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Description of a new species of *Piriana* Santos-Silva, Galileo & McClarin, and notes and new records in Cerambycinae and Lamiinae from the Neotropical Region (Coleoptera, Cerambycidae)

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Neotropical region;
South America.

Abstract. – *Piriana limai* sp. nov. (Lamiinae, Colobotheini) is described from Brazil (Acre) and compared with *P. tricusps* (Belon, 1903) and other similar species. Three new country records are provided for Brazil: *Basiptera castaneipennis* Thomson, 1864 (Cerambycinae, Basipterini); *Hemilomecopterus alienus* Martins & Galileo, 2004 (Lamiinae, Hemilophini); and *Xenofrea durantoni* Néouze & Tavakilian, 2005 (Lamiinae, Xenofreini). Five new Brazilian state records from Acre are provided: *Dodecosis saperdina* Bates, 1867 (Cerambycinae, Dodecosini); *Ozineus elongatus* Bates, 1863 (Lamiinae, Acanthocinini); *Iarucanga mimica* (Bates, 1866) (Lamiinae, Hemilophini); *Isomerida albicollis* (Laporte, 1840) (Lamiinae, Hemilophini); and *Lycidola batesi* Martins & Galileo, 1991 (Lamiinae, Hemilophini). The authorship of *Lycidola batesi* is corrected, and notes on this species and *Lycidola simulatrix* Bates, 1866 and *Lycidola palliata* (Klug, 1825) are provided. Additionally, chromatic variations in *Hemilomecopterus alienus* are reported.

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Introduction

Except for *Basiptera castaneipennis* Thomson, 1864, all specimens used in this work come from the Brazilian Amazon region. In recent years, the Brazilian Amazon has been devastated at an unprecedented pace. It is likely that some species were lost without even being known to science. Even so, this forest still reveals a large number of discoveries, whether new species or expanding the geographic distribution of already known species.

- *Basiptera* Thomson, 1864 includes only one species, known from South America;
- *Dodecosis* Bates, 1867 has only two species, distributed in the Amazon region;
- *Piriana* Santos-Silva, Galileo & McClarin, 2018 includes 10 species distributed from Nicaragua to northern South America;
- *Ozineus* Bates, 1863 is another genus with a single known species, distributed from southern Central America to northern South America;
- *Hemilomecopterus* Martins & Galileo, 2004 has only one known species, occurring in northern South America;
- *Iarucanga* Martins & Galileo, 1991 has three species distributed in northern South America;
- *Isomerida* Bates, 1866 includes 19 species known from Nicaragua to southern South America;
- *Lycidola* Thomson, 1864 has eight species distributed from Honduras to southern South America.

All the above information is from Monné (2023 a, b), Tavakilian & Chevillotte (2022), and Bezark (2022).

Material and methods

Photographs were taken at MZSP with a Canon EOS Rebel T3i DSLR camera, Canon MP-E 65 mm f/2.8 1-5X macro lens, controlled by Zerene Stacker AutoMontage software. Measurements were taken in “mm” using an ocular Hensoldt/Wetzlar - Mess 10 in the Leica MZ6 stereomicroscope, which was also used in the study of the specimens.

The references on the species are restricted to the original description. The material examined is restricted to the specimens used to establish the new records.

The acronyms used in the text are as follows:

- INPA: Instituto Nacional de Pesquisas da Amazônia, Amazonas, Manaus, Brazil.
- MZSP: Museu de Zoologia, Universidade de São Paulo, São Paulo, Brazil.

Results

CERAMBYCIDAE Latreille, 1802

CERAMBYCINAE Latreille, 1802

BASIPTERINI Fragoso, Monné & Seabra, 1987

Basiptera Thomson, 1864

Reviewers:

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***Basiptera castaneipennis* Thomson, 1864**

(Fig. 18)

Basiptera castaneipennis Thomson, 1864: 205.

Remarks. – Thomson (1864) described *B. castaneipennis* based on a single specimen from Mexico (no further details). After the original description, the species has never been recorded in Mexico, which suggests that there was an error in the type locality of the species. Burmeister (1865) described *Trachyderes sanguinolentus* from Argentina (Santiago del Estero). According to Monné (2023a) and Tavakilian & Chevillotte (2022), *Trachyderes sanguinolentus* was described based on syntypes. However, Burmeister (1865) only provided a single measure and did not suggest that there were more specimens. Dohrn (1878) described *Leptura (Sibylla) martialis* based on a single specimen from Argentina (Córdoba). Aurivillius (1912) synonymized these two species with *B. castaneipennis*. Currently, the species is known from Bolivia, Argentina, and Paraguay (Monné 2023a; Tavakilian & Chevillotte 2022).

Material examined. – BRAZIL (new country record), Mato Grosso do Sul: Corumbá, 19°34'11"S 57°01'08"W, 1 ♀, 2.XI-2.XII. 2011, Lamas and equip leg. (MZSP 52519).

DODECOSINI Aurivillius, 1912***Dodecosis* Bates, 1867*****Dodecosis saperdina* Bates, 1867**

(Fig. 19)

Dodecosis saperdina Bates, 1867: 27.

Remarks. – This species was described based on a single specimen from Brazil (Pará). Currently it is known from French Guiana and Brazil (Amapá, Amazonas, Pará, Rondônia, Mato Grosso) (Monné 2023a; Tavakilian & Chevillotte 2022).

Material examined. – BRAZIL, Acre (new state record): Senador Guimard, 1 ♀, no date and collector indicated (MZSP 52520).

LAMIINAE Latreille, 1825**ACANTHOCININI** Blanchard, 1845***Ozineus* Bates, 1863*****Ozineus elongatus* Bates, 1863**

(Fig. 17)

Ozineus elongatus Bates, 1863: 278.

Remarks. – This species was described based on a single specimen from Brazil (Amazonas, Tefê). It is currently the only species of the genus (Monné *et al.* 2020) and is known from Panama, French Guiana, and Brazil (Amazonas, Pará, Rondônia) (Monné 2023b; Tavakilian & Chevillotte 2022).

Material examined. – BRAZIL, Acre (new state record): Bujari, FES Antimary, 9°20'01"S 68°19'17"W, 1 ♂, 21.X-4.XI.2016, E.F. Morato & J.A. Rafael leg. (MZSP 52517).

COLOBOTHEINI Thomson, 1860***Piriana* Santos-Silva, Galileo & McClarin, 2018*****Piriana limai* sp. nov.**

(Fig. 1-11)

ZooBank: <http://zoobank.org/4A914B1B-A307-4020-B59E-723F5EE1F847>

Holotype, ♂, BRAZIL, Acre: Senador Guimard, no date and collector indicated (INPA).

Paratypes

BRAZIL, Acre: 1 ♀, same data as holotype (MZSP 52512); Bujari, FES Antimary, 9°20'01"S 68°19'17"W, 1 ♀, 21.X-4.XI.2016, E.F. Morato & J.A. Rafael leg. (MZSP 52513).

Description of the holotype (Fig. 1-4, 7, 9)

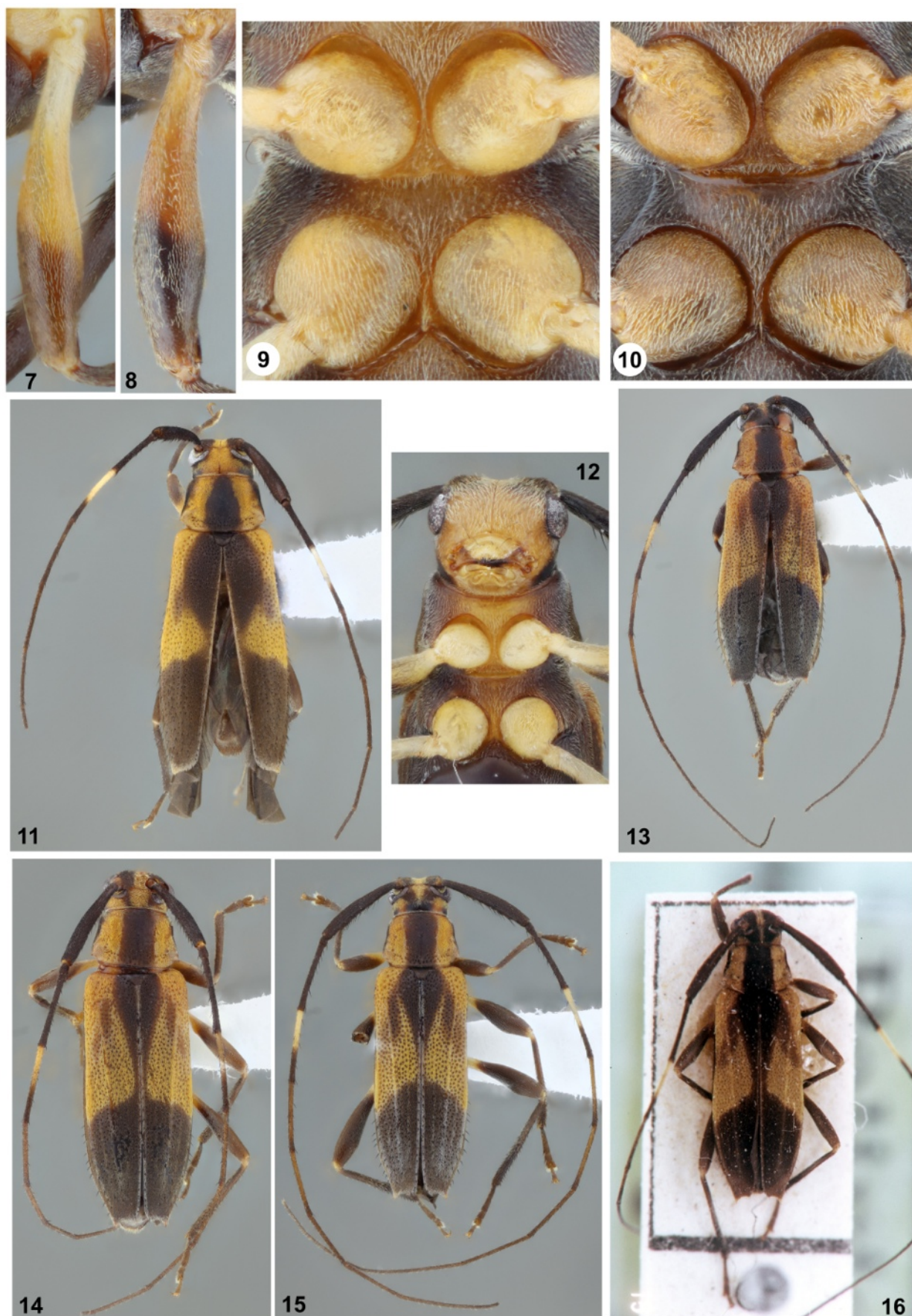
Coloration. – Head capsule orangish, except dark-brown antennal tubercles, area between eyes and base of antennae, and transverse, wide band behind lower eye lobes, and brownish superior area on frons close eyes; anteclypeus pale orange close to postclypeus, yellowish-white close to labrum; labrum pale orange except yellowish-white apex; ventral mouthparts orangish, except palpomeres yellowish-brown with apex narrowly yellowish-white; scape, pedicel, and antennomere III dark brown; antennomere IV pale yellow on basal 2/3, dark brown on apical third; antennomere V reddish brown on basal 3/4, dark brown on apical quarter; antennomere VI reddish brown on basal 2/3, dark brown on apical third; antennomeres VII–IX dark brown (missing part of X and entire XI on right antenna, and part of VIII and entire IX–XI on left antenna). Pronotum orangish brown, with wide dark-brown central band, gradually widened from anterior to posterior margin. Sides of prothorax orangish close to pronotum, dark brown on remaining surface. Prosternum dark brown laterally, orangish brown centrally; prosternal process orangish brown. Mesoventrite reddish brown laterally, orangish brown centrally; mesoventral process orangish brown. Mesanepisternum, mesepimeron, metanepisternum, and metaventrite dark brown, except dark orangish brown intercoxal process of metaventrite. Scutellum dark brown. Elytra dark orangish brown on anterior 3/5, except large dark brown macula from base to middle close to suture, with its outer margin rounded, and its apex brownish, and large, longitudinal band close to epipleural margin, macula dark brown on its anterior half, brownish on its posterior half, from near suture to about middle, with its superior margin rounded; posterior 2/5 dark brown, except yellowish apex. Pro- and mesocoxae mostly orangish brown; metacoxae mostly dark reddish brown; trochanters mostly yellowish-white; femora yellowish-white on basal quarter, orangish brown on wide central area, and brown on remaining surface; protibiae brown, slightly darker basally; meso- and metatibiae dark brown basally, slightly reddish-brown before middle, brown on remaining surface; tarsomeres I–II brown; tarsomere III brownish basally, gradually orangish toward apex; tarsomere IV and about basal 4/5 of V brownish, and apex of V orangish; tarsal claws orangish about basal half, brownish on remaining surface. Abdominal ventrites 1–2 dark brown laterally, orangish-brown with irregular dark brown areas interspersed centrobasally; 3–4 dark brown laterally, reddish brown centrobasally, orangish-brown centroapically; 5 dark brown laterally, brown centrally.

Head. – Frons densely, finely punctate; with abundant yellowish-white pubescence not obscuring integument, except glabrous median groove. Area between antennal tubercles and upper eye lobes with transverse glabrous sulcus, forming cross with median groove; with dense yellow pubescence; remaining surface of vertex and area behind upper eye lobes with dense yellow pubescence, except sparse brownish pubescence close to eye. Area behind lower eye lobes with abundant yellowish-white pubescence not obscuring integument close to eye, glabrous close to prothorax. Genae slightly longer than lower eye lobe; with abundant yellowish-white pubescence not obscuring integument, except glabrous apex; with a few long, erect yellowish-white setae interspersed. Wide central area of postclypeus with sparse yellowish-white pubescence, one long, thick, erect dark brown seta on each side, and sparse, long, erect yellowish-white setae interspersed laterally. Sides of postclypeus glabrous. Labrum with abundant yellowish-white pubescence not obscuring integument, with long, erect setae of same color interspersed, and transverse row of sparse, thick, erect dark brown setae about middle. Gulamentum glabrous, except somewhat sparse yellowish-white pubescence anteriorly. Distance between upper eye lobes 0.33 times distance between outer margins of eyes; in frontal view, distance between lower eye lobes 0.62 times distance between outer margins of eyes. Antennae (from base of scape to apex of antennomere IX) 2.6 times elytral length, reaching elytral apex at base of antennomere VI. Scape with somewhat abundant brownish pubescence not obscuring integument; with abundant, long, erect dark-brown setae ventrally, some setae with yellowish apex. Pedicel with abundant brownish pubescence not obscuring integument, and a few long, erect dark brown setae ventrally. Antennomere III with abundant brownish pubescence not obscuring integument, with short, decumbent, both yellowish and whitish setae interspersed throughout, somewhat long, erect, thick dark-brown setae ventrally, and a few somewhat short, erect, thick dark brown setae dorsally. Antennomere IV with abundant whitish pubescence not obscuring



Figures 1-6. *Piriana limai* sp. nov.

1-4. Holotype ♂. 1. Dorsal habitus. 2. Ventral habitus. 3. Head, frontal view. 4. Lateral habitus. 5-6. Paratype ♀. 5. Dorsal habitus. 6. Ventral habitus.



Figures 7-16. *Piriana* spp.

7-8. *Piriana limai* sp. nov., profemora. 7. Holotype ♂. 8. Paratype ♀. 9-11. *Piriana limai* sp. nov. 9. Holotype ♂, prosternal and mesoventral processes. 10. Paratype ♀, prosternal and mesoventral processes. 11. Paratype ♀, dorsal habitus. 12-16. *Piriana tricuspis* (Belon, 1903). 12. ♂ from Bolivia (Santa Cruz), prosternal and mesoventral processes. 13. ♀ from Bolivia (Santa Cruz), dorsal habitus. 14. ♀ from Bolivia (Santa Cruz), dorsal habitus. 15. ♂ from Bolivia (Santa Cruz), dorsal habitus. 16. Syntype, dorsal habitus, by Jesus Santiago Moure.



Figures 17-19. Cerambycidae spp., dorsal habitus.

17. *Ozineus elongatus* Bates, 1863, ♂. **18.** *Basiptera castaneipennis* Thomson, 1864, ♀. **19.** *Dodecosis saperdina* Bates, 1867, ♀.

integument, and a few short, erect, thick dark brown setae interspersed dorsally and ventrally. Antennomeres V–X with abundant yellowish-white pubescence not obscuring integument, short, erect white setae interspersed throughout, and a few short, erect dark setae interspersed on V–VIII.

Antennal formula based on length of antennomere III:

– Scape = 1.56. – Pedicel = 0.16. – IV = 1.12. – V = 1.00. – VI = 0.96. – VII = 0.94. – VIII = 1.12. – IX = 1.26.

Thorax. – Prothorax wider than long; anterior constriction narrow, well marked; sides rounded close to anterior constriction, subparallel-sided on remaining anterior third, then gradually widened toward lateral tubercles, then distinctly narrowed from lateral tubercles toward posterolateral angles; anterior region of the lateral tubercles is an enlargement of the margin of the prothorax ending in a small sharp point directed upward, located on posterior fifth. Pronotum somewhat abruptly inclined close to anterior margin, especially laterally; posterior quarter transversely depressed, abundantly, coarsely punctate; sides of area between middle and posterior quarter sparsely, coarsely punctate; remaining surface densely, minutely punctate; sides with abundant yellow pubescence almost obscuring integument, except slightly sparser pubescence close to sides of prothorax on anterior third, and abundant yellowish-white pubescence close to lateral tubercles of prothorax; dark-brown band with abundant dark-brown pubescence not obscuring integument. Sides of prothorax with anterior sulcus glabrous; superior half with dense yellow pubescence on anterior 2/3, yellowish-white on posterior third; inferior half with abundant yellowish-white pubescence partially obscuring integument; with a few long, erect dark brown setae after lateral tubercle. Prosternum with abundant yellowish-white pubescence not obscuring integument. Prosternal process (Fig. 9) almost laminiform centrally; with abundant yellowish-white pubescence not obscuring integument on anterior half, slightly sparser on posterior half. Ventral surface of meso- and metathorax with abundant grayish-white pubescence partially obscuring integument, except yellowish-white pubescence on central area of mesoventrite and on mesoventral process, and yellowish pubescence on posterocentral region of metaventrite. Mesoventral process (Fig. 9) sublaminiform, parallel-sided on posterior half. Scutellum with somewhat abundant brownish pubescence not obscuring integument.

Elytra. – Abundantly, coarsely punctate on anterior 2/3, punctures sparser, finer on posterior third; apex obliquely truncate, with outer angle spiniform and sutural angle rounded; with dense yellow pubescence on light anterior area, dense brown pubescence on dark areas, and abundant yellowish-white pubescence on apex; with somewhat abundant and long, erect dark-brown setae interspersed throughout.

Legs. – Femora with abundant yellowish-white pubescence not obscuring integument. Protibiae with somewhat sparse yellowish pubescence dorsally and laterally, except apex of sides with somewhat abundant, bristly and thick yellowish-brown pubescence, and abundant yellowish-brown pubescence ventrally, pubescence gradually bristly and denser from basal quarter; posterior third of dorsal surface with short, erect, thick dark-brown setae interspersed. Mesotibiae with abundant yellowish-white pubescence not obscuring integument on basal half, bristly, then gradually denser and yellowish-brown toward apex, except anterior region of dorsal sulcus with dense yellowish-white pubescence. Metatibiae with somewhat sparse yellowish-white pubescence on basal third, gradually denser and bristly toward apex on posterior third; with short, erect both yellowish-brown and dark brown setae interspersed, erect setae distinctly more abundant on posterior third. Dorsal surface of tarsomeres with abundant yellowish-white pubescence, except sparser pubescence on tarsomere V.

Abdomen. – Ventrites 1–4 with abundant whitish pubescence not obscuring integument; ventrite 5 with somewhat sparse whitish pubescence on basal third, pubescence distinctly sparse on remaining surface, with short, subdecumbent dark brown setae interspersed on posterior third.

Female (Fig. 5–6, 8, 10–11). – Similar to male, differing by antennae slightly shorter (both paratypes without antennomere XI), 2.3 times elytral length from base of scape to apex of antennomere IX, 2.6 times from base of scape to apex of antennomere X, reaching elytral apex slightly before middle of antennomere VI; lateral tubercles of prothorax more distinctly projected; prosternal process (Fig. 10) slightly wider, not sublaminiform centrally; mesoventral processes (Fig. 10) slightly wider on posterior half, not sublaminiform; and meso- and metafemoral club narrower (Fig. 5–6).

Chromatic variation. – Frons and vertex reddish brown; antennomeres V–XI entirely dark brown; anterolateral dark area distinctly fused with posterior dark area close to epipleural margin; dorsal dark-brown areas fused along suture; anterodorsal dark-brown macula with wide longitudinal projection on its posterior half; tibiae entirely dark; abdominal ventrites dark brown, except orangish brown apex of 1–4.

Dimensions (mm) (holotype ♂ / paratypes ♀).

– Total length, 5.15/5.60–6.10;
– Prothoracic length, 0.85/0.90–0.95;
– Anterior prothoracic width, 0.90/1.00–1.05;
– Posterior prothoracic width, 1.05/1.15–1.20;
– Maximum prothoracic width, 1.20/1.40–1.50;
– Humeral width, 1.40/1.60–1.65;
– Elytral length, 3.80/4.10–4.40.

Etymology. – The new species is dedicated to William Lima, an expert on Lampyridae (Coleoptera) for sending the specimens of the new species for study.

Remarks. – *Piriana limai* sp. nov. is similar to *P. tricusps* (Belon, 1903), but differs as follows: dorsal surface of the head without oblique brown pubescent band from upper eye lobes to vertex; longitudinal blackish central band on the pronotum distinctly widened from anterior to posterior margin; longitudinal carina on dorsal surface of the elytra barely marked; elytral apex obliquely truncate and sutural angle rounded and not projected; mesoventral process in male sublaminiform (Fig. 9). In *P. tricusps* (Fig. 12–16), the dorsal surface of the head has oblique brown pubescent band from upper eye lobes to vertex, longitudinal blackish central band on the pronotum subparallel-sided, longitudinal carina on dorsal surface of the elytra well marked, elytral apex not or slightly obliquely truncate with the sutural angle somewhat projected, and the mesoventral process is not sublaminiform (Fig. 12). *Piriana limai* sp. nov. differs from *P. birai* Santos-Silva, Galileo & McClarin, 2018 and *P. nearnsi* Santos-Silva, Galileo & McClarin, 2018, (see photographs in the original descriptions and Bezark 2023) by the elytral apex obliquely truncate and the sutural angle not projected (sinuous and with sutural angle projected in these species); and from *P. svachai* Santos-Silva, Galileo & McClarin, 2018 (see photographs in the original description and Bezark 2023) by the sutural angle of the elytra not projected (projected in *P. svachai*), and the mesoventral process in male sublaminiform (not sublaminiform in males of *P. svachai*).

HEMILOPHINI Thomson, 1868

Hemilomecopter Martins & Galileo, 2004

Hemilomecopter alienus Martins & Galileo, 2004

(Fig. 20–27)

Hemilomecopter alienus Martins & Galileo, 2004: 42.

Remarks. – *Hemilomecopter alienus* was described based on a single female from Colombia (Amazonas). Galileo & Martins (2005) recorded the species from Ecuador (Napo) based on two males. The shape of the head, with the genae very elongated and frons distinctly oblique, makes it easy to recognize the species.

According to Martins & Galileo (2004) (translated): “Head black, except macula on occipital region, on inferior region of frons, and genae which are reddish.” In the female from Brazil, the dorsal surface of the head is entirely black, frontal area of the genae is mostly brown, the apex of the genae is brownish, and the inferior region of the frons and the wide central area of the postclypeus are orangish brown. Still according to them (translated): “Antennae black” (in the female from Brazil, the dorsal apex of the scape is brown, the apical half of the dorsal surface and most of the sides of the apical third are yellowish brown, apical quarter of antennomere III is brown, antennomere IV



Figures 20-23. *Hemilomecopterus alienus* Martins & Galileo, 2004, ♀ from Brazil.
20. Ventral habitus. 21. Lateral habitus. 22. Head, lateral view. 23. Head, frontal view.

and basal half of V are brownish, and the apical half of antennomere V and antennomeres VI–VIII are dark brown); “Elytra with the basal half orangish yellow, except the humeri which are black; with transverse black band after middle, followed by another transverse yellowish band, covered with dense pubescence of same color; and reddish-brown apical fifth, covered with yellowish pubescence” (in the female from Brazil, the dark central band is dark brown, starting before middle of the elytra; the anterior region is yellowish brown; there is a dark longitudinal band from humerus to the transverse central band, not reaching epipleural margin, black basally, gradually browner on its apical region; the sides of area from the transverse dark-brown band to near apex is blackish; remaining apical half is orangish brown except the brown punctures; and the pubescence is yellowish-white, except sparser and brown pubescence on dark-brown central area); “Fore- and middle legs yellowish” (in the female from Brazil, the dorsal surface of profemora is brown basally and dorsal surface of protibiae is dark brown on wide central area; mesofemora are blackish except yellowish-brown apex of dorsal surface and sides, this area longer on sides, and yellowish-brown apical third of ventral surface; and mesotibiae is blackish basally, and most dark brown on remaining surface); “Metafemora black, except the ventral surface” (in the female from Brazil, only the apical third of the ventral surface is yellowish-brown); “Mesoventrite black with the mesoventral process yellowish” (in the female from Brazil, the mesoventral process is brown on sides of anterior half and orangish brown on remaining surface); “Mesepimeron, metanepisternum, and sides of metaventrite black” (in the female from Brazil, the mesepimeron is brownish, the metanepisternum and sides of metaventrite are reddish brown); “abdominal ventrites 1 to 3 yellowish, 4 and 5 brown (in the female from Brazil, the sides of abdominal ventrite 4 is brownish basally, gradually, dark brown toward apex, and 5 is entirely dark brown). Furthermore, in the description of the genus it was reported that the elytra in female are 3.6 times longer than the humeral width, while in the female from Brazil, the elytra are 3.2 times longer than the humeral width.

Material examined. – BRAZIL (new country record), Rondônia: Itapuã do Oeste, Floresta Nacional do Jamari, 9°15'36"S 62°54'46"W, 1 ♀, Z.F.M. Silva & J.A. Rafael leg. (MZSP 52516).

Iarucanga Martins & Galileo, 1991

Iarucanga mimica (Bates, 1866)

(Fig. 28)

Spathoptera mimica Bates, 1866: 369.

Remarks. – This species was described based on syntypes from Brazil (Amazonas). Currently it is known from Ecuador, Bolivia, French Guiana, and Brazil (Amazonas, Rondônia, Mato Grosso) (Monné 2023b; Tavakilian & Chevillotte 2022).

Material examined. – BRAZIL, Acre (new state record): Senador Guimard, 1 ♀, no date and collector indicated (MZSP 52515).

Isomerida Bates, 1866

Isomerida albicollis (Laporte, 1840)

(Fig. 29)

Hemilophus albicollis Laporte, 1840: 488.

Remarks. – This species was described based on a single specimen from French Guiana. Currently it is known from Costa Rica, Colombia, Ecuador, Peru, Bolivia, French Guiana, and Brazil (Amazonas, Pará, Mato Grosso) (Monné 2023b; Tavakilian & Chevillotte 2022). *Isomerida albicollis* shows great chromatic variation. According to Martins & Galileo (2014b) (translated): “We found in this species variability in the color of the elytral integument, in the presence or absence of dark regions on the

pronotum, and in the distribution of the white pubescence that covers the abdominal ventrites. We could not conclude whether this variability indicates forms of the same species (most likely) or closely related species.”

Material examined. – BRAZIL, Acre (new state record): Bujari, FES Antimary, 9°20'01"S 68°19'17"W, 1 ♀, 21.X-4.XI.2016, E.F. Morato & J.A. Rafael leg. (MZSP 52514).

Lycidola Thomson, 1864

Lycidola batesi Martins & Galileo, 1991

(Fig. 30)

Lycidola batesi Martins & Galileo, 1991: 554.

Remarks. – This species was described based on syntypes from Brazil (Amazonas and Pará). According to Bates (1866): “*Var.* Base of each elytron with a small fulvous spot in continuation of the thoracic stripe; lateral edge of the elytron also fulvous near the base (approaching *L. palliata*, Klug). Tapajos;” and, “The typical form not uncommon at Ega, on leaves. The var. found only on the banks of the Tapajos.” Aurivillius (1923) recorded:

“*simulatrix* Bates, Ann. Mag. Nat. Hist. (3) XVII, 1866, p. 368 (239). – Gah. Proc. South Lond. Ent. Soc. 1912–13, t. 9, f. 2, 3.”

ab. *Batesi* n. nom.

simulatrix var. Bates, Ann. Mag. Nat. Hist. (3) XVII, 1866, p. 368 (240).”

Martins & Galileo (1991) considered *L. simulatrix* ab. *batesi* as a distinct species and reported (translated): “Bates (1866) described *L. simulatrix* based on “a typical form,” from Tefê, Amazonas, with black humeri and elytra with the transverse central band white, and on a “variety,” from Tapajós, Pará, with orangish humeral macula and elytra with the transverse central band yellowish-white. Aurivillius (1923) named this variety as *Lycidola simulatrix* ab. *batesi*. The specimens now examined from Pará coincide with *batesi* and also with the original description and illustration of *L. palliata* (Klug), originally known from Bahia. It is very likely that *batesi* and *palliata* are synonyms, which can be confirmed by examining specimens from Bahia.” According to Martins & Galileo (2014a) on *L. batesi* (translated): “We did not examine specimens of *L. palliata* and found no differences, according to the original description and figure, to distinguish it from *L. batesi*. However, both have different geographical distributions, *L. palliata* was described from Bahia and assigned to Itatiaia, Rio de Janeiro (ZAJCIW, 1972a) and *L. batesi* from Pará. The description presented above for *L. palliata* agrees with *L. batesi*.” They also reported in remarks of *L. affinis* Martins & Galileo, 2012 (translated): “AURIVILLIUS (1923) changed the status of *Lycidola simulatrix* var. (BATES, 1866: 368) and named this variety as *L. batesi*, a species that occurs in Brazil (Amazonas and Pará).” However, *Lycidola simulatrix* var. (= *L. batesi*) occurs only in Brazil (Pará) and Bolivia.

It is probable that *Lycidola batesi* and *L. palliata* (Klug, 1825) are two names of the same species, although the original illustration by Klug (1825) shows the antennomere IV as very swollen. However, it is very likely that *L. simulatrix* is only a chromatic variation of *L. palliata*. To reinforce this opinion, photographs of specimens from French Guiana, identified as *L. simulatrix*, with orangish humerus were examined. It is very common to find in Bahia (Brazil) species of Cerambycidae that also occur in French Guiana. Apparently, the prothoracic and elytral shape in *L. palliata*/*L. batesi* is also somewhat variable. However, the synonymy, between the three species names, or between *L. batesi* and *L. palliata*, cannot be established without examining a larger number of specimens from the Amazon region and northeast and southeast Brazil.

It is important to note that *L. batesi* is not an available name in Aurivillius (1923) (ICZN 1999: Article 45.6.2). Therefore, *Lycidola batesi* is attributed to Martins & Galileo, 1991.

Currently, *L. simulatrix* is known from Bolivia, French Guiana and Brazil (Amapá, Amazonas, Rondônia); *L. batesi* from Bolivia and



Figures 24-27. *Hemilomecopterus alienus* Martins & Galileo, 2004.

24. ♀ from Brazil, dorsal habitus. 25-27. ♂ from Ecuador. 25. Lateral habitus. 26. Dorsal habitus. 27. Ventral habitus.

Brazil (Pará); and *L. palliata* from Brazil (Bahia, Rio de Janeiro) (Monné 2023b; Tavakilian & Chevillotte 2022).

While the eventual differences between *L. simulatrix*, *L. batesi*, and *L. palliata* remain to be found, there is no reason to consider the specimens from French Guiana as *L. simulatrix*. As, apparently, all specimens from that place have the humeri orangish or yellowish, they need to be considered as *L. batesi*. Therefore, *L. simulatrix* is excluded from the French Guiana fauna, and *L. batesi* is reported there. *L. batesi* is tentatively used instead of *L. palliata* due to the shape of the antennomere IV in Klug (1825), which is probably just an imprecise drawing.

Material examined. – BRAZIL, *Acre* (new state record): Bujari, FES Antimary, 9°20'01"S 68°19'17"W, 1 ♀, 21.X-4.XI.2016, E.F. Morato & J.A. Rafael leg. (MZSP 52518).

XENOFREINI Aurivillius, 1923

Xenofrea Bates, 1885

Xenofrea durantoni Néouze & Tavakilian, 2005

(Fig. 31)

Xenofrea durantoni Néouze & Tavakilian, 2005: 160.

Remarks. – This species was described from French Guiana and remains known only from that region of South America.

Material examined. – BRAZIL (new country record), *Amazonas*: Fonte Boa, 1 ♂, IX.1975, F.M. Oliveira leg. (Identified by G. Tavakilian) (MZSP 52521). *Pará*: Serra Norte (Serraria), 1 ♂, 21.VI.1985, H. Andrade leg. (MZSP 52522).

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Figures 28-31. Lamiinae spp., dorsal habitus.

28. *Iarucanga mimica* (Bates, 1866), ♀ from Brazil (Acre). **29.** *Isomerida albicollis* (Laporte, 1840), ♀ from Brazil (Acre). **30.** *Lycidola batesi* Martins & Galileo, 1991, ♀ from Brazil (Acre). **31.** *Xenofrea durantoni* Néouze & Tavakilian, 2005, ♂ from Brazil (Pará).

Résumé

Santos-Silva A., 2023. – Description d'une nouvelle espèce du genre *Piriana* Santos-Silva, Galileo & McClarin, et notes et nouveaux signalements de Cerambycinae et de Lamiinae de la région néotropicale (Coleoptera, Cerambycidae). *Faunitaxys*, 11(12): 1 – 12.

Piriana limai **sp. nov.** (Lamiinae, Colobotheini) est décrite du Brésil (Acre) et comparée à *P. tricuspidis* (Belon, 1903) et à d'autres espèces similaires. Trois nouvelles mentions nationales sont fournies pour le Brésil : *Basiptera castaneipennis* Thomson, 1864 (Cerambycinae, Basipterini) ; *Hemilomecopterus alienus* Martins & Galileo, 2004 (Lamiinae, Hemilophini); et *Xenofrea durantoni* Néouze & Tavakilian, 2005 (Lamiinae, Xenofreini). Cinq nouveaux enregistrements de l'état brésiliens d'Acre sont fournis : *Dodecosis saperdina* Bates, 1867 (Cerambycinae, Dodecosini) ; *Ozineus elongatus* Bates, 1863 (Lamiinae, Acanthocinini); *Iarucanga mimica* (Bates, 1866) (Lamiinae, Hemilophini); *Isomerida albicollis* (Laporte, 1840) (Lamiinae, Hemilophini); et *Lycidola batesi* Martins & Galileo, 1991 (Lamiinae, Hemilophini). La paternité de *Lycidola batesi* est corrigée et des notes sur cette espèce et *Lycidola simulatrix* Bates, 1866 et *Lycidola palliata* (Klug, 1825) sont fournies. De plus, des variations chromatiques chez *Hemilomecopterus alienus* sont signalées.

Mots-clés. – Coleoptera, Cerambycidae, Cerambycinae, Lamiinae, *Piriana*, taxonomie, morphologie, nouvelle espèce, description, région néotropicale, Amérique du Sud.

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