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Methylene blue removal from aqueous solutions using a biochar/gellan gum hydrogel composite: Effect of agitation mode on sorption kinetics

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ABSTRACT

Hydrogel membranes are prepared by casting a mixture of gellan gum (associated with PVA) and biochar pro-duced from a local Egyptian plant. The mesoporous material is characterized by a specific surface area close to 134 m² g⁻¹, a residue of 28 % (at 800 °C), and a pH_{PZC} close to 6.43. After grinding, the material is tested for Methylene Blue sorption at pH 10.5: sorption capacity reaches 1.70 mmol MB g⁻¹ (synergistic effect of the precursors). The sorption isotherms are fitted by both Langmuir and Sips eqs. MB sorption increases with tem-perature: the sorption is endothermic (Δ H°: 12.9 kJ mol⁻¹), with positive entropy (Δ S°: 125 J mol⁻¹ K⁻¹). Uptake kinetics are controlled by agitation speed (optimum ≈200 rpm) and resistance to intraparticle diffusion. The profiles are strongly affected by the mode of agitation: the equilibrium time (≈180 min) is reduced to 20–30 min under sonication (especially at frequency: 80 kHz). The mode of agitation controls the best fitting equation: pseudo-first order rate agitation for mechanical agitation contrary to pseudo-second order rate under sonication. The sorption of MB is poorly affected by ionic strength (loss <6 % in 45 g L⁻¹ NaCl solution). Desorption (faster than sorption) is completely achieved using 0.7 M HCl solution. At the sixth recycling, the loss in sorption is close to 5 % (≈ decrease in desorption efficiency). The process is successfully applied for the treatment of MB-spiked industrial solution: the color index decreases by >97 % with a sorbent dose close to 1 g L⁻¹; a higher dose is required for maximum reduction of the COD (60 % at 3 g L⁻¹).

Keywords: Mesoporous gellan gum (GG)/PVA-biochar composite Methylene blue sorption from alkaline solutions Sonication effects on kinetics Sorption/desorption kinetics

1. Introduction

The scarcity of natural water resources (especially under the effect of climate change and global warming), and the increasing demand for water in domestic, agricultural, and industrial uses explain the incentive politics for reclaiming wastewater for decontamination and re-use. One of the evident consequences consists of the reinforcement of national and regional regulations concerning the discharge of wastewater into the environment. Though the attention strongly focuses on the behavior of inorganic and metal ions in the environment, great attention is paid to organic pollutants such as drugs, pharmaceutical molecules but also dyes. Independently of their intrinsic toxicity, dyes may also affect water bodies through perturbations to photosynthetic activity (coloring

effect). These dual impacts may explain the intense research activity on the treatment of colored effluents. The processes for dye removal may involve precipitation and flotation [1–3], and solvent extraction [4–6]. Catalytic and photocatalytic degradation have been also widely documented [7–10]. It is noteworthy that the degradation of organic compounds by the catalytic process may, in certain cases, produce even more hazardous compounds than the precursors [11,12]. Sorption represents an alternative strategy for the recovery of dyes from aqueous solutions, using low-cost bio-based sorbents [13,14], chitosan, and derivatives [15–17]. Nano-based materials [18], resins [19–21], and silica [22–24] have also been referenced for their efficiency in dye removal. Activated carbon (AC) is an interesting resource for designing dye sorbents, taking advantage of its high specific surface area [25,26]. Biochar offers a cost-

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effective alternative to AC (obtained by pyrolysis of biomass) for the remediation of dyes from aqueous solutions [27-30]. Tan et al. [31] reviewed the numerous possibilities of the application of biochar for wastewater decontamination and reported some of the challenges to be ruled out for ensuring the development of biochar-based sorbents; including the practical operating conditions for sorption and desorption processing. Composite materials based on the reaction of biochar with other supports, or embedment in a polymer network represent some responses to these problems [32]. Currently, biochar and its derivatives have received special attention being intensively used in many applications in agronomy, construction, energy storage, environment, etc. [33]. These successful uses derive from their multifunctional properties (in terms of mechanical, thermal, catalytic characteristics, etc.). From the point of view of environmental applications, biochar has been used either alone or in combination with other materials to remove a wide range of contaminants, both organic and inorganic chemical species, from wastewater [34-37].

Gellan gum (GG) is a naturally occurring polysaccharide produced by Sphingomonas elodea as a linear extracellular polysaccharide. It is a linear heteropolysaccharide composed of D-glucose, D-glucuronic acid, and L-rhamnose repeating units with two acyl substituents, L-glyceryl, and acetyl, attached to the C-2 and C-6 positions of the O-3-linked 1,3-Dglucose residue [38]. Low acyl GG is produced via deacylation in an alkaline solution. Gellan polymer chains cannot create a tight double helix structure between chains because of bulky substituents. Since gellan gels are weak, soft, and elastic in their native form, reinforcements with additional materials (or complementary reactions) are usually required to achieve the desired mechanical properties [39]. It has already been tested for mass production (i.e., one company can produce over 500 tons of GG per year) [40]. Unlike other polysaccharides, GG maintains its gel state over a large range of pH in aqueous systems [41]. This feature offers exceptional acid-base stability, which makes it a good candidate to produce sorbents for the removal of organic contaminants at different pH values. In addition to being acidbase stable, GG exhibits very good thermal stability [42], performs very well in solutions with high ionic strength [43], and reacts well with organic dyes [44]. Despite these special properties, there has been little interest in using GG to remove contaminants from water until now. Examining the sorption capacity of GG or GG-based composites for contaminants and identifying the sorption mechanisms involved could pave the way for these materials to be used in green water and wastewater treatment technologies [45]. Therefore, GG hydrogel can have applications in the field of adsorption, due to its good biodegradability (at the end of its life cycle) and durability [46]. Gellan gum is a member of the broad exopolysaccharide family. In addition to hydrating in cold water with sequestrants, GG also hydrates in hot water in its low acyl form. When cooled, native high-acyl GG gives gels that are soft and elastic [45]. Combinations of GG with other materials can be used to control syneresis and form a range of textures from soft and elastic to firm and brittle.

In this context, the incorporation of biochar into the GG matrices may enhance the properties of the resulting composites because of the ability of biochar to affect the properties of the GG matrix. Gellan gum with biochar can gather the special advantages of biochar and GG including high surface area, high thermal and mechanical stability, and the flexibility of natural polymers. The use of natural materials (environmentally friendly) for the synthesis of sorbent composites has gained considerable interest from researchers in recent years. They are renewable, eco-friendly, biodegradable, and easily available.

In addition, the incorporation of polyvinyl alcohol (PVA) in the process promotes the formation of an interpenetrating network that contributes to improving mechanical properties and reducing uncontrolled swelling of polymer blends [47]. The film-forming properties are also well-known and useful for preparing membrane systems [48]. Therefore, the combination of dual polymers with the incorporation of biochar particles leads to new multifunctional materials with high

stability and promising sorption properties. These synergistic properties will be evaluated and compared to their precursors under different experimental conditions for Methylene Blue removal (as a model) for evaluating their potential in wastewater treatment.

Herein, the preferences for preparing a biochar composite are given to (a) the pyrolysis of *Orobanche aegyptiaca* biomass (endemic and Egyptian plant), and (b) the incorporation of the porous material into a biopolymer matrix (GG). Invasive *O. aegyptica* is of critical importance for local agriculture; therefore, the elimination and/or valorization of this harmful biomass is a strategic issue. Gelling of GG is processed by reaction with poly(vinyl alcohol) (PVA), which contributes to forming a polymeric network. Gellan gum has frequently been associated with synthetic polymers such as polyacrylic acid-based compounds [49–52], and amino-methacrylate derivatives [53], with eventually the incorporation of inorganic loads (montmorillonite, for example, [49]) for manufacturing composite hydrogels. The membrane is produced by casting. Its flexibility is enhanced by the incorporation of glycerol in the mixture (prior to casting).

First, the casted composite membrane (i.e., GG-BC) is characterized by scanning electron microscopy (SEM), transmission electron microscopy (TEM), N_2 adsorption/desorption (textural properties), thermogravimetric analysis (TGA, and differential scanning calorimetry, DSC), Fourier-transform infrared spectroscopy (FTIR) and zetametry.

The sorption properties are investigated for the recovery of methylene blue (MB, $C_{16}H_{18}ClN_3S$), thiazine dye, Scheme S1, see Supplementary Information) using grinded composite membranes. The influence of environmental parameters is investigated (i.e., pH, sorbent dose, agitation speed, and addition of NaCl). Uptake kinetics and sorption isotherms are also collected and modeled using conventional equations. Special attention is paid to the effect of sonication (which is compared with mechanical agitation at different speeds) on the uptake kinetics. Dye desorption and sorbent recycling are carried out before investigating the recovery of the dye from a complex medium (i.e., spiked seawater samples).

2. Materials and methods

2.1. Materials

Gellan gum (GG, low acyl) with a molecular weight of 2.4×10^5 Da) determined by gel permeation chromatography(was purchased from Alfa Aesar. Polyvinyl alcohol (PVA) (Mwt ~ 17–18 kDa) was purchased from the International Company for Chemicals (ICC, Al Khobar, Saudi Arabia). Glycerol (purity 99.5 %, Mwt ~ 92) was obtained from Al-Gomhoria Company (Zietoun, Cairo, Egypt), *Orobanche aegyptiaca* biomass was obtained locally from Damhoug, Quisna, Menofiya (Egypt).

2.2. Biochar production

Section A in Supplementary Information reports in detail the methods used for the production (and optimization) of the biochar through microwave-assisted extraction (MAE) and the proper carbonization procedure. Briefly, MAE process (Lab-made microwave equipment, with generator: 900 W and 2450 MHz frequency) was used for the extraction of polyphenolic compounds from *O. aegyptica* biomass (in aqueous suspension). Both the extraction time (0–20 min) and the power of the microwave oven (90, 180, and 270 W) were investigated and optimized [104]. The residue was recovered by filtration and used for preparing the biochar. Pyrolysis was carried out under N₂ atmosphere at 350 °C, for 3 h.

2.3. Sorbent synthesis

Gellan hydrogel membranes were prepared by the solution casting method [54]. Gellan (1 %, w/w) was dissolved in demineralized water at 80 °C under gentle stirring for 2 h (for avoiding the formation of air

bubbles). Polyvinyl alcohol (PVA, 4 %, *w*/w) was dissolved in bi-distilled water at 70 °C under stirring for 2 h. Then PVA solution was added to the gellan solution and mixed with continuous stirring until complete miscibility. The ratio of gellan and PVA in the mixture was 80/20 (%, *w*/w).

The biochar (pyrolyzed O. *aegyptica* biochar) was ground, washed with 0.1 M HCl for 5 h, washed with demineralized water to remove any impurities, and then dried in an oven at 70 °C for 24 h. Pre-treated biochar (BC, 1 %, *w*/w) was added into the Gellan/PVA mixture and sonicated for 20 min to form a homogeneous dispersed solution. Glycerol (5 %, w/w, based on the final dry weight of polymer) was added under agitation. Finally, 20 mL of the mixture was poured into a glass petri dish (15 cm) and left to dry overnight at 60 °C in a vacuum oven. The films were peeled off from the glass disc, and stored, until use, in polyethylene bags to avoid contamination. Scheme 1 reports the different steps in the synthesis of the sorbent.

2.4. Sorbent characterization

Waters Mass Spectrometers 515/2410 (Waters, USA) was used for measuring of molecular weight of the native GG using the gel permeation chromatography technique. The morphology and microstructure of the sorbent membrane (small pieces of the membrane with particle size in the range of 20–30 mm), were observed using a high-resolution transmission electron microscope (TEM, JEOL JEM 2100F, Tokyo, Japan). The images were taken at an accelerating voltage of 200kV. To prepare the TEM samples (cut and grinded), a particle-ethanol colloidal mixture was ultrasonicated for 20 min, and a drop of the suspension was placed on a grid and dried then the sample was examined. Surface morphology images have been obtained with a MEB-FEG scanning electron microscope Zeiss Sigma 300 VP – GEMINI, (Carl Zeiss Microscopy, Iena, Germany), equipped with a dispersive X-ray spectrometer (EDS). The thickness of membranes was measured with a 0–25 mm electronic outside micrometer gauge meter 0.001 mm digital tool.

The sizes of the particles (biochar and composite sorbents) were analyzed by differential light scattering using Malvern Zetasizer Ver. 6.32 (Malvern Instruments Ltd., Malvern, UK). The thermal stability properties were characterized using a Q600 SDT thermal analyzer (Waters, TA instruments, New Castle, DE, USA), under N2 atmosphere, with a heating rate of 10 $^{\circ}$ C min⁻¹ (from room temperature up to 1000 °C). The hydrophobic characteristics of membranes were determined by measurement of the contact angle (drop angle) using a T200 THETA optical tensiometer (Biolin Scientific AB, Västra Frölunda, Sweden). The X-ray diffraction analysis of the membrane was carried acquired using an X'Pert PRO X-ray diffractometer (Malvern Panalytical Ltd., Malvern, UK). The patterns were conducted using CuK_{α} radiation ($\lambda = 0.15406$ nm). The 2 θ angle varied between 4° and 70°. A Nano Zeta Sizer (Nano-ZS, Malvern Instruments, Malvern, UK) was used for the determination of the zeta potential of the composite sorbent and the native biochar and GG (sorbent dose, SD: 1 g L^{-1} ; 0.01 M KCl solution, pH₀: 1–11; contact time before analysis: 4 h). FT-IR spectra were acquired using a Nicolet iS10 FTIR spectrometer (Thermo Scientific Instruments, Waltham, MA, USA).

2.5. Sorption tests

The experiments were performed in batch systems (and duplicated). A fixed volume of solution (V, L), at C₀ concentration (mmol MB L⁻¹), was mixed, at a standard speed of 200 rpm, (unless specified) with a given amount of sorbent (m, g) during fixed contact time. The standard temperature was 25 ± 1 °C (unless specified). The sorbent was separated from the solution by filtration (1.2-µm pore size membrane) and the residual concentration (C_{eq}, mmol MB L⁻¹) was analyzed by UV–Vis spectroscopy (Palintest 7100 spectrophotometer, Palintest Ltd., Gasteshead, UK) at λ : 664 nm wavelength (after adding a micro-drop of NaOH to shift the pH close to 10 for all samples calibration points and samples).



Scheme 1. Synthesis of precursors and sorbent.

Biochar/gellan gum hydrogel composite

The mass balance equation was used for determining the sorption capacity at equilibrium ($q_{eq} = (C_0 - C_{eq}) \times V/m$, mmol MB g⁻¹). The solutions were not buffered, and the pH was not controlled during dye sorption; however, the final pH was systematically monitored using an Aqualytic AL15 pH meter (Aqualytic GmbH & Co, Dortmund, Germany).

Uptake kinetics were also obtained from batch tests; a four mL-volume sample was collected at fixed times for measuring actual dye concentration. The correction of volume was applied to deduce the sorption capacity (according to Eq. (1)).

$$q(t) = \sum_{i=1}^{n} \left(\frac{\left(C(t)_{(i-1)} - C(t)_{(i)} \right) \times V(t)_{(i-1)}}{m} \right)$$
(1)

For uptake kinetics performed under ultrasonic treatment, the sorption proceeded into a thermostatic box using an ultrasonic bath sonicator (Elmasonic P300H, Elma Schmidbauer Gmbh, Singen, Germany). The power was 380 W while two frequencies were selected at 37 and 80 kHz.

For sorption isotherms, the initial concentration was varied from 0.0313 to 3.13 mmol MB L⁻¹, and the temperature was set at 25, 35, 45, and 55 °C. A shaking incubator was used for maintaining the appropriate temperature (LSI-3016R, LabTech S.r.l., Sorisole, BG, Italy). For studying the effect of NaCl concentration, the addition of the salt led to concentrations ranging between 0.086 and 0.77 mol NaCl L⁻¹. The impact of agitation speed on the sorption capacity at equilibrium was compared for velocities in the range of 50–300 rpm. The sorbent dose, SD, was varied between 0.5 and 5 g L⁻¹. The experimental conditions are systematically reported in the caption of the figures (see below).

For dye desorption (in batch), an acidic solution (0.7 M HCl) was used. Sorption recycling was tested for six successive cycles (the waterrinsing step was systematically processed between each sorption and desorption operation).

2.6. Modeling of sorption kinetics and isotherms

Table S2a summarizes the equations used for modeling uptake kinetics: pseudo-first (PFORE), pseudo-second order rate (PSORE) equations [55], the Elovich equation [56], and the resistance to intraparticle diffusion (RIDE, the so-called Crank equation [57]). Resistance to intraparticle diffusion (RIDE) can be used for approaching the resistance to intraparticle diffusion. This equation was used for modeling the kinetic profile and approaching the determination of the effective intraparticle diffusion coefficient (D_e , $m^2 min^{-1}$). Table S2b reports the equations used for fitting sorption isotherms: Langmuir, Freundlich, and Sips [58], Temkin [59], and Dubinin-Radushkevich (D-R) [60]. The quality of fitting was measured by the determination coefficient (i.e., R^2) and the Akaike Information Criterion (AIC, see Table S2b [61]).

For the determination of thermodynamic parameters, the van't Hoff equation was applied using the methodology reported by Tran et al. [62]. They documented the importance of the adjustment of parameter units for the appropriate determination of the equilibrium constant (K°_{eq}) .

$$\Delta G^{\circ} = -RT ln K_{ea}^0 \tag{2}$$

$$\Delta G^{\circ} = \Delta H^{\circ} - T \Delta S^{\circ} \tag{3}$$

 ΔG° (kJ mol⁻¹), R is the universal gas constant (8.31 J mol⁻¹ K⁻¹), ΔH° (kJ mol⁻¹), and ΔS° (J mol⁻¹ K⁻¹) are the changes in Gibbs energy, enthalpy, and entropy, respectively; R is the universal gas constant.

The equilibrium constant must be dimensionless for being consistent with thermodynamic criteria. Tran et al. [62] convert b_L affinity coefficient (in Langmuir equation) to equilibrium constant using the following equation:

$$K_{eq}^{0} = b_{L} \times \frac{C_{Adsorbate}^{0}}{\gamma_{Adsorbate}}$$
(4)

where $C^0_{Adsorbate}$ is the unitary standard concentration of the adsorbate (~1 mol L⁻¹) and $\gamma_{Adsorbate}$ is the activity coefficient of the adsorbate (dimensionless); meaning that b_L should be converted to L mol⁻¹ unit.

3. Results and discussion

3.1. Characterization of sorbent

3.1.1. Physical characterization of sorbent particles

The thickness (τ , mm) of the composite membranes was measured using a digital micrometer gauge meter on several samples: $\tau = 0.433 \pm 0.046$ mm, the membrane size fraction collected for sorption tests was 20–30 mm. all the adsorption studies were done using these small pieces of the membrane.

Fig. 1 shows the SEM micrographs of the sorbent (before and after MB sorption) at different magnitudes. The surface of the materials is irregular with aggregates and lamellar structures. The pristine material is characterized by opened channels (representative of the structure of raw material, *O. aegyptica* [63]) that disappear (being blocked) after dye binding.

Transmission electron microscopy (Fig. 2) shows relatively homogeneous structures with a light background associated with the GG matrix. The incorporation of biochar nano-objects is highlighted by little more obscure zones. The two components of the composite are constituted by light elements, which do not offer the possibility to isolate contrasting objects (as may appear when metal nanoparticles are embedded in composites). The sorption of MB hardly changes the TEM images of the sorbent. However, in Fig. 2, the left panel for MB-loaded sorbent shows more structured objects.

Differential light scattering (DLS) may be used for determining the size of biochar and sorbent particles (Fig. S1). The size of about 77.3 % of biochar nanoparticles are close to 196 nm (average value), however, a significant part (about 22.7%) has an average value close to 0.962 μ m. This is consistent with the order of magnitude of the largest objects observed in the composites (TEM image). The average size of the composite is found close to 1 mm.

3.1.2. Thermal degradation of sorbent

Fig. S2 shows the thermal degradation of the composite sorbent (with TGA curve (a) and DTA/DSC profiles (b)). The degradation occurs through three steps:

(a) below 106 °C, free or absorbed water is released (representing a weight loss, WL \approx 6.3 %), (b) in the range 106–225 °C, most of the weight loss is registered (WL \approx 52.3 %); this phase corresponds to the release of –OH groups and the depolymerization of the GG network, and (c) above 225 °C, the formation of a char, followed by its thermal degradation WL \approx 29.2 %).

These transitions are shifted to lower temperatures compared with the thermal degradation of free GG [64] and GG/PVA blends [65]. The total degradation reaches 72 %; this is consistent with the value reported for free GG by Tyagi et al. [66].

The DTA profile shows two local minima at T: 70.0 °C and 206.1 °C, corresponding to the largest weight variations reported in TGA profiles. The DSC curve shows a weak relative minimum at 208.9 °C; this value is close to the temperature reported for minimum in DSC curve for GG-polyacrylamide composite (i.e., 237 °C, which is higher than the peak reported for GG alone) [64]. The PVA/glycerol interaction affects the thermal degradation characteristics of the composite. In the case of GG-PVA, it is noteworthy that the biochar that was pyrolyzed during the conversion of *O. aegyptica* may marginally contribute to the weight loss of the composite (consistent with conclusions raised on another type of biochar by Subratti et al. [67]).



Fig. 1. SEM images of the sorbent before (A) and after (B) MB sorption (different growing magnitudes).



Fig. 2. TEM images of the sorbent before and after MB sorption (bar: left panels, 100 nm; right panels, 200 nm).

3.1.3. FTIR spectrometry

Fig. 3 compares the FTIR spectra of the composite sorbent as produced, after conditioning the sorbent at the pH used for MB sorption, and after dye binding. Table 1 summarizes the assignments of the main bands. The comparison of these spectra allows to separate the potential contributions of pH change (conditioned sorbent at fixed pH) from the interactions with the dye. The pH control (to 10.5) hardly changes the profile of the sorbent. Two main differences are observed at 1714 cm⁻¹ and 1241 cm⁻¹ (where the weak bands disappear); these bands are assigned to ν_s (C=O) in ester carbonyl groups and ν_s (C=O) [47] (or ν (C—N) in secondary aromatic amine, [47]).

The interaction of MB induces specific variations in the FTIR spectrum of the sorbent. The shift of the broadband at 3310 cm⁻¹ to 3391 cm⁻¹ may be caused by the sorption of MB that bears numerous N-based functional groups; the band is assigned to ν_{OHbond} (N···H—O) and ν_{OHbond} (N⁺•••H-O) [68]. The band associated with the carboxyl group ($\nu(COO^-)$ is substantially changed: the intensity is decreased and superposed to the contribution of $\nu_{het}(C=N)$ and $\nu_{het}(C=C)$ bands (directly assigned to MB, [68]). In the range of 1500–1100 cm⁻¹, a typical band of MB is appearing (at 1341 cm⁻¹) while the contribution of $\nu(C-N)$ (at 1241 cm⁻¹) is reduced. The bands at 923 cm⁻¹ (ν C-O) and 850 cm⁻¹ (PVA fingerprint) are strongly reduced and replaced with two bands at 885 cm⁻¹ and 812 cm⁻¹ (associated with MB reactive groups), respectively. The weak band at 671 cm⁻¹ is another marker of the presence of MB (see Table 1).

3.1.4. Zetametry

Fig. S3 allows determining the pH_{PZC} of the composite sorbent and its components (i.e., biochar and GG). The pH_{PZC} of the sorbent is close to 6.43, while the pH_{PZC} of the native biochar and GG are close to 6.8 and 2.3, respectively. This value is consistent with the value reported for other types of biochar (i.e., 6.15 ± 0.15 [69]). Kaur and Kaur [70] investigated the association of GG with guar gum (cationic polyelectrolyte): GG remains negatively charged on a wide pH range, and the pK_a value of the carboxylic group is close to 3.72.

3.1.5. Contact angle

The contact angle was measured by the water drop procedure on the sorbent before MB sorption (Fig. S4). The value is close to $88.6 \pm 0.7^{\circ}$. This value is much higher than the data reported by Vashisth and Pruthi [71] for raw GG/PVA nanofibers. Aadil et al. [72] also reported a much lower contact angle (meaning higher hydrophilicity) for GG/PVA nanofibers (i.e., close to 35°). However, Vashisth and Pruthi [71] also observed that by playing with the crosslinking post-treatment, the

Table 1

FTIR assignments for main bands on sorbent (raw), conditioned sorbent (after pH control), and after MB sorption.

$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Assignment	Sorbent	Sorbent (pH)	Sorbent + MB	Reference
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	ν (OH), ν _{OHbond} (N···H—O), ν _{OHbond} (N ⁺ •••H-O)	3310	3310	3391	[68,72]
$\begin{array}{ccccc} \nu_{s}(CH_{3}) & 2881 & 2881 & [68] \\ \nu_{s}(C=0) \ (ester carbonyl & 1714 & [47] \\ group) & & & & & & & & \\ \nu_{as}(COO^{-}) & 1653 & 1653 & [72] \\ \nu_{het}(C=N), \nu_{het}(C=C) & 1604 & [68] \\ \nu_{het}(C=S^{+}) & 1491 & [68] \\ \nu(-CH_{2}) & 1419 & 1419 & [72,102] \\ \delta(CH_{3}) & & 1398 & [68] \\ \nu_{het}(C=S+) \ and/or \ \nu(CN) \ (in & 1341 & [68] \\ NCH_{3}) & & & & \\ \nu(C-N) & 1241 & 1213 & [47] \\ \nu(C-O), \ \nu(C-O-C) & 1107 & 1107 & 1103 & [47] \\ \nu(C-O), \ \nu(het,C=H) & 1034 & 1034 & 1042 & [68,72,102] \\ Carbohydrate \ fingerprint & 994 & 994 & [103] \\ \nu(-C-O) \ (alkyl ether) & 923 & 923 & [47] \\ \gamma_{het}(C=H) & & 885 & [68] \\ PVA \ fingerprint & 850 & 850 & [102] \\ \delta_{het}(C-C) & & 812 & [68] \\ \nu_{s}(C-S-C) + \nu(C=N) + \delta(C=H) & 671 & [68] \\ in-plane, \ \delta(C=H) \ bending \\ \dots \end{array}$	ν (-CH ₂)	2937	2937	2938	[68]
$\begin{array}{c c} \nu_{s}(C=0) \ (ester \ carbonyl \ & 1714 \ & [47] \\ group) \\ \nu_{as}(COO^{-}) & 1653 \ 1653 \ & [72] \\ \nu_{het}(C=N), \nu_{het}(C=C) & 1604 \ & [68] \\ \nu_{het}(C=S^{+}) & 1491 \ & [68] \\ \nu(-CH_2) & 1419 \ & 1419 \ & [72,102] \\ \delta(CH_3) & 1398 \ & [68] \\ \nu_{het}(C=S+) \ and/or \ \nu(CN) \ (in \ & 1341 \ & [68] \\ NCH_3) \\ \nu(C-N) & 1241 \ & 1213 \ & [47] \\ \nu(C-O), \ \nu(C-O-C) \ & 1107 \ & 1107 \ & 1103 \ & [47] \\ \nu(C-O), \ \nu_{het}(C-H) \ & 1034 \ & 1034 \ & 1042 \ & [68,72,102] \\ Carbohydrate \ fingerprint \ & 994 \ & 994 \ & [103] \\ \nu(-C-O) \ (alkyl \ ether) \ & 923 \ & 923 \ & [47] \\ \gamma_{het}(C-H) \ & 885 \ & [68] \\ PVA \ fingerprint \ & 850 \ & 850 \ & [102] \\ \delta_{het}(C-C) \ & 812 \ & [68] \\ \nu_{s}(C-S-C) + \nu(C-N) + \delta(C-H) \ & 671 \ & [68] \\ in-plane, \ \delta(C-H) \ bending \ & end \ of for \ c \ end \ & end \ end \ & end \ &$	$\nu_{\rm s}({\rm CH}_3)$	2881	2881		[68]
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	$\nu_{s}(C=O)$ (ester carbonyl		1714		[47]
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	group)				
$\begin{array}{cccc} \nu_{het}(C=N), \nu_{het}(C=C) & 1604 & [68] \\ \nu_{het}(C=S^+) & 1491 & [68] \\ \nu(CH_2) & 1419 & 1419 & [72,102] \\ \delta(CH_3) & 1398 & [68] \\ \nu_{het}(C=S+) and/or \nu(CN) (in & 1341 & [68] \\ NCH_3) & & \\ \nu(C-N) & 1241 & 1213 & [47] \\ \nu(C-O), \nu(C-O-C) & 1107 & 1107 & 1103 & [47] \\ \nu(C-O), \nu(C-O-C) & 1107 & 1107 & 1103 & [47] \\ \nu(C-O), \gamma_{het.}(C-H) & 1034 & 1034 & 1042 & [68,72,102] \\ Carbohydrate fingerprint & 994 & 994 & [103] \\ \nu(-C-O) (alkyl ether) & 923 & 923 & [47] \\ \gamma_{het.}(C-H) & 885 & [68] \\ PVA fingerprint & 850 & 850 & [102] \\ \delta_{het}(C-C) & 812 & [68] \\ \nu_s(C-S-C) + \nu(C-N) + \delta(C-H) & 671 & [68] \\ in-plane, \delta(C-H) bending \\ \nu_{st} = \delta_{10} + 0.5 \\ \mu_{st} = \delta_{$	$\nu_{as}(COO^{-})$	1653	1653		[72]
$\begin{array}{cccc} \nu_{het}(C=S^+) & 1491 & [68] \\ \nu(-CH_2) & 1419 & 1419 & [72,102] \\ \delta(CH_3) & 1398 & [68] \\ \nu_{het}(C=S+) & and/or \nu(CN) (in & 1341 & [68] \\ NCH_3) & \\ \nu(C-N) & 1241 & 1213 & [47] \\ \nu(C-O), \nu(C-O-C) & 1107 & 1107 & 1103 & [47] \\ \nu(C-O), \nu(C-O-C) & 1107 & 1107 & 1103 & [47] \\ \nu(C-O), \nu(C-O+I) & 1034 & 1034 & 1042 & [68,72,102] \\ Carbohydrate fingerprint & 994 & 994 & [103] \\ \nu(-C-O) (alkyl ether) & 923 & 923 & [47] \\ \nu_{het}(C-H) & 885 & [68] \\ PVA fingerprint & 850 & 850 & [102] \\ \delta_{het}(C-C) & 812 & [68] \\ \nu_s(C-S-C) + \nu(C-N) + \delta(C-H) & 671 & [68] \\ in-plane, \delta(C-H) bending \\ \dots & \dots$	$\nu_{het}(C=N), \nu_{het}(C=C)$			1604	[68]
$\begin{array}{cccc} \nu(\text{-CH}_2) & 1419 & 1419 & [72,102] \\ \delta(\text{CH}_3) & 1398 & [68] \\ \nu_{\text{het}}(\text{C} = \text{S}+) \text{ and/or } \nu(\text{CN}) (\text{in} & 1341 & [68] \\ \text{NCH}_3) \\ \nu(\text{C}-\text{N}) & 1241 & 1213 & [47] \\ \nu(\text{C}-\text{O}), \nu(\text{C}-\text{O}\text{C}) & 1107 & 1107 & 1103 & [47] \\ \nu(\text{C}-\text{O}), \nu(\text{C}-\text{C}\text{C}) & 1107 & 1107 & 1103 & [47] \\ \nu(\text{C}-\text{O}), \gamma_{\text{het}}(\text{C}-\text{H}) & 1034 & 1034 & 1042 & [68,72,102] \\ \text{Carbohydrate fingerprint} & 994 & 994 & [103] \\ \nu(\text{-C}-\text{O}) (alkyl ether) & 923 & 923 & [47] \\ \gamma_{\text{het}}(\text{C}-\text{H}) & \text{885} & [68] \\ \text{PVA fingerprint} & 850 & 850 & [102] \\ \delta_{\text{het}}(\text{C}-\text{C}) & 812 & [68] \\ \nu_{\text{s}}(\text{C}\text{-S}\text{C}) + \nu(\text{C}-\text{N}) + \delta(\text{C}-\text{H}) & 671 & [68] \\ \text{in-plane, } \delta(\text{C}-\text{H}) bending \\ \nu_{\text{s}} \text{ch} \text{ of } \rho_{\text{s}} \text{ of } \rho_{\text{s}} \end{array}$	$\nu_{\rm het}({ m C}={ m S}^+)$			1491	[68]
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	ν(-CH ₂)	1419	1419		[72,102]
$\begin{array}{c c} \nu_{het}(C=S+) \text{ and/or } \nu(CN) (in & 1341 & [68] \\ NCH_3) \\ \nu(C-N) & 1241 & 1213 & [47] \\ \nu(C-O), \nu(C-O-C) & 1107 & 1107 & 1103 & [47] \\ \nu(C-O), \nu(C-H) & 1034 & 1034 & 1042 & [68,72,102] \\ Carbohydrate fingerprint & 994 & 994 & [103] \\ \nu(-C-O) (alkyl ether) & 923 & 923 & [47] \\ \nu(-C-H) & 885 & [68] \\ PVA fingerprint & 850 & 850 & [102] \\ \delta_{het}(C-C) & 812 & [68] \\ \nu_s(C-S-C) + \nu(C-N) + \delta(C-H) & 671 & [68] \\ in-plane, \delta(C-H) bending \\ ret of charge S(C + C) \\ \end{array}$	δ(CH ₃)			1398	[68]
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	$\nu_{het}(C = S+)$ and/or $\nu(CN)$ (in			1341	[68]
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	NCH ₃)				
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	ν(C—N)	1241	1213		[47]
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	ν(C—O), ν(C-O-C)	1107	1107	1103	[47]
$\begin{array}{c c} Carbohydrate fingerprint & 994 & 994 & [103] \\ \nu(\text{-C-O) (alkyl ether)} & 923 & 923 & [47] \\ \gamma_{het.}(C-H) & 885 & [68] \\ PVA fingerprint & 850 & 850 & [102] \\ \delta_{hef}(C-C) & 812 & [68] \\ \nu_s(\text{C-S-C}) + \nu(\text{C-H}) + \delta(\text{C-H}) & 671 & [68] \\ \text{in-plane, } \delta(\text{C-H}) \text{ beding} \\ \text{orth of large } S(C \in C) \end{array}$	ν(C—O), γ _{het.} (C—H)	1034	1034	1042	[68,72,102]
$\begin{array}{c c} \nu(\text{-C-O) (alkyl ether)} & 923 & 923 & [47] \\ \gamma_{het.}(\text{C}-\text{H}) & 885 & [68] \\ \text{PVA fingerprint} & 850 & 850 & [102] \\ \delta_{het}(\text{C}-\text{C}) & 812 & [68] \\ \nu_s(\text{C-S-C}) + \nu(\text{C}-\text{H}) + \delta(\text{C}-\text{H}) & 671 & [68] \\ \text{in-plane, } \delta(\text{C}-\text{H}) \text{ bending} & \\ \text{sets of faces } \delta(\text{C} \in \text{C}) \end{array}$	Carbohydrate fingerprint	994	994		[103]
$\begin{array}{c c} \gamma_{het.}(C-H) & 885 & [68] \\ PVA \mbox{ fingerprint} & 850 & 850 & [102] \\ \delta_{het}(C-C) & 812 & [68] \\ \nu_s(C-S-C) + \nu(C-N) + \delta(C-H) & 671 & [68] \\ \mbox{ in-plane, } \delta(C-H) \mbox{ bedding} & \\ set \mbox{ of clarge, } S(C-C) \mbox{ constant} \end{array}$	ν(-C-O) (alkyl ether)	923	923		[47]
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	$\gamma_{het.}(C-H)$			885	[68]
$δ_{het}(C=C)$ 812 [68] $ν_s(C-S-C) + ν(C=N) + δ(C=H)$ 671 [68] in-plane, $δ(C=H)$ bending set of alore a S(C=G)	PVA fingerprint	850	850		[102]
ν_{s} (C-S-C) + ν (C-N) + δ (C-H) 671 [68] in-plane, δ (C-H) bending	$\delta_{het}(C-C)$			812	[68]
in-plane, $\delta(C-H)$ bending	$\nu_{s}(C-S-C) + \nu(C-N) + \delta(C-H)$			671	[68]
aut of plana S(C, C, C)	in-plane, δ(C—H) bending				
out-or-prane, o(C-S-C),	out-of-plane, δ(C-S-C),				
δ _{heterocycle} (C—C)	δ _{heterocycle} (C—C)				

contact angle may increase from 30° to 81°. Probably, this specific filmforming method (including glycerol and drying/casting steps) contributes to increased hydrophobicity.

After MB sorption, it was not possible to report accurate values of the contact angle: the water drop is immediately absorbed making difficult the measurement of the angle.

3.2. Sorption studies

3.2.1. Effect of pH on MB sorption

Fig. 4 shows the effect of equilibrium pH_{eq} on the sorption of MB using the composite sorbent. The sorption efficiency continuously increases with the pH. Below pH_{PZC} , the steepness of the curve is stronger than above pH 6.4. Methylene blue is a cationic dye; the equal distribution between MB⁺ and MB⁰ is reached at $pH = pK_a = 3.8$ [73]. The positive charge of the sorbent below pH 6.4 causes repulsion of MB molecules in acidic solutions; increasing the pH_0 from 1.6 to 10.5



Fig. 3. FTIR spectra of Sorbent: as-produced, after being in contact with water at controlled pH (corresponding to MB sorption), and after MB (sorption) (SD: 0.3 g L^{-1} ; C₀: 0.3 mmol MB L^{-1} ; pH₀: 10.5).



Fig. 4. Effect of pH on MB sorption (C₀: 0.3 mmol MB L^{-1} ; Sorbent dose, SD: 1.5 g L^{-1} ; time: 150 min; v: 200 rpm; T: 25 °C; duplicate series).

reduces the repulsion and the sorption capacity progressively increases from 0.155 to 0.202 mmol MB g^{-1} .

It is noteworthy that, under strongly acidic conditions (pH 1.6–1.8), there is an appreciable amount of dye that was sorbed into sorbent particles. Under these conditions, the dye molecules as well as the sorbent are protonated. This means that the binding of the dye also involves interactions different from electrostatic attraction; probably associated with electron donor-acceptor interactions; hence, Tran et al. [74] discussed the mechanisms involved in methylene green (another cationic dye) binding onto a series of hydrochars. They identified 5 different modes of interaction: (a) pore filling (negligible because of limited porous volume, contrary to the current composite), (b) electrostatic interaction (predominant), (c) n- π interaction, (d) π - π interaction, and (e) H-bonding interactions (including Yoshida H-bonding and dipoledipole H bonding). Hoslett et al. [75] also reported the contributions of electrostatic, π electron donor-acceptor, and π - π stacking interactions to the binding of MB onto biochar (produced by pyrolysis of mixed municipal discarded material). Ofomaja [76] reports the capacity of MB to form protonated monomeric and dimeric MB micelles that may bind to anionic and neutral palm kernel fibers, respectively. In the case of MB sorption using graphene oxide/hollow mesoporous silica composite, Yilmaz [77] identified the main contributions of electrostatic and π - π stacking interactions. Al-Ghouti and Al-Absi [78] assigned different mechanisms for MB sorption onto olive stones depending on the pH of the solution: (a) in acidic solutions MB is bound through hydrogen bonding with -OH groups of cellulosic fraction, (b) in neutral solutions, hydrophobic-hydrophobic interactions are involved, while (c) in basic media, electrostatic interaction may be involved.

Fig. 5a shows that the pH remains relatively constant during sorption: the pH varies by <0.5 pH unit. The log₁₀ plot of distribution coefficient D (defines as: $D = q_{eq}/C_{eq}$, L g⁻¹) vs. pH_{eq} (Fig. 5b) shows that D linearly increases with the pH. In ion-exchange mechanisms, the slope is associated with the stoichiometric ratio of proton exchange with solute ions. Herein, the slope is close to +0.14; this is not consistent with a possible ionic ratio between MB species and reactive groups on the sorbent; as another evidence that the electrostatic attraction is not the predominant or unique mechanism involved in MB binding.

3.2.2. Effect of agitation speed, sorbent dose, and NaCl concentration on equilibrium sorption

The agitation speed shows a limited effect on sorption capacity (from 0.195 to 0.204 mmol MB g⁻¹) and efficiency (from 94.4 to 97.4 %) at equilibrium (Fig. 6a). The agitation speed may influence the equilibrium time: lower agitation requires longer contact time. In these experiments, the contact time was systematically set to 150 min; this may be not enough in the case of the agitation regime; however, despite these



Fig. 5. Effect of pH on MB sorption: (a) pH variation, (b) log_{10} plot of D vs. pH_{eq} (C₀: 0.3 mmol $L^{-1};$ SD: 1.5 g $L^{-1};$ v: 200 rpm; time: 150 min).



Fig. 6. (a) Effect of agitation speed on MB sorption: Sorption capacity (q_{eq} , mmol MB g^{-1}) and sorption efficiency (%) (C₀: 0.3 mmol L⁻¹; pH₀: 10.5; SD: 1.5 g L⁻¹; v: 50–300 rpm; time: 150 min). (b) Effect of sorbent dose on MB sorption: Sorption capacity (q_{eq} , mmol MB g^{-1}) and sorption efficiency (%) (C₀: 0.3 mmol L⁻¹; pH₀: 10.5; SD: 0.5–5 g L⁻¹; v: 200 rpm; time: 150 min).

unfavorable conditions, the pseudo-equilibrium performances are hardly affected by this parameter. For further experiments, the agitation is fixed to 200 rpm, which corresponds to optimum conditions. The agitation speed is supposed to play a significant role in the control of uptake kinetics through the resistance to film diffusion.

As expected, increasing sorbent dose increases sorption efficiency (Fig. 6b). However, under selected experimental conditions, the sorption efficiency hardly varies (from 96 % to 98.5 %) when the sorbent dose increases 10 times (from 0.5 to 5 g L⁻¹). However, under these conditions, the sorption capacity is drastically reduced (from 0.596 to 0.062 mmol MB g⁻¹). For further experiments, the standard sorbent dose was set to 1.5 g L⁻¹, in order to maintain both high sorption efficiency and capacity.

The effluents from the dving industry frequently contain high concentrations of salt. This may be of critical importance because, in the case of ion-exchange mechanisms, high ionic strength may cause a dramatic decrease in sorption performance [79]. Fig. 7 shows that the addition of NaCl to the dye solution has a negligible impact on MB removal: the efficiency decreases by <8 % when NaCl concentration reaches 45 g L⁻¹. On the other side, the sorption capacity varies by <6 % (from 0.202 to 0.190 mmol MB g^{-1}). This weak effect of ionic strength confirms that the mechanism of electrostatic attraction is not playing the predominant role in MB sorption by the composite. In the case of MB sorption onto biochar, Ji et al. [80] observed a weak increase in sorption with increasing NaCl to 0.1 mM: they explain that NaCl enhances the protonation of the sorbent and the ionization of the dye; however, they did not investigate the impact of higher ionic strength. On the opposite hand, Alver et al. [81] recently reported a drastic decrease in MB sorption capacity onto magnetic alginate/rice husk composite when the concentration of NaCl increases (from 83 to 8 mg MB g^{-1} , when reaching 1 % NaCl ionic strength). They assigned this strong decrease to the competitor effect of Na⁺ ions that shield the negative charge of alginatebased material, preventing the binding of cationic MB molecules.

3.2.3. Uptake kinetics

The sorption kinetics may be controlled by several mechanisms associated with diffusion regimes (resistance to bulk, film, and/or intraparticle diffusion) and to the proper reaction rates. The contribution of these phenomena may be affected by parameters such as sorbate concentration, pH, temperature, agitation speed, porosity of the sorbent (especially in relation with the size of the sorbate), and so on [58].

3.2.3.1. Effect of agitation speed (mechanical agitation). Fig. 8 shows the kinetic profile for MB sorption using the composite sorbent under selected experimental conditions: under mechanical agitation, the speed



Fig. 7. Effect of NaCl concentration on MB sorption: sorption capacity (q_{eq} , mmol MB g⁻¹) and sorption efficiency (%) (C₀: 0.3 mmol L⁻¹; pH₀: 10.5; SD: 1.5 g L⁻¹; v: 200 rpm; time: 150 min).



Fig. 8. Effect of agitation speed on MB uptake kinetics using composite sorbent – Modeling with (a) PFORE and (b) PSORE and (c) RIDE (C_0 : 0.31 mmol MB L^{-1} ; pH₀: 10.5, SD: 1.5 g L^{-1} ; v: 50–250 rpm; T: 25 °C).

was varied between 50 and 250 rpm. With an agitation speed set at 200 rpm, the contribution of resistance to bulk and film diffusion is minimized. The $t_{50\%}$ (time required to reach half of the total sorption) varies between 30 min (at v: 50 rpm) and 18–22 min (at 200–250 rpm); the $t_{90\%}$ decreases from 94 min to 62–68 min with increasing the agitation speed. The agitation speed is usually affecting the resistance to external diffusion and bulk diffusion playing with both the thickness of the film surrounding the particles and the homogeneity in the distribution of the solute and sorbent particles in the reactor, respectively. Above 200 rpm, the variations are not very marked, and this agitation speed sounds a good compromise. This is consistent with the preliminary tests collected

at equilibrium in Fig. 6a. The agitation speed hardly influences the sorption capacity at equilibrium: $q_{eq,exp.}$ is close to 0.165 \pm 0.006 mmol MB g⁻¹.

The required contact time for reaching equilibrium is close to 180 min. This probably means that the resistance to intraparticle diffusion controls sorption kinetics, at least partially. The sorption operates through rapid sorption at the surface of the sorbent followed by slow sorption inside sorbent particles (controlled by the resistance to intraparticle diffusion). Jia et al. [82] reported the length and width of MB molecule: ≈ 14 , and 9.5, respectively; which, is larger than the values reported for pore size (as determined by N2 adsorption/desorption isotherms on dry sorbent particles). The soaking of the sorbent (when dropped into the dve solution) may expand the porous network to make internal reactive groups more accessible and available to dve molecules. However, despite this possible expansion, the respective sizes of pores and dye molecules cause kinetic restrictions to dye transfer. Experimental points are superposed with the fitted curves for PFORE (Fig. 8a) and PSORE (Fig. 8b) while Fig. 8c shows RIDE fitting (model's equations are summarized in Table S1). Table 2 reports the parameters of the models (together with statistical criteria: determination coefficient, R^2 , and Akaike Information Criterion, AIC). The quality of the fits for the different models are ranked according:

PFORE > PSORE≫RIDE

Due to the poor fit of experimental profiles by the Elovich equation, the simulated profiles are not presented. It is noteworthy that the comparison of calculated $q_{eq}\xspace$ values with experimental values shows that the PFORE values are very close to experimental values (i.e., q_{eq,exp}, the variation being <2 %) while the PSORE overestimates the reference value by 10-25 %. The PFORE offers a global better simulation of the kinetic profile. Dragan and Apopei Loghin [83] also report close fits of MB uptake kinetics by PFORE and PSORE when using modified starch/ polyacrylamide composites. Elwakeel et al. [84] obtained k1 values close to 4.1 \times 10⁻² min⁻¹, for MB binding onto 2-mercaptobenzimidazole derivative of chitosan. Herein, the apparent rate coefficient is in the range $2-5 \times 10^{-2}$ min⁻¹, which is of the same order of magnitude as the values determined by Dragan and Apopei Loghin [83]. The apparent rate coefficient increases with agitation speed above 150 rpm: from 2.11 to 2.26×10^{-2} min⁻¹ at 50–150 rpm to 3.98– 4.96×10^{-2} min⁻ Recently, Chen et al. [85] reported apparent rate coefficients in the range 0.88–1.01 \times 10⁻² min⁻¹ for MB sorption onto starch-based hydrogel. Hoslett et al. [75] reported preferential kinetic modeling by the PSORE for MB binding onto char (produced from municipal wastes); the apparent rate coefficient k_2 was 3.11 g $\text{mmol}^{-1}\ \text{min}^{-1}$ (about 10 times the values for the current sorbent; i.e., 0.12-0.42 g mmol⁻¹ min^{-1}).

The resistance to intraparticle diffusion was evaluated using the

 Table 2

 Modeling of MB uptake kinetics – Effect of agitation speed (mechanical agitation).

Model	Parameter	Agitation speed (rpm)			
		50	150	200	250
Experimental	q _{eq,exp}	0.159	0.165	0.173	0.172
PFORE	q _{eq,1}	0.162	0.167	0.172	0.168
	$k_1 \times 10^2$	2.14	2.34	3.92	4.96
	R ²	0.998	0.994	0.993	0.985
	AIC	-318	-297	-283	-260
PSORE	q _{eq,2}	0.198	0.200	0.192	0.184
	$k_2 \times 10^2$	11.6	13.3	28.7	41.7
	R ²	0.991	0.988	0.992	0.989
	AIC	-280	-277	-286	-284
RIDE	$D_e \times 10^{13}$	1.09	0.716	1.18	2.09
	R ²	0.981	0.975	0.991	0.989
	AIC	-236	-222	-219	-215

Units: q_{eq} , mmol g^{-1} ; k_1 , min⁻¹; k_2 , g mmol⁻¹ min⁻¹; D_e , m² min⁻¹.

Crank equation [86], under simplifying hypotheses such as (a) separating the contribution of this diffusion mode to the resistance to film diffusion, and (b) not taking into account the equilibrium conditions (i. e., the Langmuir or alternative equation). The application of the Crank equation supposed the sorbent to be initially free of the solute (meaning not infinite and that the concentration of the solute varies). In his reference book (The Mathematics of Diffusion), Crank defined different equations for simulating the transfer of solute within solid materials under different conditions such as finite/infinite volume, and shape of the material. Herein, the equation used for modeling kinetic profiles is based on finite volume conditions, and sorbent particles assimilated to spherical particles. This is a very simplified approach but closer to the reality of equations such as the multi-stage Weber and Morris equation [87]. This is mainly used to (a) evaluate the order of magnitude of the effective diffusion coefficient (D_e , $m^2 min^{-1}$), (b) compare with the free diffusivity of the solute in the solvent, and (c) compare the diffusion characteristics of different materials or different solutes (under similar conditions). The Crank equation globally fits the experimental profile (Fig. 8c), though less accurately in the most curved section of the kinetic profile, and less precisely than PFORE and PSORE (as confirmed by statistical criteria in Table 2). However, this model can be used for approaching the effective diffusivity coefficient D_e (defined in Table S1): varying in the range $0.7-2.1 \times 10^{-13} \text{ m}^2 \text{ min}^{-1}$. Selifonov et al. [88] reported the diffusivity coefficient of MB in water, which is close to 4.04 $\times 10^{-8} \text{ m}^2 \text{ min}^{-1}$. They observed a little decrease in D_e value considering the diffusion of dye micelle (at $1.16 \times 10^{-8} \text{ m}^2 \text{ min}^{-1}$). Leaist [89] provided slightly higher values for the diffusivity coefficient of MB in water (i.e., $5.0 \times 10^{-8} \text{ m}^2 \text{ min}^{-1}$); he also reported the decrease in the diffusivity properties with dye aggregation (i.e., down to $3.8 \times 10^{-8} \text{ m}^2$ min⁻¹) and in the presence of counter ions (which can be bound; this is the case with NaCl). The effective diffusivity coefficient calculated from Fig. 8c (in Table 2) is thus several orders of magnitude lower than the free diffusivity of MB in water; meaning that the resistance to intraparticle diffusion takes part in the overall control of dye transfer.

3.2.3.2. Effect of frequency (ultrasonic treatment). Several studies have previously illustrated the benefit of using the ultrasonic treatment for improving the sorption performances of solutes [90], including dyes MB [91]. In the case of the sorption of titan yellow and rose bengal onto a magnetic metal-oxide organic framework, the application of ultrasonic treatment allowed reducing ten times the equilibrium time [92]. Herein, the t_{50%} was reduced to 4 and 3 min (t_{90%} to 30 and 20 min) with ultrasonic treatment; increasing the frequency (F, kHz) significantly enhances the uptake kinetics. The equilibrium sorption capacity is weakly affected by the frequency (Table 3): q_{eq.exp.} close to 0.192–0.195 mmol MB g⁻¹ under selected experimental conditions; this is consistent with the conclusions raised by Rajumon et al. [91] in the case of MB sorption using reduced graphene oxide. The ultrasonic treatment increases the

Table	3	

Modeling of M	B uptake kinetics	 Effect of frequency 	(ultrasonic agitation)	•
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Model	Parameter	Ultrasonic frequency (kHz)	
		37	80
Experimental	q _{eq,exp}	0.192	0.195
PFORE	q _{eq,1}	0.185	0.189
	$k_1 imes 10^2$	13.5	19.3
	R^2	0.984	0.990
	AIC	-119	-126
PSORE	Qeq.2	0.205	0.205
	$k_2 imes 10^2$	85.8	130.8
	R ²	0.998	0.999
	AIC	-150	-158
RIDE	$\mathrm{D_e} imes 10^{13}$	6.57	6.66
	R ²	0.949	0.892
	AIC	-85	-81

Units: q_{eq} , mmol g^{-1} ; k_1 , min⁻¹; k_2 , g mmol⁻¹ min⁻¹; D_e , m² min⁻¹.

equilibrium sorption capacity by about 18 %. However, the most significant impact is associated with the apparent rate coefficient that drastically increases with ultrasonic agitation, especially at higher frequency. It is noteworthy that the ultrasonic treatment also changes the modeling of the kinetic profiles: contrary to mechanical agitation, the models can be ranked according to:

$PSORE > PFORE \gg RIDE$

In Fig. 9, the solid lines represent the fitting of experimental curves with the PSORE (using the parameters summarized in Table 3). The pseudo second order rate equation fits better the uptake kinetics under sonochemical treatment: the calculated equilibrium sorption capacity overestimates the experimental value by 6–7 %. The apparent rate coefficient k₂ increases by 3 times at F: 37 kHz and up to 4.6 times at F: 80 kHz (almost the same ratios can be reported for the variation on k₁ values). The beneficial effect of high frequency has been explained by its effect on the reduction of the cavitation bubbles generated during ultrasonic treatment [93]. During the collapse of cavitation bubbles, stronger microjets are formed that make more efficient the mechanical stress on the surface of the sorbent: the mass transfer is favored. This conclusion corroborates with the increase in the approximated value of the effective diffusion coefficient (up to $6.7-8.7 \times 10^{-13} \text{ m}^2 \text{ min}^{-1}$). The sonication of the solution improves the equilibrium performance and even more the mass transfer and reaction rate.

3.2.4. Sorption isotherms and thermodynamics

Fig. S5 compares the sorption isotherms for the biochar/GG/PVA composite with the isotherms for precursors GG/PVA and biochar. The profiles clearly demonstrate the superiority of the composite sorbent that exhibits higher sorption capacity at saturation (i.e., 1.7 vs. 1.46 and 0.53 mmol MB g⁻¹ for composite, biochar, and GG, respectively). However, the composite sorbent also shows a much higher affinity for MB as shown by the initial slope of the isotherm curves. Hence, the limit residual concentrations for reaching the saturation plateau corresponds to 0.2, 1.4, and 2.9 mmol MB L⁻¹ for composite, GG, and biochar, respectively. This figure clearly demonstrates the synergistic effect of the association of the different precursors.

The sorption isotherms are reported in Fig. 9, at different temperatures. The good overlapping of the duplicated experimental curves confirms the reproducibility of sorption tests. The profiles are characterized by a saturation plateau reached for residual concentration close to C_{eq} : 0.2 mmol MB L⁻¹ and a steep initial slope, which confirms the good affinity of the sorbent for MB. The maximum sorption capacity



Fig. 9. MB uptake kinetics – Modeling of sorption capacity vs. time for MB sorption for different frequencies of ultrasonic agitation (kHz) using the pseudo-second order rate equation (C_0 : 0.31 mmol MB L⁻¹; pH₀: 10.5, SD: 1.5 g L⁻¹; v: 200 rpm; T: 25 °C).

ranges between 1.68 and 1.84 mmol MB g⁻¹ (Table 4). Fig. S6 compares the modeling of the experimental profiles (as an example, for T: 25 °C) with the different models reported in Table S2. The asymptotic shape of the isotherm is not consistent with the exponential form of the Freundlich equation (Fig. S6a). The Temkin equation fails to fit the experimental profile at high residual concentrations and the models the most appropriate are reported in Fig. S6b. Table 4 confirms these trends. The comparison of R^2 and AIC values confirms that fitting quality follows:

Sips > Langmuir > Dubinin - Radushkevich

Langmuir equation assumes the sorption to occur through monolayer coverage, without interactions between sorbed molecules and with a homogeneous distribution of sorption energies. The Sips equation was developed to simulate adsorption in heterogeneous systems and get beyond the limitation of the rising sorbate concentration associated with the Freundlich isotherm. It is a kind of combination of the Langmuir and Freundlich equations. It reduces to the Freundlich isotherm at low sorbate concentrations but predicts monolayer sorption similarly to the Langmuir isotherm at high sorbate concentrations [94]. Therefore, the Sips isotherm shall be used to describe only monolaver adsorption systems: mathematical fit is usually improved due to the introduction of a third-adjustable parameter but with less physicochemical meaning regarding sorption mechanisms. The Dubinin-Radushkevich equation was designed for modeling the sorption of gases onto microporous sorbents: the sorption occurs by pore filling rather than by covering of internal macroporous or superficial surface (for nonporous sorbents) (as in the Langmuir or Brunauer-Emmett-Teller equations). Puccia and Avelina [60] recently questioned the application of this model for solid/ liquid sorption systems (especially for the interpretation and discrimination between physical and chemical sorption). Herein, the EDR values tend to increase with temperature (in the range of $8.48-10.4 \text{ kJ mol}^{-1}$), while the Temkin energy hardly varies between 4.96 and 5.9 kJ mol⁻¹.

The fitting of sorption isotherms with the Sips equation is shown in Fig. S7. In Fig. 10, the solid lines represent the fitting of experimental profiles with the Langmuir equation (using the parameters reported in Table 4). The calculated maximum sorption capacities (i.e., $q_{m,L}$) systematically overestimate experimental data (by about 10 %). Both the maximum sorption capacities and the affinity coefficients slightly increase with the temperature: the sorption is weakly endothermic. The application of the van' Hoff equation allows calculating the thermodynamic parameters (Fig. S8 and Table 5). The positive value of the enthalpy change (ΔH° : $\approx 12.9 \text{ kJ mol}^{-1}$) confirms the endothermic behavior of MB sorption onto the composite sorbent. The sorption is followed by increased randomness of the system as indicated by the positive value of the entropy change (ΔH° : $\approx 125 \text{ J mol}^{-1} \text{ K}^{-1}$). The negative value of the Gibbs free energy change (ΔG° : varying between -24.5 and -28.2 kJ mol⁻¹) confirms the spontaneity of the reaction; the slight increase (absolute value) is consistent with the slight improvement of sorption performance with temperature increase. In the case of MB sorption onto olive stones, Al-Gouthi and Al-Absi [78] reported the same order of magnitude for $\Delta G^\circ,$ while the values for ΔH° and ΔS° were substantially higher (i.e., 72 kJ mol⁻¹ and 297 J K⁻¹ mol⁻¹, respectively). On the opposite hand, Andrade Siqueira et al. [95] reported MB endothermic sorption onto sugarcane bagasse, with lower thermodynamic values (ΔH° : 1.93 kJ mol⁻¹ and ΔS° : 66 J K⁻¹ mol⁻¹). In the case of MB sorption onto biochar, Ji et al. [80] reported values as high as ΔH° : 42.5 kJ mol⁻¹ and ΔS° : 242 J K⁻¹ mol⁻¹. Amin et al. [96] compare the thermodynamic characteristics of biochar produced from banana and orange peel and observed significant variations in the enthalpy change (ΔH° : 8.7 and 18.8 kJ mol⁻¹, respectively).

The MB sorption properties of composite sorbent (and precursors) are compared in Table S3 with a selection of sorbents covering activated carbon-based materials, resins, biosorbents. Recently, Chen et al. [85] designed a new sorbent with outstanding sorption properties (i.e., $q_{m,L}$: superior to 8 mmol MB g⁻¹) at a comparable pH value to the pH selected

Table 4	
Fitting of sorption isotherms – Parameters of the models.	

Model	Temperature	25 °C		35 °C		45 °C		55 °C	
	series #	1	2	1	2	1	2	1	2
Exper.	q _m	1.695	1.694	1.680	1.705	1.715	1.740	1.809	1.839
Langmuir	q _{m,L}	1.926	1.958	1.887	1.935	1.868	1.924	2.030	2.042
	b _L	22.2	18.4	21.9	21.8	27.2	24.7	33.6	31.4
	\mathbb{R}^2	0.995	0.995	0.989	0.992	0.994	0.996	0.984	0.996
	AIC	-128	-130	-113	-119	-126	-133	-101	-128
Freundlich	k _F	2.58	2.56	2.46	2.56	2.55	2.68	3.15	3.23
	n _F	2.60	2.53	2.67	2.62	2.69	2.60	2.60	2.51
	R^2	0.904	0.914	0.911	0.906	0.913	0.922	0.883	0.913
	AIC	-65	-67	-67	-65	-67	-69	-59	-65
Sips	q _{m,s}	1.819	1.861	1.799	1.824	1.780	1.856	1.847	1.909
	bs	42.9	31.2	39.7	45.8	51.1	36.7	120.7	69.8
	n _s	0.864	0.883	0.872	0.843	0.875	0.918	0.789	0.856
	R^2	0.997	0.997	0.991	0.995	0.996	0.997	0.988	0.998
	AIC	-138	-135	-113	-123	-130	-134	-105	-140
Temkin	A _T	663.8	591.3	904.3	761.6	986.3	987.4	918.4	970.4
	b _T	8681	8602	9910	9307	10,094	10,035	8966	9209
	ET	5.12	5.09	5.90	5.46	5.89	5.77	4.96	5.01
	R ²	0.907	0.917	0.891	0.898	0.893	0.884	0.889	0.891
	AIC	-67	-70	-64	-65	-64	-62	-62	-62
D-R	q _{m,DR}	2.019	2.011	1.955	2.013	1.981	2.034	2.252	2.246
	$\mathrm{K}_{\mathrm{DR}} imes 10^8$	1.298	1.390	1.192	1.208	1.042	1.084	0.932	0.952
	E _{DR}	8.78	8.48	9.16	9.10	9.80	9.61	10.4	10.2
	R ²	0.988	0.991	0.984	0.985	0.986	0.990	0.970	0.985
	AIC	-109	-116	-104	-105	-107	-114	-88	-103

Units - q_m (mmol g^{-1}); b_L (L mmol⁻¹); k_F (mmol g^{-1})/(mmol L^{-1})^{nF}), n (dimensionless); b_S ((mmol L^{-1})^{nS}); b_T (J kg mol⁻²); A_T (L mol⁻¹); E (kJ mol⁻¹) is correlated to the DR constant: $K_{DR} = 1/E^2$.



Fig. 10. MB sorption isotherms at pH_0 10.5 at different temperatures – Langmuir modeling (C₀: 0.03–3 mmol MB L⁻¹; pH_0 : 10.5, SD: 1.5 g L⁻¹; v: 200 rpm; T: 25–55 °C; time: 150 min; duplicate experiments).

 Table 5

 Thermodynamic parameters for the sorption of MB (based on Langmuir constants).

Temperature (°C)	$\Delta { m H^\circ}$ (kJ mol $^{-1}$)	ΔS° (J mol ⁻¹ K ⁻¹)	$\Delta \mathrm{G}^\circ$ (kJ mol $^{-1}$)
25	12.89	125	-24.47
35			-25.72
45			-26.98
55			-28.23

in this study. Magnetic polystyrene-sulfonate/chitosan [97], mesoporous activated carbon [98], activated carbon/alginate [99], and functionalized starch/polyacrylamide hydrogel [83] show also sorption capacities in the range 2–2.7 mmol MB g⁻¹. The composite sorbent with a maximum sorption capacity of 1.93 is one of the most performing sorbents, just coming after these highly performant sorbents. It is noteworthy that the sorption capacities are much higher than those reported for synthetic commercial resins (in the range 0.01–0.3 mmol MB g^{-1} , [100,101]). However, the affinity coefficients are generally less favorable than alternative sorbents; this drawback is compensated by relatively fast kinetics. The sorbent has globally good and competitive sorption properties, as a compromise between sorption capacity and affinity, and kinetics.

3.2.5. Dye desorption and sorbent recycling

The sorption of MB being optimal in an alkaline solution, an acidic solution was selected for processing dye desorption; 0.7 M HCl solution was used for establishing desorption kinetics and testing sorbent recycling. Fig. 11 shows that the desorption proceeds within the first 60 min of contact with a desorption yield that reaches about 97 %. The first linear section of the curve (until 35–40 min) is followed by a slower desorption phase that probably corresponds to the progressive release of the dye bound in the internal porous network (which is hindered by steric hindrance). It is noteworthy that the presence of a high concentration of Cl^- in the eluent is supposed to enhance the self-aggregation of MB [89], which, in turn, is expected to decrease the diffusivity of MB. A



Fig. 11. MB desorption kinetics from dye-loaded composite sorbent using 0.7 M HCl solution (q_0 : 0.206 mmol MB g⁻¹; SD: 3 g L⁻¹; v: 200 rpm; T: 25 °C).

contrary trend is apparently observed; indeed, the desorption step is globally faster than the sorption stage.

Based on the high yield of desorption while using 0.7 M HCl solutions, this eluent was selected for comparing MB sorption and desorption efficiencies for six successive sorption/desorption cycles (with demineralized washing step between each operation). Table 6 shows the progressive decrease of the sorption capacity; however, in the sixth cycle, the loss remains below 5 % (compared with the first step). This is of the same order that the diminution of desorption efficiency, which decreases from 96.5 to 90.1 %: the cumulative desorption efficiency reaches 94 % in the sixth cycle. These results demonstrate the remarkable stability of sorption and desorption performances.

3.2.6. Application to MB-spiked wastewater

Wastewater was collected from the effluent of a local dyeing facility, in the industrial zone of Port Said, Egypt. The sample was spiked with MB at the concentration of 20.0 mg MB L^{-1} (i.e., 0.0625 mmol MB L^{-1}). The pH of the effluent was 6.7 (original pH). The conductivity was 870 μ S cm⁻¹ using a conductivity meter (4510 Bench Conductivity Meter, JENWAY, UK), while the color and the COD (Chemical oxygen demand) were 998 NTU and 242 mg $O_2 L^{-1}$, respectively, the analysis was performed using Hi 83,214 wastewater treatment photometer (HANNA instrument, USA). Fig. 12 shows the decay curves for COD and NTU with increasing sorbent dose. The composite sorbent strongly decolorizes the effluent by about 97 % with 1 g L^{-1} sorbent dose (increasing SD does not significantly the efficiency of color loss). The COD is also reduced; however, the decrease is more progressive and incomplete: the sorbent dose may be as high as 3 g L^{-1} for reaching the plateau of COD abatement (which does not exceed 62 %). A fraction of the organic contaminants remains refractory to the sorption treatment.

4. Conclusion

The comparison of sorption performances for GG/PVA and biochar with those of composite material clearly demonstrates the synergistic effect of precursors. The incorporation of the biochar (produced from a local Egyptian plant) in the composite allows for producing an efficient sorbent. The deprotonation of the sorbent in alkaline solutions improves the sorption of Methylene Blue which reaches a maximum close to pH 10.5. The fitting of sorption isotherms at different temperatures with the Langmuir equation allows calculating the enthalpy and the entropy change, which confirm the endothermic nature of MB binding onto the composite. The agitation speed influences uptake kinetics (with optimum conditions close to 200 rpm); however, alternative mode of agitation such as sonication allows drastically reducing the equilibrium time: the cavitation effects initiated by ultrasonic treatment enhance mass transfer properties (accessibility) and sorption capacities (availability of reactive groups). Applying sonication involves the reversal of fitting of experimental curves with pseudo-second order rate equation instead of the pseudo-first order rate equation. The sorption properties are remarkably maintained even in the presence of a large excess of NaCl, while the acidic desorption of the dyes allows minimized loss in sorption/desorption performance after six cycles. This bio-based material shows outstanding properties for the treatment of colored complex effluent (as confirmed by the efficiency in color removal of MB-spiked industrial effluent).

CRediT authorship contribution statement

Ahmed M. Elgarahy: Conceptualization, Methodology, Investigation, Data curation.

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Elsayed G. Zaki: Methodology, Investigation, Data curation. Shymaa M. Elsaeed: Methodology, Investigation, Data curation. Khalid Z. Elwakeel: Methodology, Investigation, Data curation.

Table 6

Sorbent recycling: sorption capacity and desorption efficiency (for 6 cycles).

Cycle	Sorption capacity $(q_{eq}, mmol g^{-1})$	Desorption efficiency (%)
#1	0.207	96.5
#2	0.206	96.0
#3	0.205	95.1
#4	0.203	93.6
#5	0.199	92.5
#6	0.197	90.1
	Loss at 6th/1st cycle: -4.8 %	Cumul. Desorption at 6th Cycle: 94.0 %

Experimental conditions – Sorption: C_0 : 0.32 mmol MB L⁻¹; pH₀: 10.5; SD: 1.5 g L⁻¹; time: 65 min; v: 200 rpm/Desorption: Eluent: 0.7 M HCl; SD: 3 g L⁻¹; time: 65 min; v: 200 rpm).



Fig. 12. Effect of sorbent dose on the treatment of industrial wastewater – COD and Color abatements (pH_0 : 6.7; time: 150 min; v: 200 rpm).

Abdullah Akhdhar: Methodology, Investigation.

Eric Guibal: Conceptualization, Review, Data curation, writing and Editing

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Data availability

Data will be made available on request.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.ijbiomac.2023.123355.

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