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1	Reversibility controls on extreme methane clumped isotope signatures from
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# 21 Abstract

Microbial anaerobic oxidation of methane (AOM) substantially mitigates atmospheric methane 22 emissions on Earth and is a process to consider for astrobiological targets where methane has been 23 detected. The measurement of doubly substituted, or "clumped", methane isotopes has proven 24 25 useful in tracing processes of methane formation and oxidation. Both near-equilibrium and extreme disequilibrium methane clumped isotope signatures can be attributed to AOM, but, to date, 26 27 understanding the mechanistic and environmental controls on those signatures have been lacking. 28 We report measurements of methane clumped isotope compositions of residual methane in AOM-29 active microbial incubations using sediment slurries from Svalbard and Santa Barbara Channel 30 methane seeps. Incubation experiments of Svalbard sediment slurries resulted in residual methane 31 with very high  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values up to 19.5‰ and 65.1‰, respectively. We found similarly high  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values in fluid samples from the Chamorro Seamount, a 32 33 serpentinite mud volcano in the Mariana forearc, suggesting that minimal reversibility of AOM intracellular reactions leads to kinetic fractionation of clumped isotopologues. When conditions 34 35 were consistent with a low thermodynamic drive for AOM, however, methane isotopologues approached intra-species quasi-equilibrium. This was clearly observed in isotope exchange 36 37 experiments with methyl-coenzyme M reductase (Mcr) and in microbial incubations of the Santa 38 Barbara Channel sediment slurries. Using an isotopologue fractionation model, we highlight the critical role of reversibility in controlling the trajectory of gases in  $\Delta^{13}$ CH<sub>3</sub>D vs.  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> space 39 during AOM. The near-equilibrium methane isotopologue signatures are generalized as a result of 40 41 the Mcr-catalyzed intracellular isotope exchange operating under near threshold free energy conditions, as shown in the deep-biosphere incubations. Our results show that the reversibility of 42

- 43 the Mcr-catalyzed reaction is central to understanding the meaning of methane isotopologue ratios
- 44 affected by microbial production and oxidation.
- 45
- 46 Keywords: methane isotopologues; methyl-coenzyme M reductase; kinetic fractionation;
- 47 equilibrium fractionation; isotopic bond re-ordering.

### 48 **1. Introduction**

Methane, the simplest of all hydrocarbons, is a gas with economic, societal, and 49 environmental relevance. It is the main component of natural gas, a key component of the global 50 carbon cycle (Reeburgh, 2007), and a primary target for understanding the relative role of geologic 51 52 processes and potential extinct/extant life elsewhere in the universe (e.g., Yung et al., 2018). As a 53 potent greenhouse gas, methane contributes to climate change and its concentration in the atmosphere has been increasing since the beginning of industrialization (IPCC, 2021). The main 54 55 "biological" processes forming methane are microbial degradation (microbialgenic) and pyrolysis 56 (thermogenic) of organic matter (e.g., Reeburgh, 2007; Repeta et al., 2016). Abiotic methane can 57 be formed through various types of water-rock interactions (Etiope and Sherwood Lollar, 2013) and references therein) under a range of both low temperature and high temperature conditions. 58 59 Identifying the sources and processes of methane emission is vital for constraining fluxes from major reservoirs contributing to climate change, and more broadly, for understanding 60 biogeochemical dynamics of the planet's critical zone. Traditionally, microbial and thermogenic 61 methane gases are interpreted by their carbon ( $\delta^{13}$ C) and hydrogen ( $\delta$ D) isotopic compositions 62 (e.g., Schoell, 1983; Whiticar, 1999). However, applying  $\delta^{13}$ C and  $\delta$ D as a diagnostic framework 63 64 alone has significant limitations, most notably circumstances in which isotopic signatures of 65 methane sources are not sharply demarcated, exhibit some degree of overlap, and such empirical 66 frameworks are inevitably redefined as additional experimental and field data come to light (Sherwood Lollar et al., 2006; Douglas et al., 2017). In addition, empirical fields in  $\delta^{13}$ C vs.  $\delta$ D 67 68 space are inherently linked to the isotopic signature of the source materials coupled with the formation processes, which in turn are strongly affected by mixing and secondary alteration 69 processes (Etiope and Sherwood Lollar, 2013 and references therein). 70

71 Recently, sophisticated instrumentation has become available to resolve the abundances of doubly substituted, "clumped", isotopologues of methane gas, including <sup>13</sup>CH<sub>3</sub>D and <sup>12</sup>CH<sub>2</sub>D<sub>2</sub> 72 (Ono et al., 2014; Stolper et al., 2014b; Young et al., 2016; Eldridge et al., 2019; Gonzalez et al., 73 74 2019; Dong et al., 2021; Zhang et al., 2021). The relative abundances of these two mass-18 methane isotopologues are expressed as  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values, representing deviations 75 76 from a stochastic standard in which distributions of isotopes across all isotopologues are effectively random. The latter is the expected state at sufficiently high temperatures (>1000 K). 77 78 The relative abundances of doubly substituted methane isotopologues differs from the stochastic distribution at lower temperatures due to the enhanced thermodynamic stability of bonds involving 79 the heavier isotopes (e.g., Ma et al., 2008; Webb and Miller, 2014; Liu and Liu, 2016). In principle, 80 81 isotopic bond ordering, or clumping, is a tracer of the processes attending the formation of the molecules, coupled with any consumption and/or re-equilibration processes, and therefore gives 82 the potential for their application to discern processes of methane formation and oxidation both on 83 Earth and beyond (Young, 2019). The utility of  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> lies in the fact that the 84 85 values should be sensitive to process, and interpretations of their significance may not dependent 86 upon long-lost isotope exchange partners (e.g., water). In practice, kinetically controlled methane 87 clumped isotope compositions can be sensitive to hydrogen sources of methane that differ in  $\delta D$ 88 (Taenzer et al., 2020).

In some cases,  $\Delta^{13}$ CH<sub>3</sub>D has been used to reconstruct the formation temperatures of thermogenic methane (Stolper et al., 2014a; Stolper et al., 2015; Wang et al., 2015; Douglas et al., 2016; Lalk et al., 2022) and the combination of  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> has been used to verify that the molecules record equilibrium at a given temperature (Young et al., 2017; Giunta et al., 2019; Gonzalez et al., 2019; Thiagarajan et al., 2020; Xie et al., 2021). The abundances of mass-

94 18 isotopologues of microbial methane, on the other hand, have been shown to be inconsistent with thermodynamic equilibrium, meaning that their distribution does not reflect the temperature 95 at which the methane was formed (Stolper et al., 2015; Wang et al., 2015; Young et al., 2017; Ash 96 et al., 2019; Giunta et al., 2019; Warr et al., 2021). The  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> of microbial 97 methane can be modeled as the result of the combination of equilibrium and kinetic steps during 98 the enzymatically facilitated formation of methane (Cao et al., 2019; Young, 2019; Gropp et al., 99 2021, 2022; Ono et al., 2022). In particular, the extremely negative  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values are most 100 likely due to the combinatorial effect of accessing multiple sources of hydrogen with significantly 101 different hydrogen isotopic signatures during the formation of methane (Röckmann et al., 2016; 102 103 Yeung, 2016; Young, 2019; Taenzer et al., 2020).

104 While the ability to define a methane isotopologue signature related to microbial 105 methanogenesis is promising, there are caveats. One is that abiotic methane and low-maturity thermogenic methane has been found with deficits in  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> compared with equilibrium values 106 107 in methane gas, which is further confirmed by pyrolysis experiments on n-octadecane (Douglas et al., 2017; Young et al., 2017; Dong et al., 2021; Warr et al., 2021; Xie et al., 2021). On the other 108 hand, abiotic or biotic consumption processes could potentially alter the relative abundances of the 109 110 mass-18 isotopologues in ways that might mimic the putative microbial methanogenesis signature. 111 Laboratory experiments and *ab initio* modeling indicate that methane oxidation in the atmosphere by OH or Cl radicals is accompanied by large shifts in methane clumping down to very negative 112  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values due to Rayleigh distillation (Haghnegahdar et al., 2017; Whitehill 113 et al., 2017; Young, 2019). Similarly, both  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values of residual methane in 114 microbial culture were observed to decrease during aerobic oxidation of methane (Wang et al., 115 116 2016; Krause et al., 2022).

117 Photocatalytic oxidation and aerobic microbial oxidation of methane occur in the atmosphere and oxygenated environments. Therefore, these processes are in principle separated 118 119 spatially from microbial methanogenesis, which operates almost exclusively under anoxic, and thus reducing, conditions. Anaerobic oxidation of methane (AOM), however, is closely related to 120 methanogenesis both spatially and metabolically. For example, concurrent methanogenesis and 121 122 AOM have been found in near-surface sediments and at the sulfate-methane transition (Xiao et al., 123 2017; Beulig et al., 2019; Krause and Treude, 2021). Anaerobic methanotrophic (ANME) archaea 124 are close relatives of methanogenic archaea and have the full enzymatic machinery of methanogenesis working in reverse (Timmers et al., 2017). Methanogens belonging to 125 Methanosarcina can reverse key reactions of methanogenesis coupled to ferric iron reduction (Yan 126 127 et al., 2018; Yu et al., 2022). The prospect of a methanotroph (ANME-1) from natural sediments conducting methanogenesis further underscores the potential versatility of methanoarchaeal 128 enzymes (Beulig et al., 2019; Kevorkian et al., 2021). Factors forcing these enzymes to operate in 129 130 either direction need further investigation. However, even when operating in full AOM mode with 131 the highest net turnover of methane, ANME archaea demonstrate some fraction (~4-15%) of net 132 reversibility, defined as the ratio of the backward to forward fluxes (Treude et al., 2007; Holler et 133 al., 2011; Wegener et al., 2021). Back flux of material during the enzymatic reactions of AOM has implications for isotope fractionation, leading to carbon isotope  $({}^{13}C/{}^{12}C)$  equilibration between 134 135 methane and intracellular metabolites, in particular when AOM is operating close to 136 thermodynamic limitations where the methyl-coenzyme M reductase (Mcr)-catalyzed reaction is 137 near equilibrium (Hoehler et al., 2000; Holler et al., 2011; Yoshinaga et al., 2014; Chuang et al., 138 2019; Wegener et al., 2021).

The precise mechanisms that are responsible for  $\Delta^{13}$ CH<sub>3</sub>D vs.  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> effects of AOM 139 140 are largely unknown, and further laboratory incubation experiments under different environmental conditions are required. The overarching goal is to develop these intra-methane isotopologue 141 signals as tracers for the origin and processing of methane in general, and AOM in particular. One 142 of the biggest challenges for understanding the isotopologue composition of natural samples is 143 144 limited knowledge of how isotopic bond re-ordering caused by AOM can overwrite formation-145 process signatures. Given the high level of reversibility of Mcr-catalyzed reactions during AOM, 146 Ash et al. (2019), Giunta et al. (2019), and Warr et al. (2021) suggested that AOM could be responsible for thermodynamic isotopic bond-order equilibrium in methane found in sulfate-147 depleted Baltic Sea sediments, sedimentary basins, and the deep subsurface, respectively. Recently, 148 149 Ono et al. (2021) found that the methane clumped isotopologue <sup>13</sup>CH<sub>3</sub>D exhibits kinetic fractionation in an ANME-1 sediment-free enrichment culture with a high sulfate concentration 150 (28 mM), where  $\Delta^{13}$ CH<sub>3</sub>D values increased by up to 5.8‰. Importantly, Wegener et al. (2021) 151 152 demonstrated a sulfate dependence of the net bulk isotope fractionation by virtue of the 153 thermodynamic drive of the intracellular reactions of AOM. Based on the earlier studies, therefore, 154 it appears that the reversibility of intracellular reactions of AOM is responsible for the observed 155 discrepancy between the kinetic isotopologue fractionation in a sulfate-replete culture (Ono et al., 156 2021) and the intra-species isotope equilibrium in sulfate-depleted settings (Ash et al., 2019; 157 Giunta et al., 2019; Warr et al., 2021).

To document the range of effects that AOM may have on modifying methane isotopologue signatures, we collected a suite of marine sediments and fracture fluids from diverse marine and terrestrial settings that showed different degrees of ANME abundance and AOM activity (Fig. 1). We performed microbial incubations using these samples to evaluate the relationship between the 162 reversibility of intracellular reactions of AOM and the methane isotopologue fractionation/reordering, and to elucidate the unknown fractionation factors for <sup>12</sup>CH<sub>2</sub>D<sub>2</sub>/<sup>12</sup>CH<sub>4</sub>. We also collected 163 natural fluids from the South Chamorro Seamount where AOM is known to substantially consume 164 abiotic source methane (Wheat et al., 2020), in order to examine whether the isotopologue 165 166 fractionation differs between laboratory incubations and endogenous AOM in natural 167 environments. In combination, our work aims to address how environmental conditions can 168 potentially affect isotopologue fractionation, as well as rates and magnitudes of isotopic bond re-169 ordering during AOM. This information will improve our ability to distinguish primary signatures of methane formation from biologically-driven overprinting to maximize the utility of 170  $^{13}$ CH<sub>3</sub>D/ $^{12}$ CH<sub>4</sub> and  $^{12}$ CH<sub>2</sub>D<sub>2</sub>/ $^{12}$ CH<sub>4</sub> as tracers of methane formation and subsequent processing. 171

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#### 173 2. MATERIALS AND METHODS

#### 174 **2.1. Incubation of methane seep sediment slurry from Svalbard**

Marine sediments were collected in summer 2011 with a video-guided multicorer from 175 176 active methane seeps covered by a dense sulfur-bacteria mat offshore northwestern Svalbard 177 during the R/V Poseidon cruise 419 (Site MUC 12; 79°00.417' N, 06°54.131' E, 1235 m water 178 depth). Details of the sampling site, including ex-situ activity of AOM (Fig. 2A) and sulfate 179 reduction and geochemical parameters in replicate sediment cores, can be found in Melaniuk et al. 180 (2022). Sediment used for the present study was collected from the top 10 cm of a replicate multicorer core (10 cm inner diameter) and filled headspace-free into 100- and 250-ml borosilicate 181 182 bottles with butyl stoppers. The sediment was stored anoxically at 4 °C in the dark for 10 years without methane or sulfate addition. The AOM community in the sediment was reactivated in the 183

laboratory at UCLA in April 2020 by mixing with anoxic artificial seawater medium at a ratio of
1:1 (v:v) (Widdel and Bak, 1992; Laso-Pérez et al., 2018) and a methane gas headspace as the sole
amended carbon source in a 2-L glass bottle. The cultivation procedures of Laso-Pérez et al. (2018)
were followed.

188 We performed a set of sulfide analyses to confirm the activity of AOM before starting the 189 actual experiments. Sulfide concentration was measured after Cord-Ruwisch (1985) using a 190 Shimadzu UV-Spectrophotometer (UV-1800). The relative standard deviation (RSD) of sulfide analysis was better than 5%. Sulfide concentration in the supernatant started to increase two 191 192 months after addition of substrates (sulfate and methane). Further, sulfide concentration increased ca. 0.07 mM per day between the 134<sup>th</sup> and 205<sup>th</sup> days since reactivation. The supernatant of the 193 194 sediment slurry was replaced with fresh anoxic medium with seawater sulfate concentration (28 195 mM) when sulfide concentration approached 11 mM. After that, sulfide concentration continued to increase ca. 0.07 mM per day between the 206<sup>th</sup> and 283<sup>rd</sup> days since reactivation. Sulfide 196 197 production from sulfate reduction with non-methane substrates was excluded by control 198 experiment (see below). The linear increase in sulfide concentration over time indicates that the ANME community was successfully reactivated in the sediment slurry. Thus, the homogenized 199 200 slurry was equally distributed to two 600-ml culture bottles in replicate and filled up completely 201 with an anoxic artificial seawater medium (Laso-Pérez et al., 2018). Sixty ml of the medium 202 volume was replaced with ca. 200 kPa of methane (Airgas), and both incubations were performed under excess sulfate (ca. 10 mmol) over methane (5.4 mmol) (cf. Ono et al., 2021). The sediment 203 slurry was then stored horizontally at 4 °C in the dark. Methane isotopologue compositions were 204 analyzed on days 1, 36, 44, 58, and 71 of the incubation. After the first experiment, we reset the 205

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slurry for longer incubation through medium replacement and bubbling with  $N_2/CO_2$  gas. Methane isotopologue compositions were analyzed on days 1, 60, 90, and 168 of the incubation.

To further confirm and quantify the activity of AOM and sulfate reduction, we measured 208 turnover rates with <sup>14</sup>C- and <sup>35</sup>S-radiotracer techniques, respectively, in the high-sulfate experiment. 209 AOM rate was determined by injecting 20 µL of <sup>14</sup>C-methane tracer (dissolved in MilliQ water, 210 activity 0.6 kBg, specific activity 1.85 GBg mmol<sup>-1</sup>) to two 13 ml headspace-free sub-samples of 211 the sediment slurry. Sulfate reduction rate was determined by injecting 10 µL of carrier-free <sup>35</sup>S-212 sulfate tracer (dissolved in MilliQ water, activity 107 kBq, specific activity 1.59 TBq mg<sup>-1</sup>) to a 213 214 10 ml sub-sample of the sediment slurry. The vials were incubated for two days at 4 °C in the dark. 215 After incubation, sulfate reduction activity was terminated by transferring the sample from the glass vial to a centrifuge tube filled with 20 ml 20% zinc acetate followed by freezing at  $-30^{\circ}$ C. 216 217 Sulfate reduction rate was analyzed and calculated according to the cold chromium distillation method (Kallmeyer et al., 2004). To terminate AOM activity, samples were transferred to 50 ml 218 219 glass vials filled with 20 ml 5% sodium hydroxide. The vials were sealed with rubber stoppers 220 immediately and shaken thoroughly. AOM rate was determined according to Treude et al. (2005) 221 and Joye et al. (2004).

After the high-sulfate experiment described above, the supernatant in the slurry was replaced with low-sulfate artificial seawater medium (1 mM) four times. The sulfate concentration in the slurry decreased over time due to medium replacement and microbial consumption. The produced sulfide was maintained at low concentration (<1 mM) through medium replacement and bubbling with  $N_2/CO_2$  gas. The culture bottle was then filled up completely with the low-sulfate medium, reaching a sulfate concentration of 0.4 mM in the slurry. Sixty ml of the medium volume was replaced with ca. 200 kPa of methane (Airgas), allowing the incubation to be performed under 229 excess methane (5.4 mmol) over sulfate (<0.2 mmol). The low-sulfate experiment was performed 230 identically to the high-sulfate experiment with sulfate concentration as the only variable at the beginning of each incubation. Towards the end of the low-sulfate experiment, sulfate was depleted 231 (0.01 mM) in the slurry. Thus, additional concentrated anoxic sulfate solution (2.5 ml) was added 232 to the culture bottle by a syringe without opening the bottle, reaching a final sulfate concentration 233 234 of 0.4 mM again in the slurry. Sulfate concentration was analyzed by ion chromatography (Metrohm 761). Alkalinity was determined by acid titration with a Metrohm 876 Dosimat Plus 235 236 (Dale et al., 2015). The concentrations were calibrated against the IAPSO standard seawater (n=10) with RSD of sulfate and alkalinity analyses better than 2%. In addition, pH was determined by a 237 pH meter (VWR sympHony B10P). 238

239 Methane concentration and isotopologue abundance were determined during the 240 incubations on a vacuum line interfaced with a gas chromatograph, and the Panorama mass spectrometer, respectively, as described below in section 2.6. Given the known volume and 241 porosity of the slurry, the amounts of methane that dissolved in the liquid phase were calculated 242 243 using Henry's law and the Bunsen solubility coefficient (Yamamoto et al., 1976). For porosity analysis, homogenized slurry was sampled using a syringe. Porosity was measured from the 244 245 volume of slurry and water weight loss after drying at 60 °C until a constant weight was reached. 246 The total amounts of methane in the culture bottle were therefore the sum of methane in the headspace and the liquid phase. 247

The liquid phase was also sampled to determine the  $\delta^{13}$ C of dissolved inorganic carbon (DIC) and the  $\delta$ D of water in the slurry (Atekwana and Krishnamurthy, 1998; Kopec et al., 2019). In brief, analysis of  $\delta^{13}$ C<sub>DIC</sub> was performed in 12 ml Labco Exetainer vials on a Thermo Scientific GasBench II coupled to a Thermo Finnigan Delta Plus XL isotope-ratio mass spectrometer (IRMS) 252 at the UC Davis Stable Isotope Facility. Isotopic values are reported in  $\delta$ -notation relative to the Vienna Pee Dee Belemnite (VPDB) standard. Analytical precision for  $\delta^{13}$ C is better than 0.1‰ 253  $(1\sigma)$  based on replicate analyses of laboratory standards. Analysis of hydrogen isotopic 254 255 composition of water was conducted at the Stable Isotope Laboratory at Dartmouth College. The  $\delta D$  values of water were measured using an H-Device, in which water was reduced by hot 256 257 chromium (850 °C), and the resulting hydrogen gas was measured by a Thermo Delta Plus XL 258 IRMS. Isotopic ratios are reported in  $\delta$ -notation relative to the Standard Mean Ocean Water 259 (VSMOW) standard. Analytical precision for  $\delta D$  is better than 0.5‰ (1 $\sigma$ ) based on replicate analyses of laboratory standards. 260

A control incubation was set up to assess potential microbial methanogenesis and 261 262 organoclastic sulfate reduction in the same sediment slurry. The control experiment was prepared by replacing the supernatant with a new methane-free anoxic artificial seawater medium and a 60-263 ml headspace was left to monitor methane concentration. The slurry was then bubbled thoroughly 264 265 with  $N_2/CO_2$  (80:20) three times (3 hours in total) to get rid of the residual dissolved methane and 266 sulfide from the previous experiments. The slurry was stirred during and between flushing to drive 267 the dissolved methane into the headspace. The control experiment slurry was stored in the dark at 4 °C for three months. The slurry supernatant was collected for sulfide and sulfate concentrations 268 269 at the start and end of the control incubation. Methane concentration in the gas phase was measured 270 by gas chromatography throughout the three-month period.

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# 272 **2.2.** Incubation of methane seep sediment slurries from the Santa Barbara Channel

273 Marine sediments were collected with four push cores from the Coal Oil Point seep field
274 (Rostocker Seep and Isla Vista Super Seep) in the Santa Barbara Channel in 2017 (Jordan et al.,

275 2020). The top 12.5 cm sediment of the cores was transferred to 300 ml culture bottles at UCLA following the same procedure as detailed in the Svalbard slurry section above. Low sulfate (< 1 276 mM) and high sulfate (> 28 mM) concentrations were achieved in the sediment slurry by either 277 dilution steps with anoxic artificial seawater (without sulfate) or additions of concentrated sodium 278 279 sulfate solution (100 mM). The sediment slurry was incubated in the dark with a 4.4 ml headspace 280 of methane at ca. 100 kPa and 20 °C on a shaker. The methane gas bubble from the sediment slurry was extracted at the end of each experiment and geochemistry analyses were performed as 281 282 described in the Svalbard slurry section. A similar control experiment followed the Svalbard slurry 283 section with a 4.4 ml Argon headspace.

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# 285 2.3. Incubations of sediment slurry and fracture fluid from the deep biosphere (Nankai 286 Trough and Beatrix Gold Mine)

287 Hole C0023A (32°22.0018' N, 134°57.9844' E), located in the Nankai Trough on the 288 subduction boundary between the Philippine Sea and Eurasian plates, was drilled during 289 International Ocean Discovery Program (IODP) Expedition 370 in 2016 (Heuer et al., 2017). 290 Marine sediments from 257 m below seafloor (mbsf) were sampled anoxically with rigorous 291 contamination control (see Heuer et al., 2017) and utilized for sediment incubation. In brief, whole-292 round cores (WRCs) were prepared from recovered sections under super-clean and anoxic 293 conditions onboard the Chikyu Drilling Vessel. X-ray computed tomography was performed on all 294 WRCs for lithological identification and to assess core quality on board. Samples without drilling 295 disturbances were designated for incubation and were subjected to secondary scraping to minimize 296 the potential for contamination.

297 At South Africa's Beatrix Gold Mine (28 °14.1' S, 26°47.7' E), fracture fluid was collected 298 in 2016 from the BE326-BH2 borehole, from 1390 m below land surface (mbls) in shaft 3, level 26. A detailed description of the study site and sample collection has been published in Lau et al. 299 (2016). Briefly, fracture fluid was filtered using a 0.2 µm hollow fiber MediaKap<sup>®</sup>-10 filter 300 (Spectrum Labs, New Brunswick, NJ USA). The microorganisms on the filter were anoxically 301 back-flushed into sterile, capped, N<sub>2</sub>-sparged 160-ml borosilicate serum vials to a final cell 302 concentration of  $\sim 10^7$  cells ml<sup>-1</sup>. Samples were stored at 4 °C upon return to the surface and used 303 304 for incubation.

305 Incubations were set up inside a clean anoxic glove bag (Coy Laboratory Products, Grass 306 Lake, MI USA) at Princeton University. Sterile aluminum foil was placed on the working surface 307 of the glove bag and sterile nitrile gloves were used over the glove bag's butyl rubber gloves to 308 minimize potential contamination of low-biomass samples. The surface of C0023A samples was scraped using a sterile scalpel to remove sediment exposed to gas-tight packaging. A total of 10 g 309 310 (wet weight) of interior sediment was weighed out from sample source core C0023A-5F02 (top 311 depth 257.3 mbsf) and transferred into combusted 160-ml borosilicate serum vials containing 100 312 ml artificial sulfate-free seawater medium modified from the recipe by Widdel and Bak (1992). 313 For the BE326-BH2 incubation, 10 ml of fracture fluid was added to 90 ml artificial sulfate-free 314 seawater medium. Serum vials were sealed with 0.1 N NaOH-boiled butyl rubber stoppers (Bellco 315 Glass, Inc., Vineland, NJ USA) and aluminum crimps (Supelco Inc., Bellefonte, PA USA), and the headspace was subsequently replaced with 100% methane. Each incubation was supplemented 316 with 10 mM of one of the following electron acceptors:  $NO_3^-$ ,  $NO_2^-$ ,  $SO_4^{2-}$ , or  $Fe^{3+}$  (in the form 317 of hydrous ferric oxide). A control without added electron acceptors was included to assess 318 319 endogenous microbial activity from remnant electron acceptors available in the inoculum, and an autoclaved sample was included as a killed control. Serum vials containing sediment slurry or
fracture fluid were incubated upside-down at an approximate *in-situ* temperature of 40 °C. At
UCLA, the collected residual methane was purified for isotopologue analysis following the method
described in section 2.6.

324 Electron acceptor depletion was monitored using Dionex IC25 ion chromatography 325 coupled to an MSQ-quadruple mass spectrometer (Thermo Scientific, Waltham, MA USA). The sediment slurry or fracture fluid was also subsampled to monitor the  $\delta^{13}$ C of DIC. Briefly, 500 µl 326 aliquots were anoxically transferred to combusted, amber borosilicate serum vials treated with 327 328 saturated HgCl<sub>2</sub> and aluminum-crimped sealed with butyl rubber stoppers. Following sample 329 transfer, vials were over-pressurized with ultra-high purity  $N_2$  gas, supplemented with 0.5 N H<sub>3</sub>PO<sub>4</sub>, and heated overnight in a water bath at 70 °C to extract all DIC out of solution. Isotopic 330 331 composition of the headspace was analyzed using a Picarro cavity ring-down spectrometer equipped with a G2101-I Isotopic CO<sub>2</sub> analyzer (Picarro, Inc., Sunnyvale, CA USA) in CO<sub>2</sub> focus 332 333 mode.

334

335 2.4. Methyl-coenzyme M reductase (Mcr) experiments

The experiment was performed as described previously with the Mcr enzyme purified from *Methanothermobacter marburgensis* (Mahlert et al., 2002; Scheller et al., 2010) at Pennsylvania State University. Mcr that was used for this assay had absorbance maxima at 387 nm, corresponding to the enzyme's active (Ni<sup>+</sup>) form. Coenzyme B (HS-CoB) was prepared from the symmetric disulfide CoB-S-S-CoB by reaction with NaBH<sub>4</sub>. Methyl-coenzyme M (CH<sub>3</sub>-S-CoM) was synthesized from coenzyme M (sodium salt) by methylation with methyl iodide. The complete reaction mixture (4 ml) contained 5 mM CH<sub>3</sub>-S-CoM, 2.5 mM HS-CoB and 32.3 mg of Mcr in 50 mM phosphate buffer (pH 7.6). This allowed isotope exchange of the methyl moiety in methylcoenzyme M and methane to occur. The reaction mixtures were incubated at 20 °C for 48 hours or 60 °C for 2–6 hours in a stoppered 10-ml serum vial with a methane headspace of ca. 100 kPa. The difference in incubation times is due to more rapid reaction at 60 °C. Control reactions contained the complete reaction mixture minus enzyme. At UCLA, the collected gas was purified for isotopologue analysis following the method detailed in section 2.6.

349

# 350 **2.5.** Methane-bearing natural fluids at the South Chamorro Seamount

351 South Chamorro Seamount is an active serpentinite mud volcano at ca. 3150 m water depth 352 in the Mariana forearc. In 2001, Hole 1200C was drilled to 266 mbsf and cased during Ocean Drilling Program (ODP) Leg 195 in the summit knoll of South Chamorro Seamount (Fryer and 353 354 Salisbury, 2006). An oceanic borehole observatory, commonly called a CORK (Circulation 355 Obviation Retrofit Kit), was deployed for subsequent fluid sampling (Wheat et al., 2008). The 356 discharged fluids were largely altered relative to seawater composition, displaying a high pH up 357 to 12.3 with abundant dissolved methane up to 33 mM (Wheat et al., 2008; Wheat et al., 2020). In 358 January 2009, pristine crustal fluids were collected directly as they discharged at Hole 1200C with 359 a remotely operated vehicle (ROV) HyperDolphin (HPD Dives 941–947) during Cruise NT09-01 360 (Wheat et al., 2020). Isobaric gas-tight samplers (Seewald et al., 2002) were used for fluid 361 sampling. Immediately upon recovery of the ROV, fluid samples were subsampled for liquid and gas analyses. Gas extraction from fluid samples was conducted using previously reported 362 techniques (Seewald et al., 2002). At UCLA, the collected gas was purified for isotopologue 363 364 analysis following the method detailed in the next section.

365

# 366 2.6. Doubly substituted isotopologue measurements and isotope notation

Methane isotopologue abundances of methane gas samples were measured using the 367 368 Panorama (Nu Instruments) high-mass-resolution gas-source isotope ratio mass spectrometer 369 housed at UCLA. Details surrounding the purification and measurement of methane gas were 370 previously published (Young et al., 2016; Young et al., 2017) and are briefly summarized here. 371 Methane sample gases were purified on a vacuum line interfaced with a gas chromatograph (GC). 372 Samples were delivered to the vacuum line through a septum by a gas-tight syringe and trapped 373 on silica gel at liquid nitrogen temperature. The Helium carrier gas was then used to flush the sample to the GC. Separation was accomplished with a 3-meter 1/8-inch OD stainless steel column 374 375 packed with 5 Å molecular sieve, followed in series by a 2-meter 1/8-inch OD stainless steel 376 column packed with HayeSep D porous polymer. Peaks were identified using an in-line, passive 377 thermal conductivity detector (TCD). Once methane collection was complete, the sample was 378 transferred to an evacuated sample tube filled with silica gel at liquid nitrogen temperature. 379 Methane in this tube was introduced to the inlet of the mass spectrometer where it was warmed to 380 60 °C and expanded into the bellow of the instrument.

The Panorama mass spectrometer was set to a mass resolving power of ~40,000 or greater, allowing the measurement of ion currents for resolved  ${}^{12}CH_4^+$ ,  ${}^{13}CH_4^+$ ,  ${}^{12}CH_3D^+$ ,  ${}^{13}CH_3D^+$ , and  ${}^{12}CH_2D_2^+$ . Isotopologues of masses 16 and 17 were measured using Faraday collectors with amplifier resistors of 10<sup>11</sup>  $\Omega$ . Both doubly substituted mass-18 isotopologues,  ${}^{13}CH_3D^+$  and  ${}^{12}CH_2D_2^+$ , were measured with an electron multiplier as the axial collector. The measured ratios of these ion currents yield values for bulk  ${}^{13}C/{}^{12}C$  and D/H as well as for both  $\Delta^{13}CH_3D$  and  $\Delta^{12}CH_2D_2$ . The isotopic compositions of carbon and hydrogen are reported as deviations from the carbon and hydrogen reference materials VPDB and VSMOW. Standard delta notation is used to
express the fractional differences in per mil units:

390 
$$\delta^{13}C = [({}^{13}C/{}^{12}C)_{\text{sample}}/({}^{13}C/{}^{12}C)_{\text{VPDB}} - 1] \times 1000$$
(1)

391 
$$\delta D = [(D/H)_{sample}/(D/H)_{VSMOW} - 1] \times 1000$$
(2)

392 The relative abundances of the two mass-18 isotopologues of methane are reported relative 393 to the stochastic reference frame expressed in per mil using the capital delta notation:

394 
$$\Delta^{13}CH_3D = \left[ ({}^{13}CH_3D / {}^{12}CH_4)_{\text{sample}} / ({}^{13}CH_3D / {}^{12}CH_4)_{\text{stochastic}} - 1 \right] \times 1000$$
(3)

395 
$$\Delta^{12}CH_2D_2 = [({}^{12}CH_2D_2/{}^{12}CH_4)_{sample}/({}^{12}CH_2D_2/{}^{12}CH_4)_{stochastic} - 1] \times 1000$$
(4)

Analytical uncertainties are estimated from the long-term reproducibility of the thermogenic Utica Shale gas through the same purification process (n=22). External precision for  $\delta^{13}$ C,  $\delta$ D,  $\Delta^{13}$ CH<sub>3</sub>D, and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> is found to be approximately 0.1‰, 0.3‰, 0.3‰ and 0.7‰, respectively (1 $\sigma$ ). The relationship between temperature and both  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> has been predicted through *ab initio* calculations based on the harmonic approximation and can be expressed by the following equations (Young et al., 2017):

402 
$$\Delta^{13}$$
CH<sub>3</sub>D (T)  $\approx 1000 \ln(1 + 0.0355502/T - 433.038/T^2 + 1270210.0/T^3 - 5.94804$ 

$$\times 10^{8}/T^{4} + 1.196630 \times 10^{11}/T^{5} - 9.07230 \times 10^{12}/T^{6})$$
<sup>(5)</sup>

404 
$$\Delta^{12}$$
CH<sub>2</sub>D<sub>2</sub> (T)  $\approx 1000 \ln(1 + 0.183798/T - 785.483/T^2 + 1056280.0/T^3 + 9.37307)$ 

405 
$$\times 10^{7}/T^{4} - 8.919480 \times 10^{10}/T^{5} + 9.901730 \times 10^{12}/T^{6})$$
 (6)

406 where T is in Kelvin. Eqs. (5) and (6) show that  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values are both positive 407 when methane is formed at thermodynamic equilibrium, and approach 0‰ at high temperatures 408 (>1000 K). 409

# 410 **2.7. Calculation of isotope fractionation factors using Rayleigh equation**

Incubations were performed in glass bottles with frequent shaking and the headspace methane was considered a well-mixed source in a closed system. Although methane in the headspace was extracted once or twice between the start and end of each experiment, the amount of methane extracted was minor (<3%) compared to the amount of methane in the whole bottle, resulting in a negligible deviation from a strictly closed system. Thus, the Rayleigh fractionation equation (Mariotti et al., 1981) was used to extract fractionation factors for the  ${}^{13}$ CH<sub>4</sub>,  ${}^{12}$ CH<sub>3</sub>D, and  ${}^{12}$ CH<sub>2</sub>D<sub>2</sub> isotopologues relative to  ${}^{12}$ CH<sub>4</sub> resulting from anaerobic fractionation:

418 
$$\frac{R_t}{R_0} = \left(\frac{\left[{}^{12}CH_4\right]_t}{\left[{}^{12}CH_4\right]_0}\right)^{\alpha-1}$$
(7)

where R refers to the ratio of isotopologues of interest in the gas phase (e.g.,  ${}^{13}CH_{3}D / {}^{12}CH_{4}$ , 419  $^{12}$ CH<sub>2</sub>D<sub>2</sub>/ $^{12}$ CH<sub>4</sub>), the subscript  $\theta$  signifies the initial property, and the ratio of methane abundance 420 at time t relative to the initial methane abundance is commonly referred to as F (fraction remaining). 421 For kinetic fractionations, the fractionation factor  $\alpha$  is simply the ratio of the rate constant 422 423 associated with the reaction of an isotopically substituted isotopologue relative to the rate constant of the unsubstituted isotopologue. These can include ratios of symmetry numbers. For the sake of 424 simplification, the isotopologue fractionation factor for <sup>13</sup>CH<sub>3</sub>D and <sup>12</sup>CH<sub>2</sub>D<sub>2</sub> relative to <sup>12</sup>CH<sub>4</sub> are 425 termed <sup>13D</sup> $\alpha$  and <sup>D2</sup> $\alpha$ , respectively. By convention,  $\alpha < 1$  signifies that the reaction favors the 426 isotopically light species, leaving the residue enriched in the heavy isotope or isotopologue. On a 427 plot of  $-\ln F$  vs.  $\ln(R/R_0)$ , the slope (often termed  $\varepsilon$ ) provides the fractionation factor ( $^{13}\varepsilon = 1 - ^{13}\alpha$ 428 or  ${}^{D}\epsilon = 1 - {}^{D}\alpha$ ). Fractionations and their uncertainties (95% confidence interval) are calculated by 429

430 the weighted least square method (York et al., 2004). Errors for methane concentrations are 431 estimated to be 0.4% of the measured value. Note that the *F* ratio was only measured in the 432 incubation experiment of the Svalbard methane seep sediments.

At zeroth order, the relationship among isotopologue fractionation factors would follow 433 434 the rule of the geometric mean (Bigeleisen, 1955), in which a stochastic distribution of isotopes occurs among the isotopologues. This would be the case where there is no energetic preference for 435 forming doubly substituted isotopologues as opposed to randomly distributing isotopes among 436 bonds (e.g.,  ${}^{13D}\alpha \approx {}^{13}\alpha \cdot {}^{D}\alpha$ ). Under these circumstances, the fractionation factors for the doubly 437 438 substituted species are simply the products of the fractionation factors for the individual isotopes 439 (i.e., the square of the geometric mean of the fractionation factors for doubly substituted species). The deviation from the rule of the geometric mean can be characterized by the kinetic clumped 440 isotopologue fractionation factor,  $\gamma$ , yielding (cf. Wang et al., 2016): 441

442 
$${}^{13\mathrm{D}}\alpha = {}^{13\mathrm{D}}\gamma \cdot {}^{13}\alpha \cdot {}^{\mathrm{D}}\alpha \tag{8}$$

443 
$${}^{D2}\alpha = {}^{D2}\gamma \cdot {}^{D}\alpha \cdot {}^{D}\alpha \tag{9}$$

444

# 445 **2.8.** Open system model — steady-state between transport and oxidation of moving methane

In natural environments, Rayleigh-type methane oxidation is rarely observed because replenishment of methane, facilitated by advection or diffusion, usually applies. We will consider two end-member environmental scenarios: a steady-state open system model presented in this section and a time-dependent closed system model with a methane source and sink in the next section. In the open system model, methane is transported into and out of the system via advection with concurrent AOM. A steady state between oxidation and supply is assumed to have been attained. A mass balance equation at steady state can be expressed as (Hayes, 2001; Wang et al., 2016):

$$455 R_0 = (1 - \varphi) R_{\rm SS} + R_{\rm SS} \varphi \alpha (10)$$

456 where  $R_0$  is the initial isotopologue ratio,  $R_{SS}$  is the steady-state value,  $\varphi$  is the fraction of methane removed via oxidation relative to that which flows through, and  $\alpha$  is the kinetic fractionation factor 457 associated with the oxidation reaction. Such a steady state will occur where the supply and sink 458 occur at constant rates. Here, advection is assumed to have no isotope effect (Alperin et al., 1988), 459 whereas AOM has associated fractionation factors  ${}^{13}\alpha$ ,  ${}^{D}\alpha$ ,  ${}^{13D}\gamma$ , and  ${}^{D2}\gamma$ . The fractional 460 contribution of AOM to the total sink,  $\varphi$ , is physically related to the Damköhler number, i.e., Da, 461 the ratio of the residence time in the flow system to the characteristic reaction time (Da =  $\tau_{flow}$  / 462  $\tau_{reaction}$ ). For a first-order reaction, the dimensionless Damköhler number can be simplified to  $\tau_{flow}$ 463 × k where  $\tau_{flow}$  is the residence time and k is the reaction rate constant with units of inverse time. 464 The  $\varphi$  values are normalized Da values, as in  $\varphi = Da/(1+Da)$ . Therefore, if k is large in comparison 465 to  $\tau$ , Da is large and  $\varphi$  approaches 1. At this extreme, a balance between production and oxidation 466 467 is achieved in a closed system as described in the next section. Conversely, if k is small in 468 comparison to  $\tau$ ,  $\varphi$  approaches 0 and the isotopologue ratio does not change. Values in between 469 these extrema cause the steady-state values to be intermediate between the initial isotopologue 470 ratios and the values at  $\varphi = 1$ .

471

## 472 **2.9.** Closed system model — formation of methane balanced against oxidation

In the closed system model, the time-dependent evolution of the moles of an isotopologue of methane can be described in terms of a constant rate of production and a rate of oxidation that behaves as a first-order reaction with an invariable rate constant (Haghnegahdar et al., 2017). In this simplest model, where the balance is only between production and consumption, the system can be described as:

$$\frac{dn_i}{dt} = E_i - k_i n_i, \qquad (11)$$

479 where  $n_i$  is the moles of the isotopic species of interest,  $E_i$  is the source term for *i* (e.g., rate of 480 abiotic or microbial methanogenesis), and  $k_i$  is the rate constant for the sink *i* (i.e., the rate constant 481 for oxidation). The solution to Eq. (11) yields:

482 
$$n_{i}(t) = n_{i}^{0} e^{-k_{i}t} + \frac{E_{i}}{k_{i}} \left(1 - e^{-k_{i}t}\right), \qquad (12)$$

483 where  $n_i^0$  is the initial moles of *i*. The moles of species *i* at steady state is obtained by evaluation 484 Eq. (12) where  $t \to \infty$ , and thus  $e^{-k_i t} \to 0$ , yielding:

485 
$$n_i(t \sim \infty) = \frac{E_i}{k_i}.$$
 (13)

486 Considering *i* to be the major isotopologue, the steady-state amount of methane is 487 controlled by the ratio of the production rate and the rate constant for oxidation. The steady-state 488 ratio of two isotopologues can therefore be expressed as:

489 
$$\frac{n_j(t \sim \infty)}{n_i(t \sim \infty)} = \frac{E_j}{E_i} \frac{k_i}{k_j}$$
(14)

490 where *n* is the moles of the two isotopologues *i* and *j*. The steady-state isotopologue ratio depends 491 on the isotopologue ratio of the methane produced  $(E_j/E_i)$  and the reciprocal of the fractionation 492 due to oxidation  $(k_i/k_j)$ , independent of the absolute rates.

493

# 494 2.10. Comparison between measured and theoretically estimated isotope fractionation 495 factors

496 To understand the meaning of the measured isotope fractionation factors from the Svalbard experiment, we used two methods to estimate the fractionation factors with mechanistic 497 implications. Firstly, if the kinetics are dominated by rupture of single carbon-hydrogen bonds 498 499 comprising the reaction coordinate, the fractionation factors can be estimated by the square root of the inverse ratios of relevant reduced masses,  $\sqrt{\mu / \mu'}$ . Here the  $\mu'$  values are the reduced masses 500 for <sup>13</sup>C-H and <sup>12</sup>C-D as required, u is the reduced mass for <sup>12</sup>C-H, and the derived fractionation 501 502 factors are those implied by these reduced masses, including the use of the rule of the geometric 503 mean for the doubly-substituted species.

Secondly, we consider the ratio of the logarithm of rotational/vibrational partition functions for a crude estimate for the transition state of methane being dismantled by the Mcr-mediated reaction. The Q\* values are estimated by calculating the partition function for methane at 4 °C with the three asymmetric stretch modes (commonly referred to as  $v_3$ ) removed. The differences among the partition functions for the different isotopologues are evidenced clearly by writing their ratios where the prime superscript refers to the isotopically-substituted species (Bigeleisen and Mayer, 1947; Urey, 1947):

511 
$$\frac{Q'}{Q} = \frac{\sigma}{\sigma'} \prod_{i} \frac{u'_{i}}{u_{i}} \frac{e^{-u'_{i}/2}}{1 - e^{-u'_{i}}} \frac{1 - e^{-u_{i}}}{e^{-u_{i}/2}}$$
(15)

512 We used the vibrational frequencies calculated by Lee et al. (1995). For our rough estimate of a suitable transition state bound to Mcr (an imprecise exercise in its own right), we removed the  $v_3$ 513 514 modes from Eq. (15). This approach provides a test of the possibility that it is the highest frequency modes, v<sub>3</sub>, that are most readily broken and dominate the kinetics (and thus are not counted in the 515 516 vibrational partition functions). Ratios of vibrational frequencies, or the square root of the inverse ratio of reduced masses, are ratios of energies associated with a vibration. Values lnQ\* are also 517 proportional to energies, but represent the weighted mean across all vibrations not involved in 518 519 bond rupture (Bigeleisen, 1952). The ratio is therefore an estimate of the relative energy associated with motions in the molecules. 520

521

# 522 **3. RESULTS**

Methane isotopologue data from laboratory experiments and natural fluids are presented in 523 Figure 3. The  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values from the Svalbard sediment slurry incubation and 524 Chamorro Seamount natural fluids are higher than thermodynamic equilibrium values at their 525 ambient temperature (>0 °C; Fig. 3A–C). Other incubations, including the Mcr-catalyzed isotope 526 exchange and incubations of the Santa Barbara Channel sediment, Nankai Trough sediment, and 527 Beatrix Gold Mine fracture fluid, show moderate increase in  $\Delta^{13}$ CH<sub>3</sub>D but minimal increase in 528  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> (Fig. 3D–F). Their apparent temperatures based on clumped isotopes are similar to the 529 530 experimental temperatures, ranging from 20 °C to 60 °C. In the following sections, we first present 531 the laboratory incubation data and then show natural fluid data for comparison.

532

# 533 **3.1. Incubation of the Svalbard methane seep sediment slurries**

Off-shore methane seep sediments from the Svalbard archipelago host high AOM activity. 534 *Ex-situ* AOM rates determined by <sup>14</sup>C-radiotracer techniques peak at 182 nmol cm<sup>-3</sup> d<sup>-1</sup> at 2–5 cm 535 below seafloor (Fig. 2A) (Melaniuk et al., 2022). In the reactivated sediment slurry with saturated 536 537 methane and high sulfate concentrations (8-24 mM), dissolved sulfide concentrations increased linearly as methane concentrations decreased linearly (Fig. 2B). In addition, sulfate concentrations 538 539 decreased, while alkalinity increased during the experiment (see the Supplementary Data). In the control experiment containing a headspace gas of N2 and CO2, no methane was detected and no 540 increase in sulfide concentration was observed, indicating negligible methanogenesis and 541 organoclastic sulfate reduction, respectively (Fig. S1A). Collectively, AOM is active in the 542 sediment slurry, and we assume that AOM is the only mechanism for methane removal in the 543 544 experiment. Accordingly, the *in-vitro* AOM rate determined by methane concentrations in the 545 slurry incubation was 90 nmol of methane per cm<sup>3</sup> sediment slurry per day (Fig. 2B), translating 546 into about 180 nmol per cm<sup>3</sup> of undiluted sediment per day. Furthermore, the *in-vitro* AOM and sulfate reduction rates determined by <sup>14</sup>C- and <sup>35</sup>S-radiotracers were 86 and 84 nmol cm<sup>-3</sup> d<sup>-1</sup>, 547 respectively, in the slurry incubation, which are equivalent to roughly 172 and 168 nmol cm<sup>-3</sup> d<sup>-1</sup> 548 549 for undiluted sediment. The in-vitro AOM rates are consistent with the ex-situ AOM rates of the same sediment measured by <sup>14</sup>C-radiotracer (Fig. 2A) and other *in-vitro* AOM enrichment cultures 550 (e.g., Ono et al., 2021; Wegener et al., 2021). 551

In two experiments with high sulfate, the bulk  $\delta^{13}$ C and  $\delta$ D values of residual methane increased by 9.6‰ and 195.4‰, respectively, with declining methane concentrations (Fig. 4A). Based on a Rayleigh distillation model, the respective carbon and hydrogen isotope fractionation 555 factors, expressed in per mil, were found to be  $4.9 \pm 0.1\%$  and  $149.0 \pm 1.5\%$  in the first experiment, and  $7.5 \pm 0.1\%$  and  $154.0 \pm 1.3\%$  in the second experiment (Fig. 5A–B). With progressive 556 methane consumption,  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values increased by 16.4‰ and 57.6‰, reaching 557 values of 19.5‰ and 65.1‰, respectively (Figs. 3A, 6A). These values are substantially higher 558 than the values expected for isotopologue equilibrium at the incubation temperature of 4 °C. The 559 kinetic clumped isotopologue fractionations ( $\gamma$ ), defined as the ratio of the mass-18 isotopologue 560 fractionation factors to the product of the bulk carbon and hydrogen fractionation factors, describe 561 562 deviations from the rule of the geometric mean (Bigeleisen, 1955; Wang et al., 2016). The respective  $\gamma$  values for <sup>13</sup>CH<sub>3</sub>D and <sup>12</sup>CH<sub>2</sub>D<sub>2</sub> defined by these data were 0.985 ± 0.002 and 0.912 563  $\pm 0.006$  in the first experiment, and  $0.985 \pm 0.002$  and  $0.909 \pm 0.005$  in the second experiment (Fig. 564 565 5C–D). With declining methane concentrations, the  $\delta^{13}$ C values of DIC decreased by 6‰ in the first experiment and 10% in the second experiment, whereas the  $\delta D$  values of water were more-566 or-less invariant (see the Supplementary Data). 567

568 The same sediment slurry was then incubated with low sulfate concentration (<0.35 mM) 569 under the same conditions (e.g., saturated methane), resulting in a lower thermodynamic driving force for AOM. Methane isotopologue compositions were analyzed on days 1, 11, 58, and 79 of 570 the incubation. In contrast to the high sulfate experiment, the bulk  $\delta^{13}$ C and  $\delta$ D values of residual 571 methane decreased by 10.1‰ and 15.2‰, respectively (Fig. 4B). The  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> 572 values, however, increased linearly by 4.6‰ and 14.8‰, respectively (Figs. 3B and 6B). There is 573 no substantial difference in  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values of residual methane taken on days 58 574 575 and 79, likely signaling a reduction in the AOM rate due to sulfate limitation (0.01 mM). To examine whether the residual methane will approach isotopologue equilibrium or overshoot it, 576 577 additional sulfate was added on day 83, to increase the concentration again to 0.39 mM in the

578 slurry. Isotopologue ratios of the residual methane were analyzed on days 92 and 178 of the incubation. Both  $\Delta^{13}CH_3D$  and  $\Delta^{12}CH_2D_2$  values increased up to 11.4‰ and 31.5‰, respectively 579 (Figs. 3B, 6B). Net isotope fractionation factors are not reported here for low sulfate incubation 580 581 because of the challenge of accurately quantifying methane consumption, limiting our ability to estimate the fraction of methane remaining in the Rayleigh process, and thus causing large errors, 582 but both  ${}^{13}\alpha$  and  ${}^{D}\alpha$  are higher than 1.0, in contrast with the high sulfate incubation in which both 583  $^{13}\alpha$  and  $^{D}\alpha$  are lower than 1.0 (Figs. 4A–B, 5). The  $\delta^{13}$ C values of DIC decreased by 2‰, less 584 pronounced than the high-sulfate experiments, whereas the  $\delta D$  values of water remained constant 585 (see the Supplementary Data). 586

587

# 588 **3.2. Incubation of the Santa Barbara Channel methane seep sediment slurries**

Analogous to the incubation of Svalbard sediment slurry, we conducted incubation with 589 590 Santa Barbara Channel sediment slurries at 20 °C. However, the volume and pressure of headspace 591 methane in the Santa Barbara slurry (4.4 ml under ca. 100 kPa) were much lower than those of the 592 Svalbard slurry (60 ml under ca. 200 kPa), presumably leading to a lower thermodynamic driving 593 force for AOM. An earlier study at the same sites of the Santa Barbara Channel found that the exsitu AOM rates determined by <sup>14</sup>C-radiotracer techniques were relatively high, on the order of tens 594 to hundreds of nmol  $cm^{-3} d^{-1}$  (Treude and Ziebis, 2010). All incubations with methane headspace 595 596 showed decreases in sulfate concentration and increases in dissolved sulfide concentration and 597 alkalinity over time (see the Supplementary Data). Along with the ex-situ AOM rates reported in the earlier study and the control experiment (Fig. S1B), these trends indicate the activity of AOM. 598 Some  $\delta D$  values of residual methane decreased by up to 15.0%, whereas the  $\delta^{13}C$  values remained 599 roughly constant over time under both high and low sulfate conditions (Fig. 4E). In the high sulfate 600

slurries (>28 mM), both  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> of residual methane increased, ranging from 3.5‰ to 5.4‰ for  $\Delta^{13}$ CH<sub>3</sub>D and from 8.5‰ to 12.1‰ for  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> (Fig. 3E). In the low sulfate slurries (<1 mM), the  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values remained nearly constant over time (6.7 ± 0.9‰), while the  $\Delta^{13}$ CH<sub>3</sub>D increased by 2.7‰, reaching a value of 5.5 ± 0.2‰ relative to stochastic (Fig. 3E). For comparison, when methane molecules are at thermodynamic equilibrium at 20 °C, the respective  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values would be 5.9‰ and 20.3‰ (Young et al., 2017).

607

# 608 **3.3. Incubation of sediment slurries and fracture fluids from the deep biosphere**

609 Marine sediments and fracture fluids were sampled from the deep subsurface of the Nankai 610 Trough and the Beatrix Gold Mine, respectively. Metagenomics, metatranscriptomics, and 611 fluorescent in situ Hybridization (FISH) of the Beatrix Gold Mine fluids were previously published 612 and the results demonstrated that ANMEs are present and active in the incubations (Lau et al., 613 2016; Harris et al., 2018; Harris et al., 2021). In parallel to the natural abundance incubations described in Section 2.3, long-term (350 day) high-pressure (40 MPa) <sup>13</sup>C-CH<sub>4</sub> tracer incubations 614 615 were performed on Nankai Trough sediment slurries (see Supplementary Methods and Fig. S2). Based on the production of <sup>13</sup>C-DIC (Fig. S3) and  $\delta^{13}C_{DIC}$  measurements (Fig. S4), trace AOM 616 617 activity was statistically distinguishable above background, albeit at very low rates (on the order of pmol cm<sup>-3</sup> day<sup>-1</sup>). These findings agree with <sup>14</sup>C-CH<sub>4</sub> radiotracer experiments reported by 618 619 Beulig et al. (2022), who found potential AOM activity near detection limits in the Nankai Trough sediments due to a high background of abiotic <sup>14</sup>C-CH<sub>4</sub> conversion to <sup>14</sup>C-CO<sub>2</sub> in the medium 620 621 controls. These results collectively highlight the difficulty in quantifying AOM activity in these 622 oligotrophic, low-biomass, deep biosphere sediments (see Heuer et al., 2020). Thus, only potential AOM activity has been observed and no explicit discussion can be made in this respect. 623

624 The bulk isotope data of incubation experiments are consistent with the sluggish nature of microbial activity in the deep subseafloor. Here, nearly all headspace methane showed no 625 substantial changes in  $\delta^{13}$ C and  $\delta$ D values (<2‰; Fig. 4F). However, substantial changes were 626 627 found for clumped isotope ratios. In the Nankai Trough slurries with nitrate or nitrite as the added electron acceptors, only  $\Delta^{13}$ CH<sub>3</sub>D increased, reaching a value of 5.7 ± 0.1‰ as incubation time 628 progressed (Figs. 3F, S5C). Similarly, when iron(III) in the form of hydrous ferric oxide was added 629 as the electron acceptor in the Beatrix Gold Mine incubation,  $\Delta^{13}$ CH<sub>3</sub>D increased up to 5.7 ± 0.1‰, 630 while there was less increase in  $\Delta^{13}$ CH<sub>3</sub>D when sulfate or no electron acceptor was added (Figs. 631 3F, S5D). Little variation in  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values was found in these experiments (Fig. 3F). 632

633

# 634 **3.4. Incubation with methyl-coenzyme M reductase (Mcr) enzyme**

We performed experiments with Mcr purified from Methanothermobacter marburgensis 635 636 that catalyzes the exchange of the methyl moiety of methyl-coenzyme M with methane. The  $\delta^{13}$ C 637 and  $\delta D$  of residual methane decreased by up to 2.3% and 6.4% in all cases, respectively (Fig. 4D). 638 Both  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> increased to different degrees relative to the initial methane 639 isotopologue ratios (Fig. 3D). In detail, at 60 °C, the incubations lasted for 2-6 hours in which a small increase of 0.5% in  $\Delta^{13}$ CH<sub>3</sub>D and 2.0% in  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> were observed. At 20 °C, the 640 incubation lasted for 48 hours and the  $\Delta^{13}$ CH<sub>3</sub>D of residual methane increased substantially by 641 4.1‰, reaching a value of 5.6  $\pm$  0.1‰ relative to stochastic while  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> showed a modest 642 increase of only 2.6‰, reaching a value of  $8.5 \pm 0.5$  ‰. 643

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# 645 **3.5. Natural methane-rich fluids from the South Chamorro Seamount**

Three fluid samples were collected from South Chamorro Seamount during HyperDolphin 646 Dives 941, 945, and 947 in 2009. The general geochemical data were previously published (Wheat 647 et al., 2020). In brief, the stagnant fluids collected during Dive 941 are rich in sulfate (12 mM) and 648 methane (33 mM), whereas discharging fluids from Dives 945 and 947 have low sulfate 649 concentrations (0.8 mM) and relatively low methane concentrations (23 mM). Further, sulfate and 650 651 methane concentrations in the fluids were negatively correlated with aqueous sulfide concentration and alkalinity, indicating that the fluids were altered by AOM to different degrees (Wheat et al., 652 2020). The bulk  $\delta^{13}$ C and  $\delta$ D values range from -37.0% to -26.1% and -110.3% to -100.6%, 653 respectively (Fig. 4C). The respective  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values are 7.5% and 25.2% in the 654 high-sulfate, high-methane fluid, but increase to 12.6‰ and 42.6‰ in low-sulfate, low-methane 655 656 fluids (Figs. 3C, 6C). These values are analogous to the results from the Svalbard slurry 657 incubations (Fig. 3A–B).

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### 659 4. DISCUSSION

# 660 4.1. Kinetic fractionations of methane clumped isotopologue during AOM

AOM was highly active in the sediment slurry from the Svalbard methane seep (Fig. 2). The high-sulfate incubation experiment showed progressive enrichment of methane isotopologues containing <sup>13</sup>C and D isotopes relative to the initial gas (Figs. 4A and 5), indicating a kinetic isotope fractionation. Although the kinetic fractionation for D/H ratios is typical, the carbon isotope fractionation is slightly lower than previous studies, which are in the range of 11‰ to 39‰, suggesting a higher degree of reversibility of the carbon pathway in our AOM incubations (Holler et al., 2009; Wegener et al., 2021). The  $\gamma$  value for <sup>13</sup>CH<sub>3</sub>D of 0.985 is lower than those obtained

for other methane oxidation reactions such as aerobic oxidation and oxidation by OH and Cl 668 radicals, while the  $\gamma$  values for <sup>12</sup>CH<sub>2</sub>D<sub>2</sub> of 0.909–0.912 are similar to those obtained for other 669 methane oxidation reactions (Wang et al., 2016; Haghnegahdar et al., 2017; Whitehill et al., 2017; 670 Ono et al., 2021; Krause et al., 2022). Lower absolute magnitudes of  $\gamma$  values imply larger 671 magnitudes of clumped isotope effects and larger deviations in the clumped isotope fractionation 672 673 factor relative to the product of the bulk isotope fractionation factors. Ono et al. (2021) proposed that the different  $\gamma$  values could be linked to the transition state structure of the reaction. For 674 example, the C-H bond length of methyl-coenzyme M reductase (2.6 Å) and soluble methane 675 monooxygenase (1.3 Å), the key enzymes involved in AOM and aerobic methanotrophy, 676 respectively, are different, necessitating changes in the <sup>13</sup>C–D stretching vibrational mode and thus 677 678 perhaps the difference in kinetic clumped isotopologue effects (Ono et al., 2021).

The extremely high  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values up to 12.6‰ and 42.6‰, respectively. 679 from the Chamorro Seamount, a serpentinite mud volcano in the Mariana forearc (Fig. 6C–F), as 680 681 well as the high values seen in the Svalbard incubations (Fig. 6A), appear to be signatures of AOM. 682 Based on samples from experiments and natural environments for which measurements currently 683 exist, AOM is the only known process that drives methane isotopologue abundances substantially above equilibrium predictions for both  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub>. In the laboratory, we treat AOM 684 685 as a Rayleigh process in which methane is consumed only by AOM in a closed system. This explains the trends in  $\Delta^{13}$ CH<sub>3</sub>D vs.  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> space for the Svalbard incubations (Fig. 6A–B). For 686 the Chamorro Seamount, an open system that involves flow, is likely to be more suitable as a 687 688 model for isotopologue fractionation. There, Wheat et al. (2020) demonstrated that methane production was abiotically fueled by hydrogen production from serpentinization and carbonate 689 dissolution in the deep-sourced fluids, and the fluid compositions were largely altered by AOM as 690

691 the fluid migrates upwards and encounters deposited pelagic sediment underlying South Chamorro 692 Seamount. Although the pH of the collected fluids is 12, the pH of deep subsurface niches where ANME archaea live and AOM occurs can be lower, as observed in neighboring serpentinite mud 693 694 volcanoes (Wheat et al., 2020). Earlier studies have shown that microorganisms can tolerate high pH fluids and perform AOM at the Chamorro Seamount (Takai et al., 2005; Curtis et al., 2013; 695 696 Kawagucci et al., 2018). These AOM-affected fluids make it to the seafloor where they discharge. 697 Therefore, the environment where fluids are traversing through the seamount represents an open 698 system, in which methane is transported into the subduction channel via advection and removed by both advection and AOM, likely resulting in a steady state, or at least a transient steady state. 699 700 A simple mass balance model can be used to investigate the effects of simultaneous advection and 701 oxidation (see Materials and Methods). Here, we assume that the serpentinite-sourced abiotic methane is in isotopologue equilibrium at its formation temperature (>250 °C; Wheat et al., 2020). 702 703 Such near-zero clumped isotope compositions are consistent with the results from abiotic silane 704 decomposition experiments at 300–600 °C reported in Young et al. (2017). We consider that the impact of diffusion in fluid transport on the  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values is negligible within 705 706 the subduction channel given the significantly smaller-scale molecular diffusion operates over 707 relative to advective processes (Wheat et al., 2020). Thus, AOM is assumed to be the only process 708 for isotope fractionation.

The combined flow and oxidation steady-state models shown in Figure 6C–D largely reproduce the observed large  $\Delta^{13}$ CH<sub>3</sub>D and  $^{12}$ CH<sub>2</sub>D<sub>2</sub> values at the Chamorro Seamount. Note that to achieve the best fit, a slightly lower  $\gamma$  value for  $^{13}$ CH<sub>3</sub>D of ~0.980 or a higher  $\gamma$  value for  $^{12}$ CH<sub>2</sub>D<sub>2</sub> of ~0.935 than obtained from the experiments is needed. While the open system model is more suitable to describe these fluids, we also considered the scenario of a closed system in which the formation of methane is balanced against oxidation. In this model, isotopologue abundances vary with time, eventually reaching a steady state that is sensitive to the precise  $\gamma$  values (Fig. 6E–F). We set the ratio of the steady-state moles to initial moles of methane, (E/k)/n<sub>0</sub>, to 1 since the modeled trajectories remain largely the same as (E/k)/n<sub>0</sub> varies. Here, <sup>13</sup>CH<sub>3</sub>D  $\gamma$  values of 0.980– 0.985 and <sup>12</sup>CH<sub>2</sub>D<sub>2</sub>  $\gamma$  values of 0.912–0.935 are needed to achieve the best fit in the closed system model. These estimated  $\gamma$  values for the Chamorro Seamount fluids from either the open- or closedsystem model are largely consistent with those obtained from the Svalbard slurry incubations.

One caveat of these two models is the, to date, largely unknown bulk isotope fractionation 721 factors. In Figure 6, we set  ${}^{13}\alpha$  and  ${}^{D}\alpha$  to be 0.995 and 0.851, respectively, based on experimental 722 observations made in the Svalbard experiment (Table 1). We then test the sensitivity of the models 723 by varying the bulk isotope fractionation factors to allow depletion in heavy isotopes of methane 724 (i.e.,  ${}^{13}\alpha$  and  ${}^{D}\alpha > 1$ ). The model outputs remain largely the same, though with different curvatures 725 (Fig. S6). This demonstrates that the trajectories in  $\Delta^{13}$ CH<sub>3</sub>D vs.  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> space are much more 726 sensitive to the relationship between the bulk  $\alpha$  values (i.e., the  $\gamma$  factors) than to the absolute  ${}^{13}\alpha$ 727 and  ${}^{D}\alpha$  values in these two models (Figs. 6, S6). Given the uncertainties in model inputs and limited 728 data points, we stress that the estimated y values are associated with large uncertainties but, overall, 729 730 are significantly lower than unity.

Taken together, both open and closed system models display similar positive trajectories in  $\Delta^{13}$ CH<sub>3</sub>D vs.  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> space, similar to the Rayleigh distillation model (Fig. 6), suggesting that extremely positive  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values are potentially a clumped isotope signature of kinetically-dominated AOM. These positive values are the result of  $\gamma < 1$  for both mass-18 isotopologues. Indeed, if  $\gamma = 1$  for both rare isotopologues, and all else being equal,  $\Delta^{13}$ CH<sub>3</sub>D and 736  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> would both decrease rather than increase with oxidation. In detail, the Rayleigh 737 fractionation equation (Eq. 7) can be simplified to (Wang et al., 2016):

738 
$$\Delta^{13}\mathrm{CH}_{3}\mathrm{D} = \Delta^{13}\mathrm{CH}_{3}\mathrm{D}_{\mathrm{init}} + ({}^{13\mathrm{D}}\gamma \cdot {}^{13}\alpha \cdot {}^{\mathrm{D}}\alpha - {}^{13}\alpha - {}^{\mathrm{D}}\alpha + 1) \cdot \ln f$$
(16)

739 
$$\Delta^{12} CH_2 D_2 = \Delta^{12} CH_2 D_{2init} + ({}^{D2}\gamma \cdot {}^{D}\alpha^2 - 2 \cdot {}^{D}\alpha + 1) \cdot \ln f$$
(17)

where  $\Delta^{13}$ CH<sub>3</sub>D<sub>init</sub> and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2init</sub> are the initial isotopologue compositions. Therefore, the 740 threshold <sup>13D</sup> $\gamma$  and <sup>D2</sup> $\gamma$  for constant  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values are  $({}^{13}\alpha + {}^{D}\alpha - 1)/({}^{13}\alpha \cdot {}^{D}\alpha)$  and 741  $(2 \cdot {}^{D}\alpha - 1)/{}^{D}\alpha^{2}$ , respectively. Using the measured  ${}^{13}\alpha$  and  ${}^{D}\alpha$  from the high-sulfate Svalbard 742 incubation, the calculated  $^{13D}\gamma$  and  $^{D2}\gamma$  thresholds are found to be 0.999 and 0.969, respectively. 743 Any measured  $\gamma$  factors lower than the threshold conditions means that both  $\Delta^{13}$ CH<sub>3</sub>D and 744  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values should evolve to more positive values in a closed system, and vice versa. 745 Although the  $\gamma$  factors appear to be linked to the transition state structure of the enzyme-mediated 746 747 reaction (Ono et al., 2021), the exact structures of transition states and their role in kinetic clumped isotopologue effect remain unknown. Further theoretical and experimental investigations are 748 needed to elucidate the fundamental mechanism. 749

750 We note that with progressive AOM from the stagnant fluids to the discharging fluids at 751 the Chamorro Seamount as indicated by alkalinity, methane, sulfate, and sulfide concentrations (Wheat et al., 2020), the  $\delta^{13}$ C of residual methane decreases by 10.9‰ and  $\delta$ D increases by 9.7‰ 752 753 in three fluid samples (Fig. 4C), resembling the ANME incubation under low-sulfate conditions 754 (<1 mM) in Wegener et al. (2021). These authors determined radiotracer-based forward and 755 backward AOM reaction rates at different sulfate concentrations and found that the net reversibility 756 of the AOM reaction increases with decreasing sulfate concentration. They concluded that the net 757 bulk isotope fractionations of AOM reflect a combination of expression of kinetic isotope effects

(KIEs) and equilibrium isotope effects (EIEs) (Yoshinaga et al., 2014; Ono et al., 2021; Wegener 758 et al., 2021). Indeed, when the Svalbard sediment slurry was incubated with even lower sulfate 759 concentrations (<0.35 mM), both  $\delta^{13}$ C and  $\delta$ D of residual methane decreased over time (Fig. 4B), 760 suggestive of a greater expression of EIEs in both carbon and hydrogen pathways of AOM due to 761 a lower net thermodynamic drive and thus relatively higher net reversibility of AOM. At 4 °C, the 762 carbon and hydrogen equilibrium isotope fractionation factors between  $HCO_3(aq)-CH_4(g)$  and 763 H<sub>2</sub>O(l)–CH<sub>4</sub>(g) are 1.091 and 1.216, respectively (Zhang et al., 1995; Turner et al., 2021). When 764 inter-species isotope equilibrium is achieved, the  $\delta^{13}$ C and  $\delta$ D of methane should be around -95‰ 765 and -241‰ based on the measured  $\delta^{13}C_{DIC}$  and  $\delta D_{H2O}$  values of -13.1‰ and -76.7‰ in the low-766 sulfate experiment. 767

768 Despite the fact that the residual methane in the low-sulfate incubation moves towards isotope equilibrium between both CH<sub>4</sub>–CO<sub>2</sub> and CH<sub>4</sub>–H<sub>2</sub>O molecules (Fig. 4B),  $\Delta^{13}$ CH<sub>3</sub>D and 769  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> increase linearly and fall on nearly the same Rayleigh fractionation line in  $\Delta^{13}$ CH<sub>3</sub>D vs. 770  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> space as the high-sulfate incubation (Fig. 6B). In particular, the  $\Delta^{13}$ CH<sub>3</sub>D reaches 7.4% 771 772 when sulfate is depleted (0.01 mM), which is higher than the value expected for intra-species isotope equilibrium at the experimental temperature ( $\Delta^{13}CH_3D_{eq.} = 6.5\%$  at 4 °C). This high 773  $\Delta^{13}$ CH<sub>3</sub>D value suggests a kinetic clumped isotopologue fractionation. We surmise that rather than 774 775 representing unidirectional consumption of methane, the isotopic effects we observed appear to 776 reflect a convolution of transition-state kinetics and equilibrium isotopic effects. Furthermore, both  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> increase linearly again after new sulfate is added to the slurry (0.39 mM), 777 indicating that the final  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values in a methane-rich closed system depend 778 on the electron acceptor concentration and availability. The last measured  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> 779 values correspond to apparent temperatures lower than -34 °C (Fig. 6B), far below the incubation 780

temperature of 4 °C. Collectively, our results from the Svalbard sediment slurry and the Chamorro Seamount show clear evidence for kinetic clumped isotopologue fractionation, with the bulk isotope ratios either increasing or decreasing with progressive methane consumption depending upon the availability of the electron acceptor.

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# 6 4.2. Equilibrium between methane isotopologues during AOM

It has been proposed that AOM drives methane isotopologue abundances to thermodynamic isotopic bond-order equilibrium in a range of marine and continental settings (Ash et al., 2019; Giunta et al., 2019; Tyne et al., 2021; Warr et al., 2021; Ono et al., 2022). The initial step of AOM is Mcr-catalyzed reversal of the final reaction in methanogenic pathways as shown for the enzyme from *M. marburgens* is (Eq. 18):

792 
$$CH_4 + CoM-S-S-CoB \rightleftharpoons CH_3-S-CoM + HS-CoB \quad (\Delta G^\circ = +30 \text{ kJ mol}^{-1})$$
 (18)

793 where CoM-S-S-CoB is the heterodisulfide of coenzymes M and B, CH<sub>3</sub>-S-CoM is methyl-794 coenzyme M, and HS-CoB is the reduced form of coenzyme B (Scheller et al., 2010). The reversibility of this reaction is evidently central to the mechanism for methane isotopologue 795 796 equilibration by AOM. The *in-vitro* experiments that comprise exchange of methane and the 797 methyl moiety in methyl-coenzyme M (CH<sub>3</sub>-S-CoM) characterize the role of the Mcr enzyme in 798 reaction Eq. (18). At 60 °C, the optimal temperature for the thermophile-derived enzyme, both  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values of residual methane increase slightly (Fig. 3D). At 20 °C, however, 799  $\Delta^{13}$ CH<sub>3</sub>D approaches the equilibrium value, while there is little change in  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub>. This implies 800 801 that carbon plays a larger role than hydrogen in the isotopologue selectivity for reaction with the 802 Mcr enzyme at a temperature below optimal.

803 Incubations of slurry from the Santa Barbara Channel, with a lower amount of methane further inform the assessment of the role of thermodynamic driving forces in determining the 804 isotopologue signatures of AOM. There is a trend towards  $\Delta^{13}$ CH<sub>3</sub>D and/or  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values 805 expected for isotopologue equilibrium at experimental temperature (Fig. 3E). Two of the high-806 sulfate incubations of the Santa Barbara sediments resemble the high-temperature Mcr 807 808 experiments under the optimal condition, whereas the low-sulfate incubations resemble the lowtemperature Mcr experiments (Fig. 3D-E). Importantly, the amounts of methane injected into the 809 810 Santa Barbara slurry (0.5 mmol) are much lower than those into the Svalbard slurry (5.4 mmol), leading to a much lower thermodynamic drive for AOM for the former and an overall lower rate 811 812 of reaction. Thus, the reversibility and isotope exchange during AOM in the Santa Barbara slurry 813 may be much higher than the Svalbard slurry under either high- or low-sulfate conditions. This vigorous exchange ultimately allows methane isotopologues to react with Mcr reversibly, 814 achieving partial intra-species isotope exchange in the Santa Barbara slurry incubations. The high 815 816 reversibility may be exacerbated by the fact that the ANME community in coastal sandy Santa 817 Barbara sediments with high depositional rates is less active with less biomass compared to that 818 from the deep-sea fine-grained Svalbard methane seep sediments. The lower sedimentation rates 819 at the Svalbard seep focus AOM in the same sediments for an extended period, allowing the 820 ANME community to grow and enrich.

All experiments of the deep biosphere sediment and fluid incubations suggest that there is a trend toward equilibrium values for  $\Delta^{13}$ CH<sub>3</sub>D but little change in  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> within experimental timescales, regardless of which electron acceptor is present (Fig. 3F). In detail, a positive correlation between incubation time (up to 500 days) and increase in  $\Delta^{13}$ CH<sub>3</sub>D values is found in incubation experiments of the Nankai Trough sediments (Fig. S5C), consistent with the well 826

documented sluggish nature of microbial activity in the deep subseafloor (Hoehler and Jørgensen,

827 2013; Heuer et al., 2020).

One may postulate that the  $\Delta^{13}$ CH<sub>3</sub>D values in our incubations that appear to approach 828 equilibrium values would in fact continue to increase beyond the equilibrium value given sufficient 829 time. However, this is probably not the case. The respective maximum observed for  $\Delta^{13}$ CH<sub>3</sub>D 830 values is 5.56‰ (2 days), 5.50‰ (3 days), 5.70‰ (500 days), and 5.66‰ (350 days) in the Mcr 831 experiment, and the incubations of the Santa Barbara Channel sediment, the Nankai Trough 832 sediment, and the Beatrix Gold Mine fracture fluid (Fig. 3D-F). The apparent temperatures based 833 on  $\Delta^{13}$ CH<sub>3</sub>D are 32 °C, 33 °C, 27 °C, and 28 °C, respectively (30 ± 3 °C, n = 4). These are all close 834 to the ambient temperature under which these different incubations took place. Although it cannot 835 be categorically ruled out that these  $\Delta^{13}$ CH<sub>3</sub>D values reflected ambient temperatures by 836 837 happenstance during an ongoing kinetically driven trend, it would be fortuitous that this would occur four times in independent experimentation. Therefore, we conclude that the  $\Delta^{13}$ CH<sub>3</sub>D values 838 839 increase over time towards equilibrium and indeed remain at thermodynamic equilibrium despite 840 a large range of incubation times of 2 to 500 days in the above four experiments.

841 Given the observed intra-species isotope quasi-equilibrium, one may similarly expect inter-842 species isotope exchange to occur as seen in the low-sulfate Svalbard experiment (Fig. 4B). The 843 residual methane in most of the Santa Barbara and deep biosphere incubations, however, showed no substantial changes in  $\delta^{13}$ C and  $\delta$ D values (Fig. 4E–F). Changes in bulk and clumped isotope 844 compositions are decoupled in these incubations. While more experiments are needed to explicitly 845 confirm the conditions under which this decoupling occurs, one tentative hypothesis is that, under 846 847 certain conditions, the Mcr enzyme only mediates the exchange of isotopes among methane molecules without expressed bulk isotope fractionation, allowing <sup>13</sup>CH<sub>3</sub>D and/or <sup>12</sup>CH<sub>2</sub>D<sub>2</sub> to be 848

re-ordered to the incubation temperature via bond breaking and reformation. This hypothesis is analogous to the heating experiments in which methane intra-species equilibration is promoted by the presence of a metal catalyst at temperatures above 200 °C (Ono et al., 2014; Stolper et al., 2014b; Young et al., 2017). In our case, the Mcr enzyme may act as the catalyst and promote isotopic bond re-ordering with limited isotope exchange between  $CH_4$ – $CO_2$  and  $CH_4$ – $H_2O$ molecules, as methane molecules are disassembled and reassembled. This hypothesis requires further experimental investigation.

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#### 4.3 A general model for methane isotopologue fractionation due to AOM

### 858 4.3.1 Model construction

To understand the mechanism of methane isotopologue fractionation and/or bond reordering during AOM, the methane isotopologue fractionation factors obtained in AOM laboratory incubation experiments can be used to construct a geochemical model with several simplifying assumptions for fractionation due to AOM. The result is a parsimonious model that explains most methane isotopologue data affected by anaerobic oxidation by ANMEs collected thus far. A crucial aspect of the model is the important role that reversibility has in controlling the trajectory of gases in  $\Delta^{13}$ CH<sub>3</sub>D vs.  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> space during AOM.

The model addresses the most important single step, the potentially reversible cleaving of CH<sub>4</sub> molecules catalyzed the Mcr enzyme to form co-enzyme hosted CH<sub>3</sub> and H in the ANME cells (e.g., Scheller et al., 2010). The relevant reaction can therefore be written succinctly as

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$$\operatorname{CH}_{4} \xrightarrow{k_{f}} \operatorname{CH}_{3} + \mathrm{H}$$
 (19)

where the forward and reverse rate constants,  $k_f$  and  $k_r$ , are controlled by the kinetics of binding and bond rupture/formation in the first step of AOM. While there are a total of 280 isotopic isomer equivalents of the reaction in Eq. (19), with one exception (see below), we will focus on a simplified reaction network of isotopologues (as opposed to all isotopic isomers). This is necessitated by the fact that the measured fractionation factors convolve a number of isotopic effects, including the symmetry effects of all isomers. The sixteen reactions in terms of 12 isotopologues are

1. 
$$CH_{4} \xrightarrow{k_{1}} CH_{3} + H$$
  
2.  $CH_{3}D \xrightarrow{k_{2}} CH_{3} + D$   
3.  $CH_{3}D \xrightarrow{k_{3}} CH_{2}D + H$   
4.  $CH_{2}D_{2} \xrightarrow{k_{4}} CH_{2}D + D$   
5.  $CH_{2}D_{2} \xrightarrow{k_{5}} CHD_{2} + H$   
6.  $^{13}CH_{4} \xrightarrow{k_{6}} ^{13}CH_{3} + H$   
7.  $^{13}CH_{3}D \xrightarrow{k_{7}} ^{13}CH_{3} + D$   
8.  $^{13}CH_{3}D \xrightarrow{k_{8}} ^{13}CH_{2}D + H$   
9.  $CH_{3} + H \xrightarrow{k_{10}} CH_{4}$   
10.  $CH_{3} + D \xrightarrow{k_{10}} CH_{3}D$   
11.  $CH_{2}D + H \xrightarrow{k_{11}} CH_{3}D$   
12.  $CH_{2}D + D \xrightarrow{k_{12}} CH_{2}D_{2}$   
13.  $CHD_{2} + H \xrightarrow{k_{13}} CH_{2}D_{2}$   
14.  $^{13}CH_{3} + H \xrightarrow{k_{14}} ^{13}CH_{4}$   
15.  $^{13}CH_{3} + D \xrightarrow{k_{15}} ^{13}CH_{3}D$   
16.  $^{13}CH_{2}D + H \xrightarrow{k_{16}} ^{13}CH_{3}D$ 

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where  $k_i$  are the relevant rate constants (first order for reactions 1 through 8, 2<sup>nd</sup> order for 9–16), and again we emphasize that the different isomers are embedded in the fractionation factors extracted from experiments.

Isotopologue fractionation factors for the first step in deconstructing methane isotopologues are in effect ratios of rate constants 1–8. We measured these fractionation factors for the Svalbard incubations in which the loss of methane gas due to AOM is consistent with an irreversible Rayleigh process. The derived fractionation factors,  $\alpha$ , are listed in Table 1 together with relevant comparisons to be discussed. Here, the 3<sup>rd</sup> column is the square root of the inverse ratios of relevant reduced masses (<sup>12</sup>C-H, <sup>13</sup>C-H, <sup>12</sup>C-D, etc.). These are the fractionation factors expected where the kinetics are dominated by rupture of single carbon-hydrogen bonds comprising the reaction coordinate. Note that the experimentally-derived fractionation factors are considerably closer to unity than those implied by the reduced masses.

The 4<sup>th</sup> column in the table of fractionation factors is the ratio of the logarithm of 890 rotational/vibrational partition functions for a crude estimate for the transition state of methane 891 892 being dismantled by the Mcr-mediated reaction. We note that the measured fractionation factors are crudely similar, but not identical, to the lnQ\*'/lnQ\* ratios; the ratios of logs of partition 893 894 functions provide better rationalizations for the measured fractionation factors than do reduced 895 masses along individual bonds. We note further that this is evidently a characteristic of AOM as 896 opposed to abiotic oxidation in the atmosphere and perhaps bacterial aerobic oxidation, where 897 fractionation factors are significantly closer to the square root of the inverse ratios of relevant 898 reduced masses (Haghnegahdar et al., 2017; Whitehill et al., 2017; Krause et al., 2022). For example, the values for  $\alpha^{13}C/^{12}C$ ,  $\alpha D/H$ ,  $\alpha^{13}CH_3D/CH_4$ , and  $\alpha^{12}CH_2D_2/CH_4$  of methane oxidation 899 900 by OH radical in the atmosphere are 0.994, 0.758, 0.752, and 0.521, respectively (Haghnegahdar 901 et al., 2017), which by inspection are seen to be closer to expectations from reduced masses along 902 a C-H reaction coordinate. We suggest that larger fractionation nearer to unity, and resembling plausible ratios of energies of methane transition states, reflect significant binding of methane to 903 904 the catalyzing enzymes (e.g., Mcr), in the case of AOM, leading to a more dispersed dependence of bond rupture on overall rotational/vibrational energy for the different isotopologues. 905

906 The model formulated to accommodate the measured fractionation factors comprises the 907 set of rate constants, with branching ratios, and fractionation factors shown in Table 2. Several assumptions are needed based on theoretical consideration because some inputs are not readily 908 available from the experiments. Taking reactions 2 and 3 as an example, the breakage of a <sup>12</sup>C-D 909 bond in <sup>12</sup>CH<sub>3</sub>D (primary kinetic isotope effect for D/H,  $\alpha_2$ ) is associated with a larger kinetic 910 isotope effect than breaking a  ${}^{12}$ C-H bond in  ${}^{12}$ CH<sub>3</sub>D (secondary kinetic isotope effect,  $\alpha_3$ ). Thus, 911 we set  $\alpha_2$  to be 0.5, which is close to the primary KIE (0.41 ± 0.04) observed in the methane 912 913 activation experiment using the Mcr enzyme (Scheller et al., 2013). The primary KIEs for doubly substituted species are derived following the rule of the geometric mean with measured  $\gamma$  values 914 915 (Eqs. 8–9). The branching ratios are set to be  $\frac{1}{4}$  for reactions 2 and 7,  $\frac{3}{4}$  for reactions 3 and 8, and 916 <sup>1</sup>/<sub>2</sub> for reactions 4 and 5, as only <sup>1</sup>/<sub>4</sub> of the time the D is removed from <sup>12</sup>CH<sub>3</sub>D or <sup>13</sup>CH<sub>3</sub>D in its destruction while <sup>3</sup>/<sub>4</sub> of the time an H is removed. Accordingly, the secondary KIEs are calculated 917 from the measured fractionation factor,  ${}^{D}\alpha$ , and the branching ratios such that  ${}^{D}\alpha = \frac{1}{4}\alpha_{2} + \frac{3}{4}\alpha_{3}$ . 918 919 We further consider bulk and clumped isotope equilibrium in the reversible reactions. At 920 equilibrium, the ratio of the reverse and forward KIEs is equal to the EIE of the reaction. Thus, the 921 KIEs for the reverse reactions are derived using the forward KIEs above and the EIEs calculated 922 by Gropp et al. (2021). When the reaction is irreversible, only the forward KIEs are expressed, 923 whereas the EIEs are expected to be more dominant when the reaction is highly reversible. In the 924 latter case, we have allowed for exchange equilibrium among methane molecules to result from methane formation. This is done by including  $^{D2}\gamma_{eq}$  and  $^{13D}\gamma_{eq}$ , the fractionation factors representing 925 equilibrium abundances of <sup>12</sup>CH<sub>2</sub>D<sub>2</sub> and <sup>13</sup>CH<sub>3</sub>D in thermodynamic equilibrium. In what follows 926 927 we use an equilibration temperature for AOM of 20 °C. We verified the branching ratios in Table

2 using the full 280 reactions involving 50 isotopomer species and simple rate constants composed
of square roots of inverse ratios of reduced masses (see below).

Reversibility is determined by the ratio  $\Phi = k_r/k_f$ . Where  $\Phi = 0$ , oxidation is "irreversible", 930 corresponding to Rayleigh fractionation with purely kinetic fractionation factors. We assume that 931 932 the high-sulfate Svalbard incubation represents this irreversible end-member. Where  $\Phi > 0$ , the 933 influences of reversibility become immediately evident for even small values (and being mindful that  $k_r$  refers to a 2<sup>nd</sup> order reaction while  $k_f$  is for a first-order reaction, with the commensurate 934 change in units for the rate constants). As  $\phi \to 1$ , the oxidation at the Mcr step approaches full 935 936 reversibility and equilibrium relative abundances of methane isotopologues result from the 937 reaction with only negligible decreases in methane abundance. In all cases, the systems reach 938 steady state, given sufficient time, with the steady state abundance of methane, and its isotopologue 939 composition, determined by the value for  $\Phi$ .

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#### 941 4.3.2 Example applications

*Irreversible Rayleigh fractionation*: This example application of the model is trivial in so far as it is the model assumed to derive the fractionation factors. Nonetheless, it is useful to illustrate that the Svalbard sediment incubation data are properly accounted for using the model. Figure 7A shows the path in  $\Delta^{13}$ CH<sub>3</sub>D vs.  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> space predicted by the model compared with the calibration data for the case of Rayleigh fractionation where  $\Phi = 0$ . The model also fits the bulk isotope data well for similar values of *F*, the fraction of methane remaining (Fig. 8A).

948 *Reversibility*: Some data for methane known to have been processed by AOM exhibit 949 significant variations in  $\Delta^{13}$ CH<sub>3</sub>D at roughly constant  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> and minimal variations in bulk isotope ratios. Our model explains these data as being the result of more reversibility in the Mcrmediated transformation between  $CH_4$  and  $CH_3$ +H. We illustrate this here by comparing our results with the sediment slurry incubation data from the Santa Barbara Channel.

Most of the data exhibit a "horizontal" trend in the mass-18 isotopologue diagram that can 953 be reproduced with our model using  $k_r/k_f \sim 5 \times 10^{-6}$ , resulting in a predicted steady-state 954 concentration of methane that is about 99% of the initial value, and with negligible changes in  $\delta D$ 955 and  $\delta^{13}$ C, in broad agreement with the data (Figs. 7B and 8B). As the  $k_r/k_f$  increases to  $10^{-1}$ , the 956 model leads to increases in both  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> (Fig. 7B). This at least qualitative 957 958 agreement between the AOM data from the Santa Barbara Channel and the model for significant reversibility in the first AOM step mediated by Mcr raises the specter of a potential monitor of 959 Mcr-catalyzed intracellular isotope exchange operating at near threshold free energy geochemical 960 961 settings, such as the deep biosphere (Fig. 7B-C).

Indeed, near-equilibrium methane isotopologue ratio of *in-situ* methane was found in the Beatrix Gold Mine with minor negative excursion in  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> relative to equilibrium (Fig. 7C; Young et al., 2017). Here, methane was produced abiotically with negative  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values that are progressively erased by the incursion of biological activity that drives the isotopic bond ordering in methane towards equilibrium (Young et al., 2017). The board consistency in isotopologue ratios between the *in-situ* gas and laboratory incubations of the Beatrix samples further confirms the modeled trajectory (Fig. 7C).

We checked that the modeled trajectories are robust, and can be derived using kinetic rate constants defined by reduced masses alone combined with the full 280 reactions for all isomers (see below). We used the measured fractionation factors from the Svalbard incubation as model input for the Santa Barbara Channel and deep biosphere incubations as well. However, this may not be the case. To test whether the model is sensitive to the input fractionation factors, all KIEs and EIEs are set to 1.0 with the  $^{D2}\gamma_{eq}$  and  $^{13D}\gamma_{eq}$  unchanged. This sensitivity test indicates that the modeled trajectory remains the same and is not sensitive to the input fractionation factors when the reversibility is relatively high (Fig. S7).

977 Exchange mediated by Mcr: In the last example we make use of a related but different model to illustrate the path in  $\Delta^{13}$ CH<sub>3</sub>D vs.  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> space due to enzymatically-mediated 978 exchange of isotopes among methane molecules. The Mcr experiments provide the data against 979 which to test this model. We do not use precisely the model shown in Table 2 in this case. The 980 981 reason is that pure exchange is likely to be dominated by symmetry effects, and the fractionation 982 factors obtained from the Svalbard experiments are an unresolvable convolution of these effects 983 with other factors. To overcome this issue, this model is analogous to the simple model above, but 984 in this case all isomers are included, producing 50 species related by 280 reactions, to explain the kinetics. Rather than the measured fractionation factors, we here use generic kinetic fractionation 985 factors corresponding to square roots of the inverse ratios of relevant reduced masses (<sup>12</sup>C-H, <sup>13</sup>C-986 H, <sup>12</sup>C-D, etc.). 987

We impose a value for  $\Phi$  of > 0.1, making the result dependent only on the relative number of isomers for each species and the two equilibrium fractionation factors  $^{D2}\gamma_{eq}$  and  $^{13D}\gamma_{eq}$ . Figure 7D shows our prediction dominated by symmetry with the 60 °C exchange experiments in which the Mcr enzyme operates at its optimal condition. Here again, the agreement is reasonably good, suggesting that the inferences about the trajectory in  $\Delta^{13}CH_3D$  vs.  $\Delta^{12}CH_2D_2$  space due to nearly complete reversibility are correct. However, at 20 °C, below the optimal condition for the Mcr enzyme, a much lower  $\Phi$  is needed to reproduce the data.

995 *Limitation*: Due to the complexity of isotopomer species involved in the reactions, the 996 models presented here only focus on the first and, arguably, the most important step of AOM but 997 do not include other intracellular pathways. As demonstrated by Wegener et al. (2021), a multistep 998 isotope model for AOM including KIEs and EIEs is useful to explain the opposing bulk isotope effects (cf. Northrop, 1981). Such multistep isotopologue models have also been used to simulate 999 1000 clumped isotope effects of microbial methanogenesis (Cao et al., 2019; Young, 2019; Gropp et al., 1001 2022; Ono et al., 2022). Indeed, this one-step model fails to yield inter-species isotope equilibrium 1002 for bulk isotopes with high reversibility, though equilibrium fractionation factors for both bulk and clumped isotopes are parameterized in the model. Therefore, we stress that the inputs and outputs 1003 1004 of our model should be considered as illustrative approximations. Furthermore, the rate of AOM 1005 itself was not directly measured in our experiments other than the Svalbard slurry incubations. The lack of rate information limits the ability to quantify the degree of reversibility on the isotopologue 1006 1007 ratios in our study. Nonetheless, the models presented here allow multiple scenarios to be tested and suggest that high reversibility and low reversibility have characteristic trajectories in  $\Delta^{13}$ CH<sub>3</sub>D 1008 vs.  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> space that are exhibited by the data. 1009

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# 1011 4.4. Kinetically-driven AOM versus equilibrium-driven AOM

Taking all of our results together, we find that the  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> signatures of AOM can be understood in terms of reversibility at the Mcr step. The degree of reversibility is presumably controlled by the tenets of disequilibrium thermodynamics in which rates of reaction are proportional to reaction affinity (deviations in chemical potentials from equilibrium). Kinetic clumped isotope fractionation is found in the Svalbard slurry in which large amounts of methane are provided to the active ANME community. This kinetic signature is also seen in natural fluids 1018 from the South Chamorro Seamount, indicating that the kinetic isotopologue fractionation does not differ substantially between laboratory incubations and natural environments (Fig. 6). When 1019 less methane is provided,  $\Delta^{13}$ CH<sub>3</sub>D tends to increase towards thermodynamic near-equilibrium 1020 values with minimal increase in  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> (Fig. 3E). For example, there is a striking difference in 1021 1022 the isotopologue evolution of residual methane between the Svalbard slurry, in which kinetics drives  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> to extreme values, and the Santa Barbara slurry incubations in 1023 which changes occur mainly in  $\Delta^{13}$ CH<sub>3</sub>D and appear to approach equilibrium, even though the 1024 sediments are both from methane seeps (Fig. 3). The simple kinetic model shown here reproduces 1025 the salient features of this behavior as being the result of reversibility (Fig. 7). Another reason for 1026 1027 this discrepancy is the biomass of ANME archaea under different conditions, such that in slurry 1028 that experienced high methane concentrations over a longer period of time, the biomass may be higher and vice-versa. We therefore infer that AOM with high sulfate concentrations may show 1029 1030 either kinetic or equilibrium behavior depending on the local ANME community, the availability 1031 of methane, and thus the overall thermodynamic drive.

We conclude that both  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values reflect kinetic isotope fractionation 1032 and increase beyond intra-methane thermodynamic equilibrium when the net AOM reaction has 1033 1034 low reversibility due to high rates, whereas largely reversible exchange between methane 1035 isotopologues facilitated by the Mcr enzyme under a lower thermodynamic drive leads to methane isotopic bond re-ordering, driving the abundances of methane isotopologues ultimately to 1036 thermodynamic quasi-equilibrium values. This observation is consistent with reaction affinity as 1037 1038 a primary determinant for the behavior of the system. These departures from equilibrium, or reaction affinities, are controlled by environmental factors, including but not limited to, 1039 temperature, concentrations of methane and electron acceptors, and perhaps the type of electron 1040

acceptors. Although it seems that <sup>13</sup>CH<sub>3</sub>D and <sup>12</sup>CH<sub>2</sub>D<sub>2</sub> equilibration are ultimately controlled by the reversibility of AOM, the driver for different trajectories in  $\Delta^{13}$ CH<sub>3</sub>D vs.  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> space is still elusive. Future efforts should explicitly focus on the environmental controls of the enzymatic activity of intracellular pathways and the reversibility of AOM, and their intrinsic link to methane isotopologue ratios.

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#### 1047 **4.5. Implications**

A schematic representation of methane clumped isotope signatures of diverse methane 1048 sources, as well as kinetically- and equilibrium-driven AOM, is shown in Figure 8. Extremely low 1049 1050  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values down to ca. -40‰ have been observed for microbial methane under 1051 experimental and natural conditions, as well as thermogenic and abiotic methane generated by laboratory experiments (e.g., Young et al., 2017; Young, 2019; Taenzer et al., 2020; Dong et al., 1052 1053 2021). Besides equilibrium-driven AOM, isotopologue data near thermodynamic equilibrium have 1054 been attributed to high-maturity thermogenic or high-temperature abiotic methane, as well as low-1055 rate methanogenesis (Stolper et al., 2015; Wang et al., 2015; Young et al., 2017; Ash et al., 2019; 1056 Giunta et al., 2019; Labidi et al., 2020; Warr et al., 2021; Xie et al., 2021; Gropp et al., 2022; Ono 1057 et al., 2022). Recently, Ono et al. (2022) demonstrated that methanogenesis does not occur or only 1058 proceeds at extremely slow rates at low pH<sub>2</sub> in energy-limited deep sedimentary environments 1059 using an isotopologue flow network model, and therefore inferred that near-equilibrium methane 1060 isotopologue signals in deep marine sediments are produced by the catalytic reversibility of the 1061 Mcr enzyme, likely from ANME archaea performing either AOM or net methanogenesis. This inference is consistent with our incubation experiments in which Mcr-catalyzed isotope exchange 1062 1063 occurs.

1064	To date, extremely positive $\Delta^{13}CH_3D$ and $\Delta^{12}CH_2D_2$ values have been observed, as
1065	demonstrated here, only under conditions where AOM occurs with a low degree of reversibility.
1066	This observation raises an intriguing question whether methane clumped isotope signature of
1067	AOM could have the potential to be diagnostic of AOM on Earth and perhaps throughout the solar
1068	system where methane has been detected (Webster et al., 2015; Waite et al., 2017; Yung et al.,
1069	2018; Thompson et al., 2022). For example, the <i>in-situ</i> discovery of isotopically depleted methane
1070	gas on Mars by Curiosity (Webster et al., 2015; House et al., 2022) indicates that methane is
1071	produced either biogenically or abiogenically (Atreya et al., 2007; Yung et al., 2018), and its
1072	emission results in the presence of methane in the atmosphere of Mars (Formisano et al., 2004).
1073	On the other hand, although the atmospheric methane sink on Mars has been quantified (Atreya et
1074	al., 2007; Yung et al., 2018), abiotic loss mechanisms would suggest a mean atmospheric residence
1075	time of $\sim$ 300 years – substantially longer than observed methane lifetimes on the order of months
1076	to years (Lefèvre, 2019). The conditions for a subsurface microbial sink on Mars are still elusive.
1077	Nevertheless, the reaction transport model by Marlow et al. (2014) demonstrated that AOM could
1078	be a feasible metabolism on ancient Mars, for example, in sulfate-rich groundwater with methane
1079	from serpentinization by-products, and acid-sulfate fluids with methane produced from basalt
1080	alteration. The surface of Mars also includes deposits of iron oxides and manganese oxides, Fe-
1081	and Mn-dependent AOM therefore has also been proposed as a possible metabolism for recent and
1082	ancient Mars (House et al., 2011). Applying these inferences to our understanding of methane
1083	cycling on Mars is speculative. Nonetheless, a robust <i>in-situ</i> approach is needed to trace processes
1084	of methane cycling on future missions such as Mars Life Explorer (National Academies of
1085	Sciences, Engineering, and Medicine, 2022).

1086 Mars is one of the most promising candidates for the application of methane isotope clumping. The reason is that doubly substituted isotopologues of methane remove the difficulties 1087 associated with using bulk  ${}^{13}C/{}^{12}C$  and D/H elsewhere in the solar system, where the geochemical 1088 context necessary for interpreting these ratios are hampered by incompletely characterized 1089 chemical cycles (Lefèvre, 2019; Young, 2019; House et al., 2022). In  $\Delta^{13}$ CH<sub>3</sub>D vs.  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> 1090 space, microbial, thermogenic, and abiotic methane gases are either below or near the 1091 1092 thermodynamic equilibrium predictions (Fig. 9). Once formed, methane could be oxidized photochemically or by microbial oxidation. These sinks have different  $\Delta^{13}$ CH<sub>3</sub>D vs.  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> 1093 signatures, depending upon whether a steady state is achieved or not. The details depend on the 1094 1095 precise  $\gamma$  values, but in general, by analogy with Earth's atmosphere, oxidation in the atmosphere 1096 (e.g., by OH radicals) should ultimately lead to either high  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> of order tens to hundreds of per mil at less variable  $\Delta^{13}$ CH<sub>3</sub>D values in residual methane at steady state, or extremely low values 1097 1098 for both where a steady state between methane formation and oxidation has not been achieved (Fig. 1099 9) (Haghnegahdar et al., 2017). The trajectories for microbial aerobic oxidation of methane are 1100 like those of the CH<sub>4</sub> + OH reaction (Krause et al., 2022). AOM, on the other hand, can lead to extremely high  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> and  $\Delta^{13}$ CH<sub>3</sub>D values up to 65.1‰ and 19.5‰, respectively, as shown 1101 1102 here. Indeed, these signatures are essentially orthogonal. Although mixing scenarios could mimic the extreme signatures from AOM, those mixing scenarios require extremely large differences in 1103 1104 bulk carbon and hydrogen isotopic values of the two endmembers of methane (e.g., some 80%) and 300% differences in endmember  $\delta^{13}C$  and  $\delta D$  values, respectively), possibly allowing 1105 kinetically-driven AOM to be distinguished from mixing with concurrent measurement of bulk 1106 1107 isotopes.

1108 While intriguing, we presently point out that clumped isotopes for methane would be an 1109 approach with a high potential for false negatives, as only anaerobic oxidation of methane, and only under conditions of low reversibility, would reasonably produce a sufficiently diagnostic 1110 1111 signal. Another caveat here is that significant technical development would be necessary to measure methane that has an atmospheric mixing ratio on the order of a few ppb. Nonetheless, 1112 1113 along with previous isotopologue studies on microbial methanogenesis (Stolper et al., 2015; Wang et al., 2015; Young et al., 2017; Taenzer et al., 2020), our work on the effects of AOM on doubly 1114 1115 substituted isotopologues of methane suggests that future missions could benefit from additional research and technique development for in-situ mission measurements of methane clumped 1116 isotopes on Mars, Enceladus, and other solar system bodies where the methane cycling is a key 1117 1118 (bio)geochemical tracer.

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### 1120 **5.** Conclusions

1121 We performed clumped isotopologue analysis on methane samples from laboratory 1122 microbial incubations and natural environments to elucidate the range of clumped isotope effects caused by AOM under different conditions. We found extremely high  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> 1123 1124 values up to 19.5‰ and 65.1‰, respectively, in microbial incubations of sediment slurries from 1125 the Svalbard methane seep and in natural fluids from the South Chamorro Seamount. We found 1126 near-equilibrium methane clumped isotope compositions in the Mcr exchange experiment and 1127 microbial incubations of sediment slurries and/or fracture fluids from the Santa Barbara Channel and deep biosphere. Thermodynamic drive and the reversibility of AOM intracellular reactions are 1128 invoked to interpret the observed differences in  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> signatures and the 1129 interpretations are consistent with results from a simple, one-step isotopologue fractionation model. 1130

Low reversibility of the AOM reaction means a pronounced expression of kinetic fractionation of clumped isotopologues, whereas the Mcr-catalyzed intracellular isotope exchange may be more prominent with higher reversibility. The latter drives the isotopic bond ordering in methane towards equilibrium and may be responsible for the widely observed near-equilibrium clumped isotope signatures in substrate-limited anoxic natural environments where microbial methane production and/or oxidation occur.

### 1137 Declaration of Competing Interest

1138 The authors declare that they have no known competing financial interests or personal 1139 relationships that could have appeared to influence the work reported in this paper.

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## 1156 Appendix A. Supplementary Material

1157 The supplementary material includes supplementary figures S1–S7, supplementary 1158 methods for the  $^{13}$ CH<sub>4</sub> tracer microcosm experiment at high hydrostatic pressure, the research data 1159 and Python scripts for the simplified reaction network of isotopologues and the full reaction 1160 network of isotopomers.

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1484 Tables

**Table 1.** Measured and estimated fractionation factors relative to  ${}^{12}$ CH<sub>4</sub>. The experimental data are from the Svalbard incubations.  $\mu$  is reduced mass. Q\* is rotational/vibrational partition functions for a crude estimate for the transition state of methane being dismantled by the Mcrmediated reaction. The prime superscript refers to the isotopically-substituted species.

Species/ <sup>12</sup> CH <sub>4</sub>	Experiment	$\sqrt{\mu/\mu'}$	$lnQ^*'/lnQ^*$
$^{13}CH_{4}$	0.995	0.997	0.997
$^{12}CH_3D$	0.851	0.734	0.832
$^{13}CH_{3}D$	0.834	0.732	0.827
$^{12}CH_2D_2$	0.660	0.539	0.694

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**Table 2**. Rate constants (*k*) and isotope fractionation factors ( $\alpha$ ) for the simplified reaction network of isotopologues. <sup>D</sup> $\alpha_{eq-P}$  and <sup>D</sup> $\alpha_{eq-S}$  are the equilibrium fractionation factors for primary and secondary fractionations, respectively. <sup>D2</sup> $\gamma_{eq(-P/S)}$  and <sup>13D</sup> $\gamma_{eq(-P/S)}$  are the fractionation factors representing equilibrium abundances of <sup>12</sup>CH<sub>2</sub>D<sub>2</sub> and <sup>13</sup>CH<sub>3</sub>D in thermodynamic equilibrium. The reader is referred to the text for interpretation of the table.

Reaction	k	α
1. $CH_4 \rightarrow CH_3 + H$	k <sub>f</sub>	$\alpha_1 = 1.000$
2. $CH_3D \rightarrow CH_3 + D$	$k_2 = \frac{1}{4} k_f \alpha_2$	$\alpha_2 = 0.500$
3. $CH_3D \rightarrow CH_2D + H$	$k_3 = \frac{3}{4} k_{\rm f}  \alpha_3$	$\alpha_3 = 0.968$
4. $CH_2D_2 \rightarrow CH_2D + D$	$k_4 = \frac{1}{2} k_{\rm f}  \alpha_4$	$\alpha_4 = 0.441$
5. $CH_2D_2 \rightarrow CHD_2 + H$	$k_5 = \frac{1}{2} k_{\rm f}  \alpha_5$	$\alpha_5 = 0.879$
$6. \ ^{13}CH_4 \rightarrow ^{13}CH_3 + H$	$k_6 = k_f \alpha_6$	$\alpha_6 = 0.995$
7. ${}^{13}CH_3D \rightarrow {}^{13}CH_3 + D$	$k_7 = \frac{1}{4} k_f \alpha_7$	$\alpha_7 = 0.490$
8. ${}^{13}CH_3D \rightarrow {}^{13}CH_2D + H$	$k_8 = \frac{3}{4} k_f \alpha_8$	$\alpha_8 = 0.949$
9. $CH_3 + H \rightarrow CH_4$	$k_9 = \frac{1}{2} k_r \alpha_9$	$\alpha_9 = \alpha_1$
$10. CH_3 + D \rightarrow CH_3D$	$k_{10} = \frac{1}{2} k_r \alpha_{10}$	$\alpha_{10} = \alpha_2 \cdot {}^{\mathrm{D}} \alpha_{\mathrm{eq-P}}$
$11. CH_2D + H \rightarrow CH_3D$	$k_{11} = \frac{1}{2} k_r \alpha_{11}$	$\alpha_{11} = \alpha_3 \cdot {}^{\mathrm{D}}\alpha_{\mathrm{eq}-\mathrm{S}}$
$12. CH_2D + D \rightarrow CH_2D_2$	$k_{12} = \frac{1}{2} k_r \alpha_{12}$	$\alpha_{12} = \alpha_4 \cdot {}^{D2}\gamma_{eq-P} \cdot {}^{D}\alpha_{eq-P} \cdot {}^{D}\alpha_{eq-S}$
13. $CHD_2 + H \rightarrow CH_2D_2$	$k_{13} = k_r  \alpha_{13}$	$\alpha_{13} = \alpha_5 \cdot {}^{D2}\gamma_{eq-S} \cdot {}^{D}\alpha_{eq-S} \cdot {}^{D}\alpha_{eq-S}$
$14. {}^{13}CH_3 + H \rightarrow {}^{13}CH_4$	$k_{14} = \frac{1}{2} k_r \alpha_{14}$	$\alpha_{14} = \alpha_6 \cdot {}^{13}\alpha_{eq}$
$15.\ ^{13}CH_3 + D \rightarrow ^{13}CH_3D$	$k_{15} = \frac{1}{2} k_{\rm r} \alpha_{15}$	$\alpha_{15} = \alpha_7 \cdot {}^{13\mathrm{D}} \gamma_{\mathrm{eq}-\mathrm{P}} \cdot {}^{13} \alpha_{\mathrm{eq}} \cdot {}^{\mathrm{D}} \alpha_{\mathrm{eq}-\mathrm{P}}$
$16. {}^{13}CH_2D + H \rightarrow {}^{13}CH_3D$	$k_{16} = k_{\rm r}  \alpha_{16}$	$\alpha_{16} = \alpha_8 \cdot {}^{13\mathrm{D}} \gamma_{\mathrm{eq}} \cdot {}^{13} \alpha_{\mathrm{eq}} \cdot {}^{\mathrm{D}} \alpha_{\mathrm{eq}} \cdot {}^{\mathrm$

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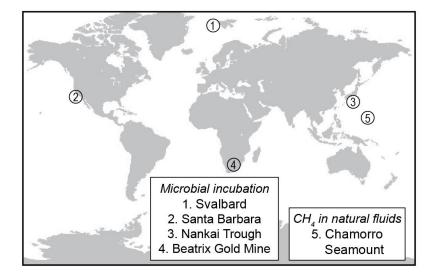




Fig. 1 Global map showing sampling sites for this study. Marine sediments or fracture fluids
were collected for slurry or fluid incubations at (1) Svalbard methane seeps, (2) Santa Barbara
Channel methane seeps, (3) Nankai Trough (International Ocean Discovery Program Hole
C0023A), and (4) Beatrix Gold Mine, South Africa. Methane in natural fluids was collected from
a sub-seafloor borehole observatory (Ocean Drilling Program Hole 1200C) on (5) South Chamorro
Seamount, a serpentinite mud volcano in the Mariana forearc.

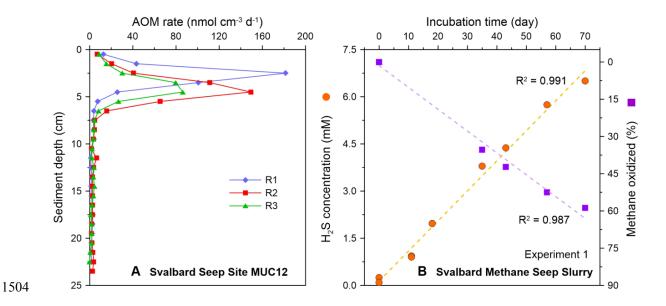
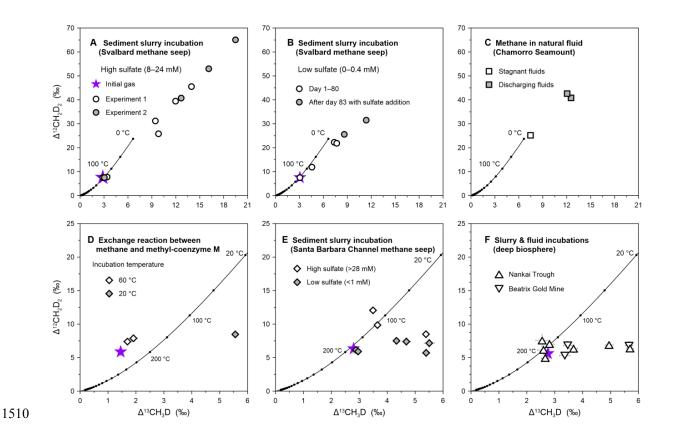


Fig. 2 Geochemistry of the Svalbard methane seep sediment and slurry. (A) *Ex-situ* AOM rates determined by <sup>14</sup>CH<sub>4</sub> radiotracer at Site MUC12 offshore Svalbard (Melaniuk et al., 2022). R1–3 represents replicate one through three. (B) Evolution of dissolved sulfide concentration and percentage of methane oxidized in the Svalbard methane seep sediment slurry with a starting sulfate concentration of 20 mM. Linear regressions are shown in panel B.



1511 Fig. 3 Clumped isotope data of residual methane in AOM incubation experiments and natural fluids. (A–B) Incubation of the Svalbard methane seep sediment slurry with high sulfate 1512 concentration (8-24 mM; panel A) and low sulfate concentration (0-0.4 mM; panel B). (C) 1513 Methane in natural fluids collected from the South Chamorro Seamount. (D) Exchange reaction 1514 1515 between methane and methyl-coenzyme M. (E) Incubation of the Santa Barbara Channel methane seep sediment slurry. (F) Incubations of the Nankai Trough sediment slurry and Beatrix Gold Mine 1516 fracture fluid from the deep biosphere. The purple stars represent the initial tank gas. The solid 1517 black line depicts theoretical thermodynamic equilibrium abundances of methane isotopologues, 1518 1519 along with corresponding temperatures. Error bars are  $1\sigma$ . Note different scales between panels 1520 A–C and D–F.

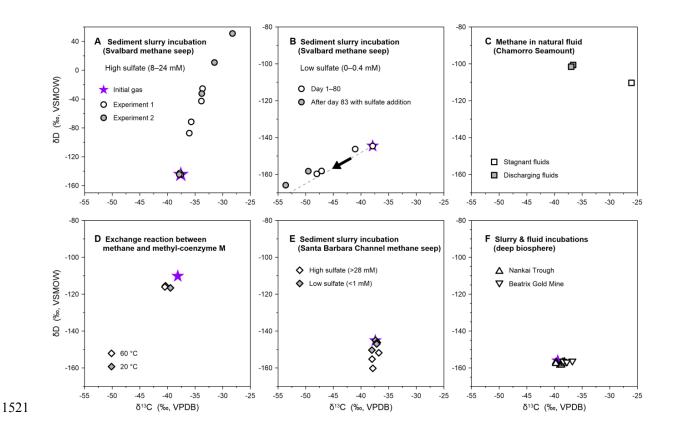


Fig. 4 Bulk isotope data of residual methane in AOM incubation experiments and natural 1522 1523 fluids. (A-B) Incubation of the Svalbard methane seep sediment slurry with high sulfate concentration (8-24 mM; panel A) and low sulfate concentration (0-0.4 mM; panel B). The arrow 1524 and dashed line in panel B indicate the evolution of residual methane towards isotope equilibrium 1525 among CH<sub>4</sub>, HCO<sub>3</sub><sup>-</sup> and H<sub>2</sub>O at 4 °C (Zhang et al., 1995; Turner et al., 2021). (C) Methane in 1526 1527 natural fluids collected from the South Chamorro Seamount. (D) Exchange reaction between methane and methyl-coenzyme M. (E) Incubation of the Santa Barbara Channel methane seep 1528 sediment slurry. (F) Incubations of the Nankai Trough sediment slurry and Beatrix Gold Mine 1529 1530 fracture fluid from the deep biosphere. The purple stars represent the initial tank gas. Uncertainties of  $\delta^{13}$ C and  $\delta$ D values are encompassed by individual data points. 1531

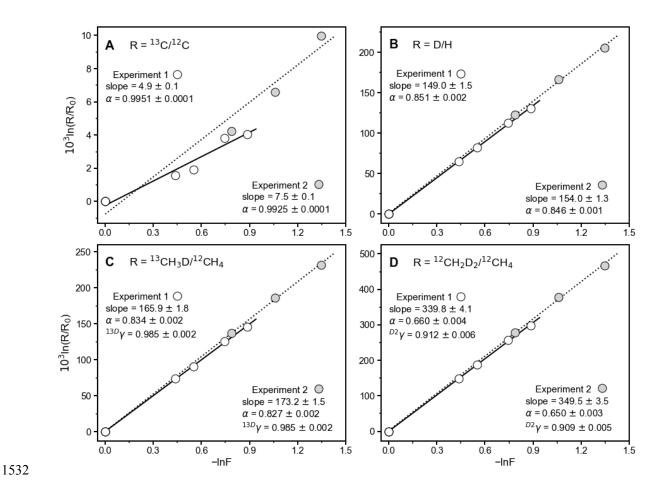
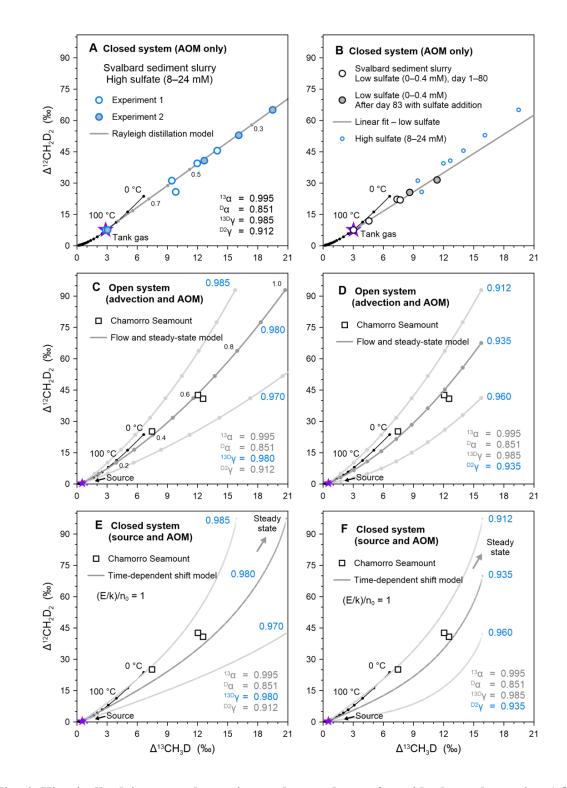
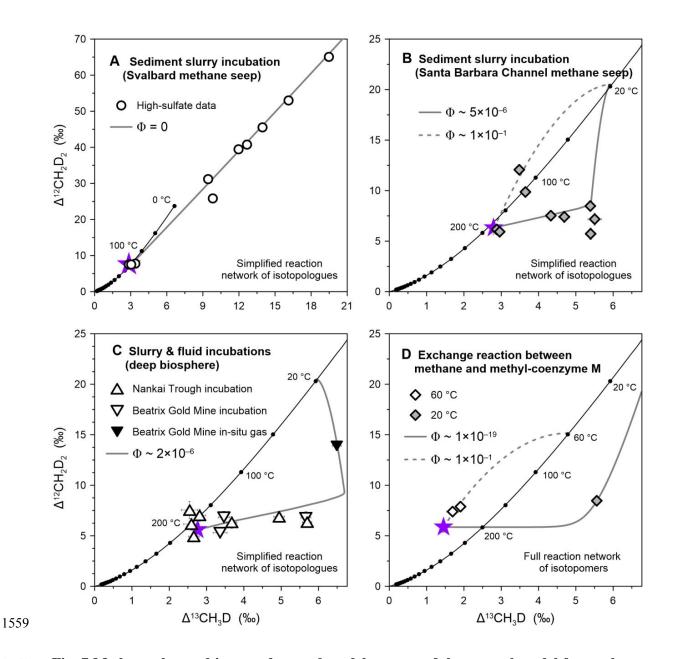


Fig. 5 Evolution of methane isotopologue ratios during methane consumption in the incubation of the Svalbard methane seep sediment slurry with high sulfate concentration (8– 24 mM). Data from experiments 1 and 2 are shown in open and filled circles, respectively. F is the fraction of methane remaining and R is the ratio of isotopologues in the gas phase.  $\alpha$  and  $\gamma$ represent the kinetic isotope fractionation factor and clumped isotopologue fractionation factor, respectively. The linear regressions and  $2\sigma$  errors are calculated by the weighted least square method of York et al. (2004).



1541 Fig. 6 Kinetically-driven methane isotopologue data of residual methane in AOM
1542 incubation experiments and methane in natural fluids. (A–B) Residual methane during

1543 incubation of the Svalbard methane seep sediment slurry with high sulfate concentration (8-24 mM; panel A) and low sulfate concentration (0–0.4 mM; panel B). (C–F) Methane in natural fluids 1544 1545 from the South Chamorro Seamount. Grey lines depict simulation outputs of the Rayleigh 1546 fractionation model (A), open-system flow and steady-state model (C-D) and closed-system timedependent shift model (E-F). The fractionation factors used in each model are shown in each 1547 1548 diagram. The purple stars represent the initial tank gas in panel A, and the source methane in panels 1549 C-F, respectively. This source methane is adopted from Young et al. (2017), representing abiotic 1550 methane gas formed through  $Si_5C_{12}H_{36}$  decomposition experiments, which resembles methane production through serpentinization. In panel A, points along the Rayleigh fractionation line are 1551 1552 marked at intervals of 0.1 in f, the fraction of initial methane remaining, while in panel C, points 1553 along the open-system model are marked at intervals of 0.1 in  $\varphi$ , the fraction of methane removed via oxidation. The modeled trajectories in panels E–F remain largely the same as  $(E/k)/n_0$  increase 1554 from  $10^{-10}$  to  $10^{10}$  (data not shown), and we use 1 as an arbitrary representation. The solid black 1555 1556 line depicts theoretical thermodynamic equilibrium abundances of methane isotopologues, along with corresponding temperatures. Uncertainties of  $\Delta^{13}$ CH<sub>3</sub>D and  $\Delta^{12}$ CH<sub>2</sub>D<sub>2</sub> values are 1557 1558 encompassed by individual data points.



**Fig. 7 Methane clumped isotope data and model outputs of the general model for methane isotopologue fractionation due to AOM.** (A) Comparison of our simplified model with incubations of Svalbard methane seep sediment slurry showing the model fits the data. (B) Comparison between Santa Barbara Channel methane seep incubations and our simplified model with inclusion of low turnover from CH<sub>4</sub> to CH<sub>3</sub> of about 1%, defined as [CH<sub>3</sub>]/[CH<sub>4</sub>] at steady state. (C) Comparison between deep biosphere incubations and our simplified model. The *in-situ* 

1566 methane gas from the Beatrix Gold Mine is shown for comparison (Young et al., 2017). The model in panels A-C uses 16 reactions involving 12 isotopologues and measured fractionation factors of 1567 1568 the Svalbard incubation (Table 2). (D) Comparison of enzymatically-mediated exchange of methane molecules with our model prediction using the full 280 reactions involving 50 isotopomer 1569 species and simple rate constants composed of square roots of inverse ratios of reduced masses. 1570 1571 Reversibility ( $\phi$ ) is determined by the ratio of the reverse rate constant to the forward rate constant 1572  $(k_r/k_f)$ . Where  $\Phi$  is closer to one, oxidation is more reversible, and vice versa. The solid black line depicts theoretical thermodynamic equilibrium abundances of methane isotopologues, along with 1573 corresponding temperatures. Error bars are  $1\sigma$ . 1574

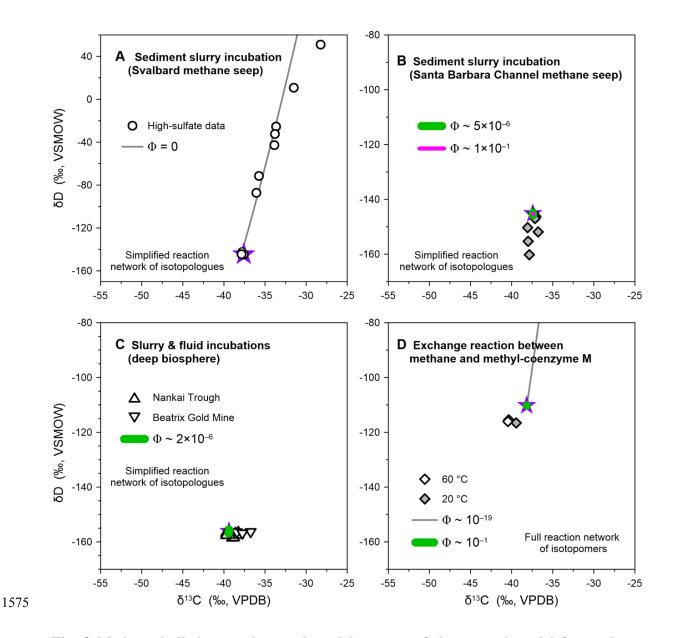
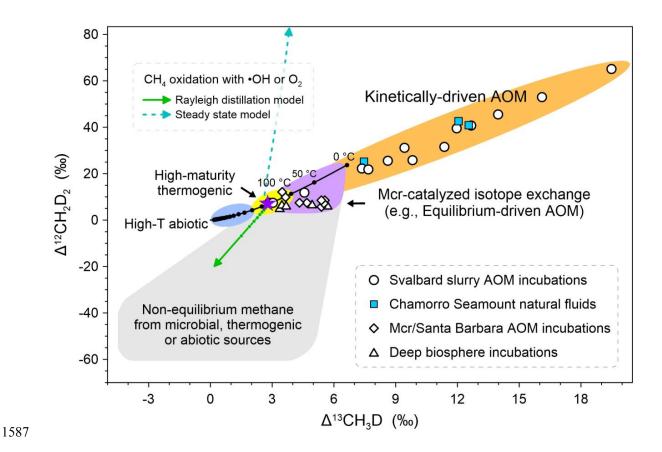


Fig. 8 Methane bulk isotope data and model outputs of the general model for methane isotopologue fractionation due to AOM. (A) Comparison of our simplified model with incubations of Svalbard methane seep sediment slurry showing the model fits the data. (B) Comparison between Santa Barbara Channel methane seep incubations and our simplified model. (C) Comparison between deep biosphere incubations and our simplified model. The model in panels A–C uses 16 reactions involving 12 isotopologues and measured fractionation factors of

1582	the Svalbard incubation (Table 2). (D) Comparison of enzymatically-mediated exchange of
1583	methane molecules with our model prediction using the full 280 reactions involving 50 isotopomer
1584	species and simple rate constants composed of square roots of inverse ratios of reduced masses.
1585	Reversibility ( $\Phi$ ) is determined by the ratio of the reverse rate constant to the forward rate constant
1586	$(k_r/k_f)$ .



1588 Fig. 9 A schematic representation of methane clumped isotope signatures of diverse 1589 methane sources, as well as kinetically- and equilibrium-driven AOM. Circle symbols depict 1590 residual methane during incubations of the Svalbard methane seep sediment slurry, while methane 1591 data of natural fluids from the South Chamorro Seamount are shown as square symbols. Rhombus 1592 symbols illustrate residual methane from the Mcr exchange experiment and incubations of the Santa Barbara Channel sediment slurry. Deep biosphere incubations are shown as triangle symbols. 1593 1594 The solid black line depicts theoretical thermodynamic equilibrium abundances of methane isotopologues, along with corresponding temperatures. The purple star represents the tank gas used 1595 in the slurry incubation experiments, and the zone of equilibrium-driven AOM is based on an 1596 1597 initial gas of thermogenic origin. For comparison, two modeled trajectories for methane oxidation 1598 with OH radical ( $\gamma$  for <sup>13</sup>CH<sub>3</sub>D = 0.999,  $\gamma$  for <sup>12</sup>CH<sub>2</sub>D<sub>2</sub> = 0.907; Haghnegahdar et al., 2017), the

1599 major methane sink in air, are shown: the solid line is the Rayleigh distillation model (cf. Fig. 6A), 1600 the dashed line is the steady-state closed-system model ((E/k)/ $n_0 = 1$ ; cf. Fig. 6E). The trajectories

- 1601 for aerobic oxidation of methane are similar to those of  $CH_4 + OH$  reaction (Krause et al., 2022).
- 1602 The zonation of methane sources is adopted from Young et al. (2017), Young (2019), Dong et al.
- (2021), etc. All the plotted data are from this study. The reader is referred to the text forinterpretation of the figure.