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Mixed Coagulant-flocculant optimization for pharmaceutical effluent pretreatment using response surface methodology and gaussian process regression

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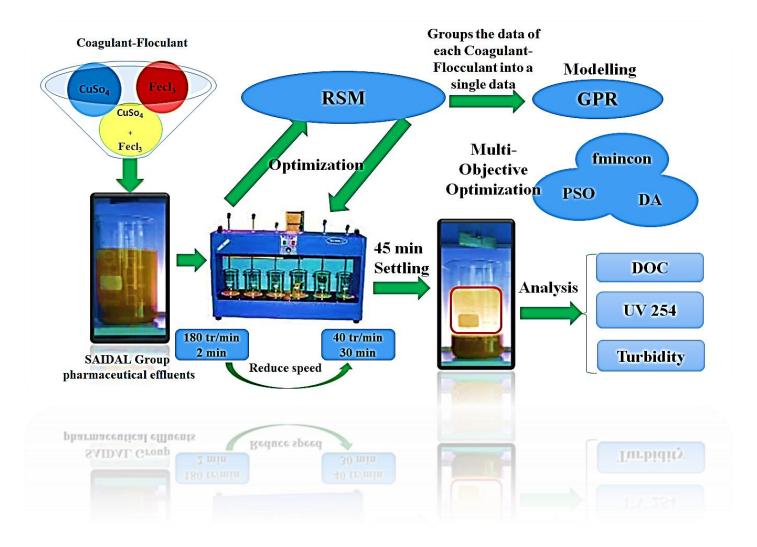
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Abstract

Wastewater from the Antibiotical-Saidal pharmaceutical plant (Medéa) was pretreated by coagulation-flocculation using copper sulfate (CuSO₄), iron chloride (FeCl₃), and mixture of the two salts combined in a 1:1 (v/v) ratio in the present study. Response surface methodology (RSM) was used to optimize pH and coagulant dosage as independent variables, while dissolved organic carbon (DOC), absorbance at 254 nm (UV 254), and turbidity were provided as dependent variables in the central composite design (CCD). Then, the databases of the three treatments were combined in a single database to create a general model valid for the three treatments at the same time, and to predict the reduction rates of DOC, UV254, and turbidity, using the Gaussian process regression coupled with the dragonfly optimization algorithm (GPR-DA).

To have the best model obtained between RMS and GPR-DA, an experimental validation was carried out after having had the optimal conditions of each type of coagulant, using the multi-objective optimization technique. The results of the experimental validation show the superiority of the GPR-DA model compared to the RSM model. Also, the results show that the mixed coagulant (CuSO₄+ FeCl₃) obtain better results than CuSO₄ or FeCl₃ alone with a treatment efficiency equal to 92.68% at pH = 5 and dosage = 600 mg/L, and the reductions in DOC, UV 254 and turbidity are 97.32%, 82.90% and 96.47%, respectively.

Graphical Abstract



Keywords

Pharmaceutical effluent, Coagulation-flocculation, Response surface methodology, Gaussian process regression, Multi-objective optimization.

1. Introduction

In recent years, the pharmaceutical industries have experienced intense development due to population growth and enormous public health needs (Gadipelly et al., 2014; Ng et al., 2016; Qian et al., 2019), which has also led to an increase in production and consequently an increase in associated wastewater discharges affecting natural resources (Ghumra et al.,

2021; Rawat, 2019). These effluents, if not adequately treated, become a significant source of micro-contaminants in the environment, resulting in harmful impacts on aquatic life and human health, such as water toxicity, pathogenic bacteria, and genotoxicity (Kumar et al., 2010; López-Fernández et al., 2012; Qian et al., 2019). Pharmaceutical wastewater contains complex mixtures of chemicals, intermediates, solvents, acids, alkalis, and many industry-specific additives used in manufacturing different pharmaceuticals (Lalwani et al., 2020). As a result, the wastewater generated may contain colors, odors, dissolved and suspended solids, inorganic chemicals, high turbidity, dissolved organic carbon, volatile organic compounds (VOCs), heavy metals, micro and macro pollutants specific to the industry; likewise, there are significant quantities of biological compounds, solvents, and catalysts that are poisonous and resistant to biodegradation (Gadipelly et al., 2014; Lalwani et al., 2020; Qian et al., 2015; Sreekanth et al., 2009). These complex mixtures are characterized by very high organic resistance (mainly provided by various organic solvents), high salinity, low biodegradability, and a wide pH range (Changotra et al., 2019a; Jose and Philip, 2021; Kaya et al., 2017). All these make pharmaceutical wastewater treatment very difficult and, as a consequence, pharmaceutical wastewater treatment methods are extensively studied with a view to effectively reduce pollution. This is part of a promising strategy to reduce water shortages and, above all, an essential step for addressing environmental issues (Chen et al., 2020), but also for emerging uses of water in the technological field (Boldrini et al., 2017; Changotra et al., 2019b; Santhosh et al., 2016; Wu et al., 2017). Thus, wastewater treatment plants have been installed in every industry to improve ecosystem and human health and help to support water harvesting (Ho et al., 2019).

According to the literature (Shahedi et al., 2020), wastewater treatment methods could be classified into three main categories: physical, chemical, and biological processes. Among

these, biological treatments remain the most widely used wastewater treatment process at pilot and industrial scale due to their environmentally friendly operations and costeffectiveness (Benitez et al., 2001; El-Din and Smith, 2002). Biological activities are successful in eliminating the majority of organic pollutants and ammoniacal nitrogen from the environment. Svojitka et al. (2017) reported a chemical oxygen demand (COD) removal efficiency of 78 % in a pilot anaerobic membrane bioreactor for postpharmaceutical wastewater treatment. Rao et al. (2005) show that fixed-film reactors can remove 60 to 70 % of the organic matter from the wastewater when the organic input rate is 10 g DOC/L/day. However, the limited treatment effectiveness of existing wastewater treatment plants in degrading recalcitrant and toxic pharmaceutical wastewater has led to the exploration of more radical pharmaceutical wastewater treatment methodologies to improve the biodegradability of wastewater before biological treatment (Changotra et al., 2019b). Indeed, multiple studies have indicated that various biological systems may be unable to break down weakly biodegradable, resistant, and refractory substances (Lapworth et al., 2012; Prieto-Rodríguez et al., 2013; Tran et al., 2014). As a result, Liu et al. (2012) reported that contents of medicines were discovered in sewage treatment plant effluents and also that pharmaceutical compounds were detected in surface water, soil and groundwater even after biological treatments (Joss et al., 2006; K'oreje et al., 2016). It is also revealed that the use of biological treatment processes in the presence of antibiotics creates long-term antimicrobial resistance (Lalwani et al., 2020; Zhou et al., 2006). Several wastewater processing processes can be combined for more efficiency to overcome this problem. However, the complete treatment of wastewater generally requires many steps (Sher et al., 2013). Due to the chemical stability and the low biodegradability of several organic pollutants, e.g. Hormones, biological treatments have generally been coupled as a complementary treatment step to attempt to overcome the reduced efficacy of conventional treatment methods (Cheng et al., 2020; Rodrigueza et al., 2016; Yarahmadi et al., 2018). Various combinations of conventional wastewater treatment processes have been used, such as coagulation (Changotra et al., 2019b), adsorption (Carvalho et al., 2012), membrane filtration (Ganiyu et al., 2015), incineration (Mu et al., 2012), and as well as biological treatments coupled with coagulation and flocculation processes for the treatment of pharmaceutical wastewater (Changotra et al., 2019b; Pal, 2018).During coagulation-flocculation treatments, the addition of chemicals modifies the physical state of the dissolved and suspended solids and promotes the removal of these solids by precipitation (Ashraf et al., 2016). The suppression mechanism of this process is mainly due to the load neutralization of the colloids charged negatively by cationic hydrolysis products, which allows the initial aggregation of colloidal particles to form micro-flakes (Sher et al., 2013). In addition, coagulation is a treatment that can reduce the turbidity and the elimination of the color and pathogens (Ashraf et al., 2016), as well as the elimination of organic matter (Ashraf et al., 2016). The effectiveness of the coagulation-flocculation process depends on several factors, including the type of coagulant, dosage, pH, temperature, ionic strength, nature and concentration of organic matter, the total dissolved solids, as well as the size and distribution of colloidal particles in suspensions (Sher et al., 2013). The most used coagulants are iron and alum salts. These coagulants favor the agglomeration of the particles by reducing the electrostatic charges of the surface of the particles in the acid pH region where the hydrolyzed metal species are plentiful (Santo et al., 2012).

Coagulation/flocculation have been reported as an effective method in the treatment of wastewater in many industries such as: the treatment of minerals, metallurgy, and textile, the treatment of Tannery effluents, yeast wastewater, textile, petroleum refinery, dyeing

bath and dyeing house, aquaculture, vinassas, municipal, lixivitations, poll page, mill with olives and many others (Sher et al., 2013).

For this purpose, many researches on the development of coagulation-flocculation methods for the treatment of industrial effluents primarily aimed at performance optimization have been in progress (Adesina et al., 2019; Tahraoui et al., 2021b; Trinh and Kang, 2011).

Various methods of optimizing the wastewater treatment methods have been used in recent decades to improve the process performance, such as Response Surface Methodology (RSM) shown in Table 1.

In this research, the potential and efficiency of a coagulation and flocculation process using copper sulfate (CuSO₄), ferric chloride (FeCl₃), and a combination of cupric sulfate and ferric chloride in a ratio of 1:1 (CuSO₄+ FeCl₃) for the pretreatment of pharmaceutical industrial effluents are investigated. The models created using RSM with central composite design technique (CCD) are used to optimize the treatment condition. Before and after the tests, the dissolved organic carbon (DOC) in the water, the absorbance at 254 Nm (UV 254), and the turbidity were measured. The two variables (pH and coagulation dosages) and the three responses (DOC, turbidity, and UV 254) were taken into account to find the best treatment condition. Then, the Gaussian process regression model coupled with the dragonfly optimization algorithm (GPR-DA) was used to model the three outputs after combining the databases from the three treatment approaches. Afterwards, multi-objective optimization procedures by using three techniques (fmincon, dragonfly and particle swarm) were carried out to find the optimal treatment conditions. Finally, an application was created using the MATLAB guide to provide a simple way to implement Multi-Objective Optimization (MOO) and predict DOC, UV254, turbidity and treatment efficiency. To our knowledge, optimizing the efficiency of coagulants flocculant used to pre-treat pharmaceutical effluents derived from RSM and GPR-DA has never been attempted before. Moreover, comparison of the MOO approaches has never been made before in this context.

The article is organized as follows. Section 2 presents the treatment of liquid effluent from the pharmaceutical plant by two individual coagulants/flocculants and their mixture and the experiments carried out using RSM and CCD. Then models are developed from the experimental data by using Gaussian process regression coupled with the dragonfly (GPR-DA). Based on the developed models, MOO based on three algorithms was performed to find the optimal treatment condition. Section 3 presents a comparison between the two individual coagulants/flocculants and their mixture in order to find the best treatment approach. The MOO results are validated through experiments. The last section concludes the paper.

Table 1 – Reviewed Research Articles on Optimizing Wastewater Treatment by RSM.

Article titles	Contributions and limitations	References
Response surface	This study focused on the use of Moringa	Adesina et al.,
methodology approach	oleifera seeds as a coagulant for surface water	2019
to optimization of	treatment using RSM. Four factors were	
process parameter for	evaluated, namely the stabilization time, the	
coagulation process of	stirring time, the stirring speed and the	
surface water using	concentration of Moringa oleifera seed extract.	
Moringa oleifera seed	The model predicts the lowest turbidity of 5.49	
	NTU with optimal conditions of stabilization	
	time of 120 min, stirring speed of 100 rpm,	
	stirring time of 10 min and coagulant	
	concentration of 3 g/L. Despite the good	
	results obtained in this study in terms of	

turbidity elimination, this method remains limited by the high costs of operation and preparation of Moringa oleifera seeds. Furthermore, the dose of Moringa oleifera used in this treatment is very high (3g/L) compared to other coagulants such as aluminum sulfate which generally does not exceed 1g/L and with an effective very high turbidity removal (Tahraoui et al., 2021b; Trinh and Kang, 2011). In addition, this study only focused on the removal of turbidity from the raw water of the Nakdong River and the method cannot be used in the treatment of pharmaceutical effluents.

Response surface methodological approach to optimize the coagulationflocculation process in drinking water treatment

RSM was used in this work to find the optimal combination of coagulant dose and pH with Kang, 2011 respect to the highest turbidity and organic carbon removal efficiency dissolved (DOC). The results obtained with polyaluminium chloride (PACl) were compared with those obtained using a conventional coagulant such as aluminum. Quadratic models developed for the two responses (turbidity removal and DOC removal) indicated that the optimal conditions for a PAC1 concentration of 0.11 mM at pH

Trinh

and

7.4 and an alum concentration of 0.15 mM at pH 6.6. The compromise to optimize both responses simultaneously resulted in 91.4% turbidity removal and 31.2% DOC removal using PACl, while removals using aluminum were 86.3% turbidity and 34.3% DOC. This study remains very weak to apply to the treatment of water loaded with organic matter, in particular for the treatment of pharmaceutical effluents. Because the two coagulants used in this study gave a very low DOC removal rate, the removal rate did not exceed 34.3% at treated water DOC values of 3.69 - 3.76 mg/L.

Optimisation and
Prediction of the
Coagulant Dose for
the Elimination of
Organic
Micropollutants
Based on Turbidity

RSM was applied to optimize and prediction (Tahraoui the coagulant dose for the removal of organic al., 2021b) micropollutants based on turbidity. The water contaminated with humic acid was treated using the aluminum sulfate coagulant according to pH, temperature, conductivity, turbidity and Total alkalimetric titre. The results show a very high treatment efficiency of 99.96% using coagulation with a dosage of 109.50 mg/L.

The results of this method are very interesting

et

apply in the field of treatment of pharmaceutical effluents. But, this method is limited by many of the inputs used., as the interaction effect of the independent variables and the collaborative analysis between a large number of independent and dependent variables becomes difficult for accurate optimization (Singh and Kumar, 2020a). This poses a difficult control problem because it is nonlinear and complex (Tahraoui et al., 2021b).

2. MATERIALS AND METHODS

2.1. Collection of wastewater samples

In Algeria, the Saidal Group Medicine Production Factory is a stock company with a capital of 2.5 billion Algerian dinars. The State holds 80% of the capital of the Saidal Group, and the remaining 20% were assigned in 1999 to institutional investors and natural persons (www.saidalgroup.dz). It contains several subsidiaries on the national territory: Biotic subsidiary, Farmall subsidiary, antibiotic subsidiary, and others. Figure 1 shows the different subsidiaries, their location, and drugs manufactured in the company (GROUPE, 2001).

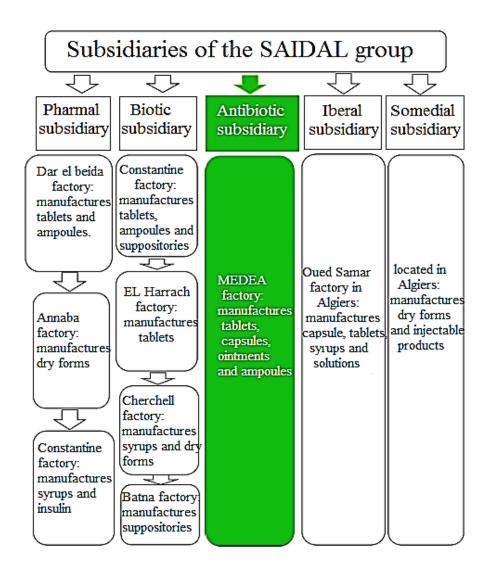


Figure 1 – Characterization of the different subsidiaries of the SAIDAL group in Algeria.

In this study, the wastewater was collected for one month from the rejection of the medéa antibiotic complex (Figure 2) before returning to to the treatment plant, and the selection of this site is because the wastewater from this site is difficult for biological degradation. The effluents from this type of plants are less biodegradable (Lapworth et al., 2012; Prieto-Rodríguez et al., 2013; Tran et al., 2014), and also the presence of antibiotics creates long-term antimicrobial resistance (Lalwani et al., 2020; Zhou et al., 2006). It should be noted that this subsidiary is located in Médéa, about 80 km west of Algeria (Iguergaziz et al., 2019), and it started in 1986 specializing in the production of penicillinic and non-penicillinic antibiotics for anti-inflammatory and antipyretic (www.saidalgroup.dz). It has

facilities needed to manufacture drugs from obtaining the active ingredient until its galenic formatting.

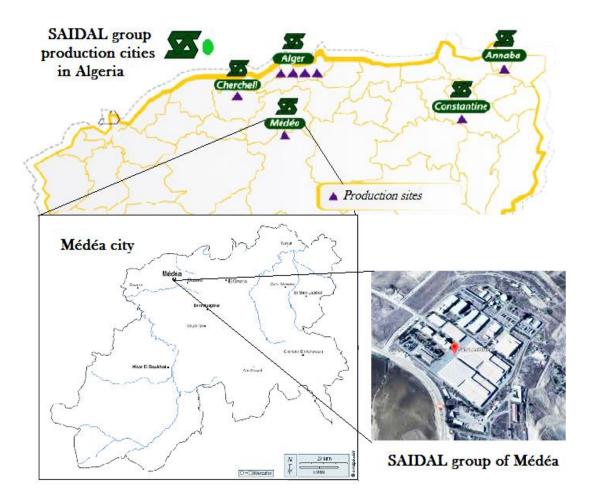


Figure 2 – Location of the sampling site of the SAIDAL group in Médéa.

The complex has an area of 25 hectares with 19 hectares of built area. The medéa antibiotic complex has several buildings and annexes including (www.saidalgroup.dz):

- A bulk raw material production building by fermentation.
- A production building of bulk raw materials by chemical synook from fermentation products.
- Two building vessels of pharmaceutical specialties, one dedicated to penicillinic products and the other to non-penicillinic products.
- A unit of production of packaging articles (printing).

• General services were necessary for the operation of these facilities.

The wastewater collected was homogenized and combined with concentrated H_2SO_4 (2 mL / L) and kept in a refrigerator at 4 °C to avoid photo-catalyzed alteration (Singh and Kumar, 2020b).

Table 2 provides a summary of the physical and chemical properties of the wastewater. It can be seen that the wastewater is complex with many components.

Table 2 – Characteristics of wastewater.

Parameters	Values	Unit
potential Hydrogen	7.54±0.09	-
Turbidity	44.92±0.06	NTU
UV 254	0.353 ± 0.01	-
DOC	18.50±0.04	mg/L
Conductivity	1968.00±0.17	μS cm ⁻¹
Total dissolved solids	462.00±0.20	-
Nitrate	23.00±0.80	mg/L
Hardness	145.00±0.50	°F
Calcium	104.21±0.90	mg/L
Magnesium	136.45±0.60	mg/L
Total alkalimetric titre	283.50±0.30	mg/L
Bicarbonate	301.45±0.80	mg/L
Chlorides	342.54±0.40	mg/L
Nitrogen dioxide	0.10 ± 0.07	mg/L
Ammonium	0.20 ± 0.09	mg/L
Phosphate	0.20 ± 0.02	mg/L
Sulfate	341.00±0.50	mg/L
Sodium	223.00±0.70	mg/L
Potassium	3.20±0.04	mg/L

Manganese	0.49 ± 0.02	mg/L
Iron	1.50±0.06	mg/L
Aluminum	0.60 ± 0.09	mg/L
Dry residues	1905.00±0.10	mg/L

2.2. **RSM**

According to Khuri and Cornell (1996), the RSM principle combines mathematical and statistical approaches for analyzing the connections between independent variables and one or more responses (Rifi et al., 2022; Tahraoui et al., 2021b). The design of experiment is required to get appropriate and reliable measurements of the responses that are of particular interest. A linear function cannot adequately reflect the connection between the response and the independent factors (Rifi et al., 2022; Singh and Kumar, 2020b). The use of a model that includes curvature is often required to estimate the response in the area near to ideal, and in most circumstances, a second-order model is sufficient to do this (Daud et al., 2018; Tahraoui et al., 2021b). A CCD is explored in detail, and it is a particularly effective design strategy for fitting second-order models that have been shown in (Rifi et al., 2022; Tahraoui et al., 2021b). This method is often employed when the output is influenced by various input elements (Rifi et al., 2022; Tahraoui et al., 2021b). In conjunction with the CCD technique, the RSM approach is used to optimize the process under investigation. In this case, the independent factors considered are pH and the dosage of the coagulant, whereas DOC, UV 254, and turbidity are chosen as the dependent variables. The JMP program (version 13 pro) is used to perform the statistical analysis. The range of independent variables for the two coagulants (CuSO₄+ FeCl₃), and the combination relative to each other with a ratio (1:1, v/v), were chosen based on preliminary studies conducted to determine the amount of DOC reduced by each coagulant. Initial studies were conducted at pH values ranging from 2 to 13 (note that sodium hydroxide (NaOH) and hydrochloric acid (HCl) were used for pH adjustment), and dosage levels ranging from 100 to 1500 mg / L. The parameters interval (pH and coagulant dosage) for both kinds of coagulants and the combination were determined based on the DOC decrease, as indicated in Table 3.

Table 3 – Coding and real levels for the CCD model.

Coagulant-floculant	pH (x1)	pH (x1) Coagulant-floculant mg/L(x2					
	-1	0	+1	-1	0	+1	
$CuSO_4$	7	9	11	600	900	1200	
$FeCl_3$	3	5	7	200	600	1000	
$CuSO_4 + FeCl_3$	5	7	9	200	600	1000	

The complete factorial design CCD gives 13 experiments for each coagulant according to equation 1 (Tahraoui et al., 2021b):

$$N = 2^k + 2k + c$$

(1)

where k is the number of independent variables, 2^k is the point factor, 2k is the axial points, and c is the number of experiments in the center points (Tahraoui et al., 2021b).

Once the range of independent variables for each incorporated coagulant and the set of different conditions have been obtained, the laboratory tests are carried out using the jar test, and the experimental results are incorporated into JMP shown in Table 4.

The jar test was carried out using beakers filled with 1 L of the collected effluent. Following the addition of the coagulant and pH adjustment by sodium hydroxide (NaOH) or hydrochloric acid (HCl) using pH meter type JP selecta, the samples were mixed at 180 rpm for 5 minutes to establish a homogeneous mixture and brought into contact between the pharmaceutical effluents and the coagulant, in order to neutralize the charge of the particles and to initiate the flocculation process by the formation of microflocs (Daud et al.,

2015; Khouni et al., 2020; Momeni et al., 2018; Verma et al., 2010). Following that, the speed was dropped to 40 rpm for 30 minutes to ensure the contacting and growth of the flocs (Daud et al., 2018; Momeni et al., 2018; Tahraoui et al., 2021b; Verma et al., 2010). Where, all experiments were conducted at a temperature of $18 \pm 2^{\circ}$ C. In addition, the time and mixing speed have been set with an automatic controller.

In each pot, the supernatant was taken after 45 minutes of decantation and filtered through pleated filter paper to determine the concentrations of DOC, UV 254, and turbidity. Conventional methods were used to perform the physicochemical analyzes. The turbidity and UV absorbance at 254 nm were measured, respectively, using a Hach 2100-year orchids turbidimeter and a Cecil Server 2 Gratin Spectrophotometer UV spectrophotometer. The water DOC dosage was carried out by the determination of potassium permanganate oxidizing (Fatombi et al., 2007; Rodier et al., 2016).

It should be noted that the DOC and UV 254 parameters are selected in this work due to the presence of the most used natural organic material (NOM) being the total organic carbon (COT) or the dissolved organic carbon (DOC), and Ultraviolet absorbance at a specific length, 254nm (UV 254) (HARRAT, 2013). These two parameters (DOC and UV 254) complement each other: the DOC provides indications on the quantity of NOM present and available for reaction, and UV 254 rather indicates its responsiveness since it is an indicator of the aromatic links contained in humic substances (HARRAT, 2013). Thus they can characterize conjugated systems (aromatic nuclei in particular), which are very abundant in the macromolecular structures that constitute the organic matter dissolved in natural waters (HARRAT, 2013).

2.3. Gaussian Process Rregression

Gaussian Process (GP) is a stochastic process with a series of random variables. Any finite range of those random variables has a joint Gaussian distribution. The probabilistic

illustration of a goal function can be used for regression and classification (Tahraoui et al., 2022b).

Gaussian Process Regression (GPR) provides a probabilistic, nonparametric strategy for solving nonlinear regression problems. GPR assumes that the measurements of the output variable *y* are generated in the following way (Park et al., 2017):

$$y = f(x(k)) + \varepsilon$$

(2)

where x stands for the measurements of input variables, f is the unknown functional dependence, and ε is a Gaussian noise with zero mean and variance σ_n^2 . GPR uses GP as a prior to describe the distribution on the target function f(x) (Park et al., 2017; Tahraoui et al., 2022b). In GPR, the function values $f_{1:n} = (f_1, ..., f_n)$ corresponding to the input values $x_{1:n} = (x_1, ..., x_n)$ are treated as random variables, where $f_i = f(x_i)$ (Park et al., 2017; Tahraoui et al., 2022b). GP is defined as a collection of random variables (stochastic process) and any finite number of which are assumed to be jointly Gaussian distributed (Park et al., 2017; Tahraoui et al., 2022b). GP can fully describe the distribution over an unknown function, f(x), by its mean function, m(x) = E[f(x)], and a kernel function, k(x,x'), that approximates the covariance E[(f(x)-m(x))(f(x')-m(x'))]. The kernel (covariance) function represents a geometrical distance measure assuming that the more closely located inputs would be more correlated in terms of their function values (Park et al., 2017; Tahraoui et al., 2022b). That is, the prior on the function values is represented as (Park et al., 2017; Tahraoui et al., 2022b):

$$(f^{1:n}) = GP(m(\cdot), k(\cdot, \cdot))$$

(3)

where:

m(.) is a mean function capturing the overall trend in the target function value and k(.,.) is a kernel function used to approximate the covariance.

In GPR, the kernel (covariance) function describes the structure of the target function. Thus, the type of kernel function k(x,x') used to build a GPR model and its parameters can strongly affect the overall representability of the GPR model and impact the accuracy of the prediction model. A wide variety of kernel functions can be used (Rasmussen and Williams, 2006).

In this article, the main objective is to create general models to predict the reduction rates of each dependent parameter for the three treatments using GPR. The dragonfly algorithm (DA) is used to optimize the GPR models by finding the best parameters of the karnel functions leading to the best performance. In this study, ten kernel functions have been optimized with Basis Function (constant, linear and zero) for each output and the one giving the best performance is selected.

The following ten karnel functions are used in this work:

Squared Exponential Kernel

$$k(x_i, x_j | \theta) = \sigma_f^2 \exp \left[-\frac{1}{2} \frac{(x_i - x_j)^T (x_i - x_j)}{\sigma_l^2} \right]$$

(4)

Where σ_l is the characteristic length scale, and σ_f is the signal standard deviation.

• Exponential Kernel

$$k(x_i, x_j | \theta) = \sigma_f^2 \exp\left[-\frac{r}{\sigma_l}\right]$$

(5)

Where σ_l is the characteristic length scale and $r = \sqrt{(x_i - x_j)^T (x_i - x_j)}$

• Matern 3/2

$$k(x_i, x_j | \theta) = \sigma_f^2 \left(1 + \frac{\sqrt{3}r}{\sigma_l} \right) \exp\left(-\frac{\sqrt{3}r}{\sigma_l} \right)$$
 where $r = \sqrt{(x_i - x_j)^T (x_i - x_j)}$

• Matern 5/2

(6)

$$k(x_{i}, x_{j} | \theta) = \sigma_{f}^{2} \left(1 + \frac{\sqrt{5}r}{\sigma_{l}} + \frac{5r^{2}}{3\sigma_{f}^{2}} \right) \exp\left(-\frac{\sqrt{5}r}{\sigma_{l}} \right) \quad \text{where} \quad r = \sqrt{(x - x_{j})^{T} (x - y_{j})^{T}}$$

$$(7)$$

• Rational Quadratic Kernel

$$k(x_i, x_j | \theta) = \sigma_f^2 \left(1 + \frac{r^2}{2\alpha \sigma_l^2} \right)^{-\alpha}$$

(8)

Where σ_l is the characteristic length scale, α is a positive scale mixture

parameter, and
$$r = \sqrt{(x_i - x_j)^T (x_i - x_j)}$$
.

• ARD Squared Exponential Kernel

$$k\left(x_{i}, x_{j} \middle| \theta\right) = \sigma_{f}^{2} \exp \left[-\frac{1}{2} \sum_{m=1}^{d} \frac{\left(x_{im} - x_{jm}\right)^{2}}{\sigma_{m}^{2}}\right]$$

(9)

• ARD Exponential Kernel

$$k(x_i, x_j | \theta) = \sigma_f^2 \exp(-r)$$
 where $r = \sqrt{\sum_{m=1}^d \frac{\left(x_{im} - x_{jm}\right)^2}{\sigma_m^2}}$ (10)

• ARD Matern 3/2

$$k(x_i, x_j | \theta) = \sigma_f^2 (1 + \sqrt{3}r) \exp(-\sqrt{3}r) \qquad \text{where} \qquad r = \sqrt{\sum_{m=1}^d \frac{(x_{im} - x_{jm})^2}{\sigma_m^2}}$$
(11)

• ARD Matern 5/2

$$k\left(x_{i}, x_{j} \middle| \theta\right) = \sigma_{f}^{2} \left(1 + \sqrt{5}r + \frac{5}{3}r^{2}\right) \exp\left(-\sqrt{5}r\right) \qquad \text{Where } r = \sqrt{\sum_{m=1}^{d} \frac{\left(x_{im} - x_{jm}\right)^{2}}{\sigma_{m}^{2}}}$$

$$(12)$$

• ARD Rational Quadratic Kernel

$$k(x_i, x_j | \theta) = \sigma_f^2 \left(1 + \frac{1}{2\alpha} \sum_{m=1}^d \frac{\left(x_{im} - x_{jm} \right)^2}{\sigma_m^2} \right)^{-\alpha}$$
(13)

2.4. Statistical evaluation criteria

Statistic analysis and the ANOVA have been used to investigate the statistical abilities of the generated models with a confidence level of 95%. The statistical adequacy of the developed models was assessed using a variety of variables, including the P-value, the value F, the degree of freedom (DF), the coefficient of determination (R^2), the adjusted determination of the coefficient (R_{adj}^2), and the Root Mean Square Error (RMSE) (Tahraoui et al., 2021b, 2022b, 2022a). The value F describes the variation of the responses, which can be checked by a regression equation, while the value P determines the statistical adequacy of the developed model. For a model to be significant, the P-value must be less than 5%, and the P-value for the inadequacy test must be greater than 5% (Singh and Kumar, 2020b).

On the other hand, the Correlation Coefficient (R), R², R_{adj}², RMSE, Mean Square Error (MSE) and Mean Absolute Error (MAE), Error Standard of Prediction (ESP), Error Prediction of Model (EPM) were used to evaluate the performance of each GPR model. These are calculated using the following equations (Belsley et al., 1980; Bousselma et al., 2021; Hong et al., 2007; Tahraoui et al., 2022b, 2022a, 2021a, 2021b, 2020).

$$R = \frac{\sum_{i=1}^{N} (y_{\text{exp}} - \overline{y}_{\text{exp}}) (y_{pred} - \overline{y}_{pred})}{\sqrt{\sum_{i=1}^{N} (y_{\text{exp}} - \overline{y}_{\text{exp}})^{2} \sum_{i=1}^{N} (y_{pred} - \overline{y}_{pred})^{2}}}$$
(14)

$$R_{adj}^{2} = 1 - \frac{(1 - R^{2})(N - 1)}{N - K - 1}$$
(15)

$$RMSE = \sqrt{\left(\frac{1}{N}\right)\left(\sum_{i=1}^{N} \left[\left(y_{\exp} - y_{pred}\right)\right]^{2}\right)}$$

$$(16 MSE = \left(\frac{1}{N}\right) \left(\sum_{i=1}^{N} \left(y_{\text{exp}} - y_{pred}\right)^{2}\right)$$

(17)
$$MAE = \left(\frac{1}{N}\right) \sum_{i=1}^{N} \left| y_{\text{exp}} - y_{\text{pred}} \right|$$

(18)

$$ESP(\%) = \frac{RMSE}{\overline{y_{exp}}} \times 100$$

(19)

$$EPM(\%) = \frac{100}{N} \sum_{i=1}^{N} \left| \frac{\left(y_{\text{exp}} - y_{\text{pred}} \right)}{y_{\text{exp}}} \right|$$

(20)

Where N is the number of data samples, K is the number of variables (inputs), y_{exp} and y_{pred} are the experimental and the predicted values respectively, \overline{y}_{exp} and \overline{y}_{pred} are respectively the average values of the experimental and the predicted values (Dolling and Varas, 2002; Manssouri et al., 2014, 2011).

3. Results and discussions

3.1. RSM

A statistical analysis of the developed RSM models was carried out using the "JMP 13 pro" program. This method found the mathematical relationship (Eq (3)) between two input parameters (pH and dosage) and three output parameters (DOC, UV 254, and turbidity) for three types of coagulants: CuSO₄, FeCl₃, and their mixture CuSO₄+ FeCl₃. The results of 13 laboratory tests for the individual coagulants and their mixture are provided in Table 4, where the best experimental values are highlighted in bold and

underlined. It should be noted that the output parameters (DOC, UV 254 and turbidity) were predicted by their final concentrations. Subsequently, the final values of each parameter were converted into percentages by equation (21), in order to know the elimination rate of each output according to the coagulants/flocculants.

Reduction rate (%)=
$$\left(\frac{Z_i - Z_f}{Z_i}\right) \times 100$$

(21)

Where Z_i is the initial value of each output (Table 2), and Z_f is the final value of each output.

Table 5 gives the statistical results necessary for the construction and interpretation of the RSM model. The equation took into account the two independent variables, the interactions between them, and their quadratic effect is given by Eq(21).

$$Y = \beta_0 + \beta_1 X_1 + \beta_2 X_2 + \beta_3 X_1 X_2 + \beta_4 X_1^2 + \beta_5 X_2^2$$
(21)

where X_1 is pH, X_2 is dosage, Y is the response variable, β_0 to β_5 are the model parameters. Subsequently, the parameters with high explanatory power were retained (PR <5%) which are bolded and underlined in Table 5 (Tahraoui et al., 2021b), On the other hand, the other parameters which have PR >5% were deleted (which are in bold and underlined in Table 5), and the models are represented by the equations given in Table 6.

In the DOC model, the pH quadratic effect with the dose of $CuSO_4$ has proven to be a non-significant parameter as p=12.32% and the same for the mixture dose ($CuSO_4+FeCl$) because their P=20.82% (Table 5.a). In the UV 254 model, it is shown that the interaction between pH and the dose of $CuSO_4$ is not significant as p=10.23% (Table 5.b).

Finally, in the turbidity model, it is found that the pH interaction with the dose of $CuSO_4$ is non-significant parameter as p=13.58% (Table 5.c). On the other hand, it is also found that the quadratic effect of the dose of $FeCl_3$ are insignificant parameters as p=10.82% (Table 5.c).

Table 4 – Set of experimental conditions of independent variables (pH and dosage) and response (DOC, UV 254, and turbidity) for CuSO₄, FeCl₃ and CuSO₄+FeCl₃ coagulants.

Run. No					CuS	04					
	n	pende ite ables	Responses								
			DOC	DOC	UV 254	UV 254	Turbidit	Turbidit			
	pН	Dosa			U V 234						
		ge	(mg/L)	Reductio		reductio	y (NITI)	y			
				n (%)		n (%)	(NTU)	reductio			
				(%)		(%)		n (%)			
1	-1	-1	8.50±0.	54.04±0.	0.125±0.	64.49±0.	20.91±0.	53.45±0.			
1	-1	-1	0.30±0.	47	0.123±0.	433	020	045			
2	-1	0	6.50±0.	64.86±0.	$0.115\pm0.$	433 67.51±0.	18.80±0.	58.16±0.			
2	-1	U	10	54	0.113±0.	911	180	401			
3	-1	1	5.50±0.	70.29±0.	$0.115\pm0.$	67.51±1.	17.39±0.	61.29±0.			
3	1	1	13	69	0.115±0.	07.31±1.	095	212			
4	0	-1	4.50±0.	75.68±0.	0.113±0.	67.89±0.	15.07±0.	66.46±0.			
•	· ·	1	05	27	002	433	105	234			
5	0	0	4.00±0.	78.38±0.	$0.104\pm0.$	70.63±0.	13.34±0.	70.31±0.			
	Ü	Ü	10	54	004	995	025	056			
6	0	0	4.00±0.	78.36±0.	0.105±0.	70.25±0.	13.38±0.	70.21±0.			
			08	42	003	850	053	118			
7	0	0	4.00±0.	78.40±0.	$0.104\pm0.$	70.44±0.	13.38±0.	70.21±0.			
			03	14	001	164	074	164			
8	0	0	4.00 ± 0 .	78.40±0.	$0.106\pm0.$	70.06±1.	13.25±0.	70.50±0.			
			07	35	005	397	066	146			
9	0	0	4.00 ± 0 .	78.38±0.	$0.106\pm0.$	70.06±1.	13.3±0.0	70.38±0.			
			12	62	004	179	45	100			
10	0	1	4.10±0.	77.84±0.	0.110±0.	68.83±1.	12.47±0.	72.25±0.			
			09	46	006	700	042	093			
11	1	-1	$2.80\pm0.$	84.88±1.	$0.093\pm0.$	73.65±0.	6.07 ± 0.0	86.49±0.			
			19	00	002	567	45	100			
12	1	0	1.78±0.	90.38±0.	0.080±0.	77.71±0.	2.46±0.0	94.52±0.			
			11	59	001	327	45	100			

13	1	1	2.50±0.	86.47±0.	0.090±0.	74.50±1.	4.96±0.0	88.96±0.
10	-	-	10	51	004	133	46	102
Run.					FeCl			
No								
	Inde	pende			Respo	onses		
		ite			1			
	vari	ables						
	рН	Dosa	DOC	DOC	UV 254	UV 254	Turbidit	Turbidit
	•	ge	(mg/L)	Reductio		reductio	у	y
		C	, ,	n		n	(NTU)	reductio
				(%)		(%)	, ,	n
								(%)
1	-1	-1	5.60 ± 0 .	69.75±0.	$0.112\pm0.$	68.27±1.	11.09±0.	75.30±0.
			18	949	006	769	123	275
2	-1	0	$4.80\pm0.$	74.04 ± 0 .	$0.109\pm0.$	69.21±0.	10.13±0.	77.45±0.
			10	541	003	713	047	105
3	-1	1	$4.40\pm0.$	$76.22\pm0.$	$0.101\pm0.$	71.38±1.	9.58 ± 0.4	78.66 ± 0 .
			06	337	004	133	10	913
4	0	-1	$7.20\pm0.$	61.08±0.	$0.134\pm0.$	61.94±1.	14.85±0.	66.94±0.
			12	623	004	145	026	059
5	0	0	$4.00\pm0.$	$78.38\pm0.$	$0.097\pm0.$	71.48±0.	9.51 ± 0.2	$78.83\pm0.$
			05	286	003	911	24	498
6	0	0	$4.00\pm0.$	$78.38\pm0.$	$0.099\pm0.$	71.95±0.	9.60 ± 0.1	$78.63\pm0.$
			16	842	002	567	16	258
7	0	0	$4.00\pm0.$	$78.38\pm0.$	$0.100\pm0.$	71.76±1.	9.56 ± 0.0	$78.72\pm0.$
			03	162	006	731	15	034
8	0	0	$4.00\pm0.$	$78.38\pm0.$	$0.098\pm0.$	72.14 ± 0 .	9.55 ± 0.1	78.74 ± 0 .
			07	354	002	433	56	348
9	0	0	$4.00\pm0.$	$78.38\pm0.$	$0.100\pm0.$	70.91±1.	9.60 ± 0.2	$78.63\pm0.$
			09	510	004	073	28	508
10	0	1	$2.50\pm0.$	86.49±0.	$0.093\pm0.$	73.56±0.	4.46±0.0	90.07±0.
			17	922	004	495	40	090
11	1	-1	12 .0±0	$35.14\pm0.$	$0.244\pm0.$	$30.87\pm1.$	$29.68\pm0.$	$33.92\pm0.$
			.12	655	007	963	184	409
12	1	0	$7.20\pm0.$	61.08±0.	$0.162\pm0.$	54.10±1.	18.96±0.	$57.78\pm0.$
			08	408	006	700	038	084
13	1	1	$3.70\pm0.$	79.98±0.	$0.130\pm0.$	$67.89\pm1.$	9.02 ± 0.0	79.91±0.
			13	695	006	560	25	056
Run.					CuSo ₄ +F	FeCl ₃		
No								
		pende			Respo	onses		
		ite						
		ables						
	pН	Dosa	DOC	DOC	UV 254	UV 254	Turbidit	Turbidit

-		ge	(mg/L)	Reductio		reductio	у	y
				n		n	(NTU)	reductio
				(%)		(%)		n
								(%)
1	-1	-1	$1.70\pm0.$	90.79±0.	$0.110\pm0.$	68.83 ± 0 .	3.60 ± 0.1	91.98±0.
			05	24	003	850	01	226
2	-1	0	$0.50\pm0.$	97.32±0.	$0.060\pm0.$	82.90±0.	1.58 ± 0.0	96.47±0.
			03	14	002	433	40	090
3	-1	1	$3.00\pm0.$	$83.78\pm0.$	$0.130\pm0.$	$63.26\pm0.$	6.20 ± 0.1	86.20 ± 0 .
			03	14	003	865	10	245
4	0	-1	$2.90\pm0.$	84.31±0.	$0.153\pm0.$	$56.65\pm1.$	5.58 ± 0.0	$87.57\pm0.$
			18	95	004	235	17	039
5	0	0	$1.42\pm0.$	$92.34\pm0.$	$0.113\pm0.$	$68.08\pm1.$	3.30 ± 0.0	92.66 ± 0 .
			35	79	005	277	42	093
6	0	0	$1.48\pm0.$	92.00 ± 0 .	$0.114\pm0.$	67.80 ± 0 .	3.20 ± 0.0	$92.86\pm0.$
			02	09	004	995	96	214
7	0	0	$1.45\pm0.$	$92.18\pm0.$	$0.110\pm0.$	$68.74 \pm 1.$	3.25 ± 0.0	$92.75\pm0.$
			14	74	004	145	15	034
8	0	0	$1.40\pm0.$	$92.43\pm0.$	$0.112\pm0.$	68.27 ± 0 .	3.30 ± 0.0	$92.65\pm0.$
			03	16	003	981	10	022
9	0	0	$1.47\pm0.$	92.05 ± 0 .	$0.109\pm0.$	69.12±1.	3.20 ± 0.1	$92.88\pm0.$
			03	19	004	021	50	335
10	0	1	$1.60\pm0.$	91.33±0.	$0.115\pm0.$	$67.42\pm1.$	3.70 ± 0.0	91.75±0.
			09	46	006	769	74	164
11	1	-1	$3.50\pm0.$	81.10±0.	$0.165\pm0.$	53.16±1.	7.18 ± 0.0	84.01±0.
			12	65	006	731	30	067
12	1	0	$2.90\pm0.$	84.31±0.	$0.155\pm0.$	$56.09\pm1.$	6.15 ± 0.0	86.31±0.
			03	17	004	235	98	218
13	1	1	$2.60\pm0.$	85.96±0.	$0.150\pm0.$	57.60 ± 0 .	5.50 ± 0.1	$87.75\pm0.$
			07	38	003	713	95	433

Table 5 shows the results of the RSM performance in terms of all errors as well as in terms of agreement vector values.

Table 5 – Results of the RSM

	$CuSo_4$					FeCl ₃			CuSo ₄ +FeCl ₃			
I Term	βi	Std	t	Prob	βi	Std	t Ratio	Prob	βi	Std	t	Prob
	a. DOC											
0 Interc	3.946	0.12	30.7	4.48	4.05	0.06	58.88	1.06	1.41	0.048	28.97	1.501
1 X_1	-	0.12	-	0.00	1.35	0.06	19.92	2.00	0.41	0.048	8.663	5.460

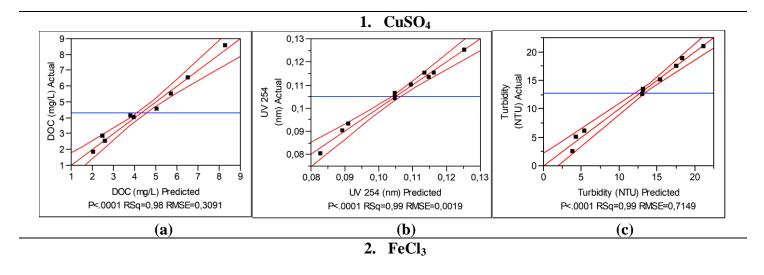
2	X_2	-	0.12	-	0.00	-	0.06	-	4.09	0.06	0.048	1.386	0.208
3	$X_1 * X_2$	0.675	0.15	$4.\hat{3}\hat{7}$	0.00	-	0.08	-	1.23	-	0.058	-	0.000
4	$X_1 * X_1$	0.325	0.18	1.75	0.12	1.79	0.09	17.96	4.08	-	0.070	-	0.003
5	X_2*X_2	$0.\hat{4}85$	0.18	2.61	0.03	0.64	0.09	6.455	0.00	$\hat{1}.\hat{5}\hat{4}$	0.070	21.86	1.057
		^					b.	UV 25	4	^ -	^		- -
0	Interc	0.104	0.00	135.	3.15	0.09	0.00	30.82	9.76	0.11	0.002	53.41	1.373
1	X_1	-	0.00	-	1.84	0.03	0.00	11.26	9.71	0.02	0.002	10.66	2.971
2	X_2	-	0.00	-	0.00	-	0.00	-	5.17	0.00	0.002	0.57	$\hat{0.583}$
3	$X_1 * X_2$	$\hat{0}.\hat{0}\hat{0}\hat{1}$	0.00	1.88	0.10	-	0.00	-	0.00	-	0.002	-	$5.\hat{2}87$
4	$X_1 * X_1$	-	0.00	-	0.00	$\hat{0}.\hat{0}\hat{3}$	0.00	$7.\overline{506}$	0.00	-	0.002	-	4.041
5	X_2*X_2	0.007	0.00	$6.\overline{55}$	0.00	0.01	0.00	2.792	0.02	$\hat{0}.\hat{04}$	0.001	15.59	3.864
		^			^ ~	• •	с.	Turbid	itv		• ^	^^	- ^
0	Interc	13.15	0.29	44.3	7.77	9.53	0.07	133.3	3.51	3.19	0.080	39.97	1.595
1	X_1		0.29	-	4.29	4.47	0.07	63.68	6.18	0.83	0.078	10.59	1.463
2	X_1		0.29	• · · ·	0.00	-	0.07		1.61	0.24	0.078	3.155	$0.01\overline{6}$
_	$X_1 * X_2$	$\hat{0}.\hat{6}\hat{0}\tilde{2}$	0.35	1.69	0.13	- ·-	0.08		1.59	-	0.096	-	1.069
	$X_1 * X_1$	-	0.43	-	0.00	$\dot{5}.\overline{0}\hat{8}$	0.10	49.03	3.83		0.115	-	0.007
		1.049	0.43	2.44	0.04	0.19	0.10	1.840	0.10	2.79	_	24.06	5.440

 $\textbf{Table 6} - RSM \ performances \ For \ CuSO_4, \ FeCl_3 \ and \ CuSO_4 + FeCl_3 \ coagulant.$

Responses	Final equation in terms of code of independent variables	P	F	\mathbb{R}^2	R ² adj	RMS E	D F
DOC	$CuSO_4$ $Y = 3.9468 - 2.2366X_1 - 0.6166X_2 + 0.675X_1X_2$ $+0.4858X_2^2$	6.453 e-6	74.5665	0.981	0.968	0.3090	5
UV 254	$Y = 0.1047 - 0.0153X_1 - 0.0026X_2 - 0.0066X_1^2 + 0.0073X_2^2$	2.729 1 e- 6	95.8741	0.985 6	0.975	0.0018	5
Turbidit y	$Y = 13.1558 - 7.2683X_1 - 1.205X_2 - 2.0905X_1^2 + 1.0494X_2^2$	8.886 5 e- 7	132.8157	0.989 5	0.982	0.7149	5
DOC	FeCl₃ $Y = 4.0586 + 1.35X_1 - 2.3666X_2 - 1.775X_1X_2$ $+1.7948X_1^2 + 0.6448X_2^2$	7.724 8 e- 9	520.6865	0.997	0.995 4	0.1660	5
UV 254	$Y = 0.0992 + 0.0356X_1 - 0.0276X_2 - 0.0257X1X$ $+0.0350X_1^2 + 0.0130X_2^2$	28.750 5 e- 6	68.1960	0.979 8	0.965 5	0.0077	5
Turbidit y	$Y = 9.5355 + 4.4766X_1 - 5.4266X_2 - 4.7875X_1X_1 + 5.0806X_1^2$	1.355 2 3 e- 11	3200.855	0.999 5	0.999	0.1721	5

The values of the coefficients of determination have decreased slightly, but the equation has become simpler after removing the low explanatory power variables. These coefficients mean that the correlations of the model were moderately positive (Figure 3). The probability was strictly below 0.5%, confirming that the model was significant (Tahraoui et al., 2021b).

The values of the significance level P and the values of the F Ratio, which provide a measure of the statistical significance of the regression models, have also been determined (Tahraoui et al., 2021b). A high value of F combined with a low value of P indicates that the equation is statistically significant (Tahraoui et al., 2021b).



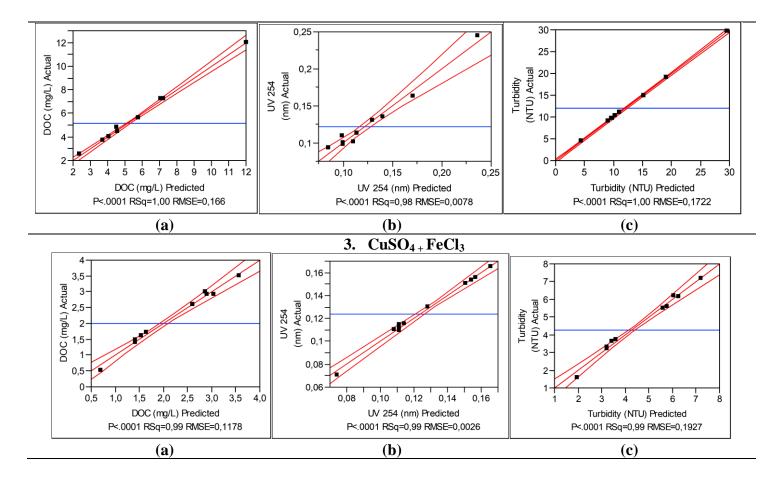


Figure 3 – Relation between the observed dose of coagulant and those estimated by the RSM model For CuSO₄, FeCl₃ and CuSO₄+FeCl₃ coagulants: (a) DOC, (b) UV 254, (c) Turbidity.

The proposed models can simultaneously infer the effect of the predictors (X_I and X_2), their interactions, and their quadratic effects. Table 5 shows the effects of independent factors, their interactions, and their quadratic terms on DOC, UV 254, and turbidity on the phenomenon of flexing coagulant. Indeed, the coefficients of each factor in the model make it possible to evaluate the impact of each factor on the answer (Lefnaoui and Moulai-Mostefa, 2014).

3.1.1. Effect of parameters on DOC

The pH factor is found to affect DOC positively under $FeCl_3$ and the mixture (CuSO₄ + $FeCl_3$) as shown in Table 5.a. On the other hand, it has a negative effect on DOC under

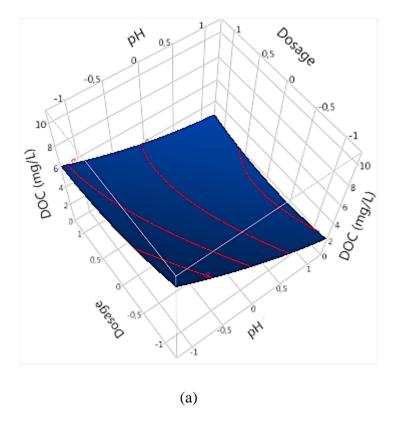
CuSO₄, i.e. each time the pH increases, the concentration of DOC decreases, as ahown in Table 5.a.

The dosage factor shows a negative effect on DOC under CuSO₄ and FeCl₃ as shown in Table 5.a. On the other hand, its effect on DOC is negligible under the mixture because it is an insignificant parameter as shown in Table 5.a.

On the other hand, Table 5.a shows that the interaction of pH and dosage has a positive effect on DOC under $CuSO_4$ and a negative effect on DOC under $FeCl_3$ and under the mixture ($CuSO_4 + FeCl_3$).

The pH has a negative quadratic effect on DOC in the presence of the mixture and a positive quadratic effect in the presence of FeCl₃ as shown in Table 5.a. However, its quadratic effect in the presence of CuSO₄ is considered negligible because prob>5% as shown in Table 4.a and Figure 5.a.

On the other hand, the dosage has positive quadratic effects in the presence of $CuSO_4$, $FeCl_3$ and the mixture as can be seen from Table 5.a.



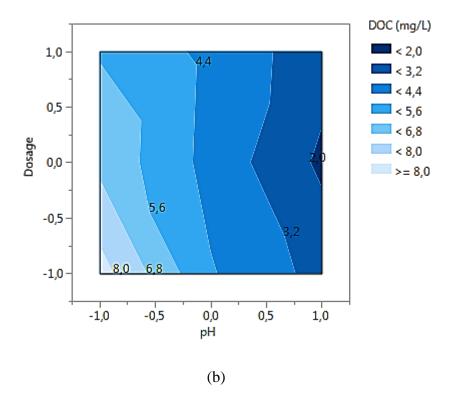
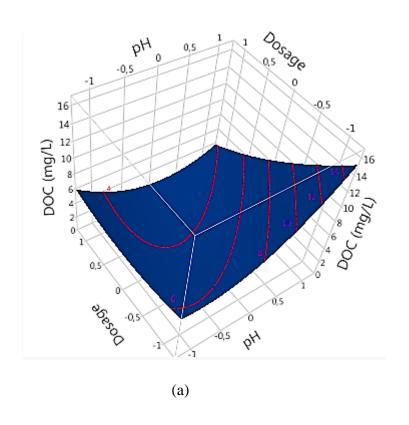


Figure 4 – Effects of operating factors and their interactions on DOC under CuSO₄: (a) interaction of coagulant dosage and pH, (b) contour plot.



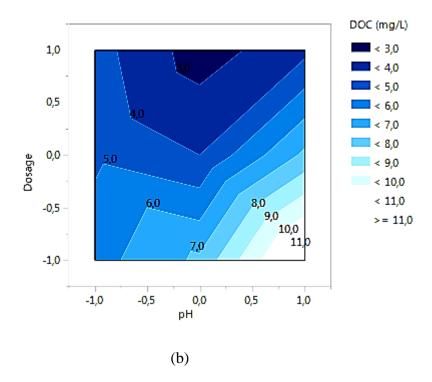
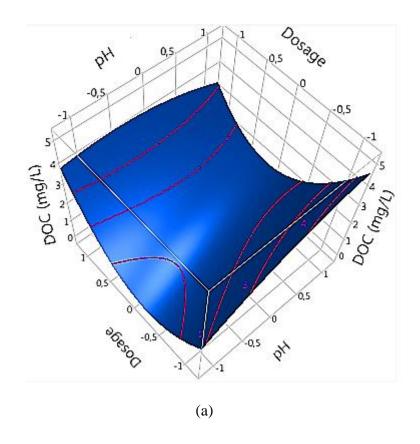


Figure 5 – Effects of operating factors and their interactions on DOC under FeCl₃: (a) interaction of coagulant dosage and pH, (b) contour plot.



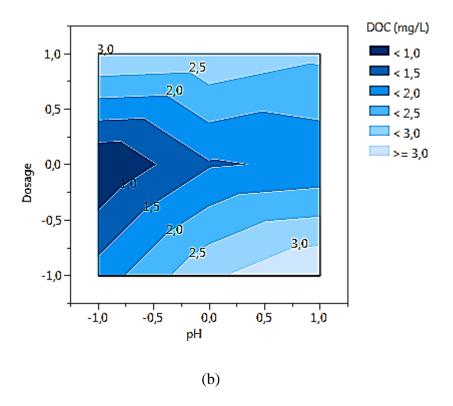


Figure 6 – Effects of operating factors and their interactions on DOC under CuSO₄

₊ FeCl₃: (a) interaction of coagulant dosage and pH, (b) contour plot.

From the obtained results shown in Figure 4.a, it appears that with increase in pH from 7 to 11, the DOC concentration decreases from 8.28 mg/L to 2.46 mg/L in the presence 600 mg/L of CuSO₄. Moreover, the pH has positive and negative quadratic effects on DOC in the presence of FeCl₃ and the mixture as shown in Figures 5 and 6 respectively. The concentration of DOC in the presence 1000 mg/L of FeCl₃ decreases from 4.55 mg/L to 2.33 mg/L for pH changing from 3 to 5 and then increases to 3.7 mg/L for pH=7 as shown in Figure 5.a. On the other hand, the concentration of DOC in the presence 1000 mg/L of the mixture increases from 2.86 to 3.04 mg/L for pH ranging from 5 to 5.875 and then decreases to 2.59 mg/L for pH = 9 as shown in Figure 6.a.

It should also be noted that the dosage of coagulant flocculant has a positive quadratic effect under CuSO₄, FeCl₃, and the mixture. The DOC concentration decreases from 2.43 to 2.01 mg/L for the CuSO₄ dosage changing from 600 to 900 mg/L at pH 11 and then

increases to 2.58 mg/L for the dosage equal to 1200 mg/L as shown in Figure 4.a. The DOC concentration decreases from 5.73 to 4.36 mg/L for the FeCl₃ dosage changing from 200 to 800 g/L at pH 3 and then increases to 4.55 mg/L at the dosage = 1000 mg/L as shown in Figure 5.a.

The DOC concentration decreases from 1.63 to 0. 63 mg/L for the mixture dose changing from 200 to 520 mg/L at pH 5 and then increases to 2.86 mg/L at dosage = 1000 mg/L as shown in Figure 6.a.

Very high DOC removal to 2.01, 2.31 and 0.63 mg/L provides a treatment efficiency equal to 89.13%, 86.48 and 97.27% respectively for $CuSO_4$, $FeCl_3$ and the mixture. The results in Figures 4 to 6 indicate the approximate optimal treatment condition are as follows: the pH must be adjusted and the coagulant-flocculant dosage injected according to $CuSO_4$, $FeCl_3$ and the mixture as follows: pH = 11, dosage = 900 mg/L (Figure 4.b); pH = 5, dosage = 1000 mg/L (Figure 5.b); pH = 5, dosage = 520 mg/L (Figure 6.b)).

From these results, it can be concluded that the coagulant-flocculant mixture ($CuSO_4 + FeCl_3$) removes DOC better than the individual coagulants. However, it should also be noted that the separate coagulants remain very practical according to the results obtained.

3.1.2. Effect of Parameters on UV 254

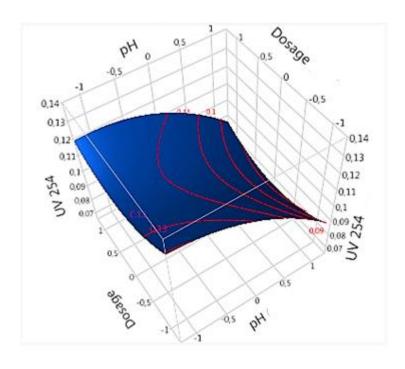
The pH factor is found to have a positive effect on UV 254 when using $FeCl_3$ and the mixture ($CuSO_4 + FeCl_3$) as shown in Table 5.b. On the other hand, it has a negative effect when $CuSO_4$ is used according to Table 5.b.

Moreover, the dosage factor shows a negative effect on UV 254 for CuSO₄ and FeCl₃ as shown in Table 5.b. However, it is considered as an insignificant parameter in presence of the mixture as shown in Table 5.b.

In contrast, the interaction of pH with dosage has a negative effect on UV 254 under $FeCl_3$ and the mixture (CuSO₄ + $FeCl_3$) according to Table 5.b. On the other hand, the effect of

the interaction of pH with the CuSO₄ dosage is negligible because it is an insignificant parameter (Table 5.b and Figure 7.a)

In addition, the results show that pH has a negative quadratic effect on UV 254 in the presence of CuSO₄ and mixture as shown in Figure 7, and a positive quadratic effect in the presence of FeCl₃ according to Figures 8 and 9. On the other hand, the dosages of CuSO₄, FeCl₃ and of the mixture shown positive quadratic effects as can be seen from Figures 7, 8 and 9.



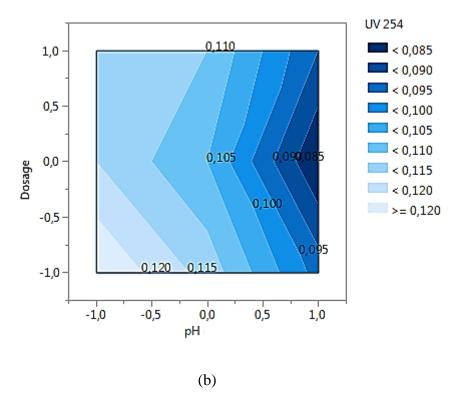
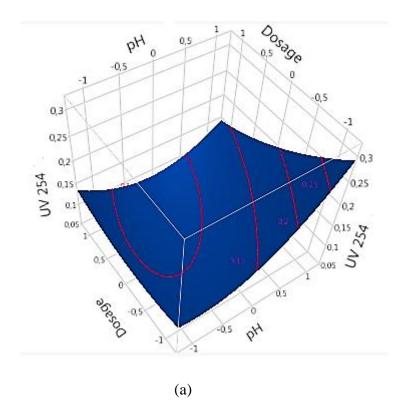


Figure 7 – Effects of operating factors and their interactions on UV 254 for CuSO₄:

(a) interaction of coagulant dosage and pH, (b) contour plot.



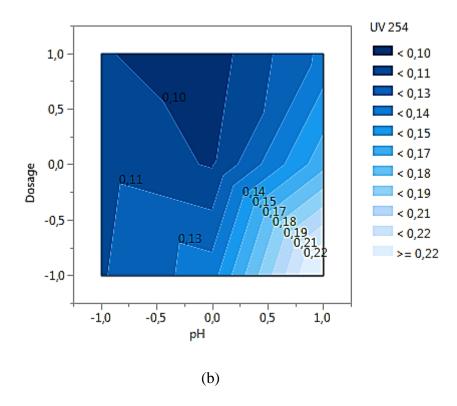
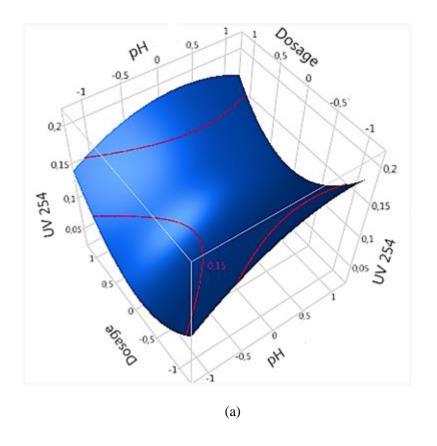


Figure 8 – Effects of operating factors and their interactions on UV 254 for FeCl₃: (a) interaction of coagulant dosage and pH, (b) contour plot.



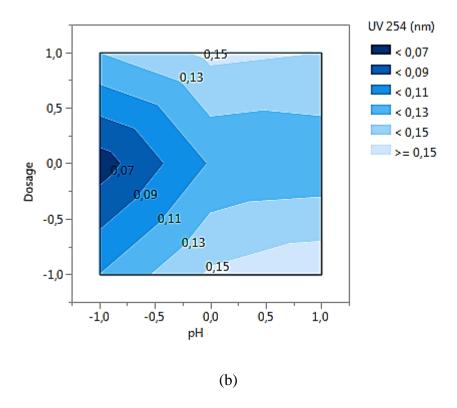


Figure 9 – Effects of operating factors and their interactions on UV 254 for CuSO₄₊ FeCl₃: (a) interaction of coagulant dosage and pH, (b) contour plot.

Given the results obtained, it appears that the pH has a negative quadratic effect on UV 254 in the presence of $CuSO_4$ and the mixture, and a positive quadratic effect for $FeCl_3$. Indeed, the value of UV 254 in the presence 600 mg/L of $CuSO_4$ for a pH lower than 7 shows a positive effect, and for a pH change from 7 to 11, the value of UV 254 decreases from 0.125 to 0.091 (Figure 7. a). On the other hand, in the presence 1000 mg/L of $FeCl_3$, the value of UV 254 goes from 0.109 to 0.084 for a pH going from 3 to 5 then increases to 0.129 at pH = 7 (Figure 8.a). However, the UV value at 254 nm in the presence of 1000 mg/L of the mixture increases from 0.125 to 0.160 when the pH is increased from 5 to 7.64, then decreases again to 0.151 when the pH reaches 9 (Figure 9.a) .

On the other hand, coagulant flocculant dosage also has a positive quadratic effect on UV 254 under $CuSO_4$, $FeCl_3$ and mixture, as UV 254 value decreases from 0.090 to 0.082 for $CuSO_4$ dosage changes from 600 to 900 mg/L at pH = 11, then a slight increase (0.089) for

the dosage = 1200 mg/L (Figure 7.a). For FeCl₃, a slight decrease in the UV 254 value is observed from 0.113 to 0.098 for dosage going from 200 to 600 mg/L at pH = 3; as well as a slight increase up to the value 0.109 for the dosage equal to 1000 mg/L (Figure 8.a).

However, in the presence of mixture, the UV 254 value decreases from 0.105 to 0.068 corresponding to the dosage increases from 200 to 520 mg/L at pH 5; then increases to 0.125 for the dosage = 1000 mg/L (Figure 9.a).

In conclusion, having aromatic nucleus removal rates as UV absorbance at wavelength of 254 nm equal to 0.082, 0.084 and 0.068 or having a treatment efficiency equal to 76.77%, 76.20% and 80.73% using respectively $CuSO_4$, $FeCl_3$ and the mixture, the pH and the coagulant-flocculant dosage must be adjusted as follows: pH = 11, dosage = 900 mg/L (Figure 7.b); pH = 5, dosage = 1000 mg/L (Figure 8.b); pH = 5, dosage = 520 mg/L (Figure 9.b).

From these results, it can be concluded that the coagulant-flocculant mixture (CuSO₄ + FeCl₃) removes the aromatic rings better than the coagulants alone. However, it should also be noted that coagulants alone remain very effective in terms of the results obtained.

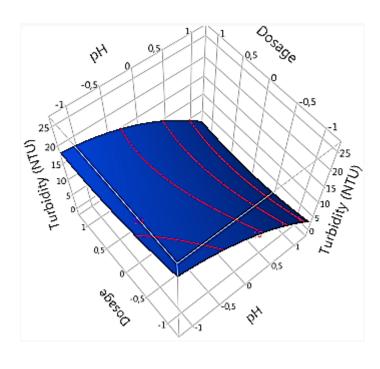
3.1.3. Effect of parameters on turbidity

The pH factor is found to have a positive effect on turbidity when FeCl₃ or the mixture is used as can be seen from Table 5.c. On the other hand, it has a negative effect on turbidity when CuSO₄ is used (Table 5.c). Moreover, the dosage factor shows a negative effect on turbidity when CuSO₄ or FeCl₃ is used and a positive effect when the mixture is used (Table 5.c).

The interaction of pH with dosage has a negative effect on turbidity under FeCl₃ or mixture (Table 5.c). On the other hand, the interaction of pH with the dosage in the presence of CuSO₄ is considered to have a negligible effect on turbidity because it is an insignificant parameter (Table 5.c and Figure 10.a).

Moreover, the results show that the dosage has a positive quadratic effect on the turbidity in the presence of CuSO₄ and the mixture (Table 5.c, Figures 10.a and 12.a), but the quadratic effect of FeCl₃ dosage on turbidity is negligible because it is an insignificant parameter (Table 5.c, Figure 11.a).

However, pH has interesting quadratic effects, indicating negative quadratic effects in the presence of CuSO4 and the mixture (Figures 10 and 12), but a positive effect in the presence of FeCl3 (Figure 11).



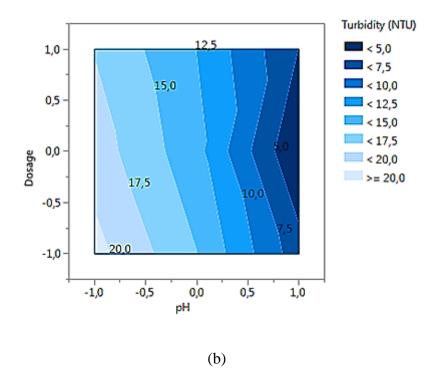
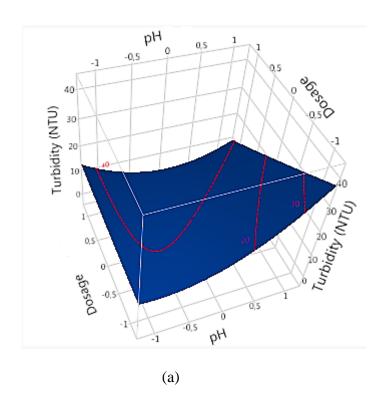


Figure 10 – Effects of operating factors and their interactions on turbidity for $CuSO_4$: (a) interaction of coagulant dosage and pH, (b) contour plot.



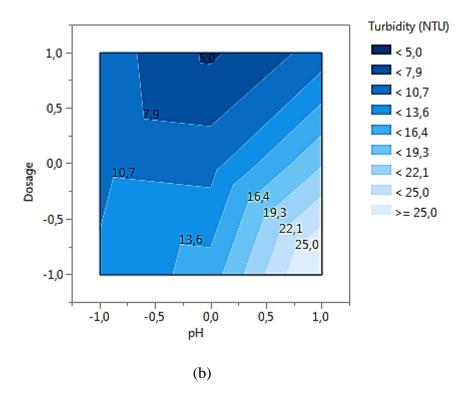
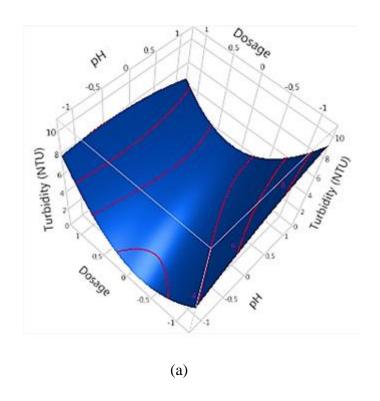


Figure 11 – Effects of operating factors and their interactions on turbidity for $FeCl_3$: (a) interaction of coagulant dosage and pH, (b) contour plot.



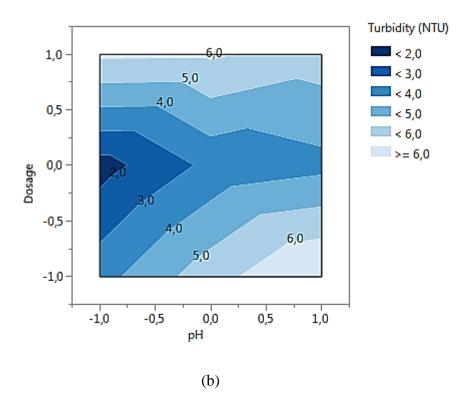


Figure 12 – Effects of operating factors and their interactions on turbidity CuSO₄₊FeCl₃:

(a) interaction of coagulant dosage and pH, (b) contour plot.

Indeed, when pH is less than 7, it has a positive effect on turbidity in the presence 600 mg/L of CuSO₄, but when pH is increased from 7 to 11, turbidity decreases from 21.19 to 5.44 NTU (Figure 10.a).

With 1000 mg/L of FeCl₃, the turbidity decreases from 9.69 to 4.29 NTU when pH changes from 3 to 5 and increases to 9.06 NTU when pH=7 (Figure 11.a). On the other hand, the turbidity in the presence 1000 mg/L of the mixture increases from 6.04 to 6.23 NTU for pH changing from 5 to 7 and then decreases to 5.56 NTU when pH = 9 (Figure 12.a).

It should also be noted that the dosage of coagulant-flocculant has a positive quadratic effect only under CuSO₄ and the mixture (Figures 10, 11 and 12). For CuSO₄, a slight decrease in the turbidity value was observed from 5.32 to 3.57 NTU for a dosage ranging

from 600 to 900 mg/L when pH is at 11; as well as a slight increase up to the value 4.24 NTU for the dosage equal to 1200 mg/L (Figure 9.a). In the presence of the mixture, the turbidity decreases from 3.40 to 1.78 NTU for a dosage changing from 200 to 520 mg/L when pH is at 5, then increases to 6.04 NTU when the dosage is equal to 1000 mg/L (Figure 12.a).

For the dosage of FeCl₃, each time the dosage increases, the turbidity decreases. When increasing the dosage from 200 to 1000 mg/L, the turbidity is reduced from 10.96 to 9.06 NTU for $CuSO_4$ at pH = 3 (Figure 11.a).

For a turbidity removal rate equal to 3.57, 4.10 and 1.78 NTU or on condition of having a treatment efficiency equal to 91.56%, 90.44% and 96.05% in using CuSO₄, FeCl₃ and the mixture respectively, it is necessary to adjust the pH and inject the dosage of coagulant-flocculant according to CuSO₄, FeCl₃ and the mixture as follows: (pH = 11, dosage = 900 mg/L (Figure 10.b); pH = 5, dosage = 1000 mg/L (Figure 11.b); pH = 5, dosage = 520 mg/L (Figure 12.b)).

Therefore, it can be concluded that the coagulant-flocculant mixture ($CuSO_4 + FeCl_3$) removes turbidity better than the single coagulants. However, it should also be noted that the single coagulants remain very effective according to the results obtained.

3.2. GPR Modeling

The main goal of this part is to create more accurate models for the prediction of DOC, UV 254, and turbidity under the three treatments. For this, the databases of the first treatment (CUSO₄), the second treatment (Fecl₃), and the third treatment (CuSo₄+Fecl₃) were combined in a single database. In this database, a variable, "type of coagulation", has been inserted as a third independent parameter representing coagulation type, which was coded as 1, 2 and 3 for CuSo₄, Fecl₃ and CuSo₄+Fecl₃ respectively.

In order to develop more accurate models, the GPR was optimised by the dragonfly (DA) optimization algorithm to find the best appropriate parameters for Kernel Scale (SigmaM and SigmaF) and Sigma.

From the start, the database was split in two: 70% for learning and 30% for validation. the database has been normalized in the interval [-1,1] using the following equation:

$$x_N = 2\left(\frac{x - x_{\min}}{x_{\max} - x_{\min}}\right) - 1 \tag{22}$$

Then, as previously indicated, Ten kernel functions (exponential, exponential squared, mother32, mother52, rational quadratic, exponential ard, exponential squared ard, ardmatern32, ardmatern52 and rational quadratic ard) were optimized with the basis function (constant, linear and zero) for each output, then the one that gives the best performance is selected based on statistical evaluation criteria.

The development method of each GPR model has been designed in detail in Figure 13, including the interval which has been fixed for each parameter of the model and also for the agoriyhme of DA.

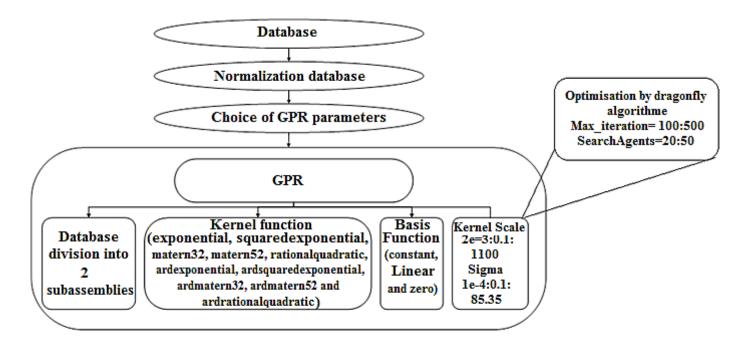


Figure 13– Organization chart for the development and optimization of the GPR architecture

The best models for the three outputs and their parameters (kernel scale and sigma) are presented in Table 7. It shows the coefficients (\mathbf{R} , \mathbf{R}^2 and \mathbf{R}^2_{adj}) and performance measures (\mathbf{RMSE} , \mathbf{MSE} , \mathbf{EPM} , \mathbf{ESP} and \mathbf{MAE}) for the learning and validation data sets, as well as the kernel parameters (Kernel Scale [sigmaM, sigmaF] and sigma. It also shows the best kernel functions with the resulting basis functions, as well as the number of dragonfly agents and the number of iterations that produced the best models. Also, it shows the number of parameters of each best model obtained.

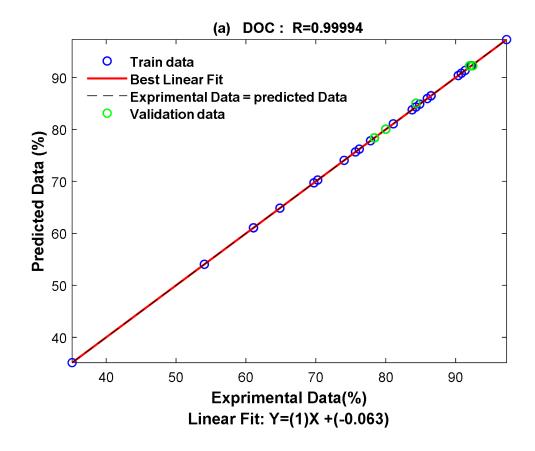
Table 7 – Performances of the different GPR models tested.

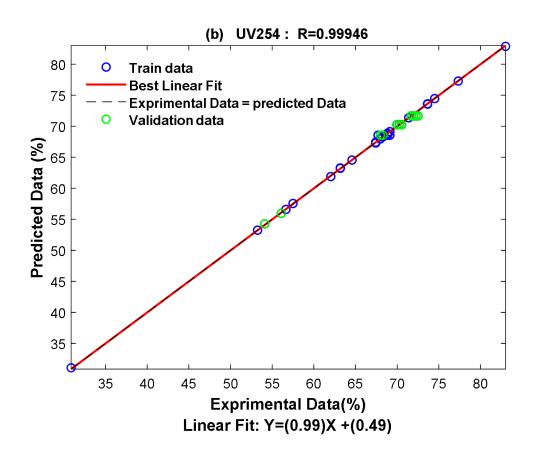
DA	Max_iteration=100 SearchAgents_no=30							
	Kernel	Basis	Kernel Scale	Sig ma	Number of		RMSE/MSE/EPM/ ESP /MAE	
Outp	function	Function	Sigm Sig		parameters	Tra VA AL	Train VAL ALL	

uts			aM	ma			in	L	L			
(%)				\mathbf{F}								
							1.0	0.9		0.0214	0.240	0.129
							000		999	0.0005	7	1
DOC							1.0	0.9		0.0078	0.057	0.016
							000	987		0.0275	9	7
				16.2	0.1		1.0	0.9		0.0067	0.143	
	Ermonont						000	981	999		2	0
	Exponent ial	Constant	2.25									0.162
	lai	Constant	40	636	350	28					6	8
												0.040
											3	2
							0.9	0.9		0.2243		
****				10.0			997	981		0.0503	8	4
UV	Squared			10.8			0.9	0.9		0.1935		0.080
254	• -			919	0.6					0.3363	7	3
	_				0.6		0.9	0.9 947		0.1230	0.449	0.265
			0.87				993	947	988		-	0.422
	Expone	Constant	0.87		558	28					0.384	9
	ntial		Uŏ		330	20					-	0.175
											4	8
							1.0	0.9	1.0	0.0421	0.214	_
							000			0.0018	0.214	1
Turbi				16.8			1.0	0.9		0.02550	-	-
dity			1.97	308			000	997		.0542	8	2
	ARD-		96		0.1		1.0	0.9	0.9	0.0205	0.227	0.082
	ARD-		5.91				000	996	999		6	5
	.		02								0.265	0.151
	Exponent	Constant	0.02		432	28					9	7
	ia		23								0.164	0.061
											8	2

From Table 7 we can see that all the models have very high statistical coefficients (all are almost 1) and also very low statistical errors (close to zero) on the training and validation data and also on all data. This shows the efficiency of the developed models.

Figure 14 presents the plots of model predictions against the experimental values of the models shown in Table 7.





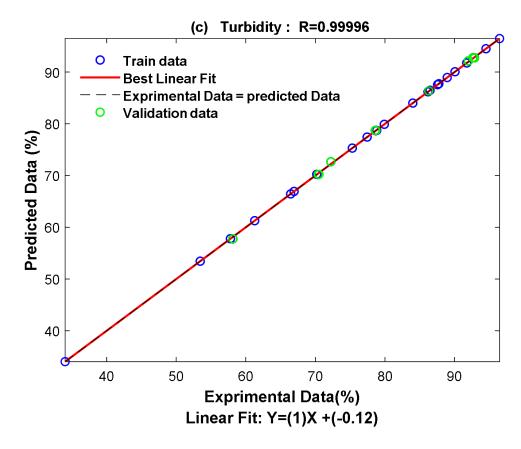
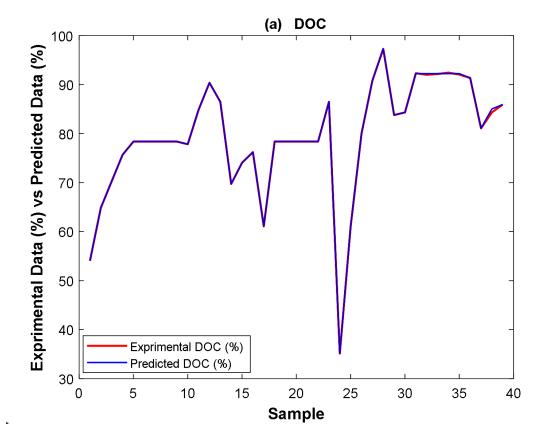
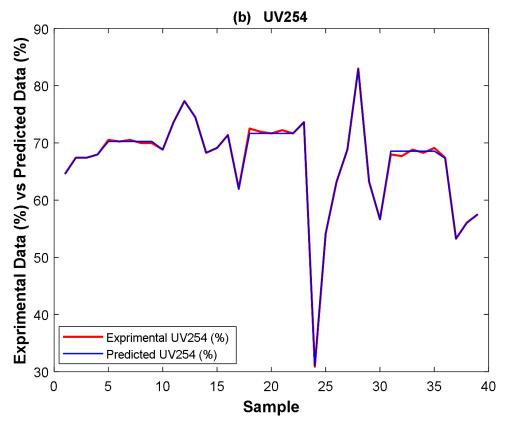


Figure 14 – Relationship between the experimental and the GPR-DA model predicted values: (a) DOC, (b) UV 254, and (c) Turbidity.

Figure 14 shows the excellent predictive capability of the developed GPR-DA models.

Figure 15 plots the experimental data and model predictions at different samples. It can be seen that the model predictions are very close to the corresponding experimental values.





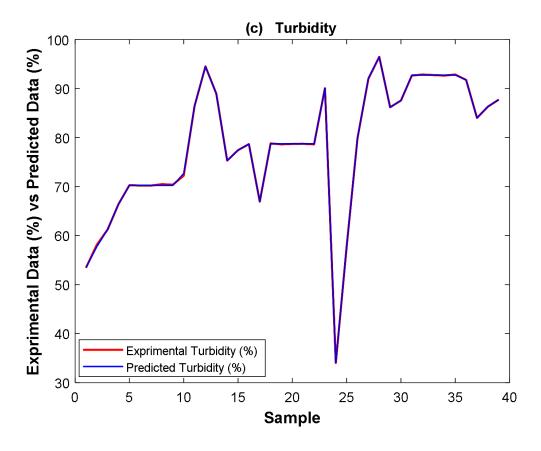


Figure 15 – experimental data and the predicted data of samples using GPR-DA model: (a) DOC, (b) UV 254, and (c) Turbidity.

3.3. Multi-objective optimization

The main objective of this optimization is to find the optimum wast water treatment condition, pH and dosage, in order to achieve the best treatment performance. For this, a MOO was used to solve this optimization problem. In this study, weightings are assigned to the 3 objectives and their weighted sum is used as a single objective which is then solved using the fmincon function available in the MATLAB Optimization Toolbox, the dragonfly algorithm (DA in MATLAB code), and the particle swarm algorithm (PSO in MATLAB code) available in the MATLAB Optimization Toolbox. For this, the maximum DOC removal rate (J₁), the maximum UV 254 removal rate (J₂) and the maximum turbidity removal rate (J₃) were considered as objective functions. They are then combined into a scalar goal as follows:

$$J = w_1 J_1 + w_2 J_2 + w_3 J_3$$

(23)

Where w_1 , w_2 and w_3 are weighting factors, which can be calculated using the rank sum method (Dobrosz-Gómez et al., 2020; Einhorn and McCoach, 1977; Masouleh et al., 2022).

In this study, w_1 , w_2 and w_3 are set to 0.34, 0.3 and 0.36 respectively. The resulting solution values of this optimization problem are given in Table 8.

Table 8 – Results of treatment efficiency (J) under optimal conditions based on DOC (J_1), UV254 (J_2) and turbidity (J_3).

\mathbf{J}_1	J_2	J_3	J
(%)	(%)	(%)	(%)
	CuSO ₄		

- Fmincon_RSM, DA_RSM, PSO_RSM: CuSO₄ (dosage = 968.77 mg/L, pH = 11)
 - Fmincon_GPR, DA_GPR, PSO_GPR: CuSO₄ (dosage = 900 mg/L, pH = 11)

RSM	90.54	76.60	92.03	86.90
GPR	90.36	77.29	94.51	87.94
		$FeCl_3$		

- Fmincon_RSM, DA_RSM, PSO_RSM: FeCl₃ (dosage = 1000 mg/L, pH = 5.01)
- Fmincon_GPR, DA_GPR, PSO_GPR: FeCl₃ (dosage = 1000 mg/L, pH = 5) RSM 87.38 76.00 90.85 85.22 GPR 86.47 73.57 90.06 83.92 CuSO₄ + FeCl₃
- Fmincon_RSM, DA_RSM, PSO_RSM: $CuSO_4 + FeCl_3 (dosage = 537.94 \text{ mg/L}, pH = 5)$
- Fmincon_GPR, DA_GPR, PSO_GPR: CuSO₄ + FeCl₃ (dosage = 600 mg/L, pH = 5)
 RSM 96.47 80.54 96.00 91.52
 GPR 97.28 82.88 96.47 92.71

The three MOO methods used (fmincon, DA and PSO) gave very similar optimization results for each treatment (CuSo₄, Fecl₃ and CuSo₄+Fecl₃) and each model (RSM and GPR). This shows the reliability of the models and the quality of the experimental results

obtained. Equally, this also shows the superiority of the mixed treatment compared to the two separate coagulants.

On the other hand, by comparing the optimization results from using the RSM model and the GPR model, it can be seen that they are very close. Identical results are obtained for the 2nd treatment (using FeCl₃). For the other two treatments, identical pH condition is obtained for optimization with both models, but there are some difference in the optimal dosage obtained. However, these slight difference between the optimal doses of coagulant did not influence the efficacy of the treatment with regard to the results obtained by the MOO methods (fmincon, DA and PSO) (Table 8). Returning to the interpretation of the results of the RSM, it can be seen that the coagulant dosage of the 1st treatment (CuSo₄) and of the 3rd treatment (CuSo₄+Fecl₃) have quadratic effects. Where, the intervals of the quadratic effects have approximately what was obtained by RSM and GPR, which confirms the proximity of the results. To confirm these results, an experimental validation method was applied.

3.4. Validation of the optimum conditions

In order to validate the optimization results, experiments were performed under the obtained optimal conditions to test the effectiveness of MOO based on RSM and GPR. These results have been presented in Table 9 in order to compare them to the predicted values and to express the error between them.

Where :
$$Error = Experimental \ response - Predicted \ response$$
 (24)

Table 9 – Comparison between actual and predicted response at optimum condition.

	Responses								
	J_1	J_1	J_2	J_2	J_3	J_3	J		
(m	ng/L)	(%)		(%)	(NTU)	(%)	(%)		

RSM: CuSO ₄ (dosage = 968.77 mg/L, pH = 11)										
	C	GPR: CuSO	4 (dosage =	900 mg/L,	pH = 11)					
Experimental (RSM)	1.81±0.0 80	90.20±0. 440	0.079±0.	77.71±0.	2.87±0.0	93.61±0.	87.60±0.			
Predicted response	1.74	90.54	0.082	76.60	3.57	92.03	86.90			
Error_Exp_R	0.07 ± 0.0	0.34 ± 0.4	$0.003\pm0.$	1.110±0.	0.7 ± 0.03	1.85 ± 0.0	0.70 ± 0.3			
Experimental (GPR)	1.78±0.1	90.38±0.	0.080±0.	77.71±0.	2.46±0.0	94.52±0.	88.06±0.			
Predicted response	1.78	90.36	0.080	77.29	2.46	94.51	87.94			
Error_Exp_	0.00 ± 0.1	0.02±0.59	0.00 ± 0.00	0.42 ± 0.32	0.00 ± 0.0	0.01 ± 0.10	0.12 ± 0.16			
~~~		- '	dosage = 1	_		)				
	GPR: $FeCl_3$ ( dosage = 1000 mg/L, pH = 5)									
Experimental (RSM/GPR)	2.50±0.1	86.49±0.	0.093±0.	73.56±0.	4.46±0.0	90.07±0.	83.92±0.			
Predicted response	2.33	87.38	0.084	76.00	4.10	90.85	85.22			
Error_Exp_R	$0.12\pm0.1$	$0.39\pm0.9$	$0.009\pm0.$	$2.44\pm0.4$	$0.36 \pm 0.0$	$0.78\pm0.0$	$1.32\pm409$			
Predicted response	2.50	86.47	0.093	73.57	4.46	90.06	83.92			
Error_Exp_	$0.00\pm0.1$	$0.02\pm0.92$	$0.00\pm0.00$	$0.01\pm0.99$	$0.00\pm0.0$	$0.01 \pm 0.09$	$0.00\pm0.40$			
~~~			eCl ₃ (dosageCl ₃	-	_		-			
Experimental (RSM)	0.55±0.0 3	97.01±0.	0.062 ± 0.003	82.43±0. 925	1.69±0.0 45	96.24±0. 001	92.35±0. 355			
Predicted response	0.65	96.47	0.068	80.54	1.79	96.00	91.52			
Error_Exp_R	0.1±0.03	0.54 ± 0.1	$0.006\pm0.$	1.89±0.9	0.1±0.04	0.24 ± 0.0	0.83 ± 0.3			
Experimental (GPR)	0.50±0.0	97.32±0.	$0.060\pm0.$	82.90±0.	1.58±0.0	96.47±0.	92.68±0.			
Predicted response	0.50	97.28	0.060	82.88	1.58	96.47	92.71			
Error_Exp_	0.00±0.0	0.04±0.14	0.00±0.00	0.02±0.43	0.00±0.0	0.00±0.00	0.03±0.39			

From Table 9, it was found that in each case the error was less than 3%. The DOC removal rate error in the three treatments is not higher than 0.54% for the two models (RSM and GPR). The maximum error of DOC elimination rate in the three treatments is 2.44% for

the two models (RSM and GPR). The error of turbidity elimination rate in the two models (RSM and GPR) based on the three treatments is not higher than 1.85%. Finally, the maximum error of the processing efficiency according to the two models used (RSM and GPR) is 1.32%.

From these experimental validation results, it can be seen that the results obtained by the two models are almost equal. This clearly reveals that the two models (RSM and GPR) developed for coagulation and flocculation are in good agreement with the experimental results, with a slight superiority of GPR model to the quadratic RSM model.

As the results are almost identical but with less errors for the GPR model, it is preferable to use the optimal conditions obtained by the GPR model. On the other hand, if one wants to reduce the operating costs, it is preferable to use the RSM model as a model which carries only five parameters at most compared to the GPR model which carries 28 parameters.

3.5. Interface for optimization and predictiont

To provide a simple way to implement MOO and to predict DOC, UV254, turbidity and treatment efficiency, an interface was designed using the MATLAB guide for the purpose of optimization and prediction (Figure 16). This interface to a tool has been converted into an exuctable application on windwos. This powerful application for direct use to predict outputs by selecting desirable pH values, coagulant dosage and even type of coagulation by RSM and also by GPR. On the other hand, the application will also make it possible to find a MOO solution to find the optimum pH and dosage for each type of coagulant according to the two models (RSM and GPR) by three methods (fmincon, DA and PSO).

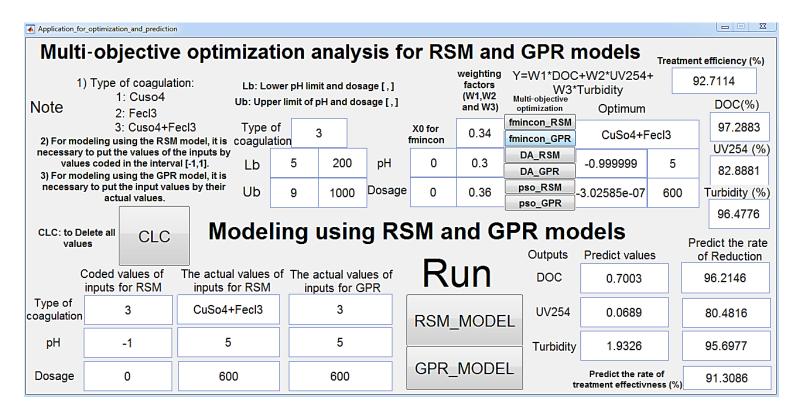


Figure 16 – Application à base une Interface de MATLAB for MOO (fmincon, DA and PSO), and predictiont DOC, UV 254, Turbidity and Treatment efficiency using RSM and GPR.

4. Conclusions

Coagulation and flocculation have been used to remove toxins from wastewater from the Pharmaceutical Effluent Treatment Plant. Coagulants copper sulfate, ferric chloride and a combination of cupric sulfate and ferric chloride in a ratio of 1:1 (CuSO4 + FeCl3) are used for the treatment process. The composite design approach has been used to improve independent process parameters such as pH and coagulant dosage in conjunction with DOC reduction, UV 254 and turbidity as dependent variables. On the other hand, the Gaussian process regression coupled with dragonfly algorithm is also used to model the three outputs, after combining the RSM databases of each coagulant in a single database. The evaluation of the best developed model is carried out by the experimental validation of optimal conditions of each treatment. For this, the MOO technique is used to determine the optimal conditions.

The experimental validations of the optimal treatment condition show the superiority of the GPR_DA model compared to RSM. The error obtained by experimental validation of the GPR_DA model is much lower than that obtained by the quadratic model of RSM. In addition, the results obtained show that the combined coagulant (CuSO4 + FeCl3) outperforms a single coagulant, CuSO4 or FeCl3. The efficiency measures of pretreatment with CuSO4, FeCl3, and (CuSO4 + FeCl3) are 88.06% (at dosage of 900 mg/L and pH of 11), 83.92% (at dosage of 1000 mg/L /L and pH of 5), and 92.68% (at a dosage of 600 mg/L and pH of 5), respectively.

From the results obtained, it can be concluded that:

- Pretreatment by coagulation and flocculation of pharmaceutical effluents is well suited to mixed coagulant (CuSO4 + FeCl3).
- The GPR-DA models show great efficiency and profermence in predicting COD,
 UV254 and turbidity reduction rates, especially they can predict the reduction rates
 of the dependent parameters for the three treatments at the same time.

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☐ The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:	