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Nunataryuk field campaigns: Understanding the origin and fate of terrestrial organic matter in the coastal waters of the Mackenzie Delta region

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Abstract. Climate warming and related drivers of soil thermal change in the Arctic are expected to modify the distribution and dynamics of carbon contained in perennially frozen grounds. Thawing of permafrost in the Mackenzie Delta region of northwestern Canada, coupled with increases in river discharge and coastal erosion, trigger the release of terrestrial organic matter (OMt) from the largest Arctic drainage basin in North America into the Arctic Ocean. While this process is ongoing, well-established, and its rate is accelerating, the fate of the newly-mobilized organic matter, as it transits from the watershed through the delta and into the marine system, remains poorly understood. In the framework of the European Horizon 2020 Nunataryuk programme, and as part of the Work Package 4 (WP4) Coastal Waters theme, four field expeditions were conducted in the Mackenzie Delta region and southern Beaufort Sea from April to September 2019. The temporal sampling design allowed the survey of ambient conditions in the coastal waters under full ice cover prior to the spring freshet, during ice break-up in summer, as well as anterior to the freeze-up period in fall. To capture the fluvial-marine transition zone, and with distinct challenges related to shallow waters and changing seasonal and meteorological conditions, the field sampling was conducted in close partnership with members of the communities of Aklavik, Inuvik and Tuktoyaktuk, using several platforms: helicopters, snowmobiles and small boats. Water column profiles of physical and optical variables were measured in situ, while surface water, groundwater and sediment samples were collected and preserved for the determination of the composition and sources of OMt, including particulate and dissolved organic carbon (POC, DOC), and chromophoric dissolved organic matter (CDOM), as well as a suite of physical, chemical and biological variables. Here we present an overview of the standardized datasets, including hydrographic profiles, remote sensing reflectance, temperature and salinity, particle absorption, nutrients, dissolved organic carbon, particulate organic carbon, particulate organic nitrogen, colored dissolved organic matter absorption, fluorescent dissolved organic matter intensity, suspended particulate matter, total particulate carbon, total particulate nitrogen, stable water isotopes, radon in water, bacterial abundance, and a string of phytoplankton pigments including total chlorophyll. Datasets and related metadata can be found in Juhls et al. 2021. <https://doi.pangaea.de/10.1594/PANGAEA.937587>.

1 Introduction

Major components of the Arctic cryosphere are currently exposed to acute changes due to accelerated climate warming rates at high latitude (Allen et al., 2018; IPCC, 2019; AMAP, 2021). In the Northern Hemisphere, nearly a quarter of the land-mass is influenced by permafrost, perennially cryotic ground (Brown et al., 1997; Gruber, 2012; Obu et al., 2019). Permafrost soils store approximately 60% of the world's soil carbon (C) in 15% of the global soil area (Schoor et al., 2015; McGuire et al., 2018; Hugelius et al., 2014). When frozen, the C contained in this reservoir is stable. But rising air temperatures and associated alterations in soil thermodynamics, as well as modifications in snow regimes (Romanovsky et al., 2010), are now leading to the release of C-rich terrestrial organic matter (OMt) in potentially climate-relevant amounts (Guo et al., 2007; Schoor et al., 2015; Schaefer et al., 2014). While permafrost thaw is ongoing, well-established, and its rate is accelerating (Camill, 2005; Biskaborn et al., 2019), the fate of the newly-mobilized OMt, as it transits from watersheds, to rivers and at the subsurface, into coastal waters of the Arctic Ocean remains poorly understood.



35 Increasing amounts of circulating organic C originating from permafrost thaw delivered to arctic coastal waters via river discharge (Cole et al., 2007; Mcguire et al., 2009; Tank et al., 2016) and rapidly eroding coastlines (Tanski et al., 2019; Wegner et al., 2015) may have significant impacts on both the cycling of C in the Arctic Ocean and the global C budget through complex physical (e.g., weathering, burial, photodegradation) and biogeochemical (e.g., microbial degradation) processes (Guo et al., 2007; Vonk et al., 2014; Stedmon et al., 2011; Tanski et al., 2017). Gaining greater insight into these processes is crucial as they may lead, among other things, to enhanced emissions of CO₂ or other greenhouse gasses (Semiletov et al., 2013; Vonk et al., 2012; Vonk and Gustafsson, 2013; Tanski et al., 2019) and modifications in the productivity of the primary producers (Kipp et al., 2018) that sustain the marine arctic food web, with unknown impacts on the communities that rely on these food sources (Fritz et al., 2017). While past studies have broadened our understanding of the riverine transport of nutrients, organic matter and suspended sediments into coastal waters of the Arctic Ocean (McClelland et al., 2006; Stedmon et al., 2011; Holmes et al., 2012), assessments of the seasonal and interannual variations in OMt fluxes at the Pan-Arctic scale are still limited.

45 During the Canada Arctic Shelf Exchange Study expeditions (CASES, 2002-2004) and the River Delta Experiment (ARDEX, 2004), field sampling and satellite observations were conducted in coastal sites near the Mackenzie River mouth and along the shelf edge into the Beaufort Sea (CASES), as well as in the River's East and Middle Channels and throughout the delta (ARDEX), to explore both the biogeochemistry and photoreactivity of colored dissolved organic matter (CDOM) transported by rivers to Arctic shelf environments (Osburn et al., 2009). Later, in 2009, the MALINA oceanographic campaign took place in the Mackenzie River estuary and the Beaufort Sea systems during summer to document the stocks and the processes controlling carbon fluxes across meridional gradients between the estuary and the open ocean (Massicotte et al., 2021). While the MALINA expedition furthered our insights into the cycling of carbon across shelf-basin systems, it left knowledge gaps regarding the spatial dynamics of OM in the delta area of the Mackenzie, where the network of rivers empties its water and sediment into the Arctic Ocean. Furthermore, lacking from the otherwise extensive MALINA, CASES and ARDEX datasets were measurements of the seasonal variations in OM sources, types and fluxes, particularly during the spring freshet when 55 river discharge rates peak.

While conducting fieldwork in key areas of the Mackenzie Delta region is challenging due to limited accessibility, estimating the fluxes of organic carbon, including particulate (POC) and dissolved (DOC) forms, to arctic coastal waters via satellite observations is also challenging due to the complexity of the optical properties of marginal environments, frequent cloud cover and low sun elevation (IOCCG, 2015). Nevertheless, knowledge of arctic shelf and coastal water optical properties has 60 recently been expanded (Matsuoka et al., 2011, 2014; Juhls et al., 2019), providing the foundation for algorithms estimating DOC and POC concentration using satellite ocean color data (Doxaran et al., 2012; Matsuoka et al., 2013, 2017). Moving forward, the expansion of ground truthing data to train remote-sensing algorithms is paramount to gain synoptic views in study regions with challenging accessibility. Such is the case for the meandering and complex network of rivers, transitional fluvial-to-marine zone, and coastal waters that characterize the Mackenzie Delta region, the largest Arctic drainage basin in North 65 America. Limited accessibility to the coastal waters adjacent to the delta watershed has resulted in poor spatial coverage of this highly dynamic zone. As such, our understanding of organic matter transformation and processing along the land-ocean aquatic continuum, and specifically in the delta where fresh and saline waters meet, has been hampered (Fritz et al., 2017).



Scientists working in the framework of the European Horizon 2020 Nunataryuk program, along with partners from the Inuvialuit Nunangit Sannaiqtuaq communities of Aklavik, Inuvik and Tuktoyaktuk (Inuvialuit Settlement Region (ISR), Northwest Territories, Canada), conducted four expeditions in the coastal waters of the Beaufort Sea adjacent to the Mackenzie Delta region, from April to September 2019. The seasonal field campaign aimed to determine the amount and quality of OM supplied to the Arctic Ocean via riverine and diffuse inputs (e.g., including submarine groundwater discharges that were investigated in July/August 2019) of the Mackenzie Delta region. Ultimately, the in situ characterization of OM (i.e., its origin, age, reactivity, lability and optical properties), will feed the development of optical remote sensing algorithms and numerical modeling endeavors. In this article, we present an overview of the broad dataset acquired during the four field surveys conducted as part of the fourth Nunataryuk Work Package - WP4 Coastal Waters - in the Mackenzie Delta region. To our knowledge, the data collected in this difficult-to-access area and included in this paper are unprecedented, both in their spatial coverage of the fluvial-to-marine transition zone and their seasonal extent.

2 Study area, environmental conditions and sampling strategy

The field campaign was conducted in the coastal waters of the Beaufort Sea adjacent to the Mackenzie Delta region (ISR, Northwest Territories, Canada), within major embayments of the area including Shallow Bay and Mackenzie Bay (in the West), as well as Kittigazuit Bay and Kugmallit Bay (in the East, see Fig. 1) in 2019 (see Fig. 2 for an overview of the sampling). When logistically possible, opportunistic sampling was also conducted in the channels of the Mackenzie River (68.26° N to 69.65° N, and 138.14° W to 133.03° W, Fig. 1). The sampling was conducted following consultations at the Inuvialuit Game Council (8-9 March 2018) as well as a tour (16-23 February 2019) with local members, organizations, committees, municipal entities, and councils of three communities: Aklavik, Inuvik and Tuktoyaktuk. The sampling periods were consensually selected in respect with traditional hunting-fishing grounds and activities of the northern community members of the ISR, and in order to capture salient features of the Mackenzie River discharge dynamics (pre-freshet, freshet, post-freshet, see Fig. 3A), salinity (Fig. 5 and Fig. C1), as well as considering a range of sea ice conditions in the coastal area (Fig. 4). Four field sampling periods were targeted: 1) Leg 1 took place from 17 April to 3 May 2019 (22 stations were sampled within this period, see Fig. 2; see section 4.4.1 for details on sediment sampling); 2) Leg 2 extended from 14 June to 4 July 2019 (40 stations sampled, Fig. 2); 3) Leg 3 took place from 25 July to 8 August 2019 (45 stations sampled, Fig. 2; see section 4.4.2 for details on the parallel groundwater field survey); 4) Leg 4 was conducted from 26 August to 9 September 2019 (39 stations sampled, Fig. 2; see section 4.4.1 for details on sediment sampling).

The sampling conducted during Leg 1 encompassed pre-freshet conditions, as shown in Fig. 3A, with low river discharge, winter freezing air temperatures (Fig. 3B), and 75 to 100% sea ice concentration in the region sampled (Fig. 4A). The second expedition (Leg 2) closely followed the breakup of a thick barrier of rubble ice (stamukhi) that is generated each year by ice convergence at the outer edge of the land-fast sea ice in the coastal area near the Mackenzie river outflow (Reimnitz et al., 1978). Additionally, Leg 2 captured the peak of the annual Mackenzie River discharge ($16,513 \text{ m}^3 \text{ s}^{-1}$, Fig. 3A) warming air temperatures ($>5^{\circ}\text{C}$ and $<10^{\circ}\text{C}$, see Fig. 3B), with on average less than 25% sea ice concentration over the sampling region



(Fig. 4B). Sea ice cover had completely receded over the study site by Leg 3 (Fig. 4C), average air temperatures were at their highest ($>10^{\circ}\text{C}$ and $<15^{\circ}\text{C}$, Fig. 3B), and river discharge was still relatively high ($11,226\text{ m}^3\text{ s}^{-1}$, Fig. 3A). Leg 3 captured the greatest extent in salinity gradient covered by the sampling grid (Figs. 5 and 6A). Environmental conditions during Leg 4 were similar to those encountered during Leg 3 with the absence of sea ice (Fig. 4), relatively high river discharge rates (Fig. 3A),
105 albeit slightly cooler air ($>5^{\circ}\text{C}$ and $<10^{\circ}\text{C}$, Fig. 3B).

Given the wide range of environmental conditions encountered between the months of April and September, flexibility in sampling strategy was adopted and various vehicles (Fig. A1) were used to ensure safe and efficient fieldwork. With the presence of consolidated sea ice in the coastal waters of the Beaufort Sea during Leg 1 (Fig. 4), helicopter flights out of Inuvik to the western sector of the coastal waters (Shallow and Mackenzie bays) were privileged due to the large distance separating
110 the sampling area and the nearest community (Aklavik) as well as the impossibility to set up camp near the coast. Snowmobile trips were made out of Tuktoyaktuk to reach the targeted stations in the eastern sector (Kittigazuit and Kugmallit bays, see Fig. 1). To access the water column underneath the sea ice cover, a hole was drilled through the ice using a battery-powered auger. After each day of sampling, the water and sediment collected were light-protected and brought to the Western Arctic Research Center (WARC) of the Aurora Research Institute in Inuvik - either by air or ground (Inuvik-Tuktoyaktuk highway) - where
115 the existing laboratories were used to immediately process, analyze and/or package samples for later shipment. During Leg 2, the presence of unconsolidated sea ice (Fig. 4) imposed helicopter sampling in stationary flight above the stations (no landing) with the use of specialized hoisting gear for sampling in the western sector. To sample the eastern sector during Leg 2, small local boats (mostly fishing boats) were hired and launched out of Tuktoyaktuk. Finally, during Legs 3 and 4, small local boats were chartered to reach the sampling sites. In the western sector, the field team made camp on Shingle Point (Fig. 1), while
120 in the eastern sector, sampling was conducted out of Tuktoyaktuk. Collected material was protected from the light as well as variations in temperature and sent by air or ground each day to the WARC laboratories in Inuvik for immediate processing. For the groundwater (GW) field survey conducted in parallel during Leg 3, sampling sites were reached using small boats and helicopter flights in the vicinity of Tuktoyaktuk (details in section 4.4.2). Samples were collected and transported to a mobile laboratory built in the Learning Center in Tuktoyaktuk for immediate processing.

125 3 Data quality control and data processing

Coherence and integrity of the data were first visually assessed to remove errors stemming from measurement procedures, methods and instruments. Cleaned data were grouped and structured into ASCII files, each constructed to gather variables of the same type (e.g., nutrients). In each of these files, a minimum number of variables (columns) were always included to help the merging of different datasets (Table 1). More than 150 different variables were measured during the WP4 Nunataryuk 2019
130 field expeditions. The complete list of variables is presented in Table 2 (section 4.4), along with statistical summaries such as average, standard deviation and ranges. The final processed, quality controlled datasets and related metadata are available in Juhls et al. 2021 (including 13 associated datasets). In the following sections, a subset of these variables, along with the methods used to collect and measure them, are presented. Data visualization shown in this article was performed with R 4.2.0

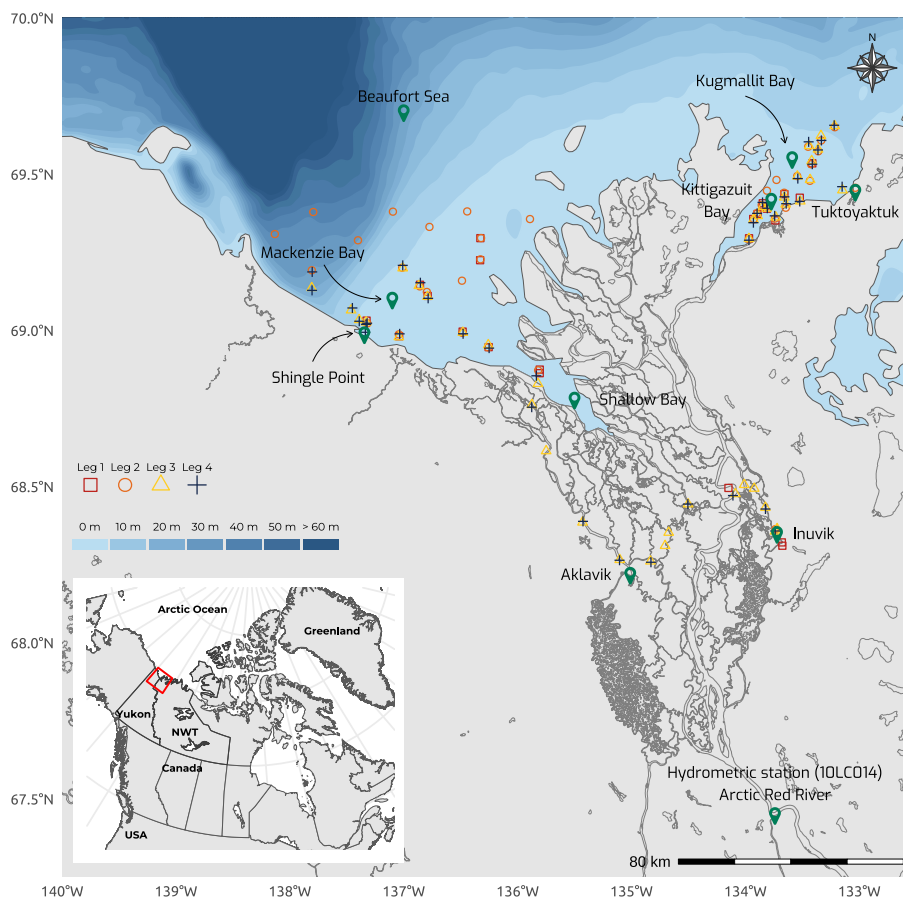


Figure 1. Map of the Mackenzie Delta region (Inuvialuit Settlement Region, Northwest Territories, Canada), with bathymetric features (from 0 to > 60 m) of the coastal waters of the southern Beaufort Sea, showing the sampling stations during the four 2019 WP4 Nunataryuk field expeditions (Legs 1, 2, 3 and 4). Note that the Arctic Red River hydrometric station (10LC014) used to retrieve discharge rates (Fig. 3A) is shown.

(R Core Team, 2022). The code used to process the figures and tables is publicly available (https://github.com/PMassicotte/nunataryuk_data_paper) under the MIT licence. The purpose of this paper is to provide an overview of the availability of data for use in future studies and not to offer an in-depth analysis of the measurements conducted.

4 Data description: an overview

4.1 Physical and optical data

Vertical profiles of underwater conductivity (Figs. 5 and 6A), conservative temperature (Fig. 6B) and depth (CTD) were measured using two RBR sensors: a Maestro model (depth rating of 100 m) during Leg 1, and a Concerto model (depth rating of

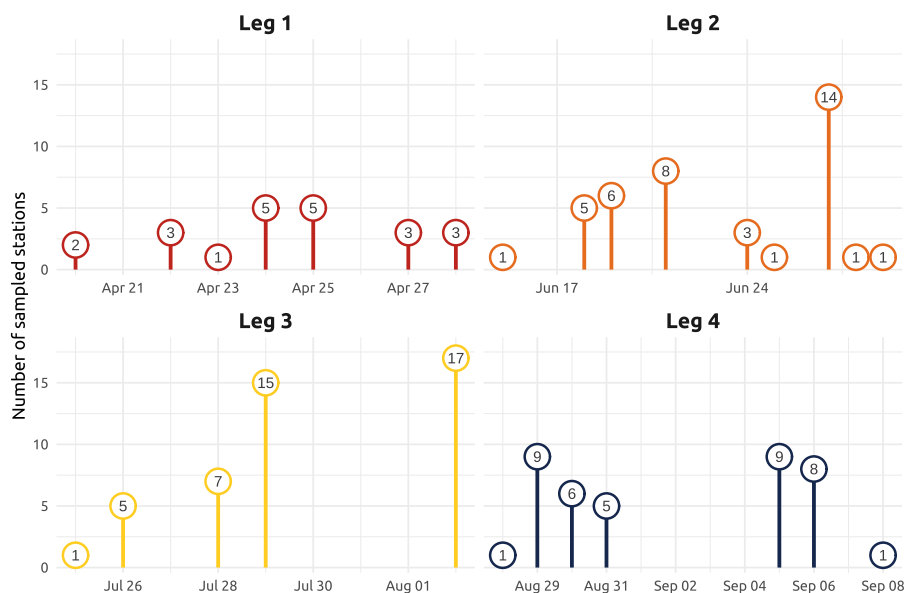


Figure 2. Number of sampled stations for each sampling day of the four main WP4 Nunataryuk expeditions (Legs 1, 2, 3 and 4) in 2019. Note that the groundwater field survey was conducted during Leg 3 in the vicinity of Tuktoyaktuk (Kugmallit Bay), where 29 water samples were collected at the nearshore (see section 4.4.2).

Table 1. Descriptions of the minimal variables included in each data set.

| Variable | Description |
|-------------|--|
| Event | Event identifier (project name_expedition_station) |
| Expedition | Expedition identifier ('1', '2', '3', '4'); also referred to as 'Leg' |
| Latitude | Latitude of the sampling site (degree decimals) |
| Longitude | Longitude of the sampling site (degree decimals) |
| Date/Time | Sampling date and time (UTC), following this format : dd.mm.YYYY HH:MM |
| Sample ID | Sample identifier corresponding to the station code |
| Depth water | Depth (m) at which the measurement was made or the sample taken |

50 m) during Legs 2, 3 and 4. The sensors, chosen for their high vertical resolution, were deployed either alone (Leg 1) or fitted to a frame together with various optical sensors (Legs 2, 3 and 4). As recommended by the instrument manufacturer, the conductivity cell was placed more than 15 cm away from any other structure. Both sensors were factory-calibrated prior to the expeditions and deployed either through the ice (Leg 1) or directly into the water column (Legs 2, 3 and 4) from helicopters or boats. As the stations were relatively shallow, from a few meters to a maximum of 28 m, the descent rate was kept slow, be-

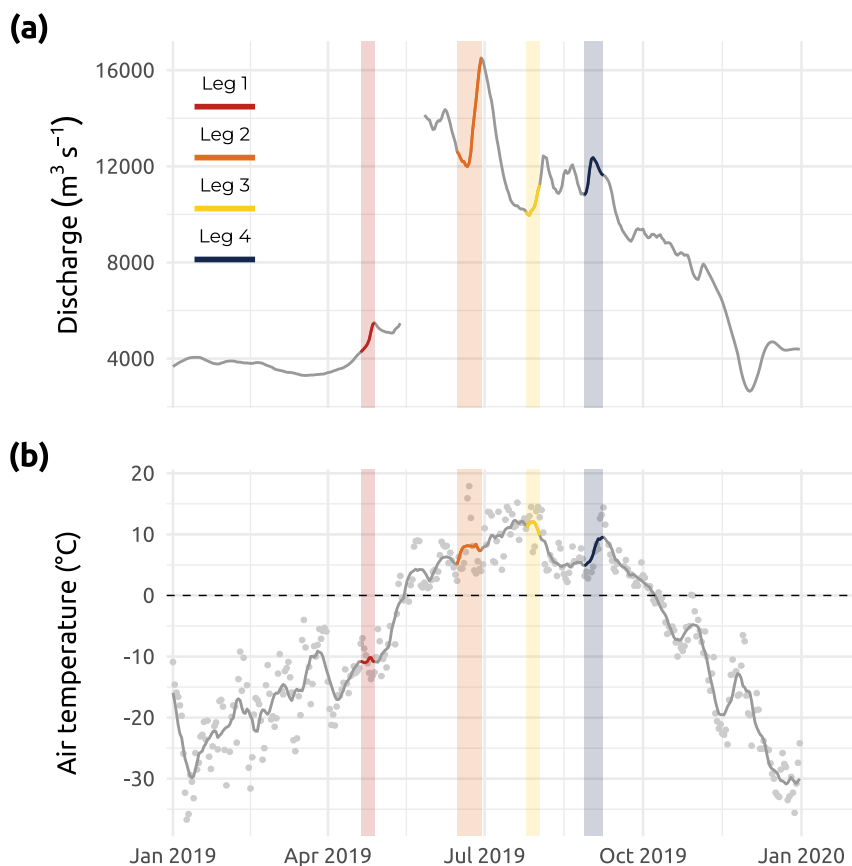


Figure 3. (A) Mackenzie River discharge ($\text{m}^3 \text{sec}^{-1}$) from January 2019 to January 2020 shown as the gray line with overlain colored segments corresponding to Legs 1, 2, 3 and 4 (data from ArcticGRO, Arctic Red River (ID 10LC014: 67.45°N , 133.74°W), Shiklomanov et al. 2021, <https://arcticgreativers.org/discharge/>); (B) Air temperature ($^\circ \text{C}$) from January 2019 to January 2020. The average (solid gray line), spread of air temperature data (gray points) and zero $^\circ \text{C}$ threshold (dotted gray line) are presented. Colored segments correspond to Legs 1, 2, 3 and 4 sampling periods (data from Government of Canada, Environment and Climate Change Canada - Meteorological Service of Canada, measured at Tuktoyaktuk (ID 2203914: 69.43°N , 133.02°W)).

tween 0.05 and 0.20 m s^{-1} . Examples of vertical Temperature-Salinity profiles are presented in Fig. C1 (Appendix C) and show two typical regimes encountered during the sampling period: stratified (Fig. C1A) and well-mixed (Fig. C1B) water columns.

The post-collection processing of the physical data was multi-step. Data were cleaned via a visual inspection of vertical temperature and salinity profile plots, and corrupt data were discarded. Poor-quality data were more frequent during Leg 1
150 sampling, as very low temperatures created frost in the sensor interstices, clogging various elements such as the depth probe opening. Only the downward casts were kept to avoid drag of underlying water masses towards the surface that occurs during upward casts. To correct for slight changes in atmospheric pressure over the span of each leg, and as the sensors do not include

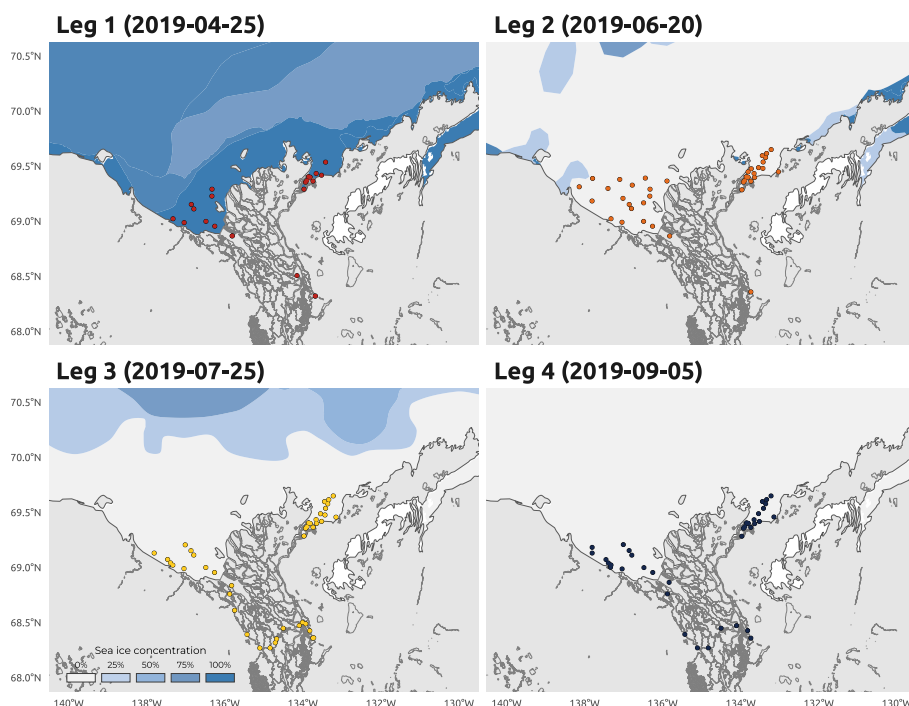


Figure 4. Sea ice concentration (SIC) in the coastal waters of the Beaufort Sea. Colored dots on each panel represent sampled stations during a specific Leg. Dates represent the middle of the sampling period of each Leg of the WP4 Nunataryuk 2019 field campaign. Data sourced from the National Snow and Ice Data Center (<https://nsidc.org/data/G10033>).

a depth tare feature, an empirical correction was applied. This was particularly relevant for shallow stations characterized by extreme physical gradients. To do so, values of atmospheric pressure measured by Environment and Climate Change Canada
155 (ECCC) weather stations near the sampling locations (Aklavik, Inuvik, Shingle Point and Tuktoyaktuk) were used to calibrate the CTD measurements. The atmospheric values collected during deployments were compared to ECCC values to compute and apply an offset (see Fig. B1 in Appendix B). Data spikes were removed (median value on a five points wide running window), and data were further smoothed by a local polynomial regression (using the function “loess” from R), and very occasionally a spline when the loess was not satisfactory) to output the data on a homogeneous 0.01 m step depth grid. Finally, multiple-
160 cast stations (mostly during Leg 1) were averaged. The raw files, metadata files, output files, code files, as well as a detailed processing summary are available on the GitHub public repository https://github.com/GuislainBecu/Nunataryuk_WP4_CTD.

4.1.1 Stable water isotopes

Unfiltered water samples for the determination of stable isotopes were collected in 10 mL HDPE vials free of any chemicals, sealed tightly, and stored in the dark at 4°C. Measurements were conducted at the laboratory facility for stable isotopes at
165 AWI Potsdam (Germany) using a Finnigan MAT Delta-S mass spectrometer equipped with equilibration units for the online

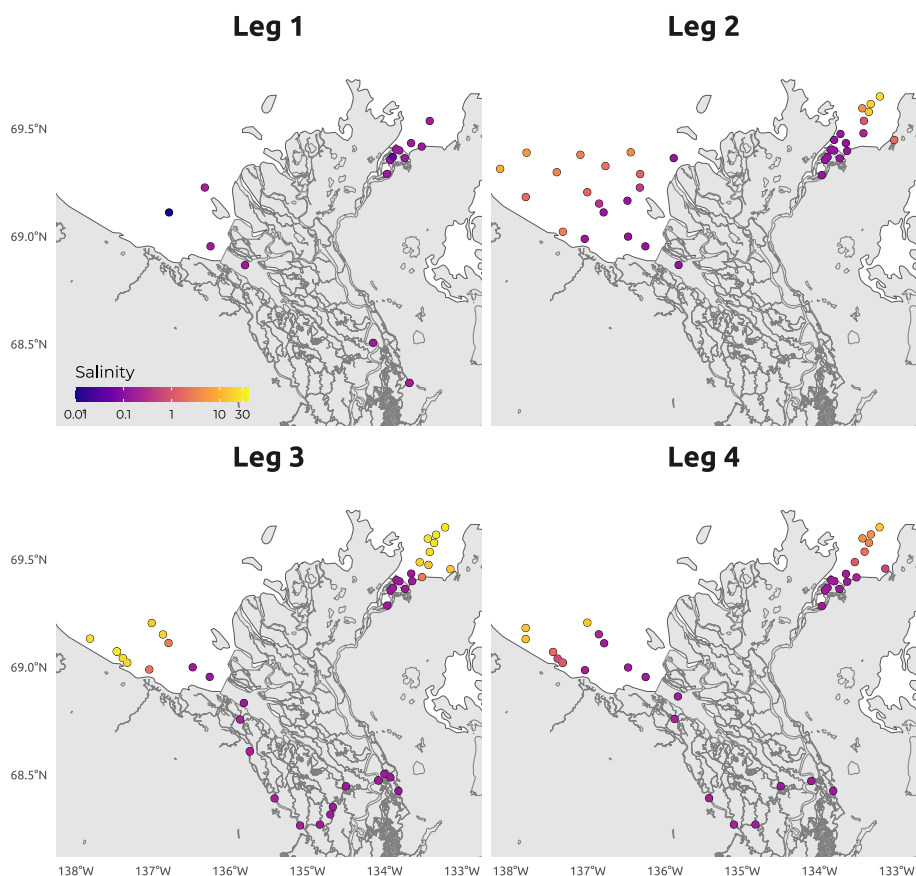


Figure 5. Spatial distribution of the CTD-measured salinity (conductivity) during the four WP4 Nunataryuk expeditions (Legs 1, 2, 3 and 4) in 2019. Note the log10 color scale used to visualize the salinity gradient.

determination of hydrogen and oxygen isotopic composition. Data were given as δD and $\delta^{18}O$ values, which are the per mille difference to standard V-SMOW. The deuterium excess (d-excess) was calculated as follows: $d\text{-excess} = \delta D - 8 * \delta^{18}O$. The measurement accuracy for hydrogen and oxygen isotopes was better than $\pm 0.8\%$ and $\pm 0.1\%$, respectively (Meyer et al., 2000).

170 Values of $\delta^{18}O$ in combination with salinity can be used to distinguish water masses (e.g., river from ocean, and meteoric from sea ice melt water). Furthermore, within freshwater, O can reveal sources of the water that is transported by rivers. Figure 6C shows the ranges of observed $\delta^{18}O$ values, indicating the spectrum of water masses (freshwater with low $\delta^{18}O$ and marine water with higher $\delta^{18}O$) that was encountered during the four expeditions in the Mackenzie Delta region and adjacent coastal waters.

4.1.2 Absorption coefficients for chromophoric dissolved organic matter

175 Water samples for the determination of absorption coefficients for chromophoric dissolved organic matter (a_{CDOM}) were filtered within 12 h of water collection using 0.2 μm GHP filters (Acrodisc Inc.) pre-rinsed with 200 ml of Milli-Q water. Filtered

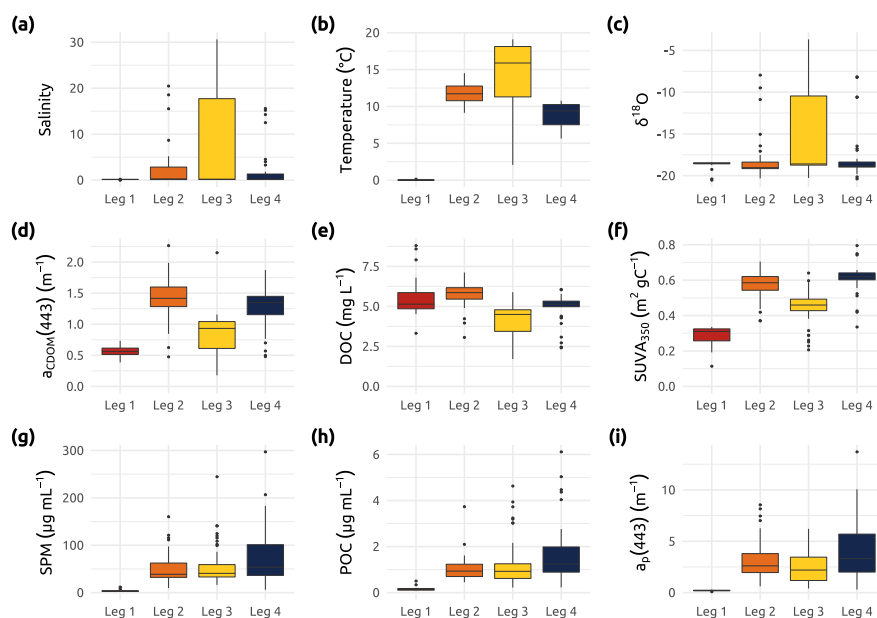


Figure 6. Boxplots showing an overview of the: (A) salinity; (B) water temperature; (C) $\delta^{18}\text{O}$; as well as a set of organic matter related parameters measured during the four Legs; (D) $a_{\text{CDOM}}(443)$ (absorption of colored dissolved organic matter measured at 443 nm); (E) DOC (dissolved organic matter); (F) SUVA_{350} (specific ultraviolet absorbance at 350 nm, i.e. $a_{\text{CDOM}}(350) / \text{DOC}$); (G) SPM (suspended particulate matter); (H) POC (particulate organic carbon); (I) $a_p(443)$ (absorption of particulate matter measured at 443 nm).

samples were then pumped into the sample cell of an UltraPath liquid waveguide system (World Precision Instruments, Inc.) using a peristaltic pump, and the absorption coefficient was measured over the wavelengths ranging from 200 to 722 nm (see Bricaud et al. 2010 and further modifications in Matsuoka et al. 2012 for the complete analytical procedures). For most samples a 200 cm pathlength, which provides data in the visible spectral domain with adequate accuracy and precision, was used. For a limited number of cases, when loss of signal in that spectral domain was observed (due to highly absorbing waters in the delta), a 10 cm pathlength was used instead. Measurements and sample processing were conducted following the protocol Ocean Optics and Biogeochemistry from the International Ocean Color Coordinating Group (Boss et al., 2018). CDOM measurements were fitted using the following equation:

$$a_{\text{CDOM}}(\lambda) = a_{\text{CDOM}}(\lambda_0) \times e^{-S(\lambda-\lambda_0)} \quad (1)$$

where S is the spectral slope of $a_{\text{CDOM}}(\lambda)$, and λ denotes wavelength in nanometers between 350 and 500 nm (Babin et al., 2003; Matsuoka et al., 2012). Coefficients of a_{CDOM} at 443 nm ranged from 0.179 m^{-1} to 2.265 m^{-1} (Table 2) over the study region and broad seasonal extent, with the lowest median value observed during Leg 1 (Fig. 6D). While the highest values are



likely associated with Mackenzie River water inputs during Leg 2, the lowest values were observed offshore beneath the ice
190 cover in April/May (Leg 1).

4.1.3 Dissolved organic carbon

To determine concentrations of dissolved organic carbon (DOC), 20-25 mL of water was collected into a sterile 30-mL syringe
void of a rubber piston. The water was then filtered through a pre-cleaned (acid-washed and MilliQ water rinsed) filter-holder
containing a 25 mm Whatman GF/F (0.7 μm), and acidified with 20 μL Suprapur HCl (10 M) on the same day of sampling. DOC
195 samples were stored and kept at 4°C in the dark during transport until further analysis. Concentration of DOC was measured
using high-temperature catalytic oxidation (TOC-VCPH, Shimadzu) at the Alfred-Wegener-Institute (AWI) Potsdam, Germany.
Blanks (Milli-Q water) and certified reference standards (Battle-02, Mauri-09 or Super-05 from the National Laboratory for
Environmental Testing, Canada) were measured for quality control. Seasonal trends in DOC reveal a range of concentrations
from a minimum of 1.7 mg L⁻¹ to a maximum of 8.8 mg L⁻¹ (Fig. 6E and Table 2) with lowest median concentrations of DOC
200 observed during summer (Leg 3).

4.1.4 Specific ultraviolet absorbance

Specific ultraviolet absorbance at 350 nm (SUVA₃₅₀, m² gC⁻¹) was calculated by dividing the absorbance at 350 nm by DOC
concentration (Weishaar et al., 2003). This index is commonly used as a proxy for assessing the chemical and the biological
reactivity of the DOM pool (see references in Massicotte et al. 2017). Higher values of SUVA₃₅₀ have been previously asso-
205 ciated with a high proportion of aromatic compounds in the DOM pools (Weishaar et al., 2003), especially in estuaries and
coastal areas where the connectivity with the surrounding terrestrial landscape is elevated. In this study, the highest values of
SUVA₃₅₀ (Fig. 6F) coincided with high discharge rates of the Mackenzie River (Fig. 3A). In contrast, low values of SUVA₃₅₀
were observed during Leg 1 at lower discharge. Lower SUVA₃₅₀ during winter and summer could indicate organic matter
sources from groundwater and/or lower soil horizons. High values during high discharge may indicate fresh and young organic
210 matter from surface plant litter (Stedmon et al., 2011; Juhls et al., 2020).

4.1.5 Suspended particulate matter

Water samples for the determination of suspended particulate matter (SPM), as well as concentrations of total particulate
carbon and nitrogen (TPC and TPN, respectively), were filtered through a glass filtration unit on pre-weighed, pre-combusted
blank Whatman GF/F (0.7 μm) 47 mm filters. After filtering volumes of water ranging from 150 to 1000 mL, the filters were
215 transferred to labeled petri dishes (previously acid-washed and MilliQ water rinsed) and placed in the oven to dry overnight
at 60°C before being vacuum sealed for storage and shipment. Analysis for SPM, TPC and TPN was conducted at Laval
University (Quebec City) at the end of 2019. Samples were weighed three times each with a Mettler Toledo microscale after a
final overnight drying at 60°C to remove any leftover moisture. Values for SPM were obtained by subtracting the initial weight
of blank filters from the final weight of the particulate matter laden filter. Due to the presence of large amounts of matter,



220 subsections of the filters were taken with a specialized punching tool. Two punched replicates were placed into tin capsules and processed in a Perkin Elmer elemental analyzer (PE 2400 Series-II CHNS/O Analyzer). Concentrations of TPC and TPN were obtained by extrapolating data from the punched subsection diameter to full filter diameter, assuming uniformity of particulate matter on the filter. As shown in Fig. 6G, Leg 1 was characterized by near zero SPM concentrations, while Legs 2, 3 and 4 exhibited higher median values around 50 $\mu\text{g SPM mL}^{-1}$.

225 4.1.6 Particulate organic matter

Water samples for the determination of particulate organic carbon (POC) and nitrogen (PON) were filtered through a glass filtration unit on pre-combusted Whatman GF/F (0.7 μm) 47 mm filters with filtration volumes ranging from 250 to 1600 mL. Resulting filters were then placed into petri dishes (previously acid-washed and MilliQ water rinsed) and transferred in the oven at 60°C to dry overnight before being vacuum sealed for storage and shipment to Laval University (Quebec City) for analysis.

230 In order to solely preserve the organic fraction of the particulate matter, all samples were acidified with pure HCl (37% w/w) placed into a container at the bottom of a desiccator. Following a 72 h exposure to HCl vapors, the filters were exposed to sodium hydroxide (NaOH) during 72 h to neutralize the samples. Using a punching tool, the filters were sectioned into two sub-samples that were individually wrapped in tin capsules prior to analysis. The 47 mm filters had to be sub-sectioned as they did not fit fully into the analysis capsules. These capsules were processed in a Perkin Elmer elemental analyzer (PE 2400

235 Series-II CHNS/O Analyzer) to obtain masses of carbon and nitrogen. A cross-product calculation was used to extrapolate POC and PON concentrations from the punched subsection diameter to full filter diameter. As shown in Fig. 6H, Leg 1 was characterized by near zero POC concentrations, while Legs 2, 3 and 4 exhibited slightly higher median values around 1 $\mu\text{g POC mL}^{-1}$. Basic information related to PON values can be found in Table 2.

4.1.7 Particulate absorption

240 Coefficients for the absorption of light by particles were obtained within ca. 12 h of water collection using a spectrophotometer (Cary 100, Agilent Technologies Inc.) equipped with a small (60 cm) integrating sphere. Samples were prepared by filtering volumes of water (from approximately 20 to 500 mL, depending on turbidity) onto 25 mm Whatman GF/F filters, with a minimum of 5 stations measured in duplicate or triplicate everyday. To minimize biases due to backscattering by particles retained on the sample filter, the transmittance and reflectance were measured from 350 to 800 nm at 1 nm increments (so-called

245 T-R method; Tassan and Ferrari 1995; Tassan 2002). A recent study demonstrated that the T-R method is particularly useful for turbid waters (Stramski et al., 2015; Boss et al., 2018), such as those found in the Mackenzie River and Delta. The derived absorbance was then converted into absorption coefficients ($a_p(\lambda)$, m^{-1}) by taking into account the volume of water filtered and the clearance area of the filter. The final $a_p(\lambda)$ was determined by applying a beta factor (Mitchell et al., 2003) specific to our instrument set-up (Tassan and Ferrari, 2003), allowing the extrapolation of the absorption of particles concentrated on the

250 filter to what would be in suspension. Figure 6I shows that median coefficients of $a_p(\lambda)$ at 443 nm were significantly higher during Legs 2, 3 and 4, following the spring freshet (after Leg 1). Trends in both $a_p(443)$ and POC concentrations were very similar, as was reflected in the significant linear relationship observed between these two variables (Fig. 7A). Both POC and

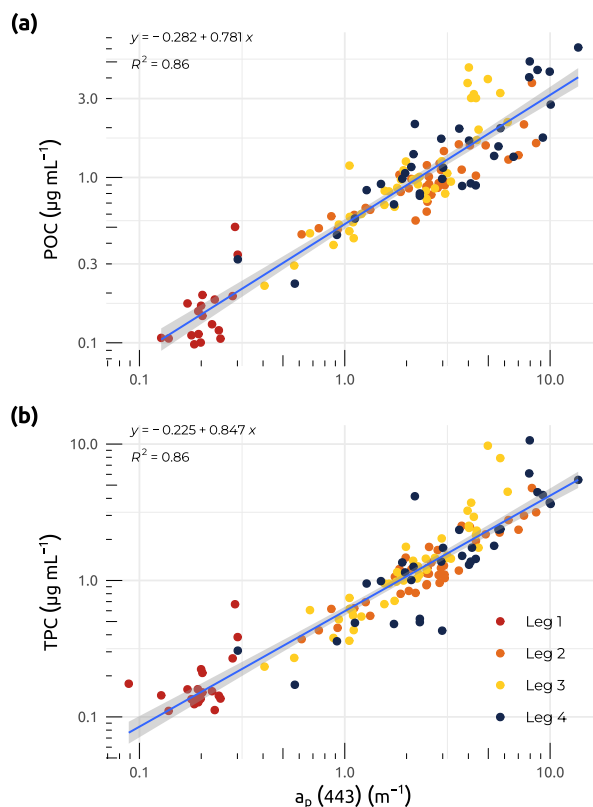


Figure 7. Linear regressions between particle absorption ($a_p(\lambda)$) at 443 nm and: (A) particulate organic carbon (POC); (B) total particulate carbon (TPC) for the four WP4 Nunataryuk expeditions (Legs 1, 2, 3 and 4) in 2019. Equations and coefficients of determination (R^2) are shown. The blue lines represent the linear regressions, whereas the shaded gray areas show the standard error around the regression lines.

total particulate carbon (TPC) showed strong linear relationships ($R^2 = 0.86$, $p < 0.001$) with particle absorption (Fig. 7A and 7B) suggesting that $a_p(443)$ may be a good proxy for particulate carbon. While Leg 1 values show a clear pattern of low POC, TPC and $a_p(443)$, the overall fit of the data points agrees with the linear relationships observed in Fig. 7A and 7B.

4.1.8 Radiometric data

In order to evaluate atmospheric correction algorithms and develop/evaluate algorithms for deriving in-water constituents from reflectance, vertical profiles of downwelling irradiance (E_d) and upwelling radiance (L_u) of the water were measured during boat sampling of Legs 2, 3 and 4 using a Compact-Optical Profiling System (C-OPS, from Biospherical instruments, Inc., see a complete description of the system in Morrow et al. 2010). Measurements of C-OPS radiometric light levels could not be acquired in the waters of Shallow Bay and Mackenzie Bay (western sector) during Leg 2 due to the lack of space aboard the helicopter, as well as for safety reasons associated with free cables outside the cockpit of the hovering aircraft. The C-OPS



system was composed of a set of highly sensitive radiometers, which acquire radiometric measurements in water and in air. In order to correct in-water E_d and L_u for changes in the incident light field during L_u profiling, above-surface downwelling incident irradiance (E_s^{0+}) was measured at about two meters above sea level and above any boat structure during the time of 265 profiling (Zibordi et al., 2019). From the E_d and L_u profiles, apparent optical properties (AOP), such as the diffuse attenuation coefficient or the remote-sensing reflectance, were computed (Bélanger et al., 2017).

The very shallow waters and the often strong currents made the usual free-fall deployment challenging and even risky for the instrument. A custom deployment method was therefore adopted using a negatively buoyant frame manually lowered 270 with a horizontal telescoping mast and a block pulley. Deploying the sensors into the direction of the sun in highly turbid waters (photons mean free path of less than 1 m) avoided any optical pollution from the small ship hull. The processing of the radiometric data was conducted according to recent protocols implemented in an open source package in R available at <https://github.com/belasi01/Cops> (Antoine et al., 2013; Bélanger et al., 2017). Due to the highly turbid waters, the so-called self-shadow correction was not estimated using the well-established Gordon and Ding 1992 method, but was rather estimated 275 using Monte Carlo simulations based on SimulO (Leymarie et al., 2010; Doxaran et al., 2016); see Fig. D1 in Appendix D for further details.

Figure 8 shows the remote sensing reflectance determined following procedures by Antoine et al. 2013 and Bélanger et al. 2017. The highest values of reflectance are present between 550 and 700 nm (green to red) and the spectra show a strong influence of CDOM absorbing light in the shorter spectral domain. This set of remote sensing reflectance measurements can 280 be useful to test and develop algorithms for retrieving water constituents using optical remote sensing and to evaluate the performance of atmospheric correction algorithms.

4.1.9 Inherent optical properties

To measure inherent optical properties (IOP) in the water column, an optical package was deployed during all four legs of the expedition. During Leg 1, the optical package included a very sensitive depth sensor (RBR model Virtuoso), a particle 285 size meter (Sequoia Scientific, model LISST-100x), as well as a data logger (Seabird Scientific, model DH4), all fitted in a cylindrical metallic frame that was manually lowered through an auger hole in the ice using a tripod and a hoisting system. During Legs 2, 3 and 4, the optical package included a different secondary CTD (RBR Concerto), the same particle size meter and data logger, 2 back-scattering meters (a HydroScat-2 from HoboLabs and a BB3 from Seabird Scientific), and a fluorescence meter (FLBBCD from Seabird Scientific). All the sensors were factory calibrated prior to the expedition. The 290 cylindrical frame was lowered at sea using a drill-operated winch during helicopter hovering flights, or using a motorized pulley with a davit when sampling from small fishing boats. In both cases, an electro-mechanical cable allowed for real-time data acquisition and minimal quality control. The instrumentation descent rate was very slow (ca. 0.05-0.20 m s⁻¹) to allow for a maximal resolution of the strong vertical gradients characteristic of several of the stations sampled. See Table 2 for details on availability of data.

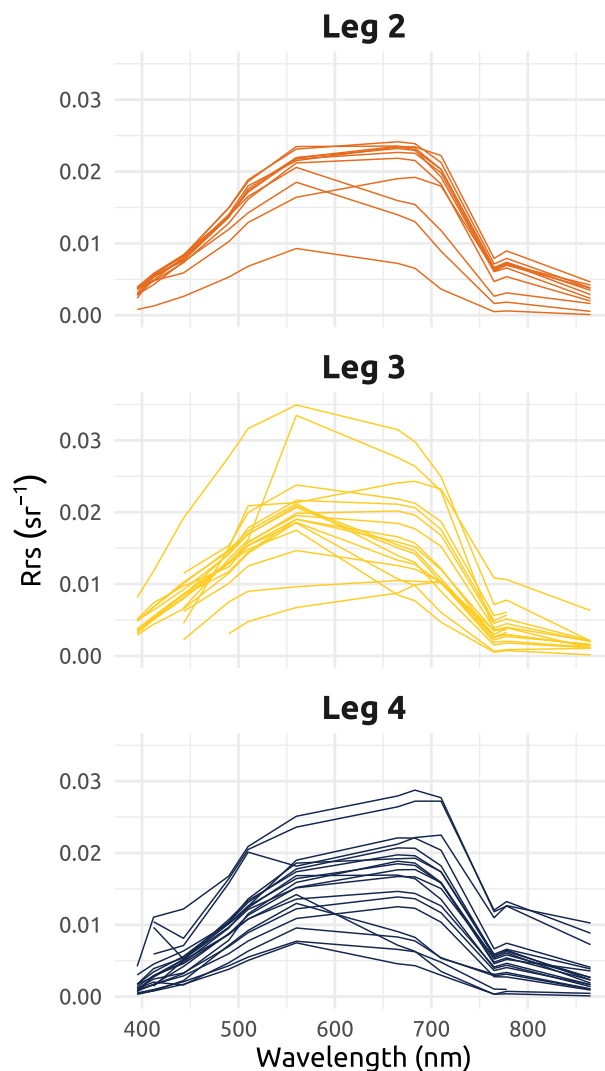


Figure 8. Remote sensing reflectance (R_{rs}) spectra measured using the C-OPS between 395 and 865 nm during Legs 2, 3 and 4. Note that R_{rs} spectra were not measured during Leg 1.

295 4.2 Nutrients

Samples for the determination of nitrate, nitrite, phosphate and silicate concentrations (Fig. 9) were obtained from water filtered through consecutive 0.7 μm Whatman GF/F filters and 0.2 μm cellulose acetate membranes. Filtrates were collected in duplicate sets of sterile 20-mL polyethylene vials, with one set being immediately stored at -20°C , while a second set was poisoned with 100 μL of mercury chloride (60 mg L^{-1}) and subsequently stored in the dark at 4°C until analysis. Nutrient
300 concentrations were determined at Laval University (Quebec City) using an automated colorimetric procedure described in

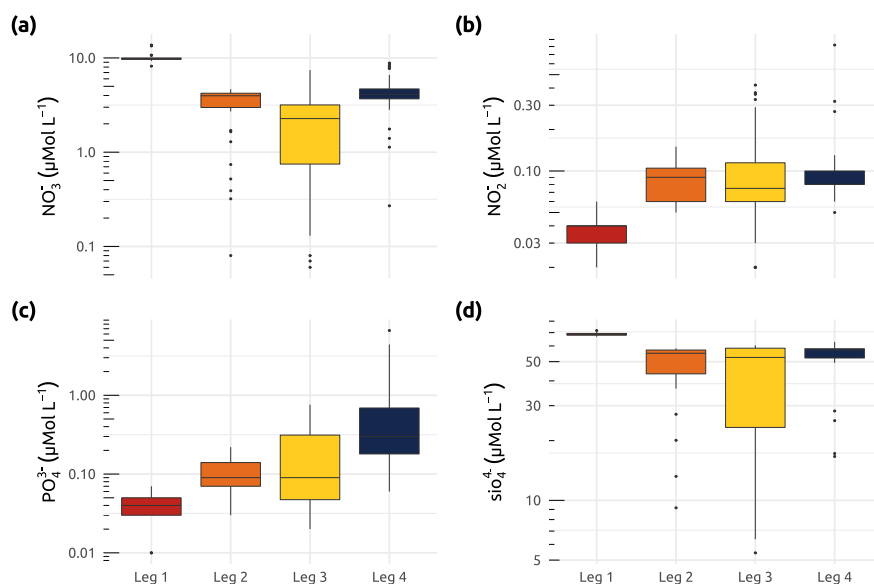


Figure 9. Box plots showing concentrations in $\mu\text{mol L}^{-1}$ of: (A) nitrate (NO_3^-); (B) nitrite (NO_2^-); (C) phosphate (PO_4^{3-}); (D) silicate (SiO_4^{4-}) for the four WP4 Nunataryuk expeditions (Legs 1, 2, 3 and 4) in 2019. Boxes represent the interquartile range (IQR), the horizontal lines show the medians, while the vertical lines show data spread with min/max whiskers.

Downes 1978 and Grasshoff et al. 1999. The winter expedition (Leg 1) was characterized by the highest median concentrations of nitrate and silicate, and the lowest median concentrations of nitrite and phosphate (Fig. 9). Overall, stations explored during Leg 3 exhibited the greatest interquartile ranges for all nutrients measured (Fig. 9).

4.3 Biological data

305 4.3.1 Phytoplankton pigments

The concentration of a suite of phytoplankton pigments, including Chlorophyll *a* (Chl *a*), a proxy for phytoplankton biomass, were determined by high-performance liquid chromatography (HPLC) following the method described in Van Heukelem and Thomas 2001, and further descriptions in Hooker et al. 2005. Volumes of water, ranging from ca. 20 to 500 mL, were filtered onto Whatman GF/F 25 mm filters that were immediately stored at -80°C until analysis at the Ocean Ecology Laboratory of the NASA-Goddard Space Flight Center in Greenbelt (USA). Samples were shipped in liquid nitrogen pre-conditioned dry shippers. Briefly, the HPLC used for pigment analysis was an Agilent RR1200 with a programmable autoinjector (900 μl syringe head), refrigerated autosampler compartment, thermostatted column compartment, quaternary pump with in-line vacuum degasser, and photo-diode array detector with deuterium and tungsten lamps, which collects in-line visible absorbance spectra for each pigment. The HPLC was controlled by Agilent Chemstation software. Calibration was performed with individual pigment standards, whose concentrations have been determined spectrophotometrically using absorption coefficients in common

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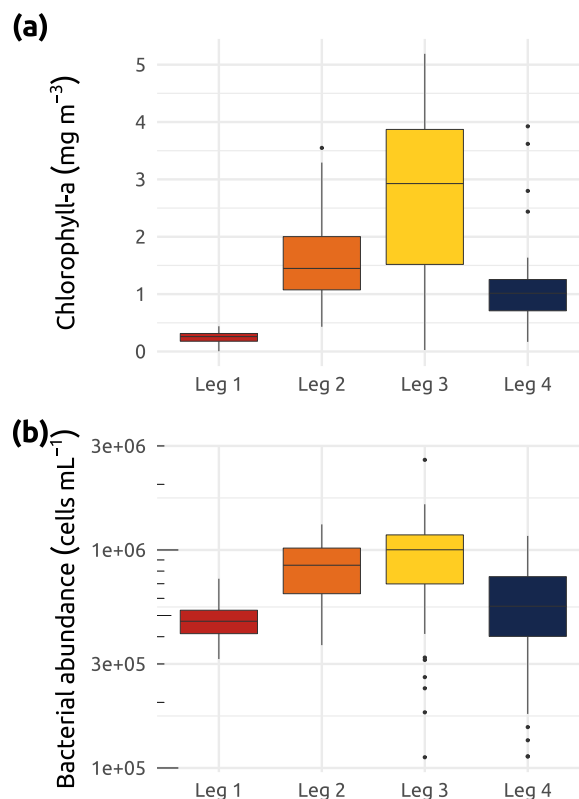


Figure 10. Box plots of : (A) Chlorophyll a (Chl a) concentration (mg m^{-3}); and (B) Bacterial abundance (cells mL^{-1}) for the four WP4 Nunataryuk expeditions (Legs 1, 2, 3 and 4) in 2019. Boxes represent the interquartile range (IQR), the horizontal lines show the medians, while the vertical lines show data spread with min/max whiskers. Notice the y-log₁₀ scale. Also note that a few data points below 100K (Leg 2) were excluded from this graphical presentation.

with those used by most other laboratories (Hooker et al., 2005) and the commercial vendor, DHI Water and Environment (Hørsholm, Denmark). Thirty-six peaks were individually quantified by HPLC, from which 26 pigments were reported (some pigments contained individual components that were summed and reported as one pigment).

320 A full list of accessory pigments analyzed in this study is available in Table 2. Median concentrations of Chl *a* over the study region were at their lowest during winter (Leg 1, Fig. 10), while the greatest interquartile range and highest median concentrations were observed throughout the summer stations (Leg 3, Fig. 10). Despite their seasonal variability, concentrations of Chl *a* were proportionally dominant over all other photosynthetic and non-photosynthetic pigments analyzed across the sampled stations (see example for three selected stations in Fig. 11).

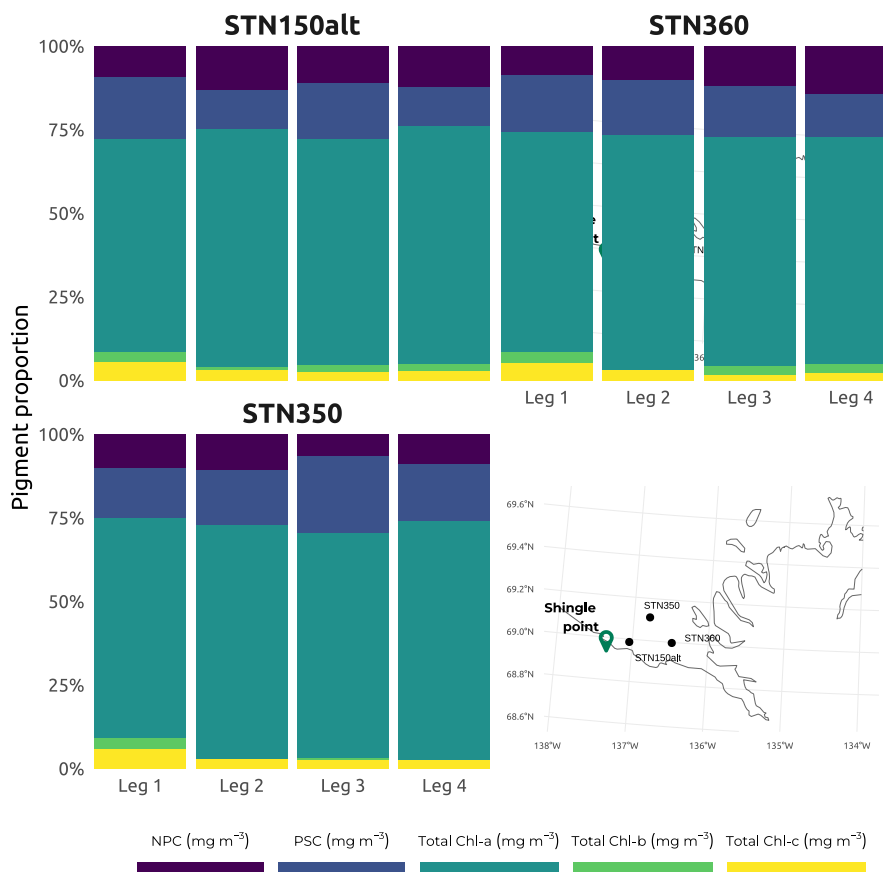


Figure 11. Subset of accessory pigments classified following Matsuoka et al. 2011: non photosynthetic carotenoids (NPC) that include zeaxanthin, diadinoxanthin, alloxanthin; photosynthetic carotenoids (PSC) that include fucoxanthin, peridinin, 19'-hexanoyloxyfucoxanthin, and 19'-butanoyloxyfucoxanthin; Total Chl *a*; Total Chl *b*; and Total Chl *c* (*c*₁, *c*₂, *c*₃) for three selected stations (150ALT, 360 and 350) visited on each of the four WP4 Nunataryuk expeditions (Legs 1, 2, 3 and 4) in 2019. Also included is a map showing the locations of the three stations in the western sector, Mackenzie Bay.

4.3.2 Bacterioplankton abundance and diversity

325 Samples for the determination of bacterial abundance were prepared immediately at the Aurora Research Institute (Inuvik)
 upon reception of the water. A volume of 1.5 mL of water was pipetted into a 2 mL Nunc Cryotube containing 15 µl of 25%
 glutaraldehyde. The Cryotubes were vortexed for 5 s, allowed to sit at room temperature for 10 min, then stored in cryoboxes
 at -80°C before analysis by flow cytometry with SYBR™ Green I (ThermoFisher Scientific) (Gasol and Del Giorgio, 2000) at
 the Institut National de la Recherche Scientifique, Quebec, Canada. Median concentrations of bacterial cells (Fig. 10B) were
 330 lowest during Leg 1 and highest during the summer months (Legs 2 and 3).



Molecular samples for the determination of bacterial RNA-DNA were processed as follows. Volumes of 1.5L of water were collected in clean pre-rinsed Corning pyrex media storage bottles. Glass filtration systems or plastic magnetic filter funnels of 47 mm diameter were cleaned with ELIMINase® and rinsed with MilliQ water. Ten milliliters of water were run through the filtration unit to condition it for each new sample, a procedure that was repeated three times. Then, a 0.2 µm polyethersulfone membrane was placed on the filtration head using tweezers cleaned with ELIMINase® and ethanol. Five hundred (500) mL of water were filtered through the glass system and the filter was then immediately placed into a cryovial. RNALater buffer was added to the cryovial to submerge the filter completely. The filtration process was conducted in triplicate to obtain 3 separate filters for later analysis. Great care was taken during these procedures, including the use of lab coats, hair nets and powder-free gloves cleaned frequently with ethanol. The cryovials were stored in -80°C freezers, shipped in liquid nitrogen pre-conditioned dry-shippers, and stored again in -80°C freezers until their analysis at the Institut National de la Recherche Scientifique in Quebec City.

Bacterial DNA extraction was performed using Qiagen PowerSoil kits following the instructions provided by the manufacturer. DNA extracts were quantified on a Qubit fluorometer and sent to Integrated Microbiome Resource (U. Dalhousie, Halifax, Canada) where library preparation, multiplexing, and sequencing were performed. The sequencing targeted the V4-V5 regions of the 16S rRNA gene using the 515FB (5'-GTGYCAGCMGCCGCGGTAA-3') and 926R (5'-CCGYCAATTYMTTTRAGTTT-3') primer sets (Parada et al., 2016; Walters et al., 2016). Sequencing was performed on an Illumina MiSeq platform, and sequences were then analyzed using the dada2 pipeline v1.16 (Callahan et al., 2016) in R v4.1.2 (R Core Team, 2021) and R-Studio (v1.2.1335). Taxonomy assignment was performed up to the Genhttps://www.overleaf.com/project/626fcc3ad4b482cb8a8e3272us level with SILVA reference database v138 (Quast et al., 2012). Details on Amplicon Sequence Variants (ASVs) of bacterial communities are given in Table 2.

For bacterial isolation, samples (0.5 mL) were placed in cryotubes containing 500 µL of 70% sterile glycerol and stored at -80°C before isolation at the laboratory. After thawing, 100 µL of each sample was spread in triplicate on R2A agar plates adjusted to the salinity of the samples, and the Petri dishes were incubated in the dark at 10°C to allow the development of aerobic heterotrophic psychrotolerant marine bacteria. At regular intervals (one, two and three weeks), the colonies were counted and categorized based on their morphological characteristics (morphotypes). Representatives of each morphotype were selected for isolation by repeated streaking. After isolation and purification, each strain was cultivated in R2A Broth medium (Neogen) at 10°C under agitation and darkness before DNA extraction using the Wizard® Genomic DNA Purification Kit (Promega) and partial sequencing of the 16S rRNA using a Sanger 16 capillary sequencer AB3130XL (Applied Biosystems) to identify the bacterial strains (Tisserand et al., 2020).

360 4.3.3 Fungal abundance and diversity

Following DNA extraction (see above), fungal abundance was evaluated by real-time quantitative PCR (qPCR) using the primer set FungiQuant-F: 5'-GGRAAACTCCACCAGGTCCAG-3' and FungiQuant-R: 5'-GSWCTATCCCCAKCACGA-3' (Liu et al., 2012) and following the protocol described in Maza-Márquez et al. 2020. Amplicon Sequence Variants (ASVs) of fungal communities were determined by Illumina MiSeq sequencing and metabarcoding analysis. After DNA extraction



365 (see above), amplification of fungal internal transcribed spacer 2 (ITS2) region, library preparation, multiplexing, and se-
quencing were performed by LGC Genomics GmbH (Berlin, Germany). Fungal ITS2 region was amplified using the primer
pair fITS7 (5'-GTGARTCATCGAATCTTTG-3') and ITS4 (5'-TCCTCCGCTTATTGATATGC-3') (Shinohara et al., 2021).
Sequences processing was performed following the R package DADA2 pipeline version 1.16 (Callahan et al., 2016) with
R software (version 4.0.5; R Core Team, 2014). Taxonomic assignment was performed using UNITE (version 8.3; Nilsson
370 et al. 2019) databases. Metadata related to fungal abundance and diversity can be found in Table 2. DNA sequences were de-
posited in the National Center for Biotechnology Information (NCBI) Sequence Read Archive (SRA) under accession number
PRJNA822885.

4.3.4 Microbial respiration

Microbial respiration rates (see metadata in Table 2) were derived from continuous dissolved oxygen (O_2) measurements using
375 a SensorDish Reader (SDR; Presens, Germany) optical sensing system equipped with 24 glass vials of 5 mL containing non-
invasive O_2 sensors (OxoDish). The sensor vials were top filled with sample water then sealed without headspace or bubbles
and inserted into the SDR plate. The entire setup was placed in an incubator at a constant temperature of 10°C. Dissolved
 O_2 concentrations were derived from 10 minute averages of quenched fluorescence measurements taken every minute over
a period of 10 hours. Linear regression analysis was performed on the dissolved O_2 concentration data from each vial to
380 determine microbial respiration ($\mu\text{mol } O_2 \text{ L}^{-1} \text{ h}^{-1}$).

4.4 Supplementary data

During the extensive 2019 Nunataryuk WP4 field campaign, the opportunity for additional sampling arose and a subset of
variables (e.g., cations, anions, sulfides, major and rare earth elements) were measured from sediments, as well as pore water,
extracted from the western and eastern sectors of the Mackenzie Delta region (see Fig. 12, and metadata presented in Table 2).
385 Additionally, two sites near Tuktoyaktuk (Tuktoyaktuk Island and Peninsula Point in the Pingo Canadian Landmark, see Fig.
13) were sampled between July 20th and August 5th 2019 for massive ice, groundwater, and melt water on permafrost slumps.
These samples served to conduct several analyses, including abundance of naturally occurring stable and radio-isotopes, dis-
solved total iron (Fe_{tot}), DOC and CDOM fluorescence measurements (Table 2). An exhaustive list of variables is presented
in Table 2, along with contact information of principal investigators associated with each measured variable or estimated
390 parameter. Full datasets associated with the Nunataryuk WP4 field campaigns can be found in Juhls et al. 2021.

4.4.1 Sediment and pore water

Sediment cores were retrieved using an UWITEC gravity corer fitted with pre-drilled liners, with holes every 1 cm for pore
water (PW) retrieval. The corer was deployed from a tripod through ice holes during Leg 1, and from a davit secured on small
boats during Leg 4. Overall, 5 and 10 stations were sampled during Leg 1 and Leg 4, respectively (Fig. 12). Sediment cores
395 from the western region (Mackenzie and Shallow bays) were brought back for processing in laboratories of the Aurora Research



Institute (Inuvik), while processing of the cores sampled in the eastern region (Kugmallit and Kittigazuit bays) was conducted in converted work spaces of the Tuktoyaktuk Learning Center. Pore water was sampled using rhizons (Rhizosphere product Inc.) with 0.2 μm polyethersulfone (PES). The PW was divided into four different sub-samples in vials prepared for specific analysis. Acid-washed high density polyethylene (HDPE) centrifuge tubes containing 200 μL of ultrapure HNO_3 were used for
400 cations. Four mL amber glass vials with polytetrafluoroethylene (PTFE)-lined caps were used for the analysis of DOC, after being acid washed in hydrochloric acid (HCl) 10% and burned at 500°C overnight. Rinsed polypropylene gas chromatography vials were used to preserve anions for later analysis. Lastly, nitrogen-filled amber vials amended with 100 μL of 10% zinc acetate solution were used to preserve dissolved sulfide.

Cations were analyzed with Agilent 8800 ICP-QQQ-MS while DOC analysis was conducted on a Shimadzu Total Organic
405 Carbon Analyzer TOC-VCPH, and sulfides were analyzed using a Horiba Scientific Aqualog and analysis of anions will be conducted by Dionex Integrion HPIC (High Performance Ion Chromatography).

The sediment cores were sliced every 1 cm. The subsamples were placed in Falcon cups and kept frozen until treatment. The samples were then freeze-dried and homogenized with an agate pestle and mortar. Ground, freeze-dried sediment was mineralized using ultra-pure nitric and hydrochloric acids using a microwave (MARS5 Microwave in EasyPrep vessels). Results were
410 validated against MESS-4 certified reference material (National Research Council Canada). The mineralisation protocol used is based on Ma et al. 2019 with further modifications detailed in Bossé-Demers et al. 2022. Major (Fe, Ca, Na, Mg, Mn and K) and rare-earth elements (REE) in the sediment were analyzed using a Thermo Scientific iCAP 7400 ICP-OES and an Agilent 8800 ICP-QQQ-MS, respectively.

4.4.2 Groundwater

415 4.4.2.1 Radon, radium and stable isotopes of water ($\delta^{18}\text{O}$, $\delta^2\text{H}$)

Naturally occurring Radon (^{222}Rn ; half-life of 3.8 days) and radium (^{223}Ra , ^{224}Ra , ^{226}Ra , ^{228}Ra ; half-lives of 11.4 days, 3.6 days, 1600 years and 5.8 years, respectively) isotopes in the environment are efficient geochemical tracers to map and quantify submarine groundwater discharge in nearshore waters. Groundwater is defined as water of any salinity that has circulated through the coastal aquifer (Moore, 1999), and groundwater is here synonymous with pore water. Two sites near Tuktoyaktuk
420 were investigated between July 20th and August 5th 2019 (Tuktoyaktuk Island, 69.4557° N, 133.0039° W; and Peninsula Point in the Pingo Canadian Landmark, 69.4552° N, 133.0069° W, see Fig. 13). Massive ice, groundwater, and melt water samples were collected on permafrost slumps. Seawater samples were also collected in front of each study site at 0.5, 1, 1.5, and 2 km from the coastline. Massive ice samples were collected and immediately stored in airtight buckets to limit radon loss. They were then kept at room temperature in the laboratory until they had melted completely. Groundwater, melt water and
425 seawater were collected using a peristaltic or a submersible pump. Water was continuously pumped at an approximate flow rate varying from 0.2 to 0.8 L min^{-1} through a Teflon tube into an inline flow cell where temperature, practical salinity and oxygen saturation were monitored with a daily calibrated multiparametric probe (YSI-600QS). For radon, 2 L of water were sampled in plastic bottles that were tightly sealed. Within the next 12 h, analyses were carried out as described in Chaillou

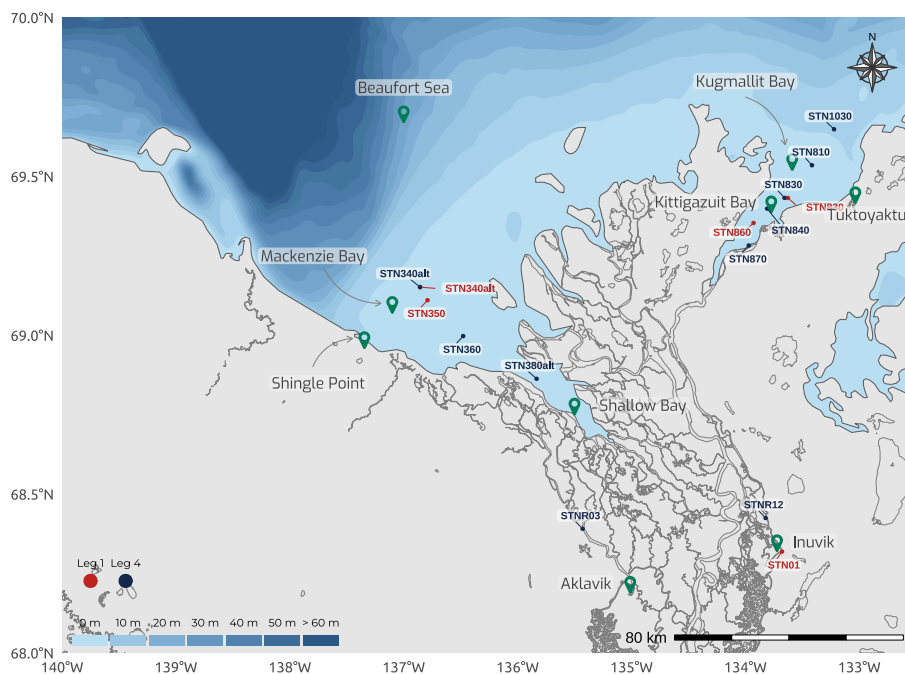


Figure 12. Map of the Mackenzie Delta region (Inuvialuit Settlement Region, Northwest Territories, Canada), with bathymetric features of the coastal waters of the southern Beaufort Sea, showing the locations for the sediment and pore water sampling during two legs of the 2019 WP4 Nunataryuk field expeditions (Legs 1 and 4).

et al. 2018. Briefly, the water was bubbled to allow ^{222}Rn degassing and the equilibrated air flowed through Drierite desiccant
430 to a radon-in-air detector (RAD7, DurrIDGE). The air volume of the closed-loop and water volume of the bottle were known
and constant over the measurement. The air recirculated through the water and continuously extracted the radon until a state of
equilibrium developed. Radon-in-water activities were finally corrected by temperature and humidity, and by radon decay that
took place between sampling and analysis. Analytical uncertainties were less than 10% (2σ).

Samples for radium were filtered through a 1 μm Hytrec cartridge (seawater) or a 0.45 μm Pall high-capacity capsule filter
435 (groundwater and melted ice) to remove suspended sediment, and then through an acrylic fiber coated in manganese oxide
(MnO_2), which quantitatively scavenges Ra (Reid et al., 1979). Fibers were thoroughly rinsed with Ra-free water to remove
any additional particles before analysis. Sample volumes ranged from 1 - 20 L for groundwater and thawed ice, and up to
120 L for seawater. The short-lived ^{224}Ra and ^{223}Ra isotopes were measured using a Radium Delayed Coincidence Counter
(RaDeCC) system (Moore and Arnold, 1996) within 3 days of sample collection. Fibers were re-analyzed after 4 weeks and
440 after 2 months to determine the activities of ^{224}Ra and ^{223}Ra supported by parent isotopes ^{228}Th (half life of 1.91 years) and
 ^{227}Ac (half life of 21.8 years), respectively. Activities of ^{224}Ra and ^{223}Ra reported in Table 2 are the activities in excess of
the parent isotopes (unsupported activities). Following RaDeCC analyses, fibers were sealed in an airtight housing for three
weeks to allow ^{222}Rn to reach equilibrium with ^{226}Ra . The activities of ^{226}Ra were then determined via ^{222}Rn emanation and



scintillation counting following the methods described in Key et al. 1979. The efficiency of RaDeCC and scintillation counting
445 was determined using MnO₂-coated fibers spiked with known activities of radium.

In addition to radon and radium isotopes in water, the mineral-bound ²²⁶Ra activity of sediments was determined on four
(4) sediment samples (two coastal permafrost cliff and two surficial Holocene inshore) collected at each site. These measure-
ments were done as a first attempt to monitor the activity of ²²⁶Ra-supported ²²²Rn in groundwater. Fifteen (15) grams of
sediment samples were dried, crushed and sealed in vials fitted for a high-purity Germanium gamma-ray spectrometer (OR-
450 TEC® GMX50) at the ISMER laboratory (UQAR, Rimouski, Canada). They were left in sealed vials for at least 23 days to
ensure radioactive re-equilibration between the ²²⁶Ra and the short-lived daughters of the ²³⁸U series (Zielinski et al., 2001).
Counting time was fixed at 4 to 5 days to provide adequate counts for the peaks of ²¹⁴Pb (using the 295.2 and 352 keV) and
²¹⁴Bi (609 keV) peaks. The counting error was <10%. Disintegrations per minute (dpm)/g were converted in Bq/m³ of wet
sediment.

455 Unfiltered water samples for the determination of stable water isotopes were collected in 30-mL HDPE vials free of any
chemicals, sealed tightly, and stored in the dark at 4°C. Stable isotopes of water ($\delta^{18}\text{O}$, $\delta^2\text{H}$) were analyzed by elemental
analysis–isotope ratio mass spectrometry (EA-IRMS) at the GÉOTOP Laboratory (UQAM, Montreal, Canada). Precisions
were $\pm 0.05\text{‰}$ and $\pm 1\text{‰}$ (at the 1σ level) for $\delta^{18}\text{O}$ and $\delta^2\text{H}$, respectively. Isotopic analyses are reported compared to the
international Vienna Standard Mean Ocean Water (VSMOW). Reference materials were used throughout the isotopic water
460 analyses to ensure high quality data.

4.4.2.2 Dissolved total iron (Fe_{tot}), DOC and CDOM fluorescence measurements

Duplicate dissolved organic carbon (DOC) samples were collected at the outlet of the Teflon tube using acid-cleaned 60-mL
polypropylene syringes and were directly filtered through combusted 0.7 μm glass fiber filters. The filtered samples were
acidified using high purity HCl (37%) to pH < 2 in combusted borosilicate EPA tubes with PTFE caps, after being acid-washed
465 in hydrochloric acid (HCl) 10%, burned at 500°C overnight, and stored in the dark at 4°C until analysis. DOC was analyzed
a few weeks post-collection using a total organic carbon analyzer (TOC-Vcpn, Shimadzu) based on the method of Wurl and
Sin (2009) and combined with a total nitrogen measuring unit (TNM-1, Shimadzu) at ISMER (UQAR, Rimouski, Canada).
The analytical uncertainties were less than 2% and the detection limit was 0.05 mg L⁻¹. Fresh acidified deionized water (blank)
and a standard solution (1.1 ± 0.03 mg C L⁻¹) were frequently analyzed during measurements to ensure the stability of the
470 instrument's performance.

Water samples for total dissolved Fe and CDOM were pumped through a Teflon tube and directly filtered using a Millipore
Opticap® XL4 capsule with a Durapore® membrane (0.22 μm porosity) connected to the outlet of the tube. Total dissolved
Fe (Fe_{tot}) samples were stored in 15-mL Falcon® tubes, acidified to pH < 2 with an ultratrace ppb grade HCl (37%) solution,
and stored at 4°C prior to total dissolved Fe analysis. Fe_{tot} was measured using the ferrozine method modified by Viollier
475 et al. 2000, with a detection limit of 0.3 μM . Measurements were done using a Genesys 20 spectrophotometer (Thermo Fisher
Scientific) using a quartz cuvette with a pathlength of 1 cm. CDOM samples were stored in acid-cleaned 15-mL glass tubes
in the dark at 4°C prior to analysis. Absorbance and fluorescence of CDOM were measured simultaneously. Before optical



analysis, samples were allowed to equilibrate at room temperature (20°C). CDOM absorbance in the UV-visible spectra was measured using a Lambda 850 UV-VIS spectrophotometer (PerkinElmer) fitted with two 1-cm pathlength quartz cuvettes, one used for the reference and one for the sample. Measurements were taken from 220 to 800 nm at 1-nm intervals with a scanning speed of 100 nm min⁻¹. Fresh Milli-Q water was used as blanks and references during the analysis. The reference water was refreshed every 30 min. Before each analysis, the quartz cuvette was flushed first with 5% HCl, then with deionized water, and finally with the sample. Absorbance metrics were then extracted from the different scans. Concomitantly, CDOM fluorescence was measured using a Varian Cary Eclipse fluorometer and a 1-cm pathlength quartz cuvette. Emission wavelengths (λ_{Em}) ranged from 230 to 600 nm and excitation wavelengths (λ_{Ex}) from 220 to 450 nm. When the absorbance was higher than 0.3 at 254 nm, the sample was diluted to avoid saturating the fluorometer (Miller and McKnight, 2010). Fresh deionized water was used as a blank and absorbance measurements of the samples were used to correct the fluorescence data for inner filter effects. All data were corrected for the Raman and Rayleigh effect, using daily fresh deionized water signals, and for the inner filter effect, using absorbance spectra. Moreover, because the low concentrations of Fe_{tot} results in low total Fe:DOC molar ratios, on the order of 10⁻², this suggested that the Fe effect on the absorbance and fluorescence of CDOM was negligible (Poulin et al., 2014). Therefore, no Fe effect correction was applied to the DOM concentration, absorbance, and fluorescence. The collected data were then described as excitation-emission matrices (EEMs) and different absorbance and fluorescence metrics were calculated (Fig. 13). This database was completed by 50 more samples collected in the same region in summer 2021 (see Flamand et al. 2022 for details).

Fluorescence spectroscopy was successfully used to determine the origin of the DOM pool. Fluorescence occurring at low excitation/emission wavelengths was associated to protein-like material originating from autochthonous production, whereas fluorescence happening at higher wavelengths was associated to humic-like material of higher molecular weight derived from terrestrial sources (Coble, 1996; Murphy et al., 2008). In Fig. 13, different DOM fluorescence signatures can be observed across stations distributed along a north-south transect near Tuktoyaktuk. Strong signals of protein-like and humic-like fluorescence can be observed at seawater stations 17 and 18, as well as at the beach groundwater station 7, which represent stations characterized by a mixture of FDOM. In contrast, there are no obvious signals of in situ derived FDOM at station 4, which was derived from the thawing of massive ice.

4.4.2.3 Dissolved inorganic carbon (DIC) and methane (CH₄)

Water sample for dissolved inorganic carbon (DIC or $\sum\text{CO}_2 = \text{CO}_2 + \text{HCO}_3^- + \text{CO}_3^{2-}$, also called total carbonate) and methane were pumped and stored in 120-mL borosilicate glass bottles hermetically closed with a Teflon rubber crimped with an aluminum ferrule. A 0.2 mL of HgCl₂ solution (7 mg L⁻¹) was added into each bottle. The DIC concentration of samples were determined using a SciTech Apollo DIC analyzer. After reaching thermal equilibration at 25°C, duplicates of 30 mL of the sample were injected into the instrument's reactor where they were acidified with 10% H₃PO₄. The CO₂ formed was then carried to a LI-COR infrared analyzer by a stream of pure nitrogen. A calibration curve was constructed using Na₂CO₃ solutions, and the accuracy of the measurements was verified using the Dikson batch 122 standard. Reproducibility was typically on the order of 0.2%. Methane (CH₄) concentrations were determined by a Peak Performer gas chromatography

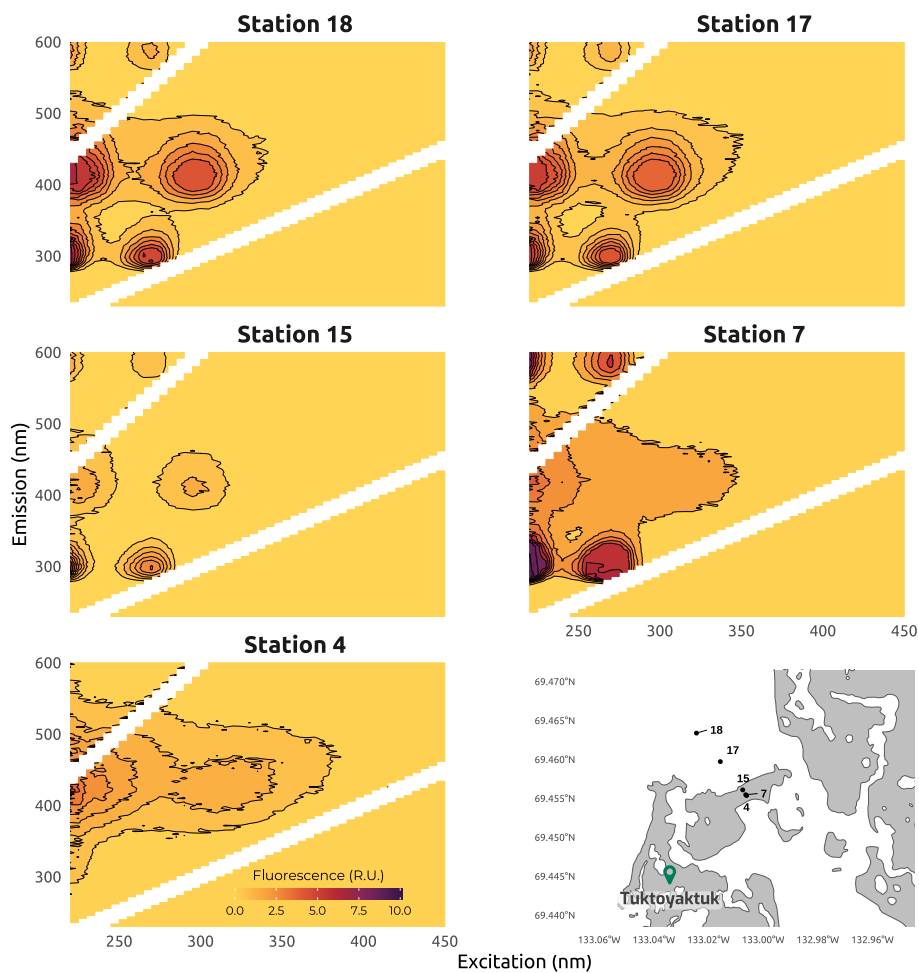


Figure 13. Examples of Excitation-emission matrices (EEMs) obtained for seawater samples (stations 15-17-18), beach groundwater (station 7), as well as thawed permafrost and massive ice (station 4) collected in Tuktoyaktuk Island site during Leg 3 of the 2019 WP4 Nunataryuk field expeditions. EEMs are all corrected for the Raman and Rayleigh scattering and inner-filter effect. Note that the intensity scale is the same for each sample. The shapefiles for the Tuktoyaktuk area were from: <https://www.geogratia.gc.ca/>.

with flame ionization detection (GC-FID, 2-mL sample loop; Peak Laboratories, USA) based on the static headspace method reported by Zhang and Xie (2015). The samples were transferred into a 50-mL glass syringe and 5 mL of CH₄-free N₂ was introduced to obtain a 1:10 gas:water ratio. The syringe was vigorously shaken for 4 min and the equilibrated headspace gas was injected into the GC-FID for CH₄ quantification. The methane concentrations in the headspace were calculated using the respective volumes of water and headspace in the vial, and the solubility coefficient of methane of Yamamoto et al. 1976 as a function of temperature and salinity. The analyzer was calibrated with a methane standard of 4.94 ppm by volume (ppmv) (Air Liquide) traceable to the National Institute of Standards and Technology, and the precision of the technique was ±4% (at 5 nmol L⁻¹).



520 5 Conclusions

Fundamental physical, optical, chemical and biological processes associated with reservoirs and fluxes of OMt were acquired, measured and processed during four WP4 Nunataryuk field campaigns in the Mackenzie Delta region and adjacent coastal waters of the Beaufort Sea (ISR, Northwest Territories, Canada). A subset of the broad dataset has been presented and described here. A full list of variables measured during the field expeditions can be found in Table 2, including authorship information for these datasets. Several of the datasets have been fed into Pangaea data publisher for earth and environmental science, in order to promote the principles of the International Council for Science - World Data System (ICSU-WDS). As far as we know, the data gathered and currently included in this paper are unparalleled in the breadth of their spatio-temporal coverage of a fluvial-to-marine transitional region in the Arctic. The scope of the dataset presented here offers contingencies towards its further exploitation in research.

Table 2: Parameters measured during the Nunataryuk surveys. Parameters are ordered by alphabetical order.

| Origin | Variable | Leg 1 | Leg 2 | Leg 3 | Leg 4 | Units | Method | Principal investigator | Contact/email address | Status | Reference DOI |
|--------|---|-------|-------|-------|-------|---------------|---|-------------------------------|---|-------------|---|
| Water | Bacterial abundance | x | x | x | x | cells/mL | Flow cytometer (FACSCanto, BD Biosciences) | Martine Lizotte / Fabien Joux | martine.lizotte@takuvik.ulaval.ca / joux@obs-banyuls.fr | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Bacterial diversity | x | x | x | NA | ASVs | Illumina MiSeq sequencing | Jérôme Comte | jerome.comte@etc.inrs.ca | On request | NA |
| Water | Bacterial isolation | NA | NA | x | NA | NA | Bacterial isolation (R2A agar and broth) and identification (Sanger sequencing) | Fabien Joux | joux@obs-banyuls.fr | In progress | NA |
| Water | Fungal abundance | x | x | x | NA | 18S copies/mL | QPCR (StepOnePlusTM, Applied Biosystems) | Fabien Joux / Lucas Tisserand | joux@obs-banyuls.fr | On request | NA |
| Water | Fungal diversity | x | x | x | NA | ASVs | Illumina MiSeq sequencing | Fabien Joux / Lucas Tisserand | joux@obs-banyuls.fr | On request | NA |
| Water | Microbial respiration | NA | x | x | NA | µmol/L/h | Optodes (Presens) | Fabien Joux / Lucas Tisserand | joux@obs-banyuls.fr | On request | NA |
| Water | CDOM absorption (aCDOM) at 254, 350, 375, 443 nm | x | x | x | x | 1/m | UltraPath liquid waveguide system (World Precision Instruments, Inc.) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | CDOM absorption spectral slope (350-500 nm) | x | x | x | x | 1/nm | UltraPath liquid waveguide system (World Precision Instruments, Inc.) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Dissolved organic carbon (DOC) | x | x | x | x | mg/L | High Temperature Catalytic Oxidation (Shimadzu TOC-VCPN) | Bennet Juhls | bennet.juhls@awi.de | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Fluorescent dissolved organic matter A Coble-peak (FDOM A-peak) | x | x | x | x | RU | Aqualog fluorescence spectrometer | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Fluorescent dissolved organic matter Biological index (FDOM BIX) | x | x | x | x | dimensionless | Aqualog fluorescence spectrometer | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Fluorescent dissolved organic matter C Coble-peak (FDOM C-peak) | x | x | x | x | RU | Aqualog fluorescence spectrometer | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |



Table 2: Parameters measured during the Nunataryuk surveys. Parameters are ordered by alphabetical order. (continued)

| Origin | Variable | Leg 1 | Leg 2 | Leg 3 | Leg 4 | Units | Method | Principal investigator | Contact/email address | Status | Reference DOI |
|--------|--|-------|-------|-------|-------|-------------------|---|-------------------------------|---|-------------|---|
| Water | Fluorescent dissolved organic matter Fluorescence index (FDOM FI) | x | x | x | x | dimensionless | Aqualog fluorescence spectrometer | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Fluorescent dissolved organic matter Humification index (FDOM HI) | x | x | x | x | dimensionless | Aqualog fluorescence spectrometer | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Fluorescent dissolved organic matter M Coble-peak (FDOM M-peak) | x | x | x | x | RU | Aqualog fluorescence spectrometer | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Fluorescent dissolved organic matter T Coble-peak (FDOM T-peak) | x | x | x | x | RU | Aqualog fluorescence spectrometer | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Intensity fluorescent dissolved organic matter component 1 (FDOM C1) | x | x | x | x | RU | Aqualog fluorescence spectrometer | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Intensity fluorescent dissolved organic matter component 2 (FDOM C2) | x | x | x | x | RU | Aqualog fluorescence spectrometer | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Intensity fluorescent dissolved organic matter component 3 (FDOM C3) | x | x | x | x | RU | Aqualog fluorescence spectrometer | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Absorption coefficient (a) at 440, 555, 630, 715, 730, 750, 767, 820, 870 nm | NA | NA | x | x | 1/m | Measured with an AC9 sensor (WetLabs/Seabird Scientific) installed within an optical frame (profiling) | David Doxaran / Guislain Bécu | doxaran@obs-vlfr.fr / guislain.becu@takuvik.ulaval.ca | In progress | NA |
| Water | Beam attenuation coefficient (c) at 440, 555, 630, 715, 730, 750, 767, 820, 870 nm | NA | NA | x | x | 1/m | Measured with an AC9 sensor (WetLabs/Seabird Scientific) installed within an optical frame (profiling) | David Doxaran / Guislain Bécu | doxaran@obs-vlfr.fr / guislain.becu@takuvik.ulaval.ca | In progress | NA |
| Water | Concentration of CDOM | NA | x | x | x | ppm | Estimated from FLBBBCD eco-pack sensor (WetLabs/Seabird Scientific) measurements profiles (instrument deployed within an optical frame) | Guislain Bécu | guislain.becu@takuvik.ulaval.ca | In progress | NA |
| Water | Concentration of Chl-a | NA | x | x | x | mg/m ³ | Estimated from FLBBBCD eco-pack sensor (WetLabs/Seabird Scientific) measurements profiles (instrument deployed within an optical frame) | Guislain Bécu | guislain.becu@takuvik.ulaval.ca | In progress | NA |
| Water | Particle size distribution, between 1.25 and 250 microns | x | x | x | x | µL/L | Measured with a LISST-100X sensor (Sequoia Scientific) installed within an optical frame (profiling) | Guislain Bécu | guislain.becu@takuvik.ulaval.ca | In progress | NA |
| Water | Particulate backscattering (bbp) at 550, 700, 720, 770, 850, 870 nm | NA | x | x | x | 1/m | Estimated from HydroScat-2 sensor (HobitLabs) measurements profiles (instrument deployed within an optical frame) | Guislain Bécu | guislain.becu@takuvik.ulaval.ca | In progress | NA |



Table 2: Parameters measured during the Nunataryuk surveys. Parameters are ordered by alphabetical order. (continued)

| Origin | Variable | Leg 1 | Leg 2 | Leg 3 | Leg 4 | Units | Method | Principal investigator | Contact/email address | Status | Reference DOI |
|--------|--|-------|-------|-------|-------|---------------------------|--|--------------------------------------|--|------------|---|
| Water | Salinity | x | x | x | x | dimensionless | in situ conductivity meter | Guislain Bécu | guislain.becu@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Salinity | x | x | x | x | dimensionless | Lab measurement on discrete water samples with a Mettler Toledo Conductivity/Salinity/pH meter | Guislain Bécu | guislain.becu@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Salinity at surface | x | x | x | x | °C | Extracted from in situ CTD RBR Maestro (Leg 1), CTD RBR Concerto (Leg 2,3,4) | Guislain Bécu | guislain.becu@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Salinity profile | x | x | x | x | dimensionless | in situ CTD RBR Maestro (Leg 1), CTD RBR Concerto (Leg 2,3,4) | Guislain Bécu | guislain.becu@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Temperature | x | x | x | x | °C | in situ conductivity meter | Guislain Bécu | guislain.becu@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Temperature at surface | x | x | x | x | °C | Extracted from in situ CTD RBR Maestro (Leg 1), CTD RBR Concerto (Leg 2,3,4) | Guislain Bécu | guislain.becu@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Temperature profile | x | x | x | x | °C | in situ CTD RBR Maestro (Leg 1), CTD RBR Concerto (Leg 2,3,4) | Guislain Bécu | guislain.becu@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Nitrate concentrations [NO ₃] | x | x | x | x | μmol/L | Automated colorimetric procedure | Martine Lizotte / Jean-Éric Tremblay | martine.lizotte@takuvik.ulaval.ca / jean-eric.tremblay@bio.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Nitrite concentrations [NO ₂] | x | x | x | x | μmol/L | Automated colorimetric procedure | Martine Lizotte / Jean-Éric Tremblay | martine.lizotte@takuvik.ulaval.ca / jean-eric.tremblay@bio.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Phosphate concentrations [PO ₄ ³⁻] | x | x | x | x | μmol/L | Automated colorimetric procedure | Martine Lizotte / Jean-Éric Tremblay | martine.lizotte@takuvik.ulaval.ca / jean-eric.tremblay@bio.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Silicate concentrations [Si(OH) ₄] | x | x | x | x | μmol/L | Automated colorimetric procedure | Martine Lizotte / Jean-Éric Tremblay | martine.lizotte@takuvik.ulaval.ca / jean-eric.tremblay@bio.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Multispectral in-air downwelling irradiance (Ed0+) | NA | x | x | x | uW/cm ² /nm | Compact-Optical Profiling System (C-OPS), ICE-Pro frame (Biospherical Instruments) | Bennet Juhls | bennet.juhls@awi.de | On request | NA |
| Water | Multispectral in-water downwelling irradiance (Ed) | NA | x | x | x | uW/cm ² /nm | Compact-Optical Profiling System (C-OPS), ICE-Pro frame (Biospherical Instruments) | Bennet Juhls | bennet.juhls@awi.de | On request | NA |
| Water | Multispectral in-water upwelling radiance (Lu) | NA | x | x | x | uW/cm ² /nm/sr | Compact-Optical Profiling System (C-OPS), ICE-Pro frame (Biospherical Instruments) | Bennet Juhls | bennet.juhls@awi.de | On request | NA |
| Water | Multispectral Remote sensing reflectance (Rs) of the surface water | NA | x | x | x | 1/sr | Calculated using Ed and Lu | Bennet Juhls | bennet.juhls@awi.de | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | aP at 360, 380, 400, 412, 443, 490, 510, 551, 555, 560, 565, 620, 645, 667, 673, 683, 709, 745, 765 nm | x | x | x | x | 1/m | Varian Cary 100 spectrophotometer (Integrating sphere attached) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Suspended particulate matter (SPM) | x | x | x | x | μg/mL | Water sample, dry filter weight per volume | Martine Lizotte | martine.lizotte@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Total particulate carbon (TPC) | x | x | x | x | μg/mL | CHN Elemental Analyzer, dry filter | Martine Lizotte | martine.lizotte@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Total particulate nitrogen (TPN) | x | x | x | x | μg/mL | CHN Elemental Analyzer, dry filter | Martine Lizotte | martine.lizotte@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |



Table 2: Parameters measured during the Nunataryuk surveys. Parameters are ordered by alphabetical order. (continued)

| Origin | Variable | Leg 1 | Leg 2 | Leg 3 | Leg 4 | Units | Method | Principal investigator | Contact/email address | Status | Reference DOI |
|--------|---|-------|-------|-------|-------|-------------------|---|------------------------|------------------------------------|-----------|---|
| Water | Particulate organic carbon (POC) | x | x | x | x | µg/mL | Acidified dry filter, CHN Elemental Analyzer | Martine Lizotte | martine.lizotte@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Particulate organic nitrogen (PON) | x | x | x | x | µg/mL | Acidified dry filter, CHN Elemental Analyzer | Martine Lizotte | martine.lizotte@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | 19'-butanoyloxyfucoxanthin concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | 19'-hexanoyloxyfucoxanthin concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Alloxanthin concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Alpha Beta Carotenes concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Chlorophyll a concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Chlorophyll b concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Chlorophyll c concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Chlorophyll c2 + chlorophyll c1 + MGDVP concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Chlorophyll c3 concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Chlorophyllide a concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Diadinoxanthin concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Diatoxanthin concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Divinyl chlorophyll a concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Divinyl chlorophyll b concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Fucoxanthin concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Gyroxanthin diester concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Lutein concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Monovinyl chlorophyll a concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Monovinyl chlorophyll b concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |



Table 2: Parameters measured during the Nunataryuk surveys. Parameters are ordered by alphabetical order. (continued)

| Origin | Variable | Leg 1 | Leg 2 | Leg 3 | Leg 4 | Units | Method | Principal investigator | Contact/email address | Status | Reference DOI |
|---------------------|--|-------|-------|-------|-------|-------------------|---|------------------------|------------------------------------|-------------|---|
| Water | Noxanthin concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Peridinin concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Photoprotective carotenoids concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Photosynthetic carotenoids concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Photosynthetic pigments concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Prasinonxanthin concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Total accessory pigments concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Total carotenoids concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Total chlorophylls concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Total diagnostic pigments concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Total pheophorbide a concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Total pheophytin a concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Total pigments concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Violaxanthin concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Zeaxanthin concentration | x | x | x | x | mg/m ³ | High performance liquid chromatography (HPLC) | Atsushi Matsuoka | atsushi.matsuoka@takuvik.ulaval.ca | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | Deuterium excess | x | x | x | x | ‰ | Calculated with $\delta^{18}O$ and δD | Bennet Juhls | bennet.juhls@awi.de | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | $\delta^{18}O$ | x | x | x | x | ‰ vs. SMOW | Finnigan MAT Delta-S mass spectrometer | Bennet Juhls | bennet.juhls@awi.de | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Water | δD | x | x | x | x | ‰ vs. SMOW | Finnigan MAT Delta-S mass spectrometer | Bennet Juhls | bennet.juhls@awi.de | Available | https://doi.pangaea.de/10.1594/PANGAEA.937587 |
| Sediment/Pore water | Anions (PO ³⁻ , NO ₃ , NO ₂ , SO ²⁻ , Cl, F) | x | NA | NA | x | NA | Ion Chromatography | Raoul-Marie Couture | raoul.couture@chm.ulaval.ca | In progress | NA |
| Sediment/Pore water | Dissolved organic carbon (DOC) | x | NA | NA | x | NA | Total Organic Carbon Analyzer | Raoul-Marie Couture | raoul.couture@chm.ulaval.ca | In progress | NA |
| Sediment/Pore water | Metal(loid)s and redox-sensitive elements | x | NA | NA | x | ng/L | ICP-QQQ-MS | Raoul-Marie Couture | raoul.couture@chm.ulaval.ca | On request | NA |
| Sediment/Pore water | Rare-earth elements | x | NA | NA | x | nmol/L | ICP-QQQ-MS | Raoul-Marie Couture | raoul.couture@chm.ulaval.ca | On request | NA |



Table 2: Parameters measured during the Nunataryuk surveys. Parameters are ordered by alphabetical order. (continued)

| Origin | Variable | Leg 1 | Leg 2 | Leg 3 | Leg 4 | Units | Method | Principal investigator | Contact/email address | Status | Reference DOI |
|---|----------------------------------|----------|----------|----------|----------|----------|--|------------------------|-----------------------------|-------------|---------------|
| Sediment/Pore water | Sulfides | x | NA | NA | x | NA | Spectrofluorimetry | Raoul-Marie Couture | raoul.couture@chm.ulaval.ca | In progress | NA |
| Sediment/Pore water | Rare-earth elements | x | NA | NA | x | mg/kg | ICP-QQ-MS | Raoul-Marie Couture | raoul.couture@chm.ulaval.ca | On request | NA |
| Sediment/Pore water | Reactive Fe/Mn/S profiles | x | NA | NA | x | g/kg | ICP-OES | Raoul-Marie Couture | raoul.couture@chm.ulaval.ca | On request | NA |
| Coastal permafrost thaw water and surficial seawater at the nearshore | Dissolved inorganic carbon (DIC) | NA | NA | x | NA | mg/L | High Temperature Catalytic Oxidation (Shimadzu,TOC-Vcpn) calibration with a Dickson std. | Gwénaëlle Chaillou | gwenaelle_chaillou@uqar.ca | On request | NA |
| Coastal permafrost thaw water and surficial seawater at the nearshore | Dissolved organic carbon (DOC) | NA | NA | x | NA | mg/L | High Temperature Catalytic Oxidation (Shimadzu,TOC-Vcpn) | Gwénaëlle Chaillou | gwenaelle_chaillou@uqar.ca | On request | NA |
| Coastal permafrost thaw water and surficial seawater at the nearshore | Methane (CH) | NA | NA | x | NA | nmol/L | Gas chromatography with flame ionization (GC-FID) | Gwénaëlle Chaillou | gwenaelle_chaillou@uqar.ca | On request | NA |
| Coastal permafrost thaw water and surficial seawater at the nearshore | Oxygen saturation | NA | NA | x | NA | % | in situ YSI600QS | Gwénaëlle Chaillou | gwenaelle_chaillou@uqar.ca | On request | NA |
| Coastal permafrost thaw water and surficial seawater at the nearshore | Total dissolved iron | NA | NA | x | NA | umol/L | Ferrozine technique - UV-VIS spectrophotometer (Lambda 850, PerkinElmer) | Gwénaëlle Chaillou | gwenaelle_chaillou@uqar.ca | On request | NA |
| Coastal permafrost thaw water and surficial seawater at the nearshore | Total dissolved nitrogen (TDN) | NA | NA | x | NA | mg/L | High Temperature Catalytic Oxidation (Shimadzu,TNM-1) | Gwénaëlle Chaillou | gwenaelle_chaillou@uqar.ca | On request | NA |
| Coastal permafrost thaw water and surficial seawater at the nearshore | CDOM (abs) | NA | NA | x | NA | NA | Fluorometer (Varian Cary Eclipse) | Gwénaëlle Chaillou | gwenaelle_chaillou@uqar.ca | On request | NA |
| Coastal permafrost thaw water and surficial seawater at the nearshore | CDOM (EEMs) | NA | NA | x | NA | NA | UV-VIS spectrophotometer (Lambda 850, PerkinElmer) | Gwénaëlle Chaillou | gwenaelle_chaillou@uqar.ca | On request | NA |
| Coastal permafrost thaw water and surficial seawater at the nearshore | Radium-223 (excess) | NA | NA | x | NA | dpm/100L | Radium Delayed Coincidence Counting (RaDeCC) | Lauren Kipp | kipp@rowan.edu | On request | NA |
| Coastal permafrost thaw water and surficial seawater at the nearshore | Radium-224 (excess) | NA | NA | x | NA | dpm/100L | Radium Delayed Coincidence Counting (RaDeCC) | Lauren Kipp | kipp@rowan.edu | On request | NA |



Table 2: Parameters measured during the Nunataryuk surveys. Parameters are ordered by alphabetical order. (continued)

| Origin | Variable | Leg | Leg | Leg | Leg | Units | Method | Principal investigator | Contact/email address | Status | Reference DOI |
|---|-------------------------------|-----|-----|-----|-----|-------------------|---|------------------------|----------------------------|-------------|---------------|
| | | 1 | 2 | 3 | 4 | | | | | | |
| Coastal permafrost thaw water and surficial seawater at the nearshore | Radium-226 | NA | NA | x | NA | dpm/100L | Radon-222 emanation and scintillation counting | Lauren Kipp | kipp@rowan.edu | On request | NA |
| Coastal permafrost thaw water and surficial seawater at the nearshore | Radium-226 (bulk sediment) | NA | NA | x | NA | NA | High-purity Germanium gamma-ray spectrometer (ORTEC® DSPEC jr. 2.0) | Gwénaëlle Chaillou | gwenaelle_chaillou@uqar.ca | In progress | NA |
| Coastal permafrost thaw water and surficial seawater at the nearshore | Radon-222 in water | NA | NA | x | NA | Bq/m ³ | RADH2O Big Bottle technique (DURRIDGE Company Inc.) | Gwénaëlle Chaillou | gwenaelle_chaillou@uqar.ca | On request | NA |
| Coastal permafrost thaw water and surficial seawater at the nearshore | $\delta^{18}\text{O}$ | NA | NA | x | NA | ‰ | Elemental analysis–isotope ratio mass spectrometry (EA-IRMS) | Gwénaëlle Chaillou | gwenaelle_chaillou@uqar.ca | On request | NA |
| Coastal permafrost thaw water and surficial seawater at the nearshore | $\delta^2\text{H}$ | NA | NA | x | NA | ‰ | Elemental analysis–isotope ratio mass spectrometry (EA-IRMS) | Gwénaëlle Chaillou | gwenaelle_chaillou@uqar.ca | On request | NA |
| Coastal permafrost thaw water and surficial seawater at the nearshore | Practical salinity | NA | NA | x | NA | dimensionless | in situ YSI600QS daily calibrated | Gwénaëlle Chaillou | gwenaelle_chaillou@uqar.ca | On request | NA |
| Coastal permafrost thaw water and surficial seawater at the nearshore | Temperature | NA | NA | x | NA | °C | in situ YSI600QS | Gwénaëlle Chaillou | gwenaelle_chaillou@uqar.ca | On request | NA |

530 6 Code and data availability

The raw data and metadata provided are hosted on Valeria (Université Laval user-specific repository) and final aligned and cleaned datasets are available on Pangaea (see Table 2, Juhls et al. 2021 <https://doi.pangaea.de/10.1594/PANGAEA.937587>). Detailed metadata are associated with each file including the principal investigator’s contact information. For specific questions, please contact the principal investigator associated with the data (see Table 2). Information about code availability. The code
 535 used to process the figures and tables is publicly available (https://github.com/PMassicotte/nunataryuk_data_paper) under the MIT license.



A Appendix A



Figure A1. Vehicles used to access sampling regions during the Nunataryuk WP4 field campaign: A) Snowmobiles during Leg 1 (Picture by Martine Lizotte); B) Helicopter during Leg 2 (Picture by Bennet Juhls); C) Small boats during Leg 3 (Picture by Joannie Ferland); D) Small boats during Leg 4 (Picture by Laurent Oziel).



B Appendix B

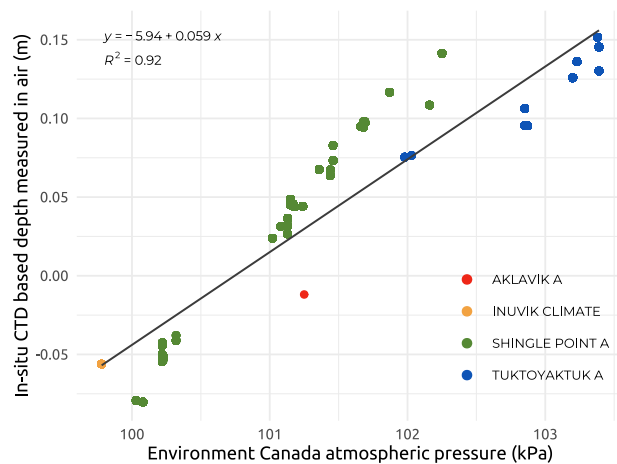


Figure B1. Depth tare correction of in situ CTD-based depth measured in air against Environment and Climate Change Canada values of atmospheric pressure with linear model fit.



C Appendix C

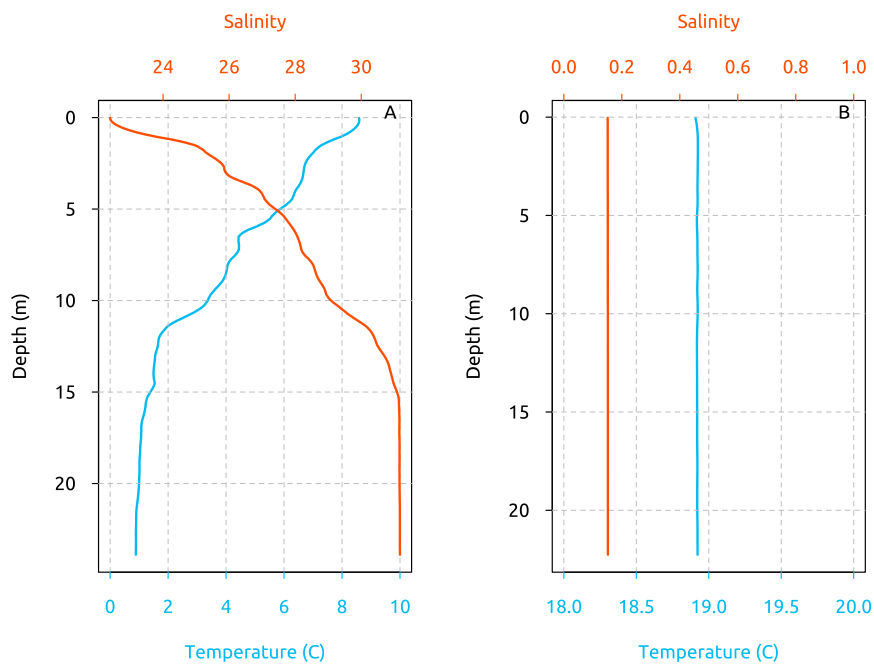


Figure C1. Two examples of vertical profiles of temperature ($^{\circ}\text{C}$) and salinity: A) STN125, 2019-07-28; B) STNR09, 2019-07-29. Note the difference in scales for temperature and salinity between the panels.



540 D Appendix D

The ranges of IOPs (light absorption (a) and scattering (b) coefficients) encountered in the Mackenzie River delta zone greatly exceeded the ones considered by Gordon and Ding (1992) when developing a well-known correction for self-shading of in-water radiometric measurements. Also, the Monte Carlo code SimulO (Leymarie et al., 2010; Doxaran et al., 2016; Voss et al., 2021) was used here to estimate and correct for the self-shading effects on in-water upwelling radiance measurements carried out using a Biospherical C-OPS radiometer deployed on the IcePro profiling platform. The exact dimensions of the IcePro Platform and radiometer (a cylinder with a radius and a length of 3.5 and 36.9 cm, respectively) were used as inputs in SimulO. The very wide ranges of a and b coefficients (up to 12 m^{-1} and 100 m^{-1} , respectively) were considered, together with two particle scattering phase functions: Fournier-Forand with backscattering ratios of 1.83% and 4% (FF183 and FF400). Black and blue sky conditions were imposed, with sun zenith angles spanning from 4° to 8° . The upwelling radiance signal at zero depth was computed using SimulO successively without, then with the sensor and IcePro platform. The difference observed, in %, was assumed to be the associated near surface (0.5m) self-shading induced by the profiling system (see Fig. D1).

Within the limited ranges of IOPs considered by Gordon and Ding (1992), a good agreement was observed between the shading simulated with the two models, with the self-shading being, as a first approximation, a function of the absorption coefficient (not shown). Considering the whole ranges of a and b coefficients, the self-shading estimated using SimulO was no longer a simple function of the absorption coefficient. Additionally considering the influence of in-water light backscattering, a function of the form $y = 1 - e^{-\alpha(a+b_b)}$ was fitted through the simulated IcePro plus radiometer self-shading factors, with b_b in m^{-1} being the total backscattering coefficient (Fig. D1). This function was used to correct the measured upwelling radiance measurements at zero depth for self-shading effects.

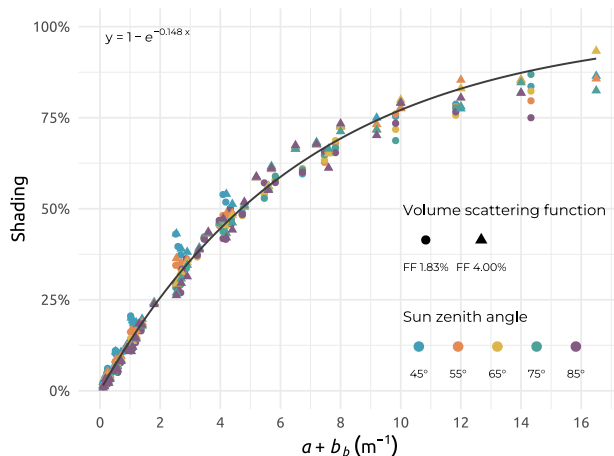


Figure D1. Relationship established between the self-shading computed using SimulO and the IOPs (namely the sum $a + b_b$, with a and b_b in m^{-1} as the absorption and backscattering coefficients, respectively), for the two particle volume scattering functions (FF 1.83% and FF 4.00%) and solar zenith angles considered.



Author contributions. Project conception and experimental design were guided by AnM, AtM, FF, GB, GC, HL, MBa, MHF, JF, and SB. Northern community consultation and engagement in the Inuvialuit Settlement Region was conducted by GC, JF, MHF, ML, and SA. Fieldwork was coordinated by AtM, GC, JF, MBa, MHF, and ML, while field sampling was conducted by BJ, CI, DOJA, EL, GB, GC, LK, LO, MD, RH, SA, TBD, and TJG. In-the-field laboratory analysis was performed by AH, ALC, AtM, CG, DD, ED, GC, JF, JM, LK, LT, MBé, ML, and TBD. Fieldwork activities were facilitated and supported by CS, DW, FF, GC, JV, LB, MF, PO, RMC, SA, and TE. Post-expedition laboratory analysis was conducted in several cities around the globe by AF, BJ, CG, CT, FB, FJ, GD, GM, JC, JET, LB, LK, LT, RMC, TBD, and TD. Dataset processing, managing and quality-controlling was performed by AtM, BJ, EL, GB, GM, ML, and PM. The creation of figures and tables was piloted by BJ, GB, GM, PM, and ML. The following co-authors wrote the main content of the paper with further contributions and suggestions from all co-authors: AtM, BJ, GC, GM, MBa, ML, PM, and TBD.

Competing interests. The authors declare that they have no conflict of interest.

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References

- 605 Allen, M., Babiker, M., Chen, Y., de Coninck, H., Connors, S., van Diemen, R., Opha, Ebi, K., Engelbrecht, F., Ferrat, M., Ford, J., Forster, P., Fuss, S., Guillén Bolaños, T., Harold, J., Hoegh-Guldberg, O., Hourcade, J.-C., Huppmann, D., and Zickfeld, K.: Summary for Policy-makers. In: *Global warming of 1.5°C. An IPCC Special Report*, 2018.
- AMAP: *Arctic Climate Change Update 2021: Key Trends and Impacts*, 2021.
- Antoine, D., Hooker, S. B., Bélanger, S., Matsuoka, A., and Babin, M.: Apparent optical properties of the Canadian Beaufort Sea - Part 1: Observational overview and water column relationships, *Biogeosciences*, 10, 4493–4509, <https://doi.org/10.5194/bg-10-4493-2013>, 2013.
- 610 Babin, M., Stramski, D., Ferrari, G. M., Claustre, H., Bricaud, A., Obolensky, G., and Hoepffner, N.: Variations in the light absorption coefficients of phytoplankton, nonalgal particles, and dissolved organic matter in coastal waters around Europe, *Journal of Geophysical Research: Oceans*, 108, 3211, <https://doi.org/10.1029/2001jc000882>, 2003.
- Bélanger, S., Carrascal-Leal, C., Jaegler, T., Larouche, P., and Galbraith, P.: Assessment of Radiometric Data from a Buoy in the St. Lawrence Estuary, *Journal of Atmospheric and Oceanic Technology*, 34, 877–896, <https://doi.org/10.1175/JTECH-D-16-0176.1>, 2017.
- 615 Biskaborn, B. K., Smith, S. L., Noetzi, J., Matthes, H., Vieira, G., Streletskiy, D. A., Schoeneich, P., Romanovsky, V. E., Lewkowicz, A. G., Abramov, A., Allard, M., Boike, J., Cable, W. L., Christiansen, H. H., Delaloye, R., Diekmann, B., Drozdov, D., Eitzmüller, B., Grosse, G., Guglielmin, M., Ingeman-Nielsen, T., Isaksen, K., Ishikawa, M., Johansson, M., Johannsson, H., Joo, A., Kaverin, D., Kholodov, A., Konstantinov, P., Kröger, T., Lambiel, C., Lanckman, J. P., Luo, D., Malkova, G., Meiklejohn, I., Moskalenko, N., Oliva, M., Phillips, M., Ramos, M., Sannel, A. B. K., Sergeev, D., Seybold, C., Skryabin, P., Vasiliev, A., Wu, Q., Yoshikawa, K., Zheleznyak, M., and Lantuit, H.: Permafrost is warming at a global scale, *Nature Communications*, 10, 264, <https://doi.org/10.1038/s41467-018-08240-4>, 2019.
- 620 Boss, E., D'Sa, E., Freeman, S., Fry, E., Mueller, J. L., Pegau, S., Reynolds, R. A., Roesler, C., Rottgers, R., Stramski, D., Twardowski, M., and Zaneveld, J. R. V.: Ocean optics and biogeochemistry protocols for satellite ocean colour sensor validation; Volume 1.0. *Inherent optical property measurements and protocols: Absorption coefficient*, 2018.
- 625 Bossé-Demers, T., Lizotte, M., A., M., Juhls, B., Oziel, L., and Couture, R. M.: Seasonal contrasts in rare-earth elements mobility in Mackenzie Delta sediment, *Manuscript in preparation*, 2022.
- Bricaud, A., Babin, M., Claustre, H., Ras, J., and Tièche, F.: Light absorption properties and absorption budget of Southeast Pacific waters, *Journal of Geophysical Research*, 115, C08 009, <https://doi.org/10.1029/2009JC005517>, 2010.
- Brown, J., Ferrians Jr., O., Heginbottom, J. A., and Melnikov, E.: *Circum-Arctic Map of Permafrost and Ground Ice Conditions*, USGS Numbered Series, p. 1, <https://doi.org/10.7265/skbg-kf16>, 1997.
- 630 Callahan, B. J., McMurdie, P. J., Rosen, M. J., Han, A. W., Johnson, A. J. A., and Holmes, S. P.: DADA2: High-resolution sample inference from Illumina amplicon data, *Nature Methods*, 13, 581–583, <https://doi.org/10.1038/nmeth.3869>, 2016.
- Camill, P.: Permafrost Thaw Accelerates in Boreal Peatlands During Late-20th Century Climate Warming, *Climatic Change*, 68, 135–152, <https://doi.org/10.1007/s10584-005-4785-y>, 2005.
- 635 Chaillou, G., Lemay-Borduas, F., Larocque, M., Couturier, M., Biehler, A., and Tommi-Morin, G.: Flow and discharge of groundwater from a snowmelt-affected sandy beach, *Journal of Hydrology*, 557, 4–15, <https://doi.org/10.1016/j.jhydrol.2017.12.010>, 2018.
- Coble, P. G.: Characterization of marine and terrestrial DOM in seawater using excitation-emission matrix spectroscopy, *Marine Chemistry*, 51, 325–346, [https://doi.org/10.1016/0304-4203\(95\)00062-3](https://doi.org/10.1016/0304-4203(95)00062-3), 1996.



- 640 Cole, J. J., Prairie, Y. T., Caraco, N. F., McDowell, W. H., Tranvik, L. J., Striegl, R. G., Duarte, C. M., Kortelainen, P., Downing, J. A., Mid-
delburg, J. J., and Melack, J.: Plumbing the global carbon cycle: Integrating inland waters into the terrestrial carbon budget, *Ecosystems*,
10, 171–184, <https://doi.org/10.1007/s10021-006-9013-8>, 2007.
- Downes, M. T.: An improved hydrazine reduction method for the automated determination of low nitrate levels in freshwater, *Water Research*,
12, 673–675, [https://doi.org/10.1016/0043-1354\(78\)90177-X](https://doi.org/10.1016/0043-1354(78)90177-X), 1978.
- Doxaran, D., Ehn, J., Bélanger, S., Matsuoka, A., Hooker, S., and Babin, M.: Optical characterisation of suspended particles in the
645 Mackenzie River plume (Canadian Arctic Ocean) and implications for ocean colour remote sensing, *Biogeosciences*, 9, 3213–3229,
<https://doi.org/10.5194/bg-9-3213-2012>, 2012.
- Doxaran, D., Leymarie, E., Nechad, B., Dogliotti, A., Ruddick, K., Gernez, P., and Knaeps, E.: Improved correction methods for field
measurements of particulate light backscattering in turbid waters, *Optics Express*, 24, 3615, <https://doi.org/10.1364/oe.24.003615>, 2016.
- Flamand, A., Lapierre, J.-F., Whalen, D., and Chaillou, G.: Origin and reactivity of permafrost dissolved organic matter, Manuscript in
650 preparation, 2022.
- Fritz, M., Vonk, J. E., and Lantuit, H.: Collapsing Arctic coastlines, *Nature Climate Change*, 7, 6–7, <https://doi.org/10.1038/nclimate3188>,
2017.
- Gasol, J. M. and Del Giorgio, P. A.: Using flow cytometry for counting natural planktonic bacteria and understanding the structure of
planktonic bacterial communities, *Scientia Marina*, 64, 197–224, <https://doi.org/10.3989/scimar.2000.64n2197>, 2000.
- 655 Gordon, H. R. and Ding, K.: Self-shading of in-water optical instruments, *Limnology and Oceanography*, 37, 491–500,
<https://doi.org/10.4319/lo.1992.37.3.0491>, 1992.
- Grasshoff, K., Kremling, K., and Ehrhardt, M.: *Methods of Seawater Analysis*, Wiley, <https://doi.org/10.1002/9783527613984>, 1999.
- Gruber, S.: Derivation and analysis of a high-resolution estimate of global permafrost zonation, *The Cryosphere*, 6, 221–233,
<https://doi.org/10.5194/tc-6-221-2012>, 2012.
- 660 Guo, Z. G., Niu, F. J., Zhan, H., and Wu, Q. B.: Changes of grassland ecosystem due to degradation of permafrost frozen soil in the Qinghai-
Tibet Plateau, *Shengtai Xuebao/ Acta Ecologica Sinica*, 27, 3294–3301, 2007.
- Holmes, R. M., McClelland, J. W., Peterson, B. J., Tank, S. E., Bulygina, E., Eglinton, T. I., Gordeev, V. V., Gurtovaya, T. Y., Raymond,
P. A., Repeta, D. J., Staples, R., Striegl, R. G., Zhulidov, A. V., and Zimov, S. A.: Seasonal and Annual Fluxes of Nutrients and Organic
Matter from Large Rivers to the Arctic Ocean and Surrounding Seas, *Estuaries and Coasts*, 35, 369–382, <https://doi.org/10.1007/s12237->
665 011-9386-6, 2012.
- Hooker, S. B., Van Heukelem, L., Thomas, C. S., Claustre, H., Ras, J., Barlow, R., Sessions, H., Schlüter, L., Perl, J., Trees, C., Stuart, V.,
Head, E., Clementson, L., Fishwick, J., Llewellyn, C., and Aiken, J.: The Second SeaWiFS HPLC Analysis Round-Robin Experiment
(SeaHARRE-2), NASA Technical Memorandum, pp. 1–112, 2005.
- Hugelius, G., Strauss, J., Zubrzycki, S., Harden, J. W., Schuur, E. A., Ping, C. L., Schirmermeister, L., Grosse, G., Michaelson, G. J., Koven,
670 C. D., O'Donnell, J. A., Elberling, B., Mishra, U., Camill, P., Yu, Z., Palmtag, J., and Kuhry, P.: Estimated stocks of circumpolar permafrost
carbon with quantified uncertainty ranges and identified data gaps, *Biogeosciences*, 11, 6573–6593, [https://doi.org/10.5194/bg-11-6573-](https://doi.org/10.5194/bg-11-6573-2014)
2014, 2014.
- IOCCG: Ocean Colour Remote Sensing in Polar Seas. Babin, M., Arrigo, K., Bélanger, S. and Forget, M.-H. (eds.), IOCCG Report Series,
No. 16, International Ocean Colour Coordinating Group, Dartmouth, Canada., No. 16, 2015.



- 675 IPCC: Climate Change and Land: An IPCC Special Report on climate change, desertification, land degradation, sustainable land management, food security, and greenhouse gas fluxes in terrestrial ecosystems. Summary for Policymakers, Tech. rep., test, https://www.ipcc.ch/site/assets/uploads/sites/4/2020/02/SPM_Updated-Jan20.pdf, 2019.
- Juhls, B., Paul Overduin, P., Hölemann, J., Hieronymi, M., Matsuoka, A., Heim, B., and Fischer, J.: Dissolved organic matter at the fluvial-marine transition in the Laptev Sea using in situ data and ocean colour remote sensing, *Biogeosciences*, 16, 2693–2713, <https://doi.org/10.5194/bg-16-2693-2019>, 2019.
- 680 Juhls, B., Stedmon, C. A., Morgenstern, A., Meyer, H., Hölemann, J., Heim, B., Povazhnyi, V., and Overduin, P. P.: Identifying Drivers of Seasonality in Lena River Biogeochemistry and Dissolved Organic Matter Fluxes, *Frontiers in Environmental Science*, 8, <https://doi.org/10.3389/fenvs.2020.00053>, 2020.
- Juhls, B., Lizotte, M., Matsuoka, A., Mével, G., Bécu, G., Overduin, P. P., Devred, E., Doxaran, D., Ferland, J., Forget, M.-H., Hilborn, A., Leymarie, E., Maury, J., Oziel, L., Tisserand, L., Miles, D., Anikina, D. O. J., Guilmette, C., Béguin, M., Couture, R.-M., Bossé-Demers, T., Laberge-Carignan, A., Chaillou, G., Bélanger, S., Bruyant, F., and Babin, M.: Hydrographical, biogeochemical and biooptical water properties in the Mackenzie Delta Region during 4 expeditions from spring to fall in 2019, <https://doi.org/10.1594/PANGAEA.937587>, 2021.
- 685 Key, R., Brewer, R., Stockwell, J., Guinasso, N., and Schink, D.: Some improved techniques for measuring radon and radium in marine sediments and in seawater, *Marine Chemistry*, 7, 251–264, [https://doi.org/10.1016/0304-4203\(79\)90042-2](https://doi.org/10.1016/0304-4203(79)90042-2), 1979.
- Kipp, L. E., Charette, M. A., Moore, W. S., Henderson, P. B., and Rigor, I. G.: Increased fluxes of shelf-derived materials to the central Arctic Ocean, *Science Advances*, 4, eaao1302, <https://doi.org/10.1126/sciadv.aao1302>, 2018.
- Leymarie, E., Doxaran, D., and Babin, M.: Uncertainties associated to measurements of inherent optical properties in natural waters, *Applied Optics*, 49, 5415–5436, <https://doi.org/10.1364/AO.49.005415>, 2010.
- 695 Liu, C. M., Kachur, S., Dwan, M. G., Abraham, A. G., Aziz, M., Hsueh, P. R., Huang, Y. T., Busch, J. D., Lamit, L. J., Gehring, C. A., Keim, P., and Price, L. B.: FungiQuant: a broad-coverage fungal quantitative real-time PCR assay., *BMC microbiology*, 12, 255, <https://doi.org/10.1186/1471-2180-12-255>, 2012.
- Ma, L., Dang, D. H., Wang, W., Evans, R. D., and Wang, W.-X.: Rare earth elements in the Pearl River Delta of China: Potential impacts of the REE industry on water, suspended particles and oysters, *Environmental Pollution*, 244, 190–201, <https://doi.org/10.1016/j.envpol.2018.10.015>, 2019.
- 700 Massicotte, P., Asmala, E., Stedmon, C., and Markager, S.: Global distribution of dissolved organic matter along the aquatic continuum: Across rivers, lakes and oceans, *Science of The Total Environment*, 609, 180–191, <https://doi.org/10.1016/j.scitotenv.2017.07.076>, 2017.
- Massicotte, P., Amon, R. M. W., Antoine, D., Archambault, P., Balzano, S., Bélanger, S., Benner, R., Boeuf, D., Bricaud, A., Bruyant, F., Chaillou, G., Chami, M., Charrière, B., Chen, J., Claustre, H., Coupel, P., Delsaut, N., Doxaran, D., Ehn, J., Fichot, C., Forget, M.-H., Fu, P., Gagnon, J., Garcia, N., Gasser, B., Ghiglione, J.-F., Gorsky, G., Gosselin, M., Gourvil, P., Gratton, Y., Guillot, P., Heipieper, H. J., Heussner, S., Hooker, S. B., Huot, Y., Jeanthon, C., Jeffrey, W., Joux, F., Kawamura, K., Lansard, B., Leymarie, E., Link, H., Lovejoy, C., Marec, C., Marie, D., Martin, J., Martín, J., Massé, G., Matsuoka, A., McKague, V., Mignot, A., Miller, W. L., Miquel, J.-C., Mucci, A., Ono, K., Ortega-Retuerta, E., Panagiotopoulos, C., Papakyriakou, T., Picheral, M., Prieur, L., Raimbault, P., Ras, J., Reynolds, R. A., Rochon, A., Rontani, J.-F., Schmechtig, C., Schmidt, S., Sempéré, R., Shen, Y., Song, G., Stramski, D., Tachibana, E., Thirouard, A., Tolosa, I., Tremblay, J.-É., Vaïtilingom, M., Vaultot, D., Vaultier, F., Volkman, J. K., Xie, H., Zheng, G., and Babin, M.: The MALINA oceanographic expedition: how do changes in ice cover, permafrost and UV radiation impact biodiversity and biogeochemical fluxes in the Arctic Ocean?, *Earth System Science Data*, 13, 1561–1592, <https://doi.org/10.5194/essd-13-1561-2021>, 2021.



- Matsuoka, A., Hill, V., Huot, Y., Babin, M., and Bricaud, A.: Seasonal variability in the light absorption properties of western Arctic waters: Parameterization of the individual components of absorption for ocean color applications, *Journal of Geophysical Research: Oceans*, 116, 715 <https://doi.org/10.1029/2009JC005594>, 2011.
- Matsuoka, A., Bricaud, A., Benner, R., Para, J., Sempéré, R., Prieur, L., Bélanger, S., and Babin, M.: Tracing the transport of colored dissolved organic matter in water masses of the Southern Beaufort Sea: relationship with hydrographic characteristics, *Biogeosciences*, 9, 925–940, <https://doi.org/10.5194/bg-9-925-2012>, 2012.
- Matsuoka, A., Hooker, S. B., Bricaud, A., Gentili, B., and Babin, M.: Estimating absorption coefficients of colored dissolved organic matter (CDOM) using a semi-analytical algorithm for southern Beaufort Sea waters: Application to deriving concentrations of dissolved organic carbon from space, *Biogeosciences*, 10, 917–927, <https://doi.org/10.5194/bg-10-917-2013>, 2013.
- Matsuoka, A., Babin, M., Doxaran, D., Hooker, S. B., Mitchell, B. G., Bélanger, S., and Bricaud, A.: A synthesis of light absorption properties of the Arctic Ocean: Application to semianalytical estimates of dissolved organic carbon concentrations from space, *Biogeosciences*, 11, 3131–3147, <https://doi.org/10.5194/bg-11-3131-2014>, 2014.
- 725 Matsuoka, A., Boss, E., Babin, M., Karp-Boss, L., Hafez, M., Chekalyuk, A., Proctor, C. W., Werdell, P. J., and Bricaud, A.: Pan-Arctic optical characteristics of colored dissolved organic matter: Tracing dissolved organic carbon in changing Arctic waters using satellite ocean color data, *Remote Sensing of Environment*, 200, 89–101, <https://doi.org/10.1016/j.rse.2017.08.009>, 2017.
- Maza-Márquez, P., Aranda, E., González-López, J., and Rodelas, B.: Evaluation of the Abundance of Fungi in Wastewater Treatment Plants Using Quantitative PCR (qPCR), in: *Methods in molecular biology* (Clifton, N.J.), vol. 2065, pp. 79–94, https://doi.org/10.1007/978-1-4939-9833-3_7, 2020.
- 730 McClelland, J. W., Déry, S. J., Peterson, B. J., Holmes, R. M., and Wood, E. F.: A pan-arctic evaluation of changes in river discharge during the latter half of the 20th century, *Geophysical Research Letters*, 33, <https://doi.org/10.1029/2006GL025753>, 2006.
- McGuire, A. D., Anderson, L. G., Christensen, T. R., Scott, D., Laodong, G., Hayes, D. J., Martin, H., Lorenson, T. D., Macdonald, R. W., and Nigel, R.: Sensitivity of the carbon cycle in the Arctic to climate change, *Ecological Monographs*, 79, 523–555, <https://doi.org/10.1890/08-2025.1>, 2009.
- 735 McGuire, A. D., Lawrence, D. M., Koven, C., Klein, J. S., Burke, E., Chen, G., Jafarov, E., MacDougall, A. H., Marchenko, S., Nicol-sky, D., Peng, S., Rinke, A., Ciais, P., Gouttevin, I., Hayes, D. J., Ji, D., Krinner, G., Moore, J. C., Romanovsky, V., Schädel, C., Schaefer, K., Schuur, E. A., and Zhuang, Q.: Dependence of the evolution of carbon dynamics in the northern permafrost region on the trajectory of climate change, *Proceedings of the National Academy of Sciences of the United States of America*, 115, 3882–3887, 740 <https://doi.org/10.1073/pnas.1719903115>, 2018.
- Meyer, H., Schönicke, L., Wand, U., Hubberten, H. W., and Friedrichsen, H.: Isotope studies of hydrogen and oxygen in ground ice - Experiences with the equilibration technique, *Isotopes in Environmental and Health Studies*, 36, 133–149, <https://doi.org/10.1080/10256010008032939>, 2000.
- Mitchell, B. G., Kahru, M., Wieland, J., and Stramska, M.: Determination of spectral absorption coefficients of particles, dissolved material and phytoplankton for discrete water samples, in: *Ocean Optics Protocols for Satellite Ocean Color Sensor Validation, Rev. 4, Vol. IV: Inherent Optical Properties: Instruments, Characterisations, Field Measurements and Data Analysis Protocols*, vol. 4, pp. 39–64, NASA, 2003.
- 745 Morrow, J. H., Hooker, S. B., Booth, C. R., Bernhard, G., Lind, R. N., and Brown, J. W.: Advances in measuring the apparent optical properties (AOPs) of optically complex waters, *NASA Tech. Memo*, 215856, 42–50, 2010.



- 750 Murphy, K. R., Stedmon, C. A., Waite, T. D., and Ruiz, G. M.: Distinguishing between terrestrial and autochthonous organic matter sources in marine environments using fluorescence spectroscopy, *Marine Chemistry*, 108, 40–58, <https://doi.org/10.1016/j.marchem.2007.10.003>, 2008.
- Nilsson, R. H., Larsson, K.-H., Taylor, A. F., Bengtsson-Palme, J., Jeppesen, T. S., Schigel, D., Kennedy, P., Picard, K., Glöckner, F. O., Tedersoo, L., Saar, I., Kõljalg, U., and Abarenkov, K.: The UNITE database for molecular identification of fungi: handling dark taxa and
755 parallel taxonomic classifications, *Nucleic Acids Research*, 47, D259–D264, <https://doi.org/10.1093/nar/gky1022>, 2019.
- Obu, J., Westermann, S., Bartsch, A., Berdnikov, N., Christiansen, H. H., Dashtseren, A., Delaloye, R., Elberling, B., Etzelmüller, B., Kholodov, A., Khomutov, A., Kääb, A., Leibman, M. O., Lewkowicz, A. G., Panda, S. K., Romanovsky, V., Way, R. G., Westergaard-Nielsen, A., Wu, T., Yamkhin, J., and Zou, D.: Northern Hemisphere permafrost map based on TTOP modelling for 2000–2016 at 1km² scale, *Earth-Science Reviews*, 193, 299–316, <https://doi.org/10.1016/j.earscirev.2019.04.023>, 2019.
- 760 Osburn, C. L., Retamal, L., and Vincent, W. F.: Photoreactivity of chromophoric dissolved organic matter transported by the Mackenzie River to the Beaufort Sea, *Marine Chemistry*, 115, 10–20, <https://doi.org/10.1016/j.marchem.2009.05.003>, 2009.
- Parada, A. E., Needham, D. M., and Fuhrman, J. A.: Every base matters: assessing small subunit rRNA primers for marine microbiomes with mock communities, time series and global field samples, *Environmental Microbiology*, 18, 1403–1414, <https://doi.org/10.1111/1462-2920.13023>, 2016.
- 765 Poulin, B. A., Ryan, J. N., and Aiken, G. R.: Effects of iron on optical properties of dissolved organic matter, *Environmental Science and Technology*, 48, 10 098–10 106, <https://doi.org/10.1021/es502670r>, 2014.
- Quast, C., Pruesse, E., Yilmaz, P., Gerken, J., Schweer, T., Yarza, P., Peplies, J., and Glöckner, F. O.: The SILVA ribosomal RNA gene database project: improved data processing and web-based tools, *Nucleic Acids Research*, 41, D590–D596, <https://doi.org/10.1093/nar/gks1219>, 2012.
- 770 R Core Team: R: A Language and Environment for Statistical Computing, R Foundation for Statistical Computing, Vienna, Austria, <https://www.r-project.org/>, 2022.
- Reid, D. F., Key, R. M., and Schink, D. R.: Radium, thorium, and actinium extraction from seawater using an improved manganese-oxide-coated fiber, *Earth and Planetary Science Letters*, 43, 223–226, [https://doi.org/10.1016/0012-821X\(79\)90205-X](https://doi.org/10.1016/0012-821X(79)90205-X), 1979.
- Reimnitz, E., Toimil, L., and Barnes, P.: Arctic continental shelf morphology related to sea-ice zonation, Beaufort Sea, Alaska, *Marine
775 Geology*, 28, 179–210, [https://doi.org/10.1016/0025-3227\(78\)90018-X](https://doi.org/10.1016/0025-3227(78)90018-X), 1978.
- Romanovsky, V. E., Smith, S. L., and Christiansen, H. H.: Permafrost thermal state in the polar Northern Hemisphere during the international polar year 2007–2009: a synthesis, *Permafrost and Periglacial Processes*, 21, 106–116, <https://doi.org/10.1002/ppp.689>, 2010.
- Schaefer, K., Lantuit, H., Romanovsky, V. E., Schuur, E. A., and Witt, R.: The impact of the permafrost carbon feedback on global climate, *Environmental Research Letters*, 9, 85 003, <https://doi.org/10.1088/1748-9326/9/8/085003>, 2014.
- 780 Schuur, E. A., McGuire, A. D., Schädel, C., Grosse, G., Harden, J. W., Hayes, D. J., Hugelius, G., Koven, C. D., Kuhry, P., Lawrence, D. M., Natali, S. M., Olefeldt, D., Romanovsky, V. E., Schaefer, K., Turetsky, M. R., Treat, C. C., and Vonk, J. E.: Climate change and the permafrost carbon feedback, *Nature*, 520, 171–179, <https://doi.org/10.1038/nature14338>, 2015.
- Semiletov, I. P., Shakhova, N. E., Pipko, I. I., Pugach, S. P., Charkin, A. N., Dudarev, O. V., Kosmach, D. A., and Nishino, S.: Space-time dynamics of carbon and environmental parameters related to carbon dioxide emissions in the Buor-Khaya Bay and adjacent part of the
785 Laptev Sea, *Biogeosciences*, 10, 5977–5996, <https://doi.org/10.5194/bg-10-5977-2013>, 2013.
- Shiklomanov, A., Holmes, R., McClelland, J., Tank, S., and Spencer, R.: Arctic Great Rivers Observatory. Discharge Dataset, Version 2021121212, <https://www.arcticrivers.org/data>, 2021.



- Shinohara, N., Woo, C., Yamamoto, N., Hashimoto, K., Yoshida-Ohuchi, H., and Kawakami, Y.: Comparison of DNA sequencing and morphological identification techniques to characterize environmental fungal communities, *Scientific Reports*, 11, 2633, 790 <https://doi.org/10.1038/s41598-021-81996-w>, 2021.
- Stedmon, C. A., Amon, R. M., Rinehart, A. J., and Walker, S. A.: The supply and characteristics of colored dissolved organic matter (CDOM) in the Arctic Ocean: Pan Arctic trends and differences, *Marine Chemistry*, 124, 108–118, <https://doi.org/10.1016/j.marchem.2010.12.007>, 2011.
- Stramski, D., Reynolds, R. A., Kaczmarek, S., Uitz, J., and Zheng, G.: Correction of pathlength amplification in the filter-pad technique for measurements of particulate absorption coefficient in the visible spectral region, *Applied Optics*, 54, 6763, 795 <https://doi.org/10.1364/ao.54.006763>, 2015.
- Tank, S. E., Striegl, R. G., McClelland, J. W., and Kokelj, S. V.: Multi-decadal increases in dissolved organic carbon and alkalinity flux from the Mackenzie drainage basin to the Arctic Ocean, *Environmental Research Letters*, 11, 054015, <https://doi.org/10.1088/1748-9326/11/5/054015>, 2016.
- 800 Tanski, G., Lantuit, H., Ruttor, S., Knoblauch, C., Radosavljevic, B., Strauss, J., Wolter, J., Irrgang, A. M., Ramage, J., and Fritz, M.: Transformation of terrestrial organic matter along thermokarst-affected permafrost coasts in the Arctic, *Science of the Total Environment*, 581–582, 434–447, <https://doi.org/10.1016/j.scitotenv.2016.12.152>, 2017.
- Tanski, G., Wagner, D., Knoblauch, C., Fritz, M., Sachs, T., and Lantuit, H.: Rapid CO₂ Release From Eroding Permafrost in Seawater, *Geophysical Research Letters*, 46, 11 244–11 252, <https://doi.org/10.1029/2019GL084303>, 2019.
- 805 Tassan, S.: A sensitivity analysis of the 'Transmittance-Reflectance' method for measuring light absorption by aquatic particles, *Journal of Plankton Research*, 24, 757–774, <https://doi.org/10.1093/plankt/24.8.757>, 2002.
- Tassan, S. and Ferrari, G. M.: An alternative approach to absorption measurements of aquatic particles retained on filters, *Limnology and Oceanography*, 40, 1358–1368, <https://doi.org/10.4319/lo.1995.40.8.1358>, 1995.
- Tassan, S. and Ferrari, G. M.: Variability of light absorption by aquatic particles in the near-infrared spectral region, *Applied Optics*, 42, 810 4802, <https://doi.org/10.1364/AO.42.004802>, 2003.
- Tisserand, L., Dadaglio, L., Intertaglia, L., Catala, P., Panagiotopoulos, C., Obernosterer, I., and Joux, F.: Use of organic exudates from two polar diatoms by bacterial isolates from the Arctic Ocean: Diatom exudates and Arctic bacteria, *Philosophical Transactions of the Royal Society A: Mathematical, Physical and Engineering Sciences*, 378, 20190356, <https://doi.org/10.1098/rsta.2019.0356>, 2020.
- Van Heukelem, L. and Thomas, C. S.: Computer-assisted high-performance liquid chromatography method development with applications 815 to the isolation and analysis of phytoplankton pigments, *Journal of Chromatography A*, 910, 31–49, [https://doi.org/10.1016/S0378-4347\(00\)00603-4](https://doi.org/10.1016/S0378-4347(00)00603-4), 2001.
- Viollier, E., Inglett, P., Hunter, K., Roychoudhury, A., and Van Cappellen, P.: The ferrozine method revisited: Fe(II)/Fe(III) determination in natural waters, *Applied Geochemistry*, 15, 785–790, [https://doi.org/10.1016/S0883-2927\(99\)00097-9](https://doi.org/10.1016/S0883-2927(99)00097-9), 2000.
- Vonk, J. E. and Gustafsson, Ö.: Permafrost-carbon complexities, *Nature Geoscience*, 6, 675–676, <https://doi.org/10.1038/ngeo1937>, 2013.
- 820 Vonk, J. E., Sanchez-Garca, L., Van Dongen, B. E., Alling, V., Kosmach, D., Charkin, A., Semiletov, I. P., Dudarev, O. V., Shakhova, N., Roos, P., Eglinton, T. I., Andersson, A., and Gustafsson, A.: Activation of old carbon by erosion of coastal and subsea permafrost in Arctic Siberia, *Nature*, 489, 137–140, <https://doi.org/10.1038/nature11392>, 2012.
- Vonk, J. E., Semiletov, I. P., Dudarev, O. V., Eglinton, T. I., Andersson, A., Shakhova, N., Charkin, A., Heim, B., and Gustafsson, Ö.: Preferential burial of permafrost-derived organic carbon in Siberian-Arctic shelf waters, *Journal of Geophysical Research: Oceans*, 119, 825 8410–8421, <https://doi.org/10.1002/2014JC010261>, 2014.



- Voss, K., Leymarie, E., Flora, S., Carol Johnson, B., Gleason, A., Yarbrough, M., Feinholz, M., and Houlihan, T.: Improved shadow correction for the marine optical buoy, MOBY, *Optics Express*, 29, 34 411, <https://doi.org/10.1364/OE.440479>, 2021.
- Walters, W., Hyde, E. R., Berg-Lyons, D., Ackermann, G., Humphrey, G., Parada, A., Gilbert, J. A., Jansson, J. K., Caporaso, J. G., Fuhrman, J. A., Apprill, A., and Knight, R.: Improved Bacterial 16S rRNA Gene (V4 and V4-5) and Fungal Internal Transcribed Spacer Marker Gene Primers for Microbial Community Surveys, *mSystems*, 1, e00 009–15, <https://doi.org/10.1128/mSystems.00009-15>, 2016.
- 830 Wegner, C., Bennett, K. E., de Vernal, A., Forwick, M., Fritz, M., Heikkilä, M., Łącka, M., Lantuit, H., Laska, M., Moskalik, M., O'Regan, M., Pawłowska, J., Promińska, A., Rachold, V., Vonk, J. E., and Werner, K.: Variability in transport of terrigenous material on the shelves and the deep Arctic Ocean during the Holocene, *Polar Research*, 34, <https://doi.org/10.3402/polar.v34.24964>, 2015.
- Weishaar, J. L., Aiken, G. R., Bergamaschi, B. A., Fram, M. S., Fujii, R., and Mopper, K.: Evaluation of specific ultraviolet absorbance as an indicator of the chemical composition and reactivity of dissolved organic carbon, *Environmental Science Technology*, 37, 4702–4708, <https://doi.org/10.1021/es030360x>, 2003.
- 835 Yamamoto, S., Alcauskas, J. B., and Crozier, T. E.: Solubility of methane in distilled water and seawater, *Journal of Chemical Engineering Data*, 21, 78–80, <https://doi.org/10.1021/je60068a029>, 1976.
- Zibordi, G., Voss, K. J., Carol Johnson, B., and Mueller, J. L.: Ocean optics and biogeochemistry protocols for satellite ocean colour sensor validation, volume 3.0: Protocols for satellite ocean colour data validation: In situ optical radiometry, Tech. rep., 2019.
- 840 Zielinski, R., Otton, J., and Budahn, J.: Use of radium isotopes to determine the age and origin of radioactive barite at oil-field production sites, *Environmental Pollution*, 113, 299–309, [https://doi.org/10.1016/S0269-7491\(00\)00188-3](https://doi.org/10.1016/S0269-7491(00)00188-3), 2001.