

The 4GFOR model – Coupling 4G early diagenesis and benthic foraminiferal ecology

Frans Jorissen, Stephen Meyers, Boris Kelly-Gerreyn, Louison Huchet, Aurélia

Mouret, Pierre Anschutz

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1	The 4GFOR Model – coupling 4G Early Diagenesis and Benthic
2	Foraminiferal Ecology
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4	Frans J. Jorissen ^{1*} . Stephen R. Meyers ² . Boris A. Kelly-Gerreyn ³ . Louison
5	Huchet ¹ Aurélia Mouret ¹ and Pierre Anschutz ⁴
5	
6	
7	¹ UMR CNRS 6112 LPG-BIAF. Angers University. France
8	² University of Wisconsin, Department of Geoscience, Madison, USA
9	³ Mount Waverley, Melbourne, Victoria 3149, Australia
10	⁴ Univ. Bordeaux, CNRS, EPOC, EPHE, UMR 5805, F-33600 Pessac, France
11	
12	*Corresponding author: Frans J. Jorissen, UMR CNRS 6112 LPG-BIAF, Université d'Angers, 2,
13	Boulevard Lavoisier, 49045 Angers Cedex, France. Email address : <u>frans.jorissen@univ-angers.fr</u>
14	
15	Author Contributions
16 17	Elli concentualization, data management, model tecting, writing, SPM: model development, writing
18	BAKG: model development writing 1H: model testing writing AM: early diagenesis modeling
19	writing. PA: early diagenesis modeling, data management, writing.
20	
21	Abstract
22	
22	with four theoretical species of heathic foraminifere. The spechamical part of the model cimulates
25	the succession of early diagonatic redev reactions in the superficial codiment layers. The four
24 25	the succession of early diagenetic redux reactions in the superficial sediment layers. The four
25	microhabitate differ by their limitations with respect to oxygen and nitrate concentration and their
20	ability to feed on different organic matter types. The foraminiferal model is a transformation model
28	in which different proportions of four types of organic matter with different reactivity are attributed
29	to each of the theoretical species. The model has been manually tuned and tested on a large data set
30	from two stations, at 1000 and 550 m depth, in the Bay of Biscay. The model adequately simulates
31	the vertical distribution of shallow, intermediate and deep infaunal species. The 4GFOR model can be
32	used to test several hypotheses concerning foraminiferal ecology. The model has already allowed us
33	to conclude that it is high unlikely that shallow infaunal taxa feed exclusively on the most labile part
34	of the organic matter flux, since it was unable to reproduce the field observations while using this
35	hypothesis. Another important conclusion based on our preliminary model results is that the absence
36	of deep infaunal taxa close to the sediment surface may be explained by their decreasing
37	competitiveness in well oxygenated environments. In the future, the inversion of the 4GFOR model
38	may yield a powerful paleoceanographic tool, enabling us to reconstruct major limiting
39	environmental factors (oxygen and nitrate concentrations; quantity and quality of the organic flux to
40	the ocean floor) from benthic foraminiferal assemblage data.

41 Keywords

42 Benthic foraminifera, ecology, microhabitats, early diagenesis, model.

43 1. Introduction

Since the pioneering work of Corliss and his colleagues in the 1980's (Corliss, 1985; Corliss & Chen 44 45 1988; Corliss and Emerson, 1990) - which has been fully confirmed by a number of subsequent 46 studies (i.e., Gooday, 1986; Mackensen and Douglas, 1989; McCorkle et al. 1990; Rathburn & Corliss, 47 1994; Jorissen et al., 1998) - we know that in open marine settings, there is a vertical succession of 48 benthic foraminiferal species in the surface sediment. This species succession, which appears to be 49 closely related with the well-known succession of redox layers in the surface sediment (e.g., Froelich 50 et al., 1979, Canfield et al., 1993) may be expanded over several centimetres (up to ~10 cm at most) 51 or be compressed in the first few millimetres of the sediment. Despite of this vertical variability, the 52 succession of foraminiferal species is always the same (Jorissen, 1999a). Consequently, foraminiferal 53 species may be considered as shallow, intermediate or deep infaunal, not so much on the basis of the 54 exact depth at which they are found in the sediment column, which can vary considerably, but rather 55 on their relative position in the vertical species succession (Jorissen et al., 2007).

56 Following earlier suggestions of Shirayama (1984), Corliss and Emerson (1990) and Loubere et al. 57 (1993), Jorissen et al. (1995) proposed the so-called TROX-model, which hypothesized that the 58 vertical penetration depth of foraminiferal species into the sediment is determined by food 59 availability in oligotrophic ecosystems, but would be limited by redox conditions (especially by pore 60 water oxygen concentration and by the presence of toxic dissolved sulphides) in eutrophic settings. 61 Different requirements with respect to these limiting factors would then explain the specific 62 microhabitats of individual species. Indirectly, the TROX model also explains biodiversity trends, 63 which are strongly affected by the availability or not of ecological niches deeper in the sediment. This conceptual model has since been confirmed by a large number of observational studies, and it has 64 65 also been proposed to explain macrofaunal distribution patterns (Carney, 2005). 66 Since 1995, several authors have tried to further elaborate upon the TROX model, by integrating 67 bioturbation (Van der Zwaan et al., 1999), or by coupling the foraminiferal distribution with several 68 redox species (Koho et al., 2015), and not only with oxygen, as in the original model. In the 25 years 69 since the publication of the TROX model, we have also learned a lot about benthic foraminiferal 70 ecology. After the ground-breaking publications of Risgaard-Petersen et al. (2006), Høgslund et al.

71 (2008), Pina-Ochoa et al. (2010) and Bernhard et al. (2012), we know that many species are capable 72 of storing nitrate in their protoplasm, and at least some of these species are capable of respiring 73 nitrate when confronted with anoxia. For other taxa, a capability to sequester chloroplasts from 74 consumed diatoms has been shown (e.g., Lee and Anderson, 1991; Bernhard and Bowser, 1999). The 75 foraminifera are able to keep these intracellular chloroplasts active for several weeks, showing 76 another foraminiferal strategy for survival under strongly adverse conditions (Jauffrais et al., 2016). 77 And finally, a last group of species may survive adverse conditions by entering into a state of 78 dormancy, with a strongly diminished metabolic rate (Bernhard and Alve, 1997; Ross and Hallock, 79 2016).

80 The presence of an alternative metabolic pathway, in facultatively anaerobic taxa, as well as the 81 other metabolic adaptations, explains why most benthic foraminiferal species are tolerant to strong 82 hypoxia (at least down to ~0.5 ml/l, or 22 μ mol/l; Jorissen et al., 2007). This also explains how 83 foraminifera can survive, and for some species at least, remain active in anoxic conditions. In this 84 context, several observational studies have shown foraminiferal survival after exposure to anoxia together with free sulphides (e.g., Langlet et al., 2013; 2014, Richirt et al., 2020), a combination 85 86 which until recently was generally considered lethal (Moodley et al., 1998). The ultimate 87 disappearance of benthic foraminifera during cases of long-term anoxia, as is for instance observed 88 in Mediterranean sapropels (e.g., Jorissen, 1999b), seems therefore not to be the consequence of

strongly increased mortality, but rather of an inhibition of reproduction under strongly adverseconditions (Richirt et al., 2020).

91 These recent advances suggest that oxygen may not be the foremost environmental parameter

92 explaining foraminiferal ecology, as was often thought in the past (e.g., Bernhard, 1886; Alve and

93 Bernhard, 1995), and was more or less hypothesized in the TROX model. Instead, it appears that it is 94 rather the quantity, and especially the quality of the food particles, which are the predominant

rather the quantity, and especially the quality of the food particles, which are the predominant
 parameters. However, the quality of the organic matter (C_{org}) flux will also affect the biological

96 oxygen demand, and thereby the bottom water and sediment redox conditions, so that the

97 combination of the organic matter flux and oxygen content forms an inextricable complex.

98 Despite the determinant role of the organic matter flux, oxygen will still be a principal controlling

99 factor when values are close to (or below) the tolerance levels of many species and/or higher

100 taxonomical categories. Although alternative metabolic pathways for organic matter decomposition

101 can be utilized under suboxic and anoxic conditions, they are energetically much less favourable,

102 with the exception of denitrification (Froelich et al., 1979; Middelburg, 2019).

109

Furthermore, we have indications that the presence of free sulphides is not directly lethal (Bernhard et al., 1993; Langlet et al., 2014), but that long-term exposure leads to a decrease of foraminiferal population densities (Moodley et al., 1998; Richirt et al., 2020). It has been suggested that this would be due to a decrease of metabolic rate, and a loss of vital functions, such as growth, and especially reproduction (Moodley et al., 1998; Richirt et al., 2020). Long-term exposure to dissolved sulphides should therefore ultimately lead to the disappearance of foraminifera.

Summarising, even if our ideas about the specific role of some environmental factors, such as oxygen
 availability, have become more nuanced, the importance of redox conditions within the foraminiferal
 microhabitat remains paramount, and consequently, the conceptual TROX model (Jorissen et al.,
 1995) still provides an excellent tool to understand foraminiferal distribution patterns.

113 1335) still provides all excellent toor to understand foraninineral distribution patients.

In this paper, we present a first attempt to translate the conceptual TROX-model into a numerical
 model, combining sediment early diagenesis (the combination of biological, chemical, and physical
 processes which change the quantity and composition of the organic matter in marine sediments
 before they become consolidated; Henrichs, 1992) and foraminiferal ecology.

119 The main aim of such a model is that it will allow us to test hypotheses concerning foraminiferal 120 ecology. We will define theoretical species, with distinct characteristics, that each represent a group

121 of real species with a similar microhabitat. We will then see how these theoretical species behave

122 (are distributed) in the vertical succession of redox zones reproduced by the model, and compare the

model results with observational data. If the model is capable of successfully reconstructing the field

observations, this means that the underlying hypotheses concerning the foraminiferal ecology may
 be realistic. If this is not possible, the way the foraminiferal species are defined is probably incorrect.

126 Ultimately, the aim is to use this model as a tool to test paleoceanographical scenarios and to

127 reconstruct environmental parameters from foraminiferal successions in fossil records.

128 For the construction of the model, we profit from an extremely favourable context. First, early

129 diagenetic models have become widely available. We decided to use the OMEXDIA model of Soetaert

130 et al. (1996), which is well established, very well documented (Soetaert and Hermann, 2009) and

131 freely available on the Comprehensive R Archive Network (https://cran.r-project.org). The OMEXDIA

132 model will be coupled to a new foraminiferal model developed in the present study. Next, we employ

a very large mixed database of sediment geochemistry (Anschutz et al., 1999, 2005; Hyacinthe et al.,

134 2001, Chaillou et al., 2002, 2003, 2006; Mouret et al., 2009, 2010; Schmidt et al., 2009) and

135 foraminiferal distribution (Fontanier et al., 2002, 2003, 2005, 2006a; Langezaal et al., 2006; Mojtahid

et al., 2010) for the Bay of Biscay. We decided to use the data collected between 1997 and 2001,

137 when a bathymetrical transect from 150 to 2000 m water depth was sampled 10 times. This data set

138 gives ample insight into seasonal and interannual variability of sediment geochemistry and

139 foraminiferal communities. We will use this very large dataset to calibrate the model, and to test it in

140 two different ecological settings.

141 This paper presents the first results of this exercise, in which the numerical model is developed, and

142 is manually tested and tuned at two sampling stations, at 1000 and 550 m depth in the Bay of Biscay.

143 In the process of developing the model, we sequentially tested a range of hypotheses regarding the

benthic environment and its link to foraminiferal ecology. To illustrate the value of quantitative models for such hypothesis testing, we present this study in a manner that emphasizes the

sequential steps in model development and calibration, highlighting the lessons learned along the

147 way.

148 2. Model characteristics

149 When we set out to develop the coupled early diagenesis – benthic foraminiferal ecology model

150 presented here, we chose to implement the model in the software R (R Core Team, 2020). R is widely

151 used for scientific programming, and is freely available (https://www.r-project.org). We decided to

use the early diagenetic model (1D) of Soetaert, which has been described in a number of

publications (Soetaert et al., 1996; Soetaert & Herman, 2009), and is freely available online in the R-

154 package 'ReacTran' (Soetaert and Meysman, 2012; http://

155 www.rforscience.com/modelling/omexdia/.

156 The next step was to introduce theoretical species of benthic foraminifera, with different ecological 157 characteristics (each theoretical species represents a group of real species), into this 1D geochemical 158 environment. At a first stage, we investigated the possibility of constructing a quantitative biological model, including the most important life functions (e.g., growth, reproduction, mortality, etc.) of the 159 different species. However, it soon turned out that this approach was severely hampered by a lack of 160 161 observational data for most of these parameters. For this reason, we decided to change our strategy, 162 and instead developed a "transformation model", in which a fraction of the available resources was 163 allocated to different members of the benthic foraminiferal community. More precisely, we assigned 164 different proportions of the various types of organic matter (Corg), more or less reactive, to the 165 foraminiferal species introduced into the model.

166 We then tested the model on our extensive geochemical and foraminiferal dataset from the Bay of

167 Biscay. After constructing the essential components of the model, we started to tune the model

using the data of stations A and B, at 1000 and 550 m depth, respectively. We first optimized the fit

between the geochemical data and the model results. Next, we tested various hypothesis concerning the ecological requirements of the benthic foraminifera, in order to obtain a maximum resemblance

171 between observational data and model output.



184 Fig. 1. Map of the bay of Biscay, with sampling stations A (1000 m) and B (550 m).

185 3. The Bay of Biscay data set

186 Bay of Biscay Stations A, at 1000 m water depth, and B, at 550 m depth, were visited 15 times 187 between October 1997 and February 2011. The surficial sediment was sampled with an interface 188 multi-corer, which allowed sampling the upper few decimetres of the sediment, the overlying 189 bottom water, and the undisturbed sediment-water interface, in a 10 cm diameter Plexiglas tube. At 190 both stations, very similar profiles of redox reactive compounds were measured in the numerous 191 short cores collected over the 14 year interval (Mouret et al., 2016). Only small differences were 192 noticed at the sediment surface, which were attributed to seasonality, bioturbation and/or spatial 193 patchiness. For the present study we decided to concentrate on the period between October 1997 194 and September 2001, when these stations were sampled 10 times, so that the seasonal and 195 interannual variability of the geochemical composition of bottom and pore waters, as well as of the 196 foraminiferal community, is particularly well constrained. The pore water geochemistry of these 197 station has been described in detail by Hyacinthe et al. (2001), Anschutz et al. (2005), Mouret et al. (2009, 2010) and Charbonnier et al. (2019). Schmidt et al. (2009) concentrated on the particle fluxes 198 199 and sediment accumulation rate. The foraminiferal community at the two stations was described by 200 Fontanier et al. (2002; 2003; 2006a) and Mojtahid et al. (2010). Important seasonal and interannual 201 variability was observed for the foraminiferal density and species composition. It appears that 202 foraminiferal densities increased to an order of magnitude after major spring and autumn 203 phytoplankton blooms, which favoured a limited number of opportunistic benthic foraminiferal taxa 204 (Fontanier et al., 2003; 2006a).

In view of the observed stability of the succession of sediment redox zones (Mouret et al., 2009), we decided to use an averaged geochemical data set to calibrate our diagenetic model. To this purpose, for every geochemical model parameter, the observations for all available sampling campaigns (their number varied for different parameters) were averaged, so that a single value was obtained for all depth levels down to 10 cm depth in the sediment. For this first modelling study, foraminiferal data

210 were averaged as well, for the 10 successive sampling campaigns.

211 4. Early Diagenesis Model Parameters

212 The first portion of our model utilizes OMEXDIA, a reactive transport model for evaluating sedimentary diagenesis (Soetaert et al., 1996), which was run to steady state. It includes multiple 213 214 organic matter remineralization pathways, incorporates key diagenetic chemical reactions, and 215 allows for depth-dependent bioturbation/biodiffusion and porosity. In the present study, many of 216 the key parameters of the model are constrained by observations derived from prior studies of 217 stations A and B. Using the dataset of stations A and B, the model was manually tuned to provide 218 initial constraints for the complete model, discussed later. The values of the main parameters used 219 as starting point for the early diagenesis part of the model were the following:

- 220 The organic carbon flux (Corg) to the sea floor: for the 1000 m deep station A, Mouret et al. 221 (2010) estimated a total Corg flux of 16.5 gC m⁻² yr⁻¹, consisting of 76% labile particulate organic carbon (POC), 9% slowly decaying POC, and 15% inert POC. At the 550 m deep 222 station B, Mouret et al. (2010) estimated a total C_{org} flux of 28.2 gC m⁻² yr⁻¹, with 53.6% labile 223 224 POC, 20.4% slowly degrading POC and 26.0% inert POC. These values were used as the basis 225 for our tuning exercise. The most labile fraction was deduced from modelled dissolved 226 oxygen consumption at the sediment surface. Other fractions were deduced from Corg 227 concentrations and mass accumulation rates.
- Bottom water temperature was always 9.5°C at station A, and 11.0°C at station B, without
 any seasonal variability (Ogawa et al., 1973; Durrieu de Madron et al., 1999 and Fontanier et
 al., 2006b).
- Bottom water oxygenation varied between 189 and 201 µmol l⁻¹ at station A and between
 205 and 221 µmol l⁻¹ at station B (Mouret et al., 2010; Anschutz and Chaillou; 2009).
- At both stations A and B, the sediment was a silty clay, with a mean grain size of 5–6 μm at station A and 10-15 μm at station B (Charbonnier et al., 2019). The sediment contained about 30% CaCO₃ at station A, versus 10-15% at station B. At both stations, sediment porosity decreased quickly in the first millimeters of the sediment, from 0.85 at the sediment-water interface to about 0.8 at 1 cm depth. Deeper down, porosity continued to decrease, until values between 0.65 and 0.7 were observed at 10 cm depth at stations A and B, respectively.
 The oxygen penetration depth into the sediment varied from 1.6 to 2.2 cm at station A, and
- from 1.1 to 1.9 cm at station B (Fontanier et al., 2003; 2006a; Mouret et al., 2010).
- 241 At station A, nitrate in the bottom waters, sampled in the supernatant water of the sediment 242 cores, varied from 8.6 to 21.1 μ mol l^{-1} , with a mean value of 14.3 μ mol l^{-1} . At station B, nitrate varied from 11.5 to 15.7 $\mu mol~l^{\text{-1}}$, with a mean value of 14.1 $\mu mol~l^{\text{-1}}$. These values are 243 244 very close to those measured on water column samples taken with Niskin bottles (Anschutz 245 and Chaillou, 2009). Maximum nitrate concentrations were generally observed in the top 2 246 cm of the sediment, with values of 24 to 33 μ mol I⁻¹ at station A, and 19 to 23 μ mol I⁻¹ at 247 station B. Deeper down, nitrate concentrations strongly decreased, with values below 23 248 $\mu mol~l^{\rm -1}$ reached between 4 and 5 cm at station A and around 2 cm at station B. Ammonia 249 always showed a strong downward increase from values around zero at the surface to mean 250 values of 30.7 µmol I⁻¹ (station A) and 61.1 µmol I⁻¹ (station B) at 10 cm depth.
- Among the reduced components, dissolved Mn²⁺ and Fe²⁺ were only determined for part of the 10 sampling campaigns (Mouret et al., 2009), whereas no data for S(-II) were available.
 The sulphate concentration was constant with depth and equal to the SO₄²⁻ concentration in seawater. No sulphide smell was evident during core processing, but a few black patches were observed near the base of the cores, suggesting H₂S production in deeper sediment layers (Chaillou et al. 2003). Since the H₂S concentration was not measured, the model output for ODU (oxygen demand units, i.e. the amount of oxygen necessary to reoxidize

- reduced counterparts from anaerobic mineralization), which regroups all reduced
 components, was always higher than the observations, which include only measured
 dissolved Mn and Fe.
- At station A, the maximum sediment accumulation rate was estimated on the basis of ²¹⁰Pb profiles at 0.068 cm/year (36 mg cm⁻² yr⁻¹) by Chaillou et al. (2002), at 0.05 to 0.06 cm/year by Schmidt et al. (2009), whereas Mouret et al., 2009 obtained a value of 0.062 cm/year based on steady state manganese diagenesis. For station B, Chaillou et al. (2002) obtained a maximum sedimentation rate of 0.15 cm/year (79.5 mg cm⁻² yr⁻¹), compared to values of 0.12 and 0.22 cm/year by Schmidt et al. (2009), and 0.12 cm/year based on steady state manganese diagenesis (Mouret et al., 2009).
- Mouret et al. (2009) obtained a large variety in biodiffusion coefficients, using ²³⁴Th profiles, for sediment cores sampled during four different campaigns, with values varying between 0.12 and 6.7 cm²/year at station A, and between 0.4 and 14.8 cm²/year at station B. They estimated a mixed layer depth of about 1cm at both stations, slightly less than the depth of the oxic layer.
- Despite the lack of observational data for some parameters (such as sulphur) in some of the
 cores, and the large variability observed for biodiffusion coefficients, the Bay of Biscay dataset
 for these two stations is remarkable by its near completeness for 10 successive sampling
 campaigns. It therefore forms an ideal dataset to develop and test our early diagenetic model.
 The main parameter that we used to tune the model was the C_{org} flux to the sea floor, or more
 precisely the proportions and reactivity constants of the various types of organic matter. A
 secondary main tuning parameter was the rate of pyrite formation in the sediment; this process
- 280 withdraws reduced components from the pore water.

282 5. Initial tuning of the preliminary (3G) early diagenetic model, using the station A data set

283 Before introducing theoretical foraminiferal species into the model, our first objective was to tune

284 the early diagenetic model to obtain an optimal fit between the averaged observed data and the

285 model output. For this initial tuning exercise, we used the dataset for station A. Fig. 2 gives an

example of an early result that employed three types of organic matter (3G).



Fig. 2. Station A - Model output for early diagenetic parameters (lines) of a preliminary version of the 4GFOR
 model, with only three types of organic matter (percent TOC = particulate Corg in weight % of dry sediment; red:
 fast degrading, blue: intermediate, black: total organic carbon (= fast degrading + intermediate + fossil),
 compared to observational data from Station A (dots).

302 It can be seen that the model results for C_{org}, oxygen and ammonia are very close to the observed

303 data. However, although the modelled ODU curve has the right form, the absolute values are much

higher than the observed ones. As explained in the previous section, this is likely due to the fact that
 only Fe and Mn were measured, and not S, a third major component contributing to ODU. As we do

306 not intend to use ODU as a limiting factor for the foraminiferal species, we do not consider this very

307 important. The main problem with this preliminary model is the nitrate curve, which in deeper

308 sediment layers shows values that are too high compared to the observed ones. This is important,

309 because we intend to use a minimum NO₃ concentration as a switch for nitrate-respiring

310 foraminifera in the subsequent foraminiferal component of the model. All our attempts to improve

the fit for nitrate systematically resulted in an upward movement of the oxygen profile, which is even

more problematic, because we intended to use oxygen concentration as a switch for obligate aerobic
 foraminiferal species. In the final version of the model we resolved this problem by adding an

additional diagenetic pathway, ferrous iron nitrification. This reaction, the anaerobic oxidation of Fe^{2+}

by nitrates (e.g., Straub et al., 1996; Kappler et al., 2021), appears to be an important process at our

sites in the Bay of Biscay (Hyacinthe et al., 2001), and was hitherto not included in the OMEXDIA

317 model. This early diagenetic pathway consumes nitrate at the lower part of its range, and diminishes

ODU. After adding this reaction to the early diagenetic part of the model, we arrived at excellent fits for all parameters except ammonia, which was overestimated (Fig. 5). Since ammonia has no impact

320 on the faunal distribution, this was considered less important.

321 6. Defining the foraminiferal species

As explained before, we decided not to include a fully biological model for the foraminiferal species, but rather a transformation model. This transformation model has two basic characteristics. First,

but rather a transformation model. This transformation model has two basic characteristics. First,
 foraminiferal activity is limited by critical environmental factors, such as oxygen and nitrate content.

325 Beyond the critical threshold values, specified foraminifera are no longer given access to the

available resources. Next, when these limiting factors are within the tolerance limits of the organism,

a specific proportion of each type of C_{org} is attributed to each of the species. In the way we defined

328 them, species living close to the sediment surface use a large proportion of reactive Corg, whereas

329 species living deeper in the sediment use a larger proportion of slowly degrading C_{org}. This

330 interspecific difference is based on two assumptions: 1) that infaunal foraminiferal taxa are better

capable than surface dwellers to feed on organic matter of a lower quality, and 2) that sediment

332 surface dwellers are more competitive than infaunal taxa when the uptake of high quality (highly

333 reactive) C_{org} is concerned.

The allocation of different proportions of various qualities of organic carbon to different species is
 reminiscent of the BFAR-concept of Herguera and Berger (1991). In fact, Herguera and Berger
 assumed that for every mg of C_{org} arriving at the sea floor, roughly one foraminiferal shell > 150 µm

337 would be preserved in the sediment record.

338 In our first attempts to model the foraminiferal distribution in the sediment (for station A), we used a

339 3G diagenetic model, with fast, slowly degrading, and inert C_{org} ; a very good fit between model 340 output and observed data was obtained with decay rates $r_{fast} = 0.01/day$, $r_{slow} = 0.00005/day$ and r_{fossil}

341 = 0. In these first runs (Fig. 3), we introduced only 2 foraminiferal species:

1) an epifaunal/shallow infaunal species, which was dependant on the presence of oxygen, at least at very low concentrations ($0_2 > 2.5 \mu mol l^{-1}$). We defined this species group so that it exclusively feeds on fast decaying C_{org} .

2) a deep infaunal species, which we inhibited by oxygen ($0_2 \le 2.5 \mu mol l^{-1}$), but which was dependent on the presence of nitrate (NO₃ > 2.5 $\mu mol l^{-1}$), and given access to both fast and slowly degrading

347 C_{org}.

348 In order to compare the model outcome (in mgC cm⁻³) with the observational data (observed number

- 349 of tests > 150 μm), we assumed that a single foraminiferal individual (> 150 μm) of the
- 350 epifaunal/shallow infaunal species would contain 0.625 μg C_{org}, whereas a deep infaunal individual (>



352 relationship between biovolume and protein content observed by Movellan et al. (2012), assuming

353 an average size of 250 µm for epifaunal/shallow infaunal taxa (considered as semi-spheres,

354 representative of most epifaunal and trochospiral taxa) and of 450 µm for deep infaunal taxa

355 (considered as a cylinder, such as Globobulimina spp.).





367 Fig. 3. First results of the coupled 3G early diagenesis (using two types of reactive organic matter) -

368 foraminiferal model. Left panel: epifaunal/shallow infaunal taxa (black) and deep infaunal taxa (red). Right

369 panel: fast degrading C_{org} (red) and intermediate degrading C_{org} (blue) Dots = observed data for station A (1000 370 m depth); lines = model output. Epifaunal/shallow infaunal taxa (black curve) only have access to fast decaying 371 organic matter.

372 7. Further developing the (3G) early diagenesis - perfecting the foraminiferal model for 373 station A

374 The first results of the coupled early diagenesis-foraminifera model for the 1000 m deep station A

375 (Fig. 3) were encouraging. To arrive at this fit, we attributed 0.2% of fast decaying Corg to

epifaunal/shallow infauna (when O₂ > 2.5 µmol I⁻¹, left panel, black curve), and 0.1% of fast decaying 376 377 C_{org} and 0.002% of slowly degrading C_{org} to deep infauna (when $O_2 \le 2.5 \ \mu mol \ l^{-1}$ and $NO_3 > 2.5 \ \mu mol \ l^{-1}$ 378 ¹, left panel, red curve).

379 These values, which were selected to obtain a reasonable fit, are in reality thought to reflect the

380 capacity of these species to feed on various types of Corg, and their competitive success compared to

381 other (foraminiferal as well as non foraminiferal) organisms feeding on the same resources. For

382 comparison, for continental shelf (37-60 m depth) and upper slope (450 m depth) environments with well oxygenated bottom waters, Geslin et al. (2011) estimated the total foraminiferal contribution to 383

384 dissolved oxygen utilisation at 0.5 to 2.5%.

385 However, although the obtained result (Fig. 3) was encouraging, it was not yet fully acceptable.

386 There were in fact three major problems. The first problem was that the epifaunal/shallow infaunal

- 387 taxon did not penetrate deep enough into the sediment; their strong decline in the first cm of the
- 388 sediment is the direct consequence of the similar decline of the fast degrading C_{org} (red curve in the 389 right panel of fig. 3). In our opinion, the model results clearly show, that our initial hypothesis, that

epifaunal/shallow infaunal taxa exclusively feed on fast degrading C_{org}, is not supported. We solved

- 391 this problem by attributing also a small fraction (0.004%) of slowly degrading $C_{\mbox{\scriptsize org}}$ to
- 392 epifaunal/shallow infaunal species (Fig. 4).



Fig. 4. First results of the coupled (3G) early diagenesis (using three types of organic matter) - foraminiferal
model for station A (1000 m depth). Left panel: epifaunal/shallow infaunal taxa (black) and deep infaunal taxa
(red). Right panel: fast degrading Corg (red) and intermediate degrading Corg (blue). Dots = observed data; lines =
model output. Epifaunal/shallow infaunal taxa now have also access to a small part of slowly degrading Corg.

The second problem was the fact that in reality, there are more than two groups of foraminiferal species. There are not only deep infaunal species (such as *Globobulimina* spp.), but also intermediate infaunal species, such as *Melonis barleeanus*, which always attain the highest density at the depth where nitrate concentration is maximal (e.g., Jorissen et al., 1998). In addition, the group of epifauna/shallow infauna seems to be more complex, with some taxa always living close to the sediment-water interface, and others living slightly deeper in the oxygenated part of the surficial sediment. The first group is dominated by trochospiral genera (*Cibicidoides, Gyroidina, Hoeglundina*,

415 *Oridorsalis,* etc.), whereas the second group is rather dominated by triserial and biserial genera

416 (Bolivina, Bulimina, Trifarina, Uvigerina, etc.).

417 In order to take into account this complexity, we decided to expand our model to four theoretical

418 foraminiferal taxa, each of which is supposed to represent a group of real species with similar

419 microhabitat characteristics. Since we think that the name "epifauna" should be reserved for taxa 420 living on elevated substrates above the sediment, we will use the terms "shallow infauna A" (SIA),

living on elevated substrates above the sediment, we will use the terms "shallow infauna A" (SIA),
"shallow infauna B" (SIB), "intermediate infauna" (II) and "deep infauna" (DI) for our four theoretical

422 species (groups) living at increasingly greater depth in the sediment.

In order to model the vertical distribution of these four theoretical species, it soon turned out that a rough subdivision of the organic carbon resources into three types (fast degrading, slowly degrading, fossil) was insufficient. For this reason, we shifted to the final 4G model, with three types of reactive

426 C_{org}: fast, intermediate and slowly degrading, and finally inert C_{org}.

The final problem was that in the first versions of the model (Figs. 3, 4), we inhibited the activity ofthe deep infaunal species with oxygen. However, Nomaki et al. (2007) observed oxic respiration for

- 429 the deep infaunal taxa *Globobulimina affinis* and *Chilostomella ovoidea*. Consequently, in most 430 studies, deep infaunal taxa are considered as facultative anaerobes, which will only shift to an
- anaerobic metabolism (using nitrate) when no oxygen is available.
- 432 We solved this problem by replacing the inhibition with an additional scaling factor. In fact, we
- 433 defined the intermediate and deep infaunal categories in such a way that the amount of fast and
- 434 intermediate degrading C_{org} allocated to them decreases exponentially towards higher oxygen
- 435 concentration. In biological terms, this means that these taxa become rapidly less competitive
- 436 (compared to shallow infaunal groups A and B) when oxygen concentrations increase. In our opinion,
- 437 this is a very realistic way to define these species for the sake of our quantitative model.

438 8. Tuning the final 4GFOR Model for stations A (1000 m) and B (550 m).

- 439 Concerning the final tuning of the early diagenesis part of the 4GFOR model, an optimal fit between
- $\label{eq:constraint} 440 \qquad \mbox{model output and observations was obtained by using a total C_{org} flux of 16.0 and 25.0 $\mbox{ gC m}^{-2}$ \mbox{yr}^{-1} at C_{org} flux of 16.0 and 25.0 $\mbox{ gC m}^{-2}$ \mbox{yr}^{-1} at C_{org} flux of 16.0 and 25.0 $\mbox{ gC m}^{-2}$ \mbox{yr}^{-1} at C_{org} flux of 16.0 and 25.0 $\mbox{ gC m}^{-2}$ \mbox{yr}^{-1} at C_{org} flux of 16.0 and 25.0 $\mbox{ gC m}^{-2}$ \mbox{yr}^{-1} at C_{org} flux of 16.0 and 25.0 $\mbox{ gC m}^{-2}$ \mbox{yr}^{-1} at C_{org} flux of 16.0 and 25.0 $\mbox{ gC m}^{-2}$ \mbox{yr}^{-1} at C_{org} flux of 16.0 and 25.0 $\mbox{ gC m}^{-2}$ \mbox{yr}^{-1} at \mbox{yr}^{-1
- 441 stations A and B, respectively (table 1). These values are very close to the values of 16.5 and 28.2 gC
- 442 $m^{-2} yr^{-1}$ given by Mouret et al., 2010).

Parameter	Description	Station A	Station B	Unit	Source
maxdep	Maximum depth for modeled sediment	24	24	cm	1
pO	Porosity in surface sediment	83.3	88.5	%	1
pinf	Porosity at maximum modeled depth	64.2	69.7	%	1
рсо	Coefficient of decrease for porosity	4.47	2.76		1
sedr	Sedimentation rate	0.055	0.15	cm/year	L ⁶
Db	Bioturbation rate	0.75	1.25	cm/year	L ³
mixl	Mixed layer depth	1.0	1.0	cm	L ³
fluxm	Total Corg flux at sediment water interface	16.0	25.0	g m ⁻² y ⁻¹	L ³ ; C
TCorginf	%Corg at infinite depth	0.79	0.83	%	L ³
rfast	1st order rate constant for fast degrading Corg	0.026	0.015	day ⁻¹	С
rint	1st order rate constant for intermediately degrading Corg	0.001	0.0005	day ⁻¹	С
rslow	1st order rate constant for slowly degrading Corg	0.00003	0.0000025	day ⁻¹	С
pfast	fraction of fast degrading Corg in total Corg flux	58.41	34.0	%	С
pint	fraction of intermediate degrading Corg in total Corg flux	20.0	21.5	%	С
pslow	fraction of slow degrading Corg in total Corg flux	6.26	18.5	%	С
NCrFdat	NC ratio fast decay det	0.16	0.16	molN/molC	L⁵; C
NCrIdat	NC ratio intermediate decay det	0.2	0.2	molN/molC	С
NCrSdat	NC ratio slow decay det	0.13	0.13	molN/molC	L5
Pdepo	Fraction of ODU production deposited as solid	0.44	0.44	% day-1	L
Temp	Bottom water temperature	9.5	11	°C	L ^{2,4}
bwO2	Oxygen concentration in bottom water	196.5	213.4	mmol m-3	L ³ ; I
bwNO3	Nitrate concentration in bottom water	16	14	mmol m-3	L ¹ ; I
bwNH3	Ammonia concentration in bottom water	0	0.6	mmol m-3	L ¹ ; I
NH3bot	Ammonia conc at bottom modeled sediment column	68.29	200	mmol m-3	1
ksO2oxic	Half-saturation O2 in oxic mineralisation	30	30	mmol m-3	L5
ksNO3denit	Half-saturation NO3 in denitrif	15	15	mmol m-3	L
kinO2denit	Half-saturation O2 inhib denitrif	0.5	0.5	mmol m-3	L5
kinNO3anox	Half-saturation NO3 inhib anoxic min	1	1	mmol m-3	L
kinO2anox	Half-saturation O2 inhib anoxic min	5	5	mmol m-3	L
ksO2nitri	Half-saturation O2 in nitrification	100	100	mmol m-3	L5
ksO2oduox	Half-saturation O2 in oxidation of ODU	100	100	mmol m-3	L
rODUox	Maximum rate oxidation of ODU	20	20	day ⁻¹	L⁵
rnit	Maximum nitrification rate	20	20	day ⁻¹	L⁵
NH3Ads	Adsorption coefficient ammonium	1.3	1.3		L ⁵
roxidNO3	Maximum oxidation of ODU by NO3	200	200	day ⁻¹	L5
ksNO3ODUox	Half-saturation NO3 in Fe oxydation	200	200	mmol m-3	L ⁵
kinO2ODUox	Half-saturation 02 in Fe oxydation	1	1	mmol m-3	L ⁵

443

444 Table 1. Parameters used in the final simulation of the early diagenesis part of the 4GFOR model. The source of

445 the parameter values is indicated as follows: C = constrained with the model using the measured data; I =

446 independently determined from field data; L = based on literature values, retrieved from: ¹Anschutz and

Chaillou (2009); ² Durrieu de Madron et al. (1999); ³ Mouret et al. (2010); ⁴ Ogawa et al. (1973); ⁵ Soetaert et al.
 (1996); ⁶ Schmidt et al. (2009).

449 Compared to Mouret et al. (2010), the fractions of inert Corg (0.153 and 0.26) were not changed. Of

450 the 0.2 and 0.215 parts (stations A and B, respectively) attributed to the newly created category

451 Corg $_{int}$, compared to the observations of Mouret et al. (2010), the large majority (87.5 and 91.2% for

452 stations A and B) was taken from the labile $C_{\text{org.}}$ and the remaining part from the slowly decaying $C_{\text{org.}}$

453 The presence of the four foraminiferal species was constrained with the boundary conditions listed in

454 Table 2, which determined whether they were present or not. In our first attempts, using the data of

455 station A, for deep infauna, we used a threshold value for nitrate of 2.5 μ mol l⁻¹. It turned out that at

456 station B, where the nitrate concentration diminishes rapidly with greater depth in the sediment, this

460			
400		Oxygen	Nitrate
461	Shallow infauna A	>2.5 µmol l ⁻¹	No nitrate limitation
	Shallow infauna B	>2.5 µmol l ⁻¹	No nitrate limitation
462	Intermediate infauna	No oxygen limitation	>2.5 µmol l ⁻¹
463	Deep infauna	No oxygen limitation	>0.05 µmol l ⁻¹

Table 2. Threshold values of oxygen and nitrate concentrations for the four theoretical species of benthic
 foraminifera.

466 Finally, foraminiferal biomass was tuned to the observational data by changing the fractions of the
 467 different types of Corg attributed to the four theoretical species of foraminifera, as shown in Table 3.

Station A – 1000 m	Corg _{fast}	Corg _{int}	Corg _{slow}
Shallow infauna A	0.0006	0.0006	0
Shallow infauna B	0.0003	0.0012	0
Intermediate infauna	0.0002	0.0004	0.00001
Deep infauna	0.0001	0.0001	0.00015

threshold truncated the downward distribution of deep infauna. In order to better simulate our field

observations, we subsequently decreased the nitrate threshold for deep infauna to 0.05 μ mol l⁻¹.

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Station B – 550 m	Corg _{fast}	Corg _{int}	Corg _{slow}
Shallow infauna A	0.0006	0.0006	0
Shallow infauna B	0.0003	0.0012	0
Intermediate infauna	0.004	0.00033	0.00001
Deep infauna	0.0001	0.00001	0.00011

469

470Table 3. Fractions of the three types of reactive organic carbon attributed to each of the 4 theoretical species of471foraminifera, at stations A and B.

The competitiveness of intermediate infauna/deep infauna with respect to highly and intermediately
reactive C_{org} further depends on oxygenation: it is low/very low under oxic conditions, and high/very

474 high under hypoxic/anoxic conditions.

475 The final results of the tuning procedure of the 4GFOR model are shown in Figs. 5 and 6. As noted

476 earlier, the fit between the observed and modelled profiles of early diagenetic parameters is very

477 satisfactory. The two shallow infaunal species (A and B) are well separated in the top 1-2 cm of the

478 sediment, suggesting that our decision to differentiate them by their ability to cope with low quality

 $479 \qquad C_{\text{org}} \text{ may be realistic. The profiles of intermediate and deep infaunal taxa correspond well with the}$

480 observed data, although in detail, the modelled distribution is slightly off for some levels.



Fig. 5. Final results for station A (1000 m depth) of the 4GFOR model with four types of organic matter (red:
fast degrading, green: intermediate, blue: slow, and black: total TOC). Foraminiferal species distribution is
represented as: Shallow Infauna A = black, Shallow Infauna B = orange, Intermediate Infauna = blue, Deep

485 Infauna = green. All dots are averaged observed data, lines are model output.



Fig. 6. Final results for station B (550 m depth) of the 4GFOR model with four types of organic matter (red: fast degrading, green: intermediate, blue: slow, and black: total TOC). Foraminiferal species distribution is
represented as: Shallow Infauna A = black, Shallow Infauna B = orange, Intermediate Infauna = blue, Deep
Infauna = green. All dots are averaged observed data, lines are model output.

491 9. Discussion

486

492 <u>9.1. Definition of organic matter</u>

493 The organic matter flux to the seafloor is composed of a large variety of particles, such as living 494 microbial cells and secondary producers, detrital material, fecal pellets, other aggregates, and 495 continental organic matter, as well as of dissolved organic carbon (Middelburg, 2018). Consequently, 496 particulate organic matter has a wide range of components, such as inert carbon, lipids, amino acids, 497 vitamins and carbohydrates. These components have very different reactivities. In models this range 498 of organic matter can be represented in different ways, by two (2G), three (3G) or four (4G) groups 499 with different reactivity, or, in the most sophisticated models, by a continuous range of organic 500 matter quality (Arndt et al., 2013). In the first version of our model, we used a 3G approach, with 501 inert, reactive and less reactive Corg types, whereas in the last (4G) version we added an 502 intermediate category, which allowed us to obtain much better simulations of the observational 503 data.

505 9.2. Characteristics of foraminiferal species

506 While developing the 4GFOR model, the characteristics of our theoretical foraminiferal species have 507 progressively evolved, to become more complex. In our first runs, we attributed only fast degrading 508 Corg to the shallow infaunal species. It turned out that because of the rapid decrease of Corg-fast with 509 depth, compared to observational data, our model species did not penetrate deep enough into the 510 sediment. This example emphasizes the great importance of theoretical models: apparently the basic 511 hypothesis (= shallow infaunal taxa feed exclusively on Corg-fast) was wrong, and consequently, the 512 model was incapable of simulating the observed data.

513 We solved this problem by separating the group of shallow infaunal taxa into two species, SIA and 514 SIB, with SIB being much more tolerant to lower quality C_{org} than SIA. The existence of two shallow 515 infaunal groups with different dominant test forms (SIA = trochospiral, planispiral; SIB = biserial, 516 triserial) is in accordance with abundant microhabitat observations in natural ecosystems and is corroborated by the interspecific differences in the δ^{13} C of their shells (e.g., McCorkle et al., 1990; 517 518 Rathburn et al., 1996; Fontanier et al., 2006b). By giving them access to different proportions of fast 519 degrading and intermediate Corg, we could obtain a nearly perfect fit with the observed data.

520 In our first trials, we used only a single deeper infaunal taxon, and we forced this species to remain at 521 depth in the sediment by limiting it to oxygen concentrations below 2.5 µmol l⁻¹. In the later versions, 522 we separated this group into intermediate and deep infaunal taxa, in accordance with the large volume of natural observations. Both groups were initially defined in such a way that they can only 523 occur when nitrate concentration is above 2.5 µmol I⁻¹. In order to obtain a deeper penetration of 524 deep infauna at station B, we had to lower this threshold to 0.05 μ mol l⁻¹ for this group. We further 525 526 differentiated intermediate and deep infauna by making DI more tolerant to low quality Corg than II. 527 To avoid that these species invade the superficial sediment layers, where large amounts of highly 528 reactive Corg are present, we added a competitive element, by attributing less Corg-fast and Corg-int to 529 these taxa when oxygen rises above 2.5 µmol I⁻¹, with an exponential decrease towards higher 530 oxygen levels. With these characteristics, the overall distribution of II and DI is very close to the 531 observed patterns. This suggests that the way we defined these taxa may be correct, and that their 532 absence in the superficial sediment layers may indeed be the consequence of a strongly diminished 533 competitiveness in well oxygenated environments. In more general terms, this suggests that 534 interspecific competition for food particles may be an important controlling parameter.

535 Finally, when tuning the model for two stations with very different characteristics, we managed to 536 obtain a good fit for the early diagenesis parameters while using the same reaction constants at both 537 stations. This underlines the robustness of this part of the model. In order to obtain good fits for the 538 foraminiferal data at both stations, we had to attribute slightly different proportions of the different 539 organic matter types to the four theoretical species (Table 3). This suggests that the foraminiferal 540 contribution to organic matter remineralisation may change as a function of the environmental

conditions. 542 9.3. Perspectives

541

543 Here, we present the newly developed 4GFOR model, and our tuning with the data of two sampling 544 station at 1000 and 550 m depth in the Bay of Biscay, on the basis of averaged data of 10 successive 545 sampling campaigns. In view of the obtained simulations, which are very close, both for geochemical 546 as for foraminiferal species, it appears that the complexity of the model is enough to correctly

547 represent the four main groups of foraminifera. The next step will be to test the model in different

548 environmental settings. We will accomplish this by modelling 1) the 10 successive sample times

individually, 2) the 8 other stations of the Bay of Biscay transect, between 140 and 4800 m depth,

550 with a wide range of environmental characteristics, 3) sampling stations from other ocean basins

551 (Indian Ocean, Cape Blanc upwelling) from which we have comparable data sets. These modelling

exercises will very probably lead to adjustments of the model, and the way the foraminiferal speciesare defined. This will oblige us to consider slightly different ecological strategies for the four species,

and thereby, to test many hypotheses we still have regarding their life strategies.

555 Once the robustness of the 4GFOR model has been fully demonstrated, the next step will be to

556 develop a formal inversion of the model, so that the main parameters controlling the foraminiferal

557 faunas, the quantity and quality of the organic flux to the sea floor and the vertical succession of

- redox species, can be reconstructed on the basis of the composition of fossil faunas in sediment
- 559 cores and outcrops.

560 **10. Conclusion**

561 The 4GFor model is a combination of an early diagenetic model, based on the OMEXDIA model of

562 Soetaert et al. (1996), and a transformation model, which transforms proportions of organic matter

563 types with different reaction rates into foraminiferal biomass. Foraminiferal communities are

simulated with four theoretical deep water species. Their different tolerance levels with respect to oxygen and nitrate concentrations decide at what depths they can occur, whereas their density is

566 mainly controlled by organic matter quality.

567 The model has been tuned to a large geochemical and foraminiferal database of two stations, at

5681000 and 550 depth in the Bay of Biscay. The outcome shows that the early diagenesis part of the

569 model correctly reproduces the vertical distribution of redox species in the top 10 cm of the 570 sediment. By attributing various proportions of the different types of organic matter to the four

sediment. By attributing various proportions of the different types of organic matter to the four
 foraminiferal species, the model adequately reproduces the vertical succession of shallow,

572 intermediate and deep infaunal species.

573 The model further separates shallow infaunal taxa into two groups. Group SIA, which is found close

to the sediment-water interface, corresponds to trochospiral (e.g., *Cibicidoides, Gyroidina*,

575 *Hoeglundina*) and planispiral species (e.g., *Cassidulina*). This group is critically sensitive to food

576 quality. Group SIB, which is found slightly deeper in the sediment, corresponds to triserial (e.g.,

577 *Bulimina, Uvigerina*) and biserial (e.g., *Bolivina*) taxa, which are less restrictive with respect to food 578 quality. The observed distribution of these groups at the two stations in the Bay of Biscay is

reproduced by the model. Also, the depth in the sediment of intermediate (e.g., *Melonis barleeanus*)

580 and deep infaunal (e.g., *Globobulimina* spp.) species is very well simulated.

581 The tuning of the model indicates that shallow infaunal taxa (neither SIA nor SIB) do not feed

exclusively on the most labile C_{org} component. Their vertical distribution can only be simulated by
 giving them access to slightly less reactive C_{org}.

584 One of the problems encountered during the tuning phase was to keep intermediate and deep

585 infaunal taxa restricted to deeper layers. Because of the presence of large quantities of high quality

586 C_{org} close to the sediment surface, this layer is also attractive for those species. To solve this problem,

587 we diminished their competitive ability exponentially towards higher oxygen levels. This solution

588 appears to be realistic, but should be tested in experimental conditions.

As is shown by these first examples, the 4GFOR model can be a powerful tool to help understand
 foraminiferal ecology. In the future, we intend to construct a formal inversion of the model, which

- 591 will make it possible to produce quantitative estimates of the main controlling faunal parameters 592 (quantity and quality of the organic flux, oxygen and nitrate concentration) on the basis of the
- 593 composition of fossil foraminiferal assemblages.

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597 12. References

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768	The 4GFOR Model – coupling 4G Early Diagenesis and Benthic
769	Foraminiferal Ecology
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771 772	Frans J. Jorissen ^{1*} , Stephen R. Meyers ² , Boris A. Kelly-Gerreyn ³ , Louison Huchet ¹ , Aurélia Mouret ¹ and Pierre Anschutz ⁴
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774 775 776 777 778 779 780 781	 ¹ UMR CNRS 6112 LPG-BIAF, Angers University, France ² University of Wisconsin, Department of Geoscience, Madison, USA ³ Mount Waverley, Melbourne, Victoria 3149, Australia ⁴ Univ. Bordeaux, CNRS, EPOC, EPHE, UMR 5805, F-33600 Pessac, France * Corresponding author: Frans J. Jorissen, UMR CNRS 6112 LPG-BIAF, Université d'Angers, 2, Boulevard Lavoisier, 49045 Angers Cedex, France. Email address : frans.jorissen@univ-angers.fr
782	Abstract
783 784 785 786 787 788 790 791 792 793 794 795 796 797 798 799 800 801	The 4GFOR model presented in this paper combines the OMEXDIA model of Soetaert et al. (1996) with four theoretical species of benthic foraminifera. The geochemical part of the model simulates the succession of early diagenetic redox reactions in the superficial sediment layers. The four theoretical foraminiferal species, which each represent a group of biological species with comparable microhabitats, differ by their limitations with respect to oxygen and nitrate concentration and their ability to feed on different organic matter types. The foraminiferal model is a transformation model in which different proportions of four types of organic matter with different reactivity are attributed to each of the theoretical species. The model has been manually tuned and tested on a large data set from two stations, at 1000 and 550 m depth, in the Bay of Biscay. The model adequately simulates the vertical distribution of shallow, intermediate and deep infaunal species. The 4GFOR model can be used to test several hypotheses concerning foraminiferal ecology. The model has already allowed us to conclude that it is high unlikely that shallow infaunal taxa feed exclusively on the most labile part of the organic matter flux, since it as unable to reproduce the field observations while using this hypothesis. Another important conclusion based on our preliminary model results is that the absence of deep infaunal taxa close to the sediment surface may be explained by their decreasing competitiveness in well oxygen and nitrate concentrations; quantity and quality of the organic flux to the ocean floor) from benthic foraminiferal assemblage data.
802	Keywords

- 803 Benthic foraminifera, ecology, microhabitats, early diagenesis, model.
- 804

805 1. Introduction

806 Since the pioneering work of Corliss and his colleagues in the 1980's (Corliss, 1985; Corliss & Chen 807 1988; Corliss and Emerson, 1990) - which has been fully confirmed by a number of subsequent 808 studies (i.e., Gooday, 1986; Mackensen and Douglas, 1989; McCorkle et al. 1990; Rathburn & Corliss, 809 1994; Jorissen et al., 1998) - we know that in open marine settings, there is a vertical succession of 810 benthic foraminiferal species in the surface sediment. This species succession, which appears to be 811 closely related with the well-known succession of redox layers in the surface sediment (e.g., Froelich 812 et al., 1979, Canfield et al., 1993) may be expanded over several centimetres (up to ~10 cm at most) 813 or be compressed in the first few millimetres of the sediment. Despite of this vertical variability, the 814 succession of foraminiferal species is always the same (Jorissen, 1999a). Consequently, foraminiferal 815 species may be considered as shallow, intermediate or deep infaunal, not so much on the basis of the 816 exact depth at which they are found in the sediment column, which can vary considerably, but rather 817 on their relative position in the vertical species succession (Jorissen et al., 2007).

818 Following earlier suggestions of Shirayama (1984), Corliss and Emerson (1990) and Loubere et al. 819 (1993), Jorissen et al. (1995) proposed the so-called TROX-model, which hypothesized that the 820 vertical penetration depth of foraminiferal species into the sediment is determined by food 821 availability in oligotrophic ecosystems, but would be limited by redox conditions (especially by pore 822 water oxygen concentration and by the presence of toxic dissolved sulphides) in eutrophic settings. 823 Different requirements with respect to these limiting factors would then explain the specific 824 microhabitats of individual species. Indirectly, the TROX model also explains biodiversity trends, 825 which are strongly affected by the availability or not of ecological niches deeper in the sediment. This 826 conceptual model has since been confirmed by a large number of observational studies, and it has 827 also been proposed to explain macrofaunal distribution patterns (Carney, 2005). 828 Since 1995, several authors have tried to further elaborate upon the TROX model, by integrating

829 bioturbation (Van der Zwaan et al., 1999), or by coupling the foraminiferal distribution with several 830 redox species (Koho et al., 2015), and not only with oxygen, as in the original model. In the 25 years 831 since the publication of the TROX model, we have also learned a lot about benthic foraminiferal 832 ecology. After the ground-breaking publications of Risgaard-Petersen et al. (2006), Høgslund et al. 833 (2008), Pina-Ochoa et al. (2010) and Bernhard et al. (2012), we know that many species are capable 834 of storing nitrate in their protoplasm, and at least some of these species are capable of respiring 835 nitrate when confronted with anoxia. For other taxa, a capability to sequester chloroplasts from 836 consumed diatoms has been shown (e.g., Lee and Anderson, 1991; Bernhard and Bowser, 1999). The 837 foraminifera are able to keep these intracellular chloroplasts active for several weeks, showing 838 another foraminiferal strategy for survival under strongly adverse conditions (Jauffrais et al., 2016). 839 And finally, a last group of species may survive adverse conditions by entering into a state of 840 dormancy, with a strongly diminished metabolic rate (Bernhard and Alve, 1997; Ross and Hallock, 841 2016).

842 The presence of an alternative metabolic pathway, in facultatively anaerobic taxa, as well as the 843 other metabolic adaptations, explains why most benthic foraminiferal species are tolerant to strong 844 hypoxia (at least down to ~0.5 ml/l, or 22 μ mol/l; Jorissen et al., 2007). This also explains how 845 foraminifera can survive, and for some species at least, remain active in anoxic conditions. In this 846 context, several observational studies have shown foraminiferal survival after exposure to anoxia together with free sulphides (e.g., Langlet et al., 2013; 2014, Richirt et al., 2020), a combination 847 which until recently was generally considered lethal (Moodley et al., 1998). The ultimate 848 849 disappearance of benthic foraminifera during cases of long-term anoxia, as is for instance observed 850 in Mediterranean sapropels (e.g., Jorissen, 1999b), seems therefore not to be the consequence of

strongly increased mortality, but rather of an inhibition of reproduction under strongly adverseconditions (Richirt et al., 2020).

These recent advances suggest that oxygen may not be the foremost environmental parameter

explaining foraminiferal ecology, as was often thought in the past (e.g., Bernhard, 1886; Alve and

855 Bernhard, 1995), and was more or less hypothesized in the TROX model. Instead, it appears that it is

rather the quantity, and especially the quality of the food particles, that are the predominant
 parameters. However, the quality of the organic matter (C_{org}) flux will also affect the biological

oxygen demand, and thereby the bottom water and sediment redox conditions, so that the

combination of the organic matter flux and oxygen content forms an inextricable complex.

860 Despite the determinant role of the organic matter flux, oxygen will still be a principal controlling

factor when values are close to (or below) the tolerance levels of many species and/or higher

862 taxonomical categories. Although alternative metabolic pathways for organic matter decomposition

can be utilized under suboxic and anoxic conditions, they are energetically much less favourable,

with the exception of denitrification (Froelich et al., 1979; Middelburg, 2019).

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Furthermore, we have indications that the presence of free sulphides is not directly lethal (Bernhard et al., 1993; Langlet et al., 2014), but that long-term exposure leads to a decrease of foraminiferal population densities (Moodley et al., 1998; Richirt et al., 2020). It has been suggested that this would be due to a decrease of metabolic rate, and a loss of vital functions, such as growth, and especially reproduction (Moodley et al., 1998; Richirt et al., 2020). Long-term exposure to dissolved sulphides should therefore ultimately lead to the disappearance of foraminifera.

Summarising, even if our ideas about the specific role of some environmental factors, such as oxygen
availability, have become more nuanced, the importance of redox conditions within the foraminiferal
microhabitat remains paramount, and consequently, the conceptual TROX model (Jorissen et al.,
1995) still provides an excellent tool to understand foraminiferal distribution patterns.

877 In this paper, we present a first attempt to translate the conceptual TROX-model into a numerical
878 model, combining sediment early diagenesis (the combination of biological, chemical, and physical
879 processes which change the quantity and composition of the organic matter before they become
880 consolidated; Henrichs, 1992) and foraminiferal ecology.

The aim of such a model is that it will allow us to test hypotheses concerning foraminiferal ecology.
We will define theoretical species, with distinct characteristics, that each represent a group of real
species with a similar microhabitat. We will then see how these theoretical species behave (are
distributed) in the vertical succession of redox zones reproduced by the model, and compare the
model results with observational data. If the model is capable of successfully reconstructing the field
observations, this means that the underlying hypotheses concerning the foraminiferal ecology may

887 be realistic. If this is not possible, the way the foraminiferal species are defined is probably incorrect.

- Ultimately, the aim is to use this model as a tool to test paleoceanographical scenarios and toreconstruct environmental parameters from foraminiferal successions in fossil records.
- 890 For the construction of the model, we profit from an extremely favourable context. First, early

diagenetic models have become widely available. We decided to use the OMEXDIA model of Soetaert

892 et al. (1996), which is well established, very well documented (Soetaert and Hermann, 2009) and

freely available on the Comprehensive R Archive Network (https://cran.r-project.org). The OMEXDIA model will be coupled to a new foraminiferal model developed in the present study. Next, we employ

a very large mixed database of sediment geochemistry (Anschutz et al., 1999, 2005; Hyacinthe et al.,

896 2001, Chaillou et al., 2002, 2003, 2006; Mouret et al., 2009, 2010; Schmidt et al., 2009) and

897 foraminiferal distribution (Fontanier et al., 2002, 2003, 2005, 2006a; Langezaal et al., 2006; Mojtahid

et al., 2010) for the Bay of Biscay. We decided to use the data collected between 1997 and 2001,

899 when a bathymetrical transect from 150 to 2000 m water depth was sampled 10 times. This data set

900 gives ample insight into seasonal and interannual variability of sediment geochemistry and

foraminiferal communities. We will use this very large dataset to calibrate the model, and to test it in
 two different ecological settings.

903 This paper presents the first results of this exercise, in which the numerical model is developed, and

is manually tested and tuned at two sampling stations, at 1000 and 550 m depth in the Bay of Biscay.
 In the process of developing the model, we sequentially tested a range of hypotheses regarding the

In the process of developing the model, we sequentially tested a range of hypotheses regarding thebenthic environment and its link to foraminiferal ecology. To illustrate the value of quantitative

907 models for such hypothesis testing, we present this study in a manner that emphasizes the

908 sequential steps in model development and calibration, highlighting the lessons learned along the

909 way.

910 2. Model characteristics

- 911 When we set out to develop the coupled early diagenesis benthic foraminiferal ecology model
- 912 presented here, we chose to implement the model in the software R (R Core Team, 2020). R is widely
- 913 used for scientific programming, and is freely available (https://www.r-project.org). We decided to
- 914 use the early diagenetic model (1D) of Soetaert, which has been described in a number of
- 915 publications (Soetaert et al., 1996; Soetaert & Herman, 2009), and is freely available online in the R-
- 916 package 'ReacTran' (Soetaert and Meysman, 2012; http://
- 917 www.rforscience.com/modelling/omexdia/.

918 The next step was to introduce theoretical species of benthic foraminifera, with different ecological 919 characteristics (each theoretical species represents a group of real species), into this 1D geochemical 920 environment. At a first stage, we investigated the possibility of constructing a quantitative biological 921 model, including the most important life functions (e.g., growth, reproduction, mortality, etc.) of the 922 different species. However, it soon turned out that this approach was severely hampered by a lack of

923 observational data for most of these parameters. For this reason, we decided to change our strategy,

- 924 and instead developed a "transformation model", in which a fraction of the available resources was
- 925 allocated to different members of the benthic foraminiferal community. More precisely, we assigned
- 926 different proportions of the various types of organic matter (C_{org}), more or less reactive, to the
- 927 foraminiferal species introduced into the model.

928 We then tested the model on our extensive geochemical and foraminiferal dataset from the Bay of

- Biscay. After constructing the essential components of the model, we started to tune the model
- 930 using the data of stations A and B, at 1000 and 550 m depth, respectively. We first optimized the fit
- 931 between the geochemical data and the model results. Next, we tested various hypothesis concerning
- 932 the ecological requirements of the benthic foraminifera, in order to obtain a maximum resemblance
- 933 between observational data and model output.



934

935 Fig. 1. Map of the bay of Biscay, with sampling stations A (1000 m) and B (550 m).

937 3. The Bay of Biscay data set

938 Bay of Biscay Stations A, at 1000 m water depth, and B, at 550 m depth, were visited 15 times between October 1997 and February 2011. The surficial sediment was sampled with an interface 939 940 multi-corer, which allowed sampling the upper few decimetres of the sediment, the overlying 941 bottom water, and the undisturbed sediment-water interface, in a 10 cm diameter Plexiglas tube. At 942 both stations, very similar profiles of redox reactive compounds were measured in the numerous 943 short cores collected over the 14 year interval (Mouret et al., 2016). Only small differences were 944 noticed at the sediment surface, which were attributed to seasonality, bioturbation and/or spatial 945 patchiness. For the present study we decided to concentrate on the period between October 1997 946 and September 2001, when these stations were sampled 10 times, so that the seasonal and 947 interannual variability of the geochemical composition of bottom and pore waters, as well as of the 948 foraminiferal community, is particularly well constrained. The pore water geochemistry of these 949 station has been described in detail by Hyacinthe et al. (2001), Anschutz et al. (2005), Mouret et al. 950 (2009, 2010) and Charbonnier et al. (2019). Schmidt et al. (2009) concentrated on the particle fluxes and sediment accumulation rate. The foraminiferal community at the two stations was described by 951 952 Fontanier et al. (2002; 2003; 2006a) and Mojtahid et al. (2010). Important seasonal and interannual 953 variability was observed for the foraminiferal density and species composition. It appears that 954 foraminiferal densities increased to an order of magnitude after major spring and autumn 955 phytoplankton blooms, which favoured a limited number of opportunistic benthic foraminiferal taxa 956 (Fontanier et al., 2003; 2006a).

957 In view of the observed stability of the succession of sediment redox zones (Mouret et al., 2009), we 958 decided to use an averaged geochemical data set to calibrate our diagenetic model. To this purpose, 959 for every geochemical model parameter, the observations for all available sampling campaigns (their 960 number varied for different parameters) were averaged, so that a single value was obtained for all 961 depth levels down to 10 cm depth in the sediment. For this first modelling study, foraminiferal data

962 were averaged as well, for the 10 successive sampling campaigns.

963 4. Early Diagenesis Model Parameters

964 The first portion of our model utilizes OMEXDIA, a reactive transport model for evaluating sedimentary diagenesis (Soetaert et al., 1996), which was run to steady state. It includes multiple 965 966 organic matter remineralization pathways, incorporates key diagenetic chemical reactions, and 967 allows for depth-dependent bioturbation/biodiffusion and porosity. In the present study, many of 968 the key parameters of the model are constrained by observations derived from prior studies of 969 stations A and B. Using the dataset of stations A and B, the model was manually tuned to provide 970 initial constraints for the complete model, discussed later. The values of the main parameters used 971 as starting point for the early diagenesis part of the model were the following:

- 972 The organic carbon flux (Corg) to the sea floor: for the 1000 m deep station A, Mouret et al. 973 (2010) estimated a total Corg flux of 16.5 gC m⁻² yr⁻¹, consisting of 76% labile particulate organic carbon (POC), 9% slowly decaying POC, and 15% inert POC. At the 550 m deep 974 station B, Mouret et al. (2010) estimated a total C_{org} flux of 28.2 gC m⁻² yr⁻¹, with 53.6% labile 975 976 POC, 20.4% slowly degrading POC and 26.0% inert POC. These values were used as the basis 977 for our tuning exercise. The most labile fraction was deduced from modelled dissolved 978 oxygen consumption at the sediment surface. Other fractions were deduced from Corg 979 concentrations and mass accumulation rates.
- Bottom water temperature was always 9.5°C at station A, and 11.0°C at station B, without
 any seasonal variability (Ogawa et al., 1973; Durrieu de Madron et al., 1999 and Fontanier et
 al., 2006b).
- Bottom water oxygenation varied between 189 and 201 µmol l⁻¹ at station A and between
 205 and 221 µmol l⁻¹ at station B (Mouret et al., 2010; Anschutz and Chaillou; 2009).
- At both stations A and B, the sediment was a silty clay, with a mean grain size of 5–6 μm at station A and 10-15 μm at station B (Charbonnier et al., 2019). The sediment contained about 30% CaCO₃ at station A, versus 10-15% at station B. At both stations, sediment porosity decreased quickly in the first millimeters of the sediment, from 0.85 at the sediment-water interface to about 0.8 at 1 cm depth. Deeper down, porosity continued to decrease, until values between 0.65 and 0.7 were observed at 10 cm depth at stations A and B, respectively.
 The oxygen penetration depth into the sediment varied from 1.6 to 2.2 cm at station A, and
- 992 from 1.1 to 1.9 cm at station B (Fontanier et al., 2003; 2006a; Mouret et al., 2010).
- 993 At station A, nitrate in the bottom waters, sampled in the supernatant water of the sediment 994 cores, varied from 8.6 to 21.1 μ mol l^{-1} , with a mean value of 14.3 μ mol l^{-1} . At station B, nitrate varied from 11.5 to 15.7 $\mu mol~l^{\text{-1}}$, with a mean value of 14.1 $\mu mol~l^{\text{-1}}$. These values are 995 996 very close to those measured on water column samples taken with Niskin bottles (Anschutz 997 and Chaillou, 2009). Maximum nitrate concentrations were generally observed in the top 2 998 cm of the sediment, with values of 24 to 33 μ mol I⁻¹ at station A, and 19 to 23 μ mol I⁻¹ at 999 station B. Deeper down, nitrate concentrations strongly decreased, with values below 23 1000 µmol I⁻¹ reached between 4 and 5 cm at station A and around 2 cm at station B. Ammonia 1001 always showed a strong downward increase from values around zero at the surface to mean 1002 values of 30.7 µmol I⁻¹ (station A) and 61.1 µmol I⁻¹ (station B) at 10 cm depth.
- Among the reduced components, dissolved Mn²⁺ and Fe²⁺ were only determined for part of the 10 sampling campaigns (Mouret et al., 2009), whereas no data for S(-II) were available.
 The sulphate concentration was constant with depth and equal to the SO₄²⁻ concentration in seawater. No sulphide smell was evident during core processing, but a few black patches were observed near the base of the cores, suggesting H₂S production in deeper sediment layers (Chaillou et al. 2003). Since the H₂S concentration was not measured, the model output for ODU (oxygen demand units, i.e. the amount of oxygen necessary to reoxidize

- 1010reduced counterparts from anaerobic mineralization), which regroups all reduced1011components, was always higher than the observations, which include only measured1012dissolved Mn and Fe.
- At station A, the maximum sediment accumulation rate was estimated on the basis of ²¹⁰Pb profiles at 0.068 cm/year (36 mg cm⁻² yr⁻¹) by Chaillou et al. (2002), at 0.05 to 0.06 cm/year by Schmidt et al. (2009), whereas Mouret et al., 2009 obtained a value of 0.062 cm/year based on steady state manganese diagenesis. For station B, Chaillou et al. (2002) obtained a maximum sedimentation rate of 0.15 cm/year (79.5 mg cm⁻² yr⁻¹), compared to values of 0.12 and 0.22 cm/year by Schmidt et al. (2009), and 0.12 cm/year based on steady state
- Mouret et al. (2009) obtained a large variety in biodiffusion coefficients, using ²³⁴Th profiles, for sediment cores sampled during four different campaigns, with values varying between 0.12 and 6.7 cm²/year at station A, and between 0.4 and 14.8 cm²/year at station B. They estimated a mixed layer depth of about 1cm at both stations, slightly less than the depth of the oxic layer.
- 1025 Despite the lack of observational data for some parameters (such as sulphur) in some of the 1026 cores, and the large variability observed for biodiffusion coefficients, the Bay of Biscay dataset 1027 for these two stations is remarkable by its near completeness for 10 successive sampling 1028 campaigns. It therefore forms an ideal dataset to develop and test our early diagenetic model. 1029 The main parameter that we used to tune the model was the C_{org} flux to the sea floor, or more 1030 precisely the proportions and reactivity constants of the various types of organic matter. A 1031 secondary main tuning parameter was the rate of pyrite formation in the sediment; this process 1032 withdraws reduced components from the pore water.

5. Initial tuning of the preliminary (3G) early diagenetic model, using the station A data set

Before introducing theoretical foraminiferal species into the model, our first objective was to tune

the early diagenetic model to obtain an optimal fit between the averaged observed data and the

model output. For this initial tuning exercise, we used the dataset for station A. Fig. 2 gives an

example of an early result that employed three types of organic matter (3G).



Fig. 2. Station A - Model output for early diagenetic parameters (lines) of a preliminary version of the 4GFOR
model, with only three types of organic matter (percent TOC = particulate C_{org} in weight % of dry sediment; red:
fast degrading, blue: intermediate, black: total organic carbon (= fast degrading + intermediate + fossil),
compared to observational data from Station A (dots).

1066 It can be seen that the model results for Corg, oxygen and ammonia are very close to the observed 1067 data. However, although the modelled ODU curve has the right form, the absolute values are much 1068 higher than the observed ones. As explained in the previous section, this is likely due to the fact that 1069 only Fe and Mn were measured, and not S, a third major component contributing to ODU. As we do 1070 not intend to use ODU as a limiting factor for the foraminiferal species, we do not consider this very 1071 important. The main problem with this preliminary model is the nitrate curve, which in deeper 1072 sediment layers showed values that are too high compared to the observed ones. This is important, 1073 because we intend to use a minimum NO₃⁻ concentration as a switch for nitrate-respiring 1074 foraminifera in the subsequent foraminiferal component of the model. All our attempts to improve 1075 the fit for nitrate systematically resulted in an upward movement of the oxygen profile, which is even 1076 more problematic, because we intend to use oxygen concentration as a switch for obligate aerobic 1077 foraminiferal species. In the final version of the model we resolved this problem by adding an

additional diagenetic pathway, ferrous iron nitrification. This reaction, the anaerobic oxidation of Fe²⁺
by nitrates (e.g., Straub et al., 1996; Kappler et al., 2021), appears to be an important process at our
sites in the Bay of Biscay (Hyacinthe et al., 2001), and was hitherto not included in the OMEXDIA
model. This early diagenetic pathway consumes nitrate at the lower part of its range, and diminishes
ODU. After adding this reaction, to the early diagenetic part of the model, we arrived at excellent fits
for all parameters except ammonia, which was overestimated (Fig. 5). Since ammonia has no impact
on the faunal distribution, this was considered less important.

1085 6. Defining the foraminiferal species

1086 As explained before, we decided not to include a fully biological model for the foraminiferal species, 1087 but rather a transformation model. This transformation model has two basic characteristics. First, foraminiferal activity is limited by critical environmental factors, such as oxygen and nitrate content. 1088 1089 Beyond the critical threshold values, specified foraminifera are no longer given access to the 1090 available resources. Next, when these limiting factors are within the tolerance limits of the organism, 1091 a specific proportion of each type of Corg is attributed to each of the species. In the way we defined 1092 them, species living close to the sediment surface use a large proportion of reactive Corg, whereas 1093 species living deeper in the sediment use a larger proportion of slowly degrading Corg. This interspecific difference is based on two assumptions: 1) that infaunal foraminiferal taxa are better 1094 1095 capable than surface dwellers to feed on organic matter of a lower quality, and 2) that sediment 1096 surface dwellers are more competitive than infaunal taxa when the uptake of high quality (highly 1097 reactive) Corg is concerned.

1098The allocation of different proportions of various qualities of organic carbon to different species is1099reminiscent of the BFAR-concept of Herguera and Berger (1991). In fact, Herguera and Berger1100assumed that for every mg of Corg arriving at the sea floor, roughly one foraminiferal shell > 150 µm

1101 would be preserved in the sediment record.

1102In our first attempts to model the foraminiferal distribution in the sediment (for station A), we used a11033G diagenetic model, with fast, slowly degrading, and inert C_{org} ; a very good fit between model1104output and observed data was obtained with decay rates $r_{fast} = 0.01/day$, $r_{slow} = 0.00005/day$ and r_{fossil} 1105= 0. In these first runs (Fig. 3), we introduced only 2 foraminiferal species:

11061) an epifaunal/shallow infaunal species, which was dependant on the presence of oxygen, at least at1107very low concentrations ($0_2 > 2.5 \ \mu mol \ l^{-1}$). We defined this species group so that it exclusively feeds1108on fast decaying $C_{org.}$

11092) a deep infaunal species, which we inhibited by oxygen ($0_2 \le 2.5 \ \mu mol \ l^{-1}$), but which was dependent1110on the presence of nitrate (NO3 > 2.5 \ \mumol mol l^{-1}), and given access to both fast and slowly degrading1111Corg.

1112 In order to compare the model outcome (in mgC cm⁻³) with the observational data (observed number
 1113 of tests > 150 μm), we assumed that a single foraminiferal individual (> 150 μm) of the

epifaunal/shallow infaunal species would contain 0.625 μg Corg, whereas a deep infaunal individual (>

1115 150 μm) would on average contain 3.6 μg C_{org}. These foraminiferal C_{org} contents are based on the

relationship between biovolume and protein content observed by Movellan et al. (2012), assuming

an average size of 250 µm for epifaunal/shallow infaunal taxa (considered as semi-spheres,

1118 representative of most epifaunal and trochospiral taxa) and of 450 µm for deep infaunal taxa

1119 (considered as a cylinder, such as *Globobulimina* spp.).



Commenté [f1]: Change slow in intermediate ?!

1132 Fig. 3. First results of the coupled 3G early diagenesis (using two types of reactive organic matter) -

1133 foraminiferal model. Left panel: epifaunal/shallow infaunal taxa (black) and deep infaunal taxa (red). Right

1134 panel: fast degrading C_{org} (red) and intermediate degrading C_{org} (blue) Dots = observed data for station A (1000 1135 m depth); lines = model output. Epifaunal/shallow infaunal taxa (black curve) only have access to fast decaying

1136 organic matter.

1137

1138 7. Further developing the (3G) early diagenesis – perfecting the foraminiferal model for 1139 station A

1140 The first results of the coupled early diagenesis-foraminifera model for the 1000 m deep station A

1141 (Fig. 3) were encouraging. To arrive at this fit, we attributed 0.2% of fast decaying C_{org} to

1142epifaunal/shallow infauna (when $O_2 > 2.5 \mu mol l^1$, left panel, black curve), and 0.1% of fast decaying1143 C_{org} and 0.002% of slowly degrading C_{org} to deep infauna (when $O_2 \le 2.5 \mu mol l^1$ and $NO_3 > 2.5 \mu mol l^2$

1144 ¹, left panel, red curve).

1145 These values, which were selected to obtain a reasonable fit, are in reality thought to reflect the

1146 capacity of these species to feed on various types of C_{org} , and their competitive success compared to

1147 other (foraminiferal as well as non foraminiferal) organisms feeding on the same resources. For

comparison, for continental shelf (37-60 m depth) and upper slope (450 m depth) environments with
 well oxygenated bottom waters, Geslin et al. (2011) estimated the total foraminiferal contribution to
 dissolved oxygen utilisation at 0.5 to 2.5%.

1151 However, although the obtained result (Fig. 3) was encouraging, it was not yet fully acceptable.

1152 There were in fact three major problems. The first problem was that the epifaunal/shallow infaunal

1153 taxon did not penetrate deep enough into the sediment; their strong decline in the first cm of the

1154 sediment is the direct consequence of the similar decline of the fast degrading C_{org} (red curve in the

right panel of fig. 3). In our opinion, the model results clearly show, that our initial hypothesis, that
 epifaunal/shallow infaunal taxa exclusively feed on fast degrading C_{org}, is not supported. We solved



1170

1171Fig. 4. First results of the coupled (3G) early diagenesis (using three types of organic matter) - foraminiferal1172model for station A (1000 m depth). Left panel: epifaunal/shallow infaunal taxa (black) and deep infaunal taxa1173(red). Right panel: fast degrading Corg (red) and intermediate degrading Corg (blue). Dots = observed data; lines =1174model output. Epifaunal/shallow infaunal taxa now have also access to a small part of slowly degrading Corg.

The second problem was the fact that in reality, there are more than two groups of foraminiferal 1175 1176 species. There are not only deep infaunal species (such as Globobulimina spp.), but also intermediate 1177 infaunal species, such as Melonis barleeanus, which always attain the highest density at the depth 1178 where nitrate concentration is maximal (e.g., Jorissen et al., 1998). In addition, the group of 1179 epifauna/shallow infauna seems to be more complex, with some taxa always living close to the 1180 sediment-water interface, and others living slightly deeper in the oxygenated part of the surficial 1181 sediment. The first group is dominated by trochospiral genera (Cibicidoides, Gyroidina, Hoeglundina, Oridorsalis, etc.), whereas the second group is rather dominated by triserial and biserial genera 1182

1183 (Bolivina, Bulimina, Trifarina, Uvigerina, etc.).

1184 In order to take into account this complexity, we decided to expand our model to four theoretical

1185 for a miniferal taxa, each of which is supposed to represent a group of real species with similar

1186 microhabitat characteristics. Since we think that the name "epifauna" should be reserved for taxa 1187 living on elevated substrates above the sediment, we will use the terms "shallow infauna A" (SIA),

1187 living on elevated substrates above the sediment, we will use the terms "shallow infauna A" (SIA),
1188 "shallow infauna B" (SIB), "intermediate infauna" (II) and "deep infauna" (DI) for our four theoretical

species (groups) living at increasingly greater depth in the sediment.

In order to model the vertical distribution of these four theoretical species, it soon turned out that a
 rough subdivision of the organic carbon resources into three types (fast degrading, slowly degrading,
 fossil) was insufficient. For this reason, we shifted to the final 4G model, with three types of reactive
 C_{org}: fast, intermediate and slowly degrading, and finally inert C_{org}.

1194 The final problem was that in the first versions of the model (Figs. 3, 4), we inhibited the activity of 1195 the deep infaunal species with oxygen. However, Nomaki et al. (2007) observed oxic respiration for 1196 the deep infaunal taxa *Globobulimina affinis* and *Chilostomella ovoidea*. Consequently, in most

- studies, deep infaunal taxa are considered as facultative anaerobes, which will only shift to ananaerobic metabolism (using nitrate) when no oxygen is available.
- 1199 We solved this problem by replacing the inhibition with an additional scaling factor. In fact, we
- 1200 defined the intermediate and deep infaunal categories in such a way that the amount of fast and
- 1201 intermediate degrading Corg allocated to them decreases exponentially towards higher oxygen
- 1202 concentration. In biological terms, this means that these taxa become rapidly less competitive
- 1203 (compared to shallow infaunal groups A and B) when oxygen concentrations increase. In our opinion,
- 1204 this is a very realistic way to define these species for the sake of our quantitative model.

1205 8. Tuning the final 4GFOR Model for stations A (1000 m) and B (550 m).

- 1206 Concerning the final tuning of the early diagenesis part of the 4GFOR model, an optimal fit between
- 1207 model output and observations was obtained by using total C_{org} flux of 16.0 and 25.0 gC m⁻² yr⁻¹ at
- 1208 stations A and B, respectively (table 1). These values are very close to the values of 16.5 and 28.2 gC
- 1209 m^{-2} yr⁻¹ given by Mouret et al., 2010).
- 1210

Parameter	Description	Station A	Station B	Unit	Source
maxdep	Maximum depth for modeled sediment	24	24	cm	1
pO	Porosity in surface sediment	83.3	88.5	%	1
pinf	Porosity at maximum modeled depth	64.2	69.7	%	1
рсо	Coefficient of decrease for porosity	4.47	2.76		1
sedr	Sedimentation rate	0.055	0.15	cm/year	L ⁶
Db	Bioturbation rate	0.75	1.25	cm/year	L ³
mixl	Mixed layer depth	1.0	1.0	cm	L ³
fluxm	Total Corg flux at sediment water interface	16.0	25.0	g m ⁻² y ⁻¹	L ³ ; C
TCorginf	%Corg at infinite depth	0.79	0.83	%	L ³
rfast	1st order rate constant for fast degrading Corg	0.026	0.015	day ⁻¹	С
rint	1st order rate constant for intermediately degrading Corg	0.001	0.0005	day ⁻¹	С
rslow	1st order rate constant for slowly degrading Corg	0.00003	0.0000025	day ⁻¹	С
pfast	fraction of fast degrading Corg in total Corg flux	58.41	34.0	%	С
pint	fraction of intermediate degrading Corg in total Corg flux	20.0	21.5	%	С
pslow	fraction of slow degrading Corg in total Corg flux	6.26	18.5	%	С
NCrFdat	NC ratio fast decay det	0.16	0.16	molN/molC	L⁵; C
NCrIdat	NC ratio intermediate decay det	0.2	0.2	molN/molC	С
NCrSdat	NC ratio slow decay det	0.13	0.13	molN/molC	L ⁵
Pdepo	Fraction of ODU production deposited as solid	0.44	0.44	% day-1	LS
Temp	Bottom water temperature	9.5	11	°C	L ^{2,4}
bwO2	Oxygen concentration in bottom water	196.5	213.4	mmol m-3	L ³ ; I
bwNO3	Nitrate concentration in bottom water	16	14	mmol m-3	L ¹ ; I
bwNH3	Ammonia concentration in bottom water	0	0.6	mmol m-3	L ¹ ; I
NH3bot	Ammonia conc at bottom modeled sediment column	68.29	200	mmol m-3	I
ksO2oxic	Half-saturation O2 in oxic mineralisation	30	30	mmol m-3	۲
ksNO3denit	Half-saturation NO3 in denitrif	15	15	mmol m-3	L
kinO2denit	Half-saturation O2 inhib denitrif	0.5	0.5	mmol m-3	L ⁵
kinNO3anox	Half-saturation NO3 inhib anoxic min	1	1	mmol m-3	L
kinO2anox	Half-saturation O2 inhib anoxic min	5	5	mmol m-3	L5
ksO2nitri	Half-saturation O2 in nitrification	100	100	mmol m-3	L ^S
ksO2oduox	Half-saturation O2 in oxidation of ODU	100	100	mmol m-3	LS
rODUox	Maximum rate oxidation of ODU	20	20	day ⁻¹	L ⁵
rnit	Maximum nitrification rate	20	20	day ⁻¹	L5
NH3Ads	Adsorption coefficient ammonium	1.3	1.3		L ⁵
roxidNO3	Maximum oxidation of ODU by NO3	200	200	day ⁻¹	LS
ksNO3ODUox	Half-saturation NO3 in Fe oxydation	200	200	mmol m-3	L ^S
kinO2ODUox	Half-saturation 02 in Fe oxydation	1	1	mmol m-3	L ⁵

¹²¹¹

1212 Table 1. Parameters used in the final simulation of the early diagenesis part of the 4GFOR model. The source of

1213 the parameter values is indicated as follows: C = constrained with the model using the measured data; I =

independently determined from field data; L = based on literature values, retrieved from: ¹Anschutz and
 Chaillou (2009); ²Durrieu de Madron et al. (1999); ³Mouret et al. (2010); ⁴Ogawa et al. (1973); ⁵Soetaert et al.

 1215
 Chaillou (2009); ² Durrieu de Madron et al. (1999); ³

 1216
 (1996); ⁶ Schmidt et al. (2009).

1217

1218 Compared to Mouret et al. (2010), the fractions of inert C_{org} (0.153 and 0.26) were not changed. Of 1219 the 0.2 and 0.215 parts (stations A and B, respectively) attributed to the newly created category 1220 Corg_{int}, compared to the observations of Mouret et al. (2010), the large majority (87.5 and 91.2% for 1221 to the newly created to the observations of Mouret et al. (2010), the large majority (87.5 and 91.2% for

1221 stations A and B) was taken from the labile $C_{org,}$ and the remaining part from the slowly decaying C_{org} .

1222 The presence of the four foraminiferal species was constrained with the boundary conditions listed in 1223 Table 2, which determined whether they were present or not. In our first attempts, using the data of

1224 station A, for deep infauna, we used a threshold value for nitrate of 2.5 μ mol l⁻¹. It turned out that at

1225 station B, where the nitrate concentration diminishes rapidly with greater depth in the sediment, this

1220			
1229		Oxygen	Nitrate
1230	Shallow infauna A	>2.5 µmol l ⁻¹	No nitrate limitation
	Shallow infauna B	>2.5 µmol l ⁻¹	No nitrate limitation
1231	Intermediate infauna	No oxygen limitation	>2.5 µmol l ⁻¹
1232	Deep infauna	No oxygen limitation	>0.05 µmol l ⁻¹

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1234 Table 2. Threshold values of oxygen and nitrate concentrations for the four theoretical species of benthic1235 foraminifera.

Finally, foraminiferal biomass was tuned to the observational data by changing the fractions of the
 different types of C_{org} attributed to the four theoretical species of foraminifera, as shown in Table 3.

Station A – 1000 m	Corg _{fast}	Corgint	Corg _{slow}
Shallow infauna A	0.0006	0.0006	0
Shallow infauna B	0.0003	0.0012	0
Intermediate infauna	0.0002	0.0004	0.00001
Deep infauna	0.0001	0.0001	0.00015

threshold truncated the downward distribution of deep infauna. In order to better simulate our field

observations, we subsequently decreased the nitrate threshold for deep infauna to 0.05 μ mol l⁻¹.

1238

Station B – 550 m	Corg _{fast}	Corgint	Corg _{slow}
Shallow infauna A	0.0006	0.0006	0
Shallow infauna B	0.0003	0.0012	0
Intermediate infauna	0.004	0.00033	0.00001
Deep infauna	0.0001	0.00001	0.00011

1239

1240 Table 3. Fractions of the three types of reactive organic carbon attributed to each of the 4 theoretical species of 1241 foraminifera, at stations A and B.

1242The competitiveness of intermediate infauna/deep infauna with respect to highly and intermediately1243reactive Corg further depends on oxygenation: it is low/very low under oxic conditions, and high/very

1244 high under hypoxic/anoxic conditions.

1245 The final results of the tuning procedure of the 4GFOR model are shown in Fig. 5 and 6. As noted

1246 earlier, the fit between the observed and modelled profiles of early diagenetic parameters is very

1247 satisfactory. The two shallow infaunal species (A and B) are well separated in the top 1-2 cm of the

1248 sediment, suggesting that our decision to differentiate them by their ability to cope with low quality

1249 C_{org} may be realistic. The profiles of intermediate and deep infaunal taxa correspond well with the
 1250 observed data, although in detail, the modelled distribution is slightly off for some levels.

1251



Fig. 5. Final results for station A (1000 m depth) of the 4GFOR model with four types of organic matter (red:
 fast degrading, green: intermediate, blue: slow, and black: total TOC). Foraminiferal species distribution is
 represented as: Shallow Infauna A = black, Shallow Infauna B = orange, Intermediate Infauna = blue, Deep
 Infauna = green. All dots are averaged observed data, lines are model output.



Fig. 6. Final results for station B (550 m depth) of the 4GFOR model with four types of organic matter (red: fast
 degrading, green: intermediate, blue: slow, and black: total TOC). Foraminiferal species distribution is
 represented as: Shallow Infauna A = black, Shallow Infauna B = orange, Intermediate Infauna = blue, Deep
 Infauna = green. All dots are averaged observed data, lines are model output.

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1265 9. Discussion

1266 <u>9.1. Definition of organic matter</u>

The organic matter flux to the seafloor is composed of a large variety of particules, such as living 1267 1268 microbial cells and secondary producers, detrital material, fecal pellets, other aggregates, and continental organic matter, as well as dissolved organic carbon (Middelburg, 1998). Consequently, 1269 1270 particulate organic matter has a wide range of components, such as inert carbon, lipids, amino acids, 1271 vitamins and carbohydrates. These components have very different reactivities (Arndt et al., 2013). In 1272 models this range of organic matter can be represented in different ways, by two (2G), three (3G) or 1273 four (4G) groups with different reactivity, or, in the most sophisticated models, by a continuous 1274 range of organic matter quality. In the first version of our model, we used a 3G approach, with inert,

reactive and less reactive Corg types, whereas in the last (4G) version we added an intermediatecategory, which allowed us to obtain much better simulations of the observational data.

1277

1278 <u>9.2. Characteristics of foraminiferal species</u>

While developing the 4GFOR model, the characteristics of our theoretical foraminiferal species have
 progressively evolved, to become more complex. In our first runs, we attributed only fast degrading
 C_{org} to the shallow infaunal species. It turned out that because of the rapid decrease of C_{org-fast} with
 depth, compared to observational data, our model species did not penetrate deep enough into the
 sediment. This example emphasizes the great importance of theoretical models: apparently the basic
 hypothesis (= shallow infaunal taxa feed exclusively on C_{org-fast}) was wrong, and consequently, the
 model was incapable of simulating the observed data.

1286 We solved this problem by separating the group of shallow infaunal taxa into two species, SIA and 1287 SIB, with SIB being much more tolerant to lower quality C_{org} than SIA. The existence of two shallow 1288 infaunal groups with different dominant test forms (SIA = trochospiral, planispiral; SIB = biserial, 1289 triserial) is in accordance with abundant microhabitat observations in natural ecosystems and is 1290 corroborated by the interspecific differences in the δ^{13} C of their shells (e.g., McCorkle et al., 1990; 1291 Rathburn et al., 1996; Fontanier et al., 2006b). By giving them access to different proportions of fast 1292 degrading and intermediate C_{org} , we could obtain a nearly perfect fit with the observed data. 1293 In our first trials, we used only a single deeper infaunal taxon, and we forced this species to remain a

In our first trials, we used only a single deeper infaunal taxon, and we forced this species to remain at 1294 depth in the sediment by limiting it to oxygen concentrations below 2.5 µmol I⁻¹. In the later versions, 1295 we separated this group into intermediate and deep infaunal taxa, in accordance with the large 1296 volume of natural observations. Both groups were initially defined in such a way that they can only 1297 occur when nitrate concentration is above 2.5 µmol l⁻¹. In order to obtain a deeper penetration of deep infauna at station B, we had to lower this threshold to 0.05 μ mol l^{-1} for this group. We further 1298 1299 differentiated intermediate and deep infauna by making DI more tolerant to low quality Corg than II. To avoid that these species invade the superficial sediment layers, where large amounts of highly 1300 reactive Corg are present, we added a competitive element, by attributing less Corg-fast and Corg-int to 1301 1302 these taxa when oxygen rises above 2.5 μmol l⁻¹, with an exponential decrease towards higher 1303 oxygen levels. With these characteristics, the overall distribution of II and DI is very close to the 1304 observed patterns. This suggests that the way we defined these taxa may be correct, and that their 1305 absence in the superficial sediment layers may indeed be the consequence of a strongly diminished 1306 competitiveness in well oxygenated environments. In more general terms, this suggests that 1307 interspecific competition for food particles may be an important controlling parameter.

Finally, when tuning the model for two stations with very different characteristics, we managed to obtain a good fit for the early diagenesis parameters while using the same reaction constants at both stations. This underlines the robustness of this part of the model. In order to obtain good fits for the foraminiferal data at both stations, we had to attribute slightly different proportions of the different organic matter types to the four theoretical species (Table 3). This suggests that the foraminiferal contribution to organic matter remineralisation may change as a function of the environmental conditions.

1315 9.3. Perspectives

Here, we present the newly developed 4GFOR model, and our tuning with the data of two sampling
stations at 1000 and 550 m depth in the Bay of Biscay, on the basis of averaged data of 10 successive
sampling campaigns. In view of the obtained simulations, which are very close, both for geochemical

as for foraminiferal species, it appears that the complexity of the model is enough to correctly 1319 1320 represent the four main groups of foraminifera. The next step will be to test the model in different 1321 environmental settings. We will accomplish this by modelling 1) the 10 successive sample times 1322 individually, 2) the 9 other stations of the Bay of Biscay transect, between 140 and 4800 m depth, 1323 with a wide range of environmental characteristics, 3) sampling stations from other ocean basins 1324 (Indian Ocean, Cape Blanc upwelling) from which we have comparable data sets. These modelling 1325 exercises will very probably lead to adjustments of the model, and the way the foraminiferal species 1326 are defined. This will oblige us to consider slightly different ecological strategies for the four species, 1327 and thereby, to test many hypotheses we still have regarding their life strategies.

Once the robustness of the 4GFOR model has been fully demonstrated, the next step will be to
develop a formal inversion of the model, so that the main parameters controlling the foraminiferal
faunas, the quantity and quality of the organic flux to the sea floor and the vertical succession of
redox species, can be reconstructed on the basis of the composition of fossil faunas in sediment
cores and outcrops.

1333 **10. Conclusion**

1334The 4GFor model is a combination of an early diagenetic model, based on the OMEXDIA model of1335Soetaert et al. (1996), and a transformation model, which transforms proportions of organic matter1336types with different reaction rates into foraminiferal biomass. Foraminiferal communities are1337simulated with four theoretical deep water species. Their different tolerance levels with respect to1338oxygen and nitrate concentrations decide at what depths they can occur, whereas their density is

- 1339 mainly controlled by organic matter quality.
- 1340

1341The model has been tuned to a large geochemical and foraminiferal database of two stations, at13421000 and 550 depth in the Bay of Biscay. The outcome shows that the early diagenesis part of the1343model correctly reproduces the vertical distribution of redox species in the top 10 cm of the1344sediment. By attributing various proportions of the different types of organic matter to the four1345foraminiferal species, the model adequately reproduces the vertical succession of shallow,1346intermediate and deep infaunal species.

1347 The model further separates shallow infaunal taxa into two groups. Group SIA, which is found close

to the sediment-water interface, corresponds to trochospiral (e.g., *Cibicidoides, Gyroidina*,
 Hoealundina) and planispiral species (e.g., *Cassidulina*). This group is critically sensitive to food

Hoeglundina) and planispiral species (e.g., *Cassidulina*). This group is critically sensitive to food
 quality. Group SIB, which is found slightly deeper in the sediment, corresponds to triserial (e.g.,

Bulimina, Uvigerina) and biserial (e.g., Bolivina) taxa, which are less restrictive with respect to food

1352 quality. The observed distribution of these groups at the two stations in the Bay of Biscay is

- 1353 reproduced by the model. Also the depth in the sediment of intermediate (e.g., Melonis barleeanus)
- and deep infaunal (e.g., *Globobulimina* spp.) species is very well simulated.

1355 The tuning of the model indicates that shallow infaunal taxa (neither SIA nor SIB) do not feed

- exclusively on the most labile C_{org} component. Their vertical distribution can only be simulated by
 giving them access to slightly less reactive C_{org}.
- 1358 One of the problems encountered during the tuning phase was to keep intermediate and deep
- 1359 infaunal taxa restricted to deeper layers. Because of the presence of large quantities of high quality
- 1360 C_{org} close to the sediment surface, this layer is also attractive for those species. To solve this problem,

we diminished their competitive ability exponentially towards higher oxygen levels. This solution
 appears to be realistic, but should be tested in experimental conditions.

- 1363 As is shown by these first examples, the 4GFOR model can be a powerful tool to help understand
- 1364 for aminiferal ecology. In the future, we intend to construct a formal inversion of the model, which
- 1365 will make it possible to produce quantitative estimates of the main controlling faunal parameters
- 1366 (quantity and quality of the organic flux, oxygen and nitrate concentration) on the basis of the
- 1367 composition of fossil foraminiferal assemblages.

1368 11. Acknowledgements

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- 1370 nitrification reactions to the early diagenesis model.

Commenté [f2]: Explain how migration is taken into account

1371 **12. References**

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