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Contamination of planktonic food webs in the Mediterranean Sea: Setting the frame for the MERITE-HIPPOCAMPE oceanographic cruise (spring 2019)

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- 48 the Mediterranean Sea: Biological pump and interactions from regional to global
- 49 *approaches*"
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51 Abstract

This paper looks at experiential feedback and the technical and scientific challenges tied to the MERITE-HIPPOCAMPE cruise that took place in the Mediterranean Sea in spring 2019. This cruise proposes an innovative approach to investigate the accumulation and transfer of inorganic and organic contaminants within the planktonic food webs. We present detailed information on how the cruise worked, including 1) the cruise track and sampling stations, 2) the overall strategy, based mainly on the collection of plankton, suspended particles and water at the deep chlorophyll maximum, and the separation of these particles and planktonic organisms into various size fractions, as well as the collection of atmospheric deposition, 3) the operations performed and material used at each station, and 4) the sequence of operations and main parameters analysed. The paper also provides the main environmental conditions that were prevailing during the campaign. Lastly, we present the types of articles produced based on work completed by the cruise that are part of this special issue.

Key words: contaminants, plankton, Mediterranean Sea, size fractions, bioaccumulation,
atmospheric deposition

1. Introduction

78	Plankton is recognised as a key gateway of inorganic and organic contaminants into the
79	marine food web (Berroralbiz et al., 2011; Tao et al., 2018; Chouvelon et al., 2019; Li et al.,
80	2021). Phytoplankton exposure to contaminants is mainly via water. Phytoplankton cells have
81	a high surface area-to-volume ratio and thus large areas for exchanges, and consequently
82	display high capacities for adsorbing and absorbing and thus bioconcentrating dissolved
83	contaminants (Martin and Knauer, 1973; Fan and Reinfelder, 2003; Heimbürger et al., 2010;
84	Lee and Fisher, 2016; Chouvelon et al., 2019). Contaminant bioconcentration in
85	phytoplankton, thought to be governed mainly by partition equilibrium processes between the
86	cells and the surrounding water (Frouin et al., 2013) even though cell growth conditions may
87	prevent contaminants from reaching thermodynamic equilibrium (Swackhamer and Skoglund,
88	1993), tends to increase with decreasing size of algal cells (Fan and Reinfelder, 2003).
89	Bioaccumulation processes in zooplankton are highly complex due to 1) entry of
90	contaminants by both the water aqueous phase (bioconcentration) and diet, 2) trophic
91	interactions and/or transfers between phytoplankton and herbivorous, carnivorous and
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92	omnivorous zooplankton, and 3) the contaminant elimination processes used by these
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92 93 94 95 96 97	omnivorous zooplankton, and 3) the contaminant elimination processes used by these organisms, including metabolization, excretion and passive release (Tiano et al., 2014; Alekseenko et al., 2018; Tao et al., 2018; Thomas et al., 2018; Li et al., 2020). The fact that all these processes can act simultaneously in the same or opposite directions makes it difficult to understand the variability of contaminant concentrations within the planktonic food web, which may ultimately display biomagnification (increasing contaminant concentrations with
92 93 94 95 96 97 98	omnivorous zooplankton, and 3) the contaminant elimination processes used by these organisms, including metabolization, excretion and passive release (Tiano et al., 2014; Alekseenko et al., 2018; Tao et al., 2018; Thomas et al., 2018; Li et al., 2020). The fact that all these processes can act simultaneously in the same or opposite directions makes it difficult to understand the variability of contaminant concentrations within the planktonic food web, which may ultimately display biomagnification (increasing contaminant concentrations with trophic level), bioreduction (decreasing contaminant concentrations with trophic level) or no

Alekseenko et al., 2018; Chouvelon et al., 2019; Tang et al., 2020; Castro-Jiménez et al.,
2021; Li et al., 2021).

One important biogeochemical implication of contaminant uptake by plankton is the 102 103 role as a "biological pump" for contaminant sequestration (Dachs et al., 2002; Galbán-Malagón et al., 2012; Duran and Cravo-Laureau, 2016; González-Gaya et al., 2019; Tang et 104 105 al., 2020). Contaminants bioconcentrated/bioaccumulated within planktonic organisms 106 subsequently get transferred to higher trophic levels or to deep waters and sediment through sinking particles. During phytoplankton blooms, contaminant uptake by plankton depletes the 107 contaminant loads in the dissolved phase of the water column and increases contaminant air-108 109 water fluxes, sinking particle fluxes and sequestration in sediments (Berrojalbiz et al., 2011; Nizzetto et al., 2012; Everaert et al., 2015; Morales et al., 2015; Casal et al., 2017; Ding et al., 110 111 2021). The phosphorus-limited Mediterranean Sea is globally classified as an oligotrophic 112 marine area (D'Ortenzio and d'Alcalà, 2009; The Mermex Group, 2011; Marañón et al., 113 114 2021) dominated by small phytoplankton (i.e., pico- and nano-sized fractions) (Uitz et al., 2006; Hunt et al., 2017; Mayot et al., 2017; Leblanc et al., 2018; Salhi et al., 2018; Ramirez-115 Romero et al., 2020), even though the occurrence of regional phytoplankton blooms (bloom 116 117 bioregions, ecoregions; D'Ortenzio and Ribera d'Alcalà, 2009; Berline et al., 2014; Reygondeau et al., 2017; Ayata et al., 2018) leads to periods of coexistence of numerous 118 microalgal groups (Siokou-Frangou et al., 2010; El Hourany et al., 2019). 119 Another feature of the Mediterranean Sea is its high exposure to chemical 120 contamination (Hinrichsen, 1990; The Mermex Group, 2011; UNEP, 2012). Indeed, the 121 intense human activities in its 23 bordering countries induce significant inputs of various 122 chemical contaminants, while its semi-closed geography limits possibilities for diluting them. 123

124 Contaminants are brought to the Mediterranean mainly by major and smaller river systems

- (Elbaz-Poulichet et al., 2001; Radakovitch et al., 2008; Sicre et al., 2008; Guigue et al., 2014;
- 126 Köck-Schulmeyer et al., 2021) but also *via* effluents, runoffs, groundwater, and maritime
- 127 activities (Tedetti et al., 2010; Oursel et al., 2013; Tornero and Hanke, 2016; Fourati et al.,
- 128 2018; Jacquet et al., 2021; Llamas-Dios et al., 2021) as well as atmospheric deposition
- 129 (Lipiatou and Albaigés, 1994; Heimbürger et al., 2011; Castro-Jiménez et al., 2017; Barhoumi
- et al., 2018; Desboeufs et al., 2022). Studies have demonstrated that atmospheric deposition is
- the major source of contaminants in remote/open sea areas (Dachs and Méjanelle, 2010;

132 González-Gaya et al., 2019; Jiskra et al., 2021; Cossa et al., 2022).

An interesting feature of the bioaccumulation of contaminants in the Mediterranean Sea 133 134 is that the recorded contamination levels are significantly higher in its predatory species (crustaceans, sharks, teleost fish) than in congeneric species of the Atlantic Ocean (Cossa and 135 Coquery, 2005; Bodiguel et al., 2009; Chouvelon et al., 2018). This difference may be related 136 (but not solely) to the enhanced ability of the Mediterranean planktonic food webs to 137 bioaccumulate certain contaminants, such as mercury (Hg) (Cossa and Coquery, 2005; 138 Harmelin-Vivien et al., 2009; Chouvelon et al., 2018, 2019; Cossa et al., 2022), which further 139 underscores the potential key role of the planktonic compartment in the transfer and 140 accumulation of contaminants in the Mediterranean Sea. Furthermore, contaminant-plankton 141 142 interactions in the Mediterranean Sea are expected to evolve in the coming years, as it has been identified as a hotspot for climate change due to its high reactivity to external forcing, 143 particularly variations in water, energy and matter fluxes that will affect its circulation, 144 biogeochemical fluxes and ecosystem functioning (Lejeusne et al., 2010; The Mermex Group, 145 2011; Ser-Giacomi et al., 2020). 146

However, there are still key gaps in our understanding around the actual ability of
plankton to accumulate and transfer contaminants (i.e., the role of plankton as a biological
pump of contaminants), especially its small size fractions, i.e., pico- and nano-plankton. This

lack of knowledge partly comes from methodological difficulties in: 1) collecting and
separating plankton into its various size fractions from pico- to macro-plankton, separating
bacterio-, phyto- from zoo-plankton, and separating non-living suspended particulate matter
from plankton, 2) obtaining sufficient material in each of these fractions to perform tracelevel chemical analyses, and 3) clearly identifying the trophic relationships between
planktonic size fractions in the presence of detritus (particularly in smaller size fractions) and
mixing of different trophic levels within each size fraction.

In this context, the MERITE-HIPPOCAMPE cruise aimed to evaluate the accumulation 157 and transfer of a hugely diverse range of inorganic and organic contaminants at the 158 159 atmosphere-water-plankton interfaces and within the planktonic food webs, i.e., phyto-, zooand bacterio-plankton, along a North-South transect in the Mediterranean Sea. This ultimately 160 161 to quantify the role of Mediterranean plankton as a biological pump of contaminants, focusing 162 on contaminant transfers into the planktonic food webs. The main collection area for plankton, suspended particles and water is the deep chlorophyll maximum (DCM), which may 163 be considered as the layer with the highest biomasses of plankton, mainly phytoplankton, but 164 also zooplankton. The plankton size fractions investigated include pico- (0.2-2 µm), nano-165 (2–20 µm), micro- (20–200 µm) and meso-plankton (200–2000 µm). The contaminants 166 167 examined are trace metals, organometals and metalloids (As, Cd, Cr, Cu, Fe, Hg, MeHg, Mn, Ni, Pb, Sb, Zn...), organic contaminants (PAHs, PCBs, PBDEs, PFASs), radionuclides 168 (^{137}Cs) , and microplastics. The specific objectives of the cruise are: 1) to determine the 169 170 contaminant concentration levels in various planktonic compartments (phyto-, zoo- and bacterio-plankton), suspended particles and water (dissolved phase), as well as atmosphere 171 and, 2) to assess the role of dry/wet atmospheric deposition as a source of contaminants for 172 marine waters, especially in offshore areas, 3) to gauge how contaminant accumulations and 173 transfers are influenced by plankton in terms of size and community structures, trophic 174

1/2	interactions and biochemical content, and 4) to establish the link between habitat
176	characteristics and concentration levels of contaminants in plankton. The questions addressed
177	by the MERITE-HIPPOCAMPE cruise are summarised in Fig. 1. In this paper, we first
178	present the implementation of the cruise project in terms of strategy, study stations, material
179	used, operations performed, sequence of operations, and main parameters analysed. We then
180	work up from a joint dataset to provide the main meteorological, hydrological, and
181	biogeochemical conditions prevailing during the cruise. Lastly, we introduce the types of
182	articles produced through the MERITE-HIPPOCAMPE cruise that are part of this special
183	issue.
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186	2. Implementation of the cruise
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188	2.1 Research strategy
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4) Collect large amounts of plankton, suspended particles, and water (dissolved phase)
in the DCM and, in a lesser extent, surface/subsurface waters (0–5 m depth) (Fig. 3). The
choice of focusing our sampling on the DCM was motivated by the need to 1) collect a high
biomass of plankton from the same location to determine the concentration levels of a wide
range of contaminants, and over a broad spectrum of planktonic size fractions, and 2) be
located in a relevant area for pelagic trophic interactions in both the coastal and marine areas.
5) Separate these large amounts of plankton and particles into various size fractions

207 (from 0.2 μ m to > 2000 μ m) by filtration or sieving (Fig. 3).

6) Share the size fractioned samples for analysis of a battery of parameters, including (i) 208 209 contaminants, with trace metals, organometals and metalloids (As, Cd, Cr, Cu, Fe, Hg, MeHg, Mn, Ni, Pb, Sb, Zn, etc.), organic contaminants (PAHs, PCBs, PBDEs, PFASs), radionuclides 210 (¹³⁷Cs) and microplastics, (ii) biomass, size structure, taxonomy, cytometry, diversity and 211 pigment composition of plankton, and (iii) biogeochemical parameters, with nutrients, total 212 chlorophyll a (TChla), suspended particulate matter (SPM), dissolved and particulate organic 213 matter, C and N isotopic ratios (δ^{13} C, δ^{15} N), and biochemical compounds (carbohydrates, 214 proteins, lipids) (Fig. 3). 215

7) Combine several methods (taxonomy, optical measurements, chemical analyses) to
tentatively distinguish the contributions of living (plankton) and non-living (detrital) materials
to contaminant accumulation and transfers.

8) Collect on-board samples of atmospheric wet deposition (rain) to assess their
potential importance as a source of contaminants for marine water and plankton. The
collection of atmospheric samples was also done in parallel on the coast by setting up two
ground stations, located in the northern part (Marseille, France) and southern part (Sfax,
Tunisia) of the Mediterranean basin (Fig. 2a), in which atmospheric total (dry and wet)

deposition were measured on a regular (bi-monthly or monthly) basis from March 2019 to
June 2020, i.e., before, during, and after the oceanographic cruise.

9) Sample small pelagic plankton-feeding fishes in Tunisian waters (see sampling
positions in Fig. 2a). The fish samples were analysed for a number of contaminants in order to
set bioaccumulation factors for higher trophic levels.

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2.2. Cruise track and study stations

The MERITE-HIPPOCAMPE cruise took place in spring 2019, from April 13 to May 231 14, on board the French Research Vessel (R/V) Antea along a North-South transect (round 232 233 trip) in the Mediterranean Sea, from the French coast (La Seyne-sur-Mer, Northwestern Mediterranean) to the Gulf of Gabès in Tunisia (Southeastern Mediterranean) (Tedetti and 234 Tronczynski, 2019) (Fig. 2a, b). Leg 1 (13–28 April 2019) ran the southward transect, 235 236 between La Seyne-sur-Mer and Tunis, with sampling at five stations: St2, St4, St3, S10, and St11 (in this chronological order). Leg 2 (30 April–14 May 2019) covered first the end of the 237 238 southward transect (from Tunis to the Gulf of Gabès), and then the return trip back northward from the Gulf of Gabès to La Seyne-sur-Mer. Five stations were sampled during leg 2: St15, 239 St17, St19, St9 and St1 (in this chronological order). Five stations (St1-St4, St9) were 240 situated in the French area, two stations (St10, St11) in the Italian area, and three stations 241 (St15, St17, St19) in the Tunisian area (Fig. 2a, b; Table 1). 242 The ten stations were chosen according to different criteria based on physical, 243 244 biogeochemical and biological conditions and anthropogenic pressures (Fig. 2a; Fig. S1; Table 1). St1 and St4 were located in the urbanised bays opposite the cities of Toulon and 245 Marseille, respectively, and thus submitted to strong anthropogenic pressures. St4 was the 246 SOLEMIO station, which is part of the French national coastal observation monitoring 247

248 network SOMLIT (<u>http://somlit.epoc.u-bordeaux1.fr/fr/</u>). St1 and St4 were "intermittent

bloom" areas or "bloom" areas according to D'Ortenzio and d'Alcalà (2009) based on 249 250 SeaWiFS satellite observations of surface TChla concentration. St2 and St3 were situated offshore of Toulon and Marseille, respectively. St2 was at the limit of the continental shelf 251 252 and the boundary of the Ligurian consensus region (Ayata et al., 2018) (Fig. S1). St3, situated at the southeast entrance to the Gulf of Lion's continental shelf, was the JULIO station, which 253 is dedicated to the study of intrusions of the Ligurian-Provencal current (Barrier et al., 2016). 254 255 St2 and St3 were considered "intermittently bloom" areas or "bloom" areas (D'Ortenzio and d'Alcalà, 2009). These stations (St1–St4) have hosted in the past or continue to host visits 256 serving several monitoring networks such as SOMLIT and ROMARIN, and projects 257 258 including MERMEX-MERITE, COSTAS and IBISCUS. St9 corresponds to offshore station 1 of the PEACETIME cruise (Guieu et al., 2020). It 259 was located north of the North Balearic thermal Front (NBF), at the boundary of the Ligurian 260 261 consensus region (Ayata et al., 2018), in the winter convection area (Fig. S1). The NBF, which is one of the main consensus frontiers of the Mediterranean Sea (Ayata et al., 2018), is 262 found between the Balearic Islands and Sardinia, but its position spans a large area and can 263 vary with wind conditions and seasons (Barral et al., 2021). St10 (offshore station 2 of the 264 PEACETIME cruise) was situated south of St9, very close to the NBF. Although St10 was 265 266 positioned south of the average position of the NBF, it was in fact located slightly north of the front which had moved further south during the sampling period (Rwawi et al., in prep.). St11 267 (offshore station 3 of the PEACETIME cruise) was plainly positioned south of the NBF, in 268 269 the Algerian consensus region (Ayata et al., 2018) (Fig. S1). St10 and St11 were characterised by the presence of intense mesoscale eddies: St10 was rather situated between one 270 271 anticyclonic and one cyclonic eddy, while St11 was located within an anticyclonic eddy (Rwawi et al., in prep.). St9, St10, and St11 are referred to as "bloom", "intermittent bloom" 272

and "no bloom" areas, respectively, according to the D'Ortenzio and d'Alcalà (2009) system
(Fig. 2a; Table 1).

St15, situated in the Gulf of Hammamet (Tunisia), was close to the Sicily Channel, 275 276 which plays host to the exchanges between the two (Western and Eastern) Mediterranean basins, and submitted to the possible entrance of the Atlantic Tunisian Current branch, also 277 called the Atlantic-Ionian stream. St17 (north of the Gulf of Gabès) and St19 (south of the 278 279 Gulf of Gabès) were located at the boundary of and within the Gabès consensus region, respectively (Ayata et al., 2018) (Fig. S1), and are typified by shallow waters, strong 280 influences of tidal pull (the highest of the Mediterranean; amplitude > 2 m) and the Atlantic 281 282 Tunisian Current. St17 and St19 are also characterised by nutrient inputs from Saharan dust deposition or sediment resuspension and the resulting high planktonic productivity levels 283 (Béjaoui et al., 2019). Moreover, given its cumulative index of warming, overfishing and 284 285 pollution effects, the Gulf of Gabès has recently been recognised as a hotspot of anthropogenic pressures within the Mediterranean Sea (Reygondeau et al., 2014). St15, St17 286 287 and St19 were zones marked by a high density of small pelagic fish and considered as "no bloom" (St15) or "coastal bloom" areas (St17, St19) based on the D'Ortenzio and d'Alcalà 288 (2009) system. These stations (St15-St19) have been investigated in the past in the framework 289 of the Tunisian POEMM and ESSATEL programs (Fig. 2a; Fig. S1; Table 1). 290 Due to rough sea conditions that occurred during sampling of St19 (south of the Gulf of 291 Gabès; Fig. 2a, b), the ship went to shelter, first off the island of Djerba and then off the city 292 of Zarzis (Fig. 2a, b), before returning to station St19 to finish work between May 3 and 4. 293 During the transit between off Djerba and off Zarzis, atmospheric deposition sampling was 294 295 carried out following an intense episode of Saharan dust deposition. Besides the ten stations prospected from the R/V Antea, two other stations (T2 and T4) 296

were sampled in the Tunisian waters during leg 2 (2–3 May 2019) on board the R/V *Hannibal*

for trawling small pelagic fishes. T2 was located in the Gulf of Gabès not far from St19,

whereas T4 was situated in the Gulf of Tunis. The duration of trawling was 40 min at T2 and

300 90 min at T4 (Fig. 2a; Table 2). The fish species captured by trawling were anchovy, sardine,

301 sardinella, and mackerel (Lajnef et al., in prep.).

- 302
- 303

2.3. Equipment and operations

At each of the ten stations, the following sampling equipment was deployed and the following operations were performed at sea and on board the R/V *Antea*.

2.3.1. Water sampling with Niskin and Go-Flo bottles and subsequent in-line

307 *filtration.* A trace metal-clean carousel equipped with ten 12-L bottles (1 Niskin, 5 Niskin X,

4 Go-Flo) and a conductivity-temperature-depth probe (CTD; Seabird SBE 911*plus*),

309 mounted with photosynthetically available radiation (PAR) (Biospherical), TChla

310 fluorescence (Aqua Tracka, Chelsea ctg), dissolved oxygen (O₂) (SBE 43), dissolved organic

311 matter (DOM) fluorescence (WETStar, WETLabs) and transmittance (C-Star, WETLabs)

sensors, was deployed from the ship's moon pool *via* the electro-mechanical (conducting)

cable (Table S1; Fig. S2a). Vertical profiles were collected over the depth range 0–250 m or

surface-to-bottom when depth was < 250 m, and seawater was sampled at two depths: 5 m or

the DCM identified based on TChla profiles. Seven bottles (4 Niskin X and 3 Go-Flo) were

dedicated to DCM sampling, while three bottles (1 Niskin, 1 Niskin X and 1 Go-Flo) were

used for 5-m-depth sampling. The Go-Flo bottles were lowered into the water in open

position. At St9 and St10, sampling was also conducted down to 500-m depth to collect

seawater in the oxygen minimum zone (OMZ) identified based on O₂ profiles. At St9, water

320 was collected at 5 m, DCM and 440 m (identified as OMZ), whereas at St10 sampling was

321 carried out at 5 m, DCM, 100, 200, 300, 400 m (identified as OMZ) and 500 m. The Niskin X

bottles used here (model 101012X) are Niskin with a completely free Teflon-coated sample

chamber, Teflon-coated and externally-located stainless steel end plug closure springs, as well 323 324 as Teflon-coated air valves and drainage taps. Before the cruise, all ten bottles were thoroughly washed with hydrochloric acid (HCl) 1 M, demineralized water and ultra-pure 325 326 water (i.e., Milli-Q water from Millipore system, final resistivity of 18.2 M Ω cm). Before sampling, they were rinsed with seawater collected at 50-m and 170-m depth at the first 327 328 station (St2). Between two casts, working on the deck of the ship, we covered the drainage 329 taps and the top (upper stoppers) of the bottles with plastic bags to avoid any sample 330 contaminations.

Once on board, after water collection at 5-m depth and in the DCM, the carousel with 331 332 all the bottles was placed in the wet laboratory for in-line filtration (Table S1; Fig. S2b). For that purpose, the bottles were pressurised to 0.5 bar with argon (UN1006, compressed, 2.2) 333 piped in using a silicon tubing system and quick-connect gas fittings replacing bottle air 334 335 valves. The upper and lower stoppers of the Niskin bottles were held tight by home-made pairs of high-density polyethylene (HDPE) rods and clamps screwed at the top and bottom. A 336 337 10-cm-long piece of acid-cleaned silicon tubing was inserted into the drainage tap of each bottle, and a perfluoroalkoxy (PFA) filter holder (Savillex®, 25 or 47-mm diameter) was 338 connected to the tubing. A graduated container collecting the filtered water served to indicate 339 340 the exact volume filtered on each filter.

In-line filtration makes it possible to effectively filter large quantities of water in clean conditions and to recover the filters and filtered water samples for a battery of analyses. Filtration was done on pre-combusted (450 °C, 6 h), pre-weighed glass fibre filters (GF/F, Whatman) of 25 or 47-mm diameter (pore size: ~ 0.7 µm) for analyses of TChl*a*, pigments, SPM, particulate organic carbon (POC), particulate organic nitrogen (PON), stable isotopes of carbon and nitrogen (δ^{13} C, δ^{15} N), biochemical compounds (carbohydrates, lipids, proteins) and Hg on the size fraction > 0.7 µm. SPM measurements were done on (pre-weighed) 25-

mm and 47-mm-diameter GF/F filters rinsed with ultra-pure water after filtration in order to 348 349 remove salts, but also on (pre-weighed) 47-mm-diameter GF/F filters that had not been rinsed with ultra-pure water (for which the amount of salts was estimated). Filtration was also done 350 351 on pre-cleaned 0.2-µm-pore-size 47-mm-diameter mixed cellulose esters (MCE) filters for the analyses of trace metals/metalloids and microbial diversity on the size fraction $> 0.2 \mu m$, and 352 on 0.8-µm-pore-size 47-mm-diameter pre-cleaned MCE filters for the analyses of copper (Cu) 353 and zinc (Zn) isotopes on the size fraction > 0.8 μ m. GF/F-filtered seawater (< 0.7 μ m) was 354 used for the analyses of nutrients, i.e., silicates $[Si(OH)_4]$, nitrates (NO_3) , nitrites (NO_2) , 355 phosphates (PO₄³⁻) and ammonium (NH₄⁺), dissolved organic carbon (DOC), and absorption 356 357 of chromophoric DOM (a_{CDOM}), whereas 0.2-µm-filtered seawater was used for the analyses of dissolved trace metals/metalloids and Hg (Fig. 3; Table S1). Volume of seawater filtered 358 ranged from 600 mL (25-mm GF/F filter for TChla) to 12 L (47-mm GF/F filter for POC, 359 PON, δ^{13} C, δ^{15} N, and biochemical compounds). 360

Raw (unfiltered) seawater was taken for the analyses of total trace metals/metalloids 361 and Hg/MeHg, while seawater pre-filtered onto 100-µm silk was collected for phytoplankton 362 taxonomy and cytometry analyses. Raw seawater from the DCM was also used to perform on-363 board experiments on microbial methylation/demethylation of Hg (at selected stations), while 364 365 raw seawater collected at 5-m depth and in the DCM at St1, St9, S15 and St19 was used to conduct dilution experiments (see section 2.3.8). Most of these filtrations were also conducted 366 on OMZ water (at St9 and St10) for the same analyses. Raw and 0.2-µm-filtered seawater at 367 the different depths was also used for shipboard analyses of Hg (see section 2.3.9) (Table S1). 368 Filter blanks were regularly run during the cruise: filters were treated as sample filters, either 369 without passing any water on them or by rinsing with ultra-pure water, depending on the 370 parameter. Detailed information on the in-line filtration from Go-Flo and Niskin bottles, 371 including storage of samples, can be found in Fig. S3. 372

2.3.2. In situ optical measurements. A small CTD unit (SBE 19*plus*) equipped with
a TChl*a* fluorescence sensor (WETStar, WETLabs) was deployed from the moon pool on the
hydrographic cable, over the same depth range as the carousel (0–250 m or 0–bottom), to
timely determine the depth of the DCM between different sampling operations, in particular
just before McLane *in situ* pumps (see section 2.3.3) and MultiNet deployments (see section
2.3.5) (Table S1).

A laser optical plankton counter (LOPC, Rolls Royce) and laser *in situ* scattering transmissometry-Holography (LISST-HOLO, Sequoia Scientific) were used together, fixed on a small carousel, to provide information on the abundances and size spectra of particles and plankton (> 20 μ m for LISST-HOLO, > 100 μ m for LOPC). Both systems were deployed from the moon pool on the hydrographic cable. Vertical profiles were performed over the depth range 0–250 m or 0–bottom (Table S1; Fig. S2c).

385 2.3.3. In situ filtration with McLane pumps. Four McLane Large Volume Water Transfer System Samplers (WTS6-142LV, 4–8 L min⁻¹), hereinafter referred to as McLane *in* 386 387 situ pumps, were used to collect large amounts of particles and plankton over different size fractions in the DCM. The four pumps were attached together with clamps and chains, and 388 deployed at the same time from the moon pool on the hydrographic cable to reach the DCM 389 390 (Fig. S2d). Pumping lasted between 40 and 60 min, giving an average ~ 240 L of water filtered by each pump. Three successive casts of the four-pump system were carried out, 391 giving a total of 12 pumping runs done at each station (Table S1). 392

Three pumps (A, B, C) were each mounted with a regular 142-mm filter-holder
("McLane holder") holding one 142-mm-diameter filter: one ~ 0.7-µm-pore-size precombusted pre-weighed GF/F filter, one 0.8-µm-pore-size pre-cleaned MCE filter, or one 20µm-pore-size pre-cleaned Nylon filter. After installing the filters, the three McLane holders
were covered with a 60-µm-pore-size sock-type pre-filter so that the filtered particles size

fractions were 0.7–60, 0.8–60 and 20–60 μ m. At three casts each, pumps A, B and C were able to sample nine filters for analysis of TChl*a*, SPM, POC, PON, δ^{13} C, δ^{15} N, biochemical compounds, PAHs, PCBs, PBDEs and Hg/MeHg on the GF/F filters (0.7–60- μ m size fraction), analysis of trace metals/metalloids, Cu and Zn isotopes and microbial diversity on the MCE filters (0.8–60- μ m size fraction), and analysis of microplastics on the Nylon filters (20–60- μ m size fraction) (Fig. 3; Table S1; Fig. S4).

404 The fourth pump (D) was mounted with a mini-Multiple Unit Large Volume in-situ Filtration System (MULVFS) filter holder composed of baffle tubes on the top followed by 405 successive baffle and filter support plates, for sequential filtration with three different filters 406 407 (142-mm diameter) (Bishop and Wood 2008; Bishop et al., 2012). The baffles were designed to straighten the flow, suppress turbulence, and distribute particles evenly across the filter. 408 409 The filter series used were either 1) one ~ 0.7-µm-pore-size pre-combusted, pre-weighed 410 GF/F filter, one 2.7-µm-pore-size pre-combusted, pre-weighed GF/D filter, and one 20-µmpore-size pre-cleaned Nylon filter, or 2) one 0.8-µm-pore-size pre-cleaned MCE filter, one 3-411 412 µm-pore-size pre-cleaned MCE filter, and one 20-µm-pore-size pre-cleaned Nylon filter. With its three casts, pump D was able to collect nine filters for the analysis of TChla, SPM, 413 POC, PON, δ^{13} C, δ^{15} N, biochemical compounds, and Hg/MeHg on the 0.7–2.7, 2.7–20, and > 414 415 20- μ m size fractions, and trace metals/metalloids on the 0.8–3, 3–20, and > 20- μ m size fractions (Fig. 3; Table S1; Fig. S4). 416

SPM measurements were made on (pre-weighed) 142-mm GF/F, GF/D and Nylon
filters rinsed with ultra-pure water after filtration in order to remove residues of the seawater
salts, but also on (pre-weighed) 142-mm GF/F filters not rinsed with ultra-pure water (for
which the amount of salts was estimated). All filters were placed in glass boxes, plastic boxes
or aluminium foil, depending on the parameter, and stored on board at -20 °C (Fig. S2e, f;
Fig. S4). A special support was purpose-designed to accommodate/fix the filter holders in the

wet laboratory to facilitate the process of preparing the filters before deployment, and then
handling, rinsing with ultra-pure water or direct drying (by connecting the filter holder to a
vacuum pump) the filters after deployment (Fig. S2e, f). Before pump deployment, all the
filter holder and filter systems were rinsed with ultra-pure water. Filter blanks were done at
regular intervals during the cruise.

2.3.4. Water sampling and subsequent in-line filtration with an ASTI pump. 428 429 Seawater was also sampled at ~ 10–20-m depth with a pneumatically-operated Teflon ASTI pump (model PFD2) set up on board and connected to Teflon tubing, which was weighted 430 down and immersed from port side with the hydrology gallows (Table S1). The pumped 431 432 seawater was filtered in-line onto a pre-combusted 142-mm GF/F filter using a 142-mm Teflon filtration holder. This filtration took place in the clean lab container (IFREMER, 433 CNXU 300022/1) installed on the rear deck of the ship. Part of the filtered seawater was 434 435 stored in three 20-L stainless steel jerrycans, while the other part was stored in a metal-free 50-L plastic container. This filtered seawater was used for sieving the large quantities of 436 437 plankton collected with the Multiple Plankton Sampler. The filtered seawater stored in two of the stainless steel jerrycans was used exclusively for sieving on a stainless steel sieve column 438 to obtain plankton samples for organic parameter analyses, and the filtered seawater stored in 439 440 the plastic container was used exclusively for sieving on a nylon sieve column for analyses of inorganic parameters (see section 2.3.5). Filtered seawater from the third jerrycan was 441 amended with 50 mL of dichloromethane, then shaken and degassed and stored in the 442 443 container at ambient temperature for subsequent analyses of dissolved ($< 0.7 \mu m$) hydrocarbons (Guigue et al., in prep.) (Fig. 3; Table S1). All seawater containers (stainless 444 steel and plastic) were rinsed several times with filtered seawater before sample collection. 445 2.3.5. Plankton sampling with a MultiNet and subsequent sieving. A Multiple 446 Plankton Sampler (Midi type, Hydro-Bios), referred to hereafter as "MultiNet", was 447

employed to collect plankton in the DCM. The MultiNet was composed of 5 individual 448 (exchangeable) 2.5-m-long nets with 0.25-m² apertures, a 60-µm mesh size, and cod ends of 449 the same mesh size, together with two Hydro-Bios flowmeters (one into the mouth and the 450 451 other on the side) to assess the volume of water filtered by the nets, plus a CTD sensor and a TChla fluorometer (Chelsea ctg). The MultiNet was connected via the electro-mechanical 452 453 cable to the operating computer to enable online monitoring of sensor feedback and opening 454 and closing of the nets, mainly based on flowmeter data and the amount of water passing through the open net. It was operated from the stern A-frame. Since the device was deployed 455 horizontally in the DCM, the usual configuration (for vertical casts) was modified with a V-456 457 fin deflector attached below the MultiNet and the five cod ends attached using a helicoidal bucket connector (Fig. S2g, h). Ship speed while towing the MultiNet was ~ 2 knots (Table 458 S1). Once the five nets were filled (after 30 to 100 min depending on the station), the 459 460 MultiNet was hauled back on board, the cod ends were rinsed out with local seawater, and their content was transferred to pre-cleaned 10-L HDPE bottles. The instrument was then 461 returned to the water. This operation ("cast") had to be repeated several times (between two 462 and eight horizontal casts, depending on the station) until we got sufficient amounts of 463 plankton. At St19, due to technical issues, the MultiNet was replaced by a square net with a 1-464 m^2 aperture mounted with single 60-µm mesh-size net. Total filtered water volume with the 465 MultiNet/square net ranged from 314 (St2) to 2373 m³ (St10). 466

In the clean on-board container lab, plankton collected in HDPE bottles was then sizefractionated on 1) a column of five stainless steel sieves (60, 200, 500, 1000 and 2000-μm
mesh-size) by wet-sieving with the GF/F filtered seawater stored in stainless steel jerrycans
for subsequent analyses of organic parameters, and 2) a column of five nylon sieves (60, 200, 200, 1000 and 2000-μm mesh-size) by wet-sieving with the GF/F filtered seawater stored in the plastic container for the subsequent analyses of inorganic parameters (Fig. S2i, j). The

filtered seawater was routed with a gentle flow to the top of the sieve column by the ASTI 473 474 pump using Teflon tubing. The plankton size fractions recovered on the stainless-steel sieves were shared out and transferred to pre-combusted glass flasks for the analyses of POC, PON, 475 δ^{13} C/ δ^{15} N, biochemical compounds, PAHs, PCBs, PBDEs and PFASs, or to plastic flasks for 476 zooplankton taxonomy and imagery, TChla (60-200-µm size fraction only) and microbial 477 diversity (60–200 and 200–500-µm size fractions only). The plankton size fractions recovered 478 479 on the nylon sieves were shared out and transferred to pre-cleaned polypropylene flasks for the analyses of trace metals/metalloids, Hg and ¹³⁷Cs (Fig. 3; Table S1; Fig. S5). Planktonic 480 biomass (in mg m⁻³ dry weight) was also determined in each of the size fractions (Fierro-481 482 González et al., 2023). All samples were stored at -20 °C except those for zooplankton taxonomy that were stored at ambient temperature after adding buffered formalin (Fig. S5). 483 2.3.6. Plankton and micro-plastic sampling with Triple and Manta nets. A triple 484 485 net, equipped with 60, 120 and 200-µm mesh-sized nets each with 0.60-m aperture diameter was used to undertake two vertical tows over the depth 0-200 m (or 0-bottom if depth < 200486 m), i.e., one tow for biomass, and one tow for taxonomy and imagery. The device was 487 deployed from port side using hydrology gallows (Table S1; Fig. S2k). After collection, the 488 cod-end contents from each of the three nets (first tow) were passed through GF/F filters that 489 490 were stored in Petri dishes at -20 °C for biomass measurements, while the cod-end contents from the second tow were passed through a 60-µm sieve for volume reduction and transferred 491 to 250-mL plastic tubes amended with 12.5-mL buffered formalin for taxonomy and imagery 492 analyses. At St1, St9, S15 and St19, the contents of the 200-µm net were also collected to 493 analyse chlorophyll gut-contents of mesozooplankton (see section 2.3.8). 494 Two Manta nets were used for horizontal tows in surface waters: one a 330-µm mesh 495 size (1 tow, 3 knots, 20 min max) for microplastic analyses, and the other a 60-µm mesh size 496

497 (1 tow, 1 knot, 10 min max) for microplastics and plankton (neuston) analyses, both deployed

from port side using hydrology gallows (Table S1). After collection, the content of the 330µm Manta net was sieved successively through 5000 and 300-µm mesh-size sieves (if large
items were present in the collectors) and transferred to a 1-L glass bottle added with formalin
(4% final concentration). The content of the 60-µm Manta net was transferred to a 1-L glass
bottle added with 50-mL buffered formalin.

2.3.7. In-line filtration onto cartridges. At each station, in-line filtration was carried 503 504 out in the wet laboratory on seawater collected continuously at 2-m depth by the shipboard pump system (Table S1). Two polypropylene cartridges (Polycap HD), the first with a 20-µm 505 pore size and the second with a 0.45-µm pore size, were mounted in series to recover the 506 507 0.45-20-µm and > 20-µm size fractions. Around 1000 L of seawater was filtered on the two cartridges at each station. The dissolved fraction (< 0.45 µm) was collected in 20-L plastic 508 containers. These samples served for radionuclide analyses (¹³⁷Cs) (Radakovitch et al., in 509 510 prep.).

2.3.8. Dilution experiments. At St1, St9, S15 and St19, seawater collected from the 511 512 carousel bottles at 5-m depth and in the DCM was used to perform dilution experiments by means of a thermostatic incubation chamber (Table S1). These experiments were 513 implemented to assess the phytoplankton production rates and microzooplankton grazing 514 515 rates. At the same stations, the contents of the 200-µm Triple nets (vertical tows) were used to analyse the chlorophyll gut contents of mesozooplankton by fluorescence in order to estimate 516 the impact of mesozooplankton grazing on large phytoplankton (i.e., nano- and micro-517 518 phytoplankton). These experiments and measurements served to determine the carbon fluxes and type and structure of the planktonic food webs (Medebb et al., in prep.). 519

520 2.3.9. Shipboard analyses. Unfiltered and 0.2-μm-filtered seawater collected from
521 the carousel bottles at 5-m depth, in the DCM and at the other depths (profiles, OMZ) served
522 for shipboard analyses of total and purgeable Hg using a Tekran[®] auto-analyser (model 2500),

523 consisting in purge and trap of volatilized Hg species followed by cold vapour atomic

fluorescence spectrometry (CVAFS) detection (Table S1; Fig. S2m).

A part of the seawater collected continuously at 2-m depth by the shipboard pump
system was subsampled and routed to a CytoSense[®] automated flow cytometer (CytoBuoy)
installed in the dry laboratory. The CytoSense was employed to analyse (in terms of
identification and abundance) individual or colonial phytoplankton cells sized between 0.8
and 800 μm. Cytometry measurements were carried out continuously at the different stations
but also all along the transect (Table S1; Fig. S2l; Boudriga et al., 2022).

2.3.10. Atmospheric deposition. During the cruise, wet atmospheric deposition 531 532 samples were collected using two rain collectors placed in PVC pipes that were fixed at the front of the ship (10 m above sea level). The collector for trace metal samples was composed 533 of an acid pre-cleaned, metal-free plastic bottle and funnel (Fig. S2n). The collector for PAH 534 535 samples was composed of a pre-combusted amber glass bottle and glass funnel (Table S1; Fig. S20). Overall, six rainwater samples were collected (Table 3): two at St10 (Rains 1 and 536 537 2), one at St11 (Rain 3), one at St19 (Rain 4), one during the transit from off Djerba island to off Zarzis city (Rain 5, south of St19; Fig. 2a, b) (for both collectors), and one at St1 (Rain 6 538 only in the PAH collector). The "transit to Zarzis" sample was collected as a result of a dry 539 540 deposition episode (intense Saharan dust event). In this case, ultra-pure water was used to rinse the plastic and glass funnels in order to retrieve all the dry fraction that had settled onto 541 them. Atmospheric samples collected during the cruise were frozen and stored at -20 °C (for 542 PAHs) or at 4 °C (for metals/metalloids), then filtered (at 0.2 or 0.7 µm) in the laboratory to 543 separate dissolved and particulate phases before treatments and analyses. Atmospheric 544 forecast bulletins were regularly sent to the on-board team to anticipate these rain events. 545 Detailed information on the rain samples collected during the cruise can be found in Table 3. 546

Glass collectors were also set up in two ground stations located in the northern part 547 548 (Marseille, France) and southern part (Sfax, Tunisia) of the Mediterranean basin (Fig. 2a; Fig. S2p) for sampling total (dry and wet) atmospheric deposition and running the subsequent 549 550 PAH analyses. This sampling was done on a regular basis (bi-monthly, monthly, or daily in the case of strong rainfall events) from March 2019 to June 2020, i.e., before, during, and 551 552 after the cruise (Poindron et al., in prep.).

553 2.3.11. On-vessel scientific equipment and continuous measurements. Subsurface temperature and salinity were recorded continuously at high frequency all along 554 the transect from flow-through pumped seawater at 2-m depth, using a thermosalinograph 555 556 (TSG, SeaBird SBE 21). A weather station (Batos 1.1 D, Météo France) continuously recorded atmospheric parameters (temperature, wind, pressure, humidity, PAR). 557

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559 2.4. The sequence of operations and main parameters analysed on the different size fractions 560

The typical sequence of at-sea/on-board operations conducted from the R/V Antea at 561 each station is given in Table S2. The whole sequence of operations at each station lasted 48 h 562 on average but varied depending on meteorological conditions encountered and occasional 563 564 technical problems. The detailed sequence and timing of the main operations performed during the whole cruise are presented in Table S3, and the total number of operations 565 completed is provided Table S4. Table S5 reports the time and depth of sampling (mostly in 566 567 the DCM) at each station for the three main operations at sea: the water sampling with the carousel/Niskin and Go-Flo bottles, in situ filtration with the McLane pumps, and plankton 568 sampling with the MultiNet. Table S6 summarises the main parameters analysed on the 569 570 different size fractions (from < 0.2 to $> 2000 \mu$ m) recovered from filtration and sieving after the collecting large amounts of plankton, particles, and water in the DCM and 571

572	surface/subsurface waters (0-5 m depth), i.e., 1) contaminants, with trace metals,
573	organometals and metalloids (As, Cd, Cr, Cu, Fe, Hg, MeHg, Mn, Ni, Pb, Sb, Zn, etc.),
574	organic contaminants (PAHs, PCBs, PBDEs, PFASs), radionuclides (137Cs) and microplastics,
575	2) plankton biomass, size structure, taxonomy, cytometry, diversity, and pigment
576	composition, and 3) biogeochemical parameters, including nutrients [Si(OH)4, NO3 ⁻ , NO2 ⁻ ,
577	NH4 ⁺ , PO4 ³⁻], TChla, SPM, dissolved and particulate organic matter (POC, PON, DOC,
578	a_{CDOM}), C and N isotopic ratios (δ^{13} C, δ^{15} N), and biochemical compounds (carbohydrates,
579	proteins, lipids).
580	
581	
582	3. Environmental context during the cruise
583	
584	3.1. Meteorological context
585	Atmospheric deposition is known to be an external source of metals and PAHs (Castro-
586	Jimenez et al., 2012; Jordi et al., 2012; Desboeufs et al., 2022) as well as of nutrients (Guieu
587	et al., 2020) for Mediterranean surface seawater. The cruise was conducted during the spring,
588	when dust deposition events were commonplace in the Western Mediterranean (Guieu et al.,
589	2020). During the cruise, the Aerosol Optical Depth (AOD) maps derived from the Spinning
590	Enhanced Visible and InfraRed Imager (SEVIRI) satellite instrument (<u>https://www.icare.univ-</u>
591	lille.fr/data-access/browse-images/geostationary-satellites/; Thieuleux et al., 2005)
592	highlighted two dust transports with high AOD (> 0.8) through the Mediterranean Sea: one
593	that occurred between 17 and 26 April 2019 (Fig. 4a; Fig. S6) and a second that occurred
594	between 3 and 4 May 2019 (Fig. 5a; Fig. S7). These events were associated with significant
595	cloud cover (Fig. 4b, 5b). The Non-hydrostatic Multiscale Model/Barcelona Supercomputing
596	Centre (NMMB/BSC)-Dust model and NASCube (http://nascube.univ-lille1.fr/; Gonzales and

Briottet, 2017) confirmed the emission and export of dust plumes from North Africa to over 597 598 the Western Mediterranean. The atmospheric dynamics and patterns of the first event are described in detail in Calidonna et al. (2020). This event was associated with a northward 599 600 atmospheric flux loaded by dust emission from Algeria and southern Morocco. The second dust plume was transported from Algeria and Tunisia by north-eastward winds (not shown). 601 602 For the April event, dust plume transport was mainly associated with wet deposition 603 from NMMB/BSC-Dust model forecasts, in agreement with the intensely cloudy conditions (Fig. 4c, d). The geographic range of dust deposition events covered a region from the 604 Balearic basin to the Tyrrhenian Sea and then the Ligurian Sea (Fig. 4a-d), i.e., the vicinity of 605 606 R/V's locations during St10 and St11 between 20 and 23 April and then on 25 April along the Western coastline of Sardinia (Table 1). The model predictions were confirmed by visual on-607 608 board observations of precipitations around and above the R/V's location, corresponding to 609 the first three rain samples (Rains 1, 2 and 3) (Table 3). Note that wet dust deposition started from the afternoon of 22 April after the R/V's arrival on St10, thus ruling out a direct impact 610 611 of dust deposition on the first cast completed at this station. However, rain events probably impacted surface waters over a large region around the R/V's location during St10 and St11. 612 For the event in May, NMMB/BSC-Dust model forecasts point mainly to dry dust 613 614 deposition occurring between 3 and 4 May over the Gulf of Gabès and the Libyan coast (Fig. 5c, d). During this event, the R/V was at St19 in the southern Gulf of Gabès then in transit to 615 Zarzis (Table 1; Table S3). Dry dust deposition was confirmed by visual observations of 616 617 dusty sky and substantial deposited dust material on the decks of the R/V between 3 and 4 May. The Rain 5 sample was collected during this event (Table 3). The inflow of desert dust 618 was also consistent with the highest temperatures and lowest pressures recorded by ship's 619 permanent instrumentation (Fig. S8). It is therefore very likely that this event represented a 620 supply of metal-bearing dust to the surface waters. 621

Outside of these desert dust transport periods, Hybrid Single-Particle Lagrangian 622 623 Integrated (HYSPLIT) back trajectories modelling showed that the air masses around the R/V position came from Europe (not shown). Nevertheless, no intense pollution event was either 624 observed or predicted during the cruise from satellite or model outputs. Moreover, no 625 volcanic emissions from Etna were recorded during the cruise. 626

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3.2. Hydrological context

Subsurface (2-m depth) temperature gradually increased from north to south of the 629 transect, whereas subsurface salinity decreased from North to South but increased again at the 630 631 extreme south of the transect (Gulf of Gabès) (Fig. 6). The lowest subsurface temperatures (~ 14.0 °C) were recorded in the Bay of Marseille, while the highest subsurface temperatures 632 (19.5 °C) were observed in the south of the Gulf of Gabès, near Djerba and Zarzis. The 633 634 highest subsurface salinity values were found in the Ligurian region (38.5) and the lowest (37.1) in the northern Tunisian coastal waters, particularly in the Gulf of Tunis (Fig. 2a, 6; 635 Fig. S1). Because of the vast and contrasted area covered by the cruise, CTD profiles 636 displayed huge variability with depth in temperature and salinity and in TChla and dissolved 637 O_2 concentrations between stations (Fig. 7, 8). Despite this high variability, some similarities 638 639 emerged. For the stations off French shores (St1-St4), temperature, salinity and TChla values over the water column ranged from 13.6–14.9 °C, 37.8–38.4 and 0.02–0.86 μ g L⁻¹, 640 respectively, with a slight decrease in temperature and a slight increase of salinity towards the 641 642 bottom (Fig. 7). Offshore stations (St9-St11) also showed a decrease in temperature and 643 increase of salinity with depth, but with a greater range of variation (compared to the stations along the French coast) in terms of temperature (13.4–15.5 °C), salinity (37.3–38.7) and 644 TChla concentrations (0.00–1.49 μ g L⁻¹) with manifestly deeper thermoclines and haloclines 645 (Fig. 7, 8). TChla concentrations were particularly high at St9 (Fig. 7). St11 was marked by a 646

significant increase in both temperature and salinity between 200 and 400 m followed by a decrease between 400 and 430 m before remaining stable down to the bottom (Fig. 8). Tunisian stations (St15–St19) showed higher water column temperatures (14.7–17.9 °C) and lower salinities (37.2–37.8) compared to stations further north, in accordance with subsurface data (Fig. 6), and lower TChl*a* concentrations (0.02–0.70 μ g L⁻¹) compared to the northernmost stations. At these Tunisian stations, thermoclines, haloclines and DCM were located relatively deep in the water column, although St17 displayed much more

homogeneous profiles (Fig. 8).

T–S diagrams allowed us to identify the main water masses encountered at each station 655 656 (Fig. 9). At the northern stations (St1–St4) with temperatures of 13.6–14.9 °C and salinities of 37.8–38.4 over 0–150-m depths, we detected (modified) Atlantic Water (AW) coming mainly 657 from the Ligurian Sea and flowing along the continental slope with Ligurian-Provencal 658 659 current. These "resident" AW (RAW), which have been in the Mediterranean for a long time, have been modified and are saltier (and warmer) than the AW that arrived more recently 660 through the Strait of Gibraltar (Millot and Taupier-Letage, 2005; Balbín et al., 2014). Moving 661 south, St9 was characterised by the presence of RAW over the depth 0-300 m and Levantine 662 Intermediate Water (LIW) over the depth 300–500 m (Fig. 9). LIW is formed in the Eastern 663 664 Mediterranean Basin by the combined effect of wintertime cooling and summertime evaporation, leading to a warm (temperatures of 13.8–16 °C) and salty (salinities of 38.3–39) 665 intermediate layer that is visible over the whole Mediterranean Sea (Lascaratos et al., 1993; 666 Balbín et al., 2014; Margirier et al., 2020). Once formed, the LIW spreads throughout the 667 entire Eastern and Western Basins and can be identified through a salinity maximum at 668 between ~ 200 and 600-m depth in the Western Basin before it eventually flows out of the 669 670 Mediterranean Sea as one of the main components of Mediterranean outflow water (Ben Ismail et al., 2012; Vargaz-Yanez et al., 2012). In addition, the presence of Western 671

Mediterranean Intermediate Water (WIW) was detected at St9 around 100-m depth with 672 temperatures of 13.2–13.9 °C and salinities of 38.4–38.6 (Fig. 7, 9). The WIW forms during 673 winter in the Northwestern Mediterranean Basin due to surface cooling of RAW and 674 intermediate convection (Salat and Font, 1987; Gasparini et al., 1999) and has been recorded 675 in the Algerian Basin by Benzohra and Millot (1995) and in the Sicily Channel by Ben Ismail 676 677 et al. (2012), flowing eastwards below the AW in the Algerian Current through the Channel 678 of Sardinia. The WIW is characterised by a relative minimum potential temperature located between 100 and 200-m depth. The circulation of WIW has only been investigated in the 679 Algerian Basin, and there is speculation that it follows the same flow paths as the overlying 680 681 AW (Millot, 1999). St10 and St11 showed weak signature of the WIW and the presence of marked LIW indicating a dominant influence of the northward advection of LIW rather than 682 winter cooling in the water column. The LIW water was well evident in St11, the closest 683 684 station to the Sardinia Channel (Fig. 8, 9). At the northernmost tip of the Tunisian coast, the water temperature progressively increased and the salinity decreased, revealing the presence 685 of fresher or "new" AW (NAW) coming from the Algerian Basin. The water column arriving 686 at St15, located in the Gulf of Hammamet, was mostly made up of RAW and NAW with 687 temperatures of 14.8–17.5 °C and low salinities of 37.1–37.8 from surface to bottom. Moving 688 689 southward toward the Gulf of Gabès (GG), the temperature and salinity increased sharply to reach 17.1 °C and 37.5 at St17 and 17.9 °C and 37.8 at St19 (Fig. 8, 9). 690

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3.3. Biogeochemical context

The analytical methods used to determine biogeochemical parameters presented in this section are detailed in Text S1. Considering the whole cruise period, the average (over 32 days) surface TChl*a* concentration derived from satellite data was higher at St9 (~ 1 μ g L⁻¹) than in the other stations (< 1 μ g L⁻¹) (Fig. 10). The higher content of phytoplankton biomass

697	at St9 was also corroborated in the TChla concentrations actually recorded during the cruise
698	at 5-m depth, either from CTD measurements or analyses on discrete samples (with a
699	significant correlation between both measurements: $r = 0.98$, $n = 10$, $p < 0.0001$; Table S7). In
700	line with the discrete samples, the highest TChla concentrations at 5-m depth were found at
701	St9 (2.90 μ g L ⁻¹) followed by St4 (1.57 μ g L ⁻¹) and St1 (0.84 μ g L ⁻¹), while the lowest TChla
702	concentrations were observed at St17 (0.15 μ g L ⁻¹) and St15 (0.10 μ g L ⁻¹) (Table S7).
703	DCM was shallowest at St4 (13 m), St1 and St9 (20 m) and deepest at St10 (51 m), St2,
704	St3 (53 m) and St15 (66 m) (Fig. 7, 8, 11a; Table S7). TChla in the DCM followed a fairly
705	similar pattern of distribution to TChla at 5-m depth, with a significant correlation between
706	both measures based on both CTD and discrete sample data (r = $0.64-0.92$, n = 10 , p =
707	0.0001–0.048). TChla concentrations measured on discrete samples from the DCM were
708	highest at St9 (1.54 μ g L ⁻¹), in line with the surface/subsurface TChla data (Fig. 10). High
709	TChla concentrations were also recorded south of the Gulf of Gabès (at St19: 1.45 μ g L ⁻¹)
710	(Fig. 11b; Table S7), even though at this station it was not really a DCM that was observed
711	but rather a bottom-lying bead, as can often be observed for shallow coastal stations. St4 and
712	St1 showed fairly high TChla concentrations of 0.98 and 0.77 μ g L ⁻¹ , respectively. The lowest
713	TChla concentrations in the DCM were found at St17 (0.21 μ g L ⁻¹), St2, and St11 (0.38 μ g L ⁻
714	¹) (Fig. 11b; Table S7).

The North-to-South distribution of other biogeochemical parameters measured in the DCM presented interesting features. POC concentration had a fairly similar pattern of distribution to TChl*a*, with decreasing values from St1 to St3, a maximal value at St9 (156.4 μ g L⁻¹), then a decrease up to St17, and finally a higher value at St19 (55.9 μ g L⁻¹) (Fig. 11c; Table S7). Si(OH)₄ concentration followed much the same pattern but with an increase from St1 to St3 and a maximal value at both St9 and St19 (~ 2.00 μ M) (Fig. 11d; Table S7). NO₃⁻ concentration increased from St1 to its peak at St3 (1.31 μ M), was still high at St9 (0.96 μ M),

722	and then decreased along to St17–St19 (Fig. 11e; Table S7). PO ₄ ³⁻ concentration was
723	particularly high at St3 (0.31 μ M) and St19 (0.55 μ M) (Fig. 11f; Table S7). Finally, DOC
724	concentration and a_{CDOM} decreased from St1 to St9 or St10 then increased up to St19 where
725	they reached maximal values (76.2 μ M and 2.05 m ⁻¹ , respectively) (Fig. 11g, h; Table S7).
726	Principal component analysis (PCA) based on the Pearson's correlation matrix was
727	applied on these biogeochemical parameters recorded in the DCM at each station (Fig. 12).
728	The first principal component (PC1), which explained 47% of total variance within samples,
729	was mainly driven by PO_4^{3-} , DOC, a_{CDOM} , and to a lesser extent TChla and Si(OH) ₄ . The
730	second principal component (PC2), which accounted for 34% of total variance, was driven by
731	POC, NO ₃ ⁻ , TChla, and Si(OH) ₄ . Four groups of samples (stations) emerged from this PCA:
732	1) St2, St3, St4, St10, characterised by moderate concentrations of nutrients and TChla and a
733	low organic matter content; 2) St1, St11, St15, St17, with the lowest concentrations in
734	nutrients and TChla (except St1); 3) St9, which showed the highest in Si(OH)4, NO ₃ ⁻ , TChla
735	and POC concentrations, and the lowest PO_4^{3-} and DOM contents; 4) St19, which showed the
736	highest concentrations in Si(OH) ₄ , TChla (equivalent to those of St9), PO ₄ ³⁻ and DOM as well
737	as high POC and low NO ₃ ⁻ concentrations (Fig. 12).
738	Overall, the levels of TChla, nutrients and other biogeochemical parameters observed
739	along the North-South transect in the subsurface waters and DCM in spring 2019 during the
740	MERITE-HIPPOCAMPE cruise are consistent with previous observations in the
741	Mediterranean Sea (The Mermex Group, 2011; Salgado-Hernanz et al., 2019; Guieu et al.,
742	2020; Marañón et al., 2021). The TChla levels in the different stations were also consistent
743	with their positioning in terms of bloom-condition areas and consensus regions defined by
744	D'Ortenzio and d'Alcalà (2009) and Ayata et al. (2018). TChla concentrations tended to be
745	higher at stations located north of the North Balearic front (St1-St4, St9) than stations located
746	south (St10, St11, St15, St17, St19).

Interestingly, two stations, St9 and St19, clearly stood apart from the others in terms of 747 748 biogeochemical content (Fig. 11, 12). St9 was situated at the boundary of the Ligurian consensus region, in the wintertime deep convection area. Time-series of water temperature 749 recorded at different depths at the LION mooring (42° 02' N, 4° 40' E) highlighted a 750 deepening of the mixed layer depth and thus the convection process down to 1500-m depth 751 within the Ligurian area in early February 2019 (Fig. S9). In late March 2019, the end of the 752 753 convection process induced the occurrence of an intense phytoplankton bloom in the same area (Fig. S9). This illustrates the fact that 2019 was a relatively convective and productive 754 year (Margirier et al., 2020; Bosse et al., 2022), and that St9, located at the border of this 755 756 convective and productive zone, still presented high concentrations of TChla, POC and some nutrients in the May 2019 sampling period. 757

The high levels of TChla, $Si(OH)_4$, PO_4^{3-} , DOM, and POC encountered at St19 south of 758 759 the Gulf of Gabès is probably related to the Saharan dust deposition event (see section 3.1), but the effect of sediment resuspension cannot be excluded due to high wind speeds during 760 this period (Fig. S8). In the Gulf of Gabès, TChla concentrations can reach > 1 μ g L⁻¹ close to 761 the coast of Djerba and Kerkennah Islands during the spring season (Bel Hassen et al., 2009). 762 The Gulf of Gabès is known to be regularly submitted to Saharan dust deposition (Béjaoui et 763 764 al., 2019). This Saharan dust, enriched in phosphorus, has been shown to induce 765 phytoplankton blooms in the Gulf (Hamza et al., 2016; Béjaoui et al., 2019), and as described above, intense Saharan dust deposition events occurred in this area during the cruise, for 766 which we collected on-board rain and particle samples (Table 3; Fig. 5; Fig. S7, S8). 767 Furthermore, in this shallow ecosystem submitted to strong tides and resulting currents, 768 disturbance and resuspension of sediments could release elements such as nutrients and 769 770 organic matter into the water column, which in turn may stimulate planktonic activity (Bel Hassen et al., 2009; Fourati et al., 2018; Zouch et al., 2018). 771

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4. Overview of the types of articles produced through the cruise

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Fig. 13 shows the types of articles that are a part of this special issue in the frame of the 776 777 MERITE-HIPPOCAMPE cruise. There are a series of articles concerning the transfer and 778 accumulation of contaminants in planktonic food webs in the DCM but also in surface/subsurface waters. These articles present: 1) contaminant concentrations in the 779 different planktonic/particulate size fractions and, for several of them, in the dissolved phase 780 781 of water (see Table S6 for the size fractions-contaminant analysis correspondence), and 2) their resulting factors of bioconcentration, bioaccumulation or food accumulation. The 782 contaminants investigated are trace metals/metalloids (Cr, Mn, Fe, Co, Ni, Cu, Zn, As, Se, 783 784 Mo, Ag, Cd, Sb, Pb...) (Chifflet et al., 2023), with a focus on Cu and Zn isotopes (Chifflet et al., 2022), Hg and MeHg (Tesán Onrubia et al., in prep.), PAHs (Guigue et al., in prep.), 785 PCBs, PBDEs, PFASs (Tronczynski et al., in prep.), and radionuclides (¹³⁷Cs) (Radakovitch 786 et al., in prep.). Another work will treat of the abundance and composition of microplastics in 787 both surface waters and the DCM (Fig. 13). 788 789 There are also a series of articles dealing with the composition and structure of planktonic food webs. Indeed, the paper by Tesán Onrubia et al. (2023) on the stable C and N 790 isotopes (δ^{13} C, δ^{15} N) and biochemical content (lipids, carbohydrates, proteins) of the different 791 792 planktonic size fractions serves to highlight the structure of planktonic food webs and the 793 transfer of organic matter within them, and to assess the factors of trophic accumulation used in the contaminant-related papers. Moreover, several articles use the cytometry and 794

taxonomy/microscope analyses, imaging (zooscan, flowcam), optical in situ measurements

796 (LOPC, LISST-HOLO) and/or 16S and 18S rRNA high-throughput sequencing analyses to

cover the abundance, biomass, distribution, size structure, and composition/diversity of the 797 798 following planktonic groups, mostly for the DCM but also for surface/subsurface waters or even within the 0-200-m depth layer (see Table S6 for the correspondence with size 799 800 fractions): pico-, nano- and micro-phytoplankton (Boudriga et al., 2022; Bellaaj Zouari et al., in prep.), micro- and meso-zooplankton and their biomass ratio with detritus (Fierro-González 801 802 et al., 2023), as well as free-living and plankton-associated bacterioplankton (heterotrophic 803 prokaryotes) (Bellaaj Zouari et al., in prep.; Quéméneur et al., in prep.), and nano- and microzooplankton related to the microbial loop (Bellaaj Zouari et al., in prep.). In addition, a 804 complementary paper proposes a methodological approach for the study of the microbiota 805 806 associated with plankton (Cabrol et al., in prep.). Also, the effect of the Saharan dust deposition event on pico- and nano-phytoplankton community in the south of Gabès is 807 808 covered by Boudriga et al. (in prep.). Overall, these articles will allow us to understand which 809 planktonic groups are present and how they influence/participate in transfers of contaminants within planktonic networks. Our understanding of the contaminant transfer processes at work 810 811 within planktonic food webs is also enhanced by the dilution experiments and the subsequent determination of phytoplankton production rates and micro- and meso-zooplankton grazing 812 rates (Meddeb et al., in prep.) (Fig. 13), as well as the grazing and excretion rates estimated 813 814 from the zooplankton size-structure according to allometric relationships allowing an estimation of grazing pressure and nutrient recycling by the metazooplankton (Fierro-815 González et al., 2023). 816

Looking beyond plankton, there is one article dedicated to δ^{13} C and δ^{15} N stable isotopes and concentrations of Hg, PAHs and PCBs in small pelagic fishes collected in Tunisian waters (Lajnef et al., in prep.). This study should provide valuable pointers to help establish a link between the contamination of plankton and the contamination of higher trophic levels. The concentrations of trace metals and PAHs in wet (rain) atmospheric deposition samples

822	collected during the cruise and on land is investigated to assess the role of dry/wet
823	atmospheric deposition as a source of contaminants in surface marine waters (Poindron et al.,
824	in prep.). Finally, the overall transport patterns and hydrodynamic context of the cruise using
825	modelling and satellite data, and their potential implication for contaminant origin and
826	distribution are addressed by Rwawi et al. (in prep.) (Fig. 13).
827	
828	
829	Data availability
830	All data from the MERITE-HIPPOCAMPE cruise (<u>https://doi.org/10.17600/18000900</u> ;
831	Tedetti and Tronczynski, 2019) is stored in the MISTRALS-SEDOO database
832	(<u>https://mistrals.sedoo.fr/MERITE/</u>) and will be made publicly accessible once all the articles
833	related to the cruise are published in the present special issue. In the meantime, data can be
834	obtained upon request from the corresponding author. In addition, navigation data and CTD
835	profiles from the MERITE-HIPPOCAMPE cruise are available via the IFREMER/SISMER
836	database (<u>https://data.ifremer.fr/SISMER</u>).
837	
838	
839	Author contribution statement
840	All the authors participated in the MERITE-HIPPOCAMPE project and design of the
841	manuscript.
842	Conception and design of study: M.T., J.T., F.C., M.P., S.B.I., C.S., M.B.H., K.D.,
843	S.C., A.B.Z., D.B., A.B., C.BP., N.B., L.C., C.C., L.C., M.N.D.Y., A.D., JC.D., C.G.,
844	L.G., A.H., LE.HB., S.J., J.K., B.M., O.P., M.Q., O.R., A.S.H., M.T., N.Z., C.G.

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916	
917	Supplementary information
918	Supplementary material related to this article is available online at: xxx
919	

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1312 Figure captions

1313

Figure 1. Conceptual scheme and questions addressed by the MERITE-HIPPOCAMPE cruise to identify the role of Mediterranean plankton as biological pump of contaminants. It considers the accumulation and transfer of metallic/metalloid/organometallic and organic contaminants at the atmosphere-water-plankton interfaces and through the planktonic food webs, i.e., phyto-, zoo- and bacterio-plankton, with the deep chlorophyll maximum (DCM) as privileged study zone.

1320

1321 Figure 2. a) Location of the ten stations (black circles) investigated during the MERITE-HIPPOCAMPE cruise (13 April-14 May 2019) along a North-South transect in the 1322 Mediterranean Sea on board the R/V Antea. The main characteristics of these stations are 1323 1324 provided in Table 1. In addition, two other stations (T2 and T4, brown circles) were sampled in the Tunisian waters during leg 2 (2–3 May 2019) on board the R/V Hannibal for trawling 1325 1326 of small pelagic fishes (see Table 2 for more details). Two ground stations, located in the northern part (Marseille, France) and southern part (Sfax, Tunisia) of the Mediterranean 1327 basin, were set up for the collection of atmospheric deposition samples from March 2019 to 1328 1329 June 2020 (i.e., before, during and after the cruise). b) Cruise track with the position of the ten stations studied on board the R/V Antea. During leg 1 (13-28 April; from La Seyne-sur-Mer 1330 to Tunis), five stations were sampled: St2, St4, St3, S10 and St11 (in this chronological 1331 order), while the five other stations were sampled during leg 2 (30 April-14 May; from Tunis 1332 to Gulf of Gabès, and then return to La Seyne-sur-Mer): St15, St17, St19, St9 and St1 (in this 1333 chronological order). 1334

1335

Figure 3. Summary of our approach with 1) the collection of large amounts of plankton,

1337 suspended particles, and water in the deep chlorophyll maximum (DCM) and

1338 surface/subsurface waters (0–5 m depth) with various collecting instruments, 2) the separation

1339 of these materials into diverse size fractions by sieving or filtration, and the sharing of the

1340 obtained size fractions for numerous biological and chemical analyses. Adapted from Alcaraz

1341 and Calbet (2003).

1342

Figure 4. Saharan dust event of April. Here is presented for 22 April 2019 which is the day
where the dust plume was the most intense: a) Daily Aerosol Optical Depth (AOD, unitless)
product from MSG-SEVIRI, b) MODIS image from NASA worldview, and BSC-DREAMDust forecast of c) dry and d) wet deposition (in mg m⁻²).

1347

Figure 5. Saharan dust event of May. Here is presented for 3 May 2019 which is the day

1349 where the dust was located over ship's position: a) Daily Aerosol Optical Depth (AOD,

1350 unitless) product from MSG-SEVIRI, b) MODIS image from NASA worldview, and BSC-

1351 DREAM-Dust forecast of c) dry and d) wet deposition (in mg m^{-2}).

1352

1353 **Figure 6.** Distribution of subsurface **a**) temperature (°C) and **b**) salinity along the North-

1354 South Mediterranean transect recorded continuously from the flow-through pumped seawater

1355 at 2-m depth. Ocean Data View (ODV) software version 4.6.5. Schlitzer, R.,

1356 http://odv.awi.de. 2014.

1357

Figure 7. Vertical profiles of temperature (°C), salinity, TChla concentration (μ g L⁻¹) and

1359 dissolved O_2 concentration (µmole kg⁻¹) issued from the main carousel CTD deployments

1360 (Seabird SBE 911*plus*) at stations St1–St9. The dotted lines represent the depth of sampling in

the deep chlorophyll maximum (DCM) for Niskin/Go-Flo bottles. In box, the depth of the
station. One or two profiles are displayed for each station. For each profile, only data acquired
during the upcast are presented.

1364

Figure 8. Vertical profiles of temperature (°C), salinity, TChl*a* concentration (μ g L⁻¹) and dissolved O₂ concentration (μ mole kg⁻¹) issued from the main carousel CTD deployments (Seabird SBE 911*plus*) at stations St10–St19. The dotted lines represent the depth of sampling in the deep chlorophyll maximum (DCM) for Niskin/Go-Flo bottles. In box, the depth of the station. One or two profiles are displayed for each station. For each profile, only data acquired during the upcast are presented.

1371

Figure 9. Temperature-salinity diagram and identification of the main waters masses

1373 encountered during the MERITE-HIPPOCAMPE cruise (RAW: resident Atlantic Water;

1374 NAW: new Atlantic Water; LIW: Levantine Intermediate Water; WIW: Western

1375 Mediterranean Intermediate Water; GG: Gulf of Gabès water). The colour code refers to a)

1376 depth (in m) and b) stations (St1–St19). Twenty-one carousel CTD casts were used for this

1377 plot. Ocean Data View (ODV) software version 4.6.5. Schlitzer, R., http://odv.awi.de. 2014.

1378

Figure 10. Composite image of average surface chlorophyll *a* (Chl*a*) concentration (in μ g L⁻¹)

1380 over the period (13 April–14 May 2019) and area of the MERITE-HIPPOCAMPE cruise

1381 from Multi Satellite products (MODIS-AQUA, NOAA20-VIIRS, NPP-VIIRS and

1382 Sentinel3A-OLCI) (L4 product level, 1-km resolution; Volpe et al., 2019). This composite

image corresponds to the average of the 32 daily images (from 13 April to 14 May 2019) for

1384 which Chla concentration was averaged during the 8 h of measurements in the area. Product

- name: OCEANCOLOUR_MED_CHL_L4_REP_OBSERVATIONS_009_078. Obtained
 from EU Copernicus Marine Service Information (CMEMS; <u>https://marine.copernicus.eu/</u>).
- 1388 Figure 11. Distribution of various biogeochemical parameters recorded at the deep chlorophyll maximum (DCM) across the ten stations, from the most coastal stations in the 1389 North (St1, St4) to the southernmost stations (St17, St19): a) depth of the DCM (in m), 1390 concentrations in b) total chlorophyll a (TChla, in $\mu g L^{-1}$), c) particulate organic carbon 1391 (POC, in μ g L⁻¹), d) silicates [Si(OH)₄, in μ M], e) nitrates (NO₃⁻, in μ M), f) phosphates (PO₄³⁻ 1392 , in μ M), g) dissolved organic carbon (DOC, in μ M), and h) absorption of chromophoric 1393 dissolved organic matter at 254 nm (a_{CDOM}, in m⁻¹). Nutrients, DOC and a_{CDOM} were 1394 measured on the fraction $< 0.7 \mu m$, while TChla and POC were measured on the fraction >1395 0.7 µm from samples collected with Niskin X bottles. 1396
- 1397

Figure 12. Principal component analysis (PCA), based on the Pearson's correlation matrix, 1398 1399 applied on the main biogeochemical parameters recorded at each stations (St1-19) in the deep 1400 chlorophyll maximum (DCM): concentrations in silicates [Si(OH)₄, in μ M], nitrates (NO₃⁻, in μ M), phosphates (PO₄³⁻, in μ M), total chlorophyll *a* (TChl*a*, in μ g L⁻¹), particulate organic 1401 carbon (POC, in μ g L⁻¹), dissolved organic carbon (DOC, in μ M) and absorption of 1402 1403 chromophoric dissolved organic matter at 254 nm (a_{CDOM}, in m⁻¹). Projection of variables (i.e., 1404 main biogeochemical parameters, in red) and distribution of samples (i.e., stations, in blue) on the first factorial plane (PC1 versus PC2). Four groups of samples (stations) are highlighted 1405 1406 from this PCA and confirmed by a hierarchical ascendant classification (dissimilarity measurement between clusters based on Ward's method): 1) St2, St3, St4, St10, 2) St1, St11, 1407 1408 St15, St17, 3) St9, and 4) St19.

1409

- **Figure 13**. Overview of the types of articles that are a part of this special issue in the frame of
- 1411 the MERITE-HIPPOCAMPE cruise.

Table 1. Main characteristics of the ten stations investigated in spring during the MERITE-HIPPOCAMPE cruise (13 April–14 May 2019) on
board the R/V *Antea* along a North-South transect in the Mediterranean Sea. The stations are presented in chronological order in leg 1 then leg 2.

Ι	Leg	Station	StationLatitude (N)Longitude (E)LocationFeatures		Area	Depth (m)	Start of operations (dd/mm in 2019)	End of operations (dd/mm in 2019)		
		St2	42° 56.020'	5° 58.041'	Offshore Toulon	Limit of the continental shelf; Boundary of the Ligurian consensus region ^d ; Intermittently bloom area or bloom area (cluster #4 or #5) ^e	French	1770	14/04	16/04
		St4	43° 14.500'	5° 17.500'	Bay of Marseille (SOLEMIO ^a station)	Urbanised bay; Intermittently bloom area or bloom area (cluster #4 or #5) ^e	French	58	16/04	18/04
1	l	St3	43° 08.150'	5° 15.280'	Offshore Marseille (JULIO ^b station)	Southeast entrance to the Gulf of Lion; Intrusions of the Ligurian-Provençal current; Intermittently bloom area or bloom area (cluster #4 or #5) ^e	French	95	18/04	20/04
		St10	40° 18.632'	7° 14.753'	Offshore (station 2 of PEACETIME cruise ^c)	Slightly north of the North Balearic Front; Intermittently bloom area (cluster #4) ^e	Italian	2791	22/04	24/04
		St11	39° 07.998'	7° 41.010'	Offshore (station 3 of PEACETIME cruise ^c)	South of the North Balearic Front; Algerian consensus region ^d ; Presence of mesoscale eddies; No bloom area (cluster #3) ^e	Italian	1378	25/04	26/04

2	St15	36° 12.883'	11° 07.641'	Gulf of Hammamet	Close to the Sicily Channel; Possible entrance of Atlantic Tunisian Current branch; No bloom area (cluster #3) ^e ; High density of small pelagic fishes	Tunisian	100	29/04	30/04
	St17	34° 30.113'	11° 43.573'	North of Gulf of Gabès	Gabès consensus region ^d boundary; Shallow area, influence of tides and Atlantic Tunisian Current; Coastal bloom area (cluster #6) ^e ; High density of small pelagic fishes	Tunisian	50	01/05	02/05
	St19	33° 51.659'	11° 18.509'	South of Gulf of Gabès	Gabès consensus region ^d ; Shallow area, influence of tides and Atlantic Tunisian Current; Coastal bloom area (cluster #6) ^e ; High density of small pelagic fishes	Tunisian	50	02/05	05/05
	St9	41° 53.508'	6° 19.998'	Offshore (station 1 of PEACETIME cruise ^c)	North of the North Balearic Front; Boundary of the Ligurian consensus region ^d ; Winter convection area; Bloom area (cluster #5) ^e	French	2575	08/05	09/05
	St1	43° 03.819'	5° 59.080'	Bay of Toulon	Nearly closed urbanised bay; Intermittently bloom area or bloom area (cluster #4 or #5) ^e	French	91	10/05	11/05

^a The SOLEMIO station *(Site d'Observation Littoral pour l'Environnement du MIO)* is part of the French national network of coastal observation SOMLIT

1416 (*Service d'Observation en Milieu LITtoral* – http://somlit.epoc.u-bordeaux1.fr/fr/).

1417 ^b The JULIO station (*JUdicious Location for Intrusions Observations*) is dedicated to the study of the intrusions of the Ligurian-Provençal current.

1418 ^c The PEACETIME cruise (*ProcEss studies at the Air-sEa Interface after dust deposition in the MEditerranean sea*) took place in May-June 2017

1419 (<u>http://peacetime-project.org/;</u> Guieu et al., 2020).

1420 ^d Consensus regions of the Mediterranean Sea as defined by Ayata et al. (2018).

1421 ^e Bloom/cluster areas as defined by D'Ortenzio and d'Alcalà (2009) from SeaWiFS satellite surface TChla concentration observations.

1422

- **Table 2.** Main characteristics of the two stations investigated during the MERITE-
- 1425 HIPPOCAMPE cruise in May 2019 on board the R/V *Hannibal* in the Tunisian waters for
- 1426 trawling of small pelagic fishes.

Station	Latitude (N)	Longitude (E)	Location	Depth (m)	Date (dd/mm in 2019)	Trawling duration
T2	34° 11.892'	10° 58.152'	Gulf of Gabès	45	02/05	0h40
T4	36° 59.142'	10° 19.734'	Gulf of Tunis	37	03/05	1h30

Table 3. Information about wet (rain) atmospheric deposition samples collected on board theR/V Antea during the MERITE-HIPPOCAMPE cruise.

Sample	Station	Start date of sampling	Start time of sampling (UT)	End date of sampling (dd/mm in 2019)	End time of sampling (UT)	Type of deposition
Rain 1	St10	22/04/19	19h15	23/04	9h45	Rain
Rain 2	St10	24/04/19	7h30	24/04	12h30	Rain
Rain 3	St11	26/04/19	3h15	26/04	12h00	Rain
Rain 4	St19	03/05/19	5h45	03/05	14h00	Rain
Rain 5	Transit from off Djerba to off Zarzis	03/05/19	16h45	04/05	9h00	Dry deposition + rain
Rain 6	St1	11/05/19	3h00	11/05	6h15	Rain



Figure 1



Figure 2



Figure 3



Figure 4



NMMB/BSC-Dust 6h Acc. Dust **Dry Deposition** 06h forecast for 18 UTC 03 May 2019





NMMB/BSC-Dust 6h Acc. Dust **Wet Deposition** 06h forecast for 18 UTC 03 May 2019



Figure 5



Figure 6



Dissolved O₂ concentration (µmole kg⁻¹)

Figure 7



Figure 8



Figure 9


Figure 10







Figure 12



Figure 13