



HAL
open science

The challenge of assessing the effects of drifting fish aggregating devices on the behaviour and biology of tropical tuna

Amaël Dupaix, Frédéric Ménard, John D Filmalter, Yannick Baidai, Nathalie Bodin, Manuela Capello, Emmanuel Chassot, Hervé Demarcq, Jean-Louis Deneubourg, Alain Fonteneau, et al.

► **To cite this version:**

Amaël Dupaix, Frédéric Ménard, John D Filmalter, Yannick Baidai, Nathalie Bodin, et al.. The challenge of assessing the effects of drifting fish aggregating devices on the behaviour and biology of tropical tuna. 2023. hal-04047298v2

HAL Id: hal-04047298

<https://hal.science/hal-04047298v2>

Preprint submitted on 20 Jul 2023 (v2), last revised 20 Dec 2023 (v4)

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

1 **Title 1: The challenge of assessing the effects of drifting fish aggregating devices on**
2 **the behaviour and biology of tropical tuna**

3 **Title 2: Effects of drifting fish aggregating devices on biology and behaviour of**
4 **tropical tuna**

5

6 **Running title:** Effects of DFADs on tropical tuna (33 characters including space < 40)

7

8 **Authors:** Dupaix Amaël¹, Ménard Frédéric², Filmlalter John D³, Baidai Yannick⁴, Bodin
9 Nathalie^{5,6}, Capello Manuela¹, Chassot Emmanuel⁷, Demarcq Hervé¹, Deneubourg Jean-
10 Louis⁸, Fonteneau Alain⁹, Forget Fabien¹, Forrestal Francesca¹⁰, Gaertner Daniel¹, Hall
11 Martin¹¹, Holland Kim N.¹², Itano David¹³, Kaplan David Michael¹, Lopez Jon¹¹, Marsac
12 Francis¹, Maufroy Alexandra¹⁴, Moreno Gala¹⁵, Muir Jeff A.¹², Murua Hilario¹⁵, Roa-Pascuali
13 Liliana¹⁶, Pérez Géraldine¹⁷, Restrepo Victor¹⁵, Robert Marianne¹⁸, Schaefer Kurt M.¹⁰, Sempo
14 Grégory¹⁹, Soria Marc¹, Dagorn Laurent¹

15

16 **Corresponding Author:** Amaël Dupaix, amael.dupaix@ird.fr

17

18 **Affiliations:**

19¹: MARBEC, Univ. Montpellier, CNRS, Ifremer, IRD, INRAE, Sète, France

20²: Aix Marseille Univ., Université de Toulon, CNRS, IRD, Mediterranean Institute of
21 Oceanography, Marseille, France

22³: South African Institute for Aquatic Biodiversity, Grahamstown, South Africa

23⁴: African Marine Expertise, Abidjan, Ivory Coast

24⁵: Seychelles Fishing Authority (SFA), Fishing Port, Victoria P.O. Box 449, Mahé, Seychelles

25⁶: Sustainable Ocean Seychelles, BeauBelle, Mahé, Seychelles

26⁷: Indian Ocean Tuna Commission (IOTC), C/O IOTC Secretariat, Blend Seychelles, Victoria,
27 Mahé, Seychelles

28⁸: CENOLI, Université Libre de Bruxelles (ULB), Bruxelles, Belgium

29⁹: Consultant, Saint-Malo, France

30¹⁰: Southeast Fisheries Science Center, National Marine Fisheries Service, National Oceanic
31and Atmospheric Administration (NOAA)

32¹¹: Inter-American Tropical Tuna Commission, La Jolla, CA 92037, USA

33¹²: Hawaii Institute of Marine Biology, University of Hawaii at Manoa, Kaneohe, HI, United
34States of America

35¹³: Opah Consulting, Honolulu, HI, United States of America

36¹⁴: Organisation des Producteurs de thon congelé et surgelé (ORTHONGEL), Concarneau,
37France

38¹⁵: International Seafood Sustainability Foundation, Washington, DC 20005, US

39¹⁶: Collecte Localisation Satellites, Ramonville Saint-Agne, France

40¹⁷: Institut océanographique Paul Ricard, Île des Embiez - 83140 Six-Fours-les-Plages,
41France

42¹⁸: UMR DECOD (Ecosystem Dynamics and Sustainability), IFREMER, INRAE, Institut Agro -
43Agrocampus Ouest, 8 rue François Toullec, 56100 Lorient, France

44¹⁹: Faculté de Médecine, Université libre de Bruxelles (ULB), Bruxelles, Belgium

45**Abstract (242 words < 250)**

46Though fisheries have intensively used drifting fish aggregating devices (DFADs) over the
47last three decades to facilitate their catch of tropical tunas, assessing the consequences of
48the presence of DFADs at sea on tuna behaviour and biology is a challenge. The use of
49DFADs has resulted in a major increase in the number of floating objects, which are spatially
50heterogeneous at sea. To date, no **converging scientific results exist** regarding the **indirect**
51effects of DFADs **(unrelated with fishing mortality)** on the large-scale movements and
52behaviour of tuna, mainly due to the difficulty of disentangling the respective roles of DFADs
53and environmental factors. Some biological indices show that tuna condition is lower when
54associated to a floating object than in a free-swimming school. It is not possible, however, to
55elucidate whether this is the cause or result of the association, or if it affects the fitness of
56individuals in the long term. Further scientific progress would require (i) the collection of time
57series of indicators to monitor habitat change, individual behaviour, individual fitness and
58population dynamics, and (ii) experimental studies to identify the underlying behavioural and
59biological processes involved **in** associative behaviour. The extent of the modification of the
60surface habitat by the massive deployment of DFADs and the current uncertainty of the
61possible long-term consequences on the individual fitness and dynamics of tuna populations
62argue for the need for increased awareness of this issue by Regional Fisheries Management
63Organisations regulating tuna fishing.

64

65**Keywords** (6 max): DFAD, ecological effects, ecological trap, fish behaviour, fisheries
66management, tuna

67

68

69 **Table of contents**

70

71 **Introduction**

72 **How much do DFADs change the habitat of tropical tunas?**

73 **Do DFADs modify the migration and the schooling behaviour of tropical tunas?**

75 *Effects on individual large-scale movements*

76 *Effects on individual fine-scale movements*

77 *Effects on schooling behaviour*

78 **Do DFADs modify the life-history parameters of tropical tunas?**

79 *Effects on feeding*

80 *Effects on body condition*

81 *Effects on reproduction and growth*

82 **What are the scientific challenges to fill the knowledge gaps?**

83 **Conclusion**

84 **Acknowledgements**

85 **Data availability statement**

86 **References**

87 **Tables**

88 **Figures**

89

90Introduction

91Many fish species are known to associate with floating objects (Castro et al., 2002; Fréon &
92Dagorn, 2000), with the first known descriptions of fishers exploiting these associations
93dating from 200 AD in the Mediterranean Sea by the Roman author Oppian (cited in Taquet,
942013). In particular, the use of floating objects to facilitate the capture of tropical tunas
95(skipjack SKJ – *Katsuwonus pelamis*; yellowfin YFT – *Thunnus albacares*; and bigeye BET –
96*T. obesus*), has undergone rapid expansion in recent decades, as a result of the growing
97importance of these floating structures to the strategy and efficiency of tropical tuna purse
98seine fleets (Dagorn et al., 2012; Fonteneau et al., 2000, 2013; Leroy et al., 2013; Miyake et
99al., 2010). Since the onset of the tropical tuna purse seine fishery, fishers took advantage of
100the associative behaviour of tunas with floating objects and actively searched for natural
101floating objects to improve their catches (Greenblatt, 1979 ; Hallier and Parajua, 1999 ; Scott
102et al. 1999). Towards the end of the 1980s, fishers began to build and deploy man-made
103drifting fish aggregating devices (DFADs), and to attach radio buoys to locate them (Ariz et
104al., 1999 ; Hallier and Parajua, 1999 ; Hall et al. 1992 ; Scott et al. 1999 ; Lopez et al., 2014;
105Marsac et al., 2014; Moreno et al., 2007; Morón, 2001; Stéquert & Marsac, 1986). DFADs
106are commonly composed of a floating structure (such as a bamboo or metal raft with
107buoyancy provided by corks, etc.) and a submerged structure (made of ropes, old netting,
108canvas, weights, etc.). During the last two decades, radio buoys have been replaced by GPS
109buoys communicating via satellite directly with fishing vessels. In the last decade (2010-
1102020), most DFADs have been equipped with echo-sounder buoys, providing estimates of
111aggregated biomass (Lopez et al., 2014). Some fleets also use supply vessels to maintain
112their DFAD array and to inform the fishing vessels of tuna aggregations, allowing these fleets
113to manage more efficiently their DFAD stock (Arrizabalaga et al., 2001; Ramos et al., 2013).
114DFADs represent very efficient fishing tools that increased the catchability of tunas, leading
115purse-seine fleets to target preferentially associated schools and expanding their fishing
116grounds (Lopez et al. 2014, Fonteneau et al. 2015, Lennert-Cody et al. 2019).

117Over time, given the growing contribution of purse seine fleets to world tuna catches and the
118increasing importance of DFAD fishing in the strategy of purse seine fleets, managing
119DFADs has become a priority for all tuna Regional Fishery Management Organisations
120(tRFMOs). In this paper we will use “operational” or “active” buoys to designate buoys
121attached to a **floating object (FOB)** that are tracked by one or several purse seine fishing
122vessel(s). Tuna RFMOs set limits of the number of operational buoys (with the very first limit
123by the IOTC, Indian Ocean Tuna Commission, in 2015) to mitigate the different risks induced
124by the deployment and use of DFADs (most recent resolutions: IOTC Res 23/02, ICCAT Rec
12522-01, IATTC Res C-21-04, WCPFC CMM 2021-01). **Fishing at FOBs was demonstrated to**
126**increase by-catch rates, compared to fishing on free-swimming schools (Amandé et al. 2012,**
127**Escalle et al. 2018), and to increase the proportion of small BET and YFT (IOTC 2022).**
128Other major DFAD-related measures concerned the design of these objects, following the
129discovery of sharks getting entangled in the netting composing the structure of DFADs
130(Filmalter et al. 2013). Limiting the pollution induced by DFADs in the ocean is also in front of
131the agendas of tRFMOs, after realizing the large quantity of plastic used in DFADs and the
132large numbers of DFAD beaching events in sensitive coastal ecosystems (**Escalle et al.**
133**2019,** Imzilen et al. 2021, 2022). However, other impacts on tuna populations (unrelated to
134fishery vulnerability) and ecosystems may be induced by the increased presence of DFADs
135in their habitat (**Marsac et al. 2000, Bromhead et al. 2003, Hallier & Gaertner 2008**). Despite
136the limits on operational buoys, DFADs number in the water has increased (Dagorn et al.
1372013a, Maufroy et al. 2017, Imzilen et al. 2021, Dupaix et al. 2021). As such, while logs and
138branches have always been components of the habitat of tropical tunas (originating from
139rivers, mangroves or shorelines), the massive use of man-made DFADs has changed their
140habitat.

141Changing a habitat can positively or negatively impact the ecology of wild animals inhabiting
142it. For example, artificial habitats could benefit some reef species (Lee et al. 2018).
143Contrarily, alterations could also reduce habitat quality, e.g. by reducing the number of
144shelters or nests for some species, or by decreasing their food resources (often through the
145alteration of the habitat of these resources themselves, Mullu 2016). In some cases, animals

146are misled by cues that were previously correlated with the habitat quality but no longer are,
147due to anthropogenic influences (Sherley et al. 2017, Swearer et al. 2021). Such impacts
148form the basis of the ecological trap theory and result in the preferential selection of low-
149quality habitats by animals, when better alternatives exist (Battin, 2004; Schlaepfer et al.,
1502002). It is worth noting that, depending on the definition, an ecological trap can occur
151without any anthropogenic influence (Robertson & Hutto, 2006; Swearer et al., 2021; Teske
152et al., 2021). In this paper, we will consider that ecological traps occur because of a sudden
153anthropogenic change in the environment, i.e. in the case of tropical tuna, the modification of
154their surface habitat by the increased deployment of DFADs (Gilroy & Sutherland, 2007;
155Hallier & Gaertner, 2008; Schlaepfer et al., 2002). While this theory has been proposed
156regularly in the face of anthropogenic environmental modifications and their impacts on
157various species, few studies have empirically demonstrated the existence of such traps
158(Battin, 2004; Swearer et al., 2021).

159Noting the increasing number of floating objects being deployed by fishers during the 1990s,
160some scientists hypothesized that the increase in the number of DFADs could lead to an
161ecological trap, altering the ecological value of floating objects for tropical tunas associated
162to DFADs (Marsac et al., 2000). It is hypothesized that large numbers of DFADs may alter
163certain biological characteristics of epipelagic populations associated with them: migration,
164schooling behaviour, growth, fish condition and bioenergetics, predation and natural mortality
165(Figures 1 & 2). TRFMOs primarily focus on developing management schemes to address
166the known effects of DFADs on catches (particularly of small YFT and BET, as well as
167sharks) or their stranding on coasts. However, there is also a need to assess whether
168DFADs, through their presence on the ocean, can alter the life history parameters and
169behaviour of tunas, so as to manage the number of DFADs deployed at sea if negative
170impacts are suspected or demonstrated.

171Almost 20 years ago, Dempster & Taquet (2004) made a systematic review of the published
172literature on FADs and concluded that further research should assess the use of DFADs by
173pelagic species, the mechanisms underlying their associative behaviour and the ecological
174consequences of the presence of DFADs at sea on pelagic fish stocks. Since then, several

175 papers reviewed existing evidence and/or proposed future research directions to address the
176 impacts of DFADs on tropical tuna (Dagorn et al. 2013b, Evans et al. 2015, Davies et al.
177 2014, Fonteneau et al. 2015, Leroy et al. 2013, Taquet 2013). Yet, most of these papers,
178 except Taquet (2013), addressed these impacts at a regional scale and all were mainly
179 focussing on the direct impacts of DFADs (induced by an increase of fishing mortality), even
180 though they mentioned potential indirect impacts (not related to fishing mortality increase).
181 The objective and originality of this paper is to review the current knowledge on the impacts
182 of DFADs on the ecology and life history traits of tropical tuna, focussing only on the indirect
183 effects (consequences of fishing on DFADs are not addressed here). The reason for this is
184 that this particular question has generated a global scientific debate for years, precluding
185 management bodies from having a complete and synthetic view of the current knowledge. An
186 in-depth literature review provides an overview of the state of the art in this area, identifies
187 knowledge gaps, and proposes future research priorities. This paper is structured around
188 four major questions:

189i) How much do DFADs change the habitat of tropical tunas?

190ii) Do DFADs modify the migration and the schooling behaviour of tropical tunas?

191iii) Do DFADs modify the life history parameters of tropical tunas?

192iv) What are the scientific challenges to fill the knowledge gaps?

193 **How much do DFADs change the habitat of tropical tunas?**

194 Natural floating objects, primarily tree trunks or branches carried by rivers, have always been
195 a component of tuna habitat. It should be noted that the current largest tuna purse seine
196 fishery in the world was first developed in the Western and Central Pacific Ocean (WCPO) by
197 Japanese exploratory fishing cruises that perfected methods for seining tuna schools found
198 in association with natural floating logs that later evolved into DFAD fisheries in all oceans
199 (Watanabe 1988). Human activities (logging, coastal development, shipping, etc.) modified
200 the number of floating objects at sea, in some cases even before modern purse-seine tuna
201 fishing began (Caddy & Majkowski, 1996; Thiel & Gutow, 2005). Some of these activities

202 may have consistently increased (coastal development, shipping), whereas others, such as
203 logging, may have varied due to increased global trade and subsequent deforestation of
204 some areas (Caddy & Majkowski, 1996). In addition, environmental changes also affect the
205 production and movement of floating objects (e.g. floods, El Niño events, tsunamis), with
206 global warming supposed to increase the frequency of extreme events. In recent years, the
207 increase in the number of DFADs deployed by fishers raised the question of the impacts of
208 this practice on tropical tuna habitat. It is therefore essential to assess the extent to which
209 DFADs have changed the habitat of tunas, in comparison to the historical pristine state when
210 only natural floating objects existed.

211 Two types of floating objects (referred to as FOBs) are commonly considered: (i) man-made
212 FADs (which can be drifting, DFADs, or anchored, AFADs) and (ii) natural objects (trees,
213 branches, etc., referred to as NLOGs) or artificial objects (wreckage, nets, washing
214 machines, etc., referred to as ALOGs) that are not deployed for the specific purpose of
215 fishing (collectively called LOGs) (Gaertner et al., 2018). Fishers fish on DFADs and LOGs
216 and can equip any of those objects with a satellite-tracking buoy, becoming therefore a
217 fishing tool monitored by a fishing vessel. For the particular question of habitat change
218 addressed in this study, only DFADs – which are the dominant type of man-made floating
219 objects used in the industrial purse seine fishery (Dagorn et al. 2013a, Maufroy et al. 2017,
220 Dupaix et al. 2021) – are considered and not AFADs.

221 Habitat changes due to DFADs can be assessed by estimating and comparing densities of
222 objects (with information on their nature: LOGs or DFADs) and distance between objects
223 (nearest neighbour), with both parameters being closely related. These parameters depend
224 upon the rates at which DFADs are added or removed from the ocean (by sinking, beaching
225 or retrieved by humans), as well as their drift. For every oceanic spatio-temporal unit (e.g.
226 region and season), comparing these parameters with those of natural floating objects and
227 for all types together (natural and artificial) is challenging, the primary concern being to
228 identify the origin of the floating object.

229The number of DFADs have regularly increased (Maufroy et al. 2017), but it is necessary to
230put this in perspective with respect to all floating objects. Using data from observers onboard
231tuna purse seine vessels in the Indian Ocean (IO), noting all FOBs encountered when the
232vessel cruises, Dupaix et al. (2021) highlighted a drastic increase in the total number of
233floating objects in the western IO, from 2006 to 2018, with multiplication factors greater than
2342 in every region and reaching as high as 60 in some areas (e.g. Somali area). The entire
235western IO is affected, with DFADs comprising over 85% of the total FOBs and DFADs
236contributing to reduce the distances between floating objects (mean distances between
237DFADs and between NLOGs of 37 km and 89 km, respectively, in 2014-2018). The impact of
238DFADs on tuna habitat reducing the distance between FOBs was observed in the study both
239when considering all DFADs or only randomly encountered DFADs (objects which do not
240belong to the vessel or its fishing company), to account for a potential sampling bias. Phillips
241et al. (2019a), using data from 2016 and 2017 and Lagrangian simulations in the Western
242and Central Pacific Ocean (WCPO), also showed an increase in FOB densities induced by
243DFAD deployments, and observed a shift of the area with the highest FOB densities, from
244the North-Eastern area of the Bismark Sea to the Tuvalu archipelago. Unfortunately, to our
245knowledge, no similar detailed study has been conducted in the other oceans, precluding
246from estimating the extent of the change of the habitat of tunas due to the addition of new
247floating objects globally.

248Most of the management effort by tRFMOs is focused on the monitoring and control of
249satellite-tracked buoys attached to floating objects (either to DFADs or to LOGs), emitting
250positions (and other variables) to vessels and qualified as operational buoys, as this variable
251is strongly related to fishing effort. This also explains why most scientific studies prioritized
252the estimate of operational buoys rather than the number of DFADs in the ocean (Table 1).
253Currently, all tRFMOs have implemented a limit on the instantaneous number of operational
254satellite buoys per vessel, and, except in the Atlantic Ocean (AO), limited the re-activation of
255buoys while at sea, but only the IOTC has limited the number of buoys purchased and in
256stock per year, per vessel (IOTC Res 23/02, ICCAT Rec 22-01, IATTC Res C-21-04,
257WCPFC CMM 2021-01). This clearly reflects a lack of concerted action worldwide to limit the

258number of new floating objects deployed in the oceans. Even under the limit of active DFADs
259at sea per vessel, the actual total numbers of DFADs in the ocean could have increased. So
260far, few studies have produced estimates of the total number of DFADs deployed annually,
261with estimations providing a range of 81,000 to 121,000 deployments worldwide, but these
262global estimates were made a decade ago (Baske et al., 2012; Gershman et al., 2015; Scott
263& Lopez, 2014). As a comparison, AFADs seem to be less numerous worldwide (13,000
264AFADs estimated in Scott & Lopez 2014), although there may be few areas with very high
265densities of AFADs, such as Indonesia (5,000-10,000, Proctor et al. 2019), the Philippines or
266Papua New Guinea.

267In practice, despite efforts by tRFMOs to require the submission of DFAD data, accurately
268determining a simple indicator such as the total number of DFADs that are drifting in the
269world's oceans is a major challenge. The easiest way would be to monitor the number of
270deployments through logbooks or onboard observers or set up a FAD register (see Res
27123/02 of the IOTC). The number of operational buoys does not correspond to the number of
272DFADs in the water (and/or deployed) as some buoys can be attached to LOGs, can be
273deactivated, and some DFADs may lack positional trackers, but it can be used as a proxy to
274illustrate the trend in numbers. Therefore, as the number of operational buoys does not
275effectively limit DFAD deployments, the number of DFADs in the water and/or deployments
276could be larger than the limits adopted by the tRFMOs. Under the assumption that the
277number of natural floating objects remains relatively constant, the increasing number of
278electronic buoys used reflects an increase of the number of FOBs. In recent years, DFAD
279deployments were stable in the WCPO (2011-2019; Escalle et al. 2020), decreased in the IO
280(2016-2021; IOTC, 2022), increased in the Eastern Pacific Ocean (EPO, 2015-2020; Lopez
281et al. 2021) and buoy deployments increased in the AO (2007-2013; Maufroy et al. 2017).
282Hence, a characterisation of DFAD deployment trends at the global scale is needed.
283However, the clear trend in the number of DFAD sets or DFAD catches (Floch et al., 2019;
284FIRMS Global Tuna Atlas cited in IOTC, 2021; Restrepo et al., 2017) suggests that DFAD
285deployment has also increased.

286 Do DFADs modify the migration and the schooling behaviour of tropical tunas?

287 DFADs may affect both the movements of tunas and their schooling behaviour. Large-scale
288 movements of tunas can be impacted in the following two ways: (i) DFADs could cause tunas
289 to relocate to new areas and (ii) they could increase residence times in some areas. Ideally,
290 the best approach for investigating such potential effects would be to compare the large-
291 scale movement patterns of tunas before and after the period in which the increase of DFAD
292 numbers occurred (i.e. before or after the 1990's). To our knowledge, historical data to
293 assess large-scale movement patterns before fishers started to massively deploy DFADs,
294 necessary for this type of analysis, exist only in the WCPO (Kim 2015) and in the EAO
295 (Cayré et al. 1986) and we do not know of any long-term study that compared movement
296 patterns before and after DFAD use increased.

297 *Effects on individual large-scale movements*

298 Wang et al. (2014) found that the spatial dynamics of free-swimming school sets in the
299 WCPO were influenced by the onset of El Niño Southern Oscillation (ENSO) events, while
300 these events had no effects on the location of floating-object-associated school sets. Catch
301 data, however, reflect the movements of the available catchable portion of the stocks and the
302 catchability of different set types (e.g. DFAD sets catching smaller individuals than the free-
303 school sets), and not the true movements of populations.

304 Hallier & Gaertner (2008) analysed conventional tagging data of SKJ and YFT in the Eastern
305 Atlantic Ocean (EAO). Different migratory directional patterns and displacement rates were
306 observed between fish recaptures associated with DFADs and those in free-swimming
307 schools. Displacement rates were significantly larger for both YFT and SKJ recaptured in
308 association with DFADs (13 and 15 nm/day, respectively) than those recaptured in free-
309 swimming schools (3 and 4.5 nm/day, respectively), which suggests that DFADs could
310 relocate tunas to new areas. In the IO, Stehfest and Dagorn (2010) found similar results for
311 SKJ, YFT and BET, but with lower displacement rates differences than in the AO. Hallier and
312 Gaertner (2008) interpreted these results as indicating significant modifications of migratory
313 patterns due to associations with DFADs, suggesting an influence of DFAD association

314 strong-enough to disturb tropical tuna migratory patterns. However, Stehfest and Dagorn
315 (2010) argue that it could only reflect an artefact of the non-uniform distribution of DFAD
316 fishing. Also, authors of both studies agree that school type at recapture might not be
317 representative of the associative history of individuals before their recapture. Using an
318 advection-diffusion model, Kim (2015) also showed that including a DFAD attraction
319 component to the model better fitted SKJ tagging data in the WCPO, suggesting an effect of
320 DFADs on SKJ migratory patterns. Comparing DFAD induced movements in the 2000s with
321 those in the 1990s, they showed that the rising DFAD density increased this modification of
322 migration patterns. These studies were the only ones to assess differences of movement
323 patterns induced by DFADs and, due to species and ocean differences, more studies would
324 be needed to interpret these results at a global scale.

325 DFADs potential to modify large-scale movements of tunas can be investigated through
326 archival tags by comparing tuna movements with the general drift patterns of DFADs. In the
327 equatorial EPO, evaluation of archival tag data sets from 96 BET (54-159 cm in length, 1-5.5
328 years of age) tagged between 2000 – 2005 (Schaefer et al., 2009; Schaefer & Fuller, 2010)
329 did not support the hypothesis that the most probable BET tracks were related to the general
330 drift patterns of DFADs. This suggests that the large-scale spatial dynamics of BET are not
331 strongly influenced by DFADs at the densities and conditions found in the EPO. However, in
332 the Central Pacific Ocean (CPO), a predominantly eastward extensive dispersion of BET
333 tagged with conventional tags and archival tags was observed (Schaefer et al., 2015). The
334 authors explain the strong regional fidelity of BET in the equatorial EPO by the high
335 concentration of food, leading to their residence and retention in that area. In the equatorial
336 CPO, the strong eastward-flowing North equatorial countercurrent and BET searching for
337 higher prey concentrations could explain the predominantly eastward dispersion of BET.

338 *Effects on individual fine-scale movements*

339 In addition, the possibility of DFADs influencing the large-scale movements of tunas could be
340 evaluated through the measure of the time tunas spend associated with DFADs and the time
341 they spend unassociated (or between two DFAD associations). It could be considered that

342the longer tunas remain associated with DFADs, the larger the influence DFADs could have
343on their large-scale movements. Acoustic tags and archival tags (only when a species
344exhibits a distinct vertical behaviour when associating with a floating object, as observed for
345BET or sometimes YFT) have been used by scientists to measure these parameters (Table
3462). Passive acoustic tagging studies on DFADs revealed that the majority of residence times
347of tunas (i.e. continuous periods of time spent associated with a given DFAD) were of a few
348days. Mean values ranged from 0.2 to 4.6 days for SKJ (Dagorn et al., 2007; Govinden et al.,
3492021; Matsumoto et al., 2014, 2016), from 1.0 to 6.6 days for YFT and 1.4 to 10 days for
350BET (Dagorn et al., 2007; Govinden et al., 2021; Matsumoto et al., 2016, Phillips et al.
3512019b). Long associations, however, have been observed on rare occasions – e.g. 27 days
352for YFT in the IO (Govinden et al., 2021) and up to 18, 50 and 30 days for SKJ, YFT and
353BET respectively, in the WCPO (Phillips et al. 2017, Phillips et al. 2019b). A recent study in
354the EAO (Tolotti et al., 2020) reported significantly larger mean residence times for the three
355tuna species, from 9 days (SKJ) to 19 days (YFT) and 25 days (BET), with record values of
35655 days and 600 km travelled associated to a DFAD for both BET and YFT.

357These studies suggest that residence times at a single DFAD vary between oceanic regions
358and species. Without more studies, it is difficult to assess whether the long DFAD
359associations observed are restricted to specific areas and time periods, or if they can often
360occur. In fact, even short DFAD residence times as those observed in the Indian and Pacific
361oceans do not prove that DFADs cannot influence large-scale movements. The short
362residence times suggest that a single DFAD does not significantly impact the behaviour of
363tunas for long enough to influence their large-scale movements. However, in an array of
364DFADs, a tuna can “switch” from one DFAD to a neighbouring one, which could retain it in
365the array. It is therefore important to also measure the time tunas spend between two
366associations (or unassociated), or in other words, the total percentage of time a tuna spends
367associated over long periods. This variable is likely to depend on the density of all floating
368objects in the area. So far, very few durations between two DFAD associations have been
369measured using acoustic tags because it is difficult to locate and exhaustively instrument
370with acoustic receivers all DFADs in an area.

371In the WCPO, 25 BET, 6 YFT and 2 SKJ displayed "homing" behaviour by returning to the
372same DFAD with absences greater than a day (Phillips et al. 2019b). Most of these absences
373were short for BET (median: 3.2 days) and longer for YFT (median: 10.5 days) but with a low
374sample size not allowing to be conclusive. In the other tropical oceans, even fewer tunas
375were observed performing such homing behaviour: one BET in the AO (out of 23 tagged fish,
376Tolotti et al., 2020), one YFT and two SKJ in the IO (out of 31 and 17 tagged fish
377respectively, Govinden et al., 2021), and these absences lasted less than two days.

378Because BET and sometimes YFT exhibit different vertical behaviour patterns when
379associated or non-associated with floating objects (Holland and Brill 1990, Abascal et al.
3802018), archival tags have been used to assess residence times at and between floating
381objects, and therefore percentage of days associated with floating objects, without the need
382to instrument all objects with acoustic receivers. Using satellite archival tagging data where
383individual BET tracks could be recorded over several months or even years, the percentage
384of time associated with floating objects was estimated to be between 4 % and 17 %
385depending on the size of the fish and the oceanic region (Fuller et al., 2015; Phillips et al.,
3862017; Schaefer & Fuller, 2010, Leroy et al. 2013, Schaefer & Fuller 2002). Associative and
387non-associative behaviour with floating objects have also been described with archival tags
388for YFT (Phillips et al., 2017; Schaefer et al., 2009; Schaefer & Fuller, 2013, Leroy et al.
3892013), with estimates of the percentage of time spent associated with floating objects
390between 10% and 23%. These percentages are much lower than those estimated from
391acoustic telemetry data (e.g. 75 % for small BET based on the measurements in Phillips, et
392al. 2019b).

393This inconsistency between studies using different tagging methods could result from the
394size of tagged individuals or the way the percentage of time spent by tuna associated and
395non-associated is calculated. Individuals monitored with archival tags were generally larger
396(fork length: 50-146 cm YFT and 46-102 cm BET in Phillips et al. 2017, 51-134 cm BET in
397Fuller et al. 2015, 88-134 cm BET in Schaefer & Fuller 2002, 54-159 cm BET in Schaefer &
398Fuller 2010) than those marked with passive acoustic tags (38-90 cm BET in Phillips et al.
3992019), even though size ranges largely overlap. This suggests that small BET spend a higher

400 proportion of their time associated with FOBs than large individuals. This agrees with
401 observed size distributions of DFAD catches, where smaller individuals are caught, and with
402 the negative correlation between BET individual length and percentage of time associated
403 found by Schaefer & Fuller (2002). However, Schaefer et al. (2009) found lower association
404 percentages with archival tags on small tunas (10.4 and 15.9 % of the time associated for
405 51-60 cm FL YFT and 65-99 cm FL BET respectively) than the work of Phillips et al. (2019)
406 with acoustic tagging. This could suggest a potential bias of the different methodologies that
407 should be further investigated, as a small percentage of time associated with floating objects
408 would indicate no or little influence of DFADs, while a high percentage could indicate a
409 potentially significant influence of DFADs on large-scale movements, which could result in an
410 ecological trap.

411 We are therefore far from understanding the effects of different densities of floating objects
412 on tuna fine-scale movements nor the link between fine-scale and large-scale movements.
413 Most electronic tagging efforts have been done on YFT and to a lesser extent BET, but more
414 behavioural data are clearly needed for all three tropical tuna species. One of the main
415 difficulty is to disentangle the effects of DFADs from the impacts of other external signals
416 (e.g. prey density) which can also influence tuna associative behaviour (Lopez et al. 2017,
417 Nooteboom et al. 2023, Schaefer et al., 2009, Schaefer & Fuller, 2010).

418 *Effects on schooling behaviour*

419 DFADs could also affect schooling behaviour, which can have a wide range of consequences
420 on the **life-history parameters** and the movements of tunas. Dagorn & Fréon (1999) and
421 Fréon & Dagorn (2000) suggested that tunas could associate with floating objects for social
422 advantages such as facilitating schooling behaviour. To date no result has been obtained **on**
423 **tropical tuna** from DFADs regarding this question. If floating objects facilitate the schooling
424 behaviour of tunas, then the deployment of large numbers of DFADs may have effects on
425 school size, either by facilitating the formation of large (but less) schools or decreasing
426 school size with DFADs offering too many aggregation sites (**Figure 2**, Dagorn et al., 2010).
427 DFADs could also modify the size structure of tuna schools, allowing the formation of large

428 aggregations composed of several unassociated schools of different size structures (Wang et
429 al. 2012). Sempo et al. (2013) modelled the impact of the increasing deployment of DFADs
430 on the distribution of social fish species such as tunas. They demonstrated that for social
431 species, increasing the number of DFADs does not necessarily lead to an increase in the
432 total amount of tuna associated with DFADs, a non-intuitive result. Capello et al. (2022) also
433 showed that the number of DFADs with associated schools and the size of associated
434 schools were not linearly related to the total number of DFADs and that this relationship
435 varied according to the considered social scenario.

436 Do DFADs modify the **life-history parameters** of tropical tunas?

437 The increasing number of DFADs at sea also raises questions regarding their effect on the
438 feeding strategy of tropical tuna, and related energy-dependent traits such as tuna health
439 (monitored for example by body condition), growth, reproduction and natural mortality.

440 *Effects on feeding*

441 In his review on DFAD impacts on tropical tunas, Taquet (2013) recommended comparative
442 analyses of stomach contents on tropical tunas. Such analyses have shown that small-sized
443 tunas may not feed while associated with DFADs in the Atlantic (Hallier & Gaertner, 2008;
444 Ménard et al., 2000), Indian (Grande et al., 2013; Jaquemet et al., 2011; Zudaire et al., 2015,
445 Malone et al. 2011, Hallier & Gaertner, 2008) and Western and Central Pacific (Machful et
446 al., 2021) oceans. Small YFT and SKJ captured in DFAD-associated schools had a higher
447 fraction of empty stomachs, lower stomach fullness or daily food rates (in the EAO; Hallier &
448 Gaertner 2008, Ménard et al. 2000; and in the IO, Hallier & Gaertner 2008), and lower prey
449 weight (in the IO; Grande et al. 2013, Zudaire et al. 2015) than those captured in free-
450 swimming schools. These results support the hypothesis that the quantity of prey present in
451 DFAD assemblages is not sufficient to sustain the dietary requirements of large aggregations
452 of small-sized tunas commonly found at DFADs (several tens of tons) (Fréon & Dagorn,
453 2000). However, except in Hallier & Gaertner (2008), the influence of the sampling time of
454 tunas on the stomach content was not taken into account. Purse seine vessels mainly fish on

455 DFADs at dawn (Forget et al., 2015) and on free-swimming schools during daytime. First,
456 feeding activities are believed to often take place in the early evening on organisms
457 performing diel vertical migration between the deep scattering layer and the surface
458 (Schaefer & Fuller, 2002), resulting in prey being fully digested by the time the fish are
459 caught and sampled, at dawn. Second, free-swimming schools of tunas are almost
460 exclusively caught when actively feeding at the sea surface, hence higher levels of stomach
461 fullness are to be expected. Nevertheless, the association of tunas with DFADs could also
462 affect the composition and quality of their diet, as shown for YFT in the Western IO (WIO)
463 and WCPO and for SKJ in the WCPO (Zudaire et al. 2015, Allain et al. 2010). Differences of
464 diet composition were observed for these species, that may be due to their associative
465 behaviour despite the above-mentioned sampling bias, with associated tuna in the WCPO
466 feeding on shallower prey than free-swimming tunas (Allain et al. 2010), which is in
467 agreement with YFT staying closer to the surface when associated (Holland and Brill, 1990,
468 Schaefer et al. 2009).

469 Independently of the trophic role of DFADs, the deployment and the drift trajectories of
470 DFADs could create new zones of high floating object densities, which may be unfavourable
471 for the foraging success of tunas (Marsac et al. 2000). Jaquemet et al. (2011) partitioned
472 their samples in “rich” (i.e., no limiting food) versus “poor” forage areas in the Indian Ocean,
473 in relation to an exceptional demographic outburst of a pelagic stomatopod (Crustacea),
474 which composed the bulk of tuna diet in this region (Potier et al., 2004, 2007). These authors
475 found that in “rich” forage areas, DFADs have no impact on the feeding pattern of tunas,
476 whereas in “poor” forage areas, tunas associated with DFADs had lower stomach fullness
477 compared to tunas in free-swimming schools. Jaquemet et al. (2011) suggested that the
478 impact of DFADs on feeding success could be location-dependent. This led the authors to
479 emphasize the possible detrimental effect on the condition of tuna associated with DFADs if
480 associated tunas drift towards areas with poor forage resources. However, such an effect
481 relies on the assumption that a tuna’s probability to depart from a DFAD is independent of
482 their local environment which seems in disagreement with behavioral studies (Fuller et al.
483 2015, Nootboom et al. 2023).

484 In the Pacific Ocean, Hunsicker et al. (2012) observed that predation on SKJ and YFT by
485 large pelagic fishes sampled from DFAD sets was greater than for those captured via other
486 fishing methods (e.g. free-swimming schools). These authors concluded that by aggregating
487 small-sized SKJ, YFT, and BET, DFADs enhance their vulnerability to predators such as
488 sharks and billfishes, and thus increase natural mortality of small sized tunas. To our
489 knowledge, this is the only study assessing the impact of DFADs on tuna vulnerability to
490 predators, hence additional data from other regions would be needed for further testing these
491 assumptions.

492 *Effects on body condition*

493 Tuna condition has been investigated using different methods: biometric condition factors
494 (e.g. plumpness), and biochemical indices (e.g., fat and water contents, lipid class
495 composition). Gaertner et al. (1999) in a preliminary investigation did not find evidence of a
496 morphometric difference between free-swimming school or DFAD-caught tunas in the EAO.
497 But Marsac et al. (2000) in the EAO and Hallier & Gaertner (2008) in the WIO found that
498 individuals associated with DFADs were in lower condition than those in free-swimming
499 schools, using morphometric indicators (thorax width or girth, plumpness of fish) as fish
500 health indicators. Robert et al. (2014) measured the condition of SKJ using BIA (Bioelectrical
501 Impedance Analysis), a non-invasive field tool that estimates body water content (inversely
502 correlated with body fat content), and determines total lipid and main lipid class
503 concentrations. They confirmed the lower condition of SKJ associated with floating objects
504 compared to those in free-swimming schools. Because the studied area (Mozambique
505 Channel, WIO) is naturally rich with NLOGs and had undergone little habitat modifications
506 due to DFADs at the time of the study (Dagorn et al., 2013a, Dupaix et al. 2021), the authors
507 concluded that before the use of DFADs, tunas associated with logs could have also been in
508 lower condition than tunas in free-swimming schools. These results can mean that (1) a
509 lower measured condition could reflect normal variations and does not necessarily imply
510 detrimental physiological consequences, or (2) some specific areas where NLOGs have

511always been in high numbers could also have negatively impacted the condition of tunas that
512passed through and stayed in these areas.

513As a lower condition measured at DFADs does not necessarily imply longer-term detrimental
514consequences for tuna, there is a need to monitor tuna condition and other biological
515parameters on a longer term and determine if they are influenced by the density of DFADs in
516the area. Dupaix et al. (2023), using length-weight data from 1987 to 2018 in the WIO, found
517no decreasing trend of YFT condition over the studied period, during which the number of
518DFADs has increased. Hence, this study, using one morphometric indicator as a proxy for
519condition (Le Cren's relative condition factor), suggests the absence of a long-term impact of
520DFADs on YFT condition, under the conditions encountered in the WIO in the last three
521decades. Nevertheless, it should be noted that other factors could also have counteracted
522potential negative effects of DFADs on tuna condition.

523Studies that investigated potential DFAD impacts on tuna condition mainly suggest that the
524condition of associated tuna is lower than that of free-swimming tuna. These results are
525reinforced by the example of the preparation of katsuobushi (shaved dried SKJ) in Japan.
526Indeed the Japanese tuna industry prefers SKJ caught on DFADs as they have less fat than
527those from free-swimming schools (Nishida, pers. comm.). However, Sardenne et al. (2016),
528when comparing biometric and biochemical indicators found inconsistencies due to a high
529variability of biometric indicators with season and ontogeny. They concluded that biometric
530indicators measured on whole tuna (e.g. thorax girth, fish plumpness, Le Cren's K_n) should
531be interpreted with caution as they may not always reflect the energetic condition measured
532in the tissues of the fish. Experimental validation of the condition factors used is needed to
533determine the potential impacts and the underlying mechanisms of the difference in tuna
534condition. For example, condition factors could be calibrated and validated by monitoring
535them during fasting experiments on captive tuna, although measuring some of them regularly
536in experimental conditions could represent a methodological challenge.

537 *Effects on reproduction and growth*

538 In the WIO, Zudaire et al. (2014, 2015) found (i) a significantly higher proportion of energy-
539 rich fish prey in the diet (stomach contents; Zudaire et al., 2015), as well as (ii) significantly
540 higher total lipid concentrations and triacylglycerol to sterol (TAG:ST) ratio, indicators of
541 energetic condition, in the gonads of YFT females caught in free-swimming schools
542 compared to females associated with DFADs (Zudaire et al., 2014). This can be interpreted
543 as simply reflecting differences in prey availability and feeding activity and thus differential
544 lipid incorporation to tissues between DFAD-associated and non-associated tunas. It could
545 also highlight higher energetic investment to reproduction in free-swimming YFT due to a
546 higher condition (i.e., better health), keeping in mind the potential bias provoked by an
547 uneven size distribution between school types in these studies. However, the study failed to
548 demonstrate a direct effect on the fecundity, most likely due to the low number of actively
549 spawning females analysed and the high inter-individual fecundity variability observed in YFT
550 (Pecoraro et al., 2017).

551 Similarly, Ashida et al. (2017) investigated the difference in reproductive traits of female SKJ,
552 of similar size distribution, between school types in the WCPO, highlighting a significant
553 higher proportion of mature females in free swimming schools, characterised by higher
554 relative condition factor, than associated with DFADs. However, as for YFT in the IO (Zudaire
555 et al. 2014), no significant effect of the school type was observed on the WCPO SKJ
556 fecundity, which corroborates previous results observed for WIO SKJ (Grande, 2013; Grande
557 et al. 2014). The lack of relationship between condition and fecundity of SKJ could be related
558 to their energy allocation and reproductive strategies. SKJ tuna females fuel their gametes
559 with energy gained concomitantly during reproduction (i.e., income breeding strategy,
560 Grande et al. 2016). However, YFT females can store additional energy reserves prior to
561 spawning, which define them as income-capital breeder (Zudaire et al., 2014) unlike SKJ.
562 Therefore, as SKJ females exhibit better condition when free swimming, it can be assumed
563 that their reproductive efficiency is lower when associated with DFADs, but the same
564 conclusion cannot be made for YFT.

565 Using tagging data collected in the EAO, Hallier & Gaertner (2008) estimated and compared
566 the growth rates of SKJ and YFT associated with DFADs versus free-swimming schools.
567 Released and recaptured SKJ associated with DFADs had a significantly lower growth rate
568 than those in free-swimming schools, but the difference was not significant for YFT (though it
569 was lower, as for SKJ). However, the history experienced by individual fish between release
570 and recapture was unknown. The “experimental” design could not be controlled as the time
571 one specimen spent associated with DFADs and in free-swimming schools is not available.
572 In addition, the authors were only able to process a small sample of free-swimming YFT (n =
573 10).

574 **What are the scientific challenges to fill the knowledge gaps?**

575 DFADs have been representing one of the key management priorities and challenges of
576 RFMOs over the last decade. Since fishers started using them, DFADs numbers
577 continuously increased until first management measures limiting the number of operational
578 buoys were adopted in the mid-2010s (Song & Shen 2022). The massive use of DFADs in all
579 oceans has been generating major concerns on the sustainability of this fishing mode.
580 DFADs increase the catchability of tropical tunas leading to large catches of small BET and
581 YFT (Dagorn et al., 2013b; Fonteneau et al., 2013), generate more bycatch, including
582 vulnerable species such as some shark species, silky (*Carcharhinus falciformis*) and oceanic
583 whitetip (*Carcharhinus longimanus*) sharks (Dagorn et al., 2013b; Fonteneau et al., 2013;
584 Leroy et al., 2013), and can strand on coastal areas causing damage to marine habitats
585 (Imzilen et al., 2021; Maufroy et al., 2015, Escalle et al. 2019). Although there is increasing
586 knowledge and literature on DFADs, the issue of their indirect impacts (not related to fishing
587 mortality) on tropical tuna remains a scientific debate. All knowledge collected and reviewed
588 on the behaviour and life-history parameters of tunas at DFADs clearly reveals a lack of
589 converging scientific results on the long term consequences on tuna (at the individual or
590 population levels) of increased numbers of floating objects. Therefore, if DFADs seem to
591 affect the short-term condition of tropical tunas, we are not currently able to conclude
592 whether DFADs affect the movements and/or other life-history parameters of tunas in a way

593that could significantly affect the fitness of individuals and the demography of their
594populations. As such, there is a need to improve the observation and understand this
595associative phenomenon to provide scientific advice on the effects of DFADs on the life-
596history parameters and behaviour of tropical tunas and other associated species.

597A major gap in tuna and DFAD science is the lack of time series of key parameters such as
598the numbers of DFADs and natural floating objects, residence and absence times at DFADs
599as well as large movements between oceanic regions, school sizes, condition and
600reproduction indices. The first research priority in this context is to initiate or continue time
601series of such indicators (Capello et al. 2023). Setting long-term monitoring programs in
602every ocean appears to be a priority, as effects of DFADs could vary depending on the
603species, the characteristics of each ecosystem and on the density of floating objects.
604Moreover, it would facilitate comparative analyses between oceans to better understand the
605drivers of tuna associative behaviour. The collection of some parameters will require
606dedicated scientific surveys (e.g. electronic tagging, biological sampling) while others (e.g.
607numbers of DFADs and natural objects, biological condition factors) have started to be
608routinely collected by tRFMOs through FAD-specific data requirements included in
609conservation and management measures (Grande et al. 2018, Báez et al. 2022, Song &
610Shen 2022) as well as government and industry initiatives (e.g., routine fishery monitoring
611and at-sea observer programs).

612Another research priority is to develop experimental studies to identify the biological and
613behavioural processes involved in the associative behaviour. The only scientific consensus is
614the fact that in a given area, conditions of tunas associated with floating objects seem to
615often be lower than those of fish in free-swimming schools. However, the different indicators
616used to assess tuna condition are not always well correlated (Sardenne et al. 2016), and
617experimental studies are needed to validate them against proper benchmarks, allowing to
618determine how representative they are of individuals' health. Then, understanding how fast
619these indicators change with the fish's associative behaviour appears essential. This could
620also be achieved through studies on captive tropical tunas (e.g., Estess et al. 2017), but non
621lethal observations should be promoted (e.g. BIA) in order to track changes throughout the

622fish lifespan. No evidence exists suggesting whether the lower condition at DFADs is the
623consequence or the cause of their association with DFADs. Often the robustness of the
624findings of **investigation on the life-history parameters** of tunas was hampered by the lack of
625knowledge of the time spent associated with a DFAD or in an array of DFADs by each
626specimen analysed. The history of each individual tuna is a hidden variable that must be
627taken into account in statistical analyses, which is a challenge. Studies combining
628behavioural observations (tagging) and condition of the individuals (e.g. BIA or biochemical
629analyses of biopsies made at the time of tagging) should then be encouraged. Ideally, tags
630equipped with physiological sensors would clearly help understanding the interplay between
631associative behaviour and tuna physiology. Such tags, however, are **only starting** to be
632developed.

633Studies on AFADs could provide insights to the questions addressed in this manuscript: as
634argued by Dagorn et al. (2010), AFADs also alter the natural environment (by adding floating
635objects to the ocean). However, it remains questionable if they are comparable due to the
636fact that AFADs are generally located nearshore, with corresponding particular
637oceanographic conditions, and they do not move with water masses. Papua New Guinea, the
638Philippines and Indonesia are examples of areas with very high numbers of AFADs (Proctor
639et al. 2019) and as such, these dense arrays of AFADs could generate the same concerns
640on tuna **life-history parameters** and behaviour that those expressed for DFADs.
641Understanding the behaviour of tunas around AFADs can also improve our general
642understanding of tunas around all types of floating objects and help design new, well focused
643studies for DFADs. **For practical reasons, more** studies have been performed on the
644behaviour of tuna at AFADs **than at DFADs** (e.g. Dagorn et al., 2007; Govinden et al., 2013;
645Holland et al., 1990; Ohta & Kakuma, 2005; Rodriguez-Tress et al., 2017). **They provided**
646estimates of residence times between two AFAD associations and therefore **of** the
647percentage of time spent associated to AFADs (e.g. Pérez et al., 2020; Robert et al., 2013b;
648Rodriguez-Tress et al., 2017), **which still needs to be further explored at DFADs**. For
649example, Pérez et al. (2020) used acoustic tagging data on AFAD arrays to demonstrate that
650when inter-AFAD distance decreases, tuna visit more AFADs, spend less time travelling

651between AFADs and more time associated with them. Concerning DFADs, as actual
652densities of drifting floating objects are difficult to obtain, studies using a modelling approach
653based on experimental data should be promoted (Pérez et al. 2022, Capello et al. 2023).
654These studies should investigate the consequences of changes in floating object density on
655tuna school sizes and associative behaviour. These modelling studies could be
656complemented and/or calibrated by studies which use data from echosounder buoys
657deployed by fishers on floating objects. Recent methodological advances allowed the
658prediction of tuna presence or absence under FOBs (Baidai et al., 2020; Orue et al., 2020).
659Using an extensive dataset from echosounder buoys in the WCPO (more than 3.8 million
660transmissions), Escalle et al. (2021c) determined different profiles of acoustic signals related
661to different types of aggregations. Other studies also show that multi-frequency echosounder
662buoys could allow the discrimination of tropical tuna species under DFADs (Moreno et al.
6632019, Sobradillo et al. 2023). These new methodological developments, in combination with
664tagging data both conventional and electronic, and modelling approaches offer promising
665perspectives for the study of tuna aggregation behaviour under FOBs and the potential
666impact of DFAD density on tuna schooling behaviour.

667Tuna RFMOs set limits on the number of operational buoys (IATTC: up to 340 depending on
668the vessel size, Res C-21-04; ICCAT: 300 in Rec 22-01; IOTC: 250 in Res 23/02; WCPFC:
669350 in CMM 2021-01). These limits are essentially set to control the fishing effort and the
670catches of tunas and non-target species, but how such limits also limit the number of
671deployed DFADs is not known. In theory, all purse seine vessels could use at least the same
672amount of DFADs than the maximum number of operational buoys authorized in each of the
673regions. Multiplying this maximum number of operational buoys authorized per vessel by the
674number of purse seine vessels in each ocean provides a global authorized limit of about
675233,000 operational buoys (Supplementary Materials 1). This number is about twice higher
676than the estimate of the global number of DFADs deployments made by Gershman et al.
677(2015), based on data from 2013. Hence, it would mean that the global purse seine fishery
678could have increased the number of DFADs in the ocean while still respecting the current
679limits on the number of active buoys. Most tRFMOs now require that DFAD identification,

680 characteristics, deployment date and deployment location are reported (e.g. ICCAT Rec 22-
681 01; IOTC Res. 23/02; IATTC C-21-04). Some studies evidenced different regional trends of
682 DFAD deployments (IOTC, 2022, Escalle et al. 2020, Lopez et al. 2021, Floch et al., 2019;
683 Maufroy et al., 2017), but no study assessed this trend on a global scale after 2013
684 (Gershman et al., 2015). Tuna RFMOs should continue to collect and make fine-scale DFAD
685 data available to scientists to allow regular estimations of the extent of the habitat
686 modifications generated by DFADs, which should be addressed at a global scale.

687 **Conclusion**

688 To summarize the questions formulated in this study:

689 (1) although the deployment of DFADs has undoubtedly modified the habitat of tropical
690 tunas, the extent of this modification still needs to be better characterized in some regions.

691 This characterization can be achieved through the continued monitoring of indicators (e.g.
692 spatialized DFAD and NLOG densities, DFAD/NLOG ratio) collected by tRFMOs.

693 (2) studies assessing the impacts of DFADs on tuna large-scale movements show
694 contradictory results. Strong ocean and species specific variability is observed for the
695 proportion of time spent associated with FOBs. However, the effect of the methodology used
696 (archival tagging vs acoustic tagging) should be investigated. To date, besides theoretical
697 studies, no evidence has been shown on the impact of DFADs on associative and schooling
698 behaviour.

699 (3) DFADs probably impact tuna short-term condition, but it does not necessarily imply a
700 longer-term detrimental effect and should be confirmed with long-term time series of
701 validated condition indicators. The results on the impacts of DFADs on other life-history
702 parameters are inconclusive.

703 (4) The main conclusion of this work is the lack of clear converging scientific results on the
704 indirect impacts of DFADs on the behaviour and life-history parameters of tropical tuna. It
705 should therefore be underlined that scientific efforts should not only focus on the direct
706 effects of DFADs on catches (target and non-target species) but should also address other

707possible impacts, such as density dependent effects on the behaviour and **life-history**
708**parameters** of tunas. This **current lack of converging results** justifies a major and urgent
709scientific effort, in terms of data collection, experimental research and modelling to tackle
710definitively whether the increased **deployment** of DFADs could lead to indirect impacts on
711tropical tuna **populations**.

712**Acknowledgements**

713The initial impetus for this work was a workshop funded by the International Seafood
714Sustainability Foundation (ISSF, 2014). The research was conducted independently by the
715authors and its results, professional opinions and conclusions are solely the work of the
716authors. There are no contractual obligations between ISSF and the authors that might
717influence the results, professional opinions and conclusions. For the purpose of Open
718Access, a CC-BY public copyright licence has been applied by the authors to the present
719document and will be applied to all subsequent versions up to the Author Accepted
720Manuscript arising from this submission.

721**Data availability statement**

722Data used in the supplementary material were obtained from Justel-Rubio et al. (2022) and
723the following website: <https://www.iattc.org/en-US/Management/Vessel-register> (accessed
7242023/03/14)

725References

- 726Abascal, F. J., Peatman, T., Leroy, B., Nicol, S., Schaefer, K., Fuller, D. W., & Hampton, J.
727(2018). Spatiotemporal variability in bigeye vertical distribution in the Pacific Ocean.
728*Fisheries Research*, 204, 371–379. <https://doi.org/10.1016/j.fishres.2018.03.013>
- 729Allain, V. (2010). *Trophic structure of the pelagic ecosystems of the western and central*
730*Pacific Ocean* (6th Regular Session of the Scientific Commity EB-IP-10). Western and
731Central Pacific Fisheries Commission. <https://meetings.wcpfc.int/index.php/node/7069>
- 732Amandè, M. J., Chassot, E., Chavance, P., Murua, H., de Molina, A. D., & Bez, N. (2012).
733Precision in bycatch estimates: The case of tuna purse-seine fisheries in the Indian Ocean.
734*ICES Journal of Marine Science*, 69(8), 1501–1510. <https://doi.org/10.1093/icesjms/fss106>
- 735Ariz, J., Delgado, A., Fonteneau, A., Gonzalez Costas, F., & Pallares, P. (1999). Logs and
736tunas in the Eastern Tropical Atlantic. A review of present knowledge and uncertainties. In
737M. D. Scott, W. H. Bayliff, C. E. Lennert-Cody, & K. M. Schaefer (Eds.), *Proceedings of the*
738*International Workshop on Fishing for Tunas Associated with Floating Objects* (Vol. 11, pp.
73921–65). Inter-American Tropical Tuna Commission.
- 740Arrizabalaga, H., Ariz, J., Mina, X., De Molina, A. D., Artetxe, I., Pallares, P., & Iriondo, A.
741(2001). *Analysis of the activities of supply vessels in the Indian Ocean from observers data*
742(Working Party on Tropical Tunas (WPTT)). Indian Ocean Tuna Commission.
743<https://iotc.org/documents/analysis-activities-supply-vessels-indian-ocean-observers-data>
- 744Ashida, H., Tanabe, T., & Suzuki, N. (2017). Difference on reproductive trait of skipjack tuna
745*Katsuo* *pelamis* female between schools (free vs FAD school) in the tropical western and
746central Pacific Ocean. *Environmental Biology of Fishes*, 100(8), 935–945.
- 747Báez, J. C., Déniz, S., Ramos, M. L., Grande, M., Ruiz, J., Murua, H., Santiago, J., Justel-
748Rubio, A., Herrera, M., Moniz, I., Lopez, J., Pascual-Alayón, P. J., Muniategi, A., Alzorric,
749N., González-Carballo, M., Rojo, V., & Abascal, F. (2022). Data Provision for Science-Based
750FAD Fishery Management: Spanish FAD Management Plan as a Case Study. *Sustainability*,
75114(6), Article 6. <https://doi.org/10.3390/su14063278>
- 752Baidai, Y., Dagorn, L., Amande, M. J., Gaertner, D., & Capello, M. (2020). Machine learning
753for characterizing tropical tuna aggregations under Drifting Fish Aggregating Devices
754(DFADs) from commercial echosounder buoys data. *Fisheries Research*, 229, 105613. <https://doi.org/10.1016/j.fishres.2020.105613>
- 756Baske, A., Gibbon, J., Benn, J., & Nickson, A. (2012). Estimating the use of drifting Fish
757Aggregation Devices (FADs) around the globe. *PEW Environment Group - Discussion Paper*,
7581–8.
- 759Battin, J. (2004). When Good Animals Love Bad Habitats: Ecological Traps and the
760Conservation of Animal Populations. *Conservation Biology*, 18(6), 1482–1491.
761<https://doi.org/10.1111/j.1523-1739.2004.00417.x>
- 762Brehmer, P., Sancho, G., Trygonis, V., Itano, D., Dalen, J., Fuchs, A., Faraj, A., & Taquet, M.
763(2019). Towards an autonomous pelagic observatory: Experiences from monitoring fish
764communities around drifting FADs. *Thalassas: An International Journal of Marine Sciences*,
76535, 177–189.
- 766Bromhead, D., Foster, J., Attard, R., Findlay, J., & Kalish, J. (2003). *A review of the impact of*
767*fish aggregating devices (fads) on tuna fisheries: Final report to the fisheries resources*
768*research fund*. Bureau of Rural Sciences.

769Caddy, J. F., & Majkowski, J. (1996). Tuna and trees: A reflection on a long-term perspective
770for tuna fishing around floating logs. *Fisheries Research*, 25(3), 369–376.
771[https://doi.org/10.1016/0165-7836\(95\)00449-1](https://doi.org/10.1016/0165-7836(95)00449-1)

772Capello, M., Merino, G., Tolotti, M., Murua, H., & Dagorn, L. (2023). Developing a science-
773based framework for the management of drifting Fish Aggregating Devices. *Marine Policy*,
774153, 105657. <https://doi.org/10.1016/j.marpol.2023.105657>

775Capello, M., Rault, J., Deneubourg, J.-L., & Dagorn, L. (2022). Schooling in habitats with
776aggregative sites: The case of tropical tuna and floating objects. *Journal of Theoretical*
777*Biology*, 547, 111163. <https://doi.org/10.1016/j.jtbi.2022.111163>

778Castro, J. J., Santiago, J. A., & Santana-Ortega, A. T. (2002). A general theory on fish
779aggregation to floating objects: An alternative to the meeting point hypothesis. *Reviews in*
780*Fish Biology and Fisheries*, 11, 255–277.

781Cayré, P., Diouf, T., Fonteneau, A., & Santa-Rita Vieira, M. H. (1986). Analyse de Données
782de Marquages et Recaptures de Listao (*Katsuwonus pelamis*) réalisées par le Sénégal et la
783République du Cap-Vert (pp. 309–316). International Commission for the Conservation of
784Atlantic Tunas. https://www.iccat.int/Documents/CVSP/SJ001_1986/SJYrProg01.html

785Chassot, E., Goujon, M., Maufroy, A., Cauquil, P., Fonteneau, A., & Gaertner, D. (2014). The
786use of artificial fish aggregating devices by the French tropical tuna purse seine fleet:
787Historical perspective and current practice in the Indian Ocean. *16ème Groupe de Travail Sur*
788*Les Thons Tropicaux*. CTOI, Victoria, p. 17p.

789Dagorn, L., Bez, N., Fauvel, T., & Walker, E. (2013a). How much do fish aggregating
790devices (FADs) modify the floating object environment in the ocean? *Fisheries*
791*Oceanography*, 22(3), 147–153. <https://doi.org/10.1111/fog.12014>

792Dagorn, L., Filmalter, J. D., Forget, F., Amandè, M. J., Hall, M. A., Williams, P., Murua, H.,
793Ariz, J., Chavance, P., & Bez, N. (2012). Targeting bigger schools can reduce ecosystem
794impacts of fisheries. *Canadian Journal of Fisheries and Aquatic Sciences*, 69(9), 1463–1467.

795Dagorn, L., & Fréon, P. (1999). Tropical tuna associated with floating objects: A simulation
796study of the meeting point hypothesis. *Canadian Journal of Fisheries and Aquatic Sciences*,
79756(6), 984–993.

798Dagorn, L., Holland, K. N., & Filmalter, J. (2010). Are drifting FADs essential for testing the
799ecological trap hypothesis? *Fisheries Research*, 106(1), 60–63.
800<https://doi.org/10.1016/j.fishres.2010.07.002>

801Dagorn, L., Holland, K. N., & Itano, D. G. (2007). Behavior of yellowfin (*Thunnus*
802*albacares*) and bigeye (*T. obesus*) tuna in a network of fish aggregating devices (FADs).
803*Marine Biology*, 151(2), 595–606.

804Dagorn, L., Holland, K. N., Restrepo, V., & Moreno, G. (2013b). Is it good or bad to fish with
805FADs? What are the real impacts of the use of drifting FADs on pelagic marine ecosystems?
806*Fish and Fisheries*, 14(3), 391–415. <https://doi.org/10.1111/j.1467-2979.2012.00478.x>

807Davies, T. K., Mees, C. C., & Milner-Gulland, E. J. (2014). The past, present and future use
808of drifting fish aggregating devices (FADs) in the Indian Ocean. *Marine Policy*, 45, 163–170.
809<https://doi.org/10.1016/j.marpol.2013.12.014>

810Delgado de Molina, A., Ariz, J., Murua, H., Santana, J. C., Ramos, L., & Soto, M. (2014).
811Spanish Fish Aggregating Device Management Plan. Preliminary data in the Indian Ocean
812(IOTC-2014-WPTT16-19). Indian Ocean Tuna Commission.
813[https://iotc.org/sites/default/files/documents/2014/11/IOTC-2014-WPTT16-19_Rev_1 -](https://iotc.org/sites/default/files/documents/2014/11/IOTC-2014-WPTT16-19_Rev_1_-_Spanish_FAD_Mgt_plan.pdf)
814 [Spanish FAD Mgt plan.pdf](https://iotc.org/sites/default/files/documents/2014/11/IOTC-2014-WPTT16-19_Rev_1_-_Spanish_FAD_Mgt_plan.pdf)

815Dempster, T., & Taquet, M. (2004). Fish aggregation device (FAD) research: Gaps in current
816knowledge and future directions for ecological studies. *Reviews in Fish Biology and*
817*Fisheries*, 14(1), 21–42.

818Dupaix, A., Capello, M., Lett, C., Andrello, M., Barrier, N., Viennois, G., & Dagorn, L.
819(2021). Surface habitat modification through industrial tuna fishery practices. *ICES Journal*
820*of Marine Science*, 78(9), 3075–3088. <https://doi.org/10.1093/icesjms/fsab175>

821Dupaix, A., Dagorn, L., Duparc, A., Guillou, A., Deneubourg, J.-L., & Capello, M. (2023).
822No evidence from long- term analysis of yellowfin tuna condition that Drifting Fish
823Aggregating Devices act as ecological traps. *Marine Ecology Progress Series*, 711, 121–127.
824<https://doi.org/10.3354/meps14313>

825Escalle, L., Gaertner, D., Chavance, P., Murua, H., Simier, M., Pascual-Alayón, P. J.,
826Ménard, F., Ruiz, J., Abascal, F., & Mérigot, B. (2018). Catch and bycatch captured by
827tropical tuna purse-seine fishery in whale and whale shark associated sets: Comparison with
828free school and FAD sets. *Biodiversity and Conservation*, 28(2), 467–499.
829<https://doi.org/10.1007/s10531-018-1672-1>

830Escalle, L., Hare, S. R., Vidal, T., Brownjohn, M., Hamer, P., & Pilling, G. (2021a).
831Quantifying drifting Fish Aggregating Device use by the world’s largest tuna fishery. *ICES*
832*Journal of Marine Science*, fsab116. <https://doi.org/10.1093/icesjms/fsab116>

833Escalle, L., Muller, B., Vidal, T., Hare, S., Hamer, P., & PNA Office. (2021b). *Report on*
834*analyses of the 2016/2021 PNA FAD tracking programme* (17th Regular Session of the
835Scientific Commity No. SC17-MI-IP-04). Western and Central Pacific Fisheries Commission.
836<https://meetings.wcpfc.int/node/12589>

837Escalle, L., Phillips, J. S., Brownjohn, M., Brouwer, S., Gupta, A. S., Sebille, E. V., Hampton,
838J., & Pilling, G. (2019). Environmental versus operational drivers of drifting FAD beaching in
839the Western and Central Pacific Ocean. *Scientific Reports*, 9(1), 1–12. <https://doi.org/10.1038/s41598-019-50364-0>

841Escalle, L., Vidal, T., Hare, S., Hamer, P., Pilling, G., & Office, P. (2020). *Estimates of the*
842*number of FAD deployments and active FADs per vessel in the WCPO* (16th Regular Session
843of the Scientific Commity No. SC16-MI-IP-13). Western and Central Pacific Fisheries
844Commission. <https://meetings.wcpfc.int/node/11729>

845Escalle, L., Vidal, T., Heuvel, B., Clarke, R., Hare, S., Hamer, P., & Pilling, G. (2021c).
846*Project 88 final report: Acoustic FAD analyses* (17th Regular Session of the Scientific
847Commity No. SC17-MI-IP-05). Western and Central Pacific Fisheries Commission.
848<https://meetings.wcpfc.int/node/12590>

849Estess, E. E., Klinger, D. H., Coffey, D. M., Gleiss, A. C., Rowbotham, I., Seitz, A. C.,
850Rodriguez, L., Norton, A., Block, B., & Farwell, C. (2017). Bioenergetics of captive
851yellowfin tuna (*Thunnus albacares*). *Aquaculture*, 468, 71–79.
852<https://doi.org/10.1016/j.aquaculture.2016.09.053>

853Evans, K., Young, J. W., Nicol, S., Kolody, D., Allain, V., Bell, J., Brown, J. N., Ganachaud,
854A., Hobday, A. J., Hunt, B., Innes, J., Gupta, A. S., van Sebille, E., Kloser, R., Patterson, T.,
855& Singh, A. (2015). Optimising fisheries management in relation to tuna catches in the
856western central Pacific Ocean: A review of research priorities and opportunities. *Marine*
857*Policy*, 59, 94–104. <https://doi.org/10.1016/j.marpol.2015.05.003>

858Filmlalter, J. D., Capello, M., Deneubourg, J.-L., Cowley, P. D., & Dagorn, L. (2013).
859Looking behind the curtain: Quantifying massive shark mortality in fish aggregating devices.
860*Frontiers in Ecology and the Environment*, 11(6), 291–296. <https://doi.org/10.1890/130045>

861Filous, A., Friedlander, A. M., Griffin, L., Lennox, R. J., Danylchuk, A. J., Mereb, G., &
862Golbuu, Y. (2020). Movements of juvenile yellowfin tuna (*Thunnus albacares*) within the

863coastal FAD network adjacent to the Palau National Marine Sanctuary: Implications for local
864fisheries development. *Fisheries Research*, 230, 105688.
865<https://doi.org/10.1016/j.fishres.2020.105688>

866Floch, L., Depetris, M., Dewals, P., & Duparc, A. (2019). Statistics of the French Purse Seine
867Fishing Fleet Targeting Tropical Tunas in the Indian Ocean (1981-2018). *Report of the 21st*
868*Session of the IOTC Working Party on Tropical Tunas*, 1–27.
869<https://www.iotc.org/documents/WPTT/21/11>

870Fonteneau, A., Chassot, E., & Bodin, N. (2013). Global spatio-temporal patterns in tropical
871tuna purse seine fisheries on drifting fish aggregating devices (DFADs): Taking a historical
872perspective to inform current challenges. *Aquatic Living Resources*, 26(1), 37–48.
873<https://doi.org/10.1051/alr/2013046>

874Fonteneau, A., Chassot, E., & Gaertner, D. (2015). Managing tropical tuna purse seine
875fisheries through limiting the number of drifting fish aggregating devices: Food for thought.
876*Collect. Vol. Sci. Pap. ICCAT*, 71(1), 460–475.
877https://www.iccat.int/Documents/CVSP/CV071_2015/n_1/CV071010460.pdf

878Fonteneau, A., Pallarés, P., & Pianet, R. (2000). *A worldwide review of purse seine fisheries*
879*on FADs*. 28, 15–35.

880Forget, F. G., Capello, M., Filmalter, J. D., Govinden, R., Soria, M., Cowley, P. D., &
881Dagorn, L. (2015). Behaviour and vulnerability of target and non-target species at drifting fish
882aggregating devices (FADs) in the tropical tuna purse seine fishery determined by acoustic
883telemetry. *Canadian Journal of Fisheries and Aquatic Sciences*, 72(9), 1398–1405.
884<https://doi.org/10.1139/cjfas-2014-0458>

885Fréon, P., & Dagorn, L. (2000). Review of fish associative behaviour: Toward a
886generalisation of the meeting point hypothesis. *Reviews in Fish Biology and Fisheries*, 10,
887183–207. <https://doi.org/10.1023/A:1016666108540>

888Fuller, D. W., Schaefer, K. M., Hampton, J., Caillot, S., Leroy, B. M., & Itano, D. G. (2015).
889Vertical movements, behavior, and habitat of bigeye tuna (*Thunnus obesus*) in the equatorial
890central Pacific Ocean. *Fisheries Research*, 172, 57–70.
891<https://doi.org/10.1016/j.fishres.2015.06.024>

892Gaertner, D., Ariz, J., Bez, N., Clermidy, S., Moreno, G., Murua, H., & Soto, M. (2018).
893Results achieved within the framework of the EU research project: Catch, effort, and
894ECOSystem impacts of FAD-fishing (CECOFAD). *Collect. Vol. Sci. Pap. ICCAT*, 74(5),
8952243–2267.

896Gaertner, D., Pagavino, M., & Marcano, J. (1999). Influence of fishers' behaviour on the
897catchability of surface tuna schools in the Venezuelan purse-seiner fishery in the Caribbean
898Sea. *Canadian Journal of Fisheries and Aquatic Sciences*, 56(3), 394–406.

899Gaertner, J. C., Taquet, M., Dagorn, L., Mérigot, B., Aumeeruddy, R., Sancho, G., & Itano,
900D. (2008). Visual censuses around drifting fish aggregating devices (FADs): A new approach
901for assessing the diversity of fish in open-ocean waters. *Marine Ecology Progress Series*, 366,
902175–186. <https://doi.org/10.3354/meps07554>

903Garcia, A., & Herrera, M. (2018). Assessing the contribution of purse seine fisheries to
904overall levels of bycatch in the Indian Ocean. *IOTC-2018-WPDCS14-26*.
905<https://www.iotc.org/documents/WPDCS/14/26-PSBycatch>

906Gershman, D., Nickson, A., & O'Toole, M. (2015). *Estimating the Use of FADs Around the*
907*World*. PEW Charitable Trusts. <http://pew.org/1XUPV4w>

908 Gilroy, J., & Sutherland, W. (2007). Beyond ecological traps: Perceptual errors and
 909 undervalued resources. *Trends in Ecology & Evolution*, 22(7), 351–356.
 910 <https://doi.org/10.1016/j.tree.2007.03.014>

911 Govinden, R., Capello, M., Forget, F., Filmalter, J. D., & Dagorn, L. (2021). Behavior of
 912 skipjack (*Katsuwonus pelamis*), yellowfin (*Thunnus albacares*), and bigeye (*T. obsesus*) tunas
 913 associated with drifting fish aggregating devices (dFADs) in the Indian Ocean, assessed
 914 through acoustic telemetry. *Fisheries Oceanography*, 30(5), 542–555. [https://doi.org/10.1111/](https://doi.org/10.1111/fog.12536)
 915 [fog.12536](https://doi.org/10.1111/fog.12536)

916 Govinden, R., Dagorn, L., Filmalter, J., & Soria, M. (2010). Behaviour of Tuna associated
 917 with drifting fish aggregating devices (FADs) in the Mozambique Channel. *IOTC*
 918 *Proceedings*. IOTC, Victoria, 22.

919 Govinden, R., Jauhary, R., Filmalter, J., Forget, F., Soria, M., Adam, S., & Dagorn, L. (2013).
 920 Movement behaviour of skipjack (*Katsuwonus pelamis*) and yellowfin (*Thunnus albacares*)
 921 tuna at anchored fish aggregating devices (FADs) in the Maldives, investigated by acoustic
 922 telemetry. *Aquatic Living Resources*, 26(1), Article 1. <https://doi.org/10.1051/alr/2012022>

923 Grande, M. (2013). *The reproductive biology, condition and feeding ecology of the skipjack,*
 924 *Katsuwonus pelamis, in the Western Indian Ocean.* <http://www.fao.org/3/bg397e/bg397e.pdf>

925 Grande, M., Murua, H., Zudaire, I., Arsenault-Pernet, E. J., Pernet, F., & Bodin, N. (2016).
 926 Energy allocation strategy of skipjack tuna *Katsuwonus pelamis* during their reproductive
 927 cycle. *Journal of Fish Biology*, 89(5), 2434–2448. <https://doi.org/10.1111/jfb.13125>

928 Grande, M., Murua, H., Zudaire, I., Goñi, N., & Bodin, N. (2014). Reproductive timing and
 929 reproductive capacity of the Skipjack Tuna (*Katsuwonus pelamis*) in the western Indian
 930 Ocean. *Fisheries Research*, 156, 14–22. <https://doi.org/10.1016/j.fishres.2014.04.011>

931 Grande, M., Ruiz, J., Báez, J. C., Ramos, M. L., Zudaire, I., Santiago, J., Pascual, P., Abascal,
 932 F., Gaertner, D., Cauquil, P., Floch, L., Maufray, A., Muniategi, A., Herrera, M., & Murua,
 933 H. (2018). *Best standards for data collection and reporting requirements on FOBs: Towards*
 934 *a science-based FOB fishery management [IOTC-2018-WPDCS14-39]*. Indian Ocean Tuna
 935 Commission.

936 Greenblatt, P., R. (1979). Associations of tuna with flotsam in the Eastern Tropical Pacific.
 937 *Fishery Bulletin*, 77(1), 147–155.

938 Hall, M. (1992). The association of tunas with floating objects and dolphins in the Eastern
 939 Pacific Ocean. 1992. Part VII. Some hypotheses on the mechanisms governing the association
 940 of tunas with floating objects and dolphins. In *International Workshop on Fishing for Tunas*
 941 *Associated with Floating Objects* (p. 7). Inter-American Tropical Tuna Commission.

942 Hallier, J.-P., & Gaertner, D. (2008). Drifting fish aggregation devices could act as an
 943 ecological trap for tropical tuna species. *Marine Ecology Progress Series*, 353, 255–264.
 944 <https://doi.org/10.3354/meps07180>

945 Hallier, J.-P., & Parajua, J. I. (1999). Review of tuna fisheries on floating objects in the Indian
 946 Ocean. In M. D. Scott, W. H. Bayliff, C. E. Lennert-Cody, & K. M. Schaefer (Eds.),
 947 *Proceedings of the International Workshop on Fishing for Tunas Associated with Floating*
 948 *Objects* (Vol. 11, pp. 195–221). Inter-American Tropical Tuna Commission.

949 Holland, K. N., Brill, R. W., & Chang, R. K. C. (1990b). Horizontal and vertical movements
 950 of yellowfin and bigeye tuna associated with fish aggregation devices. *Fishery Bulletin*, 88(3),
 951 494–507. [https://spo.nmfs.noaa.gov/content/horizontal-and-vertical-movements-yellowfin-](https://spo.nmfs.noaa.gov/content/horizontal-and-vertical-movements-yellowfin-and-bigeye-tuna-associated-fish-aggregating)
 952 [and-bigeye-tuna-associated-fish-aggregating](https://spo.nmfs.noaa.gov/content/horizontal-and-vertical-movements-yellowfin-and-bigeye-tuna-associated-fish-aggregating)

953Hunsicker, M. E., Olson, R. J., Essington, T. E., Maunder, M. N., Duffy, L. M., & Kitchell, J.
954F. (2012). Potential for top-down control on tropical tunas based on size structure of predator-
955prey interactions. *Marine Ecology Progress Series*, 445, 263–277.

956IATTC. (2021). *Resolution on the conservation measures for tropical tunas in the Eastern*
957*Pacific Ocean during 2022-2024* (Resolution C-21–04). Inter-American Tropical Tuna
958Commission. [https://www.iattc.org/GetAttachment/e3dc0a7e-e73c-4b8e-889e-a4cd2cdd7b8b/
959C-21-04-Active_Tuna-conservation-in-the-EPO-2022-2024.pdf](https://www.iattc.org/GetAttachment/e3dc0a7e-e73c-4b8e-889e-a4cd2cdd7b8b/C-21-04-Active_Tuna-conservation-in-the-EPO-2022-2024.pdf)

960ICCAT. (2022). *Recommendation by ICCAT replacing Recommendation 21-01 on a*
961*multiannual conservation and management programme for tropical tunas* (Rec 22-01).
962International Commission for the Conservation of Atlantic Tunas.
963<https://www.iccat.int/Documents/Recs/compendiopdf-e/2022-01-e.pdf>

964Imzilen, T., Lett, C., Chassot, E., & Kaplan, D. M. (2021). Spatial management can
965significantly reduce dFAD beachings in Indian and Atlantic Ocean tropical tuna purse seine
966fisheries. *Biological Conservation*, 254, 108939.
967<https://doi.org/10.1016/j.biocon.2020.108939>

968Imzilen, T., Lett, C., Chassot, E., Maufroy, A., Goujon, M., & Kaplan, D. M. (2022).
969Recovery at sea of abandoned, lost or discarded drifting fish aggregating devices. *Nature*
970*Sustainability*, 1–10. <https://doi.org/10.1038/s41893-022-00883-y>

971IOTC. (2022). *Review of data on Drifting Fish Aggregating Devices* (IOTC Ad Hoc Working
972Group on FADs (WGFAD) IOTC-2022-WGFAD03-03-Rev2). Indian Ocean Tuna
973Commission. <https://iotc.org/meetings/3rd-iotc-ad-hoc-working-group-fads-wgfad03>

974IOTC. (2023). *Resolution 23/02 on the management of drifting fish aggregating devices*
975*(DFAD) in the IOTC area of competence*. [https://iotc.org/documents/conservation-and-
976management-measures-adopted-iotc-its-6th-special-session](https://iotc.org/documents/conservation-and-management-measures-adopted-iotc-its-6th-special-session)

977ISSF. (2014). *Report of the ISSF Workshop on FADs as Ecological Traps* (ISSF Technical
978Report No. 2014–03). [https://www.issf-foundation.org/issf-downloads/download-info/issf-
979technical-report-2014-03-report-of-the-issf-workshop-on-fads-as-ecological-traps/](https://www.issf-foundation.org/issf-downloads/download-info/issf-technical-report-2014-03-report-of-the-issf-workshop-on-fads-as-ecological-traps/)

980Jaquemet, S., Potier, M., & Ménard, F. (2011). Do drifting and anchored Fish Aggregating
981Devices (FADs) similarly influence tuna feeding habits? A case study from the western
982Indian Ocean. *Fisheries Research*, 107(1), 283–290.
983<https://doi.org/10.1016/j.fishres.2010.11.011>

984Justel-Rubio, A., & Recio, L. (2022). *A snapshot of the large-scale tropical tuna purse seine*
985*fishing fleets as of July 2022 (Version 10)* [ISSF Technical Report 2022-14]. International
986Seafood Sustainability Foundation.

987Kim, E. (2015). *Effects of Fish Aggregating Devices (FADs) on tuna movement* [PhD Thesis,
988University of Hawai'i]. <https://www.soest.hawaii.edu/oceanography/PhD/2015-Kim.pdf>

989Lee, M. O., Otake, S., & Kim, J. K. (2018). Transition of artificial reefs (ARs) research and
990its prospects. *Ocean & Coastal Management*, 154, 55–65.
991<https://doi.org/10.1016/j.ocecoaman.2018.01.010>

992Lennert-Cody, C. E., Moreno, G., Restrepo, V., Román, M. H., & Maunder, M. N. (2018).
993Recent purse-seine FAD fishing strategies in the eastern Pacific Ocean: What is the
994appropriate number of FADs at sea? *ICES Journal of Marine Science*, 75(5), 1748–1757.
995<https://doi.org/10.1093/icesjms/fsy046>

996Leroy, B., Phillips, J. S., Nicol, S., Pilling, G. M., Harley, S., Bromhead, D., Hoyle, S.,
997Caillot, S., Allain, V., & Hampton, J. (2013). A critique of the ecosystem impacts of drifting
998and anchored FADs use by purse-seine tuna fisheries in the Western and Central Pacific
999Ocean. *Aquatic Living Resources*, 26(1), 49–61.

- 1000Lopez, J., Moreno, G., Ibaibarriaga, L., & Dagorn, L. (2017). Diel behaviour of tuna and non-
1001tuna species at drifting fish aggregating devices (DFADs) in the Western Indian Ocean,
1002determined by fishers' echo-sounder buoys. *Marine Biology*, 164, 44. [https://doi.org/10.1007/](https://doi.org/10.1007/s00227-017-3075-3)
1003[s00227-017-3075-3](https://doi.org/10.1007/s00227-017-3075-3)
- 1004Lopez, J., Moreno, G., Sancristobal, I., & Murua, J. (2014). Evolution and current state of the
1005technology of echo-sounder buoys used by Spanish tropical tuna purse seiners in the Atlantic,
1006Indian and Pacific Oceans. *Fisheries Research*, 155, 127–137.
1007<https://doi.org/10.1016/j.fishres.2014.02.033>
- 1008Lopez, J., Roman, M. H., Lennert-Cody, C. E., Maunder, M. N., & Vogel, N. (2021).
1009Floating-object fishery indicators: A 2020 report (FAD-05-INF-C). Inter-American Tropical
1010Tuna Commission. [https://www.iattc.org/getattachment/70b8d0d7-1639-42a5-b7d8-](https://www.iattc.org/getattachment/70b8d0d7-1639-42a5-b7d8-1011490f4a12f197/FAD-05a-INF-C_Floating-object-fishery-indicators-a-2020-report.pdf)
1011[490f4a12f197/FAD-05a-INF-C_Floating-object-fishery-indicators-a-2020-report.pdf](https://www.iattc.org/getattachment/70b8d0d7-1639-42a5-b7d8-1011490f4a12f197/FAD-05a-INF-C_Floating-object-fishery-indicators-a-2020-report.pdf)
- 1012Machful, P., Portal, A., Macdonald, J., Allain, V., Phillips, J. S., & Nicol, S. (2021). Tuna
1013stomachs: Is the glass half full, or half empty? *SPC Fisheries Newsletter*, 166, 38–44.
- 1014Malone, M. A., Buck, K. M., Moreno, G., & Sancho, G. (2011). Diet of three large pelagic
1015fishes associated with drifting fish aggregating devices (DFADs) in the western equatorial
1016Indian Ocean. *Animal Biodiversity and Conservation*, 34(2), 287–294.
1017<https://doi.org/10.32800/abc.2011.34.0287>
- 1018Marsac, F., Fonteneau, A., & Ménard, F. (2000). Drifting FADs used in tuna fisheries: An
1019ecological trap? *Pêche Thonière et Dispositifs de Concentration de Poissons*, 28, 537–552.
- 1020Marsac, F., Fonteneau, A., & Michaud, A. (2014). *L'or bleu des Seychelles: Histoire de la*
1021*pêche industrielle au thon dans l'océan Indien*.
1022<https://doi.org/10.4000/BOOKS.IRDEDITIONS.9700>
- 1023Matsumoto, T., Satoh, K., Semba, Y., & Toyonaga, M. (2016). Comparison of the behavior of
1024skipjack (*Katsuwonus pelamis*), yellowfin (*Thunnus albacares*) and bigeye (*T. obesus*) tuna
1025associated with drifting FADs in the equatorial central Pacific Ocean. *Fisheries*
1026*Oceanography*, 25(6), 565–581. <https://doi.org/10.1111/fog.12173>
- 1027Matsumoto, T., Satoh, K., & Toyonaga, M. (2014). Behavior of skipjack tuna (*Katsuwonus*
1028*pelamis*) associated with a drifting FAD monitored with ultrasonic transmitters in the
1029equatorial central Pacific Ocean. *Fisheries Research*, 157, 78–85.
- 1030Maufroy, A., Chassot, E., Joo, R., & Kaplan, D. M. (2015). Large-Scale Examination of
1031Spatio-Temporal Patterns of Drifting Fish Aggregating Devices (dFADs) from Tropical Tuna
1032Fisheries of the Indian and Atlantic Oceans. *PLOS ONE*, 10(5), e0128023.
1033<https://doi.org/10.1371/journal.pone.0128023>
- 1034Maufroy, A., Kaplan, D. M., Bez, N., De Molina, A. D., Murua, H., Floch, L., & Chassot, E.
1035(2017). Massive increase in the use of drifting Fish Aggregating Devices (dFADs) by tropical
1036tuna purse seine fisheries in the Atlantic and Indian oceans. *ICES Journal of Marine Science*,
103774(1), 215–225. <https://doi.org/10.1093/icesjms/fsw175>
- 1038Ménard, F., Stéquert, B., Rubin, A., Herrera, M., & Marchal, É. (2000). Food consumption of
1039tuna in the Equatorial Atlantic ocean: FAD-associated versus unassociated schools. *Aquatic*
1040*Living Resources*, 13(4), 233–240. [https://doi.org/10.1016/S0990-7440\(00\)01066-4](https://doi.org/10.1016/S0990-7440(00)01066-4)
- 1041Mitsunaga, Y., Endo, C., Anraku, K., Selorio, C. M., & Babaran, R. P. (2012). Association of
1042early juvenile yellowfin tuna *Thunnus albacares* with a network of payaos in the Philippines.
1043*Fisheries Science*, 78(1), 15–22. <https://doi.org/10.1007/s12562-011-0431-y>
- 1044Miyake, M., Guillotreau, P., Sun, C., & Ishimura, G. (Eds.). (2010). *Recent developments in*
1045*the tuna industry: Stocks, fisheries, management, processing, trade and markets*. Food and
1046Agriculture Organization of the United Nations.

1047Moreno, G., Boyra, G., Sancristobal, I., Itano, D., & Restrepo, V. (2019). Towards acoustic
1048discrimination of tropical tuna associated with Fish Aggregating Devices. *PLoS ONE*, *14*(6),
1049e0216353. <https://doi.org/10.1371/journal.pone.0216353>

1050Moreno, G., Dagorn, L., Sancho, G., & Itano, D. (2007). Fish behaviour from fishers'
1051knowledge: The case study of tropical tuna around drifting fish aggregating devices (DFADs).
1052*Canadian Journal of Fisheries and Aquatic Sciences*, *64*(11), 1517–1528.
1053<https://doi.org/10.1139/f07-113>

1054Morón, J. (2001). *Report on Management Measures for the European Tuna Purse Seine*
1055*Fleet*. Working Paper for the Standing Committee on Tuna and Billfish (SCTB14).

1056Mullu, D. (2016). A review on the effect of habitat fragmentation on ecosystem. *Journal of*
1057*Natural Sciences Research*, *6*(15), 1–15.

1058Nooteboom, P. D., Scutt Phillips, J., Kehl, C., Nicol, S., & van Sebille, E. (2023). Modelling
1059of tuna around fish aggregating devices: The importance of ocean flow and prey. *Ecological*
1060*Modelling*, *475*, 110188. <https://doi.org/10.1016/j.ecolmodel.2022.110188>

1061Ohta, I., & Kakuma, S. (2005). Periodic behavior and residence time of yellowfin and bigeye
1062tuna associated with fish aggregating devices around Okinawa Islands, as identified with
1063automated listening stations. *Marine Biology*, *146*(3), 581–594.

1064Orue, B., Lopez, J., Pennino, M. G., Moreno, G., Santiago, J., & Murua, H. (2020).
1065Comparing the distribution of tropical tuna associated with drifting fish aggregating devices
1066(DFADs) resulting from catch dependent and independent data. *Deep Sea Research Part II:*
1067*Topical Studies in Oceanography*, 104747.

1068Pérez, G., Dagorn, L., Deneubourg, J.-L., Forget, F., Filmalter, J. D., Holland, K., Itano, D.,
1069Adam, S., Jauharee, R., Beeharry, S. P., & Capello, M. (2020). Effects of habitat
1070modifications on the movement behavior of animals: The case study of Fish Aggregating
1071Devices (FADs) and tropical tunas. *Movement Ecology*, *8*(1), 47.
1072<https://doi.org/10.1186/s40462-020-00230-w>

1073Pérez, G., Dupaix, A., Dagorn, L., Deneubourg, J.-L., Holland, K., Beeharry, S., & Capello,
1074M. (2022). Correlated Random Walk of tuna in arrays of Fish Aggregating Devices: A field-
1075based model from passive acoustic tagging. *Ecological Modelling*, *470*, 110006.
1076<https://doi.org/10.1016/j.ecolmodel.2022.110006>

1077Phillips, J. S., Escalle, L., Pilling, G., Gupta, A. S., & Sebille, E. van. (2019a). Regional
1078connectivity and spatial densities of drifting fish aggregating devices, simulated from fishing
1079events in the Western and Central Pacific Ocean. *Environmental Research Communications*,
1080*1*(5), 055001. <https://doi.org/10.1088/2515-7620/ab21e9>

1081Phillips, J. S., Peatman, T., Escalle, L., Leroy, B., & Smith, N. (2019b). *Electronic tagging*
1082*for the mitigation of bigeye and yellowfin tuna juveniles by purse seine fisheries* (15th Regular
1083Session of the Scientific Commity No. SC15-EB-WP-08). Western and Central Pacific
1084Fisheries Commission. <https://meetings.wcpfc.int/index.php/node/11299>

1085Phillips, J. S., Pilling, G. M., Leroy, B., Evans, K., Usu, T., Lam, C. H., Schaefer, K. M., &
1086Nicol, S. (2017). Revisiting the vulnerability of juvenile bigeye (*Thunnus obesus*) and
1087yellowfin (*T. albacares*) tuna caught by purse-seine fisheries while associating with surface
1088waters and floating objects. *PloS One*, *12*(6), e0179045.

1089Potier, M., Marsac, F., Cherel, Y., Lucas, V., Sabatié, R., Maury, O., & Ménard, F. (2007).
1090Forage fauna in the diet of three large pelagic fishes (lancetfish, swordfish and yellowfin
1091tuna) in the western equatorial Indian Ocean. *Fisheries Research*, *83*(1), 60–72.

1092Potier, M., Marsac, F., Lucas, V., Sabatié, R., Hallier, J.-P., & Ménard, F. (2004). Feeding
1093partitioning among tuna taken in surface and mid-water layers: The case of yellowfin

- 1094(Thunnus albacares) and bigeye (T. obesus) in the western tropical Indian Ocean. *Western*
1095*Indian Ocean Journal of Marine Science*, 3(1), 51–62.
- 1096Proctor, C., Hobart, & Tasmania. (2019). Project final report developing research capacity for
1097management of Indonesia’s pelagic fisheries resources—Final report. *ACIAR Final Reports*,
1098FR2019/01. <https://www.cabdirect.org/cabdirect/abstract/20203534645>
- 1099Ramos, M. L., Delgado de Molina, A., & Ariz, J. (2010). *Analysis of activity data obtained*
1100*from supply vessel’s logbooks implemented by the Spanish fleet and associated in Indian*
1101*Ocean* (Working Party on Tropical Tunas (WPTT), pp. 18–25). Indian Ocean Tuna
1102Commission. [https://iotc.org/documents/analysis-activity-data-obtained-supply-vessels-](https://iotc.org/documents/analysis-activity-data-obtained-supply-vessels-1103logbooks-implemented-spanish-fleet-and)
1103[logbooks-implemented-spanish-fleet-and](https://iotc.org/documents/analysis-activity-data-obtained-supply-vessels-1103logbooks-implemented-spanish-fleet-and)
- 1104Restrepo, V., Dagorn, L., Itano, D., Justel-Rubio, A., Forget, F., & Moreno, G. (2017). A
1105Summary of Bycatch Issues and ISSF Mitigation Activities to date in purse seine fisheries,
1106with emphasis on FADs. *ISSF Technical Reports*, 41.
- 1107Robert, M., Dagorn, L., Bodin, N., Pernet, F., Arsenault-Pernet, E.-J., & Deneubourg, J.-L.
1108(2014). Comparison of condition factors of skipjack tuna (*Katsuwonus pelamis*) associated or
1109not with floating objects in an area known to be naturally enriched with logs. *Canadian*
1110*Journal of Fisheries and Aquatic Sciences*. <https://doi.org/10.1139/cjfas-2013-0389>
- 1111Robert, M., Dagorn, L., Deneubourg, J. L., Itano, D., & Holland, K. (2012). Size-dependent
1112behavior of tuna in an array of fish aggregating devices (FADs). *Marine Biology*, 159(4),
1113907–914.
- 1114Robert, M., Dagorn, L., Filmalter, J. D., Deneubourg, J.-L., Itano, D., & Holland, K. (2013).
1115Intra-individual behavioral variability displayed by tuna at fish aggregating devices (FADs).
1116*Marine Ecology Progress Series*, 484, 239–247.
- 1117Robert, M., Dagorn, L., Lopez, J., Moreno, G., & Deneubourg, J.-L. (2013). Does social
1118behavior influence the dynamics of aggregations formed by tropical tunas around floating
1119objects? An experimental approach. *Journal of Experimental Marine Biology and Ecology*,
1120440, 238–243. <https://doi.org/10.1016/j.jembe.2013.01.005>
- 1121Robertson, B. A., & Hutto, R. L. (2006). A framework for understanding ecological traps and
1122an evaluation of existing evidence. *Ecology*, 87(5), 1075–1085.
- 1123Rodriguez-Tress, P., Capello, M., Forget, F., Soria, M., Beeharry, S. P., Dussoo, N., &
1124Dagorn, L. (2017). Associative behavior of yellowfin *Thunnus albacares*, skipjack
1125*Katsuwonus pelamis*, and bigeye tuna *T. obesus* at anchored fish aggregating devices (FADs)
1126off the coast of Mauritius. *Marine Ecology Progress Series*, 570, 213–222.
- 1127Sardenne, F., Chassot, E., Fouché, E., Ménard, F., Lucas, V., & Bodin, N. (2016). Are
1128condition factors powerful proxies of energy content in wild tropical tunas? *Ecological*
1129*Indicators*, 71, 467–476. <https://doi.org/10.1016/j.ecolind.2016.06.031>
- 1130Schaefer, K. M., Fuller, D., Hampton, J., Caillot, S., Leroy, B., & Itano, D. (2015).
1131Movements, dispersion, and mixing of bigeye tuna (*Thunnus obesus*) tagged and released in
1132the equatorial Central Pacific Ocean, with conventional and archival tags. *Fisheries Research*,
1133161, 336–355.
- 1134Schaefer, K. M., & Fuller, D. W. (2002). Movements, behavior, and habitat selection of
1135bigeye tuna (*Thunnus obesus*) in the eastern equatorial Pacific, ascertained through archival
1136tags. *Fishery Bulletin*, 100(4), 765–788.
- 1137Schaefer, K. M., & Fuller, D. W. (2010). Vertical movements, behavior, and habitat of bigeye
1138tuna (*Thunnus obesus*) in the equatorial eastern Pacific Ocean, ascertained from archival tag
1139data. *Marine Biology*, 157(12), 2625–2642. <https://doi.org/10.1007/s00227-010-1524-3>

- 1140Schaefer, K. M., & Fuller, D. W. (2013). Simultaneous behavior of skipjack (*Katsuwonus*
1141*pelamis*), bigeye (*Thunnus obsesus*), and yellowfin (*T. albacares*) tunas, within large multi-
1142species aggregations associated with drifting fish aggregating devices (FADs) in the
1143equatorial eastern Pacific Ocean. *Marine Biology*, 160(11), 3005–3014.
1144<https://doi.org/10.1007/s00227-013-2290-9>
- 1145Schaefer, K. M., Fuller, D. W., & Block, B. A. (2009). Vertical movements and habitat
1146utilization of skipjack (*Katsuwonus pelamis*), yellowfin (*Thunnus albacares*), and bigeye
1147(*Thunnus obesus*) tunas in the equatorial eastern Pacific Ocean, ascertained through archival
1148tag data. In *Tagging and tracking of marine animals with electronic devices* (pp. 121–144).
1149Springer.
- 1150Schlaepfer, M. A., Runge, M. C., & Sherman, P. W. (2002). Ecological and evolutionary
1151traps. *Trends in Ecology & Evolution*, 17(10), 474–480. [https://doi.org/10.1016/S0169-11525347\(02\)02580-6](https://doi.org/10.1016/S0169-11525347(02)02580-6)
- 1153Scott, G. P., & Lopez, J. (2014). *The Use of Fads in Tuna Fisheries*. Directorate General for
1154Internal Policies, European Parliament, Committee in Fisheries.
1155[https://www.europarl.europa.eu/thinktank/en/document.html?reference=IPOL-1156PECH_NT\(2014\)514002](https://www.europarl.europa.eu/thinktank/en/document.html?reference=IPOL-1156PECH_NT(2014)514002)
- 1157Scott, M. D., Bayliff, W. H., Lennert-Cody, C. E., & Schaefer, K. M. (1999). Proceedings of
1158the international workshop on the ecology and fisheries for tunas associated with floating
1159objects. *Inter-American Tropical Tuna Commission Special Report*, 11, 478.
1160<http://hdl.handle.net/1834/23201>
- 1161Sempo, G., Dagorn, L., Robert, M., & Deneubourg, J.-L. (2013). Impact of increasing
1162deployment of artificial floating objects on the spatial distribution of social fish species.
1163*Journal of Applied Ecology*, 50(5), 1081–1092. <https://doi.org/10.1111/1365-2664.12140>
- 1164Sherley, R. B., Ludynia, K., Dyer, B. M., Lamont, T., Makhado, A. B., Roux, J.-P., Scales, K.
1165L., Underhill, L. G., & Votier, S. C. (2017). Metapopulation Tracking Juvenile Penguins
1166Reveals an Ecosystem-wide Ecological Trap. *Current Biology*, 27(4), 563–568.
1167<https://doi.org/10.1016/j.cub.2016.12.054>
- 1168Sobradillo, B., Boyra, G., Uranga, J., & Moreno, G. (2023). *TS measurments of ex-situ*
1169*yellowfin tuna (Thunnus albacares) and frequency-response discrimination for tropical tuna*
1170*species* [7th Meeting of the Ad Hoc Working Group on FADs]. Inter-American Tropical Tuna
1171Commission. https://www.iattc.org/GetAttachment/01c4217f-4f28-4c34-b6e6-e5bb7efd777e/1172FAD-07-MISC_Acoustic-discrimination-of-juvenile-yellowfin-tuna-in-an-offshore-cage.pdf
- 1173Song, L., & Shen, H. (2022). An integrated scheme for the management of drifting fish
1174aggregating devices in tuna purse seine fisheries. *Fisheries Management and Ecology*, 30(1),
117556–69. <https://doi.org/10.1111/fme.12600>
- 1176Stehfest, K. M., & Dagorn, L. (2010). *Differences in large-scale movement between free*
1177*swimming and fish aggregating device (FAD) caught tuna* (Working Party on Tropical Tuna
1178IOTC-2010-WPTT-06; p. 24). Indian Ocean Tuna Commission.
1179<https://iotc.org/meetings/12th-session-working-party-tropical-tunas>
- 1180Stéquert, B., & Marsac, F. (1986). *La pêche de surface des thonidés tropicaux dans l’Océan*
1181*Indien*. Food & Agriculture Org.
- 1182Swearer, S. E., Morris, R. L., Barrett, L. T., Sievers, M., Dempster, T., & Hale, R. (2021). An
1183overview of ecological traps in marine ecosystems. *Frontiers in Ecology and the*
1184*Environment*, 19(4), 234–242. <https://doi.org/10.1002/fee.2322>
- 1185Taquet, M. (2013). Fish aggregating devices (FADs): Good or bad fishing tools? A question
1186of scale and knowledge. *Aquatic Living Resources*, 26(1), 25–35.
1187<https://doi.org/10.1051/alr/2013043>

- 1188 Teske, P. R., Emami-Khoyi, A., Golla, T. R., Sandoval-Castillo, J., Lamont, T., Chiazzari, B.,
1189 McQuaid, C. D., Beheregaray, L. B., & van der Lingen, C. D. (2021). The sardine run in
1190 southeastern Africa is a mass migration into an ecological trap. *Science Advances*, 7(38),
1191 eabf4514. <https://doi.org/10.1126/sciadv.abf4514>
- 1192 Thiel, M., & Gutow, L. (2005). The ecology of rafting in the marine environment. I. The
1193 floating substrata. *Oceanography and Marine Biology: An Annual Review*, 42, 181–264.
- 1194 Tolotti, M. T., Forget, F., Capello, M., Filmalter, J. D., Hutchinson, M., Itano, D., Holland,
1195 K., & Dagorn, L. (2020). Association dynamics of tuna and purse seine bycatch species with
1196 drifting fish aggregating devices (FADs) in the tropical eastern Atlantic Ocean. *Fisheries*
1197 *Research*, 226, 105521. <https://doi.org/10.1016/j.fishres.2020.105521>
- 1198 Wang, X., Chen, Y., Truesdell, S., Xu, L., Cao, J., & Guan, W. (2014). The large-scale
1199 deployment of fish aggregation devices alters environmentally-based migratory behavior of
1200 skipjack tuna in the Western Pacific Ocean. *PLoS One*, 9(5), e98226.
- 1201 Wang, X., Xu, L., Chen, Y., Zhu, G., Tian, S., & Zhu, J. (2012). Impacts of fish aggregation
1202 devices on size structures of skipjack tuna *Katsuwonus pelamis*. *Aquatic Ecology*, 46(3), 343–
1203 352. <https://doi.org/10.1007/s10452-012-9405-0>
- 1204 Watanabe, Y., Tsunekawa, T., Takahashi, M., Tabuchi, M., & Sugawara, T. (1988). Results
1205 on the experimental purse seine fishing with FADs in the Indian ocean by R/V Nippon Maru.
1206 *IPTP Collect. Vol. Work. Doc.*, 3, 227–232.
- 1207 WCPFC. (2021). *Conservation and Management Measure for bigeye, yellowfin and skipjack*
1208 *tuna in the Western and Central Pacific Ocean* (CMM-2021–01). Western and Central Pacific
1209 Fisheries Commission. [https://www.wcpfc.int/doc/cmm-2018-01/conservation-and-](https://www.wcpfc.int/doc/cmm-2018-01/conservation-and-management-measure-bigeye-yellowfin-and-skipjack-tuna-western-and)
1210 [management-measure-bigeye-yellowfin-and-skipjack-tuna-western-and](https://www.wcpfc.int/doc/cmm-2018-01/conservation-and-management-measure-bigeye-yellowfin-and-skipjack-tuna-western-and)
- 1211 Weng, J.-S., Hung, M.-K., Lai, C.-C., Wu, L.-J., Lee, M.-A., & Liu, K.-M. (2013). Fine-scale
1212 vertical and horizontal movements of juvenile yellowfin tuna (*Thunnus albacares*) associated
1213 with a subsurface fish aggregating device (FAD) off southwestern Taiwan. *Journal of Applied*
1214 *Ichthyology*, 29(5), 990–1000. <https://doi.org/10.1111/jai.12265>
- 1215 Zudaire, I., Murua, H., Grande, M., Goñi, N., Potier, M., Ménard, F., Chassot, E., & Bodin,
1216 N. (2015). Variations in the diet and stable isotope ratios during the ovarian development of
1217 female yellowfin tuna (*Thunnus albacares*) in the Western Indian Ocean. *Marine Biology*,
1218 162(12), 2363–2377. <https://doi.org/10.1007/s00227-015-2763-0>
- 1219 Zudaire, I., Murua, H., Grande, M., Pernet, F., & Bodin, N. (2014). Accumulation and
1220 mobilization of lipids in relation to reproduction of yellowfin tuna (*Thunnus albacares*) in the
1221 Western Indian Ocean. *Fisheries Research*, 160, 50–59.

1222**Tables**

1223**Table 1: Summary of main findings from previous studies on the numbers of**
 1224**monitored floating objects or the number of DFADs used in large-scale tropical tuna**
 1225**purse seine fisheries.**

Area	Period	Indicator	Associated number of vessels	Estimation	Reference
All oceans	2006-2011	DFADs deployed yearly		47,000-103,000	Baske et al. (2012)
	2010s	DFADs deployed yearly		91,000	Scott & Lopez (2014)
	2013	DFADs deployed yearly		81,000-121,000	Gershman et al. (2015)
Atlantic Ocean	1998	Radio buoys	45 vessels	3,000	Ménard et al. (2000)
	2004-2014	Buoys deployed yearly	Per vessel (French PS fleet)	From 41 (2004) to 200 (2014)	Fonteneau et al. (2015)
	2007-2013	Monthly active buoys		From 1,289 (2007) to 8,856 (2013)	Maufroy et al. (2017)
Indian Ocean	2003-2005	Daily active buoys	45 vessels	2,100	Moreno et al. (2007)
	2007-2013	Monthly		From 2,679	Maufroy et al.

	active buoys	(2007)	to (2017)	10,929 (2013)	
2010-2012	Daily active buoys	34 vessels	3,750-7,500		Filmalter et al. (2013)
2010-2014	Quarterly active buoys	25 vessels	1,200		Chassot et al. (2014)
2013	Quarterly active buoys	19 vessels	6,015		Delgado de Molina et al. (2014)
2013	DFADs deployed yearly	19 vessels	12,813		Delgado de Molina et al. (2014)
2016-2021	DFADs deployed yearly	Whole ocean	10,514	to	IOTC (2022)
			24,550		
Western and Central Pacific Ocean	2011-2019 Daily active buoys	Per vessel	45-75		Escalle et al. (2021)
2011-2019	DFADs deployed yearly	268 to 322 vessels	20,000-40,000		Escalle et al. (2020, 2021a)
		(whole WCPO)			
2016-2019	Buoys deployed yearly	Whole WCPO	31,000	to	Escalle et al. (2020)
			39,500		
2016-2020	Buoys deployed	187 to 235 vessels	16,000	to	Escalle et al. (2021b)
			22,000		

		yearly						
Eastern Pacific Ocean	2018-2020	Daily active buoys	100 to 140	8,000-11,000			Lopez et al. (2021)	
	2015-2020	DFADs deployed yearly	100 to 140	20,000 to 40,000			Lopez et al. (2021)	

1226**Table 2: Summary of main findings from previous studies on tuna individual CRT and CAT assessed under anchored and drifting FADs.**

1227CRT: Continuous Residence Time – continuous bouts of time spent at the same FAD without any absence longer than 24h. CAT: Continuous Absence

1228Time – the time between two associations with a FAD. FL: fork length, YFT: *Thunnus albacares*, SKJ: *Katsuwonus pelamis*, BET: *Thunnus obesus*).

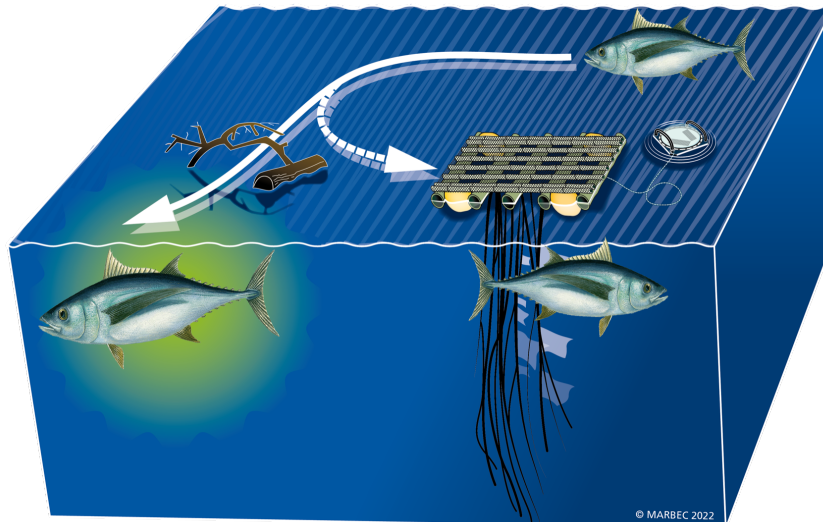
1229

FAD type	Study location	Metric	Findings	Reference
	Eastern Atlantic Ocean	CRT	YFT (34-82 cm FL): average of 19.15 days (maximum value of 55 days) SKJ (39-61 cm FL): average of 9.19 days (maximum value of 15 days) BET (45-61 cm FL): average of 25.31 days (maximum value of 55 days)	Tolotti et al. (2020)
	Mozambique Chanel (Western Indian Ocean)	CRT	YFT (29-60 cm FL): between 0.00-26.72 days with median at 9.98 days SKJ (47-57 cm FL): between 0.09-18.33 days with median at 4.47days BET (54-56 cm FL): between 0.00-6.56 days with median at 3.89 days	Govinden et al. (2010)
Drifting	Western and Central Pacific Ocean	CRT	SKJ (46-60 cm FL): median of 1 day (maximum value of 18 days) YFT (36-98 cm FL): median of 2 days (maximum value of 50 days) BET (38-90 cm FL): median of 10 days (maximum value of 30 days)	Phillips et al. (2019b)
		CAT	BET (38-90 cm FL): median of 3.2 days (maximum value of 48.2 days)	
	Equatorial Central Pacific Ocean	CRT	SKJ (36-65 cm FL) : from 0.0 to 6.4 days (with average value at 2.3 days)	Matsumoto et al. (2014)
CRT		SKJ (34.5–65.0 cm FL): less than 7 days YFT (31.6–93.5 cm FL): less than 7 days BET (33.5–85.5 cm FL): less than 7 days	Matsumoto et al. (2016)	
Anchored	Philippines (Indian Ocean)	CRT	Juvenile YFT (19–31 cm FL) : between 1 and 6 days	Mitsunaga et al., (2012)
	Maldives Islands (Indian Ocean)	CRT	SKJ (37–54 cm FL) : 0.20-3.75 days YFT (35–53 cm FL) : 0.61-0.70 days	Govinden et al. (2013)
	Mauritius islands (Indian Ocean)	CRT	SKJ (41 -59 cm FL) : 2.5 days YFT (46 -81cm FL) : 9.6 days BET (48 - 60 cm FL) : 5.2 days	Rodriguez-Tress et al. (2017)
		CAT	SKJ (41 -59 cm FL) : 2.9 days YFT (46 -81cm FL) : 1.4 days BET (48 - 60 cm FL) : 0.8 days	
Hawaii islands	CRT	Small YFT (30-39 cm FL) : 13.58 days	Robert et al. (2012)	

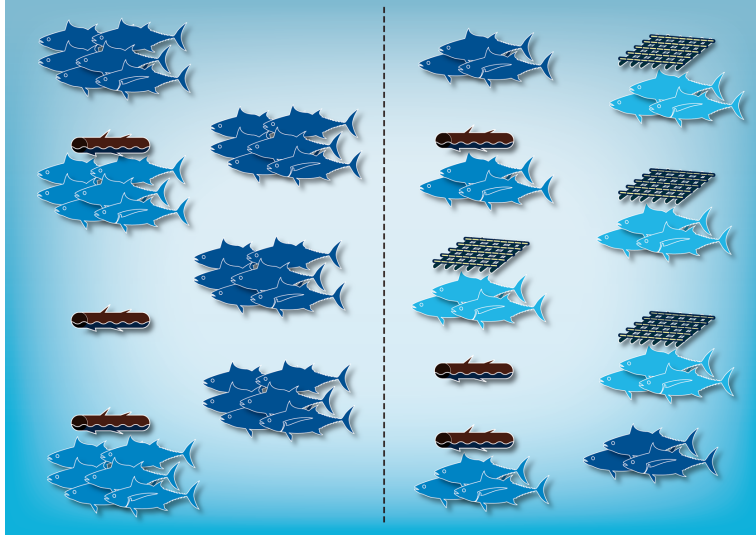
		Large YFT (63-68 cm FL): 9.44 days	
	CAT	4 days for small YFT and 1.65 days for large YFT	
(Pacific Ocean)	CRT	4 behavioural modes reported for YFT (54 to 95 cm FL) : - Brief association : 13.1 minutes - Short association: 2.9 days - Two long association modes : 13.8 and 23.2 days	Robert et al. (2013a)
	CAT	2 behavioural modes: - Short: 2.8 days - Long: infinite	
South-western Taiwan (Pacific Ocean)	CRT	YFT (35–81 cm FL) : average of 2.1 days (maximum value to 31 days)	Weng et al. (2013)
Okinawa Island (Pacific Ocean)	CRT	YFT (40-119 cm FL): median of 7.9 days (maximum value to 55 days) BET (50-77 cm FL): median of 7.0 days (maximum value to 34 days)	Ohta & Kakuma (2005)
Palau Islands (Pacific Ocean)	CRT	YFT (50-60cm FL): mean of 16 days (maximum value to 123 days) YFT (60-100cm FL): mean of 2 days (maximum value to 33 days)	Filous et al., (2020)

1230
1231
1232
1233
1234

1235 Figures



1237 **Figure 1: Schematic representation of the ecological trap hypothesis applied to**
1238 **Drifting Fish Aggregating Devices (DFADs), as originally formulated.** Under this
1239 hypothesis, before DFADs introduction, when only natural floating objects (NLOGs) were
1240 present, floating objects were indicators of productive areas. Hence, by associating with
1241 floating objects, tuna selected high quality habitats. DFAD massive deployment modified
1242 the distribution of floating objects (FOBs), which are not representative of rich areas
1243 anymore. By associating with FOBs, tunas can be attracted to or retained in habitats of
1244 lesser quality.



1246 **Figure 2: Schematic representation of potential effects of Drifting Fish Aggregating Devices**
 1247 **(DFADs) on tuna schooling behaviour. The left side represents an ocean with natural**
 1248 **floating objects (NLOGs) only (no DFAD), while the right side represents an ocean with**
 1249 **both NLOGs and DFADs, i.e. more floating objects (FOBs).** Dark blue represents tuna in free-
 1250 swimming schools, intermediate blue tuna associated with NLOGs and light blue tuna associated
 1251 with DFADs. An increase in FOB density (right panel) could lead both to (i) more tuna associated
 1252 to FOBs and less free-swimming schools, (ii) more numerous but smaller FOB-associated
 1253 schools.