
15 954 of ligands. Hu et al. [197], for example, used HEK293 cells overexpressing the epidermal
16
17 955 growth factor receptors (EGFR) to encapsulate graphene nanoparticles, which were capable
18
19 956 of fishing out luteolin and caffeic acid from the extract of *Taraxacum mongolicum* Hand.-
20
21 957 *Mazz.* Ligand fishing systems with membrane-anchored receptors are more realistic than free
22
23 958 or immobilized receptors since the cell membrane play an important role in the conformation
24
25 959 of these targets that may affect the affinity of ligands [197].
26
27 960

961 **4. Concluding Remarks**

962 Natural products are an important source of bioactive compounds. Therefore, the
963 potential for discovering new anticancer drug leads is undeniable. Despite that, classical
964 screenings are poorly efficient and quite unspecific. To address these issues, ligand fishing
965 approaches have emerged in the last two decades as a better option to accelerate the hit-to-
966 lead discovery process. In this review, we gathered detailed information from the scientific
967 literature aiming to provide a reasonable summary of findings, highlighting successful cases
968 as well as some limitations of the techniques. This approach's versatility permits various
969 suitable configurations depending on the target and its particularities. In this sense, a great
970 number of different anticancer targets, such as proteins, DNA, and cancer cells themselves,
971 were employed to fabricate robust ligand fishing systems capable of identifying new
972 bioactive compounds. Nevertheless, we observed clear tendencies among the articles, in
973 terms of natural products and the location where the research was developed. Another
974 important piece of information is that a significant number of studies do not perform
975 anticancer preclinical evaluations. Therefore, there is a need for follow-up studies designed
976 to shed light on the ligands' alleged biological activities.

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981 **Conflict of interest statement**

982 The authors have no conflict of interest to declare

983 **Data availability statement**

984 All data are already available in the current scientific literature. Therefore, data sharing does
985 not apply as the authors have not performed new experimental assays.

986

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2
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4

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16
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20 996 **REFERENCES**

- 21 997 [1] World Health Organization, Cancer fact sheet, Glob. Cancer Obs. (n.d.).
22
23 998 <https://gco.iarc.fr/today/fact-sheets-cancers> (accessed October 5, 2022).
24
- 25 999 [2] H. Sung, J. Ferlay, R.L. Siegel, M. Laversanne, I. Soerjomataram, A. Jemal, F. Bray,
26
27 1000 Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality
28
29 1001 Worldwide for 36 Cancers in 185 Countries, CA. Cancer J. Clin. 71 (2021) 209–249.
30
31 1002 <https://doi.org/10.3322/caac.21660>.
32
- 33 1003 [3] A. Nicolaysen, Nephrotoxic Chemotherapy Agents: Old and New, Adv. Chronic
34
35 1004 Kidney Dis. 27 (2020) 38–49. <https://doi.org/10.1053/j.ackd.2019.08.005>.
36
- 37 1005 [4] A.Y. Higgins, T.D. O’Halloran, J.D. Chang, Chemotherapy-induced cardiomyopathy,
38
39 1006 Heart Fail. Rev. 20 (2015) 721–730. <https://doi.org/10.1007/s10741-015-9502-y>.
40
- 41 1007 [5] K. Bukowski, M. Kciuk, R. Kontek, Mechanisms of Multidrug Resistance in Cancer
42
43 1008 Chemotherapy, Int. J. Mol. Sci. 21 (2020) 3233.
44
45 1009 <https://doi.org/10.3390/ijms21093233>.
46
- 47 1010 [6] D. Basak, S. Arrighi, Y. Darwiche, S. Deb, Comparison of Anticancer Drug
48
49 1011 Toxicities: Paradigm Shift in Adverse Effect Profile, Life. 12 (2021) 48.
50
51 1012 <https://doi.org/10.3390/life12010048>.
52
- 53 1013 [7] T.B. Steinbichler, J. Dudás, S. Skvortsov, U. Ganswindt, H. Riechelmann, I.-I.
54
55 1014 Skvortsova, Therapy resistance mediated by cancer stem cells, Semin. Cancer Biol.
56
57 1015 53 (2018) 156–167. <https://doi.org/10.1016/j.semcancer.2018.11.006>.
58
- 59 1016 [8] G.J. Yoshida, H. Saya, Molecular pathology underlying the robustness of cancer stem
60
1017 cells, Regen. Ther. 17 (2021) 38–50. <https://doi.org/10.1016/j.reth.2021.02.002>.
- 1018 [9] P. Krzyszczyk, A. Acevedo, E.J. Davidoff, L.M. Timmins, I. Marrero-Berrios, M.
1019
1020 Patel, C. White, C. Lowe, J.J. Sherba, C. Hartmanshenn, K.M. O’Neill, M.L. Balter,
1021
1022 Z.R. Fritz, I.P. Androulakis, R.S. Schloss, M.L. Yarmush, The growing role of
1023
1024 precision and personalized medicine for cancer treatment, TECHNOLOGY. 06 (2018)
79–100. <https://doi.org/10.1142/S2339547818300020>.
- 1023 [10] R.G. de Oliveira Júnior, A.F. Christiane Adrielly, J.R.G. da Silva Almeida, R.
1024
Grougnet, V. Thiéry, L. Picot, Sensitization of tumor cells to chemotherapy by natural

- 1
2
3 1025 products: A systematic review of preclinical data and molecular mechanisms,
4 1026 *Fitoterapia*. 129 (2018) 383–400. <https://doi.org/10.1016/j.fitote.2018.02.025>.
- 7 1027 [11] W. Ren, L. Han, M. Luo, B. Bian, M. Guan, H. Yang, C. Han, N. Li, T. Li, S. Li, Y.
8 1028 Zhang, Z. Zhao, H. Zhao, Multi-component identification and target cell-based
9 1029 screening of potential bioactive compounds in toad venom by UPLC coupled with
10 1030 high-resolution LTQ-Orbitrap MS and high-sensitivity Qtrap MS, *Anal. Bioanal.*
11 1031 *Chem.* 410 (2018) 4419–4435. <https://doi.org/10.1007/s00216-018-1097-4>.
- 16 1032 [12] H. Ma, H. Liang, S. Cai, B.R. O’Keefe, S.L. Mooberry, R.H. Cichewicz, B.R.
17 1033 O’Keefe, S.L. Mooberry, R.H. Cichewicz, An Integrated Strategy for the Detection,
18 1034 Dereplication, and Identification of DNA-Binding Biomolecules from Complex
19 1035 Natural Product Mixtures, *J. Nat. Prod.* 84 (2021) 750–761.
20 1036 <https://doi.org/10.1021/acs.jnatprod.0c00946>.
- 26 1037 [13] R. Zhuo, H. Liu, N. Liu, Y. Wang, Ligand Fishing: A Remarkable Strategy for
27 1038 Discovering Bioactive Compounds from Complex Mixture of Natural Products,
28 1039 *Molecules*. 21 (2016) 1516. <https://doi.org/10.3390/molecules21111516>.
- 31 1040 [14] R.A. Meryam Sardar, Enzyme Immobilization: An Overview on Nanoparticles as
32 1041 Immobilization Matrix, *Biochem. Anal. Biochem.* 04 (2015).
33 1042 <https://doi.org/10.4172/2161-1009.1000178>.
- 37 1043 [15] R.A. Wahab, N. Elias, F. Abdullah, S.K. Ghoshal, On the taught new tricks of enzymes
38 1044 immobilization: An all-inclusive overview, *React. Funct. Polym.* 152 (2020).
39 1045 <https://doi.org/10.1016/j.reactfunctpolym.2020.104613>.
- 43 1046 [16] M.C. de Moraes, C.L. Cardoso, Q.B. Cass, Solid-Supported Proteins in the Liquid
44 1047 Chromatography Domain to Probe Ligand-Target Interactions, *Front. Chem.* 7 (2019)
45 1048 1–16. <https://doi.org/10.3389/fchem.2019.00752>.
- 49 1049 [17] G. Zuo, Z. Wang, Y.N. Guillen Quispe, S.H. Hwang, H.Y. Kim, B.G. Kang, S.S. Lim,
50 1050 Target guided isolation of potential tyrosinase inhibitors from *Otholobium pubescens*
51 1051 (Poir.) J.W. Grimes by ultrafiltration, high-speed countercurrent chromatography and
52 1052 preparative HPLC, *Ind. Crops Prod.* 134 (2019) 195–205.
53 1053 <https://doi.org/10.1016/j.indcrop.2019.03.045>.
- 58 1054 [18] L. Liu, S. Shi, X. Chen, M. Peng, Analysis of tyrosinase binders from *Glycyrrhiza*
59 1055 *uralensis* root: Evaluation and comparison of tyrosinase immobilized magnetic
60 1056 fishing-HPLC and reverse ultrafiltration-HPLC, *J. Chromatogr. B.* 932 (2013) 19–25.
1057 <https://doi.org/10.1016/j.jchromb.2013.06.002>.
- 1058 [19] F. Shi, L. Xie, Q. Lin, C. Tong, Q. Fu, J. Xu, J. Xiao, S. Shi, Profiling of tyrosinase
1059 inhibitors in mango leaves for a sustainable agro-industry, *Food Chem.* 312 (2020)
1060 126042. <https://doi.org/10.1016/j.foodchem.2019.126042>.
- 1061 [20] M.-M. Wei, S.-J. Zhao, X.-M. Dong, Y.-J. Wang, C. Fang, P. Wu, G.-Q. Song, J.-N.
1062 Gao, Z.-H. Huang, T. Xie, J.-L. Zhou, A combination index and glycoproteomics-

- 1
2
3 1063 based approach revealed synergistic anticancer effects of curcuminoids of turmeric
4 1064 against prostate cancer PC3 cells, *J. Ethnopharmacol.* 267 (2021) 113467.
5 1065 <https://doi.org/10.1016/j.jep.2020.113467>.
6
7
8
9 1066 [21] S. Milica, O. Prodanovic, N. Pantic, N. Popovic, A. M. Balaz, R. Prodanovic, The
10 1067 Enzyme Immobilization: Carriers and Immobilization methods, *J. Eng. Process.*
11 1068 *Manag.* 11 (2020).
12 1069 <https://doi.org/10.7251/jepm1902089s>.
13
14
15
16 1070 [22] M. Colombo, S. Carregal-Romero, M.F. Casula, L. Gutiérrez, M.P. Morales, I.B.
17 1071 Böhm, J.T. Heverhagen, D. Prosperi, W.J. Parak, Biological applications of magnetic
18 1072 nanoparticles, *Chem. Soc. Rev.* 41 (2012) 4306. <https://doi.org/10.1039/c2cs15337h>.
19
20
21
22 1073 [23] C. Ruffert, Magnetic Bead—Magic Bullet, *Micromachines.* 7 (2016) 21.
23 1074 <https://doi.org/10.3390/mi7020021>.
24
25
26 1075 [24] I.A. Trindade Ximenes, P.C.O. de Oliveira, C.A. Wegermann, M.C. de Moraes,
27 1076 Magnetic particles for enzyme immobilization: A versatile support for ligand
28 1077 screening, *J. Pharm. Biomed. Anal.* 204 (2021) 114286.
29 1078 <https://doi.org/https://doi.org/10.1016/j.jpba.2021.114286>.
30
31
32
33
34 1079 [25] F. Zhang, H. Li, C. Liu, K. Fang, Y. Jiang, M. Wu, S. Xiao, L. Zhu, J. Yu, S. Li, G.
35 1080 Wang, Lactate Dehydrogenase-Inhibitors Isolated from Ethyl Acetate Extract of
36 1081 *Selaginella doederleinii* by Using a Rapid Screening Method with Enzyme-
37 1082 Immobilized Magnetic Nanoparticles, *Front. Biosci.* 27 (2022) 229.
38 1083 <https://doi.org/10.31083/j.fbl2708229>.
39
40
41
42
43 1084 [26] Y. Zhao, J.-J. Hu, X.-L. Bai, H.-P. Liu, X.-W. Qi, X. Liao, Fast screening of tyrosinase
44 1085 inhibitors from traditional Chinese medicinal plants by ligand fishing in combination
45 1086 with in situ fluorescent assay, *Anal. Bioanal. Chem.* 414 (2022) 2265–2273.
46 1087 <https://doi.org/10.1007/s00216-021-03864-w>.
47
48
49
50
51 1088 [27] N. Zhu, H. Ji, P. Yu, J. Niu, M. Farooq, M. Akram, I. Udego, H. Li, X. Niu, Surface
52 1089 Modification of Magnetic Iron Oxide Nanoparticles, *Nanomaterials.* 8 (2018) 810.
53 1090 <https://doi.org/10.3390/nano8100810>.
54
55
56
57 1091 [28] R. Yi, R. Fu, D. Li, J. Qi, H. Liu, Identification of ligands from natural products as
58 1092 inhibitors of glutathione S-transferases using enzyme immobilized mesoporous
59 1093 magnetic beads with high-performance liquid chromatography plus quadrupole
60 1094 time-of-flight mass spectrometry and molecular docking, *J. Sep. Sci.* 42 (2019) 3611–
1095 3620. <https://doi.org/10.1002/jssc.201900318>.
1096
1097
1098
1099 [29] C. Rajani, P. Borisa, T. Karanwad, Y. Borade, V. Patel, K. Rajpoot, R.K. Tekade,
Cancer-targeted chemotherapy: Emerging role of the folate anchored dendrimer as
drug delivery nanocarrier, in: *Pharm. Appl. Dendrimers*, Elsevier, 2020: pp. 151–198.
<https://doi.org/10.1016/B978-0-12-814527-2.00007-X>.
1100 [30] Ł. Cieśla, R. Moaddel, Comparison of analytical techniques for the identification of

- 1
2
3 1101 bioactive compounds from natural products, *Nat. Prod. Rep.* 33 (2016) 1131–1145.
4
5 1102 <https://doi.org/10.1039/C6NP00016A>.
6
7 1103 [31] R. Santos, O. Ursu, A. Gaulton, A.P. Bento, R.S. Donadi, C.G. Bologa, A. Karlsson,
8
9 1104 B. Al-Lazikani, A. Hersey, T.I. Oprea, J.P. Overington, A comprehensive map of
10
11 1105 molecular drug targets, *Nat. Rev. Drug Discov.* 16 (2017) 19–34.
12
13 1106 <https://doi.org/10.1038/nrd.2016.230>.
14
15 1107 [32] Z.C. Arituluk, J. Horne, B. Adhikari, J. Steltzner, S. Mansur, P. Ahirwar, S.E. Velu,
16
17 1108 N.E. Gray, L.M. Ciesla, Y. Bao, Identification of TrkB Binders from Complex
18
19 1109 Matrices Using a Magnetic Drug Screening Nanoplatfrom, *ACS Appl. Bio Mater.* 4
20
21 1110 (2021) 6244–6255. <https://doi.org/10.1021/acsabm.1c00552>.
22
23 1111 [33] C. Delbaldo, S. Faivre, C. Dreyer, E. Raymond, Sunitinib in advanced pancreatic
24
25 1112 neuroendocrine tumors: latest evidence and clinical potential, *Ther. Adv. Med. Oncol.*
26
27 1113 4 (2012) 9–18. <https://doi.org/10.1177/1758834011428147>.
28
29 1114 [34] N. Awasthi, R. Schwarz, Profile of nintedanib in the treatment of solid tumors: the
30
31 1115 evidence to date, *Onco. Targets. Ther.* (2015) 3691.
32
33 1116 <https://doi.org/10.2147/OTT.S78805>.
34
35 1117 [35] Y. Bu, X. Zhang, A. Zhu, L. Li, X. Xie, S. Wang, Inside-Out-Oriented Cell Membrane
36
37 1118 Biomimetic Magnetic Nanoparticles for High-Performance Drug Lead Discovery,
38
39 1119 *Anal. Chem.* 93 (2021) 7898–7907. <https://doi.org/10.1021/acs.analchem.1c00567>.
40
41 1120 [36] W. Wu, Q. He, C. Jiang, Magnetic Iron Oxide Nanoparticles: Synthesis and Surface
42
43 1121 Functionalization Strategies, *Nanoscale Res. Lett.* 3 (2008) 397.
44
45 1122 <https://doi.org/10.1007/s11671-008-9174-9>.
46
47 1123 [37] H. Park, V.H. Grassian, Commercially manufactured engineered nanomaterials for
48
49 1124 environmental and health studies: Important insights provided by independent
50
51 1125 characterization, *Environ. Toxicol. Chem.* 29 (2010) 715–721.
52
53 1126 <https://doi.org/10.1002/etc.72>.
54
55 1127 [38] M. Yasuda, D.R. Wilson, S.D. Fugmann, R. Moaddel, Synthesis and Characterization
56
57 1128 of SIRT6 Protein Coated Magnetic Beads: Identification of a Novel Inhibitor of SIRT6
58
59 1129 Deacetylase from Medicinal Plant Extracts, *Anal. Chem.* 83 (2011) 7400–7407.
60
1130 <https://doi.org/10.1021/ac201403y>.
1131 [39] N. Singh, S. Ravichandran, K. Spelman, S.D. Fugmann, R. Moaddel, The
1132
1133 1133 identification of a novel SIRT6 modulator from *Trigonella foenum-graecum* using
1134
1135 1134 ligand fishing with protein coated magnetic beads, *J. Chromatogr. B.* 968 (2014) 105–
1135
1136 1135 111. <https://doi.org/10.1016/j.jchromb.2014.03.016>.
1136
1137 [40] X. Liu, S. Hu, X. Chen, X. Bai, Hollow fiber cell fishing with high-performance liquid
1137
1138 1137 1138 chromatography for rapid screening and analysis of an antitumor-active
1138
1139 1138 protoberberine alkaloid group from *Coptis chinensis*, *J. Pharm. Biomed. Anal.* 98
(2014) 463–475. <https://doi.org/10.1016/j.jpba.2014.06.030>.

- 1
2
3 1139 [41] X. Xue, L. Li, X. Chen, S. Hu, X. Bai, Hollow fiber cell fishing with high performance
4 liquid chromatography for screening bioactive compounds from traditional Chinese
5 1140 medicines, *J. Chromatogr. A.* 1280 (2013) 75–83.
6 1141 <https://doi.org/10.1016/j.chroma.2013.01.033>.
7
8 1142
9
10 1143 [42] C.-P. Zhao, S.-J. Yin, G.-Y. Chen, Y. Wang, H. Chen, J. Zhao, F.-Q. Yang, Adsorbed
11 hollow fiber immobilized tyrosinase for the screening of enzyme inhibitors from
12 1144 *Pueraria lobata* extract, *J. Pharm. Biomed. Anal.* 193 (2021) 113743.
13 1145 <https://doi.org/10.1016/j.jpba.2020.113743>.
14 1146
15
16 1147 [43] F. Li, Y. Song, J. Wu, X. Chen, S. Hu, H. Zhao, X. Bai, Hollow fibre cell fishing and
17 hollow fibre liquid phase microextraction research on the anticancer coumarins of
18 1148 *Radix Angelicae dahuricae* in vitro and in vivo, *J. Liq. Chromatogr. Relat. Technol.*
19 1149 42 (2019) 79–88. <https://doi.org/10.1080/10826076.2019.1576141>.
20 1150
21
22 1151 [44] Y. Hu, A. Fu, Z. Miao, X. Zhang, T. Wang, A. Kang, J. Shan, D. Zhu, W. Li,
23 1152 Fluorescent ligand fishing combination with in-situ imaging and characterizing to
24 1153 screen Hsp 90 inhibitors from *Curcuma longa* L. based on InP/ZnS quantum dots
25 1154 embedded mesoporous nanoparticles, *Talanta.* 178 (2018) 258–267.
26 1155 <https://doi.org/10.1016/j.talanta.2017.09.035>.
27
28 1156 [45] Z. Miao, Y. Hu, X. Zhang, X. Yang, Y. Tang, A. Kang, D. Zhu, Screening and
29 1157 identification of ligand-protein interactions using functionalized heat shock protein
30 1158 90-fluorescent mesoporous silica-indium phosphide/zinc sulfide quantum dot
31 1159 nanocomposites, *J. Chromatogr. A.* 1562 (2018) 1–11.
32 1160 <https://doi.org/10.1016/j.chroma.2018.05.034>.
33 1161
34
35 1162 [46] Y. Hu, Z.-Y.Y. Miao, X.-J.J. Zhang, X.-T.T. Yang, Y.-Y.Y. Tang, S. Yu, C.-X.X.
36 1163 Shan, H.-M.M. Wen, D. Zhu, Preparation of Microkernel-Based Mesoporous (SiO₂-
37 1164 CdTe-SiO₂)@SiO₂ Fluorescent Nanoparticles for Imaging Screening and Enrichment
38 1165 of Heat Shock Protein 90 Inhibitors from *Tripterygium Wilfordii*, *Anal. Chem.* 90
39 1166 (2018) 5678–5686. <https://doi.org/10.1021/acs.analchem.7b05295>.
40
41
42 1167 [47] N. Xu, H. Yang, M. Cui, C. Wan, S. Liu, High-Performance Liquid Chromatography–
43 1168 Electrospray Ionization–Mass Spectrometry Ligand Fishing Assay: A Method for
44 1169 Screening Triplex DNA Binders from Natural Plant Extracts, *Anal. Chem.* 84 (2012)
45 1170 2562–2568. <https://doi.org/10.1021/ac202796v>.
46
47
48 1171 [48] M. Zhou, Y. Li, C. Huang, C. Wang, Q. Liu, Q. Deng, H. Zhang, T. Yin, G. Zhu, C.
49 1172 Wang, Y. Zhao, W. Zhang, Oriented Layered Graphene Oxide Pad Favoring High
50 1173 Loading Capacity and Stability for High-Throughput Chemical Screening, *Adv.*
51 1174 *Mater. Technol.* 7 (2022) 1–9. <https://doi.org/10.1002/admt.202101586>.
52
53
54 1175 [49] H. Yang, W. Yao, Y. Wang, L. Shi, R. Su, D. Wan, N. Xu, W. Lian, C. Chen, S. Liu,
55 1176 High-throughput screening of triplex DNA binders from complicated samples by 96-
56 1177 well plate format in conjunction with peak area-fading UHPLC-Orbitrap MS, *Analyst.*
57 142 (2017) 670–675. <https://doi.org/10.1039/C6AN01974A>.

- 1
2
3 1178 [50] H. Yang, Y. Wang, W. Yu, L. Shi, H. Wang, R. Su, C. Chen, S. Liu, Screening and
4 1179 investigation of triplex DNA binders from *Stephania tetrandra* S. Moore by a
5 1180 combination of peak area-fading ultra high-performance liquid chromatography with
6 1181 orbitrap mass spectrometry and optical spectroscopies, *J. Sep. Sci.* 41 (2018) 2878–
7 1182 2885. <https://doi.org/10.1002/jssc.201800190>.
- 1183 [51] Y. Tang, W. Qian, B. Zhang, W. Liu, X. Sun, W. Ji, L. Ma, D. Zhu, None-Loss Target
1184 Release of Biomimetic CaCO₃Nanocomposites for Screening Bioactive Components
1185 and Target Proteins, *ACS Appl. Bio Mater.* 4 (2021) 651–659.
1186 <https://doi.org/10.1021/acsabm.0c01197>.
- 1187 [52] C. Minnich, S. Hardy, S. Krämer, Stopping the Babylonian Confusion: An Updated
1188 Nomenclature for Process Analyzers in PAT Applications, *Chemie-Ingenieur-
1189 Technik.* 88 (2016) 694–697. <https://doi.org/10.1002/cite.201500188>.
- 1190 [53] E. Calleri, C. Temporini, R. Colombo, S. Tengattini, F. Rinaldi, G. Brusotti, S.
1191 Furlanetto, G. Massolini, Analytical settings for in-flow biocatalytic reaction
1192 monitoring, *TrAC - Trends Anal. Chem.* 143 (2021) 116348.
1193 <https://doi.org/10.1016/j.trac.2021.116348>.
- 1194 [54] S. Siddamurthi, G. Gutti, S. Jana, A. Kumar, S.K. Singh, Anthraquinone: a promising
1195 scaffold for the discovery and development of therapeutic agents in cancer therapy,
1196 *Future Med. Chem.* 12 (2020) 1037–1069. <https://doi.org/10.4155/fmc-2019-0198>.
- 1197 [55] E. Küpeli Akkol, Y. Genç, B. Karpuz, E. Sobarzo-Sánchez, R. Capasso, Coumarins
1198 and Coumarin-Related Compounds in Pharmacotherapy of Cancer, *Cancers (Basel).*
1199 12 (2020) 1959. <https://doi.org/10.3390/cancers12071959>.
- 1200 [56] T.A. Woyengo, V.R. Ramprasath, P.J.H. Jones, Anticancer effects of phytosterols,
1201 *Eur. J. Clin. Nutr.* 63 (2009) 813–820. <https://doi.org/10.1038/ejcn.2009.29>.
- 1202 [57] P. De, M. Baltas, F. Bedos-Belval, Cinnamic Acid Derivatives as Anticancer Agents-
1203 A Review, *Curr. Med. Chem.* 18 (2011) 1672–1703.
1204 <https://doi.org/10.2174/092986711795471347>.
- 1205 [58] C.-J. Weng, G.-C. Yen, Chemopreventive effects of dietary phytochemicals against
1206 cancer invasion and metastasis: Phenolic acids, monophenol, polyphenol, and their
1207 derivatives, *Cancer Treat. Rev.* 38 (2012) 76–87.
1208 <https://doi.org/10.1016/j.ctrv.2011.03.001>.
- 1209 [59] J. de Vasconcelos Cerqueira Braz, J.A. Carvalho Nascimento Júnior, M.R. Serafini,
1210 Terpenes with Antitumor Activity: A Patent Review, *Recent Pat. Anticancer. Drug
1211 Discov.* 15 (2020) 321–328. <https://doi.org/10.2174/1574892815666201002162315>.
- 1212 [60] E. Martino, G. Casamassima, S. Castiglione, E. Cellupica, S. Pantalone, F. Papagni,
1213 M. Rui, A.M. Siciliano, S. Collina, Vinca alkaloids and analogues as anti-cancer
1214 agents: Looking back, peering ahead, *Bioorg. Med. Chem. Lett.* 28 (2018) 2816–2826.
1215 <https://doi.org/10.1016/j.bmcl.2018.06.044>.

- 1
2
3 1216 [61] M. Abotaleb, S. Samuel, E. Varghese, S. Varghese, P. Kubatka, A. Liskova, D.
4 1217 Büsselberg, Flavonoids in Cancer and Apoptosis, *Cancers (Basel)*. 11 (2018) 28.
5 1218 <https://doi.org/10.3390/cancers11010028>.
6
7
8
9 1219 [62] Y. Zhou, J. Zheng, Y. Li, D.-P. Xu, S. Li, Y.-M. Chen, H.-B. Li, Natural Polyphenols
10 1220 for Prevention and Treatment of Cancer, *Nutrients*. 8 (2016) 515.
11 1221 <https://doi.org/10.3390/nu8080515>.
12
13
14
15 1222 [63] C. Bailly, The subgroup of 2'-hydroxy-flavonoids: Molecular diversity, mechanism of
16 1223 action, and anticancer properties, *Bioorg. Med. Chem.* 32 (2021) 116001.
17 1224 <https://doi.org/10.1016/j.bmc.2021.116001>.
18
19
20
21 1225 [64] A.N. Panche, A.D. Diwan, S.R. Chandra, Flavonoids: an overview, *J. Nutr. Sci.* 5
22 1226 (2016) e47. <https://doi.org/10.1017/jns.2016.41>.
23
24
25 1227 [65] R. Singh, R. Agarwal, Natural Flavonoids Targeting Deregulated Cell Cycle
26 1228 Progression in Cancer Cells, *Curr. Drug Targets*. 7 (2006) 345–354.
27 1229 <https://doi.org/10.2174/138945006776055004>.
28
29
30
31 1230 [66] A. Liskova, L. Koklesova, M. Samec, K. Smejkal, S.M. Samuel, E. Varghese, M.
32 1231 Abotaleb, K. Biringer, E. Kudela, J. Danko, M. Shakibaei, T.K. Kwon, D. Büsselberg,
33 1232 P. Kubatka, Flavonoids in Cancer Metastasis, *Cancers (Basel)*. 12 (2020) 1498.
34 1233 <https://doi.org/10.3390/cancers12061498>.
35
36
37
38 1234 [67] R. Gonçalves de Oliveira-Júnior, N. Marcoult-Fréville, G. Prunier, L. Beaugeard, E.
39 1235 Beserra de Alencar Filho, E.D. Simões Mourão, S. Michel, L.J. Quintans-Júnior, J.R.
40 1236 Guedes da Silva Almeida, R. Grougnet, L. Picot, Polymethoxyflavones from *Gardenia*
41 1237 *oudiepe* (Rubiaceae) induce cytoskeleton disruption-mediated apoptosis and sensitize
42 1238 BRAF-mutated melanoma cells to chemotherapy, *Chem. Biol. Interact.* 325 (2020)
43 1239 109109. <https://doi.org/10.1016/j.cbi.2020.109109>.
44
45
46
47
48
49 1240 [68] M. Bhardwaj, N.-H. Kim, S. Paul, R. Jakhar, J. Han, S.C. Kang, 5-Hydroxy-7-
50 1241 Methoxyflavone Triggers Mitochondrial-Associated Cell Death via Reactive Oxygen
51 1242 Species Signaling in Human Colon Carcinoma Cells, *PLoS One*. 11 (2016) e0154525.
52 1243 <https://doi.org/10.1371/journal.pone.0154525>.
53
54
55
56
57 1244 [69] M. Piedfer, S. Bouchet, R. Tang, C. Billard, D. Dauzonne, B. Bauvois, p70S6 kinase
58 1245 is a target of the novel proteasome inhibitor 3,3'-diamino-4'-methoxyflavone during
59 1246 apoptosis in human myeloid tumor cells, *Biochim. Biophys. Acta - Mol. Cell Res.*
60 1247 1833 (2013) 1316–1328. <https://doi.org/10.1016/j.bbamcr.2013.02.016>.
1248
1249 [70] F. Estévez-Sarmiento, M. Said, I. Brouard, F. León, C. García, J. Quintana, F. Estévez,
1250 3'-Hydroxy-3,4'-dimethoxyflavone blocks tubulin polymerization and is a potent
1251 apoptotic inducer in human SK-MEL-1 melanoma cells, *Bioorg. Med. Chem.* 25
(2017) 6060–6070. <https://doi.org/10.1016/j.bmc.2017.09.043>.
1252
1253 [71] L. Cheng, F. Wang, Y. Cao, C. Tong, Q. Wei, S. Shi, Y. Guo, Rapid profiling of
potential antitumor polymethoxylated flavonoids in natural products by integrating

- 1
2
3 1254 cell biospecific extraction with neutral loss/diagnostic ion filtering-based
4 1255 high-performance liquid chromatography–quadrupole time-of-flight tandem mass,
5 1256 *Phytochem. Anal.* 33 (2022) 895–905. <https://doi.org/10.1002/pca.3147>.
- 6
7
8
9 1257 [72] S.L. McGovern, E. Caselli, N. Grigorieff, B.K. Shoichet, A Common Mechanism
10 1258 Underlying Promiscuous Inhibitors from Virtual and High-Throughput Screening, *J.*
11 1259 *Med. Chem.* 45 (2002) 1712–1722. <https://doi.org/10.1021/jm010533y>.
- 12
13
14
15 1260 [73] D. Duan, A.K. Doak, L. Nedyalkova, B.K. Shoichet, Colloidal Aggregation and the in
16 1261 Vitro Activity of Traditional Chinese Medicines, *ACS Chem. Biol.* 10 (2015) 978–
17 1262 988. <https://doi.org/10.1021/cb5009487>.
- 18
19
20
21 1263 [74] D. Tritsch, C. Zinglé, M. Rohmer, C. Grosdemange-Billiard, Flavonoids: True or
22 1264 promiscuous inhibitors of enzyme? The case of deoxyxylulose phosphate
23 1265 reductoisomerase, *Bioorg. Chem.* 59 (2015) 140–144.
24 1266 <https://doi.org/10.1016/j.bioorg.2015.02.008>.
- 25
26
27
28 1267 [75] D. Reker, G.J.L. Bernardes, T. Rodrigues, Computational advances in combating
29 1268 colloidal aggregation in drug discovery, *Nat. Chem.* 11 (2019) 402–418.
30 1269 <https://doi.org/10.1038/s41557-019-0234-9>.
- 31
32
33
34 1270 [76] S.C. Owen, A.K. Doak, P. Wassam, M.S. Shoichet, B.K. Shoichet, Colloidal
35 1271 Aggregation Affects the Efficacy of Anticancer Drugs in Cell Culture, *ACS Chem.*
36 1272 *Biol.* 7 (2012) 1429–1435. <https://doi.org/10.1021/cb300189b>.
- 37
38
39
40 1273 [77] A. Mondal, A. Gandhi, C. Fimognari, A.G. Atanasov, A. Bishayee, Alkaloids for
41 1274 cancer prevention and therapy: Current progress and future perspectives, *Eur. J.*
42 1275 *Pharmacol.* 858 (2019) 172472. <https://doi.org/10.1016/j.ejphar.2019.172472>.
- 43
44
45
46 1276 [78] M. Tilaoui, H. Ait Mouse, A. Ziyad, Update and New Insights on Future Cancer Drug
47 1277 Candidates From Plant-Based Alkaloids, *Front. Pharmacol.* 12 (2021).
48 1278 <https://doi.org/10.3389/fphar.2021.719694>.
- 49
50
51
52 1279 [79] P. Kittakoop, C. Mahidol, S. Ruchirawat, Alkaloids as Important Scaffolds in
53 1280 Therapeutic Drugs for the Treatments of Cancer, Tuberculosis, and Smoking
54 1281 Cessation, *Curr. Top. Med. Chem.* 14 (2013) 239–252.
55 1282 <https://doi.org/10.2174/1568026613666131216105049>.
- 56
57
58
59 1283 [80] T. Isah, Anticancer alkaloids from trees: Development into drugs, *Pharmacogn. Rev.*
60 1284 10 (2016) 90. <https://doi.org/10.4103/0973-7847.194047>.
- 1285 [81] H. Malve, Exploring the ocean for new drug developments: Marine pharmacology, *J.*
1286 *Pharm. Bioallied Sci.* 8 (2016) 83. <https://doi.org/10.4103/0975-7406.171700>.
- 1287 [82] Y. Choi, K. Jermihov, S.-J. Nam, M. Sturdy, K. Maloney, X. Qiu, L.R. Chadwick, M.
1288 Main, S.-N. Chen, A.D. Mesecar, N.R. Farnsworth, G.F. Pauli, W. Fenical, J.M.
1289 Pezzuto, R.B. van Breemen, Screening Natural Products for Inhibitors of Quinone
1290 Reductase-2 Using Ultrafiltration LC–MS, *Anal. Chem.* 83 (2011) 1048–1052.

- 1
2
3 1291 <https://doi.org/10.1021/ac1028424>.
- 4
5 1292 [83] G.P. Rosa, W.R. Tavares, P.M.C. Sousa, A.K. Pagès, A.M.L. Seca, D.C.G.A. Pinto,
6 1293 Seaweed Secondary Metabolites with Beneficial Health Effects: An Overview of
7 1294 Successes in In Vivo Studies and Clinical Trials, *Mar. Drugs*. 18 (2019) 8.
8 1295 <https://doi.org/10.3390/md18010008>.
- 9
10
11
12
13 1296 [84] M.A. Hannan, A.A.M. Sohag, R. Dash, M.N. Haque, M. Mohibullah, D.F. Oktaviani,
14 1297 M.T. Hossain, H.J. Choi, I.S. Moon, Phytosterols of marine algae: Insights into the
15 1298 potential health benefits and molecular pharmacology, *Phytomedicine*. 69 (2020)
16 1299 153201. <https://doi.org/10.1016/j.phymed.2020.153201>.
- 17
18
19
20
21 1300 [85] C. Alves, M. Diederich, Marine Natural Products as Anticancer Agents, *Mar. Drugs*.
22 1301 19 (2021) 447. <https://doi.org/10.3390/md19080447>.
- 23
24
25 1302 [86] W.C. Hahn, J.S. Bader, T.P. Braun, A. Califano, P.A. Clemons, B.J. Druker, A.J.
26 1303 Ewald, H. Fu, S. Jagu, C.J. Kemp, W. Kim, C.J. Kuo, M.T. McManus, G. B. Mills, X.
27 1304 Mo, N. Sahni, S.L. Schreiber, J.A. Talamas, P. Tamayo, J.W. Tyner, B.K. Wagner,
28 1305 W.A. Weiss, D.S. Gerhard, V. Dancik, S. Gill, B. Hua, T. Sharifnia, V. Viswanathan,
29 1306 Y. Zou, F. Dela Cruz, A. Kung, B. Stockwell, J. Boehm, J. Dempster, R. Manguso, F.
30 1307 Vazquez, L.A.D. Cooper, Y. Du, A. Ivanov, S. Lonial, C.S. Moreno, Q. Niu, T.
31 1308 Owonikoko, S. Ramalingam, M. Reyna, W. Zhou, C. Grandori, I. Shmulevich, E.
32 1309 Swisher, J. Cai, I.S. Chan, M. Dunworth, Y. Ge, D. Georgess, E.M. Grasset, E.
33 1310 Henriët, H. Knútsdóttir, M.G. Lerner, V. Padmanaban, M.C. Perrone, Y. Suhail, Y.
34 1311 Tsehay, M. Warriër, Q. Morrow, T. Nechiporuk, N. Long, J. Saultz, A. Kaempf, J.
35 1312 Minnier, C.E. Tognon, S.E. Kurtz, A. Agarwal, J. Brown, K. Watanabe-Smith, T.Q.
36 1313 Vu, T. Jacob, Y. Yan, B. Robinson, E.F. Lind, Y. Kosaka, E. Demir, J. Estabrook, M.
37 1314 Grzadkowski, O. Nikolova, K. Chen, B. Deneen, H. Liang, M.C. Bassik, A.
38 1315 Bhattacharya, K. Brennan, C. Curtis, O. Gevaert, H.P. Ji, K.A.J. Karlsson, K.
39 1316 Karagyozyova, Y.-H. Lo, K. Liu, M. Nakano, A. Sathe, A.R. Smith, K. Spees, W.H.
40 1317 Wong, K. Yuki, M. Hangauer, D.S. Kaufman, A. Balmain, S.R. Bollam, W.-C. Chen,
41 1318 Q. Fan, K. Kersten, M. Krummel, Y.R. Li, M. Menard, N. Nasholm, C. Schmidt, N.K.
42 1319 Serwas, H. Yoda, A. Ashworth, S. Bandyopadhyay, T. Bivona, G. Eades, S. Oberlin,
43 1320 N. Tay, Y. Wang, J. Weissman, An expanded universe of cancer targets, *Cell*. 184
44 1321 (2021) 1142–1155. <https://doi.org/10.1016/j.cell.2021.02.020>.
- 45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60 1322 [87] T. Olivier, A. Haslam, V. Prasad, Anticancer Drugs Approved by the US Food and
1323 Drug Administration From 2009 to 2020 According to Their Mechanism of Action,
1324 *JAMA Netw. Open*. 4 (2021) e2138793.
1325 <https://doi.org/10.1001/jamanetworkopen.2021.38793>.
- 1326 [88] S. Zolghadri, A. Bahrami, M.T. Hassan Khan, J. Munoz-Munoz, F. Garcia-Molina, F.
1327 Garcia-Canovas, A.A. Saboury, A comprehensive review on tyrosinase inhibitors, *J.*
1328 *Enzyme Inhib. Med. Chem.* 34 (2019) 279–309.
1329 <https://doi.org/10.1080/14756366.2018.1545767>.

- 1
2
3 1330 [89] Z. Yang, Y. Zhang, L. Sun, Y. Wang, X. Gao, Y. Cheng, An ultrafiltration high-
4 1331 performance liquid chromatography coupled with diode array detector and mass
5 1332 spectrometry approach for screening and characterising tyrosinase inhibitors from
6 1333 mulberry leaves, *Anal. Chim. Acta.* 719 (2012) 87–95.
7 1334 <https://doi.org/10.1016/j.aca.2012.01.018>.
- 12 1335 [90] Z. Wang, S.H. Hwang, S.S. Lim, Comprehensive profiling of minor tyrosinase
13 1336 inhibitors from *Gastrodia elata* using an off-line hyphenation of ultrafiltration, high-
14 1337 speed countercurrent chromatography, and high-performance liquid chromatography,
15 1338 *J. Chromatogr. A.* 1529 (2017) 63–71. <https://doi.org/10.1016/j.chroma.2017.11.008>.
- 20 1339 [91] H. Liu, Y. Zhu, T. Wang, J. Qi, X. Liu, Enzyme-Site Blocking Combined with
21 1340 Optimization of Molecular Docking for Efficient Discovery of Potential Tyrosinase
22 1341 Specific Inhibitors from *Puerariae lobatae Radix*, *Molecules.* 23 (2018) 2612.
23 1342 <https://doi.org/10.3390/molecules23102612>.
- 27 1343 [92] Z. Wang, S.H. Hwang, B. Huang, S.S. Lim, Identification of tyrosinase specific
28 1344 inhibitors from *Xanthium strumarium* fruit extract using ultrafiltration-high
29 1345 performance liquid chromatography, *J. Chromatogr. B.* 1002 (2015) 319–328.
30 1346 <https://doi.org/10.1016/j.jchromb.2015.08.030>.
- 35 1347 [93] G. Zhang, X.-H. Guo, S.-S. Wang, Y.-Q. Li, G.-Z. Li, W.-J. Zhao, Screening and
36 1348 identification of natural ligands of tyrosinase from *Pueraria lobata* Ohwi by a
37 1349 combination of ultrafiltration and LC-MS, *Anal. Methods.* 9 (2017) 4858–4862.
38 1350 <https://doi.org/10.1039/C7AY00851A>.
- 42 1351 [94] X. Yin, X. Zhang, J. Yin, D. Kong, D. Li, Screening and identification of potential
43 1352 tyrosinase inhibitors from *Semen Oroxyli* extract by ultrafiltration LC-MS and in
44 1353 silico molecular docking, *J. Chromatogr. Sci.* 57 (2019) 838–846.
45 1354 <https://doi.org/10.1093/chromsci/bmz054>.
- 50 1355 [95] Y. Tao, D. Su, Y. Du, W. Li, B. Cai, L. Di, L. Shi, L. Hu, Magnetic solid-phase
51 1356 extraction coupled with HPLC-Q-TOF-MS for rapid analysis of tyrosinase binders
52 1357 from San-Bai decoction by Box–Behnken statistical design, *RSC Adv.* 6 (2016)
53 1358 109730–109741. <https://doi.org/10.1039/C6RA22045B>.
- 57 1359 [96] A.J. Vargas, S. Sittadjody, T. Thangasamy, E.E. Mendoza, K.H. Limesand, R. Burd,
58 1360 Exploiting Tyrosinase Expression and Activity in Melanocytic Tumors, *Integr. Cancer*
59 1361 *Ther.* 10 (2011) 328–340. <https://doi.org/10.1177/1534735410391661>.
- 1362 [97] Y. Pu, B. Zhou, H. Xiang, W. Wu, H. Yin, W. Yue, Y. Yin, H. Li, Y. Chen, H. Xu,
1363 Tyrosinase-activated prodrug nanomedicine as oxidative stress amplifier for
1364 melanoma-specific treatment, *Biomaterials.* 259 (2020) 120329.
1365 <https://doi.org/10.1016/j.biomaterials.2020.120329>.
- 1366 [98] B. Roulier, B. Pérès, R. Haudecoeur, Advances in the Design of Genuine Human
1367 Tyrosinase Inhibitors for Targeting Melanogenesis and Related Pigmentations, *J.*

- 1
2
3 1368 Med. Chem. 63 (2020) 13428–13443. <https://doi.org/10.1021/acs.jmedchem.0c00994>.
- 4
5 1369 [99] K. Subbaramaiah, L. Norton, W. Gerald, A.J. Dannenberg, Cyclooxygenase-2 Is
6 1370 Overexpressed in HER-2/neu-positive Breast Cancer, *J. Biol. Chem.* 277 (2002)
7 1371 18649–18657. <https://doi.org/10.1074/jbc.M111415200>.
- 8
9
10
11 1372 [100] X. Tang, Y.J. Sun, E. Half, M.T. Kuo, F. Sinicrope, Cyclooxygenase-2 overexpression
12 1373 inhibits death receptor 5 expression and confers resistance to tumor necrosis factor-
13 1374 related apoptosis-inducng ligand-induced apoptosis in human colon cancer cells,
14 1375 *Cancer Res.* 62 (2002) 4903–4908.
- 15
16
17
18 1376 [101] F. Hoellen, A. Waldmann, C. Banz-Jansen, A. Rody, M. Heide, F. Köster, J. Ribbat-
19 1377 Idel, C. Thorns, M. Gebhard, M. Oberländer, J.K. Habermann, M. Thill, Expression
20 1378 of cyclooxygenase-2 in cervical cancer is associated with lymphovascular invasion,
21 1379 *Oncol. Lett.* 12 (2016) 2351–2356. <https://doi.org/10.3892/ol.2016.4925>.
- 22
23
24
25 1380 [102] F.R. Khuri, H. Wu, J.J. Lee, B.L. Kemp, R. Lotan, S.M. Lippman, L. Feng, W.K.
26 1381 Hong, X.C. Xu, Cyclooxygenase-2 overexpression is a marker of poor prognosis in
27 1382 stage I non-small cell lung cancer, *Clin. Cancer Res.* 7 (2001) 861–867.
- 28
29
30
31 1383 [103] M.T. Rizzo, Cyclooxygenase-2 in oncogenesis, *Clin. Chim. Acta.* 412 (2011) 671–
32 1384 687. <https://doi.org/10.1016/j.cca.2010.12.026>.
- 33
34
35 1385 [104] N. Hashemi Goradel, M. Najafi, E. Salehi, B. Farhood, K. Mortezaee,
36 1386 Cyclooxygenase-2 in cancer: A review, *J. Cell. Physiol.* 234 (2019) 5683–5699.
37 1387 <https://doi.org/10.1002/jcp.27411>.
- 38
39
40
41 1388 [105] J. Huai, X. Zhao, S. Wang, L. Xie, Y. Li, T. Zhang, C. Cheng, R. Dai, Characterization
42 1389 and screening of cyclooxygenase-2 inhibitors from Zi-shen pill by affinity
43 1390 ultrafiltration-ultra performance liquid chromatography mass spectrometry, *J.*
44 1391 *Ethnopharmacol.* 241 (2019) 111900. <https://doi.org/10.1016/j.jep.2019.111900>.
- 45
46
47
48 1392 [106] X. Deng, S. Shi, S. Li, T. Yang, Magnetic ligand fishing combination with high-
49 1393 performance liquid chromatography–diode array detector–mass spectrometry to
50 1394 screen and characterize cyclooxygenase-2 inhibitors from green tea, *J. Chromatogr.*
51 1395 *B.* 973 (2014) 55–60. <https://doi.org/10.1016/j.jchromb.2014.10.010>.
- 52
53
54
55 1396 [107] D. Li, L. Xu, J. Qi, B. Yu, Screening and analysis of cyclooxygenase-2 inhibitors from
56 1397 the complex matrix: A case study to illustrate the important effect of immobilized
57 1398 enzyme activity in magnetic ligand fishing, *J. Pharm. Biomed. Anal.* 175 (2019)
58 1399 112795. <https://doi.org/10.1016/j.jpba.2019.112795>.
- 59
60
1400 [108] Y. Zhu, W. Wang, L. Jiang, H. Tan, Z. Liu, S. Jiang, Y. Tao, H. Wen, L. Mei, Targeted
1401 Separation of COX-2 Inhibitor from *Pterocephalus hookeri* Using Preparative High-
1402 Performance Liquid Chromatography Directed by the Affinity Solid-Phase Extraction
1403 HPLC System, *Molecules.* 26 (2021) 7395.
1404 <https://doi.org/10.3390/molecules26237395>.

- 1
2
3 1405 [109] Q. Chen, L. Zhu, K.M. Yip, Y. Tang, Y. Liu, T. Jiang, J. Zhang, Z. Zhao, T. Yi, H.
4 1406 Chen, A hybrid platform featuring nanomagnetic ligand fishing for discovering COX-
5 1407 2 selective inhibitors from aerial part of *Saussurea laniceps* Hand.-Mazz, J.
6 1408 *Ethnopharmacol.* 271 (2021) 113849. <https://doi.org/10.1016/j.jep.2021.113849>.
- 10 1409 [110] Y. Zhu, W. Wang, Z. Liu, S. Jiang, Y. Tao, L. Jiang, L. Mei, Comprehensive screening
11 1410 and separation of cyclooxygenase-2 inhibitors from *Pterocephalus hookeri* by affinity
12 1411 solid-phase extraction coupled with preparative high-performance liquid
13 1412 chromatography, *J. Chromatogr. B.* 1183 (2021) 122981.
14 1413 <https://doi.org/10.1016/j.jchromb.2021.122981>.
- 20 1414 [111] P. Saxena, P.K. Sharma, P. Purohit, A journey of celecoxib from pain to cancer,
21 1415 *Prostaglandins Other Lipid Mediat.* 147 (2020) 106379.
22 1416 <https://doi.org/10.1016/j.prostaglandins.2019.106379>.
- 26 1417 [112] M. Murakami, Y. Taketomi, Y. Miki, H. Sato, T. Hirabayashi, K. Yamamoto, Recent
27 1418 progress in phospholipase A2 research: From cells to animals to humans, *Prog. Lipid*
28 1419 *Res.* 50 (2011) 152–192. <https://doi.org/10.1016/j.plipres.2010.12.001>.
- 31 1420 [113] Z. Peng, Y. Chang, J. Fan, W. Ji, C. Su, Phospholipase A2 superfamily in cancer,
32 1421 *Cancer Lett.* 497 (2021) 165–177. <https://doi.org/10.1016/j.canlet.2020.10.021>.
- 36 1422 [114] J.N.M. Ilesley, M. Nakanishi, C. Flynn, G.S. Belinsky, S. De Guise, J.N. Adib, R.T.
37 1423 Dobrowsky, J. V. Bonventre, D.W. Rosenberg, Cytoplasmic Phospholipase A2
38 1424 Deletion Enhances Colon Tumorigenesis, *Cancer Res.* 65 (2005) 2636–2643.
39 1425 <https://doi.org/10.1158/0008-5472.CAN-04-3446>.
- 43 1426 [115] N. Wei, J. Zhao, G. Wu, W. Cao, P. Luo, Z. Zhang, G. Chen, L. Wen, Rapid Screening
44 1427 and Identification of Antitumor Ingredients from the Mangrove Endophytic Fungus
45 1428 Using an Enzyme-Immobilized Magnetic Nanoparticulate System, *Molecules.* 26
46 1429 (2021) 2255. <https://doi.org/10.3390/molecules26082255>.
- 51 1430 [116] F. Farabegoli, M. Vettrano, M. Manerba, L. Fiume, M. Roberti, G. Di Stefano,
52 1431 Galloflavin, a new lactate dehydrogenase inhibitor, induces the death of human breast
53 1432 cancer cells with different glycolytic attitude by affecting distinct signaling pathways,
54 1433 *Eur. J. Pharm. Sci.* 47 (2012) 729–738. <https://doi.org/10.1016/j.ejps.2012.08.012>.
- 58 1434 [117] K. Garber, Energy Deregulation: Licensing Tumors to Grow, *Science* (80-.). 312
59 1435 (2006) 1158–1159. <https://doi.org/10.1126/science.312.5777.1158>.
- 1436 [118] Y. Feng, Y. Xiong, T. Qiao, X. Li, L. Jia, Y. Han, Lactate dehydrogenase A: A key
1437 1438 player in carcinogenesis and potential target in cancer therapy, *Cancer Med.* 7 (2018)
1439 6124–6136. <https://doi.org/10.1002/cam4.1820>.
- 1440 [119] G. Cheng, Z. Pi, Z. Zheng, S. Liu, Z. Liu, F. Song, Magnetic nanoparticles-based
1441 1442 lactate dehydrogenase microreactor as a drug discovery tool for rapid screening
1443 inhibitors from natural products, *Talanta.* 209 (2020) 120554.
<https://doi.org/10.1016/j.talanta.2019.120554>.

- 1
2
3 1443 [120] J. Lewis, Anti-cancer effects of aloe-emodin: a systematic review, *J. Clin. Transl. Res.*
4 1444 (2017). <https://doi.org/10.18053/jctres.03.201703.001>.
- 5
6
7 1445 [121] C. Wu, H. Cao, H. Zhou, L. Sun, J. Xue, J. Li, Y. Bian, R. Sun, S. Dong, P. Liu, M.
8 1446 Sun, Research Progress on the Antitumor Effects of Rhein: Literature Review,
9 1447 *Anticancer. Agents Med. Chem.* 17 (2018).
10 1448 <https://doi.org/10.2174/1871520615666150930112631>.
- 11
12
13
14
15 1449 [122] X. Dong, Y. Zeng, Y. Liu, L. You, X. Yin, J. Fu, J. Ni, Aloe-emodin: A review of its
16 1450 pharmacology, toxicity, and pharmacokinetics, *Phyther. Res.* 34 (2020) 270–281.
17 1451 <https://doi.org/10.1002/ptr.6532>.
- 18
19
20
21 1452 [123] M.S. Lee, E.Y. Cha, J.Y. Sul, I.S. Song, J.Y. Kim, Chrysophanic acid blocks
22 1453 proliferation of colon cancer cells by inhibiting EGFR/mTOR pathway, *Phyther. Res.*
23 1454 25 (2011) 833–837. <https://doi.org/10.1002/ptr.3323>.
- 24
25
26
27 1455 [124] J. Wu, J. Wang, S. Wang, P. Rao, Lunatin, a novel lectin with antifungal and
28 1456 antiproliferative bioactivities from *Phaseolus lunatus* billb, *Int. J. Biol. Macromol.* 89
29 1457 (2016) 717–724. <https://doi.org/10.1016/j.ijbiomac.2016.04.092>.
- 30
31
32
33 1458 [125] S. Stadlbauer, C. Steinborn, A. Klemd, F. Hattori, K. Ohmori, K. Suzuki, R. Huber, P.
34 1459 Wolf, C. Gründemann, Impact of Green Tea Catechin ECG and Its Synthesized
35 1460 Fluorinated Analogue on Prostate Cancer Cells and Stimulated Immunocompetent
36 1461 Cells, *Planta Med.* 84 (2018) 813–819. <https://doi.org/10.1055/s-0044-102099>.
- 37
38
39
40 1462 [126] S.-C. Dong, H.-H. Sha, X.-Y. Xu, T.-M. Hu, R. Lou, H. Li, J.-Z. Wu, C. Dan, J. Feng,
41 1463 Glutathione S-transferase π : a potential role in antitumor therapy, *Drug Des. Devel.*
42 1464 *Ther.* Volume 12 (2018) 3535–3547. <https://doi.org/10.2147/DDDT.S169833>.
- 43
44
45
46 1465 [127] R.R. Singh, K.M. Reindl, Glutathione S-Transferases in Cancer, *Antioxidants.* 10
47 1466 (2021) 701. <https://doi.org/10.3390/antiox10050701>.
- 48
49
50
51 1467 [128] M. Cuendet, C.P. Oteham, R.C. Moon, J.M. Pezzuto, Quinone Reductase Induction as
52 1468 a Biomarker for Cancer Chemoprevention, *J. Nat. Prod.* 69 (2006) 460–463.
53 1469 <https://doi.org/10.1021/np050362q>.
- 54
55
56
57 1470 [129] B. Sun, J. Hoshino, K. Jermihov, L. Marler, J.M. Pezzuto, A.D. Mesecar, M. Cushman,
58 1471 Design, synthesis, and biological evaluation of resveratrol analogues as aromatase and
59 1472 quinone reductase 2 inhibitors for chemoprevention of cancer, *Bioorg. Med. Chem.*
60 1473 18 (2010) 5352–5366. <https://doi.org/10.1016/j.bmc.2010.05.042>.
- 1474 [130] T. -c. Hsieh, C.-J. Yang, C.-Y. Lin, Y.-S. Lee, J.M. Wu, Control of stability of cyclin
1475 D1 by quinone reductase 2 in CWR22Rv1 prostate cancer cells, *Carcinogenesis.* 33
1476 (2012) 670–677. <https://doi.org/10.1093/carcin/bgs016>.
- 1477 [131] G. Indrayanto, G.S. Putra, F. Suhud, Validation of in-vitro bioassay methods:
1478 Application in herbal drug research, in: 2021: pp. 273–307.
1479 <https://doi.org/10.1016/bs.podrm.2020.07.005>.

- 1
2
3 1480 [132] C. Boonlarppradab, C. Suriyachadkun, P. Rachtawee, W. Choowong,
4 1481 Saccharosporones A, B and C, cytotoxic antimalarial angucyclinones from
5 1482 Saccharopolyspora sp. BCC 21906, *J. Antibiot. (Tokyo)*. 66 (2013) 305–309.
6 1483 <https://doi.org/10.1038/ja.2013.16>.
- 7
8
9
10 1484 [133] S.K. Calderwood, M.A. Khaleque, D.B. Sawyer, D.R. Ciocca, Heat shock proteins in
11 1485 cancer: chaperones of tumorigenesis, *Trends Biochem. Sci.* 31 (2006) 164–172.
12 1486 <https://doi.org/10.1016/j.tibs.2006.01.006>.
- 13
14
15
16 1487 [134] C.W. Yun, H.J. Kim, J.H. Lim, S.H. Lee, Heat Shock Proteins: Agents of Cancer
17 1488 Development and Therapeutic Targets in Anti-Cancer Therapy, *Cells*. 9 (2019) 60.
18 1489 <https://doi.org/10.3390/cells9010060>.
- 19
20
21
22 1490 [135] Y. Guo, R. Wu, J.M. Gaspar, D. Sargsyan, Z.-Y. Su, C. Zhang, L. Gao, D. Cheng, W.
23 1491 Li, C. Wang, R. Yin, M. Fang, M.P. Verzi, R.P. Hart, A.-N. Kong, DNA methylome
24 1492 and transcriptome alterations and cancer prevention by curcumin in colitis-accelerated
25 1493 colon cancer in mice, *Carcinogenesis*. 39 (2018) 669–680.
26 1494 <https://doi.org/10.1093/carcin/bgy043>.
- 27
28
29
30
31 1495 [136] C. Wang, X. Song, M. Shang, W. Zou, M. Zhang, H. Wei, H. Shao, Curcumin exerts
32 1496 cytotoxicity dependent on reactive oxygen species accumulation in non-small-cell
33 1497 lung cancer cells, *Futur. Oncol.* 15 (2019) 1243–1253. [https://doi.org/10.2217/fon-](https://doi.org/10.2217/fon-2018-0708)
34 1498 [2018-0708](https://doi.org/10.2217/fon-2018-0708).
- 35
36
37
38
39 1499 [137] X. Sun, X. Zhong, W. Ma, W. Feng, Q. Huang, M. Ma, M. Lv, R. Hu, Z. Han, J. Li,
40 1500 X. Zhou, Germacrone induces caspase-3/GSDME activation and enhances ROS
41 1501 production, causing HepG2 pyroptosis, *Exp. Ther. Med.* 24 (2022) 456.
42 1502 <https://doi.org/10.3892/etm.2022.11383>.
- 43
44
45
46
47 1503 [138] R.N. Pillai, D.A. Fennell, V. Kovcin, T.-E. Ciuleanu, R. Ramlau, D. Kowalski, M.
48 1504 Schenker, I. Yalcin, F. Teofilovici, V.M. Vukovic, S.S. Ramalingam, Randomized
49 1505 Phase III Study of Ganetespib, a Heat Shock Protein 90 Inhibitor, With Docetaxel
50 1506 Versus Docetaxel in Advanced Non–Small-Cell Lung Cancer (GALAXY-2), *J. Clin.*
51 1507 *Oncol.* 38 (2020) 613–622. <https://doi.org/10.1200/JCO.19.00816>.
- 52
53
54
55
56 1508 [139] B. Birbo, E.E. Madu, C.O. Madu, A. Jain, Y. Lu, Role of HSP90 in Cancer, *Int. J.*
57 1509 *Mol. Sci.* 22 (2021) 10317. <https://doi.org/10.3390/ijms221910317>.
- 58
59
60 1510 [140] R.A.H. van de Ven, D. Santos, M.C. Haigis, Mitochondrial Sirtuins and Molecular
1511 Mechanisms of Aging, *Trends Mol. Med.* 23 (2017) 320–331.
1512 <https://doi.org/10.1016/j.molmed.2017.02.005>.
- 1513 [141] L.M. Garcia-Peterson, G. Guzmán-Pérez, C.R. Krier, N. Ahmad, The sirtuin 6: An
1514 overture in skin cancer, *Exp. Dermatol.* 29 (2020) 124–135.
1515 <https://doi.org/10.1111/exd.14057>.
- 1516 [142] J.A. Eble, S. Niland, The extracellular matrix in tumor progression and metastasis,
1517 *Clin. Exp. Metastasis*. 36 (2019) 171–198. [42](https://doi.org/10.1007/s10585-019-</p></div><div data-bbox=)

- 1
2
3 1518 09966-1.
4
5
6 1519 [143] S.J. Atkinson, T.S. Ellison, V. Steri, E. Gould, S.D. Robinson, Redefining the role(s)
7 1520 of endothelial $\alpha\beta 3$ -integrin in angiogenesis, *Biochem. Soc. Trans.* 42 (2014) 1590–
8 1521 1595. <https://doi.org/10.1042/BST20140206>.
- 11 1522 [144] R. Max, R.R.C.M. Gerritsen, P.T.G.A. Nooijen, S.L. Goodman, A. Sutter, U.
12 1523 Keilholtz, D.J. Ruiter, R.M.W. De Waal, Immunohistochemical analysis of integrin
14 1524 $\alpha\beta 3$ expression on tumor-associated vessels of human carcinomas, *Int. J. Cancer.* 71
16 1525 (1997) 320–324.
- 19 1526 [145] P.C. Brooks, A.M.P. Montgomery, M. Rosenfeld, R.A. Reisfeld, T. Hu, G. Klier, D.A.
20 1527 Cheresch, Integrin $\alpha\beta 3$ antagonists promote tumor regression by inducing apoptosis
22 1528 of angiogenic blood vessels, *Cell.* 79 (1994) 1157–1164.
24 1529 [https://doi.org/10.1016/0092-8674\(94\)90007-8](https://doi.org/10.1016/0092-8674(94)90007-8).
- 26 1530 [146] A.S. Berghoff, O. Rajky, F. Winkler, R. Bartsch, J. Furtner, J.A. Hainfellner, S.L.
28 1531 Goodman, M. Weller, J. Schittenhelm, M. Preusser, Invasion patterns in brain
29 1532 metastases of solid cancers, *Neuro. Oncol.* 15 (2013) 1664–1672.
31 1533 <https://doi.org/10.1093/neuonc/not112>.
- 34 1534 [147] T. Mikkelsen, C. Brodie, S. Finnis, M.E. Berens, J.L. Rennert, K. Nelson, N. Lemke,
35 1535 S.L. Brown, D. Hahn, B. Neuteboom, S.L. Goodman, Radiation sensitization of
37 1536 glioblastoma by cilengitide has unanticipated schedule-dependency, *Int. J. Cancer.*
39 1537 124 (2009) 2719–2727. <https://doi.org/10.1002/ijc.24240>.
- 41 1538 [148] C. V. Uliana, T.R. de Oliveira, M.R. Cominetti, R.C. Faria, Label-free evaluation of
43 1539 small-molecule–protein interaction using magnetic capture and electrochemical
45 1540 detection, *Anal. Bioanal. Chem.* 411 (2019) 2111–2119.
47 1541 <https://doi.org/10.1007/s00216-019-01636-1>.
- 49 1542 [149] A.C. Baptista Moreno Martin, R. Tomasin, L. Luna-Dulcey, A.E. Graminha, M.
51 1543 Araújo Naves, R.H.G. Teles, V.D. da Silva, J.A. da Silva, P.C. Vieira, B. Annabi,
52 1544 M.R. Cominetti, [10]-Gingerol improves doxorubicin anticancer activity and
54 1545 decreases its side effects in triple negative breast cancer models, *Cell. Oncol.* 43
56 1546 (2020) 915–929. <https://doi.org/10.1007/s13402-020-00539-z>.
- 58 1547 [150] E. Cocco, M. Scaltriti, A. Drilon, NTRK fusion-positive cancers and TRK inhibitor
59 1548 therapy, *Nat. Rev. Clin. Oncol.* 15 (2018) 731–747. [https://doi.org/10.1038/s41571-](https://doi.org/10.1038/s41571-018-0113-0)
1549 018-0113-0.
- 1550 [151] A. Lange, H.-W. Lo, Inhibiting TRK Proteins in Clinical Cancer Therapy, *Cancers*
1551 (Basel). 10 (2018) 105. <https://doi.org/10.3390/cancers10040105>.
- 1552 [152] A. Amatu, A. Sartore-Bianchi, S. Siena, NTRK gene fusions as novel targets of cancer
1553 therapy across multiple tumour types, *ESMO Open.* 1 (2016) e000023.
1554 <https://doi.org/10.1136/esmoopen-2015-000023>.

- 1
2
3 1555 [153] J. Meldolesi, Neurotrophin Trk Receptors: New Targets for Cancer Therapy, in: 2017:
4 1556 pp. 67–79. https://doi.org/10.1007/112_2017_6.
- 5
6
7 1557 [154] J.E. Frampton, Entrectinib: A Review in NTRK+ Solid Tumours and ROS1+ NSCLC,
8 1558 Drugs. 81 (2021) 697–708. <https://doi.org/10.1007/s40265-021-01503-3>.
- 9
10
11 1559 [155] A. Raja, I. Park, F. Haq, S.-M. Ahn, FGF19–FGFR4 Signaling in Hepatocellular
12 1560 Carcinoma, Cells. 8 (2019) 536. <https://doi.org/10.3390/cells8060536>.
- 13
14
15 1561 [156] S. Tang, Y. Hao, Y. Yuan, R. Liu, Q. Chen, Role of fibroblast growth factor receptor
16 1562 4 in cancer, Cancer Sci. 109 (2018) 3024–3031. <https://doi.org/10.1111/cas.13759>.
- 17
18
19 1563 [157] K.M. Levine, K. Ding, L. Chen, S. Oesterreich, FGFR4: A promising therapeutic
20 1564 target for breast cancer and other solid tumors, Pharmacol. Ther. 214 (2020) 107590.
21 1565 <https://doi.org/10.1016/j.pharmthera.2020.107590>.
- 22
23
24 1566 [158] Y. Bu, X. He, Q. Hu, C. Wang, X. Xie, S. Wang, A novel cell membrane affinity
25 1567 sample pretreatment technique for recognition and preconcentration of active
26 1568 components from traditional Chinese medicine, Sci. Rep. 7 (2017) 1–11.
27 1569 <https://doi.org/10.1038/s41598-017-03709-6>.
- 28
29
30 1570 [159] J. Huang, L. Feng, H. An, T. Zhang, Construction of Y376C-FGFR4 eukaryotic
31 1571 expression plasmid and its biological activity in HEK293 cell, Acta Biochim. Biophys.
32 1572 Sin. (Shanghai). 45 (2013) 889–892. <https://doi.org/10.1093/abbs/gmt091>.
- 33
34
35 1573 [160] A. Roidl, P. Foo, W. Wong, C. Mann, S. Bechtold, H.J. Berger, S. Streit, J.E. Ruhe,
36 1574 S. Hart, A. Ullrich, H.K. Ho, The FGFR4 Y367C mutant is a dominant oncogene in
37 1575 MDA-MB453 breast cancer cells, Oncogene. 29 (2010) 1543–1552.
38 1576 <https://doi.org/10.1038/onc.2009.432>.
- 39
40
41 1577 [161] J. Madunić, The Urokinase Plasminogen Activator System in Human Cancers: An
42 1578 Overview of Its Prognostic and Predictive Role, Thromb. Haemost. 118 (2018) 2020–
43 1579 2036. <https://doi.org/10.1055/s-0038-1675399>.
- 44
45
46 1580 [162] D.K. Morrison, MAP Kinase Pathways, Cold Spring Harb. Perspect. Biol. 4 (2012)
47 1581 a011254–a011254. <https://doi.org/10.1101/cshperspect.a011254>.
- 48
49
50 1582 [163] L. Tang, X. Han, The urokinase plasminogen activator system in breast cancer
51 1583 invasion and metastasis, Biomed. Pharmacother. 67 (2013) 179–182.
52 1584 <https://doi.org/10.1016/j.biopha.2012.10.003>.
- 53
54
55 1585 [164] H. Matthews, M. Ranson, M.J. Kelso, Anti-tumour/metastasis effects of the
56 1586 potassium-sparing diuretic amiloride: An orally active anti-cancer drug waiting for its
57 1587 call-of-duty?, Int. J. Cancer. 129 (2011) 2051–2061.
58 1588 <https://doi.org/10.1002/ijc.26156>.
- 59
60 1589 [165] L. Li, J. Kong, C. hua Yao, X. feng Liu, J. hua Liu, Rapid identification of urokinase
1590 plasminogen activator inhibitors from Traditional Chinese Medicines based on
1591 ultrafiltration, LC–MS and in silico docking, J. Pharm. Biomed. Anal. 164 (2019)

- 1
2
3 1592 241–248. <https://doi.org/10.1016/j.jpba.2018.10.036>.
- 4
5 1593 [166] O. Zitka, J. Kukacka, S. Krizkov, D. Huska, V. Adam, M. Masarik, R. Prusa, R. Kizek,
6 1594 Matrix Metalloproteinases, *Curr. Med. Chem.* 17 (2010) 3751–3768.
7 1595 <https://doi.org/10.2174/092986710793213724>.
- 8
9 1596 [167] M. Wang, T. Wang, S. Liu, D. Yoshida, A. Teramoto, The expression of matrix
10 1597 metalloproteinase-2 and-9 in human gliomas of different pathological grades, *Brain*
11 1598 *Tumor Pathol.* 20 (2003) 65–72. <https://doi.org/10.1007/BF02483449>.
- 12
13 1599 [168] H. Katori, A. Nozawa, M. Tsukuda, Increased expression of matrix metalloproteinase-
14 1600 2 and 9 and human papilloma virus infection are associated with malignant
15 1601 transformation of sinonasal inverted papilloma, *J. Surg. Oncol.* 93 (2006) 80–85.
16 1602 <https://doi.org/10.1002/jso.20386>.
- 17
18 1603 [169] A. Talvensaaari-Mattila, P. Pääkkö, T. Turpeenniemi-Hujanen, Matrix
19 1604 metalloproteinase-2 (MMP-2) is associated with survival in breast carcinoma, *Br. J.*
20 1605 *Cancer.* 89 (2003) 1270–1275. <https://doi.org/10.1038/sj.bjc.6601238>.
- 21
22 1606 [170] G.I. Murray, M.E. Duncan, P. O’Neil, J.A. McKay, W.T. Melvin, J.E. Fothergill,
23 1607 Matrix metalloproteinase-1 is associated with poor prognosis in oesophageal cancer,
24 1608 *J. Pathol.* 185 (1998) 256–261. [https://doi.org/10.1002/\(SICI\)1096-9896\(199807\)185:3<256::AID-PATH115>3.0.CO;2-A](https://doi.org/10.1002/(SICI)1096-9896(199807)185:3<256::AID-PATH115>3.0.CO;2-A).
- 25
26 1609 [171] Y. Tao, M. Pan, F. Zhu, Q. Liu, P. Wang, Construction of a Microfluidic Platform
27 1610 With Core-Shell CdSSe@ZnS Quantum Dot-Encoded Superparamagnetic Iron Oxide
28 1611 Microspheres for Screening and Locating Matrix Metalloproteinase-2 Inhibitors From
29 1612 Fruits of *Rosa roxburghii*, *Front. Nutr.* 9 (2022).
30 1613 <https://doi.org/10.3389/fnut.2022.869528>.
- 31
32 1614 [172] S.-T. Huang, C.-Y. Wang, R.-C. Yang, H.-T. Wu, S.-H. Yang, Y.-C. Cheng, J.-H.S.
33 1615 Pang, Ellagic Acid, the Active Compound of *Phyllanthus urinaria*, Exerts In Vivo
34 1616 Anti-Angiogenic Effect and Inhibits MMP-2 Activity, Evidence-Based Complement.
35 1617 *Altern. Med.* 2011 (2011) 1–10. <https://doi.org/10.1093/ecam/nep207>.
- 36
37 1618 [173] D. Wang, Q. Chen, Y. Tan, B. Liu, C. Liu, Ellagic acid inhibits human glioblastoma
38 1619 growth in vitro and in vivo, *Oncol. Rep.* 37 (2017) 1084–1092.
39 1620 <https://doi.org/10.3892/or.2016.5331>.
- 40
41 1621 [174] H.-M. Zhang, L. Zhao, H. Li, H. Xu, W.-W. Chen, L. Tao, Research progress on the
42 1622 anticarcinogenic actions and mechanisms of ellagic acid., *Cancer Biol. Med.* 11 (2014)
43 1623 92–100. <https://doi.org/10.7497/j.issn.2095-3941.2014.02.004>.
- 44
45 1624 [175] J. Losso, R. Bansode, A. Trappey, H. Bawadi, R. Truax, In vitro anti-proliferative
46 1625 activities of ellagic acid, *J. Nutr. Biochem.* 15 (2004) 672–678.
47 1626 <https://doi.org/10.1016/j.jnutbio.2004.06.004>.
- 48
49 1627 [176] M.M. Mulvihill, D.K. Nomura, Therapeutic potential of monoacylglycerol lipase
50 1628

- 1
2
3 1629 inhibitors, *Life Sci.* 92 (2013) 492–497. <https://doi.org/10.1016/j.lfs.2012.10.025>.
- 4
5 1630 [177] G.F. Grabner, R. Zimmermann, R. Schicho, U. Taschler, Monoglyceride lipase as a
6 1631 drug target: At the crossroads of arachidonic acid metabolism and endocannabinoid
7 1632 signaling, *Pharmacol. Ther.* 175 (2017) 35–46.
8 1633 <https://doi.org/10.1016/j.pharmthera.2017.02.033>.
- 9 1634 [178] J. Mei, R. Guo, F. Zhang, H. Zhang, X. Yang, B. Yu, J. Liu, X. Liu, Identification of
10 1635 bioactive natural products using yeast : Application to monoacylglycerol lipase
11 1636 inhibitor extraction from *Corydalis Rhizoma*, *Biomed. Pharmacother.* 149 (2022)
12 1637 112798. <https://doi.org/10.1016/j.biopha.2022.112798>.
- 13 1638 [179] Z. Xu, X. Chen, S. Fu, J. Bao, Y. Dang, M. Huang, L. Chen, Y. Wang,
14 1639 Dehydrocorydaline Inhibits Breast Cancer Cells Proliferation by Inducing Apoptosis
15 1640 in MCF-7 Cells, *Am. J. Chin. Med.* 40 (2012) 177–185.
16 1641 <https://doi.org/10.1142/S0192415X12500140>.
- 17 1642 [180] M. Kaushik, S. Kaushik, K. Roy, A. Singh, S. Mahendru, M. Kumar, S. Chaudhary,
18 1643 S. Ahmed, S. Kukreti, A bouquet of DNA structures: Emerging diversity, *Biochem.*
19 1644 *Biophys. Reports.* 5 (2016) 388–395. <https://doi.org/10.1016/j.bbrep.2016.01.013>.
- 20 1645 [181] J. Zhao, A. Bacolla, G. Wang, K.M. Vasquez, Non-B DNA structure-induced genetic
21 1646 instability and evolution, *Cell. Mol. Life Sci.* 67 (2010) 43–62.
22 1647 <https://doi.org/10.1007/s00018-009-0131-2>.
- 23 1648 [182] S. Mocellin, K.A. Pooley, D. Nitti, Telomerase and the search for the end of cancer,
24 1649 *Trends Mol. Med.* 19 (2013) 125–133. <https://doi.org/10.1016/j.molmed.2012.11.006>.
- 25 1650 [183] T. Tauchi, K. Shin-ya, G. Sashida, M. Sumi, S. Okabe, J.H. Ohyashiki, K. Ohyashiki,
26 1651 Telomerase inhibition with a novel G-quadruplex-interactive agent, telomestatin: in
27 1652 vitro and in vivo studies in acute leukemia, *Oncogene.* 25 (2006) 5719–5725.
28 1653 <https://doi.org/10.1038/sj.onc.1209577>.
- 29 1654 [184] T. Miyazaki, Y. Pan, K. Joshi, D. Purohit, B. Hu, H. Demir, S. Mazumder, S. Okabe,
30 1655 T. Yamori, M. Viapiano, K. Shin-ya, H. Seimiya, I. Nakano, Telomestatin Impairs
31 1656 Glioma Stem Cell Survival and Growth through the Disruption of Telomeric G-
32 1657 Quadruplex and Inhibition of the Proto-oncogene, c-Myb, *Clin. Cancer Res.* 18 (2012)
33 1658 1268–1280. <https://doi.org/10.1158/1078-0432.CCR-11-1795>.
- 34 1659 [185] Q. Shang, J.-F. Xiang, X.-F. Zhang, H.-X. Sun, L. Li, Y.-L. Tang, Fishing potential
35 1660 antitumor agents from natural plant extracts pool by dialysis and G-quadruplex
36 1661 recognition, *Talanta.* 85 (2011) 820–823.
37 1662 <https://doi.org/10.1016/j.talanta.2011.04.011>.
- 38 1663 [186] L. Yuan, P.-L. Xu, Q. Zeng, Y.-M. Liu, L.-S. Ding, X. Liao, Preparation of ds-DNA
39 1664 functionalized magnetic nanobaits for screening of bioactive compounds from
40 1665 medicinal plant, *Mater. Sci. Eng. C.* 56 (2015) 401–408.
41 1666 <https://doi.org/10.1016/j.msec.2015.06.050>.

- 1
2
3 1667 [187] Y. Gao, H. Peng, L. Li, F. Wang, J. Meng, H. Huang, S. Wang, P.C.H. Li, Y. Sun,
4 1668 Screening of high-efficiency and low-toxicity antitumor active components in
5 1669 *Macleaya cordata* seeds based on the competitive effect of drugs on double targets by
6 a new laminar flow chip, *Analyst*. 146 (2021) 4934–4944.
7 1670 <https://doi.org/10.1039/D1AN00754H>.
8 1671
9
10 1672 [188] Q. Cai, J. Meng, Y. Ge, Y. Gao, Y. Zeng, H. Li, Y. Sun, Fishing antitumor ingredients
11 1673 by G-quadruplex affinity from herbal extract on a three-phase-laminar-flow
12 microfluidic chip, *Talanta*. 220 (2020) 121368.
13 1674 <https://doi.org/10.1016/j.talanta.2020.121368>.
14 1675
15
16 1676 [189] A. Ali, S. Bhattacharya, DNA binders in clinical trials and chemotherapy, *Bioorg.*
17 1677 *Med. Chem.* 22 (2014) 4506–4521. <https://doi.org/10.1016/j.bmc.2014.05.030>.
18 1678
19
20 1678 [190] L. He, G. Yang, X. Geng, Enzymatic activity and chromatographic characteristics of
21 1679 the cell membrane immobilized on silica surface, *Chinese Sci. Bull.* 44 (1999) 826–
22 1680 831. <https://doi.org/10.1007/BF02885029>.
23
24
25 1681 [191] R. Zhang, S. Hu, X. Chen, X. Bai, Screening and Research of Anti-Cancer Matrine
26 1682 Components Based on Hollow Fiber Cell Fishing with High-Performance Liquid
27 1683 Chromatography, *Chromatographia*. 79 (2016) 125–136.
28 1684 <https://doi.org/10.1007/s10337-015-3006-8>.
29
30
31 1685 [192] Y. Li, Y. Chen, H. Zhang, C.W.K. Lam, Z. Li, C. Wang, Y. Zhao, W. Zhang, Z. Jiang,
32 1686 Immobilization of cell membrane onto a glucose-Zn-based porous coordination
33 1687 polymer and its application to rapid screening of potentially active compounds from
34 1688 *Vaccinium corymbosum* L. leaves, *Microchim. Acta*. 187 (2020).
35 1689 <https://doi.org/10.1007/s00604-020-04612-0>.
36
37
38 1690 [193] F. Wang, L. Cheng, Y. Cao, Q. Wei, C. Tong, S. Shi, Online extraction and enrichment
39 1691 coupling with high-speed counter-current chromatography for effective and target
40 1692 isolation of antitumor anthraquinones from seeds of *Cassia obtusifolia*, *J. Sep. Sci.* 45
41 1693 (2022) 938–944. <https://doi.org/10.1002/jssc.202100775>.
42
43
44 1694 [194] Y. Yan, Y. Hao, S. Hu, X. Chen, X. Bai, Hollow fibre cell fishing with high
45 1695 performance liquid chromatography for screening bioactive anthraquinones from
46 1696 traditional Chinese medicines, *J. Chromatogr. A*. 1322 (2013) 8–17.
47 1697 <https://doi.org/10.1016/j.chroma.2013.10.084>.
48
49
50 1698 [195] C. Wang, S. Hu, X. Chen, X. Bai, Screening and quantification of anticancer
51 1699 compounds in traditional Chinese medicine by hollow fiber cell fishing and hollow
52 1700 fiber liquid/solid-phase microextraction, *J. Sep. Sci.* 39 (2016) 1814–1824.
53 1701 <https://doi.org/10.1002/jssc.201600103>.
54
55
56 1702 [196] D. Wu, X. Chen, S. Hu, X. Bai, Study on major antitumor components in Yinchenhao
57 1703 decoction in vitro and in vivo based on hollow fiber cell fishing coupled with high
58 1704 performance liquid chromatography, *J. Chromatogr. B Anal. Technol. Biomed. Life*

- 1
2
3 1705 Sci. 1060 (2017) 118–125. <https://doi.org/10.1016/j.jchromb.2017.06.003>.
4
5 1706 [197] Q. Hu, X. Zhang, L. Jia, X. Zhen, X. Pan, X. Xie, S. Wang, Engineering biomimetic
6 1707 graphene nanodecoys camouflaged with the EGFR/HEK293 cell membrane for
7 1708 targeted capture of drug leads, *Biomater. Sci.* 8 (2020) 5690–5697.
8
9 1709 <https://doi.org/10.1039/D0BM00841A>.
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
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27
28
29
30
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| Species | Part | Extraction | Technique & Target | Ligands Fished | Chemical Class | Ref |
|--|-----------|---|----------------------------------|--|--------------------------------------|-------|
| <i>Aconitum szechenyianum</i> Gay (Ranunculaceae) | Roots | Reflux – EtOH 60% | Miscellaneous (FGFR4) | 1) Napellonine, 2) 12-Epinapelline, 3) Benzoylaconitine and 4) 3-Deoxyaconitine | Alkaloids | [158] |
| <i>Acorus tatarinowii</i> Schott (Acoraceae) | Undefined | Ultrasonication – EtOH 70% | Miscellaneous (MAGL) | - | - | [178] |
| <i>Actinomyces sp.</i> (Actinomycetaceae) | Whole | Liquid-liquid extraction – acetone-EtOAc-H ₂ O | Ultrafiltration (QR-2) | 1) Tetrangulol methyl ether | Anthraquinone | [82] |
| <i>Alisma orientale</i> (Sam.) Juz. (Alismataceae) | Roots | Ultrasonication – MeOH:H ₂ O (7:3) | Magnetic particle (GST) | - | - | [28] |
| <i>Alisma plantago-aquatica</i> L. (Alismataceae) | Undefined | Reflux – EtOH 85% | Miscellaneous (Hsp 90 α) | 1) Alisol F, 2) 16-oxo-11-deoxy-Alisol A, 3) Alisol F 24-acetate, 4) Alisol L 23-acetate, 5) 16-oxo-11-anhydro-Alisol A, 6) Alisol H, 7) Alisol B-23-acetate, 8) Alisol C, 9) 3-Methyl-5,6-dimethoxy-2,3-dihydro-1H-indene-1-one, 10) (1S,3 α ,7 α β)-1-Acetyloctahydro-4 α -hydroxy-7 β -isopropyl 4-methyl-1H-indene, 11) Corymbolone, 12) (-)-Spathulenol, 13) n-Hexadecanoic acid, 14) Hexadecanoic acid ethyl ester, 15) Linoleic acid ethyl ester, 16) 13,17-Seco-5 α -pregn-13(18)-en-20-one, 17) Lupeol, 18) Lupeol acetate, 19) Taraxasterol and 20) Betulinaldehyde | Fatty acids, steroids and terpenoids | [45] |
| <i>Andrographis paniculata</i> (Burm. f.) Nees (Acanthaceae) | Undefined | Ultrasonication – EtOH 70% | Miscellaneous (MAGL) | - | - | [178] |

| | | | | | | |
|--|-------------------|---|---------------------------------------|--|--|-------|
| <i>Anemarrhena asphodeloides</i> Bunge (Asparagaceae) | Rhizomes | Percolation – EtOH 95% | Ultrafiltration (COX2) | 1) Neomangiferin, 2) Phellodendrine, 3) Mangiferin, 4) Magnoflorine, 5) 3-O-feruloylquinnic acid, 6) (–)-Oblongine, 7) Menisperine, 8) Demethyleneberberine, 9) Oxoberberine, 10) Berberrubine, 11) Jatrorrhizine, 12) Timosaponin B, 13) Timosaponin B-II, 14) Timosaponin A-I, 15) Palmatine, 16) Berberine, 17) Sanleng acid, 18) Timosaponin A-III, 19) Anemarrhenasaponin I, 20) cis-Hinokiresinol | Alkaloids, xanthenes and saponins | [105] |
| <i>Angelicae dahurica</i> Fisch. (Apiaceae) | Roots | Ultrasonication – MeOH | Hollow fiber (HepG-2 cells) | 1) Xanthotoxol, 2) Xanthotoxin, 3) Psoralen, 4) Bergapten, 5) Oxypeucedanin, 6) Imperatorin, 7) Isoimperatorin. | Coumarins | [43] |
| <i>Artemisia capillaris</i> Thunb (Asteraceae) | Undefined | Decoction – H ₂ O | Hollow fiber (HepG-2 and ACHN cells) | 1) Chlorogenic acid, 2) p-Hydroxyacetophenone | Phenolic acid and terpenoid | [196] |
| <i>Asarum heterotropoides</i> F. Schmidt. (Aristolochiaceae) | Undefined | Ultrasonication – EtOH 70% | Miscellaneous (MAGL) | - | - | [178] |
| <i>Astragalus membranaceus</i> (Fisch.) Bunge (Fabaceae) | Undefined | Ultrasonication – MeOH | Magnetic particle (EGFR/HEK293 cells) | - | - | [197] |
| <i>Atractylodes macrocephala</i> Koidz (Asteraceae) | Roots | Infusion – H ₂ O | Magnetic particle (Tyrosinase) | 1) Gallic acid, 2) Albiflorin, 3) Paeoniflorin, 4) Liquiritin apioside, 5) Liquiritin, 6) Galloylpaeoniflorin, 7) Ononin, 8) Isoliquiritigenin, 9) Glycyrrhizic acid, 10) Oxypaeoniflora, 11) Benzoylpaeoniflorin, 12) Benzoyloxypaeoniflorin, 13) Mudanpioside C, 14) Paeonolide and 15) Apiopaeonoside | Phenolic acid, terpenoids saponins, and glycosides | [95] |
| <i>Bufo melanostictus</i> Schneider (Bufonidae) | Secretion (venom) | Reflux – EtOH and H ₂ O | Miscellaneous (HepG2 and MCF-7 cells) | 1) Serotonin, 2) N-methyl serotonin, 3) Bufotenidine, 4) Suberoyl arginine, 5) Arenobufagin, 6) Telocinobufatoxin, 7) Bufotalin, 8) 3-(N-pimeloyl argininyl) bufalin, 9) Cinobufotalin, 10) Bufalitoxin, 11) 3-(N-pimeloyl argininyl) resibufogenin, 12) Bufalin, 13) Resibufotoxin, 14) Cinobufotoxin, 15) 3-(N-azelayl argininyl) resibufogenin, 16) Cinobufagin and 17) Resibufogenin | Alkaloids, amino acid, bufogenins and bufotoxins | [11] |
| <i>Camellia sinensis</i> (L.) Kuntze (Theaceae) | Leaves | Ultrasonication – MeOH:H ₂ O (7:3) | Magnetic particle (GST) | 1) Caffeic acid, 2) Rosmarinic acid, 3) Gallic acid, 4) (–)-Epigallocatechin-3-O-gallate, 5) (–)-Epicatechin-3-gallate and 6) Quercetin-3-O-glucoside | Phenolic acids and flavonoids | [28] |

| | | | | | | | |
|--|---|-----------|---|---|---|--|-------|
| 1 2 3 4 5 6 | <i>Camellia sinensis</i> (L.) Kuntze (Theaceae) | Leaves | - | Magnetic particle (COX2) | 1) (-)-gallocatechin, 2) (-)-epigallocatechin, 3) (-)-epicatechin, 4) (-)-epigallocatechin-3-gallate, 5) (-)-gallocatechin-3-gallate, 6) (-)-epigallocatechin-3-(3''-O-methyl)-gallate, 7) (-)-epicatechin-3-gallate, 8) (-)-epicatechin-3-(3''-O-methyl)-gallate | Flavonoids | [106] |
| 7 8 9 10 11 12 | <i>Camellia sinensis</i> (L.) Kuntze (Theaceae) | Leaves | Ultrasonication – MeOH 70% | Magnetic particle (COX2) | 1) 3-Galloylquinic acid, 2) 5-Galloylquinic acid, 3) 3-p-Coumaroylquinic acid, 4) 5-p-Coumaroylquinic acid, 5) 5-caffeoylquinic acid, 6) (-)-Epicatechin, 7) (-)-Epicatechin 3-O-gallate, 8) Quercetin 3-O-rhamnosylgalactoside, 9) Quercetin 3-O-glucoside, 10) Kaempferol 3-O-glucosylrutinoside, 11) Kaempferol-3-O-rutinoside, 12) Kaempferol 3-O-glucoside | Phenolic acids and flavonoids | [107] |
| 13 14 15 16 17 18 19 | <i>Carthamus tinctorius</i> L. (Asteraceae) | Undefined | Ultrasonication – EtOH 70% | Miscellaneous (MAGL) | - | - | [178] |
| 20 21 22 23 24 25 26 27 | <i>Cassia obtusifolia</i> L. (Fabaceae) | Seeds | Decoction – EtOH 75% | Miscellaneous (HepG2 cells) | 1) Aurantio-obtusin, 2) 1-desmethyllaurantio-obtusin, 3) Chryso-obtusin, 4) Obtusin, and 5) Questin | Anthraquinones | [193] |
| 28 29 30 31 32 | <i>Cecropia obtusifolia</i> Bertol. (Urticaceae) | Undefined | Reflux – MeOH; Ultrasound – HCl 8%; Liquid- liquid – HCl- CHCl ₃ | Hollow fiber (HCT116 cells) | 1) Aloe-emodin and 2) Physcion | Anthraquinones | [194] |
| 33 34 35 36 | <i>Centella asiatica</i> (L.) Urban (Apiaceae) | Undefined | - | Magnetic particle (TrkB) | 1) Dicafeoylquinic acid, 2) O-cafeoylquinic acid, 3) Madecassic acid, 4) Castillicetin, 5) Quercetin, 6) Stigmasterol, 7) Pomolic acid, 8) Naringin | Phenolic acids, terpenoids, steroid and flavonoids | [32] |
| 37 38 39 40 | <i>Cibotium barometz</i> (L.) J.Sm. (Cibotiaceae) | Rhizomes | Ultrasonication – MeOH | Magnetic particle (EGFR/HEK293 cells) | - | - | [197] |
| 41 42 43 | <i>Cinnamomum cassia</i> (L.) J.Presl (Lauraceae) | Undefined | Ultrasonication – EtOH 70% | Miscellaneous (MAGL) | - | - | [178] |

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|---|-------------------|---|--|---|---|-------|
| <i>Cinnamomum cassia</i> (L.) J.Presl (Lauraceae) | Cortex | Percolation – EtOH 95% | Ultrafiltration (COX2) | 1) Neomangiferin, 2) Phellodendrine, 3) Mangiferin, 4) Magnoflorine, 5) 3-O-feruloylquinnic acid, 6) (–)-Oblongine, 7) Menisperine, 8) Demethyleneberberine, 9) Oxoberberine, 10) Berberrubine, 11) Jatrorrhizine, 12) Timosaponin B, 13) Timosaponin B-II, 14) Timosaponin A-I, 15) Palmatine, 16) Berberine, 17) Sanleng acid, 18) Timosaponin A-III, 19) Anemarrhenasaponin I, 20) cis-Hinokiresinol | Alkaloids, xanthenes and saponins | [105] |
| <i>Citrus reticulata</i> Blanco (Rutaceae) | Fruits (peels) | Percolation – EtOH 75% | Miscellaneous (MCF-7 cells) | 1) 5,7,8,30,40-Pentamethoxyflavanone, 2) 3,5,6,7,8,40-Hexamethoxyflavanone, 3) 5,7,8,30,40,50-Hexamethoxyflavone, 4) 7-Hydroxy-5,6,8,30,40-pentamethoxyflavone, 5) Isosinensetin, 6) 30-Hydroxy-5,6,7,8,40-pentamethoxyflavone or 40-Hydroxy-5,6,7,8,30-pentamethoxyflavone, 7) Sinensetin, 8) 5,6,7,8,40-Pentamethoxyflavanone, 9) 3-Hydroxy-5,6,7,8,40-pentamethoxyflavone, 10) Tetramethyl-O-isoscutellarein, 11) Tetramethyl-O-scutellarein, 12) Nobiletin, 13) 3,5,6,7,8,30,40-Heptamethoxyflavone, 14) 5,6,7,8,30,40-Hexamethoxyflavanone, 15) 5-Hydroxy-6,7,8,30,40-pentamethoxyflavone, 16) Tangeretin | Flavonoids | [71] |
| <i>Cnidium monnieri</i> (L.) Cuss. (Apiaceae) | Fruits | Reflux – MeOH | Hollow fiber (SGC7901, RAW264.7 and SHSY5Y cells) | 1) Psoralen, 2) Bergapten, 3) Imperatorin and 4) Osthole | Coumarins | [41] |
| <i>Coptis chinensis</i> Franch. (Ranunculaceae) | Roots | Solid-liquid extraction – MeOH; Liquid- liquid partition – EtOAc-H ₂ O | Ultrafiltration (DNA) | 1) Berberine, 2) Coptisine and 3) Palmitine | Alkaloids | [12] |
| <i>Coptis chinensis</i> Franch. (Ranunculaceae) | Rhizomes | Ultrasonication – EtOH 75% | Miscellaneous (DNA) | 1) Epiberberine, 2) Coptisine, 3) Jatrorrhizine, 4) Berberrubine and 5) Columbamine | Alkaloids | [49] |
| <i>Coptis chinensis</i> Franch. (Ranunculaceae) | Rhizomes | - | Magnetic particle (DNA) | 1) Columbamine, 2) Epiberberine, 3) Jatrorrhizine, 4) Palmatine and 5) Berberine | Alkaloids | [186] |
| <i>Coptis chinensis</i> Franch. (Ranunculaceae) | Undefined | Decoction – MeOH 20% | Hollow fiber (MCF7, MADB106, and SGC7901 cells) | 1) Jatrorrhizine, 2) Epiberine, 3) Coptisine, 4) Palmatine, 5) Berberine | Alkaloids | [40] |

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|--|--|-----------|---|---|--|---|-------|
| 1 2 3 | <i>Corydalis yanhusuo</i> W. T. Wang (Papaveraceae) | Rhizomes | Ultrasonication – EtOH 70% | Miscellaneous (MAGL) | 1) Tetrahydropalmatin, 2) Tetrahydrocoptisine, 3) Coptisine, 4) Fumaricine, 5) Palmatine, 6) Berberine and 7) Dehydrocorydaline | Alkaloids | [178] |
| 4 5 6 | <i>Curcuma longa</i> L. (Zingiberaceae) | Rhizomes | Maceration and ultrasonication – MeOH | Miscellaneous (PC3-cells) | 1) Curcumin, 2) Demethoxycurcumin and 3) Bisdemethoxycurcumin | Phenolic pigments | [20] |
| 7 8 9 10 | <i>Curcuma longa</i> L. (Zingiberaceae) | Rhizomes | Percolation – EtOH 95% | Miscellaneous (Hsp 90 α) | 1) Zingiberenol, 2) Ar-turmerone, 3) Curlone, 4) (6R,7R)-bisabolone, 5) (E)-Atlantone, 6) 6-(1-Hydroxymethylvinyl), 7) 4,8a-dimethyl-3,5,6,7,8,8a-hexahydro-1H-naphthalen-2-one | Terpenoids | [44] |
| 11 12 13 14 | <i>Curcuma longa</i> L. (Zingiberaceae) | Rhizomes | Ultrasonication - MeOH | Hollow fiber (HepG-2, SKOV-3, and ACHN cells) | 1) Bisdemethoxycurcumin, 2) Demethoxycurcumin and 3) Curcumin | Phenolic pigments | [195] |
| 15 16 17 18 19 | <i>Cynanchum otophyllum</i> C.K.Schneid. (Apocynaceae) | Undefined | Ultrasonication – MeOH | Magnetic particle (EGFR/HEK293 cells) | - | - | [197] |
| 20 21 22 | <i>Eucommia ulmoides</i> Oliv. (Eucommiaceae) | Undefined | Ultrasonication – EtOH 70% | Miscellaneous (MAGL) | - | - | [178] |
| 23 24 25 26 | <i>Gardenia jasminoides</i> J. Ellis (Rubiaceae) | Undefined | Decoction – H ₂ O | Hollow fiber (HepG-2 and ACHN cells) | 1) Chlorogenic acid, 2) Geniposide, 3) Crocin | Phenolic acid, glycoside and carotenoid | [196] |
| 27 28 29 30 31 32 33 34 35 36 37 | <i>Gastrodia elata</i> Blume. (Orchidiaceae) | Undefined | - | Ultrafiltration (Tyrosinase) | 1) 12-15 Heneicosadienoic acid, Octadecanoic acid, 2- hydroxylpropyl esters, 2) Unknown, 3) 3-(4-tert-butylphenyl)-2-methylpropanal, 4) 4,4'-Dihydroxybenzylsulfid, 5) Octadecanoic acid, 2-hydroxy-1-(hydroxymethyl)ethyl ester 6) 9-Octadecenamamide, (trans-), 7) Unknown, 8) Unknown, 9) 9-Octadecenamamide, (cis-), 10) 4,4'-Dihydroxydiphenyl ether, 11) 4-(1-(4-hydroxyphenyl)phenyl)phenol, 12) Octadecanoic acid, 2,3-dihydroxypropyl ester, 13) 4,4'-Dimethoxydiphenylmethane, 14) 4,4'-Methylenediphenol, 15) 2,4-Bis(4-hydroxybenzyl) phenol, 16) 4-Hydroxybenzyl methyl ether | Fatty acids, amides, polyphenols and purines | [90] |
| 38 39 40 41 42 | <i>Ginkgo biloba</i> L. (Ginkgoaceae) | Undefined | Ultrasonication – EtOH 70% | Miscellaneous (MAGL) | - | - | [178] |

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|----------------------------------|--|----------|--|---|---|--|-------|
| 1 2 3 4 5 6 | <i>Glycyrrhiza uralensis</i> Fisch. (Fabaceae) | Roots | Percolation – EtOH 95%; Liquid-liquid partition – H ₂ O- Petroleum ether- EtOAc-n-BuOH | Magnetic particle (Tyrosinase) | 1) Liquiritin apioside, 2) Neolicucuroside, 3) Liquiritigenin, 4) Licorice saponin, 5) Chrysoeriol, 6) Dihydrodaidzein, 7) Formononetin, 8) Glycyrrhisoflavanone, 9) Glycyrrhizic acid, 10) Glycycoumarin, 11) Pratensein | Flavonoids, saponin and coumarin | [18] |
| 7 8 9 10 11 12 | <i>Glycyrrhiza uralensis</i> Fisch. (Fabaceae) | Roots | Infusion – H ₂ O:Honey (85%) | Magnetic particle (Tyrosinase) | 1) Gallic acid, 2) Albiflorin, 3) Paeoniflorin, 4) Liquiritin apioside, 5) Liquiritin, 6) Galloylpaeoniflorin, 7) Ononin, 8) Isoliquiritigenin, 9) Glycyrrhizic acid, 10) Oxypaeoniflora, 11) Benzoylpaeoniflorin, 12) Benzoyloxypaeoniflorin, 13) Mudanpioside C, 14) Paeonolide and 15) Apiopaeonoside | Phenolic acid, terpenoid saponins, and glycosides | [95] |
| 13 14 15 16 | <i>Humulus lupulus</i> L. (Cannabaceae) | Flowers | Liquid-liquid extraction - MeOH-CHCl ₃ | Ultrafiltration (QR-2) | 1) Xanthohumol and 2) Xanthohumol D | Flavonoids | [82] |
| 17 18 19 20 | <i>Ligusticum chuanxiong</i> Hort. (Apiaceae) | - | Ultrasonication – EtOH 70% | Miscellaneous (MAGL) | - | - | [178] |
| 21 22 23 24 25 | <i>Ligustrum lucidum</i> W.T.Aiton (Oleaceae) | Fruits | Ultrasonication – MeOH | Magnetic particle (EGFR/HEK293 cells) | - | - | [197] |
| 26 27 28 29 30 31 | <i>Lithospermum erythrorhizon</i> Siebold & Zucc. (Boraginaceae) | Rhizomes | Reflux- Deionized H ₂ O | Miscellaneous (A549 and H1819 cells) | 1) Salvianolic acid B, 2) Deoxyshikonin and 3) Shikonin | Phenolic acid and quinones | [48] |
| 32 33 34 35 | <i>Macleaya cordata</i> (Willd.) R. Br. (Papaveraceae) | Seeds | Ultrasonication – EtOH (85%, pH 2) | Miscellaneous (DNA) | 1) Sanguinarine, 2) Chelerythrine, 3) Protopine and 4) Allocryptopine | Alkaloids | [187] |
| 36 37 38 39 40 41 | <i>Macleaya cordata</i> (Willd.) R.Br. (Papaveraceae) | Seeds | Ultrasonication – MeOH (85%, pH 2) | Ultrafiltration (DNA) | 1) Chelerythrine, 2) Sanguinarine, 3) Protopine and 4) Allocryptopine | Alkaloids | [188] |

| | | | | | | | |
|---|---|-----------|--|---|--|---|-------|
| 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 | <i>Mangifera indica</i> L. (Anacardiaceae) | Leaves | Ultrasonication – EtOH (75%)- EtOAc- <i>n</i> -BuOH | Ultrafiltration (Tyrosinase) | <p>1) Gallic acid, 2) Iriflophenone 3-C-glucoside, 3) Mangiferin, 4) Protocatechuic acid, 5) Iriflophenone 3-C-(2'-O-galloyl)-glucoside, 6) 6'-O-Galloyl-mangiferin, 7) Maclurin 3-C-(2'-O-p-hydroxybenzoyl)-glucoside, 8) Iriflophenone 3-C-(2',6'-di-O-galloyl)-glucoside, 9) Hyperoside, 10) Isoquercitrin, 11) Ethyl gallate, 12) Iriflophenone 3-C-(2'-O-p-hydroxybenzoyl)-glucoside, 13) Quercetin-3-O-xyloside, 14) 3-O-Galloyl shikimic acid, 15) 3-O-Galloyl quinic acid, 16) Maclurin 3-C-(2'-O-galloyl)-glucoside, 17) 3,5-Di-O-galloyl quinic acid, 18) 5-O-Digalloyl quinic acid, 19) Digallic acid, 20) Isomangiferin, 21) 1,4,6-Tri-O-galloyl glucoside, 22) Maclurin 3-C-(2',3'-di-O-galloyl)-glucoside, 23) 1,3-Digalloyl acetone, 24) 1,3,4,6-Tetra-O-galloyl glucoside, 25) 6'-O-(p-Hydroxybenzoyl) mangiferin, 26) Epicatechin gallate, 27) 1,2,3,4,6-Penta-O-galloyl glucoside, 28) Iriflophenone 3-C-(2',3',6'-tri-O-galloyl)-glucoside, 29) Luteolin-7-O-glucoside, 30) Quercetin-3-O-arapyranoside, 31) Quercetin-3-O-arafuranoside, 32) Kaempferol-3-O-glucoside, 33) Quercetin-3-O-rhamnoside, 34) Ethyl 2,4-dihydroxy-3-galloyl oxybenzoate, 35) Ethyl digallate, 36) 7-O-Methyl quercetin-3-O-rhamnoside</p> | Phenolic acids, xanthenes, benzophenones, flavonoids, quinic and shikimic acid derivatives | [19] |
| 26 27 28 29 30 | <i>Menispermum dauricum</i> DC. (Menispermaceae) | Undefined | Solid-liquid extraction – MeOH; Liquid- liquid partition – EtOAc-H ₂ O | Ultrafiltration (DNA) | 1) Daurisoline and 2) Dauricine | Alkaloids | [12] |
| 31 32 33 34 35 36 37 38 | <i>Morus alba</i> L. (Moraceae) | Leaves | Solid-liquid extraction – EtOH 50%; Liquid-liquid partition – H ₂ O- petroleum ether- EtOAc | Ultrafiltration (Tyrosinase) | <p>1) Neochlorogenic acid, 2) Chlorogenic acid, 3) Cryptochlorogenic acid, 4) Rutin, 5) Isoquercitrin, 6) Kaempferol-3-O-α-L-rhamnopyranosyl-(1–6)-β-D-glucopyranoside, 7) Quercetin-3-O-(6-O-malonyl)-β-D-glucopyranoside, 8) 3,4-Dicaffeoylquinic acid, 9) Astragalinalin, 10) 3,5-Dicaffeoylquinic acid, 11) 4,5-Dicaffeoylquinic acid, 12) Kaempferol-3-O-(6-O-malonyl)-β-D-glucopyranoside</p> | Phenolic acids and flavonoids | [89] |
| 39 40 41 42 | <i>Morus alba</i> L. (Moraceae) | Fruits | Ultrasonication – MeOH | Magnetic particle (EGFR/HEK293 cells) | - | - | [197] |

| | | | | | | |
|--|--------------------------|---|-----------------------------------|--|---|-------|
| <i>Oroxylum indicum</i> L. (Bignoniaceae) | Seeds | Ultrasonication – MeOH 50% | Miscellaneous (Tyrosinase) | 1) Oroxin B, 2) Kaempferid-3-O-beta-D-gentiobioside, 3) Chrysin-7-O-beta-D-gentiobioside, 4) Oroxin A, 5) Baicalein and 6) Chrysin | Flavonoids | [94] |
| <i>Otholobium pubescens</i> (Poir.) J.W. Grimes (Fabaceae) | Leaves and flowers | Percolation – CH ₂ Cl ₂ -MeOH- MeOH: H ₂ O (1:1)-H ₂ O | Miscellaneous (Tyrosinase) | 1) Daidzin, 2) Isoorientin, 3) Isovitexin, 4) Daidzein, 5) Genistein, 6) 3-(5-hydroxybenzofuran-6-yl) propanoic acid | Flavonoids | [17] |
| <i>Paeonia lactiflora</i> Pall (Paeoniaceae) | Roots | Infusion – H ₂ O | Magnetic particle (Tyrosinase) | 1) Gallic acid, 2) Albiflorin, 3) Paeoniflorin, 4) Liquiritin apioside, 5) Liquiritin, 6) Galloylpaeoniflorin, 7) Ononin, 8) Isoliquiritigenin, 9) Glycyrrhizic acid, 10) Oxypaeoniflora, 11) Benzoylpaeoniflorin, 12) Benzoyloxypaeoniflorin, 13) Mudanpioside C, 14) Paeonolide and 15) Apiopaeonoside | Phenolic acid, terpenoids saponins, and glycosides | [95] |
| <i>Paris polyphylla</i> Sm. (Melanthiaceae) | Undefined | Ultrasonication – EtOH 70% | Miscellaneous (MAGL) | - | - | [178] |
| <i>Perilla frutescens</i> (L.) Britt (Lamiaceae) | Leaves | Ultrasonication – MeOH:H ₂ O (7:3) | Magnetic particle (GST) | 1) Caffeic acid, 2) Rosmarinic acid, 3) Gallic acid, 4) (-)-Epigallocatechin-3-O-gallate, 5) (-)-Epicatechin-3-gallate and 6) Quercetin-3-O-glucoside | Phenolic acids and flavonoids | [28] |
| <i>Phellodendron chinense</i> C.K.Schneid. (Ranunculaceae) | Cortex | Ultrasonication – EtOH 75% | Miscellaneous (DNA) | 1) Epiberberine, 2) Coptisine, 3) Jatrorrhizine, 4) Berberrubine and 5) Columbamine | Alkaloids | [49] |
| <i>Phellodendron chinense</i> C.K.Schneid. (Rutaceae) | Barks | Ultrasonication – EtOH 75% | Miscellaneous (DNA) | 1) Berberine and 2) Palmatine | Alkaloids | [47] |
| <i>Phellodendron chinense</i> C.K.Schneid. (Rutaceae) | Cortex | Percolation – EtOH 95% | Ultrafiltration (COX2) | 1) Neomangiferin, 2) Phellodendrine, 3) Mangiferin, 4) Magnoflorine, 5) 3-O-feruloylquinnic acid, 6) (-)-Oblongine, 7) Menisperine, 8) Demethyleneberberine, 9) Oxoberberine, 10) Berberrubine, 11) Jatrorrhizine, 12) Timosaponin B, 13) Timosaponin B-II, 14) Timosaponin A-I, 15) Palmatine, 16) Berberine, 17) Sanleng acid, 18) Timosaponin A-III, 19) Anemarrhenasaponin I, 20) cis-Hinokiresinol | Alkaloids, xanthenes and saponins | [105] |

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|----|---|-----------|---|---|--|--|-------|
| 1 | <i>Phellodendron chinense</i> Schneid. (Rutaceae) | Barks | Infusion – EtOH:H ₂ O (1:3) | Dialysis (DNA) | 1) Berberine | Alkaloid | [185] |
| 2 | | | | | | | |
| 3 | | | | | | | |
| 4 | <i>Polygonum cuspidatum</i> Siebold & Zucc. (Polygonaceae) | Undefined | Ultrasonication – EtOH 70% | Magnetic particle (LDH) | 1) (-)-Epicatechin-3-O-gallate, 2) Trihydroxystilbene-3-O-(6''-galloyl)-glucoside, 3) Emodin-1-O-glucoside, 4) Emodin-8-O-(6-malonyl)-glucoside, 5) Torachryson sulfate, 6) Emodin | Flavonoid, stilbenoid, naphthol and anthraquinones | [119] |
| 5 | | | | | | | |
| 6 | | | | | | | |
| 7 | | | | | | | |
| 8 | | | | | | | |
| 9 | <i>Polygonum cuspidatum</i> Siebold & Zucc. (Polygonaceae) | Rhizomes | Reflux – CHCl ₃ :H ₂ SO ₄ (5:4) | Hollow fiber (HCT116 cells) | 1) Aloe-emodin, 2) Emodin and 3) Physcion | Anthraquinones | [194] |
| 10 | | | | | | | |
| 11 | | | | | | | |
| 12 | | | | | | | |
| 13 | | | | | | | |
| 14 | <i>Polygonum multiflorum</i> (Thunb.) Moldenke (Polygonaceae) | Rhizomes | Reflux – MeOH; Ultrasound – HCl 8%; Liquid-liquid – HCl-CHCl ₃ | Hollow fiber (HCT116 cells) | 1) Aloe-emodin, 2) Emodin, 3) Physcion, 4) Rhein and 5) Chrysophanol | Anthraquinones | [194] |
| 15 | | | | | | | |
| 16 | | | | | | | |
| 17 | | | | | | | |
| 18 | | | | | | | |
| 19 | <i>Pseudopithomyces sp.</i> (Didymosphaeriaceae) | Whole | Solid-liquid extraction – MeOH-EtOAc | Magnetic particle (PLA2) | 1) Cyclo-(4-hydroxyl-Pro-Leu), 2) Cyclo-(Pro-Val), 3) Fusaristatin C, 4) Ergosterol peroxide, 5) Ergosterol and 6) Cerevisterol | Cyclic peptides and Steroids | [115] |
| 20 | | | | | | | |
| 21 | | | | | | | |
| 22 | | | | | | | |
| 23 | <i>Psoralea corylifolia</i> L. (Fabaceae) | Fruits | Reflux – MeOH | Hollow fiber (SGC7901, RAW264.7 and SHSY5Y cells) | 1) Psoralen and 2) Isopsoralen | Coumarins | [41] |
| 24 | | | | | | | |
| 25 | | | | | | | |
| 26 | | | | | | | |
| 27 | <i>Pterocephalus hookeri</i> (C.B. Clarke) Diels (Caprifoliaceae) | Undefined | Percolation – EtOH 95% | Miscellaneous (COX2) | 1) Swertisin, 2) Scopoletin | Flavonoid and coumarin | [110] |
| 28 | | | | | | | |
| 29 | | | | | | | |
| 30 | | | | | | | |
| 31 | <i>Pterocephalus hookeri</i> (C.B. Clarke) Diels (Caprifoliaceae) | Whole | Percolation – EtOH 95% | Miscellaneous (COX2) | 1) Sylvestroside I | Terpenoid | [108] |
| 32 | | | | | | | |
| 33 | | | | | | | |
| 34 | | | | | | | |
| 35 | <i>Pueraria lobata</i> (Willd.) Ohwi (Fabaceae) | Undefined | Reflux – EtOH 75% | Hollow fiber (Tyrosinase) | 1) Daidzein, 2) 3'-Methoxy puerarin, 3) 3'-Hydroxy puerarin 4) Puerarin, 5) Puerarin 6''-O-xyloside 6) Puerarin-4'-O-glucoside, 7) Puerarin aposide | Flavonoids | [42] |
| 36 | | | | | | | |
| 37 | | | | | | | |
| 38 | | | | | | | |
| 39 | <i>Pueraria lobata</i> (Willd.) Ohwi (Fabaceae) | Roots | Ultrasonication – MeOH 75% | Ultrafiltration (Tyrosinase) | 1) Puerarin, 2) Mirificin, 3) Daidzin and 4) Genistin | Flavonoids | [91] |
| 40 | | | | | | | |
| 41 | | | | | | | |
| 42 | | | | | | | |
| 43 | | | | | | | |
| 44 | | | | | | | |
| 45 | | | | | | | |
| 46 | | | | | | | |

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|--|-----------------|--|--|---|--|-------|
| <i>Pueraria lobata</i> (Willd.) Ohwi (Fabaceae) | Roots | Reflux – EtOH:H ₂ O (75%); Liquid- liquid extraction – petroleum ether-EtOAc | Ultrafiltration (Tyrosinase) | 1) 3'-Hydroxypuerarin, 2) Puerarin, 3) Puerarin-6"-O-xyloside, 4) Daidzin, 5) Genistin, 6) 6"-O-acetyldaidzin and 7) Daidzein | Flavonoids | [93] |
| <i>Radix et Rhizoma Rhei</i> (Polygonaceae) | Roots | Decoction – H ₂ O | Hollow fiber (HepG-2 and ACHN cells) | 1) Rhein | Anthraquinone | [196] |
| <i>Rheum palmatum</i> L. (Polygonaceae) | Undefined | Ultrasonication – EtOH 70% | Miscellaneous (MAGL) | - | - | [178] |
| <i>Rheum rhabarbarum</i> L. (Polygonaceae) | Undefined | Ultrasonication – EtOH 70% | Magnetic particle (LDH) | 1) (-)-Epicatechin-3-O-gallate, 2) ω-Hydroxyemodin, 3) Emodin-1-O-glucoside, 4) Lunatin, 5) Chrysophanic acid, 6) Rhein-O-(acetyl)-glucoside, 7) Aloe emodin, 8) Rhein and 9) Emodin | Flavonoids and anthraquinones | [119] |
| <i>Rhus chinensis</i> Mill. (Anacardiaceae) | Undefined | Ultrasonication – MeOH 75% | Miscellaneous (uPA) | 1) Gallic acid, 2) Methyl gallate, 3) Pentagalloylglucose, 4) Ellagic acid, 5) Ziyu-glycoside I, 6) 28-O-β-D-glucopyranosyl pomolic acid, and 7) 1β-Hydroxyeuscapic acid | Phenolic acid, tannins, saponins and terpenoid | [165] |
| <i>Rosa roxburghii</i> Tratt. (Rosaceae) | Fruits | Ultrasonication – EtOH 70% | Miscellaneous (MMP-2) | 1) Oreganol A, 2) Corilagin, 3) Ellagic acid, 4) Zebirioside O, 5) Niga-ichigoside F1, 6) Peduncloside, 7) Rosamultin, 8) Kajiichigoside F1, 9) Potentillanoside A, 10) Quadranoside VIII, 11) Valerenic acid and 12) Medicagenic acid | Phenolic compound, tannin, phenolic acid, saponin and terpenoids | [171] |
| <i>Salvia miltiorrhiza</i> Bge. (Lamiaceae) | Undefined | Ultrasonication – EtOH 70% | Miscellaneous (MAGL) | - | - | [178] |
| <i>Sanguisorba officinalis</i> L. (Rosaceae) | Undefined | Ultrasonication – MeOH 75% | Miscellaneous (uPA) | 1) Gallic acid, 2) Methyl gallate, 3) Pentagalloylglucose, 4) Ellagic acid, 5) Ziyu-glycoside I, 6) 28-O-β-D-glucopyranosyl pomolic acid, and 7) 1β-Hydroxyeuscapic acid | Phenolic acid, tannins, saponins and terpenoid | [165] |
| <i>Saussurea laniceps</i> Hand.-Mazz. (Asteraceae) | Aerial parts | Reflux – H ₂ O | Magnetic particle (COX2) | 1) Skimmin, 2) Scopolin, 3) Umbelliferone, 4) Scopoletin | Coumarins | [109] |
| <i>Schisandra chinensis</i> (Turcz) | Undefined | Ultrasonication - MeOH | Hollow fiber | 1) Schisandrol A, 2) Schizandrin A, 3) Schisantherin A and 4) Schizandrin C | Lignans and tannin | [41] |

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|----|-----------------------------|-----------|--|---|--|--|-------|
| 1 | (Schisandraceae) | | | (SGC7901, RAW264.7, SHSY5Y cells) | | | |
| 2 | | | | | | | |
| 3 | | | | | | | |
| 4 | <i>Scutellaria</i> | Undefined | Ultrasonication – | Magnetic particle | 1) Baicalin, 2) Baicalein, 3) Wogonin and 4) Oroxylin A | Flavonoids | [26] |
| 5 | <i>baicalensis</i> Georgi | | EtOH 70% | (Tyrosinase) | | | |
| 6 | (Lamiaceae) | | | | | | |
| 7 | <i>Scutellaria barbata</i> | Undefined | Ultrasonication – | Magnetic particle | - | - | [197] |
| 8 | D. Don | | MeOH | (EGFR/HEK293 | | | |
| 9 | (Lamiaceae) | | | cells) | | | |
| 10 | | | | | | | |
| 11 | <i>Selaginella</i> | Undefined | Reflux – EtOH | Magnetic particle | 1) Amentoflavone and 2) Robustaflavone | Flavonoids | [25] |
| 12 | <i>doederleinii</i> Hieron. | | 70% and EtAc | (LDH) | | | |
| 13 | (Selaginellaceae) | | | | | | |
| 14 | | | | | | | |
| 15 | <i>Smilax china</i> L. | Rhizomes | Infusion – H ₂ O | Magnetic particle | 1) Gallic acid, 2) Albiflorin, 3) Paeoniflorin, 4) Liquiritin apioside, 5) Liquiritin, 6) Galloylpaeoniflorin, 7) Ononin, 8) Isoliquiritigenin, 9) Glycyrrhizic acid, 10) Oxypaeoniflora, 11) Benzoylpaeoniflorin, 12) Benzoyloxypaeoniflorin, 13) Mudanpioside C, 14) Paeonolide and 15) Apiopaeonoside | Phenolic acid, terpenoids saponins, and glycosides | [95] |
| 16 | (Smilacaceae) | | | (Tyrosinase) | | | |
| 17 | | | | | | | |
| 18 | | | | | | | |
| 19 | | | | | | | |
| 20 | <i>Sophora flavescens</i> | Roots | Infusion – H ₂ O | Hollow fiber | 1) Oxymatrine, 2) Matrine, 3) Sophoridine and 4) Oxysophocarpine | Alkaloids | [191] |
| 21 | Aiton | | | (MCF-7, ACHN, | | | |
| 22 | (Fabaceae) | | | SKOV-3, and HepG-2 cells) | | | |
| 23 | | | | | | | |
| 24 | <i>Sophora flavescens</i> | Undefined | Ultrasonication – | Magnetic particle | 1) Kurarinone and 2) Sophoraflavanone G | Flavonoids | [26] |
| 25 | Aiton | | EtOH 70% | (Tyrosinase) | | | |
| 26 | (Fabaceae) | | | | | | |
| 27 | | | | | | | |
| 28 | <i>Stephania tetrandra</i> | Undefined | Solid-liquid | Ultrafiltration | 1) Fangchinoline and 2) Tetrandrine | Alkaloids | [12] |
| 29 | S. Moore | | extraction – | | | | |
| 30 | (Menispermaceae) | | MeOH; Liquid-liquid partition – EtOAc-H ₂ O | (DNA) | | | |
| 31 | | | | | | | |
| 32 | | | | | | | |
| 33 | <i>Stephania tetrandra</i> | Undefined | Ultrasonication – | Miscellaneous | 1) Fangchinoline and 2) Tetrandrine | Alkaloids | [50] |
| 34 | S. Moore | | EtOH 75% | (DNA) | | | |
| 35 | (Menispermaceae) | | | | | | |
| 36 | | | | | | | |
| 37 | <i>Streptomyces</i> | Whole | Liquid-liquid | Ultrafiltration | 1) Actinomycin D, 2) Actinomycin V and 3) Actinomycin X0β | Actinomycins | [12] |
| 38 | <i>antibioticus</i> | | extraction - | | | | |
| 39 | (Streptomycetaceae) | | Medium-EtOAc | (DNA) | | | |
| 40 | | | | | | | |
| 41 | <i>Strychnos nux-vomica</i> | Undefined | Reflux – | Magnetic particle | 1) 10,11-dimethoxystrychnine and 2) Loganetin | Alkaloid and terpenoid | [35] |
| 42 | L. | | EtOH (60%, v/v) | (EGFR/A549 cells) | | | |
| 43 | (Loganiaceae) | | | | | | |

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|--|---|-----------|--|--|--|------------------------------------|-------|
| 1 2 3 4 | <i>Taraxacum mongolicum</i> Hand.-Mazz (Asteraceae) | Undefined | Ultrasonication – MeOH | Magnetic particle (EGFR/HEK293 cells) | 1) Luteolin and 2) Caffeic acid | Flavonoid and phenolic acid | [197] |
| 5 6 7 | <i>Trigonella foenum-graecum</i> L. (Fabaceae) | Seeds | Maceration – EtOH 72% | Magnetic particle (SIRT6) | 1) Orientin | Flavonoid | [39] |
| 8 9 10 | <i>Trigonella foenum-graecum</i> L. (Fabaceae) | Seeds | - | Magnetic particle (SIRT6) | 1) Quercetin and 2) Vitexin | Flavonoids | [38] |
| 11 12 13 14 | <i>Tripterygium wilfordii</i> Hook. f. (Celastraceae) | Undefined | Reflux – EtOH | Miscellaneous (MCF-7 cells) | 1) Genistein, 2) Tripteryl E-3, 3) Wilforlide A, 4) Cangononine, 5) Wilforine, 6) Regeol C, 7) Celastrol | Flavonoid, terpenoids and alkaloid | [51] |
| 15 16 17 18 19 20 21 | <i>Tripterygium wilfordii</i> Hook. f. (Celastraceae) | Rhizomes | Percolation and ultrasonication – EtOH 95% | Miscellaneous (Hsp 90) | 1) Demecolcine, 2) Wilforine, 3) Triptotriterpenic acid, 4) Celastrol, 5) R1-Barrigenol, 6) 24R,25-dihydroxycholecalciferol, 7) Ursodeoxycholic acid, 8) 8,14-Seco-3,19-epoxyandrostane-8,14-dione, 17-acetoxy-3 β -methoxy-4,4-dimethyl, 9) Spirost-8-en-11-one, 10) Propanoic acid, 11) Demecolcine, 12) Bufa-20,22-dienolide, 13) Ergosta-5,22-dien-3-ol, 14) β -Sitosterol and 15) Betulinaldehyde | Alkaloids, terpenoids and steroids | [46] |
| 22 23 24 | <i>Vaccinium corymbosum</i> L. (Ericaceae) | Leaves | Reflux – Petroleum ether-EtOH 70% | Miscellaneous (HepG2 cells) | 1) Chlorogenic acid, 2) Rutin, 3) Isoquercitrin, 4) Cyanidin-3-O-glucose, 5) Iridin and 6) Quercetin | Phenolic acid and flavonoids | [192] |
| 25 26 27 28 | <i>Xanthium strumarium</i> L. (Asteraceae) | Fruits | Reflux – CH ₂ Cl ₂ -MeOH | Ultrafiltration (Tyrosinase) | 1) Protocatechuic acid, 2) 3,5- Di-O-caffeoylquinic acid, 3) 1,5-Di-O-caffeoylquinic acid and 4) Chlorogenic acid | Phenolic acids | [92] |
| 29 30 31 32 | <i>Zingiber officinale</i> Roscoe. (Zingiberaceae) | Rhizomes | Vortexing - EtOH | Magnetic particle (α V β 3 integrin) | 1) [10]-gingerol | Phenolic compound | [148] |

Table S1 – Natural products screened through ligand fishing approaches sorted by species in alphabetical order. Complementary information regarding the family, the parts used, the type of extraction, the extraction solvent, the ligand fishing technique with the respective cancer-related target, the ligands fished with their chemical classification as well as the bibliographical reference is also provided.

| Ligand Fishing Technique | Ligand Fishing Medium | Target | Type of Cancer | Ligands Fished | Ref |
|--------------------------|------------------------|---------------------------------------|--|---|-------|
| Dialysis | Ultrapure water | DNA (G-quadruplex) | Various | 1) Berberine | [185] |
| Hollow Fiber | Phosphate buffer | HCT116 cells | Colorectal cancer | 1) Aloe-emodin, 2) Emodin, 3) Physcion, 4) Rhein and 5) Chrysophanol | [194] |
| | CMC-Na solution 5% | HepG-2 cells | Hepatoma | 1) Xanthotoxol, 2) Xanthotoxin, 3) Psoralen, 4) Bergapten, 5) Oxypeucedanin, 6) Imperatorin, 7) Isoimperatorin. | [43] |
| | MeOH 5% | HepG-2, SKOV-3, and ACHN cells | Hepatoma, human ovarian cancer, and human renal carcinoma | 1) Bisdemethoxycurcumin, 2) Demethoxycurcumin and 3) Curcumin | [195] |
| | Double-distilled water | MCF-7, ACHN, SKOV-3, and HepG-2 cells | Breast cancer, human renal carcinoma, human ovarian cancer, and hepatoma | 1) Oxymatrine, 2) Matrine, 3) Sophoridine and 4) Oxysophocarpine | [191] |
| | Phosphate buffer | SGC7901, RAW264.7, SHSY5Y cells | Gastric cancer, neuroblastoma and leukemia | 1) Psoralen, 2) Bergapten, 3) Imperatorin 4) Osthole 5) Isopsoralen 6) Schisandrol A, 7) Schizandrin A, 8) Schisantherin A and 9) Schizandrin C | [41] |
| | Phosphate buffer | Tyrosinase | Melanoma | 1) Daidzein, 2) 3'-Methoxy puerarin, 3) 3'-Hydroxy puerarin 4) Puerarin, 5) Puerarin 6''-O-xyloside 6) Puerarin-4'-O-glucoside, 7) Puerarin aposide | [42] |
| | Phosphate buffer | HepG-2 and ACHN cells | Hepatoma and human renal carcinoma | 1) Chlorogenic acid, 2) p-Hydroxyacetophenone 3) Geniposide, 4) Crocin 5) Rhein | [196] |
| | Double-distilled water | MCF7, MADB106, and SGC7901 cells | Breast cancer and gastric cancer | 1) Jatrorrhizine, 2) Epiberine, 3) Coptisine, 4) Palmatine, 5) Berberine | [40] |

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|---------------------------|------------------|---------------------------------|--------------------------------|---|-------|
| Magnetic particles | Phosphate buffer | EGFR/A549 cells | Lung carcinoma | 1) 10,11-dimethoxystrychnine and 2) Loganetin | [35] |
| | Phosphate buffer | Cyclooxygenase-2 (COX2) | Various | 1) (-)-gallocatechin, 2) (-)-epigallocatechin, 3) (-)-epicatechin, 4) (-)-epigallocatechin-3-gallate, 5) (-)-gallocatechin-3-gallate, 6) (-)-epigallocatechin-3-(3''-O-methyl)-gallate, 7) (-)-epicatechin-3-gallate, 8) (-)-epicatechin-3-(3''-O-methyl)-gallate | [106] |
| | Tris-HCl buffer | Cyclooxygenase-2 (COX2) | Various | 1) Skimmin, 2) Scopolin, 3) Umbelliferone, 4) Scopoletin | [109] |
| | Tris-HCl buffer | Cyclooxygenase-2 (COX2) | Various | 1) 3-Galloylquinic acid, 2) 5-Galloylquinic acid, 3) 3-p-Coumaroylquinic acid, 4) 5-p-Coumaroylquinic acid, 5) 5-caffeoylquinic acid, 6) (-)-Epicatechin, 7) (-)-Epicatechin 3-O-gallate, 8) Quercetin 3-O-rhamnosylgalactoside, 9) Quercetin 3-O-glucoside, 10) Kaempferol 3-O-glucosylrutinoside, 11) Kaempferol-3-O-rutinoside, 12) Kaempferol 3-O-glucoside | [107] |
| | Phosphate buffer | DNA (ds) | Various | 1) Columbamine, 2) Epiberberine, 3) Jatrorrhizine, 4) Palmatine and 5) Berberine | [186] |
| | Distilled water | EGFR/HEK293 cells | Glioma | 1) Luteolin and 2) Caffeic acid | [197] |
| | Phosphate buffer | Glutathione S-transferase (GST) | Various (multidrug resistance) | 1) Caffeic acid, 2) Rosmarinic acid, 3) Gallic acid, 4) (-)-Epigallocatechin-3-O-gallate, 5) (-)-Epicatechin-3-gallate and 6) Quercetin-3-O-glucoside | [28] |
| | Acetate buffer | Lactate dehydrogenase (LDH) | Various | 1) (-)-Epicatechin-3-O-gallate, 2) Trihydroxystilbene-3-O-(6''-galloyl)-glucoside, 3) Emodin-1-O-glucoside, 4) Emodin-8-O-(6-malonyl)-glucoside, 5) Torachryson sulfate, 6) Emodin, 7) ω-Hydroxyemodin, 8) Lunatin, 9) Chrysophanic acid, 10) Rhein-O-(acetyl)-glucoside, 11) Aloe emodin, 12) Rhein | [119] |

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| 6 | Tris-HCl buffer | Phospholipase A2 (PLA2) | Various | 1) Cyclo-(4-hydroxyl-Pro-Leu), 2) Cyclo-(Pro-Val), 3) Fusaristatin C, 4) Ergosterol peroxide, 5) Ergosterol and 6) Cerevisterol | [115] |
| 7 | Ammonium acetate buffer | Sirtuin 6 (SIRT6) | Various | 1) Orientin | [39] |
| 8 | | | | | |
| 9 | Phosphate buffer | Sirtuin 6 (SIRT6) | Various | 1) Quercetin and 2) Vitexin | [38] |
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| 14 | Tris-HCl buffer | Tropomyosin receptor kinase B (TrkB) | Various | 1) Dicafeoylquinic acid, 2) O-cafeoylquinic acid, 3) Madecassic acid, 4) Castillicetin, 5) Quercetin, 6) Stigmasterol, 7) Pomolic acid and 8) Naringin | [32] |
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| 20 | Phosphate buffer | Tyrosinase | Melanoma | 1) Liquiritin apioside, 2) Neolicuroside, 3) Liquiritigenin, 4) Licorice saponin, 5) Chrysoeriol, 6) Dihydrodiadzein, 7) Formononetin, 8) Glycyrrhisoflavanone, 9) Glycyrrhizic acid, 10) Glycycomarin and 11) Pratensein | [18] |
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| 28 | | | | | |
| 29 | Phosphate buffer | Tyrosinase | Melanoma | 1) Gallic acid, 2) Albiflorin, 3) Paeoniflorin, 4) Liquiritin apioside, 5) Liquiritin, 6) Galloylpaeoniflorin, 7) Ononin, 8) Isoliquiritigenin, 9) Glycyrrhizic acid, 10) Oxypaeoniflora, 11) Benzoylpaeoniflorin, 12) Benzoyloxypaeoniflorin, 13) Mudanpioside C, 14) Paeonolide and 15) Apiopaeonoside | [95] |
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| 36 | | | | | |
| 37 | Phosphate buffer | α V β 3 integrin | Various | 1) [10]-gingerol | [148] |
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|----|------------------------|--|-----------------------------|---------------------------|--|-------|--|
| 1 | | Acetate buffer | Lactate dehydrogenase (LDH) | Various | 1) Amentoflavone and 2) Robustaflavone | [25] | |
| 2 | | Tris-HCl buffer | Tyrosinase | Melanoma | 1) Baicalin, 2) Baicalein, 3) Wogonin, 4) Oroxylin A, 5) Kurarinone, 6) Sophoraflavanone G | [26] | |
| 3 | | | | | | | |
| 4 | | | | | | | |
| 5 | | | | | | | |
| 6 | Ultrafiltration | Phosphate buffer | Cyclooxygenase-2 (COX2) | Various | 1) Neomangiferin, 2) Phellodendrine, 3) Mangiferin, 4) Magnoflorine, 5) 3-O-feruloylquinnic acid, 6) (-)-Oblongine, 7) Menisperine, 8) Demethyleneberberine, 9) Oxoberberine, 10) Berberrubine, 11) Jatrorrhizine, 12) Timosaponin B, 13) Timosaponin B-II, 14) Timosaponin A-I, 15) Palmatine, 16) Berberine, 17) Sanleng acid, 18) Timosaponin A-III, 19) Anemarrhenasaponin I, 20) cis-Hinokiresinol | [105] | |
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| 11 | | | | | | | |
| 12 | | Distilled water, MeOH or MeOH-distilled water | DNA | Various | 1) Berberine, 2) Coptisine, 3) Palmitine 4) Daurisoline, 5) Dauricine, 6) Fangchinoline, 7) Tetrandrine, 8) Actinomycin D, 9) Actinomycin V and 10) Actinomycin X0β | [12] | |
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| 19 | | | | | | | |
| 20 | | Buffer (50 mM KCl, 10 mM KH ₂ PO ₄ , and 1 mM K ₂ EDTA, pH 7.4) | DNA (G-quadruplex) | Various | 1) Chelerythrine, 2) Sanguinarine, 3) Protopine and 4) Allocryptopine | [188] | |
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| 24 | | | | | | | |
| 25 | | Tris-HCl buffer | Quinone reductase-2 (QR-2) | Various (chemoprevention) | 1) Tetrangulol methyl ether, 2) Xanthohumol and 3) Xanthohumol D | [82] | |
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| 5 | Potassium phosphate | Tyrosinase | Melanoma | <p>1) 12-15 Heneicosadienoic acid, Octadecanoic acid, 2- hydroxylpropyl esters, 2) Unknown, 3) 3-(4-tert-butylphenyl)-2-methylpropanal, 4) 4,4'-Dihydroxybenzylsulfid, 5) Octadecanoic acid, 2-hydroxy-1-(hydroxymethyl)ethyl ester 6) 9-Octadecenamide, (trans-), 7) Unknown, 8) Unknown, 9) 9-Octadecenamide, (cis-), 10) 4,4'-Dihydroxydiphenyl ether, 11) 4-(1-(4-hydroxyphenyl)phenyl)phenol, 12) Octadecanoic acid, 2,3-dihydroxypropyl ester, 13) 4,4'-Dimethoxydiphenylmethane, 14) 4,4'-Methylenediphenol, 15) 2,4-Bis(4-hydroxybenzyl) phenol, 16) 4-Hydroxybenzyl methyl ether</p> | [90] |
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| 11 | | | | | |
| 12 | | | | | |
| 13 | Potassium phosphate | Tyrosinase | Melanoma | <p>1) Gallic acid, 2) Iriflophenone 3-C-glucoside, 3) Mangiferin, 4) Protocatechuic acid, 5) Iriflophenone 3-C-(2'-O-galloyl)-glucoside, 6) 6'-O-Galloyl-mangiferin, 7) Maclurin 3-C-(2'-O-p-hydroxybenzoyl)-glucoside, 8) Iriflophenone 3-C-(2',6'-di-O-galloyl)-glucoside, 9) Hyperoside, 10) Isoquercitrin, 11) Ethyl gallate, 12) Iriflophenone 3-C-(2'-O-p-hydroxybenzoyl)-glucoside, 13) Quercetin-3-O-xyloside, 14) 3-O-Galloyl shikimic acid, 15) 3-O-Galloyl quinic acid, 16) Maclurin 3-C-(2'-O-galloyl)-glucoside, 17) 3,5-Di-O-galloyl quinic acid, 18) 5-O-Digalloyl quinic acid, 19) Digallic acid, 20) Isomangiferin, 21) 1,4,6-Tri-O-galloyl glucoside, 22) Maclurin 3-C-(2',3'-di-O-galloyl)-glucoside, 23) 1,3-Digalleoyl acetone, 24) 1,3,4,6-Tetra-O-galloyl glucoside, 25) 6'-O-(p-Hydroxybenzoyl) mangiferin, 26) Epicatechin gallate, 27) 1,2,3,4,6-Penta-O-galloyl glucoside, 28) Iriflophenone 3-C-(2',3',6'-tri-O-galloyl)-glucoside, 29) Luteolin-7-O-glucoside, 30) Quercetin-3-O-arapyanoside, 31) Quercetin-3-O-arafuranoside, 32) Kaempferol-3-O-glucoside, 33) Quercetin-3-O-rhamnoside, 34) Ethyl 2,4-dihydroxy-3-galloyl oxybenzoate, 35) Ethyl digallate, 36) 7-O-Methyl quercetin-3-O-rhamnoside</p> | [19] |
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| 21 | | | | | |
| 22 | Potassium phosphate | Tyrosinase | Melanoma | | |
| 23 | buffer + Ethyl acetate | | | | |
| 24 | (fraction) | | | | |
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| 36 | Phosphate buffer | Tyrosinase | Melanoma | <p>1) Daidzin, 2) Isoorientin, 3) Isovitexin, 4) Daidzein, 5) Genistein, 6) 3-(5-hydroxybenzofuran-6-yl) propanoic acid</p> | [17] |
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|----|----------------------|---|--|--|---|-------|
| 1 | | MeOH + Potassium phosphate buffer | Tyrosinase | Melanoma | 1) Protocatechuic acid, 2) 3,5- Di-O-caffeoylquinic acid, 3) 1,5-Di-O-caffeoylquinic acid and 4) Chlorogenic acid | [92] |
| 2 | | | | | | |
| 3 | | | | | | |
| 4 | | Phosphate buffer | Tyrosinase | Melanoma | 1) 3'-Hydroxypuerarin, 2) Puerarin, 3) Puerarin-6''-O-xyloside, 4) Daidzin, 5) Genistin, 6) 6''-O-acetyldaidzin and 7) Daidzein | [93] |
| 5 | | | | | | |
| 6 | | | | | | |
| 7 | | Phosphate buffer | Tyrosinase | Melanoma | 1) Oroxin B, 2) Kaempferid-3-O-beta-D-gentiobioside, 3) Chrysin-7-O-beta-D-gentiobioside, 4) Oroxin A, 5) Baicalein and 6) Chrysin | [94] |
| 8 | | | | | | |
| 9 | | | | | | |
| 10 | | Phosphate buffer | Tyrosinase | Melanoma | 1) Puerarin, 2) Mirificin, 3) Daidzin and 4) Genistin | [91] |
| 11 | | | | | | |
| 12 | | Phosphate buffer | Urokinase-type plasminogen activator | Prognostic marker (breast, prostate, and bladder cancer) | 1) Gallic acid, 2) Methyl gallate, 3) Pentagalloylglucose, 4) Ellagic acid, 5) Ziyu-glycoside I, 6) 28-O- β -D-glucopyranosyl pomolic acid, and 7) 1 β -Hydroxyeuscapic acid | [165] |
| 13 | | | | | | |
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| 17 | | | | | | |
| 18 | Miscellaneous | Gradient EtOH-Deionized water | Cyclooxygenase-2 (COX2) | Various | 1) Sylvestroside I | [108] |
| 19 | | | | | | |
| 20 | | Gradient MeOH-Deionized water | Cyclooxygenase-2 (COX2) | Various | 1) Swertisin, 2) Scopoletin | [110] |
| 21 | | | | | | |
| 22 | | Isopropyl alcohol-Ammonium acetate buffer | Fibroblast growth factor receptor 4 (FGFR4) | Various | 1) Napellonine, 2) 12-Epinapelline, 3) Benzoylaconitine and 4) 3-Deoxyaconitine | [158] |
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| 28 | | | | | | |
| 29 | | DMSO-Tris-HCl buffer | DNA - G-quadruplex (HT24 and HT22); Duplex sequence (DNA26); DNA (Calf thymus DNA) | Various | 1) Sanguinarine, 2) Chelerythrine, 3) Protopine and 4) Allocryptopine | [187] |
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|----|--|-----------------------|----------------------------|--|-------|
| 1 | Buffer (50 mM KCl, 10 mM KH ₂ PO ₄ , and 1 mM K ₂ EDTA, pH 7.4) | DNA (G-quadruplex) | Various | 1) Chelerythrine, 2) Sanguinarine, 3) Protopine and 4) Allocryptopine | [188] |
| 2 | | | | | |
| 3 | | | | | |
| 4 | | | | | |
| 5 | Buffer II (20 mM NH ₄ OAC, 5 mM Mg(AC) ₂ , pH 5.5) | DNA (triplex) | Various | 1) Fangchinoline and 2) Tetrandrine | [50] |
| 6 | | | | | |
| 7 | | | | | |
| 8 | | | | | |
| 9 | | | | | |
| 10 | Buffer II (20 mM NH ₄ OAC, 5 mM MgCl ₂ , pH 5.5) | DNA (triplex) | Various | 1) Berberine and 2) Palmatine | [47] |
| 11 | | | | | |
| 12 | | | | | |
| 13 | | | | | |
| 14 | Buffer II (20 mM NH ₄ OAC, 5 mM Mg(AC) ₂ , pH 5.5) | DNA (triplex) | Various | 1) Epiberberine, 2) Coptisine, 3) Jatrorrhizine, 4) Berberrubine and 5) Columbamine | [49] |
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| 18 | | | | | |
| 19 | | | | | |
| 20 | Serum-free DMEM | HepG2 and MCF-7 cells | Hepatoma and breast cancer | 1) Serotonin, 2) N-methyl serotonin, 3) Bufotenidine, 4) Suberoyl arginine, 5) Arenobufagin, 6) Telocinobufatoxin, 7) Bufotalin, 8) 3-(N-pimeloyl argininy) bufalin, 9) Cinobufotalin, 10) Bufalitoxin, 11) 3-(N-pimeloyl argininy) resibufogenin, 12) Bufalin, 13) Resibufotoxin, 14) Cinobufotoxin, 15) 3-(N-azelayl argininy) resibufogenin, 16) Cinobufagin and 17) Resibufogenin | [11] |
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| 29 | Phosphate buffer | HepG2 cells | Hepatoma | 1) Chlorogenic acid, 2) Rutin, 3) Isoquercitrin, 4) Cyanidin-3-O-glucose, 5) Iridin and 6) Quercetin | [192] |
| 30 | | | | | |
| 31 | | | | | |
| 32 | Undefined | Hsp 90 | Various | 1) Demecolcine, 2) Wilforine, 3) Triptotriterpenic acid, 4) Celastrol, 5) R1-Barrigenol, 6) 24R,25-dihydroxycholecalciferol, 7) Ursodeoxycholic acid, 8) 8,14-Seco-3,19-epoxyandrostane-8,14-dione, 17-acetoxy-3β-methoxy-4,4-dimethyl, 9) Spirost-8-en-11-one, 10) Propanoic acid, 11) Demecolcine, 12) Bufa-20,22-dienolide, 13) Ergosta-5,22-dien-3-ol, 14) β-Sitosterol and 15) Betulinaldehyde | [46] |
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|---|--|------------------------------------|---|---|-------|---|---|---|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|
| 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 | 16 | 17 | 18 | 19 | 20 | 21 | 22 | 23 | 24 | 25 | 26 | 27 | 28 | 29 | 30 | 31 | 32 | 33 | 34 | 35 | 36 | 37 | 38 | 39 | 40 | 41 | 42 | 43 | 44 | 45 | 46 |
| | Undefined | Hsp 90 α | Various | 1) Alisol F, 2) 16-oxo-11-deoxy-Alisol A, 3) Alisol F 24-acetate, 4) Alisol L 23-acetate, 5) 16-oxo-11-anhydro-Alisol A, 6) Alisol H, 7) Alisol B-23-acetate, 8) Alisol C, 9) 3-Methyl-5,6- dimethoxy-2,3-dihydro-1H-indene-1-one, 10) (1S,3 α ,7 β)-1- Acetyloctahydro- 4 α -hydroxy-7 β - isopropyl 4-methyl-1H-indene, 11) Corymbolone, 12) (-)-Spathulenol, 13) n-Hexadecanoic acid, 14) Hexadecanoic acid ethyl ester, 15) Linoleic acid ethyl ester, 16) 13,17-Seco-5 α -pregn-13(18)-en-20-one, 17) Lupeol, 18) Lupeol acetate, 19) Taraxasterol and 20) Betulinaldehyde | [45] | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| | Undefined | Hsp 90 α | Various | 1) Zingiberenol, 2) Ar-turmerone, 3) Curlone, 4) (6R,7R)-bisabolone, 5) (E)-Atlantone, 6) 6-(1-Hydroxymethylvinyl), 7) 4,8a-dimethyl-3,5,6,7,8,8a-hexahydro-1H-naphthalen-2-one | [44] | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| | EtOH 50% | MCF-7 cells | Breast cancer | 1) Genistein, 2) Tripteryl E-3, 3) Wilforlide A, 4) Cangononine, 5) Wilforine, 6) Regeol C, 7) Celastrol | [51] | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| | DMSO-RPMI-1640 medium | PC3-cells | Human androgen-refractory prostate cancer | 1) Curcumin, 2) Demethoxycurcumin and 3) Bisdemethoxycurcumin | [20] | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| | Phosphate buffer | Matrix Metalloproteinase-2 (MMP-2) | Various | 1) Oreganol A, 2) Corilagin, 3) Ellagic acid, 4) Zebirioside O, 5) Nigaichigoside F1, 6) Peduncloside, 7) Rosamultin, 8) Kajiichigoside F1, 9) Potentillanoside A, 10) Quadranoside VIII, 11) Valerenic acid and 12) Medicagenic acid | [171] | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| | Tris-HCl buffer | Monoacylglycerol lipase (MAGL) | Various | 1) Tetrahydropalmatin, 2) Tetrahydrocoptisine, 3) Coptisine, 4) Fumaricine, 5) Palmatine, 6) Berberine and 7) Dehydrocorydaline | [178] | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| | Serum-free Dulbecco's modified eagle medium (DMEM) | HepG2 cells | Hepatoma | 1) Aurantio-obtusin, 2) 1-desmethyaurantio-obtusin, 3) Chryso-obtusin, 4) Obtusin, and 5) Questin | [193] | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| | Phosphate buffer | A549 and H1819 cells | Lung carcinoma | 1) Salvianolic acid B, 2) Deoxyshikonin and 3) Shikonin | [48] | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |

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|--|--|-------------|---------------|---|------|
| | Serum-free Dulbecco's modified eagle medium (DMEM) | MCF-7 cells | Breast cancer | 1) 5,7,8,30,40-Pentamethoxyflavanone, 2) 3,5,6,7,8,40-Hexamethoxyflavanone, 3) 5,7,8,30,40,50-Hexamethoxyflavone, 4) 7-Hydroxy-5,6,8,30,40-pentamethoxyflavone, 5) Isosinensetin, 6) 30-Hydroxy-5,6,7,8,40-pentamethoxyflavone or 40-Hydroxy-5,6,7,8,30-pentamethoxyflavone, 7) Sinensetin, 8) 5,6,7,8,40-Pentamethoxyflavanone, 9) 3-Hydroxy-5,6,7,8,40-pentamethoxyflavone, 10) Tetramethyl-O-isoscutellarein, 11) Tetramethyl-O-scutellarein, 12) Nobiletin, 13) 3,5,6,7,8,30,40-Heptamethoxyflavone, 14) 5,6,7,8,30,40-Hexamethoxyflavanone, 15) 5-Hydroxy-6,7,8,30,40-pentamethoxyflavone, 16) Tangeretin | [71] |
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Table S2 – Ligand fishing studies grouped by technique. Information concerning the ligand fishing medium, the cancer-related target with type of cancer involved, the ligands fished, and the respective bibliographical reference is also provided.