



HAL
open science

Matrix metalloproteinases: from molecular mechanisms to physiology, pathophysiology, and pharmacology

Luiz G.N. de Almeida, Hayley Thode, Yekta Eslambolchi, Sameeksha Chopra, Daniel Young, Sean Gill, Laurent Devel, Antoine Dufour

► To cite this version:

Luiz G.N. de Almeida, Hayley Thode, Yekta Eslambolchi, Sameeksha Chopra, Daniel Young, et al.. Matrix metalloproteinases: from molecular mechanisms to physiology, pathophysiology, and pharmacology. *Pharmacological Reviews*, 2022, 74 (3), pp.714-770. 10.1124/pharmrev.121.000349 . hal-03966470

HAL Id: hal-03966470

<https://hal.science/hal-03966470v1>

Submitted on 31 Jan 2023

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Matrix metalloproteinases – from molecular mechanisms to physiology, pathophysiology, and pharmacology

Luiz G. N. de Almeida, Hayley Thode, Yekta Eslambolchi, Sameeksha Chopra, Daniel Young, Sean Gill, Laurent Devel and Antoine Dufour

Departments of Physiology and Pharmacology and Biochemistry and Molecular Biology, University of Calgary, Calgary, Canada (L.dA., Y.E., S.C., D.Y., A.D.); Department of Physiology and Pharmacology, University of Western Ontario, London, Canada (S.G., H.T.) Université Paris-Saclay, CEA, INRAE, Medicaments et Technologies pour la Santé (MTS), SIMoS 91191 Gif-sur-Yvette, France (L.D.)

Running Title: Pharmacology of Matrix Metalloproteinases

99 Manuscript pages, 10 Figures, 2 Tables, 1 Supplementary Table

Abstract: 170 words

Introduction: 174 words

Discussion: 2,251 words

Abstract

I. Introduction

II. Matrix Metalloproteinases and their Inhibitors

A. History of Matrix Metalloproteinases (MMPs)

B. MMP Domains, Structures, and Mechanism of Action for Proteolysis

C. Endogenous Inhibitors and the Tissue Inhibitor of Metalloproteinases (TIMPs)

III. Molecular Mechanisms to Physiological Roles of Matrix Metalloproteinases

A. Extracellular matrix (ECM) Remodelling

B. Non-ECM MMP substrates

C. Intracellular roles of MMPs

D. Oxidative and nitrosative stress-activation of MMPs

IV. Pathophysiology of Matrix Metalloproteinases

A. Cancer

B. Gut and Joint Inflammatory Diseases

C. Vascular Diseases

D. Central Nervous System Diseases

E. Bacterial and Viral Infections

F. Sepsis

V. Pharmacological Inhibitors of Matrix Metalloproteinases

A. Clinical Trials for Cancer Treatment

B. Clinical Trials for Inflammatory Diseases Treatment

C. Potential Reasons for the Failure of MMP Inhibitors

D. Alternative Strategies to Inhibit MMPs

VI. Molecular tools for targeting active MMPs

- A. Substrate-based Probes for a Cleaved-Based Assay
- B. Ligand-based Probes to Bind to Active MMP
- C. Activity-based Probes to Crosslink Active MMPs

VII. Conclusions and Future Perspectives

- A. MMPs Non-Proteolytic Roles
- B. Non-ECM MMP Substrates
- C. Intracellular vs Extracellular Functions
- D. Post-Translational Modifications of MMPs
- E. Measuring MMP Activity and Diagnostic Applications
- F. The Future of MMP-directed Molecular Probes

Acknowledgements

References

Address correspondence to: Dr. Antoine Dufour, Departments of Physiology and Pharmacology and Biochemistry and Molecular Biology, University of Calgary Cumming School of Medicine, 3330 hospital dr. NW, Calgary, AB, Canada, T2N 4N1. E-mail: antoine.dufour@ucalgary.ca

Abbreviations: ABP, activity-based probe; ABPP, activity-based protein profiling; ACPPs, activatable cell penetrating peptides; ADAM, a disintegrin and metalloproteinase; ADAMTS, a disintegrin and metalloproteinase with thrombospondin motif; AG3340, prinomastat; BAY 12-9566, tanomastat; BB-94, batimastat; BB-2516, marimastat; BBB, blood-brain barrier; BM, basement membrane; BRET, Bioluminescence Resonance Energy Transfer; CA, cysteine array; CD, Crohn's disease; CNS, central nervous system; CPP, cell penetrating peptide; CSF, cerebrospinal fluid; EAE, autoimmune encephalomyelitis; ECM, extracellular matrix; EGFR, epidermal growth factor receptor; ELR, Glu-Leu-Arg; EMMPRIN, extracellular matrix metalloproteinase inducer; FRET, Förster Resonance Energy Transfer; GI, gastrointestinal; GM6001, ilomastat; GPI, glycosylphosphatidylinositol; HBV, hepatitis B virus; HIV, human immunodeficiency virus; IBD, inflammatory bowel diseases; ICU, intensive care unit; Ig, immunoglobulin; IL-1 β , interleukin-1-beta; IL8, interleukin-8; IFN α , interferon alpha; IFN γ , interferon gamma; LAP, latency-associated peptide; LRP1, lipoprotein receptor related protein; mAbs, monoclonal antibodies; MBP, myelin basic protein; mCD100, membrane-bound CD100; MMPs, matrix metalloproteinases; MRET, magnetic resonance tuning; MRI, magnetic resonance imaging; NIR, near-infrared; NMR, nuclear magnetic resonance; PET, positron emission tomography; PTM, post-translational modification; MS, multiple sclerosis; MT-MMP, membrane-type matrix metalloproteinase; MPO, myeloperoxidase; PAR1, protease-activated receptor-1; RECK, reversion-inducing cysteine-rich protein with Kazal motifs; ROS, reactive oxygen species; sCD100, soluble CD100; SPECT, single photon emission computed tomography; Tat, transactivator of transcription; TGF β , transforming growth factor-beta; TIMP, tissue inhibitor of metalloproteinase; TNF α , tumor necrosis factor alpha; VEGF, vascular endothelial growth factor; UC, ulcerative colitis.

Abstract—The first matrix metalloproteinase (MMP) was discovered in 1962 from the tail of a tadpole by its ability to degrade collagen. As their name suggests, matrix metalloproteinases are proteases capable of remodelling the extracellular matrix. More recently, MMPs have been demonstrated to play numerous additional biological roles in cell signaling, immune regulation, and transcriptional control, all of which are unrelated to the degradation of the extracellular matrix. In this review, we will present milestones and major discoveries of MMP research, including various clinical trials for the use of MMP inhibitors. We will discuss the reasons behind the failures of most MMP inhibitors for the treatment of cancer and inflammatory diseases. There are still misconceptions about the pathophysiological roles of MMPs and the best strategies to inhibit their detrimental functions. This review aims to discuss MMPs in preclinical models and human pathologies. We will discuss new biochemical tools to track their proteolytic activity *in vivo* and *ex vivo*, in addition to future pharmacological alternatives to inhibit their detrimental functions in diseases.

Significance Statement—MMPs have been implicated in most inflammatory, autoimmune, cancers and pathogen-mediated diseases. Initially overlooked, MMP contributions can be both beneficial and detrimental in disease progression and resolution. Thousands of MMP substrates have been suggested and a few hundred have been validated. After more than 60 years of MMP research, there remain intriguing enigmas to solve regarding their biological functions in diseases.

I. Introduction

Proteolysis is a key post-translational modification. Every single protein will encounter a protease, at some point in its lifetime, to either remove a signal peptide, cleave a propeptide for activation, process a protein to modify its functions, or initiate ubiquitination and degradation (Fortelny *et al.*, 2014) (Kappelhoff *et al.*, 2017). Proteases are fundamental enzymes implicated in all aspects of biology representing ~3% of the human genome and are divided into five mechanistic classes of catalysis: metallo, serine, cysteine, aspartic acid, and threonine proteases (**Fig. 1**) (Puente *et al.*, 2003); see the MEROPS database (<https://www.ebi.ac.uk/merops/>) for the lists of all known proteases (Rawlings *et al.*, 2010). Proteases can selectively process, cleave, or degrade all proteins, and our understanding of this web of interactions, known as the protease web (Fortelny *et al.*, 2014), remains largely uncharacterized. In this review, we will focus on matrix metalloproteinases (MMPs) and present our current understanding of their biological functions and roles in diseases, examine biochemical tools to study their functions, and discuss past and future clinical trials of MMP inhibitors.

II. Matrix metalloproteinases

A. History of Matrix Metalloproteinases (MMPs)

In 1962, J. Frederick Woessner published a landmark paper showing that a protease could degrade collagen in the mammalian uterus (Woessner, 1962). A few months after, Jerome Gross and Charles Lapiere published the first publication describing an MMP by demonstrating that the anuran tadpole underwent collagen turnover during metamorphosis as shown by collagen degradation in the skin, gills, and gut (Gross and Lapiere, 1962). Four years later, in 1966, the first MMP (MMP1), termed tadpole collagenase at the time, was purified from a tadpole tail fin and back skin (Nagai *et al.*, 1966). Not long after in the 1970s, MMP2 and MMP3 were identified. MMP2 was first sequenced in 1988 and was first called 72-kDa type IV collagenase/gelatinase A due to its ability to degrade denatured collagen/gelatin (Collier *et al.*, 1988). The laboratories of Guy Salvesen and Hideaki Nagase first purified MMP2 from human rheumatoid synovial fibroblasts in 1990 (Okada *et al.*, 1990). MMP3, originally termed proteoglycanase, was identified due to its lower molecular weight (54 kDa) and its ability to degrade proteoglycan and casein (Galloway *et al.*, 1983). In 1985, the laboratory of Zena Werb first purified MMP3 from rabbit synovial fibroblasts and later called it stromelysin (Chin *et al.*, 1985). The first evidence of the cross-activation of MMPs in a protease web was in 1987 by the activation of proMMP1 (latent MMP1) by MMP3 (Murphy *et al.*, 1987), and further demonstrated in 1989 by the laboratory of Gregory Golberg (He *et al.*, 1989). At the time of discovery, MMP4, -5, and -6 were believed to be novel MMPs but were later found to be redundantly identified MMP1, -2, or -3; therefore, they are not present in the current nomenclature (Overall and Sodek, 1987)(Otsuka *et al.*, 1988). As more MMPs were being identified, a single MMP was carrying multiple names; hence, in the late 1980s, the name MMP was first proposed (Okada *et al.*, 1987), and the International Union of

Biochemistry and Molecular Biology designated the family with the terminology. By 1991, there were seven known MMPs (MMP1, -2, -3, -7, -8, -9, and -10). After the completion of the human genome project, we now know that the MMP family in humans consists of 23 members, each composed of various domains (**Fig. 2**).

B. MMP Domains, Structures, and Mechanism of Action for proteolysis

MMPs belong to the superfamily of metzincin proteases, which is a family of multidomain zinc-dependent endopeptidases and can be grouped into six families: the astacins, the adamalysins (A Disintegrins And Metalloproteinases (ADAMs), the ADAM with Thrombospondin motif (ADAMTSs), the pappalysins, the serralysins and the MMPs (Sternlicht and Werb, 2001)(Sela-Passwell *et al.*, 2011)(Tokito and Jougasaki, 2016)(Wang *et al.*, 2021). The metzincins received this denomination due to a characteristic feature in their structure, a methionine (Met) residue at the active site and the use of zinc in the enzymatic reaction. They share the following conserved domain: HEXXHXXGXXH, where H is histidine, E is glutamic acid, G is Glycine, and X is any amino acid. The three histidines bind to zinc in the catalytic site (**Fig. 2-3A, B**) (Maskos, 2005). The three-dimensional structure of MMPs was elucidated by multiple groups over the years. For example, it has been demonstrated that the active site of this class of proteases has a deep cavity and its composition can be quite identical among its members, reaching up to 86% of similarity for MMP3 and -10 (Lovejoy *et al.*, 1994)(Laronha and Caldeira, 2020). Although their active sites located in the catalytic domain are highly conserved (Overall and López-Otín, 2002), the 23 members of the MMP family present various domains and exhibit some structural diversity (**Fig. 2**). MMPs can be divided into subgroups depending on their linear sequence similarity, domain organization, and substrate specificity. For example, MMP1, -3, -8, -10, -12, -13, -19, -20, and -27 are archetypal MMPs containing a signal peptide, a prodomain, a catalytic domain and a hemopexin-like C-terminal domain (**Fig. 2**). MMP2 and -9 were previously called gelatinases A and B due to their ability to cleave denatured collagen or gelatin; they contain a fibronectin type-II modules domain. MMP7 and -26, initially termed matrilysins, lack a hemopexin-like domain. There are six membrane-type (MT)-MMPs (MT1-MMP or MMP14, MT2-MMP or MMP15, MT3-MMP or MMP16, MT4-MMP or MMP17, MT5-MMP or MMP24, and MT6-MMP or MMP25) that localize to the plasma membrane via a transmembrane segment or a glycosylphosphatidylinositol (GPI) anchor domain (**Fig. 2**). MMP23 is a unique MMP that contains a cysteine array (CA) and immunoglobulin (Ig)-like domains; there is only limited knowledge in the literature on MMP23, and as such, the precise roles of these unique domains remain elusive (**Fig. 2**). MMP28 or Epilysin is the last member of the MMP to be identified and characterized, and it contains unique features not observed in other MMPs, such as the presence of threonine within its catalytic sequence (Lohi *et al.*, 2001). Therefore, MMPs contain high structural homology of their catalytic domain despite a lower overall sequence similarity due to the presence of specific domains (Stöcker *et al.*, 2008)(Sternlicht and Werb, 2001).

MMP1, -8, -13, and MT1-MMP, which were also termed collagenases in the literature, can cleave fibrillar collagen types I to IV, and also XI, resulting in the generation of two typical fragments: $\frac{1}{4}$ C-terminal and $\frac{3}{4}$ N-terminal (Visse and Nagase, 2003) (Hotary *et al.*, 2003) (Nagase *et al.*, 2006)(Sabeh *et al.*, 2009). MMP2 and -9 (gelatinases) can also cleave extracellular matrix (ECM) proteins *in vitro* including collagens and proteoglycans, resulting in changes in embryonic growth and development, tissue remodeling, inflammation, and wound healing (Sternlicht and Werb,

2001)(Overall and Kleinfeld, 2006b)(Fallata *et al.*, 2019). They previously received this denomination due to their distinct collagen-binding domain that contains three fibronectin type II repeats, conferring the ability to bind and cleave gelatin (denatured collagen) (**Fig. 2, 3**) (Allan *et al.*, 1995). MMP7 and -26 (matrilysins) can also cleave collagen and gelatin *in vitro*. They are unique due to the lack of a hemopexin domain, which is a characteristic structure found in all other MMPs (**Fig. 2**). MMP3 and -10 (stromelysins) share the same domain presented by the archetypal MMPs but cannot cleave interstitial collagen. MMP3 and -10 are closely related and share structural characteristics and substrate specificity. In addition, they can cleave numerous substrates and promote the conversion of a proMMP into an active enzyme (Nagase *et al.*, 2006). The MT-MMPs have a prodomain at the C-terminal, which contains a furin-like proprotein convertase site. Therefore, they are activated intracellularly via proteolytic processing of the furin domain, promoting activation of the enzyme that is further directed to the cell surface due to its membrane-anchoring domains (Sohail *et al.*, 2008)(Khokha *et al.*, 2013). They can be subdivided into two groups: type I transmembrane proteins (MMP14, -15, -16, and -24) and GPI anchored proteins (MMP17 and -25) (**Fig. 2**). MMP28 mRNA is typically expressed in epithelial cells of many tissues in both mice and humans, with a high expression in the lungs and skin, as well as in macrophages (Lohi *et al.*, 2001)(Rodgers *et al.*, 2009)(Gharib *et al.*, 2018). Interestingly, MMP28 appears to contain limited ability to cleave ECM proteins, as very few substrates have been identified to date, but it has been shown to stimulate chemokine expression (Ma *et al.*, 2013)(Manicone *et al.*, 2017)(Gharib *et al.*, 2018)(Long *et al.*, 2018).

MMP transcription is independently regulated as each cell type, depending on the stimulus, may produce different MMPs and at different levels (Sternlicht and Werb, 2001)(Overall and López-Otín, 2002). Most MMPs are not produced or only produced in low amounts under homeostatic conditions; however, this changes dramatically when a cell is stimulated with chemokines, cytokines, or growth factors which typically results in an elevation of MMP production. Other factors like cell shape or cell stress can also regulate MMP transcription (Kheradmand *et al.*, 1998). Some MMPs are uniquely regulated via specific transcription factor. For example, functional AP-1 site was demonstrated to mediate MMP2 transcription in cardiac cells via the binding of Fra1-JunB and FosB-JunB heterodimers (Bergman *et al.*, 2003). This regulation is often cell dependent; for example, in glomerular mesangial cells, MMP2 is regulated by YB1 transcription factor (Mertens *et al.*, 1997). Interestingly, during ischemia-reperfusion injury, elevated levels of FosB and JunB was identified whereas only JunB was identified under controls conditions (Alfonso-Jaume *et al.*, 2006). In a another study, ischemia induced the expression and binding of several transcription factors including c-FOS, c-Jun, JunB, FosB and Fra2 (Lee *et al.*, 2005). In skeletal muscle, AP1 and RE1 binding sites coupled with increased Fra1, Fra2, and AP2 (Liu *et al.*, 2010). Therefore, transcriptional regulation of MMPs appears to be cell selective, tightly regulated and is likely an adaptable process.

Changes in cell signaling via phosphorylation of various kinases such as MAPK, p38, FAK, ERK1 or AKT can induce MMP transcription and translation (Johansson *et al.*, 2000)(Dufour *et al.*, 2010)(Zarrabi *et al.*, 2011)(Madzharova *et al.*, 2019). Despite various extracellular signals and cell signaling pathways shown to regulate MMP expression, the transcription factor AP1 and its binding site are present in the promoter region of most MMPs and appears to regulate their transcription (Pendás *et al.*, 1997). AP1 contains members of the FOS and JUN family of oncoproteins which are likely involved in the regulation of MMPs in cancer cells (Overall and

López-Otín, 2002). Transcriptional control of MMPs is complex and there are multiple other nuclear factors that have been demonstrated to regulate MMPs including ETS, NF- κ B, STATs, TCF4, CIZ, p53 and CBFA1 (Bond *et al.*, 1999)(Sun *et al.*, 1999)(Nakamoto *et al.*, 2000)(Crawford *et al.*, 2001). Although most MMPs are regulated by the same transcription factors, some MMPs, like MMP26, have a unique TATA box and a transcriptional site located at 60 and 35 nucleotides upstream of its translation start site (Strongin, 2006). Only MMP12, -26 and -27 contains an unusual poly(A) site located upstream of their promoter, which, for the case of MMP26, abolishes accidental transcription; however, the functional role of this poly(A) site in MMP12 and -27 remains uncharacterized (Strongin, 2006). Overall, MMPs are transcriptionally regulated in multiple ways and tend to be closely associated with an increase in inflammation, immune cell infiltration, and tumorigenesis.

The first MMP 3D structures were published in 1994 by different groups showing the catalytic domain of MMP1 and MMP8 using X-ray crystallography, followed by the determination of the active full-length MMP1 in 1995 (Li *et al.*, 1995). To date, a multitude of new structures have been described for multiple MMPs, using tools such as X-ray crystallography and nuclear magnetic resonance (NMR) spectroscopy. These tools provided the full-length tridimensional structure of a few MMPs, including proMMP1 (Li *et al.*, 1995), proMMP2 (Morgunova, 1999), proMMP7 (Prior *et al.*, 2015), and proMMP12 (Bertini *et al.*, 2008). Based on X-ray crystallography data, MMPs are usually composed of a signal peptide, a prodomain, a catalytic site, a linker or hinge region, a hemopexin domain, and, depending on the MMP, a transmembrane region (**Fig. 3A**). Signal peptides can be of variable length and are found at the N-terminus, targeting the protein for secretion. The prodomain, typically ~80 amino acids, keeps the protease inactive and is proteolytically removed by other proteases once the enzyme has reached the respective site requiring its biological function. The catalytic site of ~170 amino acids harbors the zinc ion and is composed of five β -sheets and three α -helices. The linker or hinge region has a variable length, from 14 to 69 amino acids, and links the catalytic domain to the hemopexin-like domain. The hemopexin domain, composed of ~210 amino acids, has a four-blade propeller domain. Six MMPs, the MT-MMPs, have a transmembrane region (**Fig. 2**) (Nagase *et al.*, 2006)(Cauwe *et al.*, 2007)(Laronha and Caldeira, 2020). Notable exceptions include MMP7, -23, and -26 lacking the linker peptide and hemopexin domain (**Fig. 2**) (Nagase *et al.*, 2006). MMP2 and -9 are characterized by three repeats of a fibronectin type-II motif in the metalloprotease domain (**Fig. 2**).

MMPs are synthesized with a signal peptide located at their N-terminus (**Fig. 2**). This short peptide targets the protein to the secretory pathway before being removed in the endoplasmic reticulum, to give rise to the proMMP or zymogen (Sternlicht and Werb, 2001)(Maskos, 2005). The prodomain comprises three α -helices, connecting loops, and a cysteine switch (Sternlicht and Werb, 2001)(Visse and Nagase, 2003). The α -helices assume a nearly perpendicular shape among each other, gaining some flexibility due to the loop (Maskos, 2005). This arrangement promotes an oval structure located close to the catalytic domain and its active site cleft. The first loop, located between helix 1 and helix 2, has a protease sensitive sequence known as the “bait region”. In the case of proMMP1 and proMMP2, it is defined by EKRRN and SCNLF, respectively (Maskos, 2005)(Mannello and Medda, 2012)(Laronha and Caldeira, 2020). Cleavage within the bait region can destabilize the prodomain structure in proMMPs (Suzuki *et al.*, 1990)(Atkinson *et al.*, 1995). The flexibility that is a feature of this region imposes a challenge for proper elucidation of its

structure, which has only been described for proMMP2. There were two factors that played a role in resolving the structure of proMMP2's bait region: a shorter loop with six amino acids less when compared to other MMPs, and the presence of a disulfide bond that stabilizes the region (Maskos, 2005). In the prodomain of MMP1, a highly conserved sequence follows the three α -helices and is composed of PRCGXPD (Mannello and Medda, 2012)(Laronha and Caldeira, 2020). This region, also termed the cysteine switch, starts at amino acid 90 and ends at the cleavage site located at amino acid 99, marking the beginning of the active enzyme (Maskos, 2005). The cysteine switch rests on top of the substrate-binding site and is so-called due to its ability to inhibit the protease's function. Of note, this sequence binds from the P3' to the P2 at the substrate-binding site, which is the opposite of the actual peptide substrate and makes five hydrogen bonds with the main chain (Maskos, 2005)(Nagase *et al.*, 2006). The Arg91 and the Asp96 form a salt bridge, bending the loop structure placed above a histidine that takes part in the zinc-binding residues. Moreover, the salt bridge is protected from water by the side chains of up to three Tyr/Phe residues located in the intact prodomain (Maskos, 2005). Finally, the sulfhydryl group located on Cys92 interacts with the catalytic zinc ion, working as a fourth ligand and forming a tetrahedral coordination sphere that blocks the enzyme activity (Jacobsen *et al.*, 2010). Cleavage within the bait region allows exposure to other cleavage sites, destabilizing this Cys-Zn²⁺ interaction, allowing the binding of a water molecule to the zinc, and resulting in full activation of the enzyme (Nagase *et al.*, 2006)(Jacobsen *et al.*, 2010).

In proMMP1, the prodomain can regulate the entire enzyme configuration because it binds to the hemopexin domain and promotes a locked protein arrangement, which is the opposite of active MMP1 (Li *et al.*, 1995)(Jozic *et al.*, 2005). Upon the removal of its prodomain, MMP1 assumes an open configuration that creates a collagen-binding site. In the case of MMP11, -28, and the MT-MMPs, an increase of up to 22 residues can be observed in between the cysteine switch and the N-terminus (Maskos, 2005)(Marchenko *et al.*, 2002). Cleavage of the furin-like region located in this loop promotes enzyme activation, which takes place in the trans-Golgi network; therefore, these specific MMPs can then reach the cell surface in their active state. Lastly, MMP26 depends on specific processing mechanisms since an Arg/His mutation in the otherwise conserved cysteine motif, **P**HCGXXD, abolished its function and promoted an alternative activation pathway (Marchenko *et al.*, 2002).

The active site is largely conserved among the different MMPs, presenting as a sphere shape with a diameter of around 40 Å (Tallant *et al.*, 2010). There are two features on the surface of MMPs that are characteristic of the catalytic domain: a pocket that holds the Zn²⁺ ion and a shallow cleft for substrate binding, which further subdivides the region into an upper N-terminal subdomain and a lower C-terminal subdomain (**Fig 3A, B**). The secondary structures that constitute the catalytic domain include five β -strands (β I to β V), with four parallel strands (β I, β II, β III, β V) and one anti-parallel (β IV), three α -helices (α A to α C), and eight connecting loops (**Fig. 3C**)(Tallant *et al.*, 2010). Of note, three loops protrude from the surface of the upper subdomain: the connection between β II and β III (L β II β III), L β III β IV, and L β IV β V; these connections are a characteristic feature of MMPs, which separates them from other metzincins (Stöcker and Bode, 1995)(Gomis-Rüth, 2003)(Gomis-Rüth, 2009). Besides the Zn²⁺ ion in the catalytic site, which is required for catalysis, an additional Zn²⁺ ion and three Ca²⁺ ions promote conformational stability for the protein's structure. Starting from the N-terminus, the chain goes through β I, α A, β II, and β III (**Fig. 3A, C**)(Maskos, 2005). Next, it enters the L β III β IV loop, commonly referred as the S-loop, which has 16 amino acids and possesses two ion-binding sites (**Fig. 3C**)(Tallant *et al.*, 2010).

Zinc is the first ion housed by this loop, presenting a structural function, and tetrahedrally supported by three histidines (His147, His162, and His175, using the proMMP1 sequence), and monodentately by an aspartate (Asp149) (Tallant *et al.*, 2010). This chain composition is essential for proper MMP function, as a double-mutant presenting no metal ligands is inactive (Tallant *et al.*, 2010)(Paladini *et al.*, 2013). Calcium is the second ion supported by the S-loop, which is coordinated by the side chain of two aspartates and one glutamine (Asp154, Asp177, and Glu180), and three main chain carbonyl oxygens, all of them in a monodentate manner (Maskos, 2005)(Tallant *et al.*, 2010). The S-loop connects the chain to the β IV strand, which provides the structure for the active site cleft. Next, the main chain enters the L β IV β V loop that, in combination with L β II β III, holds another Ca^{2+} ion (Visse and Nagase, 2003)(Tallant *et al.*, 2010)(Laronha and Caldeira, 2020). Interestingly, in some structures, a third Ca^{2+} ion is housed in between the L β I α A and L β V α B loops via the carboxylate groups of two acidic groups and two main chain carbonyl groups (Maskos, 2005). After exiting strand β V, the chain goes through the L β V α B loop, which is a source of substrate specificity in the MMP family due to its high variability. This region constitutes the fibronectin type II motif, found in MMP2 and -9, spanning 177 and 178 residues, respectively (**Fig. 3C**) (Tallant *et al.*, 2010). Subsequently, the loop connects the polypeptide to helix α B, which also participates in the active site by providing two histidine residues as ligands for the catalytic zinc ion (**Fig. 3B**) (Tallant *et al.*, 2010). The glycine of the consensus sequence in the helix α B marks the end of the upper N-terminal subdomain. Moreover, this amino acid also allows the chain to perform a sharp turn closely followed by the third histidine residue that participates in the zinc ion interaction (**Fig. 3B**)(Tallant *et al.*, 2010). After the third histidine, a short loop precedes a tight 1.4-turn, named “Met-turn” due to a conserved methionine residue (Visse and Nagase, 2003)(Maskos, 2005)(Tallant *et al.*, 2010)(Laronha and Caldeira, 2020). Finally, a specificity loop that is important for substrate specificity connects the Met-turn with the α C helix, the last component of this lower C-terminal subdomain (**Fig. 3C**).

The catalytic domain is connected to the hemopexin domain via the linker or hinge region (**Fig. 2, 3A**)(Tallant *et al.*, 2010). The linker region is a proline-rich sequence of variable length (Laronha and Caldeira, 2020), spanning from 15-65 amino acids (Maskos, 2005). It is crucial for the MMP structure since it provides inter-domain flexibility (Overall and Butler, 2007)(Murphy and Nagase, 2008). Moreover, the hinge region is involved in the hydrolysis of complex substrates by MMP1, -8, and -13 since it requires the coordinated activation of both the catalytic domain and the hemopexin domains. For example, MMP1 and -8 have reduced collagenolytic function when mutations are introduced in the hinge region (Tsukada and Pourmotabbed, 2002)(Knäuper *et al.*, 1997). Lastly, the labs of Hideaki Nagase (Chung *et al.*, 2004), Chris Overall (Tam *et al.*, 2004), Irit Sagi (Rosenblum *et al.*, 2007), and Ghislain Opdenakker (Van den Steen *et al.*, 2006) have demonstrated that the linker region itself can also contribute in the binding, unwinding, and breakdown of collagen and various other MMP substrates.

The hemopexin-like domain is located at the C-terminus of MMP’s structure and was named after its similarity with hemopexin, which is a plasma heme-binding and heme-transport protein (**Fig 3A**)(Tallant *et al.*, 2010) (Faber *et al.*, 1995). The structure of the hemopexin domain can be characterized by the presence of four β -sheets (β I to β IV), or blades, which are named according to their appearance order. Importantly, the β -sheets follow a symmetrical structure that creates a central channel, which resembles a four-bladed propeller structure (Gomis-Rüth *et al.*, 1996)(Gohlke *et al.*, 1996)(Morgunova, 1999)(Cha *et al.*, 2002)(Jozic *et al.*, 2005). The β -sheets

are connected by short peptide loops and, in combination with short helices, shape the outer edges of the structure, which is connected and stabilized by disulfide bridges between blade I and blade IV (Maskos, 2005). The hemopexin domain is lacking in three MMPs: MMP7, -23, and -26 (**Fig. 2**). Interestingly, multiple MMPs that contain a hemopexin domain lose it soon after being activated. One example is MMP12; after losing its hemopexin domain, four amino acids within blade II of its hemopexin domain contain potent anti-microbial activities against both gram-negative and gram-positive bacteria (Houghton *et al.*, 2009). However, in most cases, the hemopexin domain is involved in substrate identification and specificity (Murphy *et al.*, 1992)(Dufour *et al.*, 2008) (Zucker *et al.*, 2009)(Dufour *et al.*, 2010)(Zarrabi *et al.*, 2011) (Dufour *et al.*, 2010)(Manka *et al.*, 2012)(Vandooren *et al.*, 2013).

Lastly, there is a total of six binding pockets in MMPs: S1, S2, S3, S1', S2', S3' (Rangasamy *et al.*, 2019). The S1, S2, and S3 are located on the right side of the catalytic zinc ion (Rangasamy *et al.*, 2019). Consequently, the S1', S2', and S3' are located on the left side of the zinc ion. The inhibitors or molecules interacting with the bindings pockets are named according to which pocket they interact with, such as P1, P2, P3, P1', P2', and P3' (Maskos, 2005). Among all the pockets, the S1' is the most variable and the most important since it determines substrate specificity (Nagase *et al.*, 2006). The pocket cavity, in turn, is defined by the Ω -loop, which is highly hydrophobic (Rangasamy *et al.*, 2019). Alternatively, S2 is the least variable pocket (Laronha and Caldeira, 2020). The remaining pockets, S2, S3', S1, and S3, sit in between with around the same level of variability (Nagase *et al.*, 2006). The depth of the S1' pocket can be used to subdivide the MMPs, since it can be present in a shallow, intermediate, and deep pocket configuration (Eckhard, Huesgen, Schilling, Bellac, Butler, Cox, Dufour, Goebeler, Kappelhoff, Keller, *et al.*, 2016)(Rangasamy *et al.*, 2019). MMPs are activated via various mechanisms to exert their proteolytic functions and need to be tightly regulated by endogenous inhibitors to assure an adequate physiological balance of their processing and degradative functions.

C. Endogenous inhibitors and the Tissue Inhibitor of Metalloproteinases (TIMPs)

Cloning and identification

The first tissue inhibitor of metalloproteinase (TIMP) was described in 1972 (Bauer *et al.*, 1972). Following the initial discovery and subsequent molecular characterization in 1985, TIMP1 was found to be identical to a factor with erythroid-potentiating activity (Gasson *et al.*, 1985). Importantly, due to this initial link between TIMP1 and the physiological regulation of erythropoiesis, as well as its ability to stimulate cells of the erythroid lineage, TIMPs and MMPs were initially studied separately and not in the same context of protein-protease interactions. In 1990, TIMP2 was cloned and sequenced from an A2058 human melanoma cell cDNA library (Stetler-Stevenson *et al.*, 1990). Importantly, in 1991, TIMP2 was demonstrated to inhibit collagenase activity in a 1:1 molar ratio complex, supporting a role for TIMPs in the inhibition of metalloproteinases (DeClerck *et al.*, 1991). First cloned in chicken in 1992 (Pavloff *et al.*, 1992), the human form of TIMP3 was first cloned in 1994 by multiple groups (Apte *et al.*, 1994)(Leco *et al.*, 1994)(Jay *et al.*, 2012)(Uría *et al.*, 1994)(Silbiger *et al.*, 1994)(Wilde *et al.*, 1994). Finally, human TIMP4 cDNA was cloned in 1996 (Greene *et al.*, 1996).

Structure and Overall Functions

The four TIMPs are variably glycosylated and have molecular weights between 22 and 28 kDa (Khokha *et al.*, 2013). Mammalian TIMPs exhibit basic similarities in their structure, folding to give a wedge-shaped appearance comprised of 2 main domains (Masciantonio *et al.*, 2017a). The first is an approximately 125 amino acid N-terminal domain, and the second is an approximately 65 amino acid C-terminal domain (Brew *et al.*, 2000). These two domains are connected through six conserved disulfide bonds, and each domain mediates different functions (Brew and Nagase, 2010). The N-terminal domain is critical for inhibiting metalloproteinase activity by forming a ridge that interacts with the catalytic site of metalloproteinases (Jayawardena *et al.*, 2019). This interaction occurs in a 1:1 stoichiometric inhibitor-to-enzyme ratio and blocks the active site Zn²⁺ molecule (Masciantonio *et al.*, 2017a)(Masciantonio *et al.*, 2017b)(Jayawardena *et al.*, 2019). The C-terminal domain of TIMPs was thought to have a limited role in metalloproteinase inhibition, but instead, it plays a role in mediating additional protein-protein interactions such as the interaction with the hemopexin domain of some proMMPs (Brew and Nagase, 2010)(Masciantonio *et al.*, 2017a). For example, TIMP2, TIMP3, and TIMP4 have the ability to interact with proMMP2, while TIMP1 and TIMP3 can interact with proMMP9 (Brew and Nagase, 2010).

Localization and Expression

Many mammalian tissues constitutively express the various TIMP family members, and the expression of TIMPs can be induced or inhibited in various situations, including during development, injury, and tissue repair (Masciantonio *et al.*, 2017a). Transcriptionally, TIMP levels can be altered by various growth factors, cytokines, and chemokines (Cabral-Pacheco *et al.*, 2020). TIMP1 has been identified in many tissues, including the brain, heart, arteries, colon, kidneys, liver, lungs, bladder, breasts, skin, lymph nodes, ovaries, uterus, prostate, and testis (Cabral-Pacheco *et al.*, 2020). The mRNA and protein levels of TIMP1 are generally elevated in several types of cancers and in settings of inflammation and tissue injury (Cabral-Pacheco *et al.*, 2020). For example, *TIMP1* mRNA expression can be upregulated by proinflammatory cytokines, including tumor necrosis factor-alpha (TNF α) and interleukin-1-beta (IL1 β) (Bugno *et al.*, 1999). Moreover, *TIMP1* mRNA and protein levels were found to be increased in the lungs of mice following bleomycin-induced injury (Madtes *et al.*, 2001).

TIMP2 is expressed in tissues such as the lymph nodes, brain, heart, arteries, colon, kidneys, liver, breast, ovaries, prostate, and testis (Cabral-Pacheco *et al.*, 2020). TIMP2 is generally considered to be constitutively expressed, but its levels can also be modified by stimuli including hormones, bacterial products, cytokines, and growth factors (Masciantonio *et al.*, 2017a). TIMP2 levels can also be altered in response to tissue damage or various disease states. For example, *Timp2* mRNA levels were elevated in rats after eccentric exercise-induced muscle damage (Koskinen *et al.*, 2001). Furthermore, TIMP2 was found to be overexpressed in malignant ovarian tissues (Hu *et al.*, 2004).

TIMP3 has been identified in the brain, heart, colon, kidneys, lungs, liver, breast, ovaries, prostate, and testis (Cabral-Pacheco *et al.*, 2020). The localization of TIMP3 is unique from the other TIMP family members in that it is localized to the extracellular matrix, whereas the other TIMPs are generally considered to be soluble (Stetler-Stevenson, 2008). This interaction with the extracellular matrix has been shown to occur via TIMP3 binding to heparan-sulphate and possibly chondroitin-sulphate proteoglycans (Yu *et al.*, 2000). Certain growth factors and cytokines can regulate the

levels of TIMP3, such as TNF α and transforming growth factor-beta (TGF β) (Masciantonio *et al.*, 2017a). Available TIMP3 levels can also be regulated by low-density lipoprotein receptor related protein (LRP)1, a scavenger receptor, that facilitates endocytosis of TIMP3 into the cell (Scilabra *et al.*, 2013). Furthermore, the expression of TIMP3 is frequently altered following infection and injury. For example, *Timp3* expression decreased rapidly following bleomycin-induced lung injury, a mouse model of pulmonary fibrosis (Gill *et al.*, 2010). Conversely, TIMP3 was increased in the kidneys of patients with diabetic nephropathy and chronic allograft nephropathy relative to healthy controls (Kassiri *et al.*, 2009).

TIMP4 remains the least studied member of the mammalian TIMPs. TIMP4 has been identified in the brain, heart, kidneys, breast, uterus, pancreas, colon, ovary, testes, prostate, and adipose tissue (Cabral-Pacheco *et al.*, 2020). The level of TIMP4 is altered in various types of cancer and is increased in the plasma of patients after acute myocardial infarction (Melendez-Zajgla *et al.*, 2008)(Kelly *et al.*, 2010). It is important to mention that the elevation of TIMP1, -2, -3, or -4 expression does not necessarily imply a net beneficial or detrimental pathogenic outcome. Careful validation of MMPs and other proteins linked with TIMPs expression must be evaluated in specific tissues and diseases being investigated.

MMP-Dependent TIMP Functions

TIMPs have been found to function through both metalloproteinase-dependent and metalloproteinase-independent mechanisms to regulate cellular functions such as angiogenesis, apoptosis, cell differentiation, growth, and migration (Brew and Nagase, 2010). However, the primary function for TIMPs is thought to be the regulation of metalloproteinase activity (Nagase *et al.*, 2006). The mammalian TIMPs display similar structural properties, so it is not surprising that they share overlapping metalloproteinase inhibition profiles (Masciantonio *et al.*, 2017a). Furthermore, all 4 TIMPs are collectively able to inhibit the active forms of all metalloproteinases studied to date, with binding constants being in the low picomolar range (Baker *et al.*, 2002). There are, however, some differences in the inhibition profiles of TIMPs and their affinities for different metalloproteinases. For example, TIMP1 does not appear to effectively inhibit several membrane-type MMPs (MT-MMPs), including MT1-MMP, MT3-MMP, and MT5-MMP (Brew and Nagase, 2010). TIMP1 is also a poor inhibitor of MMP19 (Brew and Nagase, 2010). Conversely, TIMP3 has a broad inhibition profile in comparison to the other TIMPs. In addition to inhibiting MMPs, TIMPs also inhibit several members of the ADAM and ADAMTS families (Brew and Nagase, 2010)(Jackson *et al.*, 2017). The *in vitro* activity of ADAM10 was demonstrated to be inhibited by TIMP1 and -3 but not TIMP2 and -4 (Amour *et al.*, 2000). TIMP3 also inhibits ADAM17 activity resulting in regulation of MAPK signaling via ERK and p38 in addition to TGF α in C α and M2 modified CHO cells, respectively (Xu *et al.*, 2012). In fact, TIMP3 appears to inhibit the greatest number of ADAMs (ADAM10, -12, -17, -28, -33) and ADAMTSs (ADAMTS1, -2, -4, -5) (Jackson *et al.*, 2017). Therefore, when assessing the biological functions of TIMPs, it is important to also consider ADAM and ADAMTS' proteolysis.

TIMP1 has been shown to promote cell survival by reducing apoptosis of hepatic stellate cells. The regulation of cell survival was at least partially MMP-dependent, as a TIMP1 mutant that selectively lacked MMP inhibitory activity had no anti-apoptotic effect (Murphy *et al.*, 2002). There is also evidence to suggest that TIMP1 negatively regulates hepatocyte growth factor activity during liver regeneration by an MMP-dependent mechanism (Mohammed *et al.*, 2005).

Additionally, TIMP1 can restrict microvascular endothelial cell migration by regulating MMP-mediated disruption of vascular endothelial-cadherin at cell-cell junctions (Akahane *et al.*, 2004). TIMP1 has also been found to regulate the process of airway epithelial cell migration by inhibiting MMP7 activity (Chen *et al.*, 2008). Specifically, TIMP1 appears to inhibit the MMP7-dependent cleavage of syndecan-1 from mediating bronchiole epithelial cell migration after injury (Chen *et al.*, 2008)(Chen *et al.*, 2009). Syndecans are heparan sulfate glycoproteins found on the surface of adherent cells, and their shedding is associated with increased cell migration (Teng *et al.*, 2012). TIMP2 has similarly been found to restrict syndecan-1 cleavage from the cell surface; however, this cleavage event was mediated by MT1-MMP (Endo *et al.*, 2003). In addition to regulating MT1-MMP, TIMP2 also regulates MMP2 and MMP9 activity (Cabral-Pacheco *et al.*, 2020). Following myocardial infarction, *Timp2*^{-/-} mice presented with greater left ventricular dilation, dysfunction, and severe inflammation. This phenotype was associated with impaired MMP2 activation but increased activity of MT1-MMP (Kandalam *et al.*, 2010). Furthermore, TIMP2 was found to inhibit both the invasion and migration of HCT-116 colon carcinoma cells via the regulation of MMP9 (Wang *et al.*, 2019).

Interestingly, TIMP3 is known to promote apoptosis. Through its N-terminal domain, TIMP3 inhibits the shedding of death receptors like Fas from the cell surface, thus promoting activation of an apoptotic signaling pathway (Ahonen *et al.*, 2003). As TIMP3 can be found localized to the ECM, it is unsurprising that TIMP3 also appears to have the ability to impact ECM-cell signaling (Gill *et al.*, 2006). Specifically, TIMP3 has been found to inhibit metalloproteinase-dependent fibronectin degradation during lung development, impairing signaling through focal adhesion kinase (Gill *et al.*, 2006). Similarly, *Timp3*^{-/-} mice showed enhanced collagen degradation in the peribronchiolar space and disorganized collagen fibrils in the alveolar interstitium that progressed as the mice aged (Leco *et al.*, 2001). This phenotype was proposed to be due to a shift of the MMP/TIMP balance, leading to enhanced metalloproteinase activity and consequent ECM degradation. Importantly, TIMP3 has also been found to promote normal function of the microvascular endothelial barrier (Arpino *et al.*, 2016). While the specific metalloproteinase remains unknown, this function of TIMP3 is at least partly mediated through metalloproteinase-mediated disruption of adherens junctions, as treatment with a broad-spectrum metalloproteinase inhibitor rescued both barrier dysfunction and endothelial cell surface localization of vascular endothelial-cadherin (Arpino *et al.*, 2016).

TIMP4 was determined to be the primary MMP inhibitor in human platelets and is involved in regulating platelet aggregation and recruitment (Radomski *et al.*, 2002). TIMP4 is known to negatively regulate MT1-MMP activity (Cabral-Pacheco *et al.*, 2020). Specifically, following ischemia-reperfusion injury in *Timp4*^{-/-} mice, there was a persistent increase in MT1-MMP activity that resulted in exacerbated diastolic dysfunction (Takawale *et al.*, 2014). TIMP4 has also been shown to regulate the activity of MMP2 by inhibiting MT1-MMP (Bigg *et al.*, 2001).

TIMPless mice

To study the biological functions of the four members in the *Timp* gene family, a quadruple *Timp* knockout termed the TIMPless mouse was generated by the lab of Rama Khokha (Shimoda *et al.*, 2014). Genetic removal of the 4 *Timps* resulted in runted pups with reduced body size and only 25% survival past 10 days postnatal (Shimoda *et al.*, 2014). TIMPless fibroblasts acquired hallmark cancer-associated fibroblast functions via an increase of α -smooth muscle actin, and

exosomes produced by these cells elevated cancer cell motility and cancer stem cell markers. In addition, the proteomes of TIMPless fibroblasts were found to be enriched in ECM proteins and ADAM10. Furthermore, knockdown of ADAM10 in TIMPless fibroblasts was sufficient to abrogate the functions of cancer-associated fibroblasts (Shimoda *et al.*, 2014). This study demonstrated the close interplay between the TIMPs, MMPs and ADAMs family members, and also highlight the difficulties in characterizing a single TIMP's biological role as the web of interactions appears to be inter-dependent among these proteins.

Latent MMP Interactions

As their name suggests, TIMPs are primarily recognized for their ability to inhibit metalloproteinases. However, there are instances of TIMPs interacting with latent (inactive) MMPs and facilitating their activation. The best-known example of this is the interaction between TIMP2 and proMMP2. First, TIMP2 binds to MT1-MMP on the cell surface, which acts to form a receptor for proMMP2; once proMMP2 is bound to this complex, another free MT1-MMP can cleave the prodomain of proMMP2, converting it to its active form (Sato *et al.*, 1994)(Brew *et al.*, 2000)(Masciantonio *et al.*, 2017a). In *Timp2*^{-/-} mice, proMMP2 activation is impaired, reflecting the unique role of TIMP2 in proMMP2 cell surface activation (Baker *et al.*, 2002). TIMP3 and TIMP4 have also been shown to interact with proMMP2, but both appear to restrict its activation (Brew and Nagase, 2010).

Other Endogenous MMP Inhibitors

MMPs have additional endogenous inhibitors, including alpha-2 macroglobulin, reversion-inducing cysteine-rich protein with Kazal motifs (RECK), tissue factor pathway inhibitor, and MMP prodomains (Sellers *et al.*, 1977) (Oh *et al.*, 2001)(Iyer *et al.*, 2012). Alpha-2 macroglobulin is a large 720 kDa glycoprotein that is regarded as the inhibitor of several proteases within the plasma (Rehman *et al.*, 2013). The protein contains four identical subunits, each with a 25 amino acid 'bait region' that is susceptible to proteolytic cleavage (Rehman *et al.*, 2013)(Serifova *et al.*, 2020). Cleavage of the bait region by active proteinases activates alpha-2 macroglobulin, triggering a conformational change that traps the protease (Rehman *et al.*, 2013)(Goulas *et al.*, 2017). This interaction is further anchored by covalent binding of an exposed reactive thioester of activated alpha-2 macroglobulin with accessible lysine residues of the protease (Rehman *et al.*, 2013). For example, Serifova *et al.* (2020) demonstrated that alpha-2 macroglobulin efficiently traps monomers of MMP9, preventing them from proteolytically cleaving large substrates such as gelatins. Additionally, following alpha-2 macroglobulin activation, receptor-binding domains can be exposed that bind to cell surface receptors, such as LRP1, which can stimulate internalization and clearance of the activated alpha-2 macroglobulin/protease complexes from the circulation (Serifova *et al.*, 2020). MMP9 alpha-2 macroglobulin complexes were shown to be removed from the extracellular environment through LRP1-mediated internalization.

RECK is a 110 kDa glycoprotein widely expressed in mammalian tissues; it was shown to mediate tissue remodeling and inhibit tumor angiogenesis and metastasis (Oh *et al.*, 2001)(Alexius-Lindgren *et al.*, 2014). The protein contains serine-protease-inhibitor like domains and is the only known MMP inhibitor that is cell membrane-bound, which occurs via a GPI anchor on RECK (Takahashi *et al.*, 1998)(Alexius-Lindgren *et al.*, 2014). Previous studies have suggested that RECK negatively regulates MMP2, -9, and MT1-MMP (Takahashi *et al.*, 1998)(Oh *et al.*, 2001)(Takagi *et al.*, 2009). RECK was also demonstrated to be a physiological inhibitor of

ADAM10, which is an upstream regulator of Notch signaling impacting brain development (Muraguchi *et al.*, 2007). RECK is also required for the development of multiple organs; *Reck*^{-/-} mice do not survive past embryonic day 10.5 and present with defects in blood vessel development, as well as compromised integrity of collagen fibers and the basal lamina, suggesting that this developmental defect may be due to aberrant metalloproteinase activity (Oh *et al.*, 2001). Therefore, despite a key role in regulating MMP functions, RECK's inhibition of other proteases, such as ADAM10, must be taken into consideration when studying its biological functions.

III. Molecular Mechanisms to Physiological Roles of Matrix Metalloproteinases

As indicated by their names, MMPs can cleave and degrade ECM proteins. However, the name matrix metalloproteinase is causing confusion in the literature in relation to its biological functions. It is estimated that only ~31% of their substrates are ECM proteins and ~69% are non-ECM proteins (Table 1, Supplementary Table 1, Fig. 4, as of December 2021)(Dufour and Overall, 2015). Therefore, in the recent decade, there has been a re-evaluation of their substrates and physiological roles.

A. Extracellular matrix (ECM) Remodelling

The ECM is a non-cellular structure that is highly dynamic and remains in close contact with cells either throughout their entire life or at important phases of their development. For this reason, the ECM is present in all tissues and organs of the body, providing structural support as one of its main functions (Hynes, 2009). The ECM has a tridimensional structure characterized by a distinct composition according to each organ; therefore, its composition is crucial since mutations in genes that encode key components of the ECM can lead to severe tissue defects or even embryonic lethality (Järveläinen *et al.*, 2009)(Bateman *et al.*, 2009). For example, gene deletion of fibronectin and collagens are often lethal at the embryonic stage, shown by multiple loss-of-function studies (Rozario and DeSimone, 2010). In the 1980s, the biochemical studies investigating the ECM composition focused on large extracellular matrices, such as those in the bone and cartilage. At the time, the biochemistry of native ECM was challenging due to its intrinsic characteristics, including insolubility and high levels of cross-linkage (Hynes and Naba, 2012). These challenges persist today, but the advancement of biochemical techniques and the availability of complete genome sequences allowed the identification of >300 ECM proteins in mammals (Hynes and Naba, 2012). These proteins constitute the core matrisome, including collagens, proteoglycans, and glycoproteins, as well as ECM-associated proteins that contribute to matrices in different scenarios such as growth factors (Hynes and Naba, 2012).

The ECM can be divided into two main types: the interstitial connective tissue matrix and the basement membrane (BM), which differ according to their localization and composition (Hynes and Naba, 2012). The connective matrix surrounds cells and provides structural support for tissues, while the BM is specialized in separating the epithelium from the neighboring stroma (Bonnans *et al.*, 2014). Importantly, the ECM function is not limited to tissue support, integrity, and elasticity. In fact, its structure is constantly being remodeled to sustain tissue homeostasis. ECM remodelling, degradation, or processing result in constant and dynamic changes in cell and tissue behavior (Lu *et al.*, 2011)(Cox, 2021). The ECM constitutes ~ 4% of the human proteome (Cox, 2021) and can regulate basic biological functions such as cell movement, shape, growth and survival via cell adhesion, cell-ECM, and cell-cell interactions (Sternlicht and Werb, 2001). Epithelial cells interact

with ECM components, which serve as ligands for cell receptors; these interactions result in signaling that controls proliferation, apoptosis, adhesion, migration, survival, or differentiation (Bonnans *et al.*, 2014). Furthermore, many cell types have a major role in remodeling and rebuilding the ECM by degrading, synthesizing, reassembling, and chemically modifying its components. These mechanisms are highly complex and require some fine-tuning to function properly; otherwise, dysregulated ECM composition is associated with a variety of pathological conditions, especially in response to injuries, and can worsen disease progression (Bonnans *et al.*, 2014). For example, uncontrolled ECM deposition or degradation is linked to fibrosis, osteoarthritis, and cancer (Zhen and Cao, 2014).

ECM remodeling is mainly accomplished by the cleavage of its components, controlling their abundance, composition, and structure. Moreover, the processing of the ECM by proteases can release bioactive molecules, like growth factors, with a variety of functions. In general, two main pathways are responsible for ECM degradation: an intracellular and an extracellular pathway (Dufour and Overall, 2015). For the intracellular processing, the ECM is degraded in the phagolysosome by lysosomal proteases, while the extracellular remodeling is performed by secreted proteases (Dufour and Overall, 2015). One misconception from the literature is that MMPs are the main acting proteases implicated in ECM degradation, especially collagens and proteoglycans. In reality, multiple other proteases other than the MMPs regulate ECM remodelling, such as the ADAMs, ADAMTSs, plasmin, cathepsins, neutrophil elastase, and several others (Brassart-Pasco *et al.*, 2020)(Cox, 2021)(Martin *et al.*, 2021). In addition, dysregulation or downregulation of protease inhibitors such as the TIMPs, cystatins, or serpins also contribute to changes in ECM remodelling (Cox, 2021). Therefore, ECM degradation is significantly more complex than initially thought and implicates much more than only the MMPs. Although collagens and proteoglycans (ECM) degradation is not carried out by MMPs alone, MMPs do play a major role in this biological process. For example, thyroid hormones during metamorphosis promote the release of several MMPs that present an active role in remodeling the intestine. Although the exact mechanism is not fully understood, active MMP11 is released by fibroblasts after hormonal stimulation and promotes epithelial cell apoptosis and growth of connective tissues (Patterton *et al.*, 1995)(Ishizuya-Oka *et al.*, 2000)(Amano *et al.*, 2005)(Bonnans *et al.*, 2014). Other MMPs are also upregulated by thyroid hormones, such as MMP2, -9, and MT1-MMP, which were shown to be increased in the classic tadpole model (Fujimoto *et al.*, 2007). In contrast to MMP11, MMP2 and MT1-MMP appear to have an important role in ECM remodeling after apoptosis has occurred during the late phases of intestinal morphogenesis (Fujimoto *et al.*, 2007)(Hasebe *et al.*, 2007).

MMPs are also important for branching morphogenesis. The ECM at the tip of ducts appears to be thinner than other regions that do not participate in the elongation process, indicating that proteases are likely required to cleave the ECM, allowing proper invasion in the surrounding tissue (Daley and Yamada, 2013). Tan *et al.* showed that MMP11 is crucial for mammary gland morphogenesis, and in *Mmp11*^{-/-} mice, a decrease in branching and downregulation of periductal collagen content occurred (Tan *et al.*, 2014). *Mmp2*^{-/-} and *Mmp3*^{-/-} mice are also defective in mammary gland branching, showing that these MMPs are implicated and important for proper gland development (Wiseman *et al.*, 2003). MT2-MMP was demonstrated to play a key role in digesting collagen IV in submandibular and mammary glands to sustain the proliferation of tip cells (Rebustini *et al.*, 2009). In lung embryogenesis, *Timp3*^{-/-} mice exhibited decreased bronchiole branching but

enhanced activity of MMPs associated with increased fibronectin degradation compared to wild-type controls (Gill *et al.*, 2006). Although mouse models of various MMP knockouts show defects in ECM remodelling, only the *MT1-MMP*^{-/-} mice exhibited craniofacial dysmorphism, osteopenia, dwarfism, and fibrosis of soft tissues resulting in death within 50-90 days after birth. Except for MT1-MMP, most MMP knockout mice resulted in fertile, viable, and relatively healthy animals suggesting that MMPs are not essential for embryogenesis and development (**Table 2**). Importantly, proteases regulate and modulate the branching process by other mechanisms besides cleavage of the ECM. For example, the hemopexin domain in MMP3 binds to WNT5B leading to an increase of mammary stem cell numbers, which results in increased branching morphogenesis (Kessenbrock *et al.*, 2013). Taken together, these results demonstrate that MMPs regulate the ECM during branching morphogenesis via proteolytic and non-proteolytic mechanisms.

In the ECM, MMPs are also involved in the degradation and processing of growth factors and their receptors, which actively participate in the wound healing process, including the modulation of angiogenic factors to promote or inhibit angiogenesis (Schultz and Wsocki, 2009). The processing of the diverse reservoir of growth factors located in the ECM can expose regions capable of activating growth factor receptors or release them to the interstitial space (Mott and Werb, 2004). Two examples of growth factors released from the ECM by MMPs include TGF β and vascular endothelial growth factor (VEGF). TGF β belongs to the transforming growth factor superfamily and it is classified as a multifunction cytokine, exerting different tasks on a wide variety of biological processes (Mott and Werb, 2004). Importantly, after TGF β is secreted, it is maintained in a latent form by interacting non-covalently and forming a complex with a latency-associated peptide (LAP). The TGF β -LAP complex remains bound to the ECM via an interaction between LAP and a protein belonging to the fibrillin family, the latent TGF β binding protein (LTBP) (Mott and Werb, 2004). TGF β becomes active when released from the complexed form and MMP2, -9, -13, and MT1-MMP can directly activate TGF β by releasing it from LAP (Yu and Stamenkovic, 2000)(Mu *et al.*, 2002). Moreover, MMP2 and -9 can cleave a soluble form of LTBP, while only MMP2 can cleave the ECM bound LTBP (Dallas *et al.*, 2002). These results indicate that MMPs are important for TGF β activation, either by directly inhibiting of the LAP interaction or releasing the TGF β complex from the matrix via LTBP cleavage.

MMPs can also activate VEGF, an important regulator of angiogenesis, as it stimulates vascular permeability and vessel growth (Park *et al.*, 1993)(Bergers *et al.*, 2000). Heparan sulfate proteoglycan in the ECM can tightly bind specific isoforms of VEGF, reducing its bioavailability (Park *et al.*, 1993). This process can also be reversed by proteolytic cleavage of the ECM, leading to the release of bound VEGF. For example, Bergers *et al.* demonstrated that MMP9 is associated with VEGF release and the development of pancreatic neuroendocrine tumors in the RIP-Tag2 insulinoma model, switching the environment from vascular quiescence to active angiogenesis (Bergers *et al.*, 2000). These studies show the importance of ECM as a rich reservoir of latent growth factors and cytokines, capable of modulating the tissue during homeostasis and disease development.

B. Non-ECM MMP substrates

N-terminomics and proteomics techniques were used to profile hundreds of cleavage sites in proteomes associated with MMP activity and identified numerous MMP substrates unrelated to

the ECM (Butler and Overall, 2009)(Starr, Bellac, *et al.*, 2012)(Dufour and Overall, 2013)(Bellac *et al.*, 2014)(Dufour and Overall, 2015)(B. Mallia-Milanes *et al.*, 2018). Data obtained from these techniques have resulted in an expansion and restructuring of our understanding of MMP biology, yet it is still underappreciated in the literature. Bioinformatics tools are essential for handling big data, and multiple programs have been developed to integrate these large datasets, such as MEROPS (Rawlings *et al.*, 2010) and TopFIND (Lange and Overall, 2011)(Fortelny *et al.*, 2015). In MEROPS (<https://www.ebi.ac.uk/merops/>), over 1 million proteases, ~92,000 cleavage sites, and ~6,500 peptidase-inhibitor interactions are included (*as of December 2021*). In TopFIND (<https://topfind.clip.msl.ubc.ca>), protein N- and C-termini, protease substrates, and proteolytic processing are presented from eight different organisms: *Homo sapiens*, *Mus musculus*, *Rattus norvegicus*, *Saccharomyces cerevisiae*, *Arabidopsis thaliana*, *Escherichia coli*, *Caenorhabditis elegans*, and *Danio rerio*. There are hundreds to thousands of potential MMP substrates, yet only a small number have been extensively validated *in vivo*, in animal models, or in humans. Apart from ECM and BM substrates, several non-ECM substrates have been identified, including chemokines, cytokines, cell-surface receptors, growth factors, metabolic proteins, and nuclear proteins (*for the full list, see Supplementary Table 1*) (Cauwe *et al.*, 2007) (Butler and Overall, 2009)(Cauwe and Opdenakker, 2010)(Dufour and Overall, 2013).

The tight regulation of MMPs is fundamental to what substrates they cleave, and substrates processing is cell-, tissue- and disease-dependent. Indeed, changes in the balance between MMP and TIMP ratio have a significant impact on disease progression and resolution of inflammation in both acute and inflammatory conditions. For example, multiple studies have demonstrated a key role of MMPs in regulating the biological functions of virtually all human chemokines functions by either an activation or inactivation cleavage resulting in a “go” or “stop” signal via chemokine receptor downstream signaling (McQuibban, 2000)(Van den Steen *et al.*, 2003)(Dean *et al.*, 2008)(Starr, Dufour, *et al.*, 2012)(Young *et al.*, 2019). MMP regulation of chemokines enhances or dampens leukocytes’ recruitment to the site of inflammation or pathogen infection. Eight CXCL chemokines have chemoattraction abilities to attract neutrophils, and, as an example, two neutrophil MMPs, MMP8 and -9, were demonstrated to cleave and activate interleukin-8 (IL8)/CXCL8 in a feedforward mechanism (Van den Steen *et al.*, 2003)(Tester *et al.*, 2007). All of the Glu-Leu-Arg (ELR)+ CXC chemokines, which are proteins able to attract neutrophils to the site of injury or inflammation, are processed by MMP12 released from macrophages, resulting in an inactivation of CXC chemokines signaling (Dean *et al.*, 2008). Other chemokines, such as CCL15 and CCL23, are also processed by multiple MMPs (MMP1, -2, -3, -7, -8, -12, 13, and MT1-MMP), resulting in their activation via a Ca²⁺ increase and an elevation of their chemotactic index (Starr, Dufour, *et al.*, 2012). MMP1 can cleave the Arg-Ser bond on protease activated receptor (PAR) 1. This substrate is also cleaved by thrombin, and proteolytic processing of PAR promotes its activation, which results, for example, in growth and invasion of breast carcinoma cells (Boire *et al.*, 2005)(Nagase *et al.*, 2006). Overall, MMPs can cleave multiple substrates including ECM and non-ECM related substrates.

C. Intracellular roles of MMPs

MMPs were first described as secreted proteases capable of cleaving ECM proteins, but we now know that their roles go beyond this initial dogma. A seminal discovery by Dr. Richard Schulz’s group in 2002 identified an intracellular form of MMP2 inside cardiac myocytes (Wang *et al.*, 2002). Later work by this group and David Lovett’s group demonstrated that there are three

intracellular isoforms (splice variants) of MMP2 since its signal peptide inefficiently targets MMP2 to the secretory pathway, thus resulting in ~50% of MMP2 remaining in the cytosol (Lovett *et al.*, 2012)(Ali *et al.*, 2012)(Lovett *et al.*, 2014)(Bassiouni *et al.*, 2021). One splice variant of MMP2, identified in the cytosol of cardiomyocytes, lacks 50 amino acids from its N-terminus and was termed MMP2_{NTT50} (Ali *et al.*, 2012). Another MMP2 intracellular variant missing 76 amino acids, termed MMP2_{NTT76}, was identified after induction of oxidative stress and was shown to activate NF- κ B and NFAT mitochondrial-nuclear stress signaling (Lovett *et al.*, 2012). MMP3 contains a proline residue that impacts its conformation and could weaken its signal sequence efficiency resulting in MMP3 localized in the nucleus in human chondrosarcoma-derived chondrocytic HCS-2/8 cells (Eguchi *et al.*, 2008). Additionally, MMP3 was found to be localized in the nucleus of human liver cancer cells (HepG2 cells) and liver myofibroblasts and was demonstrated to induce apoptosis via its catalytic activity (**Fig. 5**) (Si-Tayeb *et al.*, 2006). In dopaminergic neurons, intracellular MMP3 was also shown to induce apoptosis (Choi *et al.*, 2008). Further, intracellular MMP10 was also identified within neurons where it was demonstrated to cleave huntingtin into a toxic fragment resulting in cell death (Miller *et al.*, 2010). An MMP11 isoform of ~40 kDa was identified inside cancer cells due to an alternative gene promoter (Luo *et al.*, 2002). During viral infection, intracellular MMP12 (in HeLa cells) was demonstrated to mediate NF- κ B transcription resulting in IFN α secretion and host protection (Marchant *et al.*, 2014). Nuclear MT1-MMP was shown to modulate inflammation, independent of proteolysis, via the activation of a phosphoinositide 3-kinase δ (PI3K δ)/Akt/GSK3 β signaling cascade (**Fig. 5**) (Shimizu-Hirota, Xiong, Baxter, Kunkel, Maillard, Xiao-Wei Chen, *et al.*, 2012). MMP23 lacks a recognizable signal sequence (**Fig. 2**) and is therefore suggested to be intracellular although it has not been validated yet (Velasco *et al.*, 1999). Interestingly, most of MMP26 produced by cells remains intracellular due to MMP26's unique latency motif (Pro-His-Cys-Gly-Val-Pro-Asp) containing the conserved cysteine that is inactive (Marchenko *et al.*, 2002)(Savinov *et al.*, 2006)(Strongin, 2006).

Although multiple MMPs have been identified as intracellular proteases, it remains enigmatic how some MMPs are first secreted and can re-enter the cells at a later stage. Some MMPs have been found to enter the cells by endocytosis, either via clathrin-dependent or clathrin-independent mechanisms (Cauwe and Opdenakker, 2010). Clathrin-dependent mechanisms utilize cargo proteins that recognize adaptor proteins, resulting in packaging inside clathrin-coated vesicles subsequently absorbed by the cells (Traub, 2009)(Grant and Donaldson, 2009). Clathrin-independent mechanisms are less characterized, and function via caveolae or flotillin-dependent manner and can be linked to macropinocytosis and phagocytosis (Swanson, 2008)(Grant and Donaldson, 2009). For example, in neurons, MMP7 has been demonstrated to enter the cells via a clathrin-dependent mechanism implicating the processing of synaptosomal-associated protein of 25 kDa (Szklarczyk *et al.*, 2007). Other MMPs, such as MMP2, -9, and -13 have been demonstrated to enter the cells via the low-density lipoprotein-related protein 1 (LRP1) (Yang *et al.*, 2001)(Van den Steen *et al.*, 2006)(Raggatt *et al.*, 2006). MT1-MMP appears to be internalized via either clathrin-dependent (Jiang *et al.*, 2001) or independent mechanisms (Remacle *et al.*, 2003). Despite supporting evidence of MMPs entering via these mechanisms, it is still unclear how MMPs can exit the endosomes, enter the cytosol, and cleave substrates. Flip-flop mechanisms have been suggested to explain these mechanisms, but further experiments are needed to demonstrate these observations better (Cauwe and Opdenakker, 2010). Additionally, several MMPs contains nuclear localization signals in their sequences, which could potentially explain their entry to the

nucleus or, alternatively, could be brought into the nucleus via RNA-binding cargo proteins (Cauwe and Opdenakker, 2010)(Xie *et al.*, 2017)(Frolova *et al.*, 2020). Further characterization of the intracellular roles of MMPs will likely unravel new substrates and identify novel functions.

D. Oxidative and nitrosative stress-activation of MMPs

The production of reactive oxygen species (ROS) is an important mechanism to regulate normal metabolism but is also a key immune response to external and internal stimuli. ROS production includes numerous chemical oxidants such as hydrogen peroxide (H₂O₂), hydroxyl radicals (\bullet OH), and superoxide anion (O₂ \bullet^-) (Sies and Jones, 2020)(Muri and Kopf, 2021). During phagocytosis, ROS are generated and play important biological roles. Similarly, free radical nitric oxide (NO \bullet) is an important intra and extracellular regulatory chemical. When imbalances occur between the levels of O₂ \bullet^- and NO \bullet , peroxynitrite (ONOO $^-$) can form, resulting in nitrosative stress and increased damage to DNA, proteins, and lipids (Pacher *et al.*, 2007)(Brandes *et al.*, 2009). Reactive nitrogen species can result in three different post-translational modifications: 1) S-nitrosylation; 2) S-glutathiolation; and 3) tyrosine nitration; these can impact the latency-inducing cysteine to Zn²⁺ coordination in MMPs (Schulz, 2009)(Martínez and Andriantsitohaina, 2009).

We now know that MMPs have both extracellular and intracellular functions as MMPs have been identified in various subcellular compartments, including the cytosol, sarcomere, ER, nucleus and mitochondria (Bassiouni *et al.*, 2021). However, it is still unclear how the proteolytic activity of intracellular MMPs is regulated. One mechanism that contributes to regulating MMP activity inside the cell is the generation of reactive oxygen and nitrogen moieties (Cauwe and Opdenakker, 2010)(Bassiouni *et al.*, 2021). Via the oxidation of their thiol group within the cysteine switch, ROS has been demonstrated to activate MMPs even in the presence of the propeptide (Okamoto *et al.*, 2001)(Viappiani *et al.*, 2009). More precisely, peroxynitrite (ONOO $^-$) has been shown to activate some MMPs by cysteinyl S-glutathiolation or S-nitrosylation of cysteines 60 and 102, resulting in a disulfide S-oxide bond, as demonstrated for MMP1, -2, -8 and -9 (Okamoto *et al.*, 2001)(Viappiani *et al.*, 2009)(Jacob-Ferreira *et al.*, 2013). These types of MMP activation likely occur during an immune response or during stress. For example, the generation of hypochlorous acid from hydrogen peroxide occurs during neutrophil activation via myeloperoxidase activation and has been shown to activate MMP1, -2, -7, -8 and -9 (Weiss *et al.*, 1985)(Peppin and Weiss, 1986)(Fu *et al.*, 2001)(Meli *et al.*, 2003). Another example is oxidative stress that can occur during myocardial infarction. Specifically, the levels of peroxynitrite increase during myocardial infarction resulting in MMP2 activation and cardiac troponin I processing (Cheung *et al.*, 2000)(Wang *et al.*, 2002). In a feed-forward mechanism, inflammatory cytokines have been shown to increase peroxynitrite activating intracellular MMP2 (Ferdinandy *et al.*, 2000)(Qun Gao, 2003). Another example is the S-nitrosylation of MMP9 in cerebral ischemia that results in neuronal apoptosis (Gu *et al.*, 2002). Therefore, oxidative and nitrosative stress increases MMP activation and has been linked to pathological modifications of the biological functions of MMPs.

IV. Pathophysiology of Matrix Metalloproteinases

A. Cancer

Tumors are heterogeneous, dynamic, multicellular, and closely interact with their microenvironment (Quail and Joyce, 2013)(Maley *et al.*, 2017). Dysregulation of signalling networks in cancer impacts metabolism, growth, blood vessel formation, and immune regulation (Maley *et al.*, 2017)(Hiam-Galvez *et al.*, 2021). MMPs have been demonstrated to be implicated in most if not all of the dysregulated processes in cancer (Egeblad and Werb, 2002)(Piperigkou *et al.*, 2021). For several decades now, MMPs have been studied in the context of cancer progression, mainly for their ability to remodel the BM and to drive the cell invasion program resulting in increased metastasis (Egeblad and Werb, 2002)(Overall and Kleinfeld, 2006a)(Young *et al.*, 2019)(Piperigkou *et al.*, 2021). There is a strong relationship between cancer progression, clinical outcome, and upregulation of MMPs; however, what was originally thought about their detrimental roles in cancer has been challenged in the past two decades. The initial concept about MMPs was that broad-spectrum inhibition of their proteolytic activity would reduce ECM remodeling and prevent cell invasion and cancer metastasis. We now know that an increase of a specific MMP does not necessarily imply the promotion of tumor growth or metastasis; at least 10 MMPs have also been demonstrated to have protective roles in cancer (Dufour and Overall, 2013). Therefore, each MMP must be studied carefully as it likely has cell, tissue, or tumor-specific functions.

Matrix remodelling and cell invasion

The ECM is a fundamental component of all tissues and organs. Tissue integrity is maintained when there is a sustained equilibrium of ECM turnover, i.e., when ECM production is in balance with its degradation. However, tumors disrupt ECM homeostasis on the biochemical, biological, and structural levels due to their dynamic nature and dysregulated growth (Cox, 2021). Importantly, the ECM becomes highly dysregulated within tumors and can be both protumorigenic and antitumorigenic. For example, elevated MMP8 expression and activity is associated with good outcomes in oral squamous cell carcinoma and skin cancers (Korpi *et al.*, 2008)(Juurikka *et al.*, 2021) but is associated with poor outcomes in ovarian, digestive, and hepatocellular cancers (*reviewed by* (Juurikka *et al.*, 2019))(Cox, 2021). Several MMPs can cleave ECM proteins and selectively release cell surface-bound cytokines, growth factors, and their receptors, thereby impacting overall ECM integrity and tissue turnover (*for additional details, see* (Shimoda *et al.*, 2021)(Piperigkou *et al.*, 2021)). Multiple MMPs have been demonstrated to remodel the ECM, but there is strong evidence that either MT1-MMP, MT2-MMP, or MT3-MMP alone is sufficient to drive ECM transmigration and invasion, and could be the predominant MMPs driving ECM remodelling (Rowe and Weiss, 2008)(Rowe and Weiss, 2009). However, only the *Mt1-mmp*^{-/-} mouse suffers from severe craniofacial dysmorphism, arthritis, osteopenia, dwarfism, and fibrosis of soft tissues associated with type I collagen remodelling deficiency, which results in death within 50-90 days (**Table 2**). *Mt2-mmp*^{-/-} and *Mt3-mmp*^{-/-} mice are viable and live through adulthood with minimal defects suggesting that ECM remodelling during development is likely MT1-MMP dependent (Holmbeck *et al.*, 2004). Furthermore, MT1-MMP, but not MT2-MMP or other MMPs, was demonstrated to be the dominant MMP impacting both branching morphogenesis and carcinoma cell invasion (Hotary *et al.*, 2003)(Feinberg *et al.*, 2018).

Angiogenesis

Tumors require an adequate blood supply for their high demands in glucose and nutrients; therefore, angiogenesis, the creation of new blood vessels from pre-existing vascular networks, is a key process in cancer progression (De Palma *et al.*, 2017). Hypoxia, the low availability of

oxygen, is a key aspect of tumor angiogenesis where hypoxic cancer cells increase their secretion of multiple growth factors, including VEGFA, to increase vessel formation and reach the appropriate oxygen supply (Potente *et al.*, 2011). MMPs have also been closely implicated in regulating angiogenesis in most cancers, and broad-spectrum inhibition using batimastat (BB-94, **Fig. 6**) resulted in inhibition of the angiogenic switch in premalignant lesions (Bergers, 1999). Sprouting angiogenesis involves ECM remodelling by MMPs and other proteases produced by activated endothelial and immune cells (De Palma *et al.*, 2017). In tumors, MMPs often impact the vascular BM, which can be irregular, discontinuous, and loosely associated with endothelial cells and pericytes, resulting in an elevation of vascular leakiness; thus, an elevation in cancer cell intravasation and metastasis (Egeblad and Werb, 2002)(De Palma *et al.*, 2017)(Piperigkou *et al.*, 2021). For example, in a pancreatic neuroendocrine tumor, elevated MMP9 cleaves and releases VEGFA from the matrix, resulting in a switch between vascular quiescence to active angiogenesis initiation (Bergers *et al.*, 2000). There is evidence that multiple MMPs (MMP1, -3, -7, -9, -14, -16, and -19) can cleave and regulate VEGF bioavailability and vascularity in cancer (Piperigkou *et al.*, 2021). Processing of ECM components, such as collagen IV, XVIII, and perlecan by various MMPs (MMP1, -2, -3, -9, or -13) can initiate the production of anti-angiogenic products like tumstatin, endostatin, angiostatin, and endorepellin (Iozzo *et al.*, 2009). Despite multiple MMPs having important biological functions in the regulation of angiogenesis, individual MMPs also have tissue, cell, and tumor-specific functions. For example, Littlepage *et al.* demonstrated that MMP2, -7, and -9 have distinct roles in cancer progression and angiogenesis (Littlepage *et al.*, 2010). The authors used a rodent model of spontaneous prostate cancer and metastasis, where transgenic mice express SV40 large T antigen in their prostatic neuroendocrine cells, under the control of transcriptional regulatory elements from the mouse cryptdin-2 gene (CR2-TAg) (Littlepage *et al.*, 2010). Evaluated after 24 weeks using this model, *Mmp2^{-/-}/CR2-Tag* mice had reduced tumor burden with less lung metastases, increased survival, and decreased blood vessel density (Littlepage *et al.*, 2010). Interestingly, the survival and tumor growth of *Mmp7^{-/-}/CR2-Tag* or *Mmp9^{-/-}/CR2-Tag* mice was not impacted, but the *Mmp7^{-/-}/CR2-Tag* had reduced endothelial area coverage with decreased vessel size, and *Mmp9^{-/-}/CR2-Tag* mice had decreased tumor blood vessel size (Littlepage *et al.*, 2010). In another model, MMP9 was demonstrated to release soluble Kit ligand resulting in the transfer of endothelial and hematopoietic stem cells from a quiescent to proliferative niche and enabling the bone marrow repopulating cells to a differentiating vascular niche (Heissig *et al.*, 2002). These studies illustrate the heterogeneity of the roles that MMPs exert on tumor angiogenesis.

Apoptosis

Evading programmed cell death is key for the survival of cancer cells. Apoptosis can be initiated via extracellular receptors, such as the FAS receptors, resulting in a proteolytic cascade by intracellular caspases as well as degradation of additional substrates and nuclear DNA (Kessenbrock *et al.*, 2010)(Young *et al.*, 2019). MMPs have been demonstrated to interfere with the induction of apoptosis in cancer cells. For example, MMP7 was shown to cleave Fas ligand and in a doxorubicin-induced cell death assay, using SK-N-MC and SW-480 cancer cell lines, and MMP7 had a protective effect by reducing cell death (Mitsiades *et al.*, 2001). In a mouse model of pancreatic acinar-to-ductal metaplasia, MMP7 was demonstrated to accumulate during the metaplastic transition, increasing soluble Fas ligand (Crawford *et al.*, 2002). Additionally, *Mmp7^{-/-}* mice or mice carrying an inactive *FasL* gene demonstrated a significant defect in the development of progressive metaplasia and acinar cell apoptosis (Crawford *et al.*, 2002). When MDA-MD-231

and MDA-MB-435 human breast cancer cells were treated with sodium phenylacetate in the presence of GM6001, a broad-spectrum MMP inhibitor (**Fig. 6**), apoptosis decreased (Augustin *et al.*, 2009). Under similar conditions, the induction of autophagic vacuoles was also inhibited in MDA-MB-231 cells (Augustin *et al.*, 2009). In B cell chronic lymphocytic leukemia (B-CLL) patients, the interaction of proMMP9 via its hemopexin domain to $\alpha_4\beta_1$ integrin, and a 190 kDa variant of CD44 (CD44v), resulted in an induction of Lyn and signal transducer and activator of transcription 3 (STAT3) phosphorylation signaling, as well as prevention of apoptosis (Redondo-Muñoz *et al.*, 2010). Interestingly, the nuclear localization of various MMPs (MMP2, -9, -13) was demonstrated to induce cell death; however, no precise mechanism has yet been demonstrated (Xie *et al.*, 2017). Overall, MMPs are likely implicated in the regulation or inhibition of cell death; however, it is still unclear what the precise roles of each MMP are in these processes.

Cell migration

Cancer cells have been demonstrated to have increased motility and migratory capacity over non-cancerous cells (Paul *et al.*, 2017). In B-CLL cells, upregulation of MMP9 by the chemokine CXCL12 was demonstrated to enhance cell invasion and transendothelial migration via interaction with $\alpha_4\beta_1$ integrin (Redondo-Muñoz *et al.*, 2006). The chemokine CCL21 was also demonstrated to upregulate MMP9 and enhance cell migration of B-CLL cells via c-c chemokine receptor type 7 (CCR7) (Redondo-Muñoz, José Terol, *et al.*, 2008). Furthermore, binding of proMMP9 to $\alpha_4\beta_1$ integrin inhibited B-CLL cell migration and the proteolytic activity of MMP9 was required; importantly, this effect was not observed in normal B cells (Redondo-Muñoz, Ugarte-Berzal, *et al.*, 2008). When non-cancerous COS-1 cells were transfected with the cDNA encoding for various MMPs, their migration was enhanced, and this was independent of the MMP inhibitor CT1746 or the addition of TIMP1 or TIMP2 (Dufour *et al.*, 2008). This enhancement of cell migration by MMP9 and MT1-MMP was associated with their hemopexin domains and signaling via CD44 and epidermal growth factor receptor (EGFR) (Dufour *et al.*, 2010)(Zarrabi *et al.*, 2011). Interestingly, for MMP9, mutations in the outer amino acid sequences of blade I and IV of its hemopexin domain resulted in a reduction of cell migration using uncoated membranes in a transwell chamber assay (Dufour *et al.*, 2010). Using inhibitory peptide sequences mimicking blade I (⁵⁴⁸SRPQGPF⁵⁵⁵) and blade IV (⁶⁸⁹NQVDQVGY⁶⁹⁶) of the MMP9 hemopexin domain, HT-1080 and MDA-MB-435 cancer cells were inhibited in a dose-dependent manner (Dufour *et al.*, 2010). Using a similar approach for MT1-MMP and using peptide sequences mimicking blade I (³⁴⁹VMDGYMP³⁵⁶) and blade IV (⁴⁹⁶GYPKSALR⁵⁰³), the migration of HT-1080 cancer cells was significantly inhibited (Zarrabi *et al.*, 2011). Interestingly, in a xenograft mouse model using the MDA-MB-435 GFP labeled cancer cells that can spontaneously metastasize to the lungs, 20 mg/kg of blade I and IV mimicking peptides, injected intraperitoneally six times per week, significantly reduced the number and size of lung metastases but without any effect on tumor growth (Zarrabi *et al.*, 2011). Additionally, both peptides inhibited new blood vessel formations in a chicken chorioallantoic membrane (CAM) angiogenesis assay (Zarrabi *et al.*, 2011). Following a molecular docking study, small molecule inhibitors for MMP9 (A. Dufour *et al.*, 2011)(Das *et al.*, 2021) and MT1-MMP (Remacle *et al.*, 2012) were identified and demonstrated to inhibit cell migration.

There is unequivocal evidence that MMPs are implicated in most if not all aspects of cancer progression and metastasis (Overall and López-Otín, 2002)(Kessenbrock *et al.*, 2010)(Dufour and Overall, 2013)(Piperigkou *et al.*, 2021)(Das *et al.*, 2021). However, it is still challenging to determine what role each MMP plays, their synergies with other MMPs, other proteases, and what

substrates are cleaved leading to activation or inactivation, and, most importantly, what is the net balance between MMPs' detrimental and beneficial roles in tumors (Overall and Kleinfeld, 2006b)(Sela-Passwell *et al.*, 2011)(Dufour and Overall, 2013)(Young *et al.*, 2019). Therefore, despite over 30 years of investigating the use of MMP inhibitors for the treatment of various cancers, no broad-spectrum or selective MMP inhibitors have yet been approved.

B. Gut and Joint Inflammatory Diseases

The gastrointestinal (GI) tract, encompassing the organs from mouth to anus, breaks down food, absorbs nutrients, and eliminates waste. When homeostasis is lost in the GI tract, there is often an inflammatory response that develops and, if unresolved, can further hinder the gut's ability to perform its functions. Similarly, the joints and musculoskeletal components are susceptible to loss of function once chronic inflammation is initiated or established. Immune cells can secrete proteases and inflammatory components that ultimately result in cartilage and bone destruction. Similar destructive outcomes occur in the GI tract, and loss of epithelial lining results in a leaky gut. The inflammatory pathways that govern gut and joint diseases share common features, including a prevalence of type 3 immunity. Although its very nature remains imprecise, the gut-joint link is evident, with studies being published as early as 1958 (Wilkinson and Bywaters, 1958).

Inflammatory bowel diseases (IBD) are chronic inflammatory conditions that affect the GI, and it includes two distinct manifestations, Crohn's disease (CD) and ulcerative colitis (UC) (Ng *et al.*, 2017). CD and UC differ in the affected parts of the GI, pathophysiology, symptoms, and disease course but are centered on a strong inflammatory component that drives both diseases (Drewes *et al.*, 2020). CD can involve any part of the GI tract, and it is characterized by lesioned areas interposed between normal-appearing mucosa, affecting multiple layers of the tissue, leading to chronic pain, obstructions, and intestinal complications such as fibrotic strictures and fistulae (Drewes *et al.*, 2020). On the other hand, UC involves the rectum and colon, with lesions present in a continuous manner and an inflammatory process that is limited to the mucosal surface, leading to erosions and ulcers.

The global prevalence of IBD has been growing for the past 20 years, with Western countries, such as Canada, projecting an increase of 33.4% from 2015 to 2025 (Coward *et al.*, 2015). Importantly, the disease etiology remains elusive, but numerous studies have proposed MMPs as risk factors for IBD development and progression via proteolytic regulation or modulation of transcription factors (Dufour and Overall, 2015)(O'Sullivan *et al.*, 2015). MMPs are upregulated after cell-cell and cell-ECM interactions or as a response mechanism to proinflammatory cytokines widely expressed in IBD (Hu *et al.*, 2007)(O'Sullivan *et al.*, 2015). MMPs are produced by multiple cell types, including leukocytes, mesenchymal cells, and epithelial cells. For example, myofibroblasts can produce MMP1, -2, -3, and -9, while infiltrating neutrophils and macrophages are an important source of MMP8, -9, -10, and -12 (**Fig. 7**) (Andoh *et al.*, 2007)(Yoo *et al.*, 2011)(Drygiannakis *et al.*, 2013). Ulcerated and inflamed colon regions from UC patients show an increase of MMP1 (Wang, 2007) and MMP9 (Lakatos *et al.*, 2012) correlating with the inflammation severity. Although the exact mechanism initiating IBD is not yet known, multiple studies point towards the involvement of gut microbes. Moreover, MMPs cleave proteins and peptides that actively control the microbiota. For example, MMP7 can activate procryptidins into their active form, cryptidins, increasing their anti-microbial activity (**Fig. 7**)(Wilson, 1999)(Weeks *et al.*, 2006).

MMPs play a role in the modulation of IBD pathogenesis. Cytokines involved in the inflammatory process developed in the intestine are capable of increasing MMP levels. For example, TNF α and bradykinin can induce MMP3 expression via the signaling cascade containing PKC, PKD1, and MEK (Yoo *et al.*, 2011). Interleukin 17A (IL17A) and IL17F can upregulate the expression of MMP1 and -3 via myofibroblast, with IL17A presenting more potent effects than IL17F (**Fig. 7**). When combined with IL1 β and TNF α , both IL17 cytokines augmented the expression of MMP1 and -3 (Yagi *et al.*, 2007). In UC, TNF α induces expression of MMP1, which damages the colon by remodeling the ECM and leads to additional expression of TNF α in a feed-forward mechanism, ultimately culminating in excessive mucosal damage (**Fig. 7**) (Wang, 2007). TNF α is also dependent on proteolytic activity since it is synthesized as a membrane-anchored precursor (pro-TNF α) and requires proteolytic processing to release its soluble 17 kDa extracellular domain. The pro-TNF α cleavage is predominantly executed by the protease ADAM17, a member of the larger group of metzincin proteases (**Fig. 7**) (Horiuchi *et al.*, 2007). Interestingly, studies also indicate protective roles for MMPs in IBD, as *Mmp2^{-/-}* mice have a severe inflammatory response and higher susceptibility to disease development in a model of mucosal inflammation (**Fig. 7**) (Garg *et al.*, 2006). Therefore, additional investigation is required to better characterize better the protective functions of MMPs in IBD.

The study of MMPs in IBD is not limited to the intestine. A recent study by Majster *et al.* demonstrated an elevation of IL6 and MMP10 levels in the saliva of IBD patients, which correlated positively with the protease expression in the serum (Majster *et al.*, 2020). The identification of MMP expression in easily accessible fluids, such as saliva and serum, could be exploited to generate diagnostic tools, especially when a significant correlation with disease activity is observed. Currently, there is no single biomarker able to diagnose IBD with high accuracy and sensitivity. However, a panel of 13 blood biomarkers of CD patients has been developed to improve diagnosis (D'Haens *et al.*, 2020). The panel was evaluated in two cohorts, showing two distinct values, 0.962 and 0.693, for the area under receiver operating characteristic curve (D'Haens *et al.*, 2020). The 13 biomarkers included MMP1, -2, -3, -9, and extracellular matrix metalloproteinase inducer (EMMPRIN) (**Fig. 7**). Although not yet approved by the United States Food and Drug Administration, the test is commercially available as PROMETHEUS® Monitr™ Crohn's Disease by Prometheus Laboratories.

One of the challenges IBD patients face is the development of extraintestinal manifestations, with a prevalence ranging from 6 to 47% (Greuter and Vavricka, 2019). The most common form of extraintestinal manifestations is arthritis present as peripheral arthritis and as spondyloarthritis (Greuter and Vavricka, 2019). SpA is a group of inflammatory diseases, including ankylosing spondylitis, psoriatic arthritis, and reactive arthritis (Gracey *et al.*, 2020). Although the precise mechanism is not yet understood, the gut-joint axis is evident, which can be exemplified by the type 3 immunity, responsible for governing the immune response in the gut and joint tissue via IL17 and related cytokines (Greuter and Vavricka, 2019). SpA pathogenesis is marked by innate and adaptive immune cells infiltrating the joints, promoting chronic inflammation and tissue remodeling. In SpA, the degradation of collagens appears to be implicated, potentially associated with MMP3, -8, and -9 proteolysis (Moz *et al.*, 2017). MMP3 is elevated in ankylosing spondylitis patients compared to healthy controls, and it correlates with disease activity and C-reactive protein

(CRP) levels, a broad marker of inflammation. Less is known about MMP8 and -9 in SpA, but they also appear to be associated with disease activity (Mattey *et al.*, 2012).

In addition to SpA, studies have suggested an increased risk for the development of rheumatoid arthritis (RA) in IBD patients. Although the gut-joint axis has already been established, the link between RA and IBD remains debatable. In a systematic review and meta-analysis performed to determine if patients with IBD have a higher risk of developing RA, eight studies were included and revealed a significantly higher risk of RA in IBD patients (Relative Risk = 2.59; 95% Confidence Interval: 1.93–3.48) (Chen *et al.*, 2020). RA is a chronic inflammatory disease that has similarities to IBD and SpA but is generally characterized by rheumatoid factor, which are autoantibodies to immunoglobulin G (anti-IgG) and anti-citrullinated proteins antibodies (Steiner, 2007)(Smolen *et al.*, 2018)(Zaiss *et al.*, 2021). Importantly, the joint damage promoted by RA can lead to irreversible disability (Zaiss *et al.*, 2021). MMPs contribute to tissue damage in RA and are present starting at the early stages, as evidenced by the high levels identified within the first week after the onset of RA symptoms (Fang *et al.*, 2020). One of the key features in the synovium of RA patients is the expansion of the intimal lining due to the production of a large number of cytokines and proteases by synoviocytes (Hu *et al.*, 2007)(Hueber and McInnes, 2009). MMP1, -8, -13, and MT1-MMP are responsible for the proteolytic cleavage of important tissue components that surrounds the joint, while MMP9 can further degrade type II collagen fragments, generating important immunodominant epitopes (Vankemmelbeke *et al.*, 1998)(Van den Steen *et al.*, 2002)(Van den Steen *et al.*, 2004)(Dufour, 2015). Using monolayers and three-dimensional human RA synovial fibroblasts micro-masses, RANTES/CCL5 was demonstrated to induce MMP1 and -13 expression in a dose-dependent manner and the effect was reversed via inhibition of CCL5 (Agere *et al.*, 2017). Although MMPs are commonly associated with ECM degradation and tissue destruction, it is important to note that some MMPs, such as MMP-2, -8, and -12 have protective roles (Cox *et al.*, 2010)(Bellac *et al.*, 2014)(Dufour, 2015). For example, MMP12 secreted by macrophages can cleave interferon gamma (IFN γ) at the C-terminus (¹³⁵Glu↓Leu¹³⁶), and the truncated product of IFN γ loses the region that interacts with the IFN receptor 1 and fails to initiate the proinflammatory signaling pathway (Dufour *et al.*, 2018).

The inflammatory nature of IBD and various forms of arthritis translate to common or overlapping avenues of treatment. For example, therapies targeting TNF α have revolutionized the treatment for IBD, SpA, and RA (Smolen *et al.*, 2018)(Friedrich *et al.*, 2019)(Gracey *et al.*, 2020). One common approach is the use of anti-TNF α monoclonal antibodies (i.e., infliximab, adalimumab, golimumab, etanercept) that block the cytokine's functions. These inhibitors present different modes of action as TNF α is not limited to proinflammatory functions and can modulate cell migration, proliferation, and death (Levin *et al.*, 2016). Unfortunately, the response rate for anti-TNF α therapy remains low to moderate, and ~40% of IBD patients present primary or secondary non-responsiveness to the therapy (Friedrich *et al.*, 2019). SpA patients have a higher response rate with up to 60% of patients responding to the therapy (Gracey *et al.*, 2020). RA patients' response rate can be as high as 39% when golimumab is combined with methotrexate (Smolen *et al.*, 2018). Interestingly, MMPs have been linked to the lack of responsiveness towards monoclonal antibody therapy. When tissue homogenates from IBD patients were incubated with anti-TNF α monoclonal antibodies, MMP3 and -12 were shown to cleave infliximab, adalimumab, and etanercept (Biancheri *et al.*, 2015). Of note, a higher proportion of cleaved monoclonal antibodies is observed in non-responders to anti-TNF α therapy as compared to patients who responded better,

which could be partially explained by the higher amounts of MMP3 and -12 that are produced as a response to IL17A, a major player in the gut and joint inflammatory processes (Biancheri *et al.*, 2015). Collectively, these studies show that MMPs have important functions in IBD and arthritis as they influence disease progression and response to therapy with functions that go beyond the digestion of ECM proteins.

C. Vascular Diseases

Angiogenesis plays a role in several biological processes, including tumorigenesis, organ regeneration, wound healing, and normal development (De Palma *et al.*, 2017). As MMPs have been demonstrated to regulate angiogenesis, it is not surprising that MMPs play critical roles in vascular diseases such as atherosclerosis, myocardial infarction, hypertension, ischemia, and strokes (Hu *et al.*, 2007). Increased MMP1 levels in the heart were linked to a disruption of structural collagen and associated with cardiomyopathy (Kim *et al.*, 2000). In mice, overexpression of MMP1 in cardiac myocytes showed a decrease in cardiac interstitial collagen along with noticeable systolic and diastolic dysfunction (Kim *et al.*, 2000). Upregulation of circulating MMP2 has been associated with left ventricular remodeling after myocardial infarctions and poor prognostic outcomes (Matsunaga *et al.*, 2005)(George *et al.*, 2005). In mice, overexpression of MMP2 in hearts resulted in several physiological changes that are common with left ventricular remodeling, cardiac mitochondrial disruption, systolic heart failure, myocyte hypertrophy, and troponin I proteolysis (Bergman *et al.*, 2007). Interestingly, an intracellular splice variant of MMP2 has been identified and implicated in the development of diabetic cardiomyopathy (Lee *et al.*, 2019)(Bassiouni *et al.*, 2021). This MMP2 splice is missing the 76 amino acids at its N-terminus, which include part of the signal peptide (MMP2-NTT76) and has been observed in cardiomyocytes (Ali *et al.*, 2012)(DeCoux *et al.*, 2014)(Baghirova *et al.*, 2016)(Bassiouni *et al.*, 2021). MMP2 is localized at the sarcomere, while MMP2-NTT76 is localized in the mitochondria and in the nucleus (**Fig. 5**)(Bassiouni *et al.*, 2021). In a rat model of diabetic cardiomyopathy, treatment with doxycycline resulted in a reduction of action potential duration, calcium handling, and cardiac contractile force, suggesting that MMP2-NTT76 could be a contributing factor (Yaras *et al.*, 2008).

Following cardiovascular surgery, patients' arteries can develop scar tissue called a neointima, leading to a decrease in vascular elasticity and eventual vessel obstruction (Silvestre-Roig *et al.*, 2020). Neointima formation is often caused by the proliferation and migration of smooth muscle cells from the middle layer (tunica media) to the inner layer of the blood vessel (tunica intima), leading to scar tissue (Silvestre-Roig *et al.*, 2020). This scar tissue can lead to a lack of elasticity of the vascular tissue, which could lead to eventual plaque rupture that can obstruct smaller downstream blood vessels. This rupture of atherosclerotic plaques is the leading cause of coronary arterial thrombi in humans, and MMPs have been implicated in this process (Libby and Aikawa, 2002). There are different kinds of plaques that can develop: stable plaques that contain a thick fibrous cap that prevents the lipid core of the plaque from encountering the blood or vulnerable plaques that contain a thick fibrous cap between the plaque and the blood (Fitridge and Thompson, 2012). As the name implies, vulnerable plaques are more likely to rupture and trigger a myocardial infarction. Macrophage MMPs have been proposed to be a driving mechanism of this process (Fitridge and Thompson, 2012). In a mouse model of atherosclerosis, treatment with RXP470.1 (**Fig. 6**), a selective MMP12 inhibitor, significantly reduced atherosclerotic plaque cross-sectional

area by 2-fold (Johnson *et al.*, 2011). In the same model, MMP-13 was associated with the rupture of plaques via the degradation of the fibrous cap, as *Mmp13^{-/-}/apoE^{-/-}* mice displayed reduced collagenolytic activity within their lesions (Deguchi *et al.*, 2005)(Quillard *et al.*, 2014). MMP2 and -9 have been demonstrated to have a key role in neointima formation, and inhibition by broad-spectrum MMPs inhibitors (ilomastat or marimastat, **Fig. 6**) have been shown to reduce arterial constrictive remodeling (Sluijter *et al.*, 2006). Interestingly, *Timp1^{-/-}* mice have an increased risk for vascular scar tissue (neointima) formation (H. R. Lijnen *et al.*, 1999). *Mmp11^{-/-}* mice have an increase of neointima formation rather than a decrease suggesting that individual MMP could play distinct roles in angiogenesis (Lijnen *et al.*, 2002). Given the complex role of MMPs in angiogenesis and vascular diseases, the role of individual MMPs warrants further research and could lead to the identification of new drug targets.

D. Central Nervous System Diseases

MMPs are typically absent from a healthy central nervous system (CNS), and their upregulation may be linked with inflammation, neurological disorders, or injury (Yong *et al.*, 2001). For example, MMP9 was undetectable in the cerebrospinal fluid (CSF) of healthy individuals but elevated in multiple sclerosis (MS) and other inflammatory neurological pathologies (Gijbels *et al.*, 1992). During the pathogenesis of several CNS diseases, including multiple sclerosis (MS), bacterial meningitis, gliomas, stroke, and multiple others, CNS resident cells and invading immune cells secrete MMPs that drive blood-brain barrier (BBB) breakdown, permit leukocyte recruitment, and modulate cytokine and chemokine signaling (Leppert *et al.*, 2000)(Yong *et al.*, 2001)(Meli *et al.*, 2003). Accordingly, many studies have suggested that inhibition of MMPs is neuroprotective, although it has been hard to rule out that some MMPs could limit neuroinflammation or promote tissue repair (Chopra *et al.*, 2019). For example, MMP9 is involved in the remyelination process after a demyelinating injury. In *Mmp9^{-/-}* mice, accumulation of an inhibitory proteoglycan known as NG2 impairs remyelination by interfering with oligodendrocyte precursor cell maturation and differentiation. MMP-9 degrades NG2 deposits to facilitate the remyelination and repair response post-injury (Larsen *et al.*, 2003). Additionally, multiple studies suggest that MMP2, -7, and -9 remodel lesioned ischemic and infarct tissue to facilitate angiogenesis, vasculogenesis, and neurogenesis during post-stroke recovery (Rempe *et al.*, 2016). In the post-mortem analyses of the brains of MS patients, elevated levels of MMP2, -3, -7, and -9 were identified as compared to non-MS brains (Cuzner *et al.*, 1996)(Anthony *et al.*, 1997). In a rodent model of MS, rats with autoimmune encephalomyelitis (EAE) exhibited increased expression of MMP7, -9, and -12 (Clements *et al.*, 1997). Despite contrasting roles of various MMPs in EAE rodent models and human MS, broad-spectrum MMP inhibitors (GM6001, Ro-9790, or BB1101) alleviate or prevent EAE in rodents (Chandler *et al.*, 1997)(Wee Yong *et al.*, 1998). By digesting ECM, increasing the availability of growth factors (Zhao *et al.*, 2006), and aiding the migration of neuronal precursor cells to areas damaged by stroke, MMPs may support tissue remodelling and healing (Lee, 2006). Beyond their role in neurodegeneration and stroke, genetic polymorphisms of MMPs (particularly MMP9) have been implicated in neurological disorders such as schizophrenia, depression, and other neuropsychiatric conditions (Beroun *et al.*, 2019)(El Mouhawess *et al.*, 2020). Various mechanisms have been proposed for the involvement of MMPs in neuroinflammatory diseases. When injected in the CNS, MMPs have been demonstrated to induce demyelination; some of the MMP cleavage products of myelin basic protein (MBP) have been shown to induce rodents EAE (Opdenakker and Damme, 1994)(Chandler *et al.*, 1997). Human MMP9 can also cleave MBP, and

one of these peptide fragments is an immunodominant epitope in MS (Opdenakker and Damme, 1994).

White matter damage is a major complication in acute and chronic stroke, and MMPs have been demonstrated to play various roles associated with these injuries (Yang and Rosenberg, 2015). MMPs play a role in the acute injury that may lead to the widespread activation of other MMPs that persist in chronic stroke. In chronic injury, secondary damage to the white matter can occur, and MMPs that are expressed within the infiltrated leukocytes and blood vessels may result in disruption of the BBB and the release of toxic by-products that spread through the white matter (Yang and Rosenberg, 2015). More precisely, in the first phases after stroke, MMPs have been linked to an elevation of cerebral injury as MMP-mediated alterations often increase BBB leakage, cerebral edema, hemorrhage, and leukocyte infiltration (Yang and Rosenberg, 2015). MMPs have also been associated with damage of myelinated fibres (Rosenberg and Yang, 2007). Although MMPs can be detrimental during the early ischemic phase, they can be beneficial during the recovery phase by digesting the ECM, increasing the availability of growth factors, and aiding the migration of neuronal precursor cells to areas damaged by stroke. (Yong *et al.*, 2001)(Rempe *et al.*, 2016)(Chopra *et al.*, 2019). Overall, depending on the stage of MS and the timing post-stroke or post-neuronal injuries, the role of MMPs can be drastically different.

CNS development and stem cell niche

From early embryonic stages, MMPs play a role in the structural and functional changes during the development of the CNS. The neural stem cell niche is crucial in providing new neural cells such as the glia and neurons in the postnatal brain (Shan *et al.*, 2018). The ECM architecture and microenvironment directly influence the fate of neural stem cells and impacts their regenerative potential. Regulation and remodeling of the ECM are key processes in the development and maturation of stem cell populations during neurogenesis. Both intracellular and extracellular MMP12 have been demonstrated to regulate the generation of the brain's early postnatal ventricular-subventricular zone via ECM remodeling and inactivation of protease inhibitors (Shan *et al.*, 2018). Increased MMP12 expression was correlated with the development of ependymal cells, which line ventricles within the brain (Shan *et al.*, 2018). MMPs have been implicated in other stem cells such as the hematopoietic stem cells (reviewed by (Klein *et al.*, 2015)(Kessenbrock *et al.*, 2010)) involved in the induced mobilization process and their ability to remodel the ECM and cleave VEGF, TGF β , or TNF α ; however, their roles in controlling stem cells during neurogenesis still requires in-depth characterization.

Breakdown of the blood brain barrier

Leukocytes infiltrating the CNS parenchyma must transmigrate across multiple layers that comprise the BBB. First, immune cells penetrate the vascular endothelial cell monolayer and the underlying endothelial basement membrane. The process of transendothelial cell migration involves the binding of integrin $\alpha 4\beta 1$, expressed on leukocytes, to vascular cell adhesion molecule-1 on the endothelial surface of inflamed vessels, which induces MMP2 expression in encephalitogenic T cells and degradation of the subendothelial matrix (Graesser *et al.*, 2000). After extravasation from cerebral blood vessels, leukocytes accumulate in the perivascular space and inflammatory perivascular cuffs containing these leukocytes are one of the features found in the CNS of patients with MS and other neuroinflammatory conditions (Yong *et al.*, 2001)(Lassmann, 2019). In MS brains, perivascular macrophages exhibit an elevated expression of EMMPRIN, an

upstream inducer of MMPs (Kaushik *et al.*, 2019). EMMPRIN is also upregulated on peripheral leukocytes before the onset of clinical signs in the EAE mouse model and on infiltrating and resident cells including microglia within the CNS of symptomatic mice (Agrawal *et al.*, 2011)(Agrawal *et al.*, 2013). Using *in situ* zymography, Agrawal *et al.* previously showed that macrophage-derived MMP2 and -9 were critical for leukocytes to cross the parenchymal basement membrane and infiltrate the CNS parenchyma (Tran *et al.*, 1998)(Agrawal *et al.*, 2006). Mechanistically, MMP2 and -9 are able to cleave dystroglycan, a transmembrane receptor that anchors astrocyte endfeet to the parenchymal basement membrane, in turn permitting leukocyte infiltration (Agrawal *et al.*, 2006). Treating EAE mice with an EMMPRIN function-blocking antibody reduces MMP proteolytic activity at the glia limitans, thereby decreasing the infiltration of leukocytes into the CNS parenchyma and reducing disease severity (Agrawal *et al.*, 2011). Overall, the function and localization of MMPs vary across various areas of the brain.

Another example is the elevated expression of myeloperoxidase (MPO) observed in several autoimmune diseases, including MS. Predominantly associated with phagosomes such as neutrophils, macrophages, and dendritic cells, MPO is involved in the creation and maintenance of an alkaline milieu aimed at killing microbes and is also implicated in inflammatory disease progression (reviewed by (Arnhold, 2020)). Interestingly, in response to inflammation or microbe infection, the production of ROS via MPO can oxidatively activate proMMPs and inactivate TIMPs (Epstein and Weiss, 1989)(Wang *et al.*, 2007). In active MS, MPO was detected in microglial/macrophage cells within and surrounding CNS lesions (Strzepa *et al.*, 2017). In the EAE mouse model of MS, a magnetic resonance imaging probe detecting MPO can reveal lesions with inflammatory monocytes and neutrophils even in the absence of overt BBB breakdown (Pulli *et al.*, 2015). MPO likely contributes to MS pathogenesis through several mechanisms, one of which may involve modulating BBB integrity. For example, MPO inhibition restores BBB integrity (Zhang *et al.*, 2016) and LPS-induced BBB dysfunction is significantly lower in *Mpo*^{-/-} mice compared to wild-type (Üllen *et al.*, 2013). While MMPs have long been implicated to play a direct role in degrading the BBB, MPO may further facilitate BBB damage by oxidizing and inactivating their tissue inhibitors (Wang *et al.*, 2007), thus preventing TIMPs from keeping aberrant MMPs under control. Therefore, beyond tissue expression of MMPs and TIMPs, it is important to consider the effect of post-translational modifications, such as oxidation by MPO, on the activity of these proteins (discussed further in **Section VII. Conclusions and Future Perspectives**). Overall, MMPs play both beneficial (angiogenesis, cell survival, myelinogenesis, axonal growth, dampening of inflammation) and detrimental (neuronal death, demyelination, promotion of inflammation, tumorigenesis, disruption of BBB) functions within the CNS (Yong *et al.*, 2001); therefore, we still need to characterize the role of each MMP in specific regions of the CNS and identify what cells produce specific MMPs.

Therapeutic potential of MMP inhibition in neurological diseases

Despite several disappointing MMP clinical trials, the therapeutic potential of MMP inhibitors for neurological diseases still exists. In an open-label study, combined treatment of doxycycline (broad-spectrum MMP inhibitor) and interferon beta-1 α reduced MMP9 serum levels and brain lesions and improved Expanded Disability Status Scale values in relapsing-remitting multiple sclerosis patients (Minagar *et al.*, 2008). Tissue plasminogen activator is the only FDA-approved treatment for ischemic stroke patients; however, the neurotoxic side effects of tissue plasminogen activator are in part mediated by MMP9 upregulation, which can limit its therapeutic potential.

Combined therapy with selective MMP9 specific inhibitors that can cross the BBB may be able to address complications caused by tissue plasminogen activator (Tsuji *et al.*, 2005)(Gooyit *et al.*, 2012). For MMP targeted therapies to be effective, the timing of intervention should be carefully considered. In MS, MMP inhibition may be therapeutic during inflammatory phases of the disease, as blocking MMPs may decrease activation of proinflammatory factors and limit infiltration of immune cells into the CNS. On the other hand, promoting MMP activity later in the disease course may help facilitate tissue repair post-injury by clearing debris and other factors that inhibit remyelination processes within the lesion environment. It is tempting to speculate that MMPs inhibitors would only be useful during acute injury or early in the pathology of neuroinflammatory diseases; however, inhibiting the beneficial effects of MMPs during wound healing may impart more damage (Hsu *et al.*, 2006). Future studies should consider investigating these complex time-dependent functions of MMPs in health and neuroinflammatory diseases.

E. Bacterial and Viral Infections

The host immune response to a pathogen must be carefully coordinated to clear the infection while also sufficiently preserving tissue function. While some MMPs play a critical role in remodeling the ECM and coordinating the immune response during early infection, uncontrolled MMP activity during late or chronic stages of infection may cause host tissue damage and even death (Chopra *et al.*, 2019). The role of MMPs in bacterial or viral infection has been challenging to characterize as they play both beneficial and detrimental roles. Early in infection, MMPs facilitate leukocyte recruitment via processing cytokines, chemokines, and remodeling the ECM (Houghton *et al.*, 2009)(Marchant *et al.*, 2014)(Chopra *et al.*, 2019). However, unregulated MMP activity may contribute to immunopathology during chronic phases of the disease. For example, MMPs play a central role in tissue remodelling and fibrosis in patients with chronic infections, such as pulmonary tuberculosis. Pharmacological inhibition of phosphodiesterases has shown promise in dampening inflammation and reducing fibrosis, partly by downregulating MMPs in several disease models, including *Mycobacterium tuberculosis*-infected rabbits (Subbian *et al.*, 2016) and chorioamniotic infection that contributes to preterm premature rupture of fetal membranes (Oger *et al.*, 2005). Severe influenza pneumonia can lead to acute respiratory distress syndrome and death. Doxycycline, a broad-spectrum and unspecific MMP inhibitor, decreased the activity of MMP2 and -9 in a mouse model of virulent H3N2 influenza virus (Ng *et al.*, 2012). Doxycycline treatment decreased the expression of podoplanin (or T1 α , a membrane protein of alveolar type I epithelium) and thrombomodulin (an endothelial protein) in the bronchoalveolar lavage fluid. Moreover, it diminished inflammation and protein leakage in the lungs and reduced lung damage measured by histopathologic analyses (Ng *et al.*, 2012). This suggests that inhibiting MMPs may protect against lung injury in severe influenza infection; however, the timepoint of MMP inhibition should be carefully tested as MMPs may also play a critical role in controlling the early stages of infection. Recruitment of immune cells to the site of infection is essential for the clearance of a pathogen but infiltrating immune cells may generate excessive inflammatory responses that cause collateral tissue damage. Talmi-Frank *et al.* identified MT1-MMP as an ECM-degrading protease expressed by infiltrating myeloid cells in a model of influenza virus (Talmi-Frank *et al.*, 2016). Inhibition of MT1-MMP using a selective allosteric inhibitory antibody preserved lung tissue integrity by maintaining an abundance of collagen type I fibers and laminin during infection. Importantly, anti-MT1-MMP treatment did not modulate the immune response but protected against tissue destruction (Talmi-Frank *et al.*, 2016). Moreover, secondary bacterial infections are

the leading cause of mortality in high-risk populations with influenza and treatment using Tamiflu, an antiviral drug for influenza, does not reduce mortality rates when given post-infection. However, in mice co-infected with influenza virus and *S. pneumoniae*, preventative or therapeutic combined application of Tamiflu and MT1-MMP inhibition protected mice against sepsis by maintaining basement membrane integrity (Talmi-Frank *et al.*, 2016). This suggests that selectively inhibiting MMPs could be an effective drug target in infection if their inhibition does not impair viral clearance and supports tissue resilience to limit infection-related immunopathology. In response to a primary infection, the integrity of epithelial barriers may be compromised to allow for immune cells to enter tissues. However, failing to resolve this membrane post-infection may leave the tissue susceptible to secondary infections.

MMPs can also cleave transmembrane proteins to release soluble fragments that may exert independent functions. For example, membrane-bound CD100 (mCD100) is abundantly expressed on immune cells, such as CD4 T cells, CD8 T cells, and NK cells, and is increased upon cell activation. MMP-mediated proteolytic cleavage results in the shedding of a 120-kDa fragment corresponding to the extracellular domain of CD100, known as soluble CD100 (sCD100), which can act as a ligand to modulate the immune response. Altered mCD100/sCD100 ratios have been observed in viral infections, such as hepatitis B virus (HBV). Patients with chronic HBV infection have increased mCD100 expression on T cells, decreased serum sCD100 levels, and low serum MMP2 compared to healthy controls, suggesting insufficient cleavage of mCD100 from T cell membranes in patients with chronic HBV. In mice, inhibiting MMP2 and -9 activity during HBV infection using the synthetic peptide CTTHWGF TLC (Koivunen *et al.*, 1999) resulted in decreased serum sCD100 levels and delayed viral clearance (Yang *et al.*, 2019), possibly due to attenuated intrahepatic anti-HBV CD8 T cells responses.

Human immunodeficiency virus (HIV) infection has been associated with increased MMP9 levels in the CSF (Sporer *et al.*, 1998), and it also induces MMP expression in macrophages, causing neurotoxicity within the CNS (Yong *et al.*, 2001). Conant *et al.* showed that HIV-1 protein transactivator of transcription (Tat) increased the release of MMP1 and active MMP2 in mixed human neuron/astrocyte cultures (Conant, CS Hillaire, *et al.*, 2004), and induced MMP3 and -13 expression in primary murine astrocyte culture (Bozzelli *et al.*, 2019). HIV-1 Tat also increased the expression of MMP9 in the human astrocyte cell line CRT-MG (Ju *et al.*, 2009). Increased MMPs in the brain of HIV-infected patients may be one of the factors contributing to neurological complications, such as HIV-associated encephalitis and dementia. Tat can still be constitutively expressed by HIV-infected cells despite effective antiretroviral therapy; thus, pharmacological inhibition of Tat may be a therapeutic avenue to prevent HIV-associated neurocognitive disorders and reduce viral reservoirs (Sgadari *et al.*, 2019)(Joshi *et al.*, 2020). However, depending on the specific MMP and stage of infection, MMPs may play both protective and pathogenic roles. For example, primary human macrophages expressing a Tat sequence derived from a patient with HIV-associated dementia increased MMP2 and -7 release and activation, and supernatants from these cells caused significant neuronal death. Interestingly, this effect is Tat sequence dependent as brain-derived Tat from patients without HIV-associated dementia did not induce the same MMP upregulation. Neurotoxicity was inhibited by anti-MMP2 or -7 antibodies, suggesting that these MMPs are implicated in pathogenesis (Johnston *et al.*, 2001). However, another study suggested that MMP1 can enzymatically degrade Tat and decrease Tat-mediated neurotoxicity as well as HIV transactivation (Rumbaugh *et al.*, 2006). Thus, the protective or pathogenic fate of viral-host

interactions is MMP specific. Furthermore, MT1-MMP was demonstrated to be elevated on neuronal cell surface, resulting in proMMP2 activation from HIV-infected macrophages/microglial cells (Zhang *et al.*, 2003). Upon activation of proMMP2 by MT1-MMP, active MMP2 was demonstrated to switch receptor binding specificity from CXCR4 to CXCR3 via a four amino acid cleavage of the chemokine CXCL12/stromal cell-derived factor 1 (SDF1), resulting in neurotoxicity (McQuibban *et al.*, 2001)(Vergote *et al.*, 2006). As an example of the synergy between HIV and bacterial infections, females with bacterial vaginosis present with an altered microbiota, which colonizes the female reproductive tract and increases female susceptibility to HIV infection. Increased levels and activity of MMPs, particularly MMP1, -10, and -13, are detected in cervicovaginal fluid from women diagnosed with bacterial vaginosis compared to healthy women. *In vitro* co-culture experiments show that endocervical epithelial cells increase expression of MMPs, particularly MMP7 and -10, in response to challenge with the bacterial vaginosis-associated bacterium *Atopobium vaginae* when compared to Lactobacillus species present in healthy vaginal microflora (Cherne *et al.*, 2020). Further, the combination of MMP10 and -13 have been implicated in increasing HIV-1 transmigration through the endocervical epithelium (Cherne *et al.*, 2020). Therefore, the role of MMPs in barrier dysfunction may be one factor that influences one's susceptibility to concomitant or secondary infections. The crosstalk between pathogen-derived factors and host MMPs needs to be further characterized to understand how specific bacteria or viral agents may modulate the host immune response and contribute to pathology.

Cytokine and chemokine processing

In addition to ECM substrates, MMPs process multiple cytokines and chemokines to drive the recruitment of inflammatory cells and either promote or resolve inflammation at the site of infection (McQuibban *et al.*, 2000)(Cauwe *et al.*, 2007)(Dean *et al.*, 2008)(Starr, Dufour, *et al.*, 2012)(Proost *et al.*, 2017)(Young *et al.*, 2019). Importantly, the localization and activation of MMPs is a critical factor that dictates their function. In the antiviral response against coxsackievirus type 3 or respiratory syncytial virus, intracellular MMP12 was demonstrated to mediate NF- κ B transcription resulting in IFN α secretion and host protection (**Fig. 5**) (Marchant *et al.*, 2014). However, extracellular MMP12 was shown to cleave the IFN α receptor 2 binding site of systemic IFN α to regulate or resolve the antiviral response (Marchant *et al.*, 2014). The selective MMP12 inhibitor, RXP470.1, was demonstrated to inhibit extracellular MMP12 from inactivating IFN α but spares the beneficial activity of intracellular MMP12 and, in turn, dramatically increases systemic IFN α levels and decreases viral load in coxsackievirus type B3-infected mice (**Fig. 5**) (Marchant *et al.*, 2014). Thus, studies testing MMP inhibitors should consider the differential role intracellular and extracellular MMPs may play in immune responses. Designing specific MMP inhibitors that account for differences between intracellular and extracellular enzyme activity may prove to be more efficacious.

F. Sepsis

Sepsis is defined as a dysregulated host response to an infection leading to organ dysfunction and ultimately organ failure (Singer *et al.*, 2016). Sepsis remains one of the major causes of mortality in intensive care unit (ICU) patients and a leading cause of death from infection (Genga and Russell, 2017). Despite its importance and increasing global prevalence, safe and effective therapeutics for sepsis are lacking. Understanding the pathophysiology behind sepsis is complicated by the fact that infections can be caused by a broad range of pathogens, including

bacteria, viruses, and fungi (Dolin *et al.*, 2019). Several factors have been identified that contribute to the multiple organ dysfunction characteristically observed in septic patients. For example, a prominent feature of sepsis is a strong proinflammatory response resulting in increased levels of circulating leukocytes, cytokines, chemokines, and other mediators of tissue damage (Gyawali *et al.*, 2019). A second prominent feature of sepsis is injury and dysfunction of the microvasculature leading to extravascular fluid leakage and tissue edema (Arpino *et al.*, 2016).

Metalloproteinase Expression in Sepsis

Sepsis has been found to alter the expression of multiple MMPs. For example, a group of ICU patients with severe sepsis presented with plasma levels of MMP3, -7, -8, and -9 that were elevated more than 3-fold compared to healthy controls (Yazdan-Ashoori *et al.*, 2011). Another study found markedly elevated levels of active MMP1 in the plasma of human sepsis patients vs. healthy controls (Tressel *et al.*, 2011). Altered expression of TIMPs has also been identified in sepsis. Severe sepsis patients admitted to the ICU were found to have elevated levels of TIMP1, TIMP2, and TIMP4 relative to healthy controls (Yazdan-Ashoori *et al.*, 2011). Conversely, TIMP3 protein and mRNA levels appeared to be decreased in pulmonary endothelial cells after treatment with a mix of clinically relevant septic cytokines (Arpino *et al.*, 2016). Similarly, IL1 β and TNF α , two proinflammatory cytokines in sepsis, were determined to synergistically repress the expression of *Timp3* and increase the expression of *Timp1* in mouse brain microvascular endothelial cells (Bugno *et al.*, 1999). Importantly, these changes in the expression of certain MMPs and TIMPs during sepsis have shown to be diagnostic or prognostic, providing useful insights for both researchers and clinicians. For example, several studies have reported that lower levels of MMP9, higher levels of TIMP1, and a higher TIMP1/MMP9 ratio are associated with greater disease severity in septic patients (Lorente *et al.*, 2009)(Lorente *et al.*, 2014)(Bojic *et al.*, 2018). Moreover, MMP9 and TIMP1 have been associated with circulating cytokine levels, coagulation state, organ dysfunction, and mortality in sepsis (Bojic *et al.*, 2018).

Metalloproteinases and Septic Barrier Dysfunction

Septic microvascular barrier dysfunction results from injury and dysfunction of the microvascular endothelial cells leading to extravascular leakage of protein-rich edema and leukocyte influx into the tissue (Arpino *et al.*, 2016). MMPs have been shown to regulate vascular permeability and barrier function and thus have been examined for their role in the pathophysiology of sepsis. Disruption of microvascular endothelial intercellular junctions, including adherens and tight junctions, occurs during sepsis and contributes to a loss of barrier function (Jayawardena *et al.*, 2019). VE-cadherin, a critical adhesion molecule of adherens junctions, is known to be disrupted during sepsis via mechanisms including VE-cadherin phosphorylation and cleavage by proteases such as MMPs and ADAMs (Jayawardena *et al.*, 2019). Cleavage of VE-cadherin results in the release of soluble VE-cadherin (sVE-cadherin) fragments into the circulation and plasma levels of sVE-cadherin have been shown to positively correlate with sepsis severity (Zhang *et al.*, 2010). MMP7, which is increased under septic conditions, is known to be capable of cleaving VE-cadherin (Ichikawa *et al.*, 2006)(Yazdan-Ashoori *et al.*, 2011). These findings indirectly point to a potential role for MMPs in the breakdown of endothelial intercellular junctions during sepsis.

MMP1 may also contribute to increased vascular permeability during sepsis (Tressel *et al.*, 2011). Specifically, the mouse ortholog of MMP1, MMP1a, was released from endothelial cells of septic mice and cleaved protease-activated receptor-1 (PAR-1), resulting in its activation (Tressel *et al.*,

2011). PAR-1 activation leads to vascular leakage and leukocyte extravasation as a result of RhoGTPase activation and actin skeleton-dependent contraction of endothelial cells (Vandenbroucke *et al.*, 2011). The administration of an MMP1 inhibitor successfully reduced the increase in lung vascular permeability observed during sepsis, thereby suppressing endothelial barrier disruption (Tressel *et al.*, 2011). The MMP1 inhibitor also suppressed the proinflammatory cytokine response, reduced disseminated intravascular coagulation, and improved survival in septic mice (Tressel *et al.*, 2011). Further investigation into this MMP1/PAR1 axis may uncover a new therapeutic target in sepsis.

The BBB is vital for maintaining normal brain function and can be compromised during sepsis, resulting in cerebral edema and potentially sepsis-associated encephalopathy (Chaudhry and Duggal, 2014). Patients with sepsis-associated encephalopathy present with neurological dysfunction, altered mental status, and higher mortality rates (Nwafor *et al.*, 2019). MMP2 and -9 may have a role in mediating the increased vascular permeability observed in the brain during sepsis. Using a cecal ligation and perforation (CLP) preclinical model of sepsis, BBB permeability in Wistar rats increased in parallel with levels of MMP2 and -9 in the microvessels of the hippocampus and cortex (Dal-Pizzol *et al.*, 2013). This increase in permeability was successfully reversed by an MMP2 or MMP9 inhibitor (Dal-Pizzol *et al.*, 2013). Although the exact mechanism remains to be determined, MMP2 and MMP9 are both capable of cleaving tight junction proteins, which are crucial in maintaining BBB integrity and are disrupted during sepsis (Q Li *et al.*, 2009)(Feng *et al.*, 2011).

The integrity of the intestinal epithelial barrier can also become compromised during sepsis, with leakage of intestinal components resulting in enhanced systemic inflammation and organ dysfunction (Vandenbroucke *et al.*, 2013). MMP7 and -13 have both been implicated in regulating the permeability of the intestinal epithelial barrier during sepsis (Vandenbroucke *et al.*, 2013)(Vandenbroucke *et al.*, 2014). Specifically, *Mmp13*^{-/-} mice have reduced mortality in both LPS and CLP models of sepsis (Vandenbroucke *et al.*, 2013). This protective effect was partly attributed to the ability of MMP13 to proteolytically cleave and activate membrane-bound TNF into soluble TNF. Soluble TNF is bioactive and can induce disruption of intestinal epithelial tight junctions via caveolin-mediated endocytosis (Vandenbroucke *et al.*, 2013). In another study, *Mmp7*^{-/-} mice were protected against endotoxin-induced increases in intestinal permeability and translocation of bacteria to the mesenteric lymph nodes (Vandenbroucke *et al.*, 2014). Collectively, these studies provide evidence that MMPs are important in mediating intestinal barrier disruption and inhibiting them may result in protection against sepsis-induced organ dysfunction.

Metalloproteinases and the Septic Inflammatory Response

Activation and tissue infiltration of immune cells, such as neutrophils, are an important element of the host inflammatory response to sepsis (Rahman *et al.*, 2012). CD40L, a molecule expressed on the surface of platelets, appears to be released into the circulation during abdominal sepsis and subsequently promotes infiltration of neutrophils into the lung leading to increased tissue edema and damage (Rahman *et al.*, 2009). A recent role has been proposed for MMPs in regulating the shedding of CD40L during sepsis. Administration of GM6001/ilomastat (**Fig. 6**), a broad-spectrum MMP inhibitor, reduced CD40L shedding from platelets, Mac-1 upregulation on neutrophils, and production of CXC chemokines in the lung in a preclinical mouse sepsis model (Rahman *et al.*,

2012). Moreover, it led to decreased neutrophil infiltration and less lung damage. Further studies using *Mmp9*^{-/-} mice determined that MMP9 regulates the shedding of CD40L from platelets and the consequent neutrophil accumulation in the lung during abdominal sepsis (Rahman *et al.*, 2013). Previous work has shown that inhibiting neutrophil recruitment can be protective against septic pulmonary damage (Asaduzzaman *et al.*, 2008). Therefore, targeting the activity of MMP9 may prove to be beneficial in regulating the inflammatory response during sepsis.

MMP8 has also been identified as a critical regulator of the inflammatory response in sepsis. *MMP8* mRNA expression was significantly increased in pediatric patients with sepsis and septic shock vs. healthy control patients (Solan *et al.*, 2012). Moreover, the level of MMP8 appeared to correlate with sepsis severity, with increased *MMP8* mRNA expression found in septic shock non-survivors compared to survivors (Solan *et al.*, 2012). In a preclinical CLP model of sepsis, *Mmp8*^{-/-} mice had reduced early infiltration of neutrophils into the lung and reduced circulating levels of proinflammatory cytokines IL6 and IL1 β (Solan *et al.*, 2012). Furthermore, pharmacological inhibition of MMP8 activity by treating septic mice with (3R)-(+)-[2-(4-methoxybenzenesulfonyl)-1,2,3,4-tetrahydroisoquinolone-3-hydroxamate] significantly reduced mortality (Solan *et al.*, 2012) (**Fig. 6**, MMP8 inhibitor). In the same study, MMP8 was found to directly activate NF- κ B, an important proinflammatory transcription factor, in macrophages *in vitro* pointing to a potential mechanism to be further explored (Solan *et al.*, 2012). Collectively, these 4 lines of evidence suggest that MMP8 plays an important role in modulating the inflammatory response during sepsis and that inhibiting MMP8 activity may be a potential therapeutic strategy.

As TIMPs regulate the activity of MMPs, it is not surprising that they can impact the host response to sepsis. For example, levels of TIMP3 have been shown to be decreased under septic conditions. (Arpino *et al.*, 2016). In a CLP preclinical model of sepsis, researchers examined the impact of a TIMP3 null mutation on lung structure and function (Martin *et al.*, 2003). *Timp3*^{-/-} septic mice showed an increase in lung compliance accompanied by increased MMP2 and MMP9 activity, as well as decreased levels of collagen and fibronectin (Martin *et al.*, 2003). It was proposed that the observed septic lung abnormalities were enhanced by increased MMP degradation of ECM components. Alveolar macrophages, a major inflammatory cell population in the lung, were determined to be critical for mediating the lung changes in *Timp3*^{-/-} septic mice (Martin *et al.*, 2007). *Timp3*^{-/-} mice depleted of alveolar macrophages were protected from the sepsis-induced alterations in lung mechanics (Martin *et al.*, 2007). Moreover, MMP7 abundance was attenuated after macrophage depletion, suggesting it may be partially involved in this process (Martin *et al.*, 2007). Overall, these studies demonstrate how changes in TIMP levels during sepsis can alter the MMP/TIMP balance and thus impact the net metalloproteinase activity.

V. Pharmacological Inhibitors of Matrix Metalloproteinases

A. Clinical Trials for Cancer Treatment

The first generation of MMP inhibitors was inspired by mechanistic aspects from peptide hydrolysis. The mechanism by which all MMPs cleave substrates is driven by acid-base catalysis via a Zn²⁺ ion in the active site (Nagase and Woessner, 1999)(Visse and Nagase, 2003)(Maskos, 2005). In the 1990s, MMP inhibitors were designed based on the logical, but at the time, unproven

concept that cancer cells require MMPs to degrade ECM (predominantly collagens) to facilitate cell migration, invasion, and metastasis. The first generation of MMP inhibitors were peptide and peptide-like compounds mimicking backbone features of P1, P1' and P2' and capable of binding Zn^{2+} ion that resembled collagen (Zucker *et al.*, 2000). The goal was to develop competitive and potent reversible broad-spectrum inhibitors of MMP activity, and the first class of compounds was the hydroxamic acid zinc-binding group (Zucker *et al.*, 2000). The first compound tested in preclinical rodent cancer models and entered clinical trial was batimastat or BB-94 (developed by British Biotech, **Fig. 6**), which display broad specificity in inhibiting multiple MMPs while sparing other proteases. Batimastat was tested on lung colonization and spontaneous metastasis of the HOSP.1P rat mammary carcinoma. Intraperitoneal administration of 6 doses of 30 mg/kg batimastat inhibited 80% of lung metastases (Eccles *et al.*, 1996). When batimastat was initiated two days prior to surgical removal of the tumors, 100% of treated animals survived to 120 days, whereas the control animals all died before day 100 (Eccles *et al.*, 1996). In a mouse model of ovarian cancer, the combination of batimastat and cisplatin resulted in delayed tumor growth and increased the survival time of the mice as compared to cisplatin alone (Giavazzi *et al.*, 1998). In the RIP1-Tag2 model of pancreatic islet cell carcinogenesis, batimastat reduced angiogenesis by 49% in a prevention trial and by 83% in an intervention trial with no effect on invasive tumors (Bergers, 1999). Preclinical studies using batimastat for cancer treatment were successful, resulting in human phase I and II clinical trials. However, as batimastat was not soluble and had poor bioavailability, it had to be administered intraperitoneally and intrapleurally to patients, which resulted in sustained plasma concentrations and was found to act as a depot (Nelson *et al.*, 2000). The reported side effects included nausea, fatigue, low-grade fevers, abdominal pain when injected intraperitoneally, and pain at the injection site when injected intrapleurally (Nelson *et al.*, 2000). Thus, clinical trials using batimastat were terminated, and other MMP inhibitors were further tested (Zucker *et al.*, 2000)(Overall and Kleinfeld, 2006b)(Dufour and Overall, 2013).

Marimastat (BB-2516, **Fig. 6**) was developed after batimastat in an attempt to make the drug more bioavailable and was also a broad-spectrum MMP inhibitor (Nelson *et al.*, 2000). Phase I and II trials in patients with advanced cancers demonstrated that marimastat was well-tolerated and side effects included fatigue and cumulative inflammatory polyarthritis that was reversible when treatments were stopped (Wojtowicz-Praga *et al.*, 1998)(Nelson *et al.*, 2000)(Zucker *et al.*, 2000).

Prinomastat (AG3340, **Fig. 6**) was developed by Agouron Pharmaceuticals based on information obtained with X-ray crystallographic analysis of human MMPs and is a picomolar inhibitor of MMPs. Prinomastat was developed to be more specific than previous inhibitors and was aimed to selectively inhibit MMP2, -9, -13, and MT1-MMP (Nelson *et al.*, 2000). In a preclinical model of brain cancer, U87 glioma cells were implanted in SCID-NOD mice for three weeks before being treated with 100 mg/kg of prinomastat or vehicle intraperitoneally (Price *et al.*, 1999). In the prinomastat-treated mice, tumor volume was decreased by 78% compared to the vehicle-treated mice, resulting in less invasive and proliferative tumors and a two-fold increase in survival time (Price *et al.*, 1999). In an NCI-H460 orthotopic lung cancer model, treatment with prinomastat in combination with carboplatin resulted in longer survival compared to vehicle-treated animal, prinomastat, or carboplatin alone (Liu, 2003). When prinomastat was tested in clinical trials, similar side effects to previous MMP inhibitors were identified, such as time- and dose-dependent musculoskeletal stiffness and pain (Nelson *et al.*, 2000). Although there were benefits of combination therapy in preclinical animal models, importantly, a human phase III clinical trial for

advanced stage non-small cell lung cancer that compared standard chemotherapy with or without the addition of prinomastat did not result in increased survival between the two groups (Bissett *et al.*, 2005).

Tanomastat (BAY 12-9566, **Fig. 6**) was developed by Bayer Corporation and is a non-peptidic biphenyl inhibitor with a Zn²⁺-binding carbonyl group and was demonstrated to have a long terminal plasma half-life ranging from 90-100 hours (Gatto *et al.*, 1999). Tanomastat is a butanoic acid derivative that could account for distinct selectivity as it cannot inhibit MMP1, -8, and -13 (Nelson *et al.*, 2000). Tanomastat's distinct chemical features could be responsible for different side effects as other MMP inhibitors; there was no musculoskeletal toxicity in the phase I trial in patients with advanced tumors, but new symptoms were identified, including an asymptomatic increase of hepatic enzymes and thrombocytopenia (Nelson *et al.*, 2000). Importantly, patients with metastatic small lung cancer taking tanomastat resulted in a worse disease, suggesting that we lacked a clear understanding of specific MMP functions in cancer (Overall and Kleifeld, 2006a)(Dufour and Overall, 2013). MMP inhibitors were also tested in combination with known cancer drugs such as gemcitabine, carboplatin, or paclitaxel, but no additional benefit was clearly demonstrated (Nelson *et al.*, 2000). The failure of MMP inhibitors in randomized phase III clinical trials has generated numerous updated generations of different inhibitors to inhibit directly or indirectly MMPs (Das *et al.*, 2021).

B. Clinical Trials for Inflammatory Diseases Treatment

MMP inhibitors were also tested in various rodent models of inflammation. For example, in a model of autoimmune inflammatory arthritis in rats using complete Freund's adjuvant, injection of the compound N4-hydroxy-2-(2-methylpropyl)-N1-[2-[[2-(morpholinyl)ethyl]-,[S- (R*,S*)]] or GI168 via osmotic minipumps from days 8 to 21 significantly diminished ankle swelling in addition to bone and cartilage destruction (Conway *et al.*, 1995). In another study of adjuvant-induced arthritis, rats were treated for 21 days with 50 mg/kg of tanomastat (BAY 12-9566, **Fig. 6**) or vehicle (Hamada *et al.*, 2000). The severity of arthritis in this model was determined by measuring an arthritic index indicated by paw volume, urinary pyridinoline, and deoxypyridinoline; by examining joint inflammation; and by microscopic morphometry of articular cartilages. In this adjuvant-induced arthritis model, tanomastat was demonstrated to suppress inflammation and reduced cartilage destruction (Hamada *et al.*, 2000).

Multiple MMPs have been implicated in cardiovascular diseases having both beneficial and detrimental roles (Bassiouni *et al.*, 2021). For example, myocardial specific overexpression of constitutively active murine MMP2 resulted in cardiac defects, including increased troponin I proteolysis, mitochondrial dysregulation, left ventricular remodeling and heart failure even in the absence of an external stimulation or injury (Wang *et al.*, 2006)(Bergman *et al.*, 2007). Using a selective MMP12 phosphinic peptide inhibitor RXP470.1 (**Fig. 6**), atherosclerotic plaques cross-sectional area was diminished by over 50% in both male and females apolipoprotein E knockout mice fed on a Western diet (Johnson *et al.*, 2011). In humans, when batimastat was coated onto drug-eluting stents for the treatment of obstructive coronary artery disease, no clinical benefits were demonstrated, suggesting that a better understanding of the role of individual MMP is required (Sousa *et al.*, 2003).

In the EAE mouse model of MS, ilomastat (GM6001, **Fig. 6**) was demonstrated to suppress the development of EAE in a dose-dependent manner (Gijbels *et al.*, 1994). Another MMP inhibitor, D-penicillamine, significantly reduced the mortality and morbidity rates in the EAE mouse model (Norga *et al.*, 1995). However, when tested in humans in a double-blind, placebo-controlled pilot trial with D-penicillamine and metacycline, side effects were detected, and the trial was stopped (Dubois *et al.*, 1998). Next, minocycline, a tetracycline derivative containing some MMP9-inhibitory activity, was tested and was demonstrated to be highly effective in the EAE model (Popovic *et al.*, 2002). In a human randomized and controlled trial, 100 mg of minocycline taken twice a day was tested to see if it was able to reduce the risk of conversion from a first demyelinating event (also termed clinically isolated syndrome) to MS. In 142 patients with multiple sclerosis, it was demonstrated that minocycline treatment significantly reduced the risk of conversion from a clinically isolated syndrome to MS compared to the placebo groups (Metz *et al.*, 2017). All changes detected on magnetic resonance imaging (MRI) were significant at 6 months but not at 24 months (Metz *et al.*, 2017). Some of the side effects reported in the minocycline treated group included rash, dizziness, and dental discoloration (Metz *et al.*, 2017). It is important to mention that minocycline is not a selective MMP9 inhibitor but rather results in downregulation of MMP9 activity and likely additional MMPs.

C. Potential Reasons for the Failure of MMP Inhibitors

Over the past two decades and since the various clinical trials of MMP inhibitors, multiple lessons have been learned: 1) MMPs can be both drug targets and anti-targets depending on cell/tissue localization, type of disease, or stage of the disease; 2) MMP inhibitors are likely to be more effective for short-term dosage as opposed as long term dosage; and 3) Broad-spectrum versus selective inhibition must be carefully examined for a particular disease (reviewed in (Overall and Kleifeld, 2006b)(Dufour and Overall, 2013)(Young *et al.*, 2019). It has been demonstrated that MMPs have biological functions other than ECM remodeling: ~31% of their substrates are ECM proteins and ~69% are non-ECM proteins (**Table 1, Supplementary Table 1, Fig. 4**). Therefore, it is likely that the role MMPs play in diseases is different than what the inhibitors were intended to block during the cancer clinical trials (Dufour and Overall, 2015). Most MMP inhibitors tested were broad-spectrum and able to inhibit multiple MMPs, in addition to ADAMs and ADAMTSs for some inhibitors. Paradoxically, treating human HT-1080 fibrosarcoma cells with the broad-spectrum MMP inhibitor GI129471 increased MMP9 expression (Maquoi *et al.*, 2002). Additionally, MMP inhibitors can prevent autolytic degradation of MT1-MMP, resulting in an enhancement of proMMP2 activation and MMP2 activity (Toth *et al.*, 2000). Overall, toxicities associated with MMP inhibitors were related to musculoskeletal pain and were assumed to be due to their poor selectivity; however, this has not been fully confirmed yet (Zucker *et al.*, 2000)(Skiles *et al.*, 2004)(Fingleton, 2008)(Devy and Dransfield, 2011). Therefore, it is believed that more selective inhibitors would likely be more effective. In the past decade, several small synthetic inhibitors selectively targeting the MMP active site have been designed with some significant success for MMP2, -9, -12, and -13 (Devel *et al.*, 2010)(Mahasenan *et al.*, 2017)(Fields, 2019).

Specifically, a significant gain in selectivity was achieved by exploring zinc-binding groups than the hydroxamate function, as well as other side chains interacting within the MMP specificity loop (S₁' cavity). For example, MMP408 with a carboxylate function and an optimized P₁' side to fit within a deep S₁' cavity (**Fig. 6**, (W Li *et al.*, 2009)) has been reported as a selective inhibitor of

MMP12. In the same vein, the phosphinic pseudo peptide that displayed a high potency and selectivity for this protease (RXP470.1, **Fig. 6**, (Devel *et al.*, 2006)). In this case, it was demonstrated that the excellent selectivity of this inhibitor mainly relied on three critical features: the presence of two glutamate residues facing the MMPs S₂' and S₃' subsites (Devel *et al.*, 2006), a long and hydrophobic P₁' side-chain inserting within the large S₁' cavity of MMP12 (Czarny *et al.*, 2013), and the chemical nature of the zinc-binding moiety. On this last point, the RXP470.1 analogs with either a hydroxamate or a carboxylate function to chelate the catalytic zinc ion are much less selective for MMP12 than the parent molecule with a phosphinate moiety, highlighting the key role played by the zinc-binding group in subtly tuning inhibitor selectivity (Rouanet-Mehouas *et al.*, 2017). The phosphinate function was also incorporated within the structure of triple-helical peptide mimicking a specific region of collagen (Lauer-Fields *et al.*, 2007), thus resulting in inhibitors displaying K_i values in the low nanomolar range for MMP2 and -9. In those two examples, the high potency of phosphinic pseudo peptides was attributed to the capacity of the phosphinate function to reproduce the transition state during peptide bond hydrolysis, making those compounds potential transition state analogs (Georgiadis and Dive, 2014). In the field of small synthetic compounds, selectivity for a given MMP or a restricted set of MMPs can also be obtained by selectively targeting the S₁' cavity of MMPs. In this respect, two compounds with no zinc-binding moiety were able to target MMP8/13 ((Pochetti *et al.*, 2009), **Fig. 6**) and MMP13 ((Engel *et al.*, 2005), **Fig. 6**) with high potency and selectivity. In these cases, the inhibitors bind deeply in the S₁' cavity and extend into an additional side pocket, only present in MMP8 and -13.

Many of these inhibitors have been assessed in preclinical models of inflammatory diseases and cancer, but none have been validated in human studies to date. There is also a need to better characterize the role of individual MMP in a specific disease to establish if a specific MMP is detrimental; currently, at least 10 MMPs have been found to have protective functions and should not be inhibited (Overall and Kleifeld, 2006b)(Dufour and Overall, 2013)(Brkic *et al.*, 2015)(Chopra *et al.*, 2019).

The broad expression of multiple MMPs in various cell types (e.g., epithelial, cancer, immune, fibroblast, and others) could also imply that a local/targeted delivery is likely necessary. Also, the timing of drug delivery is important. MMPs have been shown to have opposite roles in different stages of viral infections. For example, early on, nuclear MMP12 was shown to be protective via its promotion of IFN α but was demonstrated to generate a negative feedback loop by cleaving extracellular IFN α at later time points (Marchant *et al.*, 2014). Therefore, treatment with a selective MMP12 inhibitor could result in unintended effects depending on the timing of drug treatment. The clinical use of MMP inhibitors has been more complex than initially thought. Alternatives other than inhibitors that target MMP active sites exist to inhibit and control the biological functions of MMPs. These options are now being examined in various preclinical models.

D. Alternative Strategies to inhibit MMPs

The catalytic domains of MMPs are highly conserved, whereas other domains are not present in all MMPs, or they are divergent in terms of amino acid similarities. For example, targeting MMPs exosites, the hemopexin domain, or cytoplasmic tail could result in more selective compounds. Peptides mimicking the hemopexin domains of MMPs have been designed for MMP9 to inhibit

cell migration (Dufour *et al.*, 2010), MMP12 as antibacterial drugs (Houghton *et al.*, 2009), and MT1-MMP to inhibit cell migration and invasion (Zarrabi *et al.*, 2011). Small molecule inhibitors targeting the hemopexin domain of MMP9 (Antoine Dufour *et al.*, 2011) and MT1-MMP (Remacle *et al.*, 2012) were designed, and both demonstrated anti-tumor growth activity in murine models of cancer. However, the binding affinity of exosite domains is typically much lower than the catalytic domain; therefore, it creates a new set of challenges for these types of inhibitors (Overall and Kleifeld, 2006a)(Dufour and Overall, 2013)(Young *et al.*, 2019).

Other than peptidomimetic or small molecules inhibitors, monoclonal antibodies (mAbs) have been designed with the purpose of selectively inhibit a single MMP. For example, Irit Sagi and collaborators injected a Zn-tripod to elicit an immune response against a synthetic zinc-imidazole motif, resulting in the production of mAbs that recognized MMP9 (Sela-Passwell *et al.*, 2012). Two of them generated mAbs, SDS3 and SDS4, which inhibited MMP2 and -9 (K_i values was $1 \pm 0.01 \mu\text{M}$ for SDS3 and $0.05 \pm 0.006 \mu\text{M}$ for SDS4) (Sela-Passwell *et al.*, 2012). In a murine model of dextran sulfate sodium-induced colitis, both prophylactic and therapeutic treatment with SDS3 prevented body weight loss, prevented colon shortening, decreased diarrhea, and improved colonic damage (Sela-Passwell *et al.*, 2012). Using the Zn-tripod approach, an MMP7 selective antibody (GSM-192) was recently developed and was demonstrated to induce apoptosis and reduce CFPAC-1 cell migration *in vitro* (Mohan *et al.*, 2021). Using a different approach to generate an antibody-based inhibitor against MT1-MMP called LEM-2/15, mice were immunized using a cyclic peptide of the V-B loop of MT1-MMP comprising amino acid residues 218 to 233 (Udi *et al.*, 2015). The MT1-MMP antibody LEM-2/15 was demonstrated to protect against influenza virus infection and, in combination therapy with Oseltamivir (Tamiflu), displayed a synergistic effect and resulted in complete recovery of influenza-infected mice (Talmi-Frank *et al.*, 2016). Another selective MT1-MMP inhibitor, called DX-2400, was developed using a human Fab displaying phage library (Devy *et al.*, 2009). Using a breast cancer xenograft model with MDA-MD-231 cells, DX-2400 was demonstrated to reduce tumor growth, invasion, and metastasis (Devy *et al.*, 2009). In the intravenous injection of the metastatic model using mouse B16F1 melanoma cells, DX-2400 was shown to diminish the number of lung nodules and reduce the metastasis (Devy *et al.*, 2009). Additionally, DX-2400 was demonstrated to reduce tumor growth in the 4T1 and E0771 syngeneic BALB/c mouse model, reducing TGF β and SMAD2/3 signaling (Ager *et al.*, 2015). Currently, BT1718, a hemopexin-binding MT1-MMP selective inhibitor developed by Bicycle Therapeutics containing a bicyclic peptide, a linker, and a toxin moiety, is in clinical trials for the treatment of solid tumors, projected to end in 2022 (<https://clinicaltrials.gov/ct2/show/NCT03486730>). BT1718 was demonstrated to be effective in reducing tumor volume in xenograft mouse models using HT-1080 cells (Harrison *et al.*, 2017).

Recent and ongoing MMP clinical trials include the testing of Andecaliximab (GS-5745), an MMP9 humanized monoclonal antibody designed for the treatment of cancer and inflammatory diseases by Gilead Sciences (Marshall *et al.*, 2015). In a Phase III study, Andecaliximab in combination with modified oxaliplatin, leucovorin, and fluorouracil (mFOLFOX6) were recently tested in patients with gastric or recurrent gastroesophageal junction (GEJ) adenocarcinoma ([ClinicalTrials.gov Identifier: NCT01803282](https://clinicaltrials.gov/ct2/show/NCT01803282)); however, the addition of Andecaliximab to mFOLFOX6 did not improve overall survival of patients (Shah *et al.*, 2021). Importantly, there was no associated toxicity in patients that took Andecaliximab (Shah *et al.*, 2018). Andecaliximab has also been evaluated in combination with gemcitabine and nab-paclitaxel in patients with

advanced pancreatic adenocarcinoma and was well-tolerated (Bendell *et al.*, 2020). There is also an on-going recruitment of patients for the evaluation of Andecaliximab as a treatment for glioblastoma ([ClinicalTrials.gov Identifier: NCT03631836](#)).

Andecaliximab has also been tested in inflammatory diseases such as rheumatoid arthritis ([ClinicalTrials.gov identifier: NCT02176876](#)), UC ([ClinicalTrials.gov Identifier: NCT02520284](#)), CD ([ClinicalTrials.gov Identifier: NCT02405442](#)) and chronic obstructive pulmonary disease ([ClinicalTrials.gov Identifier: NCT02077465](#)). In RA patients, Andecaliximab was administered as three infusions over 29 days and was generally safe and well-tolerated (Gossage *et al.*, 2018). However, only 15 patients were given Andecaliximab and three were given the placebo; therefore, studies in larger patient cohorts and also of increased treatment duration are needed to better characterize the efficacy of Andecaliximab (Gossage *et al.*, 2018) ([ClinicalTrials.gov Identifier: NCT02176876](#)).

In a phase II/III study in moderately to severely active UC patients, subjects were randomized [1:1:1] to receive placebo, 150 mg Andecaliximab every 2 weeks, or 150 mg andecaliximab weekly, via subcutaneous administration (Sandborn *et al.*, 2018). Although Andecaliximab was well-tolerated, eight weeks of treatment with 150 mg Andecaliximab did not induce clinical remission or response in UC patients and the trial was terminated early (Sandborn *et al.*, 2018) ([ClinicalTrials.gov Identifier: NCT02520284](#)).

In a phase II study in moderately to severely active CD patients, subjects were randomized [1:2:2:2] to receive subcutaneous injections of placebo weekly, Andecaliximab 150 mg every 2 weeks, Andecaliximab 150 mg weekly, or Andecaliximab 300 mg weekly (Schreiber *et al.*, 2018). Similar to the UC patients, Andecaliximab was well-tolerated in the CD patients, but no clinically meaningful symptomatic or endoscopic response was observed, and the trial was terminated due to lack of efficacy (Schreiber *et al.*, 2018) ([ClinicalTrials.gov Identifier: NCT02405442](#)).

Another interesting strategy is to use the biological functions of MMPs as a (Gabrielli *et al.*, 2009). For example, the combination of FCX-013 and Veledimex is currently being used in clinical trials for the treatment of localized scleroderma, a disease that hardens the skin and connective tissues (Gabrielli *et al.*, 2009) ([ClinicalTrials.gov identifier #NCT03740724](#)). This approach uses FCX-013 is a human fibroblast genetically modified using a lentivirus vector encoding for MMP1, whose expression is dependent on an agonist. This therapy consists of injecting FCX-013 subcutaneously at the location of the fibrotic lesions. The goal is that the MMP1 producing fibroblasts will break down excess collagen accumulation. In combination with FCX-013, patients will also orally take Veledimex (INXN-1001) to induce MMP1 from the injected cells. Veledimex (INXN-1001) is an oral activator and promoter for the gene therapy system (Chiocca *et al.*, 2019). Once the fibrosis is resolved, the patient stops taking Veledimex (INXN-1001), which stops MMP1 production. Therefore, the biological activity of certain MMPs could also potentially be used as a therapeutic to resolve fibrosis in specific diseases (Leong *et al.*, 2021). This approach further supports that a better characterization of the roles of individual MMPs in inflammation and fibrosis could result in the development of novel therapeutics.

VI. Molecular tools for targeting active MMPs

Conventional proteomics approaches aiming at establishing the relation of MMPs to disease states are limited by the fact that they take the total protein amount into account, whereas the protease functional (active) status is often the unique relevant parameter. To determine MMPs proteolytic activity in biological samples, two main molecular tools have been developed: 1) substrate-based probes that generate a signal upon proteolytic cleavage and 2) ligand-based probes that bind selectively to MMPs active form and spare inactive and inactivated ones. Within the family of ligand-derived tools, two classes can be further distinguished: 1) MMP inhibitors containing a reporter tag for imaging purposes and 2) affinity-based probes with a reactive motif enabling a permanent attachment of the probe to the target MMP for subsequent analyses. In this section, we will discuss the most significant advancements in those three categories of molecular tools with a special focus on covalent probes, the sole class of molecular probes capable of identifying directly and unambiguously the target MMPs through diverse analytical methods.

A. Substrate-based probes for a cleavage assay

The design and development of substrate-based probes have been inspired directly from the peptide sequence of natural MMP substrates or from combinatorial libraries of peptides obtained by phage display or chemical approaches (Knapinska and Fields, 2012). In this respect, substrate-based probes generally consist of a minimum peptide sequence containing six amino acids that extend their side chains from S_3 to S_3' subsites of MMP catalytic cleft (**Fig. 8**). Within most of the sequences, a proline residue is present in the P_3' position, thus conferring an MMP preference for peptide sequence, and variations of other side chains result in motifs targeting a specific subset of MMPs. A certain selectivity can also be achieved by incorporating additional amino acids on both sides of the minimal P_3 - P_3' sequence. By relying on such a scaffold, several types of biosensors have been developed to detect MMP activity (**Fig. 8**). This mainly includes 1) optical biosensors emitting an optical signal upon proteolytic cleavage, 2) magnetic or photoacoustic biosensors, or 3) sensors leading to the secretion of synthetic biomarkers specifically detected in urine.

Substrates-based probes emitting a fluorescent signal upon proteolytic cleavage are the most widely used optical biosensors and enable the real-time monitoring of MMP proteolytic activity in many different complex proteomes and *in vivo* (Scherer *et al.*, 2008)(Knapinska and Fields, 2012)(Hu *et al.*, 2014)(Lei *et al.*, 2020). The fluorescence-based biosensors have been designed by relying on fluorescence self-quenching (Bremer *et al.*, 2001), Förster Resonance Energy Transfer (FRET) (Knapinska and Fields, 2012)(Hu *et al.*, 2014), or Bioluminescence Resonance Energy Transfer (BRET) (Lee and Kim, 2015)(Nguyen *et al.*, 2018)(Tian *et al.*, 2021)(Park *et al.*, 2021). Benefiting from the enzyme turnover, those activatable probes detect active MMPs with high sensitivity and, if the fluorophore possesses an emission wavelength in the near-infrared (NIR) region, this detection can be achieved *in vivo* within deep tissues. For example, the fluorescence self-quenched probe *MMPsense680* with an organic NIR-Dye has been used to monitor the activity of MMP2 in tumor tissue (Bremer *et al.*, 2001) and that of MMP2 and -9 in atherosclerosis (Deguchi *et al.*, 2006). Since these seminal reports, this probe has been used in several other preclinical models including colon cancer (Hensley *et al.*, 2017), colitis (Ding *et al.*, 2014), and bacterial infections (Daghighi *et al.*, 2014).

Another example is a NIR probe developed by Tang *et al.* whose fluorescence signal was self-quenched upon self-assembly into nanoparticles, which enabled the detection of MMP2 activity *in*

in vivo (Tang *et al.*, 2018). Alternatively, several FRET-based activatable probes have been designed and became the standard in the real-time monitoring of MMPs activity in various biological matrices and *in vivo*. In this case, the peptide sequence is flanked by both a fluorescence donor and a fluorescence acceptor/quencher in close proximity. Upon proteolytic cleavage, the distance between the two partners increases, and this irreversible spatial separation induces a ratiometric change in the donor and acceptor emission spectra, resulting in the appearance of a fluorescent signal. Many different types of FRET-base probes incorporating either small organic dyes or nanomaterials as FRET donor/acceptors were designed to target MMPs in complex proteomes (see several reviews for additional information: (Garland *et al.*, 2016)(Liu *et al.*, 2018)(Oliveira-Silva *et al.*, 2020)(Soleimany and Bhatia, 2020)(Lei *et al.*, 2020). Despite their high sensitivity and low background to noise *in vivo*, the limitation of FRET-based probes with organic fluorescent dyes used for long-time imaging is a signal attenuation due to diffusion within the tissues. However, to circumvent these limitations, ultrafast-acting activatable probes enabling the visualization of MMPs expression and inhibition after only 30 minutes have been designed and validated in a tumor-bearing mouse model (Zhu *et al.*, 2011). In this case, the addition of different PolyEthylene Glycol extensions at the C-terminal end of the peptide sequence subtly modulated the activation profiles of probes *in vivo*. To limit the diffusion of cleaved fragments tethered to fluorescent emitter away from the protease active site, other effective strategies based on activatable FRET probes connected to cell-penetrating peptides have been developed (Jiang *et al.*, 2004). Specifically, an MMP-cleavable peptide sequence was inserted between complementary polycationic and polyanionic domains. Upon processing by MMP2 or -9, the polycationic entity as a cell-penetrating peptide (CPP) connected to a NIR-Dye can be released to further bind and enter the cell. Such a selective delivery of fluorescent emitter to MMP-expressing cells by Activatable Cell-Penetrating Peptides (ACPPs) has been explored in different mouse models (Hingorani *et al.*, 2017)(Chen *et al.*, 2017), notably to detect unstable atherosclerotic plaques within which certain MMPs are overexpressed under their active form (Hua *et al.*, 2015). Interestingly, the ACPP strategy was combined with the MRI technique for identifying plaques at high risk of rupture. The ACPPs has also demonstrated excellent ability to direct fluorescence-guided oncologic surgery in a preclinical model (Metildi *et al.*, 2015) and have been recently assessed in First-In-Human (Phase I) clinical trials to challenge their capacity to improve intraoperative detection of malignant tissue during breast cancer operations (Unkart *et al.*, 2017). Following an approach similar to ACPPs, Schultz *et al.* developed an MMP12 selective FRET probe containing a palmitic acid extension installed at the N-terminus end of the peptide sequence (Cobos-Correa *et al.*, 2009). Such a lipid tail allowed the fragment containing the NIR-dye to anchor onto cell membranes, therefore limiting the diffusion of the fluorescence signal. The use of FRET probes of nanometer size also limits the diffusion fluorescence signal away from the protease active site (Lei *et al.*, 2020). These FRET nanoprobe, exploiting the unique optical properties of nanomaterials, present several other advantages, including good biocompatibility and stability, high brightness with limited photobleaching, and the possibility to perform multiplexing. In this respect and taking advantage of dopant-dependent multicolor emission of upconversion nanoparticles, FRET nanoprobe with distinct emission wavelengths were designed to measure simultaneously and independently the activity of MMP2 and -7 (Cao *et al.*, 2018).

Due to the extremely low background and high sensitivity of bioluminescent reporters, BRET probes also provide great advantages for imaging MMP activity *in vivo* (Nguyen *et al.*, 2018)(Tian *et al.*, 2021)(Park *et al.*, 2021). In this case (and unlike FRET approaches), no external light is

required, but an enzyme, often luciferase, as a bioluminescent reporter is essential to emit light upon the appropriate trigger. When the emission spectrum of the bioluminescent reporter overlaps with the excitation spectrum of another fluorophore, the energy transfer results in the emission of a luminescent signal that can be associated with a specific proteolytic activity. Based on this principle, a BRET probe enabling the dynamic monitoring of MT1-MMP activity with high temporal and spatial resolution was validated in a preclinical model of lung cancer (Tian *et al.*, 2021). In this case, a membrane-bound form of luciferase was used, and the MT1-MMP recognition peptide was inserted within the sequence of this enzyme. Upon proteolytic cleavage by MT1-MMP, two luciferase subdomains get in close proximity, which results in the appearance of a high luminescent signal.

As biological matrices have a negligible magnetic background, several magnetic biosensors for the sensitive detection of MMPs in clinical samples and *in vivo* have also been developed (Lei *et al.*, 2020). Most of these biosensors exploit the properties of superparamagnetic nanoparticles capable of magnifying the magnetic signal resonance of protons in neighboring water molecules. Upon cleavage by MMPs, those sensors aggregate, which accelerates the dephasing of water protons. This change in nanoparticles state can then be detected by monitoring the variation of proton relaxation time, which is associated with MMP proteolytic activity (Harris *et al.*, 2006)(Schuerle *et al.*, 2016). Recently and in the same vein of FRET biosensors, a sensor relying on a distance-dependent magnetic resonance tuning (MRET) has been designed; the variation of distance between a paramagnetic enhancer and a superparamagnetic quencher induced the appearance of magnetic resonance imaging signal, linked to the proteolytic activity of MMP2 *in vivo* (Choi *et al.*, 2017).

Optical-based sensors can also be conjugated with other techniques such as photoacoustic imaging, which mainly aims at improving imaging resolution. The development of an activatable probe incorporating both a near-infrared dye and a quencher linked through an MMP2 peptide substrate has been done (Yin *et al.*, 2019). Upon cleavage, this probe not only emitted a fluorescence signal but also changed its aggregation state. In this respect, the responsive probe presented a dramatic MMP2-concentration-dependent absorption at around 680 nm, while that at around 730 nm was MMP2 concentration-independent. These features allowed detection of MMP2 activity via both fluorescence and photoacoustic imaging *in vitro* and in a model of breast cancer *in vivo*.

To detect MMPs activity *in vivo*, another effective approach involving nanosensors capable of generating synthetic biomarkers upon proteolytic cleavage has been reported (Soleimany and Bhatia, 2020). In this case, and unlike *in situ* imaging sensors, the peptide fragments resulting from the proteolytic cleavage diffuse away from the protease site, enter circulation, and are then secreted in the urine. Thus, the subsequent detection of those fragments by various analytical techniques such as mass spectrometry (Kwong *et al.*, 2013), ELISA (Dudani *et al.*, 2016), paper lateral flow assay (Warren *et al.*, 2014), or colorimetric test (Loynachan *et al.*, 2019) is indirectly associated to MMPs proteolytic activity at a pathologic site. Recently, those activity-based nanosensors enabled the profiling of 15 proteases, including five MMPs simultaneously, in a preclinical model of lung cancer (Kirkpatrick *et al.*, 2020).

In all those substrate-based strategies, and mainly due to the highly conserved structural topology between the different MMPs, the peptide sequence is rarely cleaved by a single MMP. To address

this issue, an activatable fluorogenic probe with enhanced specificity for MT1-MMP was recently developed (Ji *et al.*, 2020). In this case, the specificity of the FRET-Probes is increased by tethering the MT1-MMP peptide sequence to an additional recognition element that binds away from the catalytic cleft. A long linker of twelve PolyEthylene Glycol subunits was introduced between these two motifs, resulting in a significant optimization of specificity and selectivity of the fluorogenic probe for MT1-MMP *in vivo*. Beside their potential lack of selectivity within the MMPs family, the ability of substrate-based probes to target MMPs exclusively within the pathological site is also questionable (Lebel and Lepage, 2014). Indeed, in many pathological contexts, circulating active MMPs have been reported (Roy *et al.*, 2009)(Fontana *et al.*, 2012)(Hadler-Olsen *et al.*, 2013)(Goncalves *et al.*, 2015)(Peeters *et al.*, 2017)(Dofara *et al.*, 2020). This may result in substrate-based probes that are partly cleaved in the bloodstream, yielding poor target/non-target contrast when imaging diseased tissues. Preliminary work in the context of lung cancer detection suggests that a more local delivery of activatable probes, intrapulmonary delivery in this specific case, significantly increases the signal specificity while eliminating off-target activation (Kirkpatrick *et al.*, 2020).

B. Ligand-based probes to bind to active MMP

By relying on MMP inhibitors scaffold (see also the section on MMP inhibitors), numerous imaging agents have been designed for different types of imaging modalities, including positron emission tomography (PET), single photon-emission computed tomography (SPECT), magnetic resonance imaging, and optical imaging (Matusiak *et al.*, 2013)(Rangasamy *et al.*, 2019). Two types of imaging probes can be distinguished: those possessing an imaging reporter of small size (e.g., a radioisotope, such as ^{18}F , ^{11}C , or ^{123}I) that is directly inserted within the structure of the MMP inhibitors, and those incorporating a reporter of comparable size to that of the MMP inhibitor scaffold (e.g., NIR Dye, encaged ^{68}Ga or $^{99\text{m}}\text{Tc}$). In this latter case, the imaging reporter is systematically separated from the MMP binding motif by a spacer to limit eventual steric clashes (Faust *et al.*, 2008)(Razavian *et al.*, 2010)(Bordenave *et al.*, 2016)(Razavian *et al.*, 2016)(Hugenberg *et al.*, 2017)(Toczek *et al.*, 2019)(Gona *et al.*, 2020). The imaging probes contain a binding motif targeting either a wide range of MMPs or a restrictive set of MMPs. Within this second class of probes, MMP2/9 probes with a thiirane scaffold (Hohn *et al.*, 2018), MMP12 probes with either a phosphinic pseudo peptide (Bordenave *et al.*, 2016)(Razavian *et al.*, 2016)(Toczek *et al.*, 2019) or a hydroxamate-based scaffold (Gona *et al.*, 2020), and MMP13 probes have been reported (Hugenberg *et al.*, 2017) (**Fig. 8, 9** for the structure of the motif targeting selectively the MMP active site).

All these MMP inhibitor-derived imaging agents have been assessed in a wide variety of preclinical models, including cancer, atherosclerosis, myocardial infarction, aneurysm, RA, and lung inflammation and, for a rather complete overview of their scope of application (*see this review* (Rangasamy *et al.*, 2019)). In all those preclinical models, the probes accumulate within pathological sites overexpressing active MMPs, and the spatiotemporal resolution, penetration depth, as well as sensitivity are related to the imaging reporter used. Importantly, unlike the substrate-based probes benefiting from the enzyme turnover, the imaging probes bind to MMPs in a stoichiometric manner, with evident consequences on the detection limit. This may be particularly critical in the case of MMPs that are present under their active forms at very low concentrations at a pathogenic site. Another limitation of the MMP inhibitor-derived probes relies

on their structure. Indeed, most MMP inhibitors have been designed to target both the hydrophobic S₁' subsite and the catalytic zinc ion within the MMPs' active site. In this respect, they often display several aromatic units and negatively charged functions, structural features that favor unspecific binding to abundant plasma proteins (Casalini *et al.*, 2013)(Digilio *et al.*, 2014)(Bordenave *et al.*, 2016)(Razavian *et al.*, 2016). Interestingly, in the series of optical probes with a NIR-dye, it has also been shown that the dye itself can drastically modify the *in vivo* detection of the imaging probes (Razavian *et al.*, 2016). A similar impact of the reporter part has been observed with radiotracers differing in their ^{99m}Tc chelating groups (Toczek *et al.*, 2019). Beside their potential unspecific uptake ascribed to their trends to bind to serum albumin, most probes do not enable the precise documentation of the activation status of individual MMPs. Indeed, most MMP inhibitor-derived tracers are broad-spectrum and only provide a general overview of the MMPs activation pattern. Further, when a selective probe is used, the part of the whole signal corresponding to the probe/MMP complex remains difficult to attribute unambiguously, mainly due to the low concentration of active MMP *in vivo*.

C. Activity-based probes to crosslink active MMPs

The concept of activity-based protein profiling (ABPP) emerged at the end of the 1990s, demonstrated by a chemical strategy that utilizes active site-directed covalent probes to profile the functional state of enzymes in complex proteomes and *in vivo* (Evans and Cravatt, 2006). A typical activity-based probe (ABP) is composed of 1) an electrophilic "warhead" to react with a catalytic nucleophile, 2) a targeting motif that imposes selectivity upon the reactive moiety, and 3) an analytical handle for subsequent visualization and characterization of the resulting covalent adduct. Since MMPs use zinc-activated water molecules rather than a protein-bound nucleophile for catalysis, the ABPs targeting those proteases are systematically composed of a reversible inhibitor to which a photolabile group is attached (Geurink *et al.*, 2011)(Garland *et al.*, 2016). Upon photoirradiation, the photolabile group (diazirine, phenyl azido, or benzophenone) is transformed into a reactive intermediate (carbene, nitrenes, or ketyl biradical, respectively) that crosslinks amino acids in close proximity. The resulting covalently modified MMP could be distinguished from unlabeled proteins on SDS-PAGE gel or further captured for identification by mass spectrometry. Based on this principle, three types of MMP-directed ABPs have been designed: those targeting the non-primed region, those able to crosslink residues within the S₁' cavity, and ABPs reacting with amino acids present within the S₂' or S₃' primed subsites (**Fig. 9**). In 2004, two groups independently reported the two first ABPs targeting MMPs in their non-primed and primed regions, respectively (Chan *et al.*, 2004)(Saghatelian *et al.*, 2004). The "left-handed" probe with a C-terminal hydroxamic acid (probe **A**, **Fig. 9**) was able to crosslink MMP-9 (Chan *et al.*, 2004) but had a relatively low binding affinity toward MMPs, thus limiting its scope of application. Conversely, the APBs whose structure derived from marimastat (probe **B**, **Fig. 9**) were more effective to covalently modify the MMPs primed subsites. In this respect, ABPs with a benzophenone facing the S₂' subsite could crosslink several MMPs, down to 3 ng-50 fmol of MMP2 spiked into a complex proteome (Saghatelian *et al.*, 2004). The N-terminal succinyl-hydroxamate motif has been further exploited to provide ABPs with a small clickable handle (alkyne tag) for pulldown of active proteases from complex mixtures (Sieber *et al.*, 2006). The goal was to design probes with no bulky reporter groups (e.g., fluorescent) that might adversely affect probe-metalloprotease interactions. Interestingly, the use of those probes on several human cancer cells lines highlighted two major points: the succinyl hydroxamate warhead could target

metalloproteases other than MMPs, and no endogenous MMPs could have been detected, suggesting that MMPs are often found entirely in zymogen or TIMP-bound form in cancer cell proteomes. With this type of ABPs, endogenous MMP2 was detected in a zebrafish model (Keow *et al.*, 2012). More recently, similar APBs with a cleavable linker were used to characterize active MMPs in an *in vitro* model of early-stage cartilage degradation in post-traumatic osteoarthritis (Ravindra *et al.*, 2018). In this study, after pulldown, TCEP-mediated cleavage, digestion of the released proteases by trypsin, and multidimensional LC-MS/MS analysis using spectral counting, several endogenous MMPs including MMP1, -2, -3, -9, -12, and -13 were detected (Ravindra *et al.*, 2018). Importantly, the authors also showed that a significant fraction of the total secreted MMP was inactive. The marimastat-derived ABPs were also used to implement a metalloprotease activity multiplexed bead-based immunoassay as an effective approach for detecting MMP activity at picomolar range in conditioned cell culture media, peritoneal fluid aspirates, menstrual effluent, and uterine biopsies (Ahrens *et al.*, 2019).

Complementary to ABPs targeting S₂' and S₃' subsites, photoaffinity probes able to react within the hydrophobic S₁' subsite were also developed (David *et al.*, 2007)(Bregant *et al.*, 2009)(Geurink *et al.*, 2010)(Nury, Czarny, *et al.*, 2013). The first one was reported by the group of Vincent Dive in 2007 (David *et al.*, 2007). This probe contained a phosphinic pseudo peptide scaffold and a P₁' side chain with a phenyl azido moiety as a photolabile group and was shown to covalently react with several MMPs. Importantly, the incorporation of a radioactive isotope (tritium) within its structure not only enabled to reach a high detection threshold by in-gel radio imaging but also allowed to accurately quantitate the crosslinking yield for each tested MMP. In this series of phenyl azido probes and through further studies, the labeling sites within the S₁' cavity of MMP12 (Dabert-Gay *et al.*, 2008) and MMP3 (Dabert-Gay *et al.*, 2009) were also determined. This enabled a better understanding of the probe's reactivity, which resulted in the conception of the second generation of photoaffinity probes with a diazirine photolabile group (Nury, Czarny, *et al.*, 2013) (Probe C, Fig. 9). Compared to the phenyl azido probes, such a probe reacted efficiently with a larger set of MMPs and was able to detect endogenous MMP12 in bronchoalveolar lavage fluids (Nury, Bregant, *et al.*, 2013). More widely, ABPs targeting the S₁' subsite are more effective than those targeting the S₂' and S₃' subsites. This was particularly well illustrated by the work of Geurink *et al.*, that compared photoaffinity probes with a diazirine moiety in P₁' or P₂' position (Geurink *et al.*, 2010). Interestingly, the probe with the photolabile group in P₁' crosslinked all the tested MMPs (MMP1, -2, -3, -7, -8, -9, -10, -11, -12, and -13) while the probe with the same group in P₂' only efficiently reacted with MMP9 and -10. This observation has been rationalized by the fact that the P₂' photolabile group was more exposed to solvent than in P₁', which significantly reduced its reactivity towards protein amino acids. Although capable of detecting endogenous MMPs (Ravindra *et al.*, 2018)(Ahrens *et al.*, 2019), photoaffinity probes display a major limitation related to their intrinsic reactivity. Indeed, since the photoirradiation step is not straightforwardly achieved *in vivo*, the use of photoaffinity probes remains restricted to *ex vivo* samples. To address this issue, the group of Matt Bogoyo designed ABPs with an alpha-chloroacetamide electrophile in P₂' position (Probe D, Fig. 9) that can react with a cysteine residue artificially incorporated with the MMPs catalytic cleft (Morell *et al.*, 2013)(Amara *et al.*, 2018). This approach notably enabled monitoring the activity of MT1-MMP in cells and *in vivo* but required genetic manipulations incompatible with native conditions. More recently, and inspired by the ligand-directed chemistry developed by the group of Itaru Hamachi (Tamura and Hamachi, 2019), a new type of MMP-directed ABPs has been published (Kaminska *et al.*, 2021). The affinity probe harbors a phosphinic

pseudo peptide scaffold and a N-acyl-N-alkyl sulphonamide cleavable electrophile in P₃' (Probe **D**, **Fig. 9**), which can react selectively with lysine in close proximity within the MMP S₃' region. Through this strategy, MMP12 but also MMP3, -9, and -13 can be covalently tagged and detected by in-gel fluorescence imaging. This “photoactivation-free” strategy was also validated in complex proteomes and on native MMP12 under its active form, paving the way for the development of ABPs capable of reporting the MMP functional status *in vivo*.

VII. Conclusions and Future Perspectives

MMPs were first identified as ECM degrading enzymes but are not recognized as multifunctional proteases that influence cell migration, invasion, adhesion, signalling, immune responses, among others. Although it is tempting to inhibit MMPs proteolytic activities in diseases, we have learnt that they also have protective functions and participate in more processes than we initially thought (Sternlicht and Werb, 2001)(Overall and López-Otín, 2002)(Dufour and Overall, 2013) (Young *et al.*, 2019). Despite MMPs' protective functions, it is still feasible that an effective MMP inhibitor could be used to treat human diseases. There are many areas of MMP research that warrant additional investigation: 1) better characterization of their non-proteolytic roles; 2) identification of their non-ECM substrates; 3) studying the differences between their intracellular and extracellular functions; 4) profiling their post-translational modifications; 5) measuring their activity *in vivo* and *ex vivo* to distinguish between proMMP, inhibitor-bound MMP and active MMPs; and 6) development of selective activity-based probes (**Fig. 10**). Genomics, proteomics, and N-terminomics studies have not yet been performed on all human MMPs or MMP-deficient mice; therefore, there are still important basic characterization and discoveries to be made. Here, we identified six areas of future research that will be critical to complete our understanding of MMPs.

A. MMPs and their Non-Proteolytic Roles: Missed in the Myth?

ProMMPs are also capable of impacting key biological functions such as cell migration (Dufour *et al.*, 2008)(Dufour *et al.*, 2010)(Pavlaki *et al.*, 2011), cell survival (Conant, C St. Hillaire, *et al.*, 2004)(Redondo-Muñoz *et al.*, 2010), branching morphogenesis (Correia *et al.*, 2013), bacterial killing (Houghton *et al.*, 2009), and immune regulation (Shimizu-Hirota, Xiong, Baxter, Kunkel, Maillard, X.-W. Chen, *et al.*, 2012)(Marchant *et al.*, 2014). The study of the non-proteolytic roles of MMPs has so far revealed unexpected functions and is likely to uncover even more novel roles and maybe generate unique ways to control MMPs' biological functions. Another therapeutic opportunity could be to develop inhibitors that interfere with proteins that interact with MMPs' exosites or target downstream pathways that are impacted by MMPs' non-proteolytic roles.

B. Non-ECM MMP Substrates

Despite what their name indicates, MMPs can cleave more non-ECM proteins than matrix proteins (~69% *vs* ~31%, respectively, **Supplementary Table 1**). MMPs can cleave cell surface proteins, receptors, chemokines, cytokines, and other non-matrix proteins, thereby making them more fascinating than initially thought (Butler and Overall, 2009)(Dufour and Overall, 2015)(Young *et al.*, 2019). N-terminomics approaches have largely contributed to the identification of MMPs non-ECM, and there are still several human and murine MMPs to be characterized (Butler and Overall,

2009)(Bellac *et al.*, 2014)(Marchant *et al.*, 2014)(Brendan Mallia-Milanes *et al.*, 2018).

C. Intracellular vs Extracellular Functions of MMPs

MMPs are secreted proteases but, up to the time of this publication, 11 MMPs have also been demonstrated to have intracellular functions (*see this review for additional details* (Bassiouni *et al.*, 2021)). They can influence immune regulation via DNA interactions (Shimizu-Hirota, Xiong, Baxter, Kunkel, Maillard, X.-W. Chen, *et al.*, 2012)(Marchant *et al.*, 2014), oxidative stress via contractile dysfunction (Ali *et al.*, 2011)(Fan *et al.*, 2016), mammary epithelial branching morphogenesis (Correia *et al.*, 2013), neuronal signaling (Szkarczyk *et al.*, 2007)(Miller *et al.*, 2010) and many others (**Fig. 5**)(Bassiouni *et al.*, 2021). Intracellular functions of MMPs have been associated with various pathologies, including inflammatory disease, bacterial/viral infections, and cancer; however, there are no specific ways to selectively target intracellular MMPs, and still limited mechanistic information is known on how extracellular MMPs enter inside the cells despite some mechanisms have been proposed (Koppiseti *et al.*, 2014)(Bassiouni *et al.*, 2021).

D. Post-Translational Modifications of MMPs

The post-translational modifications (PTMs) of protease substrates can increase or decrease the rate of proteolysis (Madzharova *et al.*, 2019). In other instances, PTMs can protect protease substrates such as lysine methylation (Wigle *et al.*, 2010), glycosylation (King *et al.*, 2018), or phosphorylation (Turowec *et al.*, 2014)(Dix *et al.*, 2012). For example, MMP9 substrate alpha-fetoprotein (*FETA*) was demonstrated to be impacted by GalNAc-T2 O-glycosylation, and multiple additional MMP9 substrates were identified to be involved in a complex interplay between proteolysis and O-glycosylation (King *et al.*, 2018). Interestingly, MMP9 itself is also N- and O-glycosylated and contains a 64 amino acid residue linker domain implicated in orienting the hemopexin domain for inhibition by TIMP1 and regulating its interaction with cargo receptors (LRP-1 and megalin) for internalization (Van den Steen *et al.*, 2006)(Rosenblum *et al.*, 2007). Mutant MMP9 lacking the O-glycosylated linker region (OG domain) diminished MMP9's ability to increase cell migration (Dufour *et al.*, 2008). Lack of glycosylation also impacts the role of MMP9; for example, MMP9 lacking glycosylation at Asn³⁸ results in an amino acid-dependent interaction with calreticulin, likely impacting its release towards the secretory pathway (Duellman *et al.*, 2015). MMP9 can also be citrullinated by peptidyl arginine deiminases (PADs), resulting in a higher affinity for gelatin and hyperactivation in comparison to the uncitrullinated counterpart; citrullinated MMP9 was detected in neutrophils-rich sputum samples of cystic fibrosis patients suggesting a key role for PTMs in human disease (Boon *et al.*, 2021). The functions of other MMPs are also modulated by glycosylation (Boon *et al.*, 2016)(Madzharova *et al.*, 2019). For example, glycosylated MMP1(e.g., GalNAc β 1,4-(Fuc α 1,3)GlcNAc β 1,2) has higher levels of interaction with selectin on the cell surface, increasing cell migration (Saarinen *et al.*, 2001)(Boon *et al.*, 2016). MMP2, -3, -13, and MT4-MMP also contain several glycosylated sites, yet little is currently known about the impact on their functions.

MMPs have been demonstrated to be phosphorylated, but the biological significance of this PTM on MMP functions still needs to be characterized. For example, MMP2 contains 29 potential phosphorylation sites, but only five (S32, S160, Y271, T250, and S365) have been validated by mass spectrometry so far (Madzharova *et al.*, 2019). Although little is known, it appears that

dephosphorylated MMP2 contains higher enzymatic activity in comparison to phosphorylated MMP2 due to a conformational change (Jacob-Ferreira *et al.*, 2013)(Sariahmetoglu *et al.*, 2007). The phosphorylation of MT1-MMP at nine potential sites also impacts its biological function, specifically cell migration and invasion (García-Pardo and Opdenakker, 2015). Substitution of MT1-MMP's cytoplasmic domain Thr567 by an alanine residue resulted in an elevation of MT1-MMP levels at the cell surface, diminished internalization, and reduced cell migration and invasion (Williams and Coppolino, 2011). Additionally, several MMPs (MMP1, -12, -13, -14, -16, -24, and -27) can be phosphorylated extracellularly on tyrosine residues located in their hemopexin domains by the vertebrate lonesome kinase (VLK) (Bordoli *et al.*, 2014)(Tagliabracci *et al.*, 2015). Furthermore, oxidative and nitrosative stress that increase the activation of MMPs, and has been linked to cardiac and neuronal pathologies, should be investigated in additional diseases (Cheung *et al.*, 2000)(Wang *et al.*, 2002)(Gu *et al.*, 2002). Despite numerous examples of how PTMS impact MMPs, there are still multiple PTMs that have not been examined in the context of MMP functions, constituting an important area of future MMP research.

E. Measuring MMP Activity and Diagnostic Applications

As MMPs are present under their active form in several diseases, molecular probes to monitor MMPs activity must be rationally designed with this in mind, and they must be thoroughly validated in different biological fluids and preclinical models. In the perspective of developing efficient diagnostic tools, their validation in human samples will also be necessary. This will certainly require optimizing not only their specificity, but also their capacity to simultaneously detect several MMPs (multiplexing) in many different types of human matrices, this last point being essential for precision diagnostic. To this aim, microfluidic chips and microarray (Lei *et al.*, 2020) allowing automation and high throughput could constitute attractive options for the parallel analysis of multiple samples containing several MMPs. In any case, this will generate a large set of data that will need to be classified and interpreted. In such a scenario, it will become critical not only to provide robust data with a high degree of confidence but also to exploit the resources of artificial intelligence and machine learning for predictive classification (Kirkpatrick *et al.*, 2020). All those efforts would result in a new generation of activity-based diagnostics suitable in many human pathologies within which active MMPs are dysregulated.

F. The Future of MMP-directed Molecular Probes

Despite numerous signs of progress in the development of molecular probes for MMPs in the last two decades, some issues and limitations still need to be addressed to determine the MMPs activation status accurately and unambiguously both in *ex vivo* biological fluids and *in vivo*. In the case of substrate-based probes, identifying a peptide sequence cleaved exclusively by MMPs with no interferences with off-target proteases remains quite challenging. The recent efforts made to document MMPs endogenous substrates and to identify eventual cooperativity between subsites within the catalytic cleft (Schlage and auf dem Keller, 2015)(Eckhard, Huesgen, Schilling, Bellac, Butler, Cox, Dufour, Goebeler, Kappelhoff, Keller, *et al.*, 2016)(Eckhard, Huesgen, Schilling, Bellac, Butler, Cox, Dufour, Goebeler, Kappelhoff, auf dem Keller, *et al.*, 2016) would facilitate the design of sequences with high selectivity. However, due to the high homology among the different members of the MMP family, identifying substrate sequences capable of discriminating every single MMP seems difficult to achieve. As recently reported for MT1-MMP (Ji *et al.*, 2020),

such a limitation can be partly overcome by combining the MMP cleavable sequence with a binding motif interacting away from the catalytic cleft. In principle, and benefiting from recent studies pointing out the presence of discriminating allosteric sites within several MMPs (Sela-Passwell *et al.*, 2010)(Udi *et al.*, 2015)(Tokmina-Roszyk and Fields, 2018), this strategy should be readily expandable to a larger set of MMPs. A high degree of selectivity for a single MMP can also be obtained when the peptide sequence is installed on nanostructures. In this respect, a recent report showed that the morphology and size of a nanostructure have to be finely tuned to yield a nanosensor with improved selectivity (Son *et al.*, 2019). Specifically, the authors showed that the global charge of MMP (MMP9 in this study) ruled its electrostatic interaction with the nanostructure, stressing the need to take this parameter into account during the development of nanosensors. New selective MMP activatable probes could also be identified using an unbiased activity-profiling strategy with no *a priori* knowledge of the targeted MMP. This might consist in screening combinatorial substrate library directly on relevant pathological tissues, as recently validated on substrate-based probes targeting cathepsins (Tholen *et al.*, 2020).

Through the development of the first chemical probes able to covalently modify active MMPs with no photoactivation (Kaminska *et al.*, 2021), a major step has been taken, and the proteomic profiling of active MMPs *in vivo* should become amenable. However, to limit the background labelling and to expand this approach to a larger set of MMPs, several adjustments in the structure of activity-based probes will probably be necessary. This might require optimizing the structure of the binding motif as well as the positioning of the electrophile warhead within the MMP catalytic groove. In this respect, the phosphinic pseudo peptide scaffold that can project chemical residues on both sides of the MMP cleavage site would turn out particularly valuable. In addition, other types of biorthogonal chemistry could be implemented within the MMP catalytic cleft to further improve the probes' specificity and its crosslink efficiency. For instance, recent approaches enabling the chemo-selective modification of carboxyl groups in proteins (Ma *et al.*, 2020)(Martín-Gago *et al.*, 2017) could be adapted to target the MMP catalytic glutamate, which may result in ABPs capable of crosslinking a very broad set of MMPs.

Beyond the selectivity aspects and having in mind that MMPs under their active form are often present in low amounts in biological matrices, the sensitivity of the MMP-directed molecular probes could also be improved. This is particularly true in the series of ligand-based probes that do not benefit from the signal amplification due to enzyme turn-over. In this series, a multistep process involving affinity capture, on-bead digestion, and a mass spectrometry analysis remains the standard to reach a high sensitivity of detection (Sieber *et al.*, 2006)(Ravindra *et al.*, 2018). However, the recent progress made in mass spectrometry approaches, particularly in the development of derivatization strategies with mass tags to improve the detection of peptides and proteins in complex biological matrices (Qiao *et al.*, 2014)(Zhou *et al.*, 2021)(Sejalón-Cipolla *et al.*, 2021), could enable simplifying the analytical procedure. Specifically, by relying on the capacity of ABPs to selectively transfer a mass tags to MMPs and by using shotgun proteomic approaches (Zhang *et al.*, 2013), it should be possible to implement a targeted proteomic approach with a minimum of handling steps while maintaining a high sensitivity threshold. Furthermore, this would reduce potential changes coming from variabilities in sample manipulation, thus increasing the robustness of the generated data.

Finally, a better characterization of the role of individual MMP could result in novel approaches to inhibit MMP functions or, alternatively, to use the proteolytic activity of MMPs to help treat in the treatment of multiple diseases, such as localized scleroderma. Importantly, as MMPs are expressed in various tissues and immune cell types, it is important to analyze and identify local and systemic side effects according to each disease. With many MMPs having beneficial roles, it is key to analyze whether these protective functions will be affected or not by the specific MMP inhibitor being tested. Furthermore, antibodies, peptidomimetic inhibitors, or small molecule inhibitors are likely going to generate different side effects that need to be carefully monitored. Despite decades of MMP research, multiple failed clinical trials for cancer and arthritis, and a few successful clinical trials for periodontitis (Periosat®[®], doxycycline hyclate) and multiple sclerosis (minocycline), there are many opportunities that remain for the use of MMP inhibitors or MMP proteolytic activity as therapeutics.

Table 1: MMP substrates.

MMPs	Total number of putative substrates	ECM substrates	Non-ECM substrates
MMP1	41	10	31
MMP2	94	27	67
MMP3	74	26	48
MMP7	72	20	52
MMP8	38	9	29
MMP9	85	21	64
MMP10	8	4	4
MMP11	18	1	17
MMP12	69	19	50
MMP13	47	26	21
MMP14/MT1-MMP	89	26	63
MMP15/MT2-MMP	7	4	3
MMP16/MT3-MMP	12	4	8
MMP17/MT4-MMP	8	2	6
MMP19	12	8	4
MMP20	7	7	0
MMP21	0	0	0
MMP23	0	0	0
MMP24/MT5-MMP	4	1	3
MMP25/MT6-MMP	25	2	23
MMP26	8	4	4
MMP27	0	0	0
MMP28	0	0	0
Total	718	221	497
Percentages	100%	30.8%	69.2%

Table 2: MMP knockout mice phenotypes.

MMP	Phenotype of MMP knockout mouse	Impact on biological functions	References
<i>Mmp1a</i> ^{-/-}	No obvious abnormalities and both males and females were fertile.	↓ lung tumor growth and angiogenesis using LLC1 lung cancer cell line.	(Fanjul-Fernández <i>et al.</i> , 2013)(Foley <i>et al.</i> , 2014)(Fanjul-Fernández <i>et al.</i> , 2018)
<i>Mmp1b</i> ^{-/-}	<i>N/A</i>	<i>N/A</i>	<i>N/A</i>
<i>Mmp2</i> ^{-/-}	Both males and females were fertile. Smaller at birth and ~15% slower growth rate. Litters from heterozygous crosses yielded ~85% fewer than expected homozygous <i>Mmp2</i> ^{-/-} pups.	↑susceptibility to dextran-sulphate-induced colitis, ↑protection and ↓reduction in hepatocyte apoptosis in TNFα-induced hepatitis, ↑severity of antibody-induced arthritis, ↓white matter sparing and ↓serotonergic fibers in spinal-cord lesions,	(Itoh <i>et al.</i> , 1997)(Wielockx <i>et al.</i> , 2001)(Itoh <i>et al.</i> , 2002)(Campbell <i>et al.</i> , 2005)(Samolov <i>et al.</i> , 2005)(Garg <i>et al.</i> , 2006)(Hsu <i>et al.</i> , 2006)(Kuzuya <i>et al.</i> ,

		<p>↑cardiac allograft survival, Delay of neovascularization, ↓atherosclerotic plaques, ↓numbers and proliferation of osteoblasts and osteoclasts, ↑apoptosis,</p>	2006)(Martignetti <i>et al.</i> , 2014)
<i>Mmp3^{-/-}</i>	No obvious abnormalities and both males and females were fertile.	<p>↑protection and ↓reduction in hepatocyte apoptosis in TNFα-induced hepatitis, ↓immune-complex-induced lung injury and ↓neutrophils, ↑myocardial scar volume post injury, ↓skin-contact hypersensitivity, Delayed clearance of bacteria and appearance of CD4⁺ T lymphocytes into intestinal lamina propria, ↓production of macrophage chemoattractant in disc hernia <i>in vitro</i></p>	(Mudgett <i>et al.</i> , 1998)(Wang <i>et al.</i> , 1999)(Haro, Crawford, Fingleton, MacDougall, <i>et al.</i> , 2000)(Wielockx <i>et al.</i> , 2001)(Warner <i>et al.</i> , 2001)(Li <i>et al.</i> , 2004)(Mukherjee <i>et al.</i> , 2005)
<i>Mmp7^{-/-}</i>	No obvious abnormalities and both males and females were fertile. Normal lifespan.	<p>↓release of TNFα from peritoneal macrophage <i>in vitro</i>, ↓transepithelial neutrophil migration in bleomycin-induced lung injury, ↓re-epithelization post-injury in trachea, ↓processing of alpha-defensin resulting in ↓innate immunity, ↑corneal neovascularization after injury, ↓epithelial cell apoptosis linked with ↓Fas ligand processing, Protected from LPS-induced intestinal permeability and lethality</p>	(Wilson <i>et al.</i> , 1997)(Dunsmore <i>et al.</i> , 1998)(Wilson, 1999)(Powell <i>et al.</i> , 1999)(Haro, Crawford, Fingleton, Shinomiya, <i>et al.</i> , 2000)(Li <i>et al.</i> , 2002)(Kure <i>et al.</i> , 2003)(Vandenbroucke <i>et al.</i> , 2014)
<i>Mmp8^{-/-}</i>	Normal development, fertile, and no reduction in survival.	<p>↑incidence of skin tumors, ↓inflammatory cell apoptosis but ↑neutrophils in BAL in an asthma model, ↑protection in TNFα-induced hepatitis and impaired leukocyte influx, ↓neutrophil infiltration towards LPS, ↓mortality and hypothermia in sepsis and renal ischemia/reperfusion</p>	(Babín <i>et al.</i> , 2003)(Gueders <i>et al.</i> , 2005)(Van Lint <i>et al.</i> , 2005)(Tester <i>et al.</i> , 2007)(Vandenbroucke <i>et al.</i> , 2012)(Fortelny <i>et al.</i> , 2014)
<i>Mmp9^{-/-}</i>	No gross phenotypic abnormalities, fertile, and normal lifespan. However, they have abnormal development of growth plates in long bones.	<p>↑protection and ↓reduction in hepatocyte apoptosis in TNFα-induced hepatitis, ↓severity of antibody-induced arthritis, ↓cardiac allograft survival, ↓immune-complex-induced lung injury, Prolonged skin-contact hypersensitivity, ↓alveolar bronchiolization after bleomycin treatment, ↑bronchoalveolar lavage cell recruitment post allergic challenge, Impaired neutrophil infiltration and ↑early vascular permeability in a model of zymosan peritonitis, ↓resistance against <i>Escherichia coli</i> peritonitis due to ↓leukocyte recruitment, ↓dextran-sulfate-induced colitis, ↑bacterial-induced arthritis but ↓bacterial clearance, ↑brain haemorrhage and injury, ↓remyelination after spinal cord trauma, Spontaneous deficient myelination of corpus callosum with fewer oligodendrocytes, ↓experimental autoimmune encephalomyelitis in young mice, ↓vessel formation in an ischaemic limb model, ↓left-ventricle dilation and fibrosis post pressure overload, ↓capillary branching post ischaemic insult, ↑myocardial injury and foci of infection when infected with coxsackivirus B3</p>	(Vu <i>et al.</i> , 1998)(Wang <i>et al.</i> , 1999)(Dubois <i>et al.</i> , 1999)(Betsuyaku <i>et al.</i> , 2000)(Wielockx <i>et al.</i> , 2001)(Warner <i>et al.</i> , 2001)(Itoh <i>et al.</i> , 2002)(Larsen <i>et al.</i> , 2003)(McMillan <i>et al.</i> , 2004)(Johnson <i>et al.</i> , 2004)(Tang <i>et al.</i> , 2004)(Campbell <i>et al.</i> , 2005)(Heissig <i>et al.</i> , 2005)(Heymans <i>et al.</i> , 2005)(Castaneda <i>et al.</i> , 2005)(Kolaczowska <i>et al.</i> , 2006)(Renckens <i>et al.</i> , 2006)(Calander <i>et al.</i> , 2006)(Larsen, 2006)(Cheung <i>et al.</i> , 2008)
<i>Mmp10^{-/-}</i>	No overt defects in fertility, litter size, gross appearance, organ structure, or tissue histology.	<p>Muscles display ↓recruitment of endothelial cells, ↓ECM proteins, ↓fiber size, Delayed fibrinolysis, ↑collagen deposition in skin wounds, ↑infiltration of macrophages during acute infection.</p>	(Kassim <i>et al.</i> , 2007)(Orbe <i>et al.</i> , 2011)(Bobadilla <i>et al.</i> , 2014)(Rohani <i>et al.</i> , 2015)(McMahan <i>et al.</i> , 2016)
<i>Mmp11^{-/-}</i>	Viable, fertile, no behaviour differences with wild type littermates.	<p>↑neointima formation following vascular injury, ↓7,12-dimethylbenzanthracene-induced tumorigenesis, ↑subcutaneous (SC) and gonadal (GON) fat deposits, adipogenesis and adipocyte membrane alteration,</p>	(Masson <i>et al.</i> , 1998)(H. Roger Lijnen <i>et al.</i> , 1999)(Lijnen <i>et al.</i> , 2002)(Andarawewa <i>et al.</i> ,

		↑dysregulation of metabolism.	2005)(Dali-Youcef <i>et al.</i> , 2016)
<i>Mmp12</i> ^{-/-}	Normal embryonic and postnatal development in the absence of inflammatory stress. Resting hematopoiesis and myelomonocytic development are normal. <i>Mmp12</i> ^{-/-} mice have a 40% reduction in litter sizes likely due to placental abnormalities.	↓migration and invasion in macrophages, Resistance against cigarette-smoke-induced emphysema, Unable to process chemokines and cytokines like IFN α and IFN γ resulting in exacerbation of acute and chronic inflammation, Unable to resolve bacterial and virus infections, Reduced phagocytosis and hemolysis capacity, Dysregulated complement activation, ↓macrophage infiltration in ligament-injury repair, Spontaneous deficient myelination of corpus callosum with fewer oligodendrocytes, ↑experimental autoimmune encephalomyelitis severity,	(Shibley <i>et al.</i> , 1996)(Hautamaki <i>et al.</i> , 1997)(Weaver <i>et al.</i> , 2005)(Larsen, 2006)(Dean <i>et al.</i> , 2008)(Houghton <i>et al.</i> , 2009)(Marchant <i>et al.</i> , 2014)(Bellac <i>et al.</i> , 2014)(Dufour <i>et al.</i> , 2018)(B. Mallia-Milanes <i>et al.</i> , 2018)
<i>Mmp13</i> ^{-/-}	No gross phenotypic abnormalities, fertile, and normal lifespan.	Spontaneous abnormal growth plate and ↑trabecular bone	(Stickens <i>et al.</i> , 2004)(Inada <i>et al.</i> , 2004)
<i>Mmp14</i> ^{-/-} / <i>Mt1-mmp</i> ^{-/-}	Viable but display severe runting, wasting and ↑mortality. Death occurs between 50-90 days after birth.	Craniofacial dysmorphism, arthritis, osteopenia, dwarfism, and fibrosis of soft tissues, ↓lung alveolar surface area, Retarded lung alveolar development, ↓neovessel formation, Poorly differentiated kidney tubular epithelia, Lack of molar tooth eruption and root formation, Defects in dentin formation and mineralization, ↓metabolic homeostasis, ↓regulation of adipocyte fate determination in the developing mammary gland.	(Holmbeck <i>et al.</i> , 1999)(Holmbeck <i>et al.</i> , 2003)(Mai <i>et al.</i> , 2017)(Chun <i>et al.</i> , 2004)(Koshikawa <i>et al.</i> , 2004)(Irie <i>et al.</i> , 2005)(Atkinson <i>et al.</i> , 2005)(Mori <i>et al.</i> , 2016)(Xu <i>et al.</i> , 2016)(Feinberg <i>et al.</i> , 2016)
<i>Mmp15</i> ^{-/-} / <i>Mt2-mmp</i> ^{-/-}	Viable, fertile, and live through adulthood.	↑positive regulators of brown or induced brown (beige) adipocytes production.	(Feinberg <i>et al.</i> , 2016)
<i>Mmp16</i> ^{-/-} / <i>Mt3-mmp</i> ^{-/-}	Viable, fertile, but displayed retarded growth of the skeleton compared to wild types or heterozygous littermates.	↓viability of mesenchymal cells in skeletal tissues.	(Shi <i>et al.</i> , 2008)
<i>Mmp17</i> ^{-/-} / <i>Mt4-mmp</i> ^{-/-}	No apparent defects in growth, fertility, and life span.	↑protection from IL-1 β -mediated GAG release into synovial fluid in a model of joint inflammation. ↑predisposition to aortic aneurysms. Plays a role in embryonic development, brain formation, angiogenesis and limb development.	(Rikimaru <i>et al.</i> , 2007), (Clements <i>et al.</i> , 2011), (Martín-Alonso <i>et al.</i> , 2015), (Blanco <i>et al.</i> , 2017)
<i>Mmp19</i> ^{-/-}	Viable, fertile, and no apparent phenotype under homeostasis.	↑body weight under high-fat diet, Adipocytes hypertrophy, ↓susceptibility to develop skin tumors induced by a model of chemical carcinogens, In a model of contact hypersensitivity, impaired T cell-mediated immune reaction characterized by minimal influx of inflammatory cells, low proliferation of keratinocytes, ↓activated CD8 ⁺ T cells in lymph nodes, After an allergen challenge, ↑eosinophilic inflammation and ↑tenascin protein, ↑lung fibrotic response to bleomycin, ↑protection against hepatic fibrosis, ↑susceptibility to colitis in a dextran sulfate sodium-induced colitis model.	(Pendás <i>et al.</i> , 2004), (Beck <i>et al.</i> , 2008), (Gueders <i>et al.</i> , 2010), (Yu <i>et al.</i> , 2012), (Jirouskova <i>et al.</i> , 2012), (Jara <i>et al.</i> , 2015), (Brauer <i>et al.</i> , 2016)
<i>Mmp20</i> ^{-/-}	Severe tooth defects but mice were viable and had no fertility issues.	Hypomineralization of mantle dentin, Defects in processing amelogenin, Altered enamel protein and associated rod pattern.	(Caterina <i>et al.</i> , 2002), (Beniash <i>et al.</i> , 2006)
<i>Mmp21</i> ^{-/-}	N/A	N/A	N/A
<i>Mmp23</i> ^{-/-}	N/A	N/A	N/A
<i>Mmp24</i> ^{-/-} / <i>Mt5-mmp</i> ^{-/-}	Normal appearance, fertile, and have normal lifespan.	Unable to develop neuropathic pain with mechanical allodynia following a sciatic nerve injury. ↓Nerve-fibre sprouting and neural invasion. ↑Sensitivity to noxious thermal stimuli under basal conditions. Unable to develop thermal hyperalgesia under inflammatory conditions. Protected against amyloid pathology, cognitive decline, and inflammation.	(Komori <i>et al.</i> , 2004), (Folgueras <i>et al.</i> , 2009), (Baranger <i>et al.</i> , 2016)

<i>Mmp25^{-/-}/Mt6-mmp^{-/-}</i>	Viable, fertile, and no apparent phenotype under homeostasis.	Defective innate immune response via a low sensitivity to bacterial LPS, Hypergammaglobulinemia, ↓secretion of proinflammatory molecules, Impaired NF-κB activation	(Soria-Valles <i>et al.</i> , 2016)
<i>Mmp26^{-/-}</i>	N/A	N/A	N/A
<i>Mmp27^{-/-}</i>	N/A	N/A	N/A
<i>Mmp28^{-/-}</i>	Viable, fertile, and no apparent phenotype under homeostasis.	↑Macrophage recruitment, ↑macrophage migration and ↑bacterial clearance into the lung in <i>Pseudomonas aeruginosa</i> treated mice, ↓macrophage polarization, ↓collagen deposition and few myofibroblasts.	(Manicone <i>et al.</i> , 2009)(Ma <i>et al.</i> , 2013)(Gharib <i>et al.</i> , 2014)(Manicone <i>et al.</i> , 2017)

Figure legends

Figure 1: Types of protease classes. The 473 human proteases are shown by their mechanism of catalysis. There are 152 serine proteases, 148 metalloproteases, 136 cysteine proteases, 21 aspartic proteases and 16 threonine proteases. For the full list, see the MEROPS database (<https://www.ebi.ac.uk/merops/>)

Figure 2: Domains of human MMPs. Schematic representation of the various domains of the 23 human MMPs. Archetypal MMPs (MMP1, -3, -8, -10, -12, -13, -19, -20, and -27) contain a signal peptide, propeptide, a catalytic domain that binds a Zn²⁺ residue, and a hemopexin C-terminal domain. Gelatinases (MMP2 and -9) contain a fibronectin type-II modules domain. Matrilysins (MMP7 and -26) contain a signal peptide, propeptide and a catalytic domain but lack a hemopexin C-terminal domain. The convertase-activable MMPs (MMP11, -14, -15, -16, -17, -21, -23B, -24, -25, and -28) contain a convertase cleavage site in their propeptide that is cleaved by furin or furin-like proteases. Six MMPs contain a membrane linker and are localized on the cell surface. Additional domains include glycosylphosphatidylinositol (GPI), a transmembrane segment 1 or 2, a cysteine array (CA) and an immunoglobulin-like (Ig-like) domain.

Figure 3: MMP structure. (A) Ribbon representation of proMMP2 (PDB: 1GXD). MMP domains and inorganic ions are labelled and colored. Prodomain is shown in magenta, catalytic domain in teal, linker region in black, hemopexin domain in grey, zinc ion in yellow, and calcium ion in dark grey. (B) Ribbon representation of MMP1's active site (PDB: 1AYK). The overall structure is colored in teal. Important residues in the zinc interaction are labelled, colored, and represented as sticks. Zinc is shown in yellow, histidine in grey, and glutamic acid in orange. (C) Ribbon representation of proMMP2's catalytic site. Secondary structures are colored, and important segments are also labelled. Calcium is shown in dark grey, zinc in yellow, α-helices in blue (αA, αB, and αC), β-sheets in green (βI, -II, -III, -IV, -V), S-loop in navy blue, specificity loop in purple, and met-turn in pink.

Figure 4: MMP substrates. Reported ECM (black) and non-ECM (cyan) MMP substrates for all 23 human MMPs as reported from TopFIND (<https://topfind.clip.msl.ubc.ca>) (Fortelny *et al.*, 2015). TopFIND integrates information from the UniProt knowledgebase (UniProtKB), MEROPS peptidase database and experimental terminomics studies of 8 species including (*Homo sapiens*, *Mus musculus*, *Rattus norvegicus*, *Arabidopsis thaliana*, *Saccharomyces cerevisiae* and *Escherichia coli*).

Figure 5: Intracellular MMPs and localization to the cytoplasm or nucleus in three different cell types. *Left*, some intracellular MMPs associated with cardiomyocytes. Some examples of known MMP2 interactions within organelles are illustrated. *Middle*, known intracellular MMPs in cancer cells. One example is the upregulation of MMP2 by IL8-mediated signalling, as observed in melanoma cells. *Left*, known intracellular MMPs in immune cells or virus-infected cells. An example is illustrated of MMP-12 mediated $\text{INF}\alpha$ release during viral infection.

Figure 6: Structure of MMP inhibitors. The chemical structure of some representative MMP inhibitors: Col-3/Metastat, AE-941/Neovostat, BB-94/batimastat, BMS-275291/Rabimastat, Ro-32-3555/Cipemastat, BB-2516/Marimastat GM6001/ilomastat, MMI270/CGS27023A, MMP8 inhibitor, AG3340/prinomastat, BAY-12-9566/tanomastat, MMP408, RXP470.1, SB-3CT, MMP8/13 and MMP13 inhibitor. Key: when known, the zinc-binding moiety is colored in magenta within the inhibitor structure. P₁' position and P₂'/P₃' positions are identified in blue and green, respectively.

Figure 7: Matrix metalloproteinase changes in inflammatory bowel disease (IBD) and arthritis. In healthy gut tissue (**top left panel**), homeostasis is established with a balance of pro-inflammatory and anti-inflammatory signals. The local immune response can be modulated by multiple factors, including the microbiota. Proteases, such as MMP7, can shape the microbiota, and gut immunity, by aiding the release of anti-microbial peptides. Reduction of pro-inflammatory signals lowers the expression of MMP3 and -12, setting the stage for successful application of anti-TNF α monoclonal antibodies. MMPs also perform protective roles, as the absence of MMP2 results in gut inflammation, although the exact mechanism remains to be explored. In IBD (**top right panel**), multiple MMPs are upregulated. For example, increased expression of MMP8, -9, and -10 results in tissue damage, generating damage-associated molecular patterns (DAMPs) that promote further expression of additional MMPs and pro-inflammatory signals, such as TNF α . MMPs are also capable of cleaving anti-TNF α drugs, reducing treatment efficacy. High expression of certain MMPs has been used in diagnostic tests for Crohn's disease, as the protease levels and cytokines are readily detectable in the blood. The systemic nature of IBD correlates with joint inflammation, where increased expression of multiple MMPs is also observed (**lower right panel**). High expression of MMPs results in tissue damage and a feed-forward mechanism for further joint inflammation. Remarkably, MMPs also have protective roles, as exemplified by animal models lacking *Mmp2* and *Mmp8* (**lower left panel**). Lastly, MMP12 can control joint inflammation via IFN γ cleavage, generating a product incapable of promoting downstream signaling of pro-inflammatory signals.

Figure 8: Design of Substrate-based sensors from a minimal peptide sequence of six amino acids incorporating a proline in P₃' position. The peptide sequence corresponds to a fluorogenic substrate crystallized within the catalytic cleft of an inactive mutant of MMP9 (PDB code: 4IJI, (Tranchant *et al.*, 2014)). From this sequence, different substrate-based probes/sensors have been designed. This notably includes (A) Optical sensors (e.g., NIR sensors) with a quencher and emitter of fluorescence, (B) Magnetic sensors (e.g., MRET probe), and (C) Synthetic biomarkers, which, upon MMP cleavage are secreted and detected in urine. Key: MMP9 (surface), peptide sequence (pale red stick), catalytic zinc ion (magenta ball); unprimed (P₃, P₂ and P₁), P₁' and primed (P₂' and P₃') positions are identified in purple, blue and green, respectively. The cleavage site is marked with red arrow.

Figure 9: Activity-based probes (ABPs) directed to the MMP catalytic cleft. From the MMP catalytic domain (PDB code: 4IJI, (Tranchant *et al.*, 2014)) in surface representation, three regions were distinguished: the S subsites marked in purple, the S₁' subsite in blue and the primed subsites in green. To each subsite is associated a ABP capable of crosslinking it. Probe **A** target the S subsites, probe **B**, **D** and **E** the S' subsites and probe **C** the S₁' subsite. For each probe, the zinc-chelating group is marked in magenta, the photolabile group or reactive electrophile is associated to the color of site within which it reacts, and the analytical tag is in red.

Figure 10: Topics for MMP research to be studied in the next decade. Schematic representation of six areas of MMP research that warrant further investigation: the non-proteolytic roles of MMPs, their non-ECM substrates, the difference in their intra- and extracellular roles, their post-translational modifications (PTMs), their activity and the development of new activity-based probes.

Authors contributions

Wrote and contributed to the writing of the manuscript: de Almeida, Thode, Eslambolchi, Chopra, Young, Gill, Devel and Dufour.

References

- Ager EI, Kozin S V., Kirkpatrick ND, Seano G, Kodack DP, Askoxylakis V, Huang Y, Goel S, Snuderl M, Muzikansky A, Finkelstein DM, Dransfield DT, Devy L, Boucher Y, Fukumura D, and Jain RK (2015) Blockade of MMP14 Activity in Murine Breast Carcinomas: Implications for Macrophages, Vessels, and Radiotherapy. *JNCI J Natl Cancer Inst* **107**.
- Agere SA, Akhtar N, Watson JM, and Ahmed S (2017) RANTES/CCL5 Induces Collagen Degradation by Activating MMP-1 and MMP-13 Expression in Human Rheumatoid Arthritis Synovial Fibroblasts. *Front Immunol* **8**.
- Agrawal S, Anderson P, Durbeej M, van Rooijen N, Ivars F, Opdenakker G, and Sorokin LM (2006) Dystroglycan is selectively cleaved at the parenchymal basement membrane at sites of leukocyte extravasation in experimental autoimmune encephalomyelitis. *J Exp Med* **203**:1007–1019.
- Agrawal SM, Silva C, Tourtellotte WW, and Yong VW (2011) EMMPRIN: A Novel Regulator of Leukocyte Transmigration into the CNS in Multiple Sclerosis and Experimental Autoimmune Encephalomyelitis. *J Neurosci* **31**:669–677.
- Agrawal SM, Williamson J, Sharma R, Kebir H, Patel K, Prat A, and Yong VW (2013) Extracellular matrix metalloproteinase inducer shows active perivascular cuffs in multiple sclerosis. *Brain* **136**:1760–1777.
- Ahonen M, Poukkula M, Baker AH, Kashiwagi M, Nagase H, Eriksson JE, and Kähäri V-M (2003) Tissue inhibitor of metalloproteinases-3 induces apoptosis in melanoma cells by stabilization of death receptors. *Oncogene* **22**:2121–2134.
- Ahrens CC, Chiswick EL, Ravindra KC, Miller MA, Ramseier JY, Isaacson KB, Lauffenburger DA, and Griffith LG (2019) Development and Application of the Metalloprotease Activity Multiplexed Bead-Based Immunoassay (MAMBI). *Biochemistry* **58**:3938–3942.
- Akahane T, Akahane M, Shah A, Connor CM, and Thorgerirsson UP (2004) TIMP-1 inhibits microvascular endothelial cell migration by MMP-dependent and MMP-independent

- mechanisms. *Exp Cell Res* **301**:158–167.
- Alexius-Lindgren M, Andersson E, Lindstedt I, and Engström W (2014) The RECK gene and biological malignancy-its significance in angiogenesis and inhibition of matrix metalloproteinases.
- Alfonso-Jaume MA, Bergman MR, Mahimkar R, Cheng S, Jin ZQ, Karliner JS, and Lovett DH (2006) Cardiac ischemia-reperfusion injury induces matrix metalloproteinase-2 expression through the AP-1 components FosB and JunB. *Am J Physiol Circ Physiol* **291**:H1838–H1846.
- Ali MAM, Chow AK, Kandasamy AD, Fan X, West LJ, Crawford BD, Simmen T, and Schulz R (2012) Mechanisms of cytosolic targeting of matrix metalloproteinase-2. *J Cell Physiol* **227**:3397–3404.
- Ali MAM, Fan X, and Schulz R (2011) Cardiac Sarcomeric Proteins: Novel Intracellular Targets of Matrix Metalloproteinase-2 in Heart Disease. *Trends Cardiovasc Med* **21**:112–118.
- Allan JA, Docherty AJP, Barker PJ, Huskisson NS, Reynolds JJ, and Murphy G (1995) Binding of gelatinases A and B to type-I collagen and other matrix components. *Biochem J* **309**:299–306.
- Amano T, Kwak O, Fu L, Marshak A, and Shi Y-B (2005) The matrix metalloproteinase stromelysin-3 cleaves laminin receptor at two distinct sites between the transmembrane domain and laminin binding sequence within the extracellular domain. *Cell Res* **15**:150–159.
- Amara N, Tholen M, and Bogyo M (2018) Chemical Tools for Selective Activity Profiling of Endogenously Expressed MMP-14 in Multicellular Models. *ACS Chem Biol* **13**:2645–2654.
- Amour A, Knight CG, Webster A, Slocombe PM, Stephens PE, Knäuper V, Docherty AJP, and Murphy G (2000) The in vitro activity of ADAM-10 is inhibited by TIMP-1 and TIMP-3. *FEBS Lett* **473**:275–279.
- Andarawewa KL, Motrescu ER, Chenard M-P, Gansmuller A, Stoll I, Tomasetto C, and Rio M-C (2005) Stromelysin-3 Is a Potent Negative Regulator of Adipogenesis Participating to Cancer Cell-Adipocyte Interaction/Crosstalk at the Tumor Invasive Front. *Cancer Res* **65**:10862–10871.
- Andoh A, Bamba S, Brittan M, Fujiyama Y, and Wright NA (2007) Role of intestinal subepithelial myofibroblasts in inflammation and regenerative response in the gut. *Pharmacol Ther* **114**:94–106.
- Anthony DC, Ferguson B, Matyzak MK, Miller KM, Esiri MM, and Perry VH (1997) Differential matrix metalloproteinase expression in cases of multiple sclerosis and stroke. *Neuropathol Appl Neurobiol* **23**:406–415.
- Apte SS, Mattei M-G, and Olsen BR (1994) Cloning of the cDNA Encoding Human Tissue Inhibitor of Metalloproteinases-3 (TIMP-3) and Mapping of the TIMP3 Gene to Chromosome 22. *Genomics* **19**:86–90.
- Arnhold J (2020) The Dual Role of Myeloperoxidase in Immune Response. *Int J Mol Sci* **21**:8057.
- Arpino V, Mehta S, Wang L, Bird R, Rohan M, Pape C, and Gill SE (2016) Tissue inhibitor of metalloproteinases 3-dependent microvascular endothelial cell barrier function is disrupted under septic conditions. *Am J Physiol Circ Physiol* **310**:H1455–H1467.
- Asaduzzaman M, Zhang S, Lavasani S, Wang Y, and Thorlacius H (2008) LFA-1 AND MAC-1 MEDIATE PULMONARY RECRUITMENT OF NEUTROPHILS AND TISSUE DAMAGE IN ABDOMINAL SEPSIS. *Shock* **30**:254–259.

- Atkinson JJ, Holmbeck K, Yamada S, Birkedal-Hansen H, Parks WC, and Senior RM (2005) Membrane-type 1 matrix metalloproteinase is required for normal alveolar development. *Dev Dyn* **232**:1079–1090.
- Atkinson SJ, Crabbe T, Cowell S, Ward R V., Butler MJ, Sato H, Seiki M, Reynolds JJ, and Murphy G (1995) Intermolecular Autolytic Cleavage Can Contribute to the Activation of Progelatinase A by Cell Membranes. *J Biol Chem* **270**:30479–30485.
- Augustin S, Berard M, Kellaf S, Peyri N, Fauvel-LAFÈVE F, Legrand C, He L, and Crépin M (2009) Matrix metalloproteinases are involved in both type I (Apoptosis) and type II (Autophagy) cell death induced by sodium phenylacetate in MDA-MB-231 breast tumour cells. *Anticancer Res* **29**.
- Baghirova S, Hughes BG, Poirier M, Kondo MY, and Schulz R (2016) Nuclear matrix metalloproteinase-2 in the cardiomyocyte and the ischemic-reperfused heart. *J Mol Cell Cardiol* **94**:153–161.
- Baker AH, Edwards DR, and Murphy G (2002) Metalloproteinase inhibitors: biological actions and therapeutic opportunities. *J Cell Sci* **115**:3719–3727.
- Balbín M, Fueyo A, Tester AM, Pendás AM, Pitiot AS, Astudillo A, Overall CM, Shapiro SD, and López-Otín C (2003) Loss of collagenase-2 confers increased skin tumor susceptibility to male mice. *Nat Genet* **35**:252–257.
- Baranger K, Marchalant Y, Bonnet AE, Crouzin N, Carrete A, Paumier J-M, Py NA, Bernard A, Bauer C, Charrat E, Moschke K, Seiki M, Vignes M, Lichtenthaler SF, Checler F, Khrestchatsky M, and Rivera S (2016) MT5-MMP is a new pro-amyloidogenic proteinase that promotes amyloid pathology and cognitive decline in a transgenic mouse model of Alzheimer's disease. *Cell Mol Life Sci* **73**:217–236.
- Bassiouni W, Ali MAM, and Schulz R (2021) Multifunctional intracellular matrix metalloproteinases: implications in disease. *FEBS J* febs.15701.
- Bateman JF, Boot-Handford RP, and Lamandé SR (2009) Genetic diseases of connective tissues: cellular and extracellular effects of ECM mutations. *Nat Rev Genet* **10**:173–183.
- Bauer EA, Eisen AZ, and Jeffrey JJ (1972) Regulation of Vertebrate Collagenase Activity In Vivo and In Vitro. *J Invest Dermatol* **59**:50–55.
- Beck IM, Rückert R, Brandt K, Mueller MS, Sadowski T, Brauer R, Schirmacher P, Mentlein R, and Sedlacek R (2008) MMP19 Is Essential for T Cell Development and T Cell-Mediated Cutaneous Immune Responses. *PLoS One* **3**:e2343.
- Bellac CL, Dufour A, Krisinger MJ, Loonchanta A, Starr AE, auf dem Keller U, Lange PF, Goebeler V, Kappelhoff R, Butler GS, Burtnick LD, Conway EM, Roberts CR, and Overall CM (2014) Macrophage matrix metalloproteinase-12 dampens inflammation and neutrophil influx in arthritis. *Cell Rep* **9**:618–632, Elsevier B.V.
- Bendell J, Sharma S, Patel MR, Windsor KS, Wainberg ZA, Gordon M, Chaves J, Berlin J, Brachmann CB, Zavodovskaya M, Liu J, Thai D, Bhargava P, Shah MA, Khan SA, and Starodub A (2020) Safety and Efficacy of Andecaliximab (<sc>GS</sc> -5745) Plus Gemcitabine and <sc>Nab-Paclitaxel</sc> in Patients with Advanced Pancreatic Adenocarcinoma: Results from a Phase I Study. *Oncologist* **25**:954–962.
- Beniash E, Skobe Z, and Bartlett JD (2006) Formation of the dentino-enamel interface in enamelysin (MMP-20)-deficient mouse incisors. *Eur J Oral Sci* **114**:24–29.
- Bergers G (1999) Effects of Angiogenesis Inhibitors on Multistage Carcinogenesis in Mice. *Science (80-)* **284**:808–812.
- Bergers G, Brekken R, McMahon G, Vu TH, Itoh T, Tamaki K, Tanzawa K, Thorpe P, Itohara S,

- Werb Z, and Hanahan D (2000) Matrix metalloproteinase-9 triggers the angiogenic switch during carcinogenesis. *Nat Cell Biol* **2**:737–744.
- Bergman MR, Cheng S, Honbo N, Piacentini L, Karliner JS, and Lovett DH (2003) A functional activating protein 1 (AP-1) site regulates matrix metalloproteinase 2 (MMP-2) transcription by cardiac cells through interactions with JunB-Fra1 and JunB-FosB heterodimers. *Biochem J* **369**:485–496.
- Bergman MR, Teerlink JR, Mahimkar R, Li L, Zhu B-Q, Nguyen A, Dahi S, Karliner JS, and Lovett DH (2007) Cardiac matrix metalloproteinase-2 expression independently induces marked ventricular remodeling and systolic dysfunction. *Am J Physiol Circ Physiol* **292**:H1847–H1860.
- Beroun A, Mitra S, Michaluk P, Pijet B, Stefaniuk M, and Kaczmarek L (2019) MMPs in learning and memory and neuropsychiatric disorders. *Cell Mol Life Sci* **76**:3207–3228.
- Bertini I, Calderone V, Fragai M, Jaiswal R, Luchinat C, Melikian M, Mylonas E, and Svergun DI (2008) Evidence of Reciprocal Reorientation of the Catalytic and Hemopexin-Like Domains of Full-Length MMP-12. *J Am Chem Soc* **130**:7011–7021.
- Betsuyaku T, Fukuda Y, Parks WC, Shipley JM, and Senior RM (2000) Gelatinase B Is Required for Alveolar Bronchiolization after Intratracheal Bleomycin. *Am J Pathol* **157**:525–535.
- Biancheri P, Brezski RJ, Di Sabatino A, Greenplate AR, Soring KL, Corazza GR, Kok KB, Rovedatti L, Vossenkämper A, Ahmad N, Snoek SA, Vermeire S, Rutgeerts P, Jordan RE, and MacDonald TT (2015) Proteolytic Cleavage and Loss of Function of Biologic Agents That Neutralize Tumor Necrosis Factor in the Mucosa of Patients With Inflammatory Bowel Disease. *Gastroenterology* **149**:1564-1574.e3, W.B. Saunders.
- Bigg HF, Morrison CJ, Butler GS, Bogoyevitch MA, Wang Z, Soloway PD, and Overall CM (2001) Tissue inhibitor of metalloproteinases-4 inhibits but does not support the activation of gelatinase a via efficient inhibition of membrane type 1-matrix metalloproteinase. *Cancer Res* **61**.
- Bissett D, O’Byrne KJ, von Pawel J, Gatzemeier U, Price A, Nicolson M, Mercier R, Mazabel E, Penning C, Zhang MH, Collier MA, and Shepherd FA (2005) Phase III Study of Matrix Metalloproteinase Inhibitor Prinomastat in Non–Small-Cell Lung Cancer. *J Clin Oncol* **23**:842–849.
- Blanco MJ, Rodríguez-Martín I, Learte AIR, Clemente C, Montalvo MG, Seiki M, Arroyo AG, and Sánchez-Camacho C (2017) Developmental expression of membrane type 4-matrix metalloproteinase (Mt4-mmp/Mmp17) in the mouse embryo. *PLoS One* **12**:e0184767.
- Bobadilla M, Sáinz N, Rodríguez JA, Abizanda G, Orbe J, de Martino A, García Verdugo JM, Páramo JA, Prósper F, and Pérez-Ruiz A (2014) MMP-10 Is Required for Efficient Muscle Regeneration in Mouse Models of Injury and Muscular Dystrophy. *Stem Cells* **32**:447–461.
- Boire A, Covic L, Agarwal A, Jacques S, Sherifi S, and Kuliopulos A (2005) PAR1 Is a Matrix Metalloprotease-1 Receptor that Promotes Invasion and Tumorigenesis of Breast Cancer Cells. *Cell* **120**:303–313.
- Bojic S, Kotur-Stevuljevic J, Aleksic A, Gacic J, Memon L, and Simic-Ogrizovic S (2018) Matrix Metalloproteinase-9 and Tissue Inhibitor of Matrix Metalloproteinase-1 in Sepsis after Major Abdominal Surgery. *Dis Markers* **2018**:1–10.
- Bond M, Baker AH, and Newby AC (1999) Nuclear Factor κ B Activity Is Essential for Matrix Metalloproteinase-1 and -3 Upregulation in Rabbit Dermal Fibroblasts. *Biochem Biophys Res Commun* **264**:561–567.

- Bonnans C, Chou J, and Werb Z (2014) Remodelling the extracellular matrix in development and disease. *Nat Rev Mol Cell Biol* **15**:786–801.
- Boon L, Ugarte-Berzal E, Martens E, Fiten P, Vandooren J, Janssens R, Blanter M, Yu K, Boon M, Struyf S, Proost P, and Opdenakker G (2021) Citrullination as a novel posttranslational modification of matrix metalloproteinases. *Matrix Biol* **95**:68–83.
- Boon L, Ugarte-Berzal E, Vandooren J, and Opdenakker G (2016) Glycosylation of matrix metalloproteinases and tissue inhibitors: present state, challenges and opportunities. *Biochem J* **473**:1471–1482.
- Bordenave T, Helle M, Beau F, Georgiadis D, Tepshi L, Bernes M, Ye Y, Levenez L, Poquet E, Nozach H, Razavian M, Toczek J, Stura EA, Dive V, Sadeghi MM, and Devel L (2016) Synthesis and in Vitro and in Vivo Evaluation of MMP-12 Selective Optical Probes. *Bioconjug Chem*, doi: 10.1021/acs.bioconjchem.6b00377.
- Bordoli MR, Yum J, Breitkopf SB, Thon JN, Italiano JE, Xiao J, Worby C, Wong S-K, Lin G, Edenius M, Keller TL, Asara JM, Dixon JE, Yeo C-Y, and Whitman M (2014) A Secreted Tyrosine Kinase Acts in the Extracellular Environment. *Cell* **158**:1033–1044.
- Bozzelli PL, Yin T, Avdoshina V, Mocchetti I, Conant KE, and Maguire-Zeiss KA (2019) HIV-1 Tat promotes astrocytic release of CCL2 through MMP/PAR-1 signaling. *Glia* **67**:glia.23642.
- Brandes N, Schmitt S, and Jakob U (2009) Thiol-Based Redox Switches in Eukaryotic Proteins. *Antioxid Redox Signal* **11**:997–1014.
- Brassart-Pasco S, Brézillon S, Brassart B, Ramont L, Oudart J-B, and Monboisse JC (2020) Tumor Microenvironment: Extracellular Matrix Alterations Influence Tumor Progression. *Front Oncol* **10**.
- Brauer R, Tureckova J, Kanchev I, Khoylou M, Skarda J, Prochazka J, Spoutil F, Beck IM, Zbodakova O, Kasperek P, Korinek V, Chalupsky K, Karhu T, Herzig K-H, Hajdich M, Gregor M, and Sedlacek R (2016) MMP-19 deficiency causes aggravation of colitis due to defects in innate immune cell function. *Mucosal Immunol* **9**:974–985.
- Bregant S, Huillet C, Devel L, Dabert-Gay A-S, Beau F, Thai R, Czarny B, Yiotakis A, and Dive V (2009) Detection of Matrix Metalloproteinase Active Forms in Complex Proteomes: Evaluation of Affinity versus Photoaffinity Capture. *J Proteome Res* **8**:2484–2494.
- Bremer C, Tung C-H, and Weissleder R (2001) In vivo molecular target assessment of matrix metalloproteinase inhibition. *Nat Med* **7**:743–748.
- Brew K, Dinakarandian D, and Nagase H (2000) Tissue inhibitors of metalloproteinases: evolution, structure and function. *Biochim Biophys Acta - Protein Struct Mol Enzymol* **1477**:267–283.
- Brew K, and Nagase H (2010) The tissue inhibitors of metalloproteinases (TIMPs): An ancient family with structural and functional diversity. *Biochim Biophys Acta - Mol Cell Res* **1803**:55–71.
- Brkic M, Balusu S, Libert C, and Vandenbroucke RE (2015) Friends or Foes: Matrix Metalloproteinases and Their Multifaceted Roles in Neurodegenerative Diseases. *Mediators Inflamm* **2015**:1–27.
- Bugno M, Witek B, Bereta J, Bereta M, Edwards DR, and Kordula T (1999) Reprogramming of TIMP-1 and TIMP-3 expression profiles in brain microvascular endothelial cells and astrocytes in response to proinflammatory cytokines. *FEBS Lett* **448**:9–14.
- Butler GS, and Overall CM (2009) Updated Biological Roles for Matrix Metalloproteinases and New “Intracellular” Substrates Revealed by Degradomics. *Biochemistry* **48**:10830–10845.

- Cabral-Pacheco GA, Garza-Veloz I, Castruita-De la Rosa C, Ramirez-Acuña JM, Perez-Romero BA, Guerrero-Rodriguez JF, Martinez-Avila N, and Martinez-Fierro ML (2020) The Roles of Matrix Metalloproteinases and Their Inhibitors in Human Diseases. *Int J Mol Sci* **21**:9739.
- Calander A-M, Starckx S, Opdenakker G, Bergin P, Quiding-Järbrink M, and Tarkowski A (2006) Matrix metalloproteinase-9 (gelatinase B) deficiency leads to increased severity of Staphylococcus aureus-triggered septic arthritis. *Microbes Infect* **8**:1434–1439.
- Campbell LG, Ramachandran S, Liu W, Shipley JM, Itohara S, Rogers JG, Moazami N, Senior RM, and Jaramillo A (2005) Different Roles for Matrix Metalloproteinase-2 and Matrix Metalloproteinase-9 in the Pathogenesis of Cardiac Allograft Rejection. *Am J Transplant* **5**:517–528.
- Cao S, Li Z, Zhao J, Chen M, and Ma N (2018) Rational Engineering a Multichannel Upconversion Sensor for Multiplex Detection of Matrix Metalloproteinase Activities. *ACS Sensors* **3**:1522–1530.
- Casalini F, Fugazza L, Esposito G, Cabella C, Brioschi C, Cordaro A, D’Angeli L, Bartoli A, Filannino AM, Gringeri C V., Longo DL, Muzio V, Nuti E, Orlandini E, Figlia G, Quattrini A, Tei L, Digilio G, Rossello A, and Maiocchi A (2013) Synthesis and Preliminary Evaluation in Tumor Bearing Mice of New 18 F-Labeled Arylsulfone Matrix Metalloproteinase Inhibitors as Tracers for Positron Emission Tomography. *J Med Chem* **56**:2676–2689.
- Castaneda FE, Walia B, Vijay-Kumar M, Patel NR, Roser S, Kolachala VL, Rojas M, Wang L, Oprea G, Garg P, Gewirtz AT, Roman J, Merlin D, and Sitaraman S V. (2005) Targeted Deletion of Metalloproteinase 9 Attenuates Experimental Colitis in Mice: Central Role of Epithelial-Derived MMP. *Gastroenterology* **129**:1991–2008.
- Caterina JJ, Skobe Z, Shi J, Ding Y, Simmer JP, Birkedal-Hansen H, and Bartlett JD (2002) Enamelysin (Matrix Metalloproteinase 20)-deficient Mice Display an Amelogenesis Imperfecta Phenotype. *J Biol Chem* **277**:49598–49604.
- Cauwe B, and Opdenakker G (2010) Intracellular substrate cleavage: a novel dimension in the biochemistry, biology and pathology of matrix metalloproteinases. *Crit Rev Biochem Mol Biol* **45**:351–423.
- Cauwe B, Van den Steen PE, and Opdenakker G (2007) The Biochemical, Biological, and Pathological Kaleidoscope of Cell Surface Substrates Processed by Matrix Metalloproteinases. *Crit Rev Biochem Mol Biol* **42**:113–185.
- Cha H, Kopetzki E, Huber R, Lanzendörfer M, and Brandstetter H (2002) Structural Basis of the Adaptive Molecular Recognition by MMP9. *J Mol Biol* **320**:1065–1079.
- Chan EWS, Chattopadhyaya S, Panicker RC, Huang X, and Yao SQ (2004) Developing Photoactive Affinity Probes for Proteomic Profiling: Hydroxamate-based Probes for Metalloproteases. *J Am Chem Soc* **126**:14435–14446.
- Chandler S, Miller K., Clements J., Lury J, Corkill D, Anthony DC., Adams S., and Gearing AJ. (1997) Matrix metalloproteinases, tumor necrosis factor and multiple sclerosis: an overview. *J Neuroimmunol* **72**:155–161.
- Chaudhry N, and Duggal AK (2014) Sepsis Associated Encephalopathy. *Adv Med* **2014**:1–16.
- Chen P, Abacherli LE, Nadler ST, Wang Y, Li Q, and Parks WC (2009) MMP7 Shedding of Syndecan-1 Facilitates Re-Epithelialization by Affecting $\alpha 2\beta 1$ Integrin Activation. *PLoS One* **4**:e6565.
- Chen P, McGuire JK, Hackman RC, Kim K-H, Black RA, Poindexter K, Yan W, Liu P, Chen

- AJ, Parks WC, and Madtes DK (2008) Tissue Inhibitor of Metalloproteinase-1 Moderates Airway Re-Epithelialization by Regulating Matrilysin Activity. *Am J Pathol* **172**:1256–1270.
- Chen S, Cui J, Jiang T, Olson ES, Cai Q-Y, Yang M, Wu W, Guthrie JM, Robertson J, Lipton SA, Ma L, Tsien RY, and Gu Z (2017) Gelatinase activity imaged by activatable cell-penetrating peptides in cell-based and in vivo models of stroke. *J Cereb Blood Flow Metab* **37**:188–200.
- Chen Y, Chen L, Xing C, Deng G, Zeng F, Xie T, Gu L, and Yang H (2020) The risk of rheumatoid arthritis among patients with inflammatory bowel disease: a systematic review and meta-analysis. *BMC Gastroenterol* **20**:192.
- Cherne MD, Cole AL, Newberry L, Schmidt-Owens M, Deichen M, and Cole AM (2020) Matrix Metalloproteinases Expressed in Response to Bacterial Vaginosis Disrupt the Endocervical Epithelium, Increasing Transmigration of HIV. *Infect Immun* **88**:1–15.
- Cheung C, Marchant D, Walker EKY, Luo Z, Zhang J, Yanagawa B, Rahmani M, Cox J, Overall C, Senior RM, Luo H, and McManus BM (2008) Ablation of Matrix Metalloproteinase-9 Increases Severity of Viral Myocarditis in Mice. *Circulation* **117**:1574–1582.
- Cheung P-Y, Sawicki G, Wozniak M, Wang W, Radomski MW, and Schulz R (2000) Matrix Metalloproteinase-2 Contributes to Ischemia-Reperfusion Injury in the Heart. *Circulation* **101**:1833–1839.
- Chin JR, Murphy G, and Werb Z (1985) Stromelysin, a connective tissue-degrading metalloendopeptidase secreted by stimulated rabbit synovial fibroblasts in parallel with collagenase. Biosynthesis, isolation, characterization, and substrates. *J Biol Chem* **260**:12367–12376.
- Chiocca EA, Yu JS, Lukas R V., Solomon IH, Ligon KL, Nakashima H, Triggs DA, Reardon DA, Wen P, Stopa BM, Naik A, Rudnick J, Hu JL, Kumthekar P, Yamini B, Buck JY, Demars N, Barrett JA, Gelb AB, Zhou J, Lebel F, and Cooper LNJ (2019) Regulatable interleukin-12 gene therapy in patients with recurrent high-grade glioma: Results of a phase 1 trial. *Sci Transl Med* **11**.
- Choi DH, Kim E-M, Son HJ, Joh TH, Kim YS, Kim D, Flint Beal M, and Hwang O (2008) A novel intracellular role of matrix metalloproteinase-3 during apoptosis of dopaminergic cells. *J Neurochem* **106**:405–415.
- Choi J, Kim S, Yoo D, Shin T-H, Kim H, Gomes MD, Kim SH, Pines A, and Cheon J (2017) Distance-dependent magnetic resonance tuning as a versatile MRI sensing platform for biological targets. *Nat Mater* **16**:537–542.
- Chopra S, Overall CM, and Dufour A (2019) Matrix metalloproteinases in the CNS: interferons get nervous. *Cell Mol Life Sci* **76**:3083–3095.
- Chun T-H, Sabeh F, Ota I, Murphy H, McDonagh KT, Holmbeck K, Birkedal-Hansen H, Allen ED, and Weiss SJ (2004) MT1-MMP-dependent neovessel formation within the confines of the three-dimensional extracellular matrix. *J Cell Biol* **167**:757–767.
- Chung L, Dinakarandian D, Yoshida N, Lauer-Fields JL, Fields GB, Visse R, and Nagase H (2004) Collagenase unwinds triple-helical collagen prior to peptide bond hydrolysis. *EMBO J* **23**:3020–3030.
- Clements JM, Cossins JA, Wells GMA, Corkill DJ, Helfrich K, Wood LM, Pigott R, Stabler G, Ward GA, Gearing AJH, and Miller KM (1997) Matrix metalloproteinase expression during experimental autoimmune encephalomyelitis and effects of a combined matrix metalloproteinase and tumour necrosis factor- α inhibitor. *J Neuroimmunol* **74**:85–94.

- Clements KM, Flannelly JK, Tart J, Brockbank SM V, Wardale J, Freeth J, Parker AE, and Newham P (2011) Matrix metalloproteinase 17 is necessary for cartilage aggrecan degradation in an inflammatory environment. *Ann Rheum Dis* **70**:683–689.
- Cobos-Correa A, Trojanek JB, Diemer S, Mall MA, and Schultz C (2009) Membrane-bound FRET probe visualizes MMP12 activity in pulmonary inflammation. *Nat Chem Biol* **5**:628–630.
- Collier IE, Wilhelm SM, Eisen AZ, Marmer BL, Grant GA, Seltzer JL, Kronberger A, He CS, Bauer EA, and Goldberg GI (1988) H-ras oncogene-transformed human bronchial epithelial cells (TBE-1) secrete a single metalloprotease capable of degrading basement membrane collagen. *J Biol Chem* **263**:6579–6587.
- Conant K, Hillaire CS, Anderson C, Galey D, Wang J, and Nath A (2004) Human immunodeficiency virus type 1 Tat and methamphetamine affect the release and activation of matrix-degrading proteinases. *J Neurovirol* **10**:21–28.
- Conant K, Hillaire C St., Nagase H, Visse R, Gary D, Haughey N, Anderson C, Turchan J, and Nath A (2004) Matrix Metalloproteinase 1 Interacts with Neuronal Integrins and Stimulates Dephosphorylation of Akt. *J Biol Chem* **279**:8056–8062.
- Conway JG, Wakefield JA, Brown RH, Marron BE, Sekut L, Stimpson SA, McElroy A, Menius JA, Jeffreys JJ, and Clark RL (1995) Inhibition of cartilage and bone destruction in adjuvant arthritis in the rat by a matrix metalloproteinase inhibitor. *J Exp Med* **182**:449–457.
- Correia AL, Mori H, Chen EI, Schmitt FC, and Bissell MJ (2013) The hemopexin domain of MMP3 is responsible for mammary epithelial invasion and morphogenesis through extracellular interaction with HSP90. *Genes Dev* **27**:805–817.
- Coward S, Clement F, Williamson T, Hazlewood G, Ng S, Heitman S, Seow C, Panaccione R, Ghosh S, and Kaplan GG (2015) The Rising Burden of Inflammatory Bowel Disease in North America from 2015 to 2025: A Predictive Model. *Am J Gastroenterol* **110**:S829.
- Cox JH, Starr AE, Kappelhoff R, Yan R, Roberts CR, and Overall CM (2010) Matrix metalloproteinase 8 deficiency in mice exacerbates inflammatory arthritis through delayed neutrophil apoptosis and reduced caspase 11 expression. *Arthritis Rheum* **62**:3645–3655.
- Cox TR (2021) The matrix in cancer. *Nat Rev Cancer* **21**:217–238.
- Crawford HC, Fingleton B, Gustavson MD, Kurpios N, Wagenaar RA, Hassell JA, and Matrisian LM (2001) The PEA3 Subfamily of Ets Transcription Factors Synergizes with β -Catenin–LEF-1 To Activate Matrilysin Transcription in Intestinal Tumors. *Mol Cell Biol* **21**:1370–1383.
- Crawford HC, Scoggins CR, Washington MK, Matrisian LM, and Leach SD (2002) Matrix metalloproteinase-7 is expressed by pancreatic cancer precursors and regulates acinar-to-ductal metaplasia in exocrine pancreas. *J Clin Invest* **109**:1437–1444.
- Cuzner ML, Gveric D, Strand C, Loughlin AJ, Paemen L, Opdenakker G, and Newcombe J (1996) The Expression of Tissue-type Plasminogen Activator, Matrix Metalloproteases and Endogenous Inhibitors in the Central Nervous System in Multiple Sclerosis. *J Neuropathol Exp Neurol* **55**:1194–1204.
- Czarny B, Stura EA, Devel L, Vera L, Cassar-Lajeunesse E, Beau F, Calderone V, Fragai M, Luchinat C, and Dive V (2013) Molecular Determinants of a Selective Matrix Metalloprotease-12 Inhibitor: Insights from Crystallography and Thermodynamic Studies. *J Med Chem* **56**:1149–1159.
- D’Haens G, Kelly O, Battat R, Silverberg MS, Laharie D, Louis E, Savarino E, Bodini G, Yarur A, Boland BS, Afif W, Li X, Hale M, Ho J, Kondragunta V, Huang B, Kuy C, Okada L,

- Hester KD, Bray KR, Mimms L, Jain A, Singh S, Collins A, Valasek MA, Sandborn WJ, Vermeire S, and Dulai PS (2020) Development and Validation of a Test to Monitor Endoscopic Activity in Patients With Crohn's Disease Based on Serum Levels of Proteins. *Gastroenterology* **158**:515-526.e10.
- Dabert-Gay A-S, Czarny B, Devel L, Beau F, Lajeunesse E, Bregant S, Thai R, Yiotakis A, and Dive V (2008) Molecular Determinants of Matrix Metalloproteinase-12 Covalent Modification by a Photoaffinity Probe. *J Biol Chem* **283**:31058–31067.
- Dabert-Gay A-S, Czarny B, Lajeunesse E, Thai R, Nagase H, and Dive V (2009) Covalent Modification of Matrix Metalloproteinases by a Photoaffinity Probe: Influence of Nucleophilicity and Flexibility of the Residue in Position 241. *Bioconjug Chem* **20**:367–375.
- Daghighi S, Sjollem J, Dijkstra R, Jaspers V, Zaat S, van der Mei H, and Busscher H (2014) Real-time quantification of matrix metalloproteinase and integrin $\alpha\beta 3$ expression during biomaterial-associated infection in a murine model. *Eur Cells Mater* **27**:26–38.
- Dal-Pizzol F, Rojas HA, dos Santos EM, Vuolo F, Constantino L, Feier G, Pasquali M, Comim CM, Petronilho F, Gelain DP, Quevedo J, Moreira JCF, and Ritter C (2013) Matrix Metalloproteinase-2 and Metalloproteinase-9 Activities are Associated with Blood–Brain Barrier Dysfunction in an Animal Model of Severe Sepsis. *Mol Neurobiol* **48**:62–70.
- Daley WP, and Yamada KM (2013) ECM-modulated cellular dynamics as a driving force for tissue morphogenesis. *Curr Opin Genet Dev* **23**:408–414.
- Dali-Youcef N, Hnia K, Blaise S, Messaddeq N, Blanc S, Postic C, Valet P, Tomasetto C, and Rio M-C (2016) Matrix metalloproteinase 11 protects from diabetes and promotes metabolic switch. *Sci Rep* **6**:25140.
- Dallas SL, Rosser JL, Mundy GR, and Bonewald LF (2002) Proteolysis of Latent Transforming Growth Factor- β (TGF- β)-binding Protein-1 by Osteoclasts. *J Biol Chem* **277**:21352–21360.
- Das N, Benko C, Gill SE, and Dufour A (2021) The Pharmacological TAILS of Matrix Metalloproteinases and Their Inhibitors. *Pharmaceuticals* **14**:31.
- David A, Steer D, Bregant S, Devel L, Makaritis A, Beau F, Yiotakis A, and Dive V (2007) Cross-Linking Yield Variation of a Potent Matrix Metalloproteinase Photoaffinity Probe and Consequences for Functional Proteomics. *Angew Chemie Int Ed* **46**:3275–3277.
- De Palma M, Biziato D, and Petrova T V. (2017) Microenvironmental regulation of tumour angiogenesis. *Nat Rev Cancer* **17**:457–474.
- Dean RA, Cox JH, Bellac CL, Doucet A, Starr AE, and Overall CM (2008) Macrophage-specific metalloelastase (MMP-12) truncates and inactivates ELR + CXC chemokines and generates CCL2, -7, -8, and -13 antagonists: Potential role of the macrophage in terminating polymorphonuclear leukocyte influx. *Blood* **112**:3455–3464.
- DeClerck YA, Yean TD, Lu HS, Ting J, and Langley KE (1991) Inhibition of autoproteolytic activation of interstitial procollagenase by recombinant metalloproteinase inhibitor MI/TIMP-2. *J Biol Chem* **266**:3893–3899.
- DeCoux A, Lindsey ML, Villarreal F, Garcia RA, and Schulz R (2014) Myocardial matrix metalloproteinase-2: inside out and upside down. *J Mol Cell Cardiol* **77**:64–72.
- Deguchi J-O, Aikawa E, Libby P, Vachon JR, Inada M, Krane SM, Whittaker P, and Aikawa M (2005) Matrix Metalloproteinase-13/Collagenase-3 Deletion Promotes Collagen Accumulation and Organization in Mouse Atherosclerotic Plaques. *Circulation* **112**:2708–2715.
- Deguchi J, Aikawa M, Tung C-H, Aikawa E, Kim D-E, Ntziachristos V, Weissleder R, and

- Libby P (2006) Inflammation in Atherosclerosis. *Circulation* **114**:55–62.
- Devel L, Czarny B, Beau F, Georgiadis D, Stura E, and Dive V (2010) Third generation of matrix metalloprotease inhibitors: Gain in selectivity by targeting the depth of the S1' cavity. *Biochimie* **92**:1501–1508.
- Devel L, Rogakos V, David A, Makaritis A, Beau F, Cuniasse P, Yiotakis A, and Dive V (2006) Development of Selective Inhibitors and Substrate of Matrix Metalloproteinase-12. *J Biol Chem* **281**:11152–11160.
- Devy L, and Dransfield DT (2011) New Strategies for the Next Generation of Matrix-Metalloproteinase Inhibitors: Selectively Targeting Membrane-Anchored MMPs with Therapeutic Antibodies. *Biochem Res Int* **2011**:1–11.
- Devy L, Huang L, Naa L, Yanamandra N, Pieters H, Frans N, Chang E, Tao Q, Vanhove M, Lejeune A, van Gool R, Sexton DJ, Kuang G, Rank D, Hogan S, Pazmany C, Ma YL, Schoonbroodt S, Nixon AE, Ladner RC, Hoet R, Henderikx P, TenHoor C, Rabbani SA, Valentino ML, Wood CR, and Dransfield DT (2009) Selective Inhibition of Matrix Metalloproteinase-14 Blocks Tumor Growth, Invasion, and Angiogenesis. *Cancer Res* **69**:1517–1526.
- Digilio G, Tuccinardi T, Casalini F, Cassino C, Dias DM, Geraldes CFGC, Catanzaro V, Maiocchi A, and Rossello A (2014) Study of the binding interaction between fluorinated matrix metalloproteinase inhibitors and Human Serum Albumin. *Eur J Med Chem* **79**:13–23.
- Ding S, Blue RE, Morgan DR, and Lund PK (2014) Comparison of Multiple Enzyme Activatable Near-Infrared Fluorescent Molecular Probes for Detection and Quantification of Inflammation in Murine Colitis Models. *Inflamm Bowel Dis* **20**:363–377.
- Dix MM, Simon GM, Wang C, Okerberg E, Patricelli MP, and Cravatt BF (2012) Functional Interplay between Caspase Cleavage and Phosphorylation Sculpts the Apoptotic Proteome. *Cell* **150**:426–440.
- Dofara SG, Chang S-L, and Diorio C (2020) Gene Polymorphisms and Circulating Levels of MMP-2 and MMP-9: A Review of Their Role in Breast Cancer Risk. *Anticancer Res* **40**:3619–3631.
- Dolin HH, Papadimos TJ, Chen X, and Pan ZK (2019) Characterization of Pathogenic Sepsis Etiologies and Patient Profiles: A Novel Approach to Triage and Treatment. *Microbiol Insights* **12**:117863611882508.
- Drewes AM, Olesen AE, Farmer AD, Szigethy E, Rebours V, and Olesen SS (2020) Gastrointestinal pain. *Nat Rev Dis Prim* **6**:1.
- Drygiannakis I, Valatas V, Sfakianaki O, Bourikas L, Manousou P, Kambas K, Ritis K, Kolios G, and Kouroumalis E (2013) Proinflammatory cytokines induce crosstalk between colonic epithelial cells and subepithelial myofibroblasts: Implication in intestinal fibrosis. *J Crohn's Colitis* **7**:286–300.
- Dubois B, D'Hooghe MB, De Lepeleire K, Ketelaer P, Opdenakker G, and Carton H (1998) Toxicity in a double-blind, placebo-controlled pilot trial with D-penicillamine and metacycline in secondary progressive multiple sclerosis. *Mult Scler J* **4**:74–78.
- Dubois B, Masure S, Hurtenbach U, Paemen L, Heremans H, van den Oord J, Sciot R, Meinhardt T, Hämmerling G, Opdenakker G, and Arnold B (1999) Resistance of young gelatinase B-deficient mice to experimental autoimmune encephalomyelitis and necrotizing tail lesions. *J Clin Invest* **104**:1507–1515.
- Dudani JS, Buss CG, Akana RTK, Kwong GA, and Bhatia SN (2016) Sustained-Release

- Synthetic Biomarkers for Monitoring Thrombosis and Inflammation Using Point-of-Care Compatible Readouts. *Adv Funct Mater* **26**:2919–2928.
- Duellman T, Burnett J, and Yang J (2015) Functional Roles of N-Linked Glycosylation of Human Matrix Metalloproteinase 9. *Traffic* **16**:1108–1126.
- Dufour A (2015) Degradomics of matrix metalloproteinases in inflammatory diseases. *Front Biosci* **7**:430.
- Dufour A, Bellac CL, Eckhard U, Solis N, Klein T, Kappelhoff R, Fortelny N, Jobin P, Rozmus J, Mark J, Pavlidis P, Dive V, Barbour SJ, and Overall CM (2018) C-terminal truncation of IFN- γ inhibits proinflammatory macrophage responses and is deficient in autoimmune disease. *Nat Commun* **9**:2416, Nature Publishing Group.
- Dufour A, and Overall CM (2013) Missing the target: matrix metalloproteinase antitargets in inflammation and cancer. *Trends Pharmacol Sci* **34**:233–242.
- Dufour A, and Overall CM (2015) Subtracting Matrix Out of the Equation: New Key Roles of Matrix Metalloproteinases in Innate Immunity and Disease, in *Matrix Metalloproteinase Biology* pp 131–152, John Wiley & Sons, Inc, Hoboken, NJ, USA.
- Dufour Antoine, Sampson NS, Li J, Kuscu C, Rizzo RC, DeLeon JL, Zhi J, Jaber N, Liu E, and Zucker S (2011) Small-molecule anticancer compounds selectively target the hemopexin domain of matrix metalloproteinase-9. *Cancer Res* **71**:4977–4988, AACR.
- Dufour A., Sampson NS, Li J, Kuscu C, Rizzo RC, DeLeon JL, Zhi J, Jaber N, Liu E, Zucker S, and Cao J (2011) Small-molecule anticancer compounds selectively target the hemopexin domain of matrix metalloproteinase-9. *Cancer Res* **71**.
- Dufour A, Sampson NS, Zucker S, and Cao J (2008) Role of the hemopexin domain of matrix metalloproteinases in cell migration. *J Cell Physiol* **217**:643–651.
- Dufour A, Zucker S, Sampson NS, Kuscu C, and Cao J (2010) Role of matrix metalloproteinase-9 dimers in cell migration: Design of inhibitory peptides. *J Biol Chem* **285**:35944–35956.
- Dunsmore SE, Saarialho-Kere UK, Roby JD, Wilson CL, Matrisian LM, Welgus HG, and Parks WC (1998) Matrilysin expression and function in airway epithelium. *J Clin Invest* **102**:1321–1331.
- Eccles SA, Box GM, Court WJ, Bone EA, Thomas W, and Brown PD (1996) Control of lymphatic and hematogenous metastasis of a rat mammary carcinoma by the matrix metalloproteinase inhibitor batimastat (BB-94). *Cancer Res* **56**.
- Eckhard U, Huesgen PF, Schilling O, Bellac CL, Butler GS, Cox JH, Dufour A, Goebeler V, Kappelhoff R, auf dem Keller U, Klein T, Lange PF, Marino G, Morrison CJ, Prudova A, Rodriguez D, Starr AE, Wang Y, and Overall CM (2016) Active site specificity profiling datasets of matrix metalloproteinases (MMPs) 1, 2, 3, 7, 8, 9, 12, 13 and 14. *Data Br* **7**:299–310.
- Eckhard U, Huesgen PF, Schilling O, Bellac CL, Butler GS, Cox JH, Dufour A, Goebeler V, Kappelhoff R, Keller U auf dem, Klein T, Lange PF, Marino G, Morrison CJ, Prudova A, Rodriguez D, Starr AE, Wang Y, and Overall CM (2016) Active site specificity profiling of the matrix metalloproteinase family: Proteomic identification of 4300 cleavage sites by nine MMPs explored with structural and synthetic peptide cleavage analyses. *Matrix Biol* **49**:37–60.
- Egeblad M, and Werb Z (2002) New functions for the matrix metalloproteinases in cancer progression. *Nat Rev Cancer* **2**:161–174.
- Eguchi T, Kubota S, Kawata K, Mukudai Y, Uehara J, Ohgawara T, Ibaragi S, Sasaki A, Kuboki T, and Takigawa M (2008) Novel Transcription Factor-Like Function of Human Matrix

- Metalloproteinase 3 Regulating the CTGF/CCN2 Gene. *Mol Cell Biol* **28**:2391–2413.
- El Mouhawess A, Hammoud A, Zoghbi M, Hallit S, Haddad C, El Haddad K, El Khoury S, Tannous J, Obeid S, Halabi MA, and Mammari N (2020) Relationship between Toxoplasma gondii seropositivity and schizophrenia in the Lebanese population: potential implication of genetic polymorphism of MMP-9. *BMC Psychiatry* **20**:264.
- Endo K, Takino T, Miyamori H, Kinsen H, Yoshizaki T, Furukawa M, and Sato H (2003) Cleavage of Syndecan-1 by Membrane Type Matrix Metalloproteinase-1 Stimulates Cell Migration. *J Biol Chem* **278**:40764–40770.
- Engel CK, Pirard B, Schimanski S, Kirsch R, Habermann J, Klingler O, Schlotte V, Weithmann KU, and Wendt KU (2005) Structural Basis for the Highly Selective Inhibition of MMP-13. *Chem Biol* **12**:181–189.
- Epstein FH, and Weiss SJ (1989) Tissue Destruction by Neutrophils. *N Engl J Med* **320**:365–376.
- Evans MJ, and Cravatt BF (2006) Mechanism-Based Profiling of Enzyme Families. *Chem Rev* **106**:3279–3301.
- Faber HR, Groom CR, Baker HM, Morgan WT, Smith A, and Baker EN (1995) 1.8 Å crystal structure of the C-terminal domain of rabbit serum haemopexin. *Structure* **3**:551–559.
- Fallata AM, Wyatt RA, Levesque JM, Dufour A, Overall CM, and Crawford BD (2019) Intracellular localization in zebrafish muscle and conserved sequence features suggest roles for gelatinase a moonlighting in sarcomere maintenance. *Biomedicines* **7**.
- Fan X, Hughes BG, Ali MAM, Chan BYH, Launier K, and Schulz R (2016) Matrix metalloproteinase-2 in oncostatin M-induced sarcomere degeneration in cardiomyocytes. *Am J Physiol Circ Physiol* **311**:H183–H189.
- Fang Q, Zhou C, and Nandakumar KS (2020) Molecular and Cellular Pathways Contributing to Joint Damage in Rheumatoid Arthritis. *Mediators Inflamm* **2020**:1–20.
- Fanjul-Fernández M, Folgueras AR, Fueyo A, Balbín M, Suárez MF, Fernández-García MS, Shapiro SD, Freije JMP, and López-Otín C (2018) Matrix metalloproteinase Mmp-1a is dispensable for normal growth and fertility in mice and promotes lung cancer progression by modulating inflammatory responses. *J Biol Chem* **293**:11970.
- Fanjul-Fernández M, Folgueras AR, Fueyos A, Balbín M, Suárez MF, Fernández-García MS, Shapiro SD, Freije JMP, and López-Otín C (2013) Matrix metalloproteinase Mmp-1a is dispensable for normal growth and fertility in mice and promotes lung cancer progression by modulating inflammatory responses. *J Biol Chem* **288**.
- Faust A, Waschku B, Waldeck J, Höltke C, Breyholz H-J, Wagner S, Kopka K, Heindel W, Schäfers M, and Bremer C (2008) Synthesis and Evaluation of a Novel Fluorescent Photoprobe for Imaging Matrix Metalloproteinases. *Bioconjug Chem* **19**:1001–1008.
- Feinberg TY, Rowe RG, Saunders TL, and Weiss SJ (2016) Functional roles of MMP14 and MMP15 in early postnatal mammary gland development. *Development* **143**.
- Feinberg TY, Zheng H, Liu R, Wicha MS, Yu SM, and Weiss SJ (2018) Divergent Matrix-Remodeling Strategies Distinguish Developmental from Neoplastic Mammary Epithelial Cell Invasion Programs. *Dev Cell* **47**:145-160.e6.
- Feng S, Cen J, Huang Y, Shen H, Yao L, Wang Y, and Chen Z (2011) Matrix Metalloproteinase-2 and -9 Secreted by Leukemic Cells Increase the Permeability of Blood-Brain Barrier by Disrupting Tight Junction Proteins. *PLoS One* **6**:e20599.
- Ferdinandy P, Danial H, Ambrus I, Rothery RA, and Schulz R (2000) Peroxynitrite Is a Major Contributor to Cytokine-Induced Myocardial Contractile Failure. *Circ Res* **87**:241–247.

- Fields GB (2019) The Rebirth of Matrix Metalloproteinase Inhibitors: Moving Beyond the Dogma. *Cells* **8**:984.
- Fingleton B (2008) MMPs as therapeutic targets—Still a viable option? *Semin Cell Dev Biol* **19**:61–68.
- Fitridge R, and Thompson M (2012) *Mechanisms of Vascular Disease*, University of Adelaide Press, Adelaide.
- Foley CJ, Fanjul-Fernández M, Bohm A, Nguyen N, Agarwal A, Austin K, Koukos G, Covic L, López-Otín C, and Kuliopulos A (2014) Matrix metalloprotease 1a deficiency suppresses tumor growth and angiogenesis. *Oncogene* **33**:2264–2272.
- Folgueras AR, Valdes-Sanchez T, Llano E, Menendez L, Baamonde A, Denlinger BL, Belmonte C, Juarez L, Lastra A, Garcia-Suarez O, Astudillo A, Kirstein M, Pendas AM, Farinas I, and Lopez-Otin C (2009) Metalloproteinase MT5-MMP is an essential modulator of neuro-immune interactions in thermal pain stimulation. *Proc Natl Acad Sci* **106**:16451–16456.
- Fontana V, Silva PS, Gerlach RF, and Tanus-Santos JE (2012) Circulating matrix metalloproteinases and their inhibitors in hypertension. *Clin Chim Acta* **413**:656–662.
- Fortelny N, Cox JH, Kappelhoff R, Starr AE, Lange PF, Pavlidis P, and Overall CM (2014) Network Analyses Reveal Pervasive Functional Regulation Between Proteases in the Human Protease Web. *PLoS Biol* **12**:e1001869, Public Library of Science.
- Fortelny N, Yang S, Pavlidis P, Lange PF, and Overall CM (2015) Proteome TopFIND 3.0 with TopFINDER and PathFINDER: Database and analysis tools for the association of protein termini to pre- and post-translational events. *Nucleic Acids Res*, doi: 10.1093/nar/gku1012.
- Friedrich M, Pohin M, and Powrie F (2019) Cytokine Networks in the Pathophysiology of Inflammatory Bowel Disease. *Immunity* **50**:992–1006.
- Frolova AS, Petushkova AI, Makarov VA, Soond SM, and Zamyatnin AA (2020) Unravelling the Network of Nuclear Matrix Metalloproteinases for Targeted Drug Design. *Biology (Basel)* **9**:480.
- Fu X, Kassim SY, Parks WC, and Heinecke JW (2001) Hypochlorous Acid Oxygenates the Cysteine Switch Domain of Pro-matrilysin (MMP-7). *J Biol Chem* **276**:41279–41287.
- Fujimoto K, Nakajima K, and Yaoita Y (2007) Expression of matrix metalloproteinase genes in regressing or remodeling organs during amphibian metamorphosis. *Dev Growth Differ* **49**:131–143.
- Gabrielli A, Avvedimento E V., and Krieg T (2009) Scleroderma. *N Engl J Med* **360**:1989–2003.
- Galloway WA, Murphy G, Sandy JD, Gavrilovic J, Cawston TE, and Reynolds JJ (1983) Purification and characterization of a rabbit bone metalloproteinase that degrades proteoglycan and other connective-tissue components. *Biochem J* **209**:741–752.
- García-Pardo A, and Opdenakker G (2015) Nonproteolytic functions of matrix metalloproteinases in pathology and insights for the development of novel therapeutic inhibitors. *Met Med* **19**.
- Garg P, Rojas M, Ravi A, Bockbrader K, Epstein S, Vijay-Kumar M, Gewirtz AT, Merlin D, and Sitaraman S V. (2006) Selective Ablation of Matrix Metalloproteinase-2 Exacerbates Experimental Colitis: Contrasting Role of Gelatinases in the Pathogenesis of Colitis. *J Immunol* **177**:4103–4112.
- Garland M, Yim JJ, and Bogyo M (2016) A Bright Future for Precision Medicine: Advances in Fluorescent Chemical Probe Design and Their Clinical Application. *Cell Chem Biol* **23**:122–136.
- Gasson JC, Golde DW, Kaufman SE, Westbrook CA, Hewick RM, Kaufman RJ, Wong GG,

- Temple PA, Leary AC, Brown EL, Orr EC, and Clark SC (1985) Molecular characterization and expression of the gene encoding human erythroid-potentiating activity. *Nature* **315**:768–771.
- Gatto C, Rieppi M, Borsotti P, Innocenti S, Ceruti R, Drudis T, Scanziani E, Casazza AM, Taraboletti G, and Giavazzi R (1999) BAY 12-9566, a novel inhibitor of matrix metalloproteinases with antiangiogenic activity. *Clin Cancer Res* **5**.
- Genga KR, and Russell JA (2017) Update of Sepsis in the Intensive Care Unit. *J Innate Immun* **9**:441–455.
- George J, Patal S, Wexler D, Roth A, Sheps D, and Keren G (2005) Circulating matrix metalloproteinase-2 but not matrix metalloproteinase-3, matrix metalloproteinase-9, or tissue inhibitor of metalloproteinase-1 predicts outcome in patients with congestive heart failure. *Am Heart J* **150**:484–487.
- Georgiadis D, and Dive V (2014) Phosphinic Peptides as Potent Inhibitors of Zinc-Metalloproteases, in *Topics in Current Chemistry* pp 1–38.
- Geurink PP, Klein T, Prèly L, Paal K, Leeuwenburgh MA, van der Marel GA, Kauffman HF, Overkleeft HS, and Bischoff R (2010) Design of Peptide Hydroxamate-Based Photoreactive Activity-Based Probes of Zinc-Dependent Metalloproteases. *European J Org Chem* **2010**:2100–2112.
- Geurink PP, Prely LM, van der Marel GA, Bischoff R, and Overkleeft HS (2011) Photoaffinity Labeling in Activity-Based Protein Profiling, in *Topics in Current Chemistry* pp 85–113.
- Gharib SA, Johnston LK, Huizar I, Birkland TP, Hanson J, Wang Y, Parks WC, and Manicone AM (2014) MMP28 promotes macrophage polarization toward M2 cells and augments pulmonary fibrosis. *J Leukoc Biol* **95**:9–18.
- Gharib SA, Manicone AM, and Parks WC (2018) Matrix metalloproteinases in emphysema. *Matrix Biol* **73**:34–51.
- Giavazzi R, Garofalo A, Ferri C, Lucchini V, Bone EA, Chiari S, Brown PD, Nicoletti MI, and Taraboletti G (1998) Batimastat, a synthetic inhibitor of matrix metalloproteinases, potentiates the antitumor activity of cisplatin in ovarian carcinoma xenografts. *Clin Cancer Res* **4**.
- Gijbels K, Galardy RE, and Steinman L (1994) Reversal of experimental autoimmune encephalomyelitis with a hydroxamate inhibitor of matrix metalloproteases. *J Clin Invest* **94**:2177–2182.
- Gijbels K, Masure S, Carton H, and Opdenakker G (1992) Gelatinase in the cerebrospinal fluid of patients with multiple sclerosis and other inflammatory neurological disorders. *J Neuroimmunol* **41**:29–34.
- Gill SE, Huizar I, Bench EM, Sussman SW, Wang Y, Khokha R, and Parks WC (2010) Tissue Inhibitor of Metalloproteinases 3 Regulates Resolution of Inflammation following Acute Lung Injury. *Am J Pathol* **176**:64–73.
- Gill SE, Pape MC, and Leco KJ (2006) Tissue inhibitor of metalloproteinases 3 regulates extracellular matrix—Cell signaling during bronchiole branching morphogenesis. *Dev Biol* **298**:540–554.
- Gohlke U, Gomis-Rüth F-X, Crabbe T, Murphy G, Docherty AJP, and Bode W (1996) The C-terminal (haemopexin-like) domain structure of human gelatinase A (MMP2): structural implications for its function. *FEBS Lett* **378**:126–130.
- Gomis-Rüth FX (2009) Catalytic Domain Architecture of Metzincin Metalloproteases. *J Biol Chem* **284**:15353–15357.

- Gomis-Rüth FX (2003) Structural Aspects of the Metzincin Clan of Metalloendopeptidases. *Mol Biotechnol* **24**:157–202.
- Gomis-Rüth FX, Gohlke U, Betz M, Knäuper V, Murphy G, López-Otín C, and Bode W (1996) The Helping Hand of Collagenase-3 (MMP-13): 2.7 Å Crystal Structure of its C-terminal Haemopexin-like Domain. *J Mol Biol* **264**:556–566.
- Gona K, Toczek J, Ye Y, Sanzida N, Golbazi A, Boodagh P, Salarian M, Jung J-J, Rajendran S, Kukreja G, Wu TL, Devel L, and Sadeghi MM (2020) Hydroxamate-Based Selective Macrophage Elastase (MMP-12) Inhibitors and Radiotracers for Molecular Imaging. *J Med Chem* **63**:15037–15049.
- Goncalves I, Bengtsson E, Colhoun HM, Shore AC, Palombo C, Natali A, Edsfeldt A, Dunér P, Fredrikson GN, Björkbacka H, Östling G, Aizawa K, Casanova F, Persson M, Gooding K, Strain D, Khan F, Looker HC, Adams F, Belch J, Pinnoli S, Venturi E, Kozakova M, Gan L-M, Schnecke V, and Nilsson J (2015) Elevated Plasma Levels of MMP-12 Are Associated With Atherosclerotic Burden and Symptomatic Cardiovascular Disease in Subjects With Type 2 Diabetes. *Arterioscler Thromb Vasc Biol* **35**:1723–1731.
- Gooyit M, Suckow MA, Schroeder VA, Wolter WR, Mobashery S, and Chang M (2012) Selective Gelatinase Inhibitor Neuroprotective Agents Cross the Blood-Brain Barrier. *ACS Chem Neurosci* **3**:730–736.
- Gossage DL, Cieslarová B, Ap S, Zheng H, Xin Y, Lal P, Chen G, Smith V, and Sundry JS (2018) Phase 1b Study of the Safety, Pharmacokinetics, and Disease-related Outcomes of the Matrix Metalloproteinase-9 Inhibitor Andecaliximab in Patients With Rheumatoid Arthritis. *Clin Ther* **40**:156-165.e5.
- Goulas T, Garcia-Ferrer I, Marrero A, Marino-Puertas L, Duquerroy S, and Gomis-Rüth FX (2017) Structural and functional insight into pan-endopeptidase inhibition by α 2-macroglobulins. *Biol Chem* **398**:975–994.
- Gracey E, Vereecke L, McGovern D, Fröhling M, Schett G, Danese S, De Vos M, Van den Bosch F, and Elewaut D (2020) Revisiting the gut–joint axis: links between gut inflammation and spondyloarthritis. *Nat Rev Rheumatol* **16**:415–433.
- Graesser D, Mahooti S, and Madri JA (2000) Distinct roles for matrix metalloproteinase-2 and α 4 integrin in autoimmune T cell extravasation and residency in brain parenchyma during experimental autoimmune encephalomyelitis. *J Neuroimmunol* **109**:121–131.
- Grant BD, and Donaldson JG (2009) Pathways and mechanisms of endocytic recycling. *Nat Rev Mol Cell Biol* **10**:597–608.
- Greene J, Wang M, Liu YE, Raymond LA, Rosen C, and Shi YE (1996) Molecular cloning and characterization of human tissue inhibitor of metalloproteinase 4. *J Biol Chem* **271**.
- Greuter T, and Vavricka SR (2019) Extraintestinal manifestations in inflammatory bowel disease – epidemiology, genetics, and pathogenesis. *Expert Rev Gastroenterol Hepatol* **13**:307–317.
- Gross J, and Lapiere CM (1962) Collagenolytic activity in amphibian tissues: a tissue culture assay. *Proc Natl Acad Sci* **48**:1014–1022.
- Gu Z, Kaul M, Yan B, Kridel SJ, Cui J, Strongin A, Smith JW, Liddington RC, and Lipton SA (2002) S-Nitrosylation of Matrix Metalloproteinases: Signaling Pathway to Neuronal Cell Death. *Science* (80-) **297**:1186–1190.
- Gueders MM, Balbin M, Rocks N, Foidart J-M, Gosset P, Louis R, Shapiro S, Lopez-Otin C, Noël A, and Cataldo DD (2005) Matrix Metalloproteinase-8 Deficiency Promotes Granulocytic Allergen-Induced Airway Inflammation. *J Immunol* **175**:2589–2597.
- Gueders MM, Hirst SJ, Quesada-Calvo F, Paulissen G, Hacha J, Gilles C, Gosset P, Louis R,

- Foidart JM, Lopez-Otin C, Noël A, and Cataldo DD (2010) Matrix metalloproteinase-19 deficiency promotes tenascin-C accumulation and allergen-induced airway inflammation. *Am J Respir Cell Mol Biol* **43**.
- Gyawali B, Ramakrishna K, and Dhamoon AS (2019) Sepsis: The evolution in definition, pathophysiology, and management. *SAGE Open Med* **7**:205031211983504.
- Hadler-Olsen E, Winberg J-O, and Uhlin-Hansen L (2013) Matrix metalloproteinases in cancer: their value as diagnostic and prognostic markers and therapeutic targets. *Tumor Biol* **34**:2041–2051.
- Hamada T, Arima N, Shindo M, Sugama K, and Sasaguri Y (2000) Suppression of adjuvant arthritis of rats by a novel matrix metalloproteinase-inhibitor. *Br J Pharmacol* **131**:1513–1520.
- Haro H, Crawford HC, Fingleton B, MacDougall JR, Shinomiya K, Spengler DM, and Matrisian LM (2000) Matrix metalloproteinase-3-dependent generation of a macrophage chemoattractant in a model of herniated disc resorption. *J Clin Invest* **105**.
- Haro H, Crawford HC, Fingleton B, Shinomiya K, Spengler DM, and Matrisian LM (2000) Matrix metalloproteinase-7-dependent release of tumor necrosis factor- α in a model of herniated disc resorption. *J Clin Invest* **105**:143–150.
- Harris TJ, von Maltzahn G, Derfus AM, Ruoslahti E, and Bhatia SN (2006) Proteolytic Actuation of Nanoparticle Self-Assembly. *Angew Chemie Int Ed* **45**:3161–3165.
- Harrison H, Bennett G, Blakeley D, Brown A, Campbell S, Chen L, Lutz RJ, Pavan S, Rietschoten K van, Teufel D, Park PU, and Lee K (2017) Abstract 5144: BT1718, a novel bicyclic peptide-maytansinoid conjugate targeting MT1-MMP for the treatment of solid tumors: Design of bicyclic peptide and linker selection, in *Experimental and Molecular Therapeutics* pp 5144–5144, American Association for Cancer Research.
- Hasebe T, Hartman R, Fu L, Amano T, and Shi Y-B (2007) Evidence for a cooperative role of gelatinase A and membrane type-1 matrix metalloproteinase during *Xenopus laevis* development. *Mech Dev* **124**:11–22.
- Hautamaki RD, Kobayashi DK, Senior RM, and Shapiro SD (1997) Requirement for Macrophage Elastase for Cigarette Smoke-Induced Emphysema in Mice. *Science (80-)* **277**:2002–2004.
- He CS, Wilhelm SM, Pentland AP, Marmer BL, Grant GA, Eisen AZ, and Goldberg GI (1989) Tissue cooperation in a proteolytic cascade activating human interstitial collagenase. *Proc Natl Acad Sci* **86**:2632–2636.
- Heissig B, Hattori K, Dias S, Friedrich M, Ferris B, Hackett NR, Crystal RG, Besmer P, Lyden D, Moore MAS, Werb Z, and Rafii S (2002) Recruitment of Stem and Progenitor Cells from the Bone Marrow Niche Requires MMP-9 Mediated Release of Kit-Ligand. *Cell* **109**:625–637.
- Heissig B, Rafii S, Akiyama H, Ohki Y, Sato Y, Rafael T, Zhu Z, Hicklin DJ, Okumura K, Ogawa H, Werb Z, and Hattori K (2005) Low-dose irradiation promotes tissue revascularization through VEGF release from mast cells and MMP-9-mediated progenitor cell mobilization. *J Exp Med* **202**:739–750.
- Hensley H, Cooper HS, Chang W-CL, and Clapper ML (2017) Imaging Matrix Metalloproteases in Spontaneous Colon Tumors: Validation by Correlation with Histopathology, in *Methods in Molecular Biology* pp 245–255.
- Heymans S, Lupu F, Terclavers S, Vanwetswinkel B, Herbert J-M, Baker A, Collen D, Carmeliet P, and Moons L (2005) Loss or Inhibition of uPA or MMP-9 Attenuates LV

- Remodeling and Dysfunction after Acute Pressure Overload in Mice. *Am J Pathol* **166**:15–25.
- Hiam-Galvez KJ, Allen BM, and Spitzer MH (2021) Systemic immunity in cancer. *Nat Rev Cancer* **21**:345–359.
- Hingorani D V., Lemieux AJ, Acevedo JR, Glasgow HL, Kedarisetty S, Whitney MA, Molinolo AA, Tsien RY, and Nguyen QT (2017) Early detection of squamous cell carcinoma in carcinogen induced oral cancer rodent model by ratiometric activatable cell penetrating peptides. *Oral Oncol* **71**:156–162.
- Hohn M, Chang M, Meisel JE, Frost E, Schwegmann K, Hermann S, Schäfers M, Riemann B, Haufe G, Breyholz H, and Wagner S (2018) Synthesis and Preliminary In Vitro and In Vivo Evaluation of Thiirane-Based Slow-Binding MMP Inhibitors as Potential Radiotracers for PET Imaging. *ChemistrySelect* **3**:11729–11736.
- Holmbeck K, Bianco P, Caterina J, Yamada S, Kromer M, Kuznetsov SA, Mankani M, Gehron Robey P, Poole AR, Pidoux I, Ward JM, and Birkedal-Hansen H (1999) MT1-MMP-Deficient Mice Develop Dwarfism, Osteopenia, Arthritis, and Connective Tissue Disease due to Inadequate Collagen Turnover. *Cell* **99**:81–92.
- Holmbeck K, Bianco P, Chrysovergis K, Yamada S, and Birkedal-Hansen H (2003) MT1-MMP-dependent, apoptotic remodeling of unmineralized cartilage. *J Cell Biol* **163**:661–671.
- Holmbeck K, Bianco P, Yamada S, and Birkedal-Hansen H (2004) MT1-MMP: A tethered collagenase. *J Cell Physiol* **200**:11–19.
- Horiuchi K, Kimura T, Miyamoto T, Takaishi H, Okada Y, Toyama Y, and Blobel CP (2007) Cutting Edge: TNF- α -Converting Enzyme (TACE/ADAM17) Inactivation in Mouse Myeloid Cells Prevents Lethality from Endotoxin Shock. *J Immunol* **179**:2686–2689.
- Hotary KB, Allen ED, Brooks PC, Datta NS, Long MW, and Weiss SJ (2003) Membrane Type I Matrix Metalloproteinase Usurps Tumor Growth Control Imposed by the Three-Dimensional Extracellular Matrix. *Cell* **114**:33–45.
- Houghton AM, Hartzell WO, Robbins CS, Gomis-Rüth FX, and Shapiro SD (2009) Macrophage elastase kills bacteria within murine macrophages. *Nature* **460**:637–641.
- Hsu J-YC, McKeon R, Goussev S, Werb Z, Lee J-U, Trivedi A, and Noble-Haesslein LJ (2006) Matrix Metalloproteinase-2 Facilitates Wound Healing Events That Promote Functional Recovery after Spinal Cord Injury. *J Neurosci* **26**:9841–9850.
- Hu H-Y, Gehrig S, Reither G, Subramanian D, Mall MA, Plettenburg O, and Schultz C (2014) FRET-based and other fluorescent proteinase probes. *Biotechnol J* **9**:266–281.
- Hu J, Van den Steen PE, Sang Q-XA, and Opdenakker G (2007) Matrix metalloproteinase inhibitors as therapy for inflammatory and vascular diseases. *Nat Rev Drug Discov* **6**:480–498.
- Hu XX, Li L, Li DR, Zhang W, Cheng XQ, Zhang JQ, and Tang BJ (2004) Expression of matrix metalloproteinases-9,2,7, and tissue inhibitor of metalloproteinases-1,2,3 mRNA in ovarian tumors and their clinical significance. *Ai Zheng* **23**.
- Hua N, Baik F, Pham T, Phinikaridou A, Giordano N, Friedman B, Whitney M, Nguyen QT, Tsien RY, and Hamilton JA (2015) Identification of High-Risk Plaques by MRI and Fluorescence Imaging in a Rabbit Model of Atherothrombosis. *PLoS One* **10**:e0139833.
- Hueber AJ, and McInnes IB (2009) Pathogenesis in Rheumatoid Arthritis: Cytokines, in *Rheumatoid Arthritis* pp 200–208, Elsevier.
- Hugenberg V, Wagner S, Kopka K, Schäfers M, Schuit RC, Windhorst AD, and Hermann S (2017) Radiolabeled Selective Matrix Metalloproteinase 13 (MMP-13) Inhibitors:

- (Radio)Syntheses and in Vitro and First in Vivo Evaluation. *J Med Chem* **60**:307–321.
- Hynes RO (2009) The Extracellular Matrix: Not Just Pretty Fibrils. *Science* (80-) **326**:1216–1219.
- Hynes RO, and Naba A (2012) Overview of the Matrisome--An Inventory of Extracellular Matrix Constituents and Functions. *Cold Spring Harb Perspect Biol* **4**:a004903–a004903.
- Ichikawa Y, Ishikawa T, Momiyama N, Kamiyama M, Sakurada H, Matsuyama R, Hasegawa Satoshi, Chishima T, Hamaguchi Y, Fujii S, Saito S, Kubota K, Hasegawa Shingo, Ike H, Oki S, and Shimada H (2006) Matrilysin (MMP-7) degrades VE-cadherin and accelerates accumulation of beta-catenin in the nucleus of human umbilical vein endothelial cells. *Oncol Rep* **15**.
- Inada M, Wang Y, Byrne MH, Rahman MU, Miyaura C, Lopez-Otin C, and Krane SM (2004) Critical roles for collagenase-3 (Mmp13) in development of growth plate cartilage and in endochondral ossification. *Proc Natl Acad Sci* **101**:17192–17197.
- Iozzo R V., Zoeller JJ, and Nyström A (2009) Basement membrane proteoglycans: Modulators Par Excellence of cancer growth and angiogenesis. *Mol Cells* **27**:503–513.
- Irie K, Komori K, Seiki M, Tsuruga E, Sakakura Y, Kaku T, and Yajima T (2005) Impaired alveolization in mice deficient in membrane-type matrix metalloproteinase 1 (MT1-MMP). *Med Mol Morphol* **38**:43–46.
- Ishizuya-Oka A, Li Q, Amano T, Damjanovski S, Ueda S, and Shi Y-B (2000) Requirement for Matrix Metalloproteinase Stromelysin-3 in Cell Migration and Apoptosis during Tissue Remodeling in *Xenopus laevis*. *J Cell Biol* **150**:1177–1188.
- Itoh T, Ikeda T, Gomi H, Nakao S, Suzuki T, and Itohara S (1997) Unaltered Secretion of β -Amyloid Precursor Protein in Gelatinase A (Matrix Metalloproteinase 2)-deficient Mice. *J Biol Chem* **272**:22389–22392.
- Itoh T, Matsuda H, Tanioka M, Kuwabara K, Itohara S, and Suzuki R (2002) The Role of Matrix Metalloproteinase-2 and Matrix Metalloproteinase-9 in Antibody-Induced Arthritis. *J Immunol* **169**:2643–2647.
- Iyer RP, Patterson NL, Fields GB, and Lindsey ML (2012) The history of matrix metalloproteinases: milestones, myths, and misperceptions. *Am J Physiol Circ Physiol* **303**:H919–H930.
- Jackson HW, Defamie V, Waterhouse P, and Khokha R (2017) TIMPs: versatile extracellular regulators in cancer. *Nat Rev Cancer* **17**:38–53.
- Jacob-Ferreira AL, Kondo MY, Baral PK, James MNG, Holt A, Fan X, and Schulz R (2013) Phosphorylation Status of 72 kDa MMP-2 Determines Its Structure and Activity in Response to Peroxynitrite. *PLoS One* **8**:e71794.
- Jacobsen JA, Major Jourden JL, Miller MT, and Cohen SM (2010) To bind zinc or not to bind zinc: An examination of innovative approaches to improved metalloproteinase inhibition. *Biochim Biophys Acta - Mol Cell Res* **1803**:72–94.
- Jara P, Calyeca J, Romero Y, Plácido L, Yu G, Kaminski N, Maldonado V, Cisneros J, Selman M, and Pardo A (2015) Matrix metalloproteinase (MMP)-19-deficient fibroblasts display a profibrotic phenotype. *Am J Physiol Cell Mol Physiol* **308**:L511–L522.
- Järveläinen H, Sainio A, Koulu M, Wight TN, and Penttinen R (2009) Extracellular Matrix Molecules: Potential Targets in Pharmacotherapy. *Pharmacol Rev* **61**:198–223.
- Jay GD, Elsaid KA, Kelly KA, Anderson SC, Zhang L, Teeple E, Waller K, and Fleming BC (2012) Prevention of cartilage degeneration and gait asymmetry by lubricin tribosupplementation in the rat following anterior cruciate ligament transection. *Arthritis*

Rheum, doi: 10.1002/art.33461.

- Jayawardena DP, Kulkarni NP, and Gill SE (2019) The role of tissue inhibitors of metalloproteinases in microvascular endothelial cell barrier dysfunction during sepsis. *Met Med* **Volume 6**:1–12.
- Ji X, Xie S, Jiao Y, Zhang X, Sun D, Yang VC, Wang M, He H, and Sun L (2020) MT1-MMP activatable fluorogenic probes with enhanced specificity via high-affinity peptide conjugation for tumor imaging. *Biomater Sci* **8**:2308–2317.
- Jiang A, Lehti K, Wang X, Weiss SJ, Keski-Oja J, and Pei D (2001) Regulation of membrane-type matrix metalloproteinase 1 activity by dynamin-mediated endocytosis. *Proc Natl Acad Sci* **98**:13693–13698.
- Jiang T, Olson ES, Nguyen QT, Roy M, Jennings PA, and Tsien RY (2004) Tumor imaging by means of proteolytic activation of cell-penetrating peptides. *Proc Natl Acad Sci* **101**:17867–17872.
- Jirouskova M, Zbodakova O, Gregor M, Chalupsky K, Sarnova L, Hajduch M, Ehrmann J, Jirkovska M, and Sedlacek R (2012) Hepatoprotective Effect of MMP-19 Deficiency in a Mouse Model of Chronic Liver Fibrosis. *PLoS One* **7**:e46271.
- Johansson N, Ala-aho R, Uitto V, Grenman R, Fusenig NE, Lopez-Otin C, and Kahari VM (2000) Expression of collagenase-3 (MMP-13) and collagenase-1 (MMP-1) by transformed keratinocytes is dependent on the activity of p38 mitogen-activated protein kinase. *J Cell Sci* **113**:227–235.
- Johnson C, Sung H-J, Lessner SM, Fini ME, and Galis ZS (2004) Matrix Metalloproteinase-9 Is Required for Adequate Angiogenic Revascularization of Ischemic Tissues. *Circ Res* **94**:262–268.
- Johnson JL, Devel L, Czarny B, George SJ, Jackson CL, Rogakos V, Beau F, Yiotakis A, Newby AC, and Dive V (2011) A selective matrix metalloproteinase-12 inhibitor retards atherosclerotic plaque development in apolipoprotein E-knockout mice. *Arterioscler Thromb Vasc Biol* **31**:528–535, Lippincott Williams and Wilkins.
- Johnston JB, Zhang K, Silva C, Shalinsky DR, Conant K, Ni W, Corbett D, Yong VW, and Power C (2001) HIV-1 Tat neurotoxicity is prevented by matrix metalloproteinase inhibitors. *Ann Neurol* **49**:230–241.
- Joshi CR, Stacy S, Sumien N, Ghorpade A, and Borgmann K (2020) Astrocyte HIV-1 Tat Differentially Modulates Behavior and Brain MMP/TIMP Balance During Short and Prolonged Induction in Transgenic Mice. *Front Neurol* **11**.
- Jozic D, Bourenkov G, Lim N-H, Visse R, Nagase H, Bode W, and Maskos K (2005) X-ray Structure of Human proMMP-1. *J Biol Chem* **280**:9578–9585.
- Ju SM, Song HY, Lee JA, Lee SJ, Choi SY, and Park J (2009) Extracellular HIV-1 Tat up-regulates expression of matrix metalloproteinase-9 via a MAPK-NF- κ B dependent pathway in human astrocytes. *Exp Mol Med* **41**:86.
- Juurikka, Butler, Salo, Nyberg, and Åström (2019) The Role of MMP8 in Cancer: A Systematic Review. *Int J Mol Sci* **20**:4506.
- Juurikka K, Dufour A, Pehkonen K, Mainoli B, Campioni Rodrigues P, Solis N, Klein T, Nyberg P, Overall CM, Salo T, and Åström P (2021) MMP8 increases tongue carcinoma cell–cell adhesion and diminishes migration via cleavage of anti-adhesive FXYD5. *Oncogenesis* **10**:44.
- Kaminska M, Bruyat P, Malgorn C, Doladilhe M, Cassar-Lajeunesse E, Fruchart Gaillard C, De Souza M, Beau F, Thai R, Correia I, Galat A, Georgiadis D, Lequin O, Dive V, Bregant S,

- and Devel L (2021) Ligand-Directed Modification of Active Matrix Metalloproteases: Activity-based Probes with no Photolabile Group. *Angew Chemie Int Ed* 202106117.
- Kandalam V, Basu R, Abraham T, Wang X, Soloway PD, Jaworski DM, Oudit GY, and Kassiri Z (2010) TIMP2 Deficiency Accelerates Adverse Post-Myocardial Infarction Remodeling Because of Enhanced MT1-MMP Activity Despite Lack of MMP2 Activation. *Circ Res* **106**:796–808.
- Kappelhoff R, Puente XS, Wilson CH, Seth A, López-Otín C, and Overall CM (2017) Overview of transcriptomic analysis of all human proteases, non-proteolytic homologs and inhibitors: Organ, tissue and ovarian cancer cell line expression profiling of the human protease degradome by the CLIP-CHIP™ DNA microarray. *Biochim Biophys Acta Mol Cell Res* **1864**:2210–2219.
- Kassim SY, Gharib SA, Mecham BH, Birkland TP, Parks WC, and McGuire JK (2007) Individual Matrix Metalloproteinases Control Distinct Transcriptional Responses in Airway Epithelial Cells Infected with *Pseudomonas aeruginosa*. *Infect Immun* **75**:5640–5650.
- Kassiri Z, Oudit GY, Kandalam V, Awad A, Wang X, Ziou X, Maeda N, Herzenberg AM, and Scholey JW (2009) Loss of TIMP3 Enhances Interstitial Nephritis and Fibrosis. *J Am Soc Nephrol* **20**:1223–1235.
- Kaushik DK, Bhattacharya A, Mirzaei R, Rawji KS, Ahn Y, Rho JM, and Yong VW (2019) Enhanced glycolytic metabolism supports transmigration of brain-infiltrating macrophages in multiple sclerosis. *J Clin Invest* **129**:3277–3292.
- Kelly D, Squire IB, Khan SQ, Dhillon O, Narayan H, Ng KH, Quinn P, Davies JE, and Ng LL (2010) Usefulness of Plasma Tissue Inhibitors of Metalloproteinases as Markers of Prognosis After Acute Myocardial Infarction. *Am J Cardiol* **106**:477–482.
- Keow JY, Pond ED, Cisar JS, Cravatt BF, and Crawford BD (2012) Activity-Based Labeling of Matrix Metalloproteinases in Living Vertebrate Embryos. *PLoS One* **7**:e43434.
- Kessenbrock K, Dijkgraaf GJP, Lawson DA, Littlepage LE, Shahi P, Pieper U, and Werb Z (2013) A Role for Matrix Metalloproteinases in Regulating Mammary Stem Cell Function via the Wnt Signaling Pathway. *Cell Stem Cell* **13**:300–313.
- Kessenbrock K, Plaks V, and Werb Z (2010) Matrix Metalloproteinases: Regulators of the Tumor Microenvironment. *Cell* **141**:52–67.
- Kheradmand F, Werner E, Tremble P, Symons M, and Werb Z (1998) Role of Rac1 and Oxygen Radicals in Collagenase-1 Expression Induced by Cell Shape Change. *Science (80-)* **280**:898–902.
- Khokha R, Murthy A, and Weiss A (2013) Metalloproteinases and their natural inhibitors in inflammation and immunity. *Nat Rev Immunol* **13**:649–665.
- Kim HE, Dalal SS, Young E, Legato MJ, Weisfeldt ML, and D'Armiento J (2000) Disruption of the myocardial extracellular matrix leads to cardiac dysfunction. *J Clin Invest* **106**:857–866.
- King SL, Goth CK, Eckhard U, Joshi HJ, Haue AD, Vakhrushev SY, Schjoldager KT, Overall CM, and Wandall HH (2018) TAILS N-terminomics and proteomics reveal complex regulation of proteolytic cleavage by O-glycosylation. *J Biol Chem* **293**:7629–7644.
- Kirkpatrick JD, Warren AD, Soleimany AP, Westcott PMK, Voog JC, Martin-Alonso C, Fleming HE, Tammela T, Jacks T, and Bhatia SN (2020) Urinary detection of lung cancer in mice via noninvasive pulmonary protease profiling. *Sci Transl Med* **12**:eaaw0262.
- Klein G, Schmal O, and Aicher WK (2015) Matrix metalloproteinases in stem cell mobilization. *Matrix Biol* **44–46**:175–183.
- Knapinska A, and Fields GB (2012) Chemical Biology for Understanding Matrix

- Metalloproteinase Function. *ChemBioChem* **13**:2002–2020.
- Knäuper V, Docherty AJP, Smith B, Tschesche H, and Murphy G (1997) Analysis of the contribution of the hinge region of human neutrophil collagenase (HNC, MMP-8) to stability and collagenolytic activity by alanine scanning mutagenesis. *FEBS Lett* **405**:60–64.
- Koivunen E, Arap W, Valtanen H, Rainisalo A, Medina OP, Heikkilä P, Kantor C, Gahmberg CG, Salo T, Konttinen YT, Sorsa T, Ruoslahti E, and Pasqualini R (1999) Tumor targeting with a selective gelatinase inhibitor. *Nat Biotechnol* **17**:768–774.
- Kolaczowska E, Scisłowska-Czarnecka A, Chadzinska M, Plytycz B, van Rooijen N, Opendakker G, and Arnold B (2006) Enhanced early vascular permeability in gelatinase B (MMP-9)-deficient mice: putative contribution of COX-1-derived PGE2 of macrophage origin. *J Leukoc Biol* **80**:125–132.
- Komori K, Nonaka T, Okada A, Kinoh H, Hayashita-Kinoh H, Yoshida N, Yana I, and Seiki M (2004) Absence of mechanical allodynia and A β -fiber sprouting after sciatic nerve injury in mice lacking membrane-type 5 matrix metalloproteinase. *FEBS Lett* **557**:125–128.
- Koppiseti RK, Fulcher YG, Jurkevich A, Prior SH, Xu J, Lenoir M, Overduin M, and Van Doren SR (2014) Ambidextrous binding of cell and membrane bilayers by soluble matrix metalloproteinase-12. *Nat Commun* **5**:5552.
- Korpi JT, Kervinen V, Mäklin H, Väänänen A, Lahtinen M, Läärä E, Ristimäki A, Thomas G, Ylipalosaari M, Åström P, Lopez-Otin C, Sorsa T, Kantola S, Pirilä E, and Salo T (2008) Collagenase-2 (matrix metalloproteinase-8) plays a protective role in tongue cancer. *Br J Cancer* **98**:766–775.
- Koshikawa N, Schenk S, Moeckel G, Sharabi A, Miyazaki K, Gardner H, Zent R, and Quaranta V (2004) Proteolytic processing of laminin-5 by MT1-MMP in tissues and its effects on epithelial cell morphology. *FASEB J* **18**:1–22.
- Koskinen SOA, Wang W, Ahtikoski AM, Kjær M, Han XY, Komulainen J, Kovanen V, and Takala TES (2001) Acute exercise induced changes in rat skeletal muscle mRNAs and proteins regulating type IV collagen content. *Am J Physiol Integr Comp Physiol* **280**:R1292–R1300.
- Kure T, Chang J-H, Kato T, Hernández-Quintela E, Ye H, Lu PC-S, Matrisian LM, Gatinel D, Shapiro S, Ghosheh F, and Azar DT (2003) Corneal Neovascularization after Excimer Keratectomy Wounds in Matrilysin-Deficient Mice. *Investig Ophthalmology Vis Sci* **44**:137.
- Kuzuya M, Nakamura K, Sasaki T, Wu Cheng X, Itohara S, and Iguchi A (2006) Effect of MMP-2 Deficiency on Atherosclerotic Lesion Formation in ApoE-Deficient Mice. *Arterioscler Thromb Vasc Biol* **26**:1120–1125.
- Kwong GA, von Maltzahn G, Murugappan G, Abudayyeh O, Mo S, Papayannopoulos IA, Sverdlov DY, Liu SB, Warren AD, Popov Y, Schuppan D, and Bhatia SN (2013) Mass-encoded synthetic biomarkers for multiplexed urinary monitoring of disease. *Nat Biotechnol* **31**:63–70.
- Lakatos G, Sipos F, Miheller P, Hritz I, Varga MZ, Juhász M, Molnár B, Tulassay Z, and Herszényi L (2012) The Behavior of Matrix Metalloproteinase-9 in Lymphocytic Colitis, Collagenous Colitis and Ulcerative Colitis. *Pathol Oncol Res* **18**:85–91.
- Lange PF, and Overall CM (2011) TopFIND, a knowledgebase linking protein termini with function. *Nat Methods* **8**:703–704.
- Laronha H, and Caldeira J (2020) Structure and Function of Human Matrix Metalloproteinases. *Cells* **9**:1076.
- Larsen PH (2006) Myelin Formation during Development of the CNS Is Delayed in Matrix

- Metalloproteinase-9 and -12 Null Mice. *J Neurosci* **26**:2207–2214.
- Larsen PH, Wells JE, Stallcup WB, Opdenakker G, and Yong VW (2003) Matrix Metalloproteinase-9 Facilitates Remyelination in Part by Processing the Inhibitory NG2 Proteoglycan. *J Neurosci* **23**:11127–11135.
- Lassmann H (2019) Pathogenic Mechanisms Associated With Different Clinical Courses of Multiple Sclerosis. *Front Immunol* **9**.
- Lauer-Fields J, Brew K, Whitehead JK, Li S, Hammer RP, and Fields GB (2007) Triple-Helical Transition State Analogues: A New Class of Selective Matrix Metalloproteinase Inhibitors. *J Am Chem Soc* **129**:10408–10417.
- Lebel R, and Lepage M (2014) A comprehensive review on controls in molecular imaging: lessons from MMP-2 imaging. *Contrast Media Mol Imaging* **9**:187–210.
- Leco KJ, Khokha R, Pavloff N, Hawkes SP, and Edwards DR (1994) Tissue inhibitor of metalloproteinases-3 (TIMP-3) is an extracellular matrix-associated protein with a distinctive pattern of expression in mouse cells and tissues. *J Biol Chem* **269**:9352–9360.
- Leco KJ, Waterhouse P, Sanchez OH, Gowing KLM, Poole AR, Wakeham A, Mak TW, and Khokha R (2001) Spontaneous air space enlargement in the lungs of mice lacking tissue inhibitor of metalloproteinases-3 (TIMP-3). *J Clin Invest* **108**:817–829.
- Lee H, and Kim Y-P (2015) Fluorescent and bioluminescent nanoprobes for in vitro and in vivo detection of matrix metalloproteinase activity. *BMB Rep* **48**:313–318.
- Lee HW, Lee SJ, Lee MY, Park MW, Kim SS, Shin N, Lovett DH, Bae SS, Ahn J, Park J-S, Oh J-H, Choi JH, Lee HC, Cha KS, Hong TJ, and Song SH (2019) Enhanced cardiac expression of two isoforms of matrix metalloproteinase-2 in experimental diabetes mellitus. *PLoS One* **14**:e0221798.
- Lee JG, Dahi S, Mahimkar R, Tulloch NL, Alfonso-Jaume MA, Lovett DH, and Sarkar R (2005) Intronic regulation of matrix metalloproteinase-2 revealed by in vivo transcriptional analysis in ischemia. *Proc Natl Acad Sci* **102**:16345–16350.
- Lee S-R (2006) Involvement of Matrix Metalloproteinase in Neuroblast Cell Migration from the Subventricular Zone after Stroke. *J Neurosci* **26**:3491–3495.
- Lei Z, Jian M, Li X, Wei J, Meng X, and Wang Z (2020) Biosensors and bioassays for determination of matrix metalloproteinases: state of the art and recent advances. *J Mater Chem B* **8**:3261–3291.
- Leong E, Bezuhy M, and Marshall JS (2021) Distinct Metalloproteinase Expression and Functions in Systemic Sclerosis and Fibrosis: What We Know and the Potential for Intervention. *Front Physiol* **12**.
- Leppert D, Leib SL, Grygar C, Miller KM, Schaad UB, and Holländer GA (2000) Matrix Metalloproteinase (MMP)-8 and MMP-9 in Cerebrospinal Fluid during Bacterial Meningitis: Association with Blood-Brain Barrier Damage and Neurological Sequelae. *Clin Infect Dis* **31**:80–84.
- Levin AD, Wildenberg ME, and van den Brink GR (2016) Mechanism of Action of Anti-TNF Therapy in Inflammatory Bowel Disease. *J Crohn's Colitis* **10**:989–997.
- Li CKF, Pender SLF, Pickard KM, Chance V, Holloway JA, Huett A, Gonçalves NS, Mudgett JS, Dougan G, Frankel G, and MacDonald TT (2004) Impaired Immunity to Intestinal Bacterial Infection in Stromelysin-1 (Matrix Metalloproteinase-3)-Deficient Mice. *J Immunol* **173**:5171–5179.
- Li J, Brick P, O'Hare M, Skarzynski T, Lloyd L, Curry V, Clark I, Bigg H, Hazleman B, Cawston T, and Blow D (1995) Structure of full-length porcine synovial collagenase reveals

- a C-terminal domain containing a calcium-linked, four-bladed β -propeller. *Structure* **3**:541–549.
- Li Q, Park PW, Wilson CL, and Parks WC (2002) Matrilysin Shedding of Syndecan-1 Regulates Chemokine Mobilization and Transepithelial Efflux of Neutrophils in Acute Lung Injury. *Cell* **111**:635–646.
- Li Q, Zhang Q, Wang C, Liu X, Li N, and Li J (2009) Disruption of tight junctions during polymicrobial sepsis in vivo. *J Pathol* **218**:210–221.
- Li W, Li J, Wu Y, Wu J, Hotchandani R, Cunningham K, McFadyen I, Bard J, Morgan P, Schlerman F, Xu X, Tam S, Goldman SJ, Williams C, Sypek J, and Mansour TS (2009) A Selective Matrix Metalloprotease 12 Inhibitor for Potential Treatment of Chronic Obstructive Pulmonary Disease (COPD): Discovery of (S)-2-(8-(Methoxycarbonylamino)dibenzo[b , d]furan-3-sulfonamido)-3-methylbutanoic acid (MMP408). *J Med Chem* **52**:1799–1802.
- Libby P, and Aikawa M (2002) Stabilization of atherosclerotic plaques: New mechanisms and clinical targets. *Nat Med* **8**:1257–1262.
- Lijnen H. R., Soloway P, and Collen D (1999) Tissue inhibitor of matrix metalloproteinases-1 impairs arterial neointima formation after vascular injury in mice. *Circ Res* **85**.
- Lijnen HR, Van Hoef B, Frederix L, Rio M-C, and Collen D (2002) Adipocyte Hypertrophy in Stromelysin-3 Deficient Mice with Nutritionally Induced Obesity. *Thromb Haemost* **87**:530–535.
- Lijnen H. Roger, Van Hoef B, Vanlinthout I, Verstreken M, Rio M-C, and Collen D (1999) Accelerated Neointima Formation After Vascular Injury in Mice With Stromelysin-3 (MMP-11) Gene Inactivation. *Arterioscler Thromb Vasc Biol* **19**:2863–2870.
- Littlepage LE, Sternlicht MD, Rougier N, Phillips J, Gallo E, Yu Y, Williams K, Brenot A, Gordon JI, and Werb Z (2010) Matrix Metalloproteinases Contribute Distinct Roles in Neuroendocrine Prostate Carcinogenesis, Metastasis, and Angiogenesis Progression. *Cancer Res* **70**:2224–2234.
- Liu H-W, Chen L, Xu C, Li Z, Zhang H, Zhang X-B, and Tan W (2018) Recent progresses in small-molecule enzymatic fluorescent probes for cancer imaging. *Chem Soc Rev* **47**:7140–7180.
- Liu J (2003) Early combined treatment with carboplatin and the MMP inhibitor, prinomastat, prolongs survival and reduces systemic metastasis in an aggressive orthotopic lung cancer model. *Lung Cancer* **42**:335–344.
- Liu X, Manzano G, Lovett DH, and Kim HT (2010) Role of AP-1 and RE-1 binding sites in matrix metalloproteinase-2 transcriptional regulation in skeletal muscle atrophy. *Biochem Biophys Res Commun* **396**:219–223.
- Lohi J, Wilson CL, Roby JD, and Parks WC (2001) Epilysin, a Novel Human Matrix Metalloproteinase (MMP-28) Expressed in Testis and Keratinocytes and in Response to Injury. *J Biol Chem* **276**:10134–10144.
- Long ME, Gong K-Q, Volk JS, Eddy WE, Chang MY, Frevert CW, Altemeier WA, Gale M, Liles WC, and Manicone AM (2018) Matrix metalloproteinase 28 is regulated by TRIF- and type I IFN-dependent signaling in macrophages. *Innate Immun* **24**:357–365, SAGE Publications Ltd.
- Lorente L, Martín MM, Labarta L, Díaz C, Solé-Violán J, Blanquer J, Orbe J, Rodríguez JA, Jiménez A, Borreguero-León JM, Belmonte F, Medina JC, Llimiñana MC, Ferrer-Agüero JM, Ferreres J, Mora ML, Lubillo S, Sánchez M, Barrios Y, Sierra A, and Páramo JA

- (2009) Matrix metalloproteinase-9, -10, and tissue inhibitor of matrix metalloproteinases-1 blood levels as biomarkers of severity and mortality in sepsis. *Crit Care* **13**:R158.
- Lorente L, Martín MM, Solé-Violán J, Blanquer J, Labarta L, Díaz C, Borreguero-León JM, Orbe J, Rodríguez JA, Jiménez A, and Páramo JA (2014) Association of Sepsis-Related Mortality with Early Increase of TIMP-1/MMP-9 Ratio. *PLoS One* **9**:e94318.
- Lovejoy B, Cleasby A, Hassell A, Longley K, Luther M, Weigl D, McGeehan G, McElroy A, Drewry D, Lambert M, and Et A (1994) Structure of the catalytic domain of fibroblast collagenase complexed with an inhibitor. *Science (80-)* **263**:375–377.
- Lovett DH, Chu C, Wang G, Ratcliffe MB, and Baker AJ (2014) A N-terminal truncated intracellular isoform of matrix metalloproteinase-2 impairs contractility of mouse myocardium. *Front Physiol* **5**.
- Lovett DH, Mahimkar R, Raffai RL, Cape L, Maklashina E, Cecchini G, and Karliner JS (2012) A Novel Intracellular Isoform of Matrix Metalloproteinase-2 Induced by Oxidative Stress Activates Innate Immunity. *PLoS One* **7**:e34177.
- Loynachan CN, Soleimany AP, Dudani JS, Lin Y, Najer A, Bekdemir A, Chen Q, Bhatia SN, and Stevens MM (2019) Renal clearable catalytic gold nanoclusters for in vivo disease monitoring. *Nat Nanotechnol* **14**:883–890.
- Lu P, Takai K, Weaver VM, and Werb Z (2011) Extracellular Matrix Degradation and Remodeling in Development and Disease. *Cold Spring Harb Perspect Biol* **3**:a005058–a005058.
- Luo D, Mari B, Stoll I, and Anglard P (2002) Alternative Splicing and Promoter Usage Generates an Intracellular Stromelysin 3 Isoform Directly Translated as an Active Matrix Metalloproteinase. *J Biol Chem* **277**:25527–25536.
- Ma N, Hu J, Zhang Z-M, Liu W, Huang M, Fan Y, Yin X, Wang J, Ding K, Ye W, and Li Z (2020) 2 H -Azirine-Based Reagents for Chemoselective Bioconjugation at Carboxyl Residues Inside Live Cells. *J Am Chem Soc* **142**:6051–6059.
- Ma Y, Halade G V., Zhang J, Ramirez TA, Levin D, Voorhees A, Jin Y-F, Han H-C, Manicone AM, and Lindsey ML (2013) Matrix Metalloproteinase-28 Deletion Exacerbates Cardiac Dysfunction and Rupture After Myocardial Infarction in Mice by Inhibiting M2 Macrophage Activation. *Circ Res* **112**:675–688.
- Madtes DK, Elston AL, Kaback LA, and Clark JG (2001) Selective Induction of Tissue Inhibitor of Metalloproteinase-1 in Bleomycin-Induced Pulmonary Fibrosis. *Am J Respir Cell Mol Biol* **24**:599–607.
- Madzharova E, Kastl P, Sabino F, and auf dem Keller U (2019) Post-Translational Modification-Dependent Activity of Matrix Metalloproteinases. *Int J Mol Sci* **20**:3077.
- Mahasenani K V., Bastian M, Gao M, Frost E, Ding D, Zorina-Lichtenwalter K, Jacobs J, Suckow MA, Schroeder VA, Wolter WR, Chang M, and Mobashery S (2017) Exploitation of Conformational Dynamics in Imparting Selective Inhibition for Related Matrix Metalloproteinases. *ACS Med Chem Lett* **8**:654–659.
- Mai C-W, Chung FF-L, and Leong C-O (2017) Targeting Legumain As a Novel Therapeutic Strategy in Cancers. *Curr Drug Targets* **18**.
- Majster M, Lira-Junior R, Höög CM, Almer S, and Boström EA (2020) Salivary and Serum Inflammatory Profiles Reflect Different Aspects of Inflammatory Bowel Disease Activity. *Inflamm Bowel Dis* **26**:1588–1596.
- Maley CC, Aktipis A, Graham TA, Sottoriva A, Boddy AM, Janiszewska M, Silva AS, Gerlinger M, Yuan Y, Pienta KJ, Anderson KS, Gatenby R, Swanton C, Posada D, Wu C-I,

- Schiffman JD, Hwang ES, Polyak K, Anderson ARA, Brown JS, Greaves M, and Shibata D (2017) Classifying the evolutionary and ecological features of neoplasms. *Nat Rev Cancer* **17**:605–619.
- Mallia-Milanes B., Dufour A, Philp C, Solis N, Klein T, Fischer M, Bolton CE, Shapiro S, Overall CM, and Johnson SR (2018) TAILS proteomics reveals dynamic changes in airway proteolysis controlling protease activity and innate immunity during COPD exacerbations. *Am J Physiol - Lung Cell Mol Physiol* **315**.
- Mallia-Milanes Brendan, Dufour A, Philp C, Solis N, Klein T, Fischer M, Bolton CE, Shapiro S, Overall CM, and Johnson SR (2018) TAILS proteomics reveals dynamic changes in airway proteolysis controlling protease activity and innate immunity during COPD exacerbations. *Am J Physiol Cell Mol Physiol* **315**:L1003–L1014.
- Manicone AM, Birkland TP, Lin M, Betsuyaku T, van Rooijen N, Lohi J, Keski-Oja J, Wang Y, Skerrett SJ, and Parks WC (2009) Epilysin (MMP-28) Restrains Early Macrophage Recruitment in *Pseudomonas aeruginosa* Pneumonia. *J Immunol* **182**:3866–3876.
- Manicone AM, Gharib SA, Gong K-Q, Eddy WE, Long ME, Frevert CW, Altemeier WA, Parks WC, and Houghton AM (2017) Matrix Metalloproteinase-28 Is a Key Contributor to Emphysema Pathogenesis. *Am J Pathol* **187**:1288–1300.
- Manka SW, Carafoli F, Visse R, Bihan D, Raynal N, Farndale RW, Murphy G, Enghild JJ, Hohenester E, and Nagase H (2012) Structural insights into triple-helical collagen cleavage by matrix metalloproteinase 1. *Proc Natl Acad Sci* **109**:12461–12466, National Acad Sciences.
- Mannello F, and Medda V (2012) Nuclear localization of Matrix metalloproteinases. *Prog Histochem Cytochem* **47**:27–58.
- Maquoi E, Munaut C, Colige A, Lambert C, Franken F, Noël A, Grams F, Krell H-W, and Foidart J-M (2002) Stimulation of Matrix Metalloproteinase-9 Expression in Human Fibrosarcoma Cells by Synthetic Matrix Metalloproteinase Inhibitors. *Exp Cell Res* **275**:110–121.
- Marchant DJ, Bellac CL, Moraes TJ, Wadsworth SJ, Dufour A, Butler GS, Bilawchuk LM, Hendry RG, Robertson AG, Cheung CT, Ng J, Ang L, Luo Z, Heilbron K, Norris MJ, Duan W, Bucyk T, Karpov A, Devel L, Georgiadis D, Hegele RG, Luo H, Granville DJ, Dive V, McManus BM, and Overall CM (2014) A new transcriptional role for matrix metalloproteinase-12 in antiviral immunity. *Nat Med* **20**:493–502.
- Marchenko ND, Marchenko GN, and Strongin AY (2002) Unconventional Activation Mechanisms of MMP-26, a Human Matrix Metalloproteinase with a Unique PHCG XXD Cysteine-switch Motif. *J Biol Chem* **277**:18967–18972.
- Marshall DC, Lyman SK, McCauley S, Kovalenko M, Spangler R, Liu C, Lee M, O’Sullivan C, Barry-Hamilton V, Ghermazien H, Mikels-Vigdal A, Garcia CA, Jorgensen B, Velayo AC, Wang R, Adamkewicz JI, and Smith V (2015) Selective Allosteric Inhibition of MMP9 Is Efficacious in Preclinical Models of Ulcerative Colitis and Colorectal Cancer. *PLoS One* **10**:e0127063.
- Martignetti J, Mosig R, Schulz R, Kassiri Z, and Schaffler M (2014) Multicentric osteolysis with nodulosis, arthritis, and cardiac defect syndrome: loss of MMP2 leads to increased apoptosis with alteration of apoptotic regulators and caspases and embryonic lethality. *Adv Genomics Genet* **207**.
- Martín-Alonso M, García-Redondo AB, Guo D, Camafeita E, Martínez F, Alfranca A, Méndez-Barbero N, Pollán Á, Sánchez-Camacho C, Denhardt DT, Seiki M, Vázquez J, Salices M,

- Redondo JM, Milewicz D, and Arroyo AG (2015) Deficiency of MMP17/MT4-MMP Proteolytic Activity Predisposes to Aortic Aneurysm in Mice. *Circ Res* **117**.
- Martín-Gago P, Fansa EK, Winzker M, Murarka S, Janning P, Schultz-Fademrecht C, Baumann M, Wittinghofer A, and Waldmann H (2017) Covalent Protein Labeling at Glutamic Acids. *Cell Chem Biol* **24**:589-597.e5.
- Martin DR, Santamaria S, Koch CD, Ahnström J, and Apte SS (2021) Identification of novel ADAMTS1, ADAMTS4 and ADAMTS5 cleavage sites in versican using a label-free quantitative proteomics approach. *J Proteomics* **249**:104358.
- Martin EL, Moyer BZ, Pape MC, Starcher B, Leco KJ, and Veldhuizen RAW (2003) Negative impact of tissue inhibitor of metalloproteinase-3 null mutation on lung structure and function in response to sepsis. *Am J Physiol Cell Mol Physiol* **285**:L1222–L1232.
- Martin EL, Sheikh TA, Leco KJ, Lewis JF, and Veldhuizen RAW (2007) Contribution of alveolar macrophages to the response of the TIMP-3 null lung during a septic insult. *Am J Physiol Cell Mol Physiol* **293**:L779–L789.
- Martínez MC, and Andriantsitohaina R (2009) Reactive Nitrogen Species: Molecular Mechanisms and Potential Significance in Health and Disease. *Antioxid Redox Signal* **11**:669–702.
- Masciantonio MG, Lee CKS, Arpino V, Mehta S, and Gill SE (2017a) The Balance Between Metalloproteinases and TIMPs, in *Progress in Molecular Biology and Translational Science* pp 101–131.
- Masciantonio MG, Lee CKS, Arpino V, Mehta S, and Gill SE (2017b) The Balance Between Metalloproteinases and TIMPs, in pp 101–131.
- Maskos K (2005) Crystal structures of MMPs in complex with physiological and pharmacological inhibitors. *Biochimie* **87**:249–263.
- Masson R, Lefebvre O, Noël A, Fahime M El, Chenard M-P, Wendling C, Kebers F, LeMeur M, Dierich A, Foidart J-M, Basset P, and Rio M-C (1998) In Vivo Evidence That the Stromelysin-3 Metalloproteinase Contributes in a Paracrine Manner to Epithelial Cell Malignancy. *J Cell Biol* **140**:1535–1541.
- Matsunaga T, Abe N, Kameda K, Hagii J, Fujita N, Onodera H, Kamata T, Ishizaka H, Hanada H, Osanai T, and Okumura K (2005) Circulating level of gelatinase activity predicts ventricular remodeling in patients with acute myocardial infarction. *Int J Cardiol* **105**:203–208.
- Mattey DL, Packham JC, Nixon NB, Coates L, Creamer P, Hailwood S, Taylor GJ, and Bhalla AK (2012) Association of cytokine and matrix metalloproteinase profiles with disease activity and function in ankylosing spondylitis. *Arthritis Res Ther* **14**:R127.
- Matusiak N, Waarde A, Bischoff R, Oltenfreiter R, Wiele C van, Dierckx R, and Elsinga P (2013) Probes for Non-invasive Matrix Metalloproteinase-targeted Imaging with PET and SPECT. *Curr Pharm Des* **19**:4647–4672.
- McMahan RS, Birkland TP, Smigiel KS, Vandivort TC, Rohani MG, Manicone AM, McGuire JK, Gharib SA, and Parks WC (2016) Stromelysin-2 (MMP10) Moderates Inflammation by Controlling Macrophage Activation. *J Immunol* **197**:899–909.
- McMillan SJ, Kearley J, Campbell JD, Zhu X-W, Larbi KY, Shipley JM, Senior RM, Nourshargh S, and Lloyd CM (2004) Matrix Metalloproteinase-9 Deficiency Results in Enhanced Allergen-Induced Airway Inflammation. *J Immunol* **172**:2586–2594.
- McQuibban GA (2000) Inflammation Dampened by Gelatinase A Cleavage of Monocyte Chemoattractant Protein-3. *Science (80-)* **289**:1202–1206.

- McQuibban GA, Butler GS, Gong J-H, Bendall L, Power C, Clark-Lewis I, and Overall CM (2001) Matrix Metalloproteinase Activity Inactivates the CXC Chemokine Stromal Cell-derived Factor-1. *J Biol Chem* **276**:43503–43508.
- McQuibban GA, Gong JH, Tam EM, McCulloch CAG, Clark-Lewis I, and Overall CM (2000) Inflammation dampened by gelatinase a cleavage of monocyte chemoattractant protein-3. *Science* (80-), doi: 10.1126/science.289.5482.1202.
- Melendez-Zajgla J, Pozo L, Ceballos G, and Maldonado V (2008) Tissue Inhibitor of Metalloproteinases-4. The road less traveled. *Mol Cancer* **7**:85.
- Meli DN, Christen S, and Leib SL (2003) Matrix Metalloproteinase–9 in Pneumococcal Meningitis: Activation via an Oxidative Pathway. *J Infect Dis* **187**:1411–1415.
- Mertens PR, Harendza S, Pollock AS, and Lovett DH (1997) Glomerular Mesangial Cell-specific Transactivation of Matrix Metalloproteinase 2 Transcription Is Mediated by YB-1. *J Biol Chem* **272**:22905–22912.
- Metildi CA, Felsen CN, Savariar EN, Nguyen QT, Kaushal S, Hoffman RM, Tsien RY, and Bouvet M (2015) Ratiometric Activatable Cell-Penetrating Peptides Label Pancreatic Cancer, Enabling Fluorescence-Guided Surgery, Which Reduces Metastases and Recurrence in Orthotopic Mouse Models. *Ann Surg Oncol* **22**:2082–2087.
- Metz LM, Li DKB, Trabousee AL, Duquette P, Eliasziw M, Cerchiaro G, Greenfield J, Riddehough A, Yeung M, Kremenchutzky M, Vorobeychik G, Freedman MS, Bhan V, Blevins G, Marriott JJ, Grand'Maison F, Lee L, Thibault M, Hill MD, and Yong VW (2017) Trial of Minocycline in a Clinically Isolated Syndrome of Multiple Sclerosis. *N Engl J Med* **376**:2122–2133.
- Miller JP, Holcomb J, Al-Ramahi I, de Haro M, Gafni J, Zhang N, Kim E, Sanhueza M, Torcassi C, Kwak S, Botas J, Hughes RE, and Ellerby LM (2010) Matrix Metalloproteinases Are Modifiers of Huntingtin Proteolysis and Toxicity in Huntington's Disease. *Neuron* **67**:199–212.
- Minagar A, Alexander JS, Schwendimann RN, Kelley RE, Gonzalez-Toledo E, Jimenez JJ, Mauro L, Jy W, and Smith SJ (2008) Combination Therapy With Interferon Beta-1a and Doxycycline in Multiple Sclerosis. *Arch Neurol* **65**.
- Mitsiades N, Yu WH, Poulaki V, Tsokos M, and Stamenkovic I (2001) Matrix metalloproteinase-7-mediated cleavage of Fas ligand protects tumor cells from chemotherapeutic drug cytotoxicity. *Cancer Res* **61**.
- Mohammed FF, Pennington CJ, Kassiri Z, Rubin JS, Soloway PD, Ruther U, Edwards DR, and Khokha R (2005) Metalloproteinase inhibitor TIMP-1 affects hepatocyte cell cycle via HGF activation in murine liver regeneration. *Hepatology* **41**:857–867.
- Mohan V, Gaffney JP, Solomonov I, Levin M, Klepfish M, Akbareian S, Grünwald B, Dym O, Eisenstein M, Yu KH, Kelsen DP, Krüger A, Edwards DR, and Sagi I (2021) Conformation-Specific Inhibitory Anti-MMP-7 Monoclonal Antibody Sensitizes Pancreatic Ductal Adenocarcinoma Cells to Chemotherapeutic Cell Kill. *Cancers (Basel)* **13**:1679.
- Morell M, Nguyen Duc T, Willis AL, Syed S, Lee J, Deu E, Deng Y, Xiao J, Turk BE, Jessen JR, Weiss SJ, and Bogoy M (2013) Coupling Protein Engineering with Probe Design To Inhibit and Image Matrix Metalloproteinases with Controlled Specificity. *J Am Chem Soc* **135**:9139–9148.
- Morgunova E (1999) Structure of Human Pro-Matrix Metalloproteinase-2: Activation Mechanism Revealed. *Science* (80-) **284**:1667–1670.
- Mori H, Bhat R, Bruni-Cardoso A, Chen EI, Jorgens DM, Coutinho K, Louie K, Bowen B Ben,

- Inman JL, Tecca V, Lee SJ, Becker-Weimann S, Northen T, Seiki M, Borowsky AD, Auer M, and Bissell MJ (2016) New insight into the role of MMP14 in metabolic balance. *PeerJ* **4**:e2142.
- Mott JD, and Werb Z (2004) Regulation of matrix biology by matrix metalloproteinases. *Curr Opin Cell Biol* **16**:558–564.
- Moz S, Aita A, Basso D, Ramonda R, Plebani M, and Punzi L (2017) Spondyloarthritis: Matrix Metalloproteinases as Biomarkers of Pathogenesis and Response to Tumor Necrosis Factor (TNF) Inhibitors. *Int J Mol Sci* **18**:830.
- Mu D, Cambier S, Fjellbirkeland L, Baron JL, Munger JS, Kawakatsu H, Sheppard D, Broaddus VC, and Nishimura SL (2002) The integrin $\alpha\beta 8$ mediates epithelial homeostasis through MT1-MMP-dependent activation of TGF- $\beta 1$. *J Cell Biol* **157**:493–507.
- Mudgett JS, Hutchinson NI, Chartrain NA, Forsyth AJ, McDonnell J, Singer II, Bayne EK, Flanagan J, Kawka D, Shen CF, Stevens K, Chen H, Trumbauer M, and Visco DM (1998) Susceptibility of stromelysin 1-deficient mice to collagen-induced arthritis and cartilage destruction. *Arthritis Rheum* **41**:110–121.
- Mukherjee R, Bruce J, McClister D, Aallen C, Sweterlitsch S, and Saul J (2005) Time-dependent changes in myocardial structure following discrete injury in mice deficient of matrix metalloproteinase-3. *J Mol Cell Cardiol* **39**:259–268.
- Muraguchi T, Takegami Y, Ohtsuka T, Kitajima S, Chandana EPS, Omura A, Miki T, Takahashi R, Matsumoto N, Ludwig A, Noda M, and Takahashi C (2007) RECK modulates Notch signaling during cortical neurogenesis by regulating ADAM10 activity. *Nat Neurosci* **10**:838–845.
- Muri J, and Kopf M (2021) Redox regulation of immunometabolism. *Nat Rev Immunol* **21**:363–381.
- Murphy FR, Issa R, Zhou X, Ratnarajah S, Arthur MJP, Benyon C, Iredale JP, and Nagase H (2002) Inhibition of Apoptosis of Activated Hepatic Stellate Cells by Tissue Inhibitor of Metalloproteinase-1 Is Mediated via Effects on Matrix Metalloproteinase Inhibition. *J Biol Chem* **277**:11069–11076.
- Murphy G, Allan JA, Willenbrock F, Cockett MI, O’Connell JP, and Docherty AJ (1992) The role of the C-terminal domain in collagenase and stromelysin specificity. *J Biol Chem* **267**:9612–9618.
- Murphy G, Cockett MI, Stephens PE, Smith BJ, and Docherty AJP (1987) Stromelysin is an activator of procollagenase. A study with natural and recombinant enzymes. *Biochem J* **248**:265–268.
- Murphy G, and Nagase H (2008) Progress in matrix metalloproteinase research. *Mol Aspects Med* **29**:290–308.
- Nagai Y, Lapiere CM, and Gross J (1966) Tadpole Collagenase. Preparation and Purification *. *Biochemistry* **5**:3123–3130.
- Nagase H, Visse R, and Murphy G (2006) Structure and function of matrix metalloproteinases and TIMPs. *Cardiovasc Res* **69**:562–573.
- Nagase H, and Woessner JF (1999) Matrix metalloproteinases. *J Biol Chem* **274**:21491–21494, ASBMB.
- Nakamoto T, Yamagata T, Sakai R, Ogawa S, Honda H, Ueno H, Hirano N, Yazaki Y, and Hirai H (2000) CIZ, a Zinc Finger Protein That Interacts with p130 cas and Activates the Expression of Matrix Metalloproteinases. *Mol Cell Biol* **20**:1649–1658.
- Nelson AR, Fingleton B, Rothenberg ML, and Matrisian LM (2000) Matrix Metalloproteinases:

- Biologic Activity and Clinical Implications. *J Clin Oncol* **18**:1135–1135.
- Ng HH, Narasaraju T, Phoon MC, Sim MK, Seet JE, and Chow VT (2012) Doxycycline treatment attenuates acute lung injury in mice infected with virulent influenza H3N2 virus: Involvement of matrix metalloproteinases. *Exp Mol Pathol* **92**:287–295.
- Ng SC, Shi HY, Hamidi N, Underwood FE, Tang W, Benchimol EI, Panaccione R, Ghosh S, Wu JCY, Chan FKL, Sung JJY, and Kaplan GG (2017) Worldwide incidence and prevalence of inflammatory bowel disease in the 21st century: a systematic review of population-based studies. *Lancet* **390**:2769–2778.
- Nguyen D, Kim H, Kim D, Lee J, Gye M, and Kim Y-P (2018) Detection of Matrix Metalloproteinase Activity by Bioluminescence via Intein-Mediated Biotinylation of Luciferase. *Sensors* **18**:875.
- Norga K, Paemen L, Masure S, Dillen C, Heremans H, Billiau A, Carton H, Cuzner L, Olsson T, Van Damme J, and Opdenakker G (1995) Prevention of acute autoimmune encephalomyelitis and abrogation of relapses in murine models of multiple sclerosis by the protease inhibitor D-penicillamine. *Inflamm Res* **44**:529–534.
- Nury C, Bregant S, Czarny B, Berthon F, Cassar-Lajeunesse E, and Dive V (2013) Detection of Endogenous Matrix Metalloprotease-12 Active Form with a Novel Broad Spectrum Activity-based Probe*. *J Biol Chem* **288**:5636–5644.
- Nury C, Czarny B, Cassar-Lajeunesse E, Georgiadis D, Bregant S, and Dive V (2013) A Pan Photoaffinity Probe for Detecting Active Forms of Matrix Metalloproteinases. *ChemBioChem* **14**:107–114.
- Nwafor DC, Brichacek AL, Mohammad AS, Griffith J, Lucke-Wold BP, Benkovic SA, Geldenhuys WJ, Lockman PR, and Brown CM (2019) Targeting the Blood-Brain Barrier to Prevent Sepsis-Associated Cognitive Impairment. *J Cent Nerv Syst Dis* **11**:117957351984065.
- O'Sullivan S, Gilmer JF, and Medina C (2015) Matrix Metalloproteinases in Inflammatory Bowel Disease: An Update. *Mediators Inflamm* **2015**:1–19.
- Oger S, Méhats C, Dallot E, Cabrol D, and Leroy M-J (2005) Evidence for a Role of Phosphodiesterase 4 in Lipopolysaccharide-Stimulated Prostaglandin E₂ Production and Matrix Metalloproteinase-9 Activity in Human Amniochorionic Membranes. *J Immunol* **174**:8082–8089.
- Oh J, Takahashi R, Kondo S, Mizoguchi A, Adachi E, Sasahara RM, Nishimura S, Imamura Y, Kitayama H, Alexander DB, Ide C, Horan TP, Arakawa T, Yoshida H, Nishikawa S, Itoh Y, Seiki M, Itohara S, Takahashi C, and Noda M (2001) The Membrane-Anchored MMP Inhibitor RECK Is a Key Regulator of Extracellular Matrix Integrity and Angiogenesis. *Cell* **107**:789–800.
- Okada Y, Morodomi T, Enghild JJ, Suzuki K, Yasui A, Nakanishi I, Salvesen G, and Nagase H (1990) Matrix metalloproteinase 2 from human rheumatoid synovial fibroblasts. Purification and activation of the precursor and enzymic properties. *Eur J Biochem* **194**:721–730.
- Okada Y, Nagase H, and Harris ED (1987) Matrix metalloproteinases 1, 2, and 3 from rheumatoid synovial cells are sufficient to destroy joints. *J Rheumatol* **14**:41–42.
- Okamoto T, Akaike T, Sawa T, Miyamoto Y, van der Vliet A, and Maeda H (2001) Activation of Matrix Metalloproteinases by Peroxynitrite-Induced Protein S-Glutathiolation via Disulfide S-Oxide Formation. *J Biol Chem* **276**:29596–29602.
- Oliveira-Silva R, Sousa-Jerónimo M, Botequim D, Silva NJO, Paulo PMR, and Prazeres DMF

- (2020) Monitoring Proteolytic Activity in Real Time: A New World of Opportunities for Biosensors. *Trends Biochem Sci* **45**:604–618.
- Opdenakker G, and Damme J Van (1994) Cytokine-regulated proteases in autoimmune diseases. *Immunol Today* **15**:103–107.
- Orbe J, Barrenetxe J, Rodriguez JA, Vivien D, Orset C, Parks WC, Birkland TP, Serrano R, Purroy A, Martinez de Lizarrondo S, Angles-Cano E, and Páramo JA (2011) Matrix Metalloproteinase-10 Effectively Reduces Infarct Size in Experimental Stroke by Enhancing Fibrinolysis via a Thrombin-Activatable Fibrinolysis Inhibitor-Mediated Mechanism. *Circulation* **124**:2909–2919.
- Otsuka K, Pitaru S, Overall CM, Aubin JE, and Sodek J (1988) Biochemical comparison of fibroblast populations from different periodontal tissues: characterization of matrix protein and collagenolytic enzyme synthesis. *Biochem Cell Biol* **66**:167–176.
- Overall CM, and Butler GS (2007) Protease Yoga: Extreme Flexibility of a Matrix Metalloproteinase. *Structure* **15**:1159–1161.
- Overall CM, and Kleifeld O (2006a) Towards third generation matrix metalloproteinase inhibitors for cancer therapy. *Br J Cancer* **94**:941–946, Nature Publishing Group.
- Overall CM, and Kleifeld O (2006b) Validating matrix metalloproteinases as drug targets and anti-targets for cancer therapy. *Nat Rev Cancer* **6**:227–239, Nature Publishing Group.
- Overall CM, and López-Otín C (2002) Strategies for MMP inhibition in cancer: innovations for the post-trial era. *Nat Rev Cancer* **2**:657–672.
- Overall CM, and Sodek J (1987) Initial Characterization of a Neutral Metalloproteinase, Active on Native 3/4-Collagen Fragments, Synthesized by ROS 17/2.8 Osteoblastic Cells, Periodontal Fibroblasts, and Identified in Gingival Crevicular Fluid. *J Dent Res* **66**:1271–1282.
- Pacher P, Beckman JS, and Liaudet L (2007) Nitric Oxide and Peroxynitrite in Health and Disease. *Physiol Rev* **87**:315–424.
- Paladini RD, Wei G, Kundu A, Zhao Q, Bookbinder LH, Keller GA, Shepard HM, and Frost GI (2013) Mutations in the Catalytic Domain of Human Matrix Metalloproteinase-1 (MMP-1) That Allow for Regulated Activity through the Use of Ca²⁺. *J Biol Chem* **288**:6629–6639.
- Park JE, Keller GA, and Ferrara N (1993) The vascular endothelial growth factor (VEGF) isoforms: differential deposition into the subepithelial extracellular matrix and bioactivity of extracellular matrix-bound VEGF. *Mol Biol Cell* **4**:1317–1326.
- Park S-B, Jung SH, Jin H, Kim SJ, Ryu Y, Lee K-J, Kim B, Shin HJ, and Won K-J (2021) Bioluminescence Imaging of Matrix Metalloproteinases-2 and -9 Activities in Ethanol-injured Cornea of Mice. *In Vivo (Brooklyn)* **35**:1521–1528.
- Patterton D, Hayes WP, and Shi Y-B (1995) Transcriptional Activation of the Matrix Metalloproteinase Gene stromelysin-3 Coincides with Thyroid Hormone-Induced Cell Death during Frog Metamorphosis. *Dev Biol* **167**:252–262.
- Paul CD, Mistriotis P, and Konstantopoulos K (2017) Cancer cell motility: lessons from migration in confined spaces. *Nat Rev Cancer* **17**:131–140.
- Pavlaki M, Zucker S, Dufour A, Calabrese N, Bahou W, and Cao J (2011) Furin Functions as a Nonproteolytic Chaperone for Matrix Metalloproteinase-28: MMP-28 Propeptide Sequence Requirement. *Biochem Res Int* **2011**:1–12.
- Pavloff N, Staskus PW, Kishnani NS, and Hawkes SP (1992) A new inhibitor of metalloproteinases from chicken: ChIMP-3. A third member of the TIMP family. *J Biol Chem* **267**:17321–17326.

- Peeters SA, Engelen L, Buijs J, Chaturvedi N, Fuller JH, Jorsal A, Parving H-H, Tarnow L, Theilade S, Rossing P, Schalkwijk CG, and Stehouwer CDA (2017) Circulating matrix metalloproteinases are associated with arterial stiffness in patients with type 1 diabetes: pooled analysis of three cohort studies. *Cardiovasc Diabetol* **16**:139.
- Pendás AM, Balbín M, Llano E, Jiménez MG, and López-Otín C (1997) Structural Analysis and Promoter Characterization of the Human Collagenase-3 Gene (MMP13). *Genomics* **40**:222–233.
- Pendás AM, Folgueras AR, Llano E, Caterina J, Frerard F, Rodríguez F, Astudillo A, Noël A, Birkedal-Hansen H, and López-Otín C (2004) Diet-Induced Obesity and Reduced Skin Cancer Susceptibility in Matrix Metalloproteinase 19-Deficient Mice. *Mol Cell Biol* **24**:5304–5313.
- Peppin GJ, and Weiss SJ (1986) Activation of the endogenous metalloproteinase, gelatinase, by triggered human neutrophils. *Proc Natl Acad Sci* **83**:4322–4326.
- Piperigkou Z, Kyriakopoulou K, Koutsakis C, Mastronikolis S, and Karamanos NK (2021) Key Matrix Remodeling Enzymes: Functions and Targeting in Cancer. *Cancers (Basel)* **13**:1441.
- Pochetti G, Montanari R, Gege C, Chevrier C, Taveras AG, and Mazza F (2009) Extra Binding Region Induced by Non-Zinc Chelating Inhibitors into the S 1' Subsite of Matrix Metalloproteinase 8 (MMP-8). *J Med Chem* **52**:1040–1049.
- Popovic N, Schubart A, Goetz BD, Zhang S-C, Lington C, and Duncan ID (2002) Inhibition of autoimmune encephalomyelitis by a tetracycline. *Ann Neurol* **51**:215–223.
- Potente M, Gerhardt H, and Carmeliet P (2011) Basic and Therapeutic Aspects of Angiogenesis. *Cell* **146**:873–887.
- Powell WC, Fingleton B, Wilson CL, Boothby M, and Matrisian LM (1999) The metalloproteinase matrilysin proteolytically generates active soluble Fas ligand and potentiates epithelial cell apoptosis. *Curr Biol* **9**:1441–1447.
- Price A, Shi Q, Morris D, Wilcox ME, Brasher PMA, Rewcastle NB, Shalinsky D, Zou H, Appelt K, Johnston RN, Yong VW, Edwards D, and Forsyth P (1999) Marked inhibition of tumor growth in a malignant glioma tumor model by a novel synthetic matrix metalloproteinase inhibitor AG3340. *Clin Cancer Res* **5**.
- Prior SH, Fulcher YG, Koppiseti RK, Jurkevich A, and Van Doren SR (2015) Charge-Triggered Membrane Insertion of Matrix Metalloproteinase-7, Supporter of Innate Immunity and Tumors. *Structure* **23**:2099–2110.
- Proost P, Struyf S, Van Damme J, Fiten P, Ugarte-Berzal E, and Opdenakker G (2017) Chemokine isoforms and processing in inflammation and immunity. *J Autoimmun* **85**:45–57.
- Puente XS, Sánchez LM, Overall CM, and López-Otín C (2003) Human and mouse proteases: a comparative genomic approach. *Nat Rev Genet* **4**:544–558.
- Pulli B, Bure L, Wojtkiewicz GR, Iwamoto Y, Ali M, Li D, Schob S, Hsieh KL-C, Jacobs AH, and Chen JW (2015) Multiple Sclerosis: Myeloperoxidase Immunoradiology Improves Detection of Acute and Chronic Disease in Experimental Model. *Radiology* **275**:480–489.
- Qiao X, Qin X, She D, Wang R, Zhang X, Zhang L, and Zhang Y (2014) Mass spectrometry-based tag and its application to high efficient peptide analysis – A review. *Talanta* **126**:91–102.
- Quail DF, and Joyce JA (2013) Microenvironmental regulation of tumor progression and metastasis. *Nat Med* **19**:1423–1437.
- Quillard T, Araújo HA, Franck G, Tesmenitsky Y, and Libby P (2014) Matrix

- Metalloproteinase-13 Predominates Over Matrix Metalloproteinase-8 as the Functional Interstitial Collagenase in Mouse Atheromata. *Arterioscler Thromb Vasc Biol* **34**:1179–1186.
- Qun Gao C (2003) Matrix metalloproteinase-2 mediates cytokine-induced myocardial contractile dysfunction. *Cardiovasc Res* **57**:426–433.
- Radomski A, Jurasz P, Sanders EJ, Overall CM, Bigg HF, Edwards DR, and Radomski MW (2002) Identification, regulation and role of tissue inhibitor of metalloproteinases-4 (TIMP-4) in human platelets. *Br J Pharmacol* **137**:1330–1338.
- Raggatt LJ, Jefcoat SC, Choudhury I, Williams S, Tiku M, and Partridge NC (2006) Matrix metalloproteinase-13 influences ERK signalling in articular rabbit chondrocytes. *Osteoarthritis Cartil* **14**:680–689.
- Rahman M, Roller J, Zhang S, Syk I, Menger MD, Jeppsson B, and Thorlacius H (2012) Metalloproteinases regulate CD40L shedding from platelets and pulmonary recruitment of neutrophils in abdominal sepsis. *Inflamm Res* **61**:571–579.
- Rahman M, Zhang S, Chew M, Ersson A, Jeppsson B, and Thorlacius H (2009) Platelet-Derived CD40L (CD154) Mediates Neutrophil Upregulation of Mac-1 and Recruitment in Septic Lung Injury. *Ann Surg* **250**:783–790.
- Rahman M, Zhang S, Chew M, Syk I, Jeppsson B, and Thorlacius H (2013) Platelet shedding of CD40L is regulated by matrix metalloproteinase-9 in abdominal sepsis. *J Thromb Haemost* **11**:1385–1398.
- Rangasamy, Geronimo, Ortín, Coderch, Zapico, Ramos, and de Pascual-Teresa (2019) Molecular Imaging Probes Based on Matrix Metalloproteinase Inhibitors (MMPi). *Molecules* **24**:2982.
- Ravindra KC, Ahrens CC, Wang Y, Ramseier JY, Wishnok JS, Griffith LG, Grodzinsky AJ, and Tannenbaum SR (2018) Chemoproteomics of matrix metalloproteases in a model of cartilage degeneration suggests functional biomarkers associated with posttraumatic osteoarthritis. *J Biol Chem* **293**:11459–11469.
- Rawlings ND, Barrett AJ, and Bateman A (2010) MEROPS: the peptidase database. *Nucleic Acids Res* **38**:D227–D233.
- Razavian M, Bordenave T, Georgiadis D, Beau F, Zhang J, Golestani R, Toczek J, Jung J-J, Ye Y, Kim H-Y, Han J, Dive V, Devel L, and Sadeghi MM (2016) Optical imaging of MMP-12 active form in inflammation and aneurysm. *Sci Rep* **6**:38345.
- Razavian M, Zhang J, Nie L, Tavakoli S, Razavian N, Dobrucki LW, Sinusas AJ, Edwards DS, Azure M, and Sadeghi MM (2010) Molecular Imaging of Matrix Metalloproteinase Activation to Predict Murine Aneurysm Expansion In Vivo. *J Nucl Med* **51**:1107–1115.
- Rebustini IT, Myers C, Lassiter KS, Surmak A, Szabova L, Holmbeck K, Pedchenko V, Hudson BG, and Hoffman MP (2009) MT2-MMP-Dependent Release of Collagen IV NC1 Domains Regulates Submandibular Gland Branching Morphogenesis. *Dev Cell* **17**:482–493.
- Redondo-Muñoz J, Escobar-Díaz E, Samaniego R, Terol MJ, García-Marco JA, and García-Pardo A (2006) MMP-9 in B-cell chronic lymphocytic leukemia is up-regulated by $\alpha 4\beta 1$ integrin or CXCR4 engagement via distinct signaling pathways, localizes to podosomes, and is involved in cell invasion and migration. *Blood* **108**:3143–3151.
- Redondo-Muñoz J, José Terol M, García-Marco JA, and García-Pardo A (2008) Matrix metalloproteinase-9 is up-regulated by CCL21/CCR7 interaction via extracellular signal-regulated kinase-1/2 signaling and is involved in CCL21-driven B-cell chronic lymphocytic leukemia cell invasion and migration. *Blood* **111**:383–386.

- Redondo-Muñoz J, Ugarte-Berzal E, García-Marco JA, del Cerro MH, Van den Steen PE, Opdenakker G, Terol MJ, and García-Pardo A (2008) $\alpha 4\beta 1$ integrin and 190-kDa CD44v constitute a cell surface docking complex for gelatinase B/MMP-9 in chronic leukemic but not in normal B cells. *Blood* **112**:169–178.
- Redondo-Muñoz J, Ugarte-Berzal E, Terol MJ, Van den Steen PE, Hernández del Cerro M, Roderfeld M, Roeb E, Opdenakker G, García-Marco JA, and García-Pardo A (2010) Matrix Metalloproteinase-9 Promotes Chronic Lymphocytic Leukemia B Cell Survival through Its Hemopexin Domain. *Cancer Cell* **17**:160–172.
- Rehman AA, Ahsan H, and Khan FH (2013) Alpha-2-macroglobulin: A physiological guardian. *J Cell Physiol* **228**:1665–1675.
- Remacle A, Murphy G, and Roghi C (2003) Membrane type I-matrix metalloproteinase (MT1-MMP) is internalised by two different pathways and is recycled to the cell surface. *J Cell Sci* **116**:3905–3916.
- Remacle AG, Golubkov VS, Shiryayev SA, Dahl R, Stebbins JL, Chernov A V, Cheltsov A V, Pellecchia M, and Strongin AY (2012) Novel MT1-MMP small-molecule inhibitors based on insights into hemopexin domain function in tumor growth. *Cancer Res* **72**:2339–2349, AACR.
- Rempe RG, Hartz AMS, and Bauer B (2016) Matrix metalloproteinases in the brain and blood–brain barrier: Versatile breakers and makers. *J Cereb Blood Flow Metab* **36**:1481–1507.
- Renckens R, Roelofs JJTH, Florquin S, de Vos AF, Lijnen HR, van't Veer C, and van der Poll T (2006) Matrix Metalloproteinase-9 Deficiency Impairs Host Defense against Abdominal Sepsis. *J Immunol* **176**:3735–3741.
- Rikimaru A, Komori K, Sakamoto T, Ichise H, Yoshida N, Yana I, and Seiki M (2007) Establishment of an MT4-MMP-deficient mouse strain representing an efficient tracking system for MT4-MMP/MMP-17 expression in vivo using β -galactosidase. *Genes to Cells* **12**:1091–1100.
- Rodgers UR, Kevorkian L, Surridge AK, Waters JG, Swingler TE, Culley K, Illman S, Lohi J, Parker AE, and Clark IM (2009) Expression and function of matrix metalloproteinase (MMP)-28. *Matrix Biol* **28**:263–272.
- Rohani MG, McMahan RS, Razumova M V., Hertz AL, Cieslewicz M, Pun SH, Regnier M, Wang Y, Birkland TP, and Parks WC (2015) MMP-10 Regulates Collagenolytic Activity of Alternatively Activated Resident Macrophages. *J Invest Dermatol* **135**:2377–2384.
- Rosenberg GA, and Yang Y (2007) Vasogenic edema due to tight junction disruption by matrix metalloproteinases in cerebral ischemia. *Neurosurg Focus* **22**:1–9.
- Rosenblum G, Van den Steen PE, Cohen SR, Grossmann JG, Frenkel J, Sertchook R, Slack N, Strange RW, Opdenakker G, and Sagi I (2007) Insights into the Structure and Domain Flexibility of Full-Length Pro-Matrix Metalloproteinase-9/Gelatinase B. *Structure* **15**:1227–1236.
- Rouanet-Mehouas C, Czarny B, Beau F, Cassar-Lajeunesse E, Stura EA, Dive V, and Devel L (2017) Zinc–Metalloproteinase Inhibitors: Evaluation of the Complex Role Played by the Zinc-Binding Group on Potency and Selectivity. *J Med Chem* **60**:403–414.
- Rowe RG, and Weiss SJ (2008) Breaching the basement membrane: who, when and how? *Trends Cell Biol* **18**:560–574.
- Rowe RG, and Weiss SJ (2009) Navigating ECM Barriers at the Invasive Front: The Cancer Cell–Stroma Interface. *Annu Rev Cell Dev Biol* **25**:567–595.
- Roy R, Yang J, and Moses MA (2009) Matrix Metalloproteinases As Novel Biomarker s and

- Potential Therapeutic Targets in Human Cancer. *J Clin Oncol* **27**:5287–5297.
- Rozario T, and DeSimone DW (2010) The extracellular matrix in development and morphogenesis: A dynamic view. *Dev Biol* **341**:126–140.
- Rumbaugh J, Turchan-Cholewo J, Galey D, Hillaire C St., Anderson C, Conant K, Nath A, Rumbaugh J, Turchan-Cholewo J, Galey D, Hillaire C St., Anderson C, Conant K, and Nath A (2006) Interaction of HIV Tat and matrix metalloproteinase in HIV neuropathogenesis: a new host defense mechanism. *FASEB J* **20**:1736–1738.
- Saarinen J, Welgus HG, Flizar CA, Kalkkinen N, and Helin J (2001) N-Glycan structures of matrix metalloproteinase-1 derived from human fibroblasts and from HT-1080 fibrosarcoma cells. *Eur J Biochem* **259**:829–840.
- Sabeh F, Li X-Y, Saunders TL, Rowe RG, and Weiss SJ (2009) Secreted Versus Membrane-anchored Collagenases. *J Biol Chem* **284**:23001–23011.
- Saghatelian A, Jessani N, Joseph A, Humphrey M, and Cravatt BF (2004) Activity-based probes for the proteomic profiling of metalloproteases. *Proc Natl Acad Sci* **101**:10000–10005.
- Samolov B, Steen B, Seregard S, van der Ploeg I, Montan P, and Kvanta A (2005) Delayed inflammation-associated corneal neovascularization in MMP-2-deficient mice. *Exp Eye Res* **80**:159–166.
- Sandborn WJ, Bhandari BR, Randall C, Younes ZH, Romanczyk T, Xin Y, Wendt E, Chai H, McKevitt M, Zhao S, Sundry JS, Keshav S, and Danese S (2018) Andecaliximab [Anti-matrix Metalloproteinase-9] Induction Therapy for Ulcerative Colitis: A Randomised, Double-Blind, Placebo-Controlled, Phase 2/3 Study in Patients With Moderate to Severe Disease. *J Crohn's Colitis* **12**.
- Sariahmetoglu M, Crawford BD, Leon H, Sawicka J, Li L, Ballermann BJ, Holmes C, Berthiaume LG, Holt A, Sawicki G, and Schulz R (2007) Regulation of matrix metalloproteinase-2 (MMP-2) activity by phosphorylation. *FASEB J* **21**:2486–2495.
- Sato H, Takino T, Okada Y, Cao J, Shinagawa A, Yamamoto E, and Seiki M (1994) A matrix metalloproteinase expressed on the surface of invasive tumour cells. *Nature* **370**:61–65.
- Savinov AY, Remale AG, Golubkov VS, Krajewska M, Kennedy S, Duffy MJ, Rozanov D V., Krajewski S, and Strongin AY (2006) Matrix Metalloproteinase 26 Proteolysis of the NH₂-Terminal Domain of the Estrogen Receptor β Correlates with the Survival of Breast Cancer Patients. *Cancer Res* **66**:2716–2724.
- Scherer RL, McIntyre JO, and Matrisian LM (2008) Imaging matrix metalloproteinases in cancer. *Cancer Metastasis Rev* **27**:679–690.
- Schlage P, and auf dem Keller U (2015) Proteomic approaches to uncover MMP function. *Matrix Biol* **44–46**:232–238.
- Schreiber S, Siegel CA, FriedenberG KA, Younes ZH, Seidler U, Bhandari BR, Wang K, Wendt E, McKevitt M, Zhao S, Sundry JS, Lee SD, and Loftus E V. (2018) A Phase 2, Randomized, Placebo-Controlled Study Evaluating Matrix Metalloproteinase-9 Inhibitor, Andecaliximab, in Patients With Moderately to Severely Active Crohn's Disease. *J Crohn's Colitis* **12**.
- Schuerle S, Dudani JS, Christiansen MG, Anikeeva P, and Bhatia SN (2016) Magnetically Actuated Protease Sensors for in Vivo Tumor Profiling. *Nano Lett* **16**:6303–6310.
- Schultz GS, and Wysocki A (2009) Interactions between extracellular matrix and growth factors in wound healing. *Wound Repair Regen* **17**:153–162.
- Schulz R (2009) Activation of MMP-2 as a key event in oxidative stress injury to the heart. *Front Biosci* **Volume**:699.

- Scilabra SD, Troeberg L, Yamamoto K, Emonard H, Thøgersen I, Enghild JJ, Strickland DK, and Nagase H (2013) Differential Regulation of Extracellular Tissue Inhibitor of Metalloproteinases-3 Levels by Cell Membrane-bound and Shed Low Density Lipoprotein Receptor-related Protein 1. *J Biol Chem* **288**:332–342.
- Sejalon-Cipolla M, Bruyat P, Bregant S, Malgorn C, Devel L, Subra G, and Cantel S (2021) Targeting out of range biomolecules: Chemical labeling strategies for qualitative and quantitative MALDI MS-based detection. *TrAC Trends Anal Chem* **143**:116399.
- Sela-Passwell N, Kikkeri R, Dym O, Rozenberg H, Margalit R, Arad-Yellin R, Eisenstein M, Brenner O, Shoham T, and Danon T (2012) Antibodies targeting the catalytic zinc complex of activated matrix metalloproteinases show therapeutic potential. *Nat Med* **18**:143–147, Nature Publishing Group.
- Sela-Passwell N, Rosenblum G, Shoham T, and Sagi I (2010) Structural and functional bases for allosteric control of MMP activities: Can it pave the path for selective inhibition? *Biochim Biophys Acta - Mol Cell Res* **1803**:29–38.
- Sela-Passwell N, Trahtenherts A, Krüger A, and Sagi I (2011) New opportunities in drug design of metalloproteinase inhibitors: combination between structure–function experimental approaches and systems biology. *Expert Opin Drug Discov* **6**:527–542.
- Sellers A, Cartwright E, Murphy G, and Reynolds JJ (1977) Evidence that latent collagenases are enzyme-inhibitor complexes. *Biochem J* **163**:303–307.
- Serifova X, Ugarte-Berzal E, Opdenakker G, and Vandooren J (2020) Homotrimeric MMP-9 is an active hitchhiker on alpha-2-macroglobulin partially escaping protease inhibition and internalization through LRP-1. *Cell Mol Life Sci* **77**:3013–3026.
- Sgadari C, Monini P, Tripiciano A, Picconi O, Casabianca A, Orlandi C, Moretti S, Francavilla V, Arancio A, Paniccia G, Campagna M, Bellino S, Meschiari M, Nozza S, Sighinolfi L, Latini A, Muscatello A, Saracino A, Di Pietro M, Galli M, Cafaro A, Magnani M, Ensoli F, and Ensoli B (2019) Continued Decay of HIV Proviral DNA Upon Vaccination With HIV-1 Tat of Subjects on Long-Term ART: An 8-Year Follow-Up Study. *Front Immunol* **10**.
- Shah MA, Bodoky G, Starodub A, Cunningham D, Yip D, Wainberg ZA, Bendell J, Thai D, He J, Bhargava P, and Ajani JA (2021) Phase III Study to Evaluate Efficacy and Safety of Andecaliximab With mFOLFOX6 as First-Line Treatment in Patients With Advanced Gastric or GEJ Adenocarcinoma (GAMMA-1). *J Clin Oncol* **39**:990–1000.
- Shah MA, Starodub A, Sharma S, Berlin J, Patel M, Wainberg ZA, Chaves J, Gordon M, Windsor K, Brachmann CB, Huang X, Vosganian G, Maltzman JD, Smith V, Silverman JA, Lenz H-J, and Bendell JC (2018) Andecaliximab/GS-5745 Alone and Combined with mFOLFOX6 in Advanced Gastric and Gastroesophageal Junction Adenocarcinoma: Results from a Phase I Study. *Clin Cancer Res* **24**:3829–3837.
- Shan X, Tomlinson L, Yang Q, and Colognato H (2018) Distinct Requirements for Extracellular and Intracellular MMP12 in the Development of the Adult V-SVZ Neural Stem Cell Niche. *Stem Cell Reports* **10**:984–999.
- Shi J, Son M-Y, Yamada S, Szabova L, Kahan S, Chrysovergis K, Wolf L, Surmak A, and Holmbeck K (2008) Membrane-type MMPs enable extracellular matrix permissiveness and mesenchymal cell proliferation during embryogenesis. *Dev Biol* **313**:196–209.
- Shimizu-Hirota R, Xiong W, Baxter BT, Kunkel SL, Maillard I, Chen X.-W., Sabeh F, Liu R, Li X-Y, and Weiss SJ (2012) MT1-MMP regulates the PI3K {middle dot}Mi-2/NuRD-dependent control of macrophage immune function. *Genes Dev* **26**:395–413.
- Shimizu-Hirota R, Xiong W, Baxter BT, Kunkel SL, Maillard I, Chen Xiao-Wei, Sabeh F, Liu

- R, Li X-Y, and Weiss SJ (2012) MT1-MMP regulates the PI3K δ -Mi-2/NuRD-dependent control of macrophage immune function. *Genes Dev* **26**:395–413.
- Shimoda M, Ohtsuka T, Okada Y, and Kanai Y (2021) Stromal metalloproteinases: Crucial contributors to the tumor microenvironment. *Pathol Int* **71**:1–14.
- Shimoda M, Principe S, Jackson HW, Luga V, Fang H, Molyneux SD, Shao YW, Aiken A, Waterhouse PD, Karamboulas C, Hess FM, Ohtsuka T, Okada Y, Ailles L, Ludwig A, Wrana JL, Kislinger T, and Khokha R (2014) Loss of the Timp gene family is sufficient for the acquisition of the CAF-like cell state. *Nat Cell Biol* **16**:889–901.
- Shibley JM, Wesselschmidt RL, Kobayashi DK, Ley TJ, and Shapiro SD (1996) Metalloelastase is required for macrophage-mediated proteolysis and matrix invasion in mice. *Proc Natl Acad Sci* **93**:3942–3946.
- Si-Tayeb K, Monvoisin A, Mazzocco C, Lepreux S, Decossas M, Cubel G, Taras D, Blanc J-F, Robinson DR, and Rosenbaum J (2006) Matrix Metalloproteinase 3 Is Present in the Cell Nucleus and Is Involved in Apoptosis. *Am J Pathol* **169**:1390–1401.
- Sieber SA, Niessen S, Hoover HS, and Cravatt BF (2006) Proteomic profiling of metalloprotease activities with cocktails of active-site probes. *Nat Chem Biol* **2**:274–281.
- Sies H, and Jones DP (2020) Reactive oxygen species (ROS) as pleiotropic physiological signalling agents. *Nat Rev Mol Cell Biol* **21**:363–383.
- Silbiger SM, Jacobsen VL, Cupples RL, and Koski RA (1994) Cloning of cDNAs encoding human TIMP-3, a novel member of the tissue inhibitor of metalloproteinase family. *Gene* **141**:293–297.
- Silvestre-Roig C, Braster Q, Ortega-Gomez A, and Soehnlein O (2020) Neutrophils as regulators of cardiovascular inflammation. *Nat Rev Cardiol* **17**:327–340.
- Singer M, Deutschman CS, Seymour CW, Shankar-Hari M, Annane D, Bauer M, Bellomo R, Bernard GR, Chiche J-D, Cooper-Smith CM, Hotchkiss RS, Levy MM, Marshall JC, Martin GS, Opal SM, Rubenfeld GD, van der Poll T, Vincent J-L, and Angus DC (2016) The Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3). *JAMA* **315**:801.
- Skiles JW, Gonnella NC, and Jeng AY (2004) The Design, Structure, and Clinical Update of Small Molecular Weight Matrix Metalloproteinase Inhibitors. *Curr Med Chem* **11**:2911–2977.
- Sluijter J, Dekleijn D, and Pasterkamp G (2006) Vascular remodeling and protease inhibition—bench to bedside. *Cardiovasc Res* **69**:595–603.
- Smolen JS, Aletaha D, Barton A, Burmester GR, Emery P, Firestein GS, Kavanaugh A, McInnes IB, Solomon DH, Strand V, and Yamamoto K (2018) Rheumatoid arthritis. *Nat Rev Dis Prim* **4**:18001.
- Sohail A, Sun Q, Zhao H, Bernardo MM, Cho J-A, and Fridman R (2008) MT4-(MMP17) and MT6-MMP (MMP25), A unique set of membrane-anchored matrix metalloproteinases: properties and expression in cancer. *Cancer Metastasis Rev* **27**:289–302.
- Solan PD, Dunsmore KE, Denenberg AG, Odoms K, Zingarelli B, and Wong HR (2012) A novel role for matrix metalloproteinase-8 in sepsis*. *Crit Care Med* **40**:379–387.
- Soleimany AP, and Bhatia SN (2020) Activity-Based Diagnostics: An Emerging Paradigm for Disease Detection and Monitoring. *Trends Mol Med* **26**:450–468.
- Son J, Kalafatovic D, Kumar M, Yoo B, Cornejo MA, Contel M, and Ulijn R V. (2019) Customizing Morphology, Size, and Response Kinetics of Matrix Metalloproteinase-Responsive Nanostructures by Systematic Peptide Design. *ACS Nano* acsnano.8b07401.
- Soria-Valles C, Gutiérrez-Fernández A, Osorio FG, Carrero D, Ferrando AA, Colado E,

- Fernández-García MS, Bonzon-Kulichenko E, Vázquez J, Fueyo A, and López-Otín C (2016) MMP-25 Metalloprotease Regulates Innate Immune Response through NF- κ B Signaling. *J Immunol* **197**:296–302.
- Sousa JE, Serruys PW, and Costa MA (2003) New Frontiers in Cardiology. *Circulation* **107**:2274–2279.
- Sporer B, Paul R, Koedel U, Grimm R, Wick M, Goebel FD, and Pfister H (1998) Presence of Matrix Metalloproteinase-9 Activity in the Cerebrospinal Fluid of Human Immunodeficiency Virus—Infected Patients. *J Infect Dis* **178**:854–857.
- Starr AE, Bellac CL, Dufour A, Goebeler V, and Overall CM (2012) Biochemical characterization and N-terminomics analysis of leukolysin, the membrane-type 6 matrix metalloprotease (MMP25): Chemokine and vimentin cleavages enhance cell migration and macrophage phagocytic activities. *J Biol Chem* **287**:13382–13395.
- Starr AE, Dufour A, Maier J, and Overall CM (2012) Biochemical analysis of matrix metalloproteinase activation of chemokines CCL15 and CCL23 and increased glycosaminoglycan binding of CCL16. *J Biol Chem* **287**:5848–5860.
- Steiner G (2007) Auto-antibodies and autoreactive T-cells in rheumatoid arthritis. *Clin Rev Allergy Immunol* **32**:23–35.
- Sternlicht MD, and Werb Z (2001) How Matrix Metalloproteinases Regulate Cell Behavior. *Annu Rev Cell Dev Biol* **17**:463–516.
- Stetler-Stevenson WG (2008) Tissue Inhibitors of Metalloproteinases in Cell Signaling: Metalloproteinase-Independent Biological Activities. *Sci Signal* **1**:re6–re6.
- Stetler-Stevenson WG, Brown PD, Onisto M, Levy AT, and Liotta LA (1990) Tissue inhibitor of metalloproteinases-2 (TIMP-2) mRNA expression in tumor cell lines and human tumor tissues. *J Biol Chem* **265**:13933–13938.
- Stickens D, Behonick DJ, Ortega N, Heyer B, Hartenstein B, Yu Y, Fosang AJ, Schorpp-Kistner M, Angel P, and Werb Z (2004) Altered endochondral bone development in matrix metalloproteinase 13-deficient mice. *Development* **131**:5883–5895.
- Stöcker W, and Bode W (1995) Structural features of a superfamily of zinc-endopeptidases: the metzincins. *Curr Opin Struct Biol* **5**:383–390.
- Stöcker W, Grams F, Reinemer P, Bode W, Baumann U, Gomis-Rüth F-X, and McKay DB (2008) The metzincins - Topological and sequential relations between the astacins, adamalysins, serralysins, and matrixins (collagenases) define a super family of zinc-peptidases. *Protein Sci* **4**:823–840.
- Strongin AY (2006) Mislocalization and unconventional functions of cellular MMPs in cancer. *Cancer Metastasis Rev* **25**:87–98.
- Strzepa A, Pritchard KA, and Dittel BN (2017) Myeloperoxidase: A new player in autoimmunity. *Cell Immunol* **317**:1–8.
- Subbian S, Tsenova L, Holloway J, Peixoto B, O'Brien P, Dartois V, Khetani V, Zeldis JB, and Kaplan G (2016) Adjunctive Phosphodiesterase-4 Inhibitor Therapy Improves Antibiotic Response to Pulmonary Tuberculosis in a Rabbit Model. *EBioMedicine* **4**.
- Sun Yubo, Sun Yi, Wenger L, Rutter JL, Brinckerhoff CE, and Cheung HS (1999) p53 Down-regulates Human Matrix Metalloproteinase-1 (Collagenase-1) Gene Expression. *J Biol Chem* **274**:11535–11540.
- Suzuki K, Enghild JJ, Morodomi T, Salvesen G, and Nagase H (1990) Mechanisms of activation of tissue procollagenase by matrix metalloproteinase 3 (stromelysin). *Biochemistry* **29**:10261–10270.

- Swanson JA (2008) Shaping cups into phagosomes and macropinosomes. *Nat Rev Mol Cell Biol* **9**:639–649.
- Szklarczyk A, Oyler G, McKay R, Gerfen C, and Conant K (2007) Cleavage of neuronal synaptosomal-associated protein of 25 kDa by exogenous matrix metalloproteinase-7. *J Neurochem* **102**:1256–1263.
- Tagliabracci VS, Wiley SE, Guo X, Kinch LN, Durrant E, Wen J, Xiao J, Cui J, Nguyen KB, Engel JL, Coon JJ, Grishin N, Pinna LA, Pagliarini DJ, and Dixon JE (2015) A Single Kinase Generates the Majority of the Secreted Phosphoproteome. *Cell* **161**:1619–1632.
- Takagi S, Simizu S, and Osada H (2009) RECK Negatively Regulates Matrix Metalloproteinase-9 Transcription. *Cancer Res* **69**:1502–1508.
- Takahashi C, Sheng Z, Horan TP, Kitayama H, Maki M, Hitomi K, Kitaura Y, Takai S, Sasahara RM, Horimoto A, Ikawa Y, Ratzkin BJ, Arakawa T, and Noda M (1998) Regulation of matrix metalloproteinase-9 and inhibition of tumor invasion by the membrane-anchored glycoprotein RECK. *Proc Natl Acad Sci* **95**:13221–13226.
- Takawale A, Fan D, Basu R, Shen M, Parajuli N, Wang W, Wang X, Oudit GY, and Kassiri Z (2014) Myocardial Recovery From Ischemia–Reperfusion Is Compromised in the Absence of Tissue Inhibitor of Metalloproteinase 4. *Circ Hear Fail* **7**:652–662.
- Tallant C, Marrero A, and Gomis-Rüth FX (2010) Matrix metalloproteinases: Fold and function of their catalytic domains. *Biochim Biophys Acta - Mol Cell Res* **1803**:20–28.
- Talmi-Frank D, Altboum Z, Solomonov I, Udi Y, Jaitin DA, Klepfish M, David E, Zhuravlev A, Keren-Shaul H, Winter DR, Gat-Viks I, Mandelboim M, Ziv T, Amit I, and Sagi I (2016) Extracellular Matrix Proteolysis by MT1-MMP Contributes to Influenza-Related Tissue Damage and Mortality. *Cell Host Microbe*, doi: 10.1016/j.chom.2016.09.005.
- Tam EM, Moore TR, Butler GS, and Overall CM (2004) Characterization of the Distinct Collagen Binding, Helicase and Cleavage Mechanisms of Matrix Metalloproteinase 2 and 14 (Gelatinase A and MT1-MMP). *J Biol Chem* **279**:43336–43344.
- Tamura T, and Hamachi I (2019) Chemistry for Covalent Modification of Endogenous/Native Proteins: From Test Tubes to Complex Biological Systems. *J Am Chem Soc* **141**:2782–2799.
- Tan J, Buache E, Alpy F, Dagueneat E, Tomasetto C-L, Ren G-S, and Rio M-C (2014) Stromal matrix metalloproteinase-11 is involved in the mammary gland postnatal development. *Oncogene* **33**:4050–4059.
- Tang J, Liu J, Zhou C, Alexander JS, Nanda A, Granger DN, and Zhang JH (2004) MMP-9 Deficiency Enhances Collagenase-Induced Intracerebral Hemorrhage and Brain Injury in Mutant Mice. *J Cereb Blood Flow Metab* **24**:1133–1145.
- Tang W, Zhao Z, Lian Z, and Liang G (2018) Fluorogenic Disassembly of Self-Quenched Near-Infrared Nanoparticles Enables Matrix Metalloproteinase Detection. *J Biomed Nanotechnol* **14**:304–311.
- Teng YH-F, Aquino RS, and Park PW (2012) Molecular functions of syndecan-1 in disease. *Matrix Biol* **31**:3–16.
- Tester AM, Cox JH, Connor AR, Starr AE, Dean RA, Puente XS, López-Otín C, and Overall CM (2007) LPS Responsiveness and Neutrophil Chemotaxis In Vivo Require PMN MMP-8 Activity. *PLoS One* **2**:e312.
- Tholen M, Yim JJ, Groborz K, Yoo E, Martin BA, Berg NS, Drag M, and Bogoyo M (2020) Design of Optical-Imaging Probes by Screening of Diverse Substrate Libraries Directly in Disease-Tissue Extracts. *Angew Chemie Int Ed* **59**:19143–19152.

- Tian F, Chen Y, Wang W, Zhang J, Jiang T, and Lu Q (2021) Noninvasive Bioluminescence Imaging of Matrix Metalloproteinase-14 Activity in Lung Cancer Using a Membrane-Bound Biosensor. *Anal Chem* **93**:8739–8745.
- Toczek J, Bordenave T, Gona K, Kim H-Y, Beau F, Georgiadis D, Correia I, Ye Y, Razavian M, Jung J-J, Lequin O, Dive V, Sadeghi MM, and Devel L (2019) Novel Matrix Metalloproteinase 12 Selective Radiotracers for Vascular Molecular Imaging. *J Med Chem* **62**:9743–9752.
- Tokito A, and Jougasaki M (2016) Matrix metalloproteinases in non-neoplastic disorders.
- Tokmina-Roszyk M, and Fields GB (2018) Dissecting MMP P10' and P11' subsite sequence preferences, utilizing a positional scanning, combinatorial triple-helical peptide library. *J Biol Chem* **293**:16661–16676.
- Toth M, Bernardo MM, Gervasi DC, Soloway PD, Wang Z, Bigg HF, Overall CM, DeClerck YA, Tschesche H, Cher ML, Brown S, Mobashery S, and Fridman R (2000) Tissue Inhibitor of Metalloproteinase (TIMP)-2 Acts Synergistically with Synthetic Matrix Metalloproteinase (MMP) Inhibitors but Not with TIMP-4 to Enhance the (Membrane Type 1)-MMP-dependent Activation of Pro-MMP-2. *J Biol Chem* **275**:41415–41423.
- Tran EH, Hoekstra K, van Rooijen N, Dijkstra CD, and Owens T (1998) Immune invasion of the central nervous system parenchyma and experimental allergic encephalomyelitis, but not leukocyte extravasation from blood, are prevented in macrophage-depleted mice. *J Immunol* **161**.
- Tranchant I, Vera L, Czarny B, Amoura M, Cassar E, Beau F, Stura EA, and Dive V (2014) Halogen Bonding Controls Selectivity of FRET Substrate Probes for MMP-9. *Chem Biol* **21**:408–413.
- Traub LM (2009) Tickets to ride: selecting cargo for clathrin-regulated internalization. *Nat Rev Mol Cell Biol* **10**:583–596.
- Tressel SL, Kaneider NC, Kasuda S, Foley C, Koukos G, Austin K, Agarwal A, Covic L, Opal SM, and Kuliopulos A (2011) A matrix metalloprotease-PAR1 system regulates vascular integrity, systemic inflammation and death in sepsis. *EMBO Mol Med* **3**:370–384.
- Tsuji K, Aoki T, Tejima E, Arai K, Lee S-R, Atochin DN, Huang PL, Wang X, Montaner J, and Lo EH (2005) Tissue Plasminogen Activator Promotes Matrix Metalloproteinase-9 Upregulation After Focal Cerebral Ischemia. *Stroke* **36**:1954–1959.
- Tsukada H, and Pourmotabbed T (2002) Unexpected Crucial Role of Residue 272 in Substrate Specificity of Fibroblast Collagenase. *J Biol Chem* **277**:27378–27384.
- Turowec JP, Zukowski SA, Knight JDR, Smalley DM, Graves LM, Johnson GL, Li SSC, Lajoie GA, and Litchfield DW (2014) An Unbiased Proteomic Screen Reveals Caspase Cleavage Is Positively and Negatively Regulated by Substrate Phosphorylation. *Mol Cell Proteomics* **13**:1184–1197.
- Udi Y, Grossman M, Solomonov I, Dym O, Rozenberg H, Moreno V, Cuniasse P, Dive V, Arroyo AG, and Sagi I (2015) Inhibition mechanism of membrane metalloprotease by an exosite-swiveling conformational antibody. *Structure* **23**:104–115, Elsevier.
- Üllen A, Singewald E, Konya V, Fauler G, Reicher H, Nusshold C, Hammer A, Kratky D, Heinemann A, Holzer P, Malle E, and Sattler W (2013) Myeloperoxidase-Derived Oxidants Induce Blood-Brain Barrier Dysfunction In Vitro and In Vivo. *PLoS One* **8**:e64034.
- Unkart JT, Chen SL, Wapnir IL, González JE, Harootunian A, and Wallace AM (2017) Intraoperative Tumor Detection Using a Ratiometric Activatable Fluorescent Peptide: A First-in-Human Phase 1 Study. *Ann Surg Oncol* **24**:3167–3173.

- Uría JA, Ferrando AA, Velasco G, Freije JM, and López-Otín C (1994) Structure and expression in breast tumors of human TIMP-3, a new member of the metalloproteinase inhibitor family. *Cancer Res* **54**:2091–4.
- Van den Steen PE, Proost P, Brand DD, Kang AH, Van Damme J, and Opdenakker G (2004) Generation of Glycosylated Remnant Epitopes from Human Collagen Type II by Gelatinase B. *Biochemistry* **43**:10809–10816.
- Van den Steen PE, Proost P, Grillet B, Brand DD, Rang AH, Van Damme J, and Opdenakker G (2002) Cleavage of denatured natural collagen type II by neutrophil gelatinase B reveals enzyme specificity, post-translational modifications in the substrate, and the formation of remnant epitopes in rheumatoid arthritis. *FASEB J* **16**:379–389.
- Van den Steen PE, Van Aelst I, Hvidberg V, Piccard H, Fiten P, Jacobsen C, Moestrup SK, Fry S, Royle L, Wormald MR, Wallis R, Rudd PM, Dwek RA, and Opdenakker G (2006) The Hemopexin and O-Glycosylated Domains Tune Gelatinase B/MMP-9 Bioavailability via Inhibition and Binding to Cargo Receptors. *J Biol Chem* **281**:18626–18637.
- Van den Steen PE, Wuyts A, Husson SJ, Proost P, Van Damme J, and Opdenakker G (2003) Gelatinase B/MMP-9 and neutrophil collagenase/MMP-8 process the chemokines human GCP-2/CXCL6, ENA-78/CXCL5 and mouse GCP-2/LIX and modulate their physiological activities. *Eur J Biochem* **270**:3739–3749.
- Van Lint P, Wielockx B, Puimège L, Noël A, López-Otín C, and Libert C (2005) Resistance of Collagenase-2 (Matrix Metalloproteinase-8)-Deficient Mice to TNF-Induced Lethal Hepatitis. *J Immunol* **175**:7642–7649.
- Vandenbroucke RE, Dejager L, and Libert C (2011) The first MMP in sepsis. *EMBO Mol Med* **3**:367–369.
- Vandenbroucke RE, Dejonckheere E, Van Hauwermeiren F, Lodens S, De Rycke R, Van Wonterghem E, Staes A, Gevaert K, López-Otín C, and Libert C (2013) Matrix metalloproteinase 13 modulates intestinal epithelial barrier integrity in inflammatory diseases by activating TNF. *EMBO Mol Med* **5**:1000–1016.
- Vandenbroucke RE, Dejonckheere E, Van Lint P, Demeestere D, Van Wonterghem E, Vanlaere I, Puimege L, Van Hauwermeiren F, De Rycke R, Mc Guire C, Campestre C, Lopez-Otín C, Matthys P, Leclercq G, and Libert C (2012) Matrix Metalloprotease 8-Dependent Extracellular Matrix Cleavage at the Blood-CSF Barrier Contributes to Lethality during Systemic Inflammatory Diseases. *J Neurosci* **32**:9805–9816.
- Vandenbroucke RE, Vanlaere I, Van Hauwermeiren F, Van Wonterghem E, Wilson C, and Libert C (2014) Pro-inflammatory effects of matrix metalloproteinase 7 in acute inflammation. *Mucosal Immunol* **7**:579–588.
- Vandooren J, Van den Steen PE, and Opdenakker G (2013) Biochemistry and molecular biology of gelatinase B or matrix metalloproteinase-9 (MMP-9): The next decade. *Crit Rev Biochem Mol Biol* **48**:222–272.
- Vankemmelbeke M, Dekeyser MP, Hollander PA, David BJ, and Demeester J (1998) Characterization of helical cleavages in type II collagen generated by matrixins. *Biochem J* **330**:633–640.
- Velasco G, Pendás AM, Fueyo A, Knäuper V, Murphy G, and López-Otín C (1999) Cloning and Characterization of Human MMP-23, a New Matrix Metalloproteinase Predominantly Expressed in Reproductive Tissues and Lacking Conserved Domains in Other Family Members. *J Biol Chem* **274**:4570–4576.
- Vergote D, Butler GS, Ooms M, Cox JH, Silva C, Hollenberg MD, Jhamandas JH, Overall CM,

- and Power C (2006) Proteolytic processing of SDF-1 reveals a change in receptor specificity mediating HIV-associated neurodegeneration. *Proc Natl Acad Sci* **103**:19182–19187.
- Viappiani S, Nicolescu AC, Holt A, Sawicki G, Crawford BD, León H, van Mulligen T, and Schulz R (2009) Activation and modulation of 72kDa matrix metalloproteinase-2 by peroxynitrite and glutathione. *Biochem Pharmacol* **77**:826–834.
- Visse R, and Nagase H (2003) Matrix metalloproteinases and tissue inhibitors of metalloproteinases: structure, function, and biochemistry. *Circ Res* **92**:827–839, Am Heart Assoc.
- Vu TH, Shipley JM, Bergers G, Berger JE, Helms JA, Hanahan D, Shapiro SD, Senior RM, and Werb Z (1998) MMP-9/Gelatinase B Is a Key Regulator of Growth Plate Angiogenesis and Apoptosis of Hypertrophic Chondrocytes. *Cell* **93**:411–422.
- Wang G, Bergman M, Nguyen A, Turcato S, Swigart P, Rodrigo M, Simpson P, Karliner J, Lovett D, and BAKER A (2006) Cardiac transgenic matrix metalloproteinase-2 expression directly induces impaired contractility. *Cardiovasc Res* **69**:688–696.
- Wang L, Main K, Wang H, Julien O, and Dufour A (2021) Biochemical Tools for Tracking Proteolysis. *J Proteome Res* **20**:5264–5279.
- Wang M, Qin X, Mudgett JS, Ferguson TA, Senior RM, and Welgus HG (1999) Matrix metalloproteinase deficiencies affect contact hypersensitivity: Stromelysin-1 deficiency prevents the response and gelatinase B deficiency prolongs the response. *Proc Natl Acad Sci* **96**:6885–6889.
- Wang W, Li D, Xiang L, Lv M, Tao L, Ni T, Deng J, Gu X, Masatara S, Liu Y, and Zhou Y (2019) TIMP-2 inhibits metastasis and predicts prognosis of colorectal cancer via regulating MMP-9. *Cell Adh Migr* **13**:272–283.
- Wang W, Schulze CJ, Suarez-Pinzon WL, Dyck JRB, Sawicki G, and Schulz R (2002) Intracellular Action of Matrix Metalloproteinase-2 Accounts for Acute Myocardial Ischemia and Reperfusion Injury. *Circulation* **106**:1543–1549.
- Wang Y-D (2007) Expression of matrix metalloproteinase-1 and tumor necrosis factor- α in ulcerative colitis. *World J Gastroenterol* **13**:5926.
- Wang Y, Rosen H, Madtes DK, Shao B, Martin TR, Heinecke JW, and Fu X (2007) Myeloperoxidase Inactivates TIMP-1 by Oxidizing Its N-terminal Cysteine Residue. *J Biol Chem* **282**:31826–31834.
- Warner RL, Beltran L, Younkin EM, Lewis CS, Weiss SJ, Varani J, and Johnson KJ (2001) Role of Stromelysin 1 and Gelatinase B in Experimental Acute Lung Injury. *Am J Respir Cell Mol Biol* **24**:537–544.
- Warren AD, Kwong GA, Wood DK, Lin KY, and Bhatia SN (2014) Point-of-care diagnostics for noncommunicable diseases using synthetic urinary biomarkers and paper microfluidics. *Proc Natl Acad Sci* **111**:3671–3676.
- Weaver A, Da Silva AG, Nuttall RK, Edwards DR, Shapiro SD, Rivest S, and Yong VW (2005) An elevated matrix metalloproteinase (MMP) in an animal model of multiple sclerosis is protective by affecting Th1/Th2 polarization. *FASEB J* **19**:1668–1670.
- Wee Yong V, Forsyth PA, Bell R, Krekoski CA, and Edwards DR (1998) Matrix metalloproteinases and diseases of the CNS. *Trends Neurosci* **21**:75–80.
- Weeks CS, Tanabe H, Cummings JE, Crampton SP, Sheynis T, Jelinek R, Vanderlick TK, Cocco MJ, and Ouellette AJ (2006) Matrix Metalloproteinase-7 Activation of Mouse Paneth Cell Pro- α -defensins. *J Biol Chem* **281**:28932–28942.

- Weiss SJ, Peppin G, Ortiz X, Ragsdale C, and Test ST (1985) Oxidative Autoactivation of Latent Collagenase by Human Neutrophils. *Science (80-)* **227**:747–749.
- Wielockx B, Lannoy K, Shapiro SD, Itoh T, Itohara S, Vandekerckhove J, and Libert C (2001) Inhibition of matrix metalloproteinases blocks lethal hepatitis and apoptosis induced by tumor necrosis factor and allows safe antitumor therapy. *Nat Med* **7**:1202–1208.
- Wigle TJ, Provencher LM, Norris JL, Jin J, Brown PJ, Frye S V., and Janzen WP (2010) Accessing Protein Methyltransferase and Demethylase Enzymology Using Microfluidic Capillary Electrophoresis. *Chem Biol* **17**:695–704.
- Wilde C, Hawkins P, Coleman RT, Levine W, Delegeane A, Okamoto P, Ito L, Scott R, and Seilhamer J (1994) Cloning and Characterization of Human Tissue Inhibitor of Metalloproteinases-3. *DNA Cell Biol* **13**:711–718.
- Wilkinson M, and Bywaters EGL (1958) Clinical Features and Course of Ankylosing Spondylitis: As seen in a Follow-up of 222 Hospital Referred Cases. *Ann Rheum Dis* **17**:209–228.
- Williams KC, and Coppolino MG (2011) Phosphorylation of Membrane Type 1-Matrix Metalloproteinase (MT1-MMP) and Its Vesicle-associated Membrane Protein 7 (VAMP7)-dependent Trafficking Facilitate Cell Invasion and Migration. *J Biol Chem* **286**:43405–43416.
- Wilson CL (1999) Regulation of Intestinal -Defensin Activation by the Metalloproteinase Matrilysin in Innate Host Defense. *Science (80-)* **286**:113–117.
- Wilson CL, Heppner KJ, Labosky PA, Hogan BLM, and Matrisian LM (1997) Intestinal tumorigenesis is suppressed in mice lacking the metalloproteinase matrilysin. *Proc Natl Acad Sci* **94**:1402–1407.
- Wiseman BS, Sternlicht MD, Lund LR, Alexander CM, Mott J, Bissell MJ, Soloway P, Itohara S, and Werb Z (2003) Site-specific inductive and inhibitory activities of MMP-2 and MMP-3 orchestrate mammary gland branching morphogenesis. *J Cell Biol* **162**:1123–1133.
- Woessner J (1962) Catabolism of collagen and non-collagen protein in the rat uterus during post-partum involution. *Biochem J* **83**:304–314.
- Wojtowicz-Praga S, Torri J, Johnson M, Steen V, Marshall J, Ness E, Dickson R, Sale M, Rasmussen HS, Chiodo TA, and Hawkins MJ (1998) Phase I trial of Marimastat, a novel matrix metalloproteinase inhibitor, administered orally to patients with advanced lung cancer. *J Clin Oncol* **16**:2150–2156.
- Xie Y, Mustafa A, Yerzhan A, Merzhakupova D, Yerlan P, N Orakov A, Wang X, Huang Y, and Miao L (2017) Nuclear matrix metalloproteinases: functions resemble the evolution from the intracellular to the extracellular compartment. *Cell Death Discov* **3**:17036.
- Xu H, Snider TN, Wimer HF, Yamada SS, Yang T, Holmbeck K, and Foster BL (2016) Multiple essential MT1-MMP functions in tooth root formation, dentinogenesis, and tooth eruption. *Matrix Biol* **52–54**:266–283.
- Xu P, Liu J, Sakaki-Yumoto M, and Derynck R (2012) TACE Activation by MAPK-Mediated Regulation of Cell Surface Dimerization and TIMP3 Association. *Sci Signal* **5**.
- Yagi Y, Andoh A, Inatomi O, Tsujikawa T, and Fujiyama Y (2007) Inflammatory responses induced by interleukin-17 family members in human colonic subepithelial myofibroblasts. *J Gastroenterol* **42**:746–753.
- Yang S, Wang L, Pan W, Bayer W, Thoens C, Heim K, Dittmer U, Timm J, Wang Q, Yu Q, Luo J, Liu Y, Hofmann M, Thimme R, Zhang X, Chen H, Wang H, Feng X, Yang X, Lu Y, Lu M, Yang D, and Liu J (2019) MMP2/MMP9-mediated CD100 shedding is crucial for

- inducing intrahepatic anti-HBV CD8 T cell responses and HBV clearance. *J Hepatol* **71**:685–698.
- Yang Y, and Rosenberg GA (2015) Matrix metalloproteinases as therapeutic targets for stroke. *Brain Res* **1623**:30–38.
- Yang Z, Strickland DK, and Bornstein P (2001) Extracellular Matrix Metalloproteinase 2 Levels Are Regulated by the Low Density Lipoprotein-related Scavenger Receptor and Thrombospondin 2. *J Biol Chem* **276**:8403–8408.
- Yaras N, Sariahmetoglu M, Bilginoglu A, Aydemir-Koksoy A, Onay-Besikci A, Turan B, and Schulz R (2008) Protective action of doxycycline against diabetic cardiomyopathy in rats. *Br J Pharmacol* **155**:1174–1184.
- Yazdan-Ashoori P, Liaw P, Toltl L, Webb B, Kilmer G, Carter DE, and Fraser DD (2011) Elevated plasma matrix metalloproteinases and their tissue inhibitors in patients with severe sepsis. *J Crit Care* **26**:556–565.
- Yin L, Sun H, Zhang H, He L, Qiu L, Lin J, Xia H, Zhang Y, Ji S, Shi H, and Gao M (2019) Quantitatively Visualizing Tumor-Related Protease Activity in Vivo Using a Ratiometric Photoacoustic Probe. *J Am Chem Soc* **141**:3265–3273.
- Yong VW, Power C, Forsyth P, and Edwards DR (2001) Metalloproteinases in biology and pathology of the nervous system. *Nat Rev Neurosci* **2**:502–511.
- Yoo J, Perez CER, Nie W, Sinnott-Smith J, and Rozengurt E (2011) Protein kinase D1 mediates synergistic MMP-3 expression induced by TNF- α and bradykinin in human colonic myofibroblasts. *Biochem Biophys Res Commun* **413**:30–35.
- Young D, Das N, Anowai A, and Dufour A (2019) Matrix Metalloproteases as Influencers of the Cells' Social Media. *Int J Mol Sci* **20**:3847.
- Yu G, Kovkarova-Naumovski E, Jara P, Parwani A, Kass D, Ruiz V, Lopez-Otín C, Rosas IO, Gibson KF, Cabrera S, Ramírez R, Yousem SA, Richards TJ, Chensny LJ, Selman M, Kaminski N, and Pardo A (2012) Matrix Metalloproteinase-19 Is a Key Regulator of Lung Fibrosis in Mice and Humans. *Am J Respir Crit Care Med* **186**:752–762.
- Yu Q, and Stamenkovic I (2000) Cell surface-localized matrix metalloproteinase-9 proteolytically activates TGF- β and promotes tumor invasion and angiogenesis. *Genes Dev* **14**.
- Yu W-H, Yu SC, Meng Q, Brew K, and Woessner JF (2000) TIMP-3 Binds to Sulfated Glycosaminoglycans of the Extracellular Matrix. *J Biol Chem* **275**:31226–31232.
- Zaiss MM, Joyce Wu H-J, Mauro D, Schett G, and Ciccia F (2021) The gut–joint axis in rheumatoid arthritis. *Nat Rev Rheumatol* **17**:224–237.
- Zarrabi K, Dufour A, Li J, Kuscu C, Pulkoski-Gross A, Zhi J, Hu Y, Sampson NS, Zucker S, and Cao J (2011) Inhibition of matrix metalloproteinase 14 (MMP-14)-mediated cancer cell migration. *J Biol Chem* **286**:33167–33177, ASBMB.
- Zhang H, Ray A, Miller NM, Hartwig D, Pritchard KA, and Dittel BN (2016) Inhibition of myeloperoxidase at the peak of experimental autoimmune encephalomyelitis restores blood-brain barrier integrity and ameliorates disease severity. *J Neurochem* **136**:826–836.
- Zhang K, McQuibban GA, Silva C, Butler GS, Johnston JB, Holden J, Clark-Lewis I, Overall CM, and Power C (2003) HIV-induced metalloproteinase processing of the chemokine stromal cell derived factor-1 causes neurodegeneration. *Nat Neurosci* **6**:1064–1071.
- Zhang R-Y, Liu Y-Y, Li L, Cui W, Zhao K-J, Huang W-C, Gu X-W, Liu W, Wu J, Min D, Mao E-Q, and Tang Y-Q (2010) Increased Levels of Soluble Vascular Endothelial Cadherin are Associated with Poor Outcome in Severe Sepsis. *J Int Med Res* **38**:1497–1506.

- Zhang Y, Fonslow BR, Shan B, Baek M-C, and Yates JR (2013) Protein Analysis by Shotgun/Bottom-up Proteomics. *Chem Rev* **113**:2343–2394.
- Zhao B-Q, Wang S, Kim H-Y, Storrie H, Rosen BR, Mooney DJ, Wang X, and Lo EH (2006) Role of matrix metalloproteinases in delayed cortical responses after stroke. *Nat Med* **12**:441–445.
- Zhen G, and Cao X (2014) Targeting TGF β signaling in subchondral bone and articular cartilage homeostasis. *Trends Pharmacol Sci* **35**:227–236.
- Zhou Q, Fülöp A, and Hopf C (2021) Recent developments of novel matrices and on-tissue chemical derivatization reagents for MALDI-MSI. *Anal Bioanal Chem* **413**:2599–2617.
- Zhu L, Xie J, Swierczewska M, Zhang F, Quan Q, Ma Y, Fang X, Kim K, Lee S, and Chen X (2011) Real-Time Video Imaging of Protease Expression In Vivo. *Theranostics* **1**:18–27.
- Zucker S, Cao J, and Chen W-T (2000) Critical appraisal of the use of matrix metalloproteinase inhibitors in cancer treatment. *Oncogene* **19**:6642–6650.
- Zucker S, Schmidt CE, Dufour A, Kaplan RC, Park HI, and Jiang W (2009) ProMMP-2: TIMP-1 complexes identified in plasma of healthy individuals. *Connect Tissue Res* **50**.

Funding

This work was supported by by an NSERC Discovery Grant (#DGECR-2019-00112), Alberta Innovates AICE concepts (#202102341) and Canadian Institutes of Health Research (#449589).

Financial Disclosure

The authors report no conflict of interest or any financial disclosure.